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(71) Demandeur/Applicant:
GLAXOSMITHKLINE BIOLOGICALS SA, BE
(72) Inventeurs/Inventors:
AMMENDOLA, VIRGINIA, IT;
COLLOCA, STEFANO, IT;
VITELLI, ALESSANDRA, IT;
WIZEL, BENJAMIN, US
(74) Agent: NORTON ROSE FULBRIGHT CANADA
LLP/S.E.N.C.R.L., S.R.L.

(54) Titre : CONSTRUCTIONS D'ADENOVIRUS DE CHIMPANZE AVEC DES ANTIGENES DE LYSSAVIRUS
(54) Title: CHIMPANZEE ADENOVIRUS CONSTRUCTS WITH LYSSAVIRUS ANTIGENS

(57) **Abrégé/Abstract:**

The invention provides adenoviral vectors comprising transgenes encoding Lyssaviral antigens. The vectors can be used to produce vaccines for the prophylaxis, amelioration and treatment of diseases caused by Lyssaviral diseases, e.g., rabies.

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(71) Applicant: GLAXOSMITHKLINE BIOLOGICALS

SA [BE/BE]; rue de l'Institut 89, B-1330 Rixensart (BE).

(72) Inventors: AMMENDOLA, Virginia; Via di Castel Ro-

mano 100, 00128 Rome (IT). COLLOCA, Stefano; Via

Salaria 73, 00198 Rome (IT). VITELLI, Alessandra; Via

di Castel Romano 100, 00128 Rome (IT). WIZEL, Ben-

jamin; 14200 Shady Grove Road, Rockville, Maryland

20850 (US).

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(54) Title: CHIMPANZEE ADENOVIRUS CONSTRUCTS WITH LYSSAVIRUS ANTIGENS

(57) Abstract: The invention provides adenoviral vectors comprising transgenes encoding Lyssaviral antigens. The vectors can be used to produce vaccines for the prophylaxis, amelioration and treatment of diseases caused by Lyssaviral diseases, e.g., rabies.

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CHIMPANZEE ADENOVIRUS CONSTRUCTS WITH LYSSAVIRUS ANTIGENS

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on December 6, 2017, is named VU66242WO_SL.txt and is 394,117 bytes in size.

FIELD OF THE INVENTION

This invention is in the field of ameliorating disease and treating and preventing viral infections. In particular, the present invention relates to chimpanzee adenoviral vectors encoding a *Lyssavirus* antigen. It includes the use of *Lyssavirus* antigens for ameliorating *Lyssaviral* diseases and treating and preventing rabies infections.

BACKGROUND

Lyssavirus is an enveloped, single stranded RNA virus in the *Rhabdoviridae* family. Members of the *Lyssavirus* genus cause rabies and have the highest fatality rate of all known human viral pathogens. Rabies is transmitted via the saliva of infected mammals. A neurotropic virus, it enters the nervous system of its host, causing an encephalomyelitis that is almost invariably fatal. Currently there are about 60,000 rabies deaths worldwide yearly, mostly caused by dog bites in developing countries in Asia and Africa and by wildlife and bats in North America.

Rabies presents either in a furious or a paralytic form. The incubation period varies between about five days and several years but is typically between about 20 and 90 days. Clinical illness most often starts with prodromal complaints of malaise, anorexia, fatigue, headache and fever followed by pain or parathesia at the site of exposure. Anxiety, agitation or irritability may be prominent during this period, followed by hyperactivity, disorientation, seizures, hydrophobia, hypersalivation and, eventually, paralysis, coma and death.

Adenovirus has been widely used for gene transfer applications due to its ability to achieve highly efficient gene transfer in a variety of target tissues and large transgene capacity. Conventionally, adenovirus E1 genes are deleted and replaced with a transgene cassette consisting of a promoter of choice, cDNA sequence of the gene of interest and a poly A signal, resulting in a replication defective recombinant virus.

Recombinant adenoviruses are useful in both gene therapy and as vaccines. Viral vectors based on non-human simian adenovirus represent an alternative to the use of human derived vectors for the development of genetic vaccines. Certain adenoviruses isolated from non-human simians are closely related to adenoviruses isolated from humans, as demonstrated by their efficient propagation in cells of human origin.

There is a demand for vectors that can effectively deliver vaccine antigens. Specifically, rabies remains an important viral zoonosis worldwide. While prophylaxis is currently available, high numbers of doses are required both pre and post exposure, and compliance is low, diminishing the medical benefit. There is a need for an improved rabies vaccine with a simplified dosing schedule, increased safety and an enhanced manufacturing profile. Adenovirus manufacturing is safer and less expensive than the existing human rabies vaccines, which are based on inactivated rabies virus. Accordingly, there is an unmet need to develop adenoviral vectors for use in a rabies vaccine.

SUMMARY OF THE INVENTION

The present inventors provide constructs useful as components of immunogenic compositions for the induction of an immune response in a subject against Lyssaviral diseases and rabies viral infection, methods for their use in treatment, and processes for their manufacture.

There is provided an isolated polynucleotide, wherein the polynucleotide encodes a polypeptide selected from the group consisting of:

- (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 - (b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1, and
 - (c) a polypeptide having the amino acid sequence according to SEQ ID NO: 3;
- wherein the isolated polynucleotide comprises a nucleic acid sequence encoding a *Lyssavirus* antigen.

Also provided is a recombinant polynucleotide comprising a polynucleotide selected from the group consisting of:

- (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,

(b) a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1, and
 5 (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3;
 wherein the recombinant polynucleotide comprises a nucleic acid sequence encoding a *Lyssavirus* antigen.

10 Also provided is a recombinant vector comprising a polynucleotide selected from the group consisting of:
 (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 (b) a polynucleotide which encodes a functional derivative of a polypeptide
 15 having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1, and
 (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3;
 20 wherein the recombinant vector comprises a nucleic acid sequence encoding a *Lyssavirus* antigen.

Also provided is a recombinant adenovirus comprising at least one polynucleotide or polypeptide selected from the group consisting of:
 25 (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 (b) a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical
 30 over its entire length to the amino acid sequence of SEQ ID NO: 1,
 (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
 (d) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 (e) a functional derivative of a polypeptide having the amino acid sequence
 35 according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1, and

(f) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; wherein the recombinant adenovirus comprises a nucleic acid sequence encoding a *Lyssavirus* antigen; and wherein the nucleic acid sequence is operatively linked to one or more sequences which direct expression of said *Lyssavirus* antigen in a host cell.

5

The present invention provides the recombinant adenovirus comprising one polynucleotide or polypeptide selected from the group consisting of:

- (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
- 10 (b) a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1,
- (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
- 15 (d) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
- (e) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid
- 20 sequence of SEQ ID NO: 1, and
- (f) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; wherein the adenovirus comprises a nucleic acid sequence encoding a *Lyssavirus* antigen, wherein the nucleic acid sequence is operatively linked to one or more sequences which direct expression of said *Lyssavirus* antigen in a host cell. The recombinant
- 25 adenovirus can comprise one or more further polynucleotide(s) or polypeptide(s) selected from the group of (a) to (f) listed above.

Also provided is a composition comprising at least one of the following:

- 30 (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
- (b) a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1,
- 35 (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
- (d) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,

(e) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1,

- 5 (f) a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
 (g) a vector comprising a polynucleotide as described in (a), (b) or (c) above, and
 (h) a recombinant adenovirus comprising a polynucleotide as described in (a), (b) or (c) above and a pharmaceutically acceptable excipient.

wherein the composition comprises a nucleic acid sequence encoding a *Lyssavirus* antigen or a *Lyssavirus* antigen polypeptide sequence; and, optionally, the nucleic acid sequence is operatively linked to one or more sequences which direct expression of said *Lyssavirus* antigen in a host cell.

Also provided is a cell comprising at least one of the following:

- 15 (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 (b) a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical
 20 over its entire length to the amino acid sequence of SEQ ID NO: 1,
 (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
 (d) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 (e) a functional derivative of a polypeptide having the amino acid sequence
 25 according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1,
 (f) a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
 (g) a vector comprising a polynucleotide as described in (a), (b) or (c) above, and
 30 (h) a recombinant adenovirus comprising a polynucleotide as described in (a), (b) or (c) above;

wherein the cell comprises an adenovirus comprising a nucleic acid sequence encoding a *Lyssavirus* antigen; and wherein the nucleic acid sequence is operatively linked to one or more sequences which direct expression of said *Lyssavirus* antigen in a host cell.

35

Also provided is an isolated adenoviral polypeptide selected from the group consisting of:

(a) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
(b) a functional derivative of a polypeptide having the amino acid sequence
according to SEQ ID NO: 1, wherein the functional derivative has an amino acid
sequence which is at least 80% identical over its entire length to the amino acid
5 sequence of SEQ ID NO: 1, and
(c) a polypeptide having the amino acid sequence according to SEQ ID NO: 3;
and
further comprising a *Lyssavirus* antigen polypeptide sequence.

10 Also provided is an isolated polynucleotide, vector, recombinant adenovirus, composition
or cell comprising the sequence according to SEQ ID NO: 6 and further comprising a
Lyssavirus antigen.

The recombinant adenoviruses and compositions may be used as medicaments, in
15 particular for the stimulation of an immune response against *Lyssaviral* diseases, such as
a rabies infection.

Typically, the aim of the methods of the invention is to induce a protective immune
response, *i.e.* to immunize or vaccinate the subject against a related pathogen. The
20 invention may therefore be applied for the prophylaxis, treatment or amelioration of
diseases due to infection by *Lyssaviral* diseases, such as infection by a rabies virus.

In some aspects, the compositions disclosed herein are immunogenic compositions that
when administered to a subject, induce a humoral and/or cellular immune response, *i.e.*,
25 an immune response which specifically recognizes a naturally occurring *Lyssaviral*
polypeptide. For example, an immunogenic composition may induce a memory T and/or
B cell population relative to an untreated subject following viral infection, particularly in
those embodiments where the composition comprises a nucleic acid comprising a
sequence which encodes a *Lyssaviral* antigen.

30 The invention may be provided for the purpose of both pre-exposure prophylaxis and
post-exposure prophylaxis to diseases caused by *Lyssaviral* diseases. In some
embodiments, the subject has previously been vaccinated with a rabies vaccine. The
approaches of the present invention may, for example, be utilised for a subject at least
35 one year after rabies vaccination, at least two years after rabies vaccination, at least at
least five years after rabies vaccination or at least ten years after rabies vaccination.

The *Lyssavirus* antigen is an antigenic sequence, *i.e.* a sequence from a *Lyssavirus* protein which comprises at least one B or T cell epitope. Suitably the *Lyssavirus* antigen comprises at least one T cell epitope. In an embodiment of the invention the adenovirus comprises a nucleic acid sequence encoding a *Lyssavirus* antigen. In a specific
 5 embodiment of the invention, the adenovirus comprises a nucleic acid encoding a polypeptide derived from SEQ ID NO: 37. In another specific embodiment of the invention, the adenovirus comprises a nucleic acid derived from SEQ ID NO: 38.

In another embodiment of the invention, the adenovirus may comprise a nucleic acid
 10 encoding a polypeptide derived from SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, or SEQ ID NO: 45. In a further specific embodiment of the invention, the adenovirus may comprise a nucleic acid derived from SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44 or SEQ ID NO: 46.

15 The elicited immune response may be an antigen specific B cell response, which produces neutralizing antibodies. The elicited immune response may be an antigen specific T cell response, which may be a systemic and/or a local response. The antigen specific T cell response may comprise a CD4+ T cell response, such as a response involving CD4+ T cells expressing a plurality of cytokines, *e.g.* IFNgamma, tumor
 20 necrosis factor-alpha (TNFalpha) and/or IL2. Alternatively, or additionally, the antigen specific T cell response comprises a CD8+ T cell response, such as a response involving CD8+ T cells expressing a plurality of cytokines, *e.g.*, IFNgamma, TNFalpha and/or IL2.

25 DESCRIPTION OF THE DRAWINGS

FIG. 1: Diagram of the rabies glycoprotein indicating the main antigenic epitopes.

FIG. 1 discloses SEQ ID NOS 94, 64 and 78, respectively, in order of appearance.

FIG. 2A-C: Alignment of fiber protein sequences from the indicated simian
 30 adenoviruses.

ChAd3 (SEQ ID NO:27)
 PanAd3 (SEQ ID NO:28)
 ChAd17 (SEQ ID NO:29)
 ChAd19 (SEQ ID NO:30)
 35 ChAd24 (SEQ ID NO:31)
 ChAd155 (SEQ ID NO:1)
 ChAd11 (SEQ ID NO:32)

ChAd20 (SEQ ID NO:33)

ChAd31 (SEQ ID NO:34)

PanAd1 (SEQ ID NO:35)

PanAd2 (SEQ ID NO:36)

- 5 FIG. 3: Flow diagram for the production of specific ChAd155 BAC and plasmid vectors
- FIG. 4: Species C BAC Shuttle #1365 schematic
- FIG. 5: pArsChAd155 Ad5E4orf6-2 (#1490) schematic
- FIG. 6: pChAd155/RSV schematic
- FIG. 7: BAC ChAd155/RSV schematic
- 10 FIG. 8: E4 Ad5E4orf6/ TetO hCMV RG WPRE (#1509) schematic
- FIG. 9: Productivity of ChAd3 and ChAd155 vectors expressing an HIV Gag transgene
- FIG. 10: Expression levels of ChAd155 and PanAd3 vectors expressing an RSV transgene – western blot
- FIG. 11: Immunogenicity of ChAd3 and ChAd155 vectors expressing an HIV Gag transgene – IFN-gamma ELISpot
- 15 FIG. 12: Immunogenicity of PanAd3 and ChAd155 vectors expressing an RSV transgene – IFN-gamma ELISpot
- FIG. 13: Immunogenicity of ChAd155 vector expressing a rabies G protein transgene in mice – (A) antibody neutralization and (B) IFN-gamma ELISpot
- 20 FIG. 14: Stability of the immune response induced by ChAd155 vector expressing a rabies G protein transgene
- FIG. 15: Potency of ChAd155 vector expressing a rabies G protein transgene compared to commercially available vaccines
- FIG. 16: Seroconversion and protection rates of ChAd155 vector expressing a rabies G protein transgene in mice
- 25 FIG. 17: Seroconversion and protection rates of ChAd155 vector expressing a rabies G protein transgene in mice
- FIG. 18: Neutralizing antibody response to ChAd155 vector expressing a rabies G protein transgene in non-human primates
- 30 FIG. 19: T cell response to ChAd155 vector expressing a rabies G protein transgene in non-human primates

DESCRIPTION OF THE SEQUENCES

- SEQ ID NO: 1 - Polypeptide sequence of ChAd155 fiber
- 35 SEQ ID NO: 2 - Polynucleotide sequence encoding ChAd155 fiber
- SEQ ID NO: 3 - Polypeptide sequence of ChAd155 penton
- SEQ ID NO: 4 - Polynucleotide sequence encoding ChAd155 penton
- SEQ ID NO: 5 - Polypeptide sequence of ChAd155 hexon
- SEQ ID NO: 6 - Polynucleotide sequence encoding ChAd155 hexon
- 40 SEQ ID NO: 7 - Polynucleotide sequence encoding ChAd155#1434
- SEQ ID NO: 8 - Polynucleotide sequence encoding ChAd155#1390
- SEQ ID NO: 9 - Polynucleotide sequence encoding ChAd155#1375

- SEQ ID NO: 10 - Polynucleotide sequence encoding wild type ChAd155
- SEQ ID NO: 11 - Polynucleotide sequence encoding ChAd155/RSV
- SEQ ID NO: 12 - Polynucleotide sequence encoding the CASI promoter
- SEQ ID NO: 13 - Ad5orf6 primer 1 polynucleotide sequence
- 5 SEQ ID NO: 14 - Ad5orf6 primer 2 polynucleotide sequence
- SEQ ID NO: 15 - BAC/CHAd155 Δ E1_TetO hCMV RpsL-Kana primer 1 polynucleotide sequence
- SEQ ID NO: 16 - BAC/CHAd155 Δ E1_TetO hCMV RpsL-Kana (#1375) primer 2 polynucleotide sequence
- 10 SEQ ID NO: 17 - 1021-FW E4 Del Step1 primer polynucleotide sequence
- SEQ ID NO: 18 - 1022-RW E4 Del Step1 primer polynucleotide sequence
- SEQ ID NO: 19 - 1025-FW E4 Del Step2 primer polynucleotide sequence
- SEQ ID NO: 20 - 1026-RW E4 Del Step2 primer polynucleotide sequence
- SEQ ID NO: 21 - 91-SubMonte FW primer polynucleotide sequence
- 15 SEQ ID NO: 22 - 90-BghPolyA RW primer polynucleotide sequence
- SEQ ID NO: 23 - CMVfor primer polynucleotide sequence
- SEQ ID NO: 24 - CMVrev primer polynucleotide sequence
- SEQ ID NO: 25 - CMVFAM-TAMRA qPCR probe polynucleotide sequence
- SEQ ID NO: 26 - Woodchuck Hepatitis Virus Posttranscriptional Regulatory Element (WPRE) polynucleotide sequence
- 20 SEQ ID NO: 27 - Amino acid sequence for the fiber protein of ChAd3
- SEQ ID NO: 28 - Amino acid sequence for the fiber protein of PanAd3
- SEQ ID NO: 29 - Amino acid sequence for the fiber protein of ChAd17
- SEQ ID NO: 30 - Amino acid sequence for the fiber protein of ChAd19
- 25 SEQ ID NO: 31 - Amino acid sequence for the fiber protein of ChAd24
- SEQ ID NO: 32 - Amino acid sequence for the fiber protein of ChAd11
- SEQ ID NO: 33 - Amino acid sequence for the fiber protein of ChAd20
- SEQ ID NO: 34 - Amino acid sequence for the fiber protein of ChAd31
- SEQ ID NO: 35 - Amino acid sequence for the fiber protein of PanAd1
- 30 SEQ ID NO: 36 - Amino acid sequence for the fiber protein of PanAd2
- SEQ ID NO: 37 - Amino acid sequence for the RG medoid antigen
- SEQ ID NO: 38 - Nucleotide sequence for the RG medoid antigen
- SEQ ID NO: 39 - Amino acid sequence for RG AA098
- SEQ ID NO: 40 - Nucleic acid sequence for RG AA098
- 35 SEQ ID NO: 41 - Amino acid sequence for RG AA0093
- SEQ ID NO: 42 - Nucleic acid sequence for RG AA0093
- SEQ ID NO: 43 - Amino acid sequence for RG AA0094

- SEQ ID NO: 44 - Nucleic acid sequence for RG AA0094
 SEQ ID NO: 45 - Amino acid sequence for RG AA0095
 SEQ ID NO: 46 - Nucleic acid sequence for RG AA0095
 SEQ ID NO: 47 - Amino acid sequence for AdC68rab.gp – ERA strain
 5 SEQ ID NO: 48 - Nucleotide sequence for AdC68rab.gp – ERA strain
 SEQ ID NO: 49 – Poly-histidine
 SEQ ID NOS: 50-63 – Rabies G protein Site IIb antigenic epitopes
 SEQ ID NOS: 64-77 – Rabies G protein Site I antigenic epitopes
 SEQ ID NOS: 78-91 – Rabies G protein Site III antigenic epitopes
 10 SEQ ID NO: 92 – pvjTetOhCMV_WPRE_BghPolyA forward primer
 SEQ ID NO: 93 – pvjTetOhCMV_WPRE_BghPolyA reverse primer

DETAILED DESCRIPTION OF THE INVENTION

Adenoviral vectors

- 15 Adenovirus has been widely used for gene transfer applications due to its proven safety, ability to achieve highly efficient gene transfer in a variety of target tissues and large transgene capacity. Adenoviral vectors of use in the present invention may be derived from a range of mammalian hosts. Over 100 distinct serotypes of adenovirus have been isolated which infect various mammalian species. These adenoviral serotypes have
 20 been categorized into six subgenera (A–F; B is subdivided into B1 and B2) according to sequence homology and on their ability to agglutinate red blood cells.

In one embodiment, the adenoviral vector of the present invention is derived from a nonhuman simian adenovirus, also referred to simply as a simian adenovirus.

- 25 Numerous adenoviruses have been isolated from nonhuman simians such as chimpanzees, bonobos, rhesus macaques and gorillas, and vectors derived from these adenoviruses induce strong immune responses to transgenes encoded by these vectors (Colloca *et al.* (2012) *Sci. Transl. Med.* 4:1-9; Roy *et al.* (2004) *Virology* 324: 361-372; Roy *et al.* (2010) *J. of Gene Med.* 13:17-25). Certain advantages of vectors based on
 30 nonhuman simian adenoviruses include the relative lack of cross-neutralizing antibodies to these adenoviruses in the target population, thus their use overcomes the pre-existing immunity to human adenoviruses. For example, cross-reaction of certain chimpanzee adenoviruses with pre-existing neutralizing antibody responses is only present in 2% of the target population compared with 35% in the case of certain candidate human
 35 adenovirus vectors.

Specifically, the adenoviral vector may be derived from a non-human adenovirus, such as a simian adenovirus and in particular a chimpanzee adenovirus such as ChAd3, ChAd63, ChAd83, ChAd155, Pan 5, Pan 6, Pan 7 (also referred to as C7) or Pan 9 and may include, in whole or in part, a nucleotide encoding the fiber, penton or hexon of a non-human adenovirus. Examples of such strains are described in WO03/000283, WO2010/086189 and GB1510357.5 and are also available from the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110-2209, and other sources. Alternatively, adenoviral vectors may be derived from nonhuman simian adenoviruses isolated from bonobos, such as PanAd1, PanAd2 or PanAd3. Examples of such vectors described herein can be found for example in WO2005/071093 and WO2010/086189. Adenoviral vectors may also be derived from adenoviruses isolated from gorillas as described in WO2013/52799, WO2013/52811 and WO2013/52832.

Adenoviral vector structure

Adenoviruses have a characteristic morphology with an icosahedral capsid comprising three major proteins, hexon (II), penton base (III) and a knobbed fiber (IV), along with a number of other minor proteins, VI, VIII, IX, IIIa and IVa2. The hexon accounts for the majority of the structural components of the capsid, which consists of 240 trimeric hexon capsomeres and 12 penton bases. The hexon has three conserved double barrels, while the top has three towers, each tower containing a loop from each subunit that forms most of the capsid. The base of the hexon is highly conserved between adenoviral serotypes, while the surface loops are variable. The penton is another adenoviral capsid protein that forms a pentameric base to which the fiber attaches. The trimeric fiber protein protrudes from the penton base at each of the 12 vertices of the capsid and is a knobbed rod-like structure. The primary role of the fiber protein is the tethering of the viral capsid to the cell surface via the interaction of the knob region with a cellular receptor, and variations in the flexible shaft as well as knob regions of fiber are characteristic of the different serotypes.

The adenoviral genome has been well characterized. The linear, double-stranded DNA is associated with the highly basic protein VII and a small peptide pX (also termed mu). Another protein, V, is packaged with this DNA-protein complex and provides a structural link to the capsid via protein VI. There is general conservation in the overall organization of the adenoviral genome with respect to specific open reading frames being similarly positioned, e.g. the location of the E1A, E1B, E2A, E2B, E3, E4, L1, L2, L3, L4 and L5 genes of each virus. Each extremity of the adenoviral genome comprises a sequence known as an inverted terminal repeat (ITR), which is necessary for viral replication. The

- 5' end of the adenoviral genome contains the 5' cis-elements necessary for packaging and replication; i.e., the 5' ITR sequences (which function as origins of replication) and the native 5' packaging enhancer domains (that contain sequences necessary for packaging linear adenoviral genomes and enhancer elements for the E1 promoter). The
- 5 3' end of the adenoviral genome includes the 3' cis-elements (including the ITRs) necessary for packaging and encapsidation. The virus also comprises a virus-encoded protease, which is necessary for processing some of the structural proteins required to produce infectious virions.
- 10 The structure of the adenoviral genome is described on the basis of the order in which the viral genes are expressed following host cell transduction. More specifically, the viral genes are referred to as early (E) or late (L) genes according to whether transcription occurs prior to or after onset of DNA replication. In the early phase of transduction, the E1A, E1B, E2A, E2B, E3 and E4 genes of adenovirus are expressed to prepare the host
- 15 cell for viral replication. During the late phase of infection, expression of the late genes L1-L5, which encode the structural components of the virus particles, is activated.

Adenovirus capsid proteins and their encoding polynucleotides

- As outlined above, the adenoviral capsid comprises three major proteins, hexon, penton
- 20 and fiber. The hexon accounts for the majority of the structural components of the capsid, which consists of 240 trimeric hexon capsomeres and 12 penton bases. The hexon has three conserved double barrels, while the top has three towers, each tower containing a loop from each subunit that forms most of the capsid. The base of hexon is highly conserved between adenoviral serotypes, while the surface loops are variable.
- 25 The penton is another adenoviral capsid protein that forms a pentameric base to which fiber attaches. The trimeric fiber protein protrudes from the penton base at each of the 12 vertices of the capsid and is a knobbed rod-like structure. A remarkable difference in the surface of adenovirus capsids compared to that of most other icosahedral viruses is
- 30 the presence of the long, thin fiber protein. The primary role of the fiber protein is the tethering of the viral capsid to the cell surface via its interaction with a cellular receptor.

- The fiber proteins of many adenovirus serotypes share a common architecture: an N-terminal tail, a central shaft made of repeating sequences, and a C-terminal globular
- 35 knob domain (or "head"). The central shaft domain consists of a variable number of beta-repeats. The beta-repeats connect to form an elongated structure of three intertwined spiralling strands that is highly rigid and stable. The shaft connects the N-

terminal tail with the globular knob structure, which is responsible for interaction with the target cellular receptor. The globular nature of the adenovirus knob domain presents large surfaces for binding the receptor laterally and apically. The effect of this architecture is to project the receptor-binding site far from the virus capsid, thus freeing the virus from steric constraints presented by the relatively flat capsid surface.

Although fibers of many adenovirus serotypes have the same overall architecture, they have variable amino acid sequences that influence their function as well as structure. For example, a number of exposed regions on the surface of the fiber knob present an easily adaptable receptor binding site. The globular shape of the fiber knob allows receptors to bind at the sides of the knob or on top of the fiber knob. These binding sites typically lie on surface-exposed loops connecting beta-strands that are poorly conserved among human adenoviruses. The exposed side chains on these loops give the knob a variety of surface features while preserving the tertiary and quaternary structure. For example, the electrostatic potential and charge distributions at the knob surfaces can vary due to the wide range of isoelectric points in the fiber knob sequences, varying from a pI of approximately 9 for adenovirus "Ad" 8, Ad 19, and Ad 37 to approximately 5 for subgroup B adenoviruses. As a structurally complex virus ligand, the fiber protein allows the presentation of a variety of binding surfaces (knob) in a number of orientations and distances (shaft) from the viral capsid.

One of the most obvious variations between some serotypes is fiber length. Studies have shown that the length of the fiber shaft strongly influences the interaction of the knob and the virus with its target receptors. Further, fiber proteins between serotypes can also vary in their ability to bend. Although beta-repeats in the shaft form a highly stable and regular structure, electron microscopy (EM) studies have shown distinct hinges in the fiber. Analysis of the protein sequence from several adenovirus serotype fibers pinpoints a disruption in the repeating sequences of the shaft at the third beta-repeat from the N-terminal tail, which correlates strongly with one of the hinges in the shaft, as seen by EM. The hinges in the fiber allow the knob to adopt a variety of orientations relative to the virus capsid, which may circumvent steric hindrances to receptor engagement requiring the correct presentation of the receptor binding site on the knob. For example, the rigid fibers of subgroup D Ads thus require a flexible receptor or one prepositioned for virus attachment, as they are unable to bend themselves.

The identification of specific cell receptors for different Ad serotypes and the knowledge of how they contribute to tissue tropism have been achieved through the use of fiber pseudotyping technology. Although Ads of some subgroups use *Coxsackievirus* and adenovirus receptor ("CAR") as a primary receptor, it is becoming clear that many Ads use alternate primary receptors, leading to vastly different tropism *in vitro* and *in vivo*. The fibers of these serotypes show clear differences in their primary and tertiary structures, such as fiber shaft rigidity, the length of the fiber shaft, and the lack of a CAR binding site and/or the putative HSPG binding motif, together with the differences in net charge within the fiber knob. Pseudotyping Ad 5 particles with an alternate fiber shaft and knob therefore provides an opportunity to remove important cell binding domains and, in addition, may allow more efficient (and potentially more cell-selective) transgene delivery to defined cell types compared to that achieved with Ad 5. Neutralization of fiber-pseudotyped Ad particles may also be reduced if the fibers used are from Ads with lower seroprevalence in humans or experimental models, a situation that favours successful administration of the vector. Furthermore, full length fiber as well as isolated fiber knob regions, but not hexon or penton alone, are capable of inducing dendritic cell maturation and are associated with induction of a potent CD8+ T cell response. Taken together, adenoviral fiber plays an important role in at least receptor-binding and immunogenicity of adenoviral vectors.

"Low seroprevalence" may mean having a reduced pre-existing neutralizing antibody level as compared to human adenovirus 5 (Ad5). Similarly or alternatively, "low seroprevalence" may mean less than about 20% seroprevalence, less than about 15% seroprevalence, less than about 10% seroprevalence, less than about 5% seroprevalence, less than about 4% seroprevalence, less than about 3% seroprevalence, less than about 2% seroprevalence, less than about 1% seroprevalence or no detectable seroprevalence. Seroprevalence can be measured as the percentage of individuals having a clinically relevant neutralizing titer (defined as a 50% neutralisation titer >200) using methods as described in Aste-Amézaga *et al.*, *Hum. Gene Ther.* (2004) 15(3):293-304.

Illustrating the differences between the fiber proteins of Group C simian adenoviruses is the alignment provided in FIG. 1. A striking feature is that the fiber sequences of these adenoviruses can be broadly grouped into having a long fiber, such as ChAd155, or a short fiber, such as ChAd3. This length differential is due to a 36 amino acid deletion at approximately position 321 in the short fiber relative to the long fiber. In addition, there are a number of amino acid substitutions that differ between the short versus long fiber

subgroup yet are consistent within each subgroup. While the exact function of these differences have not yet been elucidated, given the function and immunogenicity of fiber, they are likely to be significant. It has been shown that one of the determinants of viral tropism is the length of the fiber shaft. It has been demonstrated that an Ad5 vector with
 5 a shorter shaft has a lower efficiency of binding to CAR receptor and a lower infectivity. It has been speculated that this impairment is the result of an increased rigidity of the shorter fiber leading to a less efficient attachment to the cell receptor. These studies may explain the improved properties of ChAd155 carrying a longer and more flexible fiber in comparison with the previously described ChAd3 and PanAd3 carrying a fiber
 10 with a shorter shaft.

In one aspect of the invention there is provided isolated fiber, penton and hexon capsid polypeptides of chimp adenovirus ChAd155 and isolated polynucleotides encoding the fiber, penton and hexon capsid polypeptides of chimp adenovirus ChAd155. An
 15 "isolated" polynucleotide is one that is removed from its original environment. For example, a naturally-occurring polynucleotide is isolated if it is separated from some or all of the coexisting materials in the natural system. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of its natural environment or if it is comprised within cDNA.

20 All three capsid proteins are expected to contribute to low seroprevalence and can, thus, be used independently from each other or in combination to suppress the affinity of an adenovirus to pre-existing neutralizing antibodies, e.g. to manufacture a recombinant adenovirus with a reduced seroprevalence. Such a recombinant adenovirus may be a
 25 chimeric adenovirus with capsid proteins from different serotypes with at least a fiber protein from ChAd155.

Transgenes

Adenoviral vectors may be used to deliver desired RNA or protein sequences, for
 30 example heterologous sequences, for *in vivo* expression. A vector may include any genetic element including naked DNA, a phage, transposon, cosmid, episome, plasmid, or virus. Such vectors contain DNA of ChAd155 as disclosed herein and an expression cassette. By "expression cassette" (or "minigene") is meant the combination of a selected heterologous gene ("transgene") and the other regulatory elements necessary
 35 to drive translation, transcription and/or expression of the gene product in a host cell.

Typically, "heterologous" means derived from a genotypically distinct entity from that of the rest of the entity to which it is being compared. A heterologous nucleic acid sequence refers to any nucleic acid sequence that is not isolated from, derived from, or based upon a naturally occurring nucleic acid sequence of the adenoviral vector.

- 5 "Naturally occurring" means a sequence found in nature and not synthetically prepared or modified. A sequence is "derived" from a source when it is isolated from a source but modified (e.g., by deletion, substitution (mutation), insertion, or other modification), suitably so as not to disrupt the normal function of the source gene.
- 10 Typically, an adenoviral vector is designed such that the expression cassette is located in a nucleic acid molecule which contains other adenoviral sequences in the region native to a selected adenoviral gene. The expression cassette may be inserted into an existing gene region to disrupt the function of that region, if desired. Alternatively, the expression cassette may be inserted into the site of a partially or fully deleted adenoviral
- 15 gene. For example, the expression cassette may be located in the site of a mutation, insertion or deletion which renders non-functional at least one gene of a genomic region selected from the group consisting of E1A, E1B, E2A, E2B, E3 and E4. The term "renders non-functional" means that a sufficient amount of the gene region is removed or otherwise disrupted, so that the gene region is no longer capable of producing functional
- 20 products of gene expression. If desired, the entire gene region may be removed (and suitably replaced with the expression cassette). Suitably, E1 genes of adenovirus are deleted and replaced with an expression cassette consisting of a promoter of choice, a cDNA sequence of the gene of interest and a poly A signal, resulting in a replication defective recombinant virus.

- 25 The transgene encoded by the adenoviral vector is a sequence encoding a product which is useful in biology and medicine, such as one or more of a therapeutic or immunogenic protein or proteins, RNA or enzymes. Desirable RNA molecules include tRNA, dsRNA, ribosomal RNA, catalytic RNAs, RNA aptamers and antisense RNAs. An
- 30 example of a useful RNA sequence is a sequence which extinguishes expression of a targeted nucleic acid sequence in the treated animal.

- The transgene is a nucleic acid sequence, heterologous to the vector sequences flanking the transgene, which encodes a protein of interest. The nucleic acid coding sequence is
- 35 operatively linked to regulatory components in a manner which permits transgene transcription, translation, and/or expression in a host cell.

The transgene may encode a polypeptide or protein used for disease treatment, amelioration or prophylaxis, for the induction of an immune response, and/or for prophylactic vaccine purposes. As used herein, induction of an immune response refers to the ability of a protein, also known as an "antigen" or "immunogen," to induce a T cell and/or a humoral immune response to the protein.

Immunogens expressed by the inventive vectors which are useful to immunize a human or non-human animal against other pathogens include, e.g., bacteria, fungi, parasitic microorganisms or multicellular parasites which infect human and non-human vertebrates, or from a cancer cell or tumor cell. For example, immunogens may be selected from a variety of viral families. In an embodiment, the immunogen is from a *Lyssavirus*, for example Mokola virus, Duvenhage virus, European bat *Lyssavirus*, European bat *Lyssavirus* 2, and Australian bat *Lyssavirus*. In an embodiment the *Lyssavirus* immunogen is from a rabies virus, for example from the CVS11, CVS-N2C, Evelyn Rokitniki Abelseth (ERA), Flury, Pitman Moore or Wistar strains. Such antigens may be derived from the rabies viral glycoprotein (G), RNA polymerase (L), matrix protein (M), nucleoprotein (N) and phosphoprotein (P), such as comprising from the rabies viral glycoprotein (G), RNA polymerase (L), matrix protein (M), nucleoprotein (N) and phosphoprotein (P) or comprising a fragment thereof (suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400, at least 500 or at least 600 amino acids).

In an embodiment, the immunogens expressed by the vectors of the invention comprise all or a fragment, suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of the glycoprotein from Mokola virus, Duvenhage virus, European bat *Lyssavirus*, European bat *Lyssavirus* 2, and Australian bat *Lyssavirus*. In an embodiment, the immunogens expressed by the vectors of the invention comprise all or a fragment, suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of the glycoprotein from a rabies virus, for example from the CVS11, CVS-N2C, Evelyn Rokitniki Abelseth (ERA), Flury, Pitman Moore or Wistar strains. In an embodiment, the immunogens expressed by the vectors of the invention comprise all or a fragment, suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of SEQ ID NO: 37.

In an embodiment, the immunogens expressed by the vectors of the invention comprise one or more of the antigenic epitopes shown in Table 1. In an embodiment, the

immunogens expressed by the vectors of the invention comprise an epitope corresponding to Site I, Site IIa, Site IIb, Site III, Site IV and/or Site a of the rabies virus strain RABV, ABLV, ARAV, BBLV, DUVV, EBLV-1, EBLV-2, IRKV, KHUV, LBV, MOKV, SHIV, WCBV or IKOV. In a particular embodiment, a vector of the invention comprises
 5 an epitope corresponding to Site I, Site IIa, Site IIb, Site III, Site IV and/or Site a found in SEQ ID NO: 37.

In an embodiment, the cross-protective breadth of a vaccine construct can be increased by comprising a medoid sequence of an antigen. By "medoid" is meant a *Lyssavirus*
 10 sequence with a minimal dissimilarity to other *Lyssavirus* sequences. In a particular embodiment, a vector of the invention comprises a medoid sequence of the G glycoprotein. In a particular embodiment, a non-human primate vector of the invention comprises a medoid sequence of the G glycoprotein. In a particular embodiment, a ChAd155 vector of the invention comprises a medoid sequence of the G glycoprotein. In
 15 a particular embodiment, the medoid sequence is derived from a natural viral strain with the highest average percent of amino acid identity among all G protein sequences annotated in the NCBI database. In a particular embodiment, the medoid sequence of the G glycoprotein is NCBI strain AGN94271.

Alternatively or in addition, a transgene sequence may include a reporter sequence, which upon expression produces a detectable signal. Such reporter sequences include, without limitation, DNA sequences encoding β -lactamase, β -galactosidase (LacZ), alkaline phosphatase, thymidine kinase, green fluorescent protein (GFP), chloramphenicol acetyltransferase (CAT), luciferase, membrane bound proteins
 25 including, for example, CD2, CD4, CD8, the influenza hemagglutinin protein, and others well known in the art, to which high affinity antibodies directed thereto exist or can be produced by conventional means, and fusion proteins comprising a membrane bound protein appropriately fused to an antigen tag domain from, among others, hemagglutinin or Myc. These coding sequences, when associated with regulatory elements which
 30 drive their expression, provide signals detectable by conventional means, including enzymatic, radiographic, colorimetric, fluorescence or other spectrographic assays, fluorescent activating cell sorting assays and immunological assays, including enzyme linked immunosorbent assay (ELISA), radioimmunoassay (RIA) and immunohistochemistry.

35 In addition to the transgene, the expression cassette also may include conventional control elements which are operably linked to the transgene in a manner that permits its

transcription, translation and/or expression in a cell transfected with the adenoviral vector. As used herein, "operably linked" sequences include both expression control sequences that are contiguous with the gene of interest and expression control sequences that act in *trans* or at a distance to control the gene of interest.

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Expression control sequences include appropriate transcription initiation, termination, promoter and enhancer sequences; efficient RNA processing signals such as splicing and polyadenylation (poly A) signals including rabbit beta-globin polyA; sequences that stabilize cytoplasmic mRNA; sequences that enhance translation efficiency (e.g., Kozak consensus sequence); sequences that enhance protein stability; and when desired, sequences that enhance secretion of the encoded product. Among other sequences, chimeric introns may be used.

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A "promoter" is a nucleotide sequence that permits binding of RNA polymerase and directs the transcription of a gene. Typically, a promoter is located in the 5' non-coding region of a gene, proximal to the transcriptional start site of the gene. Sequence elements within promoters that function in the initiation of transcription are often characterized by consensus nucleotide sequences. Examples of promoters include, but are not limited to, promoters from bacteria, yeast, plants, viruses, and mammals (including humans). A great number of expression control sequences, including promoters which are internal, native, constitutive, inducible and/or tissue-specific, are known in the art and may be utilized.

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Examples of constitutive promoters include, without limitation, the TBG promoter, the retroviral Rous sarcoma virus LTR promoter (optionally with the enhancer), the cytomegalovirus (CMV) promoter (optionally with the CMV enhancer, see, e.g., Boshart et al, Cell, 41:521-530 (1985)), the CASI promoter (WO2012/115980), the SV40 promoter, the dihydrofolate reductase promoter, the β -actin promoter, the phosphoglycerol kinase (PGK) promoter, and the EF1a promoter (Invitrogen).

25

Inducible promoters allow regulation of gene expression and can be regulated by exogenously supplied compounds, environmental factors such as temperature, or the presence of a specific physiological state, e.g., acute phase, a particular differentiation state of the cell, or in replicating cells only. Inducible promoters and inducible systems are available from a variety of commercial sources, including, without limitation, Invitrogen, Clontech and Ariad. Many other systems have been described and can be readily selected by one of skill in the art. For example, inducible promoters include the

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zinc-inducible sheep metallothioneine (MT) promoter and the dexamethasone (Dex)-inducible mouse mammary tumor virus (MMTV) promoter. Other inducible systems include the T7 polymerase promoter system; the ecdysone insect promoter, the tetracycline-repressible system and the tetracycline-inducible system. Other systems include the FK506 dimer, VP16 or p65 using castradiol, diphenol murislerone, the RU486-inducible system and the rapamycin-inducible system. The effectiveness of some inducible promoters increases over time. In such cases one can enhance the effectiveness of such systems by inserting multiple repressors in tandem, *e.g.*, TetR linked to a TetR by an IRES.

In another embodiment, the native promoter for the transgene may be used. The native promoter may be preferred when it is desired that expression of the transgene should mimic the native expression. The native promoter may be used when expression of the transgene must be regulated temporally or developmentally, or in a tissue-specific manner, or in response to specific transcriptional stimuli. In a further embodiment, other native expression control elements, such as enhancer elements, polyadenylation sites or Kozak consensus sequences may also be used to mimic the native expression.

The transgene may be operably linked to a tissue-specific promoter. For instance, if expression in skeletal muscle is desired, a promoter active in muscle should be used. These include the promoters from genes encoding skeletal β -actin, myosin light chain 2A, dystrophin, muscle creatine kinase, as well as synthetic muscle promoters with activities higher than naturally occurring promoters. Examples of promoters that are tissue-specific are known for liver; hepatitis B virus core; alpha-fetoprotein, bone osteocalcin; bone sialoprotein, lymphocytes, immunoglobulin heavy chain; T cell receptor chain), neuronal such as neuron-specific enolase (NSE) promoter, neurofilament light-chain gene, and the neuron-specific vgf gene, among others.

In some embodiments, the Woodchuck Hepatitis Virus Posttranscriptional Regulatory Element (WPRE) (Zuffrey *et al.* (1999) J. Virol.; 73(4):2886-9) may be operably linked to the transgene.

The transgene may be used for treatment, *e.g.*, as a vaccine, for induction of an immune response, and/or for prophylactic vaccine purposes. As used herein, induction of an immune response refers to the ability of a protein to induce a T cell and/or a humoral immune response to the protein.

Adenoviral vector construction

Adenoviral vectors are generated by modifying wild type adenovirus to express heterologous genes and/or delete or inactivate undesirable adenoviral sequences.

Adenoviral vectors may also have altered replication competency. For example the
 5 vector may be replication defective or have limited replication such that it has a reduced ability to replicate in non-complementing cells, compared to the wild type virus. This may be brought about by mutating the virus e.g., by deleting a gene involved in replication, for example deleting the E1A, E1B, E3 or E4 gene.

10 The adenoviral vectors in accordance with the present invention may comprise a functional E1 deletion. Thus the adenoviral vectors according to the invention may be replication defective due to the absence of the ability to express adenoviral E1A and/or E1B. The recombinant adenoviruses may also bear functional deletions in other genes for example, deletions in E3 or E4 genes. The adenovirus delayed early gene E3 may
 15 be eliminated from the adenovirus sequence which forms part of the recombinant virus. The function of E3 is not necessary to the production of the recombinant adenovirus particle. Thus, it is unnecessary to replace the function of this gene product in order to package a recombinant adenovirus useful in the invention. In one particular embodiment the recombinant adenoviruses have functionally deleted E1 and E3 genes.
 20 The construction of such vectors is described in Roy *et al.*, Human Gene Therapy 15:519-530, 2004.

Recombinant adenoviruses may also be constructed having a functional deletion of the E4 gene. In a particular embodiment, the recombinant adenoviruses have functionally
 25 deleted E1 and E4 genes as described in Colloca *et al.* (2012) Sci. Transl. Med. 4:1-9; Roy *et al.* (2004) Virol.324: 361-372. In some embodiments, it may be desirable to retain the E4 ORF6 function. In one embodiment, the E4 ORF6 region may be replaced by a heterologous E4 ORF6, such as from human adenovirus 5 (Ad5). Thus, in one particular embodiment, the adenoviral vector may be functionally deleted in E1 and have
 30 the E4 ORF6 region from Ad5. Adenovirus vectors according to the invention may also contain a functional deletion in the delayed early gene E2a. Deletions may also be made in any of the late genes L1 through to L5 of the adenovirus genome. Similarly, deletions in the intermediate genes IX and IVa may be useful.

35 Other deletions may be made in the other structural or non-structural adenovirus genes. The above deletions may be used individually, e.g. an adenovirus sequence for use in the present invention may contain deletions of E1 only. Alternatively, deletions of entire

genes or portions thereof effective to destroy their biological activity may be used in any combination. For example in one exemplary vector, the adenovirus sequences may have deletions of the E1 genes and the E4 gene, or of the E1, E2a and E3 genes, or of the E1 and E3 genes (such as functional deletions in E1a and E1b, and a deletion of at least part of E3), or of the E1, E2a and E4 genes, with or without deletion of E3 and so on. Such deletions may be partial or full deletions of these genes and may be used in combination with other mutations, such as temperature sensitive mutations to achieve a desired result.

These vectors are generated using techniques known to those of skill in the art. Such techniques include conventional cDNA cloning techniques such as those described in texts, the use of overlapping oligonucleotide sequences of the adenovirus genomes, polymerase chain reaction, and any suitable method which provides the desired nucleotide sequence. Particularly suitable methods include standard homologous recombination methods such as those provided in Colloca *et al.* (2012) *Sci. Transl. Med.* 4:1-9; Roy *et al.* (2004) *Virology* 324: 361-372; Roy *et al.* (2010) *J. of Gene Med.* 13:17-25; and WO2010/085984 or recombineering methods as described in Warming *et al.* *Nuc. Acids Res.* (2005) 33:e36.

Adenoviral vector production

The adenoviral vectors can be produced in any suitable cell line in which the virus is capable of replication. In particular, complementing cell lines which provide the factors missing from the viral vector that result in its impaired replication characteristics (such as E1) can be used. Without limitation, such a cell line may be HeLa (ATCC Accession No. CCL 2), A549 (ATCC Accession No. CCL 185), HEK 293, KB (CCL 17), Detroit (e.g., Detroit 510, CCL 72) and WI-38 (CCL 75) cells, among others. These cell lines are all available from the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110-2209, USA. Other suitable parent cell lines may be obtained from other sources, such as PGK-E1 retinoblasts, e.g., PER.C6™ cells, as represented by the cells deposited under ECACC no. 96022940 at the European Collection of Animal Cell Cultures (ECACC) at the Centre for Applied Microbiology and Research (CAMR, UK) or Her 96 cells (CruCell).

In many circumstances, a cell line expressing the one or more missing genes which are essential to the replication and infectivity of the virus, such as human E1, can be used to transcomplement a chimp adenoviral vector. This is particularly advantageous because, due to the diversity between the chimp adenovirus sequences of the invention and the

human adenovirus sequences found in currently available packaging cells, the use of the current human E1-containing cells prevents the generation of replication-competent adenoviruses during the replication and production process.

5 Alternatively, if desired, one may utilize the sequences provided herein to generate a packaging cell or cell line that expresses, at a minimum, the E1 gene from ChAd155 under the transcriptional control of a promoter for expression in a selected parent cell line. Inducible or constitutive promoters may be employed for this purpose. Examples of such promoters are described in detail elsewhere in this document. A parent cell is
10 selected for the generation of a novel cell line expressing any desired ChAd155 gene. Without limitation, such a parent cell line may be HeLa [ATCC Accession No. CCL 2], A549 [ATCC Accession No. CCL 185], HEK 293, KB [CCL 17], Detroit [e.g., Detroit 510, CCL 72] and WI-38 [CCL 75] cells, among others. These cell lines are all available from the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia
15 20110-2209, USA.

Such E1-expressing cell lines are useful in the generation of recombinant adenovirus E1 deleted vectors. Additionally, or alternatively, cell lines that express one or more adenoviral gene products, e.g., E1A, E1B, E2A, E3 and/or E4, can be constructed using
20 essentially the same procedures as used in the generation of recombinant viral vectors. Such cell lines can be utilised to transcomplement adenovirus vectors deleted in the essential genes that encode those products, or to provide helper functions necessary for packaging of a helper-dependent virus (e.g., adeno-associated virus). The preparation of a host cell involves techniques such as the assembly of selected DNA sequences.

25 In an embodiment, the essential adenoviral gene products are provided in *trans* by the adenoviral vector and/or helper virus. In such an instance, a suitable host cell can be selected from any biological organism, including prokaryotic (e.g., bacterial) cells, and eukaryotic cells, including insect cells, yeast cells and mammalian cells.

30 Host cells may be selected from among any mammalian species, including, without limitation, cells such as A549, WEHI, 3T3, 10¹/2, HEK 293 cells or Per.C6 (the latter two of which express functional adenoviral E1), Saos, C2C12, L cells, HT1080, HepG2 and primary fibroblast, hepatocyte and myoblast cells derived from mammals including
35 human, monkey, mouse, rat, rabbit, and hamster.

A particularly suitable complementation cell line is the Procell92 cell line. The Procell92 cell line is based on HEK 293 cells which express adenoviral E1 genes, transfected with the Tet repressor under control of the human phosphoglycerate kinase-1 (PGK) promoter, and the G418-resistance gene (Vitelli *et al.* PLOS One (2013) 8(e55435):1-9).

- 5 Procell92.S is adapted for growth in suspension conditions and is also useful for producing adenoviral vectors expressing toxic proteins (www.okairos.com/e/innners.php?m=00084, last accessed 13 April 2015).

Adenoviral delivery methods and dosage

- 10 The adenoviral vectors may be as administered in immunogenic compositions. An immunogenic composition as described herein is a composition comprising one or more recombinant vectors capable of inducing an immune response, for example a humoral (e.g., antibody) and/or cell-mediated (e.g., a cytotoxic T cell) response, against a transgene product delivered by the vector following delivery to a mammal, suitably a
- 15 human. A recombinant adenovirus may comprise (suitably in any of its gene deletions) a gene encoding a desired immunogen and may therefore be used in a vaccine. The recombinant adenoviruses can be used as prophylactic or therapeutic vaccines against any pathogen for which the antigen(s) crucial for induction of an immune response, is able to limit the spread of the pathogen and for which cDNA is available.

- 20 Such vaccine or other immunogenic compositions may be formulated in a suitable delivery vehicle. The levels of immunity of the selected gene can be monitored to determine the need, if any, for boosters. Following an assessment of antibody titers in the serum, optional booster immunizations may be desired.

- 25 Optionally, a vaccine or immunogenic composition of the invention may be formulated to contain other components, including, e.g., adjuvants, stabilizers, pH adjusters, preservatives and the like. Examples of suitable adjuvants are provided below under "Adjuvants." Such an adjuvant can be administered with a priming DNA vaccine
- 30 encoding an antigen to enhance the antigen-specific immune response compared with the immune response generated upon priming with a DNA vaccine encoding the antigen only. Alternatively, such an adjuvant can be administered with a polypeptide antigen which is administered in an administration regimen involving the vectors of the invention.

- 35 The adenoviral vector may be prepared for administration by being suspended or dissolved in a pharmaceutically or physiologically acceptable carrier such as isotonic saline, isotonic salt, solution or other formulations that will be apparent to those skilled in

the art. The appropriate carrier will be evident to those skilled in the art and will depend in large part upon the route of administration. The compositions described herein may be administered to a mammal in a sustained release formulation using a biodegradable biocompatible polymer, or by on-site delivery using micelles, gels and liposomes.

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In some embodiments, the recombinant adenovirus of the invention is administered to a subject by intramuscular injection, intravenous injection, intraperitoneal injection, subcutaneous injection, epicutaneous administration, intradermal administration, transdermal administration, intravaginal administration nasal administration, rectal
10 administration or oral administration.

If the therapeutic regimen involves co-administration of one or more adenoviral vectors and a further component, each formulated in different compositions, they are favorably administered co-locally at or near the same site. For example, the components can
15 be administered (*e.g.* via an administration route selected from intramuscular, transdermal, intradermal, sub-cutaneous) to the same side or extremity ("co-lateral" administration) or to opposite sides or extremities ("contra-lateral" administration).

Dosages of the viral vector will depend primarily on factors such as the condition being
20 treated, the severity of the condition being treated and the age, weight and health of the patient, thus may vary among patients. For example, a therapeutically effective adult human dosage of the viral vector generally contains 1×10^5 to 1×10^{15} viral particles, such as from 1×10^8 to 1×10^{12} (*e.g.*, 1×10^8 , 5×10^8 , 1×10^9 , 5×10^9 , 1×10^{10} , 2.5×10^{10} , 5×10^{10} , 1×10^{11} , 5×10^{11} or 1×10^{12} particles). Alternatively, a viral vector can be administered at a
25 dose that is typically from 1×10^5 to 1×10^{10} plaque forming units (PFU), such as 1×10^5 PFU, 5×10^5 PFU, 1×10^6 PFU, 5×10^6 PFU, 1×10^7 PFU, 5×10^7 PFU, 1×10^8 PFU, 5×10^8 PFU, 1×10^9 PFU, 5×10^9 PFU, or 1×10^{10} PFU. Dosages will vary depending upon the size of the subject and the route of administration. For example, a suitable human dosage (for about an 80 kg subject) for intramuscular injection is in the range of about
30 1×10^5 to about 5×10^{12} particles per ml, for a single site. Optionally, multiple sites of administration may be used. In another example, a suitable human or veterinary dosage may be in the range of about 1×10^7 to about 1×10^{15} particles for an oral formulation.

The adenoviral vector can be quantified by Quantitative PCR Analysis (Q-PCR), for
35 example with primers and probes designed based on the CMV promoter region, using as the standard curve serial dilutions of plasmid DNA containing the vector genome with the expression cassette, including the human CMV (hCMV) promoter. The copy number in

the test sample is determined by the parallel line analysis method. Alternative methods for vector particle quantification include analytical HPLC or spectrophotometric methods based on A_{260} nm.

5 An immunologically effective amount of a nucleic acid may suitably be between 1 ng and 100 mg. For example, a suitable amount can be from 1 μ g to 100 mg. By “immunologically effective amount” is meant that the administration of that amount to a subject is effective for inducing a measurable immune response against *Lyssavirus* in the subject.

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An appropriate amount of the particular nucleic acid (e.g., vector) can readily be determined by those of skill in the art.

Exemplary effective amounts of a nucleic acid component can be between 1 ng and 100
15 μ g, such as between 1 ng and 1 μ g (e.g., 100 ng-1 μ g), or between 1 μ g and 100 μ g, such as 10 ng, 50 ng, 100 ng, 150 ng, 200 ng, 250 ng, 500 ng, 750 ng, or 1 μ g. Effective amounts of a nucleic acid can also include from 1 μ g to 500 μ g, such as between 1 μ g and 200 μ g, such as between 10 and 100 μ g, for example 1 μ g, 2 μ g, 5 μ g, 10 μ g, 20 μ g, 50 μ g, 75 μ g, 100 μ g, 150 μ g, or 200 μ g. Alternatively, an exemplary
20 effective amount of a nucleic acid can be between 100 μ g and 1 mg, such as from 100 μ g to 500 μ g, for example, 100 μ g, 150 μ g, 200 μ g, 250 μ g, 300 μ g, 400 μ g, 500 μ g, 600 μ g, 700 μ g, 800 μ g, 900 μ g or 1 mg.

Generally a human dose will be in a volume of between 0.1 ml and 2ml, such as 0.5 ml
25 and 2 ml. Thus the composition described herein can be formulated in a volume of, for example 0.1, 0.25, 0.5, 1.0, 1.5 or 2.0 ml human dose per individual or combined immunogenic components.

One of skill in the art may adjust these doses, depending on the route of administration
30 and the therapeutic or vaccine application for which the recombinant vector is employed. The levels of expression of the transgene, or for an adjuvant, the level of circulating antibody, can be monitored to determine the frequency of dosage administration.

If one or more priming and/or boosting steps are used, this step may include a single
35 dose that is administered hourly, daily, weekly or monthly, or yearly. As an example, mammals may receive one or two doses containing between about 10 μ g to about 50 μ g

of plasmid in carrier. The amount or site of delivery is desirably selected based upon the identity and condition of the mammal.

The therapeutic level of, or the level of immune response against, the protein encoded by the selected transgene can be monitored to determine the need, if any, for boosters. Following an assessment of CD8+ T cell response, or optionally, antibody titers, in the serum, optional booster immunizations may be desired. Optionally, the adenoviral vector may be delivered in a single administration or in various combination regimens, e.g., in combination with a regimen or course of treatment involving other active ingredients or in a prime-boost regimen.

Recombinant adenoviruses or compositions comprising polypeptide sequences

Suitably the polynucleotides of the invention are recombinant. Recombinant means that the polynucleotide is the product of at least one of cloning, restriction or ligation steps, or other procedures that result in a polynucleotide that is distinct from a polynucleotide found in nature. A recombinant adenovirus is an adenovirus comprising a recombinant polynucleotide. A recombinant vector is a vector comprising a recombinant polynucleotide. A "recombinant virus" includes progeny of the original recombinant virus. A "recombinant vector" includes replicates of the original recombinant vector. A "recombinant polynucleotide" includes replicates of the original recombinant polynucleotide.

A "functional derivative" of a polypeptide suitably refers to a modified version of a polypeptide, e.g. wherein one or more amino acids of the polypeptide may be deleted, inserted, modified and/or substituted. A derivative of an unmodified adenoviral capsid protein is considered functional if, for example:

- (a) an adenovirus comprising the derivative capsid protein within its capsid retains substantially the same or a lower seroprevalence compared to an adenovirus comprising the unmodified capsid protein and/or
- (b) an adenovirus comprising the derivative capsid protein within its capsid retains substantially the same or a higher host cell infectivity compared to an adenovirus comprising the unmodified capsid protein and/or
- (c) an adenovirus comprising the derivative capsid protein within its capsid retains substantially the same or a higher immunogenicity compared to an adenovirus comprising the unmodified capsid protein and/or

(d) an adenovirus comprising the derivative capsid protein within its capsid retains substantially the same or a higher level of transgene productivity compared to an adenovirus comprising the unmodified capsid protein.

- 5 Suitably the recombinant adenovirus or composition of the invention comprises a polypeptide having the amino acid sequence according to SEQ ID NO: 1. Suitably the recombinant adenovirus or composition of the invention comprises a polypeptide which is a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at
- 10 least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1. Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1 has an amino acid sequence which is at least 80% identical, such as at least 85.0% identical, such as at least 90% identical, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at
- 15 least 97.0% identical, such as at least 98.0% identical, such as at least 99.0% identical, such as at least 99.2% identical, such as at least 99.4% identical, such as 99.5% identical, such as at least 99.6% identical, such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 1. Alternatively the functional derivative has no more than 130, more suitably no more than
- 20 120, more suitably no more than 110, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more
- 25 suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 1.

Suitably the recombinant adenovirus or composition according to the invention further comprises:

- 30 (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; or
(b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3, wherein the functional derivative has an amino acid sequence which is at least 50.0% identical over its entire length to the amino acid sequence of SEQ ID NO: 3,
- 35 and/or
(a) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or

(b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

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Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3 has an amino acid sequence which is at least 60.0%, such as at least 70.0%, such as at least 80.0%, such as at least 85.0%, such as at least 90.0%, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at least 97.0% identical, such as at least 98.0% identical, such as at least 99.0%, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as 99.7% identical such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 3. Alternatively the functional derivative has no more than 300, more suitably no more than 250, more suitably no more than 200, more suitably no more than 150, more suitably no more than 125, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 3.

Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 60.0%, such as at least 70.0%, such as at least 80.0%, such as at least 85.0%, such as at least 90.0%, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at least 97.0% identical, such as at least 98.0% identical, such as at least 99.0%, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as 99.7% identical such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 5. Alternatively the functional derivative has no more than 500, more suitably no more than 400, more suitably no more than 450, more suitably no more than 300, more suitably no more than 250, more suitably no more than 200, more suitably no more than 150, more suitably no more than 125, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no

more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 5.

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Suitably the recombinant adenovirus or composition of the invention comprises a polypeptide having the amino acid sequence according to SEQ ID NO: 3.

Suitably the recombinant adenovirus or composition of the invention further comprises:

- 10 (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 1; or
 (b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1
- 15 and/or
- (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
 (b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 60% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.
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Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1 has an amino acid sequence which is at least 60.0% identical, such as at least 70.0% identical, such as at least 80.0% identical, such as at least 85.0% identical, such as at least 87.0% identical, such as at least 89.0% identical, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at least 97.0% identical, such as at least 98.0% identical, such as at least 99.0% identical, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

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Alternatively the functional derivative has no more than 130, more suitably no more than 120, more suitably no more than 110, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more

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suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 1.

Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 60.0%, such as at least 70.0%, such as at least 80.0%, such as at least 85.0%, such as at least 90.0%, such as at least 95.0%, such as at least 97.0%, such as at least 99.0%, such as at least 99.0%, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO:5. Alternatively the functional derivative has no more than 500, more suitably no more than 400, more suitably no more than 450, more suitably no more than 300, more suitably no more than 250, more suitably no more than 200, more suitably no more than 150, more suitably no more than 125, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 5.

Suitably the recombinant adenovirus or composition of the invention comprises a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1. Suitably the polynucleotide has a sequence according to SEQ ID NO: 2.

Alternatively, the recombinant adenovirus or composition of the invention comprises a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1. Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1 has an amino acid sequence which is at least 80% identical, such as at least 85.0% identical, such as at least 90% identical, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at least 97.0% identical, such as at least 98.0% identical, such as at least 99.0% identical, such as at least 99% identical, such as at least 99.4% identical, such as at least 99.6% identical or such as at least 99.8% identical

- over its entire length to the amino acid sequence of SEQ ID NO: 1. Alternatively the functional derivative has no more than 130, more suitably no more than 120, more suitably no more than 110, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 1.
- Suitably the recombinant adenovirus or composition of the invention further comprises a polynucleotide encoding:
- (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; or
 - (b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3, wherein the functional derivative has an amino acid sequence which is at least 50.0% identical over its entire length to the amino acid sequence of SEQ ID NO: 3, and/or
 - (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
 - (b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.
- Suitably the functional derivative of the polypeptide having the amino acid sequence according to SEQ ID NO: 3 has an amino acid sequence which is at least 60.0%, such as at least 70.0%, such as at least 80.0%, such as at least 85.0%, such as at least 90.0%, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at least 97.0% identical, such as at least 98.0% identical, such as at least 99.0%, such as at least 99%, such as at least 99.4%, such as at least 99.6%, such as at least 99.8% identical over its entire length to the amino acid sequence of SEQ ID NO: 3. Alternatively the functional derivative has no more than 300, more suitably no more than 250, more suitably no more than 200, more suitably no more than 150, more suitably no more than 125, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no

more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 3.

- 5 Suitably the functional derivative of the polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 60.0%, such as at least 70.0%, such as at least 80.0%, such as at least 85.0%, such as at least 90.0%, such as at least 95.0%, such as at least 97.0%, such as at least 98.0%, such as at least 99.0%, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as 99.7% identical such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 5. Alternatively the functional derivative has no more than 500, more suitably no more than 400, more suitably no more than 450, more suitably no more than 300, more suitably no more than 250, more suitably no more than 200, more suitably no more than 150, more suitably no more than 125, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 5.

- Suitably the recombinant adenovirus or composition of the invention comprises a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3. Suitably the polynucleotide has a sequence according to SEQ ID NO: 4.

Suitably the recombinant adenovirus or composition of the invention further comprises a polynucleotide encoding:

- 30 (a) a polypeptide having the amino acid sequence according to SEQ ID NO: 1; or
(b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 1
35 and/or
(a) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or

(b) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

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Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1 has an amino acid sequence which is at least 60.0% identical, such as at least 70.0% identical, such as at least 80.0% identical, such as at least 85.0% identical, such as at least 87.0% identical, such as at least 89.0% identical, such as at least 91.0% identical, such as at least 93.0% identical, such as at least 95.0% identical, such as at least 97.0% identical, such as at least 98.0% identical, such as at least 99.0%, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as 99.7% identical such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

Alternatively the functional derivative has no more than 130, more suitably no more than 120, more suitably no more than 110, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than 10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 1.

Suitably the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 60.0%, such as at least 70.0%, such as at least 80.0%, such as at least 85.0%, such as at least 90.0%, such as at least 95.0%, such as at least 97.0%, such as at least 98.0%, such as at least 99.0%, such as at least 99.2%, such as at least 99.4%, such as 99.5% identical, such as at least 99.6%, such as 99.7% identical such as at least 99.8% identical, such as 99.9% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

Alternatively the functional derivative has no more than 500, more suitably no more than 400, more suitably no more than 450, more suitably no more than 300, more suitably no more than 250, more suitably no more than 200, more suitably no more than 150, more suitably no more than 125, more suitably no more than 100, more suitably no more than 90, more suitably no more than 80, more suitably no more than 70, more suitably no more than 60, more suitably no more than 50, more suitably no more than 40, more suitably no more than 30, more suitably no more than 20, more suitably no more than

10, more suitably no more than 5, more suitably no more than 4, more suitably no more than 3, more suitably no more than 2 and more suitably no more than 1 addition(s), deletion(s) and/or substitutions(s) compared to SEQ ID NO: 5.

5 *ChAd155 Backbones*

The present application describes isolated polynucleotide sequences of chimp adenovirus ChAd155, including that of wild type, unmodified ChAd155 (SEQ ID NO: 10) and modified backbone constructs of ChAd155. These modified backbone constructs include ChAd155#1434 (SEQ ID NO: 7), ChAd155#1390 (SEQ ID NO: 8) and
 10 ChAd155#1375 (SEQ ID NO: 9). ChAd155 backbones may be used in the construction of recombinant replication-competent or replication-incompetent adenoviruses for the delivery of transgenes.

The term "construct" refers to a nucleic acid that encodes polypeptide sequences
 15 described herein and may comprise DNA or non-naturally occurring nucleic acid monomers.

The term "replication-competent" adenovirus refers to an adenovirus which can replicate in a host cell in the absence of any recombinant helper proteins comprised in the cell.
 20 Suitably, a "replication-competent" adenovirus comprises the following intact or functional essential early genes: E1A, E1B, E2A, E2B, E3 and E4. Wild type adenoviruses isolated from a particular animal will be replication competent in that animal.

25 The term "replication-incompetent" or "replication-defective" adenovirus refers to an adenovirus which is incapable of replication because it has been engineered to comprise at least a functional deletion (or "loss-of-function" mutation), i.e. a deletion or mutation which impairs the function of a gene without removing it entirely, e.g. introduction of artificial stop codons, deletion or mutation of active sites or interaction domains, mutation
 30 or deletion of a regulatory sequence of a gene etc., or a complete removal of a gene encoding a gene product that is essential for viral replication, such as one or more of the adenoviral genes selected from E1A, E1B, E2A, E2B, E3 and E4 (such as E3 ORF1, E3 ORF2, E3 ORF3, E3 ORF4, E3 ORF5, E3 ORF6, E3 ORF7, E3 ORF8, E3 ORF9, E4 ORF7, E4 ORF6, E4 ORF4, E4 ORF3, E4 ORF2 and/or E4 ORF1). Particularly suitably
 35 E1, and optionally E3 and/or E4, are deleted. If deleted, the aforementioned deleted gene region will suitably not be considered in the alignment when determining % identity with respect to another sequence.

The sequences of the invention are useful as therapeutic agents and in construction of a variety of vector systems, recombinant adenovirus and host cells. Suitably the term "vector" refers to a nucleic acid that has been substantially altered (e.g., a gene or functional region that has been deleted and/or inactivated) relative to a wild type sequence and/or incorporates a heterologous sequence, *i.e.*, nucleic acid obtained from a different source (also called an "insert"), and replicating and/or expressing the inserted polynucleotide sequence, when introduced into a cell (e.g., a host cell). For example, the insert may be all or part of the ChAd155 sequences described herein. In addition or alternatively, a ChAd155 vector may be a ChAd155 adenovirus comprising one or more deletions or inactivations of viral genes, such as E1 or other viral gene or functional region described herein. Such a ChAd155, which may or may not comprise a heterologous sequence, is often called a "backbone" and may be used as is or as a starting point for additional modifications to the vector.

Annotation of the ChAd155 wild type sequence (SEQ ID NO: 10) sequence is provided below.

LOCUS	ChAd155	37830 bp	DNA	linear	10-
JUN-2015					
DEFINITION	Chimp adenovirus 155, complete genome.				
COMMENT	Annotation according to alignment of ChAd155 against the human Adenovirus 2 reference strain NC_001405				
	Two putative ORFs in the E3 region added manually				
FEATURES	Location/Qualifiers				
source	1..37830				
	/organism="Chimpanzee adenovirus 155"				
	/mol_type="genomic DNA"				
	/acronym="ChAd155"				
repeat_region	1..101				
	/standard_name="ITR"				
	/rpt_type=inverted				
gene	466..1622				
	/gene="E1A"				
TATA_signal	466..471				
	/gene="E1A"				
prim_transcript	497..1622				
	/gene="E1A"				

	CDS	join(577..1117,1231..1532)
		/gene="E1A"
		/product="E1A_280R"
5	CDS	join(577..979,1231..1532)
		/gene="E1A"
		/product="E1A_243R"
	polyA_signal	1600..1605
		/gene="E1A"
	gene	1662..4131
10		/gene="E1B"
	TATA_signal	1662..1667
		/gene="E1B"
	prim_transcript	1692..4131
		/gene="E1B"
15	CDS	1704..2267
		/gene="E1B"
		/product="E1B_19K"
	CDS	2009..3532
		/gene="E1B"
20		/product="E1B_55K"
	gene	3571..4131
		/gene="IX"
	TATA_signal	3571..3576
		/gene="IX"
25	prim_transcript	3601..4131
		/gene="IX"
	CDS	3628..4092
		/gene="IX"
		/product="IX"
30	polyA_signal	4097..4102
		/note="E1B, IX"
	gene	complement(4117..27523)
		/gene="E2B"
	prim_transcript	complement(4117..27494)
		/gene="E2B"
35	gene	complement(4117..5896)
		/gene="IVa2"
	prim_transcript	complement(4117..5896)
		/gene="IVa2"

	CDS	complement(join(4151..5487,5766..5778))
		/gene="IVa2"
		/product="E2B_IVa2"
5	polyA_signal	complement(4150..4155)
		/note="IVa2, E2B"
	CDS	complement(join(5257..8838,14209..14217))
		/gene="E2B"
		/product="E2B_polymerase"
10	gene	6078..34605
		/gene="L5"
	gene	6078..28612
		/gene="L4"
	gene	6078..22658
		/gene="L3"
15	gene	6078..18164
		/gene="L2"
	gene	6078..14216
		/gene="L1"
20	TATA_signal	6078..6083
		/note="L"
	prim_transcript	6109..34605
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	CDS	complement(36579..36971)
		/gene="E4"
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Sequence identity

Identity with respect to a sequence is defined herein as the percentage of amino acid residues in the candidate sequence that are identical with the reference amino acid sequence after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity.

Sequence identity can be determined by standard methods that are commonly used to compare the similarity in position of the amino acids of two polypeptides. Using a computer program such as BLAST or FASTA, two polypeptides are aligned for optimal matching of their respective amino acids (either along the full length of one or both sequences or along a pre-determined portion of one or both sequences). The programs provide a default opening penalty and a default gap penalty, and a scoring matrix such as PAM 250 (a standard scoring matrix can be used in conjunction with the computer program. For example, the percent identity can then be calculated as the total number of identical matches multiplied by 100 and then divided by the sum of the length of the longer sequence within the matched span and the number of gaps introduced into the shorter sequences in order to align the two sequences.

Where the present disclosure refers to a sequence by reference to a UniProt or Genbank accession code, the sequence referred to is the current version as of the filing date of the present application.

The skilled person will recognise that individual substitutions, deletions or additions to a protein which alters, adds or deletes a single amino acid or a small percentage of amino acids is an "immunogenic derivative" where the alteration(s) results in the substitution of an amino acid with a functionally similar amino acid or the substitution/deletion/addition of residues which do not substantially impact the immunogenic function.

Conservative substitution tables providing functionally similar amino acids are well known in the art. In general, such conservative substitutions will fall within one of the amino-acid groupings specified below, though in some circumstances other substitutions may be possible without substantially affecting the immunogenic properties of the antigen. The following eight groups each contain amino acids that are typically conservative substitutions for one another:

- 1) Alanine (A), Glycine (G);
- 2) Aspartic acid (D), Glutamic acid (E);
- 3) Asparagine (N), Glutamine (Q);

- 4) Arginine (R), Lysine (K);
 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V);
 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W);
 7) Serine (S), Threonine (T); and
 5 8) Cysteine (C), Methionine (M).

Suitably such substitutions do not occur in the region of an epitope, and do not therefore have a significant impact on the immunogenic properties of the antigen.

- 10 Immunogenic derivatives may also include those wherein additional amino acids are inserted compared to the reference sequence. Suitably such insertions do not occur in the region of an epitope, and do not therefore have a significant impact on the immunogenic properties of the antigen. One example of insertions includes a short stretch of histidine residues (e.g. 2-6 residues) (SEQ ID NO: 49) to aid expression and/or
 15 purification of the antigen in question.

- Immunogenic derivatives include those wherein amino acids have been deleted compared to the reference sequence. Suitably such deletions do not occur in the region of an epitope, and do not therefore have a significant impact on the immunogenic
 20 properties of the antigen.

The skilled person will recognise that a particular immunogenic derivative may comprise substitutions, deletions and additions (or any combination thereof).

***Lyssavirus* antigens and vaccines**

- 25 *Lyssavirus*, a genus in the *Rhabdoviridae* family, is an enveloped virus with a single stranded antisense RNA genome. The RNA encodes five structural proteins in the order of a nucleoprotein (N), a phosphoprotein (P), a matrix protein (M), a glycoprotein (G) and a viral RNA polymerase (L). The P protein is a structural component of the ribonucleoprotein and plays a role in the formation of viral particles and viral RNA
 30 synthesis. The G protein is thought to be important in viral pathogenicity and protective immunity; it is a major target of protective neutralizing antibodies. *Lyssavirus* is a neurotropic virus that spreads through the central nervous system causing severe inflammation of the brain and spinal cord.

- 35 The *Lyssavirus* genus comprises seven genotypes, the following six of which have been associated with cases of human rabies: rabies virus (RABV, genotype 1), Mokola virus (genotype 3), Duvenhage virus (genotype 4), European bat *Lyssavirus* (genotype 5), European bat *Lyssavirus* 2 (genotype 6), and Australian bat *Lyssavirus* (genotype 7)

(Jackson (2016) Curr Infec Dis Rep 18:38). Once symptoms develop, rabies is nearly one hundred percent fatal.

Antigenic epitopes present on the rabies G protein have been identified in multiple strains of rabies viruses. They are classified as Site I, Site IIa, Site IIb, Site III, Site IV and Site a and are listed in Table 1. This Table discloses the 'Site II b' sequences as SEQ ID NOS 50-63, the 'Site I' sequences as SEQ ID NOS 64-77, and the 'Site III' sequences as SEQ ID NOS 78-91, respectively, in order of appearance.

Table 1. Rabies G Protein Antigenic Epitopes

Virus	Phylo-group	Site II b (34-42)	Site II a (198-200)	Site I (226-231)	Site IV (263-264)	Site III (330-338)	Site 'a' (342-343)
RABV	I	GCTNLSEPF	KKA	KLCGVL	FR	KSVRWKKEI	KG
ABLV	I	GCTSLSEPF	KKA	KLCGIS	FR	KSVRWKKEI	KG
ARAV	I	GCTNLSEPF	KKA	KLCGVK	FR	KSVRWKKEV	KG
BRLV	I	GCTTLVYFS	KKA	KLCGVK	FR	KSVRWKKEI	KG
DRVV	I	GCTTLVYFS	KKA	KLCGIS	FR	KSVRWKKEI	KG
EBLV-1	I	GCTTLVYFS	KKA	KLCGVK	FR	KSVRWKKEV	KG
EBLV-2	I	GCTTLVYFS	KKA	KLCGIS	FR	KSVRWKKEV	KG
BRV	I	GCTTLVYFS	KKA	KLCGVK	FR	KSVRWKKEI	KG
EHUV	I	GCTTLSEPF	KKA	KLCGVK	FR	KSVRWKKEI	KG
LSV	II	GCHSTATFS	KKS	KLCGVK	FR	KSVRWKKEI	KG
MOKV	II	GCHSTATFS	KKA	KLCGVK	FR	KSVRWKKEI	KG
SNV	II	GCHSTATFS	KKS	KLCGVK	FR	KSVRWKKEI	KG
WCHV	III	CTCTGGGTF	KLV	KLCGVK	FR	KSVRWKKEV	KG
BEKV	?	GCHSTATFS	ALL	KLCGVK	FR	KSVRWKKEI	KG

Antigenic epitopes present on the rabies G protein corresponding to SEQ ID NO: 37 are shown in FIG. 1. Antigenic Site I harbors both conformational and linear epitopes and is located at amino acid residues 226-231. Antigenic Site II is a discontinuous conformational epitope at residues 34-42 (IIb) and 198-200 (IIa). Antigenic Site III is a continuous conformational epitope at residues 330-338. Antigenic Site IV is located at residues 263-264. Antigenic Site a is located at residues 342-343.

Rabies vaccines are currently used primarily for post-exposure prophylaxis, only a small percentage of rabies vaccine doses are used for pre-exposure prophylaxis. The intervention schedule is defined by the World Health Organization based on the seriousness and the type of the wound via which the virus gains entry and may include additional treatment with anti-rabies immunoglobulin. Pre-exposure prophylaxis typically involves two to three visits for two to three intramuscular doses with boosters timed according to the exposure risk. Post-exposure prophylaxis typically involves three to five

visits for four to five intramuscular doses or four visits for four intradermal doses. In some less developed countries, immunization is still performed by propagating rabies virus in the brains of an infected animal, inactivating the virus and providing 14-21 daily injections given subcutaneously into the abdominal wall.

5

Several rabies vaccines are currently available for human use in both pre-exposure and post-exposure prophylaxis. IMOVAX (Sanofi Pasteur) is provided as freeze-dried rabies virus prepared from strain PM-1503-3M obtained from the Wistar Institute. It is harvested from infected human diploid cells then inactivated. Both pre- and post-exposure prophylaxis consists of three doses administered intramuscularly on days 0, 7 and 21 or 28. VERORAB (Sanofi Pasteur) is provided as freeze-dried rabies virus prepared from strain PM/WI 38 1503-3M obtained from the Wistar Institute. It is harvested from Vero cells then inactivated. Pre-exposure prophylaxis consists of three doses administered intramuscularly on days 0, 7 and 21 or 28. Post-exposure prophylaxis consists of five doses administered intramuscularly on days 0, 3, 7, 14 and 28. VAXIRAB/ LYSSAVAC (Zydus Cadila/ Novavax) is provided as freeze-dried rabies virus prepared from the Pitman Moore strain of the rabies virus. It is produced in duck embryo cells then inactivated. Pre-exposure prophylaxis consists of three doses administered intramuscularly on days 0, 7 and 21 or 28. Post-exposure prophylaxis consists of five doses administered intramuscularly on days 0, 3, 7, 14 and 28. Post-exposure prophylaxis can also be administered intradermally, injected at each of two sites on days 0, 3, 7 and 28. RABIPUR/ RABAVERT (GSK) is provided as a freeze-dried rabies virus prepared from the Flury LEP (low egg passage) strain. It is grown in primary cultures of chicken fibroblasts then inactivated. Pre-exposure prophylaxis consists of three doses administered intramuscularly on days 0, 7 and 21 or 28. Post-exposure prophylaxis consists of five doses administered intramuscularly on days 0, 3, 7, 14 and 28.

Supportive pre-clinical evidence for adeno-vectored rabies vaccines has been reported in the literature. The adenoviral recombinant viral vector SAdV24, also termed AdC68 or ChAd68, modified to be replication defective and to express the full length glycoprotein (G) of the Evelyn Rokitniki Abelseth (ERA) strain of rabies showed some degree of immunogenicity in cynomologous monkeys when given prior to a rabies challenge but did not provide reliable protection after a rabies exposure (Xiang *et al.* (2014) *Virology* 450:451:243-249). A similar replication defective ChAd68 vector expressing the full length glycoprotein (G) of the Evelyn Rokitniki Abelseth (ERA) strain of rabies, given

intramuscularly, induced a degree of protection against a rabies challenge (Zhou *et al.* (2006) Mol. Ther. 14:662-672; reproduced in part in FIG. 16).

Adjuvants

- 5 An “adjuvant” as used herein refers to a composition that enhances the immune response to an immunogen. A composition according to the invention that comprises an adjuvant can be used as a vaccine, e.g. for human subjects. The adjuvant accelerates, prolongs and/or enhances the quality and/or strength of an immune response to an antigen/immunogen in comparison to the administration of the antigen alone, thus,
- 10 reduces the quantity of antigen/immunogen necessary in any given vaccine, and/or the frequency of injection necessary in order to generate an adequate immune response to the antigen/immunogen of interest.

- Examples of adjuvants that may be used in the context of the compositions of the
- 15 invention include inorganic adjuvants (e.g. inorganic metal salts such as aluminum phosphate or aluminum hydroxide), gel-like precipitates of aluminum hydroxide (alum); AlPO_4 ; alhydrogel; bacterial products from the outer membrane of Gram-negative bacteria, in particular monophosphoryl lipid A (MPLA), lipopolysaccharides (LPS), muramyl dipeptides and derivatives thereof; Freund's incomplete adjuvant; liposomes, in
- 20 particular neutral liposomes, liposomes containing the composition and optionally cytokines; AS01B, AS01E, AS02; non-ionic block copolymers; ISCOMATRIX adjuvant; unmethylated DNA comprising CpG dinucleotides (CpG motif), in particular CpG ODN with a phosphorothioate (PTO) backbone (CpG PTO ODN) or phosphodiester (PO) backbone (CpG PO ODN); synthetic lipopeptide derivatives, in particular Pam_3Cys ;
- 25 lipoarabinomannan; peptidoglycan; zymosan; heat shock proteins (HSP), in particular HSP 70; dsRNA and synthetic derivatives thereof, in particular Poly I:poly C; polycationic peptides, in particular poly-L-arginine; taxol; fibronectin; flagellin; imidazoquinoline; cytokines with adjuvant activity, in particular GM-CSF, interleukin- (IL-)2, IL-6, IL-7, IL-18, type I and II interferons, in particular interferon-gamma (IFN-gamma), TNF-alpha;
- 30 25-dihydroxyvitamin D3 (calcitriol); and synthetic oligopeptides, in particular MHCII-presented peptides. Non-ionic block polymers containing polyoxyethylene (POE) and polyoxypropylene (POP), such as POE-POP-POE block copolymers may be used as an adjuvant.

- 35 Additional examples of adjuvants include inorganic adjuvants (e.g. inorganic metal salts such as aluminium phosphate or aluminium hydroxide), organic adjuvants (e.g. saponins, such as QS21, or squalene), oil-based adjuvants (e.g. Freund's complete

adjuvant and Freund's incomplete adjuvant), cytokines (e.g. IL-1 β , IL-2, IL-7, IL-12, IL-18, GM-CSF, and INF- γ) particulate adjuvants (e.g. immuno-stimulatory complexes (ISCOMS), liposomes, biodegradable microspheres, virosomes, bacterial adjuvants (e.g. monophosphoryl lipid A, such as 3-de-O-acylated monophosphoryl lipid A (3D-MPL), or
 5 muramyl peptides), synthetic adjuvants (e.g. monophosphoryl lipid A (MPL), in particular 3-de-O-acylated monophosphoryl lipid A (3D-MPL and muramyl peptide analogues, or synthetic lipid A, and synthetic polynucleotides adjuvants, e.g., polyarginine or polylysine.

10 Saponins are also suitable adjuvants, for example, the saponin Quil A, derived from the bark of the South American tree *Quillaja Saponaria Molina*, and fractions thereof. Purified fractions of Quil A are also known as immunostimulants, such as squalene, QS21, QS17 and QS7, a non-haemolytic fraction of Quil-A. Combinations of QS21 and polysorbate or cyclodextrin are also suitable.

15 Another example of an adjuvant is an immunostimulatory oligonucleotide containing unmethylated cytosine-guanosine dinucleotide motifs present in DNA ("CpG"). CpG is known as an adjuvant when administered by both systemic and mucosal routes. When formulated into vaccines, it may be administered in free solution together with free
 20 antigen or covalently conjugated to an antigen or formulated with a carrier such as aluminium hydroxide.

Activation of specific receptors can stimulate an immune response. Such receptors are known to the skilled artisan and comprise, for example, cytokine receptors, in particular
 25 type I cytokine receptors, type II cytokine receptors, TNF receptors; and a vitamin D receptor acting as transcription factor; and the Toll-like receptors 1 (TLR1), TLR-2, TLR 3, TLR4, TLR5, TLR-6, TLR7, and TLR9. Agonists to such receptors have adjuvant activity, i.e., are immunostimulatory. Other suitable adjuvants include alkyl glucosaminide phosphates (AGPs) or pharmaceutically acceptable salts of AGPs. Some
 30 AGPs are TLR4 agonists, and some are TLR4 antagonists. An adjuvant of the composition of the present invention may be one or more Toll-like receptor agonists. In a more preferred embodiment, the adjuvant is a Toll-like receptor 4 agonist. In a particular preferred embodiment, the adjuvant is a Toll-like receptor 9 agonist.

35 Adjuvants such as those described above may be formulated together with carriers, such as liposomes, oil in water emulsions, and/or metallic salts (including aluminum salts such as aluminum hydroxide). For example, 3D-MPL may be formulated with aluminum

hydroxide or oil in water emulsions; QS21 may be formulated with cholesterol containing liposomes, oil in water emulsions or alum; CpG may be formulated with alum or with other cationic carriers.

- 5 Combinations of adjuvants may be utilized in the present invention, in particular a combination of a monophosphoryl lipid A and a saponin derivative, more particularly the combination of QS21 and 3D-MPL or a composition where the QS21 is quenched in cholesterol-containing liposomes (DQ). Alternatively, a combination of CpG plus a saponin such as QS21 is an adjuvant suitable for use in the present invention, as is a
- 10 potent adjuvant formulation involving QS21, 3D-MPL and tocopherol in an oil in water emulsion. Saponin adjuvants may be formulated in a liposome and combined with an immunostimulatory oligonucleotide. Thus, suitable adjuvant systems include, for example, a combination of monophosphoryl lipid A, preferably 3D-MPL, together with an aluminium salt. A further exemplary adjuvant comprises QS21 and/or MPL and/or CpG.
- 15 QS21 may be quenched in cholesterol-containing liposomes.

- The fusion of the invariant chain to an antigen which is comprised by an expression system used for vaccination increases the immune response against said antigen, if it is administered with an adenovirus. Accordingly, in one embodiment of the invention, the
- 20 immunogenic transgene may be co-expressed with invariant chain in a recombinant ChAd155 viral vector.

- In another embodiment, the invention provides the use of the capsid of ChAd155 (optionally an intact or recombinant viral particle or an empty capsid is used) to induce
- 25 an immunomodulatory response, or to enhance or adjuvant a cytotoxic T cell response to another active agent by delivering a ChAd155 capsid to a subject. The ChAd155 capsid can be delivered alone or in a combination regimen with an active agent to enhance the immune response thereto. Advantageously, the desired effect can be accomplished without infecting the host with an adenovirus.

30

General

- Unless otherwise explained, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure belongs. The singular terms "a," "an," and "the" include plural referents
- 35 unless context clearly indicates otherwise. Similarly, the word "or" is intended to include "and" unless the context clearly indicates otherwise. The term "plurality" refers to two or more. Additionally, numerical limitations given with respect to concentrations or levels of

a substance, such as solution component concentrations or ratios thereof, and reaction conditions such as temperatures, pressures and cycle times are intended to be approximate. The term “about” used herein is intended to mean the amount $\pm 10\%$.

- 5 The invention will be further described by reference to the following, non-limiting, examples and figures.

EXAMPLES

Example 1: Isolation of ChAd155

- 10 Wild type chimpanzee adenovirus type 155 (ChAd155) was isolated from a healthy young chimpanzee housed at the New Iberia Research Center facility (New Iberia Research Center, The University of Louisiana at Lafayette) using standard procedures as described in Colloca *et al.* (2012) Sci. Transl. Med. 4:1-9 and WO 2010086189, which is hereby incorporated by reference for the purpose of describing adenoviral isolation
15 and characterization techniques.

Example 2: ChAd155-RG Vector Construction

The ChAd155 viral genome was then cloned in a plasmid or in a BAC vector and subsequently modified as shown in FIG. 3:

- 20 a) deletion of the E1 region (from bp 449 to bp 3529) of the viral genome;
b) deletion of the E4 region (from bp 34731 to bp 37449) of the viral genome;
c) insertion of the E4orf6 derived from human Ad5; and
d) insertion of hCMV-RG-WPRE expression cassette.

- 25 *2.1: Deletion of E1 region: Construction of BAC/ChAd155 $\Delta E1$ _TetO hCMV RpsL-Kana#1375*

- The ChAd155 viral genome was cloned into a BAC vector by homologous recombination in *E. coli* strain BJ5183 electroporation competent cells (Stratagene catalog no.
30 2000154) co-transformed with ChAd155 viral DNA and Subgroup C BAC Shuttle (#1365). As shown in the schematic of FIG. 4, the Subgroup C Shuttle is a BAC vector derived from pBeloBAC11 (GenBank U51113, NEB). It is dedicated to the cloning of chimp adeno viruses belonging to species C and therefore contains the plX gene and DNA fragments derived from the right and left ends (including right and left ITRs) of
35 species C ChAd viruses.

The Species C BAC Shuttle also contains a RpsL-Kana cassette inserted between the left end and the pIX gene. In addition, an Amp-LacZ-SacB selection cassette, flanked by ISceI restriction sites, is present between the pIX gene and the right end of the viral genome. In particular, the BAC Shuttle comprised the following features: Left ITR: bp 27 to 139, hCMV(tetO) RpsL-Kana cassette: bp 493 to 3396, pIX gene: bp 3508 to 3972, ISceI restriction sites: bp 3990 and 7481, Amp-LacZ-SacB selection cassette: bp 4000 to 7471, Right ITR: bp 7805 to 7917.

BJ5183 cells were co-transformed by electroporation with ChAd155 purified viral DNA and Subgroup C BAC Shuttle vector digested with ISceI restriction enzyme and then gel purified. Homologous recombination occurring between pIX gene and right ITR sequences (present at the ends of Species C BAC Shuttle linearized DNA) and homologous sequences present in ChAd155 viral DNA lead to the insertion of ChAd155 viral genomic DNA in the BAC shuttle vector. At the same time, the viral E1 region was deleted and substituted by the RpsL-Kana cassette, generating BAC/ChAd155 Δ E1/TetO hCMV RpsL-Kana#1375.

2.2: Plasmid construction by homologous recombination in *E. coli* BJ5183

2.2.1: Deletion of E4 region - Construction of pChAd155 Δ E1, E4_Ad5E4orf6/TetO hCMV RpsL-Kana (#1434)

To improve propagation of the vector, a deletion of the E4 region spanning from nucleotide 34731-37449 (ChAd155 wild type sequence) was introduced in the vector backbone by replacing the native E4 region with Ad5 E4orf6 coding sequence using a strategy involving several steps of cloning and homologous recombination in *E. coli*. The E4 coding region was completely deleted while the E4 native promoter and polyadenylation signal were conserved. To this end, a shuttle vector was constructed to allow the insertion of Ad5orf6 by replacing the ChAd155 native E4 region by homologous recombination in *E. coli* BJ5183 as detailed below.

Construction of pARS SpeciesC Ad5E4orf6-1

A DNA fragment containing Ad5orf6 was obtained by PCR using Ad5 DNA as template, with the oligonucleotides 5'-ATACGGACTAGTGGAGAAGTACTCGCCTACATG-3' (SEQ ID NO: 13) and 5'-ATACGGAAGATCTAAGACTTCAGGAAATATGACTAC-3' (SEQ ID NO: 14). The PCR fragment was digested with BglII and SpeI and cloned into Species C RLD-EGFP shuttle digested with BglII and SpeI, generating the plasmid pARS

Species C Ad5orf6-1. Details regarding the shuttle can be found in Colloca *et al.*, Sci. Transl. Med. (2012) 4:115ra2.

Construction of pARS SpeciesC Ad5E4orf6-2

- 5 To delete the E4 region, a 177 bp DNA fragment spanning bp 34586 to bp 34730 of the ChAd155 wt sequence (SEQ ID NO: 10) was amplified by PCR using the plasmid BAC/ChAd155 ΔE1_TetO hCMV RpsL-Kana (#1375) as a template with the following oligonucleotides: 5'-ATTCAGTGTACAGGCGCGCCAAAGCATGACGCTGTTGATTTGATTC-3' (SEQ ID NO: 15) and 5'-ACTAGGACTAGTTATAAGCTAGAATGGGGCTTTGC-3' (SEQ ID NO: 16). The PCR fragment was digested with BsrGI and SpeI and cloned into pARS SubGroupC Ad5orf6-1 digested with BsrGI and SpeI, generating the plasmid pARS SpeciesC Ad5orf6-2 (#1490). A schematic diagram of this shuttle plasmid is provided in FIG. 5. In particular, the shuttle plasmid comprised the following features: Left ITR: bp 1 to 113, Species C first 460bp: bp 1 to 460, ChAd155 wt (bp 34587 to bp 34724 of SEQ ID NO:10) : bp 516 to 650, Ad5 orf6: bp 680 and 1561, Species C last 393 bp: bp 1567 to 1969, Right ITR: bp 1857 to 1969.

Construction of pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV RpsL-Kana (#1434)

- 20 The resulting plasmid pARS SubGroupC Ad5orf6-2 was then used to replace the E4 region within the ChAd155 backbone with Ad5orf6. To this end the plasmid BAC/ChAd155 ΔE1_TetO hCMV RpsL-Kana (#1375) was digested with PacI/PmeI and co-transformed into BJ5183 cells with the digested plasmid pARS SubGroupC Ad5orf6-2 BsrGI/Ascl, to obtain the pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV RpsL-Kana (#1434) pre-adenoviral plasmid.

2.2.2: Insertion of hCMV-RG Expression Cassette - Construction of pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV-RG#1481

- hCMV-RG cassette was cloned into a linearized pre-adenoviral acceptor vector via homologous recombination in *E. coli* by exploiting the homology existing between hCMV promoter and BGH polyA sequence. The plasmid pVjTetOhCMV_RG_bghpolyA, shown in FIG. 6, was cleaved with SpeI, SphI and AsiSI to excise the 2.58 Kb fragment containing the hCMV promoter with tetO, RG and BGHpolyA sequence. The resulting 2.58 Kb fragment was cloned by homologous recombination into the pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV RpsL-Kana (#1434) acceptor vector carrying the RpsL-Kana selection cassette under the control of HCMV and BGHpA. The acceptor pre-adenoviral

plasmid was linearized with the restriction endonuclease SnaBI. The resulting construct was the pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV-RG vector (#1481) (FIG. 7).

2.2.3: Insertion of hCMV-RG-WPRE Expression Cassette - Construction of pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV-RG-WPRE#1509

A WPRE sequence was cloned into a pre-adeno acceptor vector via homologous recombination in *E. coli* by exploiting the homology existing between bases 2840-2939 and 3180-3279 of pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV-RG vector (#1481). A 1031 bp DNA fragment was amplified by PCR and contains WPRE, BGHpolyA and recombination arms corresponding to bases 2840-2939 and 3180-3279 of #1481 pAdeno vector. PCR was performed using the plasmid pvjTetOhCMV_WPRE_BghPolyA (#1478) as a template and with the following oligonucleotides FW 5'-

ggaaggtcagcgtgaccagccagtcggaagtgatttcctcctgggagagctataaaagcggcgagagaccagggc

15 tgtgatgagcggccgcgatctgtaataacacctggattaca -3' (SEQ ID NO: 92) and RW 5'-

ATGGCTCCGGCGGTCTCTGCAACACAAATAAAGAGACCCTAAGACCCCCAACTTAT

ATATTTTCATGACCACCCCAGGCCACGCCCACTACCCACCTCACCATAGAGCCCA

CCGCATCC-3' (SEQ ID NO: 93). The resulting 1.03 Kb fragment was cloned by

homologous recombination into the pChAd155 ΔE1, E4_Ad5E4orf6/TetO hCMV-RG

20 vector (#1481) acceptor vector carrying the RG transgene (SEQ ID NO: 38) under the

control of hCMV promoter and BGHPA. The acceptor pre-adeno plasmid was digested

with the restriction endonuclease AsiSI. The resulting construct was the pChAd155

ΔE1, E4_Ad5E4orf6/TetO hCMV-RG-WPRE vector (#1509), shown in FIG. 8.

25 Example 3: ChAd155-RG Vector Production

The productivity of ChAd155 was evaluated in comparison to ChAd3 and PanAd3 in the Procell 92 cell line.

3.1: Production of vectors comprising an HIV Gag transgene

30 Vectors expressing the HIV Gag protein were prepared as described above (ChAd155/GAG) or previously as for ChAd3/GAG (Colloca *et al*, Sci. Transl. Med. (2012) 4:115ra2). ChAd3/GAG and ChAd155/GAG were rescued and amplified in Procell 92 until passage 3 (P3); P3 lysates were used to infect two T75 flasks of Procell 92 cells cultivated in monolayer with each vector. A multiplicity of infection (MOI) of 100 vp/cell

35 was used for both infection experiments. The infected cells were harvested when the full cytopathic effect was evident (72 hours post-infection) and pooled; the viruses were

released from the infected cells by three cycles of freeze/thaw (-70°/37°C) then the lysate was clarified by centrifugation. The clarified lysates were quantified by Quantitative PCR (QPCR) analysis with primers and probe complementary to the CMV promoter region. The oligonucleotide sequences are the following: CMVfor 5'-
 5 CATCTACGTATTAGTCATCGCTATTACCA-3' (SEQ ID NO: 23), CMVrev 5'-
 GACTTGGAAATCCCCGTGAGT-3' (SEQ ID NO: 24), CMVFAM-TAMRA probe 5'-
 ACATCAATGGGCGTGGATAGCGGTT-3' (SEQ ID NO: 25) (QPCRs were run on an ABI Prism 7900 Sequence detector – Applied Biosystem). The resulting volumetric titers (vp/ml) measured on clarified lysates and the cell specific productivity expressed in virus
 10 particles per cell (vp/cell) are provided in Table 2 below.

Table 2. Vector productivity from P3 lysates

Vector	vp/ml	Total vp (20 ml conc.)	vp/cell
ChAd3/GAG	9.82E+09	1.96E+11	6.61E+03
ChAd155/GAG	1.11E+10	2.22E+11	7.46E+03

15 To confirm the higher productivity of the ChAd155 vector expressing HIV Gag transgene, a second experiment was performed by using purified viruses as inoculum. To this end, Procell 92 cells were seeded in a T25 Flask and infected with ChAd3/GAG and ChAd155/GAG when the confluence of the cells was about 80%, using an MOI=100
 20 vp/cell. The infected cells were harvested when the full cytopathic effect was evident; the viruses were released from the infected cells by freeze/thaw and clarified by centrifugation. The clarified lysates were quantified by Quantitative PCR analysis by using the following primers and probe: CMVfor 5'-
 CATCTACGTATTAGTCATCGCTATTACCA-3' (SEQ ID NO: 23), CMV rev
 GACTTGGAAATCCCCGTGAGT (SEQ ID NO: 24), CMV FAM-TAMRA probe 5'-
 25 ACATCAATGGGCGTGGATAGCGGTT-3' (SEQ ID NO: 25) complementary to the CMV promoter region (samples were analysed on an ABI Prism 7900 Sequence detector- Applied Biosystems). The resulting volumetric titers (vp/ml) measured on clarified lysates and the cell specific productivity expressed in virus particles per cell (vp/cell) are provided in Table 3.

Table 3. Vector productivity from purified viruses

Vector	vp/ml	Total vp/T25 flask (5ml of lysate)	vp/cell
--------	-------	---------------------------------------	---------

ChAd3/GAG	1.00E+10	5.00E+10	1.67E+04
ChAd155/GAG	1.21E+10	6.05E+10	2.02E+04

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3.2: Production of vectors comprising an RSV transgene

A different set of experiments was performed to evaluate the productivity of RSV vaccine vectors in Procell 92.S cells cultivated in suspension. The experiment compared PanAd3/RSV (described in WO2012/089833) and ChAd155/RSV in parallel by infecting Procell 92.S at a cell density of 5×10^5 cells/ml. The infected cells were harvested three days post infection; the virus was released from the infected cells by three cycles of freeze/thaw and the lysate was clarified by centrifugation. The clarified lysates were then quantified by Quantitative PCR analysis as reported above. The resulting volumetric titers (vp/ml) measured on clarified lysates and the cell specific productivity expressed in virus particles per cell (vp/cell) are provided in Table 4.

Table 4. Vector productivity from purified viruses

Virus	(Vp/ml)	Total vp	(vp/cell)
PanAd3/RSV	5.82E+09	2.91E+11	1.16E+4
ChAd155/RSV	3.16E+10	1.58E+12	6.31E+04

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Example 4: Transgene Expression Levels

4.1: Expression level of HIV Gag transgene

Expression levels were compared in parallel experiments by infecting HeLa cells with ChAd3 and ChAd155 vectors comprising an HIV Gag transgene. HeLa cells were seeded in 24 well plates and infected in duplicate with ChAd3/GAG and ChAd155/GAG purified viruses using an MOI=250 vp/cell. The supernatants of HeLa infected cells were harvested 48 hours post-infection, and the production of secreted HIV Gag protein was quantified by using a commercial ELISA Kit (HIV-1 p24 ELISA Kit, PerkinElmer Life Science). The quantification was performed according to the manufacturer's instruction

by using an HIV-1 p24 antigen standard curve. The results, expressed in pg/ml of Gag protein, are illustrated in FIG. 9.

4.2: Expression level of RSV F transgene

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Expression levels were compared in parallel experiments by infecting HeLa cells with the above-described PanAd3 and ChAd155 vectors comprising an RSV F transgene. To this end, HeLa cells were seeded in 6 well plates and infected in duplicate with PanAd3/RSV and ChAd155/RSV purified viruses using an MOI=500 vp/cell. The supernatants were harvested 48 hours post-infection, and the production of secreted RSV F protein was quantified by ELISA. Five different dilutions of the supernatants were transferred to microplate wells which were coated with a commercial mouse anti-RSV F monoclonal antibody. The captured antigen was revealed using a secondary anti-RSV F rabbit antiserum followed by biotin-conjugated anti-rabbit IgG, then by adding Streptavidin-AP conjugate (BD Pharmingen cat. 554065). The quantification was performed by using an RSV F protein (Sino Biological cat. 11049-V08B) standard curve. The results obtained, expressed as ug/ml of RSV F protein, are provided in Table 5.

Table 5. Expression level of RSV F transgene

Sample	µg/ml RSV F protein
ChAd155/RSV	5.9
PanAd3/RSV	4

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A western blot analysis was also performed to confirm the higher level of transgene expression provided by the ChAd155 RSV vector relative to the PanAd3 RSV vector. HeLa cells plated in 6 well plates were infected with PanAd3/RSV and ChAd155/RSV purified viruses using an MOI=250 and 500 vp/cell. The supernatants of HeLa infected cells were harvested and the production of secreted RSV F protein were analysed by non-reducing SDS gel electrophoresis followed by western blot analysis. Equivalent quantities of supernatants were loaded onto a non-reducing SDS gel; after electrophoresis separation, the proteins were transferred to a nitrocellulose membrane to be probed with an anti-RSV F mouse monoclonal antibody (clone RSV-F-3 catalog no: ABIN308230), available at antibodies-online.com (last accessed 13 April 2015). After the incubation with primary antibody, the membrane was washed and then incubated with anti-mouse HRP conjugate secondary antibody. Finally the assay was developed by electrochemiluminescence (ECL) using standard techniques (ECL

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detection reagents Pierce catalog no W3252282). The western blot results are shown in FIG. 10. A band of about 170 kD indicated by the arrow was revealed by monoclonal antibody mAb 13 raised against the F protein, which corresponds to the expected weight of trimeric F protein. It can be seen that the ChAd155 RSV vector produced a darker band than PanAd3RSV at MOIs of both 250 and 500 vp/cell.

Example 5: Evaluation of Immunological Potency by Mouse Immunization Experiments

5.1: Immunogenicity of vectors comprising the HIV Gag transgene

The immunogenicity of the ChAd155/GAG vector was evaluated in parallel with the ChAd3/GAG vector in BALB/c mice (five per group). The experiment was performed by injecting 10^6 viral particles intramuscularly. T-cell response was measured three weeks after the immunization by *ex vivo* IFN-gamma enzyme-linked immunospot (ELISpot) using a GAG CD8+ T cell epitope mapped in BALB/c mice. The results are shown in FIG. 11, expressed as IFN-gamma Spot Forming Cells (SFC) per million splenocytes. Each dot represents the response in a single mouse, and the line corresponds to the mean for each dose group. Four out of five mice responded positively to the CD8 immunodominant peptide in response to both vectors.

5.2: Immunogenicity of vectors comprising the RSV transgene

The immunological potency of the PanAd3/RSV and ChAd155/RSV vectors was evaluated in BALB/c mice. Both vectors were injected intramuscularly at doses of 10^8 , 10^7 and 3×10^6 vp. Three weeks after vaccination the splenocytes of immunized mice were isolated and analyzed by IFN-gamma-ELISpot using as antigens immunodominant peptide F and M epitopes mapped in BALB/c mice. The levels of the immune-responses were reduced in line with decreasing dosage (as expected) but immune responses were clearly higher in the groups of mice immunized with ChAd155/RSV vector compared to the equivalent groups of mice immunized with PanAd3/RSV vaccine (FIG. 12). Symbols show individual mouse data, expressed as IFN-gamma Spot Forming Cells (SFC)/million splenocytes, calculated as the sum of responses to the three immunodominant epitopes (F₅₁₋₆₆, F₈₅₋₉₃ and M2-1₂₈₂₋₂₉₀) and corrected for background. Horizontal lines represent the mean number of IFN-gamma SFC/million splenocytes for each dose group.

Taken together the results reported above demonstrated that ChAd155 is an improved adenoviral vector in comparison to ChAd3 and PanAd3 vectors. ChAd155 was shown to

be more productive, therefore facilitating the manufacturing process, and shown to be able to express higher level of transgene both *in vitro* and *in vivo*, providing a stronger T-cell response against the antigens expressed in animal models.

5 **Example 6. ChAd155-RG Is Immunogenic and Protective Against a Rabies Challenge**

6.1: Immunogenicity of the ChAd155-AG vector

The immunological potency of the ChAd155-RG vector was evaluated in CD1 mice and the results shown in FIG. 13. The experiment was performed by injecting 10^9 vp intramuscularly. Each dot represents the response of a single mouse. FIG. 13 demonstrates that a single administration of a replication defective adenoviral vector encoding the rabies viral G protein antigen induced a potent immune response. The vector induced protective levels of neutralizing antibodies (FIG. 13A) and induced circulating rabies specific T cells (FIG. 13B).

A fluorescent antibody virus neutralization assay (FAVN) was performed as described in Cliquet F. *et al.*, J. Immunol. Methods (1998) 212:79-87. FIG. 13A demonstrates that functional neutralizing antibodies were detected in the serum within two weeks following a single administration of replication defective ChAd155-RG. Neutralizing antibodies were detected in amounts well above the protective threshold level of 0.5 IU/ml, as set forth in the World Health Organization guidelines (dotted line) by the second week post-administration showing no indication of a decline at week four. FIG. 13B demonstrates that rabies specific T cells were detected in the spleens of CD1 mice injected with 10^{-9} pfu/ml ChAd155-RG. An interferon-gamma ELISpot assay performed as described above on overlapping peptides spanning the rabies G protein sequence demonstrated the presence of rabies-specific T cells.

The antibody kinetics were followed up to 21 weeks after a single vaccine injection; the titers peaked at week 8 and then declined but remained well above the seroconversion threshold (dotted line), as shown in FIG. 14.

The immunological potency of ChAd155-RG was then compared to commercially available rabies vaccines in a single-dose regimen and the results are shown in FIG. 15. The left panel shows the results of immunizing Balb/c mice with either an estimated 1/500 of a human dose of ChAd155-RG (5×10^8 viral particles) or 1/10 of the canine dose of the veterinary rabies vaccine NOBIVAC. The right panel shows the results of

immunizing CD1 mice with either 1/1000 of an estimated human dose of ChAd155-RG (10⁸ viral particles) or 1/10 of the human dose of RABIPUR. Virus neutralizing antibody titers were measured as described above, described as IU/ML, and the titers shown at two months after the single-dose vaccination. Despite the large excesses of the commercial vaccines, the immunity induced by the ChAd155-RG vector proved superior to both the commercially available veterinary and human rabies vaccines.

6.2: Ability of the ChAd155-AG vector to protect against a rabies challenge

FIG. 16 demonstrates that a single dose of ChAd155-RG protects against a rabies challenge. Outbred ICR mice, four to six weeks of age, were injected in the gastrocnemius muscle with ChAd155-RG, ChAd155 control vector or RABIPUR at the doses shown in Table 6. Each of groups 1-6 consisted of ten mice. The mice were given three doses of RABIPUR on days 0, 7 and 21 or a single dose of ChAd155 or ChAd155-RG at the doses shown in Table 6.

Table 6. A Single Dose of ChAd155-RG Protects Against a Rabies Challenge

Group	Vector	Dose	Seroconversion Rate	Survival Rate
1	ChAd155 control	10 ⁸ virus particles	0%	60%
2	RABIPUR at days 0, 7 and 21	1/10 th human dose x 3	100%	100%
3	ChAd155-RG	10 ⁸ virus particles	100%	100%
4	ChAd155-RG	10 ⁷ virus particles	100%	100%
5	ChAd155-RG	10 ⁶ virus particles	90%	90%
6	ChAd155-RG	10 ⁵ virus particles	20%	60%

The mice were then challenged with a human isolate of a bat rabies virus variant and followed for 90 days. The challenge virus was the street RABV variant Ps P4 isolated from a fatal human case associated with exposure to a rabid bat. The challenge dose, calculated in a previous experiment in naïve unvaccinated animals, was 100% lethal. In this study, the same dose was 60% lethal. Serology was performed by a rapid fluorescent focus inhibition test for rabies (RFFIT), performed as described by Smith *et al.* (1973) Bull. World Health Organ. 48:535–541, to detect rabies-specific neutralizing antibodies. Also, direct immunofluorescence using LIGHT DIAGNOSTICS Rabies Polyclonal DFA Reagent (Millipore Cat#5199) was performed to detect viral antigen in the brain tissue.

FIG. 16 shows the level of rabies-specific neutralizing antibodies for each individual mouse. The mice in Group 1, given a negative control vector comprising no rabies antigen, did not seroconvert and 60% of the group survived. The mice in Groups 3-6

were given decreasing viral particle loads of ChAd155-RG. All of the mice seroconverted and survived when given 10^8 or 10^7 virus particles. Mice injected with 10^6 virus particles had a 90% seroconversion rate and 90% survived. Mice injected with 10^5 virus particles had a 20% seroconversion rate and 60% survived. This demonstrates that a single intramuscular vaccination of ChAd155-RG elicited neutralizing antibody titers above the threshold of 0.5 IU/ml over a wide dose range and conferred protection against a lethal rabies challenge. FIG. 16 and Table 6 therefore demonstrate that a single administration of recombinant ChAd155-RG can be at least as effective in protecting against rabies as a conventional, currently used, inactivated viral vaccine.

Example 7. ChAd155-RG is More Potent than AdC68rab.gp in Protecting Against a Rabies Challenge

The potency of the ChAd155-RG vector to protect against a rabies virus challenge was compared to the potency, as reported in the literature, for the AdC68 rab.gp vector.

Balb/c mice were immunized intramuscularly with a single dose of ChAd155-RG, as shown in Table 7. The pre-challenge viral neutralizing antibody levels were dose-dependent and are shown in FIG. 17A. These mice were then challenged with a human isolate of a bat rabies virus variant, as described in Example 6. As shown in Table 7, 60% of the mice given control ChAd155 vector survived. Balb/c mice immunized with 10^8 or 10^7 vp ChAd155-RG had a 100% survival rate, mice immunized with 10^6 vp ChAd155-RG had a 90% survival rate and mice immunized with 10^5 vp ChAd155-RG had a 60% survival rate, and the neutralizing antibody titers fell to nearly the seroconversion threshold.

These results were then correlated with the results published by Zhou (2006) Mol. Ther. 14:662 at 670 (FIG. 17B). Zhou et al. reported immunizing ICR mice intramuscularly with the adenoviral recombinant viral vector AdC68rab.gp, then challenging intranasally with CVS-N2C rabies virus. Control animals were not vaccinated and had a 100% fatality rate. Forty five percent of the mice immunized with 5×10^5 pfu AdC68rab.gp seroconverted and showed a 77% survival rate (17B left panel) while mice immunized with 5×10^4 pfu ChAd155-RG (17B right panel) showed 90% seroconversion and had a 60% survival rate.

Similar serological and protective efficacy data were obtained in the inventor's present study and the study reported by Zhou et al., when using a fifty-fold smaller dose (AdC68rab.gp at 5×10^5 pfu compared to ChAd155-RG at 10^4 pfu). ChAd155-RG is therefore about fifty times more potent than AdC68rab.gp.

Table 7. Potency of ChAd155-RG and AdC68rab.gp

Group	Vector	Dose	Seroconversion Rate	Survival Rate
1	ChAd155 control	10^8 virus particles	0%	60%
2	ChAd155-RG	10^8 virus particles	100%	100%
3	ChAd155-RG	10^7 virus particles	100%	100%
4	ChAd155-RG	10^6 virus particles	90%	90%
5	ChAd155-RG	10^5 virus particles	20%	60%
6	AdC68rab.gp	5×10^7 virus particles	44%	77%
7	AdC68rab.gp	5×10^6 virus particles	10%	60%

Example 8. ChAd155-RG Provides Long-term Immunogenicity to Non-human**5 Primates**

To evaluate the kinetics, breadth and longevity of the immunogenicity of ChAd155-RG in non-human primates, three groups of five cynomolgous monkeys (*Macaca fascicularis*) were treated as follows. Group 1 was immunized with ChAd155-RG 5×10^{10} viral particles IM followed by a booster dose of ChAd155-RG 5×10^{10} viral particles IM at week 48. Group 2 was immunized with ChAd155-RG 5×10^{10} viral particles IM, followed by a booster dose of RABIPUR vaccination at week 24 and a booster dose of ChAd155-RG 5×10^{10} viral particles IM at week 48. Group 3 received half of a human dose of RABIPUR administered intramuscularly and a booster dose of the same on days 7 and 21. Serum samples were collected at intervals and whole blood was collected for peripheral blood mononuclear cell (PBMC) analysis.

The immunogenicity, up to 48 weeks, induced by a single dose of ChAd155-RG was compared to a full course of RABIPUR. Boosts with either RABIPUR at week 24 or ChAd155 at week 48 were introduced to evaluate the compatibility of the two vaccines and the ability to boost medium to long term immune responses.

The neutralizing antibody titers induced by a single immunization with ChAd155-RG were compared to those induced with a full three dose course of RABIPUR. FIG. 18 shows the comparison of the neutralizing antibody responses, as measured by FAVN assay, of the monkeys immunized with recombinant ChAd155-RG (Groups 1 and 2) with those immunized with the fixed cell culture virus vaccine RABIPUR (Group 3) up to six months post-vaccination. A single dose of 10^{10} viral particles ChAd155-RG induced the same immune response as three doses of RABIPUR.

These results show that a single administration of ChAd155-RG was able to elicit neutralizing antibody titers well above the seroconversion threshold which are stable over at least 48 weeks and comparable to three doses of RABIPUR. The seroconversion induced by ChAd155-RG was rapid. All animals immunized with

5 ChAd155-RG exceeded the threshold two weeks after immunization, at which time the animals immunized with RABIPUR had already received a second dose.

Boosting the animals in Group 2 with RABIPUR at week 24 (FIG. 18 - squares) was highly effective in raising virus-neutralizing antibodies well above the peak level

10 achieved after the administration of ChAd155-RG at day 0. This demonstrates that the RABIPUR viral lysate antigen is fully able to boost immunity induced by the ChAd155-RG nucleic acid encoded antigen.

Boosting the animals in both Group 1 (FIG. 18 - triangles) and Group 2 (FIG. 18 – upside

15 down triangles) with ChAd155-RG was also highly effective. Animals immunized with ChAd155-RG, regardless of whether or not they were given an intermediate boost with RABIPUR, mounted a robust immune response to the ChAd155-RG boost at week 48. This demonstrates that the ChAd155-RG nucleic acid encoded antigen can be

20 effectively re-administered. It also demonstrates that the ChAd155-RG nucleic acid encoded antigen is effective in boosting the immune response after administration of the RABIPUR viral lysate antigen. In conclusion, Figure 18 demonstrates the compatibility of a simian adenovirus ChAd155 encoding the rabies G antigen with a conventional rabies vaccine comprising a viral lysate antigen.

25 **Example 9. ChAd155-RG Induces a Cellular Immune Response in Non-human Primates**

In addition to the humoral antibody response demonstrated in Example 8, ChAd155-RG induced a strong cellular immune response. FIG. 19 shows that a single dose of

30 ChAd155-RG induced a sustained level of rabies glycoprotein specific IFN γ -secreting T-cells in the peripheral blood of vaccinated animals, as detected by IFN γ -ELISpot assay. In contrast, cellular immune responses were below the limit of detection in the animals vaccinated with RABIPUR.

Animals in Group 1, immunized with ChAd155-RG and boosted with ChAd155-RG at

35 week 48, as described in Example 8, demonstrated that the boost re-amplified IFN γ levels. Animals in group 2, immunized with ChAd155-RG and boosted first with RABIPUR at week 24 then with ChAd155-RG at week 48 showed no increase in IFN

gamma levels in response to the RABIPUR boost but a robust response to the ChAd155-RG boost. This demonstrates that a ChAd155-RG boost can expand memory T cells in mature animals. No interleukin 4 responses were detected over the entire course of the follow up.

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Example 10. Dose Escalation Study for Safety in Humans

To evaluate the safety of ChAd155-RG in humans, a Phase I study will be initiated.

Subjects will be normal healthy adult men and women with no history of rabies vaccination, exposure to rabid animals or receipt of an adenovirus-based investigational vaccine. The study size will be large enough to determine the outcome of the primary study endpoint, safety. Standard statistical analyses will be performed, including 95% confidence intervals.

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Subjects will receive one or more intramuscular injections of ChAd155-RG; RABIPUR

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will be used as the comparator. A low dose of the ChAd155-RG vaccine will be administered and, following data review and approval, the dose will then be increased.

Subjects will be followed post-administration for systemic and local adverse events, including but not limited to fever, headache, nausea, vomiting, malaise and myalgia; and pain, tenderness, induration, redness or swelling at the injection site. Blood parameters will be examined and any additional unsolicited symptoms will be recorded.

20

The study may additionally evaluate immunogenicity by assessing vaccine-related immune responses. Outcome measures may include, but not be limited to, levels of serum neutralizing antibodies, quantification of circulating B-cell secreted antibodies and quantification of T-cell responses against a *Lyssaviral* antigen.

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CLAIMS

1. A recombinant adenovirus comprising at least one polynucleotide or polypeptide selected from the group consisting of:
 - 5 (a) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 - (b) a polynucleotide which encodes a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the
 - 10 amino acid sequence of SEQ ID NO: 1,
 - (c) a polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3,
 - (d) a polypeptide having the amino acid sequence according to SEQ ID NO: 1,
 - (e) a functional derivative of a polypeptide having the amino acid sequence according to
 - 15 SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1, and
 - (f) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; wherein the adenovirus comprises a nucleic acid sequence encoding a Lyssavirus
 - 20 antigen, wherein the nucleic acid sequence is operatively linked to one or more sequences which direct expression of said Lyssavirus antigen in a host cell.
2. A composition comprising the recombinant adenovirus according to claim 1 and a pharmaceutically acceptable excipient.
- 25 3. The adenovirus or composition according to either claim 1 or 2 comprising (a) the polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 1.
- 30 4. The adenovirus or composition according to claim 3, wherein the polynucleotide has a sequence according to SEQ ID NO: 2.
5. The adenovirus or composition according to either claim 1 or 2 comprising (b) the polynucleotide which encodes a functional derivative of a polypeptide having the amino
- 35 acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an

amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

6. The adenovirus or composition according to claim 5, wherein the functional derivative has an amino acid sequence which is at least 89.0% identical, such as at least 99.0% identical, such as at least 99.6% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

7. The adenovirus or composition according to any one of claims 3 to 6, further comprising a polynucleotide encoding:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3, wherein the functional derivative has an amino acid sequence which is at least 50.0% identical over its entire length to the amino acid sequence of SEQ ID NO: 3,

or

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

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8. The adenovirus or composition according to any one of claims 1 to 3, further comprising a polynucleotide encoding:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3, wherein the functional derivative has an amino acid sequence which is at least 50.0% identical over its entire length to the amino acid sequence of SEQ ID NO: 3, and

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

9. The adenovirus or composition according to either claim 7 or 8, wherein the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3 has an amino acid sequence which is at least 70.0%, such as at least 90.0%,

such as at least 99% identical over its entire length to the amino acid sequence of SEQ ID NO: 3.

10. The adenovirus or composition according to any one of claims 7 to 9, wherein the
5 functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 70.0%, such as at least 90.0%, such as at least 99% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

10 11. The adenovirus or composition according to either claim 1 or 2 comprising (c) the polynucleotide which encodes a polypeptide having the amino acid sequence according to SEQ ID NO: 3.

12. The adenovirus or composition according to claim 11, wherein the polynucleotide
15 has a polynucleotide sequence according to SEQ ID NO: 4.

13. The adenovirus or composition according to either claim 11 or 12, further comprising a polynucleotide encoding:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 1; or

20 (ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 1
or

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or

25 (ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

14. The adenovirus or composition according to either claim 11 or 12, further
30 comprising a polynucleotide encoding:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 1; or

(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 1

35 and

- (i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
- (ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

5

15. The adenovirus or composition according to either claim 13 or 14, wherein the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1 has an amino acid sequence which is at least 70.0% identical, such as at least 99.0% identical, such as at least 99.4% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

10

16. The adenovirus or composition according to any one of claims 13 to 15, wherein the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 70.0%, such as at least 90.0%, such as at least 99% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

15

17. The adenovirus or composition according to either claim 1 or 2 comprising (d) the polypeptide which has the amino acid sequence according to SEQ ID NO: 1.

20

18. The adenovirus or composition according to either claim 1 or 2 comprising (e) the polypeptide which is a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 80% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

25

19. The adenovirus or composition according to claim 18, wherein the functional derivative has an amino acid sequence which is at least 89.0% identical, such as at least 99.0% identical, such as at least 99.6% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

30

20. The adenovirus or composition according to any one of claims 17 to 19, further comprising:

- (i) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; or

(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3, wherein the functional derivative has an amino acid sequence which is at least 50.0% identical over its entire length to the amino acid sequence of SEQ ID NO: 3, or

- 5 (i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

10 21. The adenovirus or composition according to any one of claims 17 to 19, further comprising:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 3; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3, wherein the functional derivative has an amino acid sequence which is at
15 least 50.0% identical over its entire length to the amino acid sequence of SEQ ID NO: 3, and

- (i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or
(ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at
20 least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

22. The adenovirus or composition according to either claim 20 or 21, wherein the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 3 has an amino acid sequence which is at least 70.0%, such as at least 90.0%,
25 such as at least 99% identical over its entire length to the amino acid sequence of SEQ ID NO: 3.

23. The adenovirus or composition according to any one of claims 20 to 22, wherein the functional derivative of a polypeptide having the amino acid sequence according to
30 SEQ ID NO: 5 has an amino acid sequence which is at least 70.0%, such as at least 90.0%, such as at least 99% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

24. The adenovirus or composition according to either claim 1 or 2 comprising (f) the
35 polypeptide which has the amino acid sequence according to SEQ ID NO: 3.

25. The adenovirus or composition according to claim 24, further comprising:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 1; or

(ii) a functional derivative of a polypeptide having the amino acid sequence according to

5 SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 1

or

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or

(ii) a functional derivative of a polypeptide having the amino acid sequence according to

10 SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

26. The adenovirus or composition according claim 24, further comprising:

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 1; or

15 (ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 1

and

(i) a polypeptide having the amino acid sequence according to SEQ ID NO: 5; or

20 (ii) a functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5, wherein the functional derivative has an amino acid sequence which is at least 50% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

27. The adenovirus or composition according to either claim 25 or 26, wherein the

25 functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 1 has an amino acid sequence which is at least 70.0% identical, such as at least 99.0% identical, such as at least 99.4% identical over its entire length to the amino acid sequence of SEQ ID NO: 1.

30 28. The adenovirus or composition according to any one of claims 25 to 27, wherein

the functional derivative of a polypeptide having the amino acid sequence according to SEQ ID NO: 5 has an amino acid sequence which is at least 70.0%, such as at least 90.0%, such as at least 99% identical over its entire length to the amino acid sequence of SEQ ID NO: 5.

35

29. The adenovirus or composition according to any one of claims 1-28, wherein the polynucleotide comprises at least one of the following:

- an adenoviral 5'-end, preferably an adenoviral 5' inverted terminal repeat;
- an adenoviral E1A region, or a fragment thereof selected from among the E1A_280R
5 and E1A_243R regions;
- an adenoviral E1B or IX region, or a fragment thereof selected from among the group consisting of the E1B_19K, E1B_55K or IX regions;
- an adenoviral E2b region; or a fragment thereof selected from among the group consisting of the E2B_pTP, E2B_Polymerase and E2B_IVa2 regions;
- 10 • an adenoviral L1 region, or a fragment thereof, said fragment encoding an adenoviral protein selected from the group consisting of the L1_13.6k protein, L1_52k and L1_IIIa protein;
- an adenoviral L2 region, or a fragment thereof, said fragment encoding an adenoviral protein selected from the group consisting of the L2_penton protein,
15 L2_pVII, L2_V, and L2_pX protein;
- an adenoviral L3 region, or a fragment thereof, said fragment encoding an adenoviral protein selected from the group consisting of the L3_pVI protein, L3_hexon protein and L3_protease;
- an adenoviral E2A region;
- 20 • an adenoviral L4 region, or a fragment thereof said fragment encoding an adenoviral protein selected from the group consisting of the L4_100k protein, the L4_33k protein and protein L4_VIII;
- an adenoviral E3 region, or a fragment thereof selected from the group consisting of E3 ORF1, E3 ORF2, E3 ORF3, E3 ORF4, E3 ORF5, E3 ORF6, E3 ORF7, E3
25 ORF8, and E3 ORF9;
- an adenoviral L5 region, or a fragment thereof said fragment encoding the L5_fiber fiber protein;
- an adenoviral E4 region, or a fragment thereof selected from the group consisting of E4 ORF7, E4 ORF6, E4 ORF4, E4 ORF3, E4 ORF2, and E4 ORF1;
- 30 • an adenoviral 3'-end, preferably an adenoviral 3' inverted terminal repeat; and/or
- an adenoviral VAI or VAII RNA region, preferably an adenoviral VAI or VAII RNA region from an adenovirus other than ChAd155, more preferably from Ad5.

30. The adenovirus or composition according to claim 29, wherein the polynucleotide comprises at least one of the following:

- an adenoviral 5'-end, preferably an adenoviral 5' inverted terminal repeat;

- an adenoviral L1 region, or a fragment thereof, said fragment encoding an adenoviral protein selected from the group consisting of the L1_13.6k protein, L1_52k and L1_Illa protein;
- an adenoviral L2 region, or a fragment thereof, said fragment encoding an
5 adenoviral protein selected from the group consisting of the L2_penton protein, L2_pVII, L2_V, and L2_pX protein;
- an adenoviral L3 region, or a fragment thereof, said fragment encoding an adenoviral protein selected from the group consisting of the L3_pVI protein, L3_hexon hexon protein and L3_protease;
- 10 • an adenoviral L4 region, or a fragment thereof said fragment encoding an adenoviral protein selected from the group consisting of the L4_100k protein, the L4_33k protein and protein L4_VIII;
- an adenoviral L5 region, or a fragment thereof said fragment encoding the L5_fiber fiber protein;
- 15 • an adenoviral 3'-end, preferably an adenoviral 3' inverted terminal repeat.

31. The adenovirus or composition according to any one of claims 1-28, wherein the polynucleotide comprises an adenoviral VAI or VAII RNA region.

- 20 32. The adenovirus or composition according to claim 31, wherein the VAI or VAII RNA region is from an adenovirus other than ChAd155.

33. The adenovirus or composition according to claim 32, wherein the VAI or VAII RNA region is from Ad5.

25

34. The adenovirus or composition according to any one of claim 1-28, wherein the nucleic acid sequence encoding a Lyssavirus antigen is at least 98.6% identical, such as at least 99% identical, such as at least 99.5% identical over its entire length to SEQ ID NO: 38.

30

35. The adenovirus or composition according to any one of claims 34, wherein the nucleic acid sequence encoding a Lyssavirus antigen is identical over its entire length to SEQ ID NO: 38.

- 35 36. The adenovirus or composition according to any preceding claim, wherein the polynucleotide comprises a mutation or deletion which renders non-functional at least

one gene of a genomic region selected from the group consisting of E1A, E1B, E2A, E2B, E3 and E4.

37. The adenovirus or composition according to claim 36, wherein the polynucleotide
5 lacks at least one gene of a genomic region selected from the group consisting of E1A, E1B, E2A, E2B, E3 and/or E4.

38. The adenovirus or composition according to either claim 36 or 37, wherein the genomic regions are E1A and/or E1B.

10

39. The adenovirus or composition according to any one of claims 1 to 38, wherein the recombinant adenovirus is replication-incompetent.

40. The adenovirus or composition according to any one of claims 1 to 39, wherein
15 the nucleic acid sequence encodes an immunogen from Mokola virus, Duvenhage virus, European bat Lyssavirus, European bat Lyssavirus 2 or Australian bat Lyssavirus.

41. The adenovirus or composition according to claim 40, wherein the nucleic acid sequence encodes an immunogen from a rabies virus selected from the group
20 consisting of CVS11, CVS-N2C, Evelyn Rokitniki Abelseth (ERA), Flury, Pitman Moore and Wistar strains.

42. The adenovirus or composition according to any one of claims 1 to 41, wherein the nucleic acid sequence encodes an antigen derived from the rabies viral glycoprotein
25 (G), RNA polymerase (L), matrix protein (M), nucleoprotein (N) or phosphoprotein (P).

43. The adenovirus or composition according to any one of claims 1 to 42, wherein the nucleic acid sequence encodes a polypeptide comprising a rabies viral glycoprotein (G), RNA polymerase (L), matrix protein (M), nucleoprotein (N) and phosphoprotein (P)
30 or comprising a fragment thereof, such as fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400, at least 500 or at least 600 amino acids.

44. The adenovirus or composition according to any one of claims 1 to 42, wherein the nucleic acid sequence encodes a polypeptide comprising all or a fragment, suitably a
35 fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or

at least 500 amino acids, of the glycoprotein from Mokola virus, Duvenhage virus, European bat Lyssavirus, European bat Lyssavirus 2 or Australian bat Lyssavirus.

45. The adenovirus or composition according to any one of claims 1 to 42, wherein
5 the nucleic acid sequence encodes a polypeptide comprising all or a fragment, suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of the glycoprotein from a rabies virus selected from the group consisting of CVS11, CVS-N2C, Evelyn Rokitniki Abelseth (ERA), Flury, Pitman Moore and Wistar strains.

10

46. The adenovirus or composition according to any one of claims 1 to 42, wherein the nucleic acid sequence encodes a polypeptide comprising all or a fragment, suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of glycoproteins from more than one rabies virus.

15

47. The adenovirus or composition according to any one of claims 1 to 44, wherein the nucleic acid sequence encodes a polypeptide comprising all or a fragment, suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of glycoproteins from more than one rabies virus selected from
20 the group consisting of CVS11, CVS-N2C, Evelyn Rokitniki Abelseth (ERA), Flury, Pitman Moore and Wistar strains.

48. The adenovirus or composition according to any one of claims 1 to 42, wherein the nucleic acid sequence encodes a polypeptide comprising all of, or a fragment of,
25 suitably a fragment of at least 20, at least 50, at least 100, at least 200, at least 300, at least 400 or at least 500 amino acids, of SEQ ID NO: 37.

49. The adenovirus or composition according to claim 48, wherein the nucleic acid sequence encodes the polypeptide of SEQ ID NO: 37.

30

50. The adenovirus or composition according to any one of claims 1 to 49, wherein the nucleic acid sequence encodes a Lyssavirus antigen comprising at least one of Site I, Site IIa, Site IIb, Site III, Site IV and Site a.

51. The adenovirus or composition according to claim 50, wherein the nucleic acid sequence encodes a Lyssavirus antigen comprising at least one of Site I, Site IIa, Site IIb, Site III, Site IV and Site a corresponding to SEQ ID NO: 37.

5 52. The adenovirus or composition according to claim 51, wherein the subject is infected with a Lyssavirus.

53. The adenovirus or composition according to claim 52, wherein the Lyssavirus is a strain of rabies virus selected from one or more of RABV, ABLV, ARAV, BBLV, DUVV,
10 EBLV-1, EBLV-2, IRKV, KHUV, LBV, MOKV, SHIV, WCBV and IKOV.

54. The adenovirus or composition according to any one of claims 1 to 53, wherein the nucleic acid sequence encodes a Lyssavirus antigen which comprises a sequence which is an immunogenic fragment (such as comprising one or more T cell epitopes)
15 comprising at least one of Site I, Site IIa, Site IIb, Site III, Site IV and Site a.

55. The adenovirus or composition according to claim 54 wherein a Lyssavirus antigen comprises a sequence which is an immunogenic fragment of at least 20 amino acid residues.

20

56. The adenovirus or composition of any one of claims 1 to 55, wherein the one or more sequences which direct expression of said product in a host cell includes a sequence selected from one or more of the group consisting of transcription initiation, transcription termination, promoter and enhancer sequences.

25

57. The adenovirus or composition of claim 56, wherein the one or more sequences which direct expression of said product in a host cell includes a promoter sequence.

58 The adenovirus or composition of claim 57, wherein the promoter sequence is
30 selected from the group consisting of an internal promoter, a native promoter, RSV LTR promoter, CMV promoter, SV40 promoter, dihydrofolate reductase promoter, β -actin promoter, PGK promoter, EF1a promoter and CASI promoter.

59. The adenovirus or composition according to any one of claims 1 to 58, wherein the adenovirus has a seroprevalence of less than 10% in human subjects and preferably no seroprevalence in human subjects.
- 5 60. The adenovirus or composition according to any one of claims 1 to 59, wherein the adenovirus is capable of infecting a mammalian cell.
61. The composition according to any one of claims 2 to 60, comprising an adjuvant.
- 10 62. The adenovirus or composition according to any one of claims 1 to 61, for use as a medicament.
63. The adenovirus or composition according to claim 62, for use in the stimulation of an immune response.
- 15 64. The adenovirus or composition according to claim 63, for use as a vaccine.
65. The adenovirus or composition according to claim 63, for use in the prophylaxis, treatment or amelioration of a disease caused by a Lyssavirus.
- 20 66. Use of the adenovirus or composition according to any one of claims 1 to 61 in the manufacture of a medicament for the prophylaxis of a disease caused by a Lyssavirus infection.
- 25 67. Use of the adenovirus or composition according to any one of claims 1 to 61 in the manufacture of a medicament for use in humans for the prophylaxis of a disease caused by a Lyssavirus infection.
- 30 68. A method of inducing an immune response in a subject comprising administering the adenovirus or composition according to any one of claims 1 to 61 to the subject.
69. The method according to claim 68, wherein the subject is infected with a Lyssavirus.
- 35 70. The method according to claim 68 or 69, wherein the subject is a human.

71. The method according to claim 70, wherein the Lyssavirus is a strain of rabies virus selected from one or more of RABV, ABLV, ARAV, BBLV, DUVV, EBLV-1, EBLV-2, IRKV, KHUV, LBV, MOKV, SHIV, WCBV and IKOV

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72. A non-human simian adenovirus comprising the penton of SEQ ID NO: 3, the hexon of SEQ ID NO: 5 or the fiber of SEQ ID NO: 1 and also comprising a transgene encoding a Lyssavirus related antigen.

10 73. The non-human simian adenovirus according to claim 72, comprising the penton (SEQ ID NO: 3), hexon (SEQ ID NO: 5) and fiber (SEQ ID NO: 1) protein from ChAd155 and also comprising a transgene encoding a Lyssavirus related antigen.

15 74. The non-human simian adenovirus according to claim 72 or 73 wherein the encoded antigen comprises a sequence having at least 90% identity to SEQ ID NO. 37.

75. The non-human simian adenovirus according to any of claims 73-74 wherein the transgene comprises or consists of SEQ ID NO. 38.

20 76. The non-human simian adenovirus according to any of claims 70-75, which is a replication deficient adenovirus.

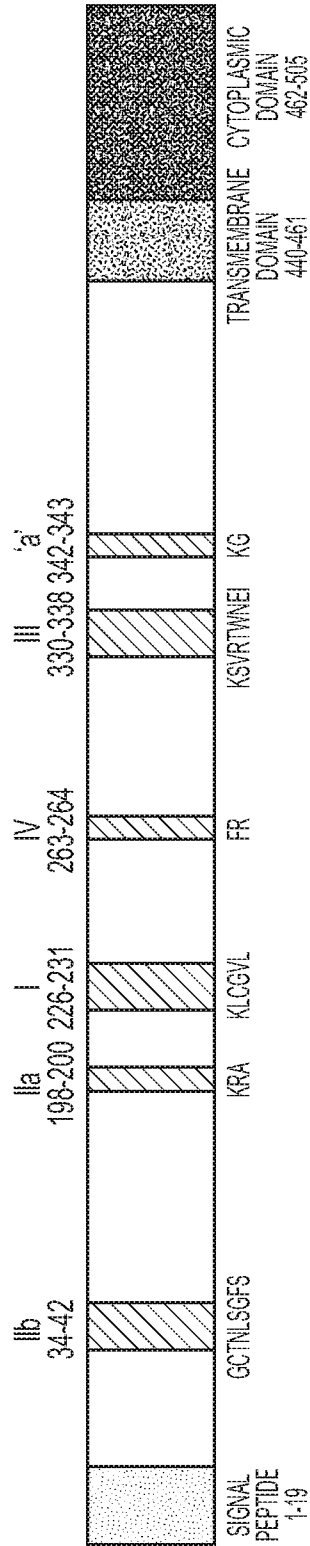
77. The non-human simian adenovirus according to claim 75 wherein the adenovirus comprises a functional inactivation (such as deletion) of the E1 gene.

25

78. The non-human simian adenovirus according to claim 75 or 77 wherein the adenovirus comprises a functional inactivation (such as deletion) of the E4 gene.

30 79. The non-human simian adenovirus according to any of claims 72 to 78 wherein the adenovirus comprises a functional inactivation (such as deletion) of the E3 gene.

80. The non-human simian adenovirus according to any one of claims 72 to 79, wherein the adenovirus comprises an Ad5E4orf6 gene substitution.



MAJOR ANTIGENIC SITES OF THE MEDOID G PROTEIN AND THEIR AMINOACID POSITION. ANTIGENIC SITE I HARBORS BOTH CONFORMATIONAL AND LINEAR EPITOPES AND IS LOCATED AT POSITION 226-231. ANTIGENIC SITE II IS A DISCONTINUOUS CONFORMATIONAL EPITOPE AT RESIDUES 34-42 (IIB) AND 198-200 (IIA). ANTIGENIC SITE III IS A CONTINUOUS CONFORMATIONAL EPITOPE AT RESIDUES 330-338. ANTIGENIC SITE IV IS LOCATED AT POSITION 263-264. MINOR SITE 'A' IS LOCATED AT POSITION 342-343.

FIG. 1

ChAd3	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>SG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQDIT <u>TA</u> SPPLKATKTNLSIE
PanAd3	(1)	MKR <u>A</u> KTSD <u>E</u> TFNFPVYPYDTE <u>NG</u> PPSPVFLTPPEVSPDGFQESPPGVLSIRISEPLVTSHGMLAIAKMG <u>NG</u> LSLDDAGNLTSQD <u>VT</u> TT <u>VT</u> PPPLKATKTNLSIQ
ChAd17	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>SG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQDITS <u>TT</u> PPPLKATKTNLSIE
ChAd19	(1)	MKR <u>TK</u> TSDK <u>SE</u> FNFPVYPYDTE <u>NG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQD <u>VT</u> TTTTPPPLKATKTNLSIE
ChAd24	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>NG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQD <u>VT</u> TTTTPPPLKATKTNLSIE
ChAd155	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>SG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQDIT <u>TA</u> SPPLKATKTNLSIE
ChAd11	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>NG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQD <u>VT</u> TTTTPPPLKATKTNLSIE
ChAd20	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>SG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQDIT <u>TA</u> SPPLKATKTNLSIE
ChAd31	(1)	MKR <u>TK</u> TSDE <u>SE</u> FNFPVYPYDTE <u>SG</u> PPSPVFLTPPEVSPDGFQESPPGVLSINIAEPLVTSHGMLAIAKMG <u>SG</u> LSLDDAGNLTSQDIT <u>TA</u> SPPLKATKTNLSIE
PanAd1	(1)	MKR <u>A</u> KTSD <u>E</u> TFNFPVYPYDTE <u>NG</u> PPSPVFLTPPEVSPDGFQESPPGVLSIRISEPLVTSHGMLAIAKMG <u>NG</u> LSLDDAGNLTSQD <u>VT</u> TT <u>VT</u> PPPLKATKTNLSIQ
PanAd2	(1)	MKR <u>A</u> KTSD <u>E</u> TFNFPVYPYDTE <u>NG</u> PPSPVFLTPPEVSPDGFQESPPGVLSIRISEPLVTSHGMLAIAKMG <u>NG</u> LSLDDAGNLTSQD <u>VT</u> TT <u>VT</u> PPPLKATKTNLSIQ
ChAd3	(101)	TSS <u>P</u> LT <u>V</u> STSGALIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
PanAd3	(101)	TS <u>A</u> PLTVS-SGSLIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>G</u> LATQ <u>G</u> PLTVSEKIL <u>T</u> LQTSAPLTAADSSLT <u>VG</u> TPPP <u>I</u> SV <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd17	(101)	TSS <u>P</u> LT <u>V</u> STSGALIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>VS</u> STPP <u>I</u> SV <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd19	(101)	TS <u>A</u> PLTVSTSGALILAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> SV <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd24	(101)	TS <u>A</u> PLTVSTSGALILAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd155	(101)	TSS <u>P</u> LT <u>V</u> STSGALIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd11	(101)	TS <u>A</u> PLTVSTSGALILAAAA <u>V</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>IS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd20	(101)	TSS <u>P</u> LT <u>V</u> STSGALIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>T</u> IAK <u>G</u> PLTVSEKIALQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
ChAd31	(101)	TSS <u>P</u> LT <u>V</u> STSGALIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>G</u> LATQ <u>G</u> PLTVSEKIL <u>T</u> LQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
PanAd1	(101)	TS <u>A</u> PLTVS-SGSLIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>G</u> LATQ <u>G</u> PLTVSEKIL <u>T</u> LQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH
PanAd2	(101)	TS <u>A</u> PLTVS-SGSLIVAAAA <u>A</u> PLAVAGTSLTMQSEAPLTVQDAKL <u>G</u> LATQ <u>G</u> PLTVSEKIL <u>T</u> LQTSAPLTAADSSLT <u>VS</u> ATPP <u>I</u> NT <u>S</u> SGS <u>G</u> IGIDMEDP <u>M</u> YTH

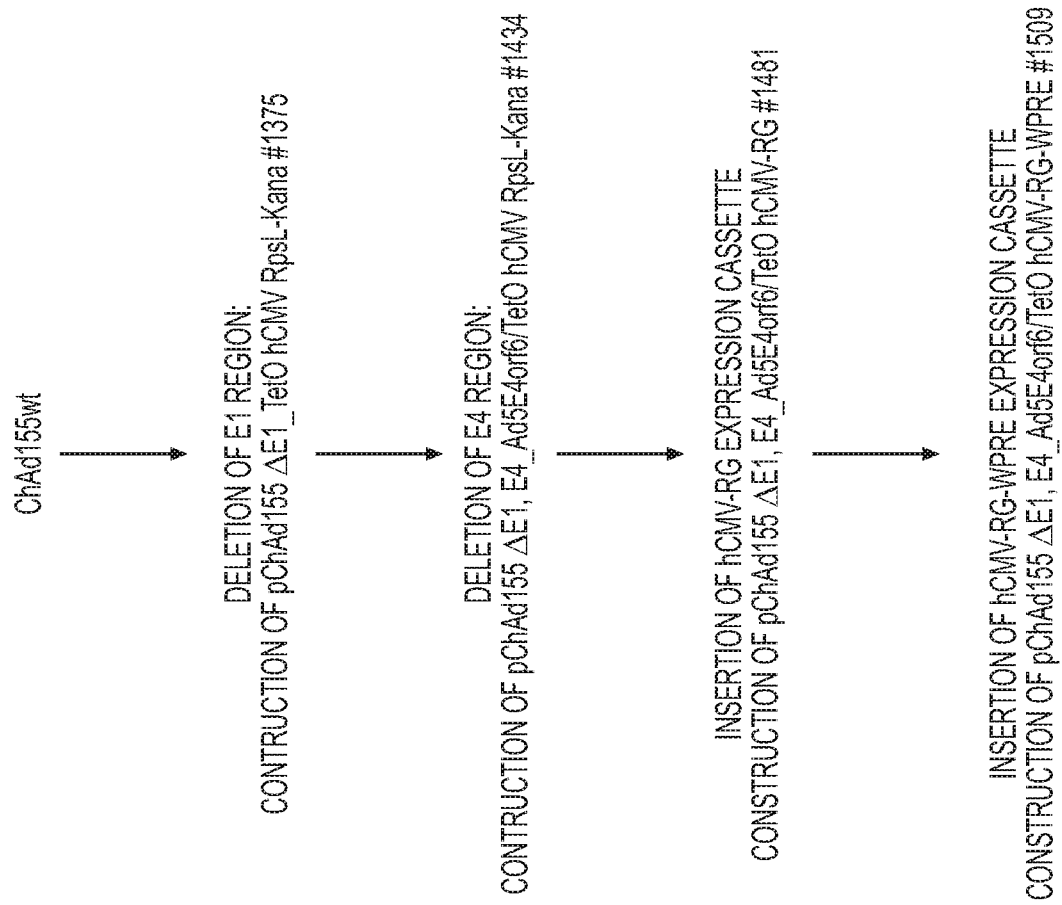
FIG. 2A

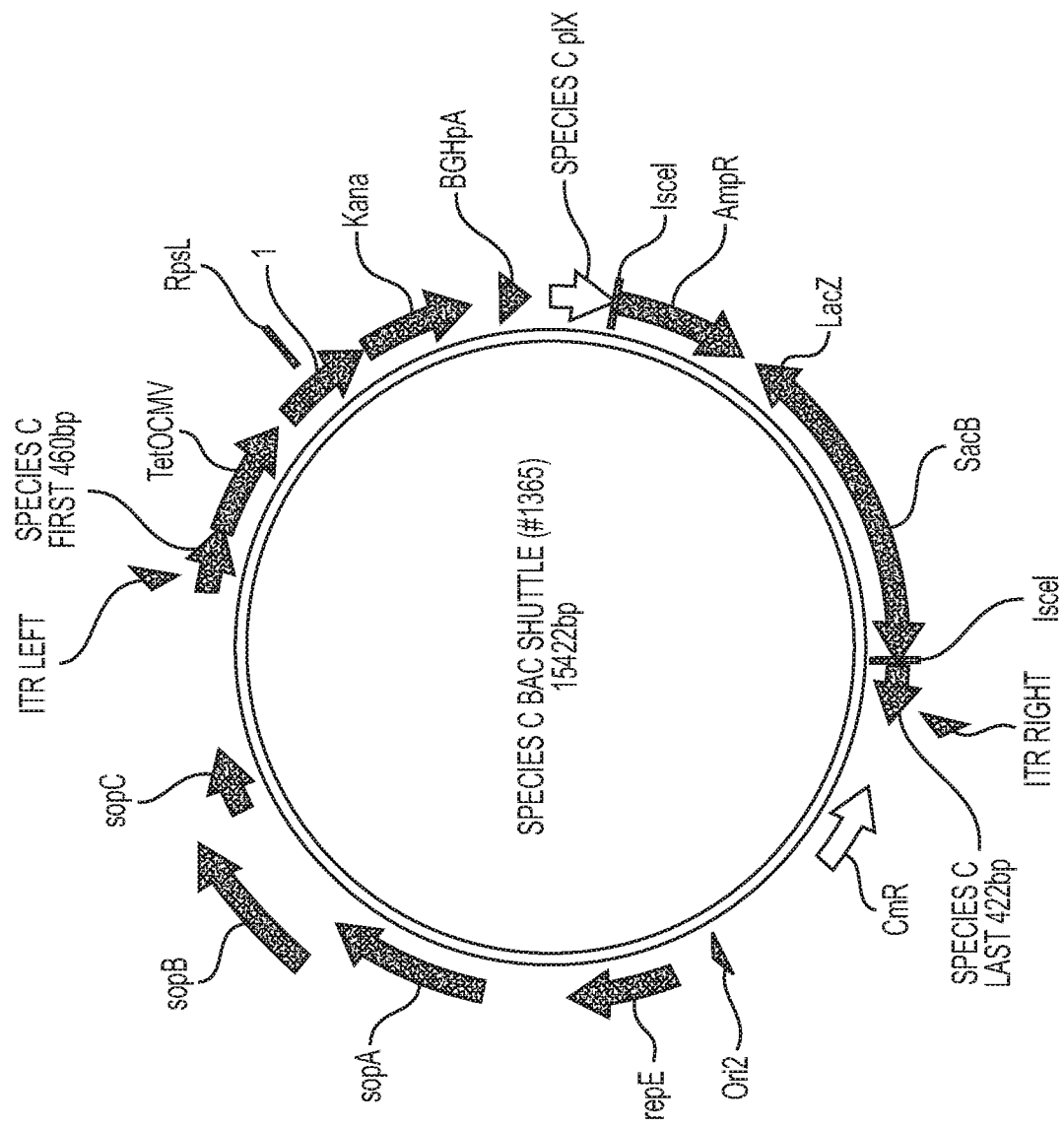
ChAd3	(201)	DGKIGIPIGGPLRVVDSLH TL TVVTGNGLTVDNNAIQTRVTCALGYDTSGNLQ LR AAGCMRIDANGQ LI LNWAYPFDQNNLSRLGQGPLY NI DHNL
PanAd3	(200)	DEKLGIPIGGPLQVDSLH TL TVVTGNGITVANNALQTRVACALGYDSSGNLELRAGCMRINTGQ LI IDVAYPFDQNNLSRLGQGPLY NI HNHL
ChAd17	(201)	DEKLGIPIGGPLRVVDSLH TL TVVTGNGLTVDNNAIQTRVTCALGYDTSGNLQ LR AAGCMRIDANGQ LI IDVAYPFDQNNLSRLGQGPLY NI DHNL
ChAd19	(201)	DEKLGIPIGGPLRVVDSLH TL TVVTGNGIAVDNNAIQTRVTCALGYDTSGNLQ LR AAGCMRIDANGQ LI IDVAYPFDQNNLSRLGQGPLY NI DHNL
ChAd24	(201)	DEKLGIPIGGPLRVVDSLH TL TVVTGNGIAVDNNAIQTRVTCALGYDTSGNLQ LR AAGCMRIDANGQ LI IDVAYPFDQNNLSRLGQGPLY NI DHNL
ChAd155	(201)	NGKLGINFCAPIHVVDLSINALTVVTGQGLTINGTALQTRVSCALWYDTSNLELRAGCMRV D ANGQ LI IDVAYPFDQNNLSRLGQGPLY NI SAHNL
ChAd11	(201)	NGKLGINFCAPIHVVDLSINALTVVTGQGLTINGTALQTRVSCALWYDSSGNLELRAGCMRV D ANGK LI IDVAYPFDQNNLSRLGQGPLY NI SAHNL
ChAd20	(201)	NGKLGINFCAPIHVVDLSINALTVVTGQGLTINGTALQTRVSCALWYDTSNLELRAGCMRV D ANGQ LI IDVAYPFDQNNLSRLGQGPLY NI SAHNL
ChAd31	(201)	NGKLGINFCAPIHVVDLSINALTVVTGQGLTINGTALQTRVSCALWYDTSNLELRAGCMRV D ANGQ LI IDVAYPFDQNNLSRLGQGPLY NI SAHNL
PanAd1	(200)	NGKIALNIGAPIHVVDLINALTVVTGQGLTINGRALQTRVTCALSYDT EG NIQ LQ AGGCMRIDNNQ LI LNWAYPFDQNNLSRLGQGPLY NI SAHNL
PanAd2	(200)	NGKIALNIGAPIHVVDLINALTVVTGQGLTINGRALQTRVTCALSYDT EG NIQ LQ AGGCMRIDNNQ LI LNWAYPFDQNNLSRLGQGPLY NI SAHNL
ChAd3	(301)	LN C NRGLTTTT NI NIKKLE T -----KISSGLDYDINGAVI I KIGTGLSFDNTGALT TV GN TG DDKLT LT WT
PanAd3	(300)	LN C NRGLTTTT SS NI TK LE T -----KIDSGLDYVANGAI I AKIGTGLTFDNTGALT TV GN TG DDKLT LT WT
ChAd17	(301)	LN C NRGLTTTT NI NIKKLE T -----KISSGLDYDINGAVI I KIGTGLSFDNTGALT TV GN TG DDKLT LT WT
ChAd19	(301)	LN C NRGLTTTT NI NIKKLE T -----KISSGLDYDINGAVI I KIGTGVSF ST CALS SV GN TG DDKLT LT WT
ChAd24	(301)	LN C NRGLTTTT NI NIKKLE T -----KISSGLDYDINGAVI I KIGTGVSF ST CALS SV GN TG DDKLT LT WT
ChAd155	(301)	LVN Y NRGLYLF TS GN TK KLEVN IK TAKGLYDPTAIAINAGDGLQ F DGSDTNPLKT IK GLGIDYDSRAI I AKLCTGLSFDNTGALT TV GN KN DDKLT LT WT
ChAd11	(301)	LVN Y NRGLYLF TS GN TK KLEVN IK TAKGLYDPTAIAINAGDGLQ F DGSDTNPLKT IK GLGIDYDSRAI I AKLCTGLSFDNTGALT TV GN KN DDKLT LT WT
ChAd20	(301)	LVN Y NRGLYLF TS GN TK KLEVN IK TAKGLYDPTAIAINAGDGLQ F DGSDTNPLKT IK GLGIDYDSRAI I AKLCTGLSFDNTGALT TV GN KN DDKLT LT WT
ChAd31	(301)	LVN Y NRGLYLF TS GN TK KLEVN IK TAKGLYDPTAIAINAGDGLQ F DGSDTNPLKT IK GLGIDYDSRAI I AKLCTGLSFDNTGALT TV GN KN DDKLT LT WT
PanAd1	(300)	LVN Y NRGLYLF TS GN TK KLEVN IK TAKGLFYDGTAAIAINAGDGLQ F DGSDTNPLQ TI IKGLGLEIYDSKAI I TKLCTGLSFDNTGALT TV GN KN DDKLT LT WT
PanAd2	(300)	LVN Y NRGLYLF TS GN TK KLEVN IK TAKGLFYDGTAAIAINAGDGLQ F DGSDTNPLQ TI IKGLGLEIYDSKAI I TKLCTGLSFDNTGALT TV GN KN DDKLT LT WT

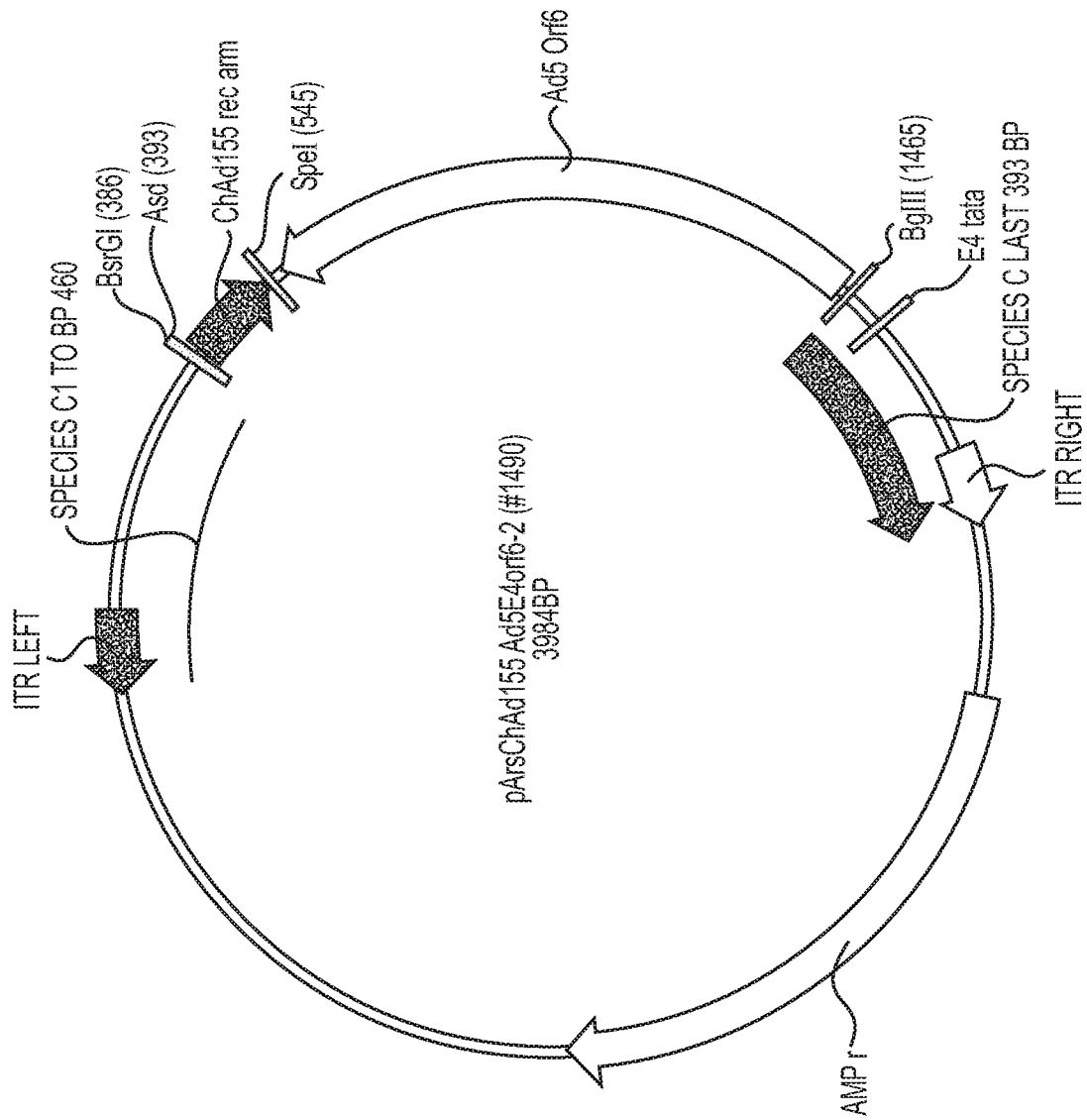
FIG. 2B

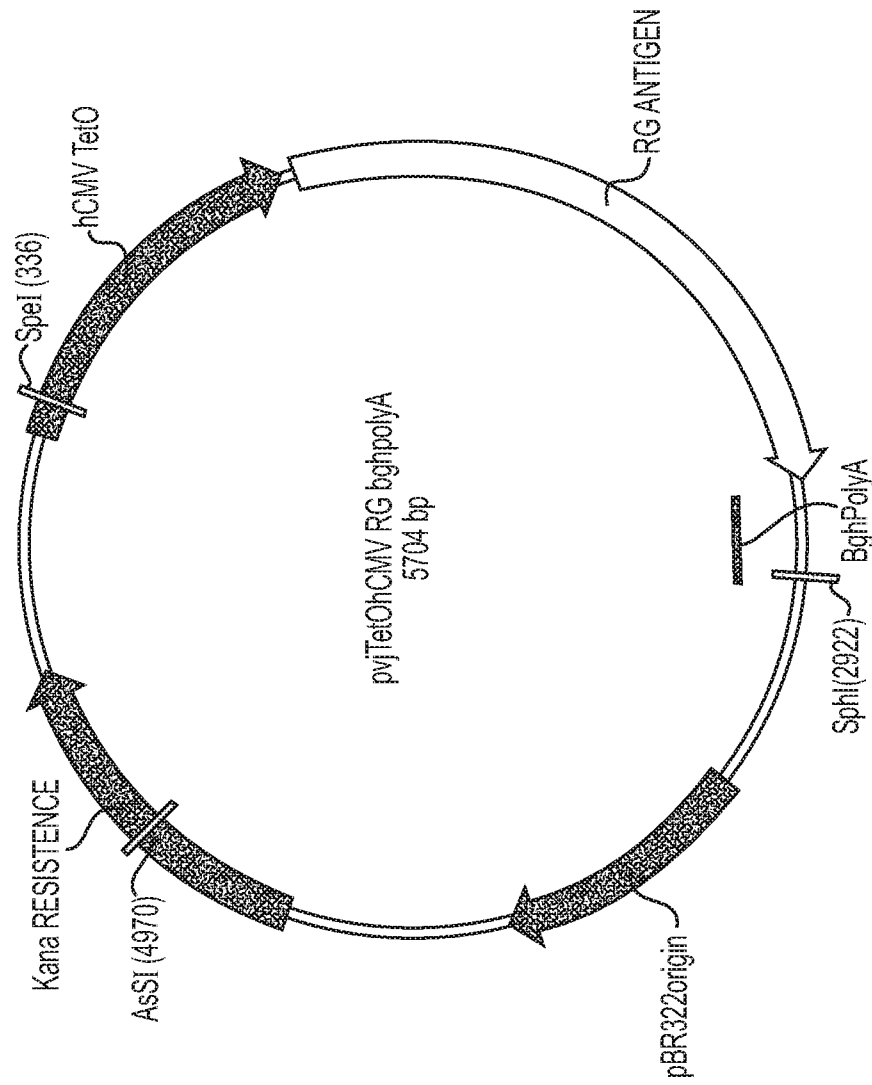
ChAd3 (365) TPDPSPNCRIHSKDCKFTLVLTKCGSQILLASVAAIAVSCNIASTIGTVASVTIFLRFDQNGVIMENSSLDQYWNFRNCNSTNAPYTNNAVGFMPNLAA
 PanAd3 (364) TPDPSPNCRIHADKCKFTLVLTKCGSQILLASVAAIAVSCNLSSTIGTVSSVTIFLRFDQNGVIMENSSLDKEYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd17 (365) TPDPSPNCRIHSKDCKFTLVLTKCGSQILLASVAAIAVSCNIASTIGTVASVTIFLRFDQNGVIMENSSLDQYWNFRNCNSTNAPYTNNAVGFMPNLAA
 ChAd19 (365) TPDPSPNCRIHSKDCKFTLVLTKCGSQILLASVAAIAVSCNIASTIGTVSSVTIFLRFDQNGVIMENSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd24 (365) TPDPSPNCRIHSKDCKFTLVLTKCGSQILLASVAAIAVSCNIASTIGTVSSVTIFLRFDQNGVIMENSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd155 (401) TPDPSPNCRIYSEKDAKFTLVLTKCGSQILLASVSVSVKGSIAPISTGTVTSAQIVLRFDENGVLISNSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd11 (401) TPDPSPNCRIYSEKDAKFTLVLTKCGSQILLASVSVSVKGSIAPISTGTVTSAQIVLRFDENGVLISNSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd20 (401) TPDPSPNCRIYSEKDAKFTLVLTKCGSQILLASVSVSVKGSIAPISTGTVTSAQIVLRFDENGVLISNSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd31 (401) TPDPSPNCRIYSEKDAKFTLVLTKCGSQILLASVSVSVKGSIAPISTGTVTSAQIVLRFDENGVLISNSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 PanAd1 (400) TPDPSPNCRIYSEKDAKFTLVLTKCGSQILLASVSVSVKGSIAPISTGTVTSAQIVLRFDENGVLISNSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 PanAd2 (400) TPDPSPNCRIYSEKDAKFTLVLTKCGSQILLASVSVSVKGSIAPISTGTVTSAQIVLRFDENGVLISNSSLDQYWNFRNCNSTNATPYTNNAVGFMPNLAA
 ChAd3 (465) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNESSETSQSHYSMSFTWAWESGQVATEFATNSTFTFSYIAEQ
 PanAd3 (464) YPKTQSQTAANNIVSEVYLHGDKSKPMTLITILNGINNESSETSQSHYSMSFTWAWESGQVATEFATNSTFTFSYIAEQ
 ChAd17 (465) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNESSETSQSHYSMSFTWAWESGQVATEFATNSTFTFSYIAEQ
 ChAd19 (465) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNESSETSQSHYSMSFTWAWESGQVATEFATNSTFTFSYIAEQ
 ChAd24 (465) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNESSETSQSHYSMSFTWAWESGQVATEFATNSTFTFSYIAEQ
 ChAd155 (501) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNETG-DATVSTYSMSFSNNWNGSNYINETFQTNSTFTFSYIAEQ
 ChAd11 (501) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNETG-DATVSTYSMSFSNNWNGSNYINETFQTNSTFTFSYIAEQ
 ChAd20 (501) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNETG-DATVSTYSMSFSNNWNGSNYINETFQTNSTFTFSYIAEQ
 ChAd31 (501) YPKTQSQTAANNIVSQVYVINGDKSKPMTLITILNGINNETG-DATVSTYSMSFSNNWNGSNYINETFQTNSTFTFSYIAEQ
 PanAd1 (500) YPKTQSQTAANNIVSQVYVINGDKTKPMTLITILNGINNETG-DATVSTYSMSFSNNWNGSNYINETFQTNSTFTFSYIAEQ
 PanAd2 (500) YPKTQSQTAANNIVSQVYVINGDKTKPMTLITILNGINNETG-DATVSTYSMSFSNNWNGSNYINETFQTNSTFTFSYIAEQ

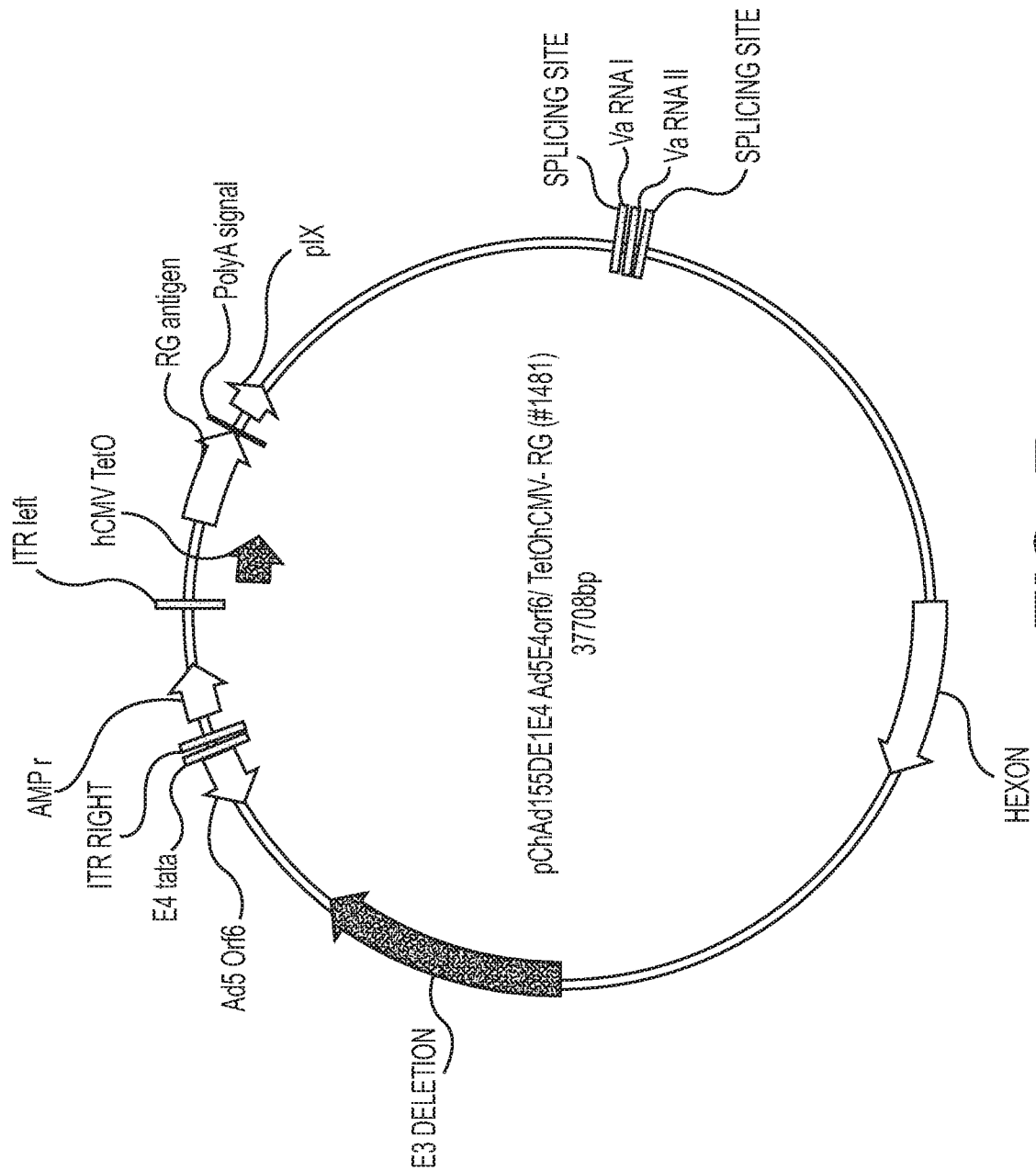
FIG. 2C

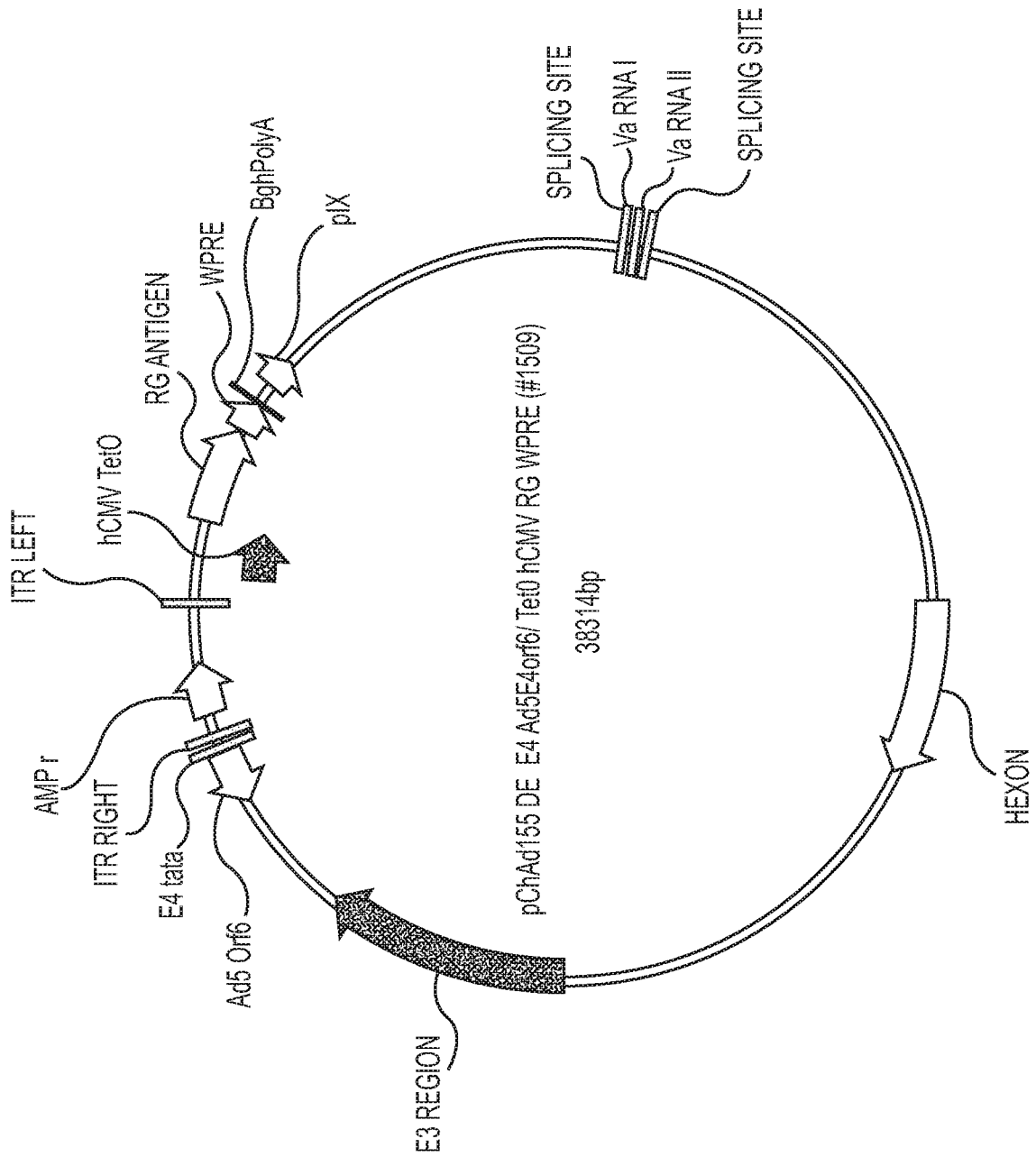
**FIG. 3**

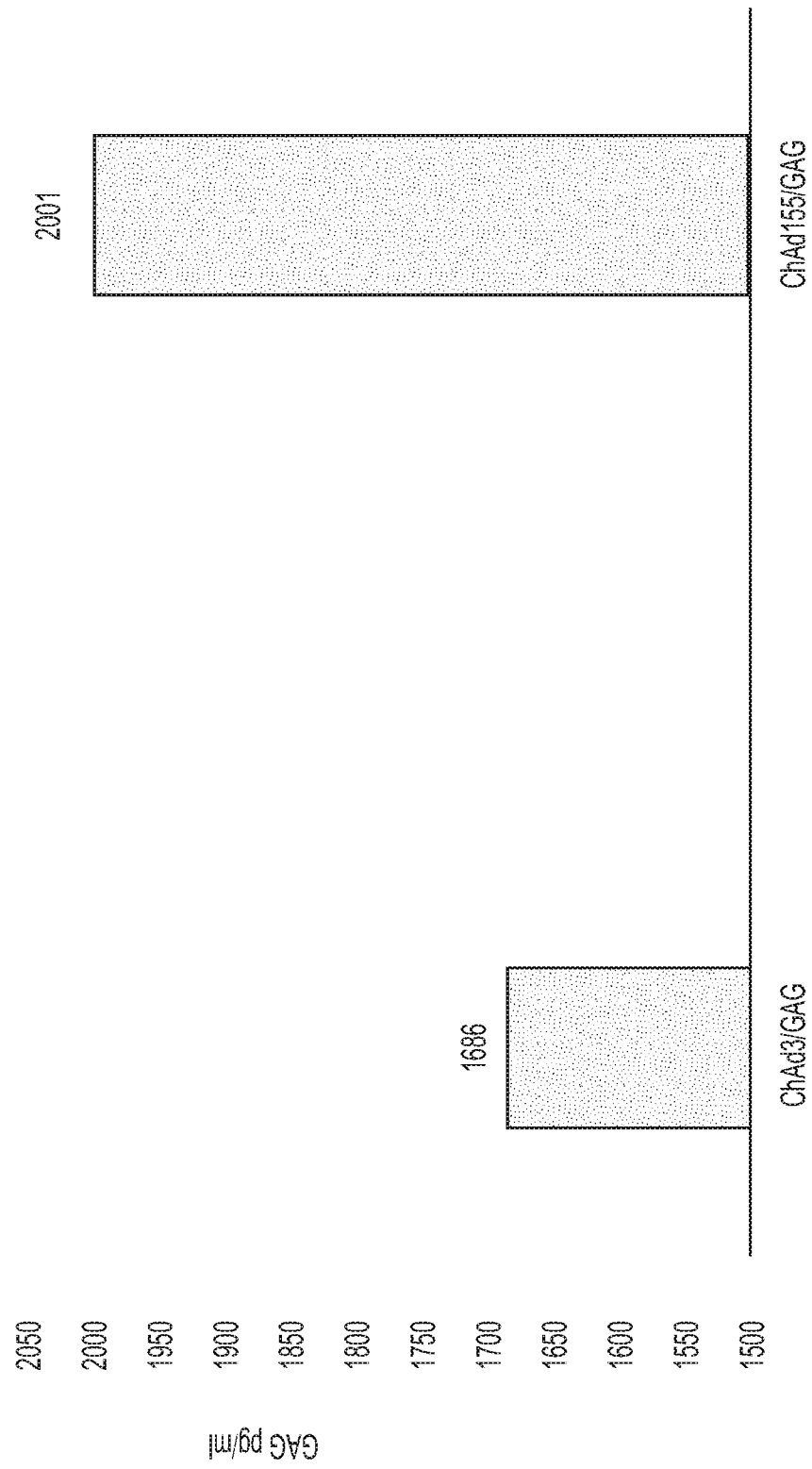
**FIG. 4**

**FIG. 5**

**FIG. 6**

**FIG. 7**

**FIG. 8**

**FIG. 9**

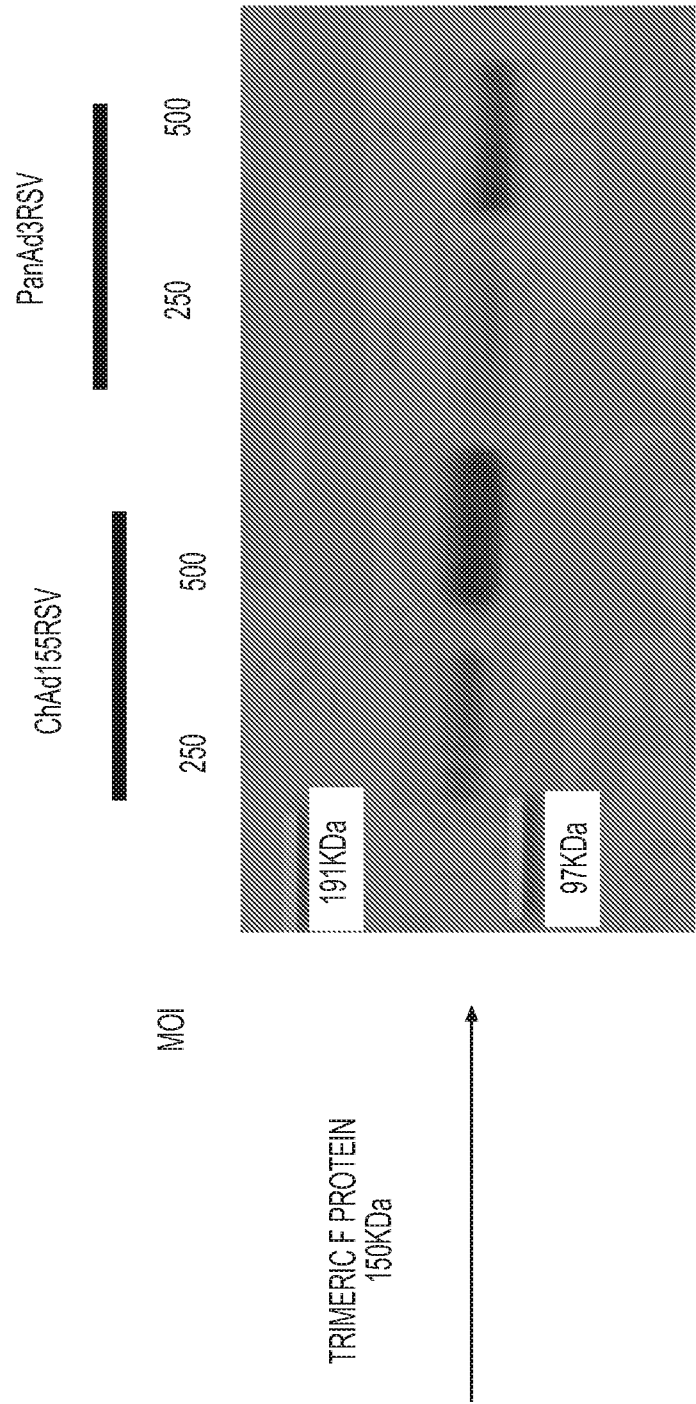
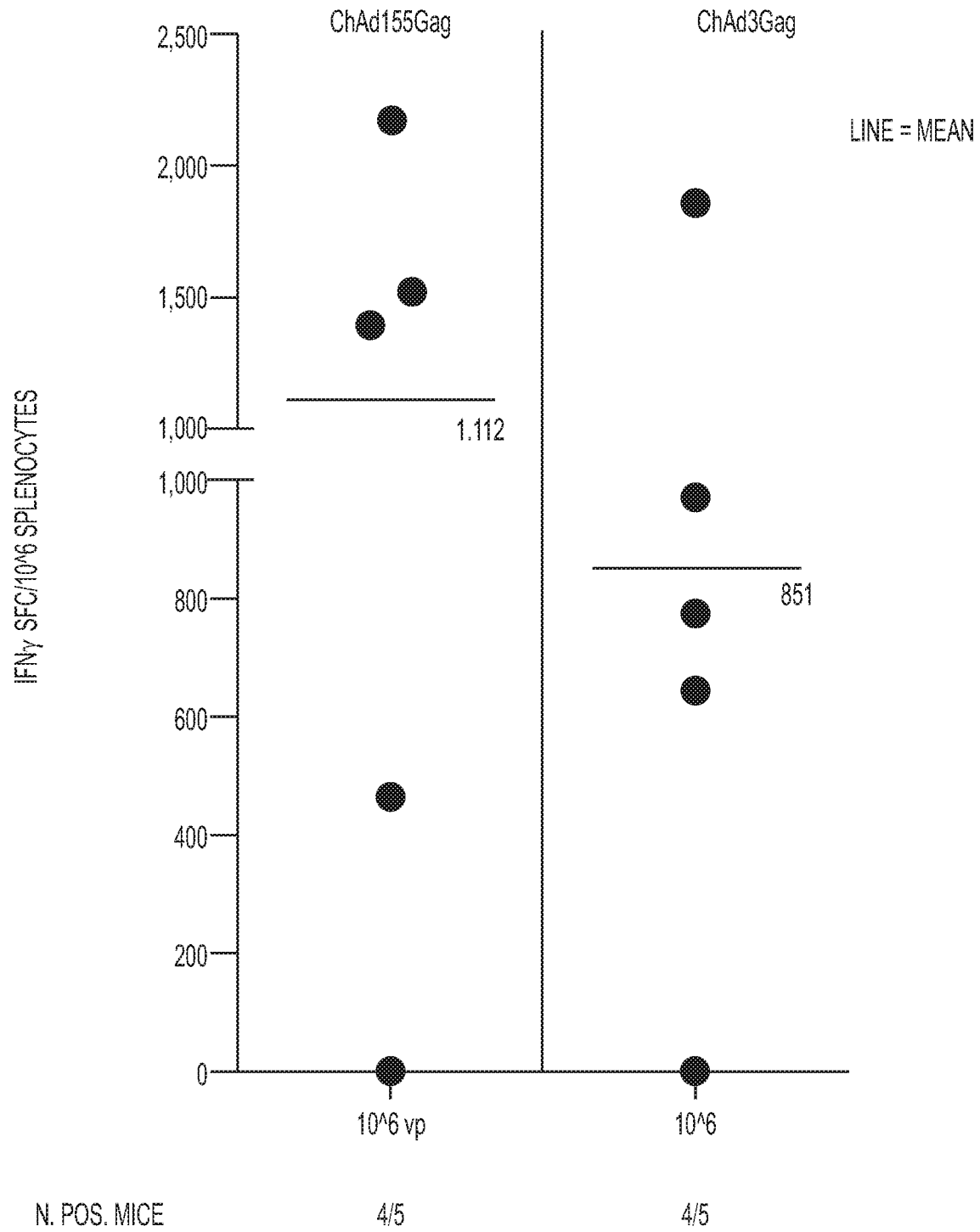
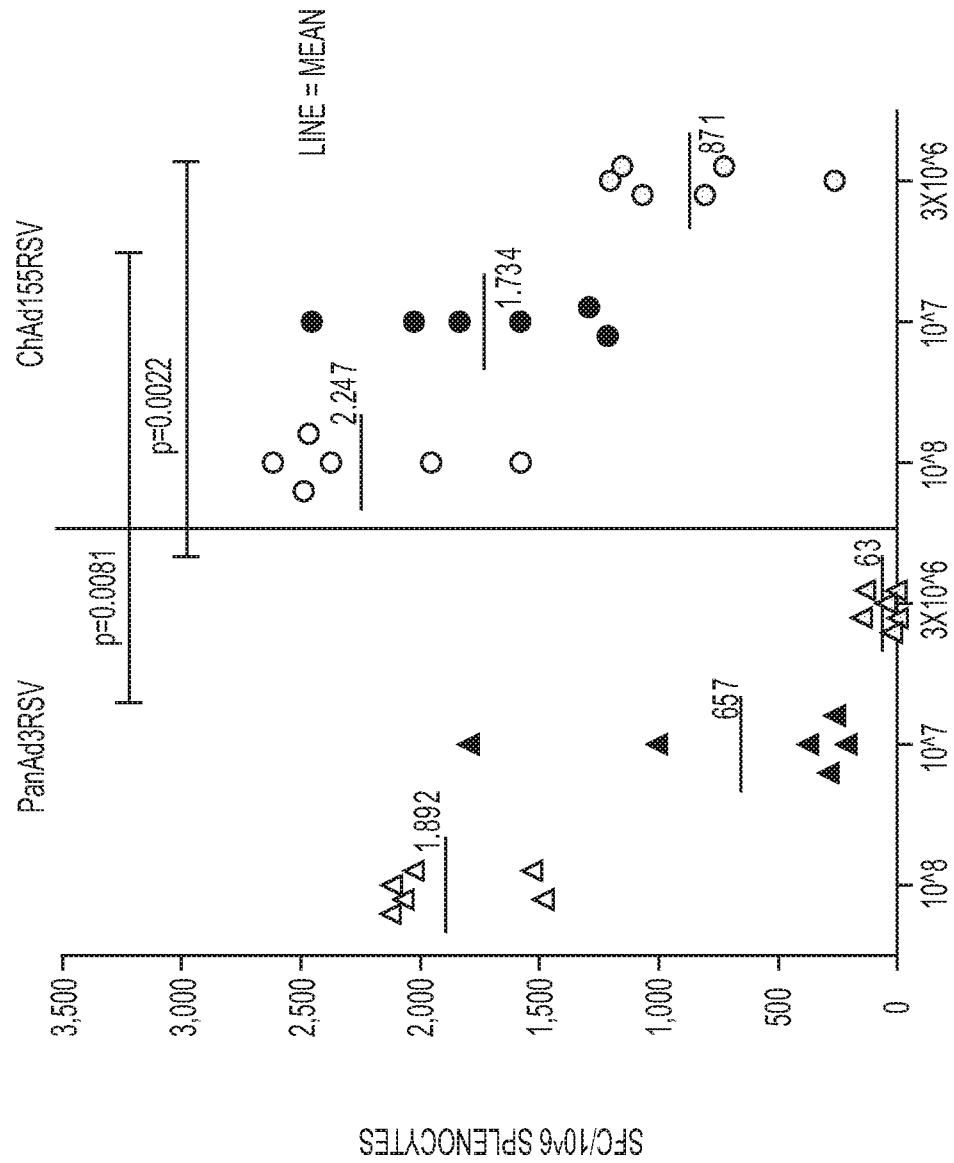


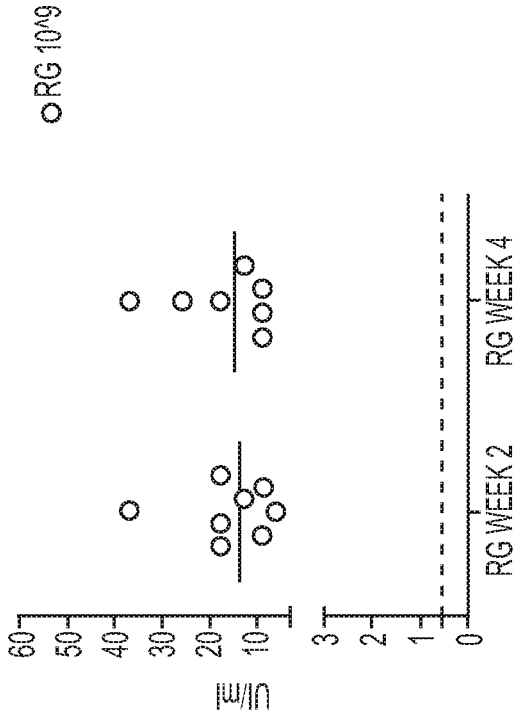
FIG. 10

**FIG. 11**

**FIG. 12**

SINGLE IM ADMINISTRATION OF 10⁹VP OF ChAd155-RG IN CD1 MICE

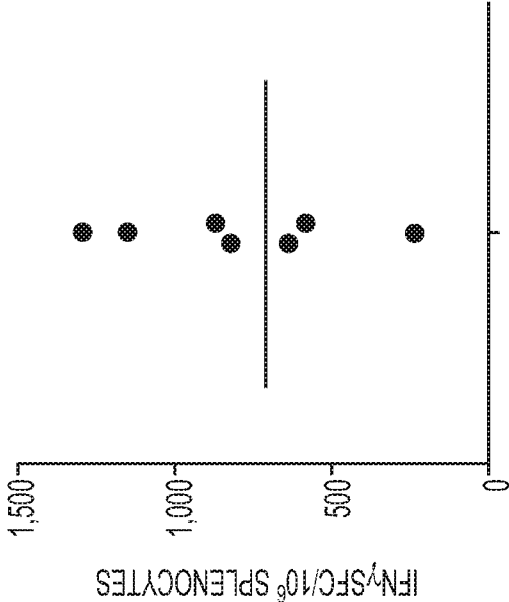
FUNTCTIONAL ANTIBODIES



ANTI-RABIES FAVN : FLURESCENT ANTIBODY VIRUS
NEUTRALISATION

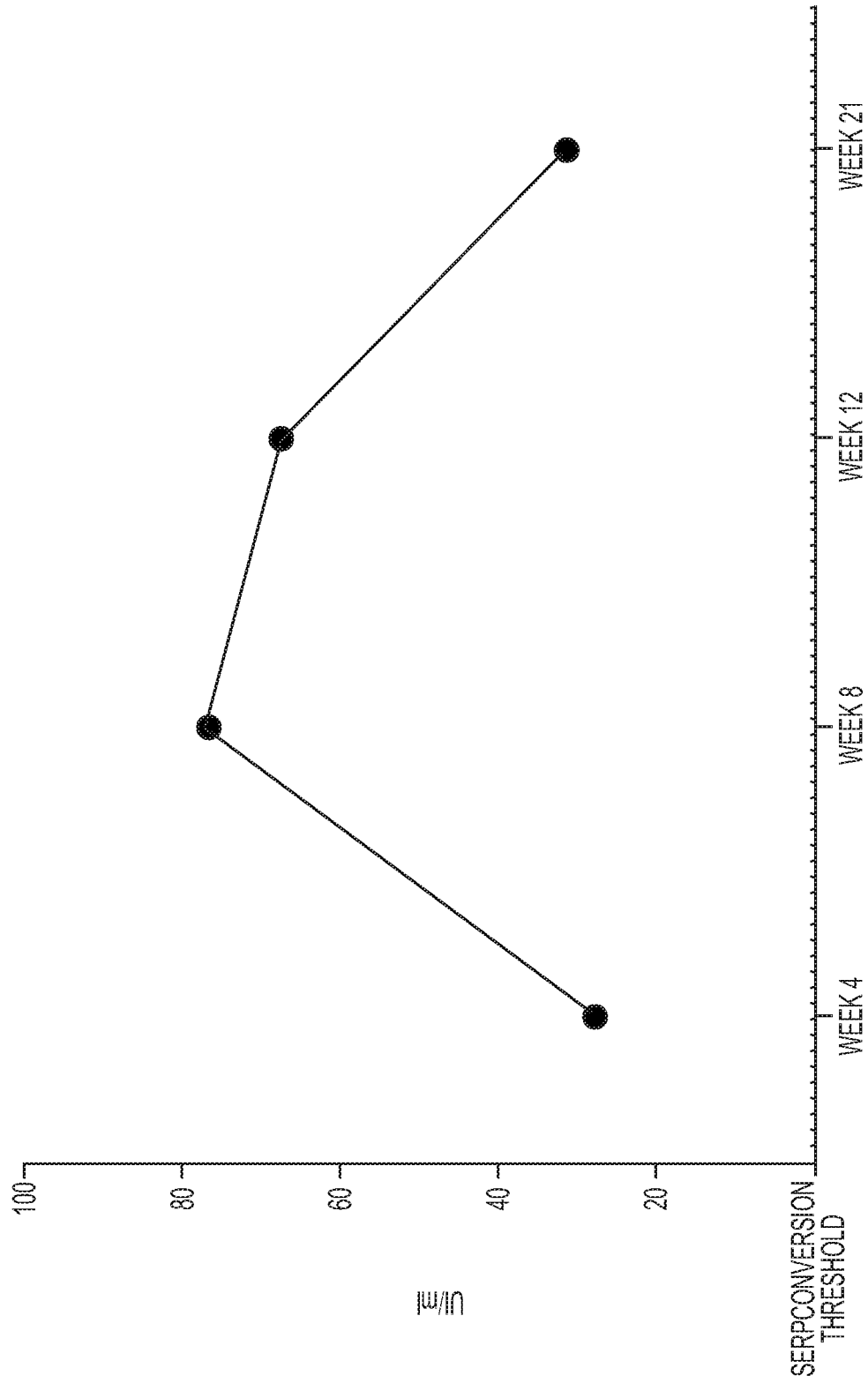
FIG. 13A

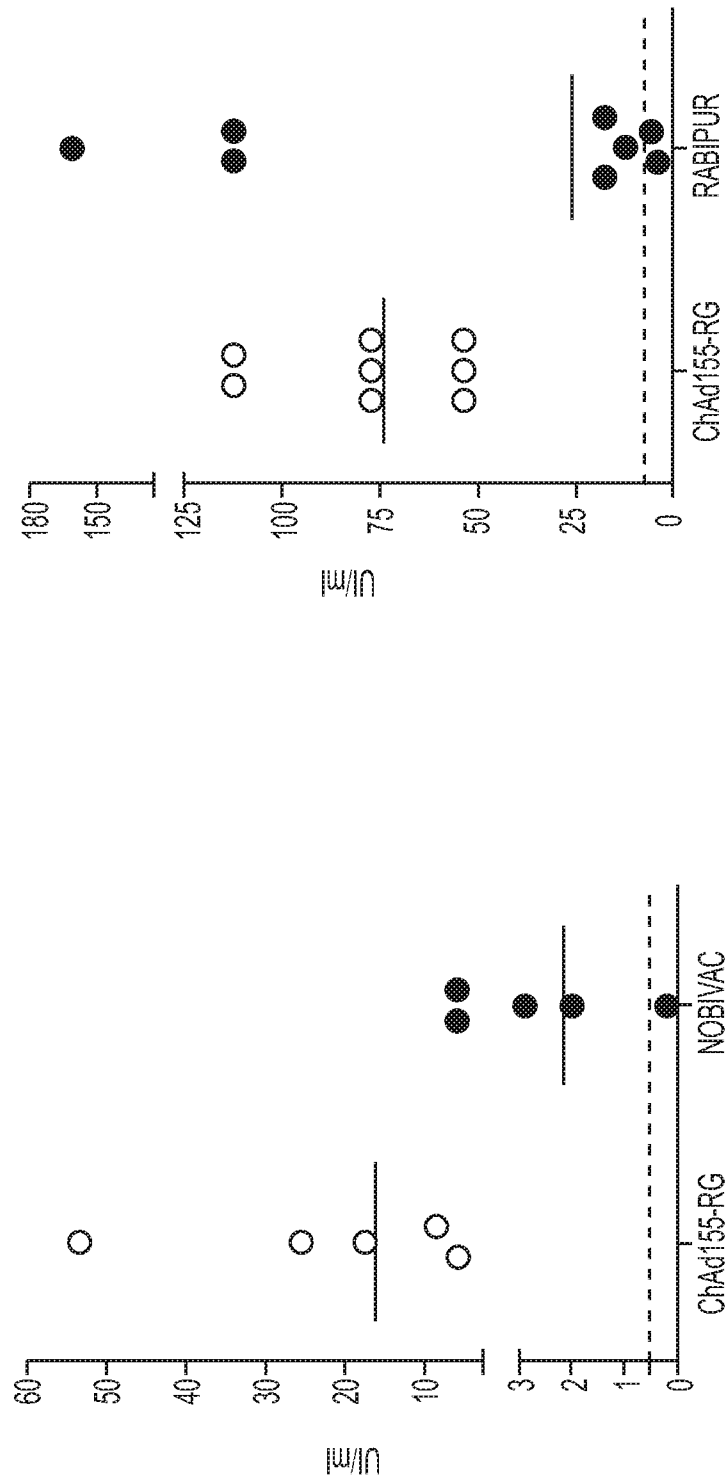
CIRCULATING RABIES-SPECIFIC T-CELLS
SPLEEN



IFN-γ ELISpot ON OVERLAPPING PEPTIDES SPANNING
THE RABIES G SEQUENCE

FIG. 13B

**FIG. 14**



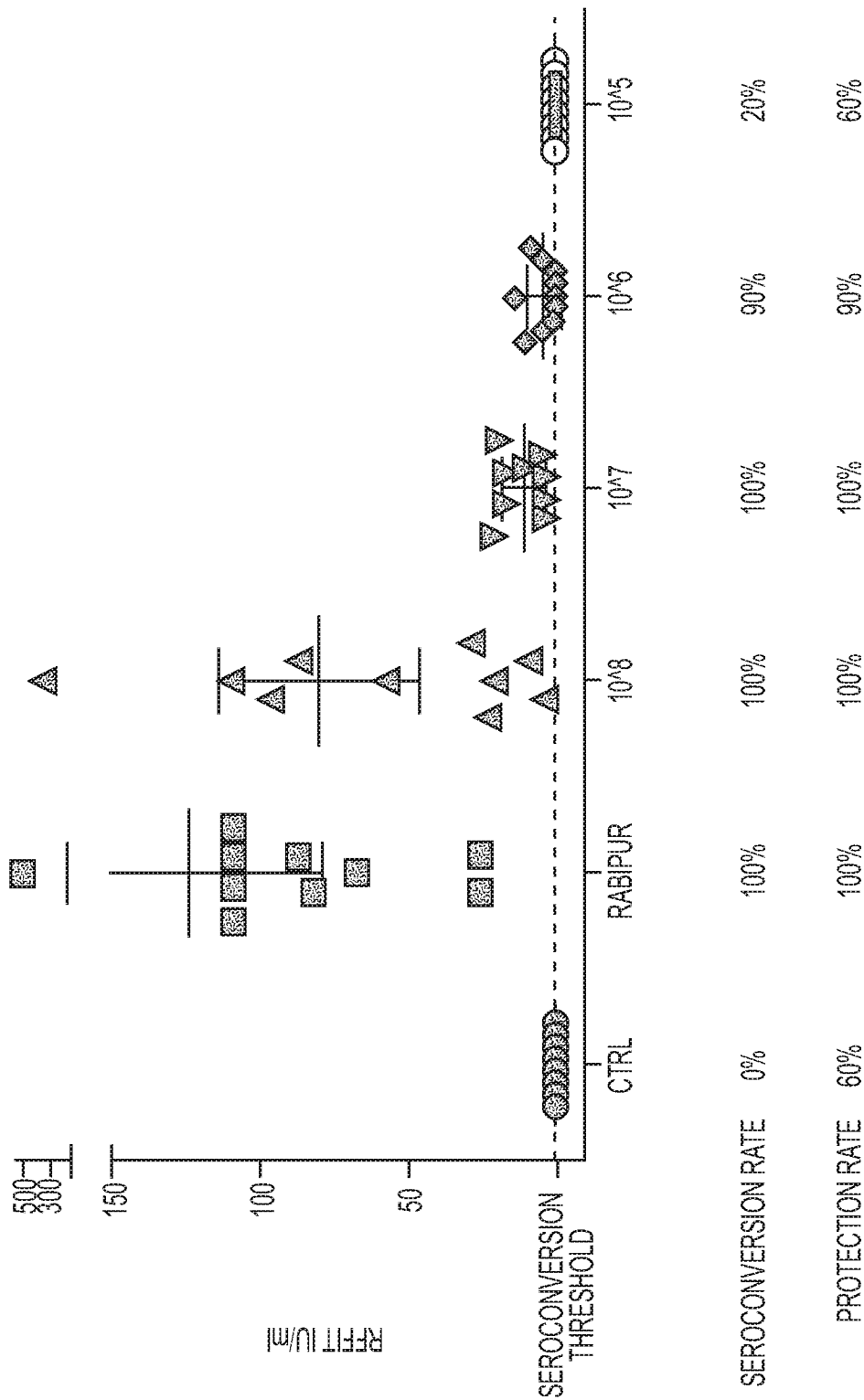


FIG. 16

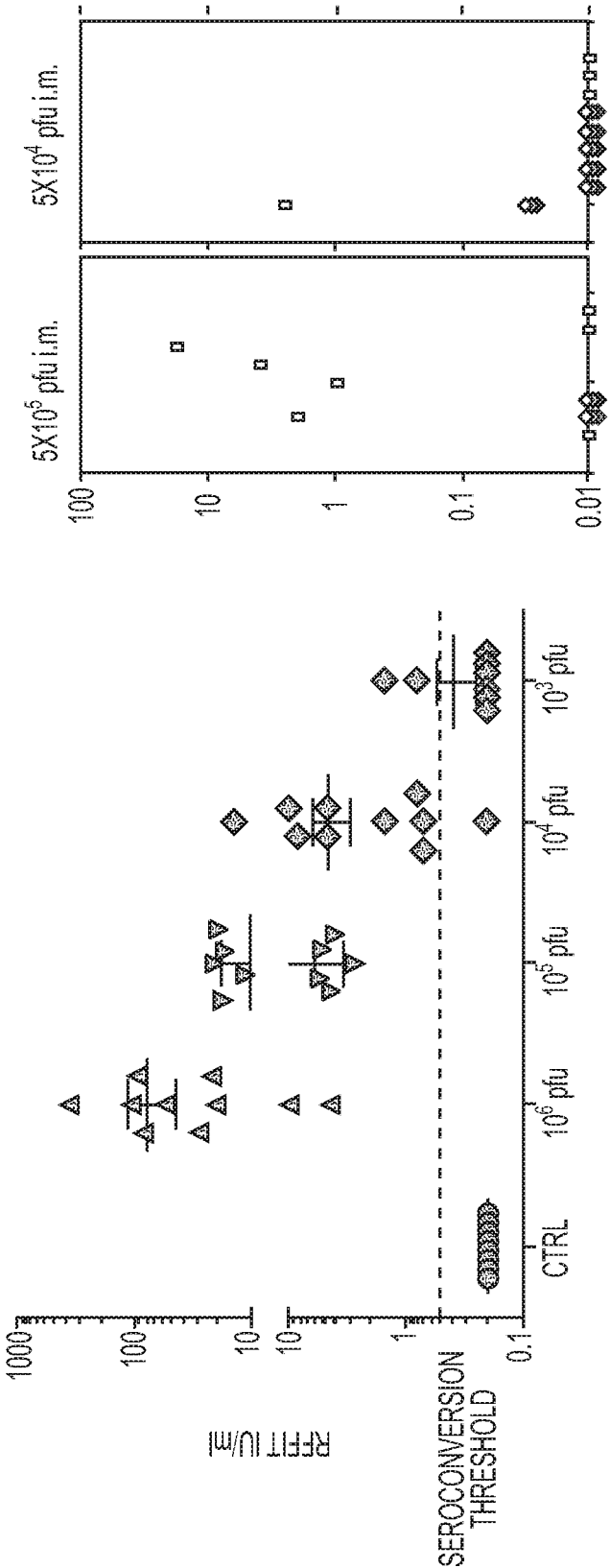
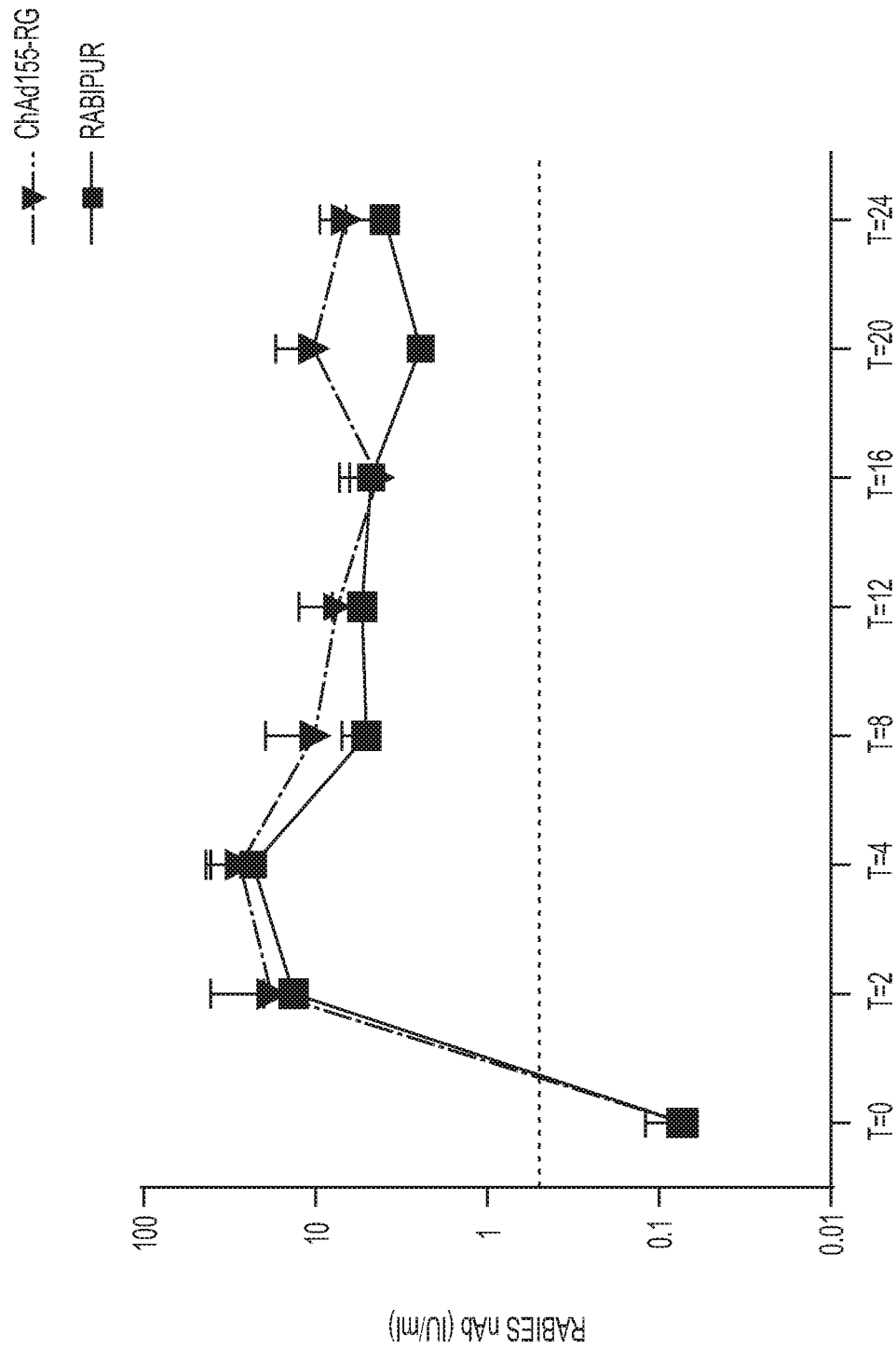
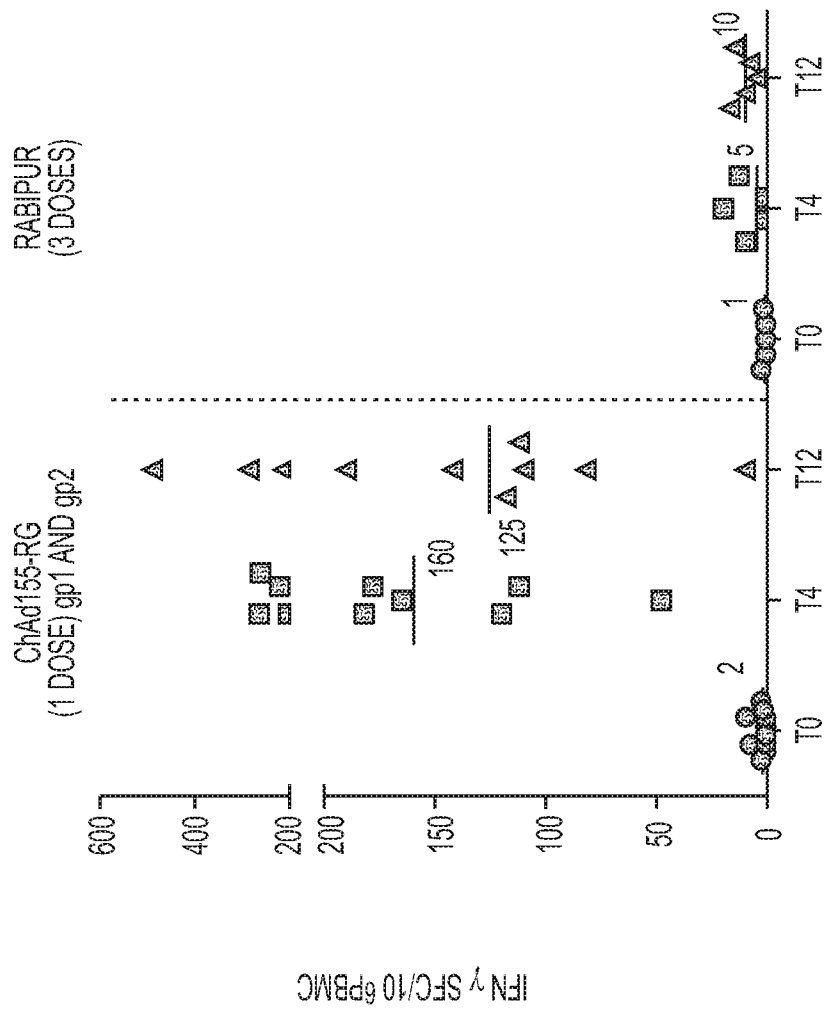


FIG. 17B

FIG. 17A

**FIG. 18**

**FIG. 19**