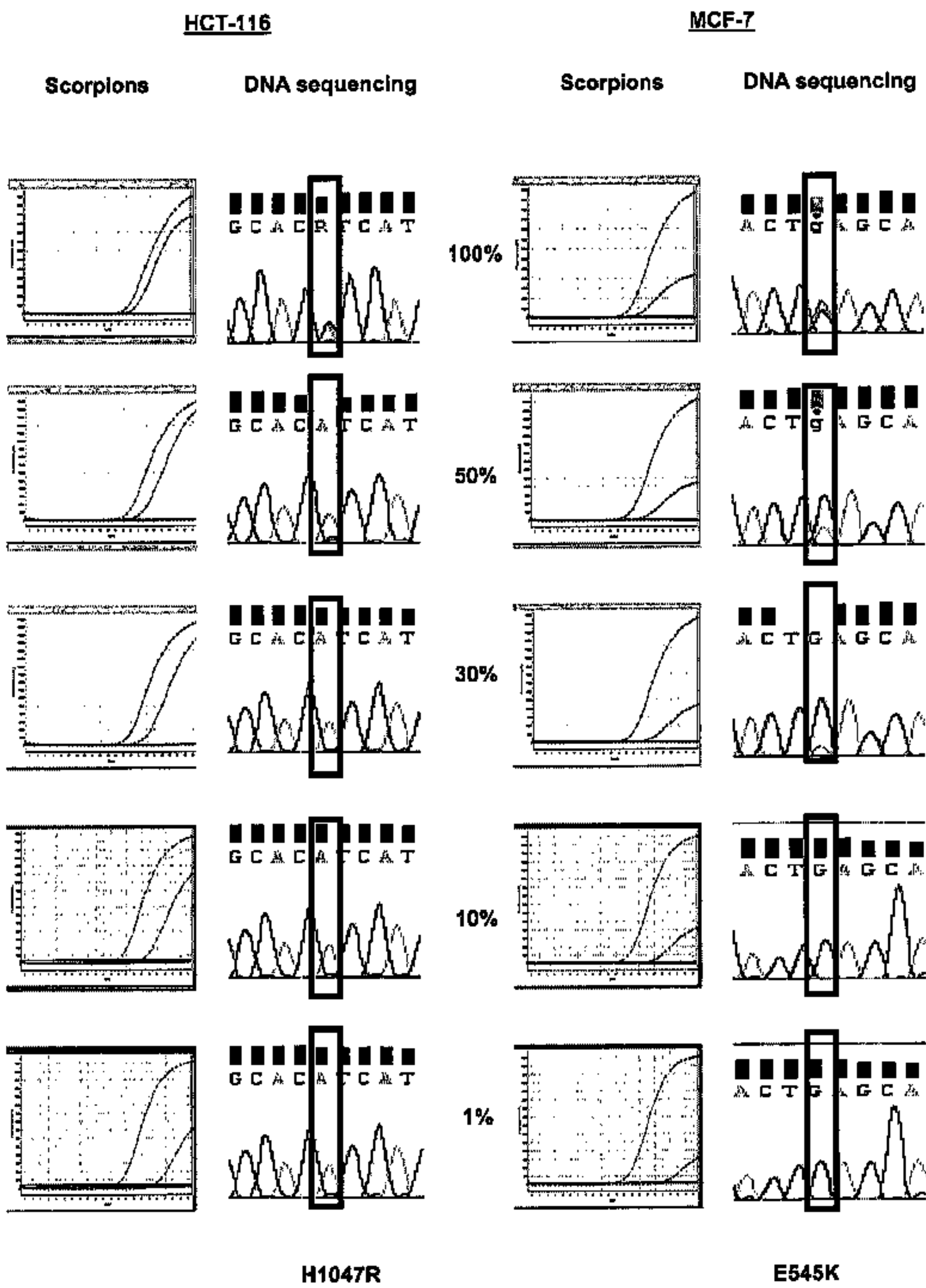




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(54) Title: POLYNUCLEOTIDE PRIMERS FOR DETECTING PIK3CA MUTATIONS



(57) Abrégé/Abstract:
A polynucleotide comprising at least the final six nucleotides of one of the following primer sequences, or a sequence complementary thereto: SEQ. ID NOS. 3 to 16, 18, 20 to 33, 35 or 37 to 39. A method of detecting the presence or absence of a mutation in the PIK3CA gene, wherein the mutation is one of H1047R, H1047L, E542K and E545K, and preferably ARMS primers are combined with Scorpion primers.

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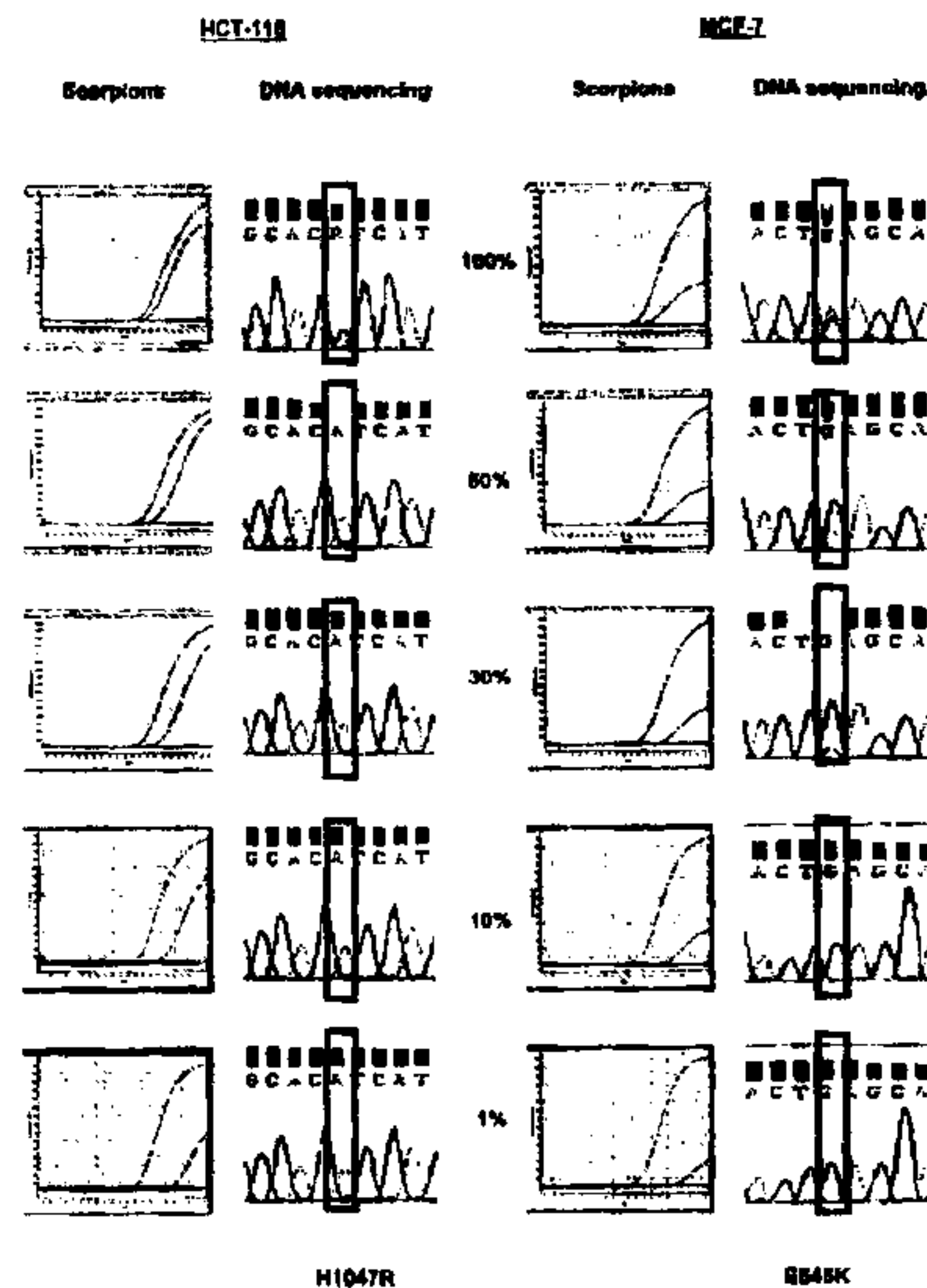


Figure 1

(57) Abstract: A polynucleotide comprising at least the final six nucleotides of one of the following primer sequences, or a sequence complementary thereto: SEQ. ID NOS. 3 to 16, 18, 20 to 33, 35 or 37 to 39. A method of detecting the presence or absence of a mutation in the PIK3CA gene, wherein the mutation is one of H1047R, H1047L, E542K and E545K, and preferably ARMS primers are combined with Scorpion primers.

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1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210 211 212 213 214 215 216 217 218 219 220 221 222 223 224 225 226 227 228 229 230 231 232 233 234 235 236 237 238 239 240 241 242 243 244 245 246 247 248 249 250 251 252 253 254 255 256 257 258 259 260 261 262 263 264 265 266 267 268 269 270 271 272 273 274 275 276 277 278 279 280 281 282 283 284 285 286 287 288 289 290 291 292 293 294 295 296 297 298 299 300 301 302 303 304 305 306 307 308 309 310 311 312 313 314 315 316 317 318 319 320 321 322 323 324 325 326 327 328 329 330 331 332 333 334 335 336 337 338 339 340 341 342 343 344 345 346 347 348 349 350 351 352 353 354 355 356 357 358 359 360 361 362 363 364 365 366 367 368 369 370 371 372 373 374 375 376 377 378 379 380 381 382 383 384 385 386 387 388 389 390 391 392 393 394 395 396 397 398 399 400 401 402 403 404 405 406 407 408 409 410 411 412 413 414 415 416 417 418 419 420 421 422 423 424 425 426 427 428 429 430 431 432 433 434 435 436 437 438 439 440 441 442 443 444 445 446 447 448 449 450 451 452 453 454 455 456 457 458 459 460 461 462 463 464 465 466 467 468 469 470 471 472 473 474 475 476 477 478 479 480 481 482 483 484 485 486 487 488 489 490 491 492 493 494 495 496 497 498 499 500 501 502 503 504 505 506 507 508 509 510 511 512 513 514 515 516 517 518 519 520 521 522 523 524 525 526 527 528 529 530 531 532 533 534 535 536 537 538 539 540 541 542 543 544 545 546 547 548 549 550 551 552 553 554 555 556 557 558 559 560 561 562 563 564 565 566 567 568 569 570 571 572 573 574 575 576 577 578 579 580 581 582 583 584 585 586 587 588 589 590 591 592 593 594 595 596 597 598 599 600 601 602 603 604 605 606 607 608 609 610 611 612 613 614 615 616 617 618 619 620 621 622 623 624 625 626 627 628 629 630 631 632 633 634 635 636 637 638 639 640 641 642 643 644 645 646 647 648 649 650 651 652 653 654 655 656 657 658 659 660 661 662 663 664 665 666 667 668 669 670 671 672 673 674 675 676 677 678 679 680 681 682 683 684 685 686 687 688 689 690 691 692 693 694 695 696 697 698 699 700 701 702 703 704 705 706 707 708 709 710 711 712 713 714 715 716 717 718 719 720 721 722 723 724 725 726 727 728 729 730 731 732 733 734 735 736 737 738 739 740 741 742 743 744 745 746 747 748 749 750 751 752 753 754 755 756 757 758 759 760 761 762 763 764 765 766 767 768 769 770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 787 788 789 790 791 792 793 794 795 796 797 798 799 800 801 802 803 804 805 806 807 808 809 810 811 812 813 814 815 816 817 818 819 820 821 822 823 824 825 826 827 828 829 830 831 832 833 834 835 836 837 838 839 840 841 842 843 844 845 846 847 848 849 850 851 852 853 854 855 856 857 858 859 860 861 862 863 864 865 866 867 868 869 870 871 872 873 874 875 876 877 878 879 880 881 882 883 884 885 886 887 888 889 890 891 892 893 894 895 896 897 898 899 900 901 902 903 904 905 906 907 908 909 910 911 912 913 914 915 916 917 918 919 920 921 922 923 924 925 926 927 928 929 930 931 932 933 934 935 936 937 938 939 940 941 942 943 944 945 946 947 948 949 950 951 952 953 954 955 956 957 958 959 960 961 962 963 964 965 966 967 968 969 970 971 972 973 974 975 976 977 978 979 980 981 982 983 984 985 986 987 988 989 990 991 992 993 994 995 996 997 998 999 1000 1001 1002 1003 1004 1005 1006 1007 1008 1009 1010 1011 1012 1013 1014 1015 1016 1017 1018 1019 1020 1021 1022 1023 1024 1025 1026 1027 1028 1029 1030 1031 1032 1033 1034 1035 1036 1037 1038 1039 1040 1

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POLYNUCLEOTIDE PRIMERS FOR DETECTING PIK3CA MUTATIONS

Technical Field

The present invention relates to a polynucleotide, a kit comprising a polynucleotide
5 and a method for detecting the presence or absence of mutations in a gene.

Background Art

Phosphatidylinositol 3-Kinases (PI3K) are a large family of lipid kinases involved in
cell signalling. The PI3K-AKT pathway is activated in a number of tumour types,
10 resulting in abnormalities of cell growth, proliferation and survival (add ref of 1 recent
review). Recently, mutations in the catalytic subunit of the class 1A PI3K (PIK3CA)
have been identified in human cancers⁽¹⁾. The precise role of these mutations in
carcinogenesis is still to be clearly defined but with ongoing development of a
number of targeted PI3K inhibitors, detection of mutations will become increasingly
15 important for patient selection. Technical challenges in the detection of such
mutations result from the limitations of tumour biopsies that may only contain small
quantities of the mutated sequences. Furthermore, DNA extracted from paraffin
embedded tissue is often degraded and of poor quality. The minimum level of mutant
DNA required for detection by sequencing is 15-25% and so there is a pressing need
20 for development of sensitive assays able to detect small amounts of mutated alleles
in a heterogenous sample and the products necessary for carrying out the assays.

The present invention seeks to address this need.

Disclosure of the Invention

25 The present invention provides sensitive and robust tests for tumour-borne PIK3CA
mutations.

According to one aspect of the present invention, there is provided a polynucleotide comprising at least the final six nucleotides of one of the following primer sequences, or a sequence complementary thereto: SEQ. ID NOS. 3 to 16, 18, 20 to 33, 35 or 37 to 39. That is, the polynucleotide comprises at least the six nucleotides at the 3' end
5 of one of the following primer sequences, or a sequence complementary thereto: SEQ. ID NOS. 3 to 16, 18, 20 to 33, 35 or 37 to 39.

Preferably, the polynucleotide comprises at least 75% of the 8, 10, 12, 14, 16, 17, 18 or 20 nucleotides at the 3' end, or the entirety, of one of the following primer sequences, a sequence complementary thereto, or a sequence having 80%, 90%,
10 95% or 99% sequence identity thereto: SEQ. ID NOS. 3 to 16, 18, 20 to 33, 35 or 37 to 39.

In some embodiments of the present invention there is provided a polynucleotide comprising at least 75% of the ten nucleotides at the 3' end of one of the following primer sequences, or a sequence complementary thereto: SEQ. ID NOS. 3 to 16,
15 18, 20 to 33, 35 or 37 to 39.

Conveniently, the polynucleotide is less than 100 nucleotides long, preferably less than 80 nucleotides long, more preferably less than 60 nucleotides long, more preferably less than 40 nucleotides, more preferably less than 30 nucleotides long.

Advantageously, the polynucleotide further comprises a quencher group and a
20 fluorophore group.

Conveniently, the quencher group and the fluorophore group are separated by a nucleotide tail sequence comprising first and second regions, the nucleotides of the first region being complementary to but in reverse order from the nucleotides of the second region, such that hybridisation of the first region to the second group results
25 in the quencher group to be sufficiently close to the fluorophore group to quench the fluorophore group.

Preferably the tail sequence further comprises a third region having a sequence complementary to a region of the PIK3CA gene.

Advantageously, the polynucleotide comprises at least the six nucleotides at the 3' end of SEQ. ID NO. 18 and the tail sequence comprises SEQ. ID NO. 17.

Alternatively, the polynucleotide comprises at least the final nucleotides at the 3' end of SEQ. ID NO. 35 and the tail sequence comprises SEQ. ID NO. 34.

- 5 Alternatively, the polynucleotide comprises at least the final nucleotides at the 3' end of SEQ. ID NO. 39 and the tail sequence comprises SEQ. ID NO. 38.

Conveniently, the quencher group comprises Dabcyl.

Preferably the fluorophore comprises Hex, Fam or Rox.

- 10 According to another aspect of the present invention, there is provided a kit comprising at least two of the polynucleotides of the invention.

- Advantageously, the kit comprises a polynucleotide comprising SEQ ID NO. 18 and a polynucleotide comprising any one of SEQ ID NOS. 3 to 16; or a polynucleotide comprising SEQ ID NO. 35 and a polynucleotide comprising any one of SEQ ID NOS. 20 to 33; or a polynucleotide comprising SEQ ID NO. 39 and a polynucleotide comprising SEQ ID NO. 37.
- 15

Conveniently, the kit further comprises nucleotide triphosphates, a polymerisation enzyme and/or a buffer solution.

- According to a further aspect of the present invention, there is provided the use of a polynucleotide or a kit of the invention; or a polynucleotide comprising four or five of the six nucleotides at the 3' end of SEQ. ID NOS. 3 to 16, 18, 20 to 33 or 35 or sequences complementary thereto for detecting a mutation in a nucleic acid sample containing at least a fragment of the PIK3CA gene.
- 20

- Advantageously, the fragment of the PIK3CA gene in the nucleic acid sample is at least 10 nucleotides long, preferably 20 nucleotides long, more preferably 30 nucleotides long and more preferably 40 nucleotides long.
- 25

According to another aspect of the present invention, there is provided a method of detecting the presence or absence of a mutation in the PIK3CA gene comprising the steps of:

a) mixing a nucleic acid sample comprising at least a fragment of the
5 PIK3CA gene with a polynucleotide comprising at least the six nucleotides at the 3' end of one of the following primer sequences, or a sequence complementary thereto: SEQ ID NOS. 3 to 16 or 20 to 33 ; and

b) detecting hybridisation of the polynucleotide to the nucleic acid sample wherein hybridisation indicates the presence of a mutation.

10 Conveniently, the polynucleotide comprises one of the following primer sequences: SEQ ID NOS. 3 to 16 or 20 to 33.

Preferably, the method further comprises the step of, prior to step a), amplifying the number of copies of the fragment of the PIK3CA gene using thermal cycling nucleic acid amplification, preferably PCR.

15 Advantageously, step b) comprises carrying out DNA polymerisation using the polynucleotide as a first primer and detecting the extension product of polymerisation.

Conveniently, step b) comprises the step of mixing the nucleic acid sample and the polynucleotide with a second primer which corresponds to a region of the fragment of
20 the PIK3CA sequence downstream of the region to which the polynucleotide is complementary and carrying out PCR on the mixture.

Preferably, the second primer comprises: SEQ. ID NO. 18 and the polynucleotide comprises at least four or five of the six nucleotides at the 3' end of SEQ. ID NOS. 3 to 16; or the second primer comprises SEQ. ID NO. 35 and the polynucleotide
25 comprises at least four or five of the six nucleotides at the 3' end of SEQ. ID NOS. 20 to 33.

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Alternatively, the method further comprises the step of carrying out PCR on the sample using control primers and comparing the amplification of the PIK3CA gene with amplification using the polynucleotide and the second primer.

5 Advantageously, the control primers comprise SEQ ID NOS. 37 and 39.

Conveniently, the polynucleotide comprise a quencher group and a fluorophore group and wherein step b) comprises exposing the mixture to light of a wavelength to which the fluorophore is responsive in the absence of the quencher group and
10 detecting light at the wavelength emitted by the fluorophore group in the absence of the quencher group.

It is preferred that the PIK3CA gene is the sequence available as GenBank accession no. NM_006218 version no. NM_006218.2 GI:54792081.

15 Where reference is made in the specification to "at least four or five of the six nucleotides at the 3' end" of a reference sequence, this means that, of the six nucleotides in the reference sequence, either one or two of the nucleotides may be missing or replaced with a different nucleotide. Of course, in some embodiments, the sequence comprises all six of the nucleotides of the reference sequence.

20

In this specification, "ARMS" is the amplification refractory mutation system disclosed in, for example, EP-A-0332435.

Where reference in this specification is made to a percentage of a polynucleotide
25 compared with a reference polynucleotide, this can be determined by algorithms known in the art.

For example the percentage identity between two sequences can be determined using the BLASTP algorithm version 2.2.2 (Altschul, Stephen F., Thomas L.
30 Madden, Alejandro A. Schäffer, Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997), "Gapped BLAST and PSI-BLAST: a new generation of

protein database search programs", Nucleic Acids Res. 25:3389-3402) using default parameters.

The present invention as claimed relates to:

- a polynucleotide with a sequence consisting of SEQ ID NO: 5;
- 5 - a PCR method of detecting the presence or absence of mutation E542K in the PIK3CA gene comprising the steps of: a) mixing a nucleic acid sample comprising at least a fragment of the PIK3CA gene with a polynucleotide primer having a sequence consisting of SEQ ID NO: 5 wherein the polynucleotide is used as a first primer and a second primer corresponds to a region of the fragment of the PIK3CA sequence
10 downstream of the region to which the polynucleotide is complementary and carrying out a PCR on the mixture; and b) detecting hybridisation of the polynucleotide to the nucleic acid sample wherein hybridisation indicates the presence of the mutation E542K; and
- a kit for detecting PIK3CA mutation E542K, the kit comprising a polynucleotide
15 consisting of SEQ ID NO: 5 and a polynucleotide consisting of SED ID NO: 18.

Brief Description of Drawings

Figure 1 shows the results of carrying out Scorpions detection and sequencing on samples containing mutant PIK3CA gene.

Detailed Description

Embodiments of the present invention provide polynucleotide primers that can be used in assays for the detection of mutations of the PIK3CA gene in a sample containing nucleic acids.

In specific embodiments, the polynucleotide primers are forward and reverse primers that hybridise with the PIK3CA gene to enable a PCR amplification reaction to take place. Thus the forward primer hybridises upstream of and to the opposite strand from the reverse primer and the forward and reverse primers together define an amplicon sequence which is amplified during PCR. The sequence of the forward primer is selected such that it is not complementary to the wild type sequence but is capable of hybridising with a mutant PIK3CA sequence.

In order to detect the presence the mutant PIK3CA gene in the sample, the primers are mixed with the sample. The necessary agents for PCR (appropriate nucleotide triphosphates, DNA polymerase enzyme and a buffer solution) are then added to the sample and PCR is carried out. If the sample contains the mutant sequence to which the forward primer is able to hybridise then the amplicon is amplified during PCR and the presence of the mutant sequence in the sample is thus indicated. If the sample does not contain the mutant sequence then the forward primer binds to the PIK3CA sequence with low efficiency and so there is little or no amplification of the amplicon sequence.

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In order to detect the mutation E542K, the forward primer sequence may be one of SEQ ID NOS. 3 to 9, preferably SEQ ID NO. 5. In order to detect the mutation E545K, the forward primer sequence may be one of SEQ ID NOS. 10 to 16, preferably SEQ ID NO. 14. In order to detect the mutation H1047R, the forward primer sequence may be one of SEQ ID NOS. 20 to 26, preferably 21. In order to detect the mutation H1047L, the forward primer sequence may be one of SEQ ID NOS. 27 to 33, preferably 28. However, it is to be appreciated that the precise sequence of the forward primer need not be identical to these sequences, provided that the forward primer hybridises to the mutant sequence more readily than to the wild type sequence. In the sequences set out above, it is the final six nucleotides (i.e. the nucleotides at the 3' end) of the primers that provide the binding specificity so these nucleotides must be identical to the given sequence.

In order to detect the presence of the amplicons formed in the sample, the reverse primer is a so called "Scorpions" primer in embodiments of the present invention.

Details of Scorpions primers are provided in WO-A-99/066071. A Scorpions primer comprises a primer sequence complementary to a first target sequence of a gene (in this invention PIK3CA) and a tail sequence comprises a probe sequence flanked by two mutually complementary sequences. A DNA polymerase blocking moiety (such as a hexethylene glycol (HEG) monomer) is provided between the primer sequence and the tail sequence. A fluorophore group is provided at one end of the tail sequence and a quencher group is provided at the other end of the tail sequence. In use, the primer sequence of the Scorpions primer acts as a reverse primer during PCR in the normal way and thus the entire Scorpions primer, including the tail sequence, becomes incorporated into each amplicon. The DNA polymerase blocking moiety prevents duplication of the tail sequence. Thus the mutually complementary sequences in the tail sequence have the tendency to hybridise with each other, bringing the fluorophore group and the quencher group into proximity and preventing emission from the fluorophore group. However, if the amplicon contains a second target sequence complementary to the probe sequence, the probe sequence preferentially binds to the second target sequence, separating the mutually complementary sequences. This results in the fluorophore group and the quencher group being spatially distanced such that the

fluorophore group emits light of one wavelength in response to incident light of another wavelength. Accordingly, the Scorpions primer enables easy detection of amplicons and moreover, avoids false positive results (caused by primer dimers, for example) because a signal is only generated when the amplicon contains the second
5 target sequence.

The fluorophore group may be Hex (4,7,2',4',5',7'-hexachloro-(3',6'-dipivaloylfluoresceinyl)-6-carboxamidoethyl]-1-O-(2-cyanoethyl)-(N,N-diisopropyl)-phosphoramidite), Fam ([[(3',6'-dipivaloylfluoresceinyl)-6-carboxamidoethyl]-1-O-(2-cyanoethyl)-(N,N-diisopropyl)-phosphoramidite) or Rox (5,6,-Carboxy-X-Rhodamine).
10 The quencher group may be Dabcyl (5'-Dimethoxytrityloxy-5-[(N-4'-carboxy-4-(dimethylamino)-azobenzene)-aminohexyl-3-acrylimido]-2'-deoxyUridine-3'-[(2-cyanoethyl)-(N,N-diisopropyl)]-phosphoramidite).

In embodiments of the present invention, a Scorpions primer is provided for detection of the E542K and E545K mutations wherein the primer sequence is SEQ ID NO. 18
15 and the probe sequence is SEQ ID NO. 17. A Scorpions primer is provided for detection of the H1047R and the H1047L mutations wherein the primer sequence is SEQ ID NO. 35 and the probe sequence is SEQ ID NO. 34.

It is to be appreciated, however, that the use of Scorpions primers is not essential to the invention and other methods of detecting the synthesis of amplicons may be
20 employed such as TaqManTM product detection, as described in patent numbers US-A-5487972 and US-A-5210015

In some embodiments, a control assay is also carried out to detect the overall concentration of the PIK3CA gene in the sample. This is achieved by carrying out a separate PCR reaction with control forward and reverse primers which define an
25 amplicon in another region of the PIK3CA gene. It is preferred that the forward primer is SEQ ID NO. 37 and the reverse primer is a Scorpions primer wherein the primer sequence is SEQ ID NO. 39 and the probe sequence is SEQ ID NO. 38. The number of PCR cycles required to generate a threshold number of control amplicons is then compared with the number of PCR cycles required to generate the threshold number
30 of amplicons containing the mutant sequence in order to assess the proportion of

mutant copies of the PIK3CA gene in the sample. Such control assays are generally carried out separately from the test assays.

The PCR assays are preferably carried out as multiplexed real time PCR assays.

5 The test sample of nucleic acid is conveniently a sample of blood, faeces, sputum, colonic lavage, bronchial lavage or other body fluid, or tissue obtained from an individual. The individual is conveniently human, preferably Homo sapiens. It will be appreciated that the test sample may equally be a nucleic acid sequence corresponding to the sequence in the test sample. That is to say that all or a part of the region in the sample nucleic acid may firstly be amplified using any convenient
10 technique such as thermal cycling nucleic acid amplification, in particular PCR, or whole genome amplification (WGA) before use in the method of the invention.

Any convenient enzyme for polymerisation may be used provided that it does not affect the ability of the DNA polymerase to discriminate between normal and mutant template sequences to any significant extent. Examples of convenient enzymes
15 include thermostable enzymes which have no significant 3'-5' exonuclease activity, for example Taq DNA polymerase, particularly "Ampli Taq Gold"TM DNA polymerase (PE Applied Biosystems), Stoffel fragment, or other appropriately N-terminal deleted modifications of Taq or Tth (Thermus thermophilus) DNA polymerases.

In further embodiments of the present invention, there are provided kits comprising
20 one or more polynucleotides of the invention and the nucleotide triphosphates, DNA polymerase enzyme and buffer solution required to carry out a PCR reaction. Preferred kits comprise forward and reverse primers for detection of a specific mutation and forward and reverse control primers.

25 EXAMPLES

Materials and Methods

Primers were designed against the 4 most common mutations in the PIK3CA gene (Accession Number: NM_006218). ARMS primers were designed to detect 2

mutations in exon 20: H1047R and H1047L; and 2 mutations in exon 9: E452K and E454K. A control primer was designed to cDNA position 2450 in the PIK3CA gene.

Scorpions were also designed. To allow multiplexing of a number of assays in each reaction the three scorpion primers were labelled with different fluorophores.

5 Primer designs

A number of ARMS primers were designed specific for each target region. The target region for the E542K and E545K mutations are shown below as SEQ ID NOS. 1 and 2 respectively (the mutant bases are shown in brackets with the normal variant first). The forward primers to the mutations are also shown below (SEQ ID NOS. 3 to 16).

- 10 To enhance the specificity of these reactions, additional primer mismatches close to the 3'-terminus were used (shown underlined in the primer sequences). The optimal primers (E542K-2 and E545K-4) were used for the experiments described. The Scorpions primer usable with the primer sequences is shown as SEQ ID NOS. 17 and 18. Regions of correspondence between the Scorpions primer and the target
- 15 regions are shown in identical highlighting or underlining.

Exon 9 Region

20 AACAGAGAATCTCCATTTTAGCACTTACCTGTGACTCCATAGAAAATCTTTCTCC
TGCTCAGTGATTT(C/T)AGAGAGAGGATCTCGTGTAGAAATTGCTTTGAGCTGTT
CTTTGTCATTTTCCCTTAATTCAATTGTCTCTAGCTAGTCTGTTACTCTGTAAAATA
AAATAATATCTTATATA (SEQ ID NO. 1)

25 AACAGAGAATCTCCATTTTAGCACTTACCTGTGACTCCATAGAAAATCTTTCTCC
TGCT(C/T)AGTGATTTCAAGAGAGAGGATCTCGTGTAGAAATTGCTTTGAGCTGTT
CTTTGTCATTTTCCCTTAATTCAATTGTCTCTAGCTAGTCTGTTACTCTGTAAAATA
AAATAATATCTTATATA (SEQ ID NO. 2)

Mutation	Primer Sequence	SEQ ID NO.
E542K-0	5'-CTTTCTCCTGCTCAGTGATT <u>T</u> -3'	3
E542K-1	5'-CTTTCTCCTGCTCAGTGATT <u>A</u> -3'	4
E542K-2	5'-CTTTCTCCTGCTCAGTGATT <u>C</u> -3'	5
E542K-3	5'-CTTTCTCCTGCTCAGTGATT <u>G</u> -3'	6
E542K-4	5'-CTTTCTCCTGCTCAGTGAT <u>A</u> -3'	7
E542K-5	5'-CTTTCTCCTGCTCAGTGAT <u>C</u> -3'	8
E542K-6	5'-CTTTCTCCTGCTCAGTGAT <u>G</u> -3'	9
E545K-0	5'-ACTCCATAGAAAATCTTTCTCCTGCT <u>T</u> -3'	10
E545K-1	5'-ACTCCATAGAAAATCTTTCTCCTGCAT <u>T</u> -3'	11
E545K-2	5'-ACTCCATAGAAAATCTTTCTCCTGCC <u>T</u> -3'	12
E545K-3	5'-ACTCCATAGAAAATCTTTCTCCTGCGT-3'	13
E545K-4	5'-ACTCCATAGAAAATCTTTCTCCTGAT <u>T</u> -3'	14
E545K-5	5'-ACTCCATAGAAAATCTTTCTCCTGGT <u>T</u> -3'	15
E545K-6	5'-ACTCCATAGAAAATCTTTCTCCTGTT <u>T</u> -3'	16
Exon 9 scorpion	Hex-CGCGCTCGTGTAGAAATTGCTTTGAGCGCG- que-heg-CAATGAATTAAAGGGAAATGACA	17 and 18

Exon 20 region

- 5 The target region for the H1047R and H1047L mutations are shown below as SEQ ID NO. 19 (the mutant bases are shown in brackets with the normal variant first). The forward primers to the mutations are also shown below (SEQ ID NOS. 20 to 33). To enhance the specificity of these reactions, additional primer mismatches close to the 3'-terminus were used (shown underlined in the primer sequences). The optimal
- 10 primers (H1047R-1 and H1047L-1) were used for the experiments described. The Scorpions primer usable with the primer sequences is shown as SEQ ID NOS. 34 and 35. Regions of correspondence between the Scorpions primer and the target regions are shown in identical highlighting or underlining.

15 AGTGCAGTGTGGAATCCAGAGTGAGCTTTTCAATTTTCTCAGTTATCTTTTCAGTTC
AATGCATGCTGTTTAAATTGTGTGGAAGATCCAATCCATTTTGTGTTGTCCAGCCAC
CATGA(T/C/A)GTGCATCATTCAATTTGTTTCATGAAATACTCCAAAGCCTCTTGCTC
AGTTTTATCTAAGGCTAGGGTCTTTTGAATGTATGCAATGTCATCAAAGATTGT
AGTTCTGGCATTCCAGAGCCAAGCATCATTGAGAAAAGATTTATGAAGAGATTG
GCATGCTGTGCAATAGCTAGATAAGCCTT (SEQ ID NO. 19)

Mutation	Primer Sequence	SEQ ID NO.
H1047R-0	5'-TGTTGTCCAGCCACCATGAC-3'	20
H1047R-1	5'-TGTTGTCCAGCCACCATGCC-3'	21
H1047R-2	5'-TGTTGTCCAGCCACCATGGC-3'	22
H1047R-3	5'-TGTTGTCCAGCCACCATGTC-3'	23
H1047R-4	5'-TGTTGTCCAGCCACCATAAC-3'	24
H1047R-5	5'-TGTTGTCCAGCCACCATCAC-3'	25
H1047R-6	5'-TGTTGTCCAGCCACCATIAC-3'	26
H1047L-0	5'-TGTTGTCCAGCCACCATGAA-3'	27
H1047L-1	5'-TGTTGTCCAGCCACCATGCA-3'	28
H1047L-2	5'-TGTTGTCCAGCCACCATGGA-3'	29
H1047L-3	5'-TGTTGTCCAGCCACCATGTA-3'	30
H1047L-4	5'-TGTTGTCCAGCCACCATAAA-3'	31
H1047L-5	5'-TGTTGTCCAGCCACCATCAA-3'	32
H1047L-6	5'-TGTTGTCCAGCCACCATTAA-3'	33
Exon 20 scorpion	Fam-CGCGGCATGAAATACTCCAAAGCCGCG-que- heg-CCCTAGGCTTAGATAAAACTGAGCAA	34 and 35

Control Primers

- 5 The control primers are shown below. Regions of correspondence between the Scorpions primer and the target regions are shown in identical highlighting or underlining.

10 AGGCTTGAAGAGTGTGGAATTATGTCCTCTGCAAAAAGGCCACTGTGGTTGAAT
TGGGAGAACCCAGACATCATGTCAGAGTTACTGTTTCAGAACAATGAGATCATC
TTTAAAAATGGGGATGG (SEQ ID NO. 36)

Mutation	Primer Sequence	SEQ ID NO.
Control primer	5'-AGATGATCTCATTGTTCTGAAACAG-3'	37
Control scorpion	Rox-CCGGCCAATTCAACCACAGTGGCCGG- que-heg-GGCTTGAAGAGTGTGGAATTAT	38 and 39

All primers were synthesised and supplied by Invitrogen. PCR Buffer, Taq and Magnesium were supplied by Eurogentec and dNTPS were purchased from Abgene Ltd. Scorpions were synthesised and supplied by ATDBio.

- 5 Assays were multiplexed into 2 reactions containing a control assay and 2 ARMS assays (1 x exon 9 and 1 x exon 20). Assays were performed in 25ul reaction volume containing 1x PCR Buffer, 4.0mM MgCl₂, 200uM dNTP mix, 0.25uM of each primer (control primer and 2 ARMS primers) and 0.25uM of each scorpion (control scorpion (SEQ ID NOS. 38 and 39), exon 20 scorpion (SEQ ID NOS. 34 and 35) and exon 9
10 scorpion (SEQ ID NOS. 17 and 18)). 2.5ul of DNA template was added to each reaction. The H1047R and E542K primers were multiplexed with 2.5 unit Taq polymerase per reaction. The H1047L and E545K primers were multiplexed with 3.0 unit Taq polymerase per reaction. The E542K primer used was E542K-2 (SEQ ID NO. 5). The E545K primer used was E545K-4 (SEQ ID NO. 14). The H1047R primer
15 used was H1047R-1 (SEQ ID NO. 21). The H1047L primer used was H1047L-1 (SEQ ID NO. 28).

In all cases the reactions were amplified on a Stratagene Mx3000P under the following conditions: 95°C for 10minutes, followed by 45 cycles of 90°C for 30 seconds and 60°C for 1 minute.

- 20 DNA cassettes harbouring point mutations to use as positive controls were constructed based on a method described by Higuchi et al.^[2]. In brief, corresponding outer and mutamer primers were used to generate half cassettes with complementary ends, each half cassette containing a mutant base. These PCR products were mixed and amplified with inner nested primers. Self priming of the
25 complementary half cassettes and subsequent amplification created a final product with a mutated base. Products were sequenced to ensure the correct sequence had been created. This process was repeated for each mutation of interest. The DNA cassette was mixed with an equal amount of genomic DNA to create a 100% positive control.

Example 1

To determine the specificity of the reactions and the primers each assay was performed with 5-50ng of genomic DNA per reaction to assess breakthrough signal caused by extension of mismatched primer. For each reaction a ΔC_t value (control Ct- mutation Ct) was defined (C_t = threshold cycle). The reactions were performed six times for each DNA concentration and repeated in triplicate on separate occasions to define a cut off ΔC_t value below which any amplification can be said to be due to the presence of mutant sequence and not due to breakthrough signal. The cut off ΔC_t value was determined to be 1 Ct below the lowest ΔC_t value seen in all reactions for each assay. For H1047R and H1047L assays the cut off ΔC_t was defined as 12, for the E542K assay the cut off ΔC_t was 9 and for the E545K assay the cut off ΔC_t was 8.

Example 2

To assess the sensitivity of the assay, 5 copies of mutant DNA were diluted in varying concentrations of genomic DNA to give final concentrations of 5, 2, 1, 0.5 and 0.1% mutant DNA to wild type. Table 1 illustrates the sensitivity of the 4 ARMS assays. The table shows the ΔC_t values for reducing concentrations of mutant DNA within a background of wild type DNA. The predefined cut off ΔC_t s are illustrated in the final column. The exon 20 assays were able to detect 5 copies of mutant DNA when this comprised only 0.1% of the total DNA (within the previously defined cut off ΔC_t). The exon 9 assays were able to detect 5 copies of DNA at 1% concentration with a ΔC_t within the predefined cut off values (Table 1).

Table 1:

WT DNA /reaction (copies)	MUT DNA /reaction (copies)	Relative % of MUT alleles	Δ CT				CUT OFF Δ Ct	
			H1047L	H1047R	E542K	E545K		
100	5	5%	5.9	4.3	5.1	5.8		
250	5	2%	7.2	6.7	7.2	6.4	H1047L	12
500	5	1%	8.3	7.6	8.4	7.0	H1047R	12
1000	5	0.5%	9.3	9.0	10.2	8.1	E542K	9
5000	5	0.1%	11.5	10.5	12.1	10.1	E545K	8

Example 3

- 5 Admixtures of cell lines containing mutation H1047R (HCT-116) and E545K (MCF-7) were used to compare the relative sensitivities of the ARMS assays compared with sequencing. Both cell lines were heterozygous for the mutation. Sequencing was performed using primers and PCR cycling conditions as described by Samuels et al^[1]. ARMS assays and sequencing were carried out at concentrations of the mutant
- 10 gene of 100%, 50%, 30%, 10% and 1% of the total mixture. The results are shown in Figure 1 in which the results under the heading "Scorpions" show the increase in amplicon copy number after successive rounds of PCR (results using control primers and mutant primers are shown as separate lines). Under the heading "DNA Sequencing" is shown the results of sequencing the reverse strand of the gene in the
- 15 mixture. Sequencing was not able to detect the presence of H1047R mutant when present at less than 50% of the total mixture and was unable to detect the presence of E545K mutant when present at less than 30% of the total mixture. Assays using

the primers of the invention, by contrast, were able to detect the presence of mutants at 1% concentration.

Example 4

5 This assay was applied to DNA extracted from fresh frozen tissue from a variety of tumour types that were assessed for the presence of PIK3CA mutations using the ARMS/Scorpion assay. In total DNA was available from 279 tumour samples. The assay reported mutations in 5 of 49 (10.2%) colorectal cancer samples, 19 of 49 (38.7%) breast cancer samples, 1 of 51 (1.9%) lung cancer samples, and 1 of 34
10 (2.9%) melanoma samples. No mutations were detected in 50 prostate or 46 ovarian cancer samples. Of the colorectal samples positive for PIK3CA mutations 3 were H1047R, 1 was H1047L and 1 was E542K; of the breast cancer samples positive for PIK3CA, 15 were H1047R, 1 was H1047L and 3 were E545K; both the mutations in the lung cancer sample and melanoma sample positive for PIK3CA mutations were
15 H1047R. Sequencing identified only 14 of the total 26 (53%) mutations detected. Sequencing detected a mutation in one breast cancer specimen which the ARMS assay was not designed to detect (c.1634 A>G; p E545G). This is not a novel mutation and has been previously described in breast and colorectal cancers^[3-5].

The incidence of PIK3CA mutations in the samples analysed was consistent with
20 previous studies with the exception of ovarian cancer^[1, 3-9]. PIK3CA mutations have been previously described in ovarian cancers but it has been suggested that there may be an association with endometrioid and clear cell cancers^[8, 10]. All the ovarian cancers tested in this study were serous adenocarcinomas which may explain the absence of any PIK3CA mutations.

25 The ARMS assay identified significantly more mutations in the clinical samples than seen by direct sequencing. The cell line admixtures confirm that this assay is more sensitive than sequencing in detecting the PI3KCA mutations of interest. It is likely that the heterogeneity of clinical samples that will contain both tumour and normal tissue will mean that in some instances the incidence of mutation will be below that
30 detectable by sequencing methods and as such the ARMS assay is more suitable for

clinical application. The drawback is that only certain ARMS specific mutations will be detected. However in this series of 279 samples only a single mutation in exon 9 or 20 of the PI3KCA gene was detected that the ARMS assay was not designed to detect.

- 5 In summary, the examples show that the present invention provides a sensitive, high throughput assay for the detection of the 4 most common mutations in the PIK3CA gene. This assay may be applied to small amounts of DNA and can detect low levels of mutant PIK3CA within a sample.

Sequence Listing Free text

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<223> E545K target region

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- <210> 15
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- <210> 19
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<210> 36
<223> Control target region

<210> 37
<223> Control primer

<210> 38
<223> Control Scorpion

<210> 39
<223> Control Scorpion 2

References:

1. Samuels Y, Wang Z, Bardelli A, et al. High frequency of mutations of the PIK3CA gene in human cancers. *Science* 2004;304(5670):554.
2. Higuchi R, Krummel B, Saiki RK. A general method of in vitro preparation and specific mutagenesis of DNA fragments: study of protein and DNA interactions. *Nucleic Acids Res* 1988;16(15):7351-67.
3. Wu G, Xing M, Mambo E, et al. Somatic mutation and gain of copy number of PIK3CA in human breast cancer. *Breast Cancer Res* 2005;7(5):R609-16.
4. Levine DA, Bogomolny F, Yee CJ, et al. Frequent mutation of the PIK3CA gene in ovarian and breast cancers. *Clin Cancer Res* 2005;11(8):2875-8.
5. Velho S, Oliveira C, Ferreira A, et al. The prevalence of PIK3CA mutations in gastric and colon cancer. *Eur J Cancer* 2005;41(11):1649-54.
6. Lee JW, Soung YH, Kim SY, et al. PIK3CA gene is frequently mutated in breast carcinomas and hepatocellular carcinomas. *Oncogene* 2005;24(8):1477-80.
7. Bachman KE, Argani P, Samuels Y, et al. The PIK3CA gene is mutated with high frequency in human breast cancers. *Cancer Biol Ther* 2004;3(8):772-5.
8. Campbell IG, Russell SE, Choong DY, et al. Mutation of the PIK3CA gene in ovarian and breast cancer. *Cancer Res* 2004;64(21):7678-81.
9. Omholt K, Krockel D, Ringborg U, Hansson J. Mutations of PIK3CA are rare in cutaneous melanoma. *Melanoma Res* 2006;16(2):197-200.
10. Wang Y, Helland A, Holm R, Kristensen GB, Borresen-Dale AL. PIK3CA mutations in advanced ovarian carcinomas. *Hum Mutat* 2005;25(3):322.

21a

SEQUENCE LISTING IN ELECTRONIC FORM

In accordance with Section 111(1) of the Patent Rules, this description contains a sequence listing in electronic form in ASCII text format (file: 54097-2 Seq 21-03-10 v1.txt).

A copy of the sequence listing in electronic form is available from the Canadian Intellectual Property Office.

The sequences in the sequence listing in electronic form are reproduced in the following table.

SEQUENCE TABLE

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<151> 2007-09-28

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21b

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22

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22

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22

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22

21c

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22

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21d

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<210> 24
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<223> H1047R-4 fwd primer

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21i

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<210> 39

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<212> DNA

<213> Artificial

<220>

<223> Control Scorpion 2

<400> 39

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21

CLAIMS:

1. A polynucleotide with a sequence consisting of SEQ ID NO: 5.
2. A PCR method of detecting the presence or absence of mutation E542K in the PIK3CA gene comprising the steps of:
 - 5 a) mixing a nucleic acid sample comprising at least a fragment of the PIK3CA gene with a polynucleotide primer having a sequence consisting of SEQ ID NO: 5 wherein the polynucleotide is used as a first primer and a second primer corresponds to a region of the fragment of the PIK3CA sequence downstream of the region to which the polynucleotide is complementary and carrying out a PCR on the mixture; and
 - 10 b) detecting hybridisation of the polynucleotide to the nucleic acid sample wherein hybridisation indicates the presence of the mutation E542K.
3. The method according to claim 2, wherein the second primer has a sequence according to SEQ ID NO: 18.
4. A kit for detecting PIK3CA mutation E542K, the kit comprising a polynucleotide
15 consisting of SEQ ID NO: 5 and a polynucleotide consisting of SED ID NO: 18.

1/1

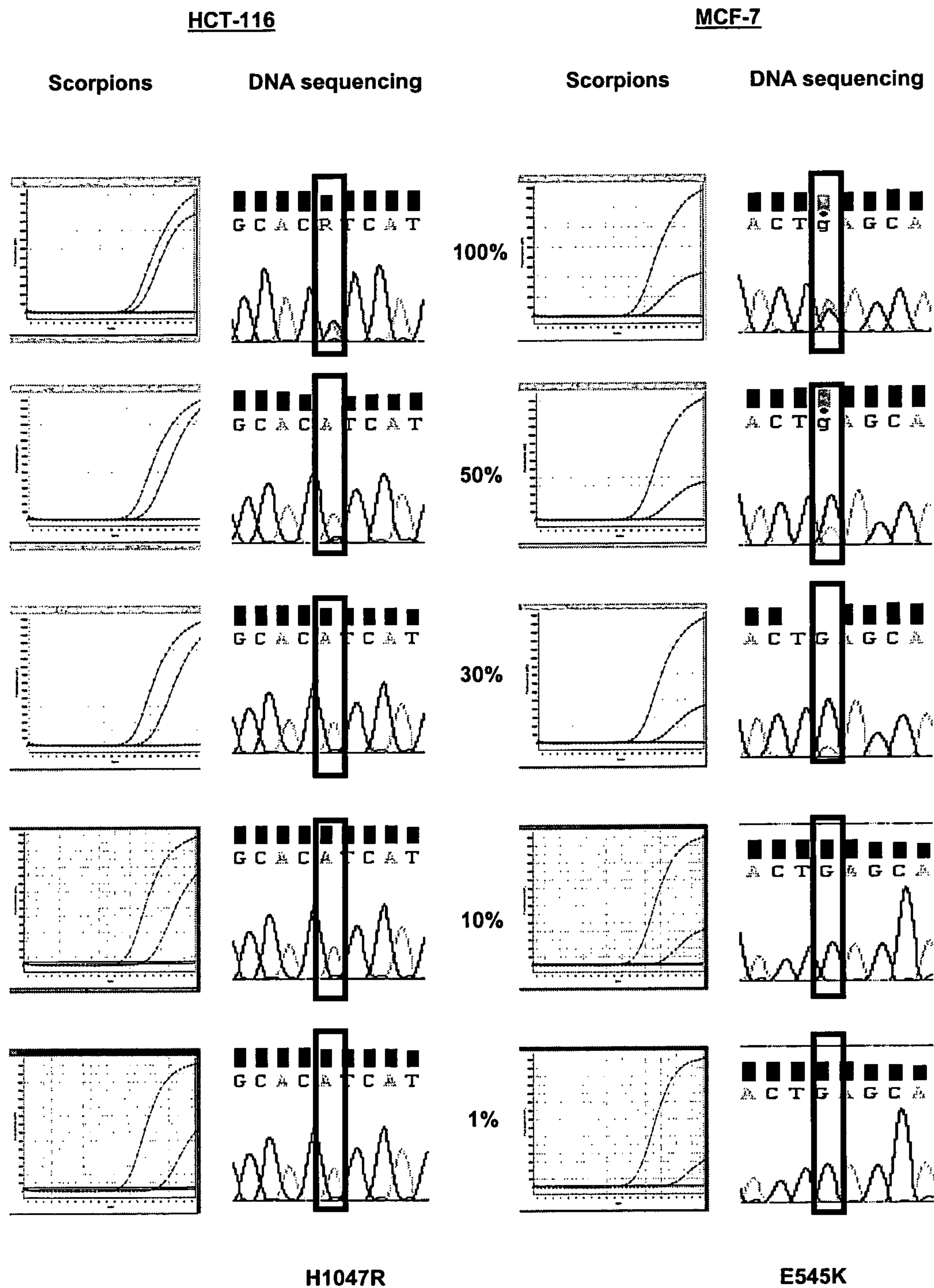


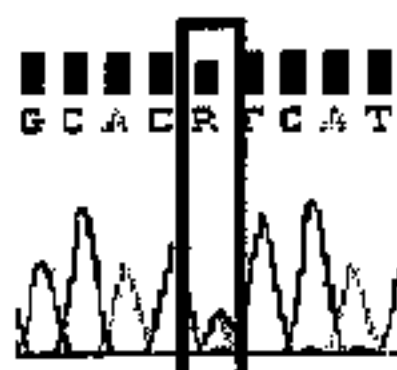
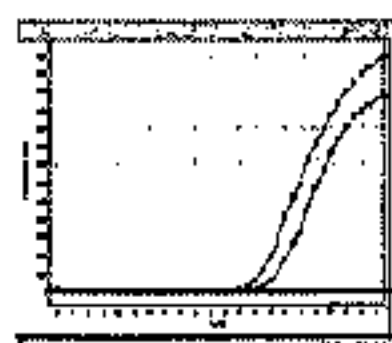
Figure 1

HCT-116

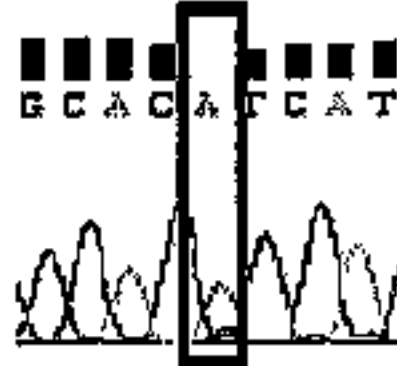
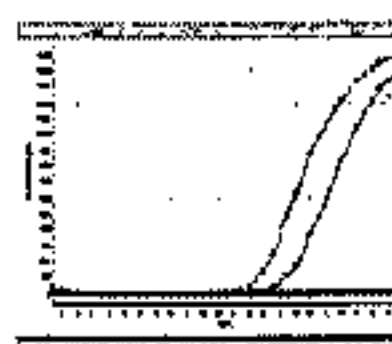
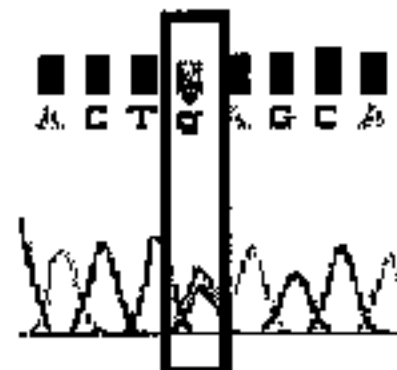
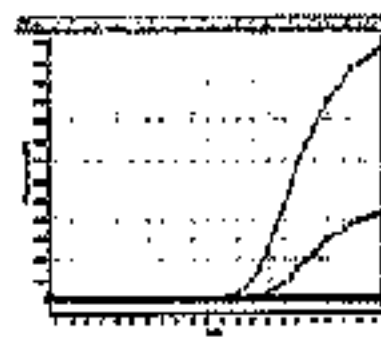
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Scorplons

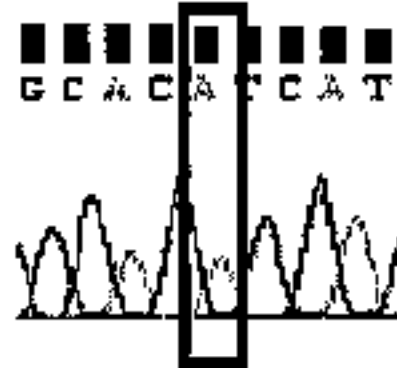
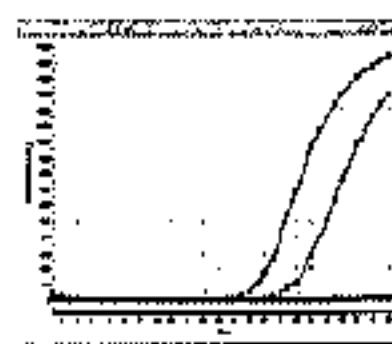
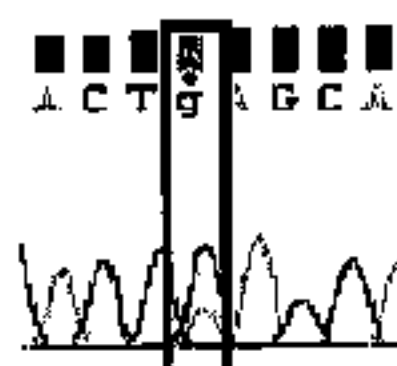
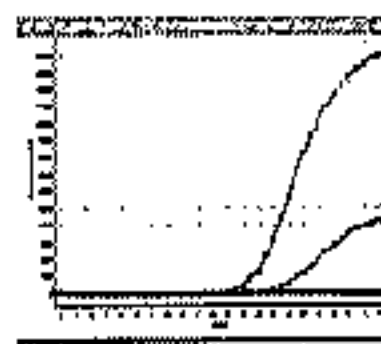
DNA sequencing



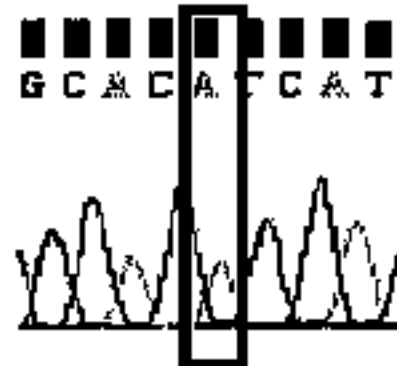
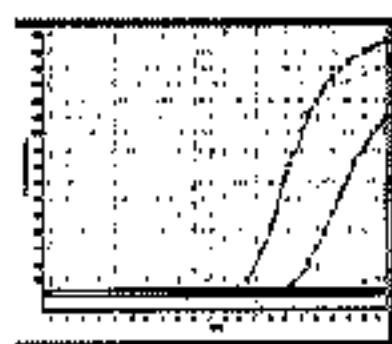
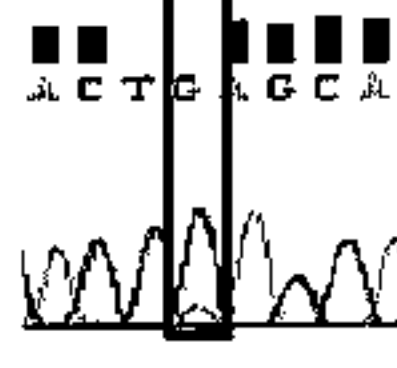
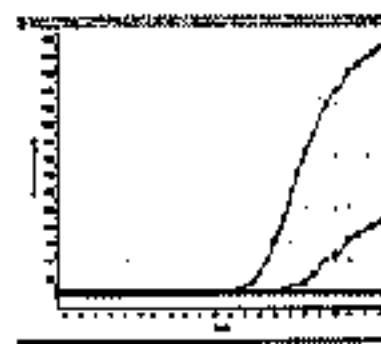
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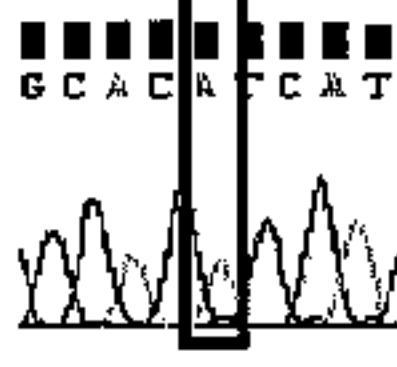
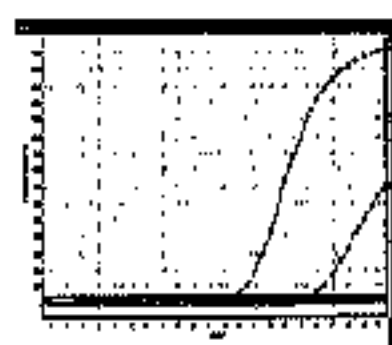
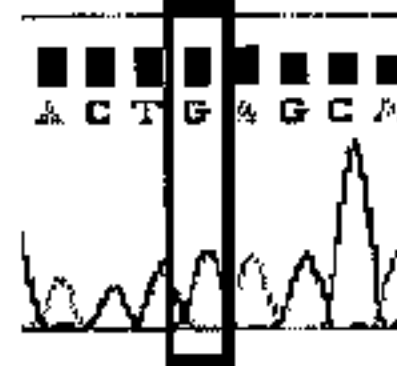
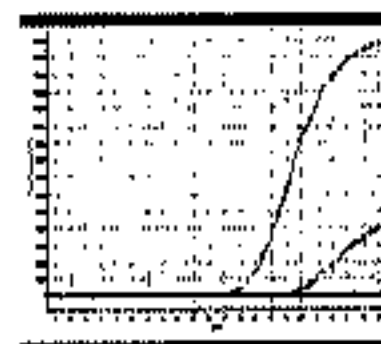
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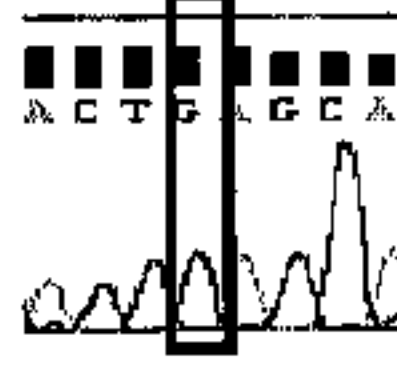
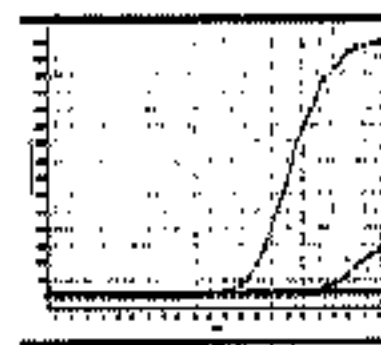
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10%



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E545K