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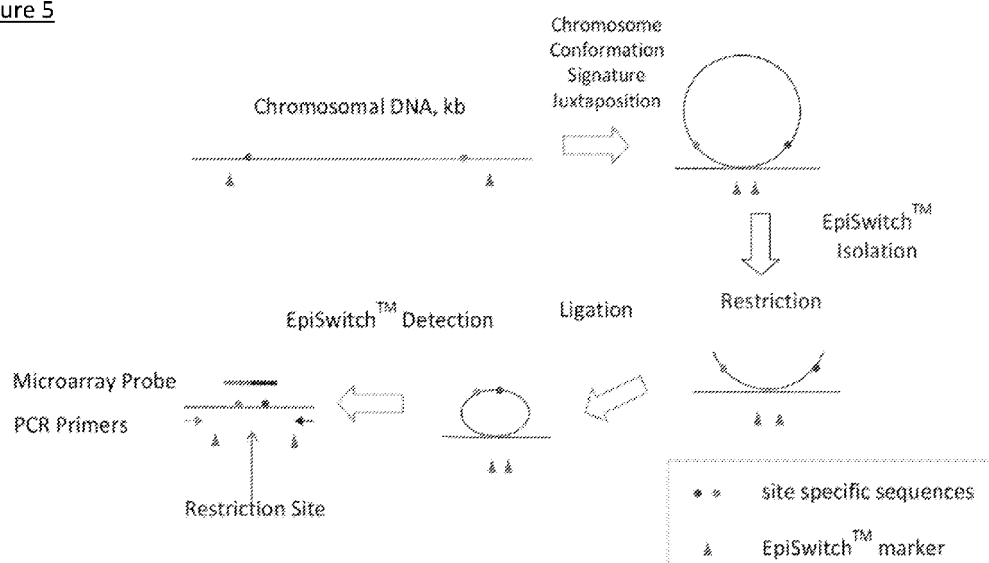
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Figure 5



(57) Abstract: A process for analysing chromosome regions and interactions relating to prognosis of prostate cancer or DLBCL.



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CHROMOSOME CONFORMATION MARKERS OF PROSTATE CANCER AND LYMPHOMA

Field of the Invention

The invention relates to disease processes.

Background of the Invention

- 5 The regulatory and causative aspects of the disease process in cancer are complex and cannot be easily elucidated using available DNA and protein typing methods.

Diffuse large B-cell lymphoma (DLBCL) is a cancer of B cells, a type of white blood cell responsible for producing antibodies. It is the most common type of non-Hodgkin lymphoma among adults, with an annual incidence of 7–8 cases per 100,000 people per year in the USA and the UK. However, there is a
 10 poor understanding of the outcomes of the disease process.

Prostate cancer is caused by the abnormal and uncontrolled growth of cells in the prostate.

Whilst prostate cancer survival rates have been improving from decade to decade, the disease is still considered largely incurable. According to the American Cancer Society, for all stages of prostate cancer combined, the one-year relative survival rate is 20%, and the five-year rate is 7%.

15 Summary of the Invention

The inventors have identified subtypes of patients in prostate cancer, diffuse large B-cell lymphoma (DLBCL) and lymphoma defined by chromosome conformation signatures.

According the invention provides a process for detecting a chromosome state which represents a subgroup in a population comprising determining whether a chromosome interaction relating to that
 20 chromosome state is present or absent within a defined region of the genome; and

- wherein said chromosome interaction has optionally been identified by a method of determining which chromosomal interactions are relevant to a chromosome state corresponding to the subgroup of the population, comprising contacting a first set of nucleic acids from subgroups with different states of the chromosome with a second set of index nucleic acids, and allowing complementary sequences to
 25 hybridise, wherein the nucleic acids in the first and second sets of nucleic acids represent a ligated product comprising sequences from both the chromosome regions that have come together in chromosomal interactions, and wherein the pattern of hybridisation between the first and second set of nucleic acids allows a determination of which chromosomal interactions are specific to the subgroup; and
- wherein the subgroup relates to prognosis for prostate cancer and the chromosome interaction either:
 30 (i) is present in any one of the regions or genes listed in Table 6; and/or

(ii) corresponds to any one of the chromosome interactions represented by any probe shown in Table 6, and/or

(iii) is present in a 4,000 base region which comprises or which flanks (i) or (ii);

or

5 - wherein the subgroup relates to prognosis for DLBCL and the chromosome interaction either:

a) is present in any one of the regions or genes listed in Table 5; and/or

b) corresponds to any one of the chromosome interactions represented by any probe shown in Table 5, and/or

c) is present in a 4,000 base region which comprises or which flanks (a) or (b);

10 or

- wherein the subgroup relates to prognosis for lymphoma and the chromosome interaction either:

(iv) is present in any one of the regions or genes listed in Table 8; and/or

(v) corresponds to any one of the chromosome interactions shown in Table 8, and/or

(vi) is present in a 4,000 base region which comprises or which flanks (iv) or (v).

15

Brief Description of the Drawings

Figure 1 shows a Principle Component Analysis (PCA) for the prostate cancer work.

Figure 2 shows a VENN comparison of the two PCA prognostic classifiers.

Figure 3 shows a PCA analysis for DLBCL.

20 Figure 4 shows a PCA for the 7 BTK markers (OBD RD051) in DLBCL.

Figure 5 shows an example of how the chromosome interaction typing may be carried out.

Figure 6 shows markers from the canine lymphoma work which can be used in the method of the invention. The Figure shows marker reduction. 70% of 38 samples were used as a training set (28) and used for marker selection. The remaining 10 were used as a test set. Multiple training and test sets were
25 used. Univariate analysis, Fisher's Exact test (column D and E results) and Multivariate analysis Penalized logistic modelling (GLMNET, columns B and C results). The markers 2 to 18 are lymphoma markers and 19 to 23 are controls. The top 11, which are all loops present in lymphoma were selected for classification.

Figure 7 shows canine markers to human genes. The table shows the top 11 canine markers mapped to the human genome (Hg38) with the closest mapping genomic region. The network adjacent is built using
30 the 11 markers (dark) the nodes which are a lighter colour and linker proteins using the NCI database.

Figure 8 shows canine markers to human genes. As before but with pathway enrichment for the network. Only the 11 canine mapping loci were used for enrichment, the linking modes were omitted from enrichment. Nodes in lighter colour belong to the KEGG CML pathway.

Figure 9 shows Training Set 1 and Test Set 1 XGBoost 11 Mark Model

5 Figure 10 shows Training Set 2 and Test Set 2 XGBoost 11 Mark Model

Figure 11 shows Training Set 3 and Test Set 3 XGBoost 11 Mark Model

Figure 12 shows Training Set 1 Logistic PCA

10 Figure 13 shows Training Set 1 and Test Set 1 Logistic PCA. The logistic PCA model was used to predict the Test set 1 (triangles). Darker triangles are lymphoma (labelled) from the test set, the lighter triangles are the controls from the test set. The training Lymphoma samples are in darker colour and Controls are in lighter colour.

Figure 14 shows Training Set 1 and Test Set 1 ROC & AUC

15 Figure 15 shows Patient PFS EpiSwitch™ Call and Loop dynamic at NFKB1. 118 patients called either ABC or GCB using EpiSwitch™ 10 marker human model, PFS modelling using this call and dynamic of loop, GCB with loop don't die, shows also that human model works well for disease prognostics.

Figure 16 shows 118 patient PFS EpiSwitch™ Call and loop dynamic at NFATC1. As before but for NFATC1, again this shows that human model for prognostics using the marker as one of the 10 human markers is a very good at classification.

20 Figure 17 shows three-step approach to identify, evaluate, and validate diagnostic and prognostic biomarkers for prostate cancer (PCa).

Figure 18 shows PCA for the five-markers applied to 78 samples containing two groups. First group, 49 known samples (24 PCa and 25 healthy controls (Cntrl)) combined with a second group of 29 samples including, 24 PCa samples and 5 healthy Cntrl samples.

Figure 19 shows the workflow to develop a classifier.

25 Figure 20 shows relevant gene groups for the classifier.

Figure 21 shows overlap of the *EpiSwitch* DLBCL-CCS and Fluidigm subtype calls and ROC Curve when applied to the Discovery cohort. A. Subtype calls made by the EpiSwitch DLBCL-CCS and the Fluidigm assays on samples of known subtypes. 60 out of 60 samples were identically called by both assays. B. The

receiver operating curve (ROC) for the DLBCL-CCS when applied to the Discovery cohort. C. Kaplan-Meier survival analysis (by progression free survival) of samples called as ABC or GCB by the DLBCL-CCS. Samples called as ABC showed a significantly poorer long-term survival than those called as GCB.

Figure 22 shows assignment of DLBCL subtypes in Type III samples by *EpiSwitch* and Fluidigm assays.

- 5 Figure 23 shows comparison of baseline DLBCL subtype calls in Type III samples using *EpiSwitch* and Fluidigm with long term survival. Kaplan-Meier survival curves for the 58 DLBCL patients classified as either ABC, GCB or Unclassified by the Fluidigm assay (A) or the *EpiSwitch* DLBCL-CCS (B). Fluidigm classified 15 samples as ABC, 22 as GCB and 21 were UNC. *EpiSwitch* classified 34 as ABC and 24 as GCB.

Figure 24 shows mean survival time by *EpiSwitch* and Fluidigm classification in the Validation cohort.

- 10 Figure 25 shows initial assessment of likely DLBCL subtype.

Figure 26 shows PCA of DLBCL patients with baseline ABC/GCB subtype calls by *EpiSwitch* in the Discovery cohort.

Detailed Description of the Invention

Aspects of the Invention

- 15 The invention concerns determining prognosis in prostate cancer, particularly in respect to whether the cancer is aggressive or indolent. This determining is by typing any of the relevant markers disclosed herein, for example in Table 6, or preferred combinations of markers, or markers in defined specific regions disclosed herein. Thus the invention relating to a method of typing a patient with prostate cancer to identify whether the cancer is aggressive or indolent.
- 20 The invention also concerns determining prognosis in DLBCL, particularly in respect to whether the prognosis is good or poor in respect of survival. This determining is by typing any of the relevant markers disclosed herein, for example in Table 5, or preferred combinations of markers, or markers in defined specific regions disclosed herein. Thus the invention relates to a method of typing a patient with DLBCL to identify whether the patient has good or poor prognosis in respect of survival, for example to
- 25 determine expected rate of development of disease and/or time to death.

Essentially in the method of the invention subpopulations of prostate cancer or DLBCL identified by typing of the markers. Therefore the invention, for example, concerns a panel of epigenetic markers which relates to prognosis in these conditions. The invention therefore allows personalised therapy to be given to the patient which accurately reflects the patient's needs.

The invention also relates to determining prognosis for lymphoma based on typing chromosome interactions defined by Tables 8 or 9.

Tables 5 to 7 preferably relate to determining prognosis in humans. Tables 8 and 9 preferably relate to determining prognosis in canines.

- 5 Any therapy, for example drug, which is mentioned herein may be administered to an individual based on the result of the method.

Marker sets are disclosed in the Tables and Figures. In one embodiment at least 10 markers from any disclosed marker set are used in the invention. In another embodiment at least 20% of the markers from any disclosed marker set are used in the invention.

10 *The Process of the Invention*

The process of the invention comprises a typing system for detecting chromosome interactions relevant to prognosis. This typing may be performed using the *EpiSwitch*[™] system mentioned herein which is based on cross-linking regions of chromosome which have come together in the chromosome interaction, subjecting the chromosomal DNA to cleavage and then ligating the nucleic acids present in the cross-linked entity to derive a ligated nucleic acid with sequence from both the regions which formed the chromosomal interaction. Detection of this ligated nucleic acid allows determination of the presence or absence of a particular chromosome interaction.

The chromosomal interactions may be identified using the above described method in which populations of first and second nucleic acids are used. These nucleic acids can also be generated using *EpiSwitch*[™] technology.

The Epigenetic Interactions Relevant to the Invention

As used herein, the term 'epigenetic' and 'chromosome' interactions typically refers to interactions between distal regions of a chromosome, said interactions being dynamic and altering, forming or breaking depending upon the status of the region of the chromosome.

- 25 In particular processes of the invention chromosome interactions are typically detected by first generating a ligated nucleic acid that comprises sequence from both regions of the chromosomes that are part of the interactions. In such processes the regions can be cross-linked by any suitable means. In a preferred aspect, the interactions are cross-linked using formaldehyde, but may also be cross-linked by any aldehyde, or D-Biotinoyl-e- aminocaproic acid-N-hydroxysuccinimide ester or Digoxigenin-3-O-methylcarbonyl-e-aminocaproic acid-N-hydroxysuccinimide ester. Para-formaldehyde can cross link DNA
- 30

chains which are 4 Angstroms apart. Preferably the chromosome interactions are on the same chromosome and optionally 2 to 10 Angstroms apart.

The chromosome interaction may reflect the status of the region of the chromosome, for example, if it is being transcribed or repressed in response to change of the physiological conditions. Chromosome interactions which are specific to subgroups as defined herein have been found to be stable, thus providing a reliable means of measuring the differences between the two subgroups.

In addition, chromosome interactions specific to a characteristic (such as prognosis) will normally occur early in a biological process, for example compared to other epigenetic markers such as methylation or changes to binding of histone proteins. Thus the process of the invention is able to detect early stages of a biological process. This allows early intervention (for example treatment) which may as a consequence be more effective. Chromosome interactions also reflect the current state of the individual and therefore can be used to assess changes to prognosis. Furthermore there is little variation in the relevant chromosome interactions between individuals within the same subgroup. Detecting chromosome interactions is highly informative with up to 50 different possible interactions per gene, and so processes of the invention can interrogate 500,000 different interactions.

Preferred Marker Sets

Herein the term 'marker' or 'biomarker' refers to a specific chromosome interaction which can be detected (typed) in the invention. Specific markers are disclosed herein, any of which may be used in the invention. Further sets of markers may be used, for example in the combinations or numbers disclosed herein. The specific markers disclosed in the tables herein are preferred as well as markers presents in genes and regions mentioned in the tables herein are preferred. These may be typed by any suitable method, for example the PCR or probe based methods disclosed herein, including a qPCR method. The markers are defined herein by location or by probe and/or primer sequences.

Location and Causes of Epigenetic Interactions

Epigenetic chromosomal interactions may overlap and include the regions of chromosomes shown to encode relevant or undescribed genes, but equally may be in intergenic regions. It should further be noted that the inventors have discovered that epigenetic interactions in all regions are equally important in determining the status of the chromosomal locus. These interactions are not necessarily in the coding region of a particular gene located at the locus and may be in intergenic regions.

The chromosome interactions which are detected in the invention could be caused by changes to the underlying DNA sequence, by environmental factors, DNA methylation, non-coding antisense RNA

transcripts, non-mutagenic carcinogens, histone modifications, chromatin remodelling and specific local DNA interactions. The changes which lead to the chromosome interactions may be caused by changes to the underlying nucleic acid sequence, which themselves do not directly affect a gene product or the mode of gene expression. Such changes may be for example, SNPs within and/or outside of the genes, gene fusions and/or deletions of intergenic DNA, microRNA, and non-coding RNA. For example, it is known that roughly 20% of SNPs are in non-coding regions, and therefore the process as described is also informative in non-coding situation. In one aspect the regions of the chromosome which come together to form the interaction are less than 5 kb, 3 kb, 1 kb, 500 base pairs or 200 base pairs apart on the same chromosome.

The chromosome interaction which is detected is preferably within any of the genes mentioned in Table 5. However it may also be upstream or downstream of the gene, for example up to 50,000, up to 30,000, up to 20,000, up to 10,000 or up to 5000 bases upstream or downstream from the gene or from the coding sequence.

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The chromosome interaction which is detected is preferably within any of the genes mentioned in Table 9. However it may also be upstream or downstream of the gene, for example up to 50,000, up to 30,000, up to 20,000, up to 10,000 or up to 5000 bases upstream or downstream from the gene or from the coding sequence.

Subgroups, Time Points and Personalised Treatment

The aim of the present invention is to determine prognosis. This may be at one or more defined time points, for example at at least 1, 2, 5, 8 or 10 different time points. The durations between at least 1, 2, 5 or 8 of the time points may be at least 5, 10, 20, 50, 80 or 100 days.

As used herein, a “subgroup” preferably refers to a population subgroup (a subgroup in a population), more preferably a subgroup in the population of a particular animal such as a particular eukaryote, or mammal (e.g. human, non-human, non-human primate, or rodent e.g. mouse or rat). Most preferably, a “subgroup” refers to a subgroup in the human population. The subgroup may be a canine subgroup, such as a dog.

The invention includes detecting and treating particular subgroups in a population. The inventors have discovered that chromosome interactions differ between subsets (for example at least two subsets) in a given population. Identifying these differences will allow physicians to categorize their patients as a part of one subset of the population as described in the process. The invention therefore provides physicians with a process of personalizing medicine for the patient based on their epigenetic chromosome interactions.

In one aspect the invention relates to testing whether an individual:

- is a fast or slow 'progressor', and/or
- has an aggressive or indolent form of disease.

10 The invention may also determine the expected survival time of the individual.

Such testing may be used to select how to subsequently treat the patient, for example the type of drug and/or its dose and/or its frequency of administration.

Generating Ligated Nucleic Acids

15 Certain aspects of the invention utilise ligated nucleic acids, in particular ligated DNA. These comprise sequences from both of the regions that come together in a chromosome interaction and therefore provide information about the interaction. The EpiSwitch™ method described herein uses generation of such ligated nucleic acids to detect chromosome interactions.

Thus a process of the invention may comprise a step of generating ligated nucleic acids (e.g. DNA) by the following steps (including a method comprising these steps):

- 20 (i) cross-linking of epigenetic chromosomal interactions present at the chromosomal locus, preferably *in vitro*;
- (ii) optionally isolating the cross-linked DNA from said chromosomal locus;
- (iii) subjecting said cross-linked DNA to cutting, for example by restriction digestion with an enzyme that cuts it at least once (in particular an enzyme that cuts at least once within said chromosomal locus);
- 25 (iv) ligating said cross-linked cleaved DNA ends (in particular to form DNA loops); and
- (v) optionally identifying the presence of said ligated DNA and/or said DNA loops, in particular using techniques such as PCR (polymerase chain reaction), to identify the presence of a specific chromosomal interaction.

These steps may be carried out to detect the chromosome interactions for any aspect mentioned herein. The steps may also be carried out to generate the first and/or second set of nucleic acids mentioned herein.

5 PCR (polymerase chain reaction) may be used to detect or identify the ligated nucleic acid, for example the size of the PCR product produced may be indicative of the specific chromosome interaction which is present, and may therefore be used to identify the status of the locus. In preferred aspects at least 1, 2 or 3 primers or primer pairs as shown in Table 5 are used in the PCR reaction. In other aspects at least 1, 10, 20, 30, 50 or 80 or the primers or primer pairs as shown in Table 6 are used in the PCR reaction. The skilled person will be aware of numerous restriction enzymes which can be used to cut the DNA within
10 the chromosomal locus of interest. It will be apparent that the particular enzyme used will depend upon the locus studied and the sequence of the DNA located therein. A non-limiting example of a restriction enzyme which can be used to cut the DNA as described in the present invention is TaqI.

EpiSwitch™ Technology

The EpiSwitch™ Technology also relates to the use of microarray EpiSwitch™ marker data in the detection
15 of epigenetic chromosome conformation signatures specific for phenotypes. Aspects such as EpiSwitch™ which utilise ligated nucleic acids in the manner described herein have several advantages. They have a low level of stochastic noise, for example because the nucleic acid sequences from the first set of nucleic acids of the present invention either hybridise or fail to hybridise with the second set of nucleic acids. This provides a binary result permitting a relatively simple way to measure a complex mechanism at the
20 epigenetic level. EpiSwitch™ technology also has fast processing time and low cost. In one aspect the processing time is 3 hours to 6 hours.

Samples and Sample Treatment

The process of the invention will normally be carried out on a sample. The sample may be obtained at a defined time point, for example at any time point defined herein. The sample will normally contain DNA
25 from the individual. It will normally contain cells. In one aspect a sample is obtained by minimally invasive means, and may for example be a blood sample. DNA may be extracted and cut up with a standard restriction enzyme. This can pre-determine which chromosome conformations are retained and will be detected with the EpiSwitch™ platforms. Due to the synchronisation of chromosome interactions between tissues and blood, including horizontal transfer, a blood sample can be used to detect the
30 chromosome interactions in tissues, such as tissues relevant to disease. For certain conditions, such as cancer, genetic noise due to mutations can affect the chromosome interaction 'signal' in the relevant tissues and therefore using blood is advantageous.

Properties of Nucleic Acids of the Invention

The invention relates to certain nucleic acids, such as the ligated nucleic acids which are described herein as being used or generated in the process of the invention. These may be the same as, or have any of the properties of, the first and second nucleic acids mentioned herein. The nucleic acids of the invention typically comprise two portions each comprising sequence from one of the two regions of the chromosome which come together in the chromosome interaction. Typically each portion is at least 8, 10, 15, 20, 30 or 40 nucleotides in length, for example 10 to 40 nucleotides in length. Preferred nucleic acids comprise sequence from any of the genes mentioned in any of the tables. Typically preferred nucleic acids comprise the specific probe sequences mentioned in Table 5; or fragments and/or homologues of such sequences. The preferred nucleic acids may comprise the specific probe sequences mentioned in Table 6; or fragments and/or homologues of such sequences.

Preferably the nucleic acids are DNA. It is understood that where a specific sequence is provided the invention may use the complementary sequence as required in the particular aspect. Preferably the nucleic acids are DNA. It is understood that where a specific sequence is provided the invention may use the complementary sequence as required in the particular aspect.

The primers shown in Table 5 may also be used in the invention as mentioned herein. In one aspect primers are used which comprise any of: the sequences shown in Table 5; or fragments and/or homologues of any sequence shown in Table 5. The primers shown in Table 6 may also be used in the invention as mentioned herein. In one aspect primers are used which comprise any of: the sequences shown in Table 6; or fragments and/or homologues of any sequence shown in Table 6. The primers shown in Table 8 may also be used in the invention as mentioned herein. In one aspect primers are used which comprise any of: the sequences shown in Table 8; or fragments and/or homologues of any sequence shown in Table 8.

The Second Set of Nucleic Acids – the ‘Index’ Sequences

The second set of nucleic acid sequences has the function of being a set of index sequences, and is essentially a set of nucleic acid sequences which are suitable for identifying subgroup specific sequence. They can represent the ‘background’ chromosomal interactions and might be selected in some way or be unselected. They are in general a subset of all possible chromosomal interactions.

The second set of nucleic acids may be derived by any suitable process. They can be derived computationally or they may be based on chromosome interaction in individuals. They typically represent a larger population group than the first set of nucleic acids. In one particular aspect, the second set of nucleic acids represents all possible epigenetic chromosomal interactions in a specific set of genes. In another particular aspect, the second set of nucleic acids represents a large proportion of all possible epigenetic chromosomal interactions present in a population described herein. In one particular aspect, the second set of nucleic acids represents at least 50% or at least 80% of epigenetic chromosomal interactions in at least 20, 50, 100 or 500 genes, for example in 20 to 100 or 50 to 500 genes.

The second set of nucleic acids typically represents at least 100 possible epigenetic chromosome interactions which modify, regulate or in any way mediate a phenotype in population. The second set of nucleic acids may represent chromosome interactions that affect a disease state (typically relevant to diagnosis or prognosis) in a species. The second set of nucleic acids typically comprises sequences representing epigenetic interactions both relevant and not relevant to a prognosis subgroup.

In one particular aspect the second set of nucleic acids derive at least partially from naturally occurring sequences in a population, and are typically obtained by *in silico* processes. Said nucleic acids may further comprise single or multiple mutations in comparison to a corresponding portion of nucleic acids present in the naturally occurring nucleic acids. Mutations include deletions, substitutions and/or additions of one or more nucleotide base pairs. In one particular aspect, the second set of nucleic acids may comprise sequence representing a homologue and/or orthologue with at least 70% sequence identity to the corresponding portion of nucleic acids present in the naturally occurring species. In another particular aspect, at least 80% sequence identity or at least 90% sequence identity to the corresponding portion of nucleic acids present in the naturally occurring species is provided.

Properties of the Second Set of Nucleic Acids

In one particular aspect, there are at least 100 different nucleic acid sequences in the second set of nucleic acids, preferably at least 1000, 2000 or 5000 different nucleic acids sequences, with up to 100,000, 1,000,000 or 10,000,000 different nucleic acid sequences. A typical number would be 100 to 1,000,000, such as 1,000 to 100,000 different nucleic acids sequences. All or at least 90% or at least 50% of these would correspond to different chromosomal interactions.

In one particular aspect, the second set of nucleic acids represent chromosome interactions in at least 20 different loci or genes, preferably at least 40 different loci or genes, and more preferably at least 100, at least 500, at least 1000 or at least 5000 different loci or genes, such as 100 to 10,000 different loci or genes. The lengths of the second set of nucleic acids are suitable for them to specifically hybridise according to Watson Crick base pairing to the first set of nucleic acids to allow identification of chromosome interactions specific to subgroups. Typically the second set of nucleic acids will comprise two portions corresponding in sequence to the two chromosome regions which come together in the chromosome interaction. The second set of nucleic acids typically comprise nucleic acid sequences which are at least 10, preferably 20, and preferably still 30 bases (nucleotides) in length. In another aspect, the nucleic acid sequences may be at the most 500, preferably at most 100, and preferably still at most 50 base pairs in length. In a preferred aspect, the second set of nucleic acids comprises nucleic acid sequences of between 17 and 25 base pairs. In one aspect at least 100, 80% or 50% of the second set of nucleic acid sequences have lengths as described above. Preferably the different nucleic acids do not have any overlapping sequences, for example at least 100%, 90%, 80% or 50% of the nucleic acids do not have the same sequence over at least 5 contiguous nucleotides.

Given that the second set of nucleic acids acts as an 'index' then the same set of second nucleic acids may be used with different sets of first nucleic acids which represent subgroups for different characteristics, i.e. the second set of nucleic acids may represent a 'universal' collection of nucleic acids which can be used to identify chromosome interactions relevant to different characteristics.

The First Set of Nucleic Acids

The first set of nucleic acids are typically from subgroups relevant to prognosis. The first nucleic acids may have any of the characteristics and properties of the second set of nucleic acids mentioned herein. The first set of nucleic acids is normally derived from samples from the individuals which have undergone treatment and processing as described herein, particularly the EpiSwitchTM cross-linking and cleaving steps. Typically the first set of nucleic acids represents all or at least 80% or 50% of the chromosome interactions present in the samples taken from the individuals.

Typically, the first set of nucleic acids represents a smaller population of chromosome interactions across the loci or genes represented by the second set of nucleic acids in comparison to the chromosome interactions represented by second set of nucleic acids, i.e. the second set of nucleic acids is representing a background or index set of interactions in a defined set of loci or genes.

Library of Nucleic Acids

Any of the types of nucleic acid populations mentioned herein may be present in the form of a library comprising at least 200, at least 500, at least 1000, at least 5000 or at least 10000 different nucleic acids of that type, such as 'first' or 'second' nucleic acids. Such a library may be in the form of being bound to an array. The library may comprise some or all of the probes or primer pairs shown in Table 5 or 6. The library may comprise all of the probe sequence from any of the tables disclosed herein.

Hybridisation

The invention requires a means for allowing wholly or partially complementary nucleic acid sequences from the first set of nucleic acids and the second set of nucleic acids to hybridise. In one aspect all of the first set of nucleic acids is contacted with all of the second set of nucleic acids in a single assay, i.e. in a single hybridisation step. However any suitable assay can be used.

Labelled Nucleic Acids and Pattern of Hybridisation

The nucleic acids mentioned herein may be labelled, preferably using an independent label such as a fluorophore (fluorescent molecule) or radioactive label which assists detection of successful hybridisation. Certain labels can be detected under UV light. The pattern of hybridisation, for example on an array described herein, represents differences in epigenetic chromosome interactions between the two subgroups, and thus provides a process of comparing epigenetic chromosome interactions and determination of which epigenetic chromosome interactions are specific to a subgroup in the population of the present invention.

The term 'pattern of hybridisation' broadly covers the presence and absence of hybridisation between the first and second set of nucleic acids, i.e. which specific nucleic acids from the first set hybridise to which specific nucleic acids from the second set, and so it not limited to any particular assay or technique, or the need to have a surface or array on which a 'pattern' can be detected.

Selecting a Subgroup with Particular Characteristics

The invention provides a process which comprises detecting the presence or absence of chromosome interactions, typically 5 to 20 or 5 to 500 such interactions, preferably 20 to 300 or 50 to 100 interactions, in order to determine the presence or absence of a characteristic relating to prognosis in an individual. Preferably the chromosome interactions are those in any of the genes mentioned herein. In one aspect

the chromosome interactions which are typed are those represented by the nucleic acids in Table 5. In another aspect the chromosome interactions are those represented in Table 6. In a further aspect the chromosome interactions which are typed are those represented by the nucleic acids in Table 8. The column titled 'Loop Detected' in the tables shows which subgroup is detected by each probe. Detection can either of the presence or absence of the chromosome interaction in that subgroup, which is what '1' and '-1' indicate.

The Individual that is Tested

Examples of the species that the individual who is tested is from are mentioned herein. In addition the individual that is tested in the process of the invention may have been selected in some way. The individual may be susceptible to any condition mentioned herein and/or may be in need of any therapy mentioned in. The individual may be receiving any therapy mentioned herein. In particular, the individual may have, or be suspected of having, prostate cancer or DLBCL. The individual may have, or be suspected of having, a lymphoma.

Preferred Gene Regions, Loci, Genes and Chromosome Interactions for Prostate Cancer

For all aspects of the invention preferred gene regions, loci, genes and chromosome interactions are mentioned in the tables, for example in Table 6. Typically in the processes of the invention chromosome interactions are detected from at least 1, 2, 3, 4 or 5 of the relevant genes listed in Table 6. Preferably the presence or absence of at least 1, 2, 3, 4 or 5 of the relevant specific chromosome interactions represented by the probe sequences in Table 6 are detected. The chromosome interaction may be upstream or downstream of any of the genes mentioned herein, for example 50 kb upstream or 20 kb downstream, for example from the coding sequence.

For all aspects of the invention preferred gene regions, loci, genes and chromosome interactions are mentioned in Table 25. Typically in the processes of the invention chromosome interactions are detected from at least 2, 4, 8, 10, 14 or all of the relevant genes listed in Table 25. Preferably the presence or absence of at least 2, 4, 8, 10, 14 or all of the relevant specific chromosome interactions shown in Table 25 are detected. The chromosome interaction may be upstream or downstream of any of the genes mentioned herein, for example 50 kb upstream or 20 kb downstream, for example from the coding sequence.

In one embodiment a combination of specific markers disclosed herein and represented by (identified by) the following combination of genes is typed: *ETS1*, *MAP3K14*, *SLC22A3* and *CASP2*. This may be to determine diagnosis. Preferably at least 2 or 3 of these markers are typed.

5 In another embodiment a combination of specific markers disclosed herein represented by (identified by) the following combination of genes is typed: *BMP6*, *ERG*, *MSR1*, *MUC1*, *ACAT1* and *DAPK1*. This may be to determine prognosis (High-risk Category 3 vs Low Risk Category 1, by Nested PCR Markers). Preferably at least 2 or 3 of these markers are typed.

10 In a further embodiment a combination of specific markers disclosed herein represented by (identified by) the following combination of genes is typed: *HSD3B2*, *VEGFC*, *APAF1*, *MUC1*, *ACAT1* and *DAPK1*. This may be to determine prognosis (High Risk Cat 3 vs Medium Risk Cat 2). Preferably at least 2 or 3 of these markers are typed.

Preferred Gene Regions, Loci, Genes and Chromosome Interactions for DLBCL

Typically at least 10, 20, 30, 50 or 80 chromosome interactions are typed from any of genes or regions disclosed the tables herein, or parts of tables disclosed herein. Preferably at least 10, 20, 30, 50 or 80
15 chromosome interactions are typed from any of the genes or regions disclosed in Table 5.

Preferably at least 2, 3, 5, 8 of the markers of Table 7 are typed.

Preferably the presence or absence of at least 10, 20, 30, 50 or 80 chromosome interactions represented by the probe sequences in Table 5 are detected. The chromosome interaction may be upstream or downstream of any of the genes mentioned herein, for example 50 kb upstream or 20 kb downstream,
20 for example from the coding sequence.

Preferably at least 1, 2, 5, 8 or all of the first 10 markers shown in Table 5 is typed. In one embodiment at least 1, 2, 3 or 6 markers from Table 5 are typed each corresponding to a different gene selected from *STAT3*, *TNFRSF13B*, *ANXA11*, *MAP3K7*, *MEF2B* and *IFNAR1*.

Preferred Gene Regions, Loci, Genes and Chromosome Interactions for Lymphoma

25 Typically at least 10, 20, 30 or 50 chromosome interactions are typed from any of the genes or regions disclosed the tables herein, or parts of tables disclosed herein. Preferably at least 10, 20, 30 or 50 chromosome interactions are typed from any of the genes or regions disclosed in Table 8.

Preferably at least 5, 10 or 15 of the markers of Table 9 are typed.

The chromosome interaction may be upstream or downstream of any of the genes mentioned herein, for example 50 kb upstream or 20 kb downstream, for example from the coding sequence.

In one embodiment at least one of the first 11 markers shown in Figure 6 is typed. In another embodiment at least 1, 2, 3 or 6 markers from Table 8 are typed each corresponding to a different gene selected from:

5 STAT3, TNFRSF13B, ANXA11, MAP3K7, MEF2B and IFNAR1.

Types of Chromosome Interaction

In one aspect the locus (including the gene and/or place where the chromosome interaction is detected) may comprise a CTCF binding site. This is any sequence capable of binding transcription repressor CTCF. That sequence may consist of or comprise the sequence CCCTC which may be present in 1, 2 or 3 copies
10 at the locus. The CTCF binding site sequence may comprise the sequence CCGCGNGGNGGCAG (in IUPAC notation). The CTCF binding site may be within at least 100, 500, 1000 or 4000 bases of the chromosome interaction or within any of the chromosome regions shown Table 5 or 6. The CTCF binding site may be within at least 100, 500, 1000 or 4000 bases of the chromosome interaction or within any of the chromosome regions shown Table 5 or 6.

15 In one aspect the chromosome interactions which are detected are present at any of the gene regions shown Table 5 or 6. In the case where a ligated nucleic acid is detected in the process then sequence shown in any of the probe sequences in Table 5 or 6 may be detected.

Thus typically sequence from both regions of the probe (i.e. from both sites of the chromosome interaction) could be detected. In preferred aspects probes are used in the process which comprise or
20 consist of the same or complementary sequence to a probe shown in any table. In some aspects probes are used which comprise sequence which is homologous to any of the probe sequences shown in the tables.

Tables Provided Herein

Tables 5 and 6 shows probe (EpiswitchTM marker) data and gene data representing chromosome
25 interactions relevant to prognosis. The probe sequences show sequence which can be used to detect a ligated product generated from both sites of gene regions that have come together in chromosome interactions, i.e. the probe will comprise sequence which is complementary to sequence in the ligated product. The first two sets of Start-End positions show probe positions, and the second two sets of Start-End positions show the relevant 4kb region. The following information is provided in the probe data table:

- HyperG_Stats: p-value for the probability of finding that number of significant EpiSwitch™ markers in the locus based on the parameters of hypergeometric enrichment
- Probe Count Total: Total number of EpiSwitch™ Conformations tested at the locus
- Probe Count Sig: Number of EpiSwitch™ Conformations found to be statistically significant at the locus
- FDR HyperG: Multi-test (Fimmunoresponse Discovery Rate) corrected hypergeometric p-value
- Percent Sig: Percentage of significant EpiSwitch™ markers relative the number of markers tested at the locus
- logFC: logarithm base 2 of Epigenetic Ratio (FC)
- AveExpr: average log2-expression for the probe over all arrays and channels
- T: moderated t-statistic
- p-value: raw p-value
- adj. p-value: adjusted p-value or q-value
- B - B-statistic (lods or B) is the log-odds that that gene is differentially expressed.
- FC - non-log Fold Change
- FC_1 - non-log Fold Change centred around zero
- LS – Binary value this relates to FC_1 values. FC_1 value below -1.1 it is set to -1 and if the FC_1 value is above 1.1 it is set to 1. Between those values the value is 0

Tables 5 and 6 shows genes where a relevant chromosome interaction has been found to occur. The p-value in the loci table is the same as the HyperG Stats (p-value for the probability of finding that number of significant EpiSwitch™ markers in the locus based on the parameters of hypergeometric enrichment). The LS column shows presence or absence of the relevant interaction with that particular subgroup (prognosis status).

For table 5, DLBCL refers to prognosis marker, indicated with 1, and healthy refers to healthy control, indicated with -1.

The probes are designed to be 30bp away from the Taq1 site. In case of PCR, PCR primers are typically designed to detect ligated product but their locations from the Taq1 site vary.

Probe locations:

Start 1 - 30 bases upstream of TaqI site on fragment 1

End 1 - TaqI restriction site on fragment 1

Start 2 - TaqI restriction site on fragment 2

End 2 - 30 bases downstream of TaqI site on fragment 2

4kb Sequence Location:

Start 1 - 4000 bases upstream of TaqI site on fragment 1

End 1 - TaqI restriction site on fragment 1

Start 2 - TaqI restriction site on fragment 2

End 2 - 4000 bases downstream of TaqI site on fragment 2

- 5 GLMNET values related to procedures for fitting the entire lasso or elastic-net regularization (Lambda set to 0.5 (elastic-net)).

In the tables herein the prostate cancer aggressive subgroup refers to class 3 patients with the following description:

- PSA level is more than 20ng/ml, and
- 10 - the Gleason score is between 8 and 10, and
- the T stage is T2c, T3 or T4

In the tables herein the prostate cancer indolent subgroup refers to class 1 patient with the following description:

- the PSA level is less than 10 ng per ml, and
- 15 - the Gleason score is no higher than 6, and
- the T stage is between T1 and T2a.

Table 7 shows preferred markers for DLBCL. Tables 8 and 9 show preferred markers for lymphoma.

Tables 5 to 7 are preferably for typing humans. Tables 8 and 9 are preferably for typing canines, for examples dogs.

20 *The Approach Taken to Identify Markers and Panels of Markers*

The invention described herein relates to chromosome conformation profile and 3D architecture as a regulatory modality in its own right, closely linked to the phenotype. The discovery of biomarkers was based on annotations through pattern recognition and screening on representative cohorts of clinical samples representing the differences in phenotypes. We annotated and screened significant parts of the genome, across coding and non-coding parts and over large sways of non-coding 5' and 3' of known genes for identification of statistically disseminating consistent conditional disseminating chromosome

conformations, which for example anchor in the non-coding sites within (intronic) or outside of open reading frames

In selection of the best markers we are driven by statistical data and p values for the marker leads. The reference to the particular genes is used for the ease of the position reference - the closest genes are usually used for the reference. It is impossible to exclude the possibility, that a chromosome conformation in the cis- position and relevant vicinity from a gene might be contributing a specific component of regulation into expression of that particular gene. At the point of marker selection or validation expression parameters are not needed on the genes referenced as location coordinates in the names of chromosome conformations. Selected and validated chromosome conformations within the signature are disseminating stratifying entities in their own right, irrespective of the expression profiles of the genes used in the reference. Further work may be done on relevant regulatory modalities, such as SNPs at the anchoring sites, changes in gene transcription profiles, changes at the level of H3K27ac.

We are taking the question of clinical phenotype differences and their stratification from the basis of fundamental biology and epigenetics controls over phenotype - including for example from the framework of network of regulation. As such, to assist stratification, one can capture changes in the network and it is preferably done through signatures of several biomarkers, for example through following a machine learning algorithm for marker reduction which includes evaluating the optimal number of markers to stratify the testing cohort with minimal noise. This usually ends with 3-17 markers, depending on case by case basis. Selection of markers for panels may be done by cross-validation statistical performance (and not for example by the functional relevance of the neighbouring genes, used for the reference name).

A panel of markers (with names of adjacent genes) is a product of clustered selection from the screening across significant parts of the genome, in non-biased way analysing statistical disseminating powers over 14,000-60,000 annotated EpiSwitch sites across significant parts of the genome. It should not be perceived as a tailored capture of a chromosome conformation on the gene of know functional value for the question of stratification. The total number of sites for chromosome interaction are 1.2 million, and so the potential number of combinations is 1.2 million to the power 1.2 million. The approach that we have followed nevertheless allows the identifying of the relevant chromosome interactions.

The specific markers that are provided by this application have passed selection, being statistically (significantly) associated with the condition. This is what the p-value in the relevant table demonstrates. Each marker can be seen as representing an event of biological epigenetic as part of network

deregulation that is manifested in the relevant condition. In practical terms it means that these markers are prevalent across groups of patients when compared to controls. On average, as an example, an individual marker may typically be present in 80% of patients tested and in 10% of controls tested.

Simple addition of all markers would not represent the network interrelationships between some of the deregulations. This is where the standard multivariate biomarker analysis GLMNET (R package) is brought in. GLMNET package helps to identify interdependence between some of the markers, that reflect their joint role in achieving deregulations leading to disease phenotype. Modelling and then testing markers with highest GLMNET scores offers not only identify the minimal number of markers that accurately identifies the patient cohort, but also the minimal number that offers the least false positive results in the control group of patients, due to background statistical noise of low prevalence in the control group. Typically a group (combination) of selected markers (such as 3 to 10) offers the best balance between both sensitivity and specificity of detection, emerging in the context of multivariate analysis from individual properties of all the selected statistical significant markers for the condition.

The tables herein show the reference names for the array probes (60-mer) for array analysis that overlaps the juncture between the long range interaction sites, the chromosome number and the start and end of two chromosomal fragments that come into juxtaposition. The tables also show standard array readouts in competitive hybridisation of disease versus control samples (labeled with two different fluorescent colours) for each of the markers. As a standard readout it shows for each marker probe:

- an average expression signal

- t test for significant difference between fluorescent colour detection for controls and for disease samples

- p value of significance of the marker readout

- adjusted p-value (using Bonferroni correction for the large data set, B - background signal, FC - fold change for the colour detection in control sample

- FC_1 - fold change for the second colour detection in the case (disease or disease type) sample, LS (Loop Status) - prevalent fluorescent signal between two colours threshold in competitive hybridisations, with -1 meaning signal is prevent in patient samples with corresponding fluorescent colour, when tested against the probe on the CGH array

- immediate genetic loci
- Prob Count Total - how many different location probes on the array were tested across that genetic locus
- Prob Count Sig - how many of them turned out to be significant in discriminating between case and control samples
- Hypergeometric Stat is statistics of enrichment of the locus with significant probes for disease detection
- FDR HyperG is the same statistics adjusted for the large data set by FDR (standard procedure)
- percentage of probes that turned to be significant in that locus
- logFC is logarithm of the fold change in array readout for that probe. Attention to the loci with high enrichment of significant probes helps selection of the top probes representing regulatory hubs with multiple inputs associated with disease providing markers with best coverage of for example network deregulation.

Preferred Aspects for Sample Preparation and Chromosome Interaction Detection

- Methods of preparing samples and detecting chromosome conformations are described herein. Optimised (non-conventional) versions of these methods can be used, for example as described in this section.
- Typically the sample will contain at least 2×10^5 cells. The sample may contain up to 5×10^5 cells. In one aspect, the sample will contain 2×10^5 to 5.5×10^5 cells
- Crosslinking of epigenetic chromosomal interactions present at the chromosomal locus is described herein. This may be performed before cell lysis takes place. Cell lysis may be performed for 3 to 7 minutes, such as 4 to 6 or about 5 minutes. In some aspects, cell lysis is performed for at least 5 minutes and for less than 10 minutes.
- Digesting DNA with a restriction enzyme is described herein. Typically, DNA restriction is performed at about 55°C to about 70°C , such as for about 65°C , for a period of about 10 to 30 minutes, such as about 20 minutes.

Preferably a frequent cutter restriction enzyme is used which results in fragments of ligated DNA with an average fragment size up to 4000 base pair. Optionally the restriction enzyme results in fragments of ligated DNA have an average fragment size of about 200 to 300 base pairs, such as about 256 base pairs. In one aspect, the typical fragment size is from 200 base pairs to 4,000 base pairs, such as 400 to 2,000 or 500 to 1,000 base pairs.

In one aspect of the EpiSwitch method a DNA precipitation step is not performed between the DNA restriction digest step and the DNA ligation step.

DNA ligation is described herein. Typically the DNA ligation is performed for 5 to 30 minutes, such as about 10 minutes.

The protein in the sample may be digested enzymatically, for example using a proteinase, optionally Proteinase K. The protein may be enzymatically digested for a period of about 30 minutes to 1 hour, for example for about 45 minutes. In one aspect after digestion of the protein, for example Proteinase K digestion, there is no cross-link reversal or phenol DNA extraction step.

In one aspect PCR detection is capable of detecting a single copy of the ligated nucleic acid, preferably with a binary read-out for presence/absence of the ligated nucleic acid.

Figure 5 shows a preferred method of detecting chromosome interactions.

Processes and Uses of the Invention

The process of the invention can be described in different ways. It can be described as a method of making a ligated nucleic acid comprising (i) *in vitro* cross-linking of chromosome regions which have come together in a chromosome interaction; (ii) subjecting said cross-linked DNA to cutting or restriction digestion cleavage; and (iii) ligating said cross-linked cleaved DNA ends to form a ligated nucleic acid, wherein detection of the ligated nucleic acid may be used to determine the chromosome state at a locus, and wherein preferably:

- the locus may be any of the loci, regions or genes mentioned in Table 5, and/or
- wherein the chromosomal interaction may be any of the chromosome interactions mentioned herein or corresponding to any of the probes disclosed in Table 5, and/or
- wherein the ligated product may have or comprise (i) sequence which is the same as or homologous to any of the probe sequences disclosed in Table 5; or (ii) sequence which is complementary to (ii).

The process of the invention can be described as a process for detecting chromosome states which represent different subgroups in a population comprising determining whether a chromosome interaction is present or absent within a defined epigenetically active region of the genome, wherein preferably:

- the subgroup is defined by presence or absence of prognosis, and/or
- the chromosome state may be at any locus, region or gene mentioned in Table 5; and/or
- the chromosome interaction may be any of those mentioned in Table 5 or corresponding to any of the probes disclosed in that table.

The process of the invention can be described as a method of making a ligated nucleic acid comprising (i) *in vitro* cross-linking of chromosome regions which have come together in a chromosome interaction; (ii) subjecting said cross-linked DNA to cutting or restriction digestion cleavage; and (iii) ligating said cross-linked cleaved DNA ends to form a ligated nucleic acid, wherein detection of the ligated nucleic acid may be used to determine the chromosome state at a locus, and wherein preferably:

- the locus may be any of the loci, regions or genes mentioned in Table 6, and/or
- wherein the chromosomal interaction may be any of the chromosome interactions mentioned herein or corresponding to any of the probes disclosed in Table 6, and/or
- wherein the ligated product may have or comprise (i) sequence which is the same as or homologous to any of the probe sequences disclosed in Table 6; or (ii) sequence which is complementary to (ii).

The process of the invention can be described as a process for detecting chromosome states which represent different subgroups in a population comprising determining whether a chromosome interaction is present or absent within a defined epigenetically active region of the genome, wherein preferably:

- the subgroup is defined by presence or absence of prognosis, and/or
- the chromosome state may be at any locus, region or gene mentioned in Table 6; and/or
- the chromosome interaction may be any of those mentioned in Table 6 or corresponding to any of the probes disclosed in that table.

The invention includes detecting chromosome interactions at any locus, gene or regions mentioned Table 5. The invention includes use of the nucleic acids and probes mentioned herein to detect chromosome interactions, for example use of at least 1, 5, 10, 20 or 50 such nucleic acids or probes to detect chromosome interactions. The nucleic acids or probes preferably detect chromosome interactions in at least 1, 5, 10, 20 or 50 different loci or genes. The invention includes detection of chromosome interactions using any of the primers or primer pairs listed in Table 5 or using variants of these primers as

described herein (sequences comprising the primer sequences or comprising fragments and/or homologues of the primer sequences).

The invention includes detecting chromosome interactions at any locus, gene or regions mentioned Table 6. The invention includes use of the nucleic acids and probes mentioned herein to detect chromosome interactions. The invention includes detection of chromosome interactions using any of the primers or primer pairs listed in Table 6 or using variants of these primers as described herein (sequences comprising the primer sequences or comprising fragments and/or homologues of the primer sequences).

When analysing whether a chromosome interaction occurs 'within' a defined gene, region or location, either both the parts of the chromosome which have together in the interaction are within the defined gene, region or location or in some aspects only one part of the chromosome is within the defined, gene, region or location.

Similarly the chromosome interactions of Tables 8 and 9 may be used in the processes and methods of the invention.

Use of the Method of the Invention to Identify New Treatments

Knowledge of chromosome interactions can be used to identify new treatments for conditions. The invention provides methods and uses of chromosomes interactions defined herein to identify or design new therapeutic agents, for example relating to therapy of prostate cancer or DLBCL.

Homologues

Homologues of polynucleotide / nucleic acid (e.g. DNA) sequences are referred to herein. Such homologues typically have at least 70% homology, preferably at least 80%, at least 85%, at least 90%, at least 95%, at least 97%, at least 98% or at least 99% homology, for example over a region of at least 10, 15, 20, 30, 100 or more contiguous nucleotides, or across the portion of the nucleic acid which is from the region of the chromosome involved in the chromosome interaction. The homology may be calculated on the basis of nucleotide identity (sometimes referred to as "hard homology").

Therefore, in a particular aspect, homologues of polynucleotide / nucleic acid (e.g. DNA) sequences are referred to herein by reference to percentage sequence identity. Typically such homologues have at least 70% sequence identity, preferably at least 80%, at least 85%, at least 90%, at least 95%, at least 97%, at least 98% or at least 99% sequence identity, for example over a region of at least 10, 15, 20, 30,

100 or more contiguous nucleotides, or across the portion of the nucleic acid which is from the region of the chromosome involved in the chromosome interaction.

For example the UWGCG Package provides the BESTFIT program which can be used to calculate homology and/or % sequence identity (for example used on its default settings) (Devereux *et al* (1984) Nucleic Acids Research 12, p387-395). The PILEUP and BLAST algorithms can be used to calculate homology and/or % sequence identity and/or line up sequences (such as identifying equivalent or corresponding sequences (typically on their default settings)), for example as described in Altschul S. F. (1993) J Mol Evol 36:290-300; Altschul, S, F *et al* (1990) J Mol Biol 215:403-10.

Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pair (HSPs) by identifying short words of length W in the query sequence that either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighbourhood word score threshold (Altschul *et al*, supra). These initial neighbourhood word hits act as seeds for initiating searches to find HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Extensions for the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLAST program uses as defaults a word length (W) of 11, the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1992) Proc. Natl. Acad. Sci. USA 89: 10915-10919) alignments (B) of 50, expectation (E) of 10, M=5, N=4, and a comparison of both strands.

The BLAST algorithm performs a statistical analysis of the similarity between two sequences; see e.g., Karlin and Altschul (1993) Proc. Natl. Acad. Sci. USA 90: 5873-5787. One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two polynucleotide sequences would occur by chance. For example, a sequence is considered similar to another sequence if the smallest sum probability in comparison of the first sequence to the second sequence is less than about 1, preferably less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

The homologous sequence typically differs by 1, 2, 3, 4 or more bases, such as less than 10, 15 or 20 bases (which may be substitutions, deletions or insertions of nucleotides). These changes may be measured

across any of the regions mentioned above in relation to calculating homology and/or % sequence identity.

Homology of a 'pair of primers' can be calculated, for example, by considering the two sequences as a single sequence (as if the two sequences are joined together) for the purpose of then comparing against the another primer pair which again is considered as a single sequence.

Arrays

The second set of nucleic acids may be bound to an array, and in one aspect there are at least 15,000, 45,000, 100,000 or 250,000 different second nucleic acids bound to the array, which preferably represent at least 300, 900, 2000 or 5000 loci. In one aspect one, or more, or all of the different populations of second nucleic acids are bound to more than one distinct region of the array, in effect repeated on the array allowing for error detection. The array may be based on an Agilent SurePrint G3 Custom CGH microarray platform. Detection of binding of first nucleic acids to the array may be performed by a dual colour system.

Therapeutic Agents (for example which are selected based on typing individuals or which are selected based on testing according to the invention)

Therapeutic agents are mentioned herein. The invention provides such agents for use in preventing or treating a disease condition in certain individuals, for example those identified by a process of the invention. This may comprise administering to an individual in need a therapeutically effective amount of the agent. The invention provides use of the agent in the manufacture of a medicament to prevent or treat a condition in certain individuals.

The formulation of the agent will depend upon the nature of the agent. The agent will be provided in the form of a pharmaceutical composition containing the agent and a pharmaceutically acceptable carrier or diluent. Suitable carriers and diluents include isotonic saline solutions, for example phosphate-buffered saline. Typical oral dosage compositions include tablets, capsules, liquid solutions and liquid suspensions. The agent may be formulated for parenteral, intravenous, intramuscular, subcutaneous, transdermal or oral administration.

The dose of an agent may be determined according to various parameters, especially according to the substance used; the age, weight and condition of the individual to be treated; the route of administration; and the required regimen. A physician will be able to determine the required route of administration and dosage for any particular agent. A suitable dose may however be from 0.1 to 100 mg/kg body weight such as 1 to 40 mg/kg body weight, for example, to be taken from 1 to 3 times daily.

The therapeutic agent may be any such agent disclosed herein, or may target any 'target' disclosed herein, including any protein or gene disclosed herein in any table (including Table 5 or 6). It is understood that any agent that is disclosed in a combination should be seen as also disclosed for administration individually.

5

Prostate Cancer Therapy

Prostate cancer treatments are recommended depending on the stage of disease progression. Radiotherapy, Hormone treatment and Chemotherapy are the three options that are often used in prostate cancer treatment. A single treatment or a combination of treatments may be used.

10

Chemotherapy

Chemotherapy is often used to treat prostate cancer that has invaded to other organs of the body (metastatic prostate cancer). Chemotherapy destroys cancer cells by interfering with the way they multiply. Chemotherapy does not cure prostate cancer, but it keeps it under control and reduce symptoms, therefore daily life is less effected.

15

Radiotherapy

This treatment may be used to cure localized and locally-advanced prostate cancer. Radiotherapy can also be used to slow the progression of metastatic prostate cancer and relieve symptoms. Patients may receive hormone therapy before undergoing chemotherapy to increase the chance of successful treatment. Hormone therapy may also be recommended after radiotherapy to reduce the chances of relapsing.

20

Hormone therapy

Hormone therapy is often used in combination with radiotherapy. Hormone therapy alone should not normally be used to treat localised prostate cancer in men who are fit and willing to receive surgery or radiotherapy. Hormone therapy can be used to slow the progression of advanced prostate cancer and relieve symptoms. Hormones control the growth of cells in the prostate. In particular, prostate cancer needs the hormone testosterone to grow. The purpose of hormone therapy is to block the effects of testosterone, either by stopping its production or by stopping patient's body to use testosterone.

25

30

Other treatments that may be used in prostate cancer therapy

- Radical prostatectomy
- High intensity focused ultrasound therapy

- Cryotherpay
- Brachytherapy
- Watchful waiting
- Trans-urethral resection of the prostate
- Treating advanced prostate cancer
- Steroid

DLBCL Therapy

The following four treatments may be used to treat DLBCL:

- Chemotherapy
- Radiotherapy
- Monocolonal antibody therapy
- Steroid therapy

Any of the above therapies may also be used to treat lymphoma.

Forms of the Substance Mentioned Herein

Any of the substances, such as nucleic acids or therapeutic agents, mentioned herein may be in purified or isolated form. They may be in a form which is different from that found in nature, for example they may be present in combination with other substance with which they do not occur in nature. The nucleic acids (including portions of sequences defined herein) may have sequences which are different to those found in nature, for example having at least 1, 2, 3, 4 or more nucleotide changes in the sequence as described in the section on homology. The nucleic acids may have heterologous sequence at the 5' or 3' end. The nucleic acids may be chemically different from those found in nature, for example they may be modified in some way, but preferably are still capable of Watson-Crick base pairing. Where appropriate the nucleic acids will be provided in double stranded or single stranded form. The invention provides all of the specific nucleic acid sequences mentioned herein in single or double stranded form, and thus includes the complementary strand to any sequence which is disclosed.

The invention provides a kit for carrying out any process of the invention, including detection of a chromosomal interaction relating to prognosis. Such a kit can include a specific binding agent capable of detecting the relevant chromosomal interaction, such as agents capable of detecting a ligated nucleic acid generated by processes of the invention. Preferred agents present in the kit include probes capable of

hybridising to the ligated nucleic acid or primer pairs, for example as described herein, capable of amplifying the ligated nucleic acid in a PCR reaction.

The invention provides a device that is capable of detecting the relevant chromosome interactions. The device preferably comprises any specific binding agents, probe or primer pair capable of detecting the chromosome interaction, such as any such agent, probe or primer pair described herein.

Detection Methods

In one aspect quantitative detection of the ligated sequence which is relevant to a chromosome interaction is carried out using a probe which is detectable upon activation during a PCR reaction, wherein said ligated sequence comprises sequences from two chromosome regions that come together in an epigenetic chromosome interaction, wherein said method comprises contacting the ligated sequence with the probe during a PCR reaction, and detecting the extent of activation of the probe, and wherein said probe binds the ligation site. The method typically allows particular interactions to be detected in a MIQE compliant manner using a dual labelled fluorescent hydrolysis probe.

The probe is generally labelled with a detectable label which has an inactive and active state, so that it is only detected when activated. The extent of activation will be related to the extent of template (ligation product) present in the PCR reaction. Detection may be carried out during all or some of the PCR, for example for at least 50% or 80% of the cycles of the PCR.

The probe can comprise a fluorophore covalently attached to one end of the oligonucleotide, and a quencher attached to the other end of the nucleotide, so that the fluorescence of the fluorophore is quenched by the quencher. In one aspect the fluorophore is attached to the 5' end of the oligonucleotide, and the quencher is covalently attached to the 3' end of the oligonucleotide.

Fluorophores that can be used in the methods of the invention include FAM, TET, JOE, Yakima Yellow, HEX, Cyanine3, ATTO 550, TAMRA, ROX, Texas Red, Cyanine 3.5, LC610, LC 640, ATTO 647N, Cyanine 5, Cyanine 5.5 and ATTO 680. Quenchers that can be used with the appropriate fluorophore include TAM, BHQ1, DAB, Eclip, BHQ2 and BBQ650, optionally wherein said fluorophore is selected from HEX, Texas Red and FAM. Preferred combinations of fluorophore and quencher include FAM with BHQ1 and Texas Red with BHQ2.

Use of the Probe in a qPCR Assay

Hydrolysis probes of the invention are typically temperature gradient optimised with concentration matched negative controls. Preferably single-step PCR reactions are optimized. More preferably a standard curve is calculated. An advantage of using a specific probe that binds across the junction of the

ligated sequence is that specificity for the ligated sequence can be achieved without using a nested PCR approach. The methods described herein allow accurate and precise quantification of low copy number targets. The target ligated sequence can be purified, for example gel-purified, prior to temperature gradient optimization. The target ligated sequence can be sequenced. Preferably PCR reactions are performed using about 10ng, or 5 to 15 ng, or 10 to 20ng, or 10 to 50ng, or 10 to 200ng template DNA. Forward and reverse primers are designed such that one primer binds to the sequence of one of the chromosome regions represented in the ligated DNA sequence, and the other primer binds to other chromosome region represented in the ligated DNA sequence, for example, by being complementary to the sequence.

10 *Choice of Ligated DNA Target*

The invention includes selecting primers and a probe for use in a PCR method as defined herein comprising selecting primers based on their ability to bind and amplify the ligated sequence and selecting the probe sequence based properties of the target sequence to which it will bind, in particular the curvature of the target sequence.

15 Probes are typically designed/chosen to bind to ligated sequences which are juxtaposed restriction fragments spanning the restriction site. In one aspect of the invention, the predicted curvature of possible ligated sequences relevant to a particular chromosome interaction is calculated, for example using a specific algorithm referenced herein. The curvature can be expressed as degrees per helical turn, e.g. 10.5° per helical turn. Ligated sequences are selected for targeting where the ligated sequence has a curvature propensity peak score of at least 5° per helical turn, typically at least 10°, 15° or 20° per helical turn, for example 5° to 20° per helical turn. Preferably the curvature propensity score per helical turn is calculated for at least 20, 50, 100, 200 or 400 bases, such as for 20 to 400 bases upstream and/or downstream of the ligation site. Thus in one aspect the target sequence in the ligated product has any of these levels of curvature. Target sequences can also be chosen based on lowest thermodynamic structure free energy.

Particular Aspects

In one aspect only intrachromosomal interactions are typed/detected, and no extrachromosomal interactions (between different chromosomes) are typed/detected.

In particular aspects certain chromosome interactions are not typed, for example any specific interaction mentioned herein (for example as defined by any probe or primer pair mentioned herein). In some aspects chromosome interactions are not typed in any of the genes mentioned herein.

The data provided herein shows that the markers are 'disseminating' ones able to differentiate cases and non-cases for the relevant disease situation. Therefore when carrying out the invention the skilled person will be able to determine by detection of the interactions which subgroup the individual is in. In one embodiment a threshold value of detection of at least 70% of the tested markers in the form they are associated with the relevant disease situation (either by absence or presence) may be used to determine whether the individual is in the relevant subgroup.

Screening method

The invention provides a method of determining which chromosomal interactions are relevant to a chromosome state corresponding to an prognosis subgroup of the population, comprising contacting a first set of nucleic acids from subgroups with different states of the chromosome with a second set of index nucleic acids, and allowing complementary sequences to hybridise, wherein the nucleic acids in the first and second sets of nucleic acids represent a ligated product comprising sequences from both the chromosome regions that have come together in chromosomal interactions, and wherein the pattern of hybridisation between the first and second set of nucleic acids allows a determination of which chromosomal interactions are specific to an prognosis subgroup. The subgroup may be any of the specific subgroups defined herein, for example with reference to particular conditions or therapies.

Publications

The contents of all publications mentioned herein are incorporated by reference into the present specification and may be used to further define the features relevant to the invention.

Specific Aspects

The EpiSwitch™ platform technology detects epigenetic regulatory signatures of regulatory changes between normal and abnormal conditions at loci. The EpiSwitch™ platform identifies and monitors the fundamental epigenetic level of gene regulation associated with regulatory high order structures of human chromosomes also known as chromosome conformation signatures. Chromosome signatures are a distinct primary step in a cascade of gene deregulation. They are high order biomarkers with a unique set of advantages against biomarker platforms that utilize late epigenetic and gene expression biomarkers, such as DNA methylation and RNA profiling.

EpiSwitch™ Array Assay

The custom EpiSwitch™ array-screening platforms come in 4 densities of, 15K, 45K, 100K, and 250K unique chromosome conformations, each chimeric fragment is repeated on the arrays 4 times, making the effective densities 60K, 180K, 400K and 1 Million respectively.

5 *Custom Designed EpiSwitch™ Arrays*

The 15K EpiSwitch™ array can screen the whole genome including around 300 loci interrogated with the EpiSwitch™ Biomarker discovery technology. The EpiSwitch™ array is built on the Agilent SurePrint G3 Custom CGH microarray platform; this technology offers 4 densities, 60K, 180K, 400K and 1 Million probes. The density per array is reduced to 15K, 45K, 100K and 250K as each EpiSwitch™ probe is presented as a quadruplicate, thus allowing for statistical evaluation of the reproducibility. The average number of potential EpiSwitch™ markers interrogated per genetic loci is 50, as such the numbers of loci that can be investigated are 300, 900, 2000, and 5000.

EpiSwitch™ Custom Array Pipeline

The EpiSwitch™ array is a dual colour system with one set of samples, after EpiSwitch™ library generation, labelled in Cy5 and the other of sample (controls) to be compared/ analyzed labelled in Cy3. The arrays are scanned using the Agilent SureScan Scanner and the resultant features extracted using the Agilent Feature Extraction software. The data is then processed using the EpiSwitch™ array processing scripts in R. The arrays are processed using standard dual colour packages in Bioconductor in R: Limma *. The normalisation of the arrays is done using the normalisedWithinArrays function in Limma * and this is done to the on chip Agilent positive controls and EpiSwitch™ positive controls. The data is filtered based on the Agilent Flag calls, the Agilent control probes are removed and the technical replicate probes are averaged, in order for them to be analysed using Limma*. The probes are modelled based on their difference between the 2 scenarios being compared and then corrected by using False Discovery Rate. Probes with Coefficient of Variation (CV) $\leq 30\%$ that are ≤ -1.1 or ≥ 1.1 and pass the $p \leq 0.1$ FDR p-value are used for further screening. To reduce the probe set further Multiple Factor Analysis is performed using the FactorMineR package in R.

* Note: LIMMA is Linear Models and Empirical Bayes Processes for Assessing Differential Expression in Microarray Experiments. Limma is an R package for the analysis of gene expression data arising from microarray or RNA-Seq.

The pool of probes is initially selected based on adjusted p-value, FC and CV <30% (arbitrary cut off point) parameters for final picking. Further analyses and the final list are drawn based only on the first two parameters (adj. p-value; FC).

Statistical Pipeline

5

EpiSwitch™ screening arrays are processed using the EpiSwitch™ Analytical Package in R in order to select high value EpiSwitch™ markers for translation on to the EpiSwitch™ PCR platform.

Step 1

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Probes are selected based on their corrected p-value (False Discovery Rate, FDR), which is the product of a modified linear regression model. Probes below p-value ≤ 0.1 are selected and then further reduced by their Epigenetic ratio (ER), probes ER have to be ≤ -1.1 or ≥ 1.1 in order to be selected for further analysis. The last filter is a coefficient of variation (CV), probes have to be below ≤ 0.3 .

Step 2

15

The top 40 markers from the statistical lists are selected based on their ER for selection as markers for PCR translation. The top 20 markers with the highest negative ER load and the top 20 markers with the highest positive ER load form the list.

Step 3

20

The resultant markers from step 1, the statistically significant probes form the bases of enrichment analysis using hypergeometric enrichment (HE). This analysis enables marker reduction from the significant probe list, and along with the markers from step 2 forms the list of probes translated on to the EpiSwitch™ PCR platform.

The statistical probes are processed by HE to determine which genetic locations have an enrichment of statistically significant probes, indicating which genetic locations are hubs of epigenetic difference.

25

The most significant enriched loci based on a corrected p-value are selected for probe list generation. Genetic locations below p-value of 0.3 or 0.2 are selected. The statistical probes mapping to these genetic locations, with the markers from step 2, form the high value markers for EpiSwitch™ PCR translation.

Array design and processing

Array Design

1. Genetic loci are processed using the SII software (currently v3.2) to:

- a. Pull out the sequence of the genome at these specific genetic loci (gene sequence with 50kb upstream and 20kb downstream)
- b. Define the probability that a sequence within this region is involved in CCs
- c. Cut the sequence using a specific RE
- 5 d. Determine which restriction fragments are likely to interact in a certain orientation
- e. Rank the likelihood of different CCs interacting together.
2. Determine array size and therefore number of probe positions available (x)
3. Pull out x/4 interactions.
4. For each interaction define sequence of 30bp to restriction site from part 1 and 30bp to restriction
- 10 site of part 2. Check those regions aren't repeats, if so exclude and take next interaction down on the list. Join both 30bp to define probe.
5. Create list of x/4 probes plus defined control probes and replicate 4 times to create list to be created on array
6. Upload list of probes onto Agilent Sure design website for custom CGH array.
- 15 7. Use probe group to design Agilent custom CGH array.

Array Processing

1. Process samples using EpiSwitch™ Standard Operating Procedure (SOP) for template production.
2. Clean up with ethanol precipitation by array processing laboratory.
- 20 3. Process samples as per Agilent SureTag complete DNA labelling kit - Agilent Oligonucleotide Array-based CGH for Genomic DNA Analysis Enzymatic labelling for Blood, Cells or Tissues
4. Scan using Agilent C Scanner using Agilent feature extraction software.

25 *EpiSwitch*™ biomarker signatures demonstrate high robustness, sensitivity and specificity in the stratification of complex disease phenotypes. This technology takes advantage of the latest breakthroughs in the science of epigenetics, monitoring and evaluation of chromosome conformation signatures as a highly informative class of epigenetic biomarkers. Current research methodologies deployed in academic environment require from 3 to 7 days for biochemical processing of cellular material in order to detect CCSs. Those procedures have limited sensitivity, and reproducibility; and furthermore, do not have the

30 benefit of the targeted insight provided by the *EpiSwitch*™ Analytical Package at the design stage.

EpiSwitch™ Array *in silico* marker identification

CCS sites across the genome are directly evaluated by the EpiSwitch™ Array on clinical samples from testing cohorts for identification of all relevant stratifying lead biomarkers. The EpiSwitch™ Array platform is used for marker identification due to its high-throughput capacity, and its ability to screen large numbers of loci rapidly. The array used was the Agilent custom-CGH array, which allows markers identified through the *in silico* software to be interrogated.

EpiSwitch™ PCR

Potential markers identified by *EpiSwitch™ Array* are then validated either by *EpiSwitch™ PCR* or DNA sequencers (i.e. Roche 454, Nanopore MinION, etc.). The top PCR markers which are statistically significant and display the best reproducibility are selected for further reduction into the final *EpiSwitch™ Signature Set*, and validated on an independent cohort of samples. *EpiSwitch™ PCR* can be performed by a trained technician following a standardised operating procedure protocol established. All protocols and manufacture of reagents are performed under ISO 13485 and 9001 accreditation to ensure the quality of the work and the ability to transfer the protocols. *EpiSwitch™ PCR* and *EpiSwitch™ Array* biomarker platforms are compatible with analysis of both whole blood and cell lines. The tests are sensitive enough to detect abnormalities in very low copy numbers using small volumes of blood.

Paragraphs showing embodiments of the invention

1. A process for detecting a chromosome state which represents a subgroup in a population comprising determining whether a chromosome interaction relating to that chromosome state is present or absent within a defined region of the genome; and
 - wherein said chromosome interaction has optionally been identified by a method of determining which chromosomal interactions are relevant to a chromosome state corresponding to the subgroup of the population, comprising contacting a first set of nucleic acids from subgroups with different states of the chromosome with a second set of index nucleic acids, and allowing complementary sequences to hybridise, wherein the nucleic acids in the first and second sets of nucleic acids represent a ligated product comprising sequences from both the chromosome regions that have come together in chromosomal interactions, and wherein the pattern of hybridisation between the first and second set of nucleic acids allows a determination of which chromosomal interactions are specific to the subgroup; and
 - wherein the subgroup relates to prognosis for prostate cancer and the chromosome interaction either:
 - (i) is present in any one of the regions or genes listed in Table 6; and/or

(ii) corresponds to any one of the chromosome interactions represented by any probe shown in Table 6, and/or

(iii) is present in a 4,000 base region which comprises or which flanks (i) or (ii);

or

5 - wherein the subgroup relates to prognosis for DLBCL and the chromosome interaction either:

a) is present in any one of the regions or genes listed in Table 5; and/or

b) corresponds to any one of the chromosome interactions represented by any probe shown in Table 5, and/or

c) is present in a 4,000 base region which comprises or which flanks (a) or (b).

10

2. A process according to paragraph 1 wherein:

- said prognosis for prostate cancer relates to whether or not the cancer is aggressive or indolent; and/or

- said prognosis for DLBCL relates to survival.

15 3. A process according to paragraph 1 or 2 wherein the subgroup relates to prostate cancer and a specific combination of chromosome interactions are typed:

(i) comprising all of the chromosome interactions represented by the probes in Table 6; and/or

(ii) comprising at least 1, 2, 3 or 4 of the chromosome interactions represented by the probes in Table 6; and/or

20 (iii) which together are present in at least 1, 2, 3 or 4 of the regions or genes listed in Table 6; and/or

(iv) wherein at least 1, 2, 3, or 4 of the chromosome interactions which are typed are present in a 4,000 base region which comprises or which flanks the chromosome interactions represented by the probes in Table 6.

25 4. A process according to paragraph 1 or 2 wherein the subgroup relates to DLBCL and a specific combination of chromosome interactions are typed:

(i) comprising all of the chromosome interactions represented by the probes in Table 5; and/or

(ii) comprising at least 10, 20, 30, 50 or 80 of the chromosome interactions represented by the probes in Table 5; and/or

30 (iii) which together are present in at least 10, 20, 30 or 50 of the regions or genes listed in Table 5; and/or

(iv) wherein at least 10, 20, 30, 50 or 80 chromosome interactions are typed which are present in a 4,000 base region which comprises or which flanks the chromosome interactions represented by the probes in Table 5.

5. A process according to paragraph 1 or 2 wherein the subgroup relates to DLBCL and a specific combination of chromosome interactions are typed:

(i) comprising all of the chromosome interactions shown in Table 7; and/or

5 (ii) comprising at least 1, 2, 5 or 8 of the chromosome interactions shown in Table 7.

6. A process according to any one of the preceding paragraphs wherein at least 10, 20, 30, 40 or 50, chromosome interactions are typed, and preferably at least 10 chromosome interactions are typed.

10 7. A process according to any one of the preceding paragraphs in which the chromosome interactions are typed:

- in a sample from an individual, and/or

- by detecting the presence or absence of a DNA loop at the site of the chromosome interactions, and/or

- detecting the presence or absence of distal regions of a chromosome being brought together in a
15 chromosome conformation, and/or

- by detecting the presence of a ligated nucleic acid which is generated during said typing and whose sequence comprises two regions each corresponding to the regions of the chromosome which come together in the chromosome interaction, wherein detection of the ligated nucleic acid is preferably by:

(i) in the case of prognosis of prostate cancer by a probe that has at least 70% identity to any of the specific
20 probe sequences mentioned in Table 6, and/or (ii) by a primer pair which has at least 70% identity to any primer pair in Table 6; or

(ii) in the case of prognosis of DLBCL a probe that has at least 70% identity to any of the specific probe sequences mentioned in Table 5, and/or (b) by a primer pair which has at least 70% identity to any primer pair in Table 5.

25

8. A process according to any one of the preceding paragraphs, wherein:

- the second set of nucleic acids is from a larger group of individuals than the first set of nucleic acids; and/or

- the first set of nucleic acids is from at least 8 individuals; and/or

30 - the first set of nucleic acids is from at least 4 individuals from a first subgroup and at least 4 individuals from a second subgroup which is preferably non-overlapping with the first subgroup; and/or

- the process is carried out to select an individual for a medical treatment.

9. A process according to any one of the preceding paragraphs wherein:

- the second set of nucleic acids represents an unselected group; and/or
- wherein the second set of nucleic acids is bound to an array at defined locations; and/or
- wherein the second set of nucleic acids represents chromosome interactions in least 100 different genes;

5 and/or

- wherein the second set of nucleic acids comprises at least 1,000 different nucleic acids representing at least 1,000 different chromosome interactions; and/or

- wherein the first set of nucleic acids and the second set of nucleic acids comprise at least 100 nucleic acids with length 10 to 100 nucleotide bases.

10

10. A process according to any one of the preceding paragraphs, wherein the first set of nucleic acids is obtainable in a process comprising the steps of: -

- (i) cross-linking of chromosome regions which have come together in a chromosome interaction;
- (ii) subjecting said cross-linked regions to cleavage, optionally by restriction digestion cleavage with an
- 15 enzyme; and
- (iii) ligating said cross-linked cleaved DNA ends to form the first set of nucleic acids (in particular comprising ligated DNA).

11. A process according to any one of the preceding paragraphs wherein said defined region of the genome:

20 (i) comprises a single nucleotide polymorphism (SNP); and/or

(ii) expresses a microRNA (miRNA); and/or

(iii) expresses a non-coding RNA (ncRNA); and/or

(iv) expresses a nucleic acid sequence encoding at least 10 contiguous amino acid residues; and/or

(v) expresses a regulating element; and/or

25 (vii) comprises a CTCF binding site.

12. A process according to any one of the preceding paragraphs which is carried out to determine whether a prostate cancer is aggressive or indolent which comprises typing at least 5 chromosome interactions as defined in Table 6.

30

13. A process according to any one of the preceding paragraphs which is carried out to determine prognosis of DLBCL which comprises typing at least 5 chromosome interactions as defined in Table 5.

14. A process according to any one of the preceding paragraphs which is carried out to identify or design a therapeutic agent for prostate cancer;

- wherein preferably said process is used to detect whether a candidate agent is able to cause a change to a chromosome state which is associated with a different level of prognosis;

5 - wherein the chromosomal interaction is represented by any probe in Table 6; and/or

- the chromosomal interaction is present in any region or gene listed in Table 6;

and wherein optionally:

- the chromosomal interaction has been identified by the method of determining which chromosomal interactions are relevant to a chromosome state as defined in paragraph 1, and/or

10 - the change in chromosomal interaction is monitored using (i) a probe that has at least 70% identity to any of the probe sequences mentioned in Table 6, and/or (ii) by a primer pair which has at least 70% identity to any primer pair in Table 6.

15. A process according to any one of preceding paragraphs 1 to 13 which is carried out to identify or design a therapeutic agent for DLBCL;

- wherein preferably said process is used to detect whether a candidate agent is able to cause a change to a chromosome state which is associated with a different level of prognosis;

- wherein the chromosomal interaction is represented by any probe in Table 5; and/or

- the chromosomal interaction is present in any region or gene listed in Table 5;

20 and wherein optionally:

- the chromosomal interaction has been identified by the method of determining which chromosomal interactions are relevant to a chromosome state as defined in paragraph 1, and/or

25 - the change in chromosomal interaction is monitored using (i) a probe that has at least 70% identity to any of the probe sequences mentioned in Table 5, and/or (ii) by a primer pair which has at least 70% identity to any primer pair in Table 5.

16. A process according to paragraph 14 or 15 which comprises selecting a target based on detection of the chromosome interactions, and preferably screening for a modulator of the target to identify a therapeutic agent for immunotherapy, wherein said target is optionally a protein.

30

17. A process according to any one of paragraphs 1 to 16, wherein the typing or detecting comprises specific detection of the ligated product by quantitative PCR (qPCR) which uses primers capable of amplifying the ligated product and a probe which binds the ligation site during the PCR reaction, wherein

said probe comprises sequence which is complementary to sequence from each of the chromosome regions that have come together in the chromosome interaction, wherein preferably said probe comprises:

an oligonucleotide which specifically binds to said ligated product, and/or

5 a fluorophore covalently attached to the 5' end of the oligonucleotide, and/or

a quencher covalently attached to the 3' end of the oligonucleotide, and optionally

said fluorophore is selected from HEX, Texas Red and FAM; and/or

said probe comprises a nucleic acid sequence of length 10 to 40 nucleotide bases, preferably a length of
10 20 to 30 nucleotide bases.

18. A process according to any one of paragraphs 1 to 17 wherein:

- the result of the process is provided in a report, and/or

- the result of the process is used to select a patient treatment schedule, and preferably to select a specific
15 therapy for the individual.

19. A therapeutic agent for use in a method of treating prostate cancer or DLBCL in an individual that has been identified as being in need of the therapeutic agent by a process according to any one of paragraphs 1 to 13 and 17.

20

The invention is illustrated by the following Examples:

Example 1

Using *EpiSwitch*[™] (chromosome conformation signature) markers

We have consistently observed highly disseminating *EpiSwitch*[™] markers with high concordance to the
25 primary and secondary affected tissues and strong validation results. *EpiSwitch*[™] biomarker signatures demonstrated high robustness and high sensitivity and specificity in the stratification of complex disease phenotypes.

The *EpiSwitch*[™] technology offers a highly effective means of screening; early detection; companion
30 diagnostic; monitoring and prognostic analysis of major diseases associated with aberrant and responsive gene expression. The major advantages of the OBD approach are that it is non-invasive, rapid, and relies on highly stable DNA based targets as part of chromosomal signatures, rather than unstable protein/RNA molecules.

CCSs form a stable regulatory framework of epigenetic controls and access to genetic information across the whole genome of the cell. Changes in CCSs reflect early changes in the mode of regulation and gene expression well before the results manifest themselves as obvious abnormalities. A simple way of thinking of CCSs is that they are topological arrangements where different distant regulatory parts of the DNA are brought in close proximity to influence each other's function. These connections are not done randomly; they are highly regulated and are well recognised as high-level regulatory mechanisms with significant biomarker stratification power.

Prognostic Stratification of Prostate Cancer

Markers were developed on the basis of retrospective annotations of Class I (low risk, indolent), Class II (intermediate), and Class III (aggressive high risk). The markers show robust classification of patients against healthy controls and also discriminate between Classes. The samples were from the United Kingdom.

To identify *EpiSwitch*TM biomarkers able to distinguish between blood from people with prostate cancer and healthy controls

A custom *EpiSwitch*TM Microarray investigation was initially used to identify and screen ~15,000 potential CCS over 425 genetic loci for discrimination between 8 Prostate Cancer (PCa) and 8 Control individuals. The top statistically significant markers were translated into Nested PCR assays and screened on a larger sample cohort of 24 PCa and 25 Healthy Control Samples. A classifier was developed using the top 5 CCS translated from the microarray which classified the PCa and Control samples with a Sensitivity and Specificity of 100% (95% CI – 86.2% to 100%) and 100% (95% CI – 86.7% to 100%) respectively.

Figure 1 shows a Principle Component Analysis of the top 5 markers on 49 samples of the development sample cohort.

The diagnostic classifier was used to classify an additional blinded independent cohort consisting of 24 PCa and 5 healthy control samples (n=29) with an accuracy of 83%. Further development of the *EpiSwitch*TM Prostate cancer assay was performed with an additional sample cohort of 95 PCa and 97 Controls (n= 192). This in turn was validated with a blinded sample cohort of 20 samples (10 PCa, 10 Controls). The results of this validation are shown in Table 1.

Table 1. Results for the classification of the blinded sample cohort (n=20)

		95% Confidence Interval (CI)
Sensitivity	80.0%	44.4%-97.5%
Specificity	80.0%	44.4%-97.5%
PPV	80.0%	44.4%-97.5%
NPV	80.0%	44.4%-97.5%

The most recent project in the PCA programme developed an alternative PCR format for the PCa diagnosis utilising hydrolysis probe based Realtime quantitative PCR (qPCR). The performance of the 6-marker model is shown in Table 2.

Table 2. Performance of 6 marker qPCR model

		95% Confidence Interval (CI)
Sensitivity	90.0%	73.47%-97.89%
Specificity	85.0%	62.11%-96.79%
PPV	90.0%	75.90%-96.26%
NPV	85.0%	65.60%-94.39%

Summary

The three independent blinded validations of the EpiSwitch™ PCa Diagnostic Signatures developed during the PCa diagnostic program, using US and UK samples of varying disease stages, achieves sensitivity and specificity of >80% for the diagnosis of Prostate Cancer. The Prostate Specific Antigen (PSA) Blood test which is the Gold Standard clinical assay for detecting PCa, which in itself relies on various other variables, typically has a sensitivity and specificity range of 32-68%. In addition a parallel research track has resulted in the development of an EpiSwitch™ assay to assess Prostate cancer prognosis to aid in the clinical management and treatment selection for individual patients diagnosed with PCA.

An additional custom EpiSwitch™ Microarray investigation was performed to identify and screen ~15,000 potential CCS over 426 genetic loci for discrimination between 8 Aggressive Prostate Cancer (Class 3) and 8 Indolent PCa (Class 1) patients, PCa class descriptions can be found in the Appendix. The top statistically significant markers were translated into Nested PCR assays and screened on a larger sample cohort of 42 Class 1, 25 Class 2 and 19 Class 3 PCa samples.

The top 6 statistically significant markers were used to develop a prognostic classifier to classify Class 1 (low risk) and Class 3 (high Risk) PCa. The performance of the classifier on an independent sample cohort of 42 Class 1 and 25 Class 3 samples (n=27) is shown in Table 3.

5

Table 3. Performance of 6 marker prognostic classifier (Class 1 vs Class 3)

		95% Confidence Interval (CI)
Sensitivity	80.0%	59.3%-93.17%
Specificity	92.86%	80.52%-98.5%
PPV	86.96%	66.41%-97.22%
NPV	88.64%	75.44%-96.21%

An alternative analysis found a further 6 markers that stratified between Class 2 and Class 3 PCa. The two classifiers share two markers, with each classifier also possessing 4 unique markers.

Figure 2 shows a VENN comparison of the two PCA prognostic classifiers.

The performance of the Class 2 vs Class 3 PCa classifier is shown in Table 4.

10

Table 4. Performance of 6 marker prognostic classifier (Class 2 vs Class 3) n =44

		95% Confidence Interval (CI)
Sensitivity	84.0%	63.92%-95.46%
Specificity	88.89%	65.29%-98.62%
PPV	91.30%	71.96%-98.93%
NPV	80.00%	56.34%-94.27%

Conclusions

The development of the diagnostic and prognostic biomarkers was achieved on multiple clinical sample cohorts. All conducted marker screening and selection was based on systemic, blood-based epigenetic changes as monitored through chromosome conformation signatures in patients with different stages of Prostate Cancer (stage 1 to 3) against healthy controls (diagnostic application), as well as patients with aggressive, high risk category 3 against indolent, low risk category 1 prostate cancers (prognostic application), or intermediate risk category 2.

15

The results of stratification development for PCa vs healthy controls showed sensitivity and specificity up to > 80% in the testing cohort and a series of blind validations. Stratification of high-risk category 3 vs low risk category 1 PCa showed sensitivity up to 80% and specificity up to 92% on cohorts of up to 67 samples, while stratification of high-risk category 3 vs intermediate-risk category 2 showed sensitivity up to 84%, and specificity up to 88% on cohorts of up to 44 samples.

Appendix

Low risk – Category 1

Localised prostate cancer is classified as low risk if

- 10 PSA level is less than 10 ng per ml, and
 Gleason score is no higher than 6, and
 The T stage is between T1 and T2a

Intermediate risk – Category 2

- 15 Localised prostate cancer is classed as intermediate risk if you have at least one of the following
 PSA level is between 10 and 20 ng/ml
 Gleason score is 7
 The T stage is T2b

High risk – Category 3

- 20 Localised prostate cancer is classed as high risk if you have at least one of the following
 PSA level is more than 20 ng/ml
 Gleason score is between 8 and 10
 The T stage is T2c, T3 or T4
- 25 If the cancer is T3 or T4 stage, this means it has broken through the outer fibrous covering (capsule) of the prostate gland, and so it is classed as locally advanced prostate cancer.

Example 2. Identifying Markers for DLBCL

Summary

- 30 This relates to identification of major groups of poor and good prognosis patients for subsequent selection of treatments (i.e. R-CHOP). The biomarkers have been developed on the basis of retrospective overall survival. Normally, patients are classified by biopsy based gene expression standards like Nanostring or

Fluidigm, according to diseases subtypes such as ABC (poor prognosis) or GCB (better prognosis). However not all patients could be classified as ABC or GCB (the so called Type III, or Unclassified patients). We identified biomarkers to provide classification for prognosis of survival at the baseline, before treatments, irrespective of ABC or GCB standard classification.

5

Identification of Markers

DLBCL shows distinct differences in patients survival (poor vs good prognosis) and is characterised by a number of molecular readouts into subtypes. Various subtypes are also treated differently in current clinical practice. This, for example includes combination of Rituximab and CHOP combination on chemotherapy. There are various approaches.

10

Currently practiced molecular readouts are based on gene expression profiling by arrays, performed on biological materials obtained by direct biopsies. Those include Nanostring and Fluidigm array-based tests for extreme types of ABC and GCB. ANC subtype normally is associated with poor prognosis. Not every patient could be classified as ABC or GCB, a number of patients remain unclassified (or Type III) in terms of the established gene expression profiles and any association with prognosis of poor survival. We built systemic biomarkers that will directly classify patients for poor vs good prognosis, irrespective of transcriptional gene expression profiling by other modalities.

15

Step one: We used the Episwitch screening array to compare the epigenetic profiles on groups of cell lines representing poor prognosis and good prognosis of survival for DLBCL. This allows identification of array based markers and designing of nested PCR primers to use for the same targets in PCR format.

20

Step two: We used top 10 nested PCR based markers read on baseline blood samples from 57-58 unclassified DLBCL patients with known retrospective survival annotations. Table 6 provides details for the markers, the final signature, and the stated performance by the classifier model.

25

Our work shows how base line calls on these patients for poor/good prognosis compared against the clinical survival data. This is a Cox estimate of hazard ratio, i.e. our baseline classification into poor prognosis shows higher probabilities for being in a poor prognosis survival group, rather than a good prognosis group by the clinical post factum annotation, with a particular value >1 . The latter is of particular value and interest for clinical teams in trial designs.

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Detailed Writeup

Diffuse large B-cell lymphoma (DLBCL) is the most common type of non-Hodgkin's lymphoma in adults. It can occur anytime between adolescence and old age, affects 7-8 people per 100,000 in the US annually, although the incidence rate increases with age. Gene expression profiling has revealed two major types of DLBCL – germinal centre B-cell like (GCB) and activated B-cell like (ABC). GCB DLBCL arises from secondary lymphoid organs e.g. lymph nodes, where naïve B-cells do not stop dividing after infection is cleared. ABC DLBCL is thought to begin in a subset of B-cells which are ready to leave the germinal centre and become plasma cells i.e. plasmablastic B-cells, but the reality is more complicated with different forms of DLBCL occurring through the whole B-cell lifecycle.

The different subtypes have varying prognoses with a 5-year survival rate of 60% for GCB DLBCL, but only 35% for ABC DLBCL. Each of the subtypes is characterized by differential gene expression. In GCB DLBCL the transcriptional repressor BCL6 is often over-expressed whereas in ABC DLBCL the NF- κ B pathway is often found to be constitutively activated. There is also a third type of DLBCL called type III which is currently less well understood but it is thought to have a gene expression profile situated between the two main types.

Current diagnostic methods involve excisional biopsy of the affected lymph node followed by immunohistochemistry (IHC). At present, treatment procedures for DLBCL are the same regardless of the subtype. Since the pathogenesis, treatment responses, and outcomes of the various subtypes differ enormously there remains a need to develop a robust, non-invasive assay to distinguish between the subtypes in order to assist in the development of differentiated treatment strategies. Although much research has been carried out to find predictive and prognostic biomarkers for DLBCL there is no consensus on a single test that can be used to distinguish between the subtypes.

To identify *EpiSwitch*[™] biomarkers able to distinguish between the different subtypes of DLBCL in blood from patients with DLBCL

We used the *EpiSwitch*[™] array platform to look at DLBCL cell lines and blood samples and identify biomarkers that were absent in healthy control patients, before confirming these biomarkers in a patient cohort consisting of 30 ABC, 30 GCB and 10 healthy control samples.

***EpiSwitch™* Array**

The *EpiSwitch™* custom array allows the screening of several thousand possible CCS's, with probes designed using pattern recognition software. Different long-range chromosomal interactions captured by *EpiSwitch™* technology reflect the epigenetic regulatory framework imposed on the loci of interest and correspond to individual different inputs from signalling pathways contributing to the co-regulation of these loci. Altogether, the combination of the different inputs modulates gene expression. Identification of an aberrant or distinct chromosomal conformation signature under specific physiological condition offers important evidence for specific contribution to deregulation before all the input signals are integrated in the gene expression profile.

Using data from several sources 98 genetic loci were selected for analysis with the proprietary software and probes for 13,332 potential chromosomal conformations were tested. Looking at one locus does not equate to looking at one marker, as there may be one, multiple, or no high-order epigenetic chromosome conformation markers in a specific locus. After manufacture cell lines and blood samples from DLBCL patients and healthy controls were processed using the *EpiSwitch* protocol, labelled, and hybridized to the array.

Samples for Diagnostic development

We used 16 cell lines, which corresponded to different subtypes, and with different levels of confidence in subtyping. The most definite ABC and GCB subtyped cell lines were used for analysis. In addition, blood samples from four DLBCL patients and 11 healthy controls were used. After biomarker identification in part one 60 further samples were provided to OBD, consisting of 30 ABC and 30 GCB blood samples, well characterised by Fluidigm testing, and this was supplemented by ten healthy control samples provided by OBD.

Results**Array analysis**

72 chromosome signature sites from the microarray were chosen to be screened based on two criteria:

- Their ability to stratify between ABC and GCB cells (highABC_highGCB)
- and/or
- A low CV value (a median value of the 5 arrays analyzed, High ABC v High GCB, DLBCL1 v Healthy Control, DLBCL2 v Healthy Control, DLBCL3 v Healthy Control and DLBCL4 v Healthy Control)

Translation of array to *EpiSwitch*TM PCR platform

- 5 After analysis of the sequence surrounding the probes of interest from the array 69 sets of primers were designed to interrogate the chromosome signature sites. These were then tested on pooled DLBCL blood samples, and of these 49 met the OBD criteria for PCR products for use in assays.

Each of these 49 potential markers were then tested on six DLBCL cell lines - three of which were ABC and
10 three of which were GCB. The cell lines used were those which were most confident were ABC or GCB, due to the same categorisation being found using multiple different identification methods. This allowed for the markers to be selected that were most useful in differentiating ABC and GCB cell subtypes. 28 *EpiSwitch*TM markers were identified for use with the PCR platform that were consistent with the *EpiSwitch*TM microarray results. In addition, the potential markers were also tested against four DLBCL
15 patients and pooled healthy controls to identify those that were present in DLBCL patients, but absent in healthy controls. 21 of the 28 *EpiSwitch*TM markers were absent in healthy control samples, but present in DLBCL samples such that it could be used as a marker of DLBCL, as well as for subtyping.

Sample Testing

- 20 The 21 markers that translated well into the *EpiSwitch*TM PCR platform were then tested amongst the 70 patient blood sample cohort. Initially, each marker was tested in six new ABC samples, and six new GCB samples, and the 21-marker set narrowed down to ten markers that showed the greatest difference. These ten markers were then tested on the remaining 24 ABC, 24 GCB and ten healthy control samples.
- 25 Each of the markers was then subjected to analysis of its power to differentiate subgroups, its collinearity with other markers, and also its ability to differentiate healthy from DLBCL. A subset of six of the markers was identified that provided the maximum possible information and these are markers in the ANXA11, IFNAR, MAP3K7, MEF2B, NFATc1, and TNFRS13C loci. Figure 3 shows the ability of these markers to differentiate the different groups of samples on a PCA plot. This six-marker panel is able to clearly
30 differentiate healthy control patients from DLBCL patients, a key characteristic of any blood-based assay for DLBCL.

Figure 3 shows a PCA plot of 60 DLBCL and 10 healthy patients based on the six EpiSwitch™ marker binary data. Samples are characterized as ABC subtype or GCB subtype by Fluidigm data, and the healthy controls are also shown.

5 Classification: Identification of ABC and GCB subtypes within DLBCL patient cohort (60 samples)

Classification was performed using the logistic regression classifier with 5-fold cross-validation, and the following results were achieved. The following results were achieved in cross-validation:

ABC subtype	83.3% (95% CI – 65.3% to 94.3%)
10 GCB subtype	83.3% (95% CI – 65.3% to 94.3%)

In addition, the resultant six-marker logistic classifier model was tested on 50 permutations of the 60-patient data set. The data was randomized each time and the accuracy statistics were calculated with a ROC curve. An area under the curve (AUC) of 0.802 and p-value 0.0000037 (H_0 = "The AUC is equal to
15 0.5"), suggests that the model is accurate and performing efficiently.

Conclusions

In this study we have demonstrated the power of their *EpiSwitch*™ technology to provide answers to difficult clinical questions, particularly the differentiation of the ABC and GCB subtypes of DLBCL. Using
20 high-throughput array methods, and translation to the simple and cost-effect PCR platform more than 13,000 potential CCS's have been tested and refined to a six marker panel for DLBCL subtype differentiation. This panel was able to distinguish DLBCL patients from healthy controls, and was able to predict subtype accurately 83.3% of the time. This test also has greater than 80% concordance for class assignment between *EpiSwitch*™ (whole blood based), LPS (cell of origin, tissue) and Fluidigm (cell of
25 origin, tissue)

EpiSwitch™ technology detects changes in long-range intergenic interactions – chromosomal conformation signatures, which result in changes in the epigenetic status and modulation of the expression mode of key genes involved in the pathogenesis of disease. The diagnostic procedure based
30 on *EpiSwitch*™ technology is a simple and rapid technique that can be transferred to other laboratories. The test consists of several molecular biology reactions, followed by detection with nested PCR. The test does not require complicated procedures and can be performed in any laboratory that runs PCR-based assays.

Example 3

Further work was performed on canines. One aim was to investigate markers for aiding in the initial diagnosis of suspected lymphoma to inform veterinary clinicians on the requirements for performing follow up biopsies. In this study, the top 75 EpiSwitch Microarray DLBCL markers (previously identified) are translated from the Human Genome Build (Grch37) to the current canine genome. In total 38 Canine samples (consisting of the 19 patients with likely lymphoma and 19 matched control samples) were screened using all 75 DLBCL markers. To carry out this work the following were performed:

- Based on 75 human DLBCL markers (associated with specific genes) orthologues in Dog genome (CanFam3.1) identified and genetic loci extracted from Biomart.
- EpiSwitch™ software run to identify potential interactions in these loci
- Primer design software and other filters added to reduce list to 75 markers for investigation.

The work and results are shown in Figures 6 to 16 and in Tables 8 and 9.

Example 4. Further Work on Prostate Cancer

Current diagnostic blood tests for prostate cancer (PCa) are unreliable for the early stage disease, resulting in numerous unnecessary prostate biopsies in men with benign disease and false reassurance of negative biopsies in men with PCa. Predicting the risk of PCa is pivotal for making an informed decision on treatment options as the five-year survival rate in the low-risk group is more than 95% and most men would benefit from less invasive therapy. Three-dimensional genome architecture and chromosome structures undergo early changes during tumorigenesis both in tumour and in circulating cells and can serve a disease biomarker.

In this prospective study we have performed chromosome conformation screening for 14,241 chromosomal loops in the loci of 425 cancer related genes in whole blood of newly diagnosed, treatment naïve PCa patients (n=140) and non-cancer controls (n=96).

Our data show that peripheral blood mononuclear cells (PBMCs) from PCa patients acquired specific chromosome conformation changes in the loci of *ETS1*, *MAP3K14*, *SLC22A3* and *CASP2* genes. Blind testing on an independent validation cohort yielded PCa detection with 80% sensitivity and 80% specificity. Further analysis between PCa risk groups yielded prognostic validation sets consisting of *BMP6*, *ERG*, *MSR1*, *MUC1*, *ACAT1* and *DAPK1* genes for high-risk category 3 vs low-risk category 1 and *HSD3B2*, *VEGFC*,

APAF1, *MUC1*, *ACAT1* and *DAPK1* genes for high-risk category 3 vs intermediate-risk category 2, which had high similarity to conformations in primary prostate tumours. These sets achieved 80% sensitivity and 92% specificity stratifying high-risk category 3 vs low risk category 1 and 84% sensitivity and 88% specificity stratifying high risk category 3 vs intermediate risk category 2 disease.

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Our results demonstrate specific chromosome conformations in the blood of PCa patients that allow PCa diagnosis and prognosis with high sensitivity and specificity. These conformations are shared between PBMCs and primary tumours. It is possible that these epigenetic signatures may potentially lead to development of a blood-based PCa diagnostic and prognostic tests.

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Introduction

In the Western world prostate cancer (PCa) is now the most commonly diagnosed non-cutaneous cancer in men and is the second leading cause of cancer-related death. Many men as young as 30 show evidence of histological PCa, most of which is microscopic and possibly will never show clinical manifestations. For the diagnosis and prognosis, prostate specific antigen (PSA), an invasive needle biopsy, Gleason score and disease stage are used. In a large multicentre study of 2,299 patients, a 12-site biopsy scheme outperformed all other schemes, with an overall PCa detection rate of only 44.4%.

The only available blood test for PCa in widespread clinical use involves measuring circulating levels of PSA (21% sensitivity and 91% specificity), however, the prostate size, benign prostatic hyperplasia and prostatitis may also increase PSA levels. At the current 4.0 ng/ml cut-off limit, only 20% of all PCa patients are being detected. In early PCa, PSA testing is not specific enough to differentiate between early-stage invasive cancers and latent, non-lethal tumours that might otherwise have remained asymptomatic during a man's lifetime. In advanced PCa, PSA kinetics are used as a clinical surrogate endpoint for outcome. However, while they do give a general prognosis they lack specificity for the individual. A number of more specific blood tests are emerging for PCa detection including 4K blood test (AUC 0.8) and PHI blood test (90% sensitivity, 17% specificity). PSA levels, disease stage and Gleason score are used to establish the severity of PCa and stratify patients to risk groups. To date, there is no prognostic blood test available that allows differentiation between low- and high-risk PCa.

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There are multiple genetic changes associated with PCa, including mutations in p53 (up to 64% of tumours), p21 (up to 55%), p73 and MMAC1/PTEN tumour suppressor genes, but these mutations do not explain all the observed effects on gene regulation. Epigenetic mechanisms involving dynamic and multi-

layered chromosomal loop interactions are powerful regulators of gene expression. Chromosome conformation capture (3C) technologies allow these signatures to be recorded. In this study, we used the EpiSwitch™ assay to screen for, define and evaluate specific chromosome conformations in the blood of PCa patients and to identify loci with potential to act as diagnostic and prognostic markers.

5

Methods

A total of 140 PCa patients and 96 controls were recruited, in two cohorts. Cohort 1: men with (n=105) or without (n=77) PCa diagnosis attending a urology clinic were prospectively recruited from October 2010 through September 2013. Cohort 2: Patients' samples (19 controls and 35 PCa) obtained from the USA.

10 Upon recruitment, a single blood sample (5 ml) was collected from PCa patients using the current practice for needle and blood collection methods into the BD Vacutainer® plastic EDTA tubes. Blood samples were passively frozen and stored at -80°C until processed. Prostate tumour samples were obtained from previously recruited patients (n=5) that subsequently underwent radical prostatectomy. Patient clinical characteristics are shown in Table 17.

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The primary endpoint of this study was to detect changes in chromosomal conformations in PBMCs from PCa patients in comparison to controls. Therefore, all treatment naïve PCa patients were eligible for this study irrespective of grade, stage and PSA levels. Patients that had previous chemotherapy or patients with other cancers were excluded from this study. PCa diagnosis was established as per clinical routine and patients were assigned to appropriate treatment. For prognostic study (secondary endpoint), patients were stratified according to the relevant NCCN risk groups (**Table 10**). No follow up study was conducted.

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Based on the preliminary findings in melanoma, an *a priori* power analysis was performed using the pwr.t.test function in the R package pwr. Testing indicated 15 patients per group should be sufficient to detect correlation between variables ($\beta=5\%$ probability type II error, significance level; 95% power; 50% confidence interval and 40% standard deviation).

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EpiSwitch™ technology platform pairs high resolution 3C results with regression analysis and a machine learning algorithm to develop disease classifications. To select epigenetic biomarkers that can diagnose cancers, samples from patients suffering from cancer, in comparison to healthy (control) samples were screened for statistically significant differences in conditional and stable profiles of genome architecture. The assay is performed on a whole blood sample by first fixing chromatin with formaldehyde to capture intrachromatin associations. The fixed chromatin is then digested into fragments with TaqI restriction

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enzyme, and the DNA strands are joined favouring cross-linked fragments. The cross-links are reversed and polymerase chain reactions (PCR) performed using the primers previously established by the EpiSwitch™ software. EpiSwitch™ was used on blood samples in a three-step process to identify, evaluate, and validate statistically-significant differences in chromosomal conformations between PCa patients and healthy controls (**Figure 17**). For the first step, sequences from 425 manually curated PCa-related genes (obtained from the public databases (www.ensembl.org)) were used as templates for this computational probabilistic identification of regulatory signals involved in chromatin interaction (**Table 18**). A customized CGH Agilent microarray (8x60k) platform was designed to test technical and biological repeats for 14,241 potential chromosome conformations across 425 genetic loci. Eight PCa and eight control samples were competitively hybridized to the array, and differential presence or absence of each locus was defined by LIMMA linear modelling, subsequent binary filtering and cluster analysis. This initially revealed 53 chromosomal interactions with the ability to best discriminate PCa patients from controls (**Figure 17**).

For the second evaluation stage, the 53 biomarkers selected from the array analysis were translated into EpiSwitch™ PCR based-detection probes and used in multiple rounds of biomarker evaluation. PCR primers were selected according to their ability to distinguish between PCa and healthy controls (n=6 in each group). The identity of PCR products generated using nested primers was confirmed by direct sequencing. Accordingly, the 53 biomarkers selected were reduced to 15 markers after the initial statistical analysis and finally a five-marker signature (**Table 11**). This selected chromosomal-conformation signature-biomarker set was then tested on a known cohort (n=49). Additionally, the five-marker signature developed from EpiSwitch™ PCR evaluation of array marker leads was tested on an independent blind validation cohort of 29 samples which were combined with the known 49 samples tested earlier (total 78 samples). Principal component analysis was also used to determine abundance levels and to identify potential outliers (**Figure 18**).

For the last step, to further validate the chromosome conformation signature used to inform PCa diagnosis, the five-marker set was tested on a blinded, independent (n=20) cohort of blood samples. The results were analysed using Bayesian Logistic modelling, p-value null hypothesis ($\Pr(N|z)$) analysis, Fisher-Exact P test and Glmnet (**Table 12**). The sample cohort sizes in the five-marker signature study were progressively increased to enable selection of the optimal markers for discriminating PCa samples from healthy controls. Cohort sizes were expanded to 95 PCa and 96 healthy control samples. Data analysis and presentation were performed in accordance with CONSORT recommendations. All measurements were

performed in a blinded manner. STARD criteria have been used to validate the analytical procedures. A similar three-step approach was followed for the identification of prognostic markers (**Table 13**).

Sequence specific oligonucleotides were designed around the chosen sites for screening potential markers by nested PCR using Primer3. All PCR amplified samples were visualized by electrophoresis in the LabChip GX, using the LabChip DNA 1K Version2 kit (Perkin Elmer, Beaconsfield, UK) and internal DNA marker was loaded on the DNA chip according to the manufacturer's protocol using fluorescent dyes. Fluorescence was detected by laser and electropherogram read-outs translated into a simulated band on gel picture using the instrument software. The threshold we set for a band to be deemed positive was 30 fluorescence units and above.

Primary tumour samples were obtained from biopsies of selected patients (n=5). The pulverized tissue samples were incubated in 0.125% collagenase at 37°C with gentle agitation for 30 minutes. The resuspended cells (250ul) were then centrifuged at 800g for 5 minutes at room temperature in a fixed arm centrifuge, supernatant removed, and the pellets resuspended in phosphate-buffered saline (PBS). Primary tumours and matching blood samples were analysed for the presence of the six-markers set for categories 3 vs 1 and 3 vs 2 at a fixed range of assay sensitivity (dilution factor 1:2). When matching PCR bands of the correct size were detected, a score of 1 was assigned, detection of no band was assigned a score of 0 (**Table 14**).

We have applied a stepwise diagnostic biomarker discovery process using EpiSwitch™ technology as described in methods. A customized CGH Agilent microarray (8x60k) platform was designed to test technical and biological repeats for 14,241 potential chromosome conformations across 425 genetic loci (**Table 18**) in eight PCa and eight control samples (**Figure 17**). The presence or absence of each locus was defined by LIMMA linear modelling, subsequent binary filtering and cluster analysis. In the second evaluation stage, nested PCR was used for the 53 selected biomarkers further reducing them to 15 markers and finally to a five-marker signature (**Figure 17**). This distinct chromosome conformational disease classification signature for PCa comprised of chromosomal interactions in five genomic loci: *ETS proto-oncogene 1, transcription factor (ETS1)*, *mitogen-activated protein kinase kinase kinase 14 (MAP3K14)*, *solute carrier family 22 member 3 (SLC22A3)* and *caspase 2 (CASP2)* (**Table 11**). The genomic locations of specific chromosomal loops in *ETS1*, *MAP3K14*, *SLC22A3* and *CASP2* genes in the chromosome conformation signature (**Table 11**) were mapped on their relative chromosomes. The two genomic sites that corresponded to the junction of each chromosome conformation signature locus for *ETS1*, *MAP3K14*,

SLC22A3 and *CASP2* genes were mapped on chromosome 11 from 128,260,682 to 128,537,926; chromosome 17 from 43,303,603 to 43,432,282; chromosome 6 from 160,744,233 to 160,944,757 and chromosome 7 from 142,935,233 to 143,008,163. Circos plots of *ETS1*, *MAP3K14*, *SLC22A3* and *CASP2* chromosome conformation signature markers showing the chromosomal loops were produced.

- 5 Principal component analysis for the five-markers was used to determine abundance levels and to identify potential outliers. This analysis was applied to 78 samples containing two groups. First group, 49 known samples (24 PCa and 25 healthy controls) combined with a second group of 29 samples including, 24 PCa samples and 5 healthy control samples (**Figure 18**). The final training set was built using 95 PCa and 96 control samples and then tested on an independent blinded validation cohort of 20 samples (10 controls and 10 PCa). The sensitivity and specificity for PCa detection using chromosomal interactions in five genomic loci were 80% (CI 44.39% to 97.48%) and 80% (CI 44.39% to 97.48%), respectively (**Table 12**).

- To select epigenetic biomarkers that can stratify PCa, the samples from PCa patients categorised into risk group categories 1-3 (low, intermediate and high, respectively, **Table 10**) were screened for statistically significant differences in conditional and stable profiles of genome architecture. EpiSwitch™ was used on blood samples in a three-step process to identify, evaluate, and validate statistically-significant differences in chromosomal conformations between PCa patients at different stages of the disease (**Figure 17**). For the first step, the array used covered 425 genetic loci, with testing probes for the total of 14,241 potential chromosomal conformations. Patients with high-risk PCa category 3 were compared to low-risk category 1 or intermediate-risk category 2. In total, 181 potential stratification marker leads for PCR evaluation were identified using enrichment statistics (**Table 19**). The top 70 top markers were then taken to the next stage of PCR detection for further evaluation of stratification of high-risk category 3, vs low-risk category 1 patient samples and finally a six-marker set for high category 3 vs low category 1 was established (**Table 13**). The best markers were identified using Chi-square and then built into a classifier on a testing set of category 1 (n=21) and category 3 (n=19). An independent cohort of category 1 (n=21) and category 3 (n=6) which were not used for any marker reduction were then used for first round of blind validation. Similarly, a six-marker set was evaluated for high-risk category 3 vs intermediate-risk category 2 on a testing set of category 3 and category 2 including, 25 and 19 samples, respectively. An independent cohort of category 2 and category 3 (n=6 in each group) which were not used for any marker reduction were then used for first round blind validation.

For the last step, to further validate the chromosome conformation signature used to inform PCa prognosis, the six-marker set for high-risk category 3 vs low-risk category 1 was tested on a larger, more

representative cohort. The original blind cohort was expanded to 67 samples, including 40 samples used in marker reduction (**Table 15**). Similarly, the six-marker set for high-risk category 3 vs intermediate-risk category 2 was tested on a on a larger, more representative cohort. The original blind cohort was expanded to 43 samples (**Table 16**).

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A six-marker set for category 3 vs category 1 was established. This set contained bone morphogenetic protein 6 (*BMP6*), *ETS transcription factor ERG* (*ERG*), macrophage scavenger receptor 1 (*MSR1*), *mucin 1* (*MUC1*), *acetyl-CoA acetyltransferase 1* (*ACAT1*) and *death-associated protein kinase 1* (*DAPK1*) genes (**Table 13**). Six-biomarkers were identified for high-risk category 3 vs intermediate-risk category 2, including *hydroxy-delta-5-steroid dehydrogenase, 3 beta- and steroid delta-isomerase 2* (*HSD3B2*), *vascular endothelial growth factor C* (*VEGFC*), *apoptotic peptidase activating factor 1* (*APAF1*), *MUC1*, *ACAT1* and *DAPK1*. Notably, the last three-biomarkers (*MUC1*, *ACAT1* and *DAPK1*) were common between categories 1 vs 3 and 3 vs 2 (**Table 13**). Stratification of high-risk category 3 vs low-risk category 1 PCa using chromosomal interactions in six genomic loci showed sensitivity of 80% (CI 59.30% to 93.17%) and specificity of 92% (CI 80.52% to 98.50%) in the blind cohort of 67 samples (**Table 15**). Similarly, the six-marker set for high-risk category 3 vs intermediate-risk category 2 was tested on a on a larger, more representative cohort of 43 samples demonstrating sensitivity of 84% (CI 63.92% to 95.46%), and specificity of 88% (CI 65.29% to 98.62%) (**Table 16**).

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Using five matching peripheral blood and primary tumour samples, we have compared the epigenetic markers identified in peripheral circulation (**Table 13**) to the tumour tissue. Our results showed that a number of deregulation markers detected in the blood as part of stratifying signatures for category 1 vs 3 and category 2 vs 3 could be detected in the tumour tissue (**Table 14**). This demonstrates that the chromosome interactions that can be detected systemically could be detected under same conditions in the primary site of tumorigenesis.

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Timely diagnosis of prostate cancer is crucial to reducing mortality. The European randomised study of screening for PCa has shown significant reduction in PCa mortality in men who underwent routine PSA screening. Total screening, however, leads to overdiagnosis of clinically insignificant disease and new less invasive tests capable of discriminating low- from high-risk disease are urgently required.

Our epigenetic analysis approach provides a potentially powerful means to address this need. The binary nature of the test (the chromosomal loop is either present or not) and the enormous combinatorial power

($>10^{10}$ combinations are possible with ~50,000 loops screened) may allow creating signatures that accurately fit clinically well-defined criteria. In PCa that would be discerning low-risk vs high-risk disease or identifying small but aggressive tumours and determining most appropriate therapeutic options. In addition, epigenetic changes are known to manifest early in tumourigenesis, making them useful for both
 5 diagnosis and prognosis.

In this study, we identified and validated chromosome conformations as distinctive biomarkers for a non-invasive blood-based epigenetic signature for PCa. Our data demonstrate the presence of stable chromatin loops in the loci of *ETS1*, *MAP3K14*, *SLC22A3* and *CASP2* genes present only in PCa patients
 10 **(Table 11)**. Validation of these markers in an independent set of 20 blinded samples showed 80% sensitivity and 80% specificity **(Table 12)**, which is remarkable for a PCa blood test. Interestingly, the expression of some of these genes has already been linked to cancer pathophysiology. *ETS1* is a member of ETS transcription factor family. *ETS1*-overexpressing prostate tumours are associated with increased cell migration, invasion and induction of epithelial-to-mesenchymal transition. *MAP3K14* (also known as
 15 nuclear factor-kappa-beta (NF- κ B)-inducing kinase (NIK)) is a member of MAP3K group (or MEKK). Physiologically, *MAP3K14*/NIK can activate noncanonical NF- κ B signalling and induce canonical NF- κ B signalling, particularly when *MAP3K14*/NIK is overexpressed. A novel role for *MAP3K14*/NIK in regulating mitochondrial dynamics to promote tumour cell invasion has been described. *SLC22A3* (also known as organic cation transporter 3 (OCT3)) is a member of SLC group of membrane transport proteins. *SLC22A3*
 20 expression is associated with PCa progression. *CASP2* is a member of caspase activation and recruitment domains group. Physiologically, *CASP2* can act as an endogenous repressor of autophagy. Two of the identified genes (*SLC22A3* and *CASP2*) were previously shown to be inversely correlated with cancer progression. Importantly, the presence of the chromatin loop can have indeterminate effect on gene expression.

25 To screen for PCa prognostic markers we performed the EpiSwitch™ custom array to analyse competitive hybridization of DNA from peripheral blood from patients with low-risk PCa (classification 1) and high risk PCa (classification 3). Six-marker set was identified for high-risk category 3 vs low-risk category 1, including *BMP6*, *ERG*, *MSR1*, *MUC1*, *ACAT1* and *DAPK1*. Six-biomarkers were identified for high-risk category 3 vs
 30 intermediate-risk category 2, including *HSD3B2*, *VEGFC*, *APAF1*, *MUC1*, *ACAT1* and *DAPK1*. Three of these biomarkers (*MUC1*, *ACAT1* and *DAPK1*) were shared between these sets. Our data show high concordance between chromosomal conformations in the primary tumour and in the blood of matched PCa patients at stages 1 and 3 **(Table 14)**. The prognostic significance and diagnostic value of some of these genes have

previously been suggested. *BMP6* plays an important role in PCa bone metastasis. In addition to *ETS1*, *ERG* is another member of the ETS family of transcription factors. Overwhelming evidence, suggesting that *ERG* is implicated in several processes relevant to PCa progression including metastasis, epithelial–mesenchymal transition, epigenetic reprogramming, and inflammation. *MSR1* may confer a moderate risk to PCa. *MUC1* is a membrane-bound glycoprotein that belongs to the mucin family. *MUC1* high expression in advanced PCa is associated with adverse clinicopathological tumour features and poor outcomes. *ACAT1* expression is elevated in high-grade and advanced PCa and acts as an indicator of reduced biochemical recurrence-free survival. *DAPK1* could function either as a tumour suppressor or as an oncogenic molecule in different cellular context. *HSD3B2* plays a crucial role in steroid hormone biosynthesis and it is up-regulated in a relevant fraction of PCa that are characterized by an adverse tumour phenotype, increased androgen receptor signalling and early biochemical recurrence. *VEGFC* is a member of VEGF family and its increased expression is associated with lymph node metastasis in PCa specimens. In a comprehensive biochemical approach, *APAF1* has been described as the core of the apoptosome.

Despite the identification of these loci, the mechanism of cancer-related epigenetic changes in PBMCs remains unidentified. The interaction, however, can be detected systemically and could be detected under same conditions in the primary site of tumorigenesis (**Table 14**). Thus for us to be able to measure the changes, chromatin conformation in PBMCs must be directed by an external factor; presumably something generated by the cells of the PCa tumour. It is known that a significant proportion of chromosomal conformations are controlled by non-coding RNAs, which regulate the tumour-specific conformations. Tumour cells have been shown to secrete non-coding RNAs that are endocytosed by neighbouring or circulating cells and may change their chromosomal conformations, and are possible regulators in this case. While RNA detection as a biomarker remains highly challenging (low stability, background drift, continuous basis for statistical stratification analysis), chromosome conformation signatures offer well recognized stable binary advantages for the biomarker targeting use, specifically when tested in the nuclei, since the circulating DNA present in plasma does not retain 3D conformational topological structures present in the intact cellular nuclei. It is important to mention, that looking at one genetic locus does not equate to looking at one marker, as there may be multiple chromosome conformations present, representing parallel pathways of epigenetic regulation over the locus of interest. One of the key challenges in the present clinical practice of PCa diagnosis is the time it takes to make a definitive diagnosis. So far, there is no single, definitive test for PCa. High levels of PSA will set the patient on a long journey of uncertainty where he will undergo magnetic resonance imaging scan followed by

biopsy, if needed. Although a biopsy is more reliable than a PSA test, it is a major procedure where missing the cancer lesions can still be an issue. The five-set biomarker panel described here is based on a relatively inexpensive and well-established molecular biology technique (PCR). The samples are based on biofluid, which is simple to collect and provides clinicians with rapidly available clinical readouts within few hours.

5 This in turn, offers a substantial time and cost savings and aids an informative diagnostic decision which fills the gap in the current protocols for assertive diagnosis of PCa.

Predicting the risk of PCa is pivotal for making an informed decision on treatment options. Five-year survival rate in the low risk group is more than 95% and most men would benefit from less invasive therapy. Currently, PCa risk stratification is based on combined assessment of circulating PSA, tumour grade (from biopsy) and tumour stage (from imaging findings). The ability to derive similar information using a simple blood test would allow significant reduction in costs and would speed up the diagnostic process. Of particular importance in PCa treatment is identifying the few tumours that initially present as low-risk, but then progress to high-risk. This subset would therefore benefit from a quicker and more-radical intervention.

In conclusion, here, we have identified subsets of chromosomal conformations in patients' PBMCs that are strongly indicative of PCa presence and prognosis. These signatures have a significant potential for the development of quick diagnostic and prognostic blood tests for PCa and significantly exceed the specificity of currently used PSA test. Preferred markers and combinations include

- *ETS1*, *MAP3K14*, *SLC22A3* and *CASP2*. This is Diagnostic, by nested PCR markers

- *BMP6*, *ERG*, *MSR1*, *MUC1*, *ACAT1* and *DAPK1*. This is Prognostic Signature (High-risk Category 3 vs Low Risk Category 1, by Nested PCR Markers)

- *HSD3B2*, *VEGFC*, *APAF1*, *MUC1*, *ACAT1* and *DAPK1*. This is Prognostic (High Risk Cat 3 vs Medium Risk Cat 2)

Example 5. Further Work on DLBCL

Diffuse large B-cell lymphoma (DLBCL) is a heterogenous blood cancer, but can be broadly classified into two main subtypes, germinal center B-cell-like (GCB) and activated B-cell-like (ABC). GCB and ABC subtypes have very different clinical courses, with ABC having a much worse survival prognosis. It has been observed that patients with different subtypes also respond differently to therapeutic intervention, in fact, some have argued that ABC and GCB can be thought of as separate diseases altogether. Due to this variability in response to therapy, having an assay to determine DLBCL subtypes has important

implications in guiding the clinical approach to the use of existing therapies, as well as in the development of new drugs. The current gold standard assay for subtyping DLBCL uses gene expression profiling on formalin fixed, paraffin embedded (FFPE) tissue to determine the “cell of origin” and thus disease subtype. However, this approach has some significant clinical limitations in that it 1) requires a biopsy 2) requires a complex, expensive and time-consuming analytical approach and 3) does not classify all DLBCL patients.

Here, we took an epigenomic approach and developed a blood-based chromosome conformation signature (CCS) for identifying DLBCL subtypes. An iterative approach using clinical samples from 118 DLBCL patients was taken to define a panel of six markers (DLBCL-CCS) to subtype the disease. The performance of the DLBCL-CCS was then compared to conventional gene expression profiling (GEX) from FFPE tissue.

The DLBCL-CCS was accurate in classifying ABC and GCB in samples of known status, providing an identical call in 100% (60/60) samples in the discovery cohort used to develop the classifier. Also, in the assessment cohort the DLBCL-CCS was able to make a DLBCL subtype call in 100% (58/58) of samples with intermediate subtypes (Type III) as defined by GEX analysis. Most importantly, when these patients were followed longitudinally throughout the course of their disease, the *EpiSwitch*[™] associated calls tracked better with the known patterns of survival rates for ABC and GCB subtypes.

This study provides an initial indication that a simple, accurate, cost-effective and clinically adoptable blood-based diagnostic for identifying DLBCL subtypes is possible.

Background

Diffuse large B-cell lymphoma (DLBCL) is the most common type of blood cancer and numerous studies using different methodologies have demonstrated it to be genetically and biologically heterogeneous. The two principal DLBCL molecular subtypes are germinal center B-cell-like (GCB) and activated B-cell-like (ABC), although more granular definitions of molecular subtypes have also been proposed. These two primary subtypes have a high degree of clinical relevance, as it has been observed that they have dramatically different disease courses, with the ABC subtype having a far worse survival prognosis. Perhaps more importantly, as novel investigational agents to treat GCB and ABC (or non-GCB) subtypes are evaluated in clinical settings and the historical observation that overall response rates in unselected patients is low, there is a pressing need to identify patient subtypes prior to the initiation of therapy. Historically, DLBCL subtypes are determined by identifying the “cell of origin” (COO). The original COO

classification was based on the observed similarity of DLBCL gene expression to activated peripheral blood B cells or normal germinal center B-cells by hierarchical clustering analysis (3). This COO-classification by whole-genome expression profiling (GEP) classifies DLBCL into activated B-cell like (ABC), germinal center B-cell like (GCB), and Type-III (unclassified) subtypes, with the ABC-DLBCL characterized by a poor prognosis and constitutive NF- κ B activation. In their seminal work, Wright et al. identified 27 genes that were most discriminative in their expression between ABC and GCB-DLBCL, and developed a linear predictor score (LPS) algorithm for COO-classification. These original studies are entirely based on retrospective investigations of fresh-frozen (FF) lymphoma tissues. A major challenge for the application of this COO-classification in clinical practice has been an establishment of a robust clinical assay amenable to routine formalin-fixed paraffin-embedded (FFPE) diagnostic biopsies. Several studies have also investigated the possibility of COO classification of DLBCL using FFPE tissues by quantitative measurement of mRNA expression, including quantitative nuclease protection assay, GEP with the Affymetrix HG U133 Plus 2.0 platform or the Illumina whole-genome DASL assay, and NanoString Lymphoma Subtyping Test (LST) technology. Several immunohistochemistry (IHC)-based algorithms have also been investigated to recapitulate the COO-classification by GEP. In general, these studies demonstrated high confidence of COO-classification of DLBCL using FFPE tissues and a robust separation in overall survival between ABC and GCB subtypes, but suffer from reproducibility issues, particularly lack of concordance between assays. In addition, any IHC-based measure requires baseline tissue, which is not always available and current turnaround times from sample collection to assay readout are long, making implementation in clinical practice a challenge.

Among the approaches that have been used historically to subtype DLBCL, one method for COO assessment uses an assay that measures the expression of 27 genes from FFPE tissue by quantitative reverse transcription PCR (qRT-PCR) using the Fluidigm BioMark HD system. While there are some advantages to this methodology over existing techniques, the approach still faces some major obstacles that limit its clinical application in that it 1) requires a tissue biopsy 2) relies on expensive, non-standard and time-consuming laboratory procedures. As such, having a blood-based assay would advance the field by providing a simple, reliable and cost-effective method for DCBCL subtyping with enhanced clinical applicability.

In this study, we used a novel blood-based assay to determine COO classification in DLBCL patients by focusing on detecting changes in genomic architecture. As part of the epigenetic regulatory framework, genomic regions can alter their 3-dimensional structure as a way of functionally regulating gene

expression. A result of this regulatory mechanism is the formation of chromatin loops at distinct genomic loci. The absence or presence of these loops can be empirically measured using chromosome conformation capture (3C). Multiple genomic regions contribute to epistatic modulation through the formation of stable, conditional long-range chromosome interactions. The collective measurement of chromosome conformations at multiple genomic loci results in a chromosome conformation signature (CCS), or a molecular barcode that reflects the genomes response to its external environment. For detection, screening and monitoring of CCS we utilized the *EpiSwitch* platform, an established, high resolution and high throughput methodology for detecting CCSs. Based on 3C, the *EpiSwitch* platform has been developed to assess changes in chromatin structure at defined genetic loci as well as long-range non-coding cis- and trans- regulatory interactions. Among the advantages of using *EpiSwitch* for patient stratification are its binary nature, reproducibility, relatively low cost, rapid turnaround time (samples can be processed in under 24 hours), the requirement of only a small amount of blood (~50 mL) and compliance with FDA standards of PCR-based detection methodologies. Thus, chromosome conformations offer a stable, binary, readout of cellular states and represent an emerging class of biomarkers.

Here, we used an approach based on the assessment of changes in chromosomal architecture to develop a blood-based diagnostic test for DLBCL COO subtyping. We hypothesized that interrogation of genomic architecture changes in blood samples from DLBCL patients could offer an alternative method to tissue-based COO classification approaches and provide a novel, non-invasive, and more clinically applicable methodology to guide clinical decision making and trial design.

A total of 118 DLBCL patients with a known COO subtype and 10 healthy controls (HC) were used in this study. The samples were a subset of those collected in a phase III, randomized, placebo-controlled, trial of rituximab plus bevacizumab in aggressive Non-Hodgkin lymphoma. Briefly, adult patients aged ≥18 years with newly-diagnosed CD20-positive DLBCL were randomized to R-CHOP or R-CHOP plus bevacizumab (RA-CHOP). Blood samples collected from 60 DLBCL patients were used as a development cohort to identify, evaluate, and refine the CCS biomarker leads. The patients from this cohort were all typed as high/strong GCB (30) or ABC (30) with a high subtype specific LPS (linear predictor scores). The remaining 58 DLBCL samples had intermediate LPS and were determined as ABC, GCB or Unclassified by Fluidigm testing (**Figure 25**). These patient samples were not used for CCSs biomarker discovery and development; but were used at a later stage to assess the resultant classifier. The Fluidigm testing was done using tissue obtained from lymph nodes (either as punch biopsies or removed during surgery), and

the *EpiSwitch* analysis was done using matched peripheral whole blood collected from the patients prior to receiving any therapy.

In addition to patient samples, 12 cell lines (six ABC and six GCB) were also used in the initial stage of the biomarker screening to identify the set of chromosome conformations that could best discriminate between ABC and GCB disease subtypes (**Table 20**). Cell lines were obtained from the American Type Culture Collection (ATCC), the German Collection of Microorganisms and Cell Cultures (DSMZ), and the Japan Health Sciences Foundation Resource Bank (JHSF).

RNA was isolated and purified from pre-treatment FFPE biopsies. DLBCL subtypes were determined by adaption of the Wright et al. algorithm to expression data from a custom Fluidigm gene expression panel containing the 27 genes of the DLBCL subtype predictor. Validation of the COO assay by comparing Fluidigm qRT-PCR to Affymetrix data in a cohort of 15 non-trial subjects revealed a high correlation between qRT-PCR measurements from matched fresh frozen (FF) and FFPE samples across 19 classifier genes used. We also found a high correlation between Affymetrix microarray and Fluidigm qRT-PCR measurements from the same FF samples. Classifier gene weights calculated from qRT-PCR data from the Fluidigm COO assay were highly concordant with weights obtained from previous microarray data in an independent patient cohort. We observed high correlation (76% concordance) between LPS derived from the Fluidigm assay, data in FFPE tumor, and LPS derived from Affymetrix microarray data in matched FF tissue in the technical registry cohort.

A pattern recognition algorithm was used to annotate the human genome for sites with the potential to form long-range chromosome conformations. The pattern recognition software operates based on Bayesian-modelling and provides a probabilistic score that a region is involved in long-range chromatin interactions. Sequences from 97 gene loci (**Table 21**) were processed through the pattern recognition software to generate a list of the 13,322 chromosomal interactions most likely to be able to discriminate between DLBCL subtypes. For the initial screening, array-based comparisons were performed. 60-mer oligonucleotide probes were designed to interrogate these potential interactions and uploaded as a custom array to the Agilent SureDesign website. Each probe was present in quadruplicate on the *EpiSwitch* microarray. To subsequently evaluate a potential CCS, nested PCR (*EpiSwitch* PCR) was performed using sequence-specific oligonucleotides designed using Primer3. Oligonucleotides were tested for specificity using oligonucleotide specific BLAST.

The top ten genomic loci that were identified as being dysregulated in DLBCL were uploaded as a protein list to the Reactome Functional Interaction Network plugin in Cytoscape to generate a network of epigenetic dysregulation in DLBCL. The ten loci were also uploaded to STRING (Search Tool for the Retrieval of Interacting Genes/Proteins DB) (<https://string-db.org/>), a database containing over 9 million known and predicted protein-protein interactions. Restricting to only human interactions, the main network (i.e. non-connected nodes were excluded) was generated. The top false discovery rate (FDR)-corrected functional enrichments were identified by Gene Ontology (GO) and the Kyoto Encyclopedia of Genes and Genomes (KEGG) databases. The top ten genomic loci were also uploaded to the KEGG Pathway Database (<https://www.genome.jp/kegg/pathway.html>) to identify specific biological pathways that exhibit dysregulation in DLBCL.

Exact and Fisher's exact test (for categorical variables) were used to identify discerning markers. The level of statistical significance was set at $p \leq 0.05$, and all tests were 2-sided. The Random Forest classifier was used to assess the ability of the *EpiSwitch* markers to identify DLBCL subtypes. Long term survival analysis was done by Kaplan-Meier analysis using the *survival* and *survminer* packages in R (38). Mean survival time was calculated using a two-tailed t-test.

We employed a step-wise approach to discover and validate a CCS biomarker panel that could differentiate between DLBCL subtypes (**Figure 19**). As a first step in the discovery of the *EpiSwitch* classifier, 97 genetic loci (**Table 21**) were selected and annotated for the predicted presence of chromosome conformation interaction sites and screened for their empirical presence using the *EpiSwitch* CGH Agilent array. The annotated array design represented 13,322 chromosome interaction candidates, with an average of 99 distinct cis-interactions tested at each locus (99 ± 64 ; mean $\pm$ SD). This discovery array was used to screen and identify a smaller pool of chromosome conformations that could differentiate between the two main DLBCL subtypes. The samples used for this step were from GCB and ABC cell lines (**Table 20**) as well as whole blood from four typed DLBCL patients (two GCB and two ABC) and four HCs. The cell lines were grouped into high ABC and GCB and low ABC and GCB based on gene expression analysis. The comparisons used on the array were: 1) individual comparisons of DLBCL patients to pooled HCs 2) pooled DLBCL samples to pooled HC samples 3) pooled high ABC compared to pooled high GCB cell lines, and 4) pooled low ABC versus pooled low GCB cell lines.

From the array analysis, we identified 1,095 statistically significant chromosomal interactions that differentiated between high ABC and GCB cell lines and were present in blood samples from DLBCL

patients, but absent in HCs. These were further reduced to the top 293 interactions using a set of statistical filters, 151 of which were associated with the ABC subtype and 143 of which were associated with the GCB subtype. The top 72 interactions from either subtype (36 interactions for ABC and 36 interactions for GCB) were selected for further refinement using the *EpiSwitch* PCR platform on 60 typed DLBCL patient samples. For all 118 DLBCL samples, initial subtype classification was assigned based on the Wright algorithm, which calculates a linear predictor score (LPS) from the expression of a panel of 27 genes. 60 samples were classified as either ABC or GCB and used to develop the *EpiSwitch* classifier (the “Discovery Cohort”) and 58 samples were of intermediate LPS scores and used to evaluate the performance of the *EpiSwitch* classifier (the “Assessment Cohort”) (**Figure 19**).

The 72 interactions identified in the initial screen were narrowed to a smaller pool using both the DLBCL patient samples during the discovery step and a second cohort of 60 DLBCL typed (30 ABC and 30 GCB) patient samples along with 12 HC (**Figure 19**). The DLBCL subtype calls made by the *EpiSwitch* assay were confirmed using the Fluidigm platform. The Fluidigm gene expression analysis was performed on tissue biopsy samples, whereas whole blood from the same patients was used for the *EpiSwitch* PCR assay. The initial steps in refinement were to confirm by PCR that the 72 chromosomal interactions identified in the initial screen were specific to DLBCL and were absent in the HC samples. This was first tested on six untyped DLBCL samples and two HCs and resulted in identification of 21 interactions that were specific for DLBCL. Next, we used *EpiSwitch* PCR to test 24 blood samples from typed DLBCL patient samples (12 ABC and 12 GCB) to identify DLBCL-specific chromosome interactions using Fisher’s test. This resulted in a set of 10 discriminating chromosome conformation interactions that could accurately discriminate between ABC and GCB subtypes and were further evaluated on blood samples from an additional set of 36 DLBCL samples (18 ABC and 18 GCB) (**Figure 19**).

To test the accuracy, performance and robustness of the 10-marker panel, we used Exact test for feature selection on 80% of the complete sample cohort (Total 48 samples: 24 ABC and 24 GCB), with the remaining 20% (12 samples, 6 ABC and 6 GCB) used for later testing of the final selected CCSs markers. The data was split 10 times and the Exact test run on each of the splits using the 80% training set of each split. The composite p-value for the 10 markers over the 10 splits was then used to rank the markers. This analysis identified six chromosome conformations in the IFNAR1, MAP3K7, STAT3, TNFRSF13B, MEF2B, and ANXA11 genetic loci. Collectively, these six interactions formed the DLBCL chromosome conformation signature (DLBCL-CCS) (**Figure 20**).

The six markers in the DLBCL-CCS were used to generate a Random forest classifier model and applied to classify the test sets for each of the data splits (12 samples, 6 ABC and 6 GCB) in the Discovery Cohort of known disease subtypes. By principal component analysis (PCA), the DLBCL-CCS classifier was able to separate ABC and GCB patients from healthy controls (**Figure 26**). The composite prediction probabilities for the DLBCL-CCS is shown in **Table 22** along with the odds ratio for each marker and the odd ratio for the model generated using logistic regression. The model provided a prediction probability score for ABC and GCB, ranging from 0.186 to 0.81 (0 =ABC, 1 = GCB). The probability cut-off values for correct classification were set at ≤ 0.30 for ABC and ≥ 0.70 for GCB. The score of ≤ 0.30 had a true positive rate (sensitivity) of 100% (95% confidence interval [95% CI] 88.4–100%), while a score of ≥ 0.70 had a true negative response rate (specificity) of 96.7% (95% CI 82.8–99.9%). With the DLBCL-CCS classifier, 60 out of 60 patients (100%) were correctly classified as either ABC or GCB, when compared to the Fluidigm calls for subtyping (**Figure 21A, Table 22**). The AUC under the receiver operating characteristic (ROC) curve for the DLBCL-CCS classifier on this sample cohort was 1 (**Figure 21B**). Last, we compared the DLBCL subtype calls made by the DLBCL-CCS to the long-term survival curves of the patients with known disease subtype. The patients called as ABC showed significantly worse survival than those patients called as GCB (**Figure 21C**).

Next, we evaluated the performance of the DLBCL-CCS the Assessment Cohort of 58 DLBCL patients with a more intermediate LPS value. We applied the DLBCL-CCS to assign these patients into DLBCL subtypes and compared the readouts to those made by Fluidigm. The DLBCL-CCS made subtyping calls for all 58 samples, whereas the Fluidigm assay made subtyping calls for 37 of the samples, leaving 21 as “unclassified” (**Figure 22**). Of the 37 samples where subtype calls for both assays was available, 15 samples (40%) were called similarly by both assays (8 ABC and 7 GCB) (**Figure 22**). Next, we evaluated the performance of the DLBCL subtype calls made by the DLBCL-CCS and Fluidigm by comparing the subtype calls made at diagnosis with the long-term survival curves of the Type III patients. As shown in the Kaplan-Meier survival curves in **Figure 23**, the ABC/GCB calls made by the DLBCL-CCS was able to separate the two populations based on the known survival trends in DLBCL, with the ABC subtype having a worse prognosis. In contrast, the ABC and GCB populations as defined by Fluidigm showed the opposite of what has been observed clinically, with samples classified as ABC having longer survival times than those classified as GCB. Though not statistically significant, the subtype calls made by the DLBCL-CCS matched historical clinical observations of survival differences between the subtypes by Hazard ratio analysis. We did find a significant difference in mean survival time between the two methods. The mean survival of patients classified as ABC and GCB by Fluidigm was 651 and 626 days, respectively ($p=0.854$), while the

mean survival of patients classified as ABC and GCB by the DLBCL-CCS assay was 550 and 801 days ($p=0.017$) (**Figure 24**).

In order to explore the relationship between the loci that were observed to be epigenetically dysregulated in this study and biological mechanisms that have previously been reported to be linked to DLBCL, we performed a series of network and pathway analyses using the top 10 dysregulated loci as inputs. First, we explored how these loci were biologically related by building a Reactome Functional Interaction Network in Cytoscape which revealed a network centred on NFKB1, STAT3 and NFATC1. A similar picture emerged when the 10 loci were used to build a network using STRING DB, with the most connected hubs centring on NFKB1, STAT3 and MAP3K7 and CD40. The top enriched GO term for biological process was “positive regulation of transcription, DNA-templated”, the top enriched GO term for molecular function was “transcriptional activator activity, RNA polymerase II transcription regulatory region sequence-specific binding” and the “Toll-like receptor signalling pathway” was the most enriched KEGG pathway (**Table 22**). When we mapped the top ten loci to the KEGG Toll-like receptor signalling pathway, we found that specific cascades related to the production of proinflammatory cytokines and costimulatory molecules through the NF- κ B and the interferon mediated JAK-STAT signalling cascades.

Due to the observed differences in disease progression for the different DLBCL subtypes, there is a pressing clinical need for a simple and reliable test that can differentiate between ABC and GCB disease subtypes. Given the aggressive nature of the disease, DLBCL requires immediate treatment. The two main subtypes have different clinical management paradigms and with several therapeutic modalities in development that target specific subtypes, having a rapid and accurate disease diagnostic is critical when clinical management depends on knowing disease subtype. The field of COO-classification in DLBCL has expanded from IHC based methodologies to DNA microarrays, parallel quantitative reverse transcription PCR (qRT-PCR) and digital gene expression. A current favoured method is based on identification of the COO by GEP on FFPE tissue and suffers from some technical and logistical limitations that limit its broad adoption in the clinical setting. In addition, there are many factors that affect the performance and reliability of COO-classification by GEP on FFPE tissue; including the nature/quality of lymphoma specimen, the experimental methods for data collection; data normalization and transformation, the type of classifier used, and the probability cut offs used for subtype assignment. Last, going from sample collection to an end readout using the Fluidigm approach is a complex and time-consuming process with many steps in between having the potential to introduce performance variability. All of these factors have an impact on the overall turnaround time of the assay and limits how it can be used clinically to diagnose

and inform treatment of the disease using existing medications as well as select patients for late stage trials for novel DLBCL therapeutics. Thus, the need for a simple, minimally invasive and reliable assay to differentiate DLBCL subtypes is needed.

- 5 Using a stepwise discovery approach, we identified a 6-marker epigenetic biomarker panel, the DLBCL-CCS, that could accurately discriminate between DLBCL subtypes. When compared to the subtype results derived from the gene expression signature there was perfect concordance; which was expected as these were samples that were used to develop the classifier. The concordance between the two assays when applied to samples with an intermediate LPS was lower (just over 40%). This is perhaps expected, as it has
- 10 been noted that there is a lack of overall concordance in DLBCL subtype calls with different methods of classification, and the Type III samples are perhaps a more heterogeneous population reflecting a more intermediate biology to begin with. However, when we evaluated the predictive classification ability of the *EpiSwitch* assay in the Type III DLBCL patients followed longitudinally as their disease progressed, baseline predictions of disease subtype using the *EpiSwitch* assay was better at predicting actual disease
- 15 subtype based on observed survival curves in patients with unclassified disease. The observation that the epigenetic readout based on regulatory 3D genomics used here is more consistent with actual clinical outcomes than the transcription-based gold-standard molecular approaches represents an actionable advance in the management of DLBCL. It is also consistent with a system biology evaluation of regulatory 3D genomics as a molecular modality closely linked to phenotypical differences in oncological conditions.
- 20 We do note that DLBCL operates on a biological continuum, with significant heterogeneity in disease biology between subtypes. By design, the DLBCL-CCS was set up to classify Type III samples into either ABC or GCB subtypes. By GEX analysis, the Type III samples were identified as having intermediate subtype biology so may represent a more heterogeneous population of patients. However, the overall observation that the DLBCL-CCS was a better predictor of disease subtype as measured by clinical progression than
- 25 using a GEX-based approach and the fact that the *EpiSwitch* assay was able to make subtype calls in all samples, provides an initial indication that this approach can be applied in a clinical setting to inform on prognostic outlook, potentially guide treatment decisions, and provide predictions for response to novel therapeutic agents currently in development.
- 30 In the network analysis, the NF-kB and STAT3 signalling cascades emerged as putative mediators that differentiate between DLBCL subtypes. The role of NF-kB signalling in DLBCL has been studied before, in fact, one of the discriminating features of the ABC subtype is constitutive expression of NF-kB target genes, a mechanism which has been hypothesized for the poor prognosis in these patients. In addition,

mutations causing constitutive signalling activation have been observed predominantly in the ABC subtype for several NF- κ B pathway genes, including TNFAIP3 and MYD88.

In addition to validating known mechanisms of DLBCL, the network analysis here identified a novel potential target for therapeutic intervention in DLBCL. For example, ANXA11, a calcium-regulated phospholipid-binding protein, has been implicated in other oncological conditions such as colorectal cancer, gastric cancer and ovarian cancer and could be a novel therapeutic intervention point in DLBCL.

One of the major clinical advantages of the approach to DLBCL subtyping described here lies in the simplified laboratory methodology and workflow. Conventional, gold-standard subtyping by GEP can be done using a variety of commercial platforms but all generally follow (and require) a four-step approach: 1) acquisition of a tissue biopsy, 2) preparation of FFPE tissue sections 3) gene expression analysis and 4) algorithmic classification of subtype. Obtaining a fine needle tissue biopsy of an enlarged, peripheral lymph node requires an inpatient visit to a clinical site and an invasive medical procedure requiring anaesthetic. Once obtained, the fresh biopsy needs to be prepared for paraffin embedding. This is a multi-step process, but generally involves immersion in liquid fixing agent (such as formalin) long enough for it to penetrate through the entire specimen, sequential dehydration through an ethanol gradient, followed by clearing in xylene, a toxic chemical. Last, the biospecimen needs to be infiltrated with paraffin wax and left to cool so that it solidifies and can be cut into micrometer sections using a microtome and mounted onto laboratory slides. The entire process of going from fresh tissue to FFPE sections on a slide can take several days. Next, in order to perform gene expression analysis, inherently unstable RNA is extracted from slide-mounted tissue sections and prepared for hybridization to microarrays according to the array manufacturer's specifications, a process that can take over a day. Following microarray hybridization, digital readouts of relative gene expression levels for the are obtained and fed into a classification algorithm to determine DLBCL subtype. All told, the process of going from a patient with suspected DLBCL to a subtype readout can take up to a week or longer, involves many different experimental steps using expensive technologies, each of which has the potential to introduce experimental variability along the way. In the approach described here, the time and the number of steps from biofluid collection to subtype readout are dramatically decreased. A patient with suspected DLBCL can present to an outpatient clinic for a routine, small volume (~ 1mL) blood draw. Fresh frozen blood can then be shipped to a central, accredited reference lab for analysis of the absence/presence of the chromosome conformations identified in this study; a process that uses an even smaller volume (~50 mL) of whole blood as input along with specific PCR primer sets and reaction conditions to detect the chromosome conformations using

simple and routine PCR instrumentation in less than 24 hours from sample receipt. The approach to DLBCL subtyping described here offers an additional advantage in that the potential for further refinement using the proposed methodology exists. In this study, final readout of the DLBCL-CCS was done using a set of nested PCR reactions to detect chromosome conformations making up the classifier. This PCR-based output can be further refined to utilize quantitative PCR as a readout and operate under the minimum information for publication of quantitative real-time PCR experiments (MIQE) guidelines, designed to enhance experimental reproducibility and reliability across reference labs and testing sites. Last, the approach described here is adaptable to the evolving understanding of the disease itself, such as the different physiologically heterogeneous forms of it.

In conclusion, here we developed a robust complementary method for non-invasive COO assignment from whole blood samples using *EpiSwitch* CCSs readouts. We demonstrated the clinical validity of this classification approach on a large cohort of DLBCL patients. The *EpiSwitch* platform has several attractive features as a biomarker modality with clinical utility. CCSs have very high biochemical stability, can be detected using very small amounts of blood (typically around 50 µl) and detection utilizes established laboratory methodologies and standard PCR readouts (including MIQE-compliant qPCR). Finally, the rapid turnaround time (~8-16 hours) of the *EpiSwitch* assay compares favourably to the over 48 hours for the Fluidigm platform.

Example 6. Further Work on Canine DLBCL

Here, we used the *EpiSwitch*TM platform technology to evaluate chromosome conformation signatures (CCS) as biomarkers for detection of canine diffuse large B-cell lymphoma (DLBCL). We examined whether established, systemic liquid biopsy biomarkers previously characterized in human DLBCL patients by *EpiSwitch*TM would translate to dogs with the homologous disease. Orthologous sequence conversion of CCS from humans to dogs was first verified and validated in control and lymphoma canine cohorts.

Blood samples from dogs with DLBCL and from apparently healthy dogs were obtained. All of the dogs diagnosed with DLBCL, were part of the LICKing Lymphoma trial. Blood samples were obtained from each dog prior to initiating treatment and at day+5 after the experimental intervention, but prior to initiating doxorubicin chemotherapy. *EpiSwitch*TM technology was used to monitor systemic epigenetic biomarkers for CCS.

A 11-marker classifier was generated with whole blood from 28 dogs, 14 diagnosed with DLBCL and 14 controls with no apparent disease, from a pool of 75 EpiSwitch CCSs identified in human DLBCL. Validation of the developed diagnostic markers was performed on a second cohort of 10 dogs: 5 with DLBCL and 5 controls. The classifier delivered stratifications for DLBCL vs. non-DLBCL with 80% accuracy, 80% sensitivity, 80% specificity, 80% positive predictive value (PPV) and 80% negative predictive value (NPV) on the second cohort.

The established EpiSwitch™ classifier contains strong systemic binary markers of epigenetic deregulation with features normally attributed to genetic markers: the binary status of these classifying markers is statistically significant for diagnosis.

	Probe	GeneLocus	Probe_Count_Total
1	STAT3_17_40446029_40448202_40557923_40558616_RR	STAT3	1108
2	ANXA11_10_81889664_81892389_81927417_81929312_FR	ANXA11	136
3	CD40_20_44739847_44744687_44767157_44770555_FR	CD40	148
4	IFNAR1_21_34696683_34697716_34777569_34779811_RF	IFNAR1	80
5	MAP3K7_6_91275515_91285706_91312237_91314731_FF	MAP3K7	308
6	MEF2B_19_19271977_19273500_19302232_19303741_RF	MEF2B	448
7	MLLT3_9_20556478_20560948_20658310_20666368_FF	MLLT3	120
8	NFATc1_18_77133931_77135912_77218993_77220063_RF	NFATc1	608
9	NFKB1_4_103425293_103430397_103512508_103516923_FR	NFKB1	96
10	TNFRSF13C_22_42302849_42305750_42342568_42346797_FR	TNFRSF13C	488
11	BAX_19_49421750_49425644_49457303_49458439_RF	BAX	92
12	BCL6_3_187438677_187439687_187454088_187455426_FF	BCL6	240
13	IL22RA1_1_24467543_24471444_24512238_24513959_RF	IL22RA1	48
14	TNFRSF13C_22_42313974_42315085_42342568_42346797_RR	TNFRSF13C	488
15	FOXO1_13_41184194_41191166_41219134_41220693_FR	FOXO1	308
16	HLF_17_53402207_53403714_53420274_53422428_FF	HLF	104
17	PAK1_11_77028527_77036211_77090325_77094591_RF	PAK1	180
18	FOS_14_75744954_75746643_75795718_75799884_FF	FOS	80
19	MTHFR_1_11807586_11814341_11843522_11845650_RF	MTHFR	52
20	WNT9A_1_228068849_228075473_228135088_228140421_RR	WNT9A	40
21	NFATc1_18_77229964_77232215_77280170_77283702_FR	NFATc1	608
22	BRCA1_17_41162341_41168331_41242678_41245761_RR	BRCA1	297
23	TET2_4_106047220_106052671_106063962_106067377_FF	TET2	104
24	TNF_6_31525914_31529267_31542458_31544282_RF	TNF	68
25	NFATc1_18_77158863_77160420_77229964_77232215_FF	NFATc1	608
26	BCL6_3_187454088_187455426_187484009_187486420_FF	BCL6	240
27	MAPK13_6_36066232_36072387_36102587_36105090_FR	MAPK13	44
28	MLLT3_9_20319606_20322797_20621547_20622617_FR	MLLT3	120
29	TOP1_20_39656117_39657610_39725920_39729106_FR	TOP1	164
30	IFNAR1_21_34696683_34697716_34717312_34717993_RF	IFNAR1	80
31	SKP1_5_133465952_133470062_133512403_133513591_RR	SKP1	136
32	FZD10_12_130601147_130601992_130676699_130678204_FR	FZD10	124
33	ITGA5_12_54787051_54795949_54806686_54808428_FR	ITGA5	80

34	TNFRSF13B_17_16842268_16844133_16924802_16926550_RR	TNFRSF13B	128
35	BCL6_3_187438677_187439687_187454088_187455426_RR	BCL6	240
36	ITPR3_6_33600698_33604388_33678436_33680494_RR	ITPR3	100
37	MAP3K7_6_91275515_91285706_91312237_91314731_FF	MAP3K7	308
38	IFNAR1_21_34696683_34697716_34777569_34779811_RF	IFNAR1	80
39	NFATc1_18_77156086_77157023_77218993_77220063_RF	NFATc1	608
40	PRDM1_6_106483435_106485826_106500642_106506822_RF	PRDM1	120
41	IL-2RB_22_37532051_37533547_37544442_37546723_FR	IL-2RB	72
42	STAT3_17_40446029_40448202_40557923_40558616_RR	STAT3	1108
43	NFKB1_4_103405171_103418579_103512508_103516923_FR	NFKB1	96
44	CABLES1_18_20774415_20775705_20863570_20868210_RF	CABLES1	136
45	JDP2_14_75883183_75893682_75936165_75936958_FF	JDP2	80
46	NFATc1_18_77133931_77135912_77218993_77220063_RF	NFATc1	608
47	CASP3_4_185504966_185506889_185543536_185552493_FR	CASP3	88
48	REL_2_61074693_61075565_61108479_61109187_FR	REL	92
49	BTK_X_100610457_100612966_100667570_100670929_RF	BTK	404
50	BCL2A1_15_80256742_80257692_80285499_80286865_RR	BCL2A1	302
51	TNFRSF13C_22_42302849_42305750_42318166_42319783_FF	TNFRSF13C	488
52	CDKN2C_1_51402271_51403526_51439728_51440611_RR	CDKN2C	72

Table 5.a

	Probe_Count_Sig	HyperG_Stats	FDR_HyperG	Percent_Sig
1	615	0.000000000125197189743782	0.0000000113929442666842	55.51
2	83	0.000391435	0.005936759	61.03
3	64	0.802231212	0.999999997	43.24
4	34	0.79036009	0.999999997	42.5
5	113	0.999793469	0.999999997	36.69
6	216	0.227265083	0.590889215	48.21
7	39	0.999297311	0.999999997	32.5
8	213	0.999999997	0.999999997	35.03
9	27	0.999920864	0.999999997	28.12
10	280	0.000000444123116904245	0.0000202076018191431	57.38
11	61	0.0000870841188293703	0.001584931	66.3
12	86	0.999659646	0.999999997	35.83
13	14	0.995140179	0.999999997	29.17
14	280	0.000000444123116904245	0.0000202076018191431	57.38
15	148	0.294116072	0.669114065	48.05
16	44	0.824486728	0.999999997	42.31
17	89	0.224299285	0.590889215	49.44
18	31	0.931715515	0.999999997	38.75
19	30	0.066847306	0.221674157	57.69
20	21	0.267230575	0.639946904	52.5
21	213	0.999999997	0.999999997	35.03
22	173	0.0000217239097176038	0.000658959	58.25
23	58	0.033733587	0.145711753	55.77
24	18	0.999770934	0.999999997	26.47
25	213	0.999999997	0.999999997	35.03
26	86	0.999659646	0.999999997	35.83
27	18	0.81000526	0.999999997	40.91
28	39	0.999297311	0.999999997	32.5
29	71	0.808882973	0.999999997	43.29

30	34	0.79036009	0.999999997	42.5
31	72	0.072624208	0.221674157	52.94
32	43	0.996927827	0.999999997	34.68
33	40	0.293962898	0.669114065	50
34	76	0.002030583	0.01918923	59.38
35	86	0.999659646	0.999999997	35.83
36	48	0.409333853	0.903779884	48
37	113	0.999793469	0.999999997	36.69
38	34	0.79036009	0.999999997	42.5
39	213	0.999999997	0.999999997	35.03
40	47	0.9542933	0.999999997	39.17
41	24	0.991079692	0.999999997	33.33
42	615	0.000000000125197189743782	0.0000000113929442666842	55.51
43	27	0.999920864	0.999999997	28.12
44	59	0.784673088	0.999999997	43.38
45	36	0.639258785	0.999999997	45
46	213	0.999999997	0.999999997	35.03
47	39	0.688804514	0.999999997	44.32
48	30	0.997411174	0.999999997	32.61
49	182	0.722716922	0.999999997	45.05
50	166	0.00150308	0.01918923	54.97
51	280	0.000000444123116904245	0.0000202076018191431	57.38
52	30	0.821366544	0.999999997	41.67

Table 5.b

	logFC	AveExpr	t	P.Value	adj.P.Val
1	0.102545415	0.102545415	2.181691533	0.06115714	0.124690581
2	0.146814815	0.146814815	3.078806942	0.015395162	0.044697142
3	0.247739738	0.247739738	4.372950932	0.002449301	0.012749359
4	0.098641538	0.098641538	1.475225491	0.178893946	0.27926686
5	0.098390923	0.098390923	2.270415909	0.053292308	0.112564482
6	0.246810388	0.246810388	5.953590771	0.000359019	0.0048119
7	0.194400918	0.194400918	2.492510608	0.037760627	0.086786653
8	0.117865744	0.117865744	1.424258285	0.19268713	0.295560888
9	0.253919456	0.253919456	1.95465634	0.086862876	0.161472968
10	0.210247736	0.210247736	2.234440593	0.056352274	0.11719604
11	-0.050897988	-0.050897988	-0.745725763	0.477453286	0.587468355
12	-0.030722825	-0.030722825	-1.143761644	0.28622615	0.400334573
13	-0.019224434	-0.019224434	-0.418322829	0.686861761	0.767615441
14	-0.014186527	-0.014186527	-0.069260125	0.946505227	0.961540839
15	-0.010289959	-0.010289959	-0.21611469	0.834379395	0.881581711
16	0.007162022	0.007162022	0.173838071	0.866368809	0.905364254
17	0.008581354	0.008581354	0.16838944	0.870512196	0.90817036
18	0.009594682	0.009594682	0.192008457	0.852583784	0.895087864
19	0.013062105	0.013062105	0.2898133	0.779427179	0.840001078
20	0.027459614	0.027459614	0.78838239	0.453500365	0.565397223
21	0.0309953	0.0309953	0.401906143	0.698417087	0.776651512
22	0.03119071	0.03119071	0.46421066	0.655032665	0.743235083
23	0.031952076	0.031952076	0.423263408	0.683401141	0.76482198
24	0.036397064	0.036397064	1.012029864	0.341541187	0.45879384
25	0.036449121	0.036449121	0.881245015	0.404223195	0.519301224

26	0.039792262	0.039792262	1.09873788	0.304267251	0.419987316
27	0.044981037	0.044981037	0.742965178	0.479031667	0.588704315
28	0.048157816	0.048157816	1.006274544	0.344135022	0.461283259
29	0.05692752	0.05692752	0.774540503	0.461182853	0.572336767
30	0.068999319	0.068999319	1.087503347	0.308907593	0.424307342
31	0.073674257	0.073674257	1.37062465	0.208203798	0.314180912
32	0.07496163	0.07496163	1.975529647	0.084116625	0.157842633
33	0.077618589	0.077618589	1.073493376	0.314772637	0.430505827
34	0.080234671	0.080234671	1.659676531	0.136077489	0.226348123
35	0.090602356	0.090602356	2.111185324	0.068216955	0.135104089
36	0.098319301	0.098319301	1.04377977	0.327501414	0.444140656
37	0.098390923	0.098390923	2.270415909	0.053292308	0.112564482
38	0.098641538	0.098641538	1.475225491	0.178893946	0.27926686
39	0.099162732	0.099162732	1.292936726	0.232594904	0.342880489
40	0.101277922	0.101277922	1.410506969	0.196565747	0.3002884
41	0.101676827	0.101676827	1.927111422	0.090619506	0.166768298
42	0.102545415	0.102545415	2.181691533	0.06115714	0.124690581
43	0.103364871	0.103364871	1.06297419	0.319233722	0.435153018
44	0.106978686	0.106978686	1.092750486	0.30673336	0.422367497
45	0.116604657	0.116604657	2.102936835	0.06909355	0.136425239
46	0.117865744	0.117865744	1.424258285	0.19268713	0.295560888
47	0.12582798	0.12582798	4.245528735	0.002904342	0.014151256
48	0.125971304	0.125971304	1.693676294	0.129294906	0.217785184
49	0.127634089	0.127634089	5.070996787	0.001004634	0.007593027
50	0.132678146	0.132678146	2.405792667	0.043193622	0.095959776
51	0.141794844	0.141794844	2.06833857	0.072892271	0.142181615
52	0.143309126	0.143309126	2.511399626	0.036672127	0.085032085

Table 5.c

	B	FC	FC_1	LS	Loop Detected
1	-4.804209212	1.073666112	1.073666112	1	DBLCL
2	-3.422371897	1.107122465	1.107122465	1	DBLCL
3	-1.514951748	1.18734545	1.18734545	1	DBLCL
4	-5.804534479	1.070764741	1.070764741	1	DBLCL
5	-4.669835237	1.070578751	1.070578751	1	DBLCL
6	0.493437892	1.186580836	1.186580836	1	DBLCL
7	-4.329420018	1.14424891	1.14424891	1	DBLCL
8	-5.869405345	1.085128386	1.085128386	1	DBLCL
9	-5.141601134	1.192442298	1.192442298	1	DBLCL
10	-4.724458483	1.156886824	1.156886824	1	DBLCL
11	-6.575035413	0.965335281	-1.035909513	-1	Ctrl
12	-6.200305189	0.978929707	-1.021523806	-1	Ctrl
13	-6.778021016	0.986763027	-1.01341454	-1	Ctrl
14	-6.87182168	0.990214838	-1.009881857	-1	Ctrl
15	-6.848536344	0.99289292	-1.007157952	-1	Ctrl
16	-6.85768141	1.004976678	1.004976678	1	DBLCL
17	-6.858716992	1.005965867	1.005965867	1	DBLCL
18	-6.85399155	1.00667269	1.00667269	1	DBLCL
19	-6.827923501	1.009095073	1.009095073	1	DBLCL
20	-6.54111486	1.019215847	1.019215847	1	DBLCL

21	-6.785369028	1.021716754	1.021716754	1	DBLCL
22	-6.755996218	1.021855153	1.021855153	1	DBLCL
23	-6.775754566	1.022394568	1.022394568	1	DBLCL
24	-6.338031258	1.025549454	1.025549454	1	DBLCL
25	-6.461788363	1.02558646	1.02558646	1	DBLCL
26	-6.248771887	1.027965796	1.027965796	1	DBLCL
27	-6.5771745	1.031669619	1.031669619	1	DBLCL
28	-6.343758805	1.033943833	1.033943833	1	DBLCL
29	-6.552299411	1.040248004	1.040248004	1	DBLCL
30	-6.260644426	1.048988833	1.048988833	1	DBLCL
31	-5.936215307	1.05239351	1.05239351	1	DBLCL
32	-5.111051252	1.053333021	1.053333021	1	DBLCL
33	-6.275323888	1.055274694	1.055274694	1	DBLCL
34	-5.559658058	1.05718999	1.05718999	1	DBLCL
35	-4.910091268	1.064814672	1.064814672	1	DBLCL
36	-6.305987164	1.070525603	1.070525603	1	DBLCL
37	-4.669835237	1.070578751	1.070578751	1	DBLCL
38	-5.804534479	1.070764741	1.070764741	1	DBLCL
39	-6.030168045	1.071151639	1.071151639	1	DBLCL
40	-5.886680521	1.072723247	1.072723247	1	DBLCL
41	-5.181746622	1.073019896	1.073019896	1	DBLCL
42	-4.804209212	1.073666112	1.073666112	1	DBLCL
43	-6.286252837	1.074276132	1.074276132	1	DBLCL
44	-6.255110449	1.076970465	1.076970465	1	DBLCL
45	-4.922419619	1.08418027	1.08418027	1	DBLCL
46	-5.869405345	1.085128386	1.085128386	1	DBLCL
47	-1.693141948	1.091133768	1.091133768	1	DBLCL
48	-5.512969318	1.091242172	1.091242172	1	DBLCL
49	-0.581917188	1.092500613	1.092500613	1	DBLCL
50	-4.462886317	1.096326979	1.096326979	1	DBLCL
51	-4.97398611	1.103276839	1.103276839	1	DBLCL
52	-4.300278466	1.104435469	1.104435469	1	DBLCL

Table 5.d

	Probe sequence	Probe Location
	60 mer	Chr
1	GGGTTTCACCATGTTGGCCAGGCTGGTCTCGAACTCCCACCTCAGGTGATCCGCCCCGCC	17
2	GAGGGGCTCTGGAGGGGGCGGGTCTCTCGATGCCTGGCCTCCACAGCACATGTGAGCA	10
3	GAGGCTTTTATGCAGGAAAGTGTCCCAGTCGAGGGACTGGCAGCAGGGGGACAGCAAGGG	20
4	ACCTCTCTTAATTTCTCAGCCATTCTTCGACCGCTCTGCCCCGCTCTCGCTCTGCAC	21
5	TGTGAAGGGAGGGGAGGAGAAAAAGAAAATCGAAACAAGCTTAGAAGCAGACACTTGCCCA	6
6	TGGGGGAGCTCTGGGGTGGGGGTAGCGGTGATGGGTCTGATGCCTCTCAGAAGGCCTT	19
7	ACATTTCAAATCCTCTCTTAGCTACCTCGAACTTCTGAGCTCAAGCAATCTTCCACCT	9
8	CTAGAGGAGAGAGGGATGCCAGGCTCTATCGAGTCTGAGTTCGTCCACGTGGTGGCCATC	18
9	TCTTTATGGTGTCTCTTATATATTTACTCGAGGCTGCAGTGAGCTATAATTGCACCACT	4
10	ACGGGCAGACAGGACCCAGCCCATGCCTCGACCACTCCCGGGGGGATCGGGACACCGC	22
11	TCCCTGCCTCTCTGGCGCTCTCGGACCCTCGAACCTCCCTTTGATCTATTCCATTCTCA	19
12	AGATCCGTGTCTGCCTGCAGATACAAAATCGAGGTGGATCGCCAGGGGCGGGCAGTCCC	3

13	GGGGGGGGGAGCGCGCCGGTCCCCGCGCTCGAAGGCTGCCCTCCTCTCTGAATTTGGGTT	1
14	ACCCAAACACGCGCAGACACCCGCACACTCGACCCACTCCCGGGGGGATCGGGACACCGC	22
15	CACACCCGCCCTACTGGATCCAAGTCACTCGAGACAACACTGAAAACACAAAGGCATTTA	13
16	TAGACTAGCGCCAGCTTTGTGCACAAGGTCGACACCCCTCTCCCCAACCCCTCTGTCAGAA	17
17	AGGGTTTACCATGTTGCCAGGCTGGTCTCGAGACCATCCTGGCTAATACGGTGAAACCC	11
18	CAACTTCATTCCCACGGTCACTGCCATCTCGACCCACCAATAGAGCAACTCCCTGAGAGG	14
19	CCATCAGCAAAGATGAACCTGGCACCTCTCGACGCCATAAGCATGGTGAGCCAGGGTGGG	1
20	TTCCAGTTCATAAAGATTTAAACAACATTGAGAAGAGAAAGGGGGGAAGCTGCTAGGT	1
21	CCCCGTGCACAGATCCACACCCAGGGTCGAAGCCCTCCGGGGCCCTCACGGGAGGGG	18
22	AATTGCTCCATTATGGCTCACTGCAGCCTCGAAGGTTTAGCTTATTCATTAATCAGTA	17
23	GCTGAAAGTTATTACTTTGTTTTCCCATCGAGGTCCCGCGCACACGCCCCGCGCGCAC	4
24	AGCTGTTCTCTTTAAGGGTGACTCCCTCGACCCCCACGTGCTGAGGGCTCCAGCCAGA	6
25	GCCATGACGGGGCTGGAGGACCAGGAGTTCGACCTGGGTGGTGGGATCTGTGCACGGGG	18
26	GGGACTGCCCCCCCCTGGGCGATCCACCTCGATGTCAAATGGTTCTTGCCTTCACTCT	3
27	GGGTTTACCGTGTAGCCAGGATGGTCTCGAGACCAGCCTGGCCAACATGGCAAAACCC	6
28	CTGTATTAGATTTTACATGCATGAGACTCGAACCAGCCCCGCAACACACTTTCAAGA	9
29	ACAGTCACCGCGCTTACCTGCGCTCCTCGACCATGAATATACTACCAAGGAAATATTT	20
30	CATGTGTTATTTCCCAATCTGGAAGACTCGACCGCTCTGCCCCGCTCTCGCTCTGCAC	21
31	TGCCCCCAAGCCCTCAGACTACAACAATCGACAACGCGATCCACCGGGCCCGAAAGAAG	5
32	ACCAGGGGGCCCAAAGAGGGGGTCAAGGCTCGAATCAAAGGGTTTCTGGATCCCTAGGTGT	12
33	TCTAGAGGGGTATCTCCCAAATCCCACTCGACCCAGCCTCTGGACCAGTGCTCCTGCCA	12
34	CCTGTGGTGCCCCATCTCACCAGGCTCTCGATGATGCCACAAGTGCCGTGCCACAGCAG	17
35	GGGTTTTGCCATGTGGGCCAGGCTGGTCTCGAGATAGGCAAAGAGAGATAGACTAAGTCTG	3
36	CACAGACACAACCCAGGCTCCATCTACTCGATCACAGTACTTATCTGTCTTACGTACAC	6
37	TGTGAAGGGAGGGGAGGAGAAAAAGAAATCGAAACAAGCTTAGAAGCAGACACTTGCCCA	6
38	ACCTCTCTTAATTTTCTCAGCCATTCTTCGACCGCTCTGCCCCGCTCTCGCTCTGCAC	21
39	CTAGAGGAGAGAGGGATGCCAGGCTCTATCGATGACTTCTCCTCCGGGGCGCGGCGCTG	18
40	TCAAGAATCATGGTTCTTAAAGATCACTCGAGGCTGCAGTGAGCTATGATAATGCCACA	6
41	CCACCATCCACCTGGGGCTGAGGGGACCTCGAGTTTGAGCACCCCTCCTGGGTCCTCAG	22
42	GGGTTTACCATGTTGGCCAGGCTGGTCTCGAACTCCCGACCTCAGGTGATCCGCCCCGCC	17
43	ATACCAACCCAGAAATAAAGTCATTCTCGAGGCTGCAGTGAGCTATAATTGCACCACT	4
44	GTTCTCACCCCTGATCACACCTGGTTTATCGAACTCTCTCAGGTTACCCAGACCAAAGA	18
45	TATCTGGCTTAGGCAGAAGGTAGGGGGCTCGAGTGATTATAGAAATCCATATATATATTG	14
46	CTAGAGGAGAGAGGGATGCCAGGCTCTATCGAGTCTGAGTTCGTCCACGTGGTGGCCATC	18
47	AACAGCAGCATTAGATTCTCATAGGAATCGAGTGTCATGAACAATCTTTTCTTTAACA	4
48	CATTCTAGTGCCAGGACCCATCTCAGGTCGACCCCTCCCAAGCCAGCCGCCGAGCAG	2
49	CACTACTACCCAGGAAAGTGATGGGAGGTCGAGATTGCAGGAAATGGAGAGTACATGCCT	X
50	AGTGGCGCAATCTTGGCTAACTGCAGCCTCGAGACCATCTACATGGTGAAACCCCGTCT	15
51	ACGGGCAGACAGGACCCAGCCCATGCCTCGAGCTGAAGGAACATGCTGGCAGGTAGCTC	22
52	ATATTAAATTGCTTACATAGAATGAAGGTGAGGATAATGAAGGGAACCTGCCCTTGAC	1

Table 5.e

	Probe Location				4 kb Sequence Location		
	Start1	End1	Start2	End2	Chr	Start1	End1
1	40446029	40446060	40557923	40557954	17	40446029	40450030
2	81892358	81892389	81927417	81927448	10	81888388	81892389
3	44744656	44744687	44767157	44767188	20	44740686	44744687
4	34696683	34696714	34779780	34779811	21	34696683	34700684
5	91285675	91285706	91314700	91314731	6	91281705	91285706
6	19271977	19272008	19303710	19303741	19	19271977	19275978
7	20560917	20560948	20666337	20666368	9	20556947	20560948

8	77133931	77133962	77220032	77220063	18	77133931	77137932
9	103430366	103430397	103512508	103512539	4	103426396	103430397
10	42305719	42305750	42342568	42342599	22	42301749	42305750
11	49421750	49421781	49458408	49458439	19	49421750	49425751
12	187439656	187439687	187455395	187455426	3	187435686	187439687
13	24467543	24467574	24513928	24513959	1	24467543	24471544
14	42313974	42314005	42342568	42342599	22	42313974	42317975
15	41191135	41191166	41219134	41219165	13	41187165	41191166
16	53403683	53403714	53422397	53422428	17	53399713	53403714
17	77028527	77028558	77094560	77094591	11	77028527	77032528
18	75746612	75746643	75799853	75799884	14	75742642	75746643
19	11807586	11807617	11845619	11845650	1	11807586	11811587
20	228068849	228068880	228135088	228135119	1	228068849	228072850
21	77232184	77232215	77280170	77280201	18	77228214	77232215
22	41162341	41162372	41242678	41242709	17	41162341	41166342
23	106052640	106052671	106067346	106067377	4	106048670	106052671
24	31525914	31525945	31544251	31544282	6	31525914	31529915
25	77160389	77160420	77232184	77232215	18	77156419	77160420
26	187455395	187455426	187486389	187486420	3	187451425	187455426
27	36072356	36072387	36102587	36102618	6	36068386	36072387
28	20322766	20322797	20621547	20621578	9	20318796	20322797
29	39657579	39657610	39725920	39725951	20	39653609	39657610
30	34696683	34696714	34717962	34717993	21	34696683	34700684
31	133465952	133465983	133512403	133512434	5	133465952	133469953
32	130601961	130601992	130676699	130676730	12	130597991	130601992
33	54795918	54795949	54806686	54806717	12	54791948	54795949
34	16842268	16842299	16924802	16924833	17	16842268	16846269
35	187438677	187438708	187454088	187454119	3	187438677	187442678
36	33600698	33600729	33678436	33678467	6	33600698	33604699
37	91285675	91285706	91314700	91314731	6	91281705	91285706
38	34696683	34696714	34779780	34779811	21	34696683	34700684
39	77156086	77156117	77220032	77220063	18	77156086	77160087
40	106483435	106483466	106506791	106506822	6	106483435	106487436
41	37533516	37533547	37544442	37544473	22	37529546	37533547
42	40446029	40446060	40557923	40557954	17	40446029	40450030
43	103418548	103418579	103512508	103512539	4	103414578	103418579
44	20774415	20774446	20868179	20868210	18	20774415	20778416
45	75893651	75893682	75936927	75936958	14	75889681	75893682
46	77133931	77133962	77220032	77220063	18	77133931	77137932
47	185506858	185506889	185543536	185543567	4	185502888	185506889
48	61075534	61075565	61108479	61108510	2	61071564	61075565
49	100610457	100610488	100670898	100670929	X	100610457	100614458
50	80256742	80256773	80285499	80285530	15	80256742	80260743
51	42305719	42305750	42319752	42319783	22	42301749	42305750
52	51402271	51402302	51439728	51439759	1	51402271	51406272

Table 5.f

	4 kb Sequence Location		Probe
	Start2	End2	
1	40557923	40561924	STAT3_17_40446029_40448202_40557923_40558616_RR
2	81927417	81931418	ANXA11_10_81889664_81892389_81927417_81929312_FR

3	44767157	44771158	CD40_20_44739847_44744687_44767157_44770555_FR
4	34775810	34779811	IFNAR1_21_34696683_34697716_34777569_34779811_RF
5	91310730	91314731	MAP3K7_6_91275515_91285706_91312237_91314731_FF
6	19299740	19303741	MEF2B_19_19271977_19273500_19302232_19303741_RF
7	20662367	20666368	MLLT3_9_20556478_20560948_20658310_20666368_FF
8	77216062	77220063	NFATc1_18_77133931_77135912_77218993_77220063_RF
9	103512508	103516509	NFKB1_4_103425293_103430397_103512508_103516923_FR
10	42342568	42346569	TNFRSF13C_22_42302849_42305750_42342568_42346797_FR
11	49454438	49458439	BAX_19_49421750_49425644_49457303_49458439_RF
12	187451425	187455426	BCL6_3_187438677_187439687_187454088_187455426_FF
13	24509958	24513959	IL22RA1_1_24467543_24471444_24512238_24513959_RF
14	42342568	42346569	TNFRSF13C_22_42313974_42315085_42342568_42346797_RR
15	41219134	41223135	FOXO1_13_41184194_41191166_41219134_41220693_FR
16	53418427	53422428	HLF_17_53402207_53403714_53420274_53422428_FF
17	77090590	77094591	PAK1_11_77028527_77036211_77090325_77094591_RF
18	75795883	75799884	FOS_14_75744954_75746643_75795718_75799884_FF
19	11841649	11845650	MTHFR_1_11807586_11814341_11843522_11845650_RF
20	228135088	228139089	WNT9A_1_228068849_228075473_228135088_228140421_RR
21	77280170	77284171	NFATc1_18_77229964_77232215_77280170_77283702_FR
22	41242678	41246679	BRCA1_17_41162341_41168331_41242678_41245761_RR
23	106063376	106067377	TET2_4_106047220_106052671_106063962_106067377_FF
24	31540281	31544282	TNF_6_31525914_31529267_31542458_31544282_RF
25	77228214	77232215	NFATc1_18_77158863_77160420_77229964_77232215_FF
26	187482419	187486420	BCL6_3_187454088_187455426_187484009_187486420_FF
27	36102587	36106588	MAPK13_6_36066232_36072387_36102587_36105090_FR
28	20621547	20625548	MLLT3_9_20319606_20322797_20621547_20622617_FR
29	39725920	39729921	TOP1_20_39656117_39657610_39725920_39729106_FR
30	34713992	34717993	IFNAR1_21_34696683_34697716_34717312_34717993_RF
31	133512403	133516404	SKP1_5_133465952_133470062_133512403_133513591_RR
32	130676699	130680700	FZD10_12_130601147_130601992_130676699_130678204_FR
33	54806686	54810687	ITGA5_12_54787051_54795949_54806686_54808428_FR
34	16924802	16928803	TNFRSF13B_17_16842268_16844133_16924802_16926550_RR
35	187454088	187458089	BCL6_3_187438677_187439687_187454088_187455426_RR
36	33678436	33682437	ITPR3_6_33600698_33604388_33678436_33680494_RR
37	91310730	91314731	MAP3K7_6_91275515_91285706_91312237_91314731_FF
38	34775810	34779811	IFNAR1_21_34696683_34697716_34777569_34779811_RF
39	77216062	77220063	NFATc1_18_77156086_77157023_77218993_77220063_RF
40	106502821	106506822	PRDM1_6_106483435_106485826_106500642_106506822_RF
41	37544442	37548443	IL-2RB_22_37532051_37533547_37544442_37546723_FR
42	40557923	40561924	STAT3_17_40446029_40448202_40557923_40558616_RR
43	103512508	103516509	NFKB1_4_103405171_103418579_103512508_103516923_FR
44	20864209	20868210	CABLES1_18_20774415_20775705_20863570_20868210_RF
45	75932957	75936958	JDP2_14_75883183_75893682_75936165_75936958_FF
46	77216062	77220063	NFATc1_18_77133931_77135912_77218993_77220063_RF
47	185543536	185547537	CASP3_4_185504966_185506889_185543536_185552493_FR
48	61108479	61112480	REL_2_61074693_61075565_61108479_61109187_FR
49	100666928	100670929	BTK_X_100610457_100612966_100667570_100670929_RF
50	80285499	80289500	BCL2A1_15_80256742_80257692_80285499_80286865_RR
51	42315782	42319783	TNFRSF13C_22_42302849_42305750_42318166_42319783_FF
52	51439728	51443729	CDKN2C_1_51402271_51403526_51439728_51440611_RR

Table 5.g

	Inner_primers		
	PCR-Primer1_ID	PCR_Primer1	PCR-Primer2_ID
1	OBD RD048.001	GGAAGACCCCTTTGTGACCTGG	OBD RD048.003
2	OBD RD048.005	CAAGACCTCACCAATGC	OBD RD048.007
3	OBD RD048.009	GAGGAAGGGTGTGCTTTG	OBD RD048.011
4	OBD RD048.013	TGGTCAGACGAGATGCCAAG	OBD RD048.015
5	OBD RD048.017	GTTTGGGACATCAGAAATACAG	OBD RD048.019
6	OBD RD048.021	CTAAGTCTTAAAGGGCCAGAG	OBD RD048.023
7	OBD RD048.025	CAGAGAGGATAGCCTTACAC	OBD RD048.027
8	OBD RD048.029	TGCTTCATGAAACTCAGATGG	OBD RD048.031
9	OBD RD048.033	ACAGCAGTCCAACAATAGTC	OBD RD048.035
10	OBD RD048.037	GTTGAGGCAGACAGAAGAG	OBD RD048.039
11	OBD RD048.041	TCGGAGGTTCTGCTCTCTGAT	OBD RD048.043
12	OBD RD048.045	TTTCTCAATAAAGATTCTCAGAT	OBD RD048.047
13	OBD RD048.049	TAGGATTCACTGAGAAGGTCCCT	OBD RD048.051
14	OBD RD048.053	CCTCTCTCTGAGTCTTGAGTTTC	OBD RD048.055
15	OBD RD048.057	GATGGAGAAAGGAGCAAGGAACCAGG	OBD RD048.059
16	OBD RD048.061	GGCTGATGGTATGGGAATGGGTGG	OBD RD048.063
17	OBD RD048.065	ACCCAGTTACTTGTTGTATTTGC	OBD RD048.067
18	OBD RD048.069	GGCTTTCCCTTCTGTTTTGTTT	OBD RD048.071
19	OBD RD048.073	CTCTGACAAGCAACTCTGAATCC	OBD RD048.075
20	OBD RD048.077	GCTTCAAAGAGTGTGATTATGTAAAA	OBD RD048.079
21	OBD RD048.081	AATAACTGTGGCATCGGAGAGGT	OBD RD048.083
22	OBD RD048.085	AAGTCTCAATGCCACCCAGGCTG	OBD RD048.087
23	OBD RD048.089	TGTATCCCTCCTGTTATCATCCC	OBD RD048.091
24	OBD RD048.093	CAGACACCTCAGGGCTAAGAGCG	OBD RD048.095
25	OBD RD048.097	GGGAGAACC GAACCCCTGGCGGC	OBD RD048.099
26	OBD RD048.101	TACCCACCCGACCACTCCGTA	OBD RD048.103
27	OBD RD048.105	GGAATACAAGTGTGTGCCACCAC	OBD RD048.107
28	OBD RD048.109	CTTTGGGCTTGAAGGCTTTGTTT	OBD RD048.111
29	OBD RD048.113	AGCCTCAGCCGTTTCTGGAGTCTCGG	OBD RD048.115
30	OBD RD048.117	TCTAACCCAGTTCTGCCAGTAA	OBD RD048.119
31	OBD RD048.121	CGGTTCTCACTTTCTTCTTTGC	OBD RD048.123
32	OBD RD048.125	CAAATGAGAGCCTCCAAGACAGC	OBD RD048.127
33	OBD RD048.129	TGGTTCACGGCAAAGTAGTCACA	OBD RD048.131
34	OBD RD048.133	TCTATCACTTTCTGGGCATCAG	OBD RD048.135
35	OBD RD048.137	CCTGCCTCAGCCTCCCAAGTAGC	OBD RD048.139
36	OBD RD048.141	TGGATGGAACCCCTGAGCCACACAGC	OBD RD048.143
37	OBD RD048.145	GGTTAGGTCTTCTGCCTTCAAAG	OBD RD048.147
38	OBD RD048.149	CAGACGAGATGCCAAGTGCTTTA	OBD RD048.151
39	OBD RD048.153	TGCTGGAGTGAAAACGCCTCTTT	OBD RD048.155
40	OBD RD048.157	TCATAATGTCAGTGTCTGTTC	OBD RD048.159
41	OBD RD048.161	GCTTTCTGAATCTTTCCTGGTG	OBD RD048.163
42	OBD RD048.165	CCTGCCTCAGCCGCCGAGTAGC	OBD RD048.167
43	OBD RD048.169	CCTCCCACTTTTGATGGCACTGC	OBD RD048.171
44	OBD RD048.173	CCCACATTTCTTCTTCTGTT	OBD RD048.175
45	OBD RD048.177	CTTCTATGGGTGATGACCTGACA	OBD RD048.179
46	OBD RD048.181	TGCTGGAGTGAAAACGCCTCTTT	OBD RD048.183
47	OBD RD048.185	CCATCGCTCACATCATTACCTGA	OBD RD048.187
48	OBD RD048.189	ACATACAGTCAGTAGGAGCCTTG	OBD RD048.191
49	OBD RD048.193	GCTCCAACACTCACATCTAACAC	OBD RD048.195
50	OBD RD048.197	GTATTTTGTGTTTGTGTTT	OBD RD048.199
51	OBD RD048.201	CTCCAAGACACCACTGCCGTTGAGGC	OBD RD048.203

52	OBD RD048.205	GCCTCATTTCTGTCCTCCTTTGA	OBD RD048.207
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Table 5.h

	Inner_primers PCR_Primer2	Gene	Marker	GLMNET
1	TCACCATTCGTTCAACACAC	STAT3	OBD RD048.001.003	2.08E-08
2	CAGTTGTGGAGGCTCAATAC	ANXA11	OBD RD048.005.007	0.00000056
3	GGAAGGAAAGCCAGTGAAG	CD40	OBD RD048.009.011	0.00000449
4	ACCCTAGAGTCTTGGACAG	IFNAR1	OBD RD048.013.015	0.000000838
5	ATCCCTAGGGCACTGAAC	MAP3K7	OBD RD048.017.019	0.00000156
6	CATACAAGGATGGAGTGACC	MEF2B	OBD RD048.021.023	0.00000137
7	AGTGTCTTGCCCTGTAATC	MLLT3	OBD RD048.025.027	0.0000046
8	AGCCTAAGCTGAGGAACTC	NFATc1	OBD RD048.029.031	0.00000181
9	AACTCCTAATGAGAAAGTCTGC	NFKB1	OBD RD048.033.035	0.00000178
10	GGTCGGGTAGTAGAGAGTG	TNFRSF13C	OBD RD048.037.039	0.000000402
11	GGACAGGTAACACGGGTCTCCC	BAX	OBD RD048.041.043	-0.000000273
12	TACCCACCCCGACCACTCCGTA	BCL6	OBD RD048.045.047	0.000000154
13	CACCTTGCGTAGAGGCAGTAGACCCC	IL22RA1	OBD RD048.049.051	-0.000000967
14	AATGTCCTCCGAGCCGCTGCTGG	TNFRSF13C	OBD RD048.053.055	2.13E-08
15	GGTGTGAGGTAAGAAGTCATAGCCAT	FOXO1	OBD RD048.057.059	0.00000117
16	CACAGAGCCTGCCATCCTCACAT	HLF	OBD RD048.061.063	0.000000324
17	ACTACAGGTGCCCCGCCACAAGGC	PAK1	OBD RD048.065.067	-4.86E-08
18	GGGATGGAGCAGGAAGGAGAGAGAGG	FOS	OBD RD048.069.071	0.00000028
19	TATGTCTTGCCCTGTGCTGCGGCTCC	MTHFR	OBD RD048.073.075	0.000000306
20	ATCAGGTCCCGACTTCTTGCGC	WNT9A	OBD RD048.077.079	0.00000596
21	AACACCGAGACACACCGAGTCCCTCC	NFATc1	OBD RD048.081.083	0.000000396
22	GACTGCTCAGGGCTATCCTCTCAG	BRCA1	OBD RD048.085.087	-0.000000314
23	AGAGGTGCCAGTGGGTGGAGGCG	TET2	OBD RD048.089.091	0.000000105
24	GCTCCTCCTCCTGCTGTCGCCAG	TNF	OBD RD048.093.095	0.00000119
25	GGGCGGCTGTGAAACTGAGGTCC	NFATc1	OBD RD048.097.099	0.00000232
26	AGGAAAGGCTTCACTGAGCATCA	BCL6	OBD RD048.101.103	0.00000193
27	TTTGATTCTTAGTAGAGACGGG	MAPK13	OBD RD048.105.107	-5.18E-08
28	GCCCGCCGCCCTGCCTTTCTGAAT	MLLT3	OBD RD048.109.111	0.00000156
29	CTCTGTGGACAGAAACCTAC	TOP1	OBD RD048.113.115	4.09E-08
30	TGAGCGACCAGACCGTTGCTGTGTGC	IFNAR1	OBD RD048.117.119	0.000000517
31	CGCCCACTGAAGTGAAGGGTCGTG	SKP1	OBD RD048.121.123	0.000000786
32	AGAAGTGCCAGTCTACATACACC	FZD10	OBD RD048.125.127	0.00000223
33	AGGCAGACACAGAGCAGAGCAGAGGC	ITGA5	OBD RD048.129.131	0.00000124
34	GGTCTCCCCTCTACCACACTGGCAT	TNFRSF13B	OBD RD048.133.135	0.000000638
35	TGAAGTTTGGTAAAGACCGAGTT	BCL6	OBD RD048.137.139	0.000000206
36	TGTTCTTGCTTTCCTCCAGGTTG	ITPR3	OBD RD048.141.143	0.000000731
37	CTGTGGGTGGAAGAGGCTCAGGCATC	MAP3K7	OBD RD048.145.147	0.00000156
38	TGAGCGACCAGACCGTTGCTGTGTGC	IFNAR1	OBD RD048.149.151	0.000000838
39	TTTCTCCTCTCCGAAGACCGCAGCC	NFATc1	OBD RD048.153.155	0.00000132
40	CTCTCTCTCTGCACCCAGGCTG	PRDM1	OBD RD048.157.159	0.00000243
41	CGTAGGCATCCGTGGGTGTGACCAGT	IL-2RB	OBD RD048.161.163	0.000000378
42	CGCCTGTAATCCAGAACTTTGG	STAT3	OBD RD048.165.167	2.08E-08
43	GTCTCACTCTGTTGCCAGGCTG	NFKB1	OBD RD048.169.171	0.00000135
44	TTCTTGATAAAATGAATCTTCTTA	CABLES1	OBD RD048.173.175	0.000000717
45	TGGAGTTTGCTGTGGGCACTGAGGCG	JDP2	OBD RD048.177.179	0.00000334
46	CCACCACCATCAGCCAGTGCCACG	NFATc1	OBD RD048.181.183	0.00000181

47	CAATGCCAGGTCTTCATACTCTA	CASP3	OBD RD048.185.187	0.00000305
48	ACCCAGCGTCGCCGTCCACCGTA	REL	OBD RD048.189.191	0.00000123
49	GGTCCACATTCTCACGAACCGCTCC	BTK	OBD RD048.193.195	0.00000409
50	CATTCTCCTGCCTCAGCCTCCTG	BCL2A1	OBD RD048.197.199	0.000000299
51	CTAAATGTGTGTGTCTTGGAGC	TNFRSF13C	OBD RD048.201.203	0.000000179
52	TGCTTCACCAGGAAGTCCACCACCCG	CDKN2C	OBD RD048.205.207	0.0000123

Table 5.i

	Probe	GeneLocus	Probe_Count_Total
53	ANXA11_10_81889664_81892389_81927417_81929312_FR	ANXA11	136
54	CD40_20_44737133_44739370_44777294_44780862_RR	CD40	148
55	CREB3L2_7_137532509_137535848_137608464_137613205_FR	CREB3L2	168
56	MyD88_3_38159544_38161117_38182050_38188284_FR	MyD88	80
57	MEF2B_19_19255724_19257122_19271977_19273500_FF	MEF2B	448
58	IL-2RB_22_37569072_37572860_37583052_37586677_RR	IL-2RB	72
59	FRAP1_1_11321482_11322337_11347781_11348658_FF	FRAP1	704
60	BCL6_3_187438677_187439687_187452395_187454091_FF	BCL6	240
61	MMP9_20_44635898_44638559_44669235_44671514_FF	MMP9	68
62	MAP3K7_6_91275515_91285706_91296544_91297579_FR	MAP3K7	308
63	MLLT3_9_20556478_20560948_20658310_20666368_FF	MLLT3	120
64	HLF_17_53404056_53408147_53420274_53422428_RF	HLF	104
65	SIRT1_10_69650583_69655218_69676432_69678199_FR	SIRT1	96
66	NFATc1_18_77124213_77127824_77280170_77283702_RF	NFATc1	608
67	TNFRSF13C_22_42302849_42305750_42342568_42346797_FR	TNFRSF13C	488
68	STAT3_17_40456120_40457219_40580136_40581714_RF	STAT3	1108
69	NFKB1_4_103512508_103516923_103561903_103565015_RF	NFKB1	96
70	MEF2B_19_19271977_19273500_19302232_19303741_RF	MEF2B	448
71	CD40_20_44739847_44744687_44767157_44770555_FR	CD40	148
72	MAPK10_4_87408248_87409426_87514697_87515355_RF	MAPK10	668
73	FRAP1_1_11190905_11194522_11269915_11272450_RR	FRAP1	704
74	NFKB1_4_103425293_103430397_103512508_103516923_FR	NFKB1	96
75	MAPK10_4_87373087_87377906_87514697_87515355_RF	MAPK10	668
76	JAK3_19_17889333_17890586_17934729_17936992_FR	JAK3	60
77	TNFRSF13C_22_42329800_42332095_42352233_42353781_FR	TNFRSF13C	488
78	TET2_4_106058602_106063965_106118157_106119978_RR	TET2	104
79	NAE1_16_66835284_66840537_66902726_66909724_RF	NAE1	64
80	TNFRSF13C_22_42335475_42336871_42362266_42363517_RR	TNFRSF13C	488
81	NFATc1_18_77151077_77154182_77274975_77276499_RR	NFATc1	608
82	BRCA1_17_41214832_41217070_41227254_41229572_RR	BRCA1	297
83	MLLT3_9_20377197_20385409_20556478_20560948_RF	MLLT3	120
84	PCDHGA6/B2/B4_5_140751685_140753982_140892508_140893138_FF	PCDHGA6/B2/B4	108
85	MAPK10_4_87166373_87167382_87408248_87409426_RR	MAPK10	668
86	BTK_X_100646274_100647902_100689454_100691928_RR	BTK	404
87	BTK_X_100625073_100626595_100689454_100691928_RR	BTK	404
88	BTK_X_100587279_100590348_100627655_100629872_FR	BTK	404
89	BTK_X_100627655_100629872_100647899_100654354_RR	BTK	404
90	BTK_X_100647899_100654354_100673203_100675145_RF	BTK	404
91	BTK_X_100602468_100603585_100647899_100654354_RR	BTK	404
92	BTK_X_100610457_100612966_100667570_100670929_RF	BTK	404

Table 5.j

	Probe_Count_Sig	HyperG_Stats	FDR_HyperG	Percent_Sig
53	83	0.000391435	0.005936759	61.03
54	64	0.802231212	0.999999997	43.24
55	47	0.999999703	0.999999997	27.98
56	27	0.991982598	0.999999997	33.75
57	216	0.227265083	0.590889215	48.21
58	24	0.991079692	0.999999997	33.33
59	359	0.006468458	0.042044978	50.99
60	86	0.999659646	0.999999997	35.83
61	25	0.957692148	0.999999997	36.76
62	113	0.999793469	0.999999997	36.69
63	39	0.999297311	0.999999997	32.5
64	44	0.824486728	0.999999997	42.31
65	64	0.0000456596437717468	0.001038757	66.67
66	213	0.999999997	0.999999997	35.03
67	280	0.000000444123116904245	0.0000202076018191431	57.38
68	615	0.000000000125197189743782	0.0000000113929442666842	55.51
69	27	0.999920864	0.999999997	28.12
70	216	0.227265083	0.590889215	48.21
71	64	0.802231212	0.999999997	43.24
72	244	0.999999947	0.999999997	36.53
73	359	0.006468458	0.042044978	50.99
74	27	0.999920864	0.999999997	28.12
75	244	0.999999947	0.999999997	36.53
76	31	0.243296934	0.615000583	51.67
77	280	0.000000444123116904245	0.0000202076018191431	57.38
78	58	0.033733587	0.145711753	55.77
79	36	0.071920785	0.221674157	56.25
80	280	0.000000444123116904245	0.0000202076018191431	57.38
81	213	0.999999997	0.999999997	35.03
82	173	0.0000217239097176038	0.000658959	58.25
83	39	0.999297311	0.999999997	32.5
84	36	0.997865052	0.999999997	33.33
85	244	0.999999947	0.999999997	36.53
86	182	0.722716922	0.999999997	45.05
87	182	0.722716922	0.999999997	45.05
88	182	0.722716922	0.999999997	45.05
89	182	0.722716922	0.999999997	45.05
90	182	0.722716922	0.999999997	45.05
91	182	0.722716922	0.999999997	45.05
92	182	0.722716922	0.999999997	45.05

Table 5.k

	logFC	AveExpr	t	P.Value	adj.P.Val
53	0.146814815	0.146814815	3.078806942	0.015395162	0.044697142
54	0.147791337	0.147791337	1.633707333	0.141477876	0.232950641
55	0.148349454	0.148349454	3.08222473	0.015316201	0.044558199
56	0.153758518	0.153758518	1.99378109	0.081784292	0.154640968
57	0.156103192	0.156103192	4.16566548	0.003235665	0.015172146
58	0.161073376	0.161073376	2.527153349	0.035788708	0.08344291

59	0.171050829	0.171050829	3.890660293	0.004727539	0.019533312
60	0.174144322	0.174144322	2.584598623	0.0327468	0.078039912
61	0.18112944	0.18112944	2.685388848	0.028032588	0.069592429
62	0.19092131	0.19092131	3.73604023	0.005879148	0.022572713
63	0.194400918	0.194400918	2.492510608	0.037760627	0.086786653
64	0.195707712	0.195707712	2.71102351	0.026948639	0.067475149
65	0.204252124	0.204252124	3.975216299	0.004202345	0.018041687
66	0.210054656	0.210054656	2.338335953	0.047960033	0.103796265
67	0.210247736	0.210247736	2.234440593	0.056352274	0.11719604
68	0.213090816	0.213090816	2.087748617	0.07073665	0.138732545
69	0.226250319	0.226250319	2.25409429	0.054659526	0.114573853
70	0.246810388	0.246810388	5.953590771	0.000359019	0.0048119
71	0.247739738	0.247739738	4.372950932	0.002449301	0.012749359
72	0.248261394	0.248261394	6.173828851	0.000282141	0.004322646
73	0.251556432	0.251556432	4.318526719	0.00263344	0.013280362
74	0.253919456	0.253919456	1.95465634	0.086862876	0.161472968
75	0.256754187	0.256754187	5.535121315	0.000577231	0.005715942
76	0.257160612	0.257160612	3.449233265	0.008887517	0.030040982
77	0.259132781	0.259132781	4.366813249	0.002469352	0.012816471
78	0.287279843	0.287279843	2.539709619	0.035100109	0.082104681
79	0.31600033	0.31600033	3.153558526	0.013761553	0.041279885
80	0.358221647	0.358221647	3.100524122	0.014900586	0.043739937
81	0.364193755	0.364193755	3.369619436	0.009987239	0.03271603
82	0.453457772	0.453457772	3.247156175	0.011968978	0.037200176
83	0.180533568	0.180533568	5.147835975	0.000914678	0.007187473
84	0.182697701	0.182697701	5.877748203	0.00039063	0.004938906
85	-0.148364769	-0.148364769	-4.986366569	0.001115061	0.008026755
86	-0.538084185	-0.538084185	-6.494881534	0.000200669	0.003807401
87	-0.545447375	-0.545447375	-6.02027801	0.000333544	0.004684915
88	-0.554745602	-0.554745602	-8.383072026	0.0000337	0.002483007
89	0.503059535	0.503059535	6.535294395	0.000192409	0.003731412
90	0.36623319	0.36623319	5.026075307	0.001061678	0.007815282
91	0.338959712	0.338959712	4.957835746	0.001155226	0.008192382
92	0.127634089	0.127634089	5.070996787	0.001004634	0.007593027

Table 5.1

	B	FC	FC_1	LS	Loop Detected
53	-3.422371897	1.107122465	1.107122465	1	DBLCL
54	-5.595015473	1.1078721	1.1078721	1	DBLCL
55	-3.417108405	1.108300771	1.108300771	1	DBLCL
56	-5.084251662	1.112463898	1.112463898	1	DBLCL
57	-1.806028568	1.114273349	1.114273349	1	DBLCL
58	-4.275957963	1.118118718	1.118118718	1	DBLCL
59	-2.201619835	1.125878252	1.125878252	1	DBLCL
60	-4.187168091	1.128295002	1.128295002	1	DBLCL
61	-4.031097147	1.133771131	1.133771131	1	DBLCL
62	-2.428494364	1.141492444	1.141492444	1	DBLCL
63	-4.329420018	1.14424891	1.14424891	1	DBLCL
64	-3.991368624	1.145285841	1.145285841	1	DBLCL
65	-2.078878648	1.152088963	1.152088963	1	DBLCL

66	-4.56625722	1.156732005	1.156732005	1	DBLCL
67	-4.724458483	1.156886824	1.156886824	1	DBLCL
68	-4.945085913	1.159168918	1.159168918	1	DBLCL
69	-4.694639254	1.169790614	1.169790614	1	DBLCL
70	0.493437892	1.186580836	1.186580836	1	DBLCL
71	-1.514951748	1.18734545	1.18734545	1	DBLCL
72	0.744313598	1.187774853	1.187774853	1	DBLCL
73	-1.590768631	1.190490767	1.190490767	1	DBLCL
74	-5.141601134	1.192442298	1.192442298	1	DBLCL
75	-0.002180366	1.194787614	1.194787614	1	DBLCL
76	-2.856981615	1.195124248	1.195124248	1	DBLCL
77	-1.523480143	1.196759104	1.196759104	1	DBLCL
78	-4.25656399	1.220337199	1.220337199	1	DBLCL
79	-3.307417374	1.24487452	1.24487452	1	DBLCL
80	-3.388938618	1.281844844	1.281844844	1	DBLCL
81	-2.977498786	1.287162103	1.287162103	1	DBLCL
82	-3.164028291	1.369318233	1.369318233	1	DBLCL
83	-0.483711076	1.13330295	1.13330295	1	DBLCL
84	0.405473595	1.135004251	1.135004251	1	DBLCL
85	-0.691112407	0.902272569	-1.108312537	-1	Ctrl
86	1.098164859	0.688684835	-1.452043008	-1	Ctrl
87	0.570114643	0.685178898	-1.459472852	-1	Ctrl
88	2.923843236	0.680777092	-1.46890959	-1	Ctrl
89	1.14173325	1.417215879	1.417215879	1	DBLCL
90	-0.639742862	1.288982958	1.288982958	1	DBLCL
91	-0.728168867	1.26484422	1.26484422	1	DBLCL
92	-0.581917188	1.092500613	1.092500613	1	DBLCL

Table 5.m

	Probe sequence	Probe Location
	60 mer	Chr
53	GAGGGGCTCTGGAGGGGCGGGTCTCTCGATGCCTGGCCTCCACAGCACATGTGAGCA	10
54	AATGAGGAAGTACAGCAGGAGGCAGCATCGAAACCTGGGATGCTAGTAACCCTACCTG	20
55	TCCAATCACCTCCCACAGGTCCCTCCCTCGATCCTGTGCTTTTCTGCTGCAGGTTTCA	7
56	AGTGGCGTGATCATGGTTCACTGAAGCCTCGAAAAGAGGTTGGCTAGAAGGCCACGGGGT	3
57	GAGGACACGGCGGGGGGCCATCACCCCTCGAACAGGAGCTGTCCCTCCCAGGAGCAGGC	19
58	GAGCCAGGTTTTGCAGGACCTGGGATATTCGAGACCAGTCTGGGCAACATAGTGAGACCC	22
59	TGGGGGTCCCGGGGAGGTGGGCGTTGCCTCGAATCTGGTCAAACCTACCCAACTCATC	1
60	AGATCCGTGTCTGCCTGCAGATACAAAATCGAGTTGGGCTGGGGAGAGGAGGAGATAGGT	3
61	CACTCGGGTGGCAGAGATGCGTGGAGAGTGCATGTGTCCCAAATTGATCTCACCTCCAC	20
62	TGTGAAGGGAGGGGAGGAGAGAAAAGAAAATCGATCATCTCACCGCCGAAGACGAGGAGGA	6
63	ACATTTCAAATCCTCTCTTCTAGCTACCTCGAACTTCTGAGCTCAAGCAATCTTCCACCT	9
64	TTCTGACAGAGGGTTGGGGAGAGGGGTGTCGACCTCCTAAAGTGCTGGGATTACAGGCGT	17
65	GGAGGATGGGGAGGGTATGTAAATATTGTGATAGAGCAAGGAAACCAGAAAGGTGTAAT	10
66	GTATGAGTGTGGGTGTGTGGATGTGGCCTCGAGATCGCGCCACTGCACTCCAGCCTGGGC	18
67	ACGGGCAGACAGGACCCAGCCCATGCCTCGACCACTCCCGGGGGGATCGGGACACCGC	22
68	AGTGGTGCATCTCAGCTTGTGAGCCTCGAGGAATTTCTAATGATAGATCCAGACCTC	17
69	TCATTCTGGGGATTATCTTTTCATTTCTCGAGGCTGCAGTGAGCTATAATTGCACCACT	4
70	TGGGGGAGCTCTGGGGTGGGGTAGCGGTGATGGTCTGATGCCTCTCAGAAGGCCTT	19

71	GAGGCTTTTATGCAGGAAAGTGTCCAGTCGAGGGACTGGCAGCAGGGGGACAGCAAGGG	20
72	GAGCTGGATGCCAGGCGGGCCAATGAGGTGCGATTGCAATGCAGGATCCTATGCTGGATTC	4
73	AACAGGCAGGAGCAGCTGTTCTCAGCATCGAACCTATTTATTTACTTATTTTTTGAGA	1
74	TCTTTATGGTGTCTCTTTATATATTTACTCGAGGCTGCAGTGAGCTATAATTGCACCACT	4
75	GAGCTGGATGCCAGGCGGGCCAATGAGGTGCAACACGATATGAACAGGACATCTGTTACA	4
76	GGGTGGAGTCAGGGAGGGGTGGGGGACGTCGAGTCTTGCTTGACCCAGAGCAGCTCCCT	19
77	GGGTTTACCGTGTTACTCAGGCTGGTCTCGAAGTCCTGGGCTCAAGCAATCCACCCGCT	22
78	CAAATACTCATGTGTATGGGCAAAAACTCGAGTAGTTGGAACCTCAAGTGTCAAAACAT	4
79	GACGGGCCGATTGCCTGAGCTCAGGAGTTCGACCCTTCTCACGTGGGCTAAGGGCCTGAC	16
80	ACTAGCTGGGTGACCTAGACAGTTTGTTCGAGGCTACAGTGAGCTGTGATAGTGCCACT	22
81	TGTTGTATCCATTATTGAAAGTGAGTATCGAGGCTGCAGTGAGCTGAGATCATTCCACT	18
82	GACAGGCAGATTGCCTGAGCTCAGGAGTTCGACATCTCTACACTCATTCTTTCTACTCAG	17
83	ACATTTCAAATCCTCTCTTCTAGTACCTCGAAACACCACTACTTGTGCTAGTTTACAATGA	9
84	CGGTGTCTGGTGAGTTTTAACATCCTTGTGCGAGCTGCAGACTTGCTTTGGAAGAATCAC	5
85	GCTCAGCAAATGAATGTTTTCAAAGCACTCGATTGCAATGCAGGATCCTATGCTGGATTC	4
86	GTGCTTCAAGCAGAGCTTCTCCCTCCGTCGAACTCCTGACCTCGTGATCCGCCTGCCTC	X
87	ATCCTAACTGCTGAAGTCTGTGTTTTATCGAACTCCTGACCTCGTGATCCGCCTGCCTC	X
88	GGCTTGCCTAAAAAGTAAACAAAACAGTCGAACTCCTGCTCATGATCCGCCTGCCTTGG	X
89	CCAAGGCAGGCGGATCATGAGCAGGAGTTCGAGACCAGCCTGGCCAAGATAGTGAAACCC	X
90	GCCGAGGCGGGTGGATCAGGTCAGGAGTTCGAGACCAGCCTGGCCAAGATAGTGAAACCC	X
91	GGCGGGTGGATCACTTGAGGTCAGGAGTTCGAGACCAGCCTGGCCAAGATAGTGAAACCC	X
92	CACTACTACCCAGGAAAGTGATGGGAGGTCGAGATTGCAAGAAATGGAGAGTACATGCCT	X

Table 5.n

	Probe Location				4 kb Sequence Location		
	Start1	End1	Start2	End2	Chr	Start1	End1
53	81892358	81892389	81927417	81927448	10	81888388	81892389
54	44737133	44737164	44777294	44777325	20	44737133	44741134
55	137535817	137535848	137608464	137608495	7	137531847	137535848
56	38161086	38161117	38182050	38182081	3	38157116	38161117
57	19257091	19257122	19273469	19273500	19	19253121	19257122
58	37569072	37569103	37583052	37583083	22	37569072	37573073
59	11322306	11322337	11348627	11348658	1	11318336	11322337
60	187439656	187439687	187454060	187454091	3	187435686	187439687
61	44638528	44638559	44671483	44671514	20	44634558	44638559
62	91285675	91285706	91296544	91296575	6	91281705	91285706
63	20560917	20560948	20666337	20666368	9	20556947	20560948
64	53404056	53404087	53422397	53422428	17	53404056	53408057
65	69655187	69655218	69676432	69676463	10	69651217	69655218
66	77124213	77124244	77283671	77283702	18	77124213	77128214
67	42305719	42305750	42342568	42342599	22	42301749	42305750
68	40456120	40456151	40581683	40581714	17	40456120	40460121
69	103512508	103512539	103564984	103565015	4	103512508	103516509
70	19271977	19272008	19303710	19303741	19	19271977	19275978
71	44744656	44744687	44767157	44767188	20	44740686	44744687
72	87408248	87408279	87515324	87515355	4	87408248	87412249
73	11190905	11190936	11269915	11269946	1	11190905	11194906
74	103430366	103430397	103512508	103512539	4	103426396	103430397
75	87373087	87373118	87515324	87515355	4	87373087	87377088
76	17890555	17890586	17934729	17934760	19	17886585	17890586
77	42332064	42332095	42352233	42352264	22	42328094	42332095

78	106058602	106058633	106118157	106118188	4	106058602	106062603
79	66835284	66835315	66909693	66909724	16	66835284	66839285
80	42335475	42335506	42362266	42362297	22	42335475	42339476
81	77151077	77151108	77274975	77275006	18	77151077	77155078
82	41214832	41214863	41227254	41227285	17	41214832	41218833
83	20377197	20377228	20560917	20560948	9	20377197	20381198
84	140753951	140753982	140893107	140893138	5	140749981	140753982
85	87166373	87166404	87408248	87408279	4	87166373	87170374
86	100646274	100646305	100689454	100689485	X	100646274	100650275
87	100625073	100625104	100689454	100689485	X	100625073	100629074
88	100590317	100590348	100627655	100627686	X	100586347	100590348
89	100627655	100627686	100647899	100647930	X	100627655	100631656
90	100647899	100647930	100675114	100675145	X	100647899	100651900
91	100602468	100602499	100647899	100647930	X	100602468	100606469
92	100610457	100610488	100670898	100670929	X	100610457	100614458

Table 5.o

	4 kb Sequence Location		Probe
	Start2	End2	
53	81927417	81931418	ANXA11_10_81889664_81892389_81927417_81929312_FR
54	44777294	44781295	CD40_20_44737133_44739370_44777294_44780862_RR
55	137608464	137612465	CREB3L2_7_137532509_137535848_137608464_137613205_FR
56	38182050	38186051	MyD88_3_38159544_38161117_38182050_38188284_FR
57	19269499	19273500	MEF2B_19_19255724_19257122_19271977_19273500_FF
58	37583052	37587053	IL-2RB_22_37569072_37572860_37583052_37586677_RR
59	11344657	11348658	FRAP1_1_11321482_11322337_11347781_11348658_FF
60	187450090	187454091	BCL6_3_187438677_187439687_187452395_187454091_FF
61	44667513	44671514	MMP9_20_44635898_44638559_44669235_44671514_FF
62	91296544	91300545	MAP3K7_6_91275515_91285706_91296544_91297579_FR
63	20662367	20666368	MLLT3_9_20556478_20560948_20658310_20666368_FF
64	53418427	53422428	HLF_17_53404056_53408147_53420274_53422428_RF
65	69676432	69680433	SIRT1_10_69650583_69655218_69676432_69678199_FR
66	77279701	77283702	NFATc1_18_77124213_77127824_77280170_77283702_RF
67	42342568	42346569	TNFRSF13C_22_42302849_42305750_42342568_42346797_FR
68	40577713	40581714	STAT3_17_40456120_40457219_40580136_40581714_RF
69	103561014	103565015	NFKB1_4_103512508_103516923_103561903_103565015_RF
70	19299740	19303741	MEF2B_19_19271977_19273500_19302232_19303741_RF
71	44767157	44771158	CD40_20_44739847_44744687_44767157_44770555_FR
72	87511354	87515355	MAPK10_4_87408248_87409426_87514697_87515355_RF
73	11269915	11273916	FRAP1_1_11190905_11194522_11269915_11272450_RR
74	103512508	103516509	NFKB1_4_103425293_103430397_103512508_103516923_FR
75	87511354	87515355	MAPK10_4_87373087_87377906_87514697_87515355_RF
76	17934729	17938730	JAK3_19_17889333_17890586_17934729_17936992_FR
77	42352233	42356234	TNFRSF13C_22_42329800_42332095_42352233_42353781_FR
78	106118157	106122158	TET2_4_106058602_106063965_106118157_106119978_RR
79	66905723	66909724	NAE1_16_66835284_66840537_66902726_66909724_RF
80	42362266	42366267	TNFRSF13C_22_42335475_42336871_42362266_42363517_RR
81	77274975	77278976	NFATc1_18_77151077_77154182_77274975_77276499_RR
82	41227254	41231255	BRCA1_17_41214832_41217070_41227254_41229572_RR
83	20556947	20560948	MLLT3_9_20377197_20385409_20556478_20560948_RF
84	140889137	140893138	PCDHGA6/B2/B4_5_140751685_140753982_140892508_140893138_FF

85	87408248	87412249	MAPK10_4_87166373_87167382_87408248_87409426_RR
86	100689454	100693455	BTK_X_100646274_100647902_100689454_100691928_RR
87	100689454	100693455	BTK_X_100625073_100626595_100689454_100691928_RR
88	100627655	100631656	BTK_X_100587279_100590348_100627655_100629872_FR
89	100647899	100651900	BTK_X_100627655_100629872_100647899_100654354_RR
90	100671144	100675145	BTK_X_100647899_100654354_100673203_100675145_RF
91	100647899	100651900	BTK_X_100602468_100603585_100647899_100654354_RR
92	100666928	100670929	BTK_X_100610457_100612966_100667570_100670929_RF

Table 5.p

	Inner primers		
	PCR-Primer1_ID	PCR_Primer1	PCR-Primer2_ID
53	OBD RD048.209	GGCTCGTAACAAACCCCTGACCCAG	OBD RD048.211
54	OBD RD048.213	TCCCCATTACCCATCAGTGCTCCCC	OBD RD048.215
55	OBD RD048.217	GGAGAGGCAGAGCAGAGAGTGAAGGG	OBD RD048.219
56	OBD RD048.221	GACAGCAGTTTCTAAGCCTGGCA	OBD RD048.223
57	OBD RD048.225	TTTGGAGGACTGGGACTTGCCGT	OBD RD048.227
58	OBD RD048.229	AACTGAAAGAAAGACCCAGAGGC	OBD RD048.231
59	OBD RD048.233	GACCCAAAGGGCAATACCAGAGC	OBD RD048.235
60	OBD RD048.237	CACGCTCGCCATCATTGAAAC	OBD RD048.239
61	OBD RD048.241	TCCCTTCATCCACAGGAATACCT	OBD RD048.243
62	OBD RD048.245	GGTTAGGTCTTCTGCCTCAAAG	OBD RD048.247
63	OBD RD048.249	GTGTAACAATCAAGTCAGGGAAT	OBD RD048.251
64	OBD RD048.253	CACAGAGCCTGCCATCCTCATAT	OBD RD048.255
65	OBD RD048.257	AAATAAGTAAGGACAAAGAGTGC	OBD RD048.259
66	OBD RD048.261	TCGCCTACGGCTTGTTACGCACAGC	OBD RD048.263
67	OBD RD048.265	GCTTATTTACAAGACGAACCCGC	OBD RD048.267
68	OBD RD048.269	TTCTGTTGTCCAGGCTTGAGTGC	OBD RD048.271
69	OBD RD048.273	CACTATTGAGTTCTAAGAGTTCT	OBD RD048.275
70	OBD RD048.277	GGAACCCACGCCCTCCCCTAAGTCTT	OBD RD048.279
71	OBD RD048.281	GGTGTGCTTTGCCAGGATAAGAA	OBD RD048.283
72	OBD RD048.285	TCTCCCTGGCGACCTCGTCCCTA	OBD RD048.287
73	OBD RD048.289	TGTTTGCTTTATGGACACACAGA	OBD RD048.291
74	OBD RD048.293	CATTTACTACTCTCATACCATA	OBD RD048.295
75	OBD RD048.297	ACTCTGCCGCTCGGTCAACACCTGA	OBD RD048.299
76	OBD RD048.301	GACAAGGGAGGGAGGAGGATGGG	OBD RD048.303
77	OBD RD048.305	CCTGCCTCAGCCTCCAAGTAGC	OBD RD048.307
78	OBD RD048.309	GTGAACTCAGCCAAGCACAGTGGTGG	OBD RD048.311
79	OBD RD048.313	TTCTTTACCCCTGTCACTCACCT	OBD RD048.315
80	OBD RD048.317	TGGTTGGAAGTAGCCCTGATTCA	OBD RD048.319
81	OBD RD048.321	GTTGCCTTGTTATCTGCCTGGTT	OBD RD048.323
82	OBD RD048.325	GTAATCCTAACACTGTGGGAGGC	OBD RD048.327
83	OBD RD048.329	GGGAGCATTGTGGGCTAACAGGAGAC	OBD RD048.331
84	OBD RD048.333	TCGTAGGCAACATCGTCAAGGAT	OBD RD048.335
85	OBD RD048.337	CTGGGCAACAGAGTGAGAGCCTG	OBD RD048.339
86	OBD RD051.001	TGCTACCTCTGACTACAGGGTGG	OBD RD051.003
87	OBD RD051.005	GCTGACTGAAGATTCTGCCTTTC	OBD RD051.007
88	OBD RD051.009	TAGGATGGCAAGCAGCATTGGCT	OBD RD051.011
89	OBD RD051.013	CACGCCTGTAATCCCAGCACTTTGG	OBD RD051.015
90	OBD RD051.017	CACGCCTGTAATCCCAGCACTCTG	OBD RD051.019
91	OBD RD051.021	ATGCCTGTAATCCCAGCACTTTGG	OBD RD051.023
92	OBD RD051.025	CCACCATTCTGTGCTCCAACACTC	OBD RD051.027

Table 5.q

	Inner primers			
	PCR_Primer2	Gene	Marker	GLMNET
53	ACAGTTGTGGAGGCTCAATACCT	ANXA11	OBD RD048.209.211	0.00000056
54	CGGTAACAGACACGGAGTGAAAT	CD40	OBD RD048.213.215	0.00000222
55	GCAGGGACTGAGAAACATAGGAT	CREB3L2	OBD RD048.217.219	3.82E-08
56	TGGACCCAGGGCAGGGCTTCAT	MyD88	OBD RD048.221.223	0.000000196
57	TCAGACCCTCCTTCCACCTCTC	MEF2B	OBD RD048.225.227	0.00000288
58	CCCCTTCTCTGCTGCTACCATCCAG	IL-2RB	OBD RD048.229.231	0.000000645
59	CTCAGGGAGACCAAGGCAGTGAC	FRAP1	OBD RD048.233.235	0.000000196
60	GGGACTGGAGGGAAGGAAGTGGG	BCL6	OBD RD048.237.239	0.00000325
61	GGAGCAGTGTAGGGCAGGGTGTGAGA	MMP9	OBD RD048.241.243	0.00000227
62	ATGTCTACAGCCTCTGCCGCTCCTC	MAP3K7	OBD RD048.245.247	0.000000566
63	GCCCTGTAATCCCAGCACTTTGG	MLLT3	OBD RD048.249.251	0.0000046
64	CCCCAGGGACTGAGGACTTGTGT	HLF	OBD RD048.253.255	0.000000743
65	AACAATCTATTTTACCAACCTAT	SIRT1	OBD RD048.257.259	0.00000188
66	CAGGTAGTGTGTTTTCCAACCTGTT	NFATc1	OBD RD048.261.263	0.000000147
67	TAGTAGAGAGTGCGGTGCCACAGGC	TNFRSF13C	OBD RD048.265.267	0.000000402
68	GGCAAGGTCTCCAGTGGTGAGGT	STAT3	OBD RD048.269.271	0.00000103
69	GTCTCACTCTGTTGCCAGGCTG	NFKB1	OBD RD048.273.275	0.00000177
70	TGGATTTTCTGCGGCTCTGTTTG	MEF2B	OBD RD048.277.279	0.00000137
71	AGTCCCCTCTCTGGGTCTCAGCCAAG	CD40	OBD RD048.281.283	0.00000449
72	TATGGCATTTCCTTCCAGTA	MAPK10	OBD RD048.285.287	0.00000213
73	CACTCCAGCCTGAGAGACAGAGC	FRAP1	OBD RD048.289.289	0.00000287
74	GTCTCACTCTGTTGCCAGGCTG	NFKB1	OBD RD048.291.293	0.00000178
75	CAGGGTTGTTGTGAGGGTTATGT	MAPK10	OBD RD048.295.297	0.00000339
76	GTCCCTGCTCTCTTAGCCCCAGA	JAK3	OBD RD048.299.301	0.000000206
77	AGACCTTTGGTTTCTACATCTAT	TNFRSF13C	OBD RD048.303.305	0.000000144
78	GGTATCAAATGTTCCACAAGTGTGC	TET2	OBD RD048.307.309	0.000000972
79	CCAGGATGTCTTACCGCCCCGTCAG	NAE1	OBD RD048.311.313	0.00000172
80	GGGTCTCACTCTGTTGCCAAGC	TNFRSF13C	OBD RD048.315.317	0.00000164
81	CGTCTTGCTCTGTCTGTTGCCAGGC	NFATc1	OBD RD048.319.321	0.00000111
82	GGCAATAGGGATGATTCTGTGAA	BRCA1	OBD RD048.323.325	0.00000046
83	GCACAGGAGGGTTACTTCACAAG	MLLT3	OBD RD048.327.329	0.0000292
84	GCTTCACGGGAGGAGGGTAGACTCTC	PCDHGA6/B2/B4	OBD RD048.331.333	0.0000208
85	TATGGCATTTCCTTCCAGTA	MAPK10	OBD RD048.335.337	-0.0000511
86	ATGTTAGTCCCTTCCACCTAT	BTK	OBD RD051.001.003	-0.000000091
87	ATGTTAGTCCCTTCCACCTAT	BTK	OBD RD051.005.007	-8.44E-08
88	ACGCCTGTAATCCCAGCACTTTG	BTK	OBD RD051.009.011	-0.0000019
89	GATTCTCCTGCCTCAGCCTCCCG	BTK	OBD RD051.013.015	9.55E-08
90	CGATTCTCCTGCCTCAGCCTCCCG	BTK	OBD RD051.017.019	5.07E-08
91	CGATTCTCCTGCCTCAGCCTCCCG	BTK	OBD RD051.021.023	2.87E-08
92	CTCACGAACCGCCTCCTTCCTC	BTK	OBD RD051.025.027	0.00000409

Table 5.r

	Probe	GeneLocus	Probe_Cou nt_Total	Probe_C ount_Sig
1	MIR98_X_53608013_53611637_53628991_53630033_RR	MIR98	16	4
2	DAPK1_9_90064560_90073617_90140806_90142738_FR	DAPK1	46	9
3	HSD3B2_1_119912462_119915175_119959754_119963670_RR	HSD3B2	20	5
4	ERG_21_39895678_39899145_39984806_39991905_RF	ERG	52	4
5	SRD5A3_4_56188038_56191526_56242301_56245314_RF	SRD5A3	12	4

6	MMP1_11_102658858_102661735_102664717_102667643_FF	MMP1	n/a	n/a
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Table 6.a

	HyperG_Stats	FDR_HyperG	Percent_Sig	logFC	AveExpr	t
1	0.064790053	0.737205743	25	0.67511652	0.67511652	13.76185645
2	0.032709022	0.548212211	19.57	0.299375751	0.299375751	7.197207444
3	0.040338404	0.548212211	25	-0.168081632	-0.168081632	-3.274998031
4	0.765503518	1	7.69	-0.425291613	-0.425291613	-11.67074071
5	0.024128503	0.483719041	33.33	0.266992266	0.266992266	4.835274287
6	n/a	n/a	n/a	n/a	4.72222828	n/a

Table 6.b

	P.Value	adj.P.Val	B	FC	FC_1	LS
1	0.000000031	0.0000143	9.558686586	1.596725728	1.596725728	1
2	0.0000184	0.000805368	3.154114326	1.230611817	1.230611817	1
3	0.007481356	0.033194645	-3.020586815	0.890025372	-1.123563476	-1
4	0.000000168	0.0000357	7.913111034	0.744688192	-1.342843905	-1
5	0.000536131	0.005815136	-0.328887879	1.203296575	1.203296575	1
6	0.04505295	0.4547981	n/a	n/a	n/a	n/a

5

Table 6.c

		Probe sequence
	Loop Detected	60 mer
1	Agressive	AGTTGTATTTTGTAGAAAGTAGTGTAAATCGATAGAAATATAACATGAAACACATATATA
2	Aggressive	ACTAATCCCCTGAAGAAGCAAATTAACCTCGAGTATCCCTTTAAGTTTGTITTTAAAATA
3	Indolent	TCAGTTTCTGCTCTCAAGAAGCTTACAGTCGAAGGTCCCAAGTTAGATTACGGCAAAGCT
4	Indolent	TCTTGAATGTGCTTAGTATTATTCAGACTCGAAAACATAATTTGAAAGGAATTCATTCTG
5	Aggressive	AGGAGGTAACGATTGGTCAGCTGCTTAATCGAGGCAGAAGTCTATTTGAAACGTAAGATA
6	n/a	GGCCTTTAAGGCCCTCTGAAATCCAGCATCGAAGAGGGAACTGCATCACA GTTGATGG

Table 6.d

	Probe Location					4 kb Sequence Location		
	Chr	Start1	End1	Start2	End2	Chr	Start1	End1
1	X	53608013	53608044	53628991	53629022	X	53608013	53612014
2	9	90073586	90073617	90140806	90140837	9	90069616	90073617
3	1	119912462	119912493	119959754	119959785	1	119912462	119916463
4	21	39895678	39895709	39991874	39991905	21	39895678	39899679
5	4	56188038	56188069	56245283	56245314	4	56188038	56192039
6	11	102661704	102661735	102667612	102667643	11	102657734	102661735

Table 6.e

	4 kb Sequence Location		Marker
	Start2	End2	
1	53628991	53632992	MIR98_X_53608013_53611637_53628991_53630033_RR
2	90140806	90144807	DAPK1_9_90064560_90073617_90140806_90142738_FR

3	119959754	119963755	HSD3B2_1_119912462_119915175_119959754_119963670_RR
4	39987904	39991905	ERG_21_39895678_39899145_39984806_39991905_RF
5	56241313	56245314	SRD5A3_4_56188038_56191526_56242301_56245314_RF
6	102663642	102667643	MMP1

Table 6.f

	<u>Primers names</u>	<u>Primer sequences</u>		
1	PCa119-245	AAGAAGGGATGGGACGGGACT	PCa119-247	GGTACACGAATTAACATATCCCTGT
2	PCa119-165	ACTGGTCACAGGGAACGATGG	PCa119-167	AGGTGTGAATGTTACTGAACACAAA
3	PCa119-130	ACTTGGAATCCCAAACGCCA	PCa119-132	CTCTCCCCGGTGAGTTTCCA
4	PCa119-065	CAGCCTACCTTGCTGACACT	PCa119-067	AAAGCCCAGTGATGGCCCAT
5	PCa119-154	TCCATTTTCCTTCCCTTTGCTCTG	PCa119-155	CCACACAGGGCCCTAATGACC
6	MMP 1-4 2F	GGGGAGTGGATGGGATAAGGTG	MMP 1F	TGGGCCTGGTTGAAAAGCAT

Table 6.g

	<u>Probe</u>	<u>Probe sequence</u>	Gene
1	OBD119F015	AGTGTTTAATCGATAGAAATATAACATGAAACACA	MIR98
2	OBD119F06	AGGGATACTCGAAGTTAATTGCTTCTT	DAPK1
3	OBD119F09	AAGAAGCTTACAGTCGAAGGTCCCAA	HSD3B2
4	OBD119F08	ATTCCTTCAAATTATGTTTTCGAGTCTGAATAATA	ERG
5	SRD5A3FAM7415RC	AAATAGACTTCTGCCTCGATTAAGCA	SRD5A3
6	MMP1F1b2	ATCCAGCATCGAAGAGGGAACTGCATCA	MMP1

5 Table 6.h

10

	Marker	GLMNET
1	PCa119-245.247	-5.91743E-06
2	PCa119-165.167	-1.57185E-05
3	PCa119-130.132	4.47291E-07
4	PCa119-065.067	6.32136E-06
5	PCa119-154.155	-8.00857E-08
6	MMP1-4 1F. MMP 1F	0

Table 6.i

Marker	GLMNET
OBD RD048.001.003	2.08E-08
OBD RD048.005.007	5.6E-07
OBD RD048.009.011	4.49E-06
OBD RD048.013.015	8.38E-07
OBD RD048.017.019	1.56E-06

OBD RD048.021.023	1.37E-06
OBD RD048.025.027	0.0000046
OBD RD048.029.031	1.81E-06
OBD RD048.033.035	1.78E-06
OBD RD048.037.039	4.02E-07

Table 7. Preferred DLBCL markers

N	EpiSwitch_ID	Inner Forward Primer_ID	Inner Forward Primer Seq
1	ORF1_1_1034282_1037357_1049484_1054771_FF	OBD169_001	GCCAGAGAACAGATGTGTGTGTCT
2	ORF5_1_1140030_1142517_1196191_1197234_RR	OBD169_005	GCCTCTCTGGTGCCACATCTTATCTT
3	ORF5_1_1182474_1185271_1270569_1273244_RF	OBD169_009	CTGCCTGTGTGTAGTCACGAGAAGC
4	ORF5_1_1182474_1185271_1196191_1197234_RR	OBD169_013	CTGACAGCAGAAGCACGAAAAGGTC
5	ORF5_1_1283682_1285577_1335341_1338794_RF	OBD169_017	CCATCCACCCACAGTTCCTATGAAA
6	ORF5_1_1147651_1150121_1196191_1197234_RF	OBD169_021	CCCAACGAGGTCAGGAAGGGAGA
7	ORF5_1_1140030_1142517_1289361_1294150_FF	OBD169_025	TGTCTCAGTATCTATTTCCCAAGTGC
8	ORF1_1_1038521_1042933_1098468_1101242_RF	OBD169_029	CAGGACCCAGACTTGCCCAAACC
9	ORF5_1_1146367_1147651_1165983_1167502_FF	OBD169_033	AGACCCAATGCCTGCCACACGGA
10	ORF5_1_1140030_1142517_1270569_1273244_RF	OBD169_037	CTGCCTGTGTGTAGTCACGAGAAGC
11	ORF5_1_1196191_1197234_1230936_1232838_RR	OBD169_041	GCATAACTCAGAGAAAGCCACTGTGA
12	ORF5_1_1182474_1185271_1209527_1216771_RR	OBD169_045	CTGACAGCAGAAGCACGAAAAGGTC
13	ORF5_1_1270569_1273244_1300933_1312034_FF	OBD169_049	CTGCCTGTGTGTAGTCACGAGAAGC
14	ORF5_1_1157878_1159517_1196191_1197234_RF	OBD169_053	CCCAACGAGGTCAGGAAGGGAGA
15	ORF5_1_1273244_1276010_1335341_1338794_RF	OBD169_057	CACCCATCCACCCACAGTTCCT
16	ORF5_1_1196191_1197234_1289361_1294150_FF	OBD169_061	CCCAACGAGGTCAGGAAGGGAGA
17	ORF5_1_1140030_1142517_1230936_1232838_RR	OBD169_065	CCTCTCTGGTGCCACATCTTATCTTA
18	ORF5_1_1142517_1146335_1270569_1273244_RR	OBD169_069	TTGACCTGGGCTCACATCGCTGA
19	ORF5_1_1230936_1232838_1273244_1276010_RR	OBD169_073	GTCTTCAAGCCACAGAGCAGGATTCC
20	ORF5_1_1157878_1159517_1300933_1312034_FF	OBD169_077	GGTCTGAAAATGTGAATGTCTTGTGT
21	ORF5_1_1147651_1150121_1273244_1276010_RR	OBD169_081	GTGCCCTTGAGTCCAGCCGTCAT
22	ORF1_1_1049484_1054771_1098468_1101242_FF	OBD169_085	TGTCTCTCTCCTAAGGTGTCCCC
23	ORF5_1_1209527_1216771_1270569_1273244_RF	OBD169_089	CTGCCTGTGTGTAGTCACGAGAAGC
24	ORF48_2_84841864_84843477_84864219_84866005_FF	OBD169_093	GCACTTTCTCTCCAGGTCACCTT
25	ORF48_2_84864219_84866005_84885415_84887815_RR	OBD169_097	CTGCTTGGGCTGGTCTTTGGTTG
26	ORF48_2_84841864_84843477_84925461_84928171_FF	OBD169_101	GGCACTTTCTCTCCAGGTCACCC
27	ORF41_2_36413514_36415342_36452868_36458269_RR	OBD169_105	TGAGCGGTCACTGCTGTTGTAGG
28	ORF48_2_84864219_84866005_84876440_84877895_RF	OBD169_109	TTCCATCCTGCTGTCCGTCTGC
29	ORF48_2_84864219_84866005_84925461_84928171_FF	OBD169_113	CGGAGAGAAGGCGGAGAAACCGT
30	ORF41_2_36413514_36415342_36468165_36471683_RR	OBD169_117	GAGCGGTCACTGCTGTTGTAGGC
31	ORF91_7_65033242_65035577_65065127_65067650_RF	OBD169_121	CATTCTGGTATCGTGTGCCGC
32	ORF91_7_65032142_65033242_65065127_65067650_FF	OBD169_125	GGACTTCCTCTCGCCTAATGCG
33	ORF91_7_65037215_65039217_65065127_65067650_RF	OBD169_129	TCCTCCATCCTCACTGGACCAC
34	ORF9_10_23456592_23460302_23494817_23496168_RR	OBD169_133	AGGGCTCTGCGTTTACTCCAGGC
35	ORF15_11_39960254_39968870_39992990_40001746_RR	OBD169_137	CTGGAGCCTGAGTAATGAATAGGAGC
36	ORF16_11_40371218_40374048_40393587_40395559_RR	OBD169_141	GCCCCAATCCCATCCAGAATCCA
37	ORF15_11_39932865_39938937_40079832_40084530_FF	OBD169_145	CTTTCTCTCTCCCTCGTCCCTGG
38	ORF15_11_39992990_40001746_40079832_40084530_RF	OBD169_149	TTTGATAATGAGGGCTGGCTGGGCAT
39	ORF16_11_40371218_40374048_40393587_40395559_RF	OBD169_153	GGATGCCTTAGTTCCTATTGACACT
40	ORF27_12_63568927_63574607_63596388_63598936_RR	OBD169_157	CTGCTGGAGGAGTGACACAAAGTTTC
41	ORF27_12_63568927_63574607_63586940_63589534_RR	OBD169_161	GCCTGCTGGAGGAGTGACACAAAGTT

42	ORF31_15_29619588_29621525_29646237_29648560_RR	OBD169_165	CCTTTCCTCTCCATCTACTCATTCC
43	ORF30_15_10476260_10484217_10545581_10548270_RR	OBD169_169	TTCTATCCCTCCACAAGATGCTCATA
44	ORF32_16_10690178_10695010_10747182_10750815_RR	OBD169_173	GGGAGACGGAGGAAAGCCTATC
45	ORF32_16_10747182_10750815_10765838_10768877_RR	OBD169_177	AACCTCCTCAAAGAGAGAGCCTTCCC
46	ORF32_16_10726068_10729293_10772875_10776021_FF	OBD169_181	AGGTCTTCAACCAACACCACCAGTG
47	ORF32_16_10747182_10750815_10792291_10794979_RF	OBD169_185	CCTCCTGTATTTCTACTTCCACTCAG
48	ORF32_16_10726068_10729293_10792291_10794979_FF	OBD169_189	GCAGGTCTTCAACCAACACCACCAG
49	ORF32_16_10747182_10750815_10772875_10776021_RR	OBD169_193	AACCTCCTCAAAGAGAGAGCCTTCCC
50	ORF32_16_10778964_10780903_10792291_10794979_FF	OBD169_197	CAGTGTGAAAGCACCTTCGCTCTTGC
51	ORF68_25_630610_633794_676143_680436_FF	OBD169_201	GGGCAATGTGAGGCTGTTATGCTTGT
52	ORF68_25_630610_633794_687567_692655_FF	OBD169_205	CCAGGGCAATGTGAGGCTGTTATGCT
53	ORF70_26_27906620_27909025_27963114_27965001_RR	OBD169_209	TTTGAGGGCAGAGCAGGAAGGGT
54	ORF70_26_27876428_27879774_27894296_27895372_RR	OBD169_213	GTCCCTGCTCCACTGCCAATGAG
55	ORF70_26_27890569_27893929_27933912_27935209_RR	OBD169_217	GTGCCCTGGATGGAGAAGCTTGCT
56	ORF70_26_27933912_27935209_27963114_27965001_RR	OBD169_221	TACAGAAAGCCCTCGCTGGGAGC
57	ORF70_26_27876428_27879774_27890569_27893929_RR	OBD169_225	AAGTGTAGCACGGACCAGAGAGC
58	ORF70_26_27894296_27895372_27963114_27965001_RR	OBD169_229	CTGCCTCCAGAAGGTGTCTCAGA
59	ORF70_26_27890569_27893929_27906620_27909025_RR	OBD169_233	GTGCCCTGGATGGAGAAGCTTGCT
60	ORF75_31_28027888_28030129_28041732_28043951_FF	OBD169_237	GGACAAGCATCCTGGTTGAGCCA
61	ORF75_31_28027888_28030129_28043951_28045576_FF	OBD169_241	GGACAAGCATCCTGGTTGAGCCA
62	ORF79_32_24013860_24017127_24039530_24040887_RF	OBD169_245	GACCCAGAAATGAACCCAAAAGATGA
63	ORF79_32_23988046_23989457_24013860_24017127_RR	OBD169_249	GCACTCCCTACACACAAATCCTTAGA
64	ORF79_32_23965697_23967743_24013860_24017127_RR	OBD169_253	GCAACAGTTCATAACCGAGTGCCAAC
65	ORF79_32_23965697_23967743_24028587_24030780_RR	OBD169_257	GCAACAGTTCATAACCGAGTGCCAAC
66	ORF79_32_23965697_23967743_24000345_24005192_RR	OBD169_261	CAGTTCATAACCGAGTGCCAACAGAA
67	ORF79_32_24013860_24017127_24028587_24030780_RR	OBD169_265	GGTGACTGATGAGACTCCAGGAAAAGT
68	ORF79_32_23965697_23967743_24039530_24040887_RF	OBD169_269	GACCCAGAAATGAACCCAAAAGATGA
69	ORF79_32_23988046_23989457_24039530_24040887_RF	OBD169_273	GACCCAGAAATGAACCCAAAAGATGA
70	ORF82_32_9652472_9664654_9692674_9698030_RR	OBD169_277	CCCACCTCCCTGCTCCAACAAGATTT
71	ORF79_32_24000345_24005192_24039530_24040887_RF	OBD169_281	GACCCAGAAATGAACCCAAAAGATGA
72	ORF79_32_23988046_23989457_24000345_24005192_RR	OBD169_285	GCAGCCTTTGGCAGCACTCTCTG
73	ORF104_X_109512943_109516164_109526507_109531763_RF	OBD169_289	CCCTTCTGGAAGTGGATGAGCCCTTA
74	ORF104_X_109508063_109510622_109526507_109531763_FF	OBD169_293	TGAGCCCTTAGTCAATGGGACCG
75	ORF106_X_75279499_75281082_75297768_75302185_RF	OBD169_297	CCAGTTCACCAAGGTTGAGTGCC

Table 8.a

N	Inner Reverse Primer ID	Inner Reverse Primer Seq	Gene	Marker	GLMNET
1	OBD169_003	AAAACTCCCACCTGTCTGTGTAC	NFATC1	OBD169_001.OBD169_003	0.150341207
2	OBD169_007	GCATAACTCAGAGAAAGCCACTGTGA	ATP9B	OBD169_005.OBD169_007	0
3	OBD169_011	GACAGCAGAAGCACGAAAAGGTCATT	ATP9B	OBD169_009.OBD169_011	-0.065057056
4	OBD169_015	TGTCCTCCAGCCTCTGTTACCC	ATP9B	OBD169_013.OBD169_015	0.011765488
5	OBD169_019	GGTCTGAAAGCACCTGTAAGTCTGGA	ATP9B	OBD169_017.OBD169_019	0
6	OBD169_023	CCCTTGAGTCCAGCCGTCATTAC	ATP9B	OBD169_021.OBD169_023	0
7	OBD169_027	ACACGATGAGACAGAGCACCAGAGTC	ATP9B	OBD169_025.OBD169_027	0
8	OBD169_031	GGTGAGTTCTGACCTGGGCTTTC	NFATC1	OBD169_029.OBD169_031	0
9	OBD169_035	TCTGAGGTCCTGATGGAGCACAG	ATP9B	OBD169_033.OBD169_035	0
10	OBD169_039	CCTCTCTGGTGCCACATCTTATCTTA	ATP9B	OBD169_037.OBD169_039	0
11	OBD169_043	GTCTTCAAGCCACAGAGCAGGATTCC	ATP9B	OBD169_041.OBD169_043	0.122625202

12	OBD169_047	CCATCTTCTGTAACCTGAACGGAGT	ATP9B	OBD169_045.OBD169_047	0
13	OBD169_051	CGTTATCTATGGTCCCACTACTGTGT	ATP9B	OBD169_049.OBD169_051	-0.050953035
14	OBD169_055	GCAGGTTATTAGAGGACCGAGGC	ATP9B	OBD169_053.OBD169_055	0
15	OBD169_059	CGCCACCAAGAATGTCATCTCCG	ATP9B	OBD169_057.OBD169_059	0
16	OBD169_063	CGATGAGACAGAGCACCAGAGTC	ATP9B	OBD169_061.OBD169_063	0.127785257
17	OBD169_067	GTCTTCAAGCCACAGAGCAGGATTCC	ATP9B	OBD169_065.OBD169_067	-6.18E-06
18	OBD169_071	GTGGCTACCTGTGGTCTCTCCT	ATP9B	OBD169_069.OBD169_071	0
19	OBD169_075	GCCACCAAGAATGTCATCTCCGATTT	ATP9B	OBD169_073.OBD169_075	0
20	OBD169_079	GGCTTCGTTATCTATGGTCCCACTAC	ATP9B	OBD169_077.OBD169_079	0
21	OBD169_083	CGCCACCAAGAATGTCATCTCCG	ATP9B	OBD169_081.OBD169_083	0
22	OBD169_087	CAGGACCCAGACTTGCCCAAACC	NFATC1	OBD169_085.OBD169_087	0
23	OBD169_091	CTGTAACCTGAACGGAGTAGAATAG	ATP9B	OBD169_089.OBD169_091	0
24	OBD169_095	GGCGGAGAAACCGTTCGTGTGTG	MTOR	OBD169_093.OBD169_095	0
25	OBD169_099	GGCAAGGGACCACTCTTAGTCTGC	MTOR	OBD169_097.OBD169_099	0
26	OBD169_103	TCCCTTATCAACCAACTCGGGC	MTOR	OBD169_101.OBD169_103	0.003937173
27	OBD169_107	TTGGTGGTCAGGACTGGAGTGCC	PCDHGC5	OBD169_105.OBD169_107	0.029250039
28	OBD169_111	CTGCTGGGCTGGTCTTTGGTTG	MTOR	OBD169_109.OBD169_111	0
29	OBD169_115	TCCCTTATCAACCAACTCGGGC	MTOR	OBD169_113.OBD169_115	0
30	OBD169_119	GAGGTCAAGGGAAGAGACAGGGA	PCDHGC5	OBD169_117.OBD169_119	0
31	OBD169_123	TGTGGAATGAGCCTCCGTCCCTG	CABLES1	OBD169_121.OBD169_123	0
32	OBD169_127	CATTCTGGTATCGTGTGGCCGC	CABLES1	OBD169_125.OBD169_127	0.005994639
33	OBD169_131	CCAGAACATCTCTCGTGGTGGG	CABLES1	OBD169_129.OBD169_131	0
34	OBD169_135	GATGCTGTCCCTGTGCTATGAGC	SREBF2	OBD169_133.OBD169_135	0.161924686
35	OBD169_139	GTCATCAACACTCTTCCCTGCTCCT	MLLT3	OBD169_137.OBD169_139	0
36	OBD169_143	CCATTGCCTGAATCCTCCCTGGC	FOCAD	OBD169_141.OBD169_143	0
37	OBD169_147	TGAGGGCTGGCTGGGCATTCATA	MLLT3	OBD169_145.OBD169_147	0
38	OBD169_151	GTCATCAACACTCTTCCCTGCTCCT	MLLT3	OBD169_149.OBD169_151	0
39	OBD169_155	CAGCCCCAATCCCATCCAGAATCCA	FOCAD	OBD169_153.OBD169_155	0
40	OBD169_159	CTGTGATTCCCTTGTATGGTTTGA	ATG5	OBD169_157.OBD169_159	0
41	OBD169_163	GCCTCTGTCTGTGTATGAAACT	ATG5	OBD169_161.OBD169_163	0
42	OBD169_167	CTACAAGGGAAGTGCCTGCTCGCTA	FAF1	OBD169_165.OBD169_167	0
43	OBD169_171	AACAGGCTTACCTCTTCGGACTGCTC	KITLG	OBD169_169.OBD169_171	0.063674679
44	OBD169_175	CTCCTCAAAGAGAGAGCCTTCCCG	CREB3L2	OBD169_173.OBD169_175	0
45	OBD169_179	GCGTGTGAGAGAGGAGATAAATGGAT	CREB3L2	OBD169_177.OBD169_179	0.013500095
46	OBD169_183	CTGGCTGGCTCTTGACTTTGCTATTG	CREB3L2	OBD169_181.OBD169_183	0
47	OBD169_187	AACCTCTCAAAGAGAGAGCCTTCCC	CREB3L2	OBD169_185.OBD169_187	0.248790766
48	OBD169_191	CCTCTGTATTTCTACTTCCACTCAG	CREB3L2	OBD169_189.OBD169_191	0
49	OBD169_195	GACTGATTGTAGGAGGACTCACAGAT	CREB3L2	OBD169_193.OBD169_195	0
50	OBD169_199	CCTCTGTATTTCTACTTCCACTCAG	CREB3L2	OBD169_197.OBD169_199	0
51	OBD169_203	ATCATTGGTTTGGAGTGACAACTACT	FOXO1	OBD169_201.OBD169_203	0
52	OBD169_207	GGTAGTGTCTGTTTCTGGACTTTAC	FOXO1	OBD169_205.OBD169_207	0
53	OBD169_211	GGTGTGGGTGTGTAAGAGGGACC	SPECC1L	OBD169_209.OBD169_211	0
54	OBD169_215	CTGCCTCCAGAAGGTGTCTCAGA	SPECC1L	OBD169_213.OBD169_215	0
55	OBD169_219	TACAGAAAGCCCTCGCTGGGAGC	SPECC1L	OBD169_217.OBD169_219	0
56	OBD169_223	AGGGTGTGGGTGTGTAAGAGGGGA	SPECC1L	OBD169_221.OBD169_223	0
57	OBD169_227	CCACTGTGCCCTGGATGGAGAAC	SPECC1L	OBD169_225.OBD169_227	0
58	OBD169_231	GGTGTGGGTGTGTAAGAGGGACC	SPECC1L	OBD169_229.OBD169_231	-0.042293888
59	OBD169_235	TTGAGGGCAGAGCAGGAAGGGTG	SPECC1L	OBD169_233.OBD169_235	0.052029568
60	OBD169_239	GGGATACCCAGAGAGAAGGGCAAG	IFNGR2	OBD169_237.OBD169_239	0

61	OBD169_243	AGACCTGAGGAAGGAGGGTGGAC	IFNGR2	OBD169_241.OBD169_243	0.043975004
62	OBD169_247	GTGAGAGGCAGAGACAGCACAGACTA	NFKB1	OBD169_245.OBD169_247	0
63	OBD169_251	GTGAGAGGCAGAGACAGCACAGACTA	NFKB1	OBD169_249.OBD169_251	0
64	OBD169_255	GGTGAAGGATGAGACTCCAGGAAAGT	NFKB1	OBD169_253.OBD169_255	0
65	OBD169_259	GCCTAAACTTTCTCTCAGTCAGCG	NFKB1	OBD169_257.OBD169_259	0.01527689
66	OBD169_263	GCCTCTGTCATTCTGCTTCCAGTGT	NFKB1	OBD169_261.OBD169_263	0
67	OBD169_267	GCCTAAACTTTCTCTCAGTCAGCG	NFKB1	OBD169_265.OBD169_267	0.141700302
68	OBD169_271	TGTTACGCACAACCTCGGCTCTG	NFKB1	OBD169_269.OBD169_271	0
69	OBD169_275	GCAGCCTTTGGCAGCACTCTCTG	NFKB1	OBD169_273.OBD169_275	0
70	OBD169_279	CCCAGAACTTTGCTAACTCCTATTG	MAPK10	OBD169_277.OBD169_279	-0.097352472
71	OBD169_283	GCCTCTGTCATTCTGCTTCCAGTGT	NFKB1	OBD169_281.OBD169_283	0
72	OBD169_287	GCCTCTGTCATTCTGCTTCCAG	NFKB1	OBD169_285.OBD169_287	0
73	OBD169_291	AAGTGCCTGTTTTATGGAGAACTGGC	F9	OBD169_289.OBD169_291	0
74	OBD169_295	CCCTTCTGGAAGTGGATGAGCCC	F9	OBD169_293.OBD169_295	0
75	OBD169_299	CACAGCCGAAGAGCCACTGAAGC	BTK	OBD169_297.OBD169_299	0

Table 8.b

N	Probe	marker	GLMNET
1	ORF1_1_1034282_1037357_1049484_1054771_FF	OBD169_001.OBD169_003	0.150341207
2	ORF5_1_1182474_1185271_1270569_1273244_RF	OBD169_009.OBD169_011	-0.065057056
3	ORF5_1_1147651_1150121_1196191_1197234_RF	OBD169_021.OBD169_023	0
4	ORF5_1_1146367_1147651_1165983_1167502_FF	OBD169_033.OBD169_035	0
5	ORF5_1_1196191_1197234_1230936_1232838_RR	OBD169_041.OBD169_043	0.122625202
6	ORF5_1_1270569_1273244_1300933_1312034_FF	OBD169_049.OBD169_051	-0.050953035
7	ORF5_1_1196191_1197234_1289361_1294150_FF	OBD169_061.OBD169_063	0.127785257
8	ORF5_1_1140030_1142517_1230936_1232838_RR	OBD169_065.OBD169_067	-6.18144E-06
9	ORF5_1_1230936_1232838_1273244_1276010_RR	OBD169_073.OBD169_075	0
10	ORF41_2_36413514_36415342_36452868_36458269_RR	OBD169_105.OBD169_107	0.029250039
11	ORF91_7_65032142_65033242_65065127_65067650_FF	OBD169_125.OBD169_127	0.005994639
12	ORF91_7_65037215_65039217_65065127_65067650_RF	OBD169_129.OBD169_131	0
13	ORF9_10_23456592_23460302_23494817_23496168_RR	OBD169_133.OBD169_135	0.161924686
14	ORF16_11_40371218_40374048_40393587_40395559_RF	OBD169_153.OBD169_155	0
15	ORF31_15_29619588_29621525_29646237_29648560_RR	OBD169_165.OBD169_167	0
16	ORF30_15_10476260_10484217_10545581_10548270_RR	OBD169_169.OBD169_171	0.063674679
17	ORF32_16_10747182_10750815_10792291_10794979_RF	OBD169_185.OBD169_187	0.248790766
18	ORF70_26_27894296_27895372_27963114_27965001_RR	OBD169_229.OBD169_231	-0.042293888
19	ORF70_26_27890569_27893929_27906620_27909025_RR	OBD169_233.OBD169_235	0.052029568
20	ORF79_32_24013860_24017127_24028587_24030780_RR	OBD169_265.OBD169_267	0.141700302
21	ORF82_32_9652472_9664654_9692674_9698030_RR	OBD169_277.OBD169_279	-0.097352472
22	ORF104_X_109508063_109510622_109526507_109531763_FF	OBD169_293.OBD169_295	0

Table 9.a

N	Freq	Rank_median	pValue_Mean	pValue_Median	Classification
1	429	14	0.061922326	0.036945939	Presence in Lymphoma
2	156	171.75	0.722387112	1	Presence in Healthy Control
3	155	29.75	0.137404255	0.119176434	Presence in Lymphoma
4	278	14.75	0.076727087	0.075561315	Presence in Lymphoma
5	300	22.25	0.11481488	0.111802994	Presence in Lymphoma
6	262	107.5	0.614169025	0.608053733	Presence in Healthy Control

7	375	16.5	0.07087785	0.048199002	Presence in Lymphoma
8	112	168.5	0.749906059	1	Presence in Healthy Control
9	115	28	0.185541633	0.104567082	Presence in Lymphoma
10	262	16.25	0.099987147	0.048199002	Presence in Lymphoma
11	300	22.75	0.163342682	0.089691605	Presence in Lymphoma
12	130	33.5	0.190563594	0.148661263	Presence in Lymphoma
13	406	18	0.093426309	0.075561315	Presence in Lymphoma
14	270	23.25	0.114536951	0.056118783	Presence in Lymphoma
15	135	23.5	0.159941064	0.104567082	Presence in Lymphoma
16	452	7	0.034141832	0.02207464	Presence in Lymphoma
17	498	2	0.009682876	0.006340396	Presence in Lymphoma
18	225	97.75	0.608040664	0.516296715	Presence in Healthy Control
19	357	9.25	0.060876258	0.035136821	Presence in Lymphoma
20	451	12	0.055525573	0.036945939	Presence in Lymphoma
21	225	94.5	0.550385123	0.521495378	Presence in Healthy Control
22	257	32.5	0.167821507	0.104567082	Presence in Lymphoma

Table 9.b

Table 10. Prostate cancer risk group categories.

Category	Risk	PSA (ng/ml)	Gleason score	T stage
1	Low risk	< 10	≤6	T1 - T2a
2	Intermediate risk	10-20	7	T2b
3	High risk	>20	8-10	T2c*, T3 or T4

* According to NCCN guidelines 2018 update T2c is considered intermediate risk.

Abbreviations. PSA: prostate specific antigen.

Table 11. Five-marker signature used for the diagnosis of prostate cancer.

Markers	Gene symbol	Gene name	P value
PCa.57.59	ETS1	ETS proto-oncogene 1, transcription factor	0.11
PCa.81.83	MAP3K14	Mitogen-activated protein kinase kinase kinase 14	0.11
PCa.73.75	SLC22A3	Solute carrier family 22 member 3	0.107
PCa.77.79	SLC22A3	Solute carrier family 22 member 3	0.005
PCa.189.191	CASP2	Caspase 2	0.137

Table 12.

Comparison of pathology and EpiSwitch™ results.

	Pathology results	
	PCa	Healthy
EpiSwitch™ diagnosis		
PCa	8	2
Healthy	2	8

Results from classification of blinded samples (n = 20).

Statistic	Value	95% CI
Sensitivity	80.00%	44.39% to 97.48%
Specificity	80.00%	44.39% to 97.48%
Positive Likelihood Ratio	4.00	1.11 to 14.35
Negative Likelihood Ratio	0.25	0.07 to 0.90
Disease prevalence (*)	50.00%	27.20% to 72.80%
Positive Predictive Value (*)	80.00%	52.71% to 93.49%
Negative Predictive Value (*)	80.00%	52.71% to 93.49%

(*) These values are dependent on disease prevalence.

Abbreviations. 95% CI: 95% confidence interval.

Table 13. Markers for high-risk category 3 vs low-risk category 1 and for high-risk category 3 vs intermediate-risk category 2.

Biomarkers for high-risk category 3 vs low-risk category 1				Biomarkers for high-risk category 3 vs intermediate-risk category 2			
Loci	Gene location	Markers	Loci	Gene location	Markers	Loci	Markers
6_7724582_7733496_7801590_7806316_FF	BMP6	PCa.119.37.39	1_119912462_119915175_119959754_119963670_RR	HSD3B2	PCa.119.129.131		
21_39895678_39899145_39984806_39991905_RF	ERG	PCa.119.65.67	4_177629821_177639626_177740221_177743175_FR	VEGFC	PCa.119.205.207		
8_16195878_16203315_16396849_16400398_FF	MSR1	PCa.119.77.79	12_99061113_99062942_99098781_99108240_FF	APAF1	PCa.119.49.51		
1_155146523_155149986_155191807_155193554_FR	MUC1	PCa.119.121.123	1_155146523_155149986_155191807_155193554_FR	MUC1	PCa.119.121.123		
11_107955219_107960166_108013361_108018367_FF	ACAT1	PCa.119.57.59	11_107955219_107960166_108013361_108018367_FF	ACAT1	PCa.119.57.59		
9_90064560_90073617_90140806_90142738_FR	DAPK1	PCa.119.165.167	9_90064560_90073617_90140806_90142738_FR	DAPK1	PCa.119.165.167		

Shaded last three markers are common.

Abbreviations. ACAT1: acetyl-CoA acetyltransferase 1; APAF1: apoptotic peptidase activating factor 1; BMP6: bone morphogenetic protein 6; DAPK1: death associated protein kinase 1; ERG: ETS transcription factor ERG; HSD3B2: hydroxy-delta-5-steroid dehydrogenase, 3 beta- and steroid delta-isomerase 2; MSR1: macrophage scavenger receptor 1; MUC1: mucin 1, cell surface associated; VEGFC: vascular endothelial growth factor C.

Table 14. Detection of similar epigenetic markers in blood and in matching primary prostate tumours at a fixed range of assay sensitivity.

Gene location	Markers	Category		Blood samples					Total number of positive markers in blood samples	Tissue samples					Total number of positive markers in tissue samples			
				1						3								
		Patient	1				3	A	B	C	D	E	A	B	C	D	E	
BMP6	PCa.119.37.39		1	1	0	1	1	4	1	1	1	0	1					4
ERG	PCa.119.65.67		1	1	1	1	1	5	1	0	1	0	1					3
MSR1	PCa.119.77.79		1	0	0	0	1	2	0	0	1	0	1					2
MUC1	PCa.119.121.123		1	1	1	1	0	4	1	1	1	1	1					5
DAPK1	PCa.119.165.167		1	0	1	1	0	3	0	0	1	0	1					2
ACAT1	PCa.119.57.59		1	1	1	1	1	5	1	1	1	1	1					5
HSD3B2	PCa.119.129.131		1	1	0	1	1	4	1	1	1	0	1					4
VEGFC	PCa.119.205.207		1	1	1	1	1	5	1	1	1	1	1					5
APAF1	PCa.119.49.51		1	1	1	1	1	5	0	1	1	1	1					4

When a PCR band of the correct size is detected, it is given a score of 1. When no band is detected, it is given a score of 0.

Abbreviations. ACAT1: acetyl-CoA acetyltransferase 1; APAF1: apoptotic peptidase activating factor 1; BMP6: bone morphogenetic protein 6; DAPK1: death associated protein kinase 1; ERG: ETS transcription factor ERG; HSD3B2: hydroxy-delta-5-steroid dehydrogenase, 3 beta- and steroid delta-isomerase 2; MSR1: macrophage scavenger receptor 1; MUC1: mucin 1, cell surface associated; VEGFC: vascular endothelial growth factor C.

Table 15.
Comparison of pathology and EpiSwitch™ results for category 3 vs 1 classifier.

EpiSwitch™ diagnosis	Pathology results	
	Category 1	Category 3
Category 1	39	5
Category 3	3	20

Results from classification of blinded samples for category 3 vs 1 classifier (n = 67).

Statistic	Value	95% CI
Sensitivity	80.00%	59.30% to 93.17%
Specificity	92.86%	80.52% to 98.50%
Positive Likelihood Ratio	11.20	3.70 to 33.91
Negative Likelihood Ratio	0.22	0.10 to 0.47
Disease prevalence (*)	37.31%	25.80% to 49.99%
Positive Predictive Value (*)	86.96%	68.77% to 95.28%
Negative Predictive Value (*)	88.64%	78.00% to 94.49%

(*) These values are dependent on disease prevalence.

Abbreviations. 95% CI: 95% confidence interval.

Table 16.
Comparison of pathology and EpiSwitch™ results for category 3 vs 2 classifier.

EpiSwitch™ diagnosis	Pathology results	
	Category 2	Category 3
Category 2	16	4
Category 3	2	21

Results from classification of blinded samples for category 3 vs 2 classifier (n = 43).

Statistic	Value	95% CI
Sensitivity	84.00%	63.92% to 95.46%
Specificity	88.89%	65.29% to 98.62%
Positive Likelihood Ratio	7.56	2.02 to 28.24
Negative Likelihood Ratio	0.18	0.07 to 0.45
Disease prevalence (*)	58.14%	42.13% to 72.99%
Positive Predictive Value (*)	91.30%	73.76% to 97.51%
Negative Predictive Value (*)	80.00%	61.62% to 90.88%

(*) These values are dependent on disease prevalence.
Abbreviations. 95% CI: 95% confidence interval.

Table 17. Clinical characteristics of the patients participated in the study.

Characteristic	Category	Number of patients
Gleason score	≤6	39
	7	54
	8-10	29
	Unknown	18
	Median	7
Stage	1	36
	2	49
	3	25
	4	14
	Unknown	16
Age	45-54	12
	55-64	21
	65-74	44
	75+	63
	Unknown	0
PSA	<10	55
	10-20	23
	>20	51
	Unknown	11
	Median	12.2
Metastatic patients		21

Abbreviation. PSA: prostate specific antigen.

Table 18. List of 425 prostate cancer-related genomic loci tested in the initial array.

Gene name	Array probe count	Gene name	Array probe count	Gene name	Array probe count	Gene name	Array probe count
ABHD3	20	CALR	15	CREBBP	20	EPS15	111
ABR	20	CAP1	20	CSE1L	20	ERBB2	20
ACAT1	20	CARS	20	CSF2	11	ERBB3	20
ACPP	20	CASP2	20	CSF2RA	3	ERBB4	200
ACTA1	20	CASP3	20	CSNK1A1	20	ERG	67
ACTR2	20	CASP9	20	CTBP1	20	ERRFI1	20
ACTR3	20	CAV1	20	CTNNA1	43	ESR1	200
ADAM9	20	CBL	20	CTNNB1	45	ESR2	20
ADRB2	20	CCDC67	31	CTNND1	20	ETS1	105
AGAP2	15	CCND1	20	CXCL16	19	ETV1	47
AIP	9	CCND2	20	CXCR1	20	ETV4	20
AKT1	20	CCNE1	20	CXCR2	19	ETV5	20
AKT2	20	CCNJ	6	CXCR4	20	EZH2	31
AKT3	122	CD244	20	CXCR6	19	FASN	20
AMACR	20	CD4	20	CYCS	18	FGD4	52
AP2M1	20	CD44	75	CYP17A1	20	FGF19	20
APAF1	26	CD82	20	CYP19A1	149	FGF2	39
APC	37	CD8A	20	CYP1B1	19	FGF6	20
AR	115	CDC25A	20	DAND5	20	FGF8	10
ARAF	20	CDC25B	20	DAPK1	56	FGFR1	20
ARNT	20	CDC25C	20	DDIT4	16	FGFR4	12
ARTN	20	CDC37	20	DOK4	20	FHL2	20
ASAH1	20	CDC45	20	DPP4	103	FLNA	9
ATM	54	CDH1	20	E2F1	20	FLT1	97
AXIN1	20	CDK2	2	E2F4	20	FLT4	10
AXL	20	CDK4	20	EDN1	20	FN1	20
BAD	20	CDK6	111	EDNRA	35	FOLH1	35
BCAR1	28	CDKN1A	20	EED	20	FOSB	11
BCL2L1	20	CDKN1B	20	EGF	26	FOXA1	18
BCORL1	20	CDKN2A	20	EGFR	200	FOXO1	21
BGLAP	11	CDKN2B	16	EIF4E	20	FOXO3	139
BIRC5	4	CDKN2C	10	EIF4EBP1	20	FOXP1	200
BMI1	20	CDKN2D	20	EIF6	20	FZD1	18
BMP6	71	CENPBD1	7	ELAC2	20	GAB1	107
BMP7	20	CHAMP1	20	ENPP2	74	GAS6	20
BRAF	20	CHEK2	20	EP300	20	GLIPR1	15
BRCA1	20	CHUK	29	EPHA2	20	GNRH1	20
BRCA2	20	CLU	28	EPHB4	20	GNRHR	3
CA1	20	COMMD3-BMI1	20	EPHB6	6	GRB2	20

Gene name	Array probe count	Gene name	Array probe count	Gene name	Array probe count	Gene name	Array probe count
GSK3B	92	KLK4	17	MIR34A	20	NFKB1	66
GSTP1	20	KRAS	20	MIR361	20	NFKBIA	20
HDAC1	20	LPAR1	30	MIR365A	5	NGF	21
HDAC3	20	LPAR2	5	MIR376A1	9	NKX3-1	20
HGF	27	LPAR3	31	MIR454	11	NOVA1	34
HIF1A	20	LPAR4	5	MIR500A	20	NOX5	43
HIPK2	20	LRP5	20	MIR582	3	NPDC1	1
HRAS	6	LRP6	35	MIR619	20	NR0B1	10
HSD3B1	20	MAGEA11	20	MIR636	20	NR3C1	25
HSD3B2	20	MAP2K1	20	MIR648	20	NR4A3	20
HSP90AA1	20	MAP2K2	20	MIR671	20	NRAS	20
HSP90AB1	20	MAP2K5	196	MIR766	15	NRIP1	69
HSPA1A	20	MAP3K14	20	MIR877	13	NTF3	31
HSPB1	20	MAP3K2	24	MIR887	18	NTRK1	20
IGF1	20	MAPK1	20	MIR93	20	PA2G4	20
IGF1R	102	MAPK3	20	MIR98	17	PAQR7	20
IGFBP3	14	MAPKAP1	31	MIRLET7G	3	PAX8	20
IGFBP5	20	MCM7	20	MLST8	2	PCBP2	20
IL16	82	MDM2	20	MMP14	20	PCYT1A	20
IL2	2	MDM4	23	MMP9	20	PDGFA	20
IL6	20	MED1	20	MSMB	20	PDGFB	20
IL6R	20	MEN1	20	MSR1	200	PDGFRA	24
IL8	17	MET	105	MTA1	1	PDGFRB	20
INPPL1	20	MIF	12	MTCH1	20	PDPK1	20
INS	17	MIR125B1	4	MTOR	32	PIAS1	89
IRAK1	20	MIR149	3	MTRR	20	PIAS2	21
IRS1	20	MIR151A	31	MUC1	7	PIAS3	20
ITK	24	MIR152	15	MYB	20	PIAS4	20
JAK1	20	MIR16-1	20	MYC	20	PIK3C2A	20
JAK2	20	MIR183	20	NCOA1	170	PIK3C2B	21
JAK3	20	MIR197	7	NCOA2	200	PIK3C2G	200
JUN	20	MIR204	20	NCOA3	20	PIK3CA	20
KAT7	20	MIR222	6	NCOA4	18	PIK3CB	21
KCNH2	20	MIR224	20	NCOR1	35	PIK3CD	20
KDM6B	20	MIR23B	20	NCOR2	26	PIK3CG	20
KIT	107	MIR24-2	20	NEDD4	85	PIK3R1	81
KLF4	20	MIR26B	3	NEDD4L	128	PIK3R2	20
KLK2	20	MIR27B	20	NET1	20	PLD1	189

KLK3	8	MIR335	20	NF1	190	PLD2	20
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Gene name	Array probe count	Gene name	Array probe count	Gene name	Array probe count	Gene name	Array probe count
PLD3	20	PXN	20	SFTPA1	20	TGFB1I1	20
PML	20	RAB9B	16	SFTPA2	8	TGFB2	75
POU2F1	26	RAD51	9	SHC1	11	TGFB3	15
POU2F2	20	RAF1	20	SIRT1	20	TGFBR1	20
PPP1CA	20	RAN	20	SKP2	16	TGFBR2	65
PPP2CA	20	RB1	82	SLC22A3	56	TIMP1	20
PRKCA	148	RCHY1	20	SMAD3	20	TMF1	20
PRKCB	176	REL	25	SMAD4	67	TMPRSS2	36
PRKCD	20	RGS6	200	SMARCE1	20	TNK2	20
PRKCG	20	RHEB	20	SOAT1	20	TOP2A	20
PRKCH	200	RHOA	20	SOS1	103	TOP2B	24
PRKCI	43	RICTOR	38	SOX9	20	TP53	15
PRKCQ	59	RNASEL	13	SP1	20	TRAF3	20
PRKCZ	20	RNF14	20	SPDEF	17	TRAF6	20
PRSS3	23	RNF20	20	SPINK1	20	TSC1	20
PRSS8	20	RNF40	20	SPOPL	20	TSC2	2
PSAP	20	ROCK1	67	SRC	20	TUBB	4
PSCA	20	ROR2	193	SRD5A1	13	VEGFA	20
PSG1	19	RPS6KA1	20	SRD5A2	79	VEGFC	57
PTEN	23	RPS6KB1	20	SRD5A3	20	VIM	9
PTGS2	20	RPTOR	68	SREBF1	20	WAS	12
PTK2B	54	RREB1	85	SRY	3	WNT1	20
PTK7	20	RYBP	20	STAT3	20	WNT2	40
PTPN11	20	S100A4	20	SUZ12	20	WNT3	20
PTPN12	20	S100P	20	SVIL	51	WNT5A	23
PTPN14	135	SAGE1	20	TBC1D8	43	ZAP70	20
PTPRF	20	SATB1	20	TERT	20	ZFAND1	20
PTPRR	200	SCGB1A1	20	TGFB1	20	ZMYND10	20
PTPRT	200					Total	14241

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	GeneLocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
ACAT1_11_107955219_107960166_108013361_108018367_FF	ACAT1	-0.436725529	-0.436725529	-11.90723067	1.37E-07	3.10E-05	8.114204006	0.738809576	-1.353528748	-1
ACTA1_1_229547333_229551721_229600994_229605798_FR	ACTA1	0.417850291	0.417850291	8.736657363	2.98E-06	0.000260244	5.025483662	1.335935441	1.335935441	1
AKT3_1_243680126_243690814_243946602_243948601_FR	AKT3	0.652970743	0.652970743	16.8960264	3.63E-09	3.08E-06	11.560433	1.572402697	1.572402697	1
AKT3_1_243680126_243690814_243915703_243918596_FR	AKT3	0.598435451	0.598435451	11.73324265	1.59E-07	3.45E-05	7.966660757	1.514073719	1.514073719	1
AKT3_1_243680126_243690814_243727939_243733240_FF	AKT3	0.520843747	0.520843747	19.9294303	6.33E-10	1.07E-06	13.10148999	1.434794129	1.434794129	1
AKT3_1_243680126_243690814_243860421_243862288_FR	AKT3	0.410316196	0.410316196	12.44251976	8.76E-08	2.55E-05	8.554504322	1.328977055	1.328977055	1
APAF1_12_990611113_990629942_99098781_99108240_FF	APAF1	-0.441488336	-0.441488336	-13.23940926	4.63E-08	1.71E-05	9.174110234	0.736374546	-1.358004571	-1
APC_5_112020873_112029146_112079758_112082452_FF	APC	0.399930381	0.399930381	6.922678201	2.63E-05	0.000993557	2.789251424	1.319444238	1.319444238	1
AR_X_66792540_66795953_66818342_66825862_RF	AR	-0.33854166	-0.33854166	-6.221155823	6.77E-05	0.001660581	1.810263084	0.790840324	-1.264477758	-1
AR_X_66736338_66750729_66875649_66881776_RR	AR	0.760948793	0.760948793	21.04618466	3.54E-10	7.78E-07	13.59167252	1.694604721	1.694604721	1
AR_X_66736338_66750729_66906874_669111452_RR	AR	0.563742659	0.563742659	20.89428568	3.82E-10	7.78E-07	13.52715233	1.478098751	1.478098751	1
AR_X_66818342_66825862_66950367_66956132_FF	AR	0.528294859	0.528294859	9.248783162	1.71E-06	0.000187011	5.588100983	1.442223604	1.442223604	1
AR_X_66911452_66916150_66950367_66956132_RR	AR	0.388697407	0.388697407	10.14296783	6.90E-07	9.11E-05	6.507167389	1.309210798	1.309210798	1
AR_X_66736338_66750729_66875649_66881776_FR	AR	0.370736884	0.370736884	5.172651841	0.000316014	0.004216579	0.216960602	1.293013093	1.293013093	1
ATM_11_108112750_108115594_108208085_108223747_FR	ATM	-0.370811815	-0.370811815	-8.707610279	3.07E-06	0.000260244	4.992731905	0.773347206	-1.293080251	-1
ATM_11_108155279_108156687_108208085_108223747_RR	ATM	0.363186489	0.363186489	6.602286154	4.02E-05	0.001238401	2.350592936	1.28626374	1.28626374	1
BMP6_6_7724582_7733496_7801590_7806316_FF	BMP6	-0.468602239	-0.468602239	-8.973309325	2.30E-06	0.00029279	5.288915333	0.722664415	-1.383768149	-1
BMP6_6_7724582_7733496_7743581_7746369_FR	BMP6	-0.388036314	-0.388036314	-5.889142945	0.000108497	0.002198909	1.322801574	0.764169025	-1.30861101	-1
CD44_11_35172600_35178637_35204720_35210484_FR	CD44	-0.398048925	-0.398048925	-5.668665177	0.000149604	0.002652749	0.990360448	0.758883891	-1.317724638	-1
CDH1_16_68794947_68799115_68857468_68863222_FR	CDH1	0.49540487	0.49540487	10.65592887	4.22E-07	6.84E-05	7.000965624	1.409716315	1.409716315	1
CTNNB1_3_41228301_41234483_41281934_41304993_FR	CTNNB1	0.427937487	0.427937487	10.8349435	3.57E-07	6.15E-05	7.167935447	1.345308914	1.345308914	1
DPP4_2_162933505_162942299_162961246_162964936_FR	DPP4	-0.512949291	-0.512949291	-14.57522664	1.71E-08	1.09E-05	10.1259903	0.700788356	-1.426964348	-1
DPP4_2_162946178_162949954_162972154_162979139_RF	DPP4	-0.445665864	-0.445665864	-3.573437421	0.004427816	0.023189841	-2.491256189	0.734245353	-1.361942565	-1
EGFR_7_55080257_55086091_55224588_55235839_RR	EGFR	-0.493404965	-0.493404965	-11.0577665	2.91E-07	5.37E-05	7.372042198	0.710346599	-1.407763479	-1

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	Genelocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
EP515_1_51804255_51813510_51945067_51946855_FF	EP515	0.386206462	0.386206462	8.698890643	3.10E-06	0.000260244	4.982882133	1.306952276	1.306952276	1
ERBB4_2_213060845_213063716_213336205_213346911_FR	ERBB4	-0.681305564	-0.681305564	-14.78217408	1.48E-08	1.00E-05	10.26456915	0.623600693	-1.603590265	-1
ERBB4_2_212789287_212798405_212962659_212969505_FR	ERBB4	-0.435496836	-0.435496836	-5.826671395	0.000118755	0.002319347	1.229315586	0.739439062	-1.352376485	-1
ERBB4_2_213052672_213059531_213336205_213346911_FR	ERBB4	-0.425728775	-0.425728775	-12.76218491	6.75E-08	2.15E-05	8.808033687	0.744462573	-1.343250872	-1
ERBB4_2_212789287_212798405_212846041_212850086_RF	ERBB4	-0.356744537	-0.356744537	-6.538422232	4.38E-05	0.001298756	2.261469357	0.780924761	-1.280533093	-1
ERBB4_2_212556994_212565232_212622803_212628844_FR	ERBB4	0.688270832	0.688270832	10.79268571	3.71E-07	6.29E-05	7.128763987	1.611351047	1.611351047	1
ERBB4_2_213182054_213190315_213317793_213323368_RR	ERBB4	0.447473513	0.447473513	10.33352546	5.73E-07	8.45E-05	6.6933320715	1.363650103	1.363650103	1
ERBB4_2_212622803_212628844_212789287_212798405_RF	ERBB4	0.416082272	0.416082272	5.631318317	0.000158077	0.002758619	0.9333355245	1.334299258	1.334299258	1
ERBB4_2_213151813_213159540_213182054_213190315_FF	ERBB4	0.378853461	0.378853461	3.217386837	0.008284402	0.035699071	-3.122932982	1.300308063	1.300308063	1
ERBB4_2_212556994_212565232_212858137_212868453_FF	ERBB4	0.359671724	0.359671724	6.923758849	2.62E-05	0.000993557	2.790707369	1.283133896	1.283133896	1
ERG_21_39895678_39899145_39984806_39991905_RF	ERG	-0.425291613	-0.425291613	-11.67074071	1.68E-07	3.57E-05	7.913111034	0.744688192	-1.342843905	-1
ESR1_6_152151654_152158599_152307023_152319013_RF	ESR1	-0.334294612	-0.334294612	-9.744626114	1.03E-06	0.000124223	6.107253454	0.793171853	-1.260760825	-1
ETS1_11_128387417_128389914_128489818_128498866_FF	ETS1	-0.529524556	-0.529524556	-8.680536392	3.17E-06	0.000260244	4.962121749	0.692783005	-1.443453423	-1
ETS1_11_128431527_128436474_128489818_128498866_RF	ETS1	-0.370215655	-0.370215655	-5.977392038	9.56E-05	0.002017625	1.453907443	0.77366684	-1.292546027	-1
ETS1_11_128342943_128345136_128399358_128409879_FF	ETS1	-0.344088814	-0.344088814	-4.169479623	0.001593588	0.012099379	-1.449395395	0.787805386	-1.269349026	-1
ETS1_11_128342943_128345136_128489818_128498866_FF	ETS1	-0.342780154	-0.342780154	-4.97505672	0.000429841	0.005055727	-0.1008332117	0.788520324	-1.26819813	-1
ETV1_7_13928482_13938998_14075713_14080964_FR	ETV1	-0.438646448	-0.438646448	-12.61661413	7.60E-08	2.29E-05	8.693428053	0.737826521	-1.355332144	-1
ETV1_7_13928482_13938998_14040827_14042620_FR	ETV1	-0.420116215	-0.420116215	-7.525930383	1.22E-05	0.000603601	3.57803973	0.747364419	-1.338035334	-1
FOLH1_11_49157869_49163274_49234427_49241370_FF	FOLH1	-0.327026922	-0.327026922	-4.300654557	0.001279787	0.010350202	-1.224472556	0.7971776	-1.254425613	-1
FOLH1_11_49214976_49217503_49234427_49241370_RF	FOLH1	0.418767287	0.418767287	6.637609483	3.83E-05	0.00120659	2.399644945	1.336784849	1.336784849	1
FOLH1_11_49157869_49163274_49193665_49198286_RR	FOLH1	0.359712744	0.359712744	10.54139329	4.70E-07	7.47E-05	6.892707121	1.28317038	1.28317038	1
GLPR1_12_75847260_75849629_75907812_75913956_FR	GLPR1	-0.345396052	-0.345396052	-5.155710032	0.000324389	0.004293149	0.189926553	0.787091873	-1.270499715	-1
GSK3B_3_119542459_119548768_119722182_119724690_FR	GSK3B	0.449273935	0.449273935	10.95895529	3.18E-07	5.68E-05	7.282033839	1.365352943	1.365352943	1
HGF_7_81320024_81325883_81430055_81434910_FF	HGF	-0.43281007	-0.43281007	-4.979466125	0.000426875	0.00504444	-0.093681464	0.740817421	-1.349860265	-1
IGFBP5_2_217560127_217567417_217584428_217589578_FR	IGFBP5	0.375003854	0.375003854	12.14650233	1.12E-07	2.78E-05	8.313515985	1.296843019	1.296843019	1

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	GeneLocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
IL16_15_81429756_81433873_81539851_815477011_FR	IL16	-0.481896248	-0.481896248	-9.127530593	1.95E-06	0.000208593	5.457383883	0.716035863	-1.396578093	-1
IL6_7_22721376_22727129_22765455_22766829_FR	IL6	0.364631786	0.364631786	4.801938715	0.00056537	0.005957969	-0.383664127	1.28755297	1.28755297	1
JUN_1_59244918_59246918_59258836_59260597_RF	JUN	-0.318607525	-0.318607525	-5.099115371	0.000354116	0.004543222	0.099326736	0.801843436	-1.247126254	-1
KIT_4_55553401_55555465_55610649_55618756_FR	KIT	0.392634635	0.392634635	7.232977499	1.76E-05	0.000779657	3.200921573	1.312788617	1.312788617	1
KRAS_12_25363357_25368892_25413300_25418096_FR	KRAS	0.414148209	0.414148209	11.99343016	1.28E-07	3.02E-05	8.186481963	1.332511707	1.332511707	1
IPAR3_1_85307233_85315383_85371057_85373099_RR	IPAR3	-0.38895037	-0.38895037	-13.49428502	3.80E-08	1.61E-05	9.363780155	0.76368502	-1.309440376	-1
IPAR3_1_85265679_85268722_85307233_85315383_RR	IPAR3	0.402766702	0.402766702	11.08898448	2.82E-07	5.32E-05	7.400314167	1.322040801	1.322040801	1
MAP2K5_15_67818519_67824995_68067482_68072379_FR	MAP2K5	0.404437029	0.404437029	9.01303797	2.20E-06	0.000221829	5.332553143	1.323572323	1.323572323	1
MAPKAP1_9_128370452_128376700_128393518_128397379_R	MAPKAP1	-0.396294954	-0.396294954	-12.18592711	1.08E-07	2.78E-05	8.345964775	0.759807072	-1.316123575	-1
MIR454_17_57199107_57202160_57227315_57228782_FF	MIR454	0.373313547	0.373313547	4.540644288	0.000861835	0.007908614	-0.818047279	1.295324487	1.295324487	1
MIR98_X_53595032_53600487_53628991_53630033_RR	MIR98	0.742460493	0.742460493	23.55441603	1.06E-10	5.41E-07	14.56868412	1.673026728	1.673026728	1
MIR98_X_53608013_53611637_53628991_53630033_RR	MIR98	0.67511652	0.67511652	13.76185645	3.10E-08	1.43E-05	9.558686586	1.596725728	1.596725728	1
MSR1_8_16213140_16220021_16405541_16412741_RR	MSR1	-0.589823054	-0.589823054	-8.317534796	4.77E-06	0.000337002	4.54380866	0.664424394	-1.50506214	-1
MSR1_8_16195878_16203315_16396849_16400398_FF	MSR1	-0.419369028	-0.419369028	-3.516010141	0.004895359	0.024952597	-2.59289798	0.747751587	-1.337342532	-1
MSR1_8_16045879_16049928_16079226_16088483_FF	MSR1	-0.385893508	-0.385893508	-10.14385541	6.89E-07	9.11E-05	6.508042059	0.765304873	-1.306668799	-1
MSR1_8_16142611_16149459_16195878_16203315_FF	MSR1	-0.337812738	-0.337812738	-6.009884545	9.13E-05	0.00198016	1.501898146	0.791239998	-1.263839041	-1
MSR1_8_16142611_16149459_16396849_16400398_RF	MSR1	-0.328903769	-0.328903769	-5.188518336	0.000308377	0.004162163	0.242242178	0.796141201	-1.256058596	-1
MSR1_8_16251114_16260512_16462527_16467449_FF	MSR1	-0.318902812	-0.318902812	-7.84658429	8.28E-06	0.000473286	3.978390344	0.801679333	-1.247381538	-1
MSR1_8_16195878_16203315_16433596_16442100_FR	MSR1	0.420145921	0.420145921	3.761840375	0.003192302	0.018914227	-2.159017761	1.338062886	1.338062886	1
NCOA1_2_24829718_248333469_24853776_24866328_RR	NCOA1	0.383599799	0.383599799	7.061388057	2.19E-05	0.000893094	2.974854782	1.304593006	1.304593006	1
NCOA1_2_24696090_24698819_24840193_24848780_RF	NCOA1	0.376458226	0.376458226	8.007884338	6.84E-06	0.000434693	4.175013545	1.298151018	1.298151018	1
NCOA1_2_24672976_24676297_24840193_24848780_RF	NCOA1	0.368303096	0.368303096	8.47574282	3.98E-06	0.000302485	4.72794807	1.290833654	1.290833654	1
NEDD4L_18_55713082_55720762_55811019_55814883_RR	NEDD4L	-0.444311776	-0.444311776	-12.22570821	1.05E-07	2.78E-05	8.378595915	0.734934827	-1.360664869	-1
NEDD4L_18_55713082_55720762_55848311_55850861_RR	NEDD4L	-0.390337066	-0.390337066	-12.14497251	1.12E-07	2.78E-05	8.312254649	0.76295133	-1.310699595	-1
NEDD4L_18_55713082_55720762_55882560_55885168_RR	NEDD4L	-0.37571287	-0.37571287	-12.60684517	7.66E-08	2.29E-05	8.685686665	0.770724485	-1.297480512	-1

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	Genelocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
NEDD4L_18_55713082_55720762_55869254_55872326_RR	NEDD4L	-0.360925867	-0.360925867	-11.91420916	1.36E-07	3.10E-05	8.120075504	0.778664702	-1.284249816	-1
NEDD4L_18_55713082_55720762_55774243_55779941_RR	NEDD4L	-0.35657074	-0.35657074	-9.2403661998	1.73E-06	0.000187011	5.579071341	0.781018843	-1.28037884	-1
NEDD4L_18_55713082_55720762_55961376_55965188_RR	NEDD4L	-0.356380382	-0.356380382	-8.945405461	2.37E-06	0.00023	5.258165908	0.781121902	-1.28020991	-1
NEDD4L_18_55713082_55720762_55784812_55787427_RR	NEDD4L	-0.354821703	-0.354821703	-12.0183087	1.25E-07	3.02E-05	8.207242683	0.781966277	-1.278827526	-1
NEDD4L_18_55713082_55720762_55986600_55989306_RR	NEDD4L	-0.349236239	-0.349236239	-11.79725824	1.51E-07	3.34E-05	8.021205936	0.784999566	-1.273886055	-1
NEDD4L_18_55713082_55720762_55927256_55929658_RR	NEDD4L	-0.343996768	-0.343996768	-8.731068109	3.00E-06	0.000260244	5.019188722	0.787855651	-1.269268043	-1
NEDD4L_18_55713082_55720762_55950636_55953432_RR	NEDD4L	-0.326323266	-0.326323266	-12.21462858	1.06E-07	2.78E-05	8.36951882	0.797566509	-1.253813932	-1
NEDD4L_18_55713082_55720762_55876126_55882560_RR	NEDD4L	-0.319472154	-0.319472154	-10.93796227	3.24E-07	5.69E-05	7.262808228	0.801363023	-1.2478739	-1
NF1_17_29477103_29483764_29709143_29714529_FF	NF1	0.640366513	0.640366513	17.71768057	2.20E-09	2.49E-06	12.01137937	1.5587251	1.5587251	1
NF1_17_29629862_29634257_29659279_29666456_FF	NF1	0.558103066	0.558103066	14.14945638	2.33E-08	1.29E-05	9.83363673	1.472332041	1.472332041	1
NFKB1_4_103436488_103442700_103548256_103555520_FF	NF1	0.451959985	0.451959985	13.17645951	4.86E-08	1.71E-05	9.126648681	1.367897363	1.367897363	1
NF1_17_29629862_29634257_29659279_29666456_FF	NF1	0.36665184	0.36665184	7.2650224	1.69E-05	0.000765315	3.242712391	1.289357057	1.289357057	1
NFKB1_4_103436488_103442700_103548256_103555520_FF	NFKB1	0.79980903	0.79980903	13.11977478	5.08E-08	1.72E-05	9.083698686	1.740870671	1.740870671	1
NFKB1_4_103425294_103430395_103548256_103555520_FF	NFKB1	0.430874236	0.430874236	14.09753934	2.42E-08	1.29E-05	9.797304101	1.348050213	1.348050213	1
NOVA1_14_26999345_27006013_27046501_27053973_FF	NOVA1	-0.461947202	-0.461947202	-11.22100125	2.51E-07	5.00E-05	7.519006961	0.726005709	-1.377399637	-1
NOVA1_14_26986332_26987866_27070837_27086602_FF	NOVA1	-0.325885172	-0.325885172	-7.120645256	2.03E-05	0.000844533	3.0533363996	0.797808737	-1.253433252	-1
NR4A3_9_102621891_102624499_102636939_102640160_FF	NR4A3	-0.326733183	-0.326733183	-6.549075716	4.32E-05	0.001283905	2.276375868	0.797339926	-1.254170233	-1
PIAS2_18_44419921_44425175_44533399_44538938_FF	PIAS2	-0.376922678	-0.376922678	-4.811482513	0.000556831	0.005918513	-0.367966827	0.770078446	-1.298569003	-1
PIK3C2A_11_17158103_17163660_17253125_17255535_FF	PIK3C2A	-0.412653759	-0.412653759	-9.581839254	1.21E-06	0.000138409	5.939488583	0.751240236	-1.331132109	-1
PIK3C2G_12_18503466_18517448_18605599_18615448_FF	PIK3C2G	-0.397193613	-0.397193613	-6.0651111351	8.44E-05	0.001886444	1.583120211	0.759333934	-1.316943647	-1
PIK3C2G_12_18682015_18689955_18755082_18765416_FF	PIK3C2G	-0.349259284	-0.349259284	-7.872086105	8.03E-06	0.000473286	4.009686303	0.784987026	-1.273906404	-1
PIK3C2G_12_18503466_18517448_18653437_18654550_FF	PIK3C2G	0.813156179	0.813156179	32.76884362	3.04E-12	3.10E-08	17.09863144	1.757051136	1.757051136	1
PIK3C2G_12_18503466_18517448_18800920_18805991_FF	PIK3C2G	0.52068763	0.52068763	19.49409228	8.00E-10	1.16E-06	12.9000512	1.434638875	1.434638875	1
PIK3C2G_12_18503466_18517448_18586459_18591749_FF	PIK3C2G	0.500422551	0.500422551	22.08623513	2.12E-10	7.17E-07	14.016338	1.414627832	1.414627832	1
PIK3C2G_12_18503466_18517448_18623979_18629934_FF	PIK3C2G	0.454500507	0.454500507	14.21973094	2.21E-08	1.29E-05	9.882576362	1.370308292	1.370308292	1

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	Genelocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
PIK3C2G_12_18466993_18474305_18503466_18517448_RF	PIK3C2G	0.418315764	0.418315764	7.846493305	8.28E-06	0.000473286	3.978278546	1.336366539	1.336366539	1
PIK3C2G_12_18407299_18408850_18503466_18517448_RF	PIK3C2G	0.417537091	0.417537091	12.19551672	1.08E-07	2.78E-05	8.35384099	1.335645449	1.335645449	1
PIK3C2G_12_18503466_18517448_18748288_18755082_FR	PIK3C2G	0.412545687	0.412545687	8.404871486	4.32E-06	0.000313738	4.645813078	1.331032398	1.331032398	1
PIK3C2G_12_18409429_18411730_18662602_18673822_RF	PIK3C2G	0.40413265	0.404135265	7.905355237	7.72E-06	0.000473286	4.050395593	1.323295505	1.323295505	1
PIK3C2G_12_18466993_18474305_18765637_18775643_FR	PIK3C2G	0.401350605	0.401350605	10.51089877	4.83E-07	7.57E-05	6.863693215	1.32074377	1.32074377	1
PRKCB_16_23929937_23938239_24143206_24145438_FR	PRKCB	0.359909066	0.359909066	12.83614979	6.36E-08	2.09E-05	8.865730305	1.283345005	1.283345005	1
PRKCH_14_61911060_61914582_62023126_62035192_FR	PRKCH	-0.411325245	-0.411325245	-5.079346062	0.000365169	0.004598091	0.067573492	0.751932339	-1.329906892	-1
PRKCH_14_61772357_61775932_61963825_61969638_RR	PRKCH	-0.335493699	-0.335493699	-7.529186724	1.22E-05	0.000603601	3.582169975	0.792512887	-1.261809134	-1
PTGS2_1_186630471_186639286_186675090_186678395_RR	PTGS2	-0.481332802	-0.481332802	-6.768676224	3.22E-05	0.001116912	2.580151439	0.716315566	-1.396032765	-1
PTPN14_1_214555543_214567111_214696754_214699528_RR	PTPN14	-0.656412698	-0.656412698	-6.179949212	7.18E-05	0.001709697	1.750618263	0.634453925	-1.576158584	-1
PTPN14_1_214555543_214567111_214590581_214592230_FF	PTPN14	-0.341216493	-0.341216493	-6.202805991	6.95E-05	0.001671497	1.783732259	0.789375423	-1.266824341	-1
PTPN14_1_214512778_214523707_214646434_214652454_FR	PTPN14	-0.318133487	-0.318133487	-4.495773109	0.000927417	0.008276791	-0.89351305	0.802106947	-1.246716543	-1
PTPN14_1_214555543_214567111_214643240_214644608_FR	PTPN14	0.743894345	0.743894345	14.95502699	1.31E-08	9.50E-06	10.37860359	1.674690326	1.674690326	1
PTPRR_12_71045661_71048060_71347632_71356891_FR	PTPRR	-0.321158046	-0.321158046	-5.612244267	0.0001626	0.002808651	0.904163728	0.80042712	-1.249332981	-1
PTPRR_12_71085097_71096639_71123929_71126257_FR	PTPRR	0.550492009	0.550492009	13.87391165	2.85E-08	1.38E-05	9.639060097	1.464585085	1.464585085	1
PTPRR_12_71085097_71096639_71150835_71153565_FR	PTPRR	0.37642789	0.37642789	7.745377455	9.35E-06	0.000508734	3.85340498	1.298123721	1.298123721	1
PTPRT_20_40761966_40770575_40995945_41003669_FR	PTPRT	0.390974701	0.390974701	5.898131356	0.000107101	0.002179292	1.336206195	1.31127902	1.31127902	1
PTPRT_20_40695490_40704819_40853486_40862226_RF	PTPRT	0.363092378	0.363092378	6.022994423	8.96E-05	0.001959674	1.521218337	1.286179837	1.286179837	1
RAN_12_131315466_131318726_131332056_131334187_RR	RAN	0.409645167	0.409645167	4.697687617	0.000668175	0.00665819	-0.555916376	1.328359062	1.328359062	1
RB1_13_48835536_48838517_49000831_49010576_FR	RB1	-0.421233583	-0.421233583	-6.218242572	6.80E-05	0.001663343	1.806054195	0.746785809	-1.339072045	-1
REL_2_61090704_61099366_61123363_61128146_FF	REL	-0.396733033	-0.396733033	-8.151247086	5.78E-06	0.000384323	4.347158457	0.759576389	-1.316523281	-1
REL_2_61090704_61099366_61149976_61161058_FF	REL	-0.36693886	-0.36693886	-7.752686136	9.27E-06	0.000506975	3.862472978	0.775426067	-1.289613597	-1
REL_2_61090704_61099366_61144132_61147262_FR	REL	0.677200702	0.677200702	15.31760281	1.02E-08	7.96E-06	10.6128694	1.599034097	1.599034097	1
RG56_14_72418571_72425681_72679959_72689252_RR	RG56	0.640246502	0.640246502	17.53962222	2.45E-09	2.49E-06	11.91593523	1.558595442	1.558595442	1
ROR2_9_94323433_94326108_94448327_94455574_FF	ROR2	-0.317797399	-0.317797399	-13.17593415	4.86E-08	1.71E-05	9.126251542	0.802293826	-1.246426144	-1

Table 19: Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	GeneLocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
SCGB1A1_11_62128712_62135211_62160970_62163465_FR	SCGB1A1	-0.657694922	-0.657694922	-10.36431148	5.56E-07	8.40E-05	6.723090349	0.633890292	-1.57756005	-1
SOS1_2_39227963_39230003_39353282_39361637_RF	SOS1	0.674466946	0.674466946	13.89064783	2.82E-08	1.38E-05	9.651002	1.596006963	1.596006963	1
SOS1_2_39209340_39220780_39276526_39280091_FR	SOS1	0.461207721	0.461207721	10.99304586	3.08E-07	5.60E-05	7.313177343	1.376693805	1.376693805	1
SRD5A2_2_31741633_31747723_31778586_31789876_FF	SRD5A2	-0.437278492	-0.437278492	-17.05212796	3.30E-09	3.05E-06	11.6482057	0.738526456	-1.354047634	-1
SRD5A2_2_31729027_31741633_31760980_31767977_FR	SRD5A2	0.394046044	0.394046044	9.411702288	1.44E-06	0.000162975	5.761374339	1.314073565	1.314073565	1
SRD5A2_2_31760980_31767977_31778586_31789876_RF	SRD5A2	0.36636811	0.36636811	11.12935664	2.72E-07	5.28E-05	7.436768688	1.289103509	1.289103509	1
TGFB2_1_218504155_218510817_218542394_218548723_RF	TGFB2	-0.401060991	-0.401060991	-18.11165268	1.74E-09	2.22E-06	12.21825228	0.757301142	-1.320478664	-1
TGFB2_1_218491029_218498929_218553354_218556593_FF	TGFB2	-0.386196098	-0.386196098	-7.834000204	8.41E-06	0.000476069	3.962917896	0.765144376	-1.306942888	-1
TMPRSS2_21_42841804_42850832_42927381_42930038_FR	TMPRSS2	0.46760546	0.46760546	8.609824216	3.43E-06	0.000272613	4.88179253	1.382812413	1.382812413	1
TOP2A_17_38547618_38549511_38613131_38616534_RR	TOP2A	-0.334752665	-0.334752665	-8.436098305	4.17E-06	0.000304987	4.682072992	0.792920063	-1.261161178	-1
TOP2A_17_38564762_38568693_38613131_38616534_RR	TOP2A	-0.33269637	-0.33269637	-9.020450371	2.18E-06	0.000221829	5.340676459	0.79405103	-1.259364905	-1
TOP2B_3_25644985_25663188_25716096_25717154_FF	TOP2B	0.677625777	0.677625777	11.54384037	1.88E-07	3.83E-05	7.803482889	1.599505304	1.599505304	1
VEGFC_4_177629821_177639626_177693384_177697283_FR	VEGFC	0.624813933	0.624813933	8.924483137	2.42E-06	0.00023044	5.235055692	1.542011936	1.542011936	1
VEGFC_4_177629821_177639626_177740221_177743175_FR	VEGFC	0.532875204	0.532875204	11.11732726	2.75E-07	5.28E-05	7.425914159	1.446809728	1.446809728	1
VEGFC_4_177629821_177639626_177693384_177697283_FF	VEGFC	0.418296493	0.418296493	13.22565823	4.68E-08	1.71E-05	9.163763582	1.336348688	1.336348688	1
EZH2_7_148496931_148503515_148602692_148606606_FF	EZH2	0.221213903	0.221213903	6.686469603	3.59E-05	0.001181426	2.467211314	1.165714021	1.165714021	1
EZH2_7_148496931_148503515_148610251_148614284_FR	EZH2	0.220468898	0.220468898	8.3111487407	4.80E-06	0.000337006	4.53671325	1.165112204	1.165112204	1
SP1_12_53752782_53754759_53771263_53775550_RF	SP1	0.197127842	0.197127842	4.00029596	0.002121214	0.014464632	-1.742110313	1.146413768	1.146413768	1
SP1_12_53771263_53775550_53824264_53827278_FR	SP1	0.193365041	0.193365041	3.676018582	0.003703744	0.02071572	-2.31009962	1.143427617	1.143427617	1
DAPK1_9_90064560_90073617_90176237_90180153_FF	DAPK1	-0.210075392	-0.210075392	-5.311660617	0.000255365	0.00370395	0.437247364	0.864492054	-1.156748631	-1
DAPK1_9_90064560_90073617_90339152_90340776_FF	DAPK1	-0.289887636	-0.289887636	-3.250481175	0.007812935	0.034291978	-3.064141545	0.817965763	-1.222545056	-1
DAPK1_9_90064560_90073617_90140806_90142738_FR	DAPK1	0.299375751	0.299375751	7.197207444	1.84E-05	0.000805368	3.154114326	1.230611817	1.230611817	1
DAPK1_9_90064560_90073617_90140806_90142738_FF	DAPK1	0.22308584	0.22308584	3.330633912	0.006781108	0.031049051	-2.92175958	1.167227549	1.167227549	1
FGD4_12_32760791_32767406_32781508_32786048_FR	FGD4	-0.274580142	-0.274580142	-5.050237381	0.000382113	0.0004709684	0.020720467	0.82669087	-1.209642004	-1
FGD4_12_32760791_32767406_32781508_32786048_RR	FGD4	-0.282349703	-0.282349703	-5.378249402	0.000230809	0.003536515	0.541798821	0.822250734	-1.216174043	-1

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	GeneLocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
FGD4_12_32714978_32722972_32735447_32738552_RR	FGD4	0.277743465	0.277743465	4.897130535	0.000486024	0.005488137	-0.227642215	1.212297233	1.212297233	1
FGD4_12_32714978_32722972_32768073_32772358_RR	FGD4	0.224649977	0.224649977	3.77524936	0.003119249	0.018634903	-2.135456788	1.168493717	1.168493717	1
GAB1_4_144235957_144242111_144369687_144374560_FF	GAB1	0.322283078	0.322283078	7.881995854	7.94E-06	0.000473286	4.021826263	1.250307607	1.250307607	1
GAB1_4_144272552_144276220_144402622_144411096_FR	GAB1	0.232807271	0.232807271	5.862864002	0.000112692	0.002243704	1.283544618	1.175119334	1.175119334	1
GAB1_4_144254752_144257034_144321110_144332903_RF	GAB1	-0.1933323	-0.1933323	-3.299161865	0.00716858	0.03218585	-2.977661138	0.874583297	-1.143401668	-1
GAB1_4_144298156_144300750_144321110_144332903_FR	GAB1	-0.203419346	-0.203419346	-6.533724514	4.41E-05	0.001301693	2.25489123	0.868489706	-1.151424126	-1
HSD3B2_1_119937390_119948935_119959754_119963670_FR	HSD3B2	0.207425802	0.207425802	6.208022809	6.90E-05	0.001671497	1.791279777	1.154626147	1.154626147	1
HSD3B2_1_119937390_119948935_119959754_119963670_FF	HSD3B2	0.167484698	0.167484698	5.70659508	0.000141491	0.002556892	1.048050136	1.123098683	1.123098683	1
HSD3B2_1_119912462_119915175_119959754_119963670_RR	HSD3B2	-0.168081632	-0.168081632	-3.274998031	0.007481356	0.033194645	-3.020586815	0.890025372	-1.123563476	-1
KLK2_19_51340459_51344004_51390533_51395187_FR	KLK2	-0.233986179	-0.233986179	-5.085188859	0.000361865	0.004584902	0.076963796	0.850282306	-1.176079983	-1
KLK2_19_51317027_51319938_51340459_51344004_FF	KLK2	0.259249519	0.259249519	3.256953029	0.007723978	0.033974819	-3.052644106	1.196855945	1.196855945	1
KLK2_19_51317027_51319938_51346270_51350944_FF	KLK2	0.251147882	0.251147882	3.297865414	0.007185018	0.032204968	-2.979964123	1.190153686	1.190153686	1
MAP3K14_17_43358197_43360790_43375304_43380378_FF	MAP3K14	0.227459078	0.227459078	4.562645956	0.000831476	0.00777086	-0.781135175	1.170771132	1.170771132	1
MAP3K14_17_43360790_43364282_43409961_43415408_FF	MAP3K14	0.198862372	0.198862372	4.316518127	0.00124648	0.010161606	-1.197399861	1.147792913	1.147792913	1
MAP3K14_17_43358197_43360790_43375304_43380378_RR	MAP3K14	-0.205626917	-0.205626917	-6.208573083	6.89E-05	0.001671497	1.792075671	0.867161784	-1.15318735	-1
MIF_22_24194490_24195811_24245843_24254074_RR	MIF	0.254727783	0.254727783	10.76065239	3.82E-07	6.37E-05	7.098970594	1.193110598	1.193110598	1
MIF_22_24206371_24208274_24245843_24254074_FR	MIF	0.207093745	0.207093745	5.104189064	0.000351337	0.004526272	0.107467258	1.154360424	1.154360424	1
MUC1_1_155176403_155179713_155191807_155193554_FR	MUC1	-0.209333417	-0.209333417	-6.160281821	7.38E-05	0.001740993	1.722065484	0.864936775	-1.156153871	-1
MUC1_1_155146523_155149986_155191807_155193554_FR	MUC1	-0.218468452	-0.218468452	-5.993061073	9.35E-05	0.001993646	1.477069135	0.859477364	-1.163497775	-1
RAD51_15_40972719_40979675_41025213_41027977_RF	RAD51	0.352545734	0.352545734	4.666716555	0.00070238	0.006893753	-0.607362881	1.276811662	1.276811662	1
RAD51_15_40937212_40938851_41025213_41027977_RF	RAD51	-0.242505623	-0.242505623	-4.646371743	0.000725849	0.007053284	-0.641225269	0.845275991	-1.183045551	-1
RAD51_15_41009919_41011826_41025213_41027977_RF	RAD51	-0.243901539	-0.243901539	-6.598967119	4.03E-05	0.001240188	2.34597507	0.844458518	-1.184190791	-1
RNASEL_1_182541376_182556600_182605098_182607856_FR	RNASEL	0.268497887	0.268497887	3.858975685	0.002700604	0.016960458	-1.988638304	1.204553012	1.204553012	1
RNASEL_1_182541376_182556600_182577916_182584530_FF	RNASEL	0.25710383	0.25710383	3.235840558	0.008018044	0.03489118	-3.090150814	1.195077211	1.195077211	1
SRC_20_35928873_35935451_35989678_35993330_FR	SRC	0.206516047	0.206516047	6.456694813	4.89E-05	0.001394478	2.146589494	1.153898277	1.153898277	1

Table 19. Markers for prognostic array stratifications category 1 vs 3 and category 2 vs 3. Top 181 markers produced from the prognostic array.

Probes	GeneLocus	logFC	AveExpr	t	P.Value	adj.P.Val	B	FC	FC_1	Binary
SRC_20_35928873_35935451_35989678_35993330_FF	SRC	0.198581558	0.198581558	6.507686654	4.56E-05	0.001334586	2.218375228	1.147569522	1.147569522	1
SRD5A3_4_56188038_56191526_56242301_56245314_RF	SRD5A3	0.266992266	0.266992266	4.835274287	0.000536131	0.005815136	-0.328887879	1.203296575	1.203296575	1
SRD5A3_4_56209429_56213336_56242301_56245314_RF	SRD5A3	0.239396914	0.239396914	4.842348143	0.000530134	0.005781472	-0.317283393	1.180499078	1.180499078	1
WNT1_12_49327866_49332429_49386082_49387249_RF	WNT1	0.171379721	0.171379721	4.015889993	0.002065728	0.014255673	-1.715014756	1.126134949	1.126134949	1
WNT1_12_49361168_49364315_49377006_49380965_RF	WNT1	-0.188758659	-0.188758659	-6.377243699	5.46E-05	0.001476292	2.0340141	0.877360306	-1.139782588	-1
WNT1_12_49327866_49332429_49364343_49365445_FF	WNT1	-0.289012147	-0.289012147	-10.04098231	7.63E-07	9.95E-05	6.406185639	0.81846229	-1.221803389	-1

Abbreviations. logFC: logarithm of the fold change; AveExpr: Average expression; adj.P.Val: Adjusted p-value; B: B-statistic (log-odds that that gene is differentially expressed); FC: Fold change; FC_1: Fold change centered around 1; Binary: Binary call for loop presence/absence.

Table 20. DLBCL cell lines used in this study. Cell lines were obtained from the American Type Culture Collection (ATCC), the German Collection of Microorganisms and Cell Cultures (DSMZ), and the Japan Health Sciences Foundation Resource Bank (JHSF).

Cell Line	Cat#	Source	DLBCL Subtype
Toledo	CRL-2631	ATCC	ABC
Pfeiffer	CRL-2632	ATCC	ABC
RC-K8	ACC 561	DSMZ	ABC
Ri-1	ACC 585	DSMZ	ABC
A4/Fukada	JCRB 0097	JHSF	ABC
A3/Kawakami	JCRB 0101	JHSF	ABC
RL	CRL 2261	ATCC	GCB
HT	CRL-2260	ATCC	GCB
DB	CRL-2289	ATCC	GCB
Karpas-422	ACC 32	DSMZ	GCB
SU-DHL-10	ACC 576	DSMZ	GCB
SU-DHL-16	ACC 577	DSMZ	GCB

Table 21. The 97 genomic loci used in the initial biomarker discovery screen.

Gene Symbol				
ABCG2	CD40	IL-10	MMP9	SFPQ
ANXA11	CD80	IL10RA	MTHFR	SIAH1
ARRB2	CDKN2C	IL-15	MyD88	SIRT1
ASPSCR1	CREB3L2	IL22RA1	NAE1	SKP1
BAX	CTNNB1	IL-2RA	NCKIPSD	SOCS7
BBS9	CXCL8	IL-2RB	NFATc1	STAT3
BCL2A1	DBF4	IL-6	NFKB1	TAL2
BCL2L10	ERC1	IL-7	NFKB2	TET2
BCL6	ETV6	ITGA5	NFKBIA	TLR4
BRCA1	FCGR2A	ITPR3	NFKBIB	TNF
BTK	FOS	JAK3	PAK1	TNFRSF12A
c13orf34	FOXO1	JDP2	PCDHGA6/B2/B4	TNFRSF13B
c15orf55	FOXP1	LRP6	PIK3CG	TNFRSF13C
c21orf45	FRAP1	MAP3K7	PIM1	TOP1
CABLES1	FZD10	MAPK10	PRDM1	WNT11
CAMK2D	GATA4	MAPK13	PRKCZ	WNT9A
CASP10	GDF6	MEF2B	PTGS2	ZMYM3
CASP3	GRAP2	MLLT3	RBL1	
CCR9	HLF	MMP14	RCA1	
CD22	IFNAR1	MMP2	REL	

Table 22. Composite prediction probabilities for the DLBCL-CCS in the Discovery cohort.

Patient_ID	Class	EpiSwitch_ABC_GBC_Prob	Patient_ID	Class	EpiSwitch_ABC_GBC_Prob
G139474	ABC	0.1932724	G580501	GCB	0.8140926
G510833	ABC	0.20029	G812590	GCB	0.813454
G779155	ABC	0.2110736	G139629	GCB	0.8133554
G425812	ABC	0.212716	G254772	GCB	0.8095988
G489616	ABC	0.2139196	G515146	GCB	0.8066254
G258685	ABC	0.2154116	G3383	GCB	0.8038896
G483540	ABC	0.2168548	G694634	GCB	0.7949872
G635399	ABC	0.216758	G158912	GCB	0.7817198
G61310	ABC	0.218381	G833688	GCB	0.780263
G935357	ABC	0.2184674	G259245	GCB	0.7739536
G232546	ABC	0.2190392	G81760	GCB	0.7730372
G783641	ABC	0.2227972	G547813	GCB	0.7723438
G421531	ABC	0.223797	G913775	GCB	0.7705852
G707842	ABC	0.225338	G474390	GCB	0.7670176
G78398	ABC	0.2266852	G213298	GCB	0.763692
G855052	ABC	0.2267892	G153040	GCB	0.762577
G985511	ABC	0.2373018	G108325	GCB	0.762146
G529835	ABC	0.2380336	G243731	GCB	0.7621028
G173235	ABC	0.2385374	G819965	GCB	0.7593464
G418054	ABC	0.2411416	G701045	GCB	0.7579728
G577623	ABC	0.2429652	G544037	GCB	0.756705
G415338	ABC	0.2478544	G370848	GCB	0.7554284
G175733	ABC	0.2580098	G739109	GCB	0.754522
G852875	ABC	0.2595216	G826574	GCB	0.7538074
G292009	ABC	0.2633318	G779214	GCB	0.7536828
G52081	ABC	0.2763932	G572374	GCB	0.7526758
G544695	ABC	0.276862	G801049	GCB	0.7491872
G880954	ABC	0.2855876	G937404	GCB	0.7285326
G181400	ABC	0.297829	G254120	GCB	0.7000562
G418564	ABC	0.2980288			

Table 23. DLBCL-CCS and Fluidigm subtype calls in the Discovery cohort. Subtype calls made by the *EpiSwitch* DLBCL-CCS and the Fluidigm assays on samples of known DLBCL subtypes. 60 out of 60 samples were identically called as ABC or GCB by both assays.

ABC			GCB		
Patient ID	Fluidigm	EpiSwitch	Patient ID	Fluidigm	EpiSwitch
RG332787	ABC	ABC	RG949161	GCB	GCB
RG857282	ABC	ABC	RG552773	GCB	GCB
RG639274	ABC	ABC	RG385960	GCB	GCB
RG227462	ABC	ABC	RG713290	GCB	GCB
RG898976	ABC	ABC	RG874071	GCB	GCB
RG469063	ABC	ABC	RG475279	GCB	GCB
RG235350	ABC	ABC	RG885516	GCB	GCB
RG341829	ABC	ABC	RG681434	GCB	GCB
RG769788	ABC	ABC	RG231526	GCB	GCB
RG401919	ABC	ABC	RG855093	GCB	GCB
RG849927	ABC	ABC	RG458634	GCB	GCB
RG714326	ABC	ABC	RG279476	GCB	GCB
RG109735	ABC	ABC	RG373871	GCB	GCB
RG563907	ABC	ABC	RG853726	GCB	GCB
RG847865	ABC	ABC	RG101525	GCB	GCB
RG698196	ABC	ABC	RG525277	GCB	GCB
RG208608	ABC	ABC	RG954268	GCB	GCB
RG126501	ABC	ABC	RG521469	GCB	GCB
RG988758	ABC	ABC	RG673708	GCB	GCB
RG436104	ABC	ABC	RG386174	GCB	GCB
RG611396	ABC	ABC	RG380741	GCB	GCB
RG565461	ABC	ABC	RG132060	GCB	GCB
RG513781	ABC	ABC	RG118710	GCB	GCB
RG549011	ABC	ABC	RG578086	GCB	GCB
RG233693	ABC	ABC	RG542280	GCB	GCB
RG192075	ABC	ABC	RG313590	GCB	GCB
RG374916	ABC	ABC	RG387871	GCB	GCB
RG410219	ABC	ABC	RG108874	GCB	GCB
RG216984	ABC	ABC	RG883839	GCB	GCB
RG538574	ABC	ABC	RG489043	GCB	GCB

Table 24. Enrichment of biological functions in the top 10 DLBCL-CCS loci.

Analysis Type	Pathway ID	Pathway Description	Gene Count	FDR	Matching Proteins
GO-Molecular Function	GO:0001228	transcriptional activator activity, RNA polymerase II transcription regulatory region	4	0.0365	MEF2B, NFATC1, NFKB1, STAT3
GO-Biological Process	GO:0045893	positive regulation of transcription, DNA-templated	7	0.00187	CD40, IFNAR1, MAP3K7, MEF2B, NFATC1, NFKB1, STAT3
KEGG Pathway	4620	Toll-like receptor signaling pathway	4	2.21E-05	CD40, IFNAR1, MAP3K7, NFKB1

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No.	Marker Details			
	Type	Genome Mapped to	Probe	GeneLocus
1	Diagnostic	GRCh37	ETS1_11_128419843_128421939_128481262_128489818_RR	ETS1
2	Diagnostic	GRCh37	SLC22A3_6_160805748_160812960_160839018_160842982_RR	SLC22A3
3	Diagnostic	GRCh37	SLC22A3_6_160805748_160812960_160884099_160888471_RR	SLC22A3
4	Diagnostic	GRCh37	MAP3K14_17_43360790_43364282_43409961_43415408_FR	MAP3K14
5	Diagnostic	GRCh37	CASP2_7_142940014_142947169_142963973_142967512_FR	CASP2
6	Prognostic 3v1	GRCh37	BMP6_6_7724582_7733496_7801590_7806316_FF	BMP6
7	Prognostic 3v1	GRCh37	ACAT1_11_107955219_107960166_108013361_108018367_FF	ACAT1
8	Prognostic 3v1	GRCh37	ERG_21_39895678_39899145_39984806_39991905_RF	ERG
9	Prognostic 3v1	GRCh37	MSR1_8_16195878_16203315_16396849_16400398_FF	MSR1
10	Prognostic 3v1	GRCh37	MUC1_1_155146523_155149986_155191807_155193554_FR	MUC1
11	Prognostic 3v1	GRCh37	DAPK1_9_90064560_90073617_90140806_90142738_FR	DAPK1
12	Prognostic 3v2	GRCh37	ACAT1_11_107955219_107960166_108013361_108018367_FF	ACAT1
13	Prognostic 3v2	GRCh37	MUC1_1_155146523_155149986_155191807_155193554_FR	MUC1
14	Prognostic 3v2	GRCh37	DAPK1_9_90064560_90073617_90140806_90142738_FR	DAPK1
15	Prognostic 3v2	GRCh37	APAF1_12_99061113_99062942_99098781_99108240_FF	APAF1
16	Prognostic 3v2	GRCh37	HSD3B2_1_119912462_119915175_119959754_119963670_RR	HSD3B2
17	Prognostic 3v2	GRCh37	VEGFC_4_177629821_177639626_177740221_177743175_FR	VEGFC

Table 25.a

No.	Hyper G array stats					Microarray output	
	Probe_Count_Total	Probe_Count_Sig	HyperG_Stats	FDR_HyperG	Percent_Sig	logFC	AveExpr
1	100	22	0.143767534	0.706164223	22	0.788832719	0.788832719
2	54	16	0.019214151	0.218625878	29.63	0.739725229	0.739725229
3	54	16	0.019214151	0.218625878	29.63	0.729027457	0.729027457
4	11	5	0.029574086	0.259389379	45.45	0.735407293	0.735407293
5	13	3	0.402919615	1	23.08	-0.469997725	-0.469997725
6	69	8	0.366815399	1	11.59	-0.468602239	-0.468602239
7	15	2	0.441893041	1	13.33	-0.436725529	-0.436725529
8	52	4	0.765503518	1	7.69	-0.425291613	-0.425291613
9	191	41	1.07E-06	0.000448644	21.47	-0.419369028	-0.419369028
10	5	3	0.008132099	0.285301135	60	-0.218468452	-0.218468452
11	46	9	0.032709022	0.548212211	19.57	0.299375751	0.299375751
12	15	2	0.441893041	1	13.33	-0.436725529	-0.436725529

13	5	3	0.008132099	0.285301135	60	-0.218468452	-0.218468452
14	46	9	0.032709022	0.548212211	19.57	0.299375751	0.299375751
15	10	1	0.644810187	1	10	-0.441488336	-0.441488336
16	20	5	0.040338404	0.548212211	25	-0.168081632	-0.168081632
17	57	16	8.02E-05	0.006755982	28.07	0.532875204	0.532875204

Table 25.b

No.	Microarray output					
	T	P.Value	adj.P.Val	B	FC	FC_1
1	15.59116667	0.0000000108	0.00000135	10.64875918	1.727676038	1.727676038
2	18.80485468	0.00000000155	0.00000124	12.53177853	1.669857773	1.669857773
3	19.34951235	0.00000000115	0.00000112	12.81371179	1.657521354	1.657521354
4	15.29282549	0.0000000131	0.00000138	10.45220415	1.664867419	1.664867419
5	-13.28933415	0.000000055	0.00000252	9.016293707	0.721965736	-1.385107284
6	-8.973309325	0.00000230	0.000229279	5.288915333	0.722664415	-1.383768149
7	-11.90723067	0.000000137	0.0000310	8.114204006	0.738809576	-1.353528748
8	-11.67074071	0.000000168	0.0000357	7.913111034	0.744688192	-1.342843905
9	-3.516010141	0.004895359	0.024952597	-2.59289798	0.747751587	-1.337342532
10	-5.993061073	0.0000935	0.001993646	1.477069135	0.859477364	-1.163497775
11	7.197207444	0.0000184	0.000805368	3.154114326	1.230611817	1.230611817
12	-11.90723067	0.000000137	0.0000310	8.114204006	0.738809576	-1.353528748
13	-5.993061073	0.0000935	0.001993646	1.477069135	0.859477364	-1.163497775
14	7.197207444	0.0000184	0.000805368	3.154114326	1.230611817	1.230611817
15	-13.23940926	0.0000000463	0.0000171	9.174110234	0.736374546	-1.358004571
16	-3.274998031	0.007481356	0.033194645	-3.020586815	0.890025372	-1.123563476
17	11.11732726	0.000000275	0.0000528	7.425914159	1.446809728	1.446809728

Table 25.c

No.	Microarray output		Probe sequence
	LS	Loop Detected	60 mer
1	1	Pca	CCATGGTGTGAGTGTGGATTAGGTGAATCGAAAGATCTAGTAGGTTCTGTCCAGACTGT
2	1	Pca	AATTCTGAGGGTGAAGGAAGGTGGGAGTGCATGGCTCTTATGCAGCATTATTATCAAT
3	1	Pca	AATTCTGAGGGTGAAGGAAGGTGGGAGTGCAGGGACTTTCAGGTAGAGGAGCCACCAAG
4	1	Pca	AGGGGCTGATCAGTTTGTGGAGTTCTGATCGAGGGAGAGGAGTGGCAGTGGGGGAGTGGGA
5	-1	HC	TCCAGAAGCTGAGCTTGAGCCAAGGTGTTCGAACTCCTGGGCTGAAGCAATCTCCTGCCT
6	-1	2_3	ACGTCGTTACAGTTTTAATTTTCTACTTCGATGTTAATCTCCTAAAAAACATCCAACCA
7	-1	1	CAATTGGTGGATATAGAAAGGTCTAAATTCGATAAGTATAGACTCAGAATGCAAAAATGT
8	-1	2_3	TCTTGAATGTGCTTAGTATTATTCAGACTCGAAAAACATAATTGAAAGGAATTCATTCTG
9	-1	2_3	CACCAGTTGGTAATTCTATGTGTAAGTTTCGAGCTTATAAGATCAATCAGGAATTATTCC
10	-1	3	GCAGGGTGGCTATAGCTCAGGAGAGTGCTCGACGGAGTCTTGCTCTTTCACCCAGGCTGG
11	1	3	ACTAATCCCCTGAAGAAGCAAATTAACCTTCGAGTATCCCTTTAAGTTTGTTTTAAAAATA
12	-1	1	CAATTGGTGGATATAGAAAGGTCTAAATTCGATAAGTATAGACTCAGAATGCAAAAATGT
13	-1	3	GCAGGGTGGCTATAGCTCAGGAGAGTGCTCGACGGAGTCTTGCTCTTTCACCCAGGCTGG
14	1	3	ACTAATCCCCTGAAGAAGCAAATTAACCTTCGAGTATCCCTTTAAGTTTGTTTTAAAAATA
15	-1	3	CCTAATTTACTTAACCAAACTCTAGTTATCGAACATCCAGGATGTTATAAGAATTCAATG
16	-1	3	TCAGTTTCTGCTCTCAAGAAGCTTACAGTCAAGGTCCCAAGTTAGATTACGGCAAAGCT
17	1	3	TTTTATGAAACATCCAACCTAAATATAATCGAATGCATTACATTACAGAACTATTTCCA

Table 25.d

No	Probe Location					4 kb Sequence Location			
	Chr	Start1	End1	Start2	End2	Chr	Start1	End1	Start2
1	11	128419843	128419873	128481262	128481292	11	128419843	128423843	128481262
2	6	160805748	160805778	160839018	160839048	6	160805748	160809748	160839018
3	6	160805748	160805778	160884099	160884129	6	160805748	160809748	160884099
4	17	43364251	43364282	43409961	43409991	17	43360281	43364282	43409961
5	7	142947138	142947169	142963973	142964003	7	142943168	142947169	142963973
6	6	7733465	7733496	7806286	7806316	6	7729496	7733496	7802316
7	11	107960135	107960166	108018337	108018367	11	107956166	107960166	108014367
8	21	39895678	39895708	39991875	39991905	21	39895678	39899678	39987905
9	8	16203284	16203315	16400368	16400398	8	16199315	16203315	16396398
10	1	155149955	155149986	155191807	155191837	1	155145986	155149986	155191807
11	9	90073586	90073617	90140806	90140836	9	90069617	90073617	90140806
12	11	107960135	107960166	108018337	108018367	11	107956166	107960166	108014367
13	1	155149955	155149986	155191807	155191837	1	155145986	155149986	155191807
14	9	90073586	90073617	90140806	90140836	9	90069617	90073617	90140806
15	12	99062911	99062942	99108209	99108240	12	99058941	99062942	99104239
16	1	119912462	119912492	119959754	119959784	1	119912462	119916462	119959754
17	4	177639595	177639626	177740221	177740251	4	177635625	177639626	177740221

Table 25.e

No.	4 kb Sequence Location		Inner primers
	End2	Probe	PCR-Primer1_ID
1	128485262	ETS1_11_128419843_128421939_128481262_128489818_RR	PCa-57
2	160843018	SLC22A3_6_160805748_160812960_160839018_160842982_RR	PCa-73
3	160888099	SLC22A3_6_160805748_160812960_160884099_160888471_RR	PCa-77
4	43413961	MAP3K14_17_43360790_43364282_43409961_43415408_FR	PCa-81
5	142967973	CASP2_7_142940014_142947169_142963973_142967512_FR	PCa-189
6	7806316	BMP6_6_7724582_7733496_7801590_7806316_FF	PCa119-37
7	108018367	ACAT1_11_107955219_107960166_108013361_108018367_FF	PCa119-57
8	39991905	ERG_21_39895678_39899145_39984806_39991905_RF	PCa119-65
9	16400398	MSR1_8_16195878_16203315_16396849_16400398_FF	PCa119-77
10	155195807	MUC1_1_155146523_155149986_155191807_155193554_FR	PCa119-121
11	90144806	DAPK1_9_90064560_90073617_90140806_90142738_FR	PCa119-165
12	108018367	ACAT1_11_107955219_107960166_108013361_108018367_FF	PCa119-57
13	155195807	MUC1_1_155146523_155149986_155191807_155193554_FR	PCa119-121
14	90144806	DAPK1_9_90064560_90073617_90140806_90142738_FR	PCa119-165
15	99108240	APAF1_12_99061113_99062942_99098781_99108240_FF	PCa119-49
16	119963754	HSD3B2_1_119912462_119915175_119959754_119963670_RR	PCa119-129

17	177744221	VEGFC_4_177629821_177639626_177740221_177743175_FR	PCa119-205
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Table 25.f

No.	Inner primers		
	PCR_Primer1	PCR-Primer2_ID	PCR_Primer2
1	CACTGCATGAGGGTAGTATAG	PCa-59	CCTCTGTCTGCATCATACC
2	TGATGAGGCACACAGATAAAG	PCa-75	ACACGCCCAGAAACAATAC
3	GAGACATGATGAGGCACAC	PCa-79	GTGTGAGTTGATAGCTGACC
4	TGGAATGGGAAGGGATGAG	PCa-83	GAGACTCCAGGCAAGAATTTG
5	ATGAAGACAGAAAGCCTATGG	PCa-191	CAGTGGAACCTCCTGAGAAC
6	CGGCCAGGAATGACTATTG	PCa119-39	GTAAGCGAGGTCATCATAGAAG
7	AGTAGTGTATCAGGACTGGGT	PCa119-59	TCTTGGTAACCTTGAAAAGTTTGAT
8	CAGCCTACCTTGCCTGACACT	PCa119-67	ATGGGCCATCACTGGGCTTT
9	AATCCTCTTGAGCACAGACC	PCa119-79	TAGGCCCAAATGGCTCAC
10	TGTTGCTAGCTCAGGAAGCC	PCa119-123	AGATCAAGCCACTGTGCTCC
11	ACTGGTCACAGGGAACGATGG	PCa119-167	AGGTGTGAATGTTACTGAACACAAA
12	AGTAGTGTATCAGGACTGGGT	PCa119-59	TCTTGGTAACCTTGAAAAGTTTGAT
13	TGTTGCTAGCTCAGGAAGCC	PCa119-123	AGATCAAGCCACTGTGCTCC
14	ACTGGTCACAGGGAACGATGG	PCa119-167	AGGTGTGAATGTTACTGAACACAAA
15	GGTATTCCAATAAATACTTGTGCCC	PCa119-51	TACTGTGCCAGATGCTCTCA
16	TCACATCAGTTTCTGCTCTCAAG	PCa119-131	GGAGGGAGGCTCAGAGAAGC
17	TCTCTGACTGCAGTGCAAAATAAT	PCa119-207	CTCCTTCTACATTACGTGCTTTCA

Table 25.g

No.	PCR Stats		
	Gene	Marker	GLMNET
1	ETS1	Pca-57.59	-0.00000007417665
2	SLC22A3	Pca-73.75	0.00000001852548
3	SLC22A3	Pca-77.79	0.00000002568381
4	MAP3K14	Pca-81.83	0.00000001902257
5	CASP2	Pca-189.191	0.0000001325828
6	BMP6	PCa-119-37.39	0.000009609007
7	ACAT1	PCa-119-57.59	0.000004371579
8	ERG	PCa-119-65.67	0.000006321361
9	MSR1	PCa-119-77.79	0.000005500154
10	MUC1	PCa-119-121.123	0.00000006234414
11	DAPK1	PCa-119-165.167	-0.00001571847
12	ACAT1	PCa-119-57.59	0.000004371579
13	MUC1	PCa-119-121.123	0.00000006234414
14	DAPK1	PCa-119-165.167	-0.00001571847
15	APAF1	PCa-119-49.51	0.000003531754
16	HSD3B2	PCa-119-129.131	0.0000004472913
17	VEGFC	PCa-119-205.207	-0.0000006807692

5 Table 25.h

CLAIMS

1. A process for detecting a chromosome state which represents a subgroup in a population comprising determining whether a chromosome interaction relating to that chromosome state is present or absent within a defined region of the genome, wherein the subgroup relates to prognosis for prostate cancer and wherein the chromosome interaction corresponds to any one of the chromosome interactions represented by any probe shown in Table 6.

2. A process according to claim 1 which further comprises detecting a chromosome state which represents a subgroup in a population comprising determining whether a chromosome interaction relating to that chromosome state is present or absent within a defined region of the genome; and
- wherein the subgroup relates to prognosis for DLBCL and the chromosome interaction corresponds to any one of the chromosome interactions represented by any probe shown in Table 5;

or

- wherein the subgroup relates to prognosis for lymphoma and the chromosome interaction corresponds to any one of the chromosome interactions shown in Table 8.

3. A process according to claim 1 or 2 wherein:

- said prognosis for prostate cancer relates to whether or not the cancer is aggressive or indolent; and/or

- said prognosis for DLBCL relates to survival.

4. A process according to any one of the preceding claims wherein the subgroup relates to prostate cancer and a specific combination of chromosome interactions are typed:

(i) comprising all of the chromosome interactions represented by the probes in Table 6; and/or

(ii) comprising at least 1, 2, 3 or 4 of the chromosome interactions represented by the probes in Table 6.

5. A process according to any one of claims 2 to 4 wherein the subgroup relates to DLBCL and a specific combination of chromosome interactions are typed:

(i) comprising all of the chromosome interactions represented by the probes in Table 5; and/or

(ii) comprising at least 10, 20, 30, 50 or 80 of the chromosome interactions represented by the probes in Table 5.

6. A process according to any one of claims 2 to 5 wherein the subgroup relates to DLBCL and a specific combination of chromosome interactions are typed:

- (i) comprising all of the chromosome interactions shown in Table 7; and/or
- (ii) comprising at least 1, 2, 5 or 8 of the chromosome interactions shown in Table 7.

7. A process according to any one of claims 2 to 6 wherein the subgroup relates to lymphoma and a specific combination of chromosome interactions are typed:

- (i) comprising all of the chromosome interactions shown in Table 8; and/or
- (ii) comprising at least 10, 20, 30 or 50 of the chromosome interactions shown in Table 8

or preferably a specific combination of chromosome interactions are typed:

- (a) comprising all of the chromosome interactions shown in Table 9; and/or
- (b) comprising at least 5, 10 or 15 of the chromosome interactions shown in Table 9.

8. A process according to any one of the preceding claims wherein at least 10, 20, 30, 40 or 50, chromosome interactions are typed, and preferably at least 10 chromosome interactions are typed.

9. A process according to any one of the preceding claims in which the chromosome interactions are typed:

- in a sample from an individual, and/or
- by detecting the presence or absence of a DNA loop at the site of the chromosome interactions, and/or
- detecting the presence or absence of distal regions of a chromosome being brought together in a chromosome conformation, and/or
- by detecting the presence of a ligated nucleic acid which is generated during said typing and whose sequence comprises two regions each corresponding to the regions of the chromosome which come together in the chromosome interaction, wherein detection of the ligated nucleic acid is preferably by:

(i) in the case of prognosis of prostate cancer by a probe that has at least 70% identity to any of the specific probe sequences mentioned in Table 6, and/or (ii) by a primer pair which has at least 70%

identity to any primer pair in Table 6; or

(ii) in the case of prognosis of DLBCL a probe that has at least 70% identity to any of the specific probe sequences mentioned in Table 5, and/or (b) by a primer pair which has at least 70% identity to any primer pair in Table 5.

10. A process according to any one of the preceding claims in which the chromosome interactions are typed by detecting the presence of a ligated nucleic acid which is generated during said typing and whose sequence comprises two regions each corresponding to the regions of the chromosome which come together in the chromosome interaction, wherein detection of the ligated nucleic acid in the case of prognosis of lymphoma is by:

- a probe that has at least 70% identity to any of the specific probe sequences mentioned in Table 5, and/or
- by a primer pair which has at least 70% identity to any primer pair in Table 5, and/or
- by a primer pair which has at least 70% identity to any primer pair in Table 8.

11. A process according to any one of the preceding claims, wherein the chromosome interaction is detected by a method comprising the steps of: -

- (i) cross-linking of chromosome regions which have come together in a chromosome interaction;
- (ii) subjecting said cross-linked regions to cleavage, optionally by restriction digestion cleavage with an enzyme;
- (iii) ligating said cross-linked cleaved DNA ends to form ligated nucleic acids and.
- (iv) detecting the presence or absence of a ligated nucleic acid corresponding to the chromosome interaction.

12. A process according to any one of the preceding claims which is carried out to determine whether a prostate cancer is aggressive or indolent which comprises typing at least 5 chromosome interactions as defined in Table 6.

13. A process according to any one of the claims 2 to 12 which determines prognosis of DLBCL and which comprises typing at least 5 chromosome interactions as defined in Table 5.

14. A process according to any one of the preceding claims which is carried out to identify or design a therapeutic agent for prostate cancer;

- wherein preferably said process is used to detect whether a candidate agent is able to cause a change to a chromosome state which is associated with a different level of prognosis;
- wherein the chromosomal interaction is represented by any probe in Table 6; and/or
- the chromosomal interaction is present in any region or gene listed in Table 6;

and wherein optionally:

- the chromosomal interaction has been identified by the method of determining which chromosomal interactions are relevant to a chromosome state as defined in claim 1, and/or
- the change in chromosomal interaction is monitored using (i) a probe that has at least 70% identity to any of the probe sequences mentioned in Table 6, and/or (ii) by a primer pair which has at least 70% identity to any primer pair in Table 6.

15. A process according to any one of preceding claims 2 to 13 which is carried out to identify or design a therapeutic agent for DLBCL;

- wherein preferably said process is used to detect whether a candidate agent is able to cause a change to a chromosome state which is associated with a different level of prognosis;
- wherein the chromosomal interaction is represented by any probe in Table 5; and/or
- the chromosomal interaction is present in any region or gene listed in Table 5;

and wherein optionally:

- the chromosomal interaction has been identified by the method of determining which chromosomal interactions are relevant to a chromosome state as defined in claim 1, and/or
- the change in chromosomal interaction is monitored using (i) a probe that has at least 70% identity to any of the probe sequences mentioned in Table 5, and/or (ii) by a primer pair which has at least 70% identity to any primer pair in Table 5.

16. A process according to any one of preceding claims 2 to 13 which is carried out to identify or design a therapeutic agent for lymphoma;

- wherein preferably said process is used to detect whether a candidate agent is able to cause a change to a chromosome state which is associated with a different level of prognosis;
- wherein the chromosomal interaction is represented by any probe in Table 8 or 9; and/or
- the chromosomal interaction is present in any region or gene listed in Table 8 or 9;

and wherein optionally:

- the chromosomal interaction has been identified by the method of determining which chromosomal interactions are relevant to a chromosome state as defined in claim 1, and/or
- the change in chromosomal interaction is monitored using (i) a probe that has at least 70% identity to any of the probe sequences mentioned in Table 5, and/or (ii) by a primer pair which has at least 70% identity to any primer pair in Table 5 or 8.

17. A process according to any one of claims 14 to 16 which comprises selecting a target based on detection of the chromosome interactions, and preferably screening for a modulator of the target to identify a therapeutic agent for immunotherapy, wherein said target is optionally a protein.

18. A process according to any one of the preceding claims wherein said prognosis is in a human or canine.

19. A process according to any one of the preceding claims, wherein the typing or detecting comprises specific detection of the ligated product by quantitative PCR (qPCR) which uses primers capable of amplifying the ligated product and a probe which binds the ligation site during the PCR reaction, wherein said probe comprises sequence which is complementary to sequence from each of the chromosome regions that have come together in the chromosome interaction, wherein preferably said probe comprises:

an oligonucleotide which specifically binds to said ligated product, and/or

a fluorophore covalently attached to the 5' end of the oligonucleotide, and/or

a quencher covalently attached to the 3' end of the oligonucleotide, and optionally

said fluorophore is selected from HEX, Texas Red and FAM; and/or

said probe comprises a nucleic acid sequence of length 10 to 40 nucleotide bases, preferably a length of 20 to 30 nucleotide bases.

20. A process according to any one of the preceding claims wherein:

- the result of the process is provided in a report, and/or

- the result of the process is used to select a patient treatment schedule, and preferably to select a specific therapy for the individual.

21. A process according any one of the preceding claims wherein:

- the subgroup relates to prostate cancer and at least one chromosome interaction from Table 25 is typed; and/or

- the subgroup relates to prostate cancer and at least one of the following combinations of interactions from Table 25 is typed:

(i) ETS1, MAP3K14, SLC22A3 and CASP2, or

(ii) BMP6, ERG, MSR1, MUC1, ACAT1 and DAPK1, or

(iii) HSD3B2, VEGFC, APAF1, MUC1, ACAT1 and DAPK1;

and/or

- the subgroup relates to DLBCL and at least one of the first 10 markers shown in Table 5 is typed, preferably corresponding to one or more of the following genes: STAT3, TNFRSF13B, ANXA11, MAP3K7, MEF2B and IFNAR1; and/or

5 - the subgroup relates to lymphoma and at least one of the first 11 markers shown in Figure 6 is typed, preferably corresponding to one or more of the following genes: STAT3, TNFRSF13B, ANXA11, MAP3K7, MEF2B and IFNAR1.

Figure 1

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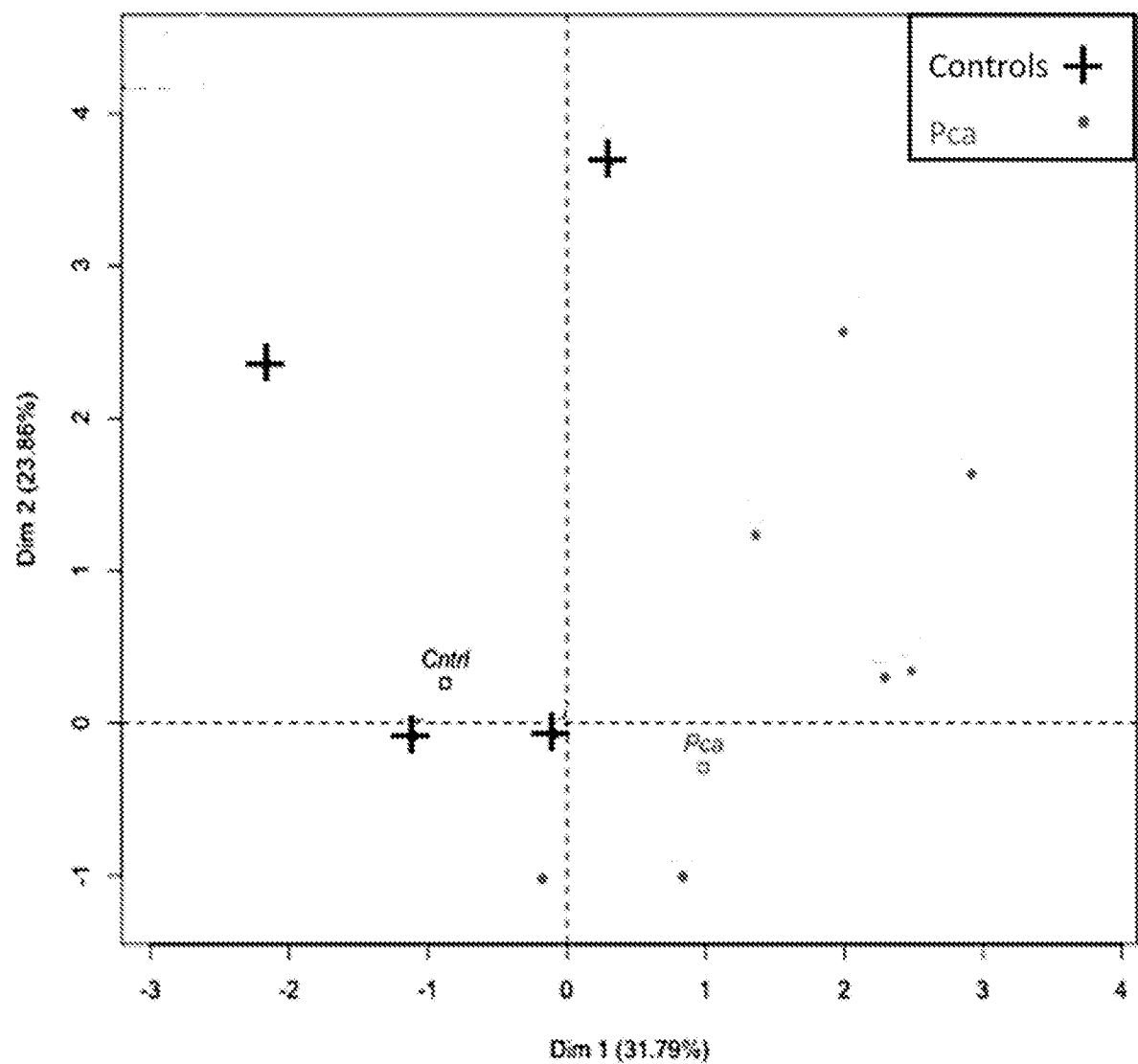


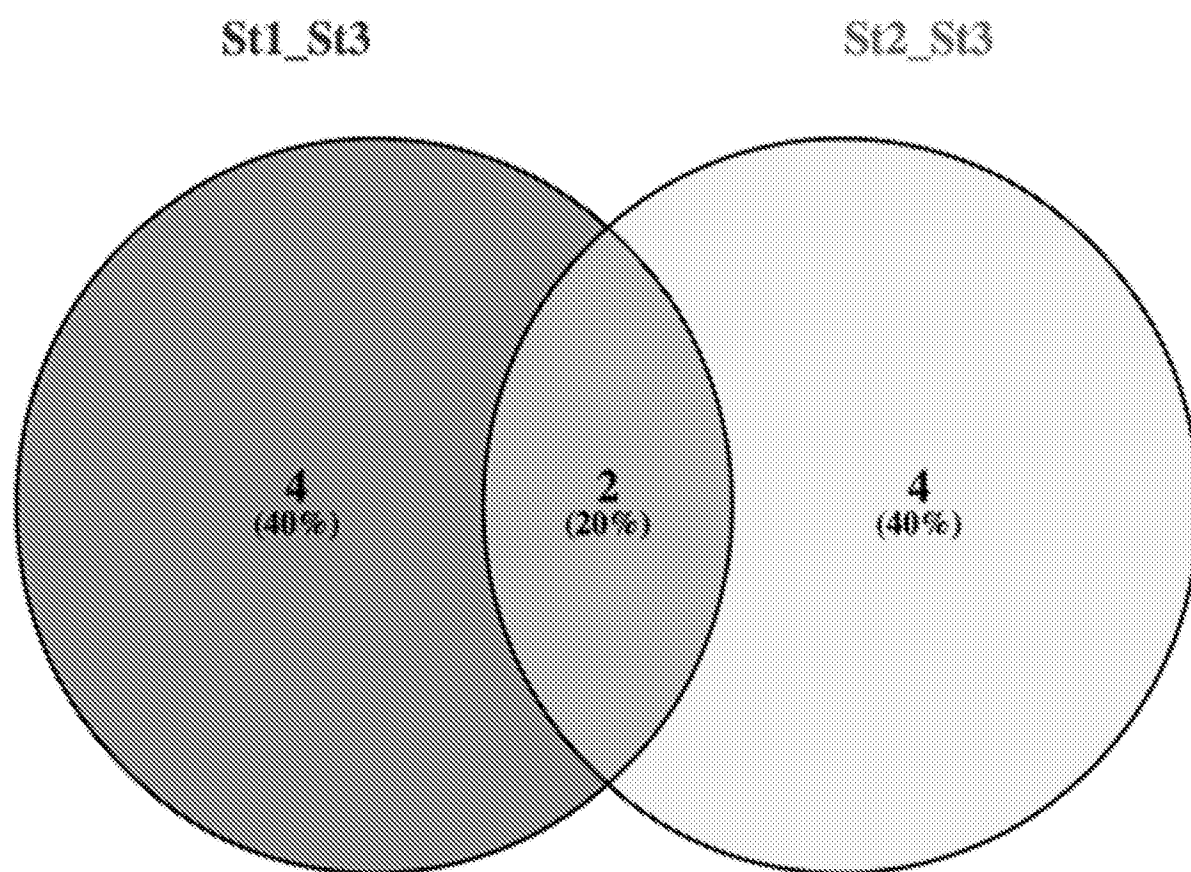
Figure 2

Figure 3

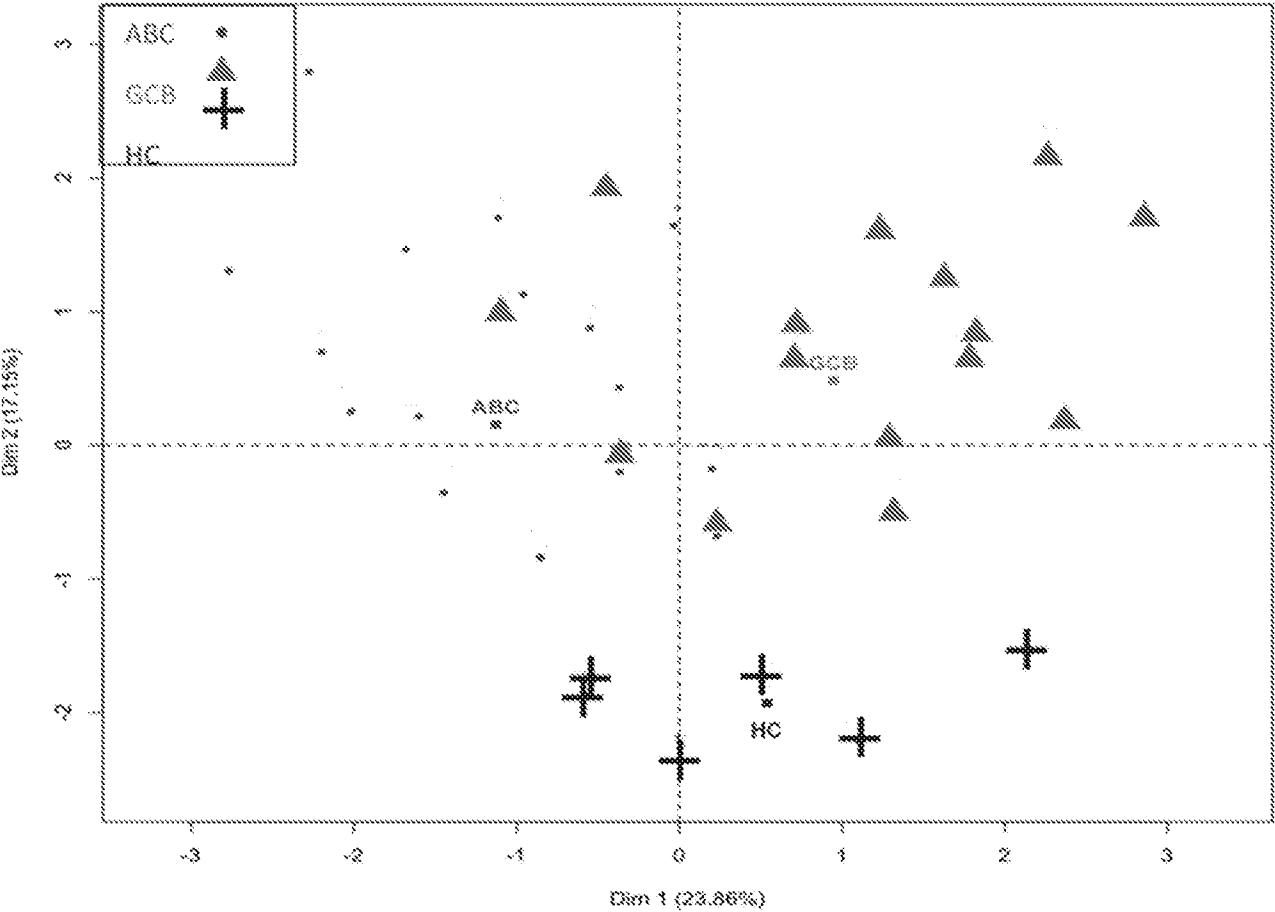


Figure 4.

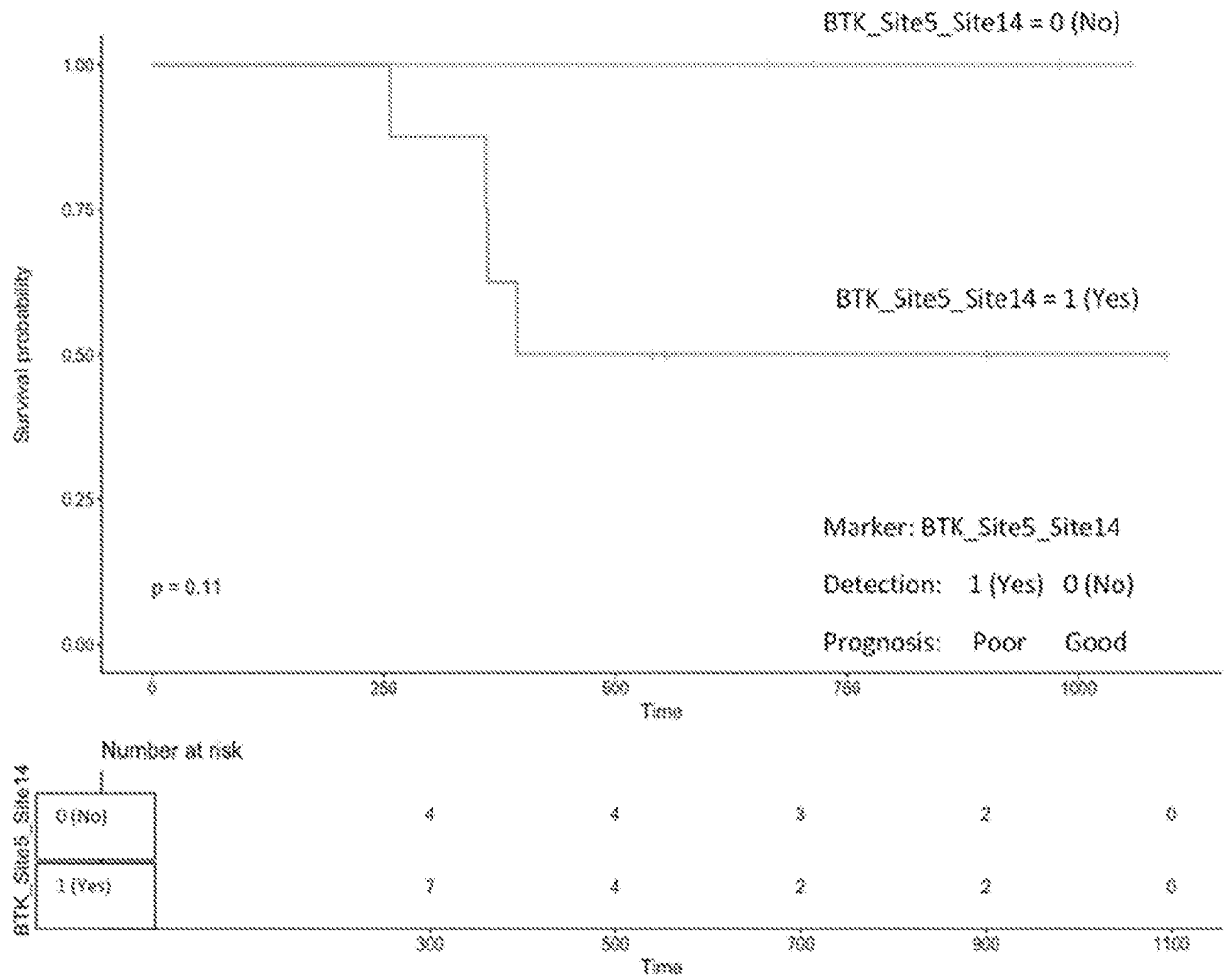


Figure 5

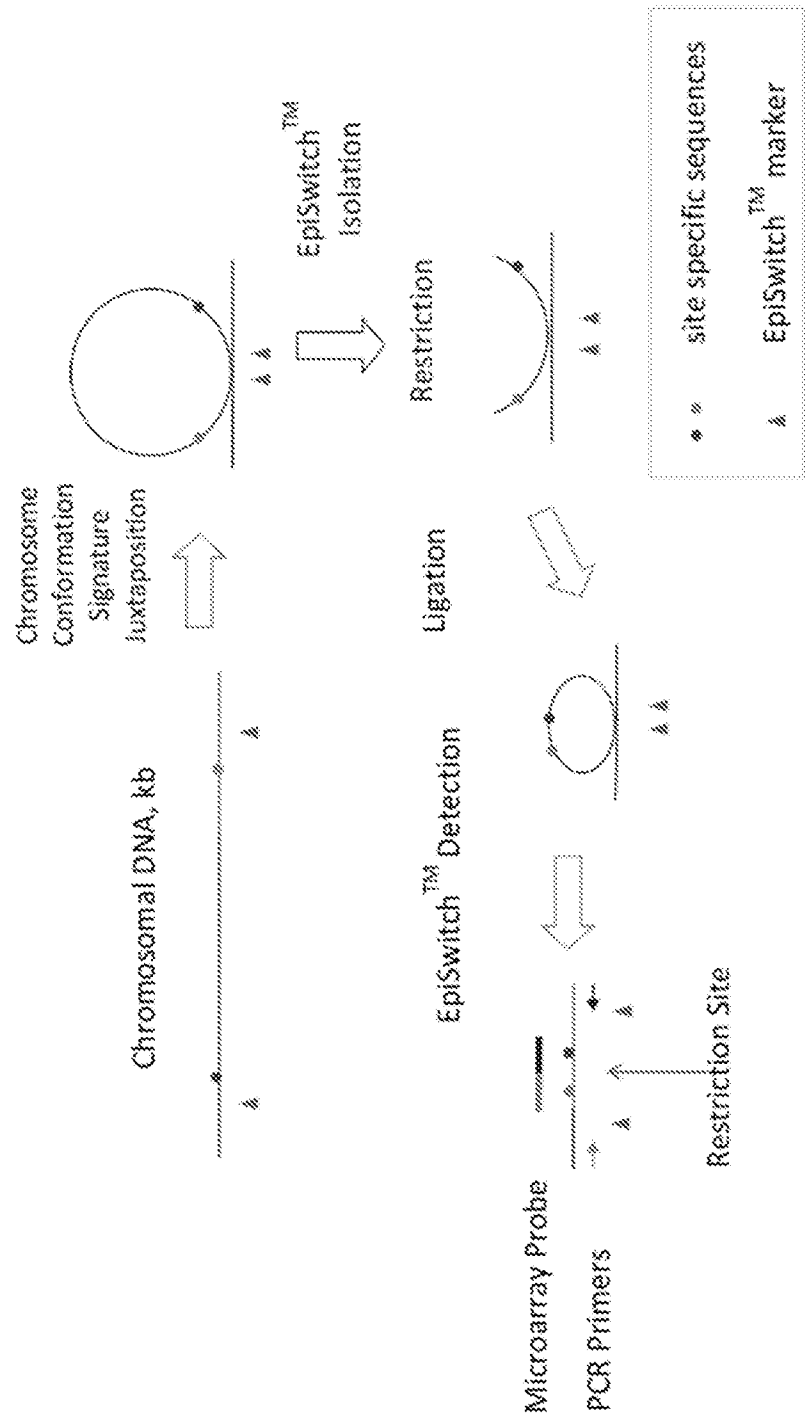


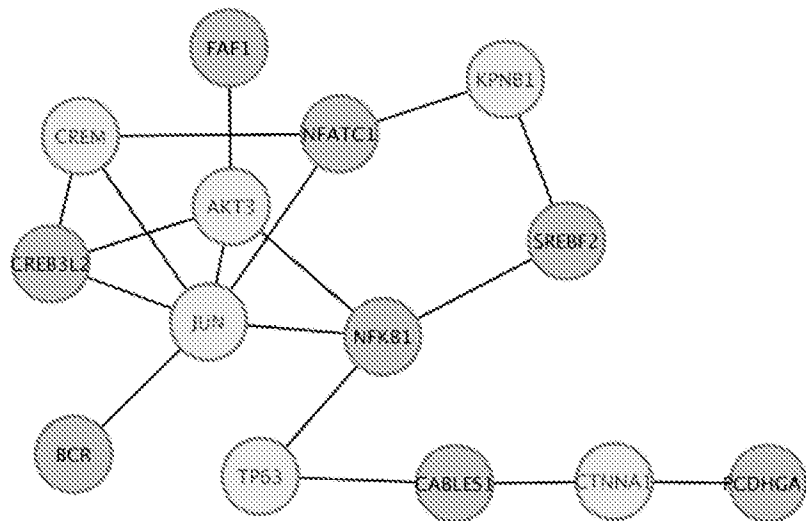
Figure 6.

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	A	B	C	D	E	F
1	marker	alpha_0.4	Freq	Rank_median	pValue_Median	Marker
2	OBD169_185.OBD169_187	0.249	498	2	0.006	OBD169_185.OBD169_187_1x
3	OBD169_169.OBD169_171	0.064	452	7	0.022	OBD169_169.OBD169_171_1x
4	OBD169_233.OBD169_235	0.052	357	9.25	0.035	OBD169_233.OBD169_235_1.4x
5	OBD169_265.OBD169_267	0.142	451	12	0.037	OBD169_265.OBD169_267_1.2x
6	OBD169_001.OBD169_003	0.15	429	14	0.037	OBD169_001.OBD169_003_1x
7	OBD169_105.OBD169_107	0.029	262	16.25	0.048	OBD169_105.OBD169_107_1.2x
8	OBD169_061.OBD169_063	0.128	375	16.5	0.048	OBD169_061.OBD169_063_1x
9	OBD169_153.OBD169_155	0	270	23.25	0.056	OBD169_153.OBD169_155_1.4x
10	OBD169_033.OBD169_035	0	278	14.75	0.076	OBD169_033.OBD169_035_1.2x
11	OBD169_133.OBD169_135	0.152	406	18	0.075	OBD169_133.OBD169_135_1x
12	OBD169_125.OBD169_127	0.006	300	22.75	0.09	OBD169_125.OBD169_127_1.2x
13	OBD169_165.OBD169_167	0	135	23.5	0.105	OBD169_165.OBD169_167_1.4x
14	OBD169_073.OBD169_075	0	115	28	0.105	OBD169_073.OBD169_075_1.2x
15	OBD169_293.OBD169_295	0	257	32.5	0.105	OBD169_293.OBD169_295_1x
16	OBD169_041.OBD169_043	0.123	300	22.25	0.112	OBD169_041.OBD169_043_1.2x
17	OBD169_021.OBD169_023	0	153	29.75	0.119	OBD169_021.OBD169_023_1x
18	OBD169_129.OBD169_131	0	130	33.5	0.149	OBD169_129.OBD169_131_1x
19	OBD169_229.OBD169_231	-0.042	225	97.75	0.516	OBD169_229.OBD169_231_1.2x
20	OBD169_277.OBD169_279	-0.097	225	94.5	0.521	OBD169_277.OBD169_279_1.4x
21	OBD169_049.OBD169_051	-0.051	262	107.5	0.608	OBD169_049.OBD169_051_1x
22	OBD169_065.OBD169_067	-6.18144E-06	112	168.5	1	OBD169_065.OBD169_067_1x
23	OBD169_009.OBD169_011	-0.055	156	171.75	1	OBD169_009.OBD169_011_1.2x

Figure 7.

Probe	Hchr	Hstart	Hstop	Gene
OBD169_033	chr18	79329076	79363050	ATP9B
OBD169_061	chr18	79162770	79279102	ATP9B
OBD169_233	chr22	23223523	23241206	BCR
OBD169_125	chr18	23194452	23232756	CABLES1
OBD169_185	chr7	137883915	137934211	CREB3L2
OBD169_169	chr1	50592143	50681003	FAF1
OBD169_153	chr9	20782946	20806283	FOCAD
OBD169_001	chr18	79485835	79516751	NFATC1
OBD169_265	chr4	102589993	102607062	NFKB1
OBD169_105	chr5	141432319	141485447	PCDHGA1
OBD169_133	chr22	41856337	41901029	SREBF2



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Figure 8.

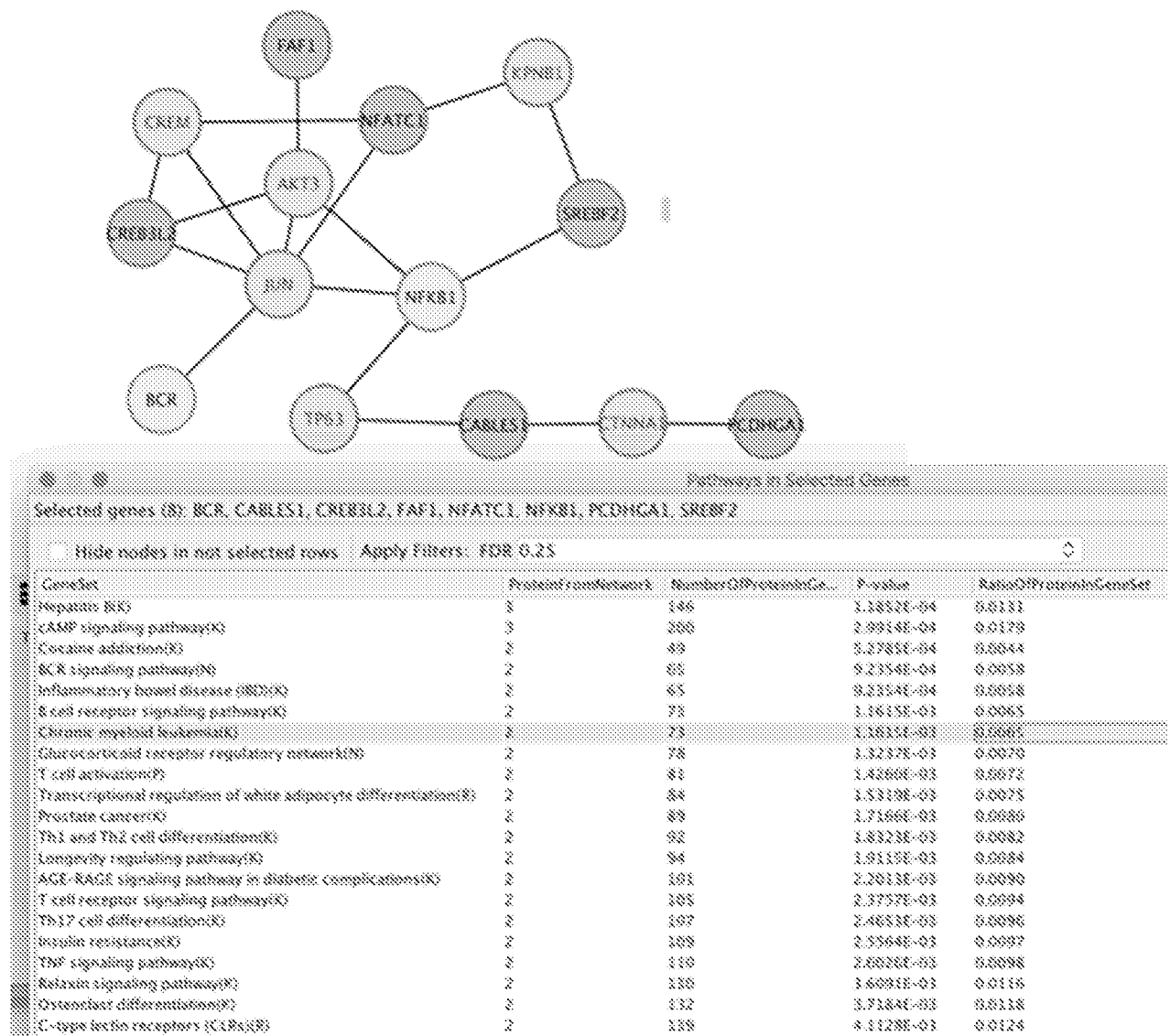


Figure 9

Confusion Matrix and Statistics		
Reference		
Prediction Ctrl Lymphoma		
Ctrl	13	1
Lymphoma	1	13
Accuracy : 0.9286		
95% CI : (0.765, 0.9912)		
No Information Rate : 0.5		
P-Value [Acc > NIR] : 1.516e-06		
Kappa : 0.8571		
McNemar's Test P-Value : 1		
Sensitivity : 0.9286		
Specificity : 0.9286		
Pos Pred Value : 0.9286		
Neg Pred Value : 0.9286		
Prevalence : 0.5000		
Detection Rate : 0.4643		
Detection Prevalence : 0.5000		
Balanced Accuracy : 0.9286		

Above shows the classification stats for training samples (28) using the 11 markers. Model was built using XGBoost in R

Confusion Matrix and Statistics		
Reference		
Prediction Ctrl Lymphoma		
Ctrl	4	1
Lymphoma	1	4
Accuracy : 0.8		
95% CI : (0.4439, 0.9749)		
No Information Rate : 0.5		
P-Value [Acc > NIR] : 0.05469		
Kappa : 0.6		
McNemar's Test P-Value : 1.00000		
Sensitivity : 0.8		
Specificity : 0.8		
Pos Pred Value : 0.8		
Neg Pred Value : 0.8		
Prevalence : 0.5		
Detection Rate : 0.4		
Detection Prevalence : 0.5		
Balanced Accuracy : 0.8		
'Positive' Class : Ctrl		

Above shows the classification stats for test samples (10) using the model built with the adjacent training samples

Figure 10

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Confusion Matrix and Statistics		
Reference		
Prediction	Ctrl	Lymphoma
Ctrl	12	2
Lymphoma	1	13
Accuracy : 0.8929		
95% CI : (0.7177, 0.9773)		
No Information Rate : 0.5357		
P-Value [Acc > NIR] : 6.278e-05		
Kappa : 0.7857		
McNemar's Test P-Value : 1		
Sensitivity : 0.9231		
Specificity : 0.8667		
Pos Pred Value : 0.8571		
Neg Pred Value : 0.9286		
Prevalence : 0.4643		
Detection Rate : 0.4286		
Detection Prevalence : 0.5000		
Balanced Accuracy : 0.8949		
'Positive' Class : Ctrl		

Above shows the classification stats for training samples (28) using the 11 markers. Model was built using XGBoost in R

Confusion Matrix and Statistics		
Reference		
Prediction	Ctrl	Lymphoma
Ctrl	5	0
Lymphoma	1	4
Accuracy : 0.9		
95% CI : (0.555, 0.9975)		
No Information Rate : 0.6		
P-Value [Acc > NIR] : 0.04636		
Kappa : 0.8		
McNemar's Test P-Value : 1.00000		
Sensitivity : 0.8333		
Specificity : 1.0000		
Pos Pred Value : 1.0000		
Neg Pred Value : 0.8000		
Prevalence : 0.6000		
Detection Rate : 0.5000		
Detection Prevalence : 0.5000		
Balanced Accuracy : 0.9167		
'Positive' Class : Ctrl		

Above shows the classification stats for test samples (10) using the model built with the adjacent training samples

Figure 11

Reference		
Prediction	Ctrl	Lymphoma
Ctrl	12	2
Lymphoma	1	13

Accuracy : 0.8929
 95% CI : (0.7177, 0.9773)
 No Information Rate : 0.5357
 P-Value [Acc > NIR] : 6.278e-05
 Kappa : 0.7857
 McNemar's Test P-Value : 1
 Sensitivity : 0.9231
 Specificity : 0.8667
 Pos Pred Value : 0.8571
 Neg Pred Value : 0.9286
 Prevalence : 0.4643
 Detection Rate : 0.4286
 Detection Prevalence : 0.5000
 Balanced Accuracy : 0.8949
 'Positive' Class : Ctrl
 Confusion Matrix and Statistics

Above shows the classification stats for training samples (28) using the 11 markers. Model was built using XGBoost in R

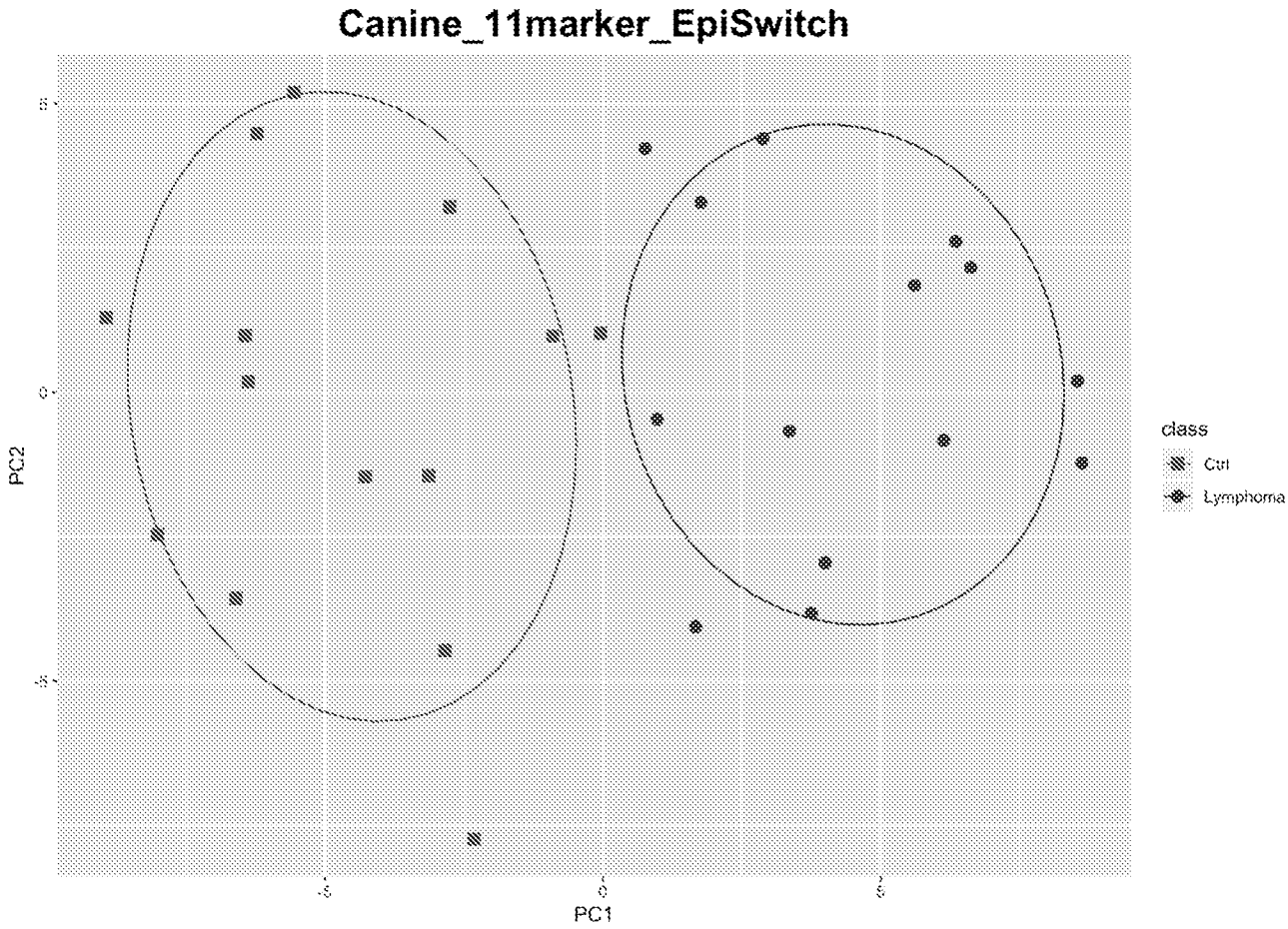
Reference		
Prediction	Ctrl	Lymphoma
Ctrl	3	2
Lymphoma	8	5

Accuracy : 0.8
 95% CI : (0.4439, 0.9748)
 No Information Rate : 0.7
 P-Value [Acc > NIR] : 0.3828
 Kappa : 0.6
 McNemar's Test P-Value : 0.4795
 Sensitivity : 1.0000
 Specificity : 0.7143
 Pos Pred Value : 0.6000
 Neg Pred Value : 1.0000
 Prevalence : 0.3000
 Detection Rate : 0.3000
 Detection Prevalence : 0.5000
 Balanced Accuracy : 0.8571
 'Positive' Class : Ctrl

Above shows the classification stats for test samples (10) using the model built with the adjacent training samples

Figure 12

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Figure 13

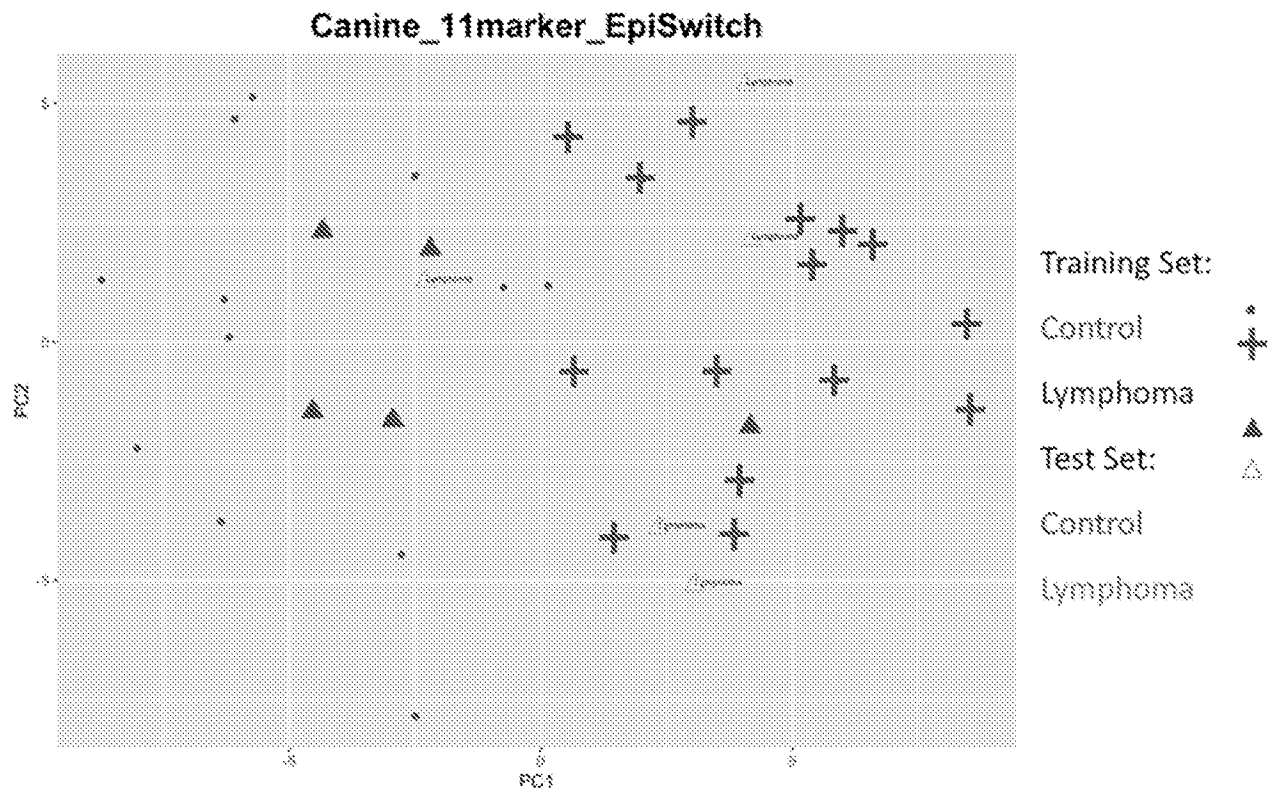
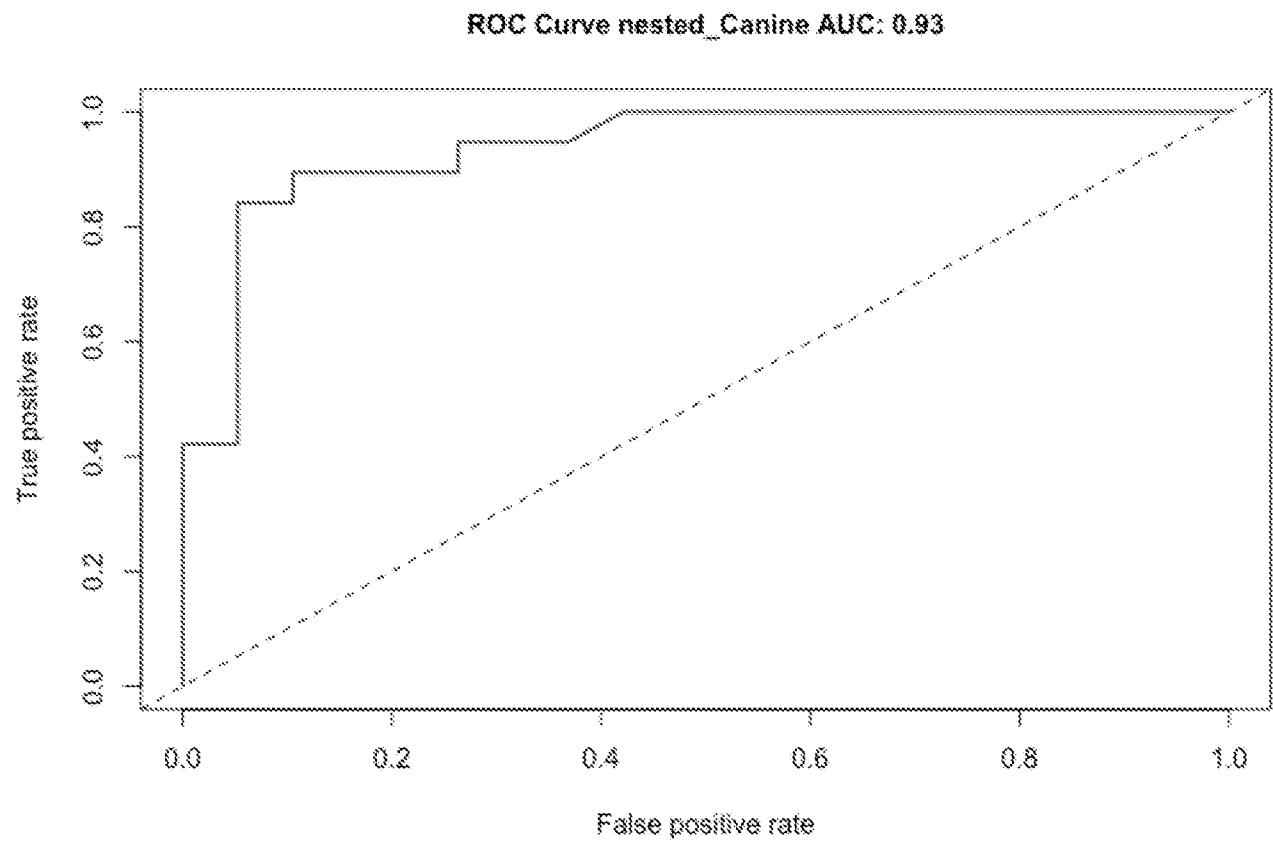
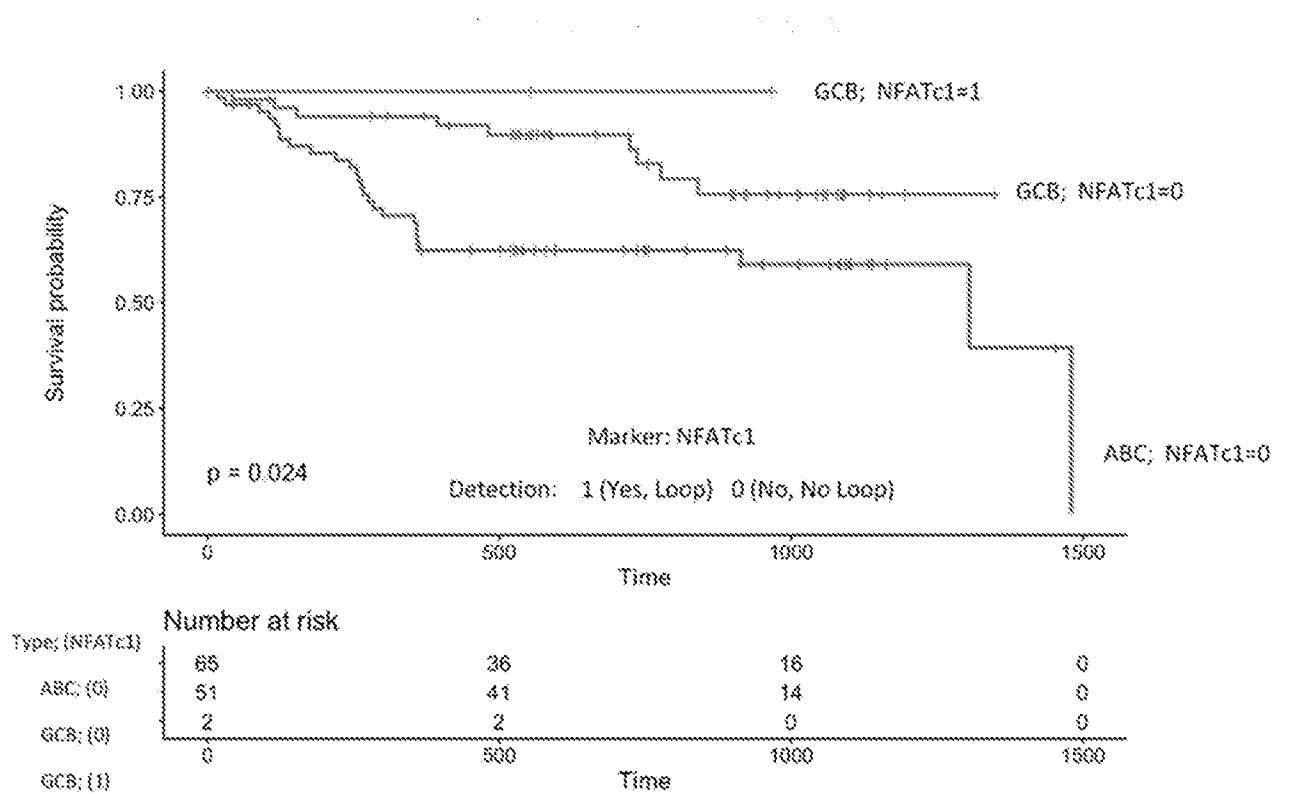


Figure 14



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Figure 15



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Figure 16

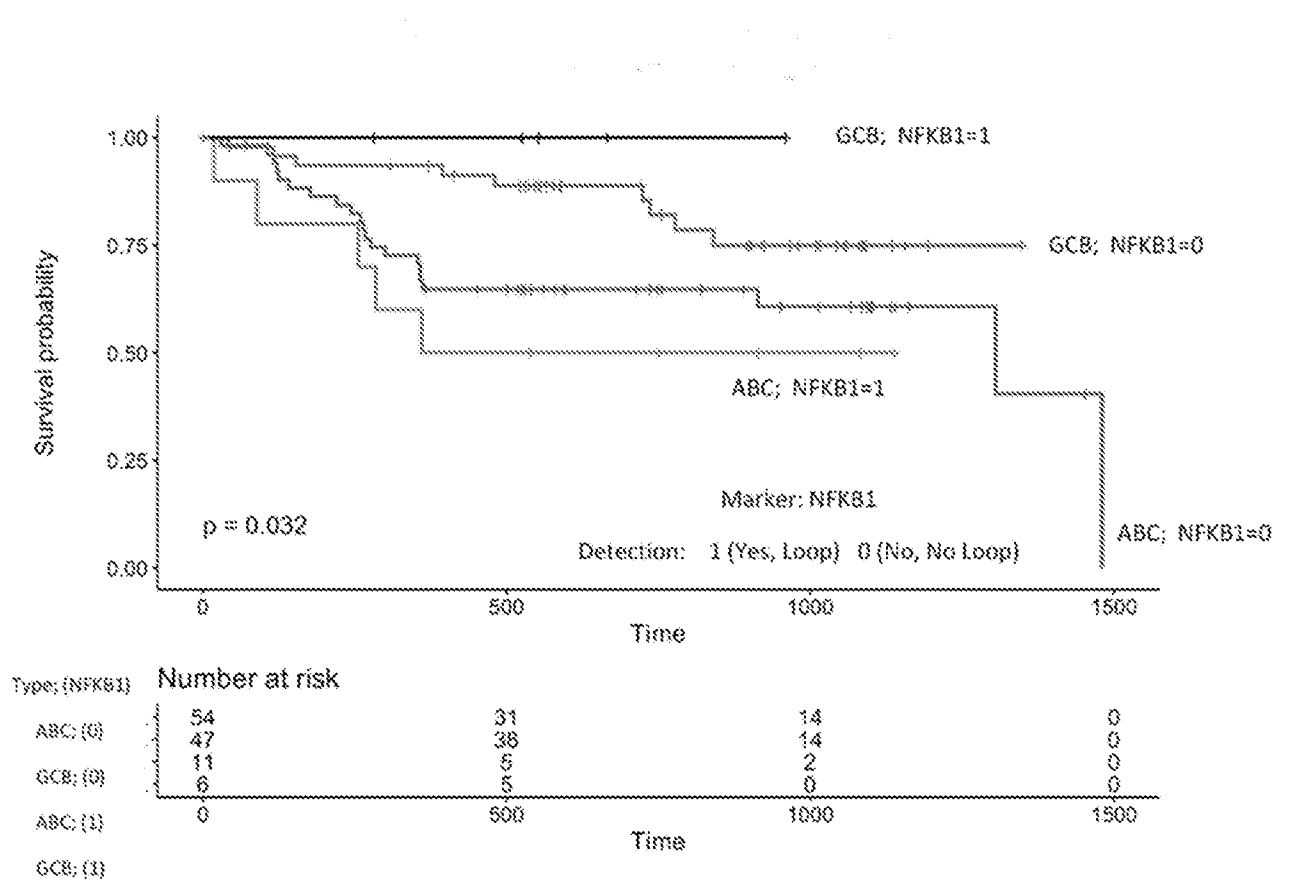
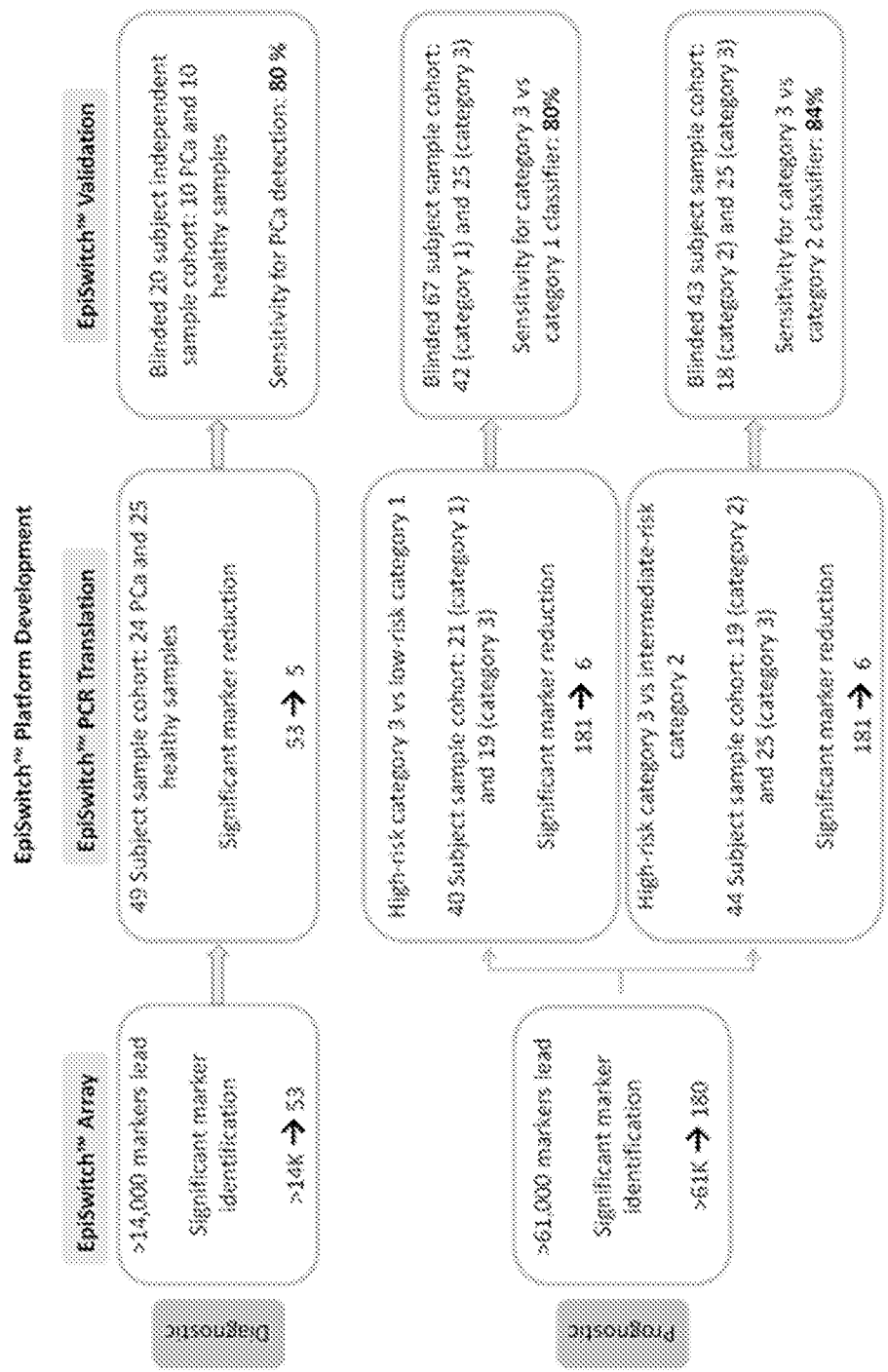


Figure 17

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Three-step approach to identify, evaluate, and validate diagnostic and prognostic biomarkers for prostate cancer (PCa).

Figure 18

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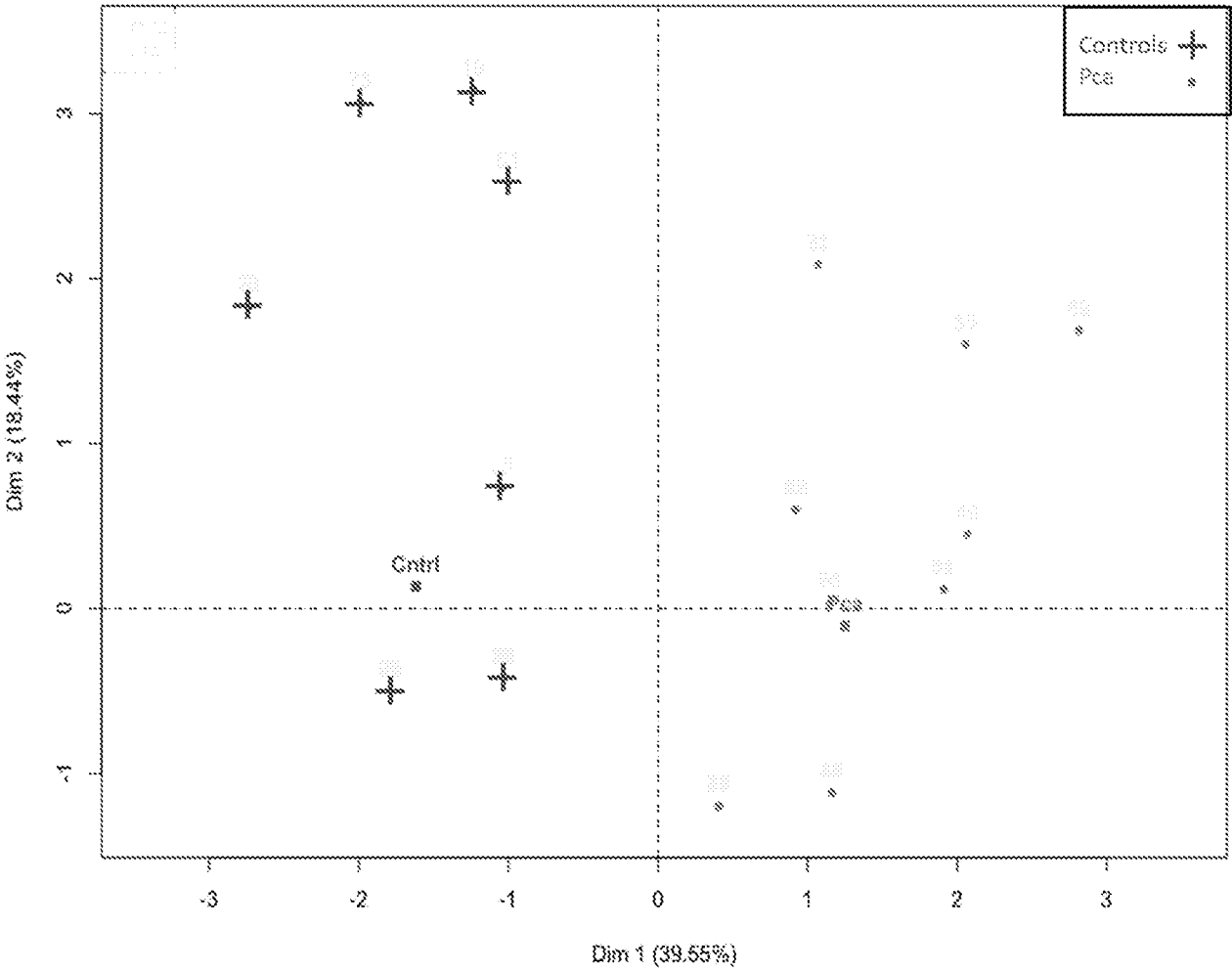
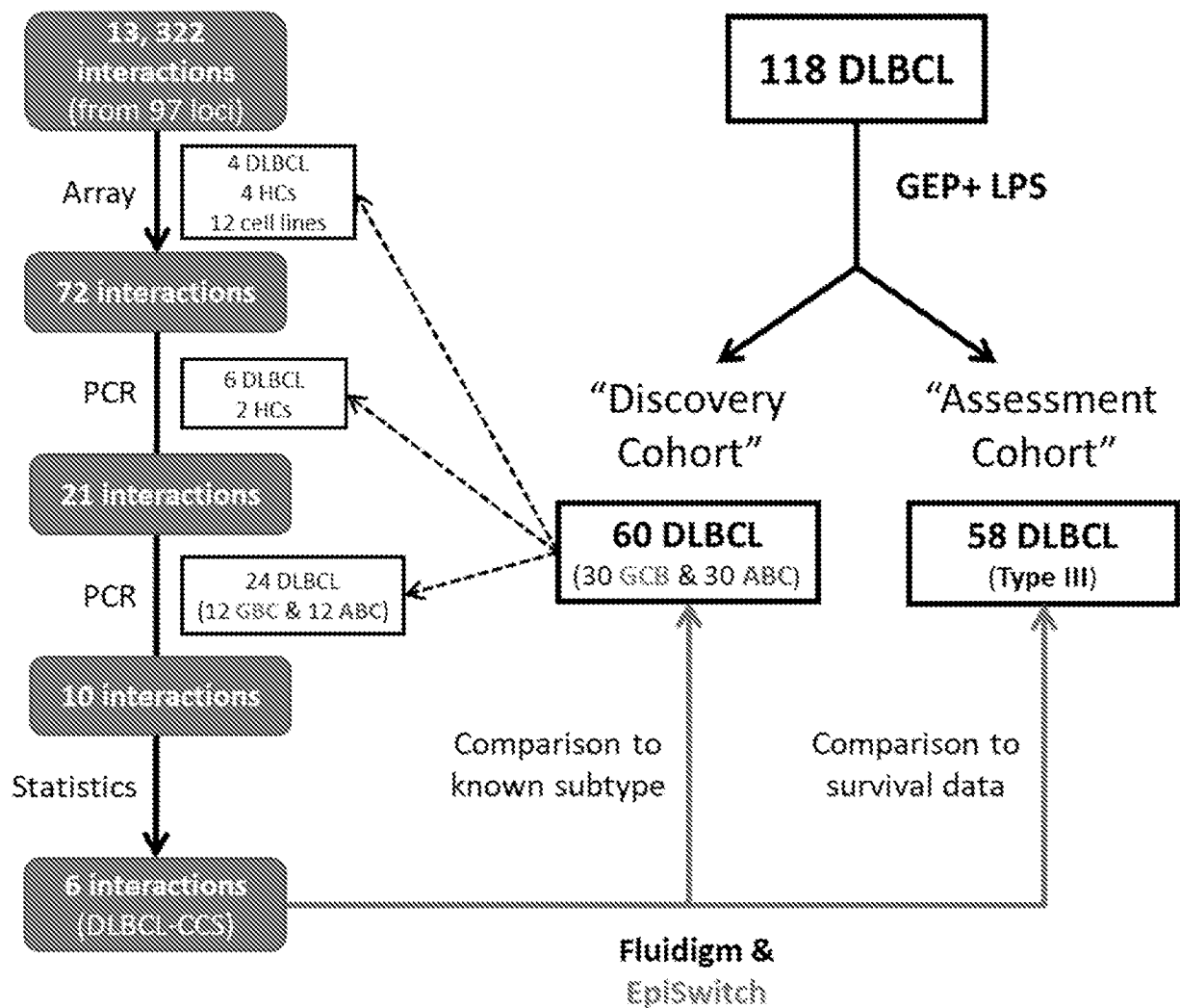


Figure 19



Workflow used to develop the DLBCL-CCS classifier. The DLBCL-CCS classifier was developed and assessed using a two-pronged, sequential approach. A total of 118 DLBCL samples were classified into disease subtypes (ABC, GCB or Type III) using gene expression profiling (GEP) and calculation of an LPS. Samples of known subtypes (30 samples on either end of the LPS spectrum) were used to identify changes in genomic architecture at six genomic loci that could best discriminate between ABC and GCB subtypes (the *EpiSwitch* DLBCL-CCS). 58 samples with intermediate LPS scores (Type III) were used to assess the performance of the *EpiSwitch* DLBCL-CCS and the Fluidigm assays.

Figure 20

Gene Locus	Subtype
STAT3	ABC
TNFRSF13B	ABC
ANXA11	ABC
MAP3K7	GCB
MEF2B	GCB
IFNAR1	GCB

The *EpiSwitch* DLBCL-CCS classifier. The DLBCL-CCS is made up of measurable changes in the genomic architecture at six genomic loci. These “gene loops” are either absent or present in different DLBCL subtypes, for example in the ABC subtype loops in STAT3, TNFRSF13N and ANXA11 are present while loops in MAP3K7, MEF2B and IFNAR1 are absent. The converse pattern is observed in GBC subtypes.

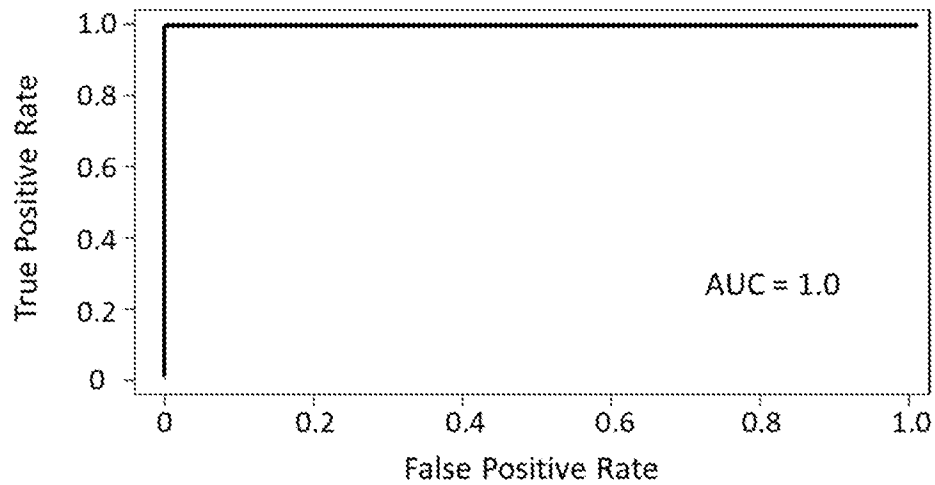
Figure 21

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A

Subtype Calls	Fluidigm	EpiSwitch	Concordance
ABC	30	30	1.0
GCB	30	30	1.0

B



C

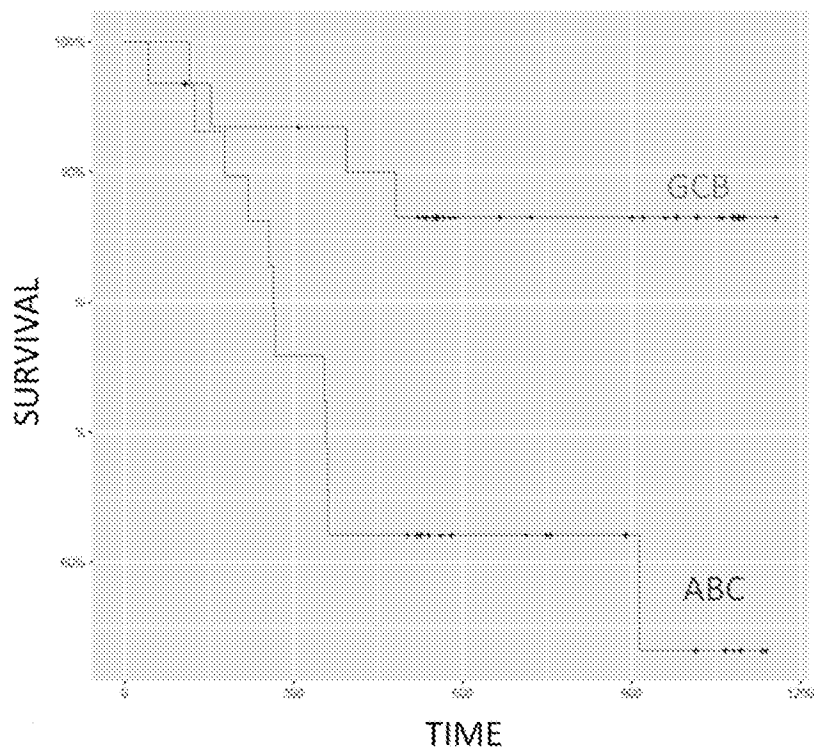


Figure 22

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A

Procedure	Called	ABC	GCB	Unclassified
Fluidigm	37	15	22	21
EpiSwitch	58	34	24	0

B

Patient ID	Fluidigm	EpiSwitch	Patient ID	Fluidigm	EpiSwitch
RG389344	ABC	ABC	RG770033	GCB	ABC
RG720922	ABC	ABC	RG260682	GCB	ABC
RG829223	ABC	ABC	RG289202	GCB	ABC
RG119158	ABC	ABC	RG626994	GCB	ABC
RG564834	ABC	ABC	RG750373	GCB	ABC
RG416179	ABC	ABC	RG789313	GCB	ABC
RG843356	ABC	ABC	RG679986	GCB	ABC
RG291088	ABC	ABC	RG728255	UNC	ABC
RG406330	GCB	GCB	RG191217	UNC	ABC
RG591532	GCB	GCB	RG242098	UNC	ABC
RG830523	GCB	GCB	RG438845	UNC	ABC
RG360351	GCB	GCB	RG235367	UNC	ABC
RG784664	GCB	GCB	RG442044	UNC	ABC
RG798672	GCB	GCB	RG431479	UNC	ABC
RG906378	GCB	GCB	RG907269	UNC	ABC
RG142143	ABC	GCB	RG162260	UNC	ABC
RG769375	ABC	GCB	RG704314	UNC	ABC
RG549340	ABC	GCB	RG832757	UNC	ABC
RG198295	ABC	GCB	RG665296	UNC	GCB
RG510145	ABC	GCB	RG427261	UNC	GCB
RG614926	ABC	GCB	RG256851	UNC	GCB
RG275043	ABC	GCB	RG431223	UNC	GCB
RG711234	(GCB)	ABC	RG762878	UNC	GCB
RG573806	GCB	ABC	RG651671	UNC	GCB
RG100774	GCB	ABC	RG388946	UNC	GCB
RG152922	GCB	ABC	RG875217	UNC	GCB
RG736755	GCB	ABC	RG123123	UNC	GCB
RG245982	GCB	ABC	RG313500	UNC	GCB
RG104142	GCB	ABC			
RG317110	GCB	ABC			

Assignment of DLBCL subtypes in Type III samples by *EpiSwitch* and *Fluidigm* assays. A. Summary of the DLBCL subtype calls made by *EpiSwitch* and *Fluidigm*. 21 of the samples were called as “unclassified” by the *Fluidigm* assay (UNC, grey). Of the 37 samples where a subtype call was made by both assays, 15 were called identically as ABC (red) or GCB (green). The remaining 22 samples were called differently by the two assays.

Figure 23

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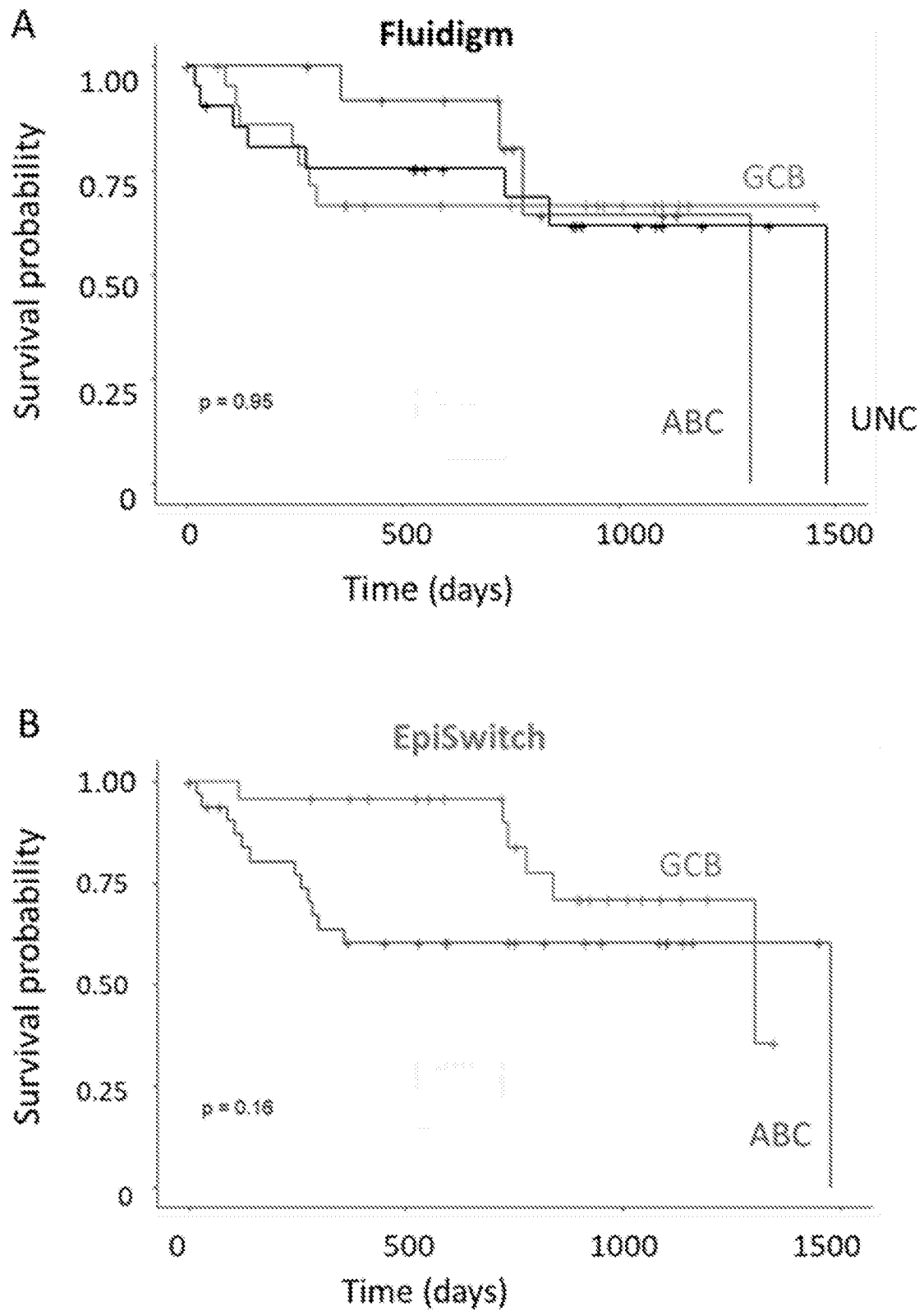
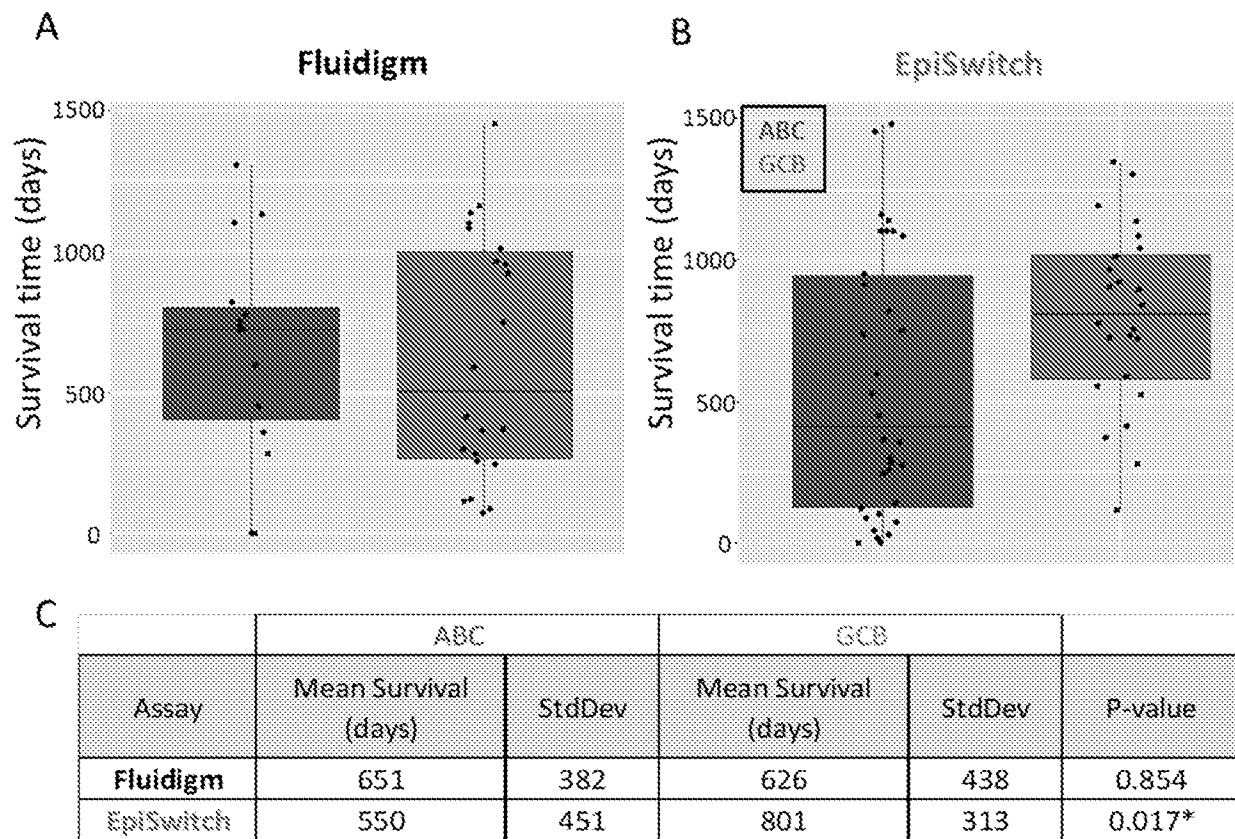


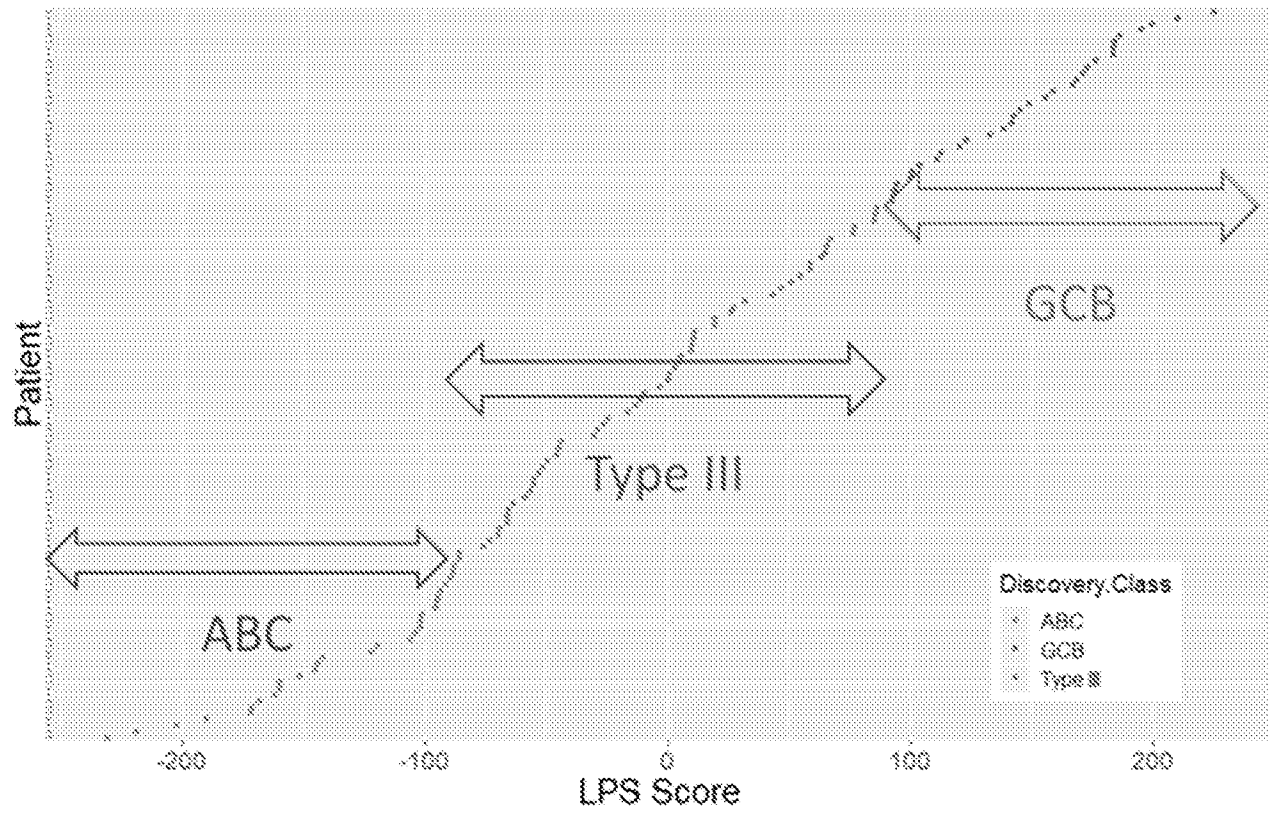
Figure 24

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Mean survival time by *EpiSwitch* and Fluidigm classification in the Validation cohort. Mean survival times of the patients classified as either ABC (red) or GCB (green) by the Fluidigm assay (A) or the *EpiSwitch* assay (green). There was a statistically significant difference in the mean survival time of the ABC (550 days) and GCB (801 days) samples called by the *EpiSwitch* classifier. StdDev = Standard deviation (days).

Figure 25



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Figure 26

