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- (73) Patenthaver: **BASF Enzymes LLC, 3550 John Hopkins Ct., San Diego, CA 92121, USA**
- (72) Opfinder: **SOLBAK, Arne, 7676 Acama Place, San Diego, CA 92126, USA**
GRAY, Kevin, 12119 Eleonore Court, San Diego, CA 92131, USA
PUROHIT, Shalaka, 8786 Park Run Road, San Diego, CA 92129, USA
GERENDASH, Joel, 4946 63rd Street, San Diego, CA 92121, USA
JANSSEN, Giselle, 10 Arch Lane, San Carlos, CA 94070, USA
DAHOD, Samun, 4651 Torrey Circle K203, San Diego, CA 92130, USA
KEROVUO, Janne, 5065 La Jolla Blvd. No. 2, San Diego, CA 92109, USA
MCCANN, Ryan, 1621 La Playa Avenue No. 9, San Diego, CA 92109, USA
- (74) Fuldmægtig i Danmark: **RWS Group, Europa House, Chiltern Park, Chiltern Hill, Chalfont St Peter, Bucks SL9 9FG, Storbritannien**
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DESCRIPTION

TECHNICAL FIELD

[0001] This invention relates to molecular and cellular biology, biochemistry and biotechnology. The invention is directed to polypeptides having a pectate lyase activity, e.g., a pectinase, polynucleotides encoding the polypeptides, and methods for making and using these polynucleotides and polypeptides. The polypeptides of the invention can be used as pectate lyases to catalyze the beta-elimination or hydrolysis of pectin and/or polygalacturonic acid, such as 1,4-linked alpha-D-galacturonic acid. They can be used in variety of industrial applications, e.g., to treat plant cell walls, such as those in cotton or other natural fibers. In another exemplary industrial application, the polypeptides of the invention can be used in textile scouring.

BACKGROUND

[0002] Cotton fiber consists of a primary and a secondary cell wall. The secondary cell wall is practically pure cellulose, whereas the primary cell wall is a complex lattice of pectin, protein, waxes, pigments, hemicellulose and cellulose. In textile scouring of cellulosic material (e.g. knitted or woven cotton fabric) alkaline conditions (up to 10% NaOH) and high temperatures (up to 100°C) are needed for effective removal of primary cell wall components. This harsh chemical treatment results in raw material losses and in substantial environmental burden. There are several different enzymes that have the ability to degrade pectin; these are the pectinases, pectin methylesterases, pectin lyases and pectate lyases.

[0003] "Size" is the name given to the substance or mixture of substances that is applied to the warp thread before weaving. The size forms a coating around the surface of the thread before weaving. This coating provides the lubrication and prevents the breakage of warp thread during the weaving operation. Some common chemicals used to prepare sizes are Polyacrylic Acid (PA), Polyvinyl Alcohol (PVA), Starch, and Modified Starch. Cellulosic fibers including cotton, rayon and blend of these with synthetic fibers such as polyester, is usually sized with starch-based sizes. Desizing process removes the size before dyeing, printing and/or finishing. Starch sizes can be removed by hot acid wash, which will hydrolyze starch. However, acid hydrolysis results in loss of raw material since cellulose is also prone to acid hydrolysis. Starch sizes can also be removed by using hydrogen peroxide to degrade starch by oxidation. Desizing can also be an enzymatic process. Amylases have been used for many years in textile industry for removal of starch sizes. Conditions (e.g., pH and temperature) for enzymatic desizing are dictated by the operating conditions of the enzyme. Most amylases used in the application are relatively thermostable, however, they are neutral or acidic optimum enzymes.

[0004] "Scouring" is a process in which desized cotton fabric is processed to solubilize and extract undesired non-cellulosic material naturally found in cotton and also to remove applied impurities such as machinery lubricants. Scouring uses highly alkaline chemicals to remove the non-cellulosic material, which has a serious environmental impact. Additionally, the chemicals partially degrade the cellulose in the cotton fiber which causes a loss of fiber strength and raw materials and as such is a non-optimal process. The final step in the cotton fabric pretreatment process is bleaching in which the natural pigments and matter present in the fiber are bleached. A thermostable alkaline pectinolytic enzyme that could target specifically the non-cellulosic material could reduce or eliminate the use of harsh chemicals lessening the burden on the environment while maintaining the integrity and strength of the cotton fibre.

[0005] Earlier disclosures cited in examination include WO 00/26464 discussing pectate lyases and their thermostability, as well as database entries AF279364 and Q9F7L3 disclosing sequences having some overlap with those of SEQ ID NO: 77 and SEQ ID NO: 78, but outside the scope of this invention.

SUMMARY

[0006] The invention provides isolated, synthetic or recombinant nucleic acids comprising a nucleic acid sequence having at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity to SEQ ID NO:77, over a region of at least about 950, 1000, 1050, 1100, 1150, 1200, 1250, 1300, 1350, 1400, 1450, 1500, 1550, 1600, 1650, 1700, 1750, 1800, 1850, 1900, 1950, 2000, 2050, 2100, 2200, 2250, 2300, 2350, 2400, 2450, 2500, or more residues, which encodes at least one polypeptide having a pectate lyase activity, and the sequence identities are determined by analysis with a sequence comparison algorithm or by a visual inspection.

[0007] Exemplary nucleic acids of the invention also include isolated or recombinant nucleic acids encoding a polypeptide having a sequence as set forth in SEQ ID NO:78, and certain subsequences thereof and variants thereof. The polypeptide has a pectate lyase activity.

[0008] In one aspect, the invention also provides said pectate lyase-encoding nucleic acids with a common novelty in that they are derived from mixed cultures.

[0009] In one aspect, the invention also provides said pectate lyase-encoding nucleic acids with a common novelty in that they are derived from environmental sources, e.g., mixed environmental sources.

[0010] In one aspect, the sequence comparison algorithm is a BLAST version 2.2.2 algorithm where a filtering setting is set to blastall -p blastp -d "nr pataa" -F F, and all other options are set to default.

[0011] The pectate lyase activity may comprise catalysis of beta-elimination (trans-elimination) or hydrolysis of pectin or polygalacturonic acid (pectate). The pectate lyase activity can comprise the breakup or dissolution of plant cell walls. The pectate lyase activity can comprise beta-elimination (trans-elimination) or hydrolysis of 1,4-linked alpha-D-galacturonic acid. The pectate lyase activity can comprise catalysis of beta-elimination (trans-elimination) or hydrolysis of methyl-esterified galacturonic acid. The pectate lyase activity can be exo-acting or endo-acting. In one aspect, the pectate lyase activity is endo-acting and acts at random sites within a polymer chain to give a mixture of oligomers. In one aspect, the pectate lyase activity is exo-acting and acts from one end of a polymer chain and produces monomers or dimers. The pectate lyase activity can catalyze the random cleavage of alpha-1,4-glycosidic linkages in pectic acid (polygalacturonic acid) by trans-elimination or hydrolysis. The pectate lyase activity can comprise activity the same or similar to pectate lyase (EC 4.2.2.2), poly(1,4-alpha-D-galacturonide) lyase, polygalacturonate lyase (EC 4.2.2.2), pectin lyase (EC 4.2.2.10), polygalacturonase (EC 3.2.1.15), exo-polygalacturonase (EC 3.2.1.67), exo-polygalacturonate lyase (EC 4.2.2.9) or exo-poly-alpha-galacturonosidase (EC 3.2.1.82). The pectate lyase activity can comprise beta-elimination (trans-elimination) or hydrolysis of galactan to galactose or galactooligomers. The pectate lyase activity can comprise beta-elimination (trans-elimination) or hydrolysis of a plant fiber. The plant fiber can comprise cotton fiber, hemp fiber or flax fiber.

[0012] The isolated or recombinant nucleic acid may encode a polypeptide having a pectate lyase activity that is thermostable. The polypeptide can retain a pectate lyase activity under conditions comprising a temperature range of between about 37°C to about 95°C; between about 55°C to about 85°C, between about 70°C to about 95°C, or, between about 90°C to about 95°C.

[0013] The isolated or recombinant nucleic acid may encode a polypeptide having a pectate lyase activity that is thermotolerant. The polypeptide can retain a pectate lyase activity after exposure to a temperature in the range from greater than 37°C to about 95°C or anywhere in the range from greater than 55°C to about 85°C. In one aspect, the polypeptide retains a pectate lyase activity after exposure to a temperature in the range from greater than 90°C to about 95°C at pH 4.5.

[0014] The invention provides isolated or recombinant nucleic acids comprising a sequence that hybridizes under stringent conditions to a nucleic acid according to the invention. The stringent conditions include a wash step comprising a wash in 0.2X SSC at a temperature of about 65°C for about 15 minutes.

[0015] The invention provides a nucleic acid probe for identifying a nucleic acid encoding a polypeptide having a pectate lyase activity, wherein the probe comprises at least about 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 or more, consecutive bases of a sequence comprising a sequence of the invention, or fragments or subsequences thereof, wherein the probe identifies the nucleic acid by binding or hybridization. The probe can comprise an oligonucleotide comprising at least about 10 to 50, about 20 to 60, about 30 to 70, about 40 to 80, or about 60 to 100 consecutive bases of a sequence comprising a sequence of the invention, or fragments or subsequences thereof.

[0016] The invention provides a nucleic acid probe for identifying a nucleic acid encoding a polypeptide having a pectate lyase activity, wherein the probe comprises a nucleic acid comprising a sequence about 60 to 100 or about 60 to 150 consecutive bases of a nucleic acid sequence of the invention: SEQ ID NO:77, SEQ ID NO: 131 or SEQ ID NO: 133.

[0017] An amplification primer sequence pair for amplifying a nucleic acid encoding a polypeptide having a pectate lyase activity may be capable of amplifying a nucleic acid comprising a sequence of the invention, or fragments or subsequences thereof. One or each member of the amplification primer sequence pair can comprise an oligonucleotide comprising at least about 10 to 50 consecutive bases of the sequence.

[0018] Methods of amplifying a nucleic acid encoding a polypeptide having a pectate lyase activity may comprise amplification of a template nucleic acid with such an amplification primer sequence pair.

[0019] The invention provides expression cassettes comprising a nucleic acid of the invention or a subsequence thereof. In one aspect, the expression cassette can comprise the nucleic acid that is operably linked to a promoter. The promoter can be a viral, bacterial, mammalian or plant promoter. In one aspect, the plant promoter can be a potato, rice, corn, wheat, tobacco or barley promoter. The promoter can be a constitutive promoter. The constitutive promoter can comprise CaMV35S. In another aspect, the promoter can be an inducible promoter. In one aspect, the promoter can be a tissue-specific promoter or an environmentally regulated or a developmentally regulated promoter. Thus, the promoter can be, e.g., a seed-specific, a leaf-specific, a root-specific, a stem-specific or an abscission-induced promoter. In one aspect, the expression cassette can further comprise a plant or plant virus expression vector.

[0020] The invention provides cloning vehicles comprising an expression cassette (e.g., a vector) of the invention or a nucleic acid of the invention. The cloning vehicle can be a viral vector, a plasmid, a phage, a phagemid, a cosmid, a fosmid, a bacteriophage or an artificial chromosome. The viral vector can comprise an adenovirus vector, a retroviral vector or an adeno-associated viral vector. The cloning vehicle can comprise a bacterial artificial chromosome (BAC), a plasmid, a bacteriophage P1-derived vector (PAC), a yeast artificial chromosome (YAC), or a mammalian artificial chromosome (MAC).

[0021] The invention provides transformed cell comprising a nucleic acid of the invention or an expression cassette (e.g., a vector) of the invention, or a cloning vehicle of the invention. In one aspect, the transformed cell can be a bacterial cell, a mammalian cell, a fungal cell, a yeast cell, an insect cell or a plant cell. In one aspect, the plant cell can be a potato, wheat, rice, corn, tobacco or barley cell.

[0022] The invention provides transgenic non-human animals comprising a nucleic acid of the invention or an expression cassette (e.g., a vector) of the invention. In one aspect, the animal is a mouse.

[0023] The invention provides transgenic plants comprising a nucleic acid of the invention or an expression cassette (e.g., a vector) of the invention. The transgenic plant can be a corn plant, a potato plant, a tomato plant, a wheat plant, an oilseed plant, a rapeseed plant, a soybean plant, a rice plant, a barley plant or a tobacco plant.

[0024] The invention provides transgenic seeds comprising a nucleic acid of the invention or an expression cassette (e.g., a vector) of the invention. The transgenic seed can be a corn seed, a wheat kernel, an oilseed, a rapeseed, a soybean seed, a palm kernel, a sunflower seed, a sesame seed, a peanut or a tobacco plant seed.

[0025] An antisense oligonucleotide may comprise a nucleic acid sequence complementary to or capable of hybridizing under stringent conditions to a nucleic acid of the invention. Methods of inhibiting the translation of a pectate lyase message in a cell may comprise administering to the cell or expressing in the cell an antisense oligonucleotide comprising a nucleic acid sequence complementary to or capable of hybridizing under stringent conditions to a nucleic acid of the invention.

[0026] The invention provides an isolated or recombinant polypeptide comprising an amino acid sequence having at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity to SEQ ID NO:78 over a region of at least 350 or more residues, or over the full length of the polypeptide, and the sequence identities are determined by analysis with a sequence comparison algorithm or by a visual inspection. A peptide of the disclosure can be, e.g., an immunogenic fragment, a motif (e.g., a binding site), a signal sequence, a prepro sequence or an active site. Exemplary polypeptide or peptide sequences of the invention include sequence encoded by a nucleic acid of the invention. Exemplary polypeptide or peptide sequences include polypeptides or peptides specifically bound by an antibody of the disclosure. The isolated or recombinant polypeptide of the invention (with or without a signal sequence) has pectate lyase activity.

[0027] In one aspect, the isolated or recombinant polypeptide can comprise the polypeptide of the invention that lacks a signal sequence. In one aspect, the isolated or recombinant polypeptide can comprise the polypeptide of the invention comprising a heterologous signal sequence, such as a heterologous pectate lyase or non-pectate lyase signal sequence.

[0028] In one aspect, the invention provides chimeric proteins comprising a first domain comprising a signal sequence of the invention (e.g., as set forth in Table 2, below) and at least a second domain. The protein can be a fusion protein. The second domain can comprise an enzyme. The enzyme can be a pectate lyase.

[0029] In one aspect, the pectate lyase activity comprises a specific activity at about 37°C in the range from about 100 to about 1000 units per milligram of protein. In another aspect, the pectate lyase activity comprises a specific activity from about 500 to about 750 units per milligram of protein. Alternatively, the pectate lyase activity comprises a specific activity at 37°C in the range from about 500 to about 1200 units per milligram of protein. In one aspect, the pectate lyase activity comprises a specific activity at 37°C in the range from about 750 to about 1000 units per milligram of protein. In another aspect, the thermotolerance comprises retention of at least half of the specific activity of the pectate lyase at 37°C after being heated to the elevated temperature. Alternatively, the thermotolerance can comprise retention of specific activity at 37°C in the range from about 500 to about 1200 units per milligram of protein after being heated to the elevated temperature.

[0030] The invention provides the isolated or recombinant polypeptide of the invention, wherein the polypeptide comprises at least one glycosylation site. In one aspect, glycosylation can be an N-linked glycosylation. In one aspect, the polypeptide can be glycosylated after being expressed in a *P. pastoris* or a *S. pombe*.

[0031] The polypeptide may retain a pectate lyase activity under conditions comprising about pH 6.5, pH 6, pH 5.5, pH 5, pH 4.5 or pH 4. Alternatively, the polypeptide may retain a pectate lyase activity under conditions comprising about pH 7, pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5 or pH 11.

[0032] In one aspect, the isolated or recombinant polypeptide can comprise the polypeptide of the invention that lacks a signal sequence and/or a prepro domain. In one aspect, the isolated or recombinant polypeptide can comprise the polypeptide of the invention comprising a heterologous signal sequence and/or prepro domain, such as a heterologous pectate lyase signal sequence.

[0033] A signal sequence may comprise a peptide comprising/ consisting of a sequence as set forth in residues 1 to 12, 1 to 13, 1 to 14, 1 to 15, 1 to 16, 1 to 17, 1 to 18, 1 to 19, 1 to 20, 1 to 21, 1 to 22, 1 to 23, 1 to 24, 1 to 25, 1 to 26, 1 to 27, 1 to 28, 1 to 28, 1 to 30, 1 to 31, 1 to 32, 1 to 33, 1 to 34, 1 to 35, 1 to 36, 1 to 37, 1 to 38, 1 to 39, 1 to 40, 1 to 41, 1 to 42, 1 to 43, 1 to 44 of a polypeptide of the invention. Chimeric proteins may comprise a first domain comprising such a signal sequence and at least a second domain. The protein can be a fusion protein. The second domain can comprise an enzyme. The enzyme can be a pectate lyase, e.g., an enzyme of the invention.

[0034] The invention provides chimeric polypeptides comprising at least a first domain comprising signal peptide (SP), a prepro domain, a catalytic domain (CD), or an active site of a pectate lyase of the invention and at least a second domain comprising a heterologous polypeptide or peptide, wherein the heterologous polypeptide or peptide is not naturally associated with the signal peptide (SP), prepro domain or catalytic domain (CD). In one aspect, the heterologous polypeptide or peptide is not a pectate lyase. The heterologous polypeptide or peptide can be amino terminal to, carboxy terminal to or on both ends of the signal peptide (SP), prepro domain or catalytic domain (CD).

[0035] The invention provides protein preparations comprising a polypeptide of the invention, wherein the protein preparation comprises a liquid, a solid or a gel.

[0036] The invention provides heterodimers comprising a polypeptide of the invention and a second domain. In one aspect, the second domain can be a polypeptide and the heterodimer can be a fusion protein. In one aspect, the second domain can be an epitope or a tag. In one aspect, the invention provides homodimers comprising a polypeptide of the invention.

[0037] The invention provides immobilized polypeptides having a pectate lyase activity, wherein the polypeptide comprises a polypeptide of the invention, a polypeptide encoded by a nucleic acid of the invention, or a polypeptide comprising a polypeptide of the invention and a second domain. In one aspect, the polypeptide can be immobilized on a cell, a metal, a resin, a polymer, a ceramic, a glass, a microelectrode, a graphitic particle, a bead, a gel, a plate, an array or a capillary tube.

[0038] Arrays may comprise an immobilized nucleic acid of the invention or an antibody of the disclosure.

[0039] Isolated or recombinant antibodies may specifically bind to a polypeptide of the invention or to a polypeptide encoded by a nucleic acid of the invention. The antibody can be a monoclonal or a polyclonal antibody. Hybridomas may comprise such an antibody.

[0040] The invention provides food supplements for an animal comprising a polypeptide of the invention, e.g., a polypeptide encoded by the nucleic acid of the invention. In one aspect, the polypeptide in the food supplement can be glycosylated. The invention provides edible enzyme delivery matrices comprising a polypeptide of the invention, e.g., a polypeptide encoded by the nucleic acid of the invention. In one aspect, the delivery matrix comprises a pellet. In one aspect, the polypeptide can be

glycosylated. In one aspect, the pectate lyase activity is thermotolerant. In another aspect, the pectate lyase activity is thermostable.

[0041] A method of isolating or identifying a polypeptide having a pectate lyase activity comprises the steps of: (a) providing an antibody of the disclosure; (b) providing a sample comprising polypeptides; and (c) contacting the sample of step (b) with the antibody of step (a) under conditions wherein the antibody can specifically bind to the polypeptide, thereby isolating or identifying a polypeptide having a pectate lyase activity.

[0042] Methods of making an anti-pectate lyase antibody comprise administering to a non-human animal a nucleic acid of the invention or a polypeptide of the invention or subsequences thereof in an amount sufficient to generate a humoral immune response, thereby making an anti-pectate lyase antibody. Methods of making an anti-pectate lyase immune comprise administering to a non-human animal a nucleic acid of the invention or a polypeptide of the invention or subsequences thereof in an amount sufficient to generate an immune response.

[0043] The invention provides methods of producing a recombinant polypeptide comprising the steps of: (a) providing a nucleic acid of the invention operably linked to a promoter; and (b) expressing the nucleic acid of step (a) under conditions that allow expression of the polypeptide, thereby producing a recombinant polypeptide. In one aspect, the method can further comprise transforming a host cell with the nucleic acid of step (a) followed by expressing the nucleic acid of step (a), thereby producing a recombinant polypeptide in a transformed cell.

[0044] Methods for identifying a polypeptide having a pectate lyase activity comprise the following steps: (a) providing a polypeptide of the invention; or a polypeptide encoded by a nucleic acid of the invention; (b) providing a pectate lyase substrate; and (c) contacting the polypeptide or a fragment or variant thereof of step (a) with the substrate of step (b) and detecting a decrease in the amount of substrate or an increase in the amount of a reaction product, wherein a decrease in the amount of the substrate or an increase in the amount of the reaction product detects a polypeptide having a pectate lyase activity.

[0045] Methods for identifying a pectate lyase substrate comprise the following steps: (a) providing a polypeptide of the invention; or a polypeptide encoded by a nucleic acid of the invention; (b) providing a test substrate; and (c) contacting the polypeptide of step (a) with the test substrate of step (b) and detecting a decrease in the amount of substrate or an increase in the amount of reaction product, wherein a decrease in the amount of the substrate or an increase in the amount of a reaction product identifies the test substrate as a pectate lyase substrate.

[0046] Methods of determining whether a test compound specifically binds to a polypeptide comprise the following steps: (a) expressing a nucleic acid or a vector comprising the nucleic acid under conditions permissive for translation of the nucleic acid to a polypeptide, wherein the nucleic acid comprises a nucleic acid of the invention, or, providing a polypeptide of the invention; (b) providing a test compound; (c) contacting the polypeptide with the test compound; and (d) determining whether the test compound of step (b) specifically binds to the polypeptide.

[0047] Methods for identifying a modulator of a pectate lyase activity comprise the following steps: (a) providing a polypeptide of the invention or a polypeptide encoded by a nucleic acid of the invention; (b) providing a test compound; (c) contacting the polypeptide of step (a) with the test compound of step (b) and measuring an activity of the pectate lyase, wherein a change in the pectate lyase activity measured in the presence of the test compound compared to the activity in the absence of the test compound provides a determination that the test compound modulates the pectate lyase activity. The pectate lyase activity can be measured by providing a pectate lyase substrate and detecting a decrease in the amount of the substrate or an increase in the amount of a reaction product, or, an increase in the amount of the substrate or a decrease in the amount of a reaction product. A decrease in the amount of the substrate or an increase in the amount of the reaction product with the test compound as compared to the amount of substrate or reaction product without the test compound identifies the test compound as an activator of pectate lyase activity. An increase in the amount of the substrate or a decrease in the amount of the reaction product with the test compound as compared to the amount of substrate or reaction product without the test compound identifies the test compound as an inhibitor of pectate lyase activity.

[0048] Computer systems may comprise a processor and a data storage device wherein said data storage device has stored thereon a polypeptide sequence or a nucleic acid sequence of the invention (e.g., a polypeptide encoded by a nucleic acid of the invention). The computer system can further comprise a sequence comparison algorithm and a data storage device having at least one reference sequence stored thereon. The sequence comparison algorithm may comprise a computer program that indicates polymorphisms. The computer system can further comprise an identifier that identifies one or more features in said sequence. Computer readable media may have stored thereon a polypeptide sequence or a nucleic acid sequence of the

invention. Methods for identifying a feature in a sequence comprise the steps of: (a) reading the sequence using a computer program which identifies one or more features in a sequence, wherein the sequence comprises a polypeptide sequence or a nucleic acid sequence of the invention; and (b) identifying one or more features in the sequence with the computer program. Methods for comparing a first sequence to a second sequence comprise the steps of: (a) reading the first sequence and the second sequence through use of a computer program which compares sequences, wherein the first sequence comprises a polypeptide sequence or a nucleic acid sequence of the invention; and (b) determining differences between the first sequence and the second sequence with the computer program. The step of determining differences between the first sequence and the second sequence can further comprise the step of identifying polymorphisms. The method can further comprise an identifier that identifies one or more features in a sequence. The method can comprise reading the first sequence using a computer program and identifying one or more features in the sequence.

[0049] Methods for isolating or recovering a nucleic acid encoding a polypeptide having a pectate lyase activity from an environmental sample comprise the steps of: (a) providing an amplification primer sequence pair for amplifying a nucleic acid encoding a polypeptide having a pectate lyase activity, wherein the primer pair is capable of amplifying a nucleic acid of the invention; (b) isolating a nucleic acid from the environmental sample or treating the environmental sample such that nucleic acid in the sample is accessible for hybridization to the amplification primer pair; and, (c) combining the nucleic acid of step (b) with the amplification primer pair of step (a) and amplifying nucleic acid from the environmental sample, thereby isolating or recovering a nucleic acid encoding a polypeptide having a pectate lyase activity from an environmental sample. One or each member of the amplification primer sequence pair can comprise an oligonucleotide comprising at least about 10 to 50 consecutive bases of a sequence of the invention.

[0050] Methods for isolating or recovering a nucleic acid encoding a polypeptide having a pectate lyase activity from an environmental sample comprise the steps of: (a) providing a polynucleotide probe comprising a nucleic acid of the invention or a subsequence thereof; (b) isolating a nucleic acid from the environmental sample or treating the environmental sample such that nucleic acid in the sample is accessible for hybridization to a polynucleotide probe of step (a); (c) combining the isolated nucleic acid or the treated environmental sample of step (b) with the polynucleotide probe of step (a); and (d) isolating a nucleic acid that specifically hybridizes with the polynucleotide probe of step (a), thereby isolating or recovering a nucleic acid encoding a polypeptide having a pectate lyase activity from an environmental sample. The environmental sample can comprise a water sample, a liquid sample, a soil sample, an air sample or a biological sample. In one aspect, the biological sample can be derived from a bacterial cell, a protozoan cell, an insect cell, a yeast cell, a plant cell, a fungal cell or a mammalian cell.

[0051] Methods of generating a variant of a nucleic acid encoding a polypeptide having a pectate lyase activity comprise the steps of: (a) providing a template nucleic acid comprising a nucleic acid of the invention; and (b) modifying, deleting or adding one or more nucleotides in the template sequence, or a combination thereof, to generate a variant of the template nucleic acid. The method can further comprise expressing the variant nucleic acid to generate a variant pectate lyase polypeptide. The modifications, additions or deletions can be introduced by a method comprising error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, *in vivo* mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly, gene site saturation mutagenesis (GSSM™), synthetic ligation reassembly (SLR) or a combination thereof. In another aspect, the modifications, additions or deletions are introduced by a method comprising recombination, recursive sequence recombination, phosphothioate-modified DNA mutagenesis, uracil-containing template mutagenesis, gapped duplex mutagenesis, point mismatch repair mutagenesis, repair-deficient host strain mutagenesis, chemical mutagenesis, radiogenic mutagenesis, deletion mutagenesis, restriction-selection mutagenesis, restriction-purification mutagenesis, artificial gene synthesis, ensemble mutagenesis, chimeric nucleic acid multimer creation and a combination thereof.

[0052] The method can be iteratively repeated until a pectate lyase having an altered or different activity or an altered or different stability from that of a polypeptide encoded by the template nucleic acid is produced. The variant pectate lyase polypeptide may be thermotolerant, and retains some activity after being exposed to an elevated temperature. The variant pectate lyase polypeptide may have increased glycosylation as compared to the pectate lyase encoded by a template nucleic acid. Alternatively, the variant pectate lyase polypeptide has a pectate lyase activity under a high temperature, wherein the pectate lyase encoded by the template nucleic acid is not active under the high temperature. The method can be iteratively repeated until a pectate lyase coding sequence having an altered codon usage from that of the template nucleic acid is produced. The method can be iteratively repeated until a pectate lyase gene having higher or lower level of message expression or stability from that of the template nucleic acid is produced.

[0053] Methods for modifying codons in a nucleic acid encoding a polypeptide having a pectate lyase activity, to increase its expression in a host cell, comprise the following steps: (a) providing a nucleic acid of the invention encoding a polypeptide having a pectate lyase activity; and, (b) identifying a non-preferred or a less preferred codon in the nucleic acid of step (a) and replacing

it with a preferred or neutrally used codon encoding the same amino acid as the replaced codon, wherein a preferred codon is a codon over-represented in coding sequences in genes in the host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell, thereby modifying the nucleic acid to increase its expression in a host cell.

[0054] Methods for modifying codons in a nucleic acid encoding a polypeptide having a pectate lyase activity comprise the following steps: (a) providing a nucleic acid of the invention; and, (b) identifying a codon in the nucleic acid of step (a) and replacing it with a different codon encoding the same amino acid as the replaced codon, thereby modifying codons in a nucleic acid encoding a pectate lyase.

[0055] Methods for modifying codons in a nucleic acid encoding a polypeptide having a pectate lyase activity to increase its expression in a host cell, comprise the following steps: (a) providing a nucleic acid of the invention encoding a pectate lyase polypeptide; and, (b) identifying a non-preferred or a less preferred codon in the nucleic acid of step (a) and replacing it with a preferred or neutrally used codon encoding the same amino acid as the replaced codon, wherein a preferred codon is a codon over-represented in coding sequences in genes in the host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell, thereby modifying the nucleic acid to increase its expression in a host cell.

[0056] Methods for modifying a codon in a nucleic acid encoding a polypeptide having a pectate lyase activity to decrease its expression in a host cell comprise the following steps: (a) providing a nucleic acid of the invention; and (b) identifying at least one preferred codon in the nucleic acid of step (a) and replacing it with a non-preferred or less preferred codon encoding the same amino acid as the replaced codon, wherein a preferred codon is a codon over-represented in coding sequences in genes in a host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell, thereby modifying the nucleic acid to decrease its expression in a host cell. The host cell can be a bacterial cell, a fungal cell, an insect cell, a yeast cell, a plant cell or a mammalian cell.

[0057] Methods for producing a library of nucleic acids encoding a plurality of modified pectate lyase active sites or substrate binding sites, wherein the modified active sites or substrate binding sites are derived from a first nucleic acid comprising a sequence encoding a first active site or a first substrate binding site, comprise the following steps: (a) providing a first nucleic acid encoding a first active site or first substrate binding site, wherein the first nucleic acid sequence comprises a sequence that hybridizes under stringent conditions to a nucleic acid of the invention, and the nucleic acid encodes a pectate lyase active site or a pectate lyase substrate binding site; (b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and, (c) using the set of mutagenic oligonucleotides to generate a set of active site-encoding or substrate binding site-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized, thereby producing a library of nucleic acids encoding a plurality of modified pectate lyase active sites or substrate binding sites. The method may comprise mutagenizing the first nucleic acid of step (a) by a method comprising an optimized directed evolution system, gene site-saturation mutagenesis (GSSM™), synthetic ligation reassembly (SLR), error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, in vivo mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly, gene site saturation mutagenesis (GSSM™), synthetic ligation reassembly (SLR) and a combination thereof. Or, the method may comprise mutagenizing the first nucleic acid of step (a) or variants by a method comprising recombination, recursive sequence recombination, phosphothioate-modified DNA mutagenesis, uracil-containing template mutagenesis, gapped duplex mutagenesis, point mismatch repair mutagenesis, repair-deficient host strain mutagenesis, chemical mutagenesis, radiogenic mutagenesis, deletion mutagenesis, restriction-selection mutagenesis, restriction-purification mutagenesis, artificial gene synthesis, ensemble mutagenesis, chimeric nucleic acid multimer creation and a combination thereof.

[0058] Methods for making a small molecule may comprise the following steps: (a) providing a plurality of biosynthetic enzymes capable of synthesizing or modifying a small molecule, wherein one of the enzymes comprises a pectate lyase enzyme encoded by a nucleic acid of the invention; (b) providing a substrate for at least one of the enzymes of step (a); and (c) reacting the substrate of step (b) with the enzymes under conditions that facilitate a plurality of biocatalytic reactions to generate a small molecule by a series of biocatalytic reactions. Methods for modifying a small molecule comprise the following steps: (a) providing a pectate lyase enzyme, wherein the enzyme comprises a polypeptide of the invention, or, a polypeptide encoded by a nucleic acid of the invention, or a subsequence thereof; (b) providing a small molecule; and (c) reacting the enzyme of step (a) with the small molecule of step (b) under conditions that facilitate an enzymatic reaction catalyzed by the pectate lyase enzyme, thereby modifying a small molecule by a pectate lyase enzymatic reaction. The method can comprise a plurality of small molecule substrates for the enzyme of step (a), thereby generating a library of modified small molecules produced by at least one enzymatic reaction catalyzed by the pectate lyase enzyme. The method can comprise a plurality of additional enzymes under

conditions that facilitate a plurality of biocatalytic reactions by the enzymes to form a library of modified small molecules produced by the plurality of enzymatic reactions. The method can further comprise the step of testing the library to determine if a particular modified small molecule that exhibits a desired activity is present within the library. The step of testing the library can further comprise the steps of systematically eliminating all but one of the biocatalytic reactions used to produce a portion of the plurality of the modified small molecules within the library by testing the portion of the modified small molecule for the presence or absence of the particular modified small molecule with a desired activity, and identifying at least one specific biocatalytic reaction that produces the particular modified small molecule of desired activity.

[0059] Methods for determining a functional fragment of a pectate lyase enzyme may comprise the steps of: (a) providing a pectate lyase enzyme, wherein the enzyme comprises a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention, or a subsequence thereof; and (b) deleting a plurality of amino acid residues from the sequence of step (a) and testing the remaining subsequence for a pectate lyase activity, thereby determining a functional fragment of a pectate lyase enzyme. The pectate lyase activity may be measured by providing a pectate lyase substrate and detecting a decrease in the amount of the substrate or an increase in the amount of a reaction product.

[0060] Methods for whole cell engineering of new or modified phenotypes by using real-time metabolic flux analysis comprise the following steps: (a) making a modified cell by modifying the genetic composition of a cell, wherein the genetic composition is modified by addition to the cell of a nucleic acid of the invention; (b) culturing the modified cell to generate a plurality of modified cells; (c) measuring at least one metabolic parameter of the cell by monitoring the cell culture of step (b) in real time; and, (d) analyzing the data of step (c) to determine if the measured parameter differs from a comparable measurement in an unmodified cell under similar conditions, thereby identifying an engineered phenotype in the cell using real-time metabolic flux analysis. The genetic composition of the cell can be modified by a method comprising deletion of a sequence or modification of a sequence in the cell, or, knocking out the expression of a gene. The method can further comprise selecting a cell comprising a newly engineered phenotype.

The method can comprise culturing the selected cell, thereby generating a new cell strain comprising a newly engineered phenotype.

[0061] Methods of increasing thermotolerance or thermostability of a pectate lyase polypeptide, may comprise glycosylating a pectate lyase polypeptide, wherein the polypeptide comprises at least thirty contiguous amino acids of a polypeptide of the invention; or a polypeptide encoded by a nucleic acid sequence of the invention, thereby increasing the thermotolerance or thermostability of the pectate lyase polypeptide. The pectate lyase specific activity can be thermostable or thermotolerant at a temperature in the range from greater than about 37°C to about 95°C.

[0062] Methods for overexpressing a recombinant pectate lyase polypeptide in a cell may comprise expressing a vector comprising a nucleic acid comprising a nucleic acid of the invention or a nucleic acid sequence of the invention, wherein the sequence identities are determined by analysis with a sequence comparison algorithm or by visual inspection, wherein overexpression is effected by use of a high activity promoter, a dicistronic vector or by gene amplification of the vector.

[0063] Methods of making a transgenic plant comprise the following steps: (a) introducing a heterologous nucleic acid sequence into the cell, wherein the heterologous nucleic sequence comprises a nucleic acid sequence of the invention, thereby producing a transformed plant cell; and (b) producing a transgenic plant from the transformed cell. Step (a) can further comprise introducing the heterologous nucleic acid sequence by electroporation or microinjection of plant cell protoplasts. Step (a) can further comprise introducing the heterologous nucleic acid sequence directly to plant tissue by DNA particle bombardment. Alternatively, the step (a) can further comprise introducing the heterologous nucleic acid sequence into the plant cell DNA using an *Agrobacterium tumefaciens* host. In one aspect, the plant cell can be a potato, corn, rice, wheat, tobacco, or barley cell.

[0064] Methods of expressing a heterologous nucleic acid sequence in a plant cell comprise the following steps: (a) transforming the plant cell with a heterologous nucleic acid sequence operably linked to a promoter, wherein the heterologous nucleic sequence comprises a nucleic acid of the invention; (b) growing the plant under conditions wherein the heterologous nucleic acids sequence is expressed in the plant cell.

[0065] Isolated or recombinant signal sequences comprise or consist of signal peptides (SP) as set forth in Table 2. Isolated or recombinant signal sequences consist of a sequence as set forth in residues 1 to 15, 1 to 16, 1 to 17, 1 to 18, 1 to 19, 1 to 20, 1 to 21, 1 to 22, 1 to 23, 1 to 24, 1 to 25, 1 to 26, 1 to 27, 1 to 28, 1 to 28, 1 to 30, 1 to 31, 1 to 32, 1 to 33, 1 to 34, 1 to 35, 1 to 36, 1 to 37, 1 to 38, 1 to 39, 1 to 40, 1 to 41, 1 to 42, 1 to 43, and/or 1 to 44, of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID

NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO: 110, SEQ ID NO:112, SEQ ID NO:114, SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120, SEQ ID NO:122, SEQ ID NO: 124, SEQ ID NO:126, SEQ ID NO:128 or SEQ ID NO: 130, SEQ ID NO:132, SEQ ID NO:134.

[0066] The invention provides isolated or recombinant peptides consisting of a pectin methyl esterase domain (PED) or a catalytic domain (CD) as set forth in Table 2.

[0067] The invention provides chimeric polypeptides comprising at least a first domain comprising signal peptide (SP), a pectin methyl esterase domain (PED) or a catalytic domain (CD) as set forth in Table 2 and at least a second domain comprising a heterologous polypeptide or peptide, wherein the heterologous polypeptide or peptide is not naturally associated with the signal peptide (SP), pectin methyl esterase domain (PED) or catalytic domain (CD). In one aspect, the heterologous polypeptide or peptide is not a pectate lyase. The heterologous polypeptide or peptide can be amino terminal to, carboxy terminal to or on both ends of the signal peptide (SP), pectin methyl esterase domain (PED) or catalytic domain (CD).

[0068] The invention provides isolated or recombinant nucleic acids encoding a chimeric polypeptide, wherein the chimeric polypeptide comprises at least a first domain comprising signal peptide (SP), a pectin methyl esterase domain (PED) or a catalytic domain (CD) as set forth in Table 2 and at least a second domain comprising a heterologous polypeptide or peptide, wherein the heterologous polypeptide or peptide is not naturally associated with the signal peptide (SP), pectin methyl esterase domain (PED) or catalytic domain (CD).

[0069] Methods of increasing thermotolerance or thermostability of a pectate lyase may comprise glycosylating a pectate lyase, wherein the polypeptide comprises at least thirty contiguous amino acids of a polypeptide of the invention, thereby increasing the thermotolerance or thermostability of the pectate lyase.

[0070] Methods for overexpressing a recombinant pectate lyase in a cell comprise expressing a vector comprising a nucleic acid of the invention, wherein overexpression is effected by use of a high activity promoter, a dicistronic vector or by gene amplification of the vector.

[0071] Methods of making a transgenic plant comprise the following steps: (a) introducing a heterologous nucleic acid sequence into the cell, wherein the heterologous nucleic sequence comprises a nucleic acid of the invention, thereby producing a transformed plant cell; (b) producing a transgenic plant from the transformed cell. In one aspect, step (a) further comprises introducing the heterologous nucleic acid sequence by electroporation or microinjection of plant cell protoplasts. Step (a) can comprise introducing the heterologous nucleic acid sequence directly to plant tissue by DNA particle bombardment or by using an *Agrobacterium tumefaciens* host.

[0072] Methods of expressing a heterologous nucleic acid sequence in a plant cell comprise the following steps: (a) transforming the plant cell with a heterologous nucleic acid sequence operably linked to a promoter, wherein the heterologous nucleic sequence comprises a sequence of the invention; (b) growing the plant under conditions wherein the heterologous nucleic acids sequence is expressed in the plant cell.

[0073] The invention provides methods for hydrolyzing, breaking up or disrupting a pectin- or pectate (polygalacturonic acid)-comprising composition comprise the following steps: (a) providing a polypeptide of the invention having a pectate lyase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a composition comprising a pectin or a pectate; and (c) contacting the polypeptide of step (a) with the composition of step (b) under conditions wherein the polypeptide hydrolyzes, breaks up or disrupts the pectin- or pectate-comprising composition. In one aspect, the composition comprises a plant cell wall or a bacterial cell wall. The plant can be a cotton plant, a hemp plant or a flax plant.

[0074] The invention provides methods for liquefying or removing a pectin or pectate (polygalacturonic acid) from a composition comprising the following steps: (a) providing a polypeptide of the invention having a pectate lyase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a composition comprising a pectin or pectate (polygalacturonic acid); and (c) contacting the polypeptide of step (a) with the composition of step (b) under conditions wherein the polypeptide removes or liquefies the pectin or pectate (polygalacturonic acid).

[0075] The invention provides detergent compositions comprising a polypeptide of the invention, or a polypeptide encoded by a

nucleic acid of the invention, wherein the polypeptide has a pectate lyase activity. In one aspect, the pectate lyase is a nonsurface-active pectate lyase or a surface-active pectate lyase. The pectate lyase can be formulated in a non-aqueous liquid composition, a cast solid, a granular form, a particulate form, a compressed tablet, a gel form, a paste or a slurry form.

[0076] The invention provides methods for washing an object comprising the following steps: (a) providing a composition comprising a polypeptide of the invention having a pectate lyase activity; (b) providing an object; and (c) contacting the polypeptide of step (a) and the object of step (b) under conditions wherein the composition can wash the object.

[0077] The invention provides textiles or fabrics comprising a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. The invention provides methods for fiber, thread, textile or fabric scouring comprising the following steps: (a) providing a polypeptide of the invention having a pectate lyase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a fiber, a thread, a textile or a fabric; and (c) contacting the polypeptide of step (a) and the textile or fabric of step (b) under conditions wherein the pectate lyase can scour the fiber, thread, textile or fabric. In one aspect, the pectate lyase is an alkaline active and thermostable pectate lyase. The desizing and scouring treatments can be combined in a single bath. The method can further comprise addition of an alkaline and thermostable amylase in the contacting of step (c). The desizing or scouring treatments can comprise conditions of between about pH 8.5 to pH 10.0 and temperatures of at about 40°C. The method can further comprise addition of a bleaching step. The desizing, scouring and bleaching treatments can be done simultaneously or sequentially in a single-bath container. The bleaching treatment can comprise hydrogen peroxide or at least one peroxy compound that can generate hydrogen peroxide when dissolved in water, or combinations thereof, and at least one bleach activator. The fiber, thread, textile or fabric can comprise a cellulosic material. The cellulosic material can comprise a crude fiber, a yarn, a woven or knit textile, a cotton, a linen, a flax, a ramie, a rayon, a hemp, a jute or a blend of natural or synthetic fibers.

[0078] Also disclosed are feeds or foods comprising a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. The invention provides methods improving the extraction of oil from an oil-rich plant material comprising the following steps: (a) providing a polypeptide of the invention having a pectate lyase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing an oil-rich plant material; and (c) contacting the polypeptide of step (a) and the oil-rich plant material. In one aspect, the oil-rich plant material comprises an oil-rich seed. The oil can be a soybean oil, an olive oil, a rapeseed (canola) oil or a sunflower oil.

[0079] The invention provides methods for preparing a fruit or vegetable juice, syrup, puree or extract comprising the following steps: (a) providing a polypeptide of the invention having a pectate lyase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a composition or a liquid comprising a fruit or vegetable material; and (c) contacting the polypeptide of step (a) and the composition, thereby preparing the fruit or vegetable juice, syrup, puree or extract.

[0080] The invention provides papers or paper products or paper pulps comprising a pectate lyase of the invention, or a polypeptide encoded by a nucleic acid of the invention. The invention provides methods for treating a paper or a paper or wood pulp comprising the following steps: (a) providing a polypeptide of the invention having a pectate lyase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a composition comprising a paper or a paper or wood pulp; and (c) contacting the polypeptide of step (a) and the composition of step (b) under conditions wherein the pectate lyase can treat the paper or paper or wood pulp.

[0081] The invention provides compositions comprising a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. The composition can act as a digestive aid.

[0082] Also disclosed are oral care products comprising a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. The oral care product can comprise a toothpaste, a dental cream, a gel or a tooth powder, an odontic, a mouth wash, a pre- or post brushing rinse formulation, a chewing gum, a lozenge or a candy.

[0083] The invention provides isolated or recombinant nucleic acids having a sequence comprising a sequence modification of SEQ ID NO:131, wherein the modification of SEQ ID NO: 131 comprises one or more of the following changes: the nucleotides at residues 352 to 354 are CAT or CAC, the nucleotides at residues 544 to 546 are GTG, GTT, GTC, or GTA, the nucleotides at residues 568 to 570 are TTG, TTA, CTT, CTC, CTA, or CTG, the nucleotides at residues 589 to 591 are GGT, GGC, GGA, or GGG, the nucleotides at residues 622 to 624 are AAG or AAA, the nucleotides at residues 655 to 657 are ATG, the nucleotides at residues 667 to 669 are GAG or GAA, the nucleotides at residues 763 to 765 are CGG, CGT, CGC, CGA, AGA, AGG, the nucleotides at residues 787 to 789 are AAG or AAA, the nucleotides at residues 823 to 825 are TAT or TAC, the nucleotides at residues 925 to 927 are TGG, or the nucleotides at residues 934 to 936 are GTT, GTG, GTC, or GTA. In one aspect, the nucleic acid encodes a polypeptide having a pectate lyase activity, which can be thermotolerant or thermostable.

[0084] The disclosure provides isolated or recombinant nucleic acids having a sequence comprising a sequence modification of a nucleic acid of the disclosure (e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO:15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:75, SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:85, SEQ ID NO:87, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, SEQ ID NO:105, SEQ ID NO:107, SEQ ID NO:109, SEQ ID NO:111, SEQ ID NO:113, SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119, SEQ ID NO:121, SEQ ID NO:123, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO: 129), wherein the sequence modification comprises one or more of the following changes: the nucleotides at the equivalent of residues 352 to 354 of SEQ ID NO: 131 are changed to CAT or CAC, the nucleotides at the equivalent of residues 544 to 546 of SEQ ID NO: 131 are changed to GTG, GTT, GTC, or GTA, the nucleotides at the equivalent of residues 568 to 570 of SEQ ID NO:131 are changed to TTG, TTA, CTT, CTC, CTA, or CTG, the nucleotides at the equivalent of residues 589 to 591 of SEQ ID NO:131 are changed to GGT, GGC, GGA, or GGG, the nucleotides at the equivalent of residues 622 to 624 of SEQ ID NO: 131 are changed to AAG or AAA, the nucleotides at the equivalent of residues 655 to 657 of SEQ ID NO: 131 are changed to ATG, the nucleotides at the equivalent of residues 667 to 669 of SEQ ID NO: 131 are GAG or GAA, the nucleotides at the equivalent of residues 763 to 765 of SEQ ID NO:131 are changed to CGG, CGT, CGC, CGA, AGA, AGG, the nucleotides at the equivalent of residues 787 to 789 of SEQ ID NO: 131 are changed to AAG or AAA, the nucleotides at the equivalent of residues 823 to 825 of SEQ ID NO:131 are changed to TAT or TAC, the nucleotides at the equivalent of residues 925 to 927 of SEQ ID NO:131 are changed to TGG, or the nucleotides at the equivalent of residues 934 to 936 of SEQ ID NO: 131 are changed to GTT, GTG, GTC, or GTA. In one aspect, the nucleic acid encodes a polypeptide having a pectate lyase activity, which can be thermotolerant or thermostable.

[0085] The invention provides isolated or recombinant polypeptides having a sequence comprising a sequence modification of SEQ ID NO: 132, wherein the modification of SEQ ID NO: 132 comprises one or more of the following mutations: the alanine at amino acid position 118 is histidine, the alanine at amino acid position 182 is valine, the threonine at amino acid position 190 is leucine, the alanine at amino acid position 197 is glycine, the serine at amino acid position 208 is lysine, the threonine at amino acid position 219 is methionine, the threonine at amino acid position 223 is glutamic acid, the serine at amino acid position 255 is arginine, the serine at amino acid position 263 is lysine, the asparagine at amino acid position 275 is tyrosine, the tyrosine at amino acid position 309 is tryptophan, or, the serine at amino acid position 312 is valine. In one aspect, the polypeptide has a pectate lyase activity, which can be thermotolerant or thermostable.

[0086] The disclosure provides isolated or recombinant polypeptides having a sequence comprising a sequence modification of a polypeptide of the disclosure (e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO: 10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO: 110, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO:116, SEQ ID NO: 118, SEQ ID NO:120, SEQ ID NO: 122, SEQ ID NO:124, SEQ ID NO: 126, SEQ ID NO:128, SEQ ID NO:130), wherein the sequence modification comprises one or more of the following changes: the amino acid at the equivalent of the alanine at residue 118 of SEQ ID NO: 132 is changed to a histidine, the amino acid at the equivalent of the alanine at residue 182 of SEQ ID NO: 132 is changed to a valine, the amino acid at the equivalent of the threonine at residue 190 of SEQ ID NO: 132 is changed to a leucine, the amino acid at the equivalent of the alanine at residue 197 of SEQ ID NO: 132 is changed to a glycine, the amino acid at the equivalent of the serine at residue 208 of SEQ ID NO: 132 is changed to a lysine, the amino acid at the equivalent of the threonine at residue 219 of SEQ ID NO: 132 is changed to a methionine, the amino acid at the equivalent of the threonine at residue 223 of SEQ ID NO: 132 is changed to a glutamic acid, the amino acid at the equivalent of the serine at residue 255 of SEQ ID NO: 132 is changed to a arginine, the amino acid at the equivalent of the serine at residue 263 of SEQ ID NO: 132 is changed to a lysine, the amino acid at the equivalent of the asparagine at residue 275 of SEQ ID NO: 132 is changed to a tyrosine, the amino acid at the equivalent of the tyrosine at residue 309 of SEQ ID NO: 132 is changed to a tryptophan, or, the amino acid at the equivalent of the serine at residue 312 of SEQ ID NO: 132 is changed to a valine. In one aspect, the polypeptide has a pectate lyase activity, which can be thermotolerant or thermostable.

[0087] Methods for generating a modified pectate-lyase encoding nucleic acid comprise making one or more sequence

modifications to a pectate-lyase encoding nucleic acid, wherein the changes in the pectate-lyase encoding nucleic acid are equivalent to one or more of the following: changing nucleotides at the equivalent of residues 352 to 354 of SEQ ID NO:131 to CAT or CAC, changing nucleotides at the equivalent of residues 544 to 546 of SEQ ID NO: 131 to GTG, GTT, GTC, or GTA, changing nucleotides at the equivalent of residues 568 to 570 of SEQ ID NO: 131 to TTG, TTA, CTT, CTC, CTA, or CTG, changing nucleotides at the equivalent of residues 589 to 591 of SEQ ID NO: 131 to GGT, GGC, GGA, or GGG, changing nucleotides at the equivalent of residues 622 to 624 of SEQ ID NO:131 to AAG or AAA, changing nucleotides at the equivalent of residues 655 to 657 of SEQ ID NO:131 to ATG, changing nucleotides at the equivalent of residues 667 to 669 of SEQ ID NO:131 to GAG or GAA, the nucleotides at the equivalent of residues 763 to 765 of SEQ ID NO: 131 to CGG, CGT, CGC, CGA, AGA, AGG, changing nucleotides at the equivalent of residues 787 to 789 of SEQ ID NO: 131 to AAG or AAA, changing nucleotides at the equivalent of residues 823 to 825 of SEQ ID NO: 131 to TAT or TAC, changing nucleotides at the equivalent of residues 925 to 927 of SEQ ID NO: 131 to TGG, or changing nucleotides at the equivalent of residues 934 to 936 of SEQ ID NO: 131 to GTT, GTG, GTC, or GTA. In one aspect, the modified pectate lyase activity has a thermotolerant or thermostable activity. The pectate-lyase encoding nucleic acid may comprise a nucleic acid having a sequence as set forth in SEQ ID NO: 1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO: 11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:75, SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:85, SEQ ID NO:87, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, SEQ ID NO:105, SEQ ID NO:107, SEQ ID NO: 109, SEQ ID NO:111, SEQ ID NO: 113, SEQ ID NO: 115, SEQ ID NO:117, SEQ ID NO:119, SEQ ID NO:121, SEQ ID NO:123, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131 or SEQ ID NO:133.

[0088] Methods for generating a modified pectate lyase comprise making one or more sequence modifications to a pectate lyase, wherein the changes in the pectate lyase are equivalent to one or more of the following changes: the amino acid at the equivalent of the alanine at residue 118 of SEQ ID NO: 132 is changed to a histidine, the amino acid at the equivalent of the alanine at residue 182 of SEQ ID NO: 132 is changed to a valine, the amino acid at the equivalent of the threonine at residue 190 of SEQ ID NO: 132 is changed to a leucine, the amino acid at the equivalent of the alanine at residue 197 of SEQ ID NO: 132 is changed to a glycine, the amino acid at the equivalent of the serine at residue 208 of SEQ ID NO: 132 is changed to a lysine, the amino acid at the equivalent of the threonine at residue 219 of SEQ ID NO: 132 is changed to a methionine, the amino acid at the equivalent of the threonine at residue 223 of SEQ ID NO: 132 is changed to a glutamic acid, the amino acid at the equivalent of the serine at residue 255 of SEQ ID NO: 132 is changed to an arginine, the amino acid at the equivalent of the serine at residue 263 of SEQ ID NO: 132 is changed to a lysine, the amino acid at the equivalent of the asparagine at residue 275 of SEQ ID NO: 132 is changed to a tyrosine, the amino acid at the equivalent of the tyrosine at residue 309 of SEQ ID NO: 132 is changed to a tryptophan, or, the amino acid at the equivalent of the serine at residue 312 of SEQ ID NO: 132 is changed to a valine. The pectate lyase may comprise a sequence of the disclosure (e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO:14, SEQ ID NO: 16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO:108, SEQ ID NO:110, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 116, SEQ ID NO:118, SEQ ID NO:120, SEQ ID NO: 122, SEQ ID NO:124, SEQ ID NO:126, SEQ ID NO: 128 or SEQ ID NO: 130). The modified pectate lyase activity may have a thermotolerant or thermostable activity.

[0089] The invention provides formulations comprising at least one enzyme of the invention comprising dosages in the range of between about 1 gram per ton and 100 or more grams per ton (per ton treated material, e.g., per ton fabric, cloth or the like), between about 10 grams per ton and 90 grams per ton, between about 20 grams per ton and 80 gram per ton, between about 30 grams per ton and 70 grams per ton, between about 40 grams per ton and 50 grams per ton. For example, exemplary formulations comprise about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, etc. to 100, 200, 300, 400, 500, etc. or more grams per ton.

[0090] Alternatively, the invention provides formulations comprising at least one enzyme of the invention comprising dosages in the range of between about 1 µg per gram and 100 or more µg per gram (per gram treated material, e.g., per gram fabric, cloth or the like), between about 10 µg per gram and 90 µg per gram, between about 20 µg per gram and 80 µg per gram, between about 30 µg per gram and 70 µg per gram, between about 40 µg per gram and 50 µg per gram. For example, exemplary formulations comprise about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29,

30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, etc. to 100, 200, 300, 400, 500, etc. or more µg per gram (e.g., per gram fabric, cloth or the like).

[0091] Alternatively, the invention provides formulations comprising at least one enzyme of the invention comprising dosages in the range of between about 0.5 mg per 0.45 kg (1 pound) and 50 or more mg per 0.45 kg (1 pound) (per pound treated material, e.g., per pound fabric, cloth or the like), between about 1 mg per pound and 45 mg per pound, between about 5 mg per pound and 40 mg per pound, between about 10 mg per pound and 35 mg per pound, between about 15 mg per pound and 30 mg per pound. For example, exemplary formulations comprise about 0.5, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, etc. to 50 or more mg per pound (e.g., per pound fabric, cloth or the like).

[0092] The invention provides formulations comprising at least one enzyme of the invention comprising dosages comprising an enzyme strength of between about 100 to 40,000 units/ml, 200 to 30,000 units/ml, 500 to 30,000 units/ml, 1000 to 20,000 units/ml, 1000 to 25,000 units/ml, 1000 to 15,000 units/ml, 1000 to 10000 units/ml, 1000 to 5000 units/ml, between about 2000 to 20000 units/ml, between about 2000 to 15000 units/ml, between about 2000 to 10000 units/ml, or between about 2000 to 4000 units/ml, or, between about 200 to 25,000 units/ml, 200 to 20,000 units/ml, 200 to 15000 units/ml, 200 to 10,000 units/ml, between about 400 to 8000 units/ml, between about 600 to 6000 units/ml, between about 800 to 4000 units/ml, or between about 1000 to 2000 units/ml., or, wherein the dosage comprises an enzyme strength of about 1000 u/ml. or, wherein the dosage comprises an enzyme strength of about 3000 units/ml.

[0093] The formulation may comprise a lyophilized enzyme (e.g., an enzyme of the invention), or, the formulation may be a water-based formulation comprising an enzyme of the invention. The formulation may comprise a lyophilized enzyme resuspended in water. It may further comprises a glycerol, sucrose, sodium chloride, dextrin, propylene glycol, sorbitol, sodium sulphate or TRIS, or an equivalent. In one aspect, a formulation of the invention further comprises a buffer, e.g., a buffer comprising pH 7, 35% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; pH 7, 35% glycerol, 300 ppm proxel; pH 7, 10% sodium chloride, 25% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; pH 7, 10% sodium chloride, 25% glycerol, 300 ppm proxel; pH 5.5, 35% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; pH 5.5, 35% glycerol, 300 ppm proxel; pH 5.5, 10% sodium chloride, 25% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; or, 20mM acetate buffer, pH 5.5, 35% glycerol; 20 mM MOPS, pH 7 or 25 mM MOPS, 50 mM NaCl, pH 7.5; pH 5.0, 40mM TRIS; pH 7.0, 40mM TRIS; pH 8.0, 40mM TRIS; pH 7.5, 50% glycerol; pH 7.5, 20% NaCl; pH 7.5, 30% propylene glycol; pH 7.5, 100mM sodium sulfate; pH 5.5, 35% glycerol; or, any combination thereof, or, equivalents thereof.

[0094] The invention provides bioscouring processes comprising the following steps: (a) providing a pectate lyase of the invention; (b) providing a pectin- or polygalacturonic acid- comprising material; (c) contacting the pectate lyase of (a) with the material of (b) under alkaline conditions, e.g., a pH great than 7.5, or, conditions comprising between about pH 8 and pH 9 or greater, e.g., pH 8.5, in bicarbonate buffer or equivalent. In one aspect, the method also comprises a non-ionic wetting agent, e.g., at about 1 g/L. In one aspect, the pectate lyase ratio is in an enzyme bath between about 10:1 to 50:1 L pectate lyase:kg of material. In one aspect, the pectate lyase dose is between about 0.1 and 0.2 ml of a concentrated extract per kg of material, or equivalent. Alternatively, the pectate lyase dose is between about 0.1 ml to 1 ml of a concentrated extract per kg of material, or equivalent. In one aspect, the temperature range is between about 50°C to 70°C. In one aspect, the treatment time is about 20 min. In one aspect of the bioscouring processes of the invention, the material comprises a fabric or a cloth. In one aspect, the pectate lyase dose is about 0.137 ml of a concentrated extract per kg of material, or equivalent. In one aspect, the contacting step further comprises use of a chelant, wherein the chelant is excluded from the enzyme bath and is added after about 20 minutes of enzyme treatment and retained for about 10 minutes before discharging bath.

[0095] Further aspects of the invention are set out in the accompanying claims.

[0096] The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF DRAWINGS

[0097] The patent or application file contains at least one drawing executed in color. Copies of this patent or patent application publication with color drawing(s) will be provided by the Office upon request and payment of the necessary fee.

Figure 1 is a block diagram of a computer system.

Figure 2 is a flow diagram illustrating one aspect of a process for comparing a new nucleotide or protein sequence with a database of sequences in order to determine the homology levels between the new sequence and the sequences in the database.

Figure 3 is a flow diagram illustrating one aspect of a process in a computer for determining whether two sequences are homologous.

Figure 4 is a flow diagram illustrating one aspect of an identifier process 300 for detecting the presence of a feature in a sequence.

Figure 5 is a chart summary of the relative substrate specificity, relative substrate specificity value, characterization activity temperature, characterization activity pH, enzyme activity, characterization description and characterization substrate of exemplary pectate lyases of the invention.

Figure 6 is a summary of pectate lyase polypeptides of the invention, characterized as "upmutants," as discussed in detail, below.

Figure 7 is a table summarizing exemplary melting temperatures and specific activities (SA) of exemplary enzymes of the invention at various temperatures.

Figure 8 summarizes data from activity assays of exemplary thermotolerant enzymes of the invention, as described in Example 4, below.

[0098] Like reference symbols in the various drawings indicate like elements.

DETAILED DESCRIPTION

[0099] The invention provides polypeptides having a pectate lyase activity, polynucleotides encoding the polypeptides, and methods for making and using these polynucleotides and polypeptides. In one aspect, the pectate lyases of the invention are used to catalyze the beta-elimination (trans-elimination) and/or hydrolysis of pectin and/or polygalacturonic acid (pectate) or other plant wall constituents, e.g., homogalacturonan or rhamnogalacturonan, including 1,4-linked alpha-D-galacturonic acid. The pectate lyases of the invention can also be used for the hydrolysis of plant cell walls, e.g., in treating natural fibers comprising pectin, for example, cotton fibers.

[0100] Use of the pectate lyases of the invention to hydrolyze primary cell wall pectin can eliminate the need for caustics and high temperatures in cotton fiber scouring. Use of the pectate lyases of the invention also can significantly reduce the amount of water used to rinse treated fibers, e.g., knitted or woven cotton fabric, after chemical scouring. Use of the pectate lyases of the invention also can reduce raw material losses in chemical scouring. In one aspect, a pectate lyase of the invention, e.g., an alkaline and/or thermostable pectate lyase, is used for bioscouring. Thus, the invention provides processes in which desized cotton fabrics are processed to solubilize and extract undesired non-cellulosic material in fabrics and other cellulosic materials using an enzyme of the invention. The processes of the invention can be used to solubilize and/or extract materials naturally found in cotton and/or to remove applied impurities, such as machinery lubricants.

[0101] Figures 5 and 7 are chart summaries of, inter alia, the relative substrate specificity, relative substrate specificity value, characterization activity temperature, characterization activity pH, enzyme activity, characterization description and characterization substrate of pectate lyases of the disclosure.

[0102] The pectate lyase preparations of the invention (including those for treating or processing feeds or foods, treating fibers and textiles, waste treatments, plant treatments, and the like) can further comprise one or more enzymes, for example, proteases, cellulases (endo-beta-1,4-glucanases), beta-glucanases (endo-beta-1,3(4)-glucanases), lipases, cutinases, peroxidases, laccases, amylases, pectate lyases, pectinases, reductases, oxidases, phenoloxidases, ligninases, pullulanases, arabinanases, hemicellulases, mannanases, xyloglucanases, xylanases, pectin acetyl esterases, rhamnogalacturonan acetyl esterases, polygalacturonases, rhamnogalacturonases, galactanases, pectin lyases, pectin methyl esterases, cellobiohydrolases, hansglutaminases; or mixtures thereof.

Definitions

[0103] The term "pectate lyase" includes all polypeptides having a pectate lyase, or pectinase, activity, including the beta-elimination (trans-elimination) and/or hydrolysis of pectin and/or polygalacturonic acid (pectate) or other plant wall constituents, e.g., homogalacturonan or rhamnogalacturonan, including 1,4-linked alpha-D-galacturonic acid. In one aspect, pectate lyase activity includes catalysis of the cleavage of glycosidic linkages of pectic substances, e.g., catalyzing the beta-elimination (trans-elimination) and/or hydrolysis of plant cell walls (e.g., the breakup or dissolution of cell walls comprising pectin, e.g., plant cell walls). In one aspect, pectate lyase activity includes catalyzing the beta-elimination (trans-elimination) and/or hydrolysis of methyl-esterified galacturonic acid, including partially or completely methyl-esterified polygalacturonic acid. In one aspect, the pectate lyase activity is mainly endo-acting, e.g., cutting the polymer (e.g., polygalacturonic acid) at random sites within a chain to give a mixture of oligomers, or the pectate lyase activity may be exo-acting, attacking from one end of the polymer and producing monomers or dimers, or, a combination thereof. In one aspect, the pectate lyase activity comprises catalyzing the random cleavage of alpha-1,4-glycosidic linkages in pectic acid (polygalacturonic acid) by trans-elimination. In one aspect, pectate lyase activity includes polypeptides having activity the same or similar to pectate lyase (EC 4.2.2.2), poly(1,4-alpha-D-galacturonide) lyase, polygalacturonate lyase (EC 4.2.2.2), pectin lyase (EC 4.2.2.10), polygalacturonase (EC 3.2.1.15), exo-polygalacturonase (EC 3.2.1.67), exo-polygalacturonate lyase (EC 4.2.2.9) and/or exo-poly-alpha-galacturonosidase (EC 3.2.1.82).

[0104] A polypeptide can be routinely assayed for pectate lyase activity (e.g., tested to see if the protein is within the scope of the invention) by any method, e.g., a PGA assay for pectate lyases. In this test pectate lyase activity is measured at desired temperature and pH using 0.2% polygalacturonic acid (Sigma, P3850) in 25mM TrisHCl - 25mM Glycine NaOH buffer. One unit of enzyme activity is defined as the amount of protein that produced 1 μmol of unsaturated oligogalacturonides per minute equivalent to 1 μmol of unsaturated digalacturonide, using molecular extinction coefficient value of $4600 \text{ M}^{-1}\text{cm}^{-1}$ at 235 nm for dimer. Protein can be determined for homogenous purified protein by measuring absorbance at 280 nm, using extinction coefficient value specific for each protein based on sequence.

[0105] The term "antibody" includes a peptide or polypeptide derived from, modeled after or substantially encoded by an immunoglobulin gene or immunoglobulin genes, or fragments thereof, capable of specifically binding an antigen or epitope, see, e.g. Fundamental Immunology, Third Edition, W.E. Paul, ed., Raven Press, N.Y. (1993); Wilson (1994) J. Immunol. Methods 175:267-273; Yarmush (1992) J. Biochem. Biophys. Methods 25:85-97. The term antibody includes antigen-binding portions, i.e., "antigen binding sites," (e.g., fragments, subsequences, complementarity determining regions (CDRs)) that retain capacity to bind antigen, including (i) a Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) a F(ab')₂ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) a Fv fragment consisting of the VL and VH domains of a single arm of an antibody, (v) a dAb fragment (Ward et al., (1989) Nature 341:544-546), which consists of a VH domain; and (vi) an isolated complementarity determining region (CDR). Single chain antibodies are also included by reference in the term "antibody."

[0106] The terms "array" or "microarray" or "biochip" or "chip" as used herein is a plurality of target elements, each target element comprising a defined amount of one or more polypeptides (including antibodies) or nucleic acids immobilized onto a defined area of a substrate surface, as discussed in further detail, below.

[0107] As used herein, the terms "computer," "computer program" and "processor" are used in their broadest general contexts and incorporate all such devices, as described in detail, below. A "coding sequence of" or a "sequence encodes" a particular polypeptide or protein, is a nucleic acid sequence which is transcribed and translated into a polypeptide or protein when placed under the control of appropriate regulatory sequences.

[0108] The term "expression cassette" as used herein refers to a nucleotide sequence which is capable of affecting expression of a structural gene (i.e., a protein coding sequence, such as a pectate lyase of the invention) in a host compatible with such sequences. Expression cassettes include at least a promoter operably linked with the polypeptide coding sequence; and, optionally, with other sequences, e.g., transcription termination signals. Additional factors necessary or helpful in effecting expression may also be used, e.g., enhancers. Thus, expression cassettes also include plasmids, expression vectors, recombinant viruses, any form of recombinant "naked DNA" vector, and the like.

[0109] "Operably linked" as used herein refers to a functional relationship between two or more nucleic acid (e.g., DNA) segments. Typically, it refers to the functional relationship of transcriptional regulatory sequence to a transcribed sequence. For example, a promoter is operably linked to a coding sequence, such as a nucleic acid of the invention, if it stimulates or modulates the transcription of the coding sequence in an appropriate host cell or other expression system. Generally, promoter transcriptional regulatory sequences that are operably linked to a transcribed sequence are physically contiguous to the transcribed sequence, i.e., they are cis-acting. However, some transcriptional regulatory sequences, such as enhancers, need

not be physically contiguous or located in close proximity to the coding sequences whose transcription they enhance.

[0110] A "vector" comprises a nucleic acid that can infect, transfect, transiently or permanently transduce a cell. It will be recognized that a vector can be a naked nucleic acid, or a nucleic acid complexed with protein or lipid. The vector optionally comprises viral or bacterial nucleic acids and/or proteins, and/or membranes (e.g., a cell membrane, a viral lipid envelope, etc.). Vectors include, but are not limited to replicons (e.g., RNA replicons, bacteriophages) to which fragments of DNA may be attached and become replicated. Vectors thus include, but are not limited to RNA, autonomous self-replicating circular or linear DNA or RNA (e.g., plasmids, viruses, and the like, see, e.g., U.S. Patent No. 5,217,879), and include both the expression and non-expression plasmids. Where a recombinant microorganism or cell culture is described as hosting an "expression vector" this includes both extra-chromosomal circular and linear DNA and DNA that has been incorporated into the host chromosome(s). Where a vector is being maintained by a host cell, the vector may either be stably replicated by the cells during mitosis as an autonomous structure, or is incorporated within the host's genome.

[0111] As used herein, the term "promoter" includes all sequences capable of driving transcription of a coding sequence in a cell, e.g., a plant cell. Thus, promoters used in the constructs of the invention include *cis*-acting transcriptional control elements and regulatory sequences that are involved in regulating or modulating the timing and/or rate of transcription of a gene. For example, a promoter can be a *cis*-acting transcriptional control element, including an enhancer, a promoter, a transcription terminator, an origin of replication, a chromosomal integration sequence, 5' and 3' untranslated regions, or an intronic sequence, which are involved in transcriptional regulation. These *cis*-acting sequences typically interact with proteins or other biomolecules to carry out (turn on/off, regulate, modulate, etc.) transcription. "Constitutive" promoters are those that drive expression continuously under most environmental conditions and states of development or cell differentiation. "Inducible" or "regulatable" promoters direct expression of the nucleic acid of the invention under the influence of environmental conditions or developmental conditions. Examples of environmental conditions that may affect transcription by inducible promoters include anaerobic conditions, elevated temperature, drought, or the presence of light.

[0112] "Tissue-specific" promoters are transcriptional control elements that are only active in particular cells or tissues or organs, e.g., in plants or animals. Tissue-specific regulation may be achieved by certain intrinsic factors that ensure that genes encoding proteins specific to a given tissue are expressed. Such factors are known to exist in mammals and plants so as to allow for specific tissues to develop.

[0113] The term "plant" includes whole plants, plant parts (e.g., leaves, stems, flowers, roots, etc.), plant protoplasts, seeds and plant cells and progeny of same. The class of plants which can be used in the method of the invention is generally as broad as the class of higher plants amenable to transformation techniques, including angiosperms (monocotyledonous and dicotyledonous plants), as well as gymnosperms. It includes plants of a variety of ploidy levels, including polyploid, diploid, haploid and hemizygous states. As used herein, the term "transgenic plant" includes plants or plant cells into which a heterologous nucleic acid sequence has been inserted, e.g., the nucleic acids and various recombinant constructs (e.g., expression cassettes) of the invention.

[0114] "Plasmids" can be commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accord with published procedures. Equivalent plasmids to those described herein are known in the art and will be apparent to the ordinarily skilled artisan.

[0115] The term "gene" includes a nucleic acid sequence comprising a segment of DNA involved in producing a transcription product (e.g., a message), which in turn is translated to produce a polypeptide chain, or regulates gene transcription, reproduction or stability. Genes can include regions preceding and following the coding region, such as leader and trailer, promoters and enhancers, as well as, where applicable, intervening sequences (introns) between individual coding segments (exons).

[0116] The phrases "nucleic acid" or "nucleic acid sequence" includes oligonucleotide, nucleotide, polynucleotide, or to a fragment of any of these, to DNA or RNA (e.g., mRNA, rRNA, tRNA) of genomic or synthetic origin which may be single-stranded or double-stranded and may represent a sense or antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material, natural or synthetic in origin, including, e.g., iRNA, ribonucleoproteins (e.g., iRNPs). The term encompasses nucleic acids, i.e., oligonucleotides, containing known analogues of natural nucleotides. The term also encompasses nucleic-acid-like structures with synthetic backbones, see e.g., Mata (1997) *Toxicol. Appl. Pharmacol.* 144:189-197; Strauss-Soukup (1997) *Biochemistry* 36:8692-8698; Samstag (1996) *Antisense Nucleic Acid Drug Dev* 6:153-156.

[0117] "Amino acid" or "amino acid sequence" include an oligopeptide, peptide, polypeptide, or protein sequence, or to a fragment, portion, or subunit of any of these, and to naturally occurring or synthetic molecules. The terms "polypeptide" and

"protein" include amino acids joined to each other by peptide bonds or modified peptide bonds, i.e., peptide isosteres, and may contain modified amino acids other than the 20 gene-encoded amino acids. The term "polypeptide" also includes peptides and polypeptide fragments, motifs and the like. The term also includes glycosylated polypeptides. The peptides and polypeptides of the invention also include all "mimetic" and "peptidomimetic" forms, as described in further detail, below.

[0118] The term "isolated" includes a material removed from its original environment, e.g., the natural environment if it is naturally occurring. For example, a naturally occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or polypeptides could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment. As used herein, an isolated material or composition can also be a "purified" composition, i.e., it does not require absolute purity; rather, it is intended as a relative definition. Individual nucleic acids obtained from a library can be conventionally purified to electrophoretic homogeneity. In alternative aspects, the invention provides nucleic acids which have been purified from genomic DNA or from other sequences in a library or other environment by at least one, two, three, four, five or more orders of magnitude.

[0119] As used herein, the term "recombinant" can include nucleic acids adjacent to a "backbone" nucleic acid to which it is not adjacent in its natural environment. In one aspect, nucleic acids represent 5% or more of the number of nucleic acid inserts in a population of nucleic acid "backbone molecules." "Backbone molecules" according to the invention include nucleic acids such as expression vectors, self-replicating nucleic acids, viruses, integrating nucleic acids, and other vectors or nucleic acids used to maintain or manipulate a nucleic acid insert of interest. In one aspect, the enriched nucleic acids represent 10%, 15%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, 98% or more of the number of nucleic acid inserts in the population of recombinant backbone molecules. "Recombinant" polypeptides or proteins refer to polypeptides or proteins produced by recombinant DNA techniques; e.g., produced from cells transformed by an exogenous DNA construct encoding the desired polypeptide or protein. "Synthetic" polypeptides or protein are those prepared by chemical synthesis, as described in further detail, below.

[0120] A promoter sequence can be "operably linked to" a coding sequence when RNA polymerase which initiates transcription at the promoter will transcribe the coding sequence into mRNA, as discussed further, below.

[0121] "Oligonucleotide" includes either a single stranded polydeoxynucleotide or two complementary polydeoxynucleotide strands which may be chemically synthesized. Such synthetic oligonucleotides have no 5' phosphate and thus will not ligate to another oligonucleotide without adding a phosphate with an ATP in the presence of a kinase. A synthetic oligonucleotide can ligate to a fragment that has not been dephosphorylated.

[0122] The phrase "substantially identical" in the context of two nucleic acids or polypeptides, can refer to two or more sequences that have, e.g., at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more nucleotide or amino acid residue (sequence) identity, when compared and aligned for maximum correspondence, as measured using one any known sequence comparison algorithm, as discussed in detail below, or by visual inspection. In alternative aspects, the invention provides nucleic acid and polypeptide sequences having substantial identity to an exemplary sequence of the invention over a region of at least about 10, 20, 30, 40, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 or more residues, or a region ranging from between about 50 residues to the full length of the nucleic acid or polypeptide. Nucleic acid sequences of the invention can be substantially identical over the entire length of a polypeptide coding region.

[0123] A "substantially identical" amino acid sequence also can include a sequence that differs from a reference sequence by one or more conservative or non-conservative amino acid substitutions, deletions, or insertions, particularly when such a substitution occurs at a site that is not the active site of the molecule, and provided that the polypeptide essentially retains its functional properties. A conservative amino acid substitution, for example, substitutes one amino acid for another of the same class (e.g., substitution of one hydrophobic amino acid, such as isoleucine, valine, leucine, or methionine, for another, or substitution of one polar amino acid for another, such as substitution of arginine for lysine, glutamic acid for aspartic acid or glutamine for asparagine). One or more amino acids can be deleted, for example, from a pectate lyase, resulting in modification of the structure of the polypeptide, without significantly altering its biological activity. For example, amino- or carboxyl-terminal amino acids that are not required for pectate lyase activity can be removed.

[0124] "Hybridization" includes the process by which a nucleic acid strand joins with a complementary strand through base pairing. Hybridization reactions can be sensitive and selective so that a particular sequence of interest can be identified even in samples in which it is present at low concentrations. Stringent conditions can be defined by, for example, the concentrations of salt or formamide in the prehybridization and hybridization solutions, or by the hybridization temperature, and are well known in

the art. For example, stringency can be increased by reducing the concentration of salt, increasing the concentration of formamide, or raising the hybridization temperature, altering the time of hybridization, as described in detail, below. In alternative aspects, nucleic acids of the invention are defined by their ability to hybridize under various stringency conditions (e.g., high, medium, and low), as set forth herein.

[0125] "Variant" includes polynucleotides or polypeptides of the invention modified at one or more base pairs, codons, introns, exons, or amino acid residues (respectively) yet still retain the biological activity of a pectate lyase of the invention. Variants can be produced by any number of means included methods such as, for example, error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, in vivo mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly, GSSM™ and any combination thereof. Techniques for producing variant pectate lyase having activity at a pH or temperature, for example, that is different from a wild-type pectate lyase, are included herein.

[0126] The term "saturation mutagenesis" or "GSSM™" includes a method that uses degenerate oligonucleotide primers to introduce point mutations into a polynucleotide, as described in detail, below.

[0127] The term "optimized directed evolution system" or "optimized directed evolution" includes a method for reassembling fragments of related nucleic acid sequences, e.g., related genes, and explained in detail, below.

[0128] The term "synthetic ligation reassembly" or "SLR" includes a method of ligating oligonucleotide fragments in a non-stochastic fashion, and explained in detail, below.

Generating and Manipulating Nucleic Acids

[0129] The invention provides isolated and recombinant nucleic acids, e.g., polynucleotides having a sequence identity to SEQ ID NO:77, SEQ ID NO:131 or SEQ ID NO: 133; nucleic acids encoding polypeptides of the invention having a sequence as set forth in SEQ ID NO:78, SEQ ID NO:132 or SEQ ID NO:134.

[0130] The nucleic acids of the invention can also comprise expression cassettes, such as expression vectors, where in one aspect they encode a polypeptide of the invention. There are disclosed methods for discovering new pectate lyase sequences using the nucleic acids of the invention, methods for inhibiting the expression of pectate lyase genes, transcripts and polypeptides using the nucleic acids of the invention, and methods for modifying the nucleic acids of the invention by, e.g., synthetic ligation reassembly, optimized directed evolution system and/or gene site saturation mutagenesis (GSSM™).

[0131] The nucleic acids of the invention can be made, isolated and/or manipulated by, e.g., cloning and expression of cDNA libraries, amplification of message or genomic DNA by PCR, and the like. Homologous genes can be modified by manipulating a template nucleic acid, as described herein. This invention can be practiced in conjunction with any method or protocol or device known in the art, which are well described in the scientific and patent literature.

General Techniques

[0132] The nucleic acids used to practice this invention, whether RNA, iRNA (i.e., RNAi), antisense nucleic acid, cDNA, genomic DNA, vectors, viruses or hybrids thereof, may be isolated from a variety of sources, genetically engineered, amplified, and/or expressed/ generated recombinantly. Recombinant polypeptides generated from these nucleic acids can be individually isolated or cloned and tested for a desired activity. Any recombinant expression system can be used, including bacterial, mammalian, yeast, insect or plant cell expression systems.

[0133] Alternatively, these nucleic acids can be synthesized *in vitro* by well-known chemical synthesis techniques, as described in, e.g., Adams (1983) J. Am. Chem. Soc. 105:661; Belousov (1997) Nucleic Acids Res. 25:3440-3444; Frenkel (1995) Free Radic. Biol. Med. 19:373-380; Blommers (1994) Biochemistry 33:7886-7896; Narang (1979) Meth. Enzymol. 68:90; Brown (1979) Meth. Enzymol. 68:109; Beaucauge (1981) Tetra. Lett. 22:1859; U.S. Patent No. 4,458,066.

[0134] Techniques for the manipulation of nucleic acids, such as, e.g., subcloning, labeling probes (e.g., random-primer labeling using Klenow polymerase, nick translation, amplification), sequencing, hybridization and the like are well described in the scientific and patent literature, see, e.g., Sambrook, ed., MOLECULAR CLONING: A LABORATORY MANUAL (2ND ED.), Vols. 1-3, Cold

Spring Harbor Laboratory, (1989); CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Ausubel, ed. John Wiley & Sons, Inc., New York (1997); LABORATORY TECHNIQUES IN BIOCHEMISTRY AND MOLECULAR BIOLOGY: HYBRIDIZATION WITH NUCLEIC ACID PROBES, Part I. Theory and Nucleic Acid Preparation, Tijssen, ed. Elsevier, N.Y. (1993).

[0135] Another useful means of obtaining and manipulating nucleic acids used to practice the methods of the invention is to clone from genomic samples, and, if desired, screen and re-clone inserts isolated or amplified from, e.g., genomic clones or cDNA clones. Sources of nucleic acid used in the methods of the invention include genomic or cDNA libraries contained in, e.g., mammalian artificial chromosomes (MACs), see, e.g., U.S. Patent Nos. 5,721,118; 6,025,155; human artificial chromosomes, see, e.g., Rosenfeld (1997) Nat. Genet. 15:333-335; yeast artificial chromosomes (YAC); bacterial artificial chromosomes (BAC); P1 artificial chromosomes, see, e.g., Woon (1998) Genomics 50:306-316; P1-derived vectors (PACs), see, e.g., Kern (1997) Biotechniques 23:120-124; cosmids, recombinant viruses, phages or plasmids.

[0136] In one aspect, a nucleic acid encoding a polypeptide of the invention is assembled in appropriate phase with a leader sequence capable of directing secretion of the translated polypeptide or fragment thereof.

[0137] The invention provides fusion proteins and nucleic acids encoding them. A polypeptide of the invention can be fused to a heterologous peptide or polypeptide, such as N-terminal identification peptides which impart desired characteristics, such as increased stability or simplified purification. Peptides and polypeptides of the invention can also be synthesized and expressed as fusion proteins with one or more additional domains linked thereto for, e.g., producing a more immunogenic peptide, to more readily isolate a recombinantly synthesized peptide, to identify and isolate antibodies and antibody-expressing B cells, and the like. Detection and purification facilitating domains include, e.g., metal chelating peptides such as polyhistidine tracts and histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle WA). The inclusion of a cleavable linker sequences such as Factor Xa or enterokinase (Invitrogen, San Diego CA) between a purification domain and the motif-comprising peptide or polypeptide to facilitate purification. For example, an expression vector can include an epitope-encoding nucleic acid sequence linked to six histidine residues followed by a thioredoxin and an enterokinase cleavage site (see e.g., Williams (1995) Biochemistry 34:1787-1797; Dobeli (1998) Protein Expr. Purif. 12:404-414). The histidine residues facilitate detection and purification while the enterokinase cleavage site provides a means for purifying the epitope from the remainder of the fusion protein. Technology pertaining to vectors encoding fusion proteins and application of fusion proteins are well described in the scientific and patent literature, see e.g., Kroll (1993) DNA Cell. Biol., 12:441-53.

Transcriptional and translational control sequences

[0138] The invention provides nucleic acid (e.g., DNA) sequences of the invention operatively linked to expression (e.g., transcriptional or translational) control sequence(s), e.g., promoters or enhancers, to direct or modulate RNA synthesis/expression. The expression control sequence can be in an expression vector. Exemplary bacterial promoters include lacI, lacZ, T3, T7, gpt, lambda PR, PL and trp. Exemplary eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein I.

[0139] Promoters suitable for expressing a polypeptide in bacteria include the *E. coli* lac or trp promoters, the lacI promoter, the lacZ promoter, the T3 promoter, the T7 promoter, the gpt promoter, the lambda PR promoter, the lambda PL promoter, promoters from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), and the acid phosphatase promoter. Eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, heat shock promoters, the early and late SV40 promoter, LTRs from retroviruses, and the mouse metallothionein-I promoter. Other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses may also be used.

Tissue-Specific Plant Promoters

[0140] The invention provides expression cassettes that can be expressed in a tissue-specific manner, e.g., that can express a pectate lyase of the invention in a tissue-specific manner. The invention also provides plants or seeds that express a pectate lyase of the invention in a tissue-specific manner. The tissue-specificity can be seed specific, stem specific, leaf specific, root specific, fruit specific and the like.

[0141] In one aspect, a constitutive promoter such as the CaMV 35S promoter can be used for expression in specific parts of the plant or seed or throughout the plant. For example, for overexpression, a plant promoter fragment can be employed which will

direct expression of a nucleic acid in some or all tissues of a plant, e.g., a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35S transcription initiation region, the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*, and other transcription initiation regions from various plant genes known to those of skill. Such genes include, e.g., *ACT11* from *Arabidopsis* (Huang (1996) Plant Mol. Biol. 33:125-139); *Cat3* from *Arabidopsis* (GenBank No. U43147, Zhong (1996) Mol. Gen. Genet. 251:196-203); the gene encoding stearylacyl carrier protein desaturase from *Brassica napus* (Genbank No. X74782, Solcombe (1994) Plant Physiol. 104:1167-1176); *Gpc1* from maize (GenBank No. X15596; Martinez (1989) J. Mol. Biol. 208:551-565); the *Gpc2* from maize (GenBank No. U45855, Manjunath (1997) Plant Mol. Biol. 33:97-112); plant promoters described in U.S. Patent Nos. 4,962,028; 5,633,440.

[0142] The invention uses tissue-specific or constitutive promoters derived from viruses which can include, e.g., the tobamovirus subgenomic promoter (Kumagai (1995) Proc. Natl. Acad. Sci. USA 92:1679-1683; the rice tungro bacilliform virus (RTBV), which replicates only in phloem cells in infected rice plants, with its promoter which drives strong phloem-specific reporter gene expression; the cassava vein mosaic virus (CVMV) promoter, with highest activity in vascular elements, in leaf mesophyll cells, and in root tips (Verdaguer (1996) Plant Mol. Biol. 31:1129-1139).

[0143] Alternatively, the plant promoter may direct expression of pectate lyase-expressing nucleic acid in a specific tissue, organ or cell type (i.e. tissue-specific promoters) or may be otherwise under more precise environmental or developmental control or under the control of an inducible promoter. Examples of environmental conditions that may affect transcription include anaerobic conditions, elevated temperature, the presence of light, or sprayed with chemicals/hormones. For example, the invention incorporates the drought-inducible promoter of maize (Busk (1997) supra); the cold, drought, and high salt inducible promoter from potato (Kirch (1997) Plant Mol. Biol. 33:897 909).

[0144] Tissue-specific promoters can promote transcription only within a certain time frame of developmental stage within that tissue. See, e.g., Blazquez (1998) Plant Cell 10:791-800, characterizing the *Arabidopsis* LEAFY gene promoter. See also Cardon (1997) Plant J 12:367-77, describing the transcription factor SPL3, which recognizes a conserved sequence motif in the promoter region of the *A. thaliana* floral meristem identity gene AP1; and Mandel (1995) Plant Molecular Biology, Vol. 29, pp 995-1004, describing the meristem promoter eIF4. Tissue specific promoters which are active throughout the life cycle of a particular tissue can be used. In one aspect, the nucleic acids of the invention are operably linked to a promoter active primarily only in cotton fiber cells. In one aspect, the nucleic acids of the invention are operably linked to a promoter active primarily during the stages of cotton fiber cell elongation, e.g., as described by Rinehart (1996) supra. The nucleic acids can be operably linked to the FbI2A gene promoter to be preferentially expressed in cotton fiber cells (Ibid). See also, John (1997) Proc. Natl. Acad. Sci. USA 89:5769-5773; John, et al., U.S. Patent Nos. 5,608,148 and 5,602,321, describing cotton fiber-specific promoters and methods for the construction of transgenic cotton plants. Root-specific promoters may also be used to express the nucleic acids of the invention. Examples of root-specific promoters include the promoter from the alcohol dehydrogenase gene (DeLisle (1990) Int. Rev. Cytol. 123:39-60). Other promoters that can be used to express the nucleic acids of the invention include, e.g., ovule-specific, embryo-specific, endosperm-specific, integument-specific, seed coat-specific promoters, or some combination thereof; a leaf-specific promoter (see, e.g., Busk (1997) Plant J. 11:1285 1295, describing a leaf-specific promoter in maize); the ORF13 promoter from *Agrobacterium rhizogenes* (which exhibits high activity in roots, see, e.g., Hansen (1997) supra); a maize pollen specific promoter (see, e.g., Guerrero (1990) Mol. Gen. Genet. 224:161 168); a tomato promoter active during fruit ripening, senescence and abscission of leaves and, to a lesser extent, of flowers can be used (see, e.g., Blume (1997) Plant J. 12:731 746); a pistil-specific promoter from the potato SK2 gene (see, e.g., Ficker (1997) Plant Mol. Biol. 35:425 431); the Blec4 gene from pea, which is active in epidermal tissue of vegetative and floral shoot apices of transgenic alfalfa making it a useful tool to target the expression of foreign genes to the epidermal layer of actively growing shoots or fibers; the ovule-specific BEL1 gene (see, e.g., Reiser (1995) Cell 83:735-742, GenBank No. U39944); and/or, the promoter in Klee, U.S. Patent No. 5,589,583, describing a plant promoter region is capable of conferring high levels of transcription in meristematic tissue and/or rapidly dividing cells.

[0145] Alternatively, plant promoters which are inducible upon exposure to plant hormones, such as auxins, are used to express the nucleic acids of the invention. For example, the invention can use the auxin-response elements E1 promoter fragment (AuxREs) in the soybean (*Glycine max* L.) (Liu (1997) Plant Physiol. 115:397-407); the auxin-responsive *Arabidopsis* GST6 promoter (also responsive to salicylic acid and hydrogen peroxide) (Chen (1996) Plant J. 10: 955-966); the auxin-inducible parC promoter from tobacco (Sakai (1996) 37:906-913); a plant biotin response element (Streit (1997) Mol. Plant Microbe Interact. 10:933-937); and, the promoter responsive to the stress hormone abscisic acid (Sheen (1996) Science 274:1900-1902).

[0146] The nucleic acids of the invention can also be operably linked to plant promoters which are inducible upon exposure to chemicals reagents which can be applied to the plant, such as herbicides or antibiotics. For example, the maize In2-2 promoter, activated by benzenesulfonamide herbicide safeners, can be used (De Veylder (1997) Plant Cell Physiol. 38:568-577);

application of different herbicide safeners induces distinct gene expression patterns, including expression in the root, hydathodes, and the shoot apical meristem. Coding sequence can be under the control of, e.g., a tetracycline-inducible promoter, e.g., as described with transgenic tobacco plants containing the *Avena sativa* L. (oat) arginine decarboxylase gene (Masgrau (1997) Plant J. 11:465-473); or, a salicylic acid-responsive element (Stange (1997) Plant J. 11:1315-1324). Using chemically- (e.g., hormone- or pesticide-) induced promoters, i.e., promoter responsive to a chemical which can be applied to the transgenic plant in the field, expression of a polypeptide of the invention can be induced at a particular stage of development of the plant. Thus, the invention also provides for transgenic plants containing an inducible gene encoding for polypeptides of the invention whose host range is limited to target plant species, such as corn, rice, barley, wheat, potato or other crops, inducible at any stage of development of the crop.

[0147] One of skill will recognize that a tissue-specific plant promoter may drive expression of operably linked sequences in tissues other than the target tissue. Thus, a tissue-specific promoter is one that drives expression preferentially in the target tissue or cell type, but may also lead to some expression in other tissues as well.

[0148] The nucleic acids of the invention can also be operably linked to plant promoters which are inducible upon exposure to chemicals reagents. These reagents include, e.g., herbicides, synthetic auxins, or antibiotics which can be applied, e.g., sprayed, onto transgenic plants. Inducible expression of the pectate lyase-producing nucleic acids of the invention will allow the grower to select plants with the optimal pectate lyase expression and/or activity. The development of plant parts can thus be controlled. In this way the invention provides the means to facilitate the harvesting of plants and plant parts. For example, in various embodiments, the maize In2-2 promoter, activated by benzenesulfonamide herbicide safeners, is used (De Veylder (1997) Plant Cell Physiol. 38:568-577); application of different herbicide safeners induces distinct gene expression patterns, including expression in the root, hydathodes, and the shoot apical meristem. Coding sequences of the invention are also under the control of a tetracycline-inducible promoter, e.g., as described with transgenic tobacco plants containing the *Avena sativa* L. (oat) arginine decarboxylase gene (Masgrau (1997) Plant J. 11:465-473); or, a salicylic acid-responsive element (Stange (1997) Plant J. 11:1315-1324).

[0149] If proper polypeptide expression is desired, a polyadenylation region at the 3'-end of the coding region should be included. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from genes in the *Agrobacterium* T-DNA.

Expression vectors and cloning vehicles

[0150] The invention provides expression vectors and cloning vehicles comprising nucleic acids of the invention, e.g., sequences encoding the pectate lyases of the invention. Expression vectors and cloning vehicles of the invention can comprise viral particles, baculovirus, phage, plasmids, phagemids, cosmids, fosmids, bacterial artificial chromosomes, viral DNA (e.g., vaccinia, adenovirus, fowl pox virus, pseudorabies and derivatives of SV40), P1-based artificial chromosomes, yeast plasmids, yeast artificial chromosomes, and any other vectors specific for specific hosts of interest (such as bacillus, *Aspergillus* and yeast). Vectors of the invention can include chromosomal, non-chromosomal and synthetic DNA sequences. Large numbers of suitable vectors are known to those of skill in the art, and are commercially available. Exemplary vectors include: bacterial: pQE vectors (Qiagen), pBluescript plasmids, pNH vectors, (lambda-ZAP vectors (Stratagene)); ptrc99a, pKK223-3, pDR540, pRIT2T (Pharmacia); Eukaryotic: pXT1, pSG5 (Stratagene), pSVK3, pBPV, pMSG, pSVLSV40 (Pharmacia). However, any other plasmid or other vector may be used so long as they are replicable and viable in the host. Low copy number or high copy number vectors may be employed with the present invention.

[0151] The expression vector can comprise a promoter, a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. Mammalian expression vectors can comprise an origin of replication, any necessary ribosome binding sites, a polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking non-transcribed sequences. In some aspects, DNA sequences derived from the SV40 splice and polyadenylation sites may be used to provide the required non-transcribed genetic elements.

[0152] In one aspect, the expression vectors contain one or more selectable marker genes to permit selection of host cells containing the vector. Such selectable markers include genes encoding dihydrofolate reductase or genes conferring neomycin resistance for eukaryotic cell culture, genes conferring tetracycline or ampicillin resistance in *E. coli*, and the *S. cerevisiae* TRP1 gene. Promoter regions can be selected from any desired gene using chloramphenicol transferase (CAT) vectors or other vectors with selectable markers.

[0153] Vectors for expressing the polypeptide or fragment thereof in eukaryotic cells can also contain enhancers to increase

expression levels. Enhancers are cis-acting elements of DNA, usually from about 10 to about 300 bp in length that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and the adenovirus enhancers.

[0154] A nucleic acid sequence can be inserted into a vector by a variety of procedures. In general, the sequence is ligated to the desired position in the vector following digestion of the insert and the vector with appropriate restriction endonucleases. Alternatively, blunt ends in both the insert and the vector may be ligated. A variety of cloning techniques are known in the art, e.g., as described in Ausubel and Sambrook. Such procedures and others are deemed to be within the scope of those skilled in the art.

[0155] The vector can be in the form of a plasmid, a viral particle, or a phage. Other vectors include chromosomal, non-chromosomal and synthetic DNA sequences, derivatives of SV40; bacterial plasmids, phage DNA, baculovirus, yeast plasmids, vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. A variety of cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by, e.g., Sambrook.

[0156] Particular bacterial vectors which can be used include the commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017), pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden), GEM1 (Promega Biotec, Madison, WI, USA) pQE70, pQE60, pQE-9 (Qiagen), pD10, psiX174 pBluescript II KS, pNH8A, pNH16a, pNH18A, pNH46A (Stratagene), ptrc99a, pKK223-3, pKK233-3, DR540, pRIT5 (Pharmacia), pKK232-8 and pCM7. Particular eukaryotic vectors include pSV2CAT, pOG44, pXT1, pSG (Stratagene) pSVK3, pBPV, pMSG, and pSVL (Pharmacia). However, any other vector may be used as long as it is replicable and viable in the host cell.

[0157] The nucleic acids of the invention can be expressed in expression cassettes, vectors or viruses and transiently or stably expressed in plant cells and seeds. One exemplary transient expression system uses episomal expression systems, e.g., cauliflower mosaic virus (CaMV) viral RNA generated in the nucleus by transcription of an episomal mini-chromosome containing supercoiled DNA, see, e.g., Covey (1990) Proc. Natl. Acad. Sci. USA 87:1633-1637. Alternatively, coding sequences, i.e., all or sub-fragments of sequences of the invention can be inserted into a plant host cell genome becoming an integral part of the host chromosomal DNA. Sense or antisense transcripts can be expressed in this manner. A vector comprising the sequences (e.g., promoters or coding regions) from nucleic acids of the invention can comprise a marker gene that confers a selectable phenotype on a plant cell or a seed. For example, the marker may encode biocide resistance, particularly antibiotic resistance, such as resistance to kanamycin, G418, bleomycin, hygromycin, or herbicide resistance, such as resistance to chlorosulfuron or Basta.

[0158] Expression vectors capable of expressing nucleic acids and proteins in plants are well known in the art, and can include, e.g., vectors from *Agrobacterium* spp., potato virus X (see, e.g., Angell (1997) EMBO J. 16:3675-3684), tobacco mosaic virus (see, e.g., Casper (1996) Gene 173:69-73), tomato bushy stunt virus (see, e.g., Hillman (1989) Virology 169:42-50), tobacco etch virus (see, e.g., Dolja (1997) Virology 234:243-252), bean golden mosaic virus (see, e.g., Morinaga (1993) Microbiol Immunol. 37:471-476), cauliflower mosaic virus (see, e.g., Cecchini (1997) Mol. Plant Microbe Interact. 10:1094-1101), maize Ac/Ds transposable element (see, e.g., Rubin (1997) Mol. Cell. Biol. 17:6294-6302; Kunze (1996) Curr. Top. Microbiol. Immunol. 204:161-194), and the maize suppressor-mutator (Spm) transposable element (see, e.g., Schlappi (1996) Plant Mol. Biol. 32:717-725); and derivatives thereof.

[0159] In one aspect, the expression vector can have two replication systems to allow it to be maintained in two organisms, for example in mammalian or insect cells for expression and in a prokaryotic host for cloning and amplification. Furthermore, for integrating expression vectors, the expression vector can contain at least one sequence homologous to the host cell genome. It can contain two homologous sequences which flank the expression construct. The integrating vector can be directed to a specific locus in the host cell by selecting the appropriate homologous sequence for inclusion in the vector. Constructs for integrating vectors are well known in the art.

[0160] Expression vectors of the invention may also include a selectable marker gene to allow for the selection of bacterial strains that have been transformed, e.g., genes which render the bacteria resistant to drugs such as ampicillin, chloramphenicol, erythromycin, kanamycin, neomycin and tetracycline. Selectable markers can also include biosynthetic genes, such as those in the histidine, tryptophan and leucine biosynthetic pathways.

Host cells and transformed cells

[0161] The invention also provides a transformed cell comprising a nucleic acid sequence of the invention, e.g., a sequence encoding a pectate lyase of the invention, or a vector of the invention. The host cell may be any of the host cells familiar to those skilled in the art, including prokaryotic cells, eukaryotic cells, such as bacterial cells, fungal cells, yeast cells, mammalian cells, insect cells, or plant cells. Exemplary bacterial cells include *E. coli*, *Streptomyces*, *Bacillus subtilis*, *Bacillus cereus*, *Salmonella typhimurium* and various species within the genera *Bacillus*, *Streptomyces*, and *Staphylococcus*. Exemplary insect cells include *Drosophila S2* and *Spodoptera Sf9*. Exemplary yeast cells include *Pichia pastoris*, *Saccharomyces cerevisiae* or *Schizosaccharomyces pombe*. Exemplary animal cells include CHO, COS or Bowes melanoma or any mouse or human cell line. The selection of an appropriate host is within the abilities of those skilled in the art. Techniques for transforming a wide variety of higher plant species are well known and described in the technical and scientific literature. See, e.g., Weising (1988) Ann. Rev. Genet. 22:421-477, U.S. Patent No. 5,750,870.

[0162] The vector can be introduced into the host cells using any of a variety of techniques, including transformation, transfection, transduction, viral infection, gene guns, or Ti-mediated gene transfer. Particular methods include calcium phosphate transfection, DEAE-Dextran mediated transfection, lipofection, or electroporation (Davis, L., Dibner, M., Battey, L., Basic Methods in Molecular Biology, (1986)).

[0163] In one aspect, the nucleic acids or vectors of the invention are introduced into the cells for screening, thus, the nucleic acids enter the cells in a manner suitable for subsequent expression of the nucleic acid. The method of introduction is largely dictated by the targeted cell type. Exemplary methods include CaPO_4 precipitation, liposome fusion, lipofection (e.g., LIPOFECTIN™), electroporation, viral infection, etc. The candidate nucleic acids may stably integrate into the genome of the host cell (for example, with retroviral introduction) or may exist either transiently or stably in the cytoplasm (i.e. through the use of traditional plasmids, utilizing standard regulatory sequences, selection markers, etc.). As many pharmaceutically important screens require human or model mammalian cell targets, retroviral vectors capable of transfecting such targets are preferred.

[0164] Where appropriate, the engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the genes of the invention. Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter may be induced by appropriate means (e.g., temperature shift or chemical induction) and the cells may be cultured for an additional period to allow them to produce the desired polypeptide or fragment thereof.

[0165] Cells can be harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract is retained for further purification. Microbial cells employed for expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents. Such methods are well known to those skilled in the art. The expressed polypeptide or fragment thereof can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the polypeptide. If desired, high performance liquid chromatography (HPLC) can be employed for final purification steps.

[0166] Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts and other cell lines capable of expressing proteins from a compatible vector, such as the C127, 3T3, CHO, HeLa and BHK cell lines.

[0167] The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Depending upon the host employed in a recombinant production procedure, the polypeptides produced by host cells containing the vector may be glycosylated or may be non-glycosylated. Polypeptides of the invention may or may not also include an initial methionine amino acid residue.

[0168] Cell-free translation systems can also be employed to produce a polypeptide of the invention. Cell-free translation systems can use mRNAs transcribed from a DNA construct comprising a promoter operably linked to a nucleic acid encoding the polypeptide or fragment thereof. In some aspects, the DNA construct may be linearized prior to conducting an in vitro transcription reaction. The transcribed mRNA is then incubated with an appropriate cell-free translation extract, such as a rabbit reticulocyte extract, to produce the desired polypeptide or fragment thereof.

[0169] The expression vectors can contain one or more selectable marker genes to provide a phenotypic trait for selection of

transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in *E. coli*.

Amplification of Nucleic Acids

[0170] In practicing the invention, nucleic acids of the invention and nucleic acids encoding the pectate lyases of the invention, or modified nucleic acids of the invention, can be reproduced by amplification. Amplification can also be used to clone or modify the nucleic acids of the invention. Thus, the disclosure provides amplification primer sequence pairs for amplifying nucleic acids of the invention. One of skill in the art can design amplification primer sequence pairs for any part of or the full length of these sequences. A nucleic acid may be amplified by a primer pair as set forth by about the first (the 5') 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 residues of a nucleic acid of the disclosure, and about the first (the 5') 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 residues of the complementary strand (e.g., of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:71, SEQ ID NO:73, SEQ ID NO:75, SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81, SEQ ID NO:83, SEQ ID NO:85, SEQ ID NO:87, SEQ ID NO:89, SEQ ID NO:91, SEQ ID NO:93, SEQ ID NO:95, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, SEQ ID NO:105, SEQ ID NO:107, SEQ ID NO:109, SEQ ID NO:111, SEQ ID NO:113, SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119, SEQ ID NO:121, SEQ ID NO:123, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133).

[0171] Amplification reactions can also be used to quantify the amount of nucleic acid in a sample (such as the amount of message in a cell sample), label the nucleic acid (e.g., to apply it to an array or a blot), detect the nucleic acid, or quantify the amount of a specific nucleic acid in a sample. Message isolated from a cell or a cDNA library may be amplified.

[0172] The skilled artisan can select and design suitable oligonucleotide amplification primers. Amplification methods are also well known in the art, and include, e.g., polymerase chain reaction, PCR (see, e.g., PCR PROTOCOLS, A GUIDE TO METHODS AND APPLICATIONS, ed. Innis, Academic Press, N.Y. (1990) and PCR STRATEGIES (1995), ed. Innis, Academic Press, Inc., N.Y., ligase chain reaction (LCR) (see, e.g., Wu (1989) Genomics 4:560; Landegren (1988) Science 241:1077; Barringer (1990) Gene 89:117); transcription amplification (see, e.g., Kwok (1989) Proc. Natl. Acad. Sci. USA 86:1173); and, self-sustained sequence replication (see, e.g., Guatelli (1990) Proc. Natl. Acad. Sci. USA 87:1874); Q Beta replicase amplification (see, e.g., Smith (1997) J. Clin. Microbiol. 35:1477-1491), automated Q-beta replicase amplification assay (see, e.g., Burg (1996) Mol. Cell. Probes 10:257-271) and other RNA polymerase mediated techniques (e.g., NASBA, Cingone, Mississauga, Ontario); see also Berger (1987) Methods Enzymol. 152:307-316; Sambrook; Ausubel; U.S. Patent Nos. 4,683,195 and 4,683,202; Sooknanan (1995) Biotechnology 13:563-564.

Determining the degree of sequence identity

[0173] The invention provides nucleic acids comprising sequences having at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity to SEQ ID NO:77 and nucleic acids encoding SEQ ID NO:78, over a region of at least about 950, 1000, 1050, 1100, 1150 or more, residues. The invention provides polypeptides comprising sequences having at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity to an exemplary polypeptide of the invention. The extent of sequence identity (homology) may be determined using any computer program and associated parameters, including those described herein, such as BLAST 2.2.2. or FASTA version 3.0t78, with the default parameters.

[0174] Homologous sequences also include RNA sequences in which uridines replace the thymines in the nucleic acid sequences. The homologous sequences may be obtained using any of the procedures described herein or may result from the correction of a sequencing error. It will be appreciated that the nucleic acid sequences as set forth herein can be represented in the traditional single character format (see, e.g., Stryer, Lubert. Biochemistry, 3rd Ed., W. H Freeman & Co., New York) or in any other format which records the identity of the nucleotides in a sequence.

[0175] Various sequence comparison programs identified herein are used. Protein and/or nucleic acid sequence identities (homologies) may be evaluated using any of the variety of sequence comparison algorithms and programs known in the art. Such algorithms and programs include, but are not limited to, TBLASTN, BLASTP, FASTA, TFASTA, and CLUSTALW (Pearson and

Lipman, Proc. Natl. Acad. Sci. USA 85(8):2444-2448, 1988; Altschul et al., J. Mol. Biol. 215(3):403-410, 1990; Thompson et al., Nucleic Acids Res. 22(2):4673-4680, 1994; Higgins et al., Methods Enzymol. 266:383-402, 1996; Altschul et al., J. Mol. Biol. 215(3):403-410, 1990; Altschul et al., Nature Genetics 3:266-272, 1993).

[0176] Homology or identity can be measured using sequence analysis software (e.g., Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, WI 53705). Such software matches similar sequences by assigning degrees of homology to various deletions, substitutions and other modifications. The terms "homology" and "identity" in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same when compared and aligned for maximum correspondence over a comparison window or designated region as measured using any number of sequence comparison algorithms or by manual alignment and visual inspection. For sequence comparison, one sequence can act as a reference sequence, e.g., a sequence of the invention, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters.

[0177] A "comparison window", as used herein, includes reference to a segment of any one of the numbers of contiguous residues. For example, contiguous residues ranging anywhere from 20 to the full length of an exemplary polypeptide or nucleic acid sequence of the invention are compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. If the reference sequence has the requisite sequence identity to an exemplary polypeptide or nucleic acid sequence of the invention, e.g., 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more sequence identity to a sequence of the invention, that sequence is within the scope of the invention. In alternative embodiments, subsequences ranging from about 20 to 600, about 50 to 200, and about 100 to 150 are compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. Methods of alignment of sequence for comparison are well known in the art. Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith & Waterman, Adv. Appl. Math. 2:482, 1981, by the homology alignment algorithm of Needleman & Wunsch, J. Mol. Biol. 48:443, 1970, by the search for similarity method of person & Lipman, Proc. Nat'l. Acad. Sci. USA 85:2444, 1988, by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by manual alignment and visual inspection. Other algorithms for determining homology or identity include, for example, in addition to a BLAST program (Basic Local Alignment Search Tool at the National Center for Biological Information), ALIGN, AMAS (Analysis of Multiply Aligned Sequences), AMPS (Protein Multiple Sequence Alignment), ASSET (Aligned Segment Statistical Evaluation Tool), BANDS, BESTSCOR, BIOSCAN (Biological Sequence Comparative Analysis Node), BLIMPS (BLocks IMProved Searcher), FASTA, Intervals & Points, BMB, CLUSTAL V, CLUSTAL W, CONSENSUS, LCONSENSUS, WCONSENSUS, Smith-Waterman algorithm, DARWIN, Las Vegas algorithm, FNAT (Forced Nucleotide Alignment Tool), Framealign, Framesearch, DYNAMIC, FILTER, FSAP (Fristensky Sequence Analysis Package), GAP (Global Alignment Program), GENAL, GIBBS, GenQuest, ISSC (Sensitive Sequence Comparison), LALIGN (Local Sequence Alignment), LCP (Local Content Program), MACAW (Multiple Alignment Construction & Analysis Workbench), MAP (Multiple Alignment Program), MBLKP, MBLKN, PIMA (Pattern-Induced Multi-sequence Alignment), SAGA (Sequence Alignment by Genetic Algorithm) and WHAT-IF. Such alignment programs can also be used to screen genome databases to identify polynucleotide sequences having substantially identical sequences. A number of genome databases are available, for example, a substantial portion of the human genome is available as part of the Human Genome Sequencing Project (Gibbs, 1995). Several genomes have been sequenced, e.g., *M. genitalium* (Fraser et al., 1995), *M. jannaschii* (Bult et al., 1996), *H. influenzae* (Fleischmann et al., 1995), *E. coli* (Blattner et al., 1997), and yeast (*S. cerevisiae*) (Mewes et al., 1997), and *D. melanogaster* (Adams et al., 2000). Significant progress has also been made in sequencing the genomes of model organism, such as mouse, *C. elegans*, and *Arabidopsis* sp. Databases containing genomic information annotated with some functional information are maintained by different organization, and are accessible via the internet.

[0178] BLAST, BLAST 2.0 and BLAST 2.2.2 algorithms are also used to practice the invention. They are described, e.g., in Altschul (1977) Nuc. Acids Res. 25:3389-3402; Altschul (1990) J. Mol. Biol. 215:403-410. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul (1990) supra). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls

off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=-4 and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength of 3, and expectations (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff (1989) Proc. Natl. Acad. Sci. USA 89:10915) alignments (B) of 50, expectation (E) of 10, M=5, N=-4, and a comparison of both strands. The BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul (1993) Proc. Natl. Acad. Sci. USA 90:5873). One measure of similarity provided by BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a references sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.2, more preferably less than about 0.01, and most preferably less than about 0.001. In one aspect, protein and nucleic acid sequence homologies are evaluated using the Basic Local Alignment Search Tool ("BLAST"). For example, five specific BLAST programs can be used to perform the following task: (1) BLASTP and BLAST3 compare an amino acid query sequence against a protein sequence database; (2) BLASTN compares a nucleotide query sequence against a nucleotide sequence database; (3) BLASTX compares the six-frame conceptual translation products of a query nucleotide sequence (both strands) against a protein sequence database; (4) TBLASTN compares a query protein sequence against a nucleotide sequence database translated in all six reading frames (both strands); and, (5) TBLASTX compares the six-frame translations of a nucleotide query sequence against the six-frame translations of a nucleotide sequence database. The BLAST programs identify homologous sequences by identifying similar segments, which are referred to herein as "high-scoring segment pairs," between a query amino or nucleic acid sequence and a test sequence which is preferably obtained from a protein or nucleic acid sequence database. High-scoring segment pairs are preferably identified (i.e., aligned) by means of a scoring matrix, many of which are known in the art. Preferably, the scoring matrix used is the BLOSUM62 matrix (Gonnet et al., Science 256:1443-1445, 1992; Henikoff and Henikoff, Proteins 17:49-61, 1993). Less preferably, the PAM or PAM250 matrices may also be used (see, e.g., Schwartz and Dayhoff, eds., 1978, Matrices for Detecting Distance Relationships: Atlas of Protein Sequence and Structure, Washington: National Biomedical Research Foundation).

[0179] To determine if a nucleic acid has the requisite sequence identity to be within the scope of the invention, the NCBI BLAST 2.2.2 programs is used, default options to blastp. There are about 38 setting options in the

[0180] BLAST 2.2.2 program. In this example, all default values are used except for the default filtering setting (i.e., all parameters set to default except filtering which is set to OFF); in its place a "-F F" setting is used, which disables filtering. Use of default filtering often results in Karlin-Altschul violations due to short length of sequence.

[0181] The default values used in this exemplary aspect of the invention include:

"Filter for low complexity: ON

Word Size: 3

Matrix: Blosum62

Gap Costs: Existence: 11

Extension:1"

[0182] Other default settings can be: filter for low complexity OFF, word size of 3 for protein, BLOSUM62 matrix, gap existence penalty of -11 and a gap extension penalty of -1. An exemplary NCBI BLAST 2.2.2 program setting has the "-W" option default to 0. This means that, if not set, the word size defaults to 3 for proteins and 11 for nucleotides.

Computer systems and computer program products

[0183] To determine and identify sequence identities, structural homologies, motifs and the like in silico, the sequence of the invention can be stored, recorded, and manipulated on any medium which can be read and accessed by a computer. Accordingly, computers, computer systems, computer readable mediums, computer programs products and the like may have recorded or stored thereon the nucleic acid and polypeptide sequences of the invention. As used herein, the words "recorded" and "stored"

refer to a process for storing information on a computer medium. A skilled artisan can readily adopt any known methods for recording information on a computer readable medium to generate manufactures comprising one or more of the nucleic acid and/or polypeptide sequences of the invention.

[0184] A computer readable medium may have recorded thereon at least one nucleic acid and/or polypeptide sequence of the invention. Computer readable media include magnetically readable media, optically readable media, electronically readable media and magnetic/optical media. For example, the computer readable media may be a hard disk, a floppy disk, a magnetic tape, CD-ROM, Digital Versatile Disk (DVD), Random Access Memory (RAM), or Read Only Memory (ROM) as well as other types of other media known to those skilled in the art.

[0185] An example system 100, which stores and manipulates the sequences and sequence information described herein, is illustrated in block diagram form in Figure 1. As used herein, "a computer system" refers to the hardware components, software components, and data storage components used to analyze a nucleotide or polypeptide sequence of the invention. The computer system 100 can include a processor for processing, accessing and manipulating the sequence data. The processor 105 can be any well-known type of central processing unit, such as, for example, the Pentium III from Intel Corporation, or similar processor from Sun, Motorola, Compaq, AMD or International Business Machines. The computer system 100 is a general purpose system that comprises the processor 105 and one or more internal data storage components 110 for storing data, and one or more data retrieving devices for retrieving the data stored on the data storage components. A skilled artisan can readily appreciate that any one of the currently available computer systems are suitable.

[0186] In one aspect, the computer system 100 includes a processor 105 connected to a bus which is connected to a main memory 115 (preferably implemented as RAM) and one or more internal data storage devices 110, such as a hard drive and/or other computer readable media having data recorded thereon. The computer system 100 can further include one or more data retrieving device 118 for reading the data stored on the internal data storage devices 110. The data retrieving device 118 may represent, for example, a floppy disk drive, a compact disk drive, a magnetic tape drive, or a modem capable of connection to a remote data storage system (e.g., via the internet) etc. In some embodiments, the internal data storage device 110 is a removable computer readable medium such as a floppy disk, a compact disk, a magnetic tape, etc. containing control logic and/or data recorded thereon. The computer system 100 may advantageously include or be programmed by appropriate software for reading the control logic and/or the data from the data storage component once inserted in the data retrieving device. The computer system 100 includes a display 120 which is used to display output to a computer user. It should also be noted that the computer system 100 can be linked to other computer systems 125a-c in a network or wide area network to provide centralized access to the computer system 100. Software for accessing and processing the nucleotide or amino acid sequences of the invention can reside in main memory 115 during execution. In some aspects, the computer system 100 may further comprise a sequence comparison algorithm for comparing a nucleic acid sequence of the invention. The algorithm and sequence(s) can be stored on a computer readable medium. A "sequence comparison algorithm" refers to one or more programs which are implemented (locally or remotely) on the computer system 100 to compare a nucleotide sequence with other nucleotide sequences and/or compounds stored within a data storage means. For example, the sequence comparison algorithm may compare the nucleotide sequences of the invention stored on a computer readable medium to reference sequences stored on a computer readable medium to identify homologies or structural motifs.

[0187] The parameters used with the above algorithms may be adapted depending on the sequence length and degree of homology studied. In some aspects, the parameters may be the default parameters used by the algorithms in the absence of instructions from the user. Figure 2 is a flow diagram illustrating one aspect of a process 200 for comparing a new nucleotide or protein sequence with a database of sequences in order to determine the homology levels between the new sequence and the sequences in the database. The database of sequences can be a private database stored within the computer system 100, or a public database such as GENBANK that is available through the Internet. The process 200 begins at a start state 201 and then moves to a state 202 wherein the new sequence to be compared is stored to a memory in a computer system 100. As discussed above, the memory could be any type of memory, including RAM or an internal storage device. The process 200 then moves to a state 204 wherein a database of sequences is opened for analysis and comparison. The process 200 then moves to a state 206 wherein the first sequence stored in the database is read into a memory on the computer. A comparison is then performed at a state 210 to determine if the first sequence is the same as the second sequence. It is important to note that this step is not limited to performing an exact comparison between the new sequence and the first sequence in the database. Well-known methods are known to those of skill in the art for comparing two nucleotide or protein sequences, even if they are not identical. For example, gaps can be introduced into one sequence in order to raise the homology level between the two tested sequences. The parameters that control whether gaps or other features are introduced into a sequence during comparison are normally entered by the user of the computer system. Once a comparison of the two sequences has been performed at the state 210, a determination is made at a decision state 210 whether the two sequences are the same. Of course, the term "same" is not limited to sequences that are absolutely identical. Sequences that are within the homology parameters entered by the user will be

marked as "same" in the process 200. If a determination is made that the two sequences are the same, the process 200 moves to a state 214 wherein the name of the sequence from the database is displayed to the user. This state notifies the user that the sequence with the displayed name fulfills the homology constraints that were entered. Once the name of the stored sequence is displayed to the user, the process 200 moves to a decision state 218 wherein a determination is made whether more sequences exist in the database. If no more sequences exist in the database, then the process 200 terminates at an end state 220. However, if more sequences do exist in the database, then the process 200 moves to a state 224 wherein a pointer is moved to the next sequence in the database so that it can be compared to the new sequence. In this manner, the new sequence is aligned and compared with every sequence in the database. It should be noted that if a determination had been made at the decision state 212 that the sequences were not homologous, then the process 200 would move immediately to the decision state 218 in order to determine if any other sequences were available in the database for comparison. A computer system may comprise a processor, a data storage device having stored thereon a nucleic acid sequence of the invention and a sequence comparer for conducting the comparison. The sequence comparer may indicate a homology level between the sequences compared or identify structural motifs, or it may identify structural motifs in sequences which are compared to these nucleic acid codes and polypeptide codes. Figure 3 is a flow diagram illustrating one embodiment of a process 250 in a computer for determining whether two sequences are homologous. The process 250 begins at a start state 252 and then moves to a state 254 wherein a first sequence to be compared is stored to a memory. The second sequence to be compared is then stored to a memory at a state 256. The process 250 then moves to a state 260 wherein the first character in the first sequence is read and then to a state 262 wherein the first character of the second sequence is read. It should be understood that if the sequence is a nucleotide sequence, then the character would normally be either A, T, C, G or U. If the sequence is a protein sequence, then it can be a single letter amino acid code so that the first and sequence sequences can be easily compared. A determination is then made at a decision state 264 whether the two characters are the same. If they are the same, then the process 250 moves to a state 268 wherein the next characters in the first and second sequences are read. A determination is then made whether the next characters are the same. If they are, then the process 250 continues this loop until two characters are not the same. If a determination is made that the next two characters are not the same, the process 250 moves to a decision state 274 to determine whether there are any more characters either sequence to read. If there are not any more characters to read, then the process 250 moves to a state 276 wherein the level of homology between the first and second sequences is displayed to the user. The level of homology is determined by calculating the proportion of characters between the sequences that were the same out of the total number of sequences in the first sequence. Thus, if every character in a first 100 nucleotide sequence aligned with an every character in a second sequence, the homology level would be 100%.

[0188] Alternatively, the computer program can compare a reference sequence to a sequence of the invention to determine whether the sequences differ at one or more positions. The program can record the length and identity of inserted, deleted or substituted nucleotides or amino acid residues with respect to the sequence of either the reference or the invention. The computer program may be a program which determines whether a reference sequence contains a single nucleotide polymorphism (SNP) with respect to a sequence of the invention, or, whether a sequence of the invention comprises a SNP of a known sequence. Thus, in some aspects, the computer program is a program which identifies SNPs. The method may be implemented by the computer systems described above and the method illustrated in Figure 3. The method can be performed by reading a sequence of the invention and the reference sequences through the use of the computer program and identifying differences with the computer program.

[0189] The computer based system may comprise an identifier for identifying features within a nucleic acid or polypeptide of the invention. An "identifier" refers to one or more programs which identifies certain features within a nucleic acid sequence. For example, an identifier may comprise a program which identifies an open reading frame (ORF) in a nucleic acid sequence. Figure 4 is a flow diagram illustrating one aspect of an identifier process 300 for detecting the presence of a feature in a sequence. The process 300 begins at a start state 302 and then moves to a state 304 wherein a first sequence that is to be checked for features is stored to a memory 115 in the computer system 100. The process 300 then moves to a state 306 wherein a database of sequence features is opened. Such a database would include a list of each feature's attributes along with the name of the feature. For example, a feature name could be "Initiation Codon" and the attribute would be "ATG". Another example would be the feature name "TAATAA Box" and the feature attribute would be "TAATAA". An example of such a database is produced by the University of Wisconsin Genetics Computer Group. Alternatively, the features may be structural polypeptide motifs such as alpha helices, beta sheets, or functional polypeptide motifs such as enzymatic active sites, helix-turn-helix motifs or other motifs known to those skilled in the art. Once the database of features is opened at the state 306, the process 300 moves to a state 308 wherein the first feature is read from the database. A comparison of the attribute of the first feature with the first sequence is then made at a state 310. A determination is then made at a decision state 316 whether the attribute of the feature was found in the first sequence. If the attribute was found, then the process 300 moves to a state 318 wherein the name of the found feature is displayed to the user. The process 300 then moves to a decision state 320 wherein a determination is made whether move features exist in the database. If no more features do exist, then the process 300 terminates at an end state 324. However, if

more features do exist in the database, then the process 300 reads the next sequence feature at a state 326 and loops back to the state 310 wherein the attribute of the next feature is compared against the first sequence. If the feature attribute is not found in the first sequence at the decision state 316, the process 300 moves directly to the decision state 320 in order to determine if any more features exist in the database. Thus, in one aspect, the invention provides a computer program that identifies open reading frames (ORFs).

[0190] A polypeptide or nucleic acid sequence of the invention can be stored and manipulated in a variety of data processor programs in a variety of formats. For example, a sequence can be stored as text in a word processing file, such as MicrosoftWORD or WORDPERFECT or as an ASCII file in a variety of database programs familiar to those of skill in the art, such as DB2, SYBASE, or ORACLE. In addition, many computer programs and databases may be used as sequence comparison algorithms, identifiers, or sources of reference nucleotide sequences or polypeptide sequences to be compared to a nucleic acid sequence of the invention. The programs and databases used to practice the invention include, but are not limited to: MacPattern (EMBL), DiscoveryBase (Molecular Applications Group), GeneMine (Molecular Applications Group), Look (Molecular Applications Group), MacLook (Molecular Applications Group), BLAST and BLAST2 (NCBI), BLASTN and BLASTX (Altschul et al, J. Mol. Biol. 215: 403, 1990), FASTA (Pearson and Lipman, Proc. Natl. Acad. Sci. USA, 85: 2444, 1988), FASTDB (Brutlag et al. Comp. App. Biosci. 6:237-245, 1990), Catalyst (Molecular Simulations Inc.), Catalyst/SHAPE (Molecular Simulations Inc.), Cerius2.DBAccess (Molecular Simulations Inc.), HypoGen (Molecular Simulations Inc.), Insight II, (Molecular Simulations Inc.), Discover (Molecular Simulations Inc.), CHARMM (Molecular Simulations Inc.), Felix (Molecular Simulations Inc.), DelPhi, (Molecular Simulations Inc.), QuanteMM, (Molecular Simulations Inc.), Homology (Molecular Simulations Inc.), Modeler (Molecular Simulations Inc.), ISIS (Molecular Simulations Inc.), Quanta/Protein Design (Molecular Simulations Inc.), WebLab (Molecular Simulations Inc.), WebLab Diversity Explorer (Molecular Simulations Inc.), Gene Explorer (Molecular Simulations Inc.), SeqFold (Molecular Simulations Inc.), the MDL Available Chemicals Directory database, the MDL Drug Data Report data base, the Comprehensive Medicinal Chemistry database, Derwent's World Drug Index database, the BioByteMasterFile database, the Genbank database, and the Genseq database. Many other programs and data bases would be apparent to one of skill in the art given the present disclosure.

[0191] Motifs which may be detected using the above programs include sequences encoding leucine zippers, helix-turn-helix motifs, glycosylation sites, ubiquitination sites, alpha helices, and beta sheets, signal sequences encoding signal peptides which direct the secretion of the encoded proteins, sequences implicated in transcription regulation such as homeoboxes, acidic stretches, enzymatic active sites, substrate binding sites, and enzymatic cleavage sites.

Hybridization of nucleic acids

[0192] The invention provides isolated or recombinant nucleic acids that hybridize under stringent conditions to a sequence of the invention or a nucleic acid that encodes a polypeptide of the invention.

The stringent conditions can be highly stringent conditions, medium stringent conditions and/or low stringent conditions, including the high and reduced stringency conditions described herein. It may be the stringency of the wash conditions that set forth the conditions which determine whether a nucleic acid is within the scope of the invention, as discussed below.

[0193] Nucleic acids as defined by their ability to hybridize under stringent conditions can be between about five residues and the full length of nucleic acid of the invention; e.g., they can be at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 55, 60, 65, 70, 75, 80, 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, or more, residues in length. Nucleic acids shorter than full length are also included. These nucleic acids can be useful as, e.g., hybridization probes, labeling probes, PCR oligonucleotide probes, iRNA, antisense or sequences encoding antibody binding peptides (epitopes), motifs, active sites and the like.

[0194] Nucleic acids may be defined by their ability to hybridize under high stringency comprising conditions of about 50% formamide at about 37°C to 42°C, or by their ability to hybridize under reduced stringency comprising conditions in about 35% to 25% formamide at about 30°C to 35°C.

[0195] Alternatively, nucleic acids may be defined by their ability to hybridize under high stringency comprising conditions at 42°C in 50% formamide, 5X SSPE, 0.3% SDS, and a repetitive sequence blocking nucleic acid, such as cot-1 or salmon sperm DNA (e.g., 200 n/ml sheared and denatured salmon sperm DNA), or by their ability to hybridize under reduced stringency conditions comprising 35% formamide at a reduced temperature of 35°C.

[0196] Following hybridization, the filter may be washed with 6X SSC, 0.5% SDS at 50°C. These conditions are considered to be "moderate" conditions above 25% formamide and "low" conditions below 25% formamide. A specific example of "moderate"

hybridization conditions is when the above hybridization is conducted at 30% formamide. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 10% formamide.

[0197] The temperature range corresponding to a particular level of stringency can be further narrowed by calculating the purine to pyrimidine ratio of the nucleic acid of interest and adjusting the temperature accordingly. Nucleic acids of the invention are also defined by their ability to hybridize under high, medium, and low stringency conditions as set forth in Ausubel and Sambrook. Variations on the above ranges and conditions are well known in the art. Hybridization conditions are discussed further, below.

[0198] The above procedure may be modified to identify nucleic acids having decreasing levels of homology to the probe sequence. For example, to obtain nucleic acids of decreasing homology to the detectable probe, less stringent conditions may be used. For example, the hybridization temperature may be decreased in increments of 5°C from 68°C to 42°C in a hybridization buffer having a Na⁺ concentration of approximately 1M. Following hybridization, the filter may be washed with 2X SSC, 0.5% SDS at the temperature of hybridization. These conditions are considered to be "moderate" conditions above 50°C and "low" conditions below 50°C. A specific example of "moderate" hybridization conditions is when the above hybridization is conducted at 55°C. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 45°C.

[0199] Alternatively, the hybridization may be carried out in buffers, such as 6X SSC, containing formamide at a temperature of 42°C. In this case, the concentration of formamide in the hybridization buffer may be reduced in 5% increments from 50% to 0% to identify clones having decreasing levels of homology to the probe. Following hybridization, the filter may be washed with 6X SSC, 0.5% SDS at 50°C. These conditions are considered to be "moderate" conditions above 25% formamide and "low" conditions below 25% formamide. A specific example of "moderate" hybridization conditions is when the above hybridization is conducted at 30% formamide. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 10% formamide.

[0200] However, the selection of a hybridization format is not critical - it is the stringency of the wash conditions that set forth the conditions which determine whether a nucleic acid is within the scope of the invention. Wash conditions used to identify nucleic acids within the scope of the invention include, e.g.: a salt concentration of about 0.02 molar at pH 7 and a temperature of at least about 50°C or about 55°C to about 60°C; or, a salt concentration of about 0.15 M NaCl at 72°C for about 15 minutes; or, a salt concentration of about 0.2X SSC at a temperature of at least about 50°C or about 55°C to about 60°C for about 15 to about 20 minutes; or, the hybridization complex is washed twice with a solution with a salt concentration of about 2X SSC containing 0.1% SDS at room temperature for 15 minutes and then washed twice by 0.1X SSC containing 0.1% SDS at 68°C for 15 minutes; or, equivalent conditions. See Sambrook, Tijssen and Ausubel for a description of SSC buffer and equivalent conditions.

[0201] These methods may be used to isolate nucleic acids of the invention.

Oligonucleotides probes and methods for using them

[0202] The invention also provides nucleic acid probes that can be used, e.g., for identifying nucleic acids encoding a polypeptide with a pectate lyase activity or fragments thereof or for identifying pectate lyase genes. The probe comprises at least 60 to 100 or 60 to 150 consecutive bases of a sequence as set forth in a nucleic acid of the invention. The probes identify a nucleic acid by binding and/or hybridization. The probes can be used in arrays, see discussion below, including, e.g., capillary arrays. The probes of the invention can also be used to isolate other nucleic acids or polypeptides.

[0203] The probes of the invention can be used to determine whether a biological sample, such as a soil sample, contains an organism having a nucleic acid sequence of the invention or an organism from which the nucleic acid was obtained. In such procedures, a biological sample potentially harboring the organism from which the nucleic acid was isolated is obtained and nucleic acids are obtained from the sample. The nucleic acids are contacted with the probe under conditions which permit the probe to specifically hybridize to any complementary sequences present in the sample. Where necessary, conditions which permit the probe to specifically hybridize to complementary sequences may be determined by placing the probe in contact with complementary sequences from samples known to contain the complementary sequence, as well as control sequences which do not contain the complementary sequence. Hybridization conditions, such as the salt concentration of the hybridization buffer, the formamide concentration of the hybridization buffer, or the hybridization temperature, may be varied to identify conditions which allow the probe to hybridize specifically to complementary nucleic acids (see discussion on specific hybridization conditions).

[0204] If the sample contains the organism from which the nucleic acid was isolated, specific hybridization of the probe is then detected. Hybridization may be detected by labeling the probe with a detectable agent such as a radioactive isotope, a

fluorescent dye or an enzyme capable of catalyzing the formation of a detectable product. Many methods for using the labeled probes to detect the presence of complementary nucleic acids in a sample are familiar to those skilled in the art. These include Southern Blots, Northern Blots, colony hybridization procedures, and dot blots. Protocols for each of these procedures are provided in Ausubel and Sambrook.

[0205] Alternatively, more than one probe (at least one of which is capable of specifically hybridizing to any complementary sequences which are present in the nucleic acid sample), may be used in an amplification reaction to determine whether the sample contains an organism containing a nucleic acid sequence of the invention (e.g., an organism from which the nucleic acid was isolated). In one aspect, the probes comprise oligonucleotides. In one aspect, the amplification reaction may comprise a PCR reaction. PCR protocols are described in Ausubel and Sambrook (see discussion on amplification reactions). In such procedures, the nucleic acids in the sample are contacted with the probes, the amplification reaction is performed, and any resulting amplification product is detected. The amplification product may be detected by performing gel electrophoresis on the reaction products and staining the gel with an intercalator such as ethidium bromide. Alternatively, one or more of the probes may be labeled with a radioactive isotope and the presence of a radioactive amplification product may be detected by autoradiography after gel electrophoresis.

[0206] Probes derived from sequences near the 3' or 5' ends of a nucleic acid sequence of the invention can also be used in chromosome walking procedures to identify clones containing additional, e.g., genomic sequences. Such methods allow the isolation of genes which encode additional proteins of interest from the host organism.

[0207] In one aspect, nucleic acid sequences of the invention are used as probes to identify and isolate related nucleic acids. In some aspects, the so-identified related nucleic acids may be cDNAs or genomic DNAs from organisms other than the one from which the nucleic acid of the invention was first isolated. In such procedures, a nucleic acid sample is contacted with the probe under conditions which permit the probe to specifically hybridize to related sequences. Hybridization of the probe to nucleic acids from the related organism is then detected using any of the methods described above.

[0208] In nucleic acid hybridization reactions, the conditions used to achieve a particular level of stringency can vary, depending on the nature of the nucleic acids being hybridized. For example, the length, degree of complementarity, nucleotide sequence composition (e.g., GC v. AT content), and nucleic acid type (e.g., RNA v. DNA) of the hybridizing regions of the nucleic acids can be considered in selecting hybridization conditions. An additional consideration is whether one of the nucleic acids is immobilized, for example, on a filter. Hybridization can be carried out under conditions of low stringency, moderate stringency or high stringency. As an example of nucleic acid hybridization, a polymer membrane containing immobilized denatured nucleic acids is first prehybridized for 30 minutes at 45°C in a solution consisting of 0.9 M NaCl, 50 mM NaH₂PO₄, pH 7.0, 5.0 mM Na₂EDTA, 0.5% SDS, 10X Denhardt's, and 0.5 mg/ml polyriboadenylic acid. Approximately 2 X 10⁷ cpm (specific activity 4-9 X 10⁸ cpm/μg) of ³²P end-labeled oligonucleotide probe can then be added to the solution. After 12-16 hours of incubation, the membrane is washed for 30 minutes at room temperature (RT) in 1X SET (150 mM NaCl, 20 mM Tris hydrochloride, pH 7.8, 1 mM Na₂EDTA) containing 0.5% SDS, followed by a 30 minute wash in fresh 1X SET at T_m-10°C for the oligonucleotide probe. The membrane is then exposed to auto-radiographic film for detection of hybridization signals.

[0209] By varying the stringency of the hybridization conditions used to identify nucleic acids, such as cDNAs or genomic DNAs, which hybridize to the detectable probe, nucleic acids having different levels of homology to the probe can be identified and isolated. Stringency may be varied by conducting the hybridization at varying temperatures below the melting temperatures of the probes. The melting temperature, T_m, is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly complementary probe. Very stringent conditions are selected to be equal to or about 5°C lower than the T_m for a particular probe. The melting temperature of the probe may be calculated using the following exemplary formulas. For probes between 14 and 70 nucleotides in length the melting temperature (T_m) is calculated using the formula: T_m=81.5+16.6(log [Na⁺])+0.41(fraction G+C)-(600/N) where N is the length of the probe. If the hybridization is carried out in a solution containing formamide, the melting temperature may be calculated using the equation: T_m=81.5+16.6(log [Na⁺])+0.41(fraction G+C)-(0.63% formamide)-(600/N) where N is the length of the probe. Prehybridization may be carried out in 6X SSC, 5X Denhardt's reagent, 0.5% SDS, 100μg denatured fragmented salmon sperm DNA or 6X SSC, 5X Denhardt's reagent, 0.5% SDS, 100μg denatured fragmented salmon sperm DNA, 50% formamide. Formulas for SSC and Denhardt's and other solutions are listed, e.g., in Sambrook.

[0210] Hybridization is conducted by adding the detectable probe to the prehybridization solutions listed above. Where the probe comprises double stranded DNA, it is denatured before addition to the hybridization solution. The filter is contacted with the hybridization solution for a sufficient period of time to allow the probe to hybridize to cDNAs or genomic DNAs containing sequences complementary thereto or homologous thereto. For probes over 200 nucleotides in length, the hybridization may be

carried out at 15-25°C below the T_m . For shorter probes, such as oligonucleotide probes, the hybridization may be conducted at 5-10°C below the T_m . In one aspect, hybridizations in 6X SSC are conducted at approximately 68°C. In one aspect, hybridizations in 50% formamide containing solutions are conducted at approximately 42°C. All of the foregoing hybridizations would be considered to be under conditions of high stringency.

[0211] Following hybridization, the filter is washed to remove any non-specifically bound detectable probe. The stringency used to wash the filters can also be varied depending on the nature of the nucleic acids being hybridized, the length of the nucleic acids being hybridized, the degree of complementarity, the nucleotide sequence composition (e.g., GC v. AT content), and the nucleic acid type (e.g., RNA v. DNA). Examples of progressively higher stringency condition washes are as follows: 2X SSC, 0.1% SDS at room temperature for 15 minutes (low stringency); 0.1X SSC, 0.5% SDS at room temperature for 30 minutes to 1 hour (moderate stringency); 0.1X SSC, 0.5% SDS for 15 to 30 minutes at between the hybridization temperature and 68°C (high stringency); and 0.15M NaCl for 15 minutes at 72°C (very high stringency). A final low stringency wash can be conducted in 0.1X SSC at room temperature. The examples above are merely illustrative of one set of conditions that can be used to wash filters. One of skill in the art would know that there are numerous recipes for different stringency washes.

[0212] Nucleic acids which have hybridized to the probe can be identified by autoradiography or other conventional techniques. The above procedure may be modified to identify nucleic acids having decreasing levels of homology to the probe sequence. For example, to obtain nucleic acids of decreasing homology to the detectable probe, less stringent conditions may be used. For example, the hybridization temperature may be decreased in increments of 5°C from 68°C to 42°C in a hybridization buffer having a Na^+ concentration of approximately 1M. Following hybridization, the filter may be washed with 2X SSC, 0.5% SDS at the temperature of hybridization. These conditions are considered to be "moderate" conditions above 50°C and "low" conditions below 50°C. An example of "moderate" hybridization conditions is when the above hybridization is conducted at 55°C. An example of "low stringency" hybridization conditions is when the above hybridization is conducted at 45°C.

[0213] Alternatively, the hybridization may be carried out in buffers, such as 6X SSC, containing formamide at a temperature of 42°C. In this case, the concentration of formamide in the hybridization buffer may be reduced in 5% increments from 50% to 0% to identify clones having decreasing levels of homology to the probe. Following hybridization, the filter may be washed with 6X SSC, 0.5% SDS at 50°C. These conditions are considered to be "moderate" conditions above 25% formamide and "low" conditions below 25% formamide. A specific example of "moderate" hybridization conditions is when the above hybridization is conducted at 30% formamide. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 10% formamide.

[0214] These probes and methods described can be used to isolate nucleic acids having a sequence with at least about 99%, 98%, 97%, at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 65%, at least 60%, at least 55%, or at least 50% homology to a nucleic acid sequence of the invention comprising at least about 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 250, 300, 350, 400, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, or more consecutive bases thereof, and the sequences complementary thereto. Homology may be measured using an alignment algorithm, as discussed herein. For example, the homologous polynucleotides may have a coding sequence which is a naturally occurring allelic variant of one of the coding sequences described herein. Such allelic variants may have a substitution, deletion or addition of one or more nucleotides when compared to a nucleic acid of the invention.

[0215] Additionally, the probes and methods can be used to isolate nucleic acids which encode polypeptides having at least about 99%, at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 65%, at least 60%, at least 55%, or at least 50% sequence identity (homology) to a polypeptide of the invention comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids, as determined using a sequence alignment algorithm (e.g., such as the FASTA version 3.0t78 algorithm with the default parameters, or a BLAST 2.2.2 program with exemplary settings as set forth herein).

Inhibiting Expression of Pectate lyase

[0216] The invention provides nucleic acids complementary to (e.g., antisense sequences to) the nucleic acid sequences of the invention. Antisense sequences are capable of inhibiting the transport, splicing or transcription of pectate lyase-encoding genes. The inhibition can be effected through the targeting of genomic DNA or messenger RNA. The transcription or function of targeted nucleic acid can be inhibited, for example, by hybridization and/or cleavage. One particularly useful set of inhibitors provided includes oligonucleotides which are able to either bind pectate lyase gene or message, in either case preventing or inhibiting the production or function of pectate lyase. The association can be through sequence specific hybridization. Another useful class of inhibitors includes oligonucleotides which cause inactivation or cleavage of pectate lyase message. The oligonucleotide can have

enzyme activity which causes such cleavage, such as ribozymes. The oligonucleotide can be chemically modified or conjugated to an enzyme or composition capable of cleaving the complementary nucleic acid. A pool of many different such oligonucleotides can be screened for those with the desired activity. Thus, the invention provides various compositions for the inhibition of pectate lyase expression on a nucleic acid and/or protein level, e.g., antisense, iRNA and ribozymes comprising pectate lyase sequences of the invention and the anti-pectate lyase antibodies described herein.

[0217] Inhibition of pectate lyase expression can have a variety of industrial applications. For example, inhibition of pectate lyase expression can slow or prevent "soft-rot" spoilage. "Soft-rot" spoilage occurs when pectin, a major structural polysaccharide in the plant cell wall, is enzymatically degraded. This can lead to the deterioration, or rot, of fruits and vegetables. In one aspect, use of compositions of the invention that inhibit the expression and/or activity of pectate lyases, e.g., antibodies, antisense oligonucleotides, ribozymes and RNAi, are used to slow or prevent "soft-rot" spoilage. Thus, methods and compositions comprise application onto a plant or plant product (e.g., a fruit, seed, root, leaf, etc.) of antibodies, antisense oligonucleotides, ribozymes and RNAi of the disclosure to slow or prevent "soft-rot" spoilage. These compositions also can be expressed by the plant (e.g., a transgenic plant) or another organism (e.g., a bacterium or other microorganism transformed with a pectate lyase gene of the invention).

[0218] Inhibition of pectate lyase expression also can prevent or slow the normal growth of the powdery mildew pathogen *Erysiphe cichoracearum*. This powdery mildew resistance represents a form of disease resistance based on the loss of a gene required during a compatible interaction rather than the activation of known host defense pathways. See, e.g., Vogel (2002) Plant Cell 14:2095-2106. Thus, methods and compositions comprise application onto a plant or plant product (e.g., a fruit, seed, root, leaf, etc.) of antibodies, antisense oligonucleotides, ribozymes and RNAi of the disclosure to slow or prevent growth of the powdery mildew pathogen.

Antisense Oligonucleotides

[0219] The disclosure provides antisense oligonucleotides capable of binding pectate lyase message which can inhibit proteolytic activity by targeting mRNA. Strategies for designing antisense oligonucleotides are well described in the scientific and patent literature, and the skilled artisan can design such pectate lyase oligonucleotides using the novel reagents of the invention. For example, gene walking/ RNA mapping protocols to screen for effective antisense oligonucleotides are well known in the art, see, e.g., Ho (2000) Methods Enzymol. 314:168-183, describing an RNA mapping assay, which is based on standard molecular techniques to provide an easy and reliable method for potent antisense sequence selection. See also Smith (2000) Eur. J. Pharm. Sci. 11:191-198.

[0220] Naturally occurring nucleic acids are used as antisense oligonucleotides. The antisense oligonucleotides can be of any length; for example, in alternative aspects, the antisense oligonucleotides are between about 5 to 100, about 10 to 80, about 15 to 60, about 18 to 40. The optimal length can be determined by routine screening. The antisense oligonucleotides can be present at any concentration. The optimal concentration can be determined by routine screening. A wide variety of synthetic, non-naturally occurring nucleotide and nucleic acid analogues are known which can address this potential problem. For example, peptide nucleic acids (PNAs) containing non-ionic backbones, such as N-(2-aminoethyl) glycine units can be used. Antisense oligonucleotides having phosphorothioate linkages can also be used, as described in WO 97/03211; WO 96/39154; Mata (1997) Toxicol Appl Pharmacol 144:189-197; Antisense Therapeutics, ed. Agrawal (Humana Press, Totowa, N.J., 1996). Antisense oligonucleotides having synthetic DNA backbone analogues provided by the invention can also include phosphoro-dithioate, methylphosphonate, phosphoramidate, alkyl phosphotriester, sulfamate, 3'-thioacetal, methylene(methylimino), 3'-N-carbamate, and morpholino carbamate nucleic acids, as described above.

[0221] Combinatorial chemistry methodology can be used to create vast numbers of oligonucleotides that can be rapidly screened for specific oligonucleotides that have appropriate binding affinities and specificities toward any target, such as the sense and antisense pectate lyase sequences of the invention (see, e.g., Gold (1995) J. of Biol. Chem. 270:13581-13584).

Inhibitory Ribozymes

[0222] The disclosure provides ribozymes capable of binding pectate lyase message. These ribozymes can inhibit pectate lyase activity by, e.g., targeting mRNA. Strategies for designing ribozymes and selecting the pectate lyase-specific antisense sequence for targeting are well described in the scientific and patent literature, and the skilled artisan can design such ribozymes using the novel reagents of the invention. Ribozymes act by binding to a target RNA through the target RNA binding portion of a ribozyme

which is held in close proximity to an enzymatic portion of the RNA that cleaves the target RNA. Thus, the ribozyme recognizes and binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cleave and inactivate the target RNA. Cleavage of a target RNA in such a manner will destroy its ability to direct synthesis of an encoded protein if the cleavage occurs in the coding sequence. After a ribozyme has bound and cleaved its RNA target, it can be released from that RNA to bind and cleave new targets repeatedly.

[0223] In some circumstances, the enzymatic nature of a ribozyme can be advantageous over other technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its transcription, translation or association with another molecule) as the effective concentration of ribozyme necessary to effect a therapeutic treatment can be lower than that of an antisense oligonucleotide. This potential advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, a ribozyme is typically a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding, but also on the mechanism by which the molecule inhibits the expression of the RNA to which it binds. That is, the inhibition is caused by cleavage of the RNA target and so specificity is defined as the ratio of the rate of cleavage of the targeted RNA over the rate of cleavage of non-targeted RNA. This cleavage mechanism is dependent upon factors additional to those involved in base pairing. Thus, the specificity of action of a ribozyme can be greater than that of antisense oligonucleotide binding the same RNA site.

[0224] The ribozyme, e.g., an enzymatic ribozyme RNA molecule, can be formed in a hammerhead motif, a hairpin motif, as a hepatitis delta virus motif, a group I intron motif and/or an RNaseP-like RNA in association with an RNA guide sequence. Examples of hammerhead motifs are described by, e.g., Rossi (1992) *Aids Research and Human Retroviruses* 8:183; hairpin motifs by Hampel (1989) *Biochemistry* 28:4929, and Hampel (1990) *Nuc. Acids Res.* 18:299; the hepatitis delta virus motif by Perrotta (1992) *Biochemistry* 31:16; the RNaseP motif by Guerrier-Takada (1983) *Cell* 35:849; and the group I intron by Cech U.S. Pat. No. 4,987,071. The recitation of these specific motifs is not intended to be limiting. Those skilled in the art will recognize that a ribozyme, e.g., an enzymatic RNA molecule can have a specific substrate binding site complementary to one or more of the target gene RNA regions. A ribozyme of the invention can have a nucleotide sequence within or surrounding that substrate binding site which imparts an RNA cleaving activity to the molecule.

RNA interference (RNAi)

[0225] The disclosure provides an RNA inhibitory molecule, a so-called "RNAi" molecule, comprising a pectate lyase sequence of the invention. The RNAi molecule comprises a double-stranded RNA (dsRNA) molecule. The RNAi can inhibit expression of a pectate lyase gene. In one aspect, the RNAi is about 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or more duplex nucleotides in length. While not limited by any particular mechanism of action, the RNAi can enter a cell and cause the degradation of a single-stranded RNA (ssRNA) of similar or identical sequences, including endogenous mRNAs. When a cell is exposed to double-stranded RNA (dsRNA), mRNA from the homologous gene is selectively degraded by a process called RNA interference (RNAi). A possible basic mechanism behind RNAi is the breaking of a double-stranded RNA (dsRNA) matching a specific gene sequence into short pieces called short interfering RNA, which trigger the degradation of mRNA that matches its sequence. In one aspect, the RNAi's are used in gene-silencing therapeutics, see, e.g., Shuey (2002) *Drug Discov. Today* 7:1040-1046. Methods may selectively degrade RNA using the RNAi's. The process may be practiced *in vitro*, *ex vivo* or *in vivo*. The RNAi molecules can be used to generate a loss-of-function mutation in a cell, an organ or an animal. Methods for making and using RNAi molecules for selectively degrade RNA are well known in the art, see, e.g., U.S. Patent No. 6,506,559; 6,511,824; 6,515,109; 6,489,127.

Modification of Nucleic Acids

[0226] Methods may generate variants of the nucleic acids of the invention, the variant nucleic acids generated by these methods (e.g., SEQ ID NO:133) and polypeptides encoded by them (e.g., SEQ ID NO: 134) are discussed below. These methods can be repeated or used in various combinations to generate pectate lyases having an altered or different activity or an altered or different stability from that of a pectate lyase encoded by the template nucleic acid. These methods also can be repeated or used in various combinations, e.g., to generate variations in gene/ message expression, message translation or message stability. In another aspect, the genetic composition of a cell is altered by, e.g., modification of a homologous gene *in vitro*, *in vivo* or *ex vivo*, followed by its reinsertion into the cell.

[0227] A nucleic acid of the invention can be altered by any means. For example, random or stochastic methods, or, non-stochastic, or "directed evolution," methods, see, e.g., U.S. Patent No. 6,361,974. Methods for random mutation of genes are well known in the art, see, e.g., U.S. Patent No. 5,830,696. For example, mutagens can be used to randomly mutate a gene. Mutagens

include, e.g., ultraviolet light or gamma irradiation, or a chemical mutagen, e.g., mitomycin, nitrous acid, photoactivated psoralens, alone or in combination, to induce DNA breaks amenable to repair by recombination. Other chemical mutagens include, for example, sodium bisulfite, nitrous acid, hydroxylamine, hydrazine or formic acid. Other mutagens are analogues of nucleotide precursors, e.g., nitrosoguanidine, 5-bromouracil, 2-aminopurine, or acridine. These agents can be added to a PCR reaction in place of the nucleotide precursor, thereby mutating the sequence. Intercalating agents such as proflavine, acriflavine, quinacrine and the like can be used.

[0228] Any technique in molecular biology can be used, e.g., random PCR mutagenesis, see, e.g., Rice (1992) *Proc. Natl. Acad. Sci. USA* 89:5467-5471; or, combinatorial multiple cassette mutagenesis, see, e.g., Cramer (1995) *Biotechniques* 18:194-196. Alternatively, nucleic acids, e.g., genes, can be reassembled after random, or "stochastic," fragmentation, see, e.g., U.S. Patent Nos. 6,291,242; 6,287,862; 6,287,861; 5,955,358; 5,830,721; 5,824,514; 5,811,238; 5,605,793. In alternative aspects, modifications, additions or deletions are introduced by error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, *in vivo* mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly, gene site saturation mutagenesis (GSSM™), synthetic ligation reassembly (SLR), recombination, recursive sequence recombination, phosphothioate-modified DNA mutagenesis, uracil-containing template mutagenesis, gapped duplex mutagenesis, point mismatch repair mutagenesis, repair-deficient host strain mutagenesis, chemical mutagenesis, radiogenic mutagenesis, deletion mutagenesis, restriction-selection mutagenesis, restriction-purification mutagenesis, artificial gene synthesis, ensemble mutagenesis, chimeric nucleic acid multimer creation, and/or a combination of these and other methods.

[0229] The following publications describe a variety of recursive recombination procedures and/or methods which can be incorporated into the methods of the invention: Stemmer (1999) "Molecular breeding of viruses for targeting and other clinical properties" *Tumor Targeting* 4:1-4; Ness (1999) *Nature Biotechnology* 17:893-896; Chang (1999) "Evolution of a cytokine using DNA family shuffling" *Nature Biotechnology* 17:793-797; Minshull (1999) "Protein evolution by molecular breeding" *Current Opinion in Chemical Biology* 3:284-290; Christians (1999) "Directed evolution of thymidine kinase for AZT phosphorylation using DNA family shuffling" *Nature Biotechnology* 17:259-264; Cramer (1998) "DNA shuffling of a family of genes from diverse species accelerates directed evolution" *Nature* 391:288-291; Cramer (1997) "Molecular evolution of an arsenate detoxification pathway by DNA shuffling," *Nature Biotechnology* 15:436-438; Zhang (1997) "Directed evolution of an effective fucosidase from a galactosidase by DNA shuffling and screening" *Proc. Natl. Acad. Sci. USA* 94:4504-4509; Patten et al. (1997) "Applications of DNA Shuffling to Pharmaceuticals and Vaccines" *Current Opinion in Biotechnology* 8:724-733; Cramer et al. (1996) "Construction and evolution of antibody-phage libraries by DNA shuffling" *Nature Medicine* 2:100-103; Gates et al. (1996) "Affinity selective isolation of ligands from peptide libraries through display on a lac repressor 'headpiece dimer'" *Journal of Molecular Biology* 255:373-386; Stemmer (1996) "Sexual PCR and Assembly PCR" In: *The Encyclopedia of Molecular Biology*. VCH Publishers, New York. pp.447-457; Cramer and Stemmer (1995) "Combinatorial multiple cassette mutagenesis creates all the permutations of mutant and wildtype cassettes" *Biotechniques* 18:194-195; Stemmer et al. (1995) "Single-step assembly of a gene and entire plasmid from large numbers of oligodeoxyribonucleotides" *Gene*, 164:49-53; Stemmer (1995) "The Evolution of Molecular Computation" *Science* 270: 1510; Stemmer (1995) "Searching Sequence Space" *Bio/Technology* 13:549-553; Stemmer (1994) "Rapid evolution of a protein in vitro by DNA shuffling" *Nature* 370:389-391; and Stemmer (1994) "DNA shuffling by random fragmentation and reassembly: In vitro recombination for molecular evolution." *Proc. Natl. Acad. Sci. USA* 91:10747-10751.

[0230] Mutational methods of generating diversity include, for example, site-directed mutagenesis (Ling et al. (1997) "Approaches to DNA mutagenesis: an overview" *Anal Biochem.* 254(2): 157-178; Dale et al. (1996) "Oligonucleotide-directed random mutagenesis using the phosphorothioate method" *Methods Mol. Biol.* 57:369-374; Smith (1985) "In vitro mutagenesis" *Ann. Rev. Genet.* 19:423-462; Botstein & Shortle (1985) "Strategies and applications of in vitro mutagenesis" *Science* 229:1193-1201; Carter (1986) "Site-directed mutagenesis" *Biochem. J.* 237:1-7; and Kunkel (1987) "The efficiency of oligonucleotide directed mutagenesis" in *Nucleic Acids & Molecular Biology* (Eckstein, F. and Lilley, D. M. J. eds., Springer Verlag, Berl in)); mutagenesis using uracil containing templates (Kunkel (1985) "Rapid and efficient site-specific mutagenesis without phenotypic selection" *Proc. Natl. Acad. Sci. USA* 82:488-492; Kunkel et al. (1987) "Rapid and efficient site-specific mutagenesis without phenotypic selection" *Methods in Enzymol.* 154, 367-382; and Bass et al. (1988) "Mutant Trp repressors with new DNA-binding specificities" *Science* 242:240-245); oligonucleotide-directed mutagenesis (Methods in Enzymol. 100: 468-500 (1983); Methods in Enzymol. 154: 329-350 (1987); Zoller & Smith (1982) "Oligonucleotide-directed mutagenesis using M13-derived vectors: an efficient and general procedure for the production of point mutations in any DNA fragment" *Nucleic Acids Res.* 10:6487-6500; Zoller & Smith (1983) "Oligonucleotide-directed mutagenesis of DNA fragments cloned into M13 vectors" *Methods in Enzymol.* 100:468-500; and Zoller & Smith (1987) Oligonucleotide-directed mutagenesis: a simple method using two oligonucleotide primers and a single-stranded DNA template" *Methods in Enzymol.* 154:329-350); phosphorothioate-modified DNA mutagenesis (Taylor et al. (1985) "The use of phosphorothioate-modified DNA in restriction enzyme reactions to prepare nicked DNA" *Nucl. Acids Res.* 13: 8749-8764; Taylor et al. (1985) "The rapid generation of oligonucleotide-directed mutations at high frequency using phosphorothioate-modified DNA" *Nucl. Acids Res.* 13: 8765-8787 (1985); Nakamaye (1986) "Inhibition of restriction endonuclease

Nci I cleavage by phosphorothioate groups and its application to oligonucleotide-directed mutagenesis" Nucl. Acids Res. 14: 9679-9698; Sayers et al. (1988) "Y-T Exonucleases in phosphorothioate-based oligonucleotide-directed mutagenesis" Nucl. Acids Res. 16:791-802; and Sayers et al. (1988) "Strand specific cleavage of phosphorothioate-containing DNA by reaction with restriction endonucleases in the presence of ethidium bromide" Nucl. Acids Res. 16: 803-814); mutagenesis using gapped duplex DNA (Kramer et al. (1984) "The gapped duplex DNA approach to oligonucleotide-directed mutation construction" Nucl. Acids Res. 12: 9441-9456; Kramer & Fritz (1987) Methods in Enzymol. "Oligonucleotide-directed construction of mutations via gapped duplex DNA" 154:350-367; Kramer et al. (1988) "Improved enzymatic in vitro reactions in the gapped duplex DNA approach to oligonucleotide-directed construction of mutations" Nucl. Acids Res. 16: 7207; and Fritz et al. (1988) "Oligonucleotide-directed construction of mutations: a gapped duplex DNA procedure without enzymatic reactions in vitro" Nucl. Acids Res. 16: 6987-6999).

[0231] Additional protocols that can be used to practice the invention include point mismatch repair (Kramer (1984) "Point Mismatch Repair" Cell 38:879-887), mutagenesis using repair-deficient host strains (Carter et al. (1985) "Improved oligonucleotide site-directed mutagenesis using M13 vectors" Nucl. Acids Res. 13: 4431-4443; and Carter (1987) "Improved oligonucleotide-directed mutagenesis using M13 vectors" Methods in Enzymol. 154: 382-403), deletion mutagenesis (Eghtedarzadeh (1986) "Use of oligonucleotides to generate large deletions" Nucl. Acids Res. 14: 5115), restriction-selection and restriction-selection and restriction-purification (Wells et al. (1986) "Importance of hydrogen-bond formation in stabilizing the transition state of subtilisin" Phil. Trans. R. Soc. Lond. A 317: 415-423), mutagenesis by total gene synthesis (Nambiar et al. (1984) "Total synthesis and cloning of a gene coding for the ribonuclease S protein" Science 223: 1299-1301; Sakamar and Khorana (1988) "Total synthesis and expression of a gene for the α -subunit of bovine rod outer segment guanine nucleotide-binding protein (transducin)" Nucl. Acids Res. 14: 6361-6372; Wells et al. (1985) "Cassette mutagenesis: an efficient method for generation of multiple mutations at defined sites" Gene 34:315-323; and Grundstrom et al. (1985) "Oligonucleotide-directed mutagenesis by microscale 'shot-gun' gene synthesis" Nucl. Acids Res. 13: 3305-3316), double-strand break repair (Mandecki (1986); Arnold (1993) "Protein engineering for unusual environments" Current Opinion in Biotechnology 4:450-455. "Oligonucleotide-directed double-strand break repair in plasmids of Escherichia coli: a method for site-specific mutagenesis" Proc. Natl. Acad. Sci. USA, 83:7177-7181). Additional details on many of the above methods can be found in Methods in Enzymology Volume 154, which also describes useful controls for trouble-shooting problems with various mutagenesis methods.

[0232] Protocols that can be used to practice the invention are described, e.g., in U.S. Patent Nos. 5,605,793 to Stemmer (Feb. 25, 1997), "Methods for In Vitro Recombination," U.S. Pat. No. 5,811,238 to Stemmer et al. (Sep. 22, 1998) "Methods for Generating Polynucleotides having Desired Characteristics by Iterative Selection and Recombination," U.S. Pat. No. 5,830,721 to Stemmer et al. (Nov. 3, 1998), "DNA Mutagenesis by Random Fragmentation and Reassembly," U.S. Pat. No. 5,834,252 to Stemmer, et al. (Nov. 10, 1998) "End-Complementary Polymerase Reaction," U.S. Pat. No. 5,837,458 to Minshull, et al. (Nov. 17, 1998), "Methods and Compositions for Cellular and Metabolic Engineering," WO 95/22625, Stemmer and Cramer, "Mutagenesis by Random Fragmentation and Reassembly," WO 96/33207 by Stemmer and Lipschutz "End Complementary Polymerase Chain Reaction," WO 97/20078 by Stemmer and Cramer "Methods for Generating Polynucleotides having Desired Characteristics by Iterative Selection and Recombination," WO 97/35966 by Minshull and Stemmer, "Methods and Compositions for Cellular and Metabolic Engineering," WO 99/41402 by Punnonen et al. "Targeting of Genetic Vaccine Vectors," WO 99/41383 by Punnonen et al. "Antigen Library Immunization," WO 99/41369 by Punnonen et al. "Genetic Vaccine Vector Engineering," WO 99/41368 by Punnonen et al. "Optimization of Immunomodulatory Properties of Genetic Vaccines," EP 752008 by Stemmer and Cramer, "DNA Mutagenesis by Random Fragmentation and Reassembly," EP 0932670 by Stemmer "Evolving Cellular DNA Uptake by Recursive Sequence Recombination," WO 99/23107 by Stemmer et al., "Modification of Virus Tropism and Host Range by Viral Genome Shuffling," WO 99/21979 by Apt et al., "Human Papillomavirus Vectors," WO 98/31837 by del Cardayre et al. "Evolution of Whole Cells and Organisms by Recursive Sequence Recombination," WO 98/27230 by Patten and Stemmer, "Methods and Compositions for Polypeptide Engineering," WO 98/27230 by Stemmer et al., "Methods for Optimization of Gene Therapy by Recursive Sequence Shuffling and Selection," WO 00/00632, "Methods for Generating Highly Diverse Libraries," WO 00/09679, "Methods for Obtaining in Vitro Recombined Polynucleotide Sequence Banks and Resulting Sequences," WO 98/42832 by Arnold et al., "Recombination of Polynucleotide Sequences Using Random or Defined Primers," WO 99/29902 by Arnold et al., "Method for Creating Polynucleotide and Polypeptide Sequences," WO 98/41653 by Vind, "An in Vitro Method for Construction of a DNA Library," WO 98/41622 by Borchert et al., "Method for Constructing a Library Using DNA Shuffling," and WO 98/42727 by Pati and Zarlign, "Sequence Alterations using Homologous Recombination."

[0233] Protocols that can be used to practice the invention (providing details regarding various diversity generating methods) are described, e.g., in U.S. Patent application serial no. (USSN) 09/407,800, "SHUFFLING OF CODON ALTERED GENES" by Patten et al. filed Sep. 28, 1999; "EVOLUTION OF WHOLE CELLS AND ORGANISMS BY RECURSIVE SEQUENCE RECOMBINATION" by del Cardayre et al., United States Patent No. 6,379,964; "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION" by Cramer et al., United States Patent Nos. 6,319,714; 6,368,861; 6,376,246; 6,423,542; 6,426,224 and PCT/US00/01203; "USE OF CODON-VARIED OLIGONUCLEOTIDE SYNTHESIS FOR SYNTHETIC SHUFFLING" by Welch et al.,

United States Patent No. 6,436,675; "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" by Selifonov et al., filed Jan. 18, 2000, (PCT/US00/01202) and, e.g. "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" by Selifonov et al., filed Jul. 18, 2000 (U.S. Ser. No. 09/618,579); "METHODS OF POPULATING DATA STRUCTURES FOR USE IN EVOLUTIONARY SIMULATIONS" by Selifonov and Stemmer, filed Jan. 18, 2000 (PCT/US00101138); and "SINGLE-STRANDED NUCLEIC ACID TEMPLATE-MEDIATED RECOMBINATION AND NUCLEIC ACID FRAGMENT ISOLATION" by Affholter, filed Sep. 6, 2000 (U.S. Ser. No. 09/656,549); and United States Patent Nos. 6,177,263; 6,153,410.

[0234] Non-stochastic, or "directed evolution," methods include, e.g., saturation mutagenesis (GSSM™), synthetic ligation reassembly (SLR), or a combination thereof are used to modify the nucleic acids of the invention to generate pectate lyases with new or altered properties (e.g., activity under highly acidic or alkaline conditions, high temperatures, and the like). Polypeptides encoded by the modified nucleic acids can be screened for an activity before testing for proteolytic or other activity. Any testing modality or protocol can be used, e.g., using a capillary array platform. See, e.g., U.S. Patent Nos. 6,361,974; 6,280,926; 5,939,250.

Saturation mutagenesis, or, GSSM™

[0235] In one aspect, codon primers containing a degenerate N,N,G/T sequence are used to introduce point mutations into a polynucleotide, e.g., a pectate lyase or an antibody of the invention, so as to generate a set of progeny polypeptides in which a full range of single amino acid substitutions is represented at each amino acid position, e.g., an amino acid residue in an enzyme active site or ligand binding site targeted to be modified. These oligonucleotides can comprise a contiguous first homologous sequence, a degenerate N,N,G/T sequence, and, optionally, a second homologous sequence. The downstream progeny translational products from the use of such oligonucleotides include all possible amino acid changes at each amino acid site along the polypeptide, because the degeneracy of the N,N,G/T sequence includes codons for all 20 amino acids. In one aspect, one such degenerate oligonucleotide (comprised of, e.g., one degenerate N,N,G/T cassette) is used for subjecting each original codon in a parental polynucleotide template to a full range of codon substitutions. In another aspect, at least two degenerate cassettes are used - either in the same oligonucleotide or not, for subjecting at least two original codons in a parental polynucleotide template to a full range of codon substitutions. For example, more than one N,N,G/T sequence can be contained in one oligonucleotide to introduce amino acid mutations at more than one site. This plurality of N,N,G/T sequences can be directly contiguous, or separated by one or more additional nucleotide sequence(s). In another aspect, oligonucleotides serviceable for introducing additions and deletions can be used either alone or in combination with the codons containing an N,N,G/T sequence, to introduce any combination or permutation of amino acid additions, deletions, and/or substitutions.

[0236] In one aspect, simultaneous mutagenesis of two or more contiguous amino acid positions is done using an oligonucleotide that contains contiguous N,N,G/T triplets, i.e. a degenerate (N,N,G/T)_n sequence. In another aspect, degenerate cassettes having less degeneracy than the N,N,G/T sequence are used. For example, it may be desirable in some instances to use (e.g. in an oligonucleotide) a degenerate triplet sequence comprised of only one N, where said N can be in the first second or third position of the triplet. Any other bases including any combinations and permutations thereof can be used in the remaining two positions of the triplet. Alternatively, it may be desirable in some instances to use (e.g. in an oligo) a degenerate N,N,N triplet sequence.

[0237] In one aspect, use of degenerate triplets (e.g., N,N,G/T triplets) allows for systematic and easy generation of a full range of possible natural amino acids (for a total of 20 amino acids) into each and every amino acid position in a polypeptide (in alternative aspects, the methods also include generation of less than all possible substitutions per amino acid residue, or codon, position). For example, for a 100 amino acid polypeptide, 2000 distinct species (i.e. 20 possible amino acids per position X 100 amino acid positions) can be generated. Through the use of an oligonucleotide or set of oligonucleotides containing a degenerate N,N,G/T triplet, 32 individual sequences can code for all 20 possible natural amino acids. Thus, in a reaction vessel in which a parental polynucleotide sequence is subjected to saturation mutagenesis using at least one such oligonucleotide, there are generated 32 distinct progeny polynucleotides encoding 20 distinct polypeptides. In contrast, the use of a non-degenerate oligonucleotide in site-directed mutagenesis leads to only one progeny polypeptide product per reaction vessel. Nondegenerate oligonucleotides can optionally be used in combination with degenerate primers disclosed; for example, nondegenerate oligonucleotides can be used to generate specific point mutations in a working polynucleotide. This provides one means to generate specific silent point mutations, point mutations leading to corresponding amino acid changes, and point mutations that cause the generation of stop codons and the corresponding expression of polypeptide fragments.

[0238] In one aspect, each saturation mutagenesis reaction vessel contains polynucleotides encoding at least 20 progeny

polypeptide (e.g., pectate lyases) molecules such that all 20 natural amino acids are represented at the one specific amino acid position corresponding to the codon position mutagenized in the parental polynucleotide (other aspects use less than all 20 natural combinations). The 32-fold degenerate progeny polypeptides generated from each saturation mutagenesis reaction vessel can be subjected to clonal amplification (e.g. cloned into a suitable host, e.g., E. coli host, using, e.g., an expression vector) and subjected to expression screening. When an individual progeny polypeptide is identified by screening to display a favorable change in property (when compared to the parental polypeptide, such as increased proteolytic activity under alkaline or acidic conditions), it can be sequenced to identify the correspondingly favorable amino acid substitution contained therein.

[0239] In one aspect, upon mutagenizing each and every amino acid position in a parental polypeptide using saturation mutagenesis as disclosed herein, favorable amino acid changes may be identified at more than one amino acid position. One or more new progeny molecules can be generated that contain a combination of all or part of these favorable amino acid substitutions. For example, if 2 specific favorable amino acid changes are identified in each of 3 amino acid positions in a polypeptide, the permutations include 3 possibilities at each position (no change from the original amino acid, and each of two favorable changes) and 3 positions. Thus, there are $3 \times 3 \times 3$ or 27 total possibilities, including 7 that were previously examined - 6 single point mutations (i.e. 2 at each of three positions) and no change at any position.

[0240] In another aspect, site-saturation mutagenesis can be used together with another stochastic or non-stochastic means to vary sequence, e.g., synthetic ligation reassembly (see below), shuffling, chimerization, recombination and other mutagenizing processes and mutagenizing agents. This invention provides for the use of any mutagenizing process(es), including saturation mutagenesis, in an iterative manner.

Synthetic Ligation Reassembly (SLR)

[0241] A non-stochastic gene modification system termed "synthetic ligation reassembly," or simply "SLR," a "directed evolution process," may generate polypeptides, e.g., pectate lyases or antibodies, with new or altered properties. SLR is a method of ligating oligonucleotide fragments together non-stochastically. This method differs from stochastic oligonucleotide shuffling in that the nucleic acid building blocks are not shuffled, concatenated or chimerized randomly, but rather are assembled non-stochastically. See, e.g., U.S. Patent Application Serial No. (USSN) 09/332,835 entitled "Synthetic Ligation Reassembly in Directed Evolution" and filed on June 14, 1999 ("USSN 09/332,835"). In one aspect, SLR comprises the following steps: (a) providing a template polynucleotide, wherein the template polynucleotide comprises sequence encoding a homologous gene; (b) providing a plurality of building block polynucleotides, wherein the building block polynucleotides are designed to cross-over reassemble with the template polynucleotide at a predetermined sequence, and a building block polynucleotide comprises a sequence that is a variant of the homologous gene and a sequence homologous to the template polynucleotide flanking the variant sequence; (c) combining a building block polynucleotide with a template polynucleotide such that the building block polynucleotide cross-over reassembles with the template polynucleotide to generate polynucleotides comprising homologous gene sequence variations.

[0242] SLR does not depend on the presence of high levels of homology between polynucleotides to be rearranged. Thus, this method can be used to non-stochastically generate libraries (or sets) of progeny molecules comprised of over 10100 different chimeras. SLR can be used to generate libraries comprised of over 101000 different progeny chimeras. Thus, non-stochastic methods may produce a set of finalized chimeric nucleic acid molecule having an overall assembly order that is chosen by design. This method includes the steps of generating by design a plurality of specific nucleic acid building blocks having serviceable mutually compatible ligatable ends, and assembling these nucleic acid building blocks, such that a designed overall assembly order is achieved.

[0243] The mutually compatible ligatable ends of the nucleic acid building blocks to be assembled are considered to be "serviceable" for this type of ordered assembly if they enable the building blocks to be coupled in predetermined orders. Thus, the overall assembly order in which the nucleic acid building blocks can be coupled is specified by the design of the ligatable ends. If more than one assembly step is to be used, then the overall assembly order in which the nucleic acid building blocks can be coupled is also specified by the sequential order of the assembly step(s). In one aspect, the annealed building pieces are treated with an enzyme, such as a ligase (e.g. T4 DNA ligase), to achieve covalent bonding of the building pieces.

[0244] In one aspect, the design of the oligonucleotide building blocks is obtained by analyzing a set of progenitor nucleic acid sequence templates that serve as a basis for producing a progeny set of finalized chimeric polynucleotides. These parental oligonucleotide templates thus serve as a source of sequence information that aids in the design of the nucleic acid building blocks that are to be mutagenized, e.g., chimerized or shuffled. In one aspect of this method, the sequences of a plurality of parental nucleic acid templates are aligned in order to select one or more demarcation points. The demarcation points can be

located at an area of homology, and are comprised of one or more nucleotides. These demarcation points are preferably shared by at least two of the progenitor templates. The demarcation points can thereby be used to delineate the boundaries of oligonucleotide building blocks to be generated in order to rearrange the parental polynucleotides. The demarcation points identified and selected in the progenitor molecules serve as potential chimerization points in the assembly of the final chimeric progeny molecules. A demarcation point can be an area of homology (comprised of at least one homologous nucleotide base) shared by at least two parental polynucleotide sequences. Alternatively, a demarcation point can be an area of homology that is shared by at least half of the parental polynucleotide sequences, or, it can be an area of homology that is shared by at least two thirds of the parental polynucleotide sequences. Even more preferably a serviceable demarcation points is an area of homology that is shared by at least three fourths of the parental polynucleotide sequences, or, it can be shared by at almost all of the parental polynucleotide sequences. In one aspect, a demarcation point is an area of homology that is shared by all of the parental polynucleotide sequences.

[0245] In one aspect, a ligation reassembly process is performed exhaustively in order to generate an exhaustive library of progeny chimeric polynucleotides. In other words, all possible ordered combinations of the nucleic acid building blocks are represented in the set of finalized chimeric nucleic acid molecules. At the same time, in another aspect, the assembly order (i.e. the order of assembly of each building block in the 5' to 3' sequence of each finalized chimeric nucleic acid) in each combination is by design (or non-stochastic) as described above. Because of the non-stochastic nature of this, the possibility of unwanted side products is greatly reduced.

[0246] The ligation reassembly method may be performed systematically. For example, the method is performed in order to generate a systematically compartmentalized library of progeny molecules, with compartments that can be screened systematically, e.g. one by one. This provides that, through the selective and judicious use of specific nucleic acid building blocks, coupled with the selective and judicious use of sequentially stepped assembly reactions, a design can be achieved where specific sets of progeny products are made in each of several reaction vessels. This allows a systematic examination and screening procedure to be performed. Thus, these methods allow a potentially very large number of progeny molecules to be examined systematically in smaller groups. Because of its ability to perform chimerizations in a manner that is highly flexible yet exhaustive and systematic as well, particularly when there is a low level of homology among the progenitor molecules, these methods provide for the generation of a library (or set) comprised of a large number of progeny molecules. Because of the non-stochastic nature of the instant ligation reassembly, the progeny molecules generated preferably comprise a library of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design. The saturation mutagenesis and optimized directed evolution methods also can be used to generate different progeny molecular species. It is appreciated that this provides freedom of choice and control regarding the selection of demarcation points, the size and number of the nucleic acid building blocks, and the size and design of the couplings. It is appreciated, furthermore, that the requirement for intermolecular homology is highly relaxed for the operability of this process. In fact, demarcation points can even be chosen in areas of little or no intermolecular homology. For example, because of codon wobble, i.e. the degeneracy of codons, nucleotide substitutions can be introduced into nucleic acid building blocks without altering the amino acid originally encoded in the corresponding progenitor template. Alternatively, a codon can be altered such that the coding for an originally amino acid is altered. This provides that such substitutions can be introduced into the nucleic acid building block in order to increase the incidence of intermolecular homologous demarcation points and thus to allow an increased number of couplings to be achieved among the building blocks, which in turn allows a greater number of progeny chimeric molecules to be generated.

[0247] In another aspect, the synthetic nature of the step in which the building blocks are generated allows the design and introduction of nucleotides (e.g., one or more nucleotides, which may be, for example, codons or introns or regulatory sequences) that can later be optionally removed in an in vitro process (e.g. by mutagenesis) or in an in vivo process (e.g. by utilizing the gene splicing ability of a host organism). It is appreciated that in many instances the introduction of these nucleotides may also be desirable for many other reasons in addition to the potential benefit of creating a serviceable demarcation point.

[0248] In one aspect, a nucleic acid building block is used to introduce an intron. Thus, functional introns are introduced into a man-made gene manufactured according to the methods described herein. The artificially introduced intron(s) can be functional in a host cells for gene splicing much in the way that naturally-occurring introns serve functionally in gene splicing.

Optimized Directed Evolution System

[0249] A non-stochastic gene modification system termed "optimized directed evolution system" may generate polypeptides, e.g., pectate lyases or antibodies, with new or altered properties. Optimized directed evolution is directed to the use of repeated cycles of reductive reassortment, recombination and selection that allow for the directed molecular evolution of nucleic acids through

recombination. Optimized directed evolution allows generation of a large population of evolved chimeric sequences, wherein the generated population is significantly enriched for sequences that have a predetermined number of crossover events.

[0250] A crossover event is a point in a chimeric sequence where a shift in sequence occurs from one parental variant to another parental variant. Such a point is normally at the juncture of where oligonucleotides from two parents are ligated together to form a single sequence. This method allows calculation of the correct concentrations of oligonucleotide sequences so that the final chimeric population of sequences is enriched for the chosen number of crossover events. This provides more control over choosing chimeric variants having a predetermined number of crossover events.

[0251] In addition, this method provides a convenient means for exploring a tremendous amount of the possible protein variant space in comparison to other systems. Previously, if one generated, for example, 10^{13} chimeric molecules during a reaction, it would be extremely difficult to test such a high number of chimeric variants for a particular activity. Moreover, a significant portion of the progeny population would have a very high number of crossover events which resulted in proteins that were less likely to have increased levels of a particular activity. By using these methods, the population of chimeric molecules can be enriched for those variants that have a particular number of crossover events. Thus, although one can still generate 10^{13} chimeric molecules during a reaction, each of the molecules chosen for further analysis most likely has, for example, only three crossover events. Because the resulting progeny population can be skewed to have a predetermined number of crossover events, the boundaries on the functional variety between the chimeric molecules is reduced. This provides a more manageable number of variables when calculating which oligonucleotide from the original parental polynucleotides might be responsible for affecting a particular trait.

[0252] One method for creating a chimeric progeny polynucleotide sequence is to create oligonucleotides corresponding to fragments or portions of each parental sequence. Each oligonucleotide preferably includes a unique region of overlap so that mixing the oligonucleotides together results in a new variant that has each oligonucleotide fragment assembled in the correct order. Additional information can also be found, e.g., in USSN 09/332,835; U.S. Patent No. 6,361,974.

[0253] The number of oligonucleotides generated for each parental variant bears a relationship to the total number of resulting crossovers in the chimeric molecule that is ultimately created. For example, three parental nucleotide sequence variants might be provided to undergo a ligation reaction in order to find a chimeric variant having, for example, greater activity at high temperature. As one example, a set of 50 oligonucleotide sequences can be generated corresponding to each portions of each parental variant. Accordingly, during the ligation reassembly process there could be up to 50 crossover events within each of the chimeric sequences. The probability that each of the generated chimeric polynucleotides will contain oligonucleotides from each parental variant in alternating order is very low. If each oligonucleotide fragment is present in the ligation reaction in the same molar quantity it is likely that in some positions oligonucleotides from the same parental polynucleotide will ligate next to one another and thus not result in a crossover event. If the concentration of each oligonucleotide from each parent is kept constant during any ligation step in this example, there is a 1/3 chance (assuming 3 parents) that an oligonucleotide from the same parental variant will ligate within the chimeric sequence and produce no crossover.

[0254] Accordingly, a probability density function (PDF) can be determined to predict the population of crossover events that are likely to occur during each step in a ligation reaction given a set number of parental variants, a number of oligonucleotides corresponding to each variant, and the concentrations of each variant during each step in the ligation reaction. The statistics and mathematics behind determining the PDF is described below. By utilizing these methods, one can calculate such a probability density function, and thus enrich the chimeric progeny population for a predetermined number of crossover events resulting from a particular ligation reaction. Moreover, a target number of crossover events can be predetermined, and the system then programmed to calculate the starting quantities of each parental oligonucleotide during each step in the ligation reaction to result in a probability density function that centers on the predetermined number of crossover events. These methods are directed to the use of repeated cycles of reductive reassortment, recombination and selection that allow for the directed molecular evolution of a nucleic acid encoding a polypeptide through recombination. This system allows generation of a large population of evolved chimeric sequences, wherein the generated population is significantly enriched for sequences that have a predetermined number of crossover events. A crossover event is a point in a chimeric sequence where a shift in sequence occurs from one parental variant to another parental variant. Such a point is normally at the juncture of where oligonucleotides from two parents are ligated together to form a single sequence. The method allows calculation of the correct concentrations of oligonucleotide sequences so that the final chimeric population of sequences is enriched for the chosen number of crossover events. This provides more control over choosing chimeric variants having a predetermined number of crossover events.

[0255] In addition, these methods provide a convenient means for exploring a tremendous amount of the possible protein variant space in comparison to other systems. By using the methods described herein, the population of chimeric molecules can be enriched for those variants that have a particular number of crossover events. Thus, although one can still generate 10^{13}

chimeric molecules during a reaction, each of the molecules chosen for further analysis most likely has, for example, only three crossover events. Because the resulting progeny population can be skewed to have a predetermined number of crossover events, the boundaries on the functional variety between the chimeric molecules is reduced. This provides a more manageable number of variables when calculating which oligonucleotide from the original parental polynucleotides might be responsible for affecting a particular trait.

[0256] In one aspect, the method creates a chimeric progeny polynucleotide sequence by creating oligonucleotides corresponding to fragments or portions of each parental sequence. Each oligonucleotide preferably includes a unique region of overlap so that mixing the oligonucleotides together results in a new variant that has each oligonucleotide fragment assembled in the correct order. See also USSN 09/332,835.

Determining Crossover Events

[0257] A system and software may receive a desired crossover probability density function (PDF), the number of parent genes to be reassembled, and the number of fragments in the reassembly as inputs. The output of this program is a "fragment PDF" that can be used to determine a recipe for producing reassembled genes, and the estimated crossover PDF of those genes. The processing described herein is preferably performed in MATLAB™ (The Mathworks, Natick, Massachusetts) a programming language and development environment for technical computing.

Iterative Processes

[0258] These processes can be iteratively repeated. For example, a nucleic acid (or, the nucleic acid) responsible for an altered or new pectate lyase phenotype is identified, re-isolated, again modified, re-tested for activity. This process can be iteratively repeated until a desired phenotype is engineered. For example, an entire biochemical anabolic or catabolic pathway can be engineered into a cell, including, e.g., epoxide hydrolysis activity.

[0259] Similarly, if it is determined that a particular oligonucleotide has no affect at all on the desired trait (e.g., a new pectate lyase phenotype), it can be removed as a variable by synthesizing larger parental oligonucleotides that include the sequence to be removed. Since incorporating the sequence within a larger sequence prevents any crossover events, there will no longer be any variation of this sequence in the progeny polynucleotides. This iterative practice of determining which oligonucleotides are most related to the desired trait, and which are unrelated, allows more efficient exploration all of the possible protein variants that might be provide a particular trait or activity.

In vivo shuffling

[0260] *In vivo* shuffling of molecules is used in methods that provide variants of polypeptides of the invention. *In vivo* shuffling can be performed utilizing the natural property of cells to recombine multimers. While recombination *in vivo* has provided the major natural route to molecular diversity, genetic recombination remains a relatively complex process that involves 1) the recognition of homologies; 2) strand cleavage, strand invasion, and metabolic steps leading to the production of recombinant chiasma; and finally 3) the resolution of chiasma into discrete recombined molecules. The formation of the chiasma requires the recognition of homologous sequences.

[0261] A method may produce a hybrid polynucleotide from at least a first polynucleotide (e.g., a pectate lyase of the invention) and a second polynucleotide (e.g., an enzyme, such as a pectate lyase of the invention or any other pectate lyase, or, a tag or an epitope). There may be produced a hybrid polynucleotide by introducing at least a first polynucleotide and a second polynucleotide which share at least one region of partial sequence homology into a suitable host cell. The regions of partial sequence homology promote processes which result in sequence reorganization producing a hybrid polynucleotide. The term "hybrid polynucleotide", as used herein, is any nucleotide sequence which results from the method described and contains sequence from at least two original polynucleotide sequences. Such hybrid polynucleotides can result from intermolecular recombination events which promote sequence integration between DNA molecules. In addition, such hybrid polynucleotides can result from intramolecular reductive reassortment processes which utilize repeated sequences to alter a nucleotide sequence within a DNA molecule.

Producing sequence variants

[0262] There are additional methods for making sequence variants of the nucleic acid (e.g., pectate lyase) sequences of the invention and additional methods for isolating pectate lyases using the nucleic acids and polypeptides of the invention. Variants of a pectate lyase coding sequence (e.g., a gene, cDNA or message) of the invention can be altered by any means, including, e.g., random or stochastic methods, or, non-stochastic, or "directed evolution," methods, as described above.

[0263] The isolated variants may be naturally occurring. Variant can also be created *in vitro*. Variants may be created using genetic engineering techniques such as site directed mutagenesis, random chemical mutagenesis, Exonuclease III deletion procedures, and standard cloning techniques. Alternatively, such variants, fragments, analogs, or derivatives may be created using chemical synthesis or modification procedures. Other methods of making variants are also familiar to those skilled in the art. These include procedures in which nucleic acid sequences obtained from natural isolates are modified to generate nucleic acids which encode polypeptides having characteristics which enhance their value in industrial or laboratory applications. In such procedures, a large number of variant sequences having one or more nucleotide differences with respect to the sequence obtained from the natural isolate are generated and characterized. These nucleotide differences can result in amino acid changes with respect to the polypeptides encoded by the nucleic acids from the natural isolates.

[0264] For example, variants may be created using error prone PCR. In error prone PCR, PCR is performed under conditions where the copying fidelity of the DNA polymerase is low, such that a high rate of point mutations is obtained along the entire length of the PCR product. Error prone PCR is described, e.g., in Leung, D.W., et al., Technique, 1:11-15, 1989) and Caldwell, R. C. & Joyce G.F., PCR Methods Applic., 2:28-33, 1992. Briefly, in such procedures, nucleic acids to be mutagenized are mixed with PCR primers, reaction buffer, MgCl₂, MnCl₂, Taq polymerase and an appropriate concentration of dNTPs for achieving a high rate of point mutation along the entire length of the PCR product. For example, the reaction may be performed using 20 fmoles of nucleic acid to be mutagenized, 30 pmole of each PCR primer, a reaction buffer comprising 50mM KCl, 10mM Tris HCl (pH 8.3) and 0.01% gelatin, 7mM MgCl₂, 0.5mM MnCl₂, 5 units of Taq polymerase, 0.2mM dGTP, 0.2mM dATP, 1mM dCTP, and 1mM dTTP. PCR may be performed for 30 cycles of 94°C for 1 min, 45°C for 1 min, and 72°C for 1 min. However, it will be appreciated that these parameters may be varied as appropriate. The mutagenized nucleic acids are cloned into an appropriate vector and the activities of the polypeptides encoded by the mutagenized nucleic acids is evaluated.

[0265] Variants may also be created using oligonucleotide directed mutagenesis to generate site-specific mutations in any cloned DNA of interest. Oligonucleotide mutagenesis is described, e.g., in Reidhaar-Olson (1988) Science 241:53-57. Briefly, in such procedures a plurality of double stranded oligonucleotides bearing one or more mutations to be introduced into the cloned DNA are synthesized and inserted into the cloned DNA to be mutagenized. Clones containing the mutagenized DNA are recovered and the activities of the polypeptides they encode are assessed.

[0266] Another method for generating variants is assembly PCR. Assembly PCR involves the assembly of a PCR product from a mixture of small DNA fragments. A large number of different PCR reactions occur in parallel in the same vial, with the products of one reaction priming the products of another reaction. Assembly PCR is described in, e.g., U.S. Patent No. 5,965,408.

[0267] Still another method of generating variants is sexual PCR mutagenesis. In sexual PCR mutagenesis, forced homologous recombination occurs between DNA molecules of different but highly related DNA sequence *in vitro*, as a result of random fragmentation of the DNA molecule based on sequence homology, followed by fixation of the crossover by primer extension in a PCR reaction. Sexual PCR mutagenesis is described, e.g., in Stemmer (1994) Proc. Natl. Acad. Sci. USA 91:10747-10751. Briefly, in such procedures a plurality of nucleic acids to be recombined are digested with DNase to generate fragments having an average size of 50-200 nucleotides. Fragments of the desired average size are purified and resuspended in a PCR mixture. PCR is conducted under conditions which facilitate recombination between the nucleic acid fragments. For example, PCR may be performed by resuspending the purified fragments at a concentration of 10-30ng/μl in a solution of 0.2mM of each dNTP, 2.2mM MgCl₂, 50mM KCL, 10mM Tris HCl, pH 9.0, and 0.1% Triton X-100. 2.5 units of Taq polymerase per 100:1 of reaction mixture is added and PCR is performed using the following regime: 94°C for 60 seconds, 94°C for 30 seconds, 50-55°C for 30 seconds, 72°C for 30 seconds (30-45 times) and 72°C for 5 minutes. However, it will be appreciated that these parameters may be varied as appropriate. In some aspects, oligonucleotides may be included in the PCR reactions. In other aspects, the Klenow fragment of DNA polymerase I may be used in a first set of PCR reactions and Taq polymerase may be used in a subsequent set of PCR reactions. Recombinant sequences are isolated and the activities of the polypeptides they encode are assessed.

[0268] Variants may also be created by *in vivo* mutagenesis. In some aspects, random mutations in a sequence of interest are generated by propagating the sequence of interest in a bacterial strain, such as an *E. coli* strain, which carries mutations in one or more of the DNA repair pathways. Such "mutator" strains have a higher random mutation rate than that of a wild-type parent.

Propagating the DNA in one of these strains will eventually generate random mutations within the DNA. Mutator strains suitable for use for in vivo mutagenesis are described, e.g., in PCT Publication No. WO 91/16427.

[0269] Variants may also be generated using cassette mutagenesis. In cassette mutagenesis a small region of a double stranded DNA molecule is replaced with a synthetic oligonucleotide "cassette" that differs from the native sequence. The oligonucleotide often contains completely and/or partially randomized native sequence.

[0270] Recursive ensemble mutagenesis may also be used to generate variants. Recursive ensemble mutagenesis is an algorithm for protein engineering (protein mutagenesis) developed to produce diverse populations of phenotypically related mutants whose members differ in amino acid sequence. This method uses a feedback mechanism to control successive rounds of combinatorial cassette mutagenesis. Recursive ensemble mutagenesis is described, e.g., in Arkin (1992) Proc. Natl. Acad. Sci. USA 89:7811-7815.

[0271] In some aspects, variants are created using exponential ensemble mutagenesis. Exponential ensemble mutagenesis is a process for generating combinatorial libraries with a high percentage of unique and functional mutants, wherein small groups of residues are randomized in parallel to identify, at each altered position, amino acids which lead to functional proteins. Exponential ensemble mutagenesis is described, e.g., in Delegrave (1993) Biotechnology Res. 11:1548-1552. Random and site-directed mutagenesis are described, e.g., in Arnold (1993) Current Opinion in Biotechnology 4:450-455.

[0272] In some aspects, the variants are created using shuffling procedures wherein portions of a plurality of nucleic acids which encode distinct polypeptides are fused together to create chimeric nucleic acid sequences which encode chimeric polypeptides as described in, e.g., U.S. Patent Nos. 5,965,408; 5,939,250 (see also discussion, above).

[0273] The invention also provides variants of polypeptides of the invention (e.g., pectate lyases) comprising sequences in which one or more of the amino acid residues (e.g., of an exemplary polypeptide of the invention) are substituted with a conserved or non-conserved amino acid residue (e.g., a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code. Conservative substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Thus, polypeptides of the invention include those with conservative substitutions of sequences of the invention, e.g., the exemplary polypeptides of the invention, including but not limited to the following replacements: replacements of an aliphatic amino acid such as Alanine, Valine, Leucine and Isoleucine with another aliphatic amino acid; replacement of a Serine with a Threonine or vice versa; replacement of an acidic residue such as Aspartic acid and Glutamic acid with another acidic residue; replacement of a residue bearing an amide group, such as Asparagine and Glutamine, with another residue bearing an amide group; exchange of a basic residue such as Lysine and Arginine with another basic residue; and replacement of an aromatic residue such as Phenylalanine, Tyrosine with another aromatic residue. Other variants are those in which one or more of the amino acid residues of the polypeptides of the invention includes a substituent group.

[0274] Other variants are those in which the polypeptide is associated with another compound, such as a compound to increase the half-life of the polypeptide, for example, polyethylene glycol.

[0275] Additional variants those in which additional amino acids are fused to the polypeptide, such as a leader sequence, a secretory sequence, a proprotein sequence or a sequence which facilitates purification, enrichment, or stabilization of the polypeptide.

[0276] In some aspects, the variants, fragments, derivatives and analogs of the polypeptides of the invention retain the same biological function or activity as the exemplary polypeptides, e.g., pectate lyase activity, as described herein. In other aspects, the variant, fragment, derivative, or analog includes a proprotein, such that the variant, fragment, derivative, or analog can be activated by cleavage of the proprotein portion to produce an active polypeptide.

Optimizing codons to achieve high levels of protein expression in host cells

[0277] Methods may modify pectate lyase-encoding nucleic acids to modify codon usage, for example modifying codons in a nucleic acid encoding a pectate lyase to increase or decrease its expression in a host cell. The method comprises identifying a "non-preferred" or a "less preferred" codon in pectate lyase-encoding nucleic acid and replacing one or more of these non-preferred or less preferred codons with a "preferred codon" encoding the same amino acid as the replaced codon and at least one non-preferred or less preferred codon in the nucleic acid has been replaced by a preferred codon encoding the same amino acid. A preferred codon is a codon over-represented in coding sequences in genes in the host cell and a non-preferred or less

preferred codon is a codon under-represented in coding sequences in genes in the host cell.

[0278] Host cells for expressing the nucleic acids, expression cassettes and vectors of the invention include bacteria, yeast, fungi, plant cells, insect cells and mammalian cells. Methods for optimizing codon usage in all of these cells are disclosed. Exemplary host cells include gram negative bacteria, such as *Escherichia coli*; gram positive bacteria, such as any *Streptomyces*, *Lactobacillus gasseri*, *Lactococcus lactis*, *Lactococcus cremoris*, any *Bacillus*, e.g., *Bacillus subtilis*, *Bacillus cereus*. Exemplary host cells also include eukaryotic organisms, e.g., various yeast, such as *Saccharomyces* sp., including *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Pichia pastoris*, and *Kluyveromyces lactis*, *Hansenula polymorpha*, *Aspergillus niger*, and mammalian cells and cell lines and insect cells and cell lines. Thus, the disclosure also includes nucleic acids and polypeptides optimized for expression in these organisms and species.

[0279] For example, the codons of a nucleic acid encoding a pectate lyase isolated from a bacterial cell are modified such that the nucleic acid is optimally expressed in a bacterial cell different from the bacteria from which the pectate lyase was derived, a yeast, a fungi, a plant cell, an insect cell or a mammalian cell. Methods for optimizing codons are well known in the art, see, e.g., U.S. Patent No. 5,795,737; Baca (2000) Int. J. Parasitol. 30:113-118; Hale (1998) Protein Expr. Purif. 12:185-188; Narum (2001) Infect. Immun. 69:7250-7253. See also Narum (2001) Infect. Immun. 69:7250-7253, describing optimizing codons in mouse systems; Outchkourov (2002) Protein Expr. Purif. 24:18-24, describing optimizing codons in yeast; Feng (2000) Biochemistry 39:15399-15409, describing optimizing codons in *E. coli*; Humphreys (2000) Protein Expr. Purif. 20:252-264, describing optimizing codon usage that affects secretion in *E. coli*.

Synthetic gene reassembly

[0280] A non-stochastic method termed synthetic gene reassembly (e.g., GeneReassembly™, see, e.g., U.S. Patent No. 6,537,776) may be for modifying pectate lyases of the invention or building new pectate lyases within the scope of the invention. GeneReassembly™ differs from stochastic shuffling in that the nucleic acid building blocks are not shuffled or concatenated or chimerized randomly, but rather are assembled non-stochastically.

[0281] The synthetic gene reassembly method does not depend on the presence of a high level of homology between polynucleotides to be shuffled. The invention can be used to non-stochastically generate libraries (or sets) of progeny molecules comprised of over 10^{100} different chimeras. Conceivably, synthetic gene reassembly can even be used to generate libraries comprised of over 10^{1000} different progeny chimeras.

[0282] Such a non-stochastic method may produce a set of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design. It is comprised of the steps of generating by design a plurality of specific nucleic acid building blocks having serviceable mutually compatible ligatable ends and assembling these nucleic acid building blocks, such that a designed overall assembly order is achieved.

[0283] In one aspect, synthetic gene reassembly comprises a method of: 1) preparing a progeny generation of molecule(s) (including a molecule comprising a polynucleotide sequence, e.g., a molecule comprising a polypeptide coding sequence), that is mutagenized to achieve at least one point mutation, addition, deletion, and/or chimerization, from one or more ancestral or parental generation template(s); 2) screening the progeny generation molecule(s), e.g., using a high throughput method, for at least one property of interest (such as an improvement in an enzyme activity); 3) optionally obtaining and/or cataloguing structural and/or functional information regarding the parental and/or progeny generation molecules; and 4) optionally repeating any of steps 1) to 3). In one aspect, there is generated (e.g., from a parent polynucleotide template), in what is termed "codon site-saturation mutagenesis," a progeny generation of polynucleotides, each having at least one set of up to three contiguous point mutations (i.e. different bases comprising a new codon), such that every codon (or every family of degenerate codons encoding the same amino acid) is represented at each codon position. Corresponding to, and encoded by, this progeny generation of polynucleotides, there is also generated a set of progeny polypeptides, each having at least one single amino acid point mutation. In a one aspect, there is generated, in what is termed "amino acid site-saturation mutagenesis", one such mutant polypeptide for each of the 19 naturally encoded polypeptide-forming alpha-amino acid substitutions at each and every amino acid position along the polypeptide. This yields, for each and every amino acid position along the parental polypeptide, a total of 20 distinct progeny polypeptides including the original amino acid, or potentially more than 21 distinct progeny polypeptides if additional amino acids are used either instead of or in addition to the 20 naturally encoded amino acids

[0284] This approach is also serviceable for generating mutants containing, in addition to and/or in combination with the 20 naturally encoded polypeptide-forming alpha-amino acids, other rare and/or not naturally-encoded amino acids and amino acid

derivatives. This approach is also serviceable for generating mutants by the use of, in addition to and/or in combination with natural or unaltered codon recognition systems of suitable hosts, altered, mutagenized, and/or designer codon recognition systems (such as in a host cell with one or more altered tRNA molecules).

[0285] The disclosure relates to recombination and more specifically to a method for preparing polynucleotides encoding a polypeptide by a method of *in vivo* reassortment of polynucleotide sequences containing regions of partial homology, assembling the polynucleotides to form at least one polynucleotide and screening the polynucleotides for the production of polypeptide(s) having a useful property.

[0286] The disclosure is serviceable for analyzing and cataloguing, with respect to any molecular property (e.g. an enzymatic activity) or combination of properties allowed by current technology, the effects of any mutational change achieved (including particularly saturation mutagenesis). Thus, a comprehensive method is provided for determining the effect of changing each amino acid in a parental polypeptide into each of at least 19 possible substitutions. This allows each amino acid in a parental polypeptide to be characterized and catalogued according to its spectrum of potential effects on a measurable property of the polypeptide.

[0287] An intron may be introduced into a chimeric progeny molecule by way of a nucleic acid building block. Introns often have consensus sequences at both termini in order to render them operational. In addition to enabling gene splicing, introns may serve an additional purpose by providing sites of homology to other nucleic acids to enable homologous recombination. For this purpose, and potentially others, it may be sometimes desirable to generate a large nucleic acid building block for introducing an intron. If the size is overly large easily generating by direct chemical synthesis of two single stranded oligos, such a specialized nucleic acid building block may also be generated by direct chemical synthesis of more than two single stranded oligos or by using a polymerase-based amplification reaction

[0288] The mutually compatible ligatable ends of the nucleic acid building blocks to be assembled are considered to be "serviceable" for this type of ordered assembly if they enable the building blocks to be coupled in predetermined orders. Thus, the overall assembly order in which the nucleic acid building blocks can be coupled is specified by the design of the ligatable ends and, if more than one assembly step is to be used, then the overall assembly order in which the nucleic acid building blocks can be coupled is also specified by the sequential order of the assembly step(s). The annealed building pieces may be treated with an enzyme, such as a ligase (e.g., T4 DNA ligase) to achieve covalent bonding of the building pieces.

[0289] Coupling can occur in a manner that does not make use of every nucleotide in a participating overhang. The coupling is particularly lively to survive (e.g. in a transformed host) if the coupling reinforced by treatment with a ligase enzyme to form what may be referred to as a "gap ligation" or a "gapped ligation". This type of coupling can contribute to generation of unwanted background product(s), but it can also be used advantageously increase the diversity of the progeny library generated by the designed ligation reassembly. Certain overhangs are able to undergo self-coupling to form a palindromic coupling. A coupling is strengthened substantially if it is reinforced by treatment with a ligase enzyme. Lack of 5' phosphates on these overhangs can be used advantageously to prevent this type of palindromic self-ligation. Accordingly, nucleic acid building blocks can be chemically made (or ordered) that lack a 5' phosphate group. Alternatively, they can be removed, e.g. by treatment with a phosphatase enzyme, such as a calf intestinal alkaline phosphatase (CIAP), in order to prevent palindromic self-ligations in ligation reassembly processes.

Transgenic non-human animals

[0290] The invention provides transgenic non-human animals comprising a nucleic acid, a polypeptide (e.g., a pectate lyase), an expression cassette or vector or a transfected or transformed cell of the invention. The invention also provides methods of making and using these transgenic non-human animals.

[0291] The transgenic non-human animals can be, e.g., goats, rabbits, sheep, pigs, cows, rats and mice, comprising the nucleic acids of the invention. These animals can be used, e.g., as *in vivo* models to study pectate lyase activity, or, as models to screen for agents that change the pectate lyase activity *in vivo*. The coding sequences for the polypeptides to be expressed in the transgenic non-human animals can be designed to be constitutive, or, under the control of tissue-specific, developmental-specific or inducible transcriptional regulatory factors. Transgenic non-human animals can be designed and generated using any method known in the art; see, e.g., U.S. Patent Nos. 6,211,428; 6,187,992; 6,156,952; 6,118,044; 6,111,166; 6,107,541; 5,959,171; 5,922,854; 5,892,070; 5,880,327; 5,891,698; 5,639,940; 5,573,933; 5,387,742; 5,087,571, describing making and using transformed cells and eggs and transgenic mice, rats, rabbits, sheep, pigs and cows. See also, e.g., Pollock (1999) J. Immunol.

Methods 231:147-157, describing the production of recombinant proteins in the milk of transgenic dairy animals; Baguisi (1999) Nat. Biotechnol. 17:456-461, demonstrating the production of transgenic goats. U.S. Patent No. 6,211,428, describes making and using transgenic non-human mammals which express in their brains a nucleic acid construct comprising a DNA sequence. U.S. Patent No. 5,387,742, describes injecting cloned recombinant or synthetic DNA sequences into fertilized mouse eggs, implanting the injected eggs in pseudo-pregnant females, and growing to term transgenic mice whose cells express proteins related to the pathology of Alzheimer's disease. U.S. Patent No. 6,187,992, describes making and using a transgenic mouse whose genome comprises a disruption of the gene encoding amyloid precursor protein (APP).

[0292] "Knockout animals" can also be used to practice the methods of the invention. For example, in one aspect, the transgenic or modified animals of the invention comprise a "knockout animal," e.g., a "knockout mouse," engineered not to express an endogenous gene, which is replaced with a gene expressing a pectate lyase of the invention, or, a fusion protein comprising a pectate lyase of the invention.

Transgenic Plants and Seeds

[0293] The invention provides transgenic plants and seeds comprising a nucleic acid, a polypeptide (e.g., a pectate lyase), an expression cassette or vector or a transfected or transformed cell of the invention. The invention also provides plant products, e.g., oils, seeds, leaves, extracts and the like, comprising a nucleic acid and/or a polypeptide (e.g., a pectate lyase) of the invention. The transgenic plant can be dicotyledonous (a dicot) or monocotyledonous (a monocot). The invention also provides methods of making and using these transgenic plants and seeds. The transgenic plant or plant cell expressing a polypeptide of the present invention may be constructed in accordance with any method known in the art. See, for example, U.S. Patent No. 6,309,872.

[0294] Nucleic acids and expression constructs of the invention can be introduced into a plant cell by any means. For example, nucleic acids or expression constructs can be introduced into the genome of a desired plant host, or, the nucleic acids or expression constructs can be episomes. Introduction into the genome of a desired plant can be such that the host's pectate lyase production is regulated by endogenous transcriptional or translational control elements. The invention also provides "knockout plants" where insertion of gene sequence by, e.g., homologous recombination, has disrupted the expression of the endogenous gene. Means to generate "knockout" plants are well-known in the art, see, e.g., Strepp (1998) Proc Natl. Acad. Sci. USA 95:4368-4373; Miao (1995) Plant J 7:359-365. See discussion on transgenic plants, below.

[0295] The nucleic acids of the invention can be used to confer desired traits on essentially any plant, e.g., on starch-producing plants, such as potato, wheat, rice, barley, and the like. Nucleic acids of the invention can be used to manipulate metabolic pathways of a plant in order to optimize or alter host's expression, of pectate lyase. The can change pectate lyase activity in a plant. Alternatively, a pectate lyase of the invention can be used in production of a transgenic plant to produce a compound not naturally produced by that plant. This can lower production costs or create a novel product.

[0296] In one aspect, the first step in production of a transgenic plant involves making an expression construct for expression in a plant cell. These techniques are well known in the art. They can include selecting and cloning a promoter, a coding sequence for facilitating efficient binding of ribosomes to mRNA and selecting the appropriate gene terminator sequences. One exemplary constitutive promoter is CaMV35S, from the cauliflower mosaic virus, which generally results in a high degree of expression in plants. Other promoters are more specific and respond to cues in the plant's internal or external environment. An exemplary light-inducible promoter is the promoter from the cab gene, encoding the major chlorophyll a/b binding protein.

[0297] In one aspect, the nucleic acid is modified to achieve greater expression in a plant cell. For example, a sequence of the invention is likely to have a higher percentage of A-T nucleotide pairs compared to that seen in a plant, some of which prefer G-C nucleotide pairs. Therefore, A-T nucleotides in the coding sequence can be substituted with G-C nucleotides without significantly changing the amino acid sequence to enhance production of the gene product in plant cells.

[0298] Selectable marker gene can be added to the gene construct in order to identify plant cells or tissues that have successfully integrated the transgene. This may be necessary because achieving incorporation and expression of genes in plant cells is a rare event, occurring in just a few percent of the targeted tissues or cells. Selectable marker genes encode proteins that provide resistance to agents that are normally toxic to plants, such as antibiotics or herbicides. Only plant cells that have integrated the selectable marker gene will survive when grown on a medium containing the appropriate antibiotic or herbicide. As for other inserted genes, marker genes also require promoter and termination sequences for proper function.

[0299] In one aspect, making transgenic plants or seeds comprises incorporating sequences of the invention and, optionally, marker genes into a target expression construct (e.g., a plasmid), along with positioning of the promoter and the terminator sequences. This can involve transferring the modified gene into the plant through a suitable method. For example, a construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation and microinjection of plant cell protoplasts, or the constructs can be introduced directly to plant tissue using ballistic methods, such as DNA particle bombardment. For example, see, e.g., Christou (1997) *Plant Mol. Biol.* 35:197-203; Pawłowski (1996) *Mol. Biotechnol.* 6:17-30; Klein (1987) *Nature* 327:70-73; Takumi (1997) *Genes Genet. Syst.* 72:63-69, discussing use of particle bombardment to introduce transgenes into wheat; and Adam (1997) *supra*, for use of particle bombardment to introduce YACs into plant cells. For example, Rinehart (1997) *supra*, used particle bombardment to generate transgenic cotton plants. Apparatus for accelerating particles is described U.S. Pat. No. 5,015,580; and, the commercially available BioRad (Biolistics) PDS-2000 particle acceleration instrument; see also, John, U.S. Patent No. 5,608,148; and Ellis, U.S. Patent No. 5,681,730, describing particle-mediated transformation of gymnosperms.

[0300] In one aspect, protoplasts can be immobilized and injected with a nucleic acids, e.g., an expression construct. Although plant regeneration from protoplasts is not easy with cereals, plant regeneration is possible in legumes using somatic embryogenesis from protoplast derived callus. Organized tissues can be transformed with naked DNA using gene gun technique, where DNA is coated on tungsten microprojectiles, shot 1/100th the size of cells, which carry the DNA deep into cells and organelles. Transformed tissue is then induced to regenerate, usually by somatic embryogenesis. This technique has been successful in several cereal species including maize and rice.

[0301] Nucleic acids, e.g., expression constructs, can also be introduced in to plant cells using recombinant viruses. Plant cells can be transformed using viral vectors, such as, e.g., tobacco mosaic virus derived vectors (Rouwendal (1997) *Plant Mol. Biol.* 33:989-999), see Porta (1996) "Use of viral replicons for the expression of genes in plants," *Mol. Biotechnol.* 5:209-221.

[0302] Alternatively, nucleic acids, e.g., an expression construct, can be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent marker into the plant cell DNA when the cell is infected by the bacteria. *Agrobacterium tumefaciens*-mediated transformation techniques, including disarming and use of binary vectors, are well described in the scientific literature. See, e.g., Horsch (1984) *Science* 233:496-498; Fraley (1983) *Proc. Natl. Acad. Sci. USA* 80:4803 (1983); *Gene Transfer to Plants*, Potrykus, ed. (Springer-Verlag, Berlin 1995). The DNA in an *A. tumefaciens* cell is contained in the bacterial chromosome as well as in another structure known as a Ti (tumor-inducing) plasmid. The Ti plasmid contains a stretch of DNA termed T-DNA (~20 kb long) that is transferred to the plant cell in the infection process and a series of vir (virulence) genes that direct the infection process. *A. tumefaciens* can only infect a plant through wounds: when a plant root or stem is wounded it gives off certain chemical signals, in response to which, the vir genes of *A. tumefaciens* become activated and direct a series of events necessary for the transfer of the T-DNA from the Ti plasmid to the plant's chromosome. The T-DNA then enters the plant cell through the wound. One speculation is that the T-DNA waits until the plant DNA is being replicated or transcribed, then inserts itself into the exposed plant DNA. In order to use *A. tumefaciens* as a transgene vector, the tumor-inducing section of T-DNA have to be removed, while retaining the T-DNA border regions and the vir genes. The transgene is then inserted between the T-DNA border regions, where it is transferred to the plant cell and becomes integrated into the plant's chromosomes.

[0303] The invention provides for the transformation of monocotyledonous plants using the nucleic acids of the invention, including important cereals, see Hiei (1997) *Plant Mol. Biol.* 35:205-218. See also, e.g., Horsch, *Science* (1984) 233:496; Fraley (1983) *Proc. Natl. Acad. Sci. USA* 80:4803; Thykjaer (1997) *supra*; Park (1996) *Plant Mol. Biol.* 32:1135-1148, discussing T-DNA integration into genomic DNA. See also D'Halluin, U.S. Patent No. 5,712,135, describing a process for the stable integration of a DNA comprising a gene that is functional in a cell of a cereal, or other monocotyledonous plant.

[0304] In one aspect, the third step can involve selection and regeneration of whole plants capable of transmitting the incorporated target gene to the next generation. Such regeneration techniques rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker that has been introduced together with the desired nucleotide sequences. Plant regeneration from cultured protoplasts is described in Evans et al., *Protoplasts Isolation and Culture*, *Handbook of Plant Cell Culture*, pp. 124-176, MacMillan Publishing Company, New York, 1983; and Binding, *Regeneration of Plants, Plant Protoplasts*, pp. 21-73, CRC Press, Boca Raton, 1985. Regeneration can also be obtained from plant callus, explants, organs, or parts thereof. Such regeneration techniques are described generally in Klee (1987) *Ann. Rev. of Plant Phys.* 38:467-486. To obtain whole plants from transgenic tissues such as immature embryos, they can be grown under controlled environmental conditions in a series of media containing nutrients and hormones, a process known as tissue culture. Once whole plants are generated and produce seed, evaluation of the progeny begins.

[0305] After the expression cassette is stably incorporated in transgenic plants, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed. Since transgenic expression of the nucleic acids of the invention leads to phenotypic changes, plants comprising the recombinant nucleic acids of the invention can be sexually crossed with a second plant to obtain a final product. Thus, the seed of the invention can be derived from a cross between two transgenic plants of the invention, or a cross between a plant of the invention and another plant. The desired effects (e.g., expression of the polypeptides of the invention to produce a plant in which flowering behavior is altered) can be enhanced when both parental plants express the polypeptides (e.g., a pectate lyase) of the invention. The desired effects can be passed to future plant generations by standard propagation means.

[0306] The nucleic acids and polypeptides of the invention are expressed in or inserted in any plant or seed. Transgenic plants of the invention can be dicotyledonous or monocotyledonous. Examples of monocot transgenic plants of the invention are grasses, such as meadow grass (blue grass, *Poa*), forage grass such as fescue, *Lolium*, temperate grass, such as *Agrostis*, and cereals, e.g., wheat, oats, rye, barley, rice, sorghum, and maize (corn). Examples of dicot transgenic plants of the invention are tobacco, legumes, such as lupins, potato, sugar beet, pea, bean and soybean, and cruciferous plants (family *Brassicaceae*), such as cauliflower, rape seed, and the closely related model organism *Arabidopsis thaliana*. Thus, the transgenic plants and seeds of the invention include a broad range of plants, including, but not limited to, species from the genera *Anacardium*, *Arachis*, *Asparagus*, *Atropa*, *Avena*, *Brassica*, *Citrus*, *Citrullus*, *Capsicum*, *Carthamus*, *Cocos*, *Coffea*, *Cucumis*, *Cucurbita*, *Daucus*, *Elaeis*, *Fragaria*, *Glycine*, *Gossypium*, *Helianthus*, *Heterocallis*, *Hordeum*, *Hyoscyamus*, *Lactuca*, *Linum*, *Lolium*, *Lupinus*, *Lycopersicon*, *Malus*, *Manihot*, *Majorana*, *Medicago*, *Nicotiana*, *Olea*, *Oryza*, *Panicum*, *Pennisetum*, *Persea*, *Phaseolus*, *Pistachia*, *Pisum*, *Pyrus*, *Prunus*, *Raphanus*, *Ricinus*, *Secale*, *Senecio*, *Sinapis*, *Solanum*, *Sorghum*, *Theobromus*, *Trigonella*, *Triticum*, *Vicia*, *Vitis*, *Vigna*, and *Zea*.

[0307] In alternative embodiments, the nucleic acids of the invention are expressed in plants which contain fiber cells, including, e.g., cotton, silk cotton tree (Kapok, *Ceiba pentandra*), desert willow, creosote bush, winterfat, balsa, ramie, kenaf, hemp, roselle, jute, sisal abaca and flax. In alternative embodiments, the transgenic plants of the invention can be members of the genus *Gossypium*, including members of any *Gossypium* species, such as *G. arboreum*; *G. herbaceum*, *G. barbadense*, and *G. hirsutum*.

[0308] The invention also provides for transgenic plants to be used for producing large amounts of the polypeptides (e.g., a pectate lyase or antibody) of the invention. For example, see Palmgren (1997) Trends Genet. 13:348; Chong (1997) Transgenic Res. 6:289-296 (producing human milk protein beta-casein in transgenic potato plants using an auxin-inducible, bidirectional mannopine synthase (*mas1'*,*2'*) promoter with *Agrobacterium tumefaciens*-mediated leaf disc transformation methods).

[0309] Using known procedures, one of skill can screen for plants of the invention by detecting the increase or decrease of transgene mRNA or protein in transgenic plants. Means for detecting and quantitation of mRNAs or proteins are well known in the art.

Polypeptides and peptides

[0310] In one aspect, the invention provides isolated or recombinant polypeptides having a sequence identity (e.g., at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity) to SEQ ID NO:78. The polypeptide has a pectate lyase (e.g., pectinase) activity.

[0311] The identity can be over the full length of the polypeptide, or, the identity can be over a region of at least 300, 350, 400, 450, 500, 550, 600, 650 or more residues. Polypeptides can also be shorter than the full length of exemplary polypeptides. Polypeptides (peptides, fragments) may range in size between about 5 and the full length of a polypeptide, e.g., an enzyme, such as a pectate lyase; exemplary sizes being of about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 100, 125, 150, 175, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650 or more residues, e.g., contiguous residues of an exemplary pectate lyase of the invention. Such peptides can be useful as, e.g., labeling probes, antigens, toleragens, motifs, pectate lyase active sites, carbohydrate binding domains, and the like. Such polypeptides also include antibodies capable of binding to an enzyme of the invention.

[0312] The polypeptides of the invention include pectate lyases in an active or inactive form. For example, the polypeptides of the invention include proproteins before "maturation" or processing of prepro sequences, e.g., by a proprotein-processing enzyme, such as a proprotein convertase to generate an "active" mature protein. The polypeptides of the invention include pectate lyases inactive for other reasons, e.g., before "activation" by a post-translational processing event, e.g., an endo- or exo-

peptidase or proteinase action, a phosphorylation event, an amidation, a glycosylation or a sulfation, a dimerization event, and the like. Methods for identifying "prepro" domain sequences and signal sequences are well known in the art, see, e.g., Van de Ven (1993) Crit. Rev. Oncog. 4(2): 115-136. For example, to identify a prepro sequence, the protein is purified from the extracellular space and the N-terminal protein sequence is determined and compared to the unprocessed form.

[0313] The polypeptides of the invention include all active forms, including active subsequences, e.g., catalytic domains or active sites, of an enzyme of the invention. In one aspect, the invention provides catalytic domains or active sites as set forth below. In one aspect, the invention provides a peptide or polypeptide comprising or consisting of an active site domain as predicted through use of a database such as Pfam (which is a large collection of multiple sequence alignments and hidden Markov models covering many common protein families, The Pfam protein families database, A. Bateman, E. Birney, L. Cerruti, R. Durbin, L. Eweller, S.R. Eddy, S. Griffiths-Jones, K.L. Howe, M. Marshall, and E.L.L. Sonnhammer, Nucleic Acids Research, 30(1):276-280, 2002) or equivalent.

[0314] The invention includes polypeptides with or without a signal sequence and/or a prepro sequence. The invention includes polypeptides with heterologous signal sequences and/or prepro sequences. The prepro sequence (including a sequence of the invention used as a heterologous prepro domain) can be located on the amino terminal or the carboxy terminal end of the protein. The invention also includes isolated or recombinant signal sequences, prepro sequences and catalytic domains (e.g., "active sites") comprising sequences of the invention.

[0315] Polypeptides and peptides of the invention can be isolated from natural sources, be synthetic, or be recombinantly generated polypeptides. Peptides and proteins can be recombinantly expressed *in vitro* or *in vivo*. The peptides and polypeptides of the invention can be made and isolated using any method known in the art. Polypeptide and peptides of the invention can also be synthesized, whole or in part, using chemical methods well known in the art. See e.g., Caruthers (1980) Nucleic Acids Res. Symp. Ser. 215-223; Horn (1980) Nucleic Acids Res. Symp. Ser. 225-232; Banga, A.K., Therapeutic Peptides and Proteins, Formulation, Processing and Delivery Systems (1995) Technomic Publishing Co., Lancaster, PA. For example, peptide synthesis can be performed using various solid-phase techniques (see e.g., Roberge (1995) Science 269:202; Merrifield (1997) Methods Enzymol. 289:3-13) and automated synthesis may be achieved, e.g., using the ABI 431A Peptide Synthesizer (Perkin Elmer) in accordance with the instructions provided by the manufacturer.

[0316] The peptides and polypeptides of the invention can also be glycosylated. The glycosylation can be added post-translationally either chemically or by cellular biosynthetic mechanisms, wherein the later incorporates the use of known glycosylation motifs, which can be native to the sequence or can be added as a peptide or added in the nucleic acid coding sequence. The glycosylation can be O-linked or N-linked.

[0317] The peptides and polypeptides of the invention, as defined above, include all "mimetic" and "peptidomimetic" forms. The terms "mimetic" and "peptidomimetic" refer to a synthetic chemical compound which has substantially the same structural and/or functional characteristics of the polypeptides of the invention. The mimetic can be either entirely composed of synthetic, non-natural analogues of amino acids, or, is a chimeric molecule of partly natural peptide amino acids and partly non-natural analogs of amino acids. The mimetic can also incorporate any amount of natural amino acid conservative substitutions as long as such substitutions also do not substantially alter the mimetic's structure and/or activity. As with polypeptides of the invention which are conservative variants, routine experimentation will determine whether a mimetic is within the scope of the invention, i.e., that its structure and/or function is not substantially altered. Thus, in one aspect, a mimetic composition is within the scope of the invention if it has a peptidase activity.

[0318] Polypeptide mimetic compositions of the invention can contain any combination of non-natural structural components. In alternative aspect, mimetic compositions of the invention include one or all of the following three structural groups: a) residue linkage groups other than the natural amide bond ("peptide bond") linkages; b) non-natural residues in place of naturally occurring amino acid residues; or c) residues which induce secondary structural mimicry, i.e., to induce or stabilize a secondary structure, e.g., a beta turn, gamma turn, beta sheet, alpha helix conformation, and the like. For example, a polypeptide of the invention can be characterized as a mimetic when all or some of its residues are joined by chemical means other than natural peptide bonds. Individual peptidomimetic residues can be joined by peptide bonds, other chemical bonds or coupling means, such as, e.g., glutaraldehyde, N-hydroxysuccinimide esters, bifunctional maleimides, N,N'-dicyclohexylcarbodiimide (DCC) or N,N'-diisopropylcarbodiimide (DIC). Linking groups that can be an alternative to the traditional amide bond ("peptide bond") linkages include, e.g., ketomethylene (e.g., -C(=O)-CH₂- for -C(=O)-NH-), aminomethylene (CH₂-NH), ethylene, olefin (CH=CH), ether (CH₂-O), thioether (CH₂-S), tetrazole (CN₄-), thiazole; retroamide, thioamide, or ester (see, e.g., Spatola (1983) in Chemistry and Biochemistry of Amino Acids, Peptides and Proteins, Vol. 7, pp 267-357, "Peptide Backbone Modifications," Marcell Dekker, NY).

[0319] A polypeptide of the invention can also be characterized as a mimetic by containing all or some non-natural residues in place of naturally occurring amino acid residues. Non-natural residues are well described in the scientific and patent literature; a few exemplary non-natural compositions useful as mimetics of natural amino acid residues and guidelines are described below. Mimetics of aromatic amino acids can be generated by replacing by, e.g., D- or L- naphylalanine; D- or L- phenylglycine; D- or L-2 thieneylalanine; D- or L-1, -2, 3-, or 4- pyreneylalanine; D- or L-3 thieneylalanine; D- or L-(2-pyridinyl)-alanine; D- or L-(3-pyridinyl)-alanine; D- or L-(2-pyrazinyl)-alanine; D- or L-(4-isopropyl)-phenylglycine; D-(trifluoromethyl)-phenylglycine; D-(trifluoromethyl)-phenylalanine; D-p-fluoro-phenylalanine; D- or L-p-biphenylphenylalanine; D- or L-p-methoxy-biphenylphenylalanine; D- or L-2-indole(alkyl)alanines; and, D- or L-alkylainines, where alkyl can be substituted or unsubstituted methyl, ethyl, propyl, hexyl, butyl, pentyl, isopropyl, iso-butyl, sec-isotyl, iso-pentyl, or a non-acidic amino acids. Aromatic rings of a non-natural amino acid include, e.g., thiazolyl, thiophenyl, pyrazolyl, benzimidazolyl, naphthyl, furanyl, pyrrolyl, and pyridyl aromatic rings.

[0320] Mimetics of acidic amino acids can be generated by substitution by, e.g., non-carboxylate amino acids while maintaining a negative charge; (phosphono)alanine; sulfated threonine. Carboxyl side groups (e.g., aspartyl or glutamyl) can also be selectively modified by reaction with carbodiimides (R'-N-C-N-R') such as, e.g., 1-cyclohexyl-3(2-morpholinyl-(4-ethyl) carbodiimide or 1-ethyl-3(4-azonia- 4,4-dimethylpentyl) carbodiimide. Aspartyl or glutamyl can also be converted to asparaginyl and glutaminyl residues by reaction with ammonium ions. Mimetics of basic amino acids can be generated by substitution with, e.g., (in addition to lysine and arginine) the amino acids ornithine, citrulline, or (guanidino)-acetic acid, or (guanidino)alkyl-acetic acid, where alkyl is defined above. Nitrile derivative (e.g., containing the CN-moiety in place of COOH) can be substituted for asparagine or glutamine. Asparaginyl and glutaminyl residues can be deaminated to the corresponding aspartyl or glutamyl residues. Arginine residue mimetics can be generated by reacting arginyl with, e.g., one or more conventional reagents, including, e.g., phenylglyoxal, 2,3-butanedione, 1,2-cyclohexanedione, or ninhydrin, preferably under alkaline conditions. Tyrosine residue mimetics can be generated by reacting tyrosyl with, e.g., aromatic diazonium compounds or tetranitromethane. N-acetylimidazol and tetranitromethane can be used to form O-acetyl tyrosyl species and 3-nitro derivatives, respectively. Cysteine residue mimetics can be generated by reacting cysteinyl residues with, e.g., alpha-haloacetates such as 2-chloroacetic acid or chloroacetamide and corresponding amines; to give carboxymethyl or carboxyamidomethyl derivatives. Cysteine residue mimetics can also be generated by reacting cysteinyl residues with, e.g., bromo-trifluoroacetone, alpha-bromo-beta-(5-imidozoyl) propionic acid; chloroacetyl phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide; methyl 2-pyridyl disulfide; p-chloromercuribenzoate; 2-chloromercuri-4 nitrophenol; or, chloro-7-nitrobenzo-oxa-1,3-diazole. Lysine mimetics can be generated (and amino terminal residues can be altered) by reacting lysinyl with, e.g., succinic or other carboxylic acid anhydrides. Lysine and other alpha-amino-containing residue mimetics can also be generated by reaction with imidoesters, such as methyl picolinimide, pyridoxal phosphate, pyridoxal, chloroborohydride, trinitro-benzenesulfonic acid, O-methylisourea, 2,4, pentanedione, and transamidase-catalyzed reactions with glyoxylate. Mimetics of methionine can be generated by reaction with, e.g., methionine sulfoxide. Mimetics of proline include, e.g., pipecolic acid, thiazolidine carboxylic acid, 3- or 4- hydroxy proline, dehydroproline, 3- or 4-methylproline, or 3,3,-dimethylproline. Histidine residue mimetics can be generated by reacting histidyl with, e.g., diethylprocarbonate or para-bromophenacyl bromide. Other mimetics include, e.g., those generated by hydroxylation of proline and lysine; phosphorylation of the hydroxyl groups of seryl or threonyl residues; methylation of the alpha-amino groups of lysine, arginine and histidine; acetylation of the N-terminal amine; methylation of main chain amide residues or substitution with N-methyl amino acids; or amidation of C-terminal carboxyl groups.

[0321] A residue, e.g., an amino acid, of a polypeptide of the invention can also be replaced by an amino acid (or peptidomimetic residue) of the opposite chirality. Thus, any amino acid naturally occurring in the L-configuration (which can also be referred to as the R or S, depending upon the structure of the chemical entity) can be replaced with the amino acid of the same chemical structural type or a peptidomimetic, but of the opposite chirality, referred to as the D- amino acid, but also can be referred to as the R- or S- form.

[0322] Methods may modify the polypeptides of the invention by either natural processes, such as post-translational processing (e.g., phosphorylation, acylation, etc), or by chemical modification techniques, and the resulting modified polypeptides. Modifications can occur anywhere in the polypeptide, including the peptide backbone, the amino acid side-chains and the amino or carboxyl termini. It will be appreciated that the same type of modification may be present in the same or varying degrees at several sites in a given polypeptide. Also a given polypeptide may have many types of modifications. Modifications include acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of a phosphatidylinositol, cross-linking cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, pegylation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, and transfer-RNA mediated addition of amino acids to protein such as arginylation. See, e.g., Creighton, T.E., *Proteins - Structure and Molecular Properties* 2nd Ed., W.H. Freeman and Company, New York (1993); Posttranslational

Covalent Modification of Proteins, B.C. Johnson, Ed., Academic Press, New York, pp. 1-12(1983).

[0323] Solid-phase chemical peptide synthesis methods can also be used to synthesize the polypeptide or fragments of the invention. Such method have been known in the art since the early 1960's (Merrifield, R. B., J. Am. Chem. Soc., 85:2149-2154, 1963) (See also Stewart, J. M. and Young, J. D., Solid Phase Peptide Synthesis, 2nd Ed., Pierce Chemical Co., Rockford, Ill., pp. 11-12)) and have recently been employed in commercially available laboratory peptide design and synthesis kits (Cambridge Research Biochemicals). Such commercially available laboratory kits have generally utilized the teachings of H. M. Geysen et al, Proc. Natl. Acad. Sci., USA, 81:3998 (1984) and provide for synthesizing peptides upon the tips of a multitude of "rods" or "pins" all of which are connected to a single plate. When such a system is utilized, a plate of rods or pins is inverted and inserted into a second plate of corresponding wells or reservoirs, which contain solutions for attaching or anchoring an appropriate amino acid to the pin's or rod's tips. By repeating such a process step, i.e., inverting and inserting the rod's and pin's tips into appropriate solutions, amino acids are built into desired peptides. In addition, a number of available Fmoc peptide synthesis systems are available. For example, assembly of a polypeptide or fragment can be carried out on a solid support using an Applied Biosystems, Inc. Model 431A™ automated peptide synthesizer. Such equipment provides ready access to the peptides of the invention, either by direct synthesis or by synthesis of a series of fragments that can be coupled using other known techniques.

[0324] The invention includes pectate lyases of the invention with and without signal. The polypeptide comprising a signal sequence of the invention can be a pectate lyase of the invention or another pectate lyase or another enzyme or other polypeptide.

[0325] The disclosure includes immobilized pectate lyases, anti-pectate lyase antibodies and fragments thereof. The disclosure provides methods for inhibiting pectate lyase activity, e.g., using dominant negative mutants or anti-pectate lyase antibodies. The invention includes heterocomplexes, e.g., fusion proteins, heterodimers, etc., comprising the pectate lyases of the invention.

[0326] Polypeptides of the invention can have a pectate lyase activity under various conditions, e.g., extremes in pH and/or temperature, oxidizing agents, and the like. Methods may lead to alternative pectate lyase preparations with different catalytic efficiencies and stabilities, e.g., towards temperature, oxidizing agents and changing wash conditions. In one aspect, pectate lyase variants can be produced using techniques of site-directed mutagenesis and/or random mutagenesis. In one aspect, directed evolution can be used to produce a great variety of pectate lyase variants with alternative specificities and stability.

[0327] The proteins of the invention are also useful as research reagents to identify pectate lyase modulators, e.g., activators or inhibitors of pectate lyase activity. Briefly, test samples (compounds, broths, extracts, and the like) are added to pectate lyase assays to determine their ability to inhibit substrate cleavage. Inhibitors identified in this way can be used in industry and research to reduce or prevent undesired proteolysis. As with pectate lyases, inhibitors can be combined to increase the spectrum of activity.

[0328] The disclosure also provides methods of discovering new pectate lyases using the nucleic acids, polypeptides and antibodies described herein, lambda phage libraries may be screened for expression-based discovery of pectate lyases, to allow detection of toxic clones; improved access to substrate; reduced need for engineering a host, by-passing the potential for any bias resulting from mass excision of the library; and, faster growth at low clone densities. Screening of lambda phage libraries can be in liquid phase or in solid phase. In one aspect, the invention provides screening in liquid phase. This gives a greater flexibility in assay conditions; additional substrate flexibility; higher sensitivity for weak clones; and ease of automation over solid phase screening.

[0329] Screening method may use the proteins and nucleic acids of the invention and robotic automation enables the execution of many thousands of biocatalytic reactions and screening assays in a short period of time, e.g., per day, as well as ensuring a high level of accuracy and reproducibility (see discussion of arrays, below). As a result, a library of derivative compounds can be produced in a matter of weeks. For further teachings on modification of molecules, including small molecules, see PCT/US94/09174.

[0330] The present invention includes pectate lyase enzymes which are non-naturally occurring carbonyl hydrolase variants (e.g., pectate lyase variants) having a different proteolytic activity, stability, substrate specificity, pH profile and/or performance characteristic as compared to the precursor carbonyl hydrolase from which the amino acid sequence of the variant is derived. Specifically, such pectate lyase variants have an amino acid sequence not found in nature, which is derived by substitution of a plurality of amino acid residues of a precursor pectate lyase with different amino acids. The precursor pectate lyase may be a naturally-occurring pectate lyase or a recombinant pectate lyase. The useful pectate lyase variants encompass the substitution of any of the naturally occurring L-amino acids at the designated amino acid residue positions.

Gene site saturation mutagenesis (GSSM™) variants

[0331] The invention provides pectate lyase variants and the nucleic acids that encode them. In one aspect, the invention provides SEQ ID NO: 134, encoded by SEQ ID NO: 133, respectively. SEQ ID NO: 133 is a nucleic acid variant generated by gene site saturation mutagenesis (GSSM™) of SEQ ID NO: 131 (which encodes SEQ ID NO: 132). SEQ ID NO:131 and SEQ ID NO:132 are truncated variations of the nucleic acid as set forth in SEQ ID NO:77, encoding SEQ ID NO:78, respectively. The following Table 1 summarizes the amino acid changes resulting from the GSSM™- generated variations in their respective encoding nucleic acids (the full length SEQ ID NO:78 encoded by SEQ ID NO:77, and the truncated "parent" SEQ ID NO:132 encoded by SEQ ID NO:131:

Table 1			
Mutation - including amino acid position in SEQ ID NOS:131, 132	Nucleotide position in truncated wild-type gene (SEQ ID NOS:131, 132)	Nucleotide position in full length wild-type gene (SEQ ID NOS:77, 78)	Amino acid position in full length wild-type gene (SEQ ID NOS:77, 78)
A118H	352-354	1423-1425	475
A182V	544-546	1615-1617	539
T190L	568-570	1639-1641	547
A197G	589-591	1660-1662	554
S208K	622-624	1693-1695	565
T219M	655-657	1726-1728	576
T223E	667-669	1738-1740	580
S255R	763-765	1834-1836	612
S263K	787-789	1858-1860	620
N275Y	823-825	1894-1896	632
Y309W	925-927	1996-1998	666
S312V	934-936	2005-2007	669

[0332] Figure 6 is a table summarizing exemplary sequence changes in pectate lyase polypeptides of the invention, characterized as "upmutants." The upmutants identified as A-S are combinatorial upmutants (each have several GSSM™-generated changes). The upmutants identified as AA-LL are single upmutants (one GSSM™-generated change each).

[0333] Figure 7 is a table summarizing exemplary melting temperatures and specific activities (SA) of exemplary enzymes of the invention at various temperatures. Specific activity (U/mg pure enzyme) was measured at different temperatures at pH 9.5 in 25 mM Glycine NaOH 25 mM TrisHCl buffer. One unit of enzymatic activity was defined as the amount of enzyme that produced 1 μ mol of unsaturated oligogalacturonides equivalent to 1 μ mol of unsaturated digalacturonide per minute. Protein concentrations of the pure enzyme preparations were measured at A280 using a molar extinction coefficient of 73800 M⁻¹cm⁻¹ (1 A280 eq. to 0.50 mg/mL). Melting temperatures were determined with a differential scanning calorimeter.

[0334] In these Figures, mutant "N" has a sequence as set forth in SEQ ID NO: 134, encoded by SEQ ID NO:133.

Pectate lyase signal sequences, pectin methyl esterase domains and catalytic domains, carbohydrate binding modules and prepro domains

[0335] The disclosure provides signal sequences (e.g., signal peptides (SPs)), prepro domains and catalytic domains (CDs). The SPs, prepro domains and/or CDs can be isolated or recombinant peptides or can be part of a fusion protein, e.g., as a heterologous domain in a chimeric protein. The disclosure provides nucleic acids encoding these catalytic domains (CDs), prepro domains and signal sequences (SPs, e.g., a peptide having a sequence comprising/ consisting of amino terminal residues of a polypeptide of the invention).

[0336] The disclosure provides pectate lyase signal sequences (e.g., signal peptides (SPs)) and nucleic acids encoding these signal sequences, e.g., a peptide having a sequence comprising/ consisting of amino terminal residues of a polypeptide of the

invention, e.g., signal peptides (SPs) as set forth in Table 2, below. A signal sequence may comprise a peptide comprising/consisting of a sequence as set forth in residues 1 to 15, 1 to 16, 1 to 17, 1 to 18, 1 to 19, 1 to 20, 1 to 21, 1 to 22, 1 to 23, 1 to 24, 1 to 25, 1 to 26, 1 to 27, 1 to 28, 1 to 28, 1 to 30, 1 to 31, 1 to 32, 1 to 33, 1 to 34, 1 to 35, 1 to 36, 1 to 37, 1 to 38, 1 to 39, 1 to 40, 1 to 41, 1 to 42, 1 to 43, 1 to 44 of a polypeptide of the disclosure, e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO: 10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:98, SEQ ID NO: 100, SEQ ID NO:102, SEQ ID NO:104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO: 110, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 116, SEQ ID NO:118, SEQ ID NO: 120, SEQ ID NO:122, SEQ ID NO:124, SEQ ID NO:126, SEQ ID NO:128, SEQ ID NO: 130, SEQ ID NO: 132, SEQ ID NO: 134.

[0337] The invention also provides pectate lyase pectin methyl esterase domains (PEDs) and catalytic domains (CDs) as set forth in Table 2, below.

[0338] The pectate lyase signal sequences (SPs), CDs, and/or prepro sequences can be isolated peptides, or, sequences joined to another hydrolase or a non-pectate lyase polypeptide, e.g., as a fusion (chimeric) protein. Polypeptides may comprise such pectate lyase signal sequences. Polypeptides may comprise such pectate lyase signal sequences SPs, CDs, and/or prepro and comprise sequences heterologous to pectate lyases of the invention (e.g., a fusion protein comprising an SP, CD, and/or prepro of the polypeptide of the invention and sequences from another pectate lyase or a non-pectate lyase protein). In one aspect, the invention provides pectate lyases of the invention with heterologous SPs, CDs, and/or prepro sequences, e.g., sequences with a yeast signal sequence. A pectate lyase of the invention can comprise a heterologous SP and/or prepro in a vector, e.g., a pPIC series vector (Invitrogen, Carlsbad, CA).

[0339] Table 2 summarizes signal sequences (i.e., signal peptides in their isolated form), catalytic domains, carbohydrate binding modules and pectin methyl esterase domains. For example, Table 2 describes: in row 1, a signal peptide (SP) at residues 1 to 28 of SEQ ID NO: 102 (encoded by SEQ ID NO:101) and a catalytic domain (CD) at residues 78-459 of SEQ ID NO: 102; in row 2, a signal peptide (SP) at residues 1 to 21 of SEQ ID NO:2 (encoded by SEQ ID NO:1), a pectin methyl esterase domain (PED) at residues 28-308, and a catalytic domain (CD) at residues 309-638; at row 3, etc.

Table 2

SEQ ID NO:	Modules (SP =signal peptide, CD=catalytic domain, CBM=carbohydrate binding module, PED=pectin methyl esterase domain)
101, 102	SP;1-28, CD;78-459
1, 2	SP;1-21, PED;28-308, CD;309-638
103, 104	SP;1-26, CD;27-366
105, 106	SP;1-43, CD;44-400
107, 108	SP;1-31, CD;32-357
109, 110	SP;1-21, PED;28-308, CD309-637
11, 12	CD;1-388
111, 112	SP;1-27, CD;82-461
113, 114	SP;1-18, CD;19-388
115, 116	CD; 1-331
117, 118	SP;1-24, CD;25-574
119, 120	CBM;1-61, CBM;134-257, CD;258-615
121, 122	SP;1-29, CD;30-348
123, 124	SP;1-21, CD;22-390
125, 126	CD;24-325
127, 128	SP;1-24, CD;125-482
129, 130	CD;38-326
13, 14	SP;1-22, CD;23-354

SEQ ID NO:	Modules (SP =signal peptide, CD=catalytic domain, CBM=carbohydrate binding module, PED=pectin methyl esterase domain)
15, 16	SP;1-33, CD;34-359
17, 18	CD;1-348
19, 20	CD;1-373
21, 22	SP;1-23, CD;24-422
23, 24	SP;1-18, CD;19-393
25, 26	SP;1-15, CD;16-397
27, 28	SP;1-21, PED;28-308, CD;309-638
29, 30	SP;1-27, CD;77-459
3, 4	SP;1-28, CD;81-476
31, 32	CD;1-348
33, 34	SP;1-18, CD;19-346
35, 36	CD;1-356
37, 38	SP;1-35, CD;36-387
39, 40	SP;1-32, CD; 33-358
41, 42	SP;1-21, CD;22-359
43, 44	CBM;4-89, CBM;152-275, CD;277-633
45, 46	SP;1-20, CD;21-328
47, 48	SP;1-21, CD;22-358
49, 50	SP;1-16, CD;17-340
5, 6	CD;1-358
51, 52	CD;1-376
53, 54	SP;1-31, CBM;32-124, CBM;180-303, CD;304-658
55, 56	CD;1-374
57, 58	CD;1-389
59, 60	SP;1-24, CD;25-359
61, 62	CD;90-407
63, 64	SP;1-16, CD;17-340
65, 66	SP;1-28, CD;29-436
67, 68	SP;1-32, CBM;33-126, CBM;184-307, CD;308-664
69, 70	SP;1-22, CD;23-344
7, 8	CD;1-374
71, 72	SP;1-20, CD;21-345
73, 74	SP;1-22, CD;23-406
75, 76	SP;1-34, CD;110-555
77, 78	SP;1-33, CBM;34-126, CBM;199-322, CD;323-680
79, 80	SP;1-28, CD;81-458
81, 82	SP;1-30, CD;31-354
83, 84	PED;268-556, CD;782-1164
85, 86	CD;1-383
87, 88	SP;1-32, CD; 33-375
89, 90	SP;1-31, CD;32-459
9, 10	SP;1-29, CD;30-371
91, 92	CD;1-374

SEQ ID NO:	Modules (SP =signal peptide, CD=catalytic domain, CBM=carbohydrate binding module, PED=pectin methyl esterase domain)
93, 94	CD;1-353
95, 96	SP;1-31, CD;32-357
97, 98	PED;45-333, CD;336-698
99, 100	SP;1-35, CD;36-593

[0340] In one aspect, SPs, CDs, and/or prepro sequences are identified following identification of novel pectate lyase polypeptides. The pathways by which proteins are sorted and transported to their proper cellular location are often referred to as protein targeting pathways. One of the most important elements in all of these targeting systems is a short amino acid sequence at the amino terminus of a newly synthesized polypeptide called the signal sequence. This signal sequence directs a protein to its appropriate location in the cell and is removed during transport or when the protein reaches its final destination. Most lysosomal, membrane, or secreted proteins have an amino-terminal signal sequence that marks them for translocation into the lumen of the endoplasmic reticulum. The signal sequences can vary in length from 13 to 45 or more amino acid residues. Various methods of recognition of signal sequences are known to those of skill in the art. For example, in one aspect, novel pectate lyase signal peptides are identified by a method referred to as SignalP. SignalP uses a combined neural network which recognizes both signal peptides and their cleavage sites. (Nielsen, et al., "Identification of prokaryotic and eukaryotic signal peptides and prediction of their cleavage sites." Protein Engineering, vol. 10, no. 1, p. 1-6 (1997).

[0341] In some aspects pectate lyases of the invention do not have SPs and/or prepro sequences, and/or catalytic domains (CDs). In one aspect, the invention provides polypeptides (e.g., pectate lyases) lacking all or part of an SP, a CD and/or a prepro domain. In one aspect, the invention provides a nucleic acid sequence encoding a signal sequence (SP), a CD, and/or prepro from one pectate lyase operably linked to a nucleic acid sequence of a different pectate lyase or, optionally, a signal sequence (SPs) and/or prepro domain from a non-pectate lyase protein may be desired.

[0342] The invention also provides isolated or recombinant polypeptides comprising signal sequences (SPs), prepro domains, pectin methyl esterase domains (PEDs) and catalytic domains (CDs) of the invention and heterologous sequences. The heterologous sequences are sequences not naturally associated (e.g., to a pectate lyase) with an SP, prepro domain, PED, and/or CD. The sequence to which the SP, prepro domains, PED and/or CD are not naturally associated can be on the SP's, prepro domain's, PED's, and/or CD's amino terminal end, carboxy terminal end, and/or on both ends of the SP, prepro domain, PED and/or CD. In one aspect, the invention provides an isolated or recombinant polypeptide comprising (or consisting of) a polypeptide comprising a signal sequence (SP), prepro domain, pectin methyl esterase domain (PED) and/or catalytic domain (CD) of the invention with the proviso that it is not associated with any sequence to which it is naturally associated (e.g., a pectate lyase sequence). Similarly in one aspect, the invention provides isolated or recombinant nucleic acids encoding these polypeptides. Thus, in one aspect, the isolated or recombinant nucleic acid of the invention comprises coding sequence for a signal sequence (SP), prepro domain, pectin methyl esterase domain (PED) and/or catalytic domain (CD) of the invention and a heterologous sequence (i.e., a sequence not naturally associated with the a signal sequence (SP), prepro domain, pectin methyl esterase domain (PED) and/or catalytic domain (CD) of the invention). The heterologous sequence can be on the 3' terminal end, 5' terminal end, and/or on both ends of the SP, prepro domain, PED and/or CD coding sequence.

Glycosylation

[0343] The peptides and polypeptides of the invention (e.g., pectate lyases, antibodies) can also be glycosylated, for example, in one aspect, comprising at least one glycosylation site, e.g., an N-linked or O-linked glycosylation. In one aspect, the polypeptide can be glycosylated after being expressed in a *P. pastoris* or a *S. pombe*. The glycosylation can be added post-translationally either chemically or by cellular biosynthetic mechanisms, wherein the later incorporates the use of known glycosylation motifs, which can be native to the sequence or can be added as a peptide or added in the nucleic acid coding sequence.

Hybrid (chimeric) pectate lyases and peptide libraries

[0344] In one aspect, the invention provides hybrid pectate lyases and fusion proteins, including peptide libraries, comprising sequences of the invention. The peptide libraries of the invention can be used to isolate peptide modulators (e.g., activators or

inhibitors) of targets, such as pectate lyase substrates, receptors, enzymes. The peptide libraries of the invention can be used to identify formal binding partners of targets, such as ligands, e.g., cytokines, hormones and the like. In one aspect, the invention provides chimeric proteins comprising a signal sequence (SP), pectin methyl esterase domain (PED) and/or catalytic domain (CD) of the polypeptide of the invention and a heterologous sequence (see above).

[0345] In one aspect, the fusion proteins of the invention (e.g., the peptide moiety) are conformationally stabilized (relative to linear peptides) to allow a higher binding affinity for targets. The invention provides fusions of pectate lyases of the invention and other peptides, including known and random peptides. They can be fused in such a manner that the structure of the pectate lyases is not significantly perturbed and the peptide is metabolically or structurally conformationally stabilized. This allows the creation of a peptide library that is easily monitored both for its presence within cells and its quantity.

[0346] Amino acid sequence variants of the invention can be characterized by a predetermined nature of the variation, a feature that sets them apart from a naturally occurring form, e.g., an allelic or interspecies variation of a pectate lyase sequence. In one aspect, the variants of the invention exhibit the same qualitative biological activity as the naturally occurring analogue. Alternatively, the variants can be selected for having modified characteristics. In one aspect, while the site or region for introducing an amino acid sequence variation is predetermined, the mutation per se need not be predetermined. For example, in order to optimize the performance of a mutation at a given site, random mutagenesis may be conducted at the target codon or region and the expressed pectate lyase variants screened for the optimal combination of desired activity. Techniques for making substitution mutations at predetermined sites in DNA having a known sequence are well known, as discussed herein for example, M13 primer mutagenesis and PCR mutagenesis. Screening of the mutants can be done using assays of proteolytic activities. In alternative aspects, amino acid substitutions can be single residues; insertions can be on the order of from about 1 to 20 amino acids, although considerably larger insertions can be done. Deletions can range from about 1 to about 20, 30, 40, 50, 60, 70 residues or more. To obtain a final derivative with the optimal properties, substitutions, deletions, insertions or any combination thereof may be used. Generally, these changes are done on a few amino acids to minimize the alteration of the molecule. However, larger changes may be tolerated in certain circumstances.

[0347] The invention provides pectate lyases where the structure of the polypeptide backbone, the secondary or the tertiary structure, e.g., an alpha-helical or beta-sheet structure, has been modified. In one aspect, the charge or hydrophobicity has been modified. In one aspect, the bulk of a side chain has been modified. Substantial changes in function or immunological identity are made by selecting substitutions that are less conservative. For example, substitutions can be made which more significantly affect: the structure of the polypeptide backbone in the area of the alteration, for example a alpha-helical or a beta-sheet structure; a charge or a hydrophobic site of the molecule, which can be at an active site; or a side chain. The invention provides substitutions in polypeptide of the invention where (a) a hydrophilic residues, e.g. seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g. leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g. lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g. glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g. phenylalanine, is substituted for (or by) one not having a side chain, e.g. glycine. The variants can exhibit the same qualitative biological activity (i.e. pectate lyase activity) although variants can be selected to modify the characteristics of the pectate lyases as needed.

[0348] In one aspect, pectate lyases of the invention comprise epitopes or purification tags, signal sequences or other fusion sequences, etc. In one aspect, the pectate lyases of the invention can be fused to a random peptide to form a fusion polypeptide. By "fused" or "operably linked" herein is meant that the random peptide and the pectate lyase are linked together, in such a manner as to minimize the disruption to the stability of the pectate lyase structure, e.g., it retains pectate lyase activity. The fusion polypeptide (or fusion polynucleotide encoding the fusion polypeptide) can comprise further components as well, including multiple peptides at multiple loops.

[0349] In one aspect, the peptides and nucleic acids encoding them are randomized, either fully randomized or they are biased in their randomization, e.g. in nucleotide/residue frequency generally or per position. "Randomized" means that each nucleic acid and peptide consists of essentially random nucleotides and amino acids, respectively. In one aspect, the nucleic acids which give rise to the peptides can be chemically synthesized, and thus may incorporate any nucleotide at any position. Thus, when the nucleic acids are expressed to form peptides, any amino acid residue may be incorporated at any position. The synthetic process can be designed to generate randomized nucleic acids, to allow the formation of all or most of the possible combinations over the length of the nucleic acid, thus forming a library of randomized nucleic acids. The library can provide a sufficiently structurally diverse population of randomized expression products to affect a probabilistically sufficient range of cellular responses to provide one or more cells exhibiting a desired response. An interaction library may be large enough so that at least one of its members will have a structure that gives it affinity for some molecule, protein, or other factor.

Screening Methodologies and "On-line" Monitoring Devices

[0350] In practicing these methods, a variety of apparatus and methodologies can be used to in conjunction with the polypeptides and nucleic acids of the invention, e.g., to screen polypeptides for pectate lyase activity, to screen compounds as potential modulators, e.g., activators or inhibitors, of a pectate lyase activity, for antibodies that bind to a polypeptide of the invention, for nucleic acids that hybridize to a nucleic acid of the invention, to screen for cells expressing a polypeptide of the invention and the like.

Capillary Arrays

[0351] Capillary arrays, such as the GIGAMATRIX™, Diversa Corporation, San Diego, CA, can be used to in the methods of the invention. Nucleic acids or polypeptides of the invention can be immobilized to or applied to an array, including capillary arrays. Arrays can be used to screen for or monitor libraries of compositions (e.g., small molecules, antibodies, nucleic acids, etc.) for their ability to bind to or modulate the activity of a nucleic acid or a polypeptide of the invention. Capillary arrays provide another system for holding and screening samples. For example, a sample screening apparatus can include a plurality of capillaries formed into an array of adjacent capillaries, wherein each capillary comprises at least one wall defining a lumen for retaining a sample. The apparatus can further include interstitial material disposed between adjacent capillaries in the array, and one or more reference indicia formed within of the interstitial material. A capillary for screening a sample, wherein the capillary is adapted for being bound in an array of capillaries, can include a first wall defining a lumen for retaining the sample, and a second wall formed of a filtering material, for filtering excitation energy provided to the lumen to excite the sample.

[0352] A polypeptide or nucleic acid, e.g., a ligand, can be introduced into a first component into at least a portion of a capillary of a capillary array. Each capillary of the capillary array can comprise at least one wall defining a lumen for retaining the first component. An air bubble can be introduced into the capillary behind the first component. A second component can be introduced into the capillary, wherein the second component is separated from the first component by the air bubble. A sample of interest can be introduced as a first liquid labeled with a detectable particle into a capillary of a capillary array, wherein each capillary of the capillary array comprises at least one wall defining a lumen for retaining the first liquid and the detectable particle, and wherein the at least one wall is coated with a binding material for binding the detectable particle to the at least one wall. The method can further include removing the first liquid from the capillary tube, wherein the bound detectable particle is maintained within the capillary, and introducing a second liquid into the capillary tube.

[0353] The capillary array can include a plurality of individual capillaries comprising at least one outer wall defining a lumen. The outer wall of the capillary can be one or more walls fused together. Similarly, the wall can define a lumen that is cylindrical, square, hexagonal or any other geometric shape so long as the walls form a lumen for retention of a liquid or sample. The capillaries of the capillary array can be held together in close proximity to form a planar structure. The capillaries can be bound together, by being fused (e.g., where the capillaries are made of glass), glued, bonded, or clamped side-by-side. The capillary array can be formed of any number of individual capillaries, for example, a range from 100 to 4,000,000 capillaries. A capillary array can form a micro titer plate having about 100,000 or more individual capillaries bound together.

Arrays, or "Biochips"

[0354] Nucleic acids or polypeptides of the invention can be immobilized to or applied to an array. Arrays can be used to screen for or monitor libraries of compositions (e.g., small molecules, antibodies, nucleic acids, etc.) for their ability to bind to or modulate the activity of a nucleic acid or a polypeptide of the invention. For example, in one aspect of the invention, a monitored parameter is transcript expression of a pectate lyase gene. One or more, or, all the transcripts of a cell can be measured by hybridization of a sample comprising transcripts of the cell, or, nucleic acids representative of or complementary to transcripts of a cell, by hybridization to immobilized nucleic acids on an array, or "biochip." By using an "array" of nucleic acids on a microchip, some or all of the transcripts of a cell can be simultaneously quantified. Alternatively, arrays comprising genomic nucleic acid can also be used to determine the genotype of a newly engineered strain made by the methods disclosed herein. Polypeptide arrays" can also be used to simultaneously quantify a plurality of proteins. The present invention can be practiced with any known "array," also referred to as a "microarray" or "nucleic acid array" or "polypeptide array" or "antibody array" or "biochip," or variation thereof. Arrays are generically a plurality of "spots" or "target elements," each target element comprising a defined amount of one or more biological molecules, e.g., oligonucleotides, immobilized onto a defined area of a substrate surface for specific binding to a sample molecule, e.g., mRNA transcripts.

[0355] In practicing the methods, any known array and/or method of making and using arrays can be incorporated in whole or in part, or variations thereof, as described, for example, in U.S. Patent Nos. 6,277,628; 6,277,489; 6,261,776; 6,258,606; 6,054,270; 6,048,695; 6,045,996; 6,022,963; 6,013,440; 5,965,452; 5,959,098; 5,856,174; 5,830,645; 5,770,456; 5,632,957; 5,556,752; 5,143,854; 5,807,522; 5,800,992; 5,744,305; 5,700,637; 5,556,752; 5,434,049; see also, e.g., WO 99/51773; WO 99/09217; WO 97/46313; WO 96/17958; see also, e.g., Johnston (1998) *Curr. Biol.* 8:R171-R174; Schummer (1997) *Biotechniques* 23:1087-1092; Kern (1997) *Biotechniques* 23:120-124; Solinas-Toldo (1997) *Genes, Chromosomes & Cancer* 20:399-407; Bowtell (1999) *Nature Genetics Supp.* 21:25-32. See also published U.S. patent applications Nos. 20010018642; 20010019827; 20010016322; 20010014449; 20010014448; 20010012537; 20010008765.

Antibodies and Antibody-based screening methods

[0356] The disclosure provides isolated or recombinant antibodies that specifically bind to a pectate lyase of the invention. These antibodies can be used to isolate, identify or quantify the pectate lyases of the invention or related polypeptides. These antibodies can be used to isolate other polypeptides within the scope the invention or other related pectate lyases. The antibodies can be designed to bind to an active site of a pectate lyase. Methods of inhibiting pectate lyases may use these antibodies.

[0357] The disclosure provides fragments of the enzymes of the invention, including immunogenic fragments of a polypeptide of the invention which can further comprise adjuvants, carriers and the like.

[0358] The antibodies can be used in immunoprecipitation, staining, immunoaffinity columns, and the like. If desired, nucleic acid sequences encoding for specific antigens can be generated by immunization followed by isolation of polypeptide or nucleic acid, amplification or cloning and immobilization of polypeptide onto an array. Alternatively, these methods can be used to modify the structure of an antibody produced by a cell to be modified, e.g., an antibody's affinity can be increased or decreased. Furthermore, the ability to make or modify antibodies can be a phenotype engineered into a cell by these methods.

[0359] Methods of immunization, producing and isolating antibodies (polyclonal and monoclonal) are known to those of skill in the art and described in the scientific and patent literature, see, e.g., Coligan, *CURRENT PROTOCOLS IN IMMUNOLOGY*, Wiley/Greene, NY (1991); Stites (eds.) *BASIC AND CLINICAL IMMUNOLOGY* (7th ed.) Lange Medical Publications, Los Altos, CA ("Stites"); Goding, *MONOCLONAL ANTIBODIES: PRINCIPLES AND PRACTICE* (2d ed.) Academic Press, New York, NY (1986); Kohler (1975) *Nature* 256:495; Harlow (1988) *ANTIBODIES, A LABORATORY MANUAL*, Cold Spring Harbor Publications, New York. Antibodies also can be generated *in vitro*, e.g., using recombinant antibody binding site expressing phage display libraries, in addition to the traditional *in vivo* methods using animals. See, e.g., Hoogenboom (1997) *Trends Biotechnol.* 15:62-70; Katz (1997) *Annu. Rev. Biophys. Biomol. Struct.* 26:27-45.

[0360] Polypeptides or peptides can be used to generate antibodies which bind specifically to the polypeptides, e.g., the pectate lyases, of the invention. The resulting antibodies may be used in immunoaffinity chromatography procedures to isolate or purify the polypeptide or to determine whether the polypeptide is present in a biological sample. In such procedures, a protein preparation, such as an extract, or a biological sample is contacted with an antibody capable of specifically binding to one of the polypeptides of the invention.

[0361] In immunoaffinity procedures, the antibody is attached to a solid support, such as a bead or other column matrix. The protein preparation is placed in contact with the antibody under conditions in which the antibody specifically binds to one of the polypeptides of the invention. After a wash to remove non-specifically bound proteins, the specifically bound polypeptides are eluted.

[0362] The ability of proteins in a biological sample to bind to the antibody may be determined using any of a variety of procedures familiar to those skilled in the art. For example, binding may be determined by labeling the antibody with a detectable label such as a fluorescent agent, an enzymatic label, or a radioisotope. Alternatively, binding of the antibody to the sample may be detected using a secondary antibody having such a detectable label thereon. Particular assays include ELISA assays, sandwich assays, radioimmunoassays, and Western Blots.

[0363] Polyclonal antibodies generated against the polypeptides of the invention can be obtained by direct injection of the polypeptides into an animal or by administering the polypeptides to a non-human animal. The antibody so obtained will then bind the polypeptide itself. In this manner, even a sequence encoding only a fragment of the polypeptide can be used to generate antibodies which may bind to the whole native polypeptide. Such antibodies can then be used to isolate the polypeptide from cells

expressing that polypeptide.

[0364] For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique, the trioma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique (see, e.g., Cole (1985) in *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96).

[0365] Techniques described for the production of single chain antibodies (see, e.g., U.S. Patent No. 4,946,778) can be adapted to produce single chain antibodies to the polypeptides of the invention. Alternatively, transgenic mice may be used to express humanized antibodies to these polypeptides or fragments thereof.

[0366] Antibodies generated against the polypeptides of the invention may be used in screening for similar polypeptides (e.g., pectate lyases) from other organisms and samples. In such techniques, polypeptides from the organism are contacted with the antibody and those polypeptides which specifically bind the antibody are detected. Any of the procedures described above may be used to detect antibody binding.

Kits

[0367] Kits may comprise the compositions, e.g., nucleic acids, expression cassettes, vectors, cells, transgenic seeds or plants or plant parts, polypeptides (e.g., pectate lyases) and/or antibodies disclosed herein. The kits also can contain instructional material teaching the methodologies and industrial uses of the invention, as described herein.

Measuring Metabolic Parameters

[0368] Whole cell evolution, or whole cell engineering, of a cell may develop a new cell strain having a new phenotype, e.g., a new or modified pectate lyase activity, by modifying the genetic composition of the cell. The genetic composition can be modified by addition to the cell of a nucleic acid of the invention. To detect the new phenotype, at least one metabolic parameter of a modified cell is monitored in the cell in a "real time" or "on-line" time frame. A plurality of cells, such as a cell culture, may be monitored in "real time" or "on-line." A plurality of metabolic parameters may be monitored in "real time" or "on-line." Metabolic parameters can be monitored using the pectate lyases of the invention.

[0369] Metabolic flux analysis (MFA) is based on a known biochemistry framework. A linearly independent metabolic matrix is constructed based on the law of mass conservation and on the pseudo-steady state hypothesis (PSSH) on the intracellular metabolites. In practicing these methods, metabolic networks are established, including the:

- identity of all pathway substrates, products and intermediary metabolites
- identity of all the chemical reactions interconverting the pathway metabolites, the stoichiometry of the pathway reactions,
- identity of all the enzymes catalyzing the reactions, the enzyme reaction kinetics,
- the regulatory interactions between pathway components, e.g. allosteric interactions, enzyme-enzyme interactions etc,
- intracellular compartmentalization of enzymes or any other supramolecular organization of the enzymes, and,
- the presence of any concentration gradients of metabolites, enzymes or effector molecules or diffusion barriers to their movement.

[0370] Once the metabolic network for a given strain is built, mathematic presentation by matrix notion can be introduced to estimate the intracellular metabolic fluxes if the on-line metabolome data is available. Metabolic phenotype relies on the changes of the whole metabolic network within a cell. Metabolic phenotype relies on the change of pathway utilization with respect to environmental conditions, genetic regulation, developmental state and the genotype, etc. After the on-line MFA calculation, the dynamic behavior of the cells, their phenotype and other properties are analyzed by investigating the pathway utilization. For example, if the glucose supply is increased and the oxygen decreased during the yeast fermentation, the utilization of respiratory pathways will be reduced and/or stopped, and the utilization of the fermentative pathways will dominate. Control of physiological state of cell cultures will become possible after the pathway analysis. The methods of the invention can help determine how to manipulate the fermentation by determining how to change the substrate supply, temperature, use of inducers, etc. to control the physiological state of cells to move along desirable direction. The MFA results can also be compared with transcriptome and proteome data to design experiments and protocols for metabolic engineering or gene shuffling, etc.

[0371] Any modified or new phenotype can be conferred and detected, including new or improved characteristics in the cell. Any aspect of metabolism or growth can be monitored.

Monitoring expression of an mRNA transcript

[0372] The engineered phenotype may comprise increasing or decreasing the expression of an mRNA transcript (e.g., a pectate lyase message) or generating new (e.g., pectate lyase) transcripts in a cell. This increased or decreased expression can be traced by testing for the presence of a pectate lyase of the invention or by pectate lyase activity assays. mRNA transcripts, or messages, also can be detected and quantified by any method known in the art, including, e.g., Northern blots, quantitative amplification reactions, hybridization to arrays, and the like. Quantitative amplification reactions include, e.g., quantitative PCR, including, e.g., quantitative reverse transcription polymerase chain reaction, or RT-PCR; quantitative real time RT-PCR, or "real-time kinetic RT-PCR" (see, e.g., Kreuzer (2001) Br. J. Haematol. 114:313-318; Xia (2001) Transplantation 72:907-914).

[0373] The engineered phenotype may be generated by knocking out expression of a homologous gene. The gene's coding sequence or one or more transcriptional control elements can be knocked out, e.g., promoters or enhancers. Thus, the expression of a transcript can be completely ablated or only decreased.

[0374] The engineered phenotype may comprise increasing the expression of a homologous gene. This can be effected by knocking out of a negative control element, including a transcriptional regulatory element acting in cis- or trans-, or, mutagenizing a positive control element. One or more, or, all the transcripts of a cell can be measured by hybridization of a sample comprising transcripts of the cell, or, nucleic acids representative of or complementary to transcripts of a cell, by hybridization to immobilized nucleic acids on an array.

Monitoring expression of a polypeptides, peptides and amino acids

[0375] The engineered phenotype may comprise increasing or decreasing the expression of a polypeptide (e.g., a pectate lyase) or generating new polypeptides in a cell. This increased or decreased expression can be traced by determining the amount of pectate lyase present or by pectate lyase activity assays. Polypeptides, peptides and amino acids also can be detected and quantified by any method known in the art, including, e.g., nuclear magnetic resonance (NMR), spectrophotometry, radiography (protein radiolabeling), electrophoresis, capillary electrophoresis, high performance liquid chromatography (HPLC), thin layer chromatography (TLC), hyperdiffusion chromatography, various immunological methods, e.g. immunoprecipitation, immunodiffusion, immuno-electrophoresis, radioimmunoassays (RIAs), enzyme-linked immunosorbent assays (ELISAs), immuno-fluorescent assays, gel electrophoresis (e.g., SDS-PAGE), staining with antibodies, fluorescent activated cell sorter (FACS), pyrolysis mass spectrometry, Fourier-Transform Infrared Spectrometry, Raman spectrometry, GC-MS, and LC-Electrospray and cap-LC-tandem-electrospray mass spectrometries, and the like. Novel bioactivities can also be screened using methods, or variations thereof, described in U.S. Patent No. 6,057,103. Furthermore, as discussed below in detail, one or more, or, all the polypeptides of a cell can be measured using a protein array.

Industrial Applications

Detergent Compositions

[0376] Detergent compositions may comprise one or more polypeptides (e.g., pectate lyases) of the invention. For methods of making and using detergent compositions, see, e.g., U.S. Patent No. 6,413,928; 6,399,561; 6,365,561; 6,380,147. The detergent compositions can be a one and two part aqueous composition, a non-aqueous liquid composition, a cast solid, a granular form, a particulate form, a compressed tablet, a gel and/or a paste and a slurry form. The pectate lyases of the invention can also be used as a detergent additive product in a solid or a liquid form. Such additive products are intended to supplement or boost the performance of conventional detergent compositions and can be added at any stage of the cleaning process.

[0377] The invention also provides methods capable of removing gross food soils, films of food residue and other minor food compositions using these detergent compositions. Pectate lyases of the invention can facilitate the removal of starchy stains by

means of catalytic hydrolysis or trans-elimination of pectins, including the disruption of plant and bacterial cell walls. Pectate lyases of the invention can be used in dishwashing detergents in textile laundering detergents.

[0378] The actual active enzyme content depends upon the method of manufacture of a detergent composition and is not critical, assuming the detergent solution has the desired enzymatic activity. In one aspect, the amount of pectate lyase present in the final solution ranges from about 0.001 mg to 0.5 mg per gram of the detergent composition. The particular enzyme chosen for use in the process and products of this invention depends upon the conditions of final utility, including the physical product form, use pH, use temperature, and soil types to be degraded or altered. The enzyme can be chosen to provide optimum activity and stability for any given set of utility conditions. In one aspect, the pectate lyases of the present invention are active in the pH ranges of from about 4 to about 12 and in the temperature range of from about 20°C to about 95°C. The detergents of the invention can comprise cationic, semi-polar nonionic or zwitterionic surfactants; or, mixtures thereof.

[0379] Pectate lyases of the invention can be formulated into powdered and liquid detergents having pH between 4.0 and 12.0 at levels of about 0.01 to about 5% (preferably 0.1% to 0.5%) by weight. These detergent compositions can also include other enzymes such as proteases, cellulases, lipases or endoglycosidases, endo-beta.-1,4-glucanases, beta-glucanases, endo-beta-1,3(4)-glucanases, cutinases, peroxidases, laccases, amylases, glucoamylases, pectinases, reductases, oxidases, phenoloxidases, ligninases, pullulanases, arabinanases, hemicellulases, mannanases, xyloglucanases, xylanases, pectin acetyl esterases, rhamnogalacturonan acetyl esterases, polygalacturonases, rhamnogalacturonases, galactanases, pectin lyases, pectin methylesterases, cellobiohydrolases and/or transglutaminases. These detergent compositions can also include builders and stabilizers.

[0380] The addition of pectate lyases of the invention to conventional cleaning compositions does not create any special use limitation. In other words, any temperature and pH suitable for the detergent is also suitable for the compositions of the invention as long as the enzyme is active at or tolerant of the pH and/or temperature of the intended use. In addition, the pectate lyases of the invention can be used in a cleaning composition without detergents, again either alone or in combination with builders and stabilizers.

[0381] The present invention provides cleaning compositions including detergent compositions for cleaning hard surfaces, detergent compositions for cleaning fabrics, dishwashing compositions, oral cleaning compositions, denture cleaning compositions, and contact lens cleaning solutions.

[0382] In one aspect, the invention provides a method for washing an object comprising contacting the object with a polypeptide of the invention under conditions sufficient for washing. A pectate lyase of the invention may be included as a detergent additive. The detergent composition may, for example, be formulated as a hand or machine laundry detergent composition comprising a polypeptide of the invention. A laundry additive suitable for pre-treatment of stained fabrics can comprise a polypeptide of the invention. A fabric softener composition can comprise a pectate lyase of the invention. Alternatively, a pectate lyase of the invention can be formulated as a detergent composition for use in general household hard surface cleaning operations. In alternative aspects, detergent additives and detergent compositions may comprise one or more other enzymes such as a protease, a lipase, a cutinase, another pectate lyase, a carbohydrase, a cellulase, a pectinase, a mannanase, an arabinase, a galactanase, a xylanase, an oxidase, e.g., a lactase, and/or a peroxidase (see also, above). The properties of the enzyme(s) of the invention are chosen to be compatible with the selected detergent (i.e. pH-optimum, compatibility with other enzymatic and non-enzymatic ingredients, etc.) and the enzyme(s) is present in effective amounts. In one aspect, pectate lyase enzymes of the invention are used to remove malodorous materials from fabrics. Various detergent compositions and methods for making them that can be used in practicing the invention are described in, e.g., U.S. Patent Nos. 6,333,301; 6,329,333; 6,326,341; 6,297,038; 6,309,871; 6,204,232; 6,197,070; 5,856,164.

[0383] When formulated as compositions suitable for use in a laundry machine washing method, the pectate lyases of the invention can comprise both a surfactant and a builder compound. They can additionally comprise one or more detergent components, e.g., organic polymeric compounds, bleaching agents, additional enzymes, suds suppressors, dispersants, lime-soap dispersants, soil suspension and anti-redeposition agents and corrosion inhibitors. Laundry compositions of the invention can also contain softening agents, as additional detergent components. Such compositions containing carbohydrase can provide fabric cleaning, stain removal, whiteness maintenance, softening, color appearance, dye transfer inhibition and sanitization when formulated as laundry detergent compositions.

[0384] The density of the laundry detergent compositions of the invention can range from about 200 to 1500 g/liter, or, about 400 to 1200 g/liter, or, about 500 to 950 g/liter, or, 600 to 800 g/liter, of composition; this can be measured at about 20°C.

[0385] The "compact" form of laundry detergent compositions is best reflected by density and, in terms of composition, by the

amount of inorganic filler salt. Inorganic filler salts are conventional ingredients of detergent compositions in powder form. In conventional detergent compositions, the filler salts are present in substantial amounts, typically 17% to 35% by weight of the total composition. In one aspect of the compact compositions, the filler salt is present in amounts not exceeding 15% of the total composition, or, not exceeding 10%, or, not exceeding 5% by weight of the composition. The inorganic filler salts can be selected from the alkali and alkaline-earth-metal salts of sulphates and chlorides, e.g., sodium sulphate.

[0386] Liquid detergent compositions can also be in a "concentrated form." In one aspect, the liquid detergent compositions can contain a lower amount of water, compared to conventional liquid detergents. In alternative aspects, the water content of the concentrated liquid detergent is less than 40%, or, less than 30%, or, less than 20% by weight of the detergent composition. Detergent compounds of the invention can comprise formulations as described in WO 97/01629.

Treating fibers and textiles

[0387] The invention provides methods of treating fibers, fabrics or any pectate-or polygalacturonic acid-comprising material using one or more pectate lyases of the invention. The pectate lyases can be used in any fiber- or fabric-treating method, which are well known in the art, see, e.g., U.S. Patent No. 6,261,828; 6,077,316; 6,024,766; 6,021,536; 6,017,751; 5,980,581; US Patent Publication No. 20020142438 A1. For example, pectate lyases of the invention can be used in fiber and/or fabric scouring. In one aspect, the feel and appearance of a fabric is improved by a method of the invention comprising contacting the fabric with a pectate lyase of the invention in a solution. In one aspect, the fabric is treated with the solution under pressure. For example, pectate lyases of the invention can be used in the removal of stains.

[0388] In one aspect, pectate lyases of the invention are applied during or after the weaving of textiles, or during the desizing stage, or during one or more additional fabric processing steps. During the weaving of textiles, the threads are exposed to considerable mechanical strain. Prior to weaving on mechanical looms, warp yarns are often coated with sizing starch or starch derivatives in order to increase their tensile strength and to prevent breaking. After the textiles have been woven, a fabric can proceed to a desizing stage. This can be followed by one or more additional fabric processing steps. Desizing is the act of removing "size" from textiles. After weaving, the size coating must be removed before further processing the fabric in order to ensure a homogeneous and wash-proof result.

[0389] The enzymes of the invention can be used to scour fabrics or any pectate-or polygalacturonic acid-comprising material, including cotton-containing fabrics, as detergent additives, e.g., in aqueous compositions. For the manufacture of clothes, the fabric can be cut and sewn into clothes or garments. These can be finished before or after the treatment. In particular, for the manufacture of denim jeans, different enzymatic finishing methods have been developed. The finishing of denim garment normally is initiated with an enzymatic desizing step, during which garments are subjected to the action of amylolytic enzymes in order to provide softness to the fabric and make the cotton more accessible to the subsequent enzymatic finishing steps. The invention provides methods of finishing denim garments, enzymatic desizing and providing softness to fabrics by using any combination of enzymes, such as amylases, endoglucanases, and a pectate lyase of the invention.

[0390] In one aspect, an alkaline and thermostable amylase and pectate lyase are combined in a single bath desizing and bioscouring. Among advantages of combining desizing and scouring in one step are cost reduction and lower environmental impact due to savings in energy and water usage and lower waste production. Application conditions for desizing and bioscouring can be between about pH 8.5 to pH 10.0 and temperatures at about 40°C and up. Low enzyme dosages (e.g., about 5 g per a ton of cotton) and short reaction times (e.g., about 15 minutes) can be used to obtain efficient desizing and scouring with out added calcium.

[0391] The pectate lyases of the invention can be used in combination with other carbohydrate degrading enzymes, e.g., cellulase, arabinanase, xyloglucanase, pectinase, xylanase, and the like, for the preparation of fibers or for cleaning of fibers. Proteases can also be used in combination. These can be used in combination with detergents. In one aspect, pectate lyases of the invention can be used in treatments to prevent the graying of a textile.

[0392] The pectate lyases of the invention can be used to treat any cellulosic material, including fibers (e.g., fibers from cotton, hemp, flax or linen), sewn and unsewn fabrics, e.g., knits, wovens, denims, yarns, and toweling, made from cotton, cotton blends or natural or manmade cellulose (e.g. originating from xylan-containing cellulose fibers such as from wood pulp) or blends thereof. Examples of blends are blends of cotton or rayon/viscose with one or more companion material such as wool, synthetic fibers (e.g. polyamide fibers, acrylic fibers, polyester fibers, polyvinyl alcohol fibers, polyvinyl chloride fibers, polyvinylidene chloride fibers, polyurethane fibers, polyurea fibers, aramid fibers), and cellulose-containing fibers (e.g. rayon/viscose, ramie,

hemp, flax/linen, jute, cellulose acetate fibers, lyocell).

[0393] The textile treating processes of the invention (for example, scouring using pectate lyases of the invention) can be used in conjunction with other textile treatments, e.g., desizing and bleaching. Scouring is the removal of non-cellulosic material from the cotton fiber, e.g., the cuticle (mainly consisting of waxes) and primary cell wall (mainly consisting of pectin, protein and xyloglucan). A proper wax removal is necessary for obtaining a high wettability. This is needed for dyeing. Removal of the primary cell walls by the processes of the invention improves wax removal and ensures a more even dyeing. Treating textiles with the processes of the invention can improve whiteness in the bleaching process. The main chemical used in scouring is sodium hydroxide in high concentrations and at high temperatures. Bleaching comprises oxidizing the textile. Bleaching typically involves use of hydrogen peroxide as the oxidizing agent in order to obtain either a fully bleached (white) fabric or to ensure a clean shade of the dye.

[0394] The invention provides a single-bath process for desizing, scouring and bleaching of cellulosic materials. In one aspect, desizing, scouring and bleaching are carried in a single-bath by contacting the cellulosic materials simultaneously or sequentially in a container (a "single-bath") with an enzyme system and a bleaching system comprising hydrogen peroxide or at least one peroxy compound which can generate hydrogen peroxide when dissolved in water, or combinations thereof, and at least one bleach activator. Cellulosic materials including crude fibers, yarn, or woven or knit textiles, made of cotton, linen, flax, ramie, rayon, hemp, jute, or blends of these fibers with each other or with other natural or synthetic fibers, can be treated by the processes of the invention.

[0395] The invention also provides alkaline pectinases (pectate lyases active under alkaline conditions). These have wide-ranging applications in textile processing, degumming of plant fibers (e.g., plant bast fibers), treatment of pectic wastewaters, paper making, and coffee and tea fermentations. See, e.g., Hoondal (2002) *Applied Microbiology and Biotechnology* 59:409-418.

Treating foods and food processing

[0396] The pectate lyases of the invention have numerous applications in food processing industry. For example, in one aspect, the pectate lyases of the invention are used to improve the extraction of oil from oil-rich plant material, e.g., oil-rich seeds, for example, soybean oil from soybeans, olive oil from olives, rapeseed oil from rapeseed and/or sunflower oil from sunflower seeds.

[0397] The pectate lyases of the invention can be used for separation of components of plant cell materials. For example, pectate lyases of the invention can be used in the separation of pectin-rich material (e.g., cell walls), sugar or starch-rich plant material into components, e.g., sucrose from sugar beet or starch or sugars from potato, pulp or hull fractions. In one aspect, pectate lyases of the invention can be used to separate protein-rich or oil-rich crops into valuable protein and oil and hull fractions. The separation process may be performed by use of methods known in the art.

[0398] The pectate lyases of the invention can be used in the preparation of fruit or vegetable juices, syrups, extracts and the like to increase yield. The pectate lyases of the invention can be used in the enzymatic treatment (e.g., hydrolysis of pectins and/or polygalacturonic acid, such as 1,4-linked alpha-D-galacturonic acid) of various plant cell wall-derived materials or waste materials, e.g. from wine or juice production, or agricultural residues such as vegetable hulls, bean hulls, sugar beet pulp, olive pulp, potato pulp, and the like. The pectate lyases of the invention can be used to modify the consistency and appearance of processed fruit or vegetables. For example, the pectate lyases of the invention can be used in the production of clear juices, e.g., from apples, pears or berries; to cloud stable juices, e.g., from apples, pears, berries, citrus or tomatoes; and to treat purees, e.g., from carrots and tomatoes, and to treat date syrup (see, e.g., Sidhu (2002) *Food Chemistry* 79:215-220). In these processes, the pectate lyases of the invention can be used with other enzymes (e.g., cellulases, amylases, etc.) or other compositions. For example, in one aspect, pectinase and cellulase enzymes are used to improve juice yield, stability and quality from a fruit, e.g., prickly pear fruit. A pectinase of the invention can improve the yield, stability and color (color-assayed as release of anthocyanins or carotenoids) and clarity of a juice. In one aspect, a combination of pectinase and cellulase is used; pectinase at 0.50% v/w can produce a high yield, a sediment-free clear juice and high-quality juice. See, e.g., Essa, Hesham A., et. al., 2002, *Nahrung*, 46(4):245-250.

[0399] In one aspect, an enzyme or enzyme preparation of the invention is used for de-pectinization and viscosity reduction in vegetable and/or fruit juice, e.g., in apple or pear juices or other apple or pear food preparations (e.g., sauces). In one aspect, the fruit or vegetable juice is treated with an enzyme preparation of the invention in an amount effective for degrading pectin-containing material contained in the fruit or vegetable juice.

[0400] In one aspect, the enzyme or enzyme preparation is used in the treatment of mash from fruits and vegetables in order to improve the extractability or degradability of the mash. The enzyme preparation can be used in the treatment of mash from apples and pears for juice production, and in the mash treatment of grapes for wine production.

[0401] The pectate lyases of the invention can be used to treat plant material to facilitate processing of plant material, including foods, facilitate purification or extraction of plant components such as galactans, pectins and/or polygalacturonic acids. The pectate lyases of the invention can be used to purify pectins from citrus, improve feed value, decrease the water binding capacity, improve the degradability in waste water plants and/or improve the conversion of plant material to ensilage, and the like.

Animal feeds and food or feed additives

[0402] The invention provides methods for treating animal feeds and foods and food or feed additives using pectate lyases of the invention, animals including mammals (e.g., humans), birds, fish and the like. The invention provides animal feeds, foods, and additives comprising pectate lyases of the invention. In one aspect, treating animal feeds, foods and additives using pectate lyases of the invention can help in the availability of nutrients, e.g., starch, in the animal feed or additive. This can result in release of readily digestible and easily absorbed nutrients and sugars.

[0403] Pectate lyases of the present invention, in the modification of animal feed or a food, can process the food or feed either *in vitro* (by modifying components of the feed or food) or *in vivo*. Pectate lyases can be added to animal feed or food compositions containing high amounts of arabinogalactans or galactans, e.g. feed or food containing plant material from soy bean, rape seed, lupin and the like. When added to the feed or food the pectate lyase significantly improves the *in vivo* break-down of plant cell wall material, whereby a better utilization of the plant nutrients by the animal (e.g., human) is achieved. In one aspect, the growth rate and/or feed conversion ratio (i.e. the weight of ingested feed relative to weight gain) of the animal is improved. For example the indigestible galactan is degraded by a pectate lyase of the invention, e.g. in combination with beta-galactosidase, to galactose or galactooligomers. These enzyme digestion products are more digestible by the animal. Thus, they can contribute to the available energy of the feed. Also, by the degradation of galactan the pectate lyase of the invention can improve the digestibility and uptake of non-carbohydrate feed constituents such as protein, fat and minerals.

[0404] In another aspect, pectate lyase of the invention can be supplied by expressing the enzymes directly in transgenic feed crops (as, e.g., transgenic plants, seeds and the like), such as corn, soy bean, rape seed, lupin and the like. As discussed above, the invention provides transgenic plants, plant parts and plant cells comprising a nucleic acid sequence encoding a polypeptide of the invention. In one aspect, the nucleic acid is expressed such that the pectate lyase of the invention is produced in recoverable quantities. The pectate lyase can be recovered from any plant or plant part. Alternatively, the plant or plant part containing the recombinant polypeptide can be used as such for improving the quality of a food or feed, e.g., improving nutritional value, palatability, and rheological properties, or to destroy an antinutritive factor.

Paper or pulp treatment

[0405] The pectate lyases of the invention can be in paper or pulp treatment or paper deinking. For example, in one aspect, the invention provides a paper treatment process using pectate lyases of the invention. In one aspect, the pectate lyases can be used to modify pectin and/or polygalacturonic acid, such as 1,4-linked alpha-D-galacturonic acid. In another aspect, paper components of recycled photocopied paper during chemical and enzymatic deinking processes. In one aspect, pectate lyases of the invention can be used in combination with cellulases. The paper can be treated by the following three processes: 1) disintegration in the presence of pectate lyases of the invention, 2) disintegration with a deinking chemical and pectate lyases of the invention, and/or 3) disintegration after soaking with pectate lyases of the invention. The recycled paper treated with pectate lyases can have a higher brightness due to removal of toner particles as compared to the paper treated with just cellulase. While the invention is not limited by any particular mechanism, the effect of pectate lyases of the invention may be due to its behavior as surface-active agents in pulp suspension.

[0406] The invention provides methods of treating paper and paper pulp using one or more pectate lyases of the invention. The pectate lyases of the invention can be used in any paper- or pulp-treating method, which are well known in the art, see, e.g., U.S. Patent No. 6,241,849; 6,066,233; 5,582,681. For example, in one aspect, the invention provides a method for deinking and decolorizing a printed paper containing a dye, comprising pulping a printed paper to obtain a pulp slurry, and dislodging an ink from the pulp slurry in the presence of pectate lyases of the invention (other enzymes can also be added). In another aspect, the invention provides a method for enhancing the freeness of pulp, e.g., pulp made from secondary fiber, by adding an enzymatic

mixture comprising pectate lyases of the invention (can also include other enzymes, e.g., cellulase, amylase or glucoamylase enzymes) to the pulp and treating under conditions to cause a reaction to produce an enzymatically treated pulp. The freeness of the enzymatically treated pulp is increased from the initial freeness of the secondary fiber pulp without a loss in brightness.

Repulping: treatment of lignocellulosic materials

[0407] The invention also provides a method for the treatment of lignocellulosic fibers, wherein the fibers are treated with pectate lyases of the invention, in an amount which is efficient for improving the fiber properties. The pectate lyases of the invention may also be used in the production of lignocellulosic materials such as pulp, paper and cardboard, from starch-reinforced waste paper and cardboard, especially where repulping occurs at pH above 7 and where pectate lyases can facilitate the disintegration of the waste material through degradation of cell walls. The pectate lyases of the invention can be useful in a process for producing a papermaking pulp from starch-coated printed paper. The process may be performed as described in, e.g., WO 95/14807.

[0408] An exemplary process comprises disintegrating the paper to produce a pulp, treating with a pectin-degrading enzyme of the invention before, during or after the disintegrating, and separating ink particles from the pulp after disintegrating and enzyme treatment. See also U.S. Patent No. 6,309,871 and other US patents cited herein. Thus, the invention includes a method for enzymatic deinking of recycled paper pulp, wherein pectate lyases are applied in an amount which is efficient for effective deinking of the fiber surface.

Waste treatment

[0409] The pectate lyases of the invention can be used in a variety of other industrial applications, e.g., in waste treatment. For example, in one aspect, the invention provides a solid waste digestion process using pectate lyases of the invention. The methods can comprise reducing the mass and volume of substantially untreated solid waste. Solid waste can be treated with an enzymatic digestive process in the presence of an enzymatic solution (including pectate lyases of the invention) at a controlled temperature. This results in a reaction without appreciable bacterial fermentation from added microorganisms. The solid waste is converted into a liquefied waste and any residual solid waste. The resulting liquefied waste can be separated from said any residual solidified waste. See e.g., U.S. Patent No. 5,709,796.

Oral care products

[0410] The invention provides oral care product comprising pectate lyases of the invention. Exemplary oral care products include toothpastes, dental creams, gels or tooth powders, odontics, mouth washes, pre- or post brushing rinse formulations, chewing gums, lozenges, or candy. See, e.g., U.S. Patent No. 6,264,925.

Brewing and fermenting

[0411] The invention provides methods of brewing (e.g., fermenting) beer comprising pectate lyases of the invention. In one exemplary process, starch-containing raw materials are disintegrated and processed to form a malt. A pectate lyase of the invention is used at any point in the fermentation process. For example, pectate lyases of the invention can be used in the processing of barley malt. The major raw material of beer brewing is barley malt. This can be a three stage process. First, the barley grain can be steeped to increase water content, e.g., to around about 40%. Second, the grain can be germinated by incubation at 15 to 25°C for 3 to 6 days when enzyme synthesis is stimulated under the control of gibberellins. In one aspect, pectate lyases of the invention are added at this (or any other) stage of the process. The action of pectate lyases results in an increase in fermentable reducing sugars. This can be expressed as the diastatic power, DP, which can rise from around 80 to 190 in 5 days at 12°C. Pectate lyases of the invention can be used in any beer or alcoholic beverage producing process, as described, e.g., in U.S. Patent No. 5,762,991; 5,536,650; 5,405,624; 5,021,246; 4,788,066.

Other industrial applications

[0412] The invention also includes a method of increasing the flow of production fluids from a subterranean formation by

removing a viscous, pectin-containing, damaging fluid formed during production operations and found within the subterranean formation which surrounds a completed well bore comprising allowing production fluids to flow from the well bore; reducing the flow of production fluids from the formation below expected flow rates; formulating an enzyme treatment by blending together an aqueous fluid and a polypeptide of the invention; pumping the enzyme treatment to a desired location within the well bore; allowing the enzyme treatment to degrade the viscous, pectin-containing, damaging fluid, whereby the fluid can be removed from the subterranean formation to the well surface; and wherein the enzyme treatment is effective to attack the pectin in cell walls.

[0413] The invention will be further described with reference to the following examples; however, it is to be understood that the invention is not limited to such examples.

EXAMPLES

EXAMPLE 1: Pectate lyase activity assays

[0414] The following example describes exemplary pectate lyase activity assays to determine the catalytic activity of a pectate lyase. These exemplary assays can be used to determine if a polypeptide is within the scope of the invention.

APSU unit viscosity assay

[0415] APSU units: The APSU unit assay is a viscosity measurement using the substrate polygalacturonic acid with no added calcium.

[0416] The substrate 5% polygalacturonic acid sodium salt (Sigma P-1879) is solubilized in 0.1 M glycine buffer pH 10. The 4 ml substrate is preincubated for 5 min at 40°C. The enzyme is added (in a volume of 250 µl) and mixed for 10 sec on a mixer at maximum speed, it is then incubated for 20 min at 40°C. For a standard curve double determination of a dilution of enzyme concentration in the range of 5 APSU/ml to above 100 APSU/ml with minimum of 4 concentrations between 10 and 60 APSU per ml. The viscosity can be measured using a MVI600™ (Sofraser, Villemendeur, France). The viscosity can be measured as mV after 10 sec. The GRAFPAD PRISM™ Prism program, using a non linear fit with a one phase exponential decay with a plateau, can be used for calculations. The plateau plus span is the mV obtained without enzyme. See, e.g., U.S. Patent No. 6,368,843.

Beta-elimination assay

[0417] An exemplary lyase assay (at 235 nm) for the determination of the beta-elimination activity measures increases in absorbance at 235 nm. The substrate 0.1% polygalacturonic acid sodium salt (Sigma P-1879) is solubilized in 0.1 M Glycine buffer pH 10. For calculation of the catalytic rate an increase of 5.2 absorbency at 235 units per min corresponds to formation of 1 µmol of unsaturated product (see, e.g., Nasuna (1966) J. Biol. Chem. 241:5298-5306; Bartling (1995) Microbiology 141:873-881). Steady state condition is measured using a 0.5 ml cuvette with a 1 cm light path on a HP diode array spectrophotometer in a temperature controlled cuvette holder with continuous measurement of the absorbency at 235 nm. For steady state a linear increase for at least 200 sec can be used for calculation of the rate. It is used for converting pmol per min product. See, e.g., U.S. Patent No. 6,368,843.

Agar Assay

[0418] Pectate lyase activity can be measured by an agar assay. A test solution is applied to 4 mm holes punched out in agar plates (e.g., LB agar), containing 0.7% w/v sodium polygalacturonate (Sigma P 1879). The plates are then incubated for 6 h at a particular temperature (e.g., 75°C.). The plates are then soaked in either (i) 1M CaCl₂ for 0.5 h or (ii) 1% mixed alkyl trimethylammonium Br (MTAB, Sigma M-7635) for 1 h. Both of these procedures cause the precipitation of polygalacturonate within the agar. Pectate lyase activity can be detected by the appearance of clear zones within a background of precipitated polygalacturonate. Sensitivity of the assay is calibrated using dilution of a standard preparation of pectate lyase.

[0419] Endpoint Analysis--Trans-elimination at 235 nm for Pectate Lyases (High Calcium Method: 1 mM Calcium in the Final

Incubation Mixture). In this method, the substrate and enzyme is incubated for 20 min at 37°C followed by measurement at 235 nm of the formation of double bounds. Finally, the rate of the degradation is calculated based on the molar extinction coefficient in terms of Trans Units.

[0420] Procedure: Mixing of 0.5 ml enzyme dilution with 0.5 ml substrate solution. Substrate: Polygalacturonic acid from Sigma P-1879 lot 77H3784. Buffer 2x 0.1M Glycine pH 10+, 2.0 mmol CaCl₂, Stop reagent: 0.02 M H₃ PO₄, Temperature of incubation 37°C, Reaction time 20 min. Extinction coefficient of the trans-elimination 0.0052 $\mu\text{mol cm}^{-1}$. Enzyme diluted in ion-free water to 0.5 to 5 APSU per ml. Main value in duplicate 0.5 ml. The 2% w/v substrate in 2x.buffer is mixed with 0.5 ml diluted enzyme. Both pre-incubated 5 min on water bath at 37°C. Incubate for 20 min. Stop using 5 ml stop reagent and mix. Blank mix enzyme and stop reagent first and then add substrate all in the same volume.

Enzyme	0.5 ml
Substrate	0.5 ml
Stop	5 ml
Total volume	6 ml

Measure the absorbency at 235 nm in a 1 cm cuvette. Calculate the formation of trans-elimination per min using the extinction coefficient of 0.0052 $\mu\text{mol cm}^{-1}$. See, e.g., U.S. Patent No. 6,368,843.

EXAMPLE 2: Cotton Bio-Scouring Application Assay

[0421] The following example describes an exemplary Cotton Scouring Application Assay using the pectate lyase enzymes of the invention. Use of the pectate lyases of the invention to hydrolyze primary cell wall pectin ("bioscouring") can eliminate the need for caustics and high temperatures in cotton fiber scouring.

Materials/ Preparation:

[0422]

- Requires 50 mM Sodium-Bicarbonate buffer at optimum pH
- 1:10 dilution of Calloway 1663 surfactant
- 50 mM Phosphate buffer pH 6
Mix 43.3 mL of 1.0 M Na-P monobasic, 6.6 mL of 1.0 M Na-P dibasic, adjust volume to 1 L with D.I. water. Adjust pH to 6
- Ruthenium Red (R-2751 SIGMA)
Add 0.5g of Ruthenium Red to the 1L Phosphate Buffer producing a final concentration of 0.05%.
- NaOAc (5g/L pH 5)
- Cotton fabric 400R (Testfabrics Inc.) which is desized prior scouring

Scouring Procedure:

[0423]

1. Place 1.0 g of desized cotton fabric (into each Labomat beaker).
2. Each experiment should use a blank, untreated cotton (no enzyme added).
3. Add 50 mL of 50 mM Sodium-Bicarbonate buffer at pH 8.5-9 to each beaker and 2.5 mL of 1:10 dilution of Calloway Surfactant 1663.
4. Tighten the lids using an Allen wrench and install the beakers into the Labomat. making sure that the beakers are distributed evenly on the rotary rack. Connect beaker 1 with the temperature detecting cable to the connector in the middle of the rack.
5. Ramp up the heat to the desired temperature and hold for 10 minutes.
6. Add 50-200 μL of enzyme (e.g., a pectate lyase of the invention) at a concentration previously diluted to 0.1 $\mu\text{g}/\mu\text{L}$ through septum in the beaker using a syringe. Total enzyme concentration used to scour 1 gram cotton fabric can be between 5-20 μg .

7. 7. Run the reaction in the Labomat at temperature for 15 minutes.
8. 8. Rinse the cotton fabric twice by pouring the cotton fabric into the hand and squeezing the cotton dry, place the cotton back into the beaker and filling the beaker with D.I. water and repeating this step again, finish with squeezing the excess water out of the cotton.
9. 9. Soak the cotton fabric in NaOAc (5g/L pH 5) for 2 minutes.
10. 10. Repeat the 2X rinse cycle in step 8.
11. 11. Place the cotton fabric on weigh-boats and allow the fabric to dry overnight in the laminar flow biohoods.

Dyeing Procedure:

[0424]

1. 1. Place the treated cotton fabric in the Labomat beakers.
2. 2. Add 100 mL of 0.05% Ruthenium Red, Na-P buffer pH 6 to each beaker.
3. 3. Tighten the lids using an Allen wrench and install the beakers into the Labomat making sure that the beakers are distributed evenly on the rotary rack. Connect beaker 1 with the temperature detecting cable to the connector in the middle of the rack.
4. 4. Ramp up the heat to 50°C and hold for 30 minutes.
5. 5. Rinse the fabric twice by pouring the cotton into the hand and squeezing the cotton dry, place the fabric back into the beaker and filling the beaker with D.I. water and repeating this step again, finish with squeezing the excess water out of the fabric.
6. 6. Place the dyed fabric into the beaker and add 100 mL of D.I. water.
7. 7. Tighten the lids using Allen wrench and install the beakers into the Labomat making sure that the beakers are distributed evenly on the rotary rack.
8. 8. Ramp up the heat to 100°C and hold for 10 minutes; cool the beakers down to 60°C.
9. 9. Repeat the 2Xrinse cycle in step 5.
10. 10. Place the dyed fabric on weigh-boats and allow the fabric to dry overnight in the laminar flow biohoods.

Enzyme Scouring Quantification:

[0425]

1. 1. Calibrate the GretagMacbeth Color Eye 7000A by selecting the Color Eye Icon on the desk top of the computer.
2. 2. Place the black lens over the orifice and hit enter when the program request the calibration of the negative thresh hold.
3. 3. Place the white filter over the orifice and hit enter when the program request the calibration of the white balance.
4. 4. Place the dry dyed fabric over the orifice and push F4 to read the fabric whiteness.
5. 5. Record the L* number, turn the fabric over to read the other side and record the L* number. Compute the average L* number for each sample.
6. 6. Graph the delta L for each cotton scoured sample by subtracting the samples L* number with the untreated fabric L*.

EXAMPLE 3: A single-bath process for desizing and scouring

[0426] The following example describes an exemplary single-bath process for desizing and scouring. The invention provides methods and compositions for desizing, scouring and bleaching of cellulosic materials by contacting the cellulosic materials simultaneously or sequentially in a single-bath process with an enzyme system comprising a pectate lyase of the invention. The single-bath process can further comprise a bleaching system comprising hydrogen peroxide or at least one peroxy compound which generates hydrogen peroxide when dissolved in water, or combinations thereof, and at least one bleach activator.

[0427] Cellulosic materials including crude fibers, yarn, or woven or knit textiles, made of cotton, linen, flax, ramie, rayon, hemp,

jute, or blends of these fibers with each other or with other natural or synthetic fibers, can be treated by this single-bath process of the invention. In one aspect, a fabric weighing is loaded into a container, which is subsequently filled with a buffer solution (e.g., 20 mM Na phosphate buffer, pH 9.2) comprising a pectate lyase of the invention (e.g., 3000 APSU/kg-fiber of pectate lyase), wetting agent (e.g., 0.5 g/L), H₂O₂ (e.g., 1.7 g/L) and stabilizer (e.g., 0.75 g/L). The fabric can be treated, e.g., at 55°C for about 15 min, after which temperature was raised at 5°C/min to 70°C for 1 h. The fabric is then washed thoroughly with water to remove the residual chemicals and dried at room temperature overnight.

EXAMPLE 4: Assay for detecting thermotolerant enzymes

[0428] The following example describes an exemplary assay for detecting thermotolerant enzymes that can be used to determine if an enzyme is within the scope of the invention. This example describes an absorbance based screening ("discovery") assay for detecting thermotolerant enzymes, which, in one aspect, can be characterized as "upmutants" from a "parental" pectate lyase gene. This exemplary protocol can be used for variants, or mutants, generated by either the GSSM™ or combinatorial methods.

Materials and preparations

[0429]

1. a. Polygalacturonic Acid (PGA), and 2% [Sigma P-3889]
2. b. UV (friendly) 96 Well Flat Bottom Plates [Thomson Instrument 931801B]
3. c. COSTAR 96 Well Plates
4. d. Adhesive PCR Foil Seals [Marsh AB-0626]
5. e. B-PER [PIERCE 78248]
6. f. LBamp100 or LBarb100
7. g. TRIS pH 8.0 (250 mM, 10X)
8. h. Glycine (250 mM, 10X)
9. i. 0.2% Polygalacturonic Acid Substrate for enzyme activity detection: 100 mL of each of the following: 10X Tris, 10X Glycine, and 2% PGA, plus 700 mL of holy water.
10. j. Plates: aliquot 200 µL of medium, LBamp100 or LBarb100, into the wells of both the COSTAR and the UV friendly 96 well flat bottom plates

Colony Picking and plate replication

[0430] GSSM™ or combinatorial mutant clones colonies were picked with an Autogen (Framingham, MA) colony picker and the cells were inoculated into Lamp100 medium. A total of 168 GSSM™ clones were screened per residue site or 13,000 clones from the combinatorial library 2328 were screened. The mutated clones were picked into rows A, B, C, E, F, G, and H of the 96 well plate. Wild Type (wt) clone (SEQ ID NO:132, encoded by SEQ ID NO:131) were picked into row D as a control. After completing the colony picking, the plates were incubated overnight (approximately 18 hrs) at 37°C, shaking at 150 RPM. These plates will be referred to as the master plates from now on.

[0431] Copies of each master plate were made into UV friendly plates (now called "assay plates") using the automated plate replicator. Once replication was complete, the assay plates were placed in a humidified 30°C incubator overnight.

Primary Assay

[0432] Cell densities in each well of all plates were determined at OD₆₀₀ using a SPECTRAMAX™ (Molecular Devices Corporation, Sunnyvale CA) system. All assay plates were then sealed with PCR Foil Seals and then spun at 2200 rpm in an Eppendorf centrifuge for 10 minutes. Using the PowerWasher system, the supernatant was then aspirated out of the assay plates leaving only the cells behind. 20 µL of B-PER™ (Pierce Biotechnology, Rockford, IL) was then added to each well and the assay plates were resealed. The plates were then placed on a plate shaker for 10 minutes in order to ensure proper cell lyses. The plates were then placed in an incubator preheated to 50°C for 50 minutes for the GSSM™ assay or 70°C for 25 minutes for the

combinatorial up-mutant assay. The assay plates were then removed from the incubator after the proper heat challenge time and quickly cooled to room temperature. The SPECTRAMAX™ was used to read kinetics at wavelength 235 nm over a 2 minute period. Any putative hit that performed better than wild type was broken out for a secondary assay. Figure 8 illustrates a residue with multiple positive hits. In Figure 8, Row D contains the residual activity of the wild type (wt), SEQ ID NO: 132, and rows A, B, C, E, F, G, H are the GSSM™ clones of mutation site 182.

Secondary Assay

[0433] All wells that showed an improved enzymatic rate compared to the wild type performance were identified and the clones from the respective master plate were broken out. Using aseptic techniques, a sterile toothpick was used in the well of a putative hit from the master plate. Cells adhering to the toothpick were transferred to a new plate selecting a new well with 200 μ L LBamp100. Also, in the same manner, row D was filled with WT for each break out plate. The secondary master plates were placed in the 30°C humidified incubator overnight. The secondary master plates were then pin tooled into UV friendly 96 well plates. The secondary assay plates were then placed in a 30°C humidified incubator overnight. Cell densities in each well of all plates were determined at OD₆₀₀ using the SpectraMax systems. All assay plates were then sealed with PCR Foil Seals and then spun at 2200 rpm in an Eppendorf Centrifuge for 10 minutes. The remaining steps of the secondary assay were the same as indicated for the primary assay. Any confirmed hits that performed better than wild type were broken out and tested again in the tertiary assay.

Tertiary Assay

[0434] 5 μ L of culture from wells that confirmed improved thermotolerance activity from the wild type clone were aliquotted onto a small LBcarb100 petri dish to make streak plates. 5 μ L of one of the control "wild type" (wt) (SEQ ID NO: 132, encoded by SEQ ID NO:131) wells was also used to make a streak plate. The streak plates were incubated at 37°C overnight. A small section of an individual colony was scraped and the cells were inoculated 5 mL of LBcarb100. The culture was allowed to grow overnight at 37°C at 200 RPM. The confirmed hit was then diluted to OD₆₀₀ = 0.2. 200 μ L of a confirmed clone was aliquotted into a well, filling an entire row on the 96 well UV friendly plate. The same was done for a wt control. All plates were then sealed and spun at 2000 rpm in the Eppendorf Centrifuge for 10 minutes. The remaining steps of the tertiary assay were the same as indicated for the primary assay. At the end, all putative hits that performed better than wild type were sent for sequencing. Glycerol stocks were also prepared.

EXAMPLE 5: Processes and formulations for enzymes of the invention

[0435] The following example describes exemplary processes (e.g., a bioscouring process) and formulations of the invention. Compositions and processes of the invention were tested using the exemplary pectate lyase having a sequence as set forth in SEQ ID NO: 134, encoded by, e.g., SEQ ID NO:133 ("SEQ ID NO: 134").

Definition of unit:

[0436] Pectate lyase activity (of SEQ ID NO:134) was routinely measured using 0.2% (w/v) polygalacturonic acid (Sigma, P3850) in 25mM TrisHCl - 25mM Glycine NaOH buffer. One unit of enzyme activity was defined as the amount of protein that produced 1 μ mol of unsaturated oligogalacturonides per minute equivalent to 1 μ mol of unsaturated digalacturonide, using molecular extinction coefficient value of 4600 M⁻¹cm⁻¹ at 235 nm for dimer.

[0437] SpectraMax instrumentation in 96-well UV plates was used.

Formulation strength:

[0438] The enzymes of the invention can be formulated in any dosage to suit a particular need; assays for determining the optimal dosage for any particular formulation are known in the art, and several are described herein. In alternative aspects,

formulations of the invention can have a low strength of between about 2000 to 4000 u/ml (where u = unit). This is comparable to the other products on the market. In one aspect, the formulation minimum is about 1000 u/ml, and, in another aspect, the formulation maximum is about 10,000 u/ml, e.g., the formulation can comprise an enzyme of the invention in an amount of between about 1000 u/ml and 10,000 u/ml.

[0439] Solubility studies with lyophilized product (SEQ ID NO: 134) resuspended in water indicated that the solubility of the enzyme can be as high as 25000 u/ml at 4°C. Therefore, in one aspect, the invention provides formulations having a level as high as about 25000 u/ml, or more. In one aspect the invention provides formulations comprising an enzyme of the invention in an amount of between about 100 u/ml and 25000 u/ml, 30000 u/ml, 35000 u/ml or 40000 u/ml, or more.

Formulation design:

[0440] The invention provides formulations comprising at least one enzyme of the invention, and, in alternative aspects, further comprising any additive(s). Formulations of the invention can be based on known additives in other, e.g., analogous, enzyme formulations. For example, formulations of the invention can comprise the additives and/or conditions set forth in Tables 3, 4, 5 and 6, below, or any variation thereof. For example, formulations of the invention can comprise glycerol, sucrose, sodium chloride, dextrin, propylene glycol, sorbitol, sodium sulphate or TRIS, or an equivalent.

[0441] In one aspect, a formulation of the invention can be a water based formulation, or, an oil-based formulation.

[0442] Two phases of formulation stability studies were conducted; these studies used the exemplary enzyme SEQ ID NO: 134:

Accelerated stability study at 37°C

[0443] Note: these are buffer based formulations.

- Screen various additives
- Test different pH values
- Formulations at approximately 2000 u/ml.
- SEQ ID NO:134 was the exemplary enzyme tested

Table 3 (SN = sample number)

SN	pH	Glycerol	Sucrose	Sodium chloride	Dextrin	Propylene glycol	sorbitol	Sodium sulphate	Effective TRIS Conc.
1	5								40mM
2	6								40mM
3	7								40mM
4	8								40mM
5	7.5	35%							20mM
6	7.5	50%							20mM
7	7.5		35%						20mM
8	7.5			20%					20mM
9	7.5				10%				20mM
10	7.5					30%			20mM
11	7.5							100mM	20mM
12	7.5						35%		20mM
13	5.5	35%							40mM
14	5.5	50%							40mM

[0444] Best performing formulations (using SEQ ID NO: 134 as an exemplary enzyme of the invention) based on physical

appearance and retention of greater than 80% activity:

Table 4

Formulation	Additive
1	pH 5.0, 40mM TRIS
3	pH 7.0, 40mM TRIS
4	pH 8.0, 40mM TRIS
6	pH 7.5, 50% glycerol
8	pH 7.5, 20% NaCl
10	pH 7.5, 30% propylene glycol
11	pH 7.5, 100mM sodium sulfate
13	pH 5.5, 35% glycerol

[0445] In alternative aspects, the formulations of the invention can be at approximately 10,000 u/ml, or in an amount of between about 100 u/ml, 200 u/ml, 300 u/ml, 400 u/ml or 500 u/ml and 10,000 u/ml, 15,000 u/ml, 20,000 u/ml, 25000 u/ml, 30000 u/ml, 35000 u/ml or 40000 u/ml, or more. In alternative aspects, the formulations of the invention can be at approximately 500 to 30,000 units/ml, 1000 to 25,000 units/ml, or, between about 1000 to 20,000 units/ml, 1000 to 15,000 units/ml, 1000 to 10000 units/ml, 1000 to 5000 units/ml, between about 2000 to 20000 units/ml, between about 2000 to 15000 units/ml, between about 2000 to 10000 units/ml, or between about 2000 to 4000 units/ml. In alternative aspects, the formulations of the invention can comprise a water-based formulation, e.g., when no buffer is feasible; any water-based buffer system can be used.

Table 5:

PECTATE LYASE FORMULATION STABILITY STUDY PHASE II					
Formulation No.	pH	Buffer	ADDITIVES		
1	pH 7.0	NA			0.1% sodium benzoate, 0.1% potassium sorbate
2	pH 7.0	NA			300 ppm proxel
3	pH 7.0	NA	sodium chloride 15%		0.1% sodium benzoate, 0.1% potassium sorbate
4	pH 7.0	NA	sodium chloride 15%		300 ppm proxel
5	pH 7.0	NA		glycerol 35%	0.1% sodium benzoate, 0.1% potassium sorbate
6	pH 7.0	NA		glycerol 35%	300 ppm proxel
7	pH 7.0	NA	sodium chloride 10%	glycerol 25%	0.1% sodium benzoate, 0.1% potassium sorbate
8	pH 7.0	NA	sodium chloride 10%	glycerol 25%	300 ppm proxel
9	pH 5.5	NA			0.1% sodium benzoate, 0.1% potassium sorbate
10	pH 5.5	NA		300 ppm proxel	
11	pH 5.5	NA	sodium chloride 15%		0.1% sodium benzoate, 0.1% potassium sorbate
12	pH 5.5	NA	sodium chloride 15%		300 ppm proxel
13	pH 5.5	NA		glycerol 35%	0.1% sodium benzoate, 0.1% potassium sorbate
14	pH 5.5	NA		glycerol 35%	300 ppm proxel
15	pH 5.5	NA	sodium chloride 10%	glycerol 25%	0.1% sodium benzoate, 0.1% potassium sorbate
16	pH 5.5	NA	sodium chloride 10%	glycerol 25%	300 ppm proxel
CONTROLS					
17	pH 7.0	TRIS		glycerol 35%	0.1% sodium benzoate, 0.1% potassium sorbate
18	pH 5.5	TRIS		glycerol 35%	0.1% sodium benzoate, 0.1% potassium sorbate
19	pH 7.0	Acetate		glycerol 35%	0.1% sodium benzoate, 0.1% potassium sorbate
20	pH 5.5	Acetate		glycerol 35%	0.1% sodium benzoate, 0.1% potassium sorbate

[0446] Additional buffers that can be used in a formulation of the invention: 20 mM MOPS, pH 7 or 25 mM MOPS, 50 mM NaCl, pH 7.5.

[0447] Best performing formulations (using SEQ ID NO: 134 as an exemplary enzyme of the invention) based on physical appearance and retention of greater than 80% activity:

Table 6	
Formulation No	Details
5	pH 7, 35% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate
6	pH 7, 35% glycerol, 300 ppm proxel
7	pH 7, 10% sodium chloride, 25% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate
8	pH 7, 10% sodium chloride, 25% glycerol, 300 ppm proxel
13	pH 5.5, 35% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate
14	pH 5.5, 35% glycerol, 300 ppm proxel
15	pH 5.5, 10% sodium chloride, 25% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate
20*	20mM acetate buffer, pH 5.5, 35% glycerol

[0448] For example, the invention provides formulations comprising at least one enzyme of the invention and comprising a buffer (formulation) of: pH 7, 35% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; pH 7, 35% glycerol, 300 ppm proxel; pH 7, 10% sodium chloride, 25% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; pH 7, 10% sodium chloride, 25% glycerol, 300 ppm proxel; pH 5.5, 35% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; pH 5.5, 35% glycerol, 300 ppm proxel; pH 5.5, 10% sodium chloride, 25% glycerol, 0.1% sodium benzoate, 0.1% potassium sorbate; or, 20mM acetate buffer, pH 5.5, 35% glycerol; 20 mM MOPS, pH 7 or 25 mM MOPS, 50 mM NaCl, pH 7.5; pH 5.0, 40mM TRIS; pH 7.0, 40mM TRIS; pH 8.0, 40mM TRIS; pH 7.5, 50% glycerol; pH 7.5, 20% NaCl; pH 7.5, 30% propylene glycol; pH 7.5, 100mM sodium sulfate; pH 5.5, 35% glycerol; or, any combination thereof, or, with equivalents thereof.

Exemplary Bioscouring Application

[0449]

- In one aspect, pH is pH 8.5 (bicarbonate buffer)
- Non-ionic wetting agent (1 g/L) [e.g.: Apollowet NFW]
- Liquor ratio in the enzyme bath: 10:1 to 50:1 (L liquor:kg fabric)
- Enzyme dose: 0.137 ml of the concentrated extract per kg of fabric
- Temperature range: between about 50°C to 70°C
- Treatment time about 20 min
- Chelants should be excluded from the enzyme bath, and should only be added after 20 minutes of enzyme treatment and retained for 10 minutes before discharging bath

[0450] Thus, in the invention provides a bioscouring process using at least one enzyme of the invention comprising at least one, several or all of the following steps/ limitations: pH is pH 8.5, in bicarbonate buffer, comprising a non-ionic wetting agent (at, e.g., 1 g/L), where the liquor ratio in the enzyme bath is between about 10:1 to 50:1 (L liquor:kg fabric), where the enzyme dose is between about 0.1 and 0.2 ml, e.g., at about 0.137 ml of the concentrated extract per kg of fabric, at a temperature range: between about 50°C to 70°C; with a treatment time about 20 min; and, in one aspect, comprising chelants, which should be excluded from the enzyme bath and should only be added after 20 minutes of enzyme treatment and retained for 10 minutes before discharging bath.

[0451] The enzyme SEQ ID NO: 134 performed well in the range of 5 to 25 grams of pure enzyme per ton of treated fabric.

SEQUENCE LISTING

[0452]

<110> Diversa Corporation
Kerovuo, Janne
Solbak, Arne

Gray, Kevin
 McCann, Ryan
 Purohit, Shalaka
 Gerendash, Joel
 Janssen, Giselle
 Dahod, Samun

<120> PECTATE LYASES, NUCLEIC ACIDS ENCODING THEM AND METHODS FOR MAKING AND USING THEM

<130> 564462009640

<140> To Be Assigned

<141> Concurrently herewith

<150> 60/460,842 <151> 2003-04-04

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<212> DNA

<213> Unknown

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gtgcgtaaga	agaacgattc	cacgatcgac	aatggcgcca	cttacacgca	actctcgttt	1200
ctggcgcggg	tttacacggc	gcaaaagcag	gagcggcctc	gcgagtcgtt	tctgaaggga	1260
ctcgattacc	tgttgaaggc	gcagtatcca	aacggaggct	ggccgcagtt	ctatcccaac	1320
ctcaacggct	attacaaca	catcactttc	aacgacaacg	ccatgatcgg	cgtgatgaaa	1380
ctgctgcgcg	acgtagcgac	agcgaaaacc	gcgtatgcgt	tcgtcgacga	agcaccgacg	1440
acgagtgccg	cgaaggcggg	cgaaaaagga	atcgagtcca	tactgaagac	gcaggtggtt	1500
gtgaatggcc	ggcgcaccgt	gtggtgtgcy	caacatgacg	aagtcaacgt	cgcgcctgcc	1560
ccggcgagga	cgtttgaatt	agtgttcgct	agtggtgtgt	aaagcgttga	gatcgtgcgc	1620
tttttgatgt	cgatcaagaa	cccgtcgccg	gcggttgtcg	aggcgatcga	gtcggcgggt	1680
gcgtggttcg	agcaatcgca	agtgaagat	cccgcggcca	aacctgcgtg	ggcgcgattt	1740
tatgagatcg	gcactaatcg	tccgatcttc	gccgggcgtg	acggcgctcg	taagtatgat	1800
gtgaaacaga	tcgatgagga	acgacgaaag	aattacgcat	ggtacgttga	cgacgcagcg	1860
aaactactga	aaaccgacta	tcttgagtgg	aaagaaaaga	acgccaaaga	tcaatga	1917

<210> 2

<211> 638

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(21)

<221> DOMAIN
 <222> (28)...(308)
 <223> Pectin methyl esterase domain

<221> DOMAIN
 <222> (309)...(638)
 <223> Catalytic domain

<400> 2

```
Met Ser Leu Phe Arg Lys Leu Ala Leu Leu Val Leu Cys Gly Leu Leu
 1      5      10      15
Leu Ser Val Gly Ala Glu Thr Arg Ala Ser Lys Arg Ile Val Val Ala
 20      25      30
Ala Asp Gly Ser Gly Asp Val Arg Thr Ile Gln Gln Ala Val Asp Gln
 35      40      45
Val Pro Lys Asp Asn Thr His Pro Val Leu Ile Gln Ile Lys Pro Gly
 50      55      60
Val Tyr Gln Glu Gln Val Arg Val Ala Ala Gly Lys Arg Phe Ile Thr
 65      70      75
Phe Arg Gly Asp Asp Ala Ser Lys Thr Val Ile Thr Tyr Arg Leu Ser
 85      90      95
Ala Leu Gln Ala Gly Asn Thr Arg Leu Ala Phe Thr Thr Phe Val Asn
100      105      110
Ala Asp Asp Phe Arg Ala Glu Asn Leu Thr Phe Glu Asn Ser Phe Gly
115      120      125
Thr Gly Ser Gln Ala Val Ala Leu Phe Val Asp Ala Asp Arg Ala Thr
130      135      140
Phe Glu Asn Cys Arg Phe Leu Gly Trp Gln Asp Thr Leu Phe Val Asn
145      150      155
Gly Ser Arg His Phe Phe Lys Asp Cys Tyr Val Glu Gly His Val Asp
165      170      175
Phe Ile Phe Gly Thr Ala Ser Ala Val Phe Glu Asn Cys Thr Ile His
180      185      190
Ser Lys Gly Glu Gly Tyr Val Thr Ala His Tyr Arg Thr Ser Asp Glu
```

```
195      200      205
Met Asp Thr Gly Phe Val Phe His Arg Cys Arg Leu Thr Gly Arg Asp
210      215      220
Thr Gly Arg Gly Val Tyr Leu Gly Arg Pro Trp Arg Pro Tyr Ala Arg
225      230      235
Val Val Phe Ile Asp Cys Trp Leu Asp Ala His Ile Arg Pro Glu Gly
245      250      255
Trp Asp Asn Trp Arg Asp Pro Glu Arg Glu Lys Thr Ala Trp Phe Ala
260      265      270
Glu Tyr Lys Ser Lys Gly Pro Gly Ala Asn Pro Val Ala Arg Val Ala
275      280      285
Trp Ser Arg Gln Leu Thr Thr Glu Gln Ala Ala Glu Phe Ser Arg Glu
290      295      300
Arg Phe Phe Ser Arg Ala Val Arg Gly Leu Ser Gly Gln Ala Asn Gln
305      310      315
Ala Val Gly Thr Ile Ala Trp Asp Asp Ala Gln Lys Lys Pro Asn Glu
325      330      335
Trp Tyr Ala Ser Ala Glu Ala Leu Arg Ile Ala Asp Asn Val Val Leu
340      345      350
Tyr Gln Arg Asp Ser Gly Gly Trp Pro Lys Asn Ile Asp Met Gly Lys
355      360      365
Pro Leu Asp Glu Lys Gly Arg Ala Gly Leu Leu Arg Val Arg Lys Lys
370      375      380
Asn Asp Ser Thr Ile Asp Asn Gly Ala Thr Tyr Thr Gln Leu Ser Phe
385      390      395
Leu Ala Arg Val Tyr Thr Ala Gln Lys Gln Glu Arg His Arg Glu Ser
405      410      415
Phe Leu Lys Gly Leu Asp Tyr Leu Leu Lys Ala Gln Tyr Pro Asn Gly
420      425      430
Gly Trp Pro Gln Phe Tyr Pro Asn Leu Asn Gly Tyr Tyr Lys His Ile
435      440      445
Thr Phe Asn Asp Asn Ala Met Ile Gly Val Met Lys Leu Leu Arg Asp
450      455      460
Val Ala Thr Ala Lys Pro Ala Tyr Ala Phe Val Asp Glu Ala Arg Arg
465      470      475
Thr Ser Ala Ala Lys Ala Val Glu Lys Gly Ile Glu Cys Ile Leu Lys
485      490      495
Thr Gln Val Val Val Asn Gly Arg Arg Thr Val Trp Cys Ala Gln His
500      505      510
Asp Glu Val Thr Leu Ala Pro Ala Pro Ala Arg Thr Phe Glu Leu Val
515      520      525
Ser Leu Ser Gly Gly Glu Ser Val Glu Ile Val Arg Phe Leu Met Ser
530      535      540
Ile Lys Asn Pro Ser Pro Ala Val Val Glu Ala Ile Glu Ser Ala Val
545      550      555
Ala Trp Phe Glu Gln Ser Gln Val Lys Asp Pro Ala Gly Lys Pro Ala
565      570      575
Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg Pro Ile Phe Ala Gly
580      585      590
Arg Asp Gly Val Val Lys Tyr Asp Val Lys Gln Ile Asp Glu Glu Arg
595      600      605
Arg Lys Asn Tyr Ala Trp Tyr Val Asp Asp Ala Ala Lys Leu Leu Lys
610      615      620
Thr Asp Tyr Pro Glu Trp Lys Glu Lys Asn Ala Lys Asp Gln
625      630      635
```

<210> 3

<211> 1416

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 3

```

atgtcgtcac gacgcgagtt cattagagat ctgttgactg gccggcgcaact gatcgccgtc 60
gcgcgcgcgtc tgtctgcgtt tgcagcggag gagaatccgt gggaacggt gatgccttcg 120
atcgtgaaac gcataaagcg acctcgtttc ccgatgcgca cgtttgatct caccgagttt 180
ggagcgaag gtgatggacg aacagattgc acgttggtt tccgtcgcgc gatcgatcga 240
tgcacgaacg ccggtggtgg gagagtagtt gttccaccgg gtctgtatct cactggcgcc 300
attcatttga agagcaacgt cgaccttcatt atctcagaag gtactacggt caagttcagc 360
cagaacccga aagactacct gcccgttgtt ttctcgcgtt gggaaggcgt cgaggtgttc 420
aactactcgc cttttatcta cgccttcgaa caaacgaaca ttgcgatcac tggcaaggcg 480
acgctcaacg gtcaaaacga caacgaacac tggtgccctt ggaacggacg tgcgcgtac 540
ggctggaaag aagggatgag caatcagcgt cccgatcgaa atgcgctgtt tgcgatggcc 600
gaaaaaggtg tcccggttca ggagcgcat tttggtgagg gccattactt aaggccgcag 660
ttcattcaac cttatcgttg tgagaacgtg ctgatcgaag gtgtcactat tcgaaactcg 720
ccgatgtggg aaattcatcc ggtgctctgc cggaatgtca tcgtccaaaa tgtgatcatc 780
aacagtcatt gtccaaacaa cgacgggtgt aatcctgagt cgtgcacgga tgtgttgatt 840
aaggattgtg acttcgacac tggtgacgat tgtatcgga tcaagtcagg ccgaaatgca 900
gatgggcggc gactgaaggc tctactgaa aacattatcg tgactggttg tcgcatgaaa 960
gatggtcacg gccggattac ggtgggcagc gagatttcgg gtggggtgcg aaatcttttc 1020
gcataccaact gccgggtcga cagtcggaac ctggaccatg cattgcgggt taagaataac 1080
gctatcgctg gccgggtgtt ggagaatctg cacttccgaa atatcgacgt cgggcaagt 1140
gcgcacgcgg tgatcacgat cgatttcaat tatgaggaa gcgcgaagg atcgttcacg 1200
ccagtcgttc gtgattacac cgtcgatggc ctctgcagca cgaagaagta gtacgcgtc 1260
gatgtgcagg gcttggcgac ggcccgatc gtgaatctgc gtctaacc aa ctgcatcttc 1320
gacaatgtcg ctgaaggaaa tgtgtgaag aacgtaaa atgcaactat cgagaatgtc 1380
aaaatcaatg gaaaaagcgt tgatgcagt cgttag 1416

```

<210> 4

<211> 471

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(28)

<221> DOMAIN

<222> (81)...(476)

<223> Catalytic domain

<400> 4

```

Met Ser Ser Arg Glu Phe Ile Arg Asp Leu Leu Thr Gly Gly Ala
1      5      10      15
Leu Ile Ala Val Ala Pro Arg Leu Ser Ala Phe Ala Ala Glu Glu Asn
20     25     30
Pro Trp Glu Thr Val Met Pro Ser Ile Val Lys Arg Ile Lys Arg Pro
35     40     45
Arg Phe Pro Met Arg Thr Phe Asp Leu Thr Glu Phe Gly Ala Lys Gly
50     55     60
Asp Gly Arg Thr Asp Cys Thr Leu Ala Phe Arg Arg Ala Ile Asp Arg
65     70     75     80
Cys Thr Asn Ala Gly Gly Gly Arg Val Val Pro Pro Gly Ser Tyr
85     90     95

```

Leu Thr Gly Ala Ile His Leu Lys Ser Asn Val Asp Leu His Ile Ser
 100 105 110
 Glu Gly Thr Thr Val Lys Phe Ser Gln Asn Pro Lys Asp Tyr Leu Pro
 115 120 125
 Val Val Phe Ser Arg Trp Glu Gly Val Glu Val Phe Asn Tyr Ser Pro
 130 135 140
 Phe Ile Tyr Ala Phe Glu Gln Thr Asn Ile Ala Ile Thr Gly Lys Gly
 145 150 155 160
 Thr Leu Asn Gly Gln Ser Asp Asn Glu His Trp Trp Pro Trp Asn Gly
 165 170 175
 Arg Ala Ala Tyr Gly Trp Lys Glu Gly Met Ser Asn Gln Arg Pro Asp
 180 185 190
 Arg Asn Ala Leu Phe Ala Met Ala Glu Lys Gly Val Pro Val Gln Glu
 195 200 205
 Arg Ile Phe Gly Glu Gly His Tyr Leu Arg Pro Gln Phe Ile Gln Pro
 210 215 220
 Tyr Arg Cys Glu Asn Val Leu Ile Glu Gly Val Thr Ile Arg Asn Ser
 225 230 235 240
 Pro Met Trp Glu Ile His Pro Val Leu Cys Arg Asn Val Ile Val Gln
 245 250 255
 Asn Val Ile Ile Asn Ser His Gly Pro Asn Asn Asp Gly Cys Asn Pro
 260 265 270
 Glu Ser Cys Thr Asp Val Leu Ile Lys Asp Cys Asp Phe Asp Thr Gly
 275 280 285
 Asp Asp Cys Ile Ala Ile Lys Ser Gly Arg Asn Ala Asp Gly Arg Arg
 290 295 300
 Leu Lys Ala Pro Thr Glu Asn Ile Ile Val Thr Gly Cys Arg Met Lys
 305 310 315 320
 Asp Gly His Gly Gly Ile Thr Val Gly Ser Glu Ile Ser Gly Gly Val
 325 330 335
 Arg Asn Leu Phe Ala Ser Asn Cys Arg Leu Asp Ser Pro Asn Leu Asp
 340 345 350
 His Ala Leu Arg Val Lys Asn Asn Ala Met Arg Gly Gly Leu Leu Glu
 355 360 365
 Asn Leu His Phe Arg Asn Ile Asp Val Gly Gln Val Ala His Ala Val
 370 375 380
 Ile Thr Ile Asp Phe Asn Tyr Glu Glu Gly Ala Lys Gly Ser Phe Thr
 385 390 395 400
 Pro Val Val Arg Asp Tyr Thr Val Asp Gly Leu Arg Ser Thr Lys Ser
 405 410 415
 Lys Tyr Ala Leu Asp Val Gln Gly Leu Ala Thr Ala Pro Ile Val Asn
 420 425 430
 Leu Arg Leu Thr Asn Cys Ile Phe Asp Asn Val Ala Glu Gly Asn Val
 435 440 445
 Val Lys Asn Val Lys Asp Ala Thr Ile Glu Asn Val Lys Ile Asn Gly
 450 455 460
 Lys Ser Val Asp Ala Val Pro
 465 470

<210> 5

<211> 1077

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 5

ttggacgaca agtgggctga gcggacatcg cccgatttca atctcgtctc gtggaatgaa 60

attctaaagc agccgaaact ttggtacgcg gtcgacgaag cgacgaggat cgcaaatcag 120
 gtgatccctt atcaacgcga caacggtggt tggccgaaga atatcgacat ggccgcatg 180
 ctcatgcagc cagaacgcga aaaacttagt cgcgagaaga gcgagaccga cagcacaatc 240
 gacaacggcg cgacgacaac ccagctcgcg tatctggcga aggtcatcac ggccaagaat 300
 atcgaaagcc atcgctcgcg gtttttcaaa ggctctgatt ttcttttcgc catgcagtac 360
 gggaaatggcg gcttcccgca atttttccct ctgctgacg attattcgcg cgagattacg 420
 ttcaacgaca acgcatgatg aaatgtgctt cgtttgctcc gcgacatagc cgatcgaaag 480
 aacgattatg tgtttgtcga tgaagagcgg cgagcgaagg ccgagcaggc tgtaaggcgt 540
 gcgaccccggt tgatccctcag cagcagggtc gtcgtcgatg gaaagaaaac cgtctgggct 600
 gcgcagtatg atgagaagac attgaagccg gccgcggcgc gaaagttcga gccggcatca 660
 ttgaccgcgg gcgagagcgt tggcatcgtc cgttttttga tgctagaaaa accaacaccc 720
 gagatcatta acgcatcgca atccgccatc gcttggtaca aggcgaacaa catctcgga 780
 cttaggtggg agaggcgaaa cggcgagaaac atttgatca aagacaagaa cgcgcgccc 840
 gtcctgggccc gcttttatca gatcgaaacg atgaggccga tcttcgccgg tcgcgatcgc 900
 gtcacgatg acgatgtgat gcagatcgag tcggaacgct gaaacggata tgcattgtac 960
 gtatccgaac cgaatgagtt gttgaatgaa gattatccga agtgaggac aaggagtgcg 1020
 aagcgtgccc agatctttca acgtccgcct cttggttcga gatttcggac cgtgtag 1077

<210> 6

<211> 358

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN
 <222> (1)...(358)
 <223> Catalytic domain

<400> 6

```

Met Asp Asp Lys Trp Ala Glu Arg Thr Ser Pro Asp Phe Asn Leu Val
 1          5          10          15
Ser Trp Asn Glu Ile Leu Lys Gln Pro Lys Leu Trp Tyr Ala Val Asp
 20          25          30
Glu Ala Thr Arg Ile Ala Asn Gln Val Ile Leu Tyr Gln Arg Asp Asn
 35          40          45
Gly Gly Trp Pro Lys Asn Ile Asp Met Ala Ala Met Leu Met Gln Ala
 50          55          60
Glu Arg Glu Lys Leu Ser Arg Glu Lys Ser Glu Thr Asp Thr Thr Ile
 65          70          75          80
Asp Asn Gly Ala Thr Thr Thr Gln Leu Ala Tyr Leu Ala Lys Val Ile
 85          90          95
Thr Ala Lys Asn Ile Glu Ser His Arg Val Ala Phe Phe Lys Gly Leu
100          105          110
Asp Phe Leu Phe Ala Met Gln Tyr Gly Asn Gly Gly Phe Pro Gln Phe
115          120          125
Phe Pro Leu Arg Asp Asp Tyr Ser Arg Glu Ile Thr Phe Asn Asp Asn
130          135          140
Ala Met Ile Asn Val Leu Arg Leu Leu Arg Asp Ile Ala Asp Arg Lys
145          150          155          160
Asn Asp Tyr Val Phe Val Asp Glu Glu Arg Arg Ala Lys Ala Glu Gln
165          170          175
Ala Val Arg Arg Ala Ile Pro Leu Ile Leu Ser Thr Gln Val Val Val
180          185          190
Asp Gly Lys Lys Thr Val Trp Ala Ala Gln Tyr Asp Glu Lys Thr Leu
195          200          205
Lys Pro Ala Ala Ala Arg Lys Phe Glu Pro Ala Ser Leu Thr Ala Gly
210          215          220

```

```

Glu Ser Val Gly Ile Val Arg Phe Leu Met Leu Glu Lys Pro Thr Pro
225          230          235          240
Glu Ile Ile Asn Ala Ile Glu Ser Ala Ile Ala Trp Tyr Lys Ala Asn
245          250          255
Asn Ile Ser Gly Leu Arg Trp Glu Arg Arg Asn Gly Glu Asn Ile Val
260          265          270
Ile Lys Asp Lys Asn Ala Pro Val Trp Ala Arg Phe Tyr Gln Ile
275          280          285
Glu Thr Met Arg Pro Ile Phe Ala Gly Arg Asp Ala Val Ile Arg Tyr
290          295          300
Asp Val Met Gln Ile Glu Ser Glu Arg Arg Asn Gly Tyr Ala Trp Tyr
305          310          315          320
Val Ser Glu Pro Asn Glu Leu Leu Asn Glu Asp Tyr Pro Lys Trp Arg
325          330          335
Thr Arg Ser Ala Lys Arg Ala Gln Ile Phe Gln Arg Pro Pro Leu Gly
340          345          350
Ser Arg Phe Arg Thr Val
355

```

<210> 7
 <211> 1125
 <212> DNA
 <213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 7

```

gtgcatgcgg gcgcgaataa cgtgagccga tggcgcggaag agttcctgcg cgacttcgcc      60
gcgcggctct cccgaaccat tccgtcctcg ccggcgcgaga gcgctgcggt cagcgggggt      120
ccggcgcgga tccgctgggg agcggagctc ctgcggcgaga agccggagtg gtatgcctcg      180
cgagagcgga ggacgatcgc cgacagcgtc atccagtacc aggcggcgga cgcgcgctgg      240
cccaagaada cgcacacctg gactccgccc acggctgaat cacgcgcccg cgcgcgcgcc      300
gacgtgacgt cgagcaccat cgacaacaac ggcaacgaga tgccgatgca gttccttgcc      360
ctggtggcgg acgcgaccgg cgaggctcgc tatcgccgct cgttcctccg cggcttcgac      420
tacctgctcg ccgcgcagta tcccaacggc ggctggccgc agttctttcc gtcgcccgcc      480
gggtattaca cccacatcac cttcaacgac aacgcgatgg tcaactgctg gacgtgctg      540
cgcgatgccg cggcgggtca ggggccatag gccttcgtgg acgagccccg ccgcgcgaag      600
gcccgcgccc ccgtgtcccg ggggatcgac gtcactctga agaccgaagt gaaacagaac      660
ggcaagctga cggcgtggtg cggcgagcac gacgagaaga ccctcgcgcc ggcgtggggc      720
cgcgcttacg agccgcacac gctctccggc agcgaaacgg tcggcatcgt ccgcttcctg      780
atggagatcg agaagccgtc acgggagatc gtgcgcgcga tcgaaggggc cgtcgccctg      840
ctgaagtcct tggcgattcc ggggctgcgc tacgagtcct tcaccggcgc ggacggacag      900
agggaccgcg gcgtcgttcc ggatccatcg gccggactcc tgtggggcgc gttctacgag      960
ctcggcacca accggccgat cttcctcggc cgcgactccg tggttcgcgc cgcgctcagt      1020
gacatcgaac gcgagcgccg cgcgggctac gcctactacg gtacgtggcc ggcgagccct      1080
attgctgcgg actaccgcgg ctggcgcttc acgctccggc gctga      1125

```

<210> 8
 <211> 374
 <212> PRT
 <213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(374)

<223> Catalytic domain

<400> 8

```

Met His Ala Gly Ala Lys His Val Ser Arg Trp Arg Glu Glu Phe Leu
 1      5      10      15
Arg Asp Phe Ala Ala Arg Leu Ser Arg Thr Ile Pro Ser Ser Pro Ala
 20      25      30
Gln Ser Ala Ala Val Ser Gly Val Pro Ala Ala Ile Arg Trp Gly Ala
 35      40      45
Asp Val Leu Arg Gln Lys Pro Glu Trp Tyr Ala Ser Arg Glu Ala Arg
 50      55      60
Thr Ile Ala Asp Ser Val Ile Gln Tyr Gln Ala Ala Asp Gly Gly Trp
 65      70      75      80
Pro Lys Asn Thr Asp Leu Gly Thr Pro Pro Thr Ala Glu Ser Arg Ala
 85      90      95
Gly Ala Ala Ala Asp Val Thr Ser Ser Thr Ile Asp Asn Asn Gly Thr
100      105      110
Thr Met Pro Met Gln Phe Leu Ala Leu Val Ala Asp Ala Thr Gly Glu
115      120      125
Ala Arg Tyr Arg Ala Ser Phe Leu Arg Gly Phe Asp Tyr Leu Leu Ala
130      135      140
Ala Gln Tyr Pro Asn Gly Gly Trp Pro Gln Phe Phe Pro Leu Arg Arg
145      150      155      160
Gly Tyr Tyr Thr His Ile Thr Phe Asn Asp Asn Ala Met Val Asn Val
165      170      175
Leu Thr Val Leu Arg Asp Ala Ala Ala Gly Gln Ala Pro Tyr Ala Phe
180      185      190
Val Asp Glu Pro Arg Arg Ala Lys Ala Arg Ala Val Ser Arg Gly
195      200      205
Ile Asp Val Ile Leu Lys Thr Gln Val Lys Gln Asn Gly Lys Leu Thr
210      215      220
Ala Trp Cys Ala Gln His Asp Glu Lys Thr Leu Ala Pro Ala Trp Ala
225      230      235      240
Arg Ala Tyr Glu Pro Pro Ser Leu Ser Gly Ser Glu Thr Val Gly Ile
245      250      255
Val Arg Phe Leu Met Glu Ile Glu Lys Pro Ser Pro Glu Ile Val Ala
260      265      270
Ala Ile Glu Gly Ala Val Ala Trp Leu Lys Ser Val Ala Ile Pro Gly
275      280      285
Leu Arg Tyr Glu Ser Phe Thr Gly Ala Asp Gly Gln Arg Asp Arg Arg
290      295      300
Val Val Pro Asp Pro Ser Ala Gly Leu Leu Trp Ala Arg Phe Tyr Glu
305      310      315      320
Leu Gly Thr Asn Arg Pro Ile Phe Leu Gly Arg Asp Ser Val Val Arg
325      330      335
Ala Ala Leu Ser Asp Ile Glu Arg Glu Arg Arg Ala Gly Tyr Ala Tyr
340      345      350
Tyr Gly Thr Trp Pro Ala Ser Leu Ile Ala Ala Asp Tyr Pro Arg Trp
355      360      365
Arg Ser Thr Leu Arg Arg
370

```

<210> 9

<211> 1116

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 9

```

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ctgctctttg cggcagccac acagggaaac aagccgcccg aagtgcggtg gaatgagtgc 120
ctaaaccaaa aacctgcctg gtacggcagc cgggaagcgg tgcgcattgc tgacaacctg 180
ttgctttacc aacgcgacca cggcggttgg cacaagaata tcgaaatggc tgcggtcttg 240
accgaacagc aacaggcaga gttgaaagcg caaaaggcaa ccgacgattc gacgattgat 300
aacggcgcgga cctataccca ggtgatttat ctggcgcgcg tcttcaatgc gacgaagcag 360
gagcgattca aaacccggtt tctcaaagga ttcgattatc tgctcaagcg tcagtatgcg 420
aacggcggtc ggccgcagta ttaccgcgt ttgcagggtt attacaaaca catcacgttc 480
aacgatgacg cgtggttcgg cgtgcttgat cttctgcgcg atgttgcgcg cggcgattcc 540
ggttatcggt tcgtggacag cagccggcgc gcccgcgcca gccaggccgt gcaaaaagga 600
attgagtcca tcttgaaatg ccagatcgtg gtcccgggga aaaagaccgc ctggtgcgcg 660
caacacgagt aagtgcattt cggcccgcg cggcgacgca cctacgagaa aatttcgctg 720
agcggcagcg aatcggttgg cctgatccgc ttctgatgg gcattgaaca accggacgcg 780
cgtgtagttg aggcgattga gtccgcggtt gcctggctca agcaagccaa gctgaccgcg 840
atcaaatggg ttcagaagcg ggtgcttcg aagcccaatg gcttcgacgg ggtcgctggt 900
gaagatgcac aagccgggcc attgtgggcg cgttttacg agatcggtac gggccgcccg 960
atcttttccg gacgtgacgg catcgtcaaa tacagcttgg cggaaatcga acacgaacgg 1020
cgacgggctt acgggtgta cacgaatgcg cccgcgaaat tgctggaaca agattatccg 1080
gcctggcaaa tcaaacgcgg gggaagaaa aagtaa 1116

```

<210> 10

<211> 371

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(29)

<221> DOMAIN

<222> (30)...(371)

<223> Catalytic domain

<400> 10

```

Met Ile Gly Ser Met Lys Thr Ile Leu Ser Asn Leu Asn Ala Ala Leu
 1           5           10           15
Leu Ser Cys Ala Leu Leu Phe Ala Ala Thr Gln Gly Thr Lys Pro
 20           25           30
Pro Glu Val Arg Trp Asn Glu Cys Leu Asn Gln Lys Pro Ala Trp Tyr
 35           40           45
Gly Ser Pro Glu Ala Val Arg Ile Ala Asp Asn Leu Leu Leu Tyr Gln
 50           55           60
Arg Asp His Gly Gly Trp His Lys Asn Ile Glu Met Ala Ala Val Leu
 65           70           75           80
Thr Glu Gln Gln Gln Ala Glu Leu Lys Ala Gln Lys Ala Thr Asp Asp
 85           90           95
Ser Thr Ile Asp Asn Gly Ala Thr Tyr Thr Gln Val Ile Tyr Leu Ala
100          105          110
Arg Val Phe Asn Ala Thr Lys Gln Glu Arg Phe Lys Thr Ala Phe Leu
115          120          125
Lys Gly Phe Asp Tyr Leu Leu Lys Ala Gln Tyr Ala Asn Gly Gly Trp
130          135          140
Pro Gln Tyr Tyr Pro Arg Leu Gln Gly Tyr Tyr Lys His Ile Thr Phe
145          150          155          160
Asn Asp Asp Ala Met Val Gly Val Leu Asp Leu Leu Arg Asp Val Ala
165          170          175

```

```

Arg Gly Asp Ser Gly Tyr Arg Phe Val Asp Ser Asp Arg Arg Ala Arg
180          185          190
Ala Ser Gln Ala Val Gln Lys Gly Ile Glu Cys Ile Leu Lys Cys Gln
195          200          205
Ile Val Val Ala Gly Lys Lys Thr Ala Trp Cys Ala Gln His Asp Glu
210          215          220
Val Thr Phe Ala Pro Ala Pro Ala Arg Thr Tyr Glu Lys Ile Ser Leu
225          230          235          240
Ser Gly Ser Glu Ser Val Gly Leu Ile Arg Phe Leu Met Gly Ile Glu
245          250          255
Gln Pro Asp Ala Arg Val Val Glu Ala Ile Glu Ser Ala Val Ala Trp
260          265          270
Leu Lys Gln Ala Lys Leu Thr Gly Ile Lys Val Val Gln Lys Ala Asp
275          280          285
Ala Ser Lys Pro Asn Gly Phe Asp Arg Val Val Val Glu Asp Ala Gln
290          295          300
Ala Gly Pro Leu Trp Ala Arg Phe Tyr Glu Ile Gly Thr Gly Arg Pro
305          310          315          320
Ile Phe Ser Gly Arg Asp Gly Ile Val Lys Tyr Ser Leu Ala Glu Ile
325          330          335
Glu His Glu Arg Arg Thr Gly Tyr Gly Trp Tyr Thr Asn Ala Pro Ala
340          345          350
Lys Leu Leu Glu Gln Asp Tyr Pro Ala Trp Gln Ile Lys Arg Gly Gly
355          360          365
Lys Lys Lys
370

```

<210> 11

<211> 1167

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 11

```

atgtcgttgg gaccagggtgc taatccgaaa gctcgcgttc cctgggtccaa acaactatcg      60
gggtgttgagg caaagtgtgtt cgtatcgcgag cggttcttca gcctcgtgc ggaacgaacc      120
tctaagaaga atgaccagca agtcggcgcc atcgcgtgga aagatgcaca cggaaaggca      180
gatgagtggt atgcgagcgt tgaagcactt cgtatagccg ataactcgt tttctatcaa      240
cgtgactcag gtggtcgcc caagaatatt gagatggcga agacgttgag cgtatcgtag      300
aaggctcgca ttctccgcga gaagaaaaag aatgactcaa caatcgacaa tggcgcgact      360
cacactcagt tatcttttct ggccgcgctc tatacagcac aacagcagga gcgacatcgc      420
gagtcatttt taaaaggact ggattactta ctgaaggcgc agtattcaaa tgggtggtgg      480
ccacagtctt atccaaactt gaatggctac tacaacgga tcacgtacaa cgtatggcgcg      540
atgatcggtg tgatgaagct tctgcgtgat gttgcgcgag cgaacctga atacgcgttt      600
gtcgtgaaaa ctcgcgctgc gaaggctcgc aacgcgtgg aaaaaggcat cgtgtgcatt      660
ttgaaaacgc aggtggttgt tgatggcgt cgcactgttt ggtgtgcaca acacgacgaa      720
gtgacgtttg cgcgcgcgcc tgcaagaaaag tttgagttag cttcgttgag cggcggtag      780
agcgtcgata ttgttcgatt tctaattgct ataaaggatc catcgcgtaa cgtggttgaa      840
tcgattgaat cggcagttaa atggtttgag cagtccgagc taaaaggcgt taagtgggtc      900
aagaaaacgc acgctactca acctaatggg ttcgattgtg tcgttggtta agatccggag      960
agctctgttt ggccgcgctt ttacgagatt ggcacgaacc gcccgatctt tgccgggcgt      1020
gatggagtgc ctaagtatga cgtcgcgcag atcgaacacg agcgcgaac gggttacgaa      1080
tggtagcttg atgagcgagc aaaactcgtg aaaaaagatt atccggcgtg gaagaaacga      1140
catgtcgtca cgacgcgagt tcattag      1167

```

<210> 12

<211> 388

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(388)

<223> Catalytic domain

<400> 12

```

Met Ser Leu Gly Pro Gly Ala Asn Pro Lys Ala Arg Val Pro Trp Ser
  1          5          10          15
Lys Gln Leu Ser Gly Val Glu Ala Lys Leu Phe Asp Arg Glu Arg Phe
  20          25          30
Phe Ser Leu Ala Ala Glu Arg Thr Ser Lys Lys Asn Asp Gln Gln Val
  35          40          45
Gly Ala Ile Ala Trp Lys Asp Ala His Gly Lys Ala Asp Glu Trp Tyr
  50          55          60
Ala Ser Val Glu Ala Leu Arg Ile Ala Asp Asn Val Val Phe Tyr Gln
  65          70          75          80
Arg Asp Ser Gly Gly Trp Pro Lys Asn Ile Glu Met Ala Lys Thr Leu
  85          90          95
Ser Asp Arg Glu Lys Ala Ala Ile Leu Arg Glu Lys Lys Lys Asn Asp
  100          105          110
Ser Thr Ile Asp Asn Gly Ala Thr His Thr Gln Leu Ser Phe Leu Ala
  115          120          125
Arg Val Tyr Thr Ala Gln Gln Gln Glu Arg His Arg Glu Ser Phe Leu
  130          135          140
Lys Gly Leu Asp Tyr Leu Leu Lys Ala Gln Tyr Ser Asn Gly Gly Trp
  145          150          155          160
Pro Gln Phe Tyr Pro Asn Leu Asn Gly Tyr Tyr Lys Arg Ile Thr Tyr
  165          170          175
Asn Asp Gly Ala Met Ile Gly Val Met Lys Leu Leu Arg Asp Val Ala
  180          185          190
Ala Ala Lys Pro Glu Tyr Ala Phe Val Asp Glu Thr Arg Arg Ala Lys
  195          200          205
Ala Ala Asn Ala Val Glu Lys Gly Ile Val Cys Ile Leu Lys Thr Gln
  210          215          220
Val Val Val Asp Gly Arg Arg Thr Val Trp Cys Ala Gln His Asp Glu
  225          230          235          240
Val Thr Phe Ala Pro Ala Pro Ala Arg Lys Phe Glu Leu Ala Ser Leu
  245          250          255
Ser Gly Gly Glu Ser Val Asp Ile Val Arg Phe Leu Met Ser Ile Lys
  260          265          270
Asp Pro Ser Arg Asn Val Val Glu Ser Ile Glu Ser Ala Val Lys Trp
  275          280          285
Phe Glu Gln Ser Glu Leu Lys Gly Val Lys Trp Val Lys Lys Thr Asp
  290          295          300
Ala Thr Gln Pro Asn Gly Phe Asp Cys Val Val Val Lys Asp Pro Glu
  305          310          315          320
Ser Ser Val Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg Pro Ile
  325          330          335
Phe Ala Gly Arg Asp Gly Val Pro Lys Tyr Asp Val Ala Gln Ile Glu
  340          345          350
His Glu Arg Arg Thr Gly Tyr Glu Trp Tyr Val Asp Glu Ala Ala Lys
  355          360          365
Leu Leu Lys Lys Asp Tyr Pro Ala Trp Lys Lys Arg His Val Val Thr
  370          375          380

```

Thr Arg Val His
385

<210> 13

<211> 1065

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 13

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atgaaaacga tcagccttat ttgcctcgca atctctgctg ggattctgga ttcggttgcg      60
gcggcacgct ggaacgaatt cggccagaag gcggatgatt ggtatcgagg tgacgaaggc      120
aggcgcggtt cttcgaatat tctttctcac caatcactgc aagggaagctg gcccaagaat      180
accgatacca ccgcgagatt cttcaatgga gatctagcga agattcaggg cacttcgac      240
aacggtgcga cgacggacga gttgcgtttc ctggcccgcg cgtttgtcgc caggaagaa      300
aaaaactacg agtcagcggt ccgaaaaggc ttcgaaacaca ttctcgcggc gcaatacgcg      360
aacggcggat ggccgcaata ttcgcccgcg cccaaaaagt accaccgaca cattacctc      420
aacgataatt cgatgggtgc gctgatgatt ttcttctcgc aggtcacgac ttcgaatctc      480
tactcgttcg tcgaagcgcc gctgcgaaca caagcccgcg aaagtctcga tcgcggtgtg      540
cgggtgcattc ttaagtgccg gatcgtcgtg aacgggcaca agaccgctg gtgcgcgcaa      600
catgatgaaa cggatttcag ccccgatcc gcgcgtagtt acgaactgcc ttcgctgagc      660
ggttctgaat cagtcggcat tgtcgcgttg ctgatgagcc tcgatcagcc gagcccgga      720
gtgatcgatg ccatcaccac cgccgtagcg tggttcgaat cggcgaagct gcccgggatc      780
aaaaccgttc aagagaccga tccgaattcg cccaaaggct ggaatcgctg cgtcgtaaaa      840
gatgaaaagt cccgaccgat gtgggcgcgt ttctacgaca tcaacaccaa caaaccttc      900
ttttgtgatc gcgatggtgt gccaaagccg agtcttgccg agatcggtta tgaacggcgg      960
aacggttatg cgtgggtcgg atactggcct gaagacttgc tcgcaagaga gtatccagcg      1020
tggaagatga agtggttgaa gcccaagag cgccagcat ttgga      1065

```

<210> 14

<211> 354

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(22)

<221> DOMAIN

<222> (23)...(354)

<223> Catalytic domain

<400> 14

```

Met Lys Thr Ile Ser Leu Ile Cys Leu Ala Ile Ser Ala Gly Ile Leu
 1          5          10          15
Asp Ser Val Ala Ala Ala Arg Trp Asn Glu Phe Ala Gln Lys Ala Asp
 20          25          30
Asp Trp Tyr Arg Gly Asp Glu Gly Arg Arg Val Ala Ser Asn Ile Leu
 35          40          45
Ser His Gln Ser Leu Gln Gly Ser Trp Pro Lys Asn Thr Asp Thr Thr
 50          55          60
Ala Arg Phe Phe Asn Gly Asp Leu Ala Lys Ile Gln Gly Thr Phe Asp
 65          70          75          80
Asn Gly Ala Thr Thr Asp Glu Leu Arg Phe Leu Ala Arg Ala Phe Val
 85          90          95

```

Ala Thr Lys Glu Lys Asn Tyr Glu Ser Ala Phe Arg Lys Gly Phe Glu
 100 105 110
 His Ile Leu Ala Ala Gln Tyr Ala Asn Gly Gly Trp Pro Gln Tyr Ser
 115 120 125
 Pro Pro Pro Lys Ser Tyr His Arg His Ile Thr Phe Asn Asp Asn Ser
 130 135 140
 Met Val Arg Leu Met Ile Phe Leu Arg Glu Val Thr Thr Ser Asn Leu
 145 150 155 160
 Tyr Ser Phe Val Glu Ala Pro Leu Arg Thr Gln Ala Arg Glu Ser Phe
 165 170 175
 Asp Arg Gly Val Arg Cys Ile Leu Lys Cys Gln Ile Val Val Asn Gly
 180 185 190
 His Lys Thr Ala Trp Cys Ala Gln His Asp Glu Thr Asp Phe Ser Pro
 195 200 205
 Arg Ser Ala Arg Ser Tyr Glu Leu Pro Ser Leu Ser Gly Ser Glu Ser
 210 215 220
 Val Gly Ile Val Arg Leu Leu Met Ser Leu Asp Gln Pro Ser Arg Gly
 225 230 235 240
 Val Ile Asp Ala Ile Thr Asn Ala Val Ala Trp Phe Glu Ser Ala Lys
 245 250 255
 Leu Pro Gly Ile Lys Thr Val Gln Glu Thr Asp Pro Asn Ser Pro Lys
 260 265 270
 Gly Trp Asn Arg Val Val Val Lys Asp Glu Ser Ala Arg Pro Met Trp
 275 280 285
 Ala Arg Phe Tyr Asp Ile Asn Thr Asn Lys Pro Phe Phe Cys Asp Arg
 290 295 300
 Asp Gly Val Pro Lys Pro Ser Leu Ala Glu Ile Gly Tyr Glu Arg Arg
 305 310 315 320
 Asn Gly Tyr Ala Trp Leu Gly Tyr Trp Pro Glu Asp Leu Leu Ala Arg
 325 330 335
 Glu Tyr Pro Ala Trp Lys Met Lys Trp Leu Lys Pro Lys Glu Arg Pro
 340 345 350
 Ala Phe

<210> 15

<211> 1575

<212> DNA

<213> Bacteria

<400> 15

atgagacgac cagtcgcact ccggctccac gcggcactgg ccacctgggc cctggcggcc 60
 gcgacggcg tgggtctctc gatccccag gcatcggcgg cggccggcgg cgccaccggc 120
 tacgccggcc agaaccggcg caccaccggc ggtgcggcgg gccagaccgt acgggcccacc 180
 acgggcaccg ccattccacg gccctgtg gcacggggcca gcagcagcac cccgatcacg 240
 atcgaggtcg agggaaacgat caaccacgcc aacaccgcca aggtgtccgg cccagctgc 300
 aacaccgccg ccggagtgat cagctgaag cagatcagca acgtcacgt cgtcggggtc 360
 ggctccggcg cgtcttcga ccaactcggc atccacatcc gcgagtcag caacatcatc 420
 atccagaacg tgacgggtccg gaacgtcaag aagtcgggct cgcgctgtc caacggcggc 480
 gacgccatcg gcattggagag cgacgtccgc aacgtctggg tcgaccactc caccctggag 540
 gcctcggcg gcgagtcgga gggtacgac ggcctcttcg acatgaagga caacaccggg 600
 tacgtgacc tgctgtacag catctgcgc aaatccgggc gcggcgccct cgtggggtc 660
 agcgagaccg aactctcgaa cagcttcac acgtaccacc acaacctgta cgagaacatc 720
 gactcgcgcg cgccctgct gcgcggcggg accgcccaca tgtacaacaa ccactacctg 780
 cggatacaag agtcggcat caactccgt gccggagccc acgccaaggt ggacaacaac 840
 tacttcgagg actccaagga cgtctcggc acctctaca ccgacgccg cgggtactgg 900
 caggtcagcg gcaacgtcta cgacaacgt acctggtccg cccggggcac cgacaacaac 960
 ccggcggggc cggaccggca gtccaacacc accgtctcca tccctacgc cttcagcctc 1020
 gaccgggcca cctgcgtgcc ggacgtcgtg agccgaacgg cgggtgccgg caagggaactt 1080

caggtgtcga acggcagctg ctcccgcgag acaccacgc ccacgcgcac gggcacgcgg 1140
 accacaccg cgccgacgac tcccaccgg agcccgacgc cctccacgcc cggaccgacc 1200
 cagcccgcg ggaagaacct cagcatcggt gccgggtccg acggttcgag caaggccgac 1260
 ggcaccagct acggcaacgt ccgggacggg gacctcggca cccactggtc tccggccggt 1320
 tcgaccgct ccgtgtcgat caagtggggc agcgcaccca cgtctcccg catcgtcatc 1380
 cgcgagggcg cgggcgcgac gggcgtcatc ggctcctggc tcgtcctgaa cggcgacacc 1440
 ggcgccgtgc tgacctccgg cagcggggcg gggacgatct ccgtccccc gacggccctg 1500
 aagaagatca ccttcgagat caccggggcg agcggcacgc cagggatcgc cgagttcgag 1560
 acgtacggc gctag 1575

<210> 16

<211> 524

<212> PRT

<213> Bacteria

<220>

<221> SIGNAL

<222> (1)...(33)

<221> DOMAIN

<222> (34)...(359)

<223> Catalytic domain

<400> 16

```

Met Arg Arg Pro Val Ala Leu Arg Leu His Ala Ala Leu Ala Thr Leu
1      5      10      15
Ala Leu Ala Ala Ala Thr Gly Val Val Leu Ser Ile Pro Gln Ala Ser
20      25      30
Ala Ala Ala Gly Gly Ala Thr Gly Tyr Ala Gly Gln Asn Gly Gly Thr
35      40      45
Thr Gly Gly Ala Gly Gly Gln Thr Val Arg Ala Thr Thr Gly Thr Ala
50      55      60
Ile His Ala Ala Leu Cys Gly Arg Ala Ser Ser Ser Thr Pro Ile Thr
65      70      75      80
Ile Glu Val Glu Gly Thr Ile Asn His Ala Asn Thr Ala Lys Val Ser
85      90      95
Gly Pro Ser Cys Asn Thr Ala Ala Gly Val Ile Glu Leu Lys Gln Ile
100     105     110
Ser Asn Val Thr Leu Val Gly Val Gly Ser Gly Ala Val Phe Asp Gln
115     120     125
Leu Gly Ile His Ile Arg Glu Ser Ser Asn Ile Ile Ile Gln Asn Val
130     135     140
Thr Val Arg Asn Val Lys Lys Ser Gly Ser Pro Leu Ser Asn Gly Gly
145     150     155     160
Asp Ala Ile Gly Met Glu Ser Asp Val Arg Asn Val Trp Val Asp His
165     170     175
Ser Thr Leu Glu Ala Ser Gly Gly Glu Ser Glu Gly Tyr Asp Gly Leu
180     185     190
Phe Asp Met Lys Asp Asn Thr Arg Tyr Val Thr Leu Ser Tyr Ser Ile
195     200     205
Leu Arg Lys Ser Gly Arg Gly Gly Leu Val Gly Ser Ser Glu Thr Glu
210     215     220
Leu Ser Asn Ser Phe Ile Thr Tyr His His Asn Leu Tyr Glu Asn Ile
225     230     235     240
Asp Ser Arg Ala Pro Leu Leu Arg Gly Gly Thr Ala His Met Tyr Asn
245     250     255
Asn His Tyr Leu Arg Ile Asn Glu Ser Gly Ile Asn Ser Arg Ala Gly
260     265     270

Ala His Ala Lys Val Asp Asn Asn Tyr Phe Glu Asp Ser Lys Asp Val
275     280     285
Leu Gly Thr Phe Tyr Thr Asp Ala Ala Gly Tyr Trp Gln Val Ser Gly
290     295     300
Asn Val Tyr Asp Asn Val Thr Trp Ser Ala Arg Gly Thr Asp Asn Asn
305     310     315     320
Pro Ala Gly Pro Asp Pro Gln Ser Asn Thr Thr Val Ser Ile Pro Tyr
325     330     335
Ala Phe Ser Leu Asp Pro Ala Thr Cys Val Pro Asp Val Val Ser Arg
340     345     350
Thr Ala Gly Ala Gly Lys Gly Leu Gln Val Ser Asn Gly Ser Cys Ser
355     360     365
Pro Gln Thr Pro Thr Pro Thr Pro Thr Gly Thr Pro Thr Thr Pro Ala
370     375     380
Pro Thr Thr Pro Thr Pro Ser Pro Thr Pro Ser Thr Pro Gly Pro Thr
385     390     395     400
Gln Pro Gly Gly Thr Asn Leu Ser Ile Gly Ala Gly Ser Asp Gly Ser
405     410     415
Ser Lys Ala Asp Gly Thr Ser Tyr Gly Asn Val Arg Asp Gly Asp Leu
420     425     430
Gly Thr His Trp Ser Pro Ala Gly Ser Thr Gly Ser Val Ser Ile Lys
435     440     445
Trp Gly Ser Ala Thr Thr Val Ser Arg Ile Val Ile Arg Glu Ala Ala
450     455     460
Gly Ala Thr Gly Val Ile Gly Ser Trp Leu Val Leu Asn Gly Asp Thr
465     470     475     480
Gly Ala Val Leu Thr Ser Gly Ser Gly Ala Gly Thr Ile Ser Val Pro
485     490     495
Arg Thr Ala Leu Lys Lys Ile Thr Phe Glu Ile Thr Gly Ala Ser Gly
500     505     510
Thr Pro Arg Ile Ala Glu Phe Glu Thr Tyr Ala Gly
515     520

```

<210> 17

<211> 1047

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 17

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ttgccgcgtg cgcccggtgg tgagtcgtca tcgccagcgc agacgtcatc ggttgoggtc 60
tcctggggtc agatcctccg tcagcctcgc gcctgggtacg gcggtgcgga ggcgttgcca 120
gtcgtcgaga acgtgctcct gtatcagcgc gcggcaggag ggtggccgaa gaacatcaac 180
atggcggcgc cgatgaccgc cgtgaccgt gcgaaagtca cggacgagcg cgcgcagaaac 240
gacgccacga tcgacaacac gtcaacgacg acgcagatcc gttttcttgc gctcgttctt 300
cgcggcaccg ccgacgcacg attcaaggac gcggcgctga agggcatcga cttcctgctg 360
gctgcgcaat acgcgaatgg aggtcggcct cagtattttc ccotgcgcga cgactactcg 420
cggcgcatca cgttcaatga cgacgcgatg gtgaatgtga tgacgctgct gcgcgagact 480
tcgcagggcc agacgcgctt caggttcgtc gacgcctcgc gcgcgggccc ggcggcgag 540
tctgtctcac gcgcgctcga cgtcatgctg cgcacgcaga ttcgagtcaa cggcgtgctg 600
accggctggt gccagcagca cgacgagcgg aactttcagc cggggaaggc gcgcgcgtac 660
gaacatccgt cgattgccag caaggaaacc gcgagcatcg caagattcct gatggggatt 720
gaacggccgt cgccggagat cgtgtccgcg gtggatggcg cagtcgcgtg gttgcgagcg 780
gcgcagattt caggtgtgcg gacggagcgc cggcccgacg gatcgaatcc ggcggcgagc 840
gtcgtggcgg tgcaggactc cgcgcgcgcc ccaatctggg cccgcttcta cgagattggc 900
accaaccggc cgatgttttc gggtcgcgac ggcgtcatca agtacagcct cagcgagatc 960
gagatcgagc ggcgcgctgg atacagctgg tacggcgact acgcgcgcag actgctcaga 1020

```

gacgactatc cgaagtggaa gaaatga

1047

<210> 18

<211> 348

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(348)

<223> Catalytic domain

<400> 18

```

Met Pro Arg Ala Pro Gly Gly Glu Ser Ser Ser Pro Ala Gln Thr Ser
1      5      10      15
Ser Val Ala Val Ser Trp Asp Gln Ile Leu Arg Gln Pro Ala Ala Trp
20      25      30
Tyr Gly Gly Ala Glu Ala Leu Arg Val Ala Glu Asn Val Leu Leu Tyr
35      40      45
Gln Arg Ala Ala Gly Gly Trp Pro Lys Asn Ile Asn Met Ala Ala Pro
50      55      60
Met Thr Ala Ala Asp Arg Ala Lys Val Thr Asp Glu Arg Ala Gln Asn
65      70      75      80
Asp Ala Thr Ile Asp Asn Thr Ser Thr Thr Thr Gln Ile Arg Phe Leu
85      90      95
Ala Leu Val Leu Arg Gly Thr Ala Asp Ala Arg Phe Lys Asp Ala Ala
100     105     110
Leu Lys Gly Ile Asp Phe Leu Leu Ala Ala Gln Tyr Ala Asn Gly Gly
115     120     125
Trp Pro Gln Tyr Phe Pro Leu Arg Asp Asp Tyr Ser Arg Arg Ile Thr
130     135     140
Phe Asn Asp Asp Ala Met Val Asn Val Met Thr Leu Leu Arg Glu Thr
145     150     155     160
Ser Gln Gly Gln Thr Pro Phe Glu Phe Val Asp Ala Ser Arg Arg Gly
165     170     175
Arg Ala Ala Gln Ser Val Ser Arg Gly Val Asp Val Met Leu Arg Thr
180     185     190
Gln Ile Arg Val Asn Gly Val Leu Thr Gly Trp Cys Gln Gln His Asp
195     200     205
Glu Arg Asn Phe Gln Pro Val Lys Ala Arg Ala Tyr Glu His Pro Ser
210     215     220
Ile Ala Ser Lys Glu Thr Ala Ser Ile Ala Arg Phe Leu Met Gly Ile
225     230     235     240
Glu Arg Pro Ser Pro Glu Ile Val Ser Ala Val Asp Gly Ala Val Ala
245     250     255
Trp Leu Arg Ala Ala Gln Ile Ser Gly Val Arg Thr Glu Arg Arg Pro
260     265     270
Asp Gly Ser Asn Pro Gly Gly Asp Val Val Ala Val Gln Asp Ser Ala
275     280     285
Ala Pro Pro Ile Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg Pro
290     295     300
Met Phe Ser Gly Arg Asp Gly Val Ile Lys Tyr Ser Leu Ser Glu Ile
305     310     315     320
Glu Ile Glu Arg Arg Ala Gly Tyr Ser Trp Tyr Gly Asp Tyr Ala Ala
325     330     335
Arg Leu Leu Arg Asp Asp Tyr Pro Lys Trp Lys Lys
340     345

```

<210> 19

<211> 1122

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 19

```
gtgaacaggt ggccggaaga cttcttgccg gacttcgcgg cccgcatgct ccggtgcatg 60
gttccccggc cgcagatcca ctggggcgcc ggtgtcatcc ggcaggaacc ggaatggtac 120
ggctcggccg aggcgcgtgc gatcgccgac agcgtttctc aataccagtc gaccgctggc 180
ggctggccca agaaccacga cttgacggtc tcgccaccgt ccgccaatt ccttgccgat 240
gcggatggtc tcacgaacac gatcgacaac gacgccacca cgttgccgat gcatattctc 300
gctctggtgg cgcacgcgac cggcgccatc aagtaccgcy ccgcttcca acgcggtctg 360
gactacctgc tcgccgtcca gtatcccaat ggcggctggc ctacgtattt tccctgcgt 420
gacggctatt actcgacat caccataaac gacaatcgca tggtaacgt cctcaccgtt 480
ctgcgcgatg cggcgccggg ccggccccc tactcgttcg tcgacagggc ccggcgccgc 540
agagcagaaa cggccatcgc tcggcgcatc gacatcatcg tcgcactca ggtgagacgg 600
gccggcggtc tgaccgcatg gtgcgccag cagcagaaa agacgctcga gccggcggtg 660
gcgcgcaact acgaaccgcc gacactctcc gggcagaaa gcgtcggcat cgtgcgttt 720
ctcatgggaa tcgaaaagcc cagcccgagg atcgtcgcgg cggtgcaagg cgcgcgtgac 780
tggttgagag ccgtcgcgat cagcgggttg cgtctcgagg aattcaccga ccgcatggc 840
aggcgcgaca ggcgcgtcgt cgcgatccg gcagcgcgc tcctgtgggc gcgcttctac 900
gagcttgaca cggaccgtcc cgtcttcacc ggccgcgaca aggtgatccg gtactcgtc 960
agcgaatcg agcagcagcg ccggaacggg tatgcctact atggcacatg gccggccacg 1020
ctcctcagcg aggagtacc cgttgccgc gcgaacacc tggctcgacg gagcgtcagg 1080
caggtagagg agggaatcgc gatacgcgtc cctaaccct ga 1122
```

<210> 20

<211> 373

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(373)

<223> Catalytic domain

<400> 20

```
Met Asn Arg Trp Arg Glu Asp Phe Leu Arg Asp Phe Ala Ala Arg Met
 1          5          10          15
Leu Arg Cys Met Val Pro Arg Pro Gln Ile His Trp Gly Gly Gly Val
 20          25          30
Ile Arg Gln Glu Pro Glu Trp Tyr Gly Ser Ala Glu Ala Arg Ala Ile
 35          40          45
Ala Asp Ser Val Leu Gln Tyr Gln Ser Thr Ala Gly Gly Trp Pro Lys
 50          55          60
Asn Thr Asp Leu Thr Val Ser Pro Pro Ser Ala Glu Phe Leu Ala Asp
 65          70          75          80
Ala Asp Gly Leu Thr Asn Thr Ile Asp Asn Asp Ala Thr Thr Leu Pro
 85          90          95
Met Arg Phe Leu Ala Leu Val Ala His Ala Thr Gly Gly Ile Lys Tyr
100          105          110
Arg Ala Ala Phe Glu Arg Gly Leu Asp Tyr Leu Leu Ala Ala Gln Tyr
115          120          125

Pro Asn Gly Gly Trp Pro Gln Tyr Phe Pro Leu Arg Asp Gly Tyr Tyr
130          135          140
Ser His Ile Thr Tyr Asn Asp Asn Ala Met Val Asn Val Leu Thr Val
145          150          155          160
Leu Arg Asp Ala Ala Ala Gly Arg Pro Pro Tyr Ser Phe Val Asp Arg
165          170          175
Ala Arg Arg Ala Arg Ala Glu Thr Ala Ile Ala Arg Gly Ile Asp Ile
180          185          190
Ile Val Arg Thr Gln Val Arg Arg Ala Gly Val Leu Thr Ala Trp Cys
195          200          205
Ala Gln His Asp Glu Lys Thr Leu Glu Pro Ala Trp Ala Arg Asn Tyr
210          215          220
Glu Pro Pro Thr Leu Ser Gly His Glu Ser Val Gly Ile Val Arg Phe
225          230          235          240
Leu Met Gly Ile Glu Lys Pro Thr Pro Arg Ile Val Ala Ala Val Gln
245          250          255
Gly Ala Ala Asp Trp Leu Arg Ala Val Ala Ile Ser Gly Leu Arg Leu
260          265          270
Glu Glu Phe Thr Asp Ala Asp Gly Arg Arg Asp Arg Val Val Ala
275          280          285
Asp Pro Ala Ala Pro Leu Leu Trp Ala Arg Phe Tyr Glu Leu Gly Thr
290          295          300
Asp Arg Pro Val Phe Thr Gly Arg Asp Lys Val Ile Arg Tyr Ser Leu
305          310          315          320
Ser Glu Ile Glu His Glu Arg Arg Asn Gly Tyr Ala Tyr Tyr Gly Thr
325          330          335
Trp Pro Ala Thr Leu Leu Ser Glu Glu Tyr Pro Arg Trp Arg Ala Lys
340          345          350
His Leu Ala Arg Arg Ser Val Arg Gln Val Glu Glu Gly Ile Ala Ile
355          360          365
Arg Val Pro Asn Pro
370
```

<210> 21

<211> 1269

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 21

atgcgtaaat	cgaactgggc	cgtcacaacg	gccatcctgc	tgcgctgag	cgccgcaccg	60
ctggcgga	agcccatcg	acagatcacc	ctcgccgtgc	cgctcagccc	ggcgcgctg	120
accgaaacgc	cgcttgagca	gcgggcgcaa	tggcaggcct	atctcgccac	caccgaggca	180
cagcttaagg	cagacaaggc	ggcgctggct	gccgagcgcg	ccggtctggc	cgaaatcccc	240
gccaagccga	agaccggcag	cgccaacacc	atgcgctcg	acaagccgct	ggaatggtag	300
gcgtcgctcg	agggcggtct	ggtcgccgat	aatatcgta	gctatcagac	tccggcaggc	360
ggctggggca	aaaatcaggc	ccgcaacgaa	cccacgggt	tgaagggtca	ggcctacact	420
atcgatgacg	cgatccac	cggttcgggc	aaatggaact	tgcgcggcac	catcgacaac	480
gacgccacca	tcgtagaat	tgcctttctc	gcccgcgtag	cgcgcgcgcg	cacgggcccg	540
gaaggcgacg	tctatcgcg	ctccgccacg	cgcggcatca	cctacttgct	ggcgcgcgag	600
taccccaatg	gcggctggcc	gcaggtcttg	ccgcttcagg	gcggctatca	cgacgccatc	660
accctcaatg	acggcgcgat	gatccatgtg	ctgaactgt	ttgacgacat	cgccagcgga	720
caggcgcgact	tgccttctc	gcctgagccg	ctgcgcgaca	aggtcgaggc	cgacaggca	780
aagggtcaga	aggtgcttct	cgatcttcag	cttaagcgca	acggcggaacg	cacctgtgg	840
gcgcagcagt	acgatccgat	tacctctttg	cccagcgcg	cgcgtaacta	cgagccgtcg	900
tcgatcagca	ccggtgaaag	cgccggtgtg	ctgatctacc	tcatgtccct	gcccaacccc	960
tcgcctgaag	tgcgcgacgc	catcgaaaaa	ggcgtggccc	tgtgatcaa	acttcagatc	1020

aacggcatgg	catgggaaaa	ggacggcatg	cgcaaacgtc	tggtcgcaa	ggctgacgcc	1080
tcgccgctgt	ggtcgcgcta	tcacgactcg	gaaacgctgc	tgcccatctt	cggtgaccgc	1140
gacatgcgca	tcttcgacga	cgtcaacgac	atcagcgacg	aacgcagccg	cggtatgcc	1200
tggtaggca	caagcccgcc	acgggccatc	gccgaatacg	aaaaatggaa	acagggcaac	1260
ggcaaatga						1269

<210> 22

<211> 422

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(23)

<221> DOMAIN

<222> (24)...(422)

<223> Catalytic domain

<400> 22

```

Met Arg Lys Ser Asn Trp Ala Val Thr Thr Ala Ile Leu Leu Ala Leu
1      5      10      15
Ser Ala Ala Pro Leu Ala Ala Lys Pro Ile Gly Gln Ile Thr Leu Ala
20     25     30
Val Pro Leu Ser Pro Ala Arg Leu Thr Glu Thr Pro Pro Glu Gln Arg
35     40     45
Ala Gln Trp Gln Ala Tyr Leu Ala Thr Thr Glu Ala Gln Leu Lys Ala
50     55     60
Asp Lys Ala Ala Leu Ala Ala Glu Arg Ala Gly Leu Ala Glu Ile Pro
65     70     75     80
Ala Lys Pro Lys Thr Gly Ser Ala Asn Thr Met Pro Leu Asp Lys Pro
85     90     95
Leu Glu Trp Tyr Ala Ser Ser Glu Ala Arg Leu Val Ala Asp Asn Ile
100    105    110
Val Ser Tyr Gln Thr Pro Ala Gly Gly Trp Gly Lys Asn Gln Ala Arg
115    120    125
Asn Glu Pro Thr Arg Leu Lys Gly Gln Ala Tyr Thr Ile Asp Asp Ala
130    135    140
Asp Pro Thr Gly Ser Gly Lys Trp Asn Phe Val Gly Thr Ile Asp Asn
145    150    155    160
Asp Ala Thr Ile Val Glu Ile Arg Phe Leu Ala Arg Val Ala Ala Ala
165    170    175
Ala Thr Gly Pro Glu Gly Asp Val Tyr Arg Ala Ser Ala Thr Arg Gly
180    185    190
Ile Thr Tyr Leu Leu Ala Ala Gln Tyr Pro Asn Gly Gly Trp Pro Gln
195    200    205
Val Trp Pro Leu Gln Gly Gly Tyr His Asp Ala Ile Thr Leu Asn Asp
210    215    220
Gly Ala Met Ile His Val Leu Glu Leu Phe Asp Ile Ala Ser Gly
225    230    235    240
Gln Gly Asp Phe Ala Phe Leu Pro Glu Pro Leu Arg Asp Lys Val Glu
245    250    255
Ala Ala Gln Ala Lys Gly Gln Lys Val Leu Leu Asp Leu Gln Leu Lys
260    265    270
Arg Asn Gly Glu Arg Thr Leu Trp Ala Gln Gln Tyr Asp Pro Ile Thr
275    280    285
Leu Leu Pro Ser Ala Ala Arg Asn Tyr Glu Pro Ser Ser Ile Ser Thr

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290      295      300
Gly Glu Ser Ala Gly Val Leu Ile Tyr Leu Met Ser Leu Pro Asn Pro
305    310    315    320
Ser Pro Glu Val Arg Asp Ala Ile Glu Lys Gly Val Ala Leu Leu Ile
325    330    335
Lys Leu Gln Ile Asn Gly Met Ala Trp Glu Lys Asp Gly Met Arg Lys
340    345    350
Arg Leu Val Ala Lys Ala Asp Ala Ser Pro Leu Trp Ser Arg Tyr His
355    360    365
Asp Ser Glu Thr Leu Leu Pro Ile Phe Gly Asp Arg Asp Met Arg Ile
370    375    380
Phe Asp Asp Val Asn Asp Ile Ser Asp Glu Arg Ser Arg Gly Tyr Ala
385    390    395    400
Trp Tyr Gly Thr Ser Pro Ala Arg Ala Ile Ala Glu Tyr Glu Lys Trp
405    410    415
Lys Gln Gly Asn Gly Lys
420

```

<210> 23

<211> 1182

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 23

```

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ctgggggtata tgacgcctgc gcagcgttg accgagggcg gcattgccc gctgccggcg 120
tcggagcagg gcgcctggcg gggctacct gccgcctccc gcgcagccat ggacgccgac 180
aaggcccgccc tggccgcccga gcgcgcgcgc ctgcgccacc taccgccggc gccgccgcat 240
ggcgggtgtg atggcgggat ggcgcgcac cgtccgacgg ctgtgtatgg gacgccggaa 300
gcgcggcaca tcgcggacaa tatcgtcagc ttccagacgc cgtccggcgg ctggggcaag 360
aacgtggacc gcacgggacc tgtgcgccag cgcggacagc attacgtttc cttcgatggc 420
aaggagtcct ggaacttcat cggcacgata gacaacaacg ccacaacgag cgaagtga 480
ttcctggcgc gcgtgcaggc gcaaatgccc ggcgcggcgg gcgacgaata ccggaaggcc 540
gccctgcgcg gcatcagcta cctgttgaa tcacaatat ccaacggcgg cttcccgca 600
gtctatccgc tgcaaggcgg ctaccacgac gccatcacct tcaacgacga tgccttcgcc 660
aacgtgctgc aagtgtctgt ggaagtggcg aaccgaggg gcgactatgc cttcgtcccc 720
gaaaccgtgg caaccgatgc ccgcgcggcc gcggacaagg cgctccaagt cctgctggcg 780
agccagatca tcgtcggcgg cgtacgcacc gcctggtgcc agcagcacga tgcgatcacg 840
ctggcggccc tcggcggccc caatttcgaa ccggccgcgc tgaccagcac ggaagcgcg 900
cgctgctga tgcgttgat gctgtgcc ccgagcgcc cggagctgag agcgtcaatc 960
catgccccga tggcctggct gcagaaagcg gcgctgcggg gggatgtgtg gtcgcgtac 1020
tatgacctga acacgatgag gccgatcttt ggggatcggt accgcagtat ccacgatgat 1080
gtgaaggaaat tgagcgaggg gaggcaaaaa ggctatggct gggttcagtaa cggaccagcc 1140
agagctaaac aggccttttga ggccctggacg cgcaaacctt ga 1182

```

<210> 24

<211> 393

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(18)

<221> DOMAIN

<222> (19)...(393)

<223> Catalytic domain

<400> 24

```

Met Asn Arg Gly Val Ile Val Leu Leu Ala Ala Ala Pro Ala Ala Ala
 1          5          10          15
His Gly Ala Val Leu Gly Tyr Met Thr Pro Ala Gln Pro Leu Thr Glu
 20          25          30
Ala Arg Ile Ala Ala Leu Pro Ala Ser Glu Gln Gly Ala Trp Arg Gly
 35          40          45
Tyr Leu Ala Arg Ser Arg Ala Ala Met Asp Ala Asp Lys Ala Ala Leu
 50          55          60
Ala Ala Glu Arg Ala Ala Leu Ala Thr Val Pro Pro Ala Pro Pro His
 65          70          75          80
Gly Gly Gly Asp Gly Gly Met Ala Arg Asn Arg Pro Thr Ala Trp Tyr
 85          90          95
Gly Thr Pro Glu Ala Arg His Ile Ala Asp Asn Ile Val Ser Phe Gln
100          105          110
Thr Pro Ser Gly Gly Trp Gly Lys Asn Val Asp Arg Thr Gly Pro Val
115          120          125
Arg Gln Arg Gly Gln His Tyr Val Ser Phe Asp Gly Lys Glu Ser Trp
130          135          140
Asn Phe Ile Gly Thr Ile Asp Asn Asn Ala Thr Thr Ser Glu Leu Lys
145          150          155          160
Phe Leu Ala Arg Val Gln Ala Gln Met Pro Gly Ala Ala Gly Asp Glu
165          170          175
Tyr Arg Lys Ala Ala Leu Arg Gly Ile Ser Tyr Leu Leu Asn Ser Gln
180          185          190
Tyr Pro Asn Gly Gly Phe Pro Gln Val Tyr Pro Leu Gln Gly Gly Tyr
195          200          205
His Asp Ala Ile Thr Phe Asn Asp Asp Ala Phe Ala Asn Val Leu Gln
210          215          220
Val Leu Leu Glu Val Ala Asn Arg Arg Gly Asp Tyr Ala Phe Val Pro
225          230          235          240
Glu Thr Val Ala Thr Asp Ala Arg Ala Ala Asp Lys Ala Leu Gln
245          250          255
Val Leu Leu Ala Ser Gln Ile Ile Val Gly Gly Val Arg Thr Ala Trp
260          265          270
Cys Gln Gln His Asp Ala Ile Thr Leu Ala Pro Val Gly Ala Arg Asn
275          280          285
Phe Glu Pro Ala Ala Leu Thr Ser Thr Glu Ser Ala Arg Leu Leu Met
290          295          300
Leu Leu Met Leu Leu Pro Asp Pro Ser Pro Glu Leu Arg Ala Ser Ile
305          310          315          320
His Ala Gly Met Ala Trp Leu Gln Lys Ala Ala Leu Pro Gly Asp Val
325          330          335
Trp Ser Arg Tyr Tyr Asp Leu Asn Thr Met Arg Pro Ile Phe Gly Asp
340          345          350
Arg Asp Arg Ser Ile His Asp Asp Val Lys Glu Leu Ser Glu Glu Arg
355          360          365
Gln Lys Gly Tyr Gly Trp Phe Ser Asn Gly Pro Ala Arg Ala Lys Gln
370          375          380
Ala Phe Glu Ala Trp Thr Arg Lys Pro
385          390

```

<210> 25

<211> 1194

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 25

```

ttggtcgctg cccatttaag ctgcggcagc gccaatctct atgcagaatc aaccgcaaaa 60
tcggttacgc aatcagcagc cacaatcaa ttgcaaatg aaaaaagcag ttgggacagc 120
tattacgccc catcaaaaa aatacatcag gcagaccagg attttctcgc cgctgaatta 180
aaaaaactcg gtcagaaaa accaacattg cccgcacaca ccaaagattt tggttttgat 240
gttaagcagg taaatgcaga ttggtttaa agtgacgaag gcaaacgtgt gatggagatt 300
attctctcct tccaaacccc gtccggcggg ttgtcaaagc gtaccgacat ggccaaggcg 360
gtgcgacaac ctgggcaagc ctttggcggt gaaaaaggct atatcccaac atttgataat 420
ggcgctacca gcactcaatt gatgttgctc gcgcaagcac accaagccac cgcgatcac 480
cgcttttagc acgcatttgg gcgcggcttg caattaattt tgactgcgca ataccogaat 540
ggtggctggc caaaaaactt tccactaacc ggtagctacc acgattacat cacctacaac 600
gacaatctta cgcgcgacct gatggtagt ctgcacaaaa cagcgcaggc aaaaaatgat 660
tttgcatctg tgaccaaaagc gcagcaaatc gcagcgtag ctagcctcgc gcgtgcactt 720
gattgcgtat tgaatcaca agttgtcgtc aatggcacac gcacactctg ggcgcacag 780
cacgatgta aaacactgca accaaccaaa gcgcgcgcac ttgaaatggt gtcactcact 840
accactgaaa gcgcagccat gctcagtttt ctgatggata tcaaaaatcc cagcgcggat 900
attattcaat ccatacatgc agccatagcc tggtagtagc aaaataaat cgtcggaaaa 960
acctggacac gtggtgatgc ggaattaaaa gataataaaa attcgcagcc actctgggcg 1020
cggttttatg agataggcac taataagcct atatttgggg atcgcgatga cactgtgtat 1080
tacgatttgg caaaagtgtc faaagagcgt cgcgagggtt atgcgttgta ctccactgac 1140
ccgaataaga cgctaaaaaa atatgctgaa tgggtctaaa aatatcccaa ataa 1194

```

<210> 26

<211> 397

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(15)

<221> DOMAIN

<222> (16)...(397)

<223> Catalytic domain

<400> 26

```

Met Val Ala Ala Leu Leu Ser Cys Gly Ser Ala Asn Leu Tyr Ala Glu
1      5      10      15
Ser Thr Ala Lys Ser Val Thr Gln Ser Ala Ala Thr Asn Gln Leu Gln
20     25     30
Asn Glu Lys Ser Ser Trp Asp Ser Tyr Tyr Ala Ala Ser Lys Lys Ile
35     40     45
His Gln Ala Asp Gln Asp Phe Leu Ala Ala Glu Leu Lys Lys Leu Gly
50     55     60
Gln Lys Lys Pro Thr Leu Pro Ala His Thr Lys Asp Phe Gly Phe Asp
65     70     75     80
Val Lys Gln Val Asn Ala Asp Trp Phe Lys Ser Asp Glu Gly Lys Arg
85     90     95
Val Met Glu Ile Ile Leu Ser Phe Gln Thr Pro Ser Gly Gly Trp Ser
100    105    110
Lys Arg Thr Asp Met Ala Lys Ala Val Arg Gln Pro Gly Gln Ala Phe
115    120    125

```

Gly Val Glu Lys Gly Tyr Ile Pro Thr Phe Asp Asn Gly Ala Thr Ser
 130 135 140
 Thr Gln Leu Met Leu Leu Ala Gln Ala His Gln Ala Thr Gly Asp His
 145 150 155 160
 Arg Phe Ser Asp Ala Phe Gly Arg Gly Leu Gln Leu Ile Leu Thr Ala
 165 170 175
 Gln Tyr Pro Asn Gly Gly Trp Pro Gln Asn Phe Pro Leu Thr Gly Ser
 180 185 190
 Tyr His Asp Tyr Ile Thr Tyr Asn Asp Asn Leu Thr Arg Asp Leu Met
 195 200 205
 Val Val Leu His Lys Thr Ala Gln Ala Lys Asn Asp Phe Ala Phe Val
 210 215 220
 Thr Lys Ala Gln Gln Ile Ala Ala Ser Ala Ser Leu Ala Arg Ala Leu
 225 230 235 240
 Asp Cys Val Leu Lys Ser Gln Val Val Val Asn Gly Thr Arg Thr Leu
 245 250 255
 Trp Gly Ala Gln His Asp Val Lys Thr Leu Gln Pro Thr Lys Ala Arg
 260 265 270
 Ala Phe Glu Met Val Ser Leu Thr Thr Thr Glu Ser Ala Ala Met Leu
 275 280 285
 Ser Phe Leu Met Asp Ile Lys Asn Pro Ser Ala Asp Ile Ile Gln Ser
 290 295 300
 Ile His Ala Ala Ile Ala Trp Tyr Glu Gln Asn Lys Ile Val Gly Lys
 305 310 315 320
 Thr Trp Thr Arg Gly Asp Ala Glu Leu Lys Asp Asn Lys Asn Ser Gln
 325 330 335
 Pro Leu Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Lys Pro Ile Phe
 340 345 350
 Gly Asp Arg Asp Asp Thr Val Tyr Tyr Asp Leu Ala Lys Val Ser Lys
 355 360 365
 Glu Arg Arg Glu Gly Tyr Ala Trp Tyr Ser Thr Asp Pro Asn Lys Thr
 370 375 380
 Leu Lys Lys Tyr Ala Glu Trp Ser Lys Lys Tyr Pro Lys
 385 390 395

<210> 27

<211> 1917

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 27

gtgtctctct	ttagaaaaact	cgcactgccc	gttctgtgccc	gtctactgct	ttctgtcgga	60
gcagaaaacc	gagcgtcgaa	gcgcattgtc	gtggccgctg	atggatcggg	tgacgtcagg	120
acgattcaac	aagcgttgga	ccaggttccc	aaagacaata	cacaccocgt	cttgattcag	180
atcaagccgg	gtgtgtatca	ggaacaagt	cgtgtgcgcc	ccggcaaacg	ctttatcact	240
cttcgcggcg	acgacgcgag	caagaccgtc	atcacctatc	gattgagcgc	actacaagcg	300
ggaataacc	gggtggcatt	caccacetta	attaatgcag	acgactttcg	cgccgagAAC	360
ctgacgtttg	aaaactccct	cggcaccggt	tcacaagcgg	ttgctttgtt	tgtcgatcgg	420
aaccgcgcga	cgtttgaaaa	ctgcgggttc	ctcgggtggc	aggacacttt	gtttgtgaac	480
ggcagccgcc	actctctcaa	agactgtctc	gtcgaaggcc	atgtcgattt	cattttcggc	540
acggcctccg	cagtgtttga	gaactgcacc	attcacagca	aaggcgaagg	ttatgtgacc	600
gcgcactatc	gcaccagcga	tgagatggat	accggttttg	tctttcatcg	ttgtcgtttg	660
accggacgag	acacggggcg	cggagtttat	ctcgggaaggc	cgtggcgacc	ttacgcgcgc	720
gtcgtcttta	tcgattgctg	gctggacgca	cacatcagac	ctgaaggctg	ggataattgg	780
agagatcccg	aacgagagaa	gaccgcgtgg	tttgccgagt	acaagtcaaa	agggcccggt	840
gctaataccc	tagctcgtgt	cgcgtggtcc	aggcagttga	cgacagaaca	agccgccgag	900

ttttcgcggg	aacgcttttt	cagccgcgct	gttcgcgggc	tctctgggca	ggccaaccag	960
gcagtcggaa	cgatcgcgtg	ggacgatgcg	cagaaaaaac	cgaacgagtg	gtatgcgagc	1020
gccgaggcgt	tgcccatcgc	cgacaacgtt	gttctttatc	aacgtgactc	cgccgctggt	1080
cccaagaaca	tcgacatggg	gaagccgctc	gacgacaagg	gtcagccggg	tctctgcgc	1140
gtgcgtaaga	agaacgattc	caccatcgat	aacggcgcca	cttacacgca	actctcgttt	1200
ctagcgcggg	tttacacggc	gcaaaagcag	gagcggcatc	gcgagtcgtt	tctgaaggga	1260
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ctcaacggct	attacaaaca	catcactttc	aacgacaacg	cgatgatcgg	cgtgatgaaa	1380
ctgctgcgcg	acgtagcggc	agcgaaaccg	gcgtatgcgt	ttgtcgacga	agcacgacga	1440
acgagtcggg	cgaaggcggg	cgaaaaagga	atcgagtcca	tactgaagac	gcaggtggtt	1500
gtgaatggcc	ggcgcaccgt	gtggtgtgcg	caacatgacg	aagtcacgct	cgcgcctgcc	1560
ccggcgaggga	cgtttgaatt	agtttcgctg	agtgtgtgtg	aaagcgttga	gatcgtgcgc	1620
tttttgatgt	cgatcaagaa	cccgtcgccg	gcggtgtgct	aggcgatcga	gtcggcgggt	1680
gcgtggttcg	agcaatcgca	agtgaagat	cccgcggcca	aacctgcgtg	ggcgcgattt	1740
tatgagatcg	gcactaatcg	tccgatcttc	gccggggcgtg	acggcgtcgt	taagtatgat	1800
gtgaaacaga	tcgatgagga	acgacgaaag	aattacgcat	ggtacgttga	cgacgcagcg	1860
aaactactga	agaccgacta	tcctgagtg	aaagaaaaga	acgcacaaag	tcaatga	1917

<210> 28

<211> 638

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(21)

<221> DOMAIN

<222> (28)...(308)

<223> Pectin methyl esterase domain

<221> DOMAIN

<222> (309)...(638)

<223> Catalytic domain

<400> 28

```

Met Ser Leu Phe  Arg Lys Leu Ala Leu Pro Val Leu Cys Gly Leu Leu
 1      5      10      15
Leu Ser Val Gly Ala Glu Thr Arg Ala Ser Lys Arg Ile Val Val Ala
 20      25      30
Ala Asp Gly Ser Gly Asp Val Arg Thr Ile Gln Gln Ala Val Asp Gln
 35      40      45
Val Pro Lys Asp Asn Thr His Pro Val Leu Ile Gln Ile Lys Pro Gly
 50      55      60
Val Tyr Gln Glu Gln Val Arg Val Ala Ala Gly Lys Arg Phe Ile Thr
 65      70      75      80
Leu Arg Gly Asp Asp Ala Ser Lys Thr Val Ile Thr Tyr Arg Leu Ser
 85      90      95
Ala Leu Gln Ala Gly Asn Thr Arg Leu Ala Phe Thr Thr Leu Ile Asn
100      105      110
Ala Asp Asp Phe Arg Ala Glu Asn Leu Thr Phe Glu Asn Ser Phe Gly
115      120      125
Thr Gly Ser Gln Ala Val Ala Leu Phe Val Asp Ala Asn Arg Ala Thr
130      135      140
Phe Glu Asn Cys Arg Phe Leu Gly Trp Gln Asp Thr Leu Phe Val Asn
145      150      155      160
Gly Ser Arg His Phe Phe Lys Asp Cys Tyr Val Glu Gly His Val Asp

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165      170      175
Phe Ile Phe Gly Thr Ala Ser Ala Val Phe Glu Asn Cys Thr Ile His
180      185      190
Ser Lys Gly Glu Gly Tyr Val Thr Ala His Tyr Arg Thr Ser Asp Glu
195      200      205
Met Asp Thr Gly Phe Val Phe His Arg Cys Arg Leu Thr Gly Arg Asp
210      215      220
Thr Gly Arg Gly Val Tyr Leu Gly Arg Pro Trp Arg Pro Tyr Ala Arg
225      230      235      240
Val Val Phe Ile Asp Cys Trp Leu Asp Ala His Ile Arg Pro Glu Gly
245      250      255
Trp Asp Asn Trp Arg Asp Pro Glu Arg Glu Lys Thr Ala Trp Phe Ala
260      265      270
Glu Tyr Lys Ser Lys Gly Pro Gly Ala Asn Pro Val Ala Arg Val Ala
275      280      285
Trp Ser Arg Gln Leu Thr Thr Glu Gln Ala Ala Glu Phe Ser Arg Glu
290      295      300
Arg Phe Phe Ser Arg Ala Val Arg Gly Leu Ser Gly Gln Ala Asn Gln
305      310      315      320
Ala Val Gly Thr Ile Ala Trp Asp Asp Ala Gln Lys Lys Pro Asn Glu
325      330      335
Trp Tyr Ala Ser Ala Glu Ala Leu Arg Ile Ala Asp Asn Val Val Leu
340      345      350
Tyr Gln Arg Asp Ser Gly Gly Trp Pro Lys Asn Ile Asp Met Gly Lys
355      360      365
Pro Leu Asp Asp Lys Gly Arg Ala Gly Leu Leu Arg Val Arg Lys Lys
370      375      380
Asn Asp Ser Thr Ile Asp Asn Gly Ala Thr Tyr Thr Gln Leu Ser Phe
385      390      395      400
Leu Ala Arg Val Tyr Thr Ala Gln Lys Gln Glu Arg His Arg Glu Ser
405      410      415
Phe Leu Lys Gly Leu Asp Tyr Leu Leu Lys Ala Gln Tyr Pro Asn Gly
420      425      430
Gly Trp Pro Gln Phe Tyr Pro Asn Leu Asn Gly Tyr Tyr Lys His Ile
435      440      445
Thr Phe Asn Asp Asn Ala Met Ile Gly Val Met Lys Leu Leu Arg Asp
450      455      460
Val Ala Ala Ala Lys Pro Ala Tyr Ala Phe Val Asp Glu Ala Arg Arg
465      470      475      480
Thr Ser Ala Ala Lys Ala Val Glu Lys Gly Ile Glu Cys Ile Leu Lys
485      490      495
Thr Gln Val Val Val Asn Gly Arg Arg Thr Val Trp Cys Ala Gln His
500      505      510
Asp Glu Val Thr Leu Ala Pro Ala Pro Ala Arg Thr Phe Glu Leu Val
515      520      525
Ser Leu Ser Gly Gly Glu Ser Val Glu Ile Val Arg Phe Leu Met Ser
530      535      540
Ile Lys Asn Pro Ser Pro Ala Val Val Glu Ala Ile Glu Ser Ala Val
545      550      555      560
Ala Trp Phe Glu Gln Ser Gln Val Lys Asp Pro Ala Gly Lys Pro Ala
565      570      575
Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg Pro Ile Phe Ala Gly
580      585      590
Arg Asp Gly Val Val Lys Tyr Asp Val Lys Gln Ile Asp Glu Glu Arg
595      600      605
Arg Lys Asn Tyr Ala Trp Tyr Val Asp Asp Ala Ala Lys Leu Leu Lys
610      615      620
Thr Asp Tyr Pro Glu Trp Lys Glu Lys Asn Ala Lys Asp Gln
625      630      635

```

<210> 29

<211> 1398

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 29

```

atgattaacc gtcgagattt cataaaagac ctcacatca cctccgcgg agtcgcggtt 60
ctcccgcaac tggcggtcgg acaaaacgat ccctggaaaa ctcaataccc gcagatcctc 120
gcgcggatca aaccgcgaa atttcgaag cgcgatttcg tcatcacgaa gtccggcgcg 180
aaggcgggaa ccgatatcac gcaagcgatc gctaaagccc tcgacgcttg cgcgaaagcc 240
ggcgccggac gcgtcgtcgt accgcgcggc gaatttctca ccggtgcgat ccactctcaag 300
tcgaacacca atctctacgt ctcaaaaggc gcgactctga agttttcgac cgaccccgaa 360
aaatatctgc cgategttca cacgcggtgg gaaggatgg agttgatgca tctctcgccg 420
ttcatctacg cgtacgagca gacgaacatc gcgatcacgg gcgagggcac gtcgacggc 480
caaggcaaat cgttcttttg gaagtggcac ggcaaccgcg gatacggcgg caaccccgaa 540
gtgatcagtc agcaaaaagc gcgggcgcga ctttacgaga tgatggacaa gaacgtaccc 600
gtcgcggagc gcgtgttcgg tatcgggcac tatctccggc cgcagttcat ccagccgtac 660
aaatgtaaga acgtcttgat cgaaggcgtg acgatcatcg actcgccgat gtgggaagtt 720
catcgggtgc tttgcgagaa tgtcacgcgc cgaatcttc acatctcgtc gcacggtccg 780
aacaacgacg gctgcgatcc cgagtcgtgc aaagacgtcc tgatcgacaa ctgcttcttc 840
gacaccggtg acgactgcat cgcgatcaag tcgggtcgca ataacgacgg tcgtcgtctg 900
aacaccgcca ccgagaacat catcgtccgc aactgcacga tgaagacgg tcacggtggt 960
atcaccggtc gaagcgagat ctccggcggc gtgcgaaact tgttcgcaca cgattgcaag 1020
atggacagtg cggatctgtg gaccgcgctc cgggtaaaaga acaacgcacg gcggggcggc 1080
atgctggaga atttctattt ccgcaacatc accgtcgggc aagtcgcgcg tgctgtggtc 1140
gagatcgatt tcaactatga agaaggcgcg aagggtcgt acacaccggt catcgccaac 1200
tacgtggtg aggatctgac gtgcaccagc gggaaccggc ccgtcgatct gcaagatta 1260
gacaacgcgc caatttacga tgtgtcgctg cgtaacacga ccttcggcgc gatgaagaac 1320
aagagcgctg tgaagaatgt ccgaggactg aagatcgaaa acggtaccgt cagcggcacg 1380
cgcgtggaga gtttatga 1398

```

<210> 30

<211> 465

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(27)

<221> DOMAIN

<222> (77)...(459)

<223> Catalytic domain

<400> 30

```

Met Ile Asn Arg Arg Asp Phe Ile Lys Asp Leu Ile Ile Thr Ser Ala
 1           5           10           15
Gly Val Ala Val Leu Pro Gln Leu Ala Phe Gly Gln Asn Asp Pro Trp
          20           25           30
Lys Thr Gln Tyr Pro Gln Ile Leu Ala Arg Ile Lys Pro Pro Lys Phe
          35           40           45
Pro Lys Arg Asp Phe Val Ile Thr Lys Phe Gly Ala Lys Ala Gly Thr
          50           55           60

```

```

Asp Ser Thr Gln Ala Ile Ala Lys Ala Leu Asp Ala Cys Ala Lys Ala
65      70      75      80
Gly Gly Gly Arg Val Val Val Pro Ala Gly Glu Phe Leu Thr Gly Ala
85      90      95
Ile His Leu Lys Ser Asn Thr Asn Leu Tyr Val Ser Lys Gly Ala Thr
100     105     110
Leu Lys Phe Ser Thr Asp Pro Glu Lys Tyr Leu Pro Ile Val His Thr
115     120     125
Arg Trp Glu Gly Met Glu Leu Met His Leu Ser Pro Phe Ile Tyr Ala
130     135     140
Tyr Glu Gln Thr Asn Ile Ala Ile Thr Gly Glu Gly Thr Leu Asp Gly
145     150     155
Gln Gly Lys Ser Phe Phe Trp Lys Trp His Gly Asn Pro Arg Tyr Gly
165     170     175
Gly Asn Pro Glu Val Ile Ser Gln Gln Lys Ala Arg Ala Arg Leu Tyr
180     185     190
Glu Met Met Asp Lys Asn Val Pro Val Ala Glu Arg Val Phe Gly Ile
195     200     205
Gly His Tyr Leu Arg Pro Gln Phe Ile Gln Pro Tyr Lys Cys Lys Asn
210     215     220
Val Leu Ile Glu Gly Val Thr Ile Ile Asp Ser Pro Met Trp Glu Val
225     230     235
His Pro Val Leu Cys Glu Asn Val Thr Val Arg Asn Leu His Ile Ser
245     250     255
Ser His Gly Pro Asn Asn Asp Gly Cys Asp Pro Glu Ser Cys Lys Asp
260     265     270
Val Leu Ile Asp Asn Cys Phe Phe Asp Thr Gly Asp Asp Cys Ile Ala
275     280     285
Ile Lys Ser Gly Arg Asn Asn Asp Gly Arg Arg Leu Asn Thr Pro Thr
290     295     300
Glu Asn Ile Ile Val Arg Asn Cys Thr Met Lys Asp Gly His Gly Gly
305     310     315
Ile Thr Val Gly Ser Glu Ile Ser Gly Gly Val Arg Asn Leu Phe Ala
325     330     335
His Asp Cys Lys Met Asp Ser Ala Asp Leu Trp Thr Ala Leu Arg Val
340     345     350
Lys Asn Asn Ala Ser Arg Gly Gly Met Leu Glu Asn Phe Tyr Phe Arg
355     360     365
Asn Ile Thr Val Gly Gln Val Ala Arg Ala Val Val Glu Ile Asp Phe
370     375     380
Asn Tyr Glu Glu Gly Ala Lys Gly Ser Tyr Thr Pro Val Met Arg Asn
385     390     395
Tyr Val Val Glu Asp Leu Thr Cys Thr Ser Gly Asn Arg Pro Val Asp
405     410     415
Leu Gln Gly Leu Asp Asn Ala Pro Ile Tyr Asp Val Ser Leu Arg Asn
420     425     430
Thr Thr Phe Gly Ala Met Lys Asn Lys Ser Val Val Lys Asn Val Arg
435     440     445
Gly Leu Lys Ile Glu Asn Val Thr Val Ser Gly Thr Arg Val Glu Ser
450     455     460
Leu
465

```

<210> 31

<211> 1401

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 31

```

atgatcaatc tttatggcgt ctttgacatc cggacctttg gggcccaacc ggacggagaa 60
acgccttcca ctgcggcgat tacggcggcc atcgaaactt gtgccgcggc cgggggagga 120
gtgggtctaca tcccgccggg acggttcttc accggtcccc tccgcctcaa aagccacgtc 180
cggctccatc tcgaggccgg agcgcaactt ctctttagtc aggaccgggc cgattatcct 240
gttctggaga cgaggtggga ggggaaggag gtcttgacct atgcacacca gatctacggc 300
gaggacctcg aaggggctcg gattaccggt cgggggacca tcgacggccg gggcgagact 360
tggtggcgac tcttccgcgc caaagccttc acccatcccc gaccccgccct catcgccctt 420
accgctcgca agggacatcct gatagaagga gtaaccctcg tcaattcacc ggcctggacc 480
atcaatcctg tgatgtgcga gcgggtgacc atcgataagg tgactatcat caaccggccc 540
gactcgccca acaccgacgg gatcgacccc gattcctccc ggaacgtcta tatcactaac 600
tgctacattg acgtaggcga tgactgcac gccatcaaag cgggccgaga ggactccctt 660
tatcggaacg cttgtgaaaa cattgtcatc gccaaactgcc tcatgcgcca cggtcacggc 720
gggtgtgtca tcggcagcga gaccagcggg ggtattcgca aggtatcat taccactgac 780
atcttcgagg acaccgaccg gggcattaga cttaaagtcg ggcgcggacg cggcgggttc 840
gtcaggagacc tccggggcga gaattattatc atggaaaagg tgctctgtcc ctctgtcttc 900
aacatgtact atgataccgg gggaggcgtg atcgacgagc gcgcgcataga cttagaaccc 960
catccggtaa gcgaggctac accctccttc cgccgcctct ccttcagtca cattactgcc 1020
cgggaagtgc aggccgcgc ggccttcctc tacggcctgc ccgaacagcc tctggaggac 1080
gtctattttg acgatattct gatagagctg gccgcgacg cttctcctgc ccgtccggcc 1140
atgatgcggg ccgtcccgcc catgagccaa ggtggtgtgc tctgctacgg tgcgcggcgg 1200
atctccttcc ggcacatgca cctccgcggg caccgcggtc cggccttcca gatcgaacgc 1260
gcggaggcgg tgcagttgat gggctgctcg accgacggca gtgaagaccc ccagcttgct 1320
ttgggtcaag cggaggaggt caccatccgt gactgcacct ttaccgccca gcaggacccc 1380
gcaaaagaaa ggcaaaatta a 1401

```

<210> 32

<211> 466

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(348)

<223> Catalytic domain

<400> 32

```

Met Ile Asn Leu Tyr Gly Val Phe Asp Ile Arg Thr Phe Gly Ala Gln
 1      5      10      15
Pro Asp Gly Glu Thr Pro Ser Thr Ala Ala Ile Thr Ala Ala Ile Glu
 20      25      30
Thr Cys Ala Ala Ala Gly Gly Gly Val Val Tyr Ile Pro Ala Gly Arg
 35      40      45
Phe Leu Thr Gly Pro Leu Arg Leu Lys Ser His Val Arg Leu His Leu
 50      55      60
Glu Ala Gly Ala His Leu Leu Phe Ser Gln Asp Pro Ala Asp Tyr Pro
 65      70      75
Val Leu Glu Thr Arg Trp Glu Gly Lys Glu Val Leu Thr Tyr Ala His
 85      90      95
Gln Ile Tyr Gly Glu Asp Leu Glu Gly Val Ala Ile Thr Gly Arg Gly
100      105      110
Thr Ile Asp Gly Arg Gly Glu Thr Trp Trp Arg Leu Phe Arg Ala Lys
115      120      125
Ala Phe Thr His Pro Arg Pro Arg Leu Ile Ala Phe Thr Arg Cys Lys
130      135      140

```

```

Asp Ile Leu Ile Glu Gly Val Thr Leu Val Asn Ser Pro Ala Trp Thr
145      150      155      160
Ile Asn Pro Val Met Cys Glu Arg Val Thr Ile Asp Lys Val Thr Ile
165      170      175
Ile Asn Pro Pro Asp Ser Pro Asn Thr Asp Gly Ile Asp Pro Asp Ser
180      185      190
Ser Arg Asn Val Tyr Ile Thr Asn Cys Tyr Ile Asp Val Gly Asp Asp
195      200      205
Cys Ile Ala Ile Lys Ala Gly Arg Glu Asp Ser Leu Tyr Arg Thr Pro
210      215      220
Cys Glu Asn Ile Val Ile Ala Asn Cys Leu Met Arg His Gly His Gly
225      230      235      240
Gly Val Val Ile Gly Ser Glu Thr Ser Gly Gly Ile Arg Lys Val Val
245      250      255
Ile Thr Asn Cys Ile Phe Glu Asp Thr Asp Arg Gly Ile Arg Leu Lys
260      265      270
Ser Arg Arg Gly Arg Gly Gly Phe Val Glu Asp Leu Arg Ala Thr Asn
275      280      285
Ile Ile Met Glu Lys Val Leu Cys Pro Phe Val Leu Asn Met Tyr Tyr
290      295      300
Asp Thr Gly Gly Gly Val Ile Asp Glu Arg Ala His Asp Leu Glu Pro
305      310      315      320
His Pro Val Ser Glu Ala Thr Pro Ser Phe Arg Arg Leu Ser Phe Ser
325      330      335
His Ile Thr Ala Arg Glu Val Gln Ala Ala Ala Phe Leu Tyr Gly
340      345      350
Leu Pro Glu Gln Pro Leu Glu Asp Val Leu Phe Asp Asp Ile Trp Ile
355      360      365
Glu Leu Ala Ala Asp Ala Ser Pro Ala Arg Pro Ala Met Met Arg Ala
370      375      380
Val Pro Pro Met Ser Gln Gly Gly Val Leu Cys Tyr Gly Ala Arg Arg
385      390      395      400
Ile Ser Phe Arg His Met His Leu Arg Gly His Arg Gly Pro Ala Phe
405      410      415
Gln Ile Glu Arg Ala Glu Ala Val Gln Leu Met Gly Cys Ser Thr Asp
420      425      430
Gly Ser Glu Asp Pro Gln Leu Val Leu Gly Gln Ala Glu Glu Val Thr
435      440      445
Ile Arg Asp Cys Thr Phe Thr Ala Gln Gln Asp Pro Ala Lys Glu Arg
450      455      460
Gln Asn
465

```

<210> 33

<211> 1041

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 33

```

atgaaacttc gatgtctgat gctcaccctg cttctttgcg gcagcgccett cgcgcgcgac 60
cggattacgg cgcacaagat caacaacaag cccgactcct ggcttaccag cgacgaaggc 120
atcaagctga tcgacaacat catcacctgg cagaaccccg aggggtggctg ggccaagtac 180
tacgacgcga ccaatccgca caaacaaggc gaagtctacg gcgactggga cggcgctcggc 240
accatcgaca accggtacac ctacaccgag ctgaatctcc tggcgcaogt ctacaccctc 300
accaagcgcc cggagatcct cgattcgttc aacaagggcc tggagtttct gctcaaagcc 360
caatacccca gcggcggtg gccgcaacg tttcgggtgc ccaacaacta cgggcaagtgc 420

```

```

atcacgctca acgacaacgc gatggtgaac gtgatgcagt tcctgcagaa cgtcgcaaa 480
ggcaagggaag acttcgcttt cgtcgacgag cagcgctcgc ccaaagcgaa ggaggcgttt 540
gaccgcggga tcgactgcct tctgaagctc cagattaccg tgaacggcaa gcttaccgcc 600
tgggcccagc agtatgaccc gaagacactc gccgcggcgc ccgccgggc gtacgagctc 660
ccgggctcga cgggctcga aagcgcgccc gtcgtgcgct tgttcagtgc tttggagaac 720
cccagtcgcc aagttcagcg cgcggtccac gcggcgcgcg cttggtacga ggcgtcgaag 780
atcacggcca agaagctggt gcgcgagaac aacgacgtga cactggccga cgaccccaac 840
ggcgagccgc tttggggcgc cttctaogac atcgaaacca accgcccgtt ctattgcggt 900
cgcgacggcg tgaagaagtg gtcgctggac gagatcgagc ccgaacgcgc caagggctac 960
gcttgggtcc gccctgggc gacgagcgta ctggagcagt atcgcaagtg ggcgcggaag 1020
caccacccg tgaacagttg a 1041

```

<210> 34

<211> 346

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(18)

<221> DOMAIN

<222> (19)...(346)

<223> Catalytic domain

<400> 34

```

Met Lys Leu Arg Cys Leu Met Leu Thr Leu Leu Leu Cys Gly Ser Ala
 1          5          10          15
Phe Ala Ala Asp Arg Ile Thr Ala Asp Lys Ile Asn Asn Lys Pro Asp
 20          25          30
Ser Trp Leu Thr Ser Asp Glu Gly Ile Lys Leu Ile Asp Asn Ile Ile
 35          40          45
Thr Trp Gln Asn Pro Glu Gly Trp Ala Lys Tyr Tyr Asp Ala Thr
 50          55          60
Asn Pro His Lys Gln Gly Glu Val Tyr Gly Asp Trp Asp Gly Val Gly
 65          70          75          80
Thr Ile Asp Asn Gly Tyr Thr Tyr Thr Leu Asn Leu Leu Ala His
 85          90          95
Val Tyr Thr Leu Thr Lys Arg Pro Glu Ile Leu Asp Ser Phe Asn Lys
100          105          110
Gly Leu Glu Phe Leu Leu Lys Ala Gln Tyr Pro Ser Gly Gly Trp Pro
115          120          125
Gln Arg Phe Pro Val Pro Asn Asn Tyr Gly Lys Cys Ile Thr Leu Asn
130          135          140
Asp Asn Ala Met Val Asn Val Met Gln Phe Leu Gln Asn Val Ala Lys
145          150          155          160
Gly Lys Glu Asp Phe Ala Phe Val Asp Glu Gln Arg Arg Ala Lys Ala
165          170          175
Lys Glu Ala Phe Asp Arg Gly Ile Asp Cys Leu Leu Lys Leu Gln Ile
180          185          190
Thr Val Asn Gly Lys Leu Thr Ala Trp Ala Gln Gln Tyr Asp Pro Lys
195          200          205
Thr Leu Ala Ala Ala Pro Ala Arg Ala Tyr Glu Leu Pro Gly Leu Ser
210          215          220
Gly Cys Glu Ser Ala Pro Val Met Arg Leu Phe Met Ser Leu Glu Asn
225          230          235          240
Pro Ser Pro Glu Val Gln Arg Ala Val His Ala Ala Ala Ala Trp Tyr

```

```

          245          250          255
Glu Ala Ser Lys Ile Thr Gly Lys Lys Leu Val Arg Glu Asn Asn Asp
260          265          270
Val Thr Leu Ala Asp Asp Pro Asn Gly Glu Pro Leu Trp Ala Arg Phe
275          280          285
Tyr Asp Ile Glu Thr Asn Arg Pro Phe Tyr Cys Gly Arg Asp Gly Val
290          295          300
Lys Lys Trp Ser Leu Asp Glu Ile Glu Pro Glu Arg Arg Lys Gly Tyr
305          310          315          320
Ala Trp Val Arg Pro Trp Ala Thr Ser Val Leu Glu Gln Tyr Arg Lys
325          330          335
Trp Ala Ala Lys His Pro Pro Val Asn Ser
340          345

```

<210> 35

<211> 1071

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 35

```

atgccaaaaa attccgacga cgcgtggcgg gaaaagactc cgcccgattg gagtcttgtc    60
acatggagcg acgtattcaa acagaagcct ctctgggtacc aaaccgacga ggcgggtcga    120
gtcgcggacc aactcctcat ctatcaaaaa gagaacggcg ggtttgagaa gaatgtcgac    180
atggcgttga tgcgtacgca gaaggaaaaa gaagagctca ccgcaaaagcg gtcagacgtc    240
tccgaaaacga cgtacgacaa ccggaccacg tatcctcagg tcgcgtatct cggtcgagta    300
atcacccgcaa gccttcttaa accttcgccc ccggcggaatc ttccgaaata caaagacgcc    360
ttcaacaaag gtcttgatta cctgcttgcc tcccagtatg agaacggagg atttccgcaa    420
ttctatccgt tgaaaaaagg ctattacaca cacatcacct tcaacgacga cgcgatgatc    480
ggcgtcctga aggtgcttcg cgacatcgca aataagaaaag aggattacgt gttcgtggat    540
gaagcgcgaa gacttcgccc cgagcaagcg gtcgccaaag cgtgcctctt tattctgaag    600
cttcaggttg tcgtcgacgg aaagaaaacc gtctgggctg cgagtatga cgagactacg    660
ctggcgctcg cagcggctcg caagtgtgag cccgtgtcgt tgaccgctgg tgagagcgctc    720
ggcatcgtcc gatacctgat gcaggaaaaa ccgacgcccgg agatcaccca tgcgatcgag    780
tctgcgatcg attggtatcg aaagaacaag atcgacggaa tacgttggga ggcgatcaaa    840
ggcgagaaca cggttgtgaa agacaaatcg gctccccta tatgggcacg gttctatcag    900
atcgaaacga tgcgtccgat cttcatcgga cgtgattcgg ttatcaagta tgacgtgacg    960
caggtcgaa cggagcgtcg gaatgggttac gcctgggtacg tcaccgcacc gaatgaattg   1020
gtgaacgagg attatttgaa gtggaagggg aaaagcgccc gagccaagta g    1071

```

<210> 36

<211> 356

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(356)

<223> Catalytic domain

<400> 36

```

Met Pro Lys Asn Ser Asp Asp Ala Trp Arg Glu Lys Thr Pro Pro Asp
 1      5      10      15
Trp Ser Leu Val Thr Trp Ser Asp Val Phe Lys Gln Lys Pro Leu Trp
      20      25      30

Tyr Gln Thr Asp Glu Ala Ala Arg Val Ala Asp Gln Leu Leu Ile Tyr
 35      40      45
Gln Lys Glu Asn Gly Gly Phe Glu Lys Asn Val Asp Met Ala Leu Met
 50      55      60
Leu Thr Gln Lys Glu Lys Glu Glu Leu Thr Ala Lys Arg Ser Asp Val
 65      70      75      80
Ser Glu Thr Thr Ile Asp Asn Arg Thr Thr Tyr Pro Gln Val Ala Tyr
 85      90      95
Leu Gly Arg Val Ile Thr Ala Ser Leu Leu Lys Pro Ser Pro Pro Ala
100      105      110
Asn Leu Pro Lys Tyr Lys Asp Ala Phe Asn Lys Gly Leu Asp Tyr Leu
115      120      125
Leu Ala Ser Gln Tyr Glu Asn Gly Gly Phe Pro Gln Phe Tyr Pro Leu
130      135      140
Lys Lys Gly Tyr Tyr Thr His Ile Thr Phe Asn Asp Asp Ala Met Ile
145      150      155      160
Gly Val Leu Lys Val Leu Arg Asp Ile Ala Asn Lys Lys Glu Asp Tyr
165      170      175
Val Phe Val Asp Glu Ala Arg Arg Leu Arg Ala Glu Gln Ala Val Ala
180      185      190
Lys Ala Leu Pro Leu Ile Leu Lys Leu Gln Val Val Val Asp Gly Lys
195      200      205
Lys Thr Val Trp Ala Ala Gln Tyr Asp Glu Thr Thr Leu Ala Pro Ala
210      215      220
Ala Ala Arg Lys Phe Glu Pro Val Ser Leu Thr Ala Gly Glu Ser Val
225      230      235      240
Gly Ile Val Arg Tyr Leu Met Gln Glu Lys Pro Thr Pro Glu Ile Thr
245      250      255
Asp Ala Ile Glu Ser Ala Ile Asp Trp Tyr Arg Lys Asn Lys Ile Asp
260      265      270
Gly Ile Arg Trp Glu Arg Ile Lys Gly Glu Asn Thr Val Val Lys Asp
275      280      285
Lys Ser Ala Pro Pro Ile Trp Ala Arg Phe Tyr Gln Ile Glu Thr Met
290      295      300
Arg Pro Ile Phe Ile Gly Arg Asp Ser Val Ile Lys Tyr Asp Val Thr
305      310      315      320
Gln Val Glu Ala Glu Arg Arg Asn Gly Tyr Ala Trp Tyr Val Thr Ala
325      330      335
Pro Asn Glu Leu Val Asn Glu Asp Tyr Leu Lys Trp Lys Gly Lys Ser
340      345      350
Ala Gly Ala Lys
355

```

<210> 37

<211> 1860

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 37

```

atgttacta ctactggctc tcattgcgcc cgggaattccg cgcgtttttc cottactcgc      60
atagcagccg ctgttgcgtt gatggcaggc acttcagcat ttgcagctgc gacgggtggc      120
ttctctacca ctgatgggtg caacgtatcg ggcgcccggt cgtttactgc atcgacttac      180
cagcaaatca acaccattat tgccaacgca aaactggatg atgcaggtaa aaaagtcact      240
gggggtgctt acccgcttat cattacctac accggtaatg aagactcgcg gattaaccag      300
atgatcaaa accacacggt gaattcatcg ggcaactgcc ctaacccgcg ttggagcgaa      360
gcctatcgct acgtggaagt taaagagttt accaagggtt ttaccattca aggcgcgaat      420

```

```

ggttcttcag caaacttcgg cattgtgatt aataaatctg acaatgtgat tgtgcgtaat      480
atgaaaatcg gtgcgcttgc tgggtcggagt aacgatgcgg atatgattcg tatcgacacc      540
ggcgtaaacg tgtggattga tcacaaggaa ttgtttgcgg taaataatga atgtaaaagt      600
tcaccocgat gtgctgtcac atttgaaagt gcgattgata ttaaaaaagc atcgcaaaat      660
attacggtgt cctacaacat tatccgcgat agtaaaaaag tagggctcga tggttcgagt      720
agcagtgata ttgcaggtgg ccgtaagatt acgttccatc acaatattta tcgcaatgtt      780
gggtgcacgtt taccgttgca acgcggtggg tggacacaca tgtataacaa tctttacgac      840
ggagttacca gctcgggtat taacgttcgt caaggtggct acgcgctaag cgagaacaac      900
tggttccaaa atgctgtcaa ccgcggtacc tgccgttttg acagtagtaa ctgcggttac      960
tgggactcgc gcaacaacaa cgtgcgcaac cctggtgatt tctccacctt caacattacc      1020
tggaccagcg tgggcaccat cgacgccacc aactggacta ccactcaacc tttcccgatt      1080
agcattccct acagctactc gcctgttagc ccgcagtgtg tcaaagacaa gttggcaaat      1140
tatgctgggt tcggtaaaaa caatgcgcaa ttaacggcgt ctgcgtgcag cggaaatact      1200
tcacggtagt caccttcacg agtgccagca tcacggcgcg caccctcaag ccgttcaccc      1260
agcagtgcag cgccatccag cacaccaact acatcaagct cgagttcagt tgccgcaacc      1320
ggttcaattt cgctcgggtc aacggcaacc aacaacagca ttgtgttgag ttggtcacc      1380
aacaatgga cgctcgggtc gcaagaagtg tatcgcgata ccgacgctga tccatcgggg      1440
cgtgtgcgta tcgcatccct ggctgcttca gcgcgtatgt ataccgatag cacagcgga      1500
tcgggcaaaa cctattacta ctggattaaa aataccactt ctggtgttgt caccaattcc      1560
aatgctgcat cagcgcgtat tggtagcacg gcgtccagtt ctggtgcacg aagcagctca      1620
agttcaagcg gcggcgcgcc cgtattaggt ggtactggtg attatccaag cggcttctcc      1680
aagtgcgctg atttggcgcg gacttgttca gtgtcatcgg gcgatggctg ggttgcgttt      1740
ggtcgcaaa gcaagtgggt taccagaaga gtatcggtag gtatgtcaat cgctgtacc      1800
gttcggcgat ttggttcgga tccacagggc aaccctaaca agtgttctta caaacgttaa      1860

```

<210> 38

<211> 619

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(35)

<221> DOMAIN

<222> (36)...(387)

<223> Catalytic domain

<400> 38

```

Met Phe Thr Thr Gly Ser His Cys Ala Arg Asn Ser Ala Arg Phe
 1      5      10      15
Ser Leu Thr Ala Ile Ala Ala Ala Val Ala Leu Met Ala Gly Thr Ser
 20      25      30
Ala Phe Ala Ala Ala Thr Gly Gly Phe Ser Thr Thr Asp Gly Gly Asn
 35      40      45
Val Ser Gly Ala Arg Ser Phe Thr Ala Ser Thr Tyr Gln Gln Ile Asn
 50      55      60
Thr Ile Ile Ala Asn Ala Lys Leu Asp Asp Ala Gly Lys Lys Val Thr
 65      70      75      80
Gly Gly Ala Tyr Pro Leu Ile Ile Thr Tyr Thr Gly Asn Glu Asp Ser
 85      90      95
Leu Ile Asn Gln Met Ile Lys Asp His Thr Val Asn Ser Ser Gly Asn
100     105     110
Cys Pro Asn Pro Arg Trp Ser Glu Ala Tyr Arg Tyr Val Glu Ile Lys
115     120     125
Glu Phe Thr Lys Gly Ile Thr Ile Gln Gly Ala Asn Gly Ser Ser Ala
130     135     140

```

Asn Phe Gly Ile Val Ile Asn Lys Ser Asp Asn Val Ile Val Arg Asn
 145 150 155 160
 Met Lys Ile Gly Ala Leu Ala Gly Ala Ser Asn Asp Ala Asp Met Ile
 165 170 175
 Arg Ile Asp Thr Gly Val Asn Val Trp Ile Asp His Asn Glu Leu Phe
 180 185 190
 Ala Val Asn Asn Glu Cys Lys Gly Ser Pro Asp Gly Asp Leu Thr Phe
 195 200 205
 Glu Ser Ala Ile Asp Ile Lys Lys Ala Ser Gln Asn Ile Thr Val Ser
 210 215 220
 Tyr Asn Ile Ile Arg Asp Ser Lys Lys Val Gly Leu Asp Gly Ser Ser
 225 230 235 240
 Ser Ser Asp Ile Ala Gly Gly Arg Lys Ile Thr Phe His His Asn Ile
 245 250 255
 Tyr Arg Asn Val Gly Ala Arg Leu Pro Leu Gln Arg Gly Gly Trp Thr
 260 265 270
 His Met Tyr Asn Asn Leu Tyr Asp Gly Val Thr Ser Ser Gly Ile Asn
 275 280 285
 Val Arg Gln Gly Gly Tyr Ala Leu Ile Glu Asn Asn Trp Phe Gln Asn
 290 295 300
 Ala Val Asn Pro Val Thr Cys Arg Phe Asp Ser Ser Asn Cys Gly Tyr
 305 310 315 320
 Trp Asp Leu Arg Asn Asn Val Arg Asn Pro Gly Asp Phe Ser Thr
 325 330 335
 Tyr Asn Ile Thr Trp Thr Ser Gly Gly Thr Ile Asp Ala Thr Asn Trp
 340 345 350
 Thr Thr Thr Gln Pro Phe Pro Ile Ser Ile Pro Tyr Ser Tyr Ser Pro
 355 360 365
 Val Ser Pro Gln Cys Val Lys Asp Lys Leu Ala Asn Tyr Ala Gly Val
 370 375 380
 Gly Lys Asn Asn Ala Gln Leu Thr Ala Ser Ala Cys Ser Gly Asn Thr
 385 390 395 400
 Ser Ser Val Ala Pro Ser Ser Val Pro Ala Ser Ser Ala Ala Pro Ser
 405 410 415
 Ser Arg Ser Ser Ser Ser Ala Ala Pro Ser Ser Thr Pro Thr Thr Ser
 420 425 430
 Ser Ser Ser Ser Val Ala Ala Thr Gly Ser Ile Ser Leu Gly Ala Thr
 435 440 445
 Ala Thr Asn Asn Ser Ile Val Leu Ser Trp Ser Pro Asn Asn Val Thr
 450 455 460
 Leu Gly Ser Gln Glu Val Tyr Arg Asp Thr Asp Ala Asp Pro Ser Gly
 465 470 475 480
 Arg Val Arg Ile Ala Ser Leu Ala Ala Ser Ala Arg Met Tyr Thr Asp
 485 490 495
 Ser Thr Ala Ala Ser Gly Gln Thr Tyr Tyr Trp Ile Lys Asn Thr
 500 505 510
 Thr Ser Gly Val Val Thr Asn Ser Asn Ala Ala Ser Ala Arg Ile Gly
 515 520 525
 Ser Thr Ala Ser Ser Ser Val Ala Ser Ser Ser Ser Ser Ser Gly
 530 535 540
 Gly Ala Pro Val Leu Gly Gly Thr Gly Asp Tyr Pro Ser Gly Phe Ser
 545 550 555 560
 Lys Cys Ala Asp Leu Gly Gly Thr Cys Ser Val Ser Ser Gly Asp Gly
 565 570 575
 Trp Val Ala Phe Gly Arg Lys Gly Lys Trp Val Thr Lys Lys Val Ser
 580 585 590
 Val Gly Ser Ser Ile Ala Cys Thr Val Ala Ala Phe Gly Ser Asp Pro
 595 600 605
 Gln Gly Asn Pro Asn Lys Cys Ser Tyr Lys Arg

610

615

<210> 39

<211> 1077

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 39

atggcgccga	tctccgacc	caacctcctt	tgcacttacg	cgctctgcat	ggccttgctc	60
gccgtggtga	gctgcgcggc	ggggccgggtg	tcagcgcagc	agccggcgcc	atggagcacg	120
gccatcgtgg	agcaggagga	gagcgcgttc	gcctcccgt	cgatgcgcag	cgtcgccgac	180
aacgtcgtgc	gccatcagtc	ggccgaaggc	ggctggccta	agaacaccaa	tctggcggcg	240
ccgccatcgg	ggccggcgcc	ggaggcgctc	gccaatcaga	tcgacaatga	tcgcacgacg	300
ctgccgatgg	agtttctggc	gcgtgtgata	cacgcggcgc	gcgtccgata	caagccggcc	360
ttcgagcgcg	ggctggatta	tctgcttgcg	gctcagtacg	cgaacggcgg	ctggccgcag	420
ttctatccgc	tgcgcggggg	ctattacgat	cacgtgacgt	tcaacgacga	cgccatgata	480
cggtgatga	ttctgtctcg	cgcagtggcg	cgcggcgggg	cgccctatga	atttgtcgac	540
gccgggcggc	gcgcgcgcgc	tgacagccgc	gtcagcggg	gcctggcgct	catcctgcgc	600
acgcagatcc	ggcaggcgcg	ggcgctgacg	gtctggtgcg	cgcagtatga	cagcggccacc	660
ttgcagcccg	cctgggcgcg	cgctatgag	cgcgcgtccc	tgtccggcgc	ggaaagtgtg	720
gggatcgtgc	gctatctcat	gtcgatcgac	catccctcgc	ccgaagtcgt	cgccgcgcgtc	780
gacggcgctg	tggcatggct	gcgcgcggcc	gccattgccc	gcgtgcgcgt	ggagaatttc	840
acggacgccg	acggccgcgc	tgaccgcgcg	gcctggcccg	acgcggggcg	gccgcgatac	900
tgggcgcggt	tctacagatt	cggcgccaac	cggccgatct	tcttggggcg	tgattccgtt	960
tttcactaca	cgttcggaga	aatcgagcgc	gagcgcgcgc	caggctacaa	ttattacgga	1020
tactgggcgc	gctccgtgct	ggaagactat	cgcgcctggc	gcgcgcgcgt	gcgatga	1077

<210> 40

<211> 358

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(32)

<221> DOMAIN

<222> (33)...(358)

<223> Catalytic domain

<400> 40

```

Met Ala Pro Ile Leu Arg Pro Asn Leu Leu Cys Thr Tyr Ala Leu Cys
 1      5      10      15
Met Gly Leu Leu Ala Val Val Ser Cys Ala Ala Gly Pro Val Ser Ala
 20      25      30
Gln Gln Pro Ala Pro Trp Ser Thr Ala Ile Val Glu Gln Glu Glu Ser
 35      40      45
Ala Phe Ala Ser Pro Ser Met Arg Ser Val Ala Asp Asn Val Val Arg
 50      55      60
His Gln Ser Ala Glu Gly Gly Trp Pro Lys Asn Thr Asn Leu Ala Ala
 65      70      75      80
Pro Pro Ser Gly Pro Ala Pro Glu Gly Val Ala Asn Thr Ile Asp Asn
 85      90      95
Asp Ala Thr Thr Leu Pro Met Glu Phe Leu Ala Arg Val Ile His Ala

```

```

      100      105      110
Gly Gly Val Arg Tyr Lys Pro Ala Phe Glu Arg Gly Leu Asp Tyr Leu
 115      120      125
Leu Ala Ala Gln Tyr Ala Asn Gly Gly Trp Pro Gln Phe Tyr Pro Leu
 130      135      140
Arg Gly Gly Tyr Tyr Asp His Val Thr Phe Asn Asp Asp Ala Met Ile
 145      150      155      160
Arg Val Met Ile Leu Leu Gly Ala Val Ala Arg Gly Gly Ala Pro Tyr
 165      170      175
Glu Phe Val Asp Ala Gly Arg Arg Ala Arg Ala Ala Ala Val Glu
 180      185      190
Arg Gly Leu Ala Leu Ile Leu Arg Thr Gln Ile Arg Gln Gly Gly Ala
 195      200      205
Leu Thr Val Trp Cys Ala Gln Tyr Asp Ser Ala Thr Leu Gln Pro Ala
 210      215      220
Trp Ala Arg Ala Tyr Glu Pro Pro Ser Leu Ser Gly Ala Glu Ser Val
 225      230      235      240
Gly Ile Val Arg Tyr Leu Met Ser Ile Asp His Pro Ser Pro Glu Val
 245      250      255
Val Ala Ala Val Asp Gly Ala Val Ala Trp Leu Arg Ala Ala Ala Ile
 260      265      270
Ala Gly Val Arg Val Glu Asn Phe Thr Asp Ala Asp Gly Arg Pro Asp
 275      280      285
Arg Arg Ala Val Ala Asp Ala Gly Ala Pro Pro Ile Trp Ala Arg Phe
 290      295      300
Tyr Glu Phe Gly Ala Asn Arg Pro Ile Phe Leu Gly Arg Asp Ser Val
 305      310      315      320
Phe His Tyr Thr Phe Gly Glu Ile Glu Arg Glu Arg Arg Ala Gly Tyr
 325      330      335
Asn Tyr Tyr Gly Tyr Trp Ala Arg Ser Val Leu Glu Asp Tyr Pro Ala
 340      345      350
Trp Arg Ala Arg Val Arg
 355

```

<210> 41

<211> 1080

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 41

```

atgaaaaatt taaaatacag tttagtttca tttgtactac tcattactat gaatgttttt 60
acgcaagaaa aaaaagtaac ttggaaaagc atcacagaaa ataacgatga aaattgggtt 120
gtaagcgaaag aagccaaaaa aatagccgaa aatgttttgt tataatcaacg cgatattggt 180
ggttggccaa aaaaactga aattcaaaat gaactttcag aaaaagaaaa actaacatta 240
aaagaattaa aatcggtacc aaaaggatgt accatcgaca atgggtgcaac gtgtcaggaa 300
ttacttttct tatccaaaat atataaatcc aatccagatg agcgatataa aatggctttc 360
ttaaagggtg tgatttaact gattacagct caatacaaaa atgggtggtg gccacaatat 420
taccctttga gagaaggata ttacactcat attacttaca acgataatgc aatgggtgaat 480
gttttaaaagt tgttgaaga agttaaagat aaactctgatt actactcaat tcaagcacc 540
gatgaaattt ccaaaatggc tgaagtatca tttataaaag gagtcgattg catattaaaa 600
acacagtaca aacaaaatgg aatattaacc gcttgggtgt cacaacatga cagggaaaca 660
ttgaaacctg ctaagcaag agcttatgaa ttgccttcgt taagcggaaa agaatacagc 720
aaaattgtgt tgttattaat gtcaatcgaa aatccatcta aagaagtaat tactgccgta 780
aattcagcag ttaattggtt tgaaaaaaca aaatcaacg gaattaaaat tgaaccatt 840
tccaccggga aaaaggatga aaaagataga attgtgtgtg aaagtccgta tgcctccg 900
ctttgggcaa gatttatgga attaagtgc aacaaacct tttttgtga tctgacgga 960

```

```

aagaaaaaat acagcatgtc agaaattagt caagagcgta gaaccggcta tgcattgtac 1020
accaacgaac caaaagaagt tttaaaaaaa tacgatgatt ggaagtcac attaaaaata 1080

```

<210> 42

<211> 359

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(21)

<221> DOMAIN

<222> (22)...(359)

<223> Catalytic domain

<400> 42

```

Met Lys Asn Leu Lys Tyr Ser Leu Val Ser Phe Val Leu Leu Ile Thr
 1           5           10           15
Met Asn Val Phe Thr Gln Glu Lys Lys Val Thr Trp Lys Ser Ile Thr
 20           25           30
Glu Asn Asn Asp Glu Asn Trp Phe Val Ser Glu Glu Ala Lys Lys Ile
 35           40           45
Ala Glu Asn Val Leu Leu Tyr Gln Arg Asp Ile Gly Gly Trp Pro Lys
 50           55           60
Asn Thr Glu Ile Gln Asn Glu Leu Ser Glu Lys Glu Lys Leu Thr Leu
 65           70           75           80
Lys Glu Leu Lys Ser Asp Pro Lys Gly Cys Thr Ile Asp Asn Gly Ala
 85           90           95
Thr Cys Gln Glu Leu Leu Phe Leu Ser Lys Ile Tyr Lys Ser Asn Pro
100           105           110
Asp Glu Arg Tyr Lys Met Ala Phe Leu Lys Gly Val Ile Tyr Leu Ile
115           120           125
Thr Ala Gln Tyr Lys Asn Gly Gly Trp Pro Gln Tyr Tyr Pro Leu Arg
130           135           140
Glu Gly Tyr Tyr Thr His Ile Thr Tyr Asn Asp Asn Ala Met Val Asn
145           150           155           160
Val Leu Lys Leu Leu Lys Glu Val Lys Asp Lys Ser Asp Tyr Tyr Ser
165           170           175
Ile Gln Ala Pro Asp Glu Ile Ser Lys Met Ala Glu Val Ser Phe Asn
180           185           190
Lys Gly Val Asp Cys Ile Leu Lys Thr Gln Tyr Lys Gln Asn Gly Ile
195           200           205
Leu Thr Ala Trp Cys Ala Gln His Asp Arg Glu Thr Leu Lys Pro Ala
210           215           220
Lys Ala Arg Ala Tyr Glu Leu Pro Ser Leu Ser Gly Lys Glu Ser Ala
225           230           235           240
Lys Ile Val Leu Leu Leu Met Ser Ile Glu Asn Pro Ser Lys Glu Val
245           250           255
Ile Thr Ala Val Asn Ser Ala Val Asn Trp Phe Glu Lys Thr Lys Ile
260           265           270
Asn Gly Ile Lys Ile Glu Thr Ile Ser Thr Gly Lys Lys Asp Glu Lys
275           280           285
Asp Arg Ile Val Val Glu Ser Pro Asp Ala Pro Pro Leu Trp Ala Arg
290           295           300
Phe Met Glu Leu Ser Asp Asn Lys Pro Phe Phe Cys Asp Arg Asp Gly
305           310           315           320

Lys Lys Lys Tyr Ser Met Ser Glu Ile Ser Gln Glu Arg Arg Thr Gly
325           330           335
Tyr Ala Trp Tyr Thr Asn Glu Pro Lys Glu Val Leu Lys Lys Tyr Asp
340           345           350
Asp Trp Lys Ser Ser Leu Asn
355

```

<210> 43

<211> 1902

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 43

```

gtggatccaa agaattgggg .cagcggattt accggcgaaa tcaaagtaac taacaacaca      60
agccaaacag tcaatagctg gtctgtgtca tggcaagagg caggagccag tgtaactaat      120
tcctggaatg caaccttggg agggacgaat ccttataccg caaccgggtt aggatggaac      180
tcaaccctgg cgcccgagc ctctgccagt tttggtttcc aagcaaacgg cactgcgggg      240
gcaccaaaag taaatggcag tttgtgtggt gcgactgcat catctgcagc gaccagcaaa      300
tcagtgcgga gtgttgcgag ttcaaaagatt gcaagttcaa ttcaatcaag tgcaactagc      360
agttcaaaat cgtccagttc tgctgcacct tcaagcacgc caaaatccag tagctctgct      420
ccaacggctg catcattcac tattcaagaa gagcaagcgg gttttgccc tgtagacggt      480
attgcaacgg aaagtaccaa caccggattc accggcaacg gctacacca ttccaataat      540
gtacaagggt ctgcoattgt gtgggcggta aatgcaacta ccagtgcacg ccatacaatt      600
actttccgct tcgctaattg tggcactgcg aatcgcaatg gctcgctagt cattaacggc      660
ggcagcaatg gtaattacac ggtgcaatta ccacgcaccg cgagctgggc tgactggcaa      720
acagtaagtc tggaaattga tttggtacaa ggcaataaca atttgcaact caccgcattg      780
actgcagatg gcctcgcaaa tatcgacttc atcaaaattg aaggagcadc aaccaaaagc      840
ggaacctgtg caggtgcggt cagcagtagc agtgttgcc ctccggtaaa atccagtgtc      900
agcgcgccaa gcagttctgt accaacgaac accggcgcca tgctaacttt ggatggcaac      960
cctgccgcaa gctggcctaa caaatcgctt acaaagtgga gcgcacgcg cgctgacatt     1020
gttgctctct atcaacagtc caacggcggc tggcaaaaaa atctggatta caattcagtg     1080
agcgctggtt atggcggcag tgcaagcggc accatcgata atggtgcaac tattactgaa     1140
atggttttat tcgctgaggt ttacaaaacc ggaacaata ccaagtaccg cgatgcagtt     1200
cgccgtgcag caaactttat cgtgagttcg caatatagca ctggcgcggt gccgcaattt     1260
tatccgctca aaggtggcta tgcagaccac gccaccttta atgataacgg catggccttac     1320
gcattaactg tattggattt cgctgcaaac aagcgcgccg cttttgatac ggatgtcttt     1380
aatgacacag accgcgcaaa atttaaaaca gcggtaacca aaggtgttga ttacatttta     1440
aaagcgcaat ggaacaaaaa tggaaaatta acagcctggt gcgcacaaca tggcgcgact     1500
gactatcaac ctaaaaaagc acgcgcttat gaattggaat cactgagtg tagcgagtc     1560
gttggtgtaa ttgcattttt aatgacgcag cgcagacag cacaatatcca aacggccggt     1620
aaagcaggcc tcaactgggt caatagcccg agcaacctatt tggaaaggta cacctacgat     1680
tcattccaaag cgtccactaa tcccatagtg cagaaagcgg gaagtagaat gtggtatcgc     1740
ttttacgatt taaataccaa ccgtggtttt ttcagcgacc gggacggcag caaattctat     1800
gacattacca aaatgtctga agaacgtcgc acgggttata gttggggtg cgcttatggt     1860
gagagcatca tcgccttttg caaaaaagtg ggctatctat aa                               1902

```

<210> 44

<211> 633

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> BINDING

<222> (4)...(89)

<223> Carbohydrate binding module

<221> BINDING

<222> (152)...(275)

<223> Carbohydrate binding module

<221> DOMAIN

<222> (277)...(633)

<223> Catalytic domain

<400> 44

```

Met Asp Pro Lys Asn Trp Gly Ser Gly Phe Thr Gly Glu Ile Lys Val
1      5      10      15
Thr Asn Asn Thr Ser Gln Thr Val Asn Ser Trp Ser Val Ser Trp Gln
20      25      30
Glu Ala Gly Ala Ser Val Thr Asn Ser Trp Asn Ala Thr Leu Gly Gly
35      40      45
Thr Asn Pro Tyr Thr Ala Thr Gly Leu Gly Trp Asn Ser Thr Leu Ala
50      55      60
Pro Gly Ala Ser Ala Ser Phe Gly Phe Gln Ala Asn Gly Thr Ala Gly
65      70      75      80
Ala Pro Lys Val Asn Gly Ser Leu Cys Gly Ala Thr Ala Ser Ser Ala
85      90      95
Ala Thr Ser Lys Ser Ser Ala Ser Val Ala Ser Ser Lys Ile Ala Ser
100     105     110
Ser Ile Gln Ser Ser Ala Thr Ser Ser Ser Lys Ser Ser Ser Ala
115     120     125
Ala Pro Ser Ser Thr Pro Lys Ser Ser Ser Ala Pro Thr Ala Ala
130     135     140
Ser Phe Thr Ile Gln Glu Gln Ala Gly Phe Cys Arg Val Asp Gly
145     150     155     160
Ile Ala Thr Glu Ser Thr Asn Thr Gly Phe Thr Gly Asn Gly Tyr Thr
165     170     175
Asn Ser Asn Asn Val Gln Gly Ala Ala Ile Val Trp Ala Val Asn Ala
180     185     190
Thr Thr Ser Ala Arg His Thr Ile Thr Phe Arg Phe Ala Asn Gly Gly
195     200     205
Thr Ala Asn Arg Asn Gly Ser Leu Val Ile Asn Gly Gly Ser Asn Gly
210     215     220
Asn Tyr Thr Val Gln Leu Pro Arg Thr Ala Ser Trp Ala Asp Trp Gln
225     230     235     240
Thr Val Ser Leu Glu Ile Asp Leu Val Gln Gly Asn Asn Asn Leu Gln
245     250     255
Leu Thr Ala Leu Thr Ala Asp Gly Leu Ala Asn Ile Asp Phe Ile Lys
260     265     270
Ile Glu Gly Ala Ser Thr Lys Ala Gly Thr Cys Ala Gly Ala Val Ser
275     280     285
Ser Ser Ser Val Ala Ser Ser Val Lys Ser Ser Ala Ser Ala Ala Ser
290     295     300
Ser Ser Val Pro Thr Asn Thr Gly Ala Met Leu Thr Leu Asp Gly Asn
305     310     315     320
Pro Ala Ala Ser Trp Leu Asn Lys Ser Arg Thr Lys Trp Ser Ala Ser
325     330     335
Arg Ala Asp Ile Val Ala Ser Tyr Gln Gln Ser Asn Gly Gly Trp Pro
340     345     350
Lys Asn Leu Asp Tyr Asn Ser Val Ser Ala Gly Asn Gly Gly Ser Ala
355     360     365
Ser Gly Thr Ile Asp Asn Gly Ala Thr Ile Thr Glu Met Val Tyr Leu
370     375     380

```

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Ala Glu Val Tyr Lys Thr Gly Asn Asn Thr Lys Tyr Arg Asp Ala Val
385     390     395     400
Arg Arg Ala Ala Asn Phe Ile Val Ser Ser Gln Tyr Ser Thr Gly Ala
405     410     415
Leu Pro Gln Phe Tyr Pro Leu Lys Gly Gly Tyr Ala Asp His Ala Thr
420     425     430
Phe Asn Asp Asn Gly Met Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala
435     440     445
Ala Asn Lys Arg Ala Pro Phe Asp Thr Asp Val Phe Asn Asp Thr Asp
450     455     460
Arg Ala Lys Phe Lys Thr Ala Val Thr Lys Gly Val Asp Tyr Ile Leu
465     470     475     480
Lys Ala Gln Trp Lys Gln Asn Gly Lys Leu Thr Ala Trp Cys Ala Gln
485     490     495
His Gly Ala Thr Asp Tyr Gln Pro Lys Lys Ala Arg Ala Tyr Glu Leu
500     505     510
Glu Ser Leu Ser Gly Ser Glu Ser Val Gly Val Ile Ala Phe Leu Met
515     520     525
Thr Gln Pro Gln Thr Ala Gln Ile Gln Thr Ala Val Lys Ala Gly Leu
530     535     540
Asn Trp Phe Asn Ser Pro Ser Thr Tyr Leu Glu Gly Tyr Thr Tyr Asp
545     550     555     560
Ser Ser Lys Ala Ser Thr Asn Pro Ile Val Gln Lys Ala Gly Ser Arg
565     570     575
Met Trp Tyr Arg Phe Tyr Asp Leu Asn Thr Asn Arg Gly Phe Phe Ser
580     585     590
Asp Arg Asp Gly Ser Lys Phe Tyr Asp Ile Thr Lys Met Ser Glu Glu
595     600     605
Arg Arg Thr Gly Tyr Ser Trp Gly Gly Ala Tyr Gly Glu Ser Ile Ile
610     615     620
Ala Phe Gly Lys Lys Val Gly Tyr Leu
625     630

```

<210> 45

<211> 987

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 45

```

atgactagac ggcgccttcac cgcgggttatc tgtttcttcg cggccgtctg cgcgcacgcg      60
cagtcacacc tgcgctcgaa ggacgtgctc gagcagtcgg agggctggta ttccacgacc      120
gccgcgcacg tcgtcgccga caccgtgctg ctgtatcaac gtccatccgg tggatggccg      180
aaggacatcg acatgacggc gccgcggcgg gaccgcactc ctcccgccgg tccagacgcg      240
accatcgaca accggcgccac gaccacgcag atccgcctgc tcgctcgtgc gccctcgggc      300
gcaccggcgg ctgcgcgcca cacctacacg gcggcggggc ttccggggat cgattacctg      360
ctcgagcgcg agtatcccaa cggcggtctg ccgcagttct tcccccctgg caaggactat      420
tcgcgctacg tcacgttcaa cgcgcgcgag atgatgaacg tgatgttcct gctggacgag      480
gtctcggcgg gagatgcgcc gtccacgttc gtggacgaac aacgcgcgca ccgcgcgcgc      540
gctgccgtcg ccaagggggt ctccgtctac ctgaagtgcg aggtccggat cgacggggacg      600
ctgaccgcct ggtgcgcgca acacgaacgag atcacctggg caccgcgtcc gccgcgcacc      660
ttcgagcacg cgtcgctcag cggcaacgag tctgtcgcga tcgtgcgctt cctgatgacc      720
cgtccgcgca cgcacagcat cgtcgccgcg gtcgatgcgg cggtcgcctg gctcagacgc      780
gtccgcctcc ctgacggacg gtgggcccga ttctacgagt tcggtaccaa tcgtccgatc      840
ttctcggggc gagacagtgt cgtgcgctac aaactcgagg agatcgaaca ggaacgtcag      900
gagggctacg cgtggtacgg cacgtggccg aggcgccttg ttgagaagat gtaccctgca      960
tggaagtcgc ggtttccggg caagtag                                     987

```

<210> 46

<211> 328

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(20)

<221> DOMAIN

<222> (21)...(328)

<223> Catalytic domain

<400> 46

```

Met Thr Arg Arg Ala Phe Ile Ala Val Ile Cys Phe Phe Ala Ala Val
 1          5          10          15
Cys Ala His Ala Gln Ser Thr Val Arg Trp Lys Asp Val Leu Glu Gln
 20          25          30
Ser Glu Gly Trp Tyr Ser Thr Thr Ala Ala His Val Val Ala Asp Thr
 35          40          45
Val Leu Leu Tyr Gln Arg Pro Ser Gly Gly Trp Pro Lys Asp Ile Asp
 50          55          60
Met Thr Ala Pro Pro Ala Asp Arg Thr Pro Pro Ala Arg Pro Asp Ala
 65          70          75          80
Thr Ile Asp Asn Gly Ala Thr Thr Thr Gln Ile Arg Leu Leu Ala Arg
 85          90          95
Ala Ala Ser Gly Ala Pro Ala Ala Ala Ala His Thr Tyr Thr Ala Ala
100          105          110
Ala Leu Arg Gly Ile Asp Tyr Leu Leu Glu Ala Gln Tyr Pro Asn Gly
115          120          125
Gly Trp Pro Gln Phe Phe Pro Leu Arg Lys Asp Tyr Ser Arg Tyr Val
130          135          140
Thr Phe Asn Asp Asp Ala Met Met Asn Val Met Phe Leu Leu Asp Glu
145          150          155          160
Val Ser Ala Gly Asp Ala Pro Phe Thr Phe Val Asp Glu Gln Arg Arg
165          170          175
Asp Arg Ala Arg Ala Ala Val Ala Lys Gly Val Ser Val Ile Leu Lys
180          185          190
Ser Gln Val Arg Ile Asp Gly Thr Leu Thr Ala Trp Cys Ala Gln His
195          200          205
Asp Glu Ile Thr Leu Ala Pro Arg Pro Ala Arg Thr Phe Glu His Ala
210          215          220
Ser Leu Ser Gly Asn Glu Ser Val Ala Ile Val Arg Phe Leu Met Thr
225          230          235          240
Arg Pro Pro Thr Pro Ala Ile Val Ala Ala Val Asp Ala Val Ala
245          250          255
Trp Leu Arg Arg Val Arg Leu Pro Asp Gly Arg Trp Ala Arg Phe Tyr
260          265          270
Glu Phe Gly Thr Asn Arg Pro Ile Phe Ser Gly Arg Asp Ser Val Val
275          280          285
Arg Tyr Lys Leu Glu Glu Ile Glu Gln Glu Arg Gln Glu Gly Tyr Ala
290          295          300
Trp Tyr Gly Thr Trp Pro Arg Thr Leu Val Glu Lys Met Tyr Pro Ala
305          310          315          320
Trp Lys Ser Arg Leu Pro Gly Lys
325

```

<210> 47

<211> 1077

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 47

```

atgaaaaatt ttaaaaaat ttaggagcg ttacttatat ctgtaacgtt ttgtgtgcac 60
gggcaggtaa acaaaaaatc tggcgggct attacacagt ctaacgcaga tgcattggtt 120
gcattctgat gagctgcaca gattgcagat aatgtattac tctatcagcg caatgttggc 180
ggatggccta aaaaatttga aatgcaggaa ccgcttagtg aggccgacaa aaaaaagctg 240
atagatctta agtctacggc caaagaaagt actacagata atggggctac gtgtcaggaa 300
atggtattcc tctctaagat atataaaca aagcccgaag agaagtataa agaggctttt 360
ttaaaaggac ttaattattt gcttgaagca cagtataaaa atggtgggatg gccacagttc 420
taccctttaa aaaaaggtta ttatacccac attacctata atgacgattc tatggtaaac 480
attcttatga tottaaagaa tattaaggaa gatgccaact attacagtat tacgccaaagc 540
gataaagttt taaagcaggt atcgacagct tttagacagag gcattgactg cattctaaaa 600
acacagtaca agcaaaaggg tgtgcttaca agctgggtg cccagcacga tgaggttaca 660
ttagaacctg caaatgcaag ggcttttgag ttggcatcac taagtggtaa agaactgct 720
aaaataacgt tgttgctaag gtctgtaaaa aatccgtcta aagaggttgt tgctgctgta 780
gatgctgctg tggcgtggtt tgaaaaaaca aaaattgaag gcattaaagt agaagaagta 840
accggagctg atggcaaaaa ggatagggta gtagtacaaa gggctgatgc cgaaccattg 900
tgggcgcgtt ttatggaact ggataccaac aggccatttt tttagcagag ggcaggtata 960
aaaaaatatt cgcttgctga gataggtcat gaacgccgta acggatatgg ctggtacacc 1020
aacgaaccaa aagaagtttt aaagaaatac accaaatgga aaaaacagtct taaatag 1077

```

<210> 48

<211> 358

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(21)

<221> DOMAIN

<222> (22)...(358)

<223> Catalytic domain

<400> 48

```

Met Lys Asn Phe Lys Asn Ile Val Gly Ala Leu Leu Ile Ser Val Thr
1 5 10 15
Phe Cys Val His Gly Gln Val Asn Lys Lys Ser Trp Arg Ala Ile Thr
20 25 30
Gln Ser Asn Asp Asp Ala Trp Phe Ala Ser Asp Gly Ala Ala Gln Ile
35 40 45
Ala Asp Asn Val Leu Leu Tyr Gln Arg Asn Val Gly Gly Trp Pro Lys
50 55 60
Asn Ile Glu Met Gln Glu Pro Leu Ser Glu Ala Asp Lys Lys Lys Leu
65 70 75 80
Ile Asp Leu Lys Ser Thr Ala Lys Glu Ser Thr Thr Asp Asn Gly Ala
85 90 95
Thr Cys Gln Glu Met Val Phe Leu Ser Lys Ile Tyr Lys Gln Lys Pro
100 105 110

```

```

Glu Glu Lys Tyr Lys Glu Ala Phe Leu Lys Gly Leu Asn Tyr Leu Leu
115 120 125
Glu Ala Gln Tyr Lys Asn Gly Gly Trp Pro Gln Phe Tyr Pro Leu Lys
130 135 140
Lys Gly Tyr Tyr Thr His Ile Thr Tyr Asn Asp Ser Met Val Asn
145 150 155 160
Ile Leu Met Ile Leu Lys Asn Ile Lys Glu Asp Ala Asn Tyr Tyr Ser
165 170 175
Ile Thr Pro Ser Asp Lys Val Leu Lys Gln Val Ser Thr Ala Phe Asp
180 185 190
Arg Gly Ile Asp Cys Ile Leu Lys Thr Gln Tyr Lys Gln Lys Gly Val
195 200 205
Leu Thr Ser Trp Cys Ala Gln His Asp Glu Val Thr Leu Glu Pro Ala
210 215 220
Asn Ala Arg Ala Phe Glu Leu Ala Ser Leu Ser Gly Lys Glu Ser Ala
225 230 235 240
Lys Ile Thr Leu Leu Leu Met Ser Val Lys Asn Pro Ser Lys Glu Val
245 250 255
Val Ala Ala Val Asp Ala Ala Val Ala Trp Phe Glu Lys Thr Lys Ile
260 265 270
Glu Gly Ile Lys Val Glu Glu Val Thr Gly Ala Asp Gly Lys Lys Asp
275 280 285
Arg Val Val Val Gln Arg Ala Asp Ala Glu Pro Leu Trp Ala Arg Phe
290 295 300
Met Glu Leu Asp Thr Asn Arg Pro Phe Phe Cys Asp Arg Asp Gly Ile
305 310 315 320
Lys Lys Tyr Ser Leu Ala Glu Ile Gly His Glu Arg Arg Asn Gly Tyr
325 330 335
Gly Trp Tyr Thr Asn Glu Pro Lys Glu Val Leu Lys Lys Tyr Thr Lys
340 345 350
Trp Lys Asn Ser Leu Lys
355

```

<210> 49

<211> 1023

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 49

```

atgttaaagtt tcacgcggt atcagtggtt cataattact gcacaggcca gacagcgtcc 60
accaaaaaatt cagtgccga aaagatgctt cagtaccagt tgtaaatgg cgcctggccc 120
aaacagttgg tagacaaaag tgcgttgat tacagtcttc cattaacgaa agagcgcccta 180
cagcagatca agaaaaacaga tattgatcat gctacgctcg acaacagtgc gacaaccggg 240
gaaataactg aattgatcaa ggcttttaag gacactaaaa ataaggcata ttgactgct 300
gtagaaaagg ggattgcata tattttatcg gctcaatatg agaattggcg atttccacaa 360
tactacccaa ataaattata ctatagagct gagataacat acaacgatga tgcgatgac 420
aatgcattac tagtgcttta caaagtagcc aataagcgag aggggtttga ggctatcaat 480
cccatatttg tgtcaaaagc gcaaaaagca gttgaaaagg gtataacctg tatcctaaaa 540
acacaggtca tacaagacgg aaaaaggagt atttgggctg cgcaatacga tcagaacact 600
ttacaacctg ctcaggcaag aaagtgtgaa ccagcttcac tgagcacaaag tgaatctgtt 660
tccatcggtc gctttctcat gctacagcct gcaaccactg aaattaagca agcgatcgaa 720
catgcaatac aatggttcga acagcatgat attgaaggtt accgtttcga ccgcatacaa 780
gatagggtga ctggaaaata tcaacggcaa cttgtcgaag atcggacttc cagcatttgg 840
gcgcgatttt ataactcga agacaaccgc ccattgtttg gagatcgga caatacaatc 900
aaatacaact ttgaggaggt ttcagaggag cgtagaaatg gctatgcttg gttcggcaac 960
tgcccgaaa agctgatcca aaaggactat ccaaaatgga aaaaacaata caaaattaaa 1020

```

taa 1023

<210> 50

<211> 340

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(16)

<221> DOMAIN

<222> (17)...(340)

<223> Catalytic domain

<400> 50

```

Met Leu Ser Phe Ile Ala Val Ser Val Phe His Asn Tyr Cys Thr Gly
1      5      10      15
Gln Thr Ala Ser Thr Lys Asn Ser Val Ala Glu Lys Met Leu Gln Tyr
20      25      30
Gln Leu Ser Asn Gly Ala Trp Pro Lys Gln Leu Val Asp Lys Ser Val
35      40      45
Val Asp Tyr Ser Leu Pro Leu Thr Lys Glu Arg Leu Gln Gln Ile Lys
50      55      60
Lys Thr Asp Ile Asp His Ala Thr Leu Asp Asn Ser Ala Thr Thr Arg
65      70      75      80
Glu Ile Thr Glu Leu Ile Lys Ala Phe Lys Asp Thr Lys Asn Lys Ala
85      90      95
Tyr Leu Thr Ala Val Glu Lys Gly Ile Ala Tyr Ile Leu Ser Ala Gln
100     105     110
Tyr Glu Asn Gly Gly Phe Pro Gln Tyr Tyr Pro Asn Lys Leu Tyr Tyr
115     120     125
Arg Ala Glu Ile Thr Tyr Asn Asp Asp Ala Met Ile Asn Ala Leu Leu
130     135     140
Val Leu Tyr Lys Val Ala Asn Lys Arg Glu Gly Phe Glu Ala Ile Asn
145     150     155     160
Pro Ile Phe Val Ser Lys Ala Gln Lys Ala Val Glu Lys Gly Ile Thr
165     170     175
Cys Ile Leu Lys Thr Gln Val Ile Gln Asp Gly Lys Arg Ser Ile Trp
180     185     190
Ala Ala Gln Tyr Asp Gln Asn Thr Leu Gln Pro Ala Gln Ala Arg Lys
195     200     205
Phe Glu Pro Ala Ser Leu Ser Thr Ser Glu Ser Val Ser Ile Val Arg
210     215     220
Phe Leu Met Leu Gln Pro Ala Thr Thr Glu Ile Lys Gln Ala Ile Glu
225     230     235     240
His Ala Ile Gln Trp Phe Glu Gln His Asp Ile Glu Gly Tyr Arg Phe
245     250     255
Asp Arg Ile Gln Asp Arg Val Thr Gly Lys Tyr Gln Arg Gln Leu Val
260     265     270
Ala Asp Arg Thr Ser Thr Ile Trp Ala Arg Phe Tyr Asn Leu Glu Asp
275     280     285
Asn Arg Pro Leu Phe Gly Asp Arg Asp Asn Thr Ile Lys Tyr Asn Phe
290     295     300
Glu Glu Val Ser Glu Glu Arg Arg Asn Gly Tyr Ala Trp Phe Gly Asn
305     310     315     320
Trp Pro Glu Lys Leu Ile Gln Lys Asp Tyr Pro Lys Trp Lys Lys Gln

```

```

325      330      335
Tyr Lys Ile Lys
340

```

<210> 51

<211> 1131

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 51

```

gtgacgtggg atcagatcct tcgtcagcct gccgcctggt acggcgggtcc ggaagcgaga 60
cggatcgcca atctgggtcct gctgtaccag cgcgcgacgg ggggctggcc caagaacatc 120
gacatggcgc ggtcgttgtc tcgcgacgat cgcacgacgc tcgcggcgga acgggcccctc 180
accgactcga cgtatcgaca tggatcgacg acgacgcagt tgcggtttct cgcgatgggtg 240
cagcacgccc agcaggcacc cgtgcgcgac gccatcacgc acggcctgga ctatctgctg 300
aacgcgcaat actcgaacgg cggatggcgg cagtactttc cgctccgaga cgactactcg 360
cgtcacatca cgttcaacga cgcgcgatg atcaatgtaa tgacgggtgt acgcgatgtc 420
gcagaagctc gcatgccctt cgaaggatc gacgcgggtcc gtcgggacgg ggcgctgtc 480
gccatcacgc gtggcatcga cgtgattctc gggacgcaaa tcgcgctcgg ggaccgtctg 540
acgggctggt gccagcagca tgacgagcgc tccctcgccc ccaccaaggc tcgcgcctac 600
gagcaccatc cgtatcgccg caaggaaacg gtaaccatca cgcgttctct catgaccctc 660
gatcgcccca gtcagcagat catcgcgggc atcgaggcgg ctgtcagatg gttgcgcgtg 720
gcgacctgtg cgggtgtgag agttgagcgt cggccggacc cggcgagtcc gaccgatat 780
gacgtcgtcg cgcgcgggga tgccgcgcga cctccgacct gggcacgggt ctacgagatc 840
ggcacgaacc gcccaatggt ttccggccgc gacggcgtga tcagattccg gctcgcggac 900
atcgagattg agcgcgcac cggctacagc tggatggcgg actatgccgc gaggttgctg 960
aacgaggagt atccggcgtg ggcgaggcta cggcggcgga gctttcagaa cgccgagctg 1020
cacaaggagt ccggtgaagt cgtacacacg gcgatcgtgc acgatcttgc cttccttgat 1080
gtcgaagaca aagaccagcc gcagccgaaa gtgcttttcg ctgggcggta g 1131

```

<210> 52

<211> 376

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(376)

<223> Catalytic domain

<400> 52

```

Met Thr Trp Asp Gln Ile Leu Arg Gln Pro Ala Ala Trp Tyr Gly Gly
1      5      10      15
Pro Glu Ala Arg Arg Ile Ala Asn Leu Val Leu Leu Tyr Gln Arg Ala
20     25     30
Thr Gly Gly Trp Pro Lys Asn Ile Asp Met Ala Arg Ser Leu Ser Pro
35     40     45
Asp Asp Arg Thr Thr Leu Ala Ala Glu Arg Ala Leu Thr Asp Ser Thr
50     55     60
Ile Asp Asn Gly Ser Thr Thr Gln Leu Arg Phe Leu Ala Met Val
65     70     75     80
Gln His Ala Gln Gln Ala Pro Val Arg Asp Ala Ile Thr His Gly Leu
85     90     95
Asp Tyr Leu Leu Asn Ala Gln Tyr Ser Asn Gly Gly Trp Pro Gln Tyr

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100      105      110
Phe Pro Leu Arg Asp Asp Tyr Ser Arg His Ile Thr Phe Asn Asp Asp
115     120     125
Ala Met Ile Asn Val Met Thr Val Leu Arg Asp Val Ala Glu Ala Arg
130     135     140
Met Pro Phe Glu Gly Ile Asp Ala Val Arg Arg Asp Arg Ala Arg Val
145     150     155     160
Ala Ile Thr Arg Gly Ile Asp Val Ile Leu Gly Thr Gln Ile Arg Val
165     170     175
Gly Asp Arg Leu Thr Gly Trp Cys Gln Gln His Asp Glu Arg Ser Leu
180     185     190
Ala Pro Thr Lys Ala Arg Ala Tyr Glu His Pro Ser Ile Ala Ser Lys
195     200     205
Glu Thr Val Thr Ile Thr Arg Phe Leu Met Thr Leu Asp Arg Pro Ser
210     215     220
Gln Gln Ile Ile Ala Ala Ile Glu Ala Ala Val Glu Trp Leu Arg Val
225     230     235     240
Ala Thr Leu Ser Gly Val Arg Val Glu Arg Arg Pro Asp Pro Ala Ser
245     250     255
Pro Thr Gly Tyr Asp Val Val Ala Ala Pro Asp Ala Ala Pro Pro
260     265     270
Thr Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg Pro Met Phe Ser
275     280     285
Gly Arg Asp Gly Val Ile Arg Phe Arg Leu Ala Asp Ile Glu Ile Glu
290     295     300
Arg Arg Thr Gly Tyr Ser Trp Met Gly Asp Tyr Ala Ala Arg Leu Leu
305     310     315     320
Asn Glu Glu Tyr Pro Ala Trp Ala Arg Leu Arg Arg Ala Ser Phe Gln
325     330     335
Asn Ala Glu Leu His Lys Glu Ser Gly Glu Val Val His Thr Ala Ile
340     345     350
Val His Asp Leu Ala Phe Leu Asp Val Glu Asp Lys Asp Gln Pro Gln
355     360     365
Pro Lys Val Leu Phe Ala Gly Arg
370     375

```

<210> 53

<211> 1977

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 53

```

atgaataact caacaaaaa aatgattcgg ccaactcaagg catcttttgc cttgggcgct 60
ctcgcaactg caatcgcatc accctcatgg gcggcttgct cttacagcgt aaccaataat 120
tggggctctg gctttaccgg agaataataa gtaaccaacg atacaacatc gactgtaaat 180
aattggtctg tgtcttggca ggaatcaggc gtgaccgtca ctaacgcacg gaatgcaaca 240
ctgagcggat caaatcotta taccgcaaca tcaactcgggt ggaacggaac tctcgtcca 300
aaagcttcag caagttttgg ttttcaagca aatggaacag cgggcgcacc gaaagtaaat 360
ggaaccttgt gtggtaccag cacatcatca acaggtacat cctcagttgc acctcatcc 420
gtagcgagta gcgttgctgt atcaagcagt aaatcatcaa gctctgttgc aaccatcagt 480
agctctaaat ccagcagcag tgtgccgaca gtttcatcat tcactattca ggaagaccaa 540
gccggtttct gccgtgtaga tggcattgca actgaaagta ctaacactgg ctatacaggt 600
aatggctaca ccaacaccac taatgcgcaa ggcgctgcaa ttgaatgggc aattaatgct 660
cccaacagca gccgctacac cctcaccttc cgttatgcca atgctgtgtac cgtaatcgc 720
aatggttcgt tattaattaa cgacggaagc aatggtaact acacagtgca attgccaaagt 780
accggcgcat gggcaacctg gcaaacctgc agtggtgaag tggatttggg gcaaggcaat 840

```

```

aatattttga aactcgcttc gcttactgct gatggccttg cgaatataga ttcattaaaa 900
attgaaggcg cacaagccaa agctgggtga tgcagcacta cggtaagtag cagctcttctg 960
tcaattaaat caagttccag ttcattcatcg tccagctcaa ctgcagcagt aaaaacatta 1020
acactggatg gtaaccctgc tgcaaacctg ttaataaat ccagaaccaa gtggaatgtc 1080
agcagagctg acatcgctact ttcgtatcag caatcaaatg gtggctggcc aaaaatttg 1140
gactacaact cggtaggctc aggtaatggt ggtagcgaca ggggcactat tgataatggt 1200
gcaaccataa ccgaaatggt gtacotcgtc gaagtgtata aaaaatggcg gaataccaaa 1260
taccgcgacg ccgtgcgcag agcagcgaat tttattgtga gttcacata cagcactggt 1320
gctttaccgc agttttatcc gctgaaaggt ggttacgcag atcacgctac ctttaatgat 1380
aatggtatgg cttacgcgtt gactgttctg gatttcgcgg taaataaacg cgcgccattt 1440
gataacgata ttttctctga ctctgaccgc agcaaattta aaactgctgt taccaaaggg 1500
gtcgattaca tattaaaagc gcaatggaaa cagaatggaa aattaaccgt atggtgcgca 1560
caacacgggt ctaatgatta tcaaccgaaa aaagcgcgtg cttacgagtt agaatacttg 1620
agtgttagtg aatctgtcgg tgtactcgtc ttcttaatga ctcaaccaca aaccacgcaa 1680
attgaagcag ctgtgcgtgc aggtgtggcc tggtttaata gcccaagcac ctacttgaat 1740
aattacactt acgattcttc caaagcttcg accaatccaa tctgtccaaa atccggaagc 1800
aaaatgttgt atcgctttta tgacctgaat accaaccgcg gtttcttcag tgatcgtgac 1860
ggcagcaagt tctacgacat caccctaatg tcagaagagc gtcgcactgg ttacagttgg 1920
ggtggtgact acgcgagctc gattatcagc ttcgcacaaa aagtgggata tctctaa 1977

```

<210> 54

<211> 658

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(31)

<221> BINDING

<222> (32)...(124)

<223> Carbohydrate binding module

<221> BINDING

<222> (180)...(303)

<223> Carbohydrate binding module

<221> DOMAIN

<222> (304)...(658)

<223> Catalytic domain

<400> 54

```

Met Asn Asn Ser Thr Lys Lys Met Ile Arg Pro Leu Lys Ala Ser Phe
 1          5          10          15
Ala Leu Gly Ala Leu Ala Leu Ala Ile Ala Ser Pro Ser Trp Ala Ala
 20          25          30
Cys Ser Tyr Ser Val Thr Asn Asn Trp Gly Ser Gly Phe Thr Gly Glu
 35          40          45
Ile Lys Val Thr Asn Asp Thr Thr Ser Thr Val Asn Asn Trp Ser Val
 50          55          60
Ser Trp Gln Glu Ser Gly Val Thr Val Thr Asn Ala Trp Asn Ala Thr
 65          70          75
Leu Ser Gly Ser Asn Pro Tyr Thr Ala Thr Ser Leu Gly Trp Asn Gly
 80          85          90          95
Thr Leu Ala Pro Lys Ala Ser Ala Ser Phe Gly Phe Gln Ala Asn Gly
100          105          110
Thr Ala Gly Ala Pro Lys Val Asn Gly Thr Leu Cys Gly Thr Ser Thr

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      115      120      125
Ser Ser Thr Gly Thr Ser Ser Val Ala Pro Ser Ser Val Ala Ser Ser
130      135      140
Val Ala Val Ser Ser Ser Lys Ser Ser Ser Val Ala Thr Ile Ser
145      150      155      160
Ser Ser Lys Ser Ser Ser Val Pro Thr Val Ser Ser Phe Thr Ile
      165      170      175
Gln Glu Glu Gln Ala Gly Phe Cys Arg Val Asp Gly Ile Ala Thr Glu
180      185      190
Ser Thr Asn Thr Gly Tyr Thr Gly Asn Gly Tyr Thr Asn Thr Thr Asn
195      200      205
Ala Gln Gly Ala Ala Ile Glu Trp Ala Ile Asn Ala Pro Asn Ser Ser
210      215      220
Arg Tyr Thr Leu Thr Phe Arg Tyr Ala Asn Ala Gly Thr Ala Asn Arg
225      230      235      240
Asn Gly Ser Leu Leu Ile Asn Asp Gly Ser Asn Gly Asn Tyr Thr Val
      245      250      255
Gln Leu Pro Ser Thr Gly Ala Trp Ala Thr Trp Gln Thr Val Ser Val
260      265      270
Glu Val Asp Leu Val Gln Gly Asn Asn Ile Leu Lys Leu Ala Ser Leu
275      280      285
Thr Ala Asp Gly Leu Ala Asn Ile Asp Ser Leu Lys Ile Glu Gly Ala
290      295      300
Gln Ala Lys Ala Gly Val Cys Ser Thr Thr Val Ser Ser Ser Ser
305      310      315      320
Ser Ile Lys Ser Ser Ser Ser Ser Ser Ser Ser Ser Thr Ala Ala
      325      330      335
Val Lys Thr Leu Thr Leu Asp Gly Asn Pro Ala Ala Asn Trp Phe Asn
340      345      350
Lys Ser Arg Thr Lys Trp Asn Val Ser Arg Ala Asp Ile Val Leu Ser
355      360      365
Tyr Gln Gln Ser Asn Gly Gly Trp Pro Lys Asn Leu Asp Tyr Asn Ser
370      375      380
Val Gly Ser Gly Asn Gly Gly Ser Asp Ser Gly Thr Ile Asp Asn Gly
385      390      395      400
Ala Thr Ile Thr Glu Met Val Tyr Leu Ala Glu Val Tyr Lys Asn Gly
      405      410      415
Gly Asn Thr Lys Tyr Arg Asp Ala Val Arg Arg Ala Ala Asn Phe Ile
420      425      430
Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu Pro Gln Phe Tyr Pro Leu
435      440      445
Lys Gly Gly Tyr Ala Asp His Ala Thr Phe Asn Asp Asn Gly Met Ala
450      455      460
Tyr Ala Leu Thr Val Leu Asp Phe Ala Val Asn Lys Arg Ala Pro Phe
465      470      475      480
Asp Asn Asp Ile Phe Ser Asp Ser Asp Arg Ser Lys Phe Lys Thr Ala
      485      490      495
Val Thr Lys Gly Val Asp Tyr Ile Leu Lys Ala Gln Trp Lys Gln Asn
500      505      510
Gly Lys Leu Thr Val Trp Cys Ala Gln His Gly Ala Asn Asp Tyr Gln
515      520      525
Pro Lys Lys Ala Arg Ala Tyr Glu Leu Glu Ser Leu Ser Gly Ser Glu
530      535      540
Ser Val Gly Val Leu Ala Phe Leu Met Thr Gln Pro Gln Thr Thr Gln
545      550      555      560
Ile Glu Ala Ala Val Arg Ala Gly Val Ala Trp Phe Asn Ser Pro Ser
      565      570      575
Thr Tyr Leu Asn Asn Tyr Thr Tyr Asp Ser Ser Lys Ala Ser Thr Asn
580      585      590

Pro Ile Val Pro Lys Ser Gly Ser Lys Met Trp Tyr Arg Phe Tyr Asp
595      600      605
Leu Asn Thr Asn Arg Gly Phe Phe Ser Asp Arg Asp Gly Ser Lys Phe
610      615      620
Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg Arg Thr Gly Tyr Ser Trp
625      630      635      640
Gly Gly Asp Tyr Gly Ser Ser Ile Ile Ser Phe Ala Gln Lys Val Gly
      645      650      655
Tyr Leu

```

<210> 55

<211> 1125

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 55

```

gtggtcctag gtaataacgg cggcagcttg agttgcgtcc aatatattgt gattgtgaaa 60
ggacccggtg gacctcgacc gccggtgaaa ccggccgtcc aggcgcccggt tagggttacc 120
tggagcgcat gcctagtcca gcggcccgaa tggtagcgga gtgacgaagc gatccgcctc 180
gcggacaacg tctctctcta ccagcgcaac accggcgggt ggccgaagga catagatatg 240
gccgagccca tcccggaaca caggaagtcc ttttctctca ccgagaagga gcggaccgat 300
gactcgacca tgcacaacgg tgccaccgtg acccagctca agtatctcgc ccggtctctc 360
aaggcgacca ggctggaacg gttcaaggag ggcttctctc aaggctctga ctacctcttg 420
gccgcccagt acccgaaacg cggctggccc cagtattatc ctaacttgag gggctactac 480
gccaacatca cttataacga caatgccatg gtgaacgtgc tcacctctct ccagagcatc 540
gccaaaaagg ccccggaagta cgacttcgtc gaccggcgcc gccgggagaa ggccgcccgg 600
gccgtggcga aagggatcga ctgcatctct aagaccaga tccgtgtcaa tggaaaactt 660
accgcctggt gcgcccagca tgaccccaag acgctggcgc ccgcgccggc ccgttcgtat 720
gagcttgagt ccatcagcgg tttcgagagc gtcgggatcg tccggttctt aatgagcctc 780
gagaatccga gcccgaaagg catcgaggcg gttagaggcc cgtgaaaatg gttcgaggag 840
gtcaagctta ccgggatcaa ggtggtcgag aaacccgacc cgtcccttcc gggcggttac 900
gaccgctggt tggtcgaaga ccccaacgcg ccgcccatct gggcccggtt ctacgagatc 960
ggcaccaacc gtcccttctt ctgcggccgc gatggtatca aaaaatacag cctggcggag 1020
atcgaaacag aacgcccggg cggttactcc tggtaacca atgccccggc ctacctcatc 1080
gagaaggagt atccgctctg gcgggcaaaa caccctacca agtaa 1125

```

<210> 56

<211> 374

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(374)

<223> Catalytic domain

<400> 56

```

Met Val Leu Gly Asn Gly Gly Ser Leu Ser Cys Val Gln Tyr Ile
1      5      10      15
Val Ile Val Lys Gly Pro Gly Gly Pro Arg Pro Pro Val Lys Pro Ala
20     25     30
Val Gln Ala Pro Val Arg Val Thr Trp Ser Ala Cys Leu Val Gln Arg
35     40     45

```

```

Pro Glu Trp Tyr Gly Ser Asp Glu Ala Ile Arg Ile Ala Asp Asn Val
50      55      60
Leu Leu Tyr Gln Arg Asn Thr Gly Gly Trp Pro Lys Asp Ile Asp Met
65      70      75      80
Ala Glu Pro Ile Pro Glu His Arg Lys Ser Phe Phe Leu Thr Glu Lys
85      90      95
Glu Arg Thr Asp Asp Ser Thr Ile Asp Asn Gly Ala Thr Val Thr Gln
100     105     110
Leu Lys Tyr Leu Ala Arg Val Tyr Lys Ala Thr Arg Leu Glu Arg Phe
115     120     125
Lys Glu Gly Phe Leu Lys Gly Leu Asp Tyr Leu Leu Ala Ala Gln Tyr
130     135     140
Pro Asn Gly Gly Trp Pro Gln Tyr Tyr Pro Asn Leu Arg Gly Tyr Tyr
145     150     155     160
Ala Asn Ile Thr Tyr Asn Asp Asn Ala Met Val Asn Val Leu Thr Leu
165     170     175
Leu Gln Ser Ile Ala Lys Lys Ala Pro Glu Tyr Asp Phe Val Asp Pro
180     185     190
Ala Arg Arg Glu Lys Ala Ala Arg Ala Val Ala Lys Gly Ile Asp Cys
195     200     205
Ile Leu Lys Thr Gln Ile Arg Val Asn Gly Lys Leu Thr Ala Trp Cys
210     215     220
Ala Gln His Asp Pro Lys Thr Leu Ala Pro Ala Pro Ala Arg Ser Tyr
225     230     235
Glu Leu Glu Ser Ile Ser Gly Phe Glu Ser Val Gly Ile Val Arg Phe
245     250     255
Leu Met Ser Leu Glu Asn Pro Ser Pro Lys Val Ile Glu Ala Val Glu
260     265     270
Ala Ala Val Lys Trp Phe Glu Glu Val Lys Leu Thr Gly Ile Lys Val
275     280     285
Val Glu Lys Pro Asp Pro Ser Leu Pro Gly Tyr Asp Arg Val Val
290     295     300
Val Glu Asp Pro Asn Ala Pro Pro Ile Trp Ala Arg Phe Tyr Glu Ile
305     310     315
Gly Thr Asn Arg Pro Phe Phe Cys Gly Asp Gly Ile Lys Lys Tyr
325     330     335
Ser Leu Ala Glu Ile Glu His Glu Arg Arg Val Gly Tyr Ser Trp Tyr
340     345     350
Thr Asn Ala Pro Ala Tyr Leu Ile Glu Lys Glu Tyr Pro Leu Trp Arg
355     360     365
Ala Lys His Pro Thr Lys
370

```

<210> 57

<211> 1170

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 57

```

atggacaaac gcgtcaaatg gattcatcag ctttcaaaag aagaagcaaa gcagttcgag    60
cccgaaaatt tcctcaaaag caaagacggc tggaatccga aaaaggcggg tgaccgctgg    120
ctcgaaaaaa caaaacctga ctggcagctc gttacgtgga acgacgcgtt acgccaggcg    180
ccgctctggt atcaaaccca tgaagcggcg cgcattgccg accagggtgat ttgtaccag    240
aaagacaacg gcggctggga aaaaaatctc gatatgacgg ccatgctcac gcaagccgaa    300
cgcgaaaagc tcgccaaga aaaaatcgaa acgtcggaaa cgacgatcga caaccgcacg    360
acctacacgc aagtcgcttt tctcgcaaaa gtcattacgg gcagcttgca gaaaacgact    420

```

```

ccgcccacca atttccgaa acataaggaa gcttttttca agggcttggg ttacctgctc    480
gcgtcgcagt acgaatcggg cggctttccg .cagttttatc cgctcaaaaa aggttattac    540
acgcacatca cgttcaacga cgaatgcgatg attggcgttt tgaagggttt gcgcgaaatc    600
gccaaaaaga aggaagacta tctttttggt gacgaagaac gcgcctgaa agcggaaaaa    660
tcggtcgaaa aagcgtgccc gctgattctg aaattgcagg ttgaagtcgg cggcaaaaaa    720
acgggtttggg cggcgcagta tgacgaaaaa acittttaaac ccgcagcggc gcgaaaagttt    780
gaaccgggtt ctttaacggc gggcgaatcg gtcggcatcg tccggttttt aatgtacgat    840
tcaaagcccg accagcgac gattgacgag attgaatctg ccattcagtg gtatcgcgcg    900
aacaataatcg aaggcattcg atgggtgcgc gaaaacggcg aaaaccgcgt cgtcaaggac    960
aaaaacgcgc cgccgatttg gcgcggtttt tacgaaatcg aaacgatgaa gccgattttc    1020
atcggggcgcg acgccatcat tcgttacgac gtgtctgaaa tcgaagccga gcgccgcaac    1080
ggctacgcgt ggtacgtctc ggagccgaac gagctgcttg aaaaagatta cccgaaatgg    1140
ctggaaaaaa ttaaaaaatc agtaaagtaa    1170

```

<210> 58

<211> 389

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(389)

<223> Catalytic domain

<400> 58

```

Met Asp Lys Arg Val Lys Trp Ile His Gln Leu Ser Lys Glu Glu Ala
 1          5          10          15
Lys Gln Phe Glu Pro Glu Asn Phe Leu Lys Gly Lys Asp Gly Trp Asn
 20          25          30
Pro Lys Lys Ala Asp Asp Arg Trp Leu Glu Lys Thr Lys Pro Asp Trp
 35          40          45
Gln Leu Val Thr Trp Asn Asp Ala Leu Arg Gln Ala Pro Leu Trp Tyr
 50          55          60
Gln Thr Asp Glu Ala Ala Arg Ile Ala Asp Gln Val Ile Leu Tyr Gln
 65          70          75          80
Lys Asp Asn Gly Gly Trp Glu Lys Asn Leu Asp Met Thr Ala Met Leu
 85          90          95
Thr Gln Ala Glu Arg Glu Lys Leu Ala Lys Glu Lys Ser Asn Thr Ser
100          105          110
Glu Thr Thr Ile Asp Asn Arg Thr Thr Tyr Thr Gln Val Ala Phe Leu
115          120          125
Ala Lys Val Ile Thr Gly Ser Leu Gln Lys Thr Thr Pro Pro Thr Asn
130          135          140
Phe Pro Lys His Lys Glu Ala Phe Phe Lys Gly Leu Asp Tyr Leu Leu
145          150          155          160
Ala Ser Gln Tyr Glu Ser Gly Gly Phe Pro Gln Phe Tyr Pro Leu Lys
165          170          175
Lys Gly Tyr Tyr Thr His Ile Thr Phe Asn Asp Asp Ala Met Ile Gly
180          185          190
Val Leu Lys Val Leu Arg Glu Ile Ala Lys Lys Lys Glu Asp Tyr Leu
195          200          205
Phe Val Asp Glu Glu Arg Arg Leu Lys Ala Glu Lys Ser Val Glu Lys
210          215          220
Ala Leu Pro Leu Ile Leu Lys Leu Gln Val Glu Val Gly Gly Lys Lys
225          230          235          240
Thr Val Trp Ala Ala Gln Tyr Asp Glu Asn Thr Phe Lys Pro Ala Ala
245          250          255

```

```

Ala Arg Lys Phe Glu Pro Val Ser Leu Thr Ala Gly Glu Ser Val Gly
      260      265      270
Ile Val Arg Phe Leu Met Tyr Asp Ser Lys Pro Asp Gln Ala Thr Ile
      275      280      285
Asp Ala Ile Glu Ser Ala Ile Gln Trp Tyr Arg Ala Asn Lys Ile Glu
      290      295      300
Gly Ile Arg Trp Val Arg Glu Asn Gly Glu Asn Arg Val Val Lys Asp
      305      310      315      320
Lys Asn Ala Pro Pro Ile Trp Ala Arg Phe Tyr Glu Ile Glu Thr Met
      325      330      335
Lys Pro Ile Phe Ile Gly Arg Asp Ala Ile Ile Arg Tyr Asp Val Ser
      340      345      350
Glu Ile Glu Ala Glu Arg Arg Asn Gly Tyr Ala Trp Tyr Val Ser Glu
      355      360      365
Pro Asn Glu Leu Leu Glu Lys Asp Tyr Pro Lys Trp Leu Glu Lys Ile
      370      375      380
Lys Lys Ser Val Lys
      385

```

<210> 59

<211> 1080

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 59

```

atgagaatcc ggtcctcttc aatcgcgttc ggcctgattt gcagtcctggc gctaagggtg      60
cctgcgcaag cgcaggtcac cgtgcgtgg gggagctcc tgaaccagcc cgccgcctgg      120
tatggcaccg atgaagcccg tcgaattgcc gaccacgtgc tcgagcatca acgagcggaa      180
ggcggatggc caaagaacac ggacatgacc gcagcgcccg atccggcggt gctcacagcc      240
gcgcgagtga agccagactc gacgatcgat aacggcgcca ccgtcactga aatgcgcgtc      300
ctcgcgcgcg tctaccgttc atcaccgat ccccgttatc gcgatgcgt gctcaagggt      360
ctcgactatc tgttggcagc gcagtatgcc aacggcggtt ggccgcagtt ctaccgctc      420
cggcaggact attcgcgcta tatcacgttc aacgacaacg cgatgatcaa tgcgtgacg      480
ctgctctcag acgtcgctgc cggaaatggc gactgggctt ttgctgatgc cagccggcgc      540
gagaaaagcc ggacggctgt agagaaggcc gtagaagtca tcctgcgcgc gcaggtgaga      600
gttgacggcc ggctgaccgc gtggtgcgcc caacacgacg aggtgacact cgagccgcgc      660
aaggcccgcg cctacgaaca tccgtcgtg agcggacagg agacggtggg gatcatccgg      720
ttttcatga cccgcgataa accggatcag agagtcgtcg atgcaatoga ggcgtcagtg      780
gcatggctga aggcggtgca gctcaaagga cttcgctcg accagcgccg cgatccctcg      840
ctgcggaggg ggcgtgacgt ggtgaccgtc gctgaccogt cggcgccgcc gctctgggcg      900
cgcttctacg aaatcgggac caatcgcccg atcttctctg gacgcgacgg cgtgatccga      960
tactcgctgg cagagatcga gcacgaacgc cggatagggt acgcctggct cggaacctgg      1020
ccgcggaagc tgctcgatac cgaataccca tcttgcgac ggactcaaca aaggccgtga      1080

```

<210> 60

<211> 359

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(24)

<221> DOMAIN

<222> (25)...(359)

<223> Catalytic domain

<400> 60

```

Met Arg Ile Arg Ser Ser Ser Ile Ala Phe Gly Leu Ile Cys Ser Leu
1      5      10      15
Ala Leu Arg Val Pro Ala Gln Ala Gln Val Thr Val Arg Trp Ala Asp
20      25      30
Val Leu Asn Gln Pro Ala Ala Trp Tyr Gly Thr Asp Glu Ala Arg Arg
35      40      45
Ile Ala Asp His Val Leu Glu His Gln Arg Ala Glu Gly Gly Trp Pro
50      55      60
Lys Asn Thr Asp Met Thr Ala Ala Pro Asp Pro Ala Val Leu Thr Ala
65      70      75      80
Ala Arg Val Lys Pro Asp Ser Thr Ile Asp Asn Gly Ala Thr Val Thr
85      90      95
Glu Met Arg Val Leu Ala Arg Val Tyr Arg Ser Ser Pro Asp Pro Arg
100      105      110
Tyr Arg Asp Ala Leu Leu Lys Gly Leu Asp Tyr Leu Leu Ala Ala Gln
115      120      125
Tyr Ala Asn Gly Gly Trp Pro Gln Phe Tyr Pro Leu Arg Gln Asp Tyr
130      135      140
Ser Arg Tyr Ile Thr Phe Asn Asp Asn Ala Met Ile Asn Val Val Thr
145      150      155      160
Leu Leu Ser Asp Val Ala Ala Gly Asn Gly Asp Trp Ala Phe Ala Asp
165      170      175
Ala Ser Arg Arg Glu Lys Ser Arg Thr Ala Val Glu Lys Ala Val Glu
180      185      190
Val Ile Leu Arg Ala Gln Val Arg Val Asp Gly Arg Leu Thr Ala Trp
195      200      205
Cys Ala Gln His Asp Glu Val Thr Leu Glu Pro Arg Lys Ala Arg Ala
210      215      220
Tyr Glu His Pro Ser Leu Ser Gly Gln Glu Thr Val Gly Ile Ile Arg
225      230      235      240
Phe Leu Met Thr Arg Asp Lys Pro Asp Gln Arg Val Val Asp Ala Ile
245      250      255
Glu Ala Ser Val Ala Trp Leu Lys Ala Val Gln Leu Lys Gly Leu Arg
260      265      270
Val Asp Gln Arg Arg Asp Pro Ser Leu Pro Glu Gly Arg Asp Val Val
275      280      285
Thr Val Ala Asp Pro Ser Ala Pro Pro Leu Trp Ala Arg Phe Tyr Glu
290      295      300
Ile Gly Thr Asn Arg Pro Ile Phe Ser Gly Arg Asp Gly Val Ile Arg
305      310      315      320
Tyr Ser Leu Ala Glu Ile Glu His Glu Arg Arg Ile Gly Tyr Ala Trp
325      330      335
Leu Gly Thr Trp Pro Ala Lys Leu Leu Asp Thr Glu Tyr Pro Ser Trp
340      345      350
Arg Arg Thr Gln Gln Arg Pro
355

```

<210> 61

<211> 1224

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 61

```

gtggaattac cagtaaccgg cgcattgggca acctggcaaa ccgcaactgt tgaaattgat      60
ttggtgcaag gtaacaacct gttaaaactt tctgcgatca cggctgatgg ttggcgaat      120
atcgattcgt tgaataatga cggcgacaaa accaaagccg cgtgtgacag cactgtggca      180
agcagcagct cttcatccgt tgcttcatcg attaaatcaa gctccagttc atcctcttcc      240
agttcaacga cgacggtaaaa aacattaaca ctggatggca accccgcagc aaactggttt      300
aacaatcca gaacaaaatg gaataccagc agagccgatg ttgtactttc ctatcaacaa      360
tccaacggcg gctggccaaa aaatctcgat tacaattcag taagcgcagg taatggcggc      420
agcgatagcg gcaccatcga taacggtgca accattactg aaatggttta tctcgcggaa      480
gtttacaaaa atggcaacaa caccaagtat cgcgatgcgg tgcgcagagc cgcaaatatt      540
attgtcagct cgcaatacag cactggtgca ttaccacaat tttatccatt gaaaggcggc      600
tatgcagacc acgccacctt taacgataac ggcatggcat atgcattaac ggtattggat      660
tttgcaatca acaaacgcgc ccatttggat actgatgttt tctccgattc tgatcgccgc      720
aaattcaaaa ccgctgttgc caaagggtgt gattacattt tgaagcgcaa gtggaacaaa      780
aacggaaaaa taaccgtgtg gtgtgcacaa catgggtgta ccgattatca accgaaaaaa      840
gcgcgcgcct atgaattgga atcactgagt ggcagcgaat ctgttggtgt actcgctttc      900
ttgatgaccc aaccgcaaac cgcacaaatt gaagccgctg taaaagccgg ttagcctgg      960
ttcaatagcc ccaacacgta ttgaacaat tacacttacg actcttcaaa agcgtcaact      1020
aatccaatag ttgcaagtc tggaaacaaa atgtggtatc gcttttaaga tttaaatacc      1080
aatcgtggct tcttcagtga tcgcgatggc agcaaatctc atgacatcac ccagatgtca      1140
gaagagcgtc gcactggata tagctggggt ggtgattacg gcacgtcgat tatctccttc      1200
gcgcaaaaag tggatatctt gtaa
1224

```

<210> 62

<211> 407

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (0)...(407)

<223> Catalytic domain

<400> 62

```

Met Glu Leu Pro Val Thr Gly Ala Trp Ala Thr Trp Gln Thr Ala Thr
 1      5      10      15
Val Glu Ile Asp Leu Val Gln Gly Asn Asn Leu Leu Lys Leu Ser Ala
 20     25     30
Ile Thr Ala Asp Gly Leu Ala Asn Ile Asp Ser Leu Lys Ile Asp Gly
 35     40     45
Ala Gln Thr Lys Ala Gly Val Cys Ser Thr Val Ala Ser Ser Ser Ser
 50     55     60
Ser Ser Val Ala Ser Ser Ile Lys Ser Ser Ser Ser Ser Ser Ser
 65     70     75     80
Ser Ser Thr Thr Thr Val Lys Thr Leu Thr Leu Asp Gly Asn Pro Ala
 85     90     95
Ala Asn Trp Phe Asn Lys Ser Arg Thr Lys Trp Asn Thr Ser Arg Ala
100    105    110
Asp Val Val Leu Ser Tyr Gln Gln Ser Asn Gly Gly Trp Pro Lys Asn
115    120    125
Leu Asp Tyr Asn Ser Val Ser Ala Gly Asn Gly Gly Ser Asp Ser Gly
130    135    140
Thr Ile Asp Asn Gly Ala Thr Ile Thr Glu Met Val Tyr Leu Ala Glu
145    150    155    160
Val Tyr Lys Asn Gly Asn Asn Thr Lys Tyr Arg Asp Ala Val Arg Arg
165    170    175
Ala Ala Asn Phe Ile Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu Pro

```

```

      180      185      190
Gln Phe Tyr Pro Leu Lys Gly Gly Tyr Ala Asp His Ala Thr Phe Asn
195    200    205
Asp Asn Gly Met Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala Val Asn
210    215    220
Lys Arg Ala Pro Phe Asp Thr Asp Val Phe Ser Asp Ser Asp Arg Ala
225    230    235    240
Lys Phe Lys Thr Ala Val Ala Lys Gly Val Asp Tyr Ile Leu Lys Ala
245    250    255
Gln Trp Lys Gln Asn Gly Lys Leu Thr Val Trp Cys Ala Gln His Gly
260    265    270
Ala Thr Asp Tyr Gln Pro Lys Lys Ala Arg Ala Tyr Glu Leu Glu Ser
275    280    285
Leu Ser Gly Ser Glu Ser Val Gly Val Leu Ala Phe Leu Met Thr Gln
290    295    300
Pro Gln Thr Ala Gln Ile Glu Ala Ala Val Lys Ala Gly Val Ala Trp
305    310    315    320
Phe Asn Ser Pro Asn Thr Tyr Leu Asn Asn Tyr Thr Tyr Asp Ser Ser
325    330    335
Lys Ala Ser Thr Asn Pro Ile Val Ala Lys Ser Gly Ser Lys Met Trp
340    345    350
Tyr Arg Phe Tyr Asp Leu Asn Thr Asn Arg Gly Phe Phe Ser Asp Arg
355    360    365
Asp Gly Ser Lys Phe Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg Arg
370    375    380
Thr Gly Tyr Ser Trp Gly Gly Asp Tyr Gly Thr Ser Ile Ile Ser Phe
385    390    395    400
Ala Gln Lys Val Gly Tyr Leu
405

```

<210> 63

<211> 1023

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 63

```

atgttaagtt tcatcgcggt atcagtggtt cataattact gtacagggca gactgcgtcc 60
accaaaaatt cagtgccgga aaagatgctt cagtaccagt tgtcaaatgg cgcctggccc 120
aaacagttgg tagacaaaag tgtcgttgat tacagcttcc cattaacgaa agagctccta 180
cagcagatca agaaaacaga tattgatcat gctacgctcg acaacagtcg gacaacccgg 240
gaaataactg aattgatcaa ggcttttaag gacactaaaa ataaggcata ttgactgct 300
gcagaaaagg ggattgcata tattttatcg gctcaatatg agaatggcgg attccacaa 360
tactacccaa ataaattata ctatagagct gagataacat acaacgatga tgcgatgatc 420
aatgcattac tagtgcttta caaagtagcc aataagcgag aggggtttga ggctatcaat 480
cccatatttg tgtcaaaagc gcaaaaagca gttgaaaagg gtataacctg tatcctaataa 540
acacaggtca tacaagacgg aaaaaggagt attgggctg cgcaatacga tcagaacact 600
ttacaacctg ctacaggcaag aaagtttgaa ccagcttcat tgagcacaa tgaaatctgtt 660
tccatcgctc gctttctcat gctacagcct gcaaccactg aaattaagca agcgatcgaa 720
catgcaatac aatgggtcga acagcatgat attgaaggtt accgtttcga ccgcatacaa 780
gatagggtga ctggaaaata tcaacggcag cttgtcgctg atcggacttc cagatttgg 840
gcgcgatttt ataactctga agacaaccgt ccattgtttg gagatcgggg caatacaatc 900
aaatacaact ttgaggaggt ttacagaggag cgtagaaatg gctatgcttg gttcggcaac 960
tgcccgaaaa agctgatcca aaaggactat ccaaaatgga aaaaacaata caaatttcaa 1020
taa 1023

```

<210> 64

<211> 340

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(16)

<221> DOMAIN

<222> (17)...(340)

<223> Catalytic domain

<400> 64

```

Met Leu Ser Phe Ile Ala Val Ser Val Phe His Asn Tyr Cys Thr Gly
 1      5      10      15
Gln Thr Ala Ser Thr Lys Asn Ser Val Ala Glu Lys Met Leu Gln Tyr
 20     25     30
Gln Leu Ser Asn Gly Ala Trp Pro Lys Gln Leu Val Asp Lys Ser Val
 35     40     45
Val Asp Tyr Ser Leu Pro Leu Thr Lys Glu Leu Leu Gln Gln Ile Lys
 50     55     60
Lys Thr Asp Ile Asp His Ala Thr Leu Asp Asn Ser Ala Thr Thr Arg
 65     70     75     80
Glu Ile Thr Glu Leu Ile Lys Ala Phe Lys Asp Thr Lys Asn Lys Ala
 85     90     95
Tyr Leu Thr Ala Ala Glu Lys Gly Ile Ala Tyr Ile Leu Ser Ala Gln
100    105    110
Tyr Glu Asn Gly Gly Phe Pro Gln Tyr Tyr Pro Asn Lys Leu Tyr Tyr
115    120    125
Arg Ala Glu Ile Thr Tyr Asn Asp Asp Ala Met Ile Asn Ala Leu Leu
130    135    140
Val Leu Tyr Lys Val Ala Asn Lys Arg Glu Gly Phe Glu Ala Ile Asn
145    150    155    160
Pro Ile Phe Val Ser Lys Ala Gln Lys Ala Val Glu Lys Gly Ile Thr
165    170    175
Cys Ile Leu Lys Thr Gln Val Ile Gln Asp Gly Lys Arg Ser Ile Trp
180    185    190
Ala Ala Gln Tyr Asp Gln Asn Thr Leu Gln Pro Ala Gln Ala Arg Lys
195    200    205
Phe Glu Pro Ala Ser Leu Ser Thr Ser Glu Ser Val Ser Ile Val Arg
210    215    220
Phe Leu Met Leu Gln Pro Ala Thr Thr Glu Ile Lys Gln Ala Ile Glu
225    230    235    240
His Ala Ile Gln Trp Phe Glu Gln His Asp Ile Glu Gly Tyr Arg Phe
245    250    255
Asp Arg Ile Gln Asp Arg Val Thr Gly Lys Tyr Gln Arg Gln Leu Val
260    265    270
Ala Asp Arg Thr Ser Thr Ile Trp Ala Arg Phe Tyr Asn Leu Glu Asp
275    280    285
Asn Arg Pro Leu Phe Gly Asp Arg Asp Asn Thr Ile Lys Tyr Asn Phe
290    295    300
Glu Glu Val Ser Glu Glu Arg Arg Asn Gly Tyr Ala Trp Phe Gly Asn
305    310    315    320
Trp Pro Glu Lys Leu Ile Gln Lys Asp Tyr Pro Lys Trp Lys Lys Gln
325    330    335
Tyr Lys Ile Gln
340

```

<210> 65

<211> 1311

<212> DNA

<213> Bacteria

<400> 65

```

gtgaaccgac gtaccgcctt gggagcggtc gccgcgaccg ccctcgccct gacggtcacc 60
gccccgcgcg ccggtgccca cgcgcgcgct cccacgcgcg cgccacgccc ggtcgccgat 120
ccggctcgcg ccacgtgccc cgcgcgcgac ggctggggtt ccgaggggac cggcacgacc 180
ggtggggcgcg ccgcccaggc ctcccgggtc ttcaccgtcg ccacctggga ggagtcccg 240
gccgcgctcg cgtgcccggg ctccgagccc aggatcgtca aggtggtggg cagctgaac 300
gccaccgcgcg ccggctcgcg cgccttcgag gcgcggggct acgacttcgc ccgtacctc 360
gccgactacg acccggcgct gtgggggtac gagaaggagg tcagcgccc gcaggaggag 420
ctgcggggcg cgtccgcgac cgcgcagggc caggccatca aggtcaaggt gccggcgaac 480
accacgatcg tcggggtcgg caggcacgcg gggatcacgg gcggcagcct ccaggtgcag 540
ggcgtcgaca acgtcgtggt ccgcaacctg acgctggaga gcccgctcga ctgcttcccg 600
cagtggggacc cgaccgacgg cgcgaccggg gcgtggaact ccgagtacga cagcctcgtc 660
gtgtacggct ccacccatgt ctggatcgac cacaacacct tcaccgacgg cgcccaccgg 720
gacagttcgc tgcctcgta ctacggcgag gtctaccagc agcacgacgg cgaactggac 780
gtcgtgcggg gcgcggacct cgtcacggtc tegtggaaag ccttcaccga ccacgacaag 840
accctgatga tcggcaacag cgacagcgcg gccgccaccg accggggcaa gctgcgggtc 900
accctgcacc acaacctgtt cgagaacgtc gtcgagcggg cgcggcggtt caggttcggg 960
caggtcgacg cgtacaacaa ccacttcgtc gtgccgagtt cggcctacgc gtacagcctg 1020
ggcgtcgggc aggagtcaca gctcttcgcg gagaagaacg cgttcacctt cgccgggggc 1080
gtgcggcgcg ggaagatcct caagaagtgg aaggacgcgc ccgtcaccac cgtcggaac 1140
tacgtgaacg gcaggcggtt cgacctgctc gccgtccaca acaccagtt ccgggaggag 1200
cagttgcggg ccgacgcggg ctggaccccc gtccctgcgc caaggggtcga ccaccggag 1260
gcgtccccg cgtcgtcga ccaccgcgcg gccgcggccc gctcctgctg a 1311

```

<210> 66

<211> 436

<212> PRT

<213> Bacteria

<220>

<221> SIGNAL

<222> (1)...(28)

<221> DOMAIN

<222> (29)...(436)

<223> Catalytic domain

<400> 66

```

Met Asn Arg Arg Thr Arg Leu Gly Ala Val Ala Ala Thr Ala Leu Ala
1      5      10      15
Leu Thr Val Thr Ala Pro Ala Ala Gly Ala His Ala Ala Ala Pro His
20     25     30
Ala Ala Pro Arg Pro Val Ala Asp Pro Ala Arg Ala Thr Leu Pro Ala
35     40     45
Gly Asp Gly Trp Ala Ser Glu Gly Thr Gly Thr Thr Gly Gly Ala Ala
50     55     60
Ala Glu Ala Ser Arg Val Phe Thr Val Ala Thr Trp Glu Glu Phe Arg
65     70     75     80
Ala Ala Leu Ala Val Pro Gly Ser Glu Pro Arg Ile Val Lys Val Val
85     90     95
Gly Thr Leu Asn Ala Thr Ala Ala Gly Cys Gly Ala Phe Glu Ala Pro
100    105    110

```

Gly Tyr Asp Phe Ala Arg Tyr Leu Ala Asp Tyr Asp Pro Ala Val Trp
 115 120 125
 Gly Tyr Glu Lys Glu Val Ser Gly Pro Gln Glu Glu Leu Arg Ala Ala
 130 135 140
 Ser Ala Thr Ala Gln Gly Gln Ala Ile Lys Val Lys Val Pro Ala Asn
 145 150 155 160
 Thr Thr Ile Val Gly Val Gly Arg His Ala Gly Ile Thr Gly Gly Ser
 165 170 175
 Leu Gln Val Gln Gly Val Asp Asn Val Val Val Arg Asn Leu Thr Leu
 180 185 190
 Glu Ser Pro Leu Asp Cys Phe Pro Gln Trp Asp Pro Thr Asp Gly Ala
 195 200 205
 Thr Gly Ala Trp Asn Ser Glu Tyr Asp Ser Leu Val Val Tyr Gly Ser
 210 215 220
 Thr His Val Trp Ile Asp His Asn Thr Phe Thr Asp Gly Ala His Pro
 225 230 235 240
 Asp Ser Ser Leu Pro Ser Tyr Tyr Gly Glu Val Tyr Gln Gln His Asp
 245 250 255
 Gly Glu Leu Asp Val Val Arg Gly Ala Asp Leu Val Thr Val Ser Trp
 260 265 270
 Asn Ala Phe Thr Asp His Asp Lys Thr Leu Met Ile Gly Asn Ser Asp
 275 280 285
 Ser Ala Gly Ala Thr Asp Arg Gly Lys Leu Arg Val Thr Leu His His
 290 295 300
 Asn Leu Phe Glu Asn Val Val Glu Arg Ala Pro Arg Val Arg Phe Gly
 305 310 315 320
 Gln Val Asp Ala Tyr Asn Asn His Phe Val Val Pro Ser Ser Ala Tyr
 325 330 335
 Ala Tyr Ser Leu Gly Val Gly Gln Glu Ser Gln Leu Phe Ala Glu Lys
 340 345 350
 Asn Ala Phe Thr Leu Ala Gly Gly Val Pro Ala Gly Lys Ile Leu Lys
 355 360 365
 Lys Trp Lys Asp Ala Pro Val Thr Thr Val Gly Asn Tyr Val Asn Gly
 370 375 380
 Arg Pro Val Asp Leu Leu Ala Val His Asn Thr Gln Phe Pro Glu Glu
 385 390 395 400
 Gln Leu Arg Ala Asp Ala Gly Trp Thr Pro Val Leu Arg Thr Arg Val
 405 410 415
 Asp His Pro Arg Ala Val Pro Ala Leu Val Asp His Arg Ala Gly Ala
 420 425 430
 Gly Arg Ser Cys
 435

<210> 67

<211> 1995

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 67

atgaaaaatt	caaaaactgt	ttttactgca	caaaaaaac	tcatgcaactc	ttgcattggcc	60
gocgctatcg	gcttggcgat	aagttcaggt	gcttgggtcag	cttgtactta	cactgtcaacc	120
aataattggg	gttctggcgt	caccgggtgaa	atcaaagtta	ccaacaacac	atcatcggtct	180
gtaaatgggt	ggtctgtgtc	ttggcaggaa	tcaggcgcat	cagtcacca	ctcatggaac	240
gcaactctga	gcggatcaaa	tccttatacy	gcagccgcct	taggttggaa	tgcaactctc	300
gcacccgggtg	cttctgcag	ttttggcttt	caagcaaatg	gcactgctag	cgcacotaaa	360
gtgaatggca	ctttatgtgg	aacagctact	tcatacaac	ctgcgtcatc	cagcagtgtt	420
gcgagttcgg	ttaaatcaag	cgacccggtt	tcgtccagca	gtaaatcatc	cagctcaatc	480
actgtgagta	gtagttctat	cgccagcagc	agcgcaacaa	gtgtttcttc	attaacaatt	540
caggaaagagc	aagctggcct	ctgtcgtggt	gatggcattg	caacagaaag	caccaacacc	600
ggctttaccg	gcaacggcta	taccaatgca	aacaacgcac	aaggtgcagc	gattgaatgg	660
gcggtaaatg	caccgagcag	tggccgctac	acactcacat	tccgttttgc	aaatggcggc	720
actgcagcac	gcaatgggtc	actgttaatt	aacggcggtg	gcaatggtaa	ctacaccgtg	780
gatttaccac	taaccggcgc	atgggcgact	tggcaaacag	cgactgtaga	aatcgatttg	840
gtacaaggca	ccaacacgct	gaaactttct	gcattaaccc	cagatggcct	agctaataatc	900
gattcoattaa	aaattgatgg	caaccaaccg	aaagcaggca	cttgacgcaa	tacatcaagc	960
agtgttgcca	cgagtctctc	atccgttaaa	tccagttcaa	gttcttcac	aaagctcatcc	1020
accactgcaa	aaatgctgac	tcttgatggc	aaccccgccg	caagttgggt	caacaaatcc	1080
agaaccaaat	ggaatagcag	tcgtgcggac	attgtgttgt	cttaccagca	agctaaccggc	1140
ggctggccaa	aaaatctgga	ttacaactca	gtaagcgcag	gtaatgggtg	tagcgacagc	1200
ggcactatcg	acaacggcgc	aaccatcacg	gaaatgggtt	atctcgcaga	agtttataaa	1260
aacgggggca	atacaaaata	tcgcgatgca	gtacgtaaag	cggaacactt	tattgtgagt	1320
tcgcaatata	gcaactgtgc	gttaccacaa	ttttaccat	tgaaaggtgg	ttatgcagat	1380
cacgccacct	ttaacgataa	cggcatggct	tacgcattaa	cggtattgga	ttttgcggtg	1440
aacaaacgtg	cgccggttga	taacgatgta	ttttctgatg	cagaccgcgc	aaaattcaaa	1500
actgccgtga	caaaaggtat	tgattacatt	ttgaaagccc	aatggaaaaa	aaatggaaaa	1560
ctcaccgcat	ggtgtgcgca	acacggagca	aacgactatc	aacaaaaagc	ggcgcggtgct	1620
tatgagttag	tatctttaag	cggcagcgaa	tcggttgcca	tcactgcgtt	cctgatgacc	1680
caaccacaaa	ctgcgcaaat	cgaagcagcg	gttaaagccg	gtgtaaacgtg	gttcgctagc	1740
ccgaatacat	acttggctaa	ttacacctac	gactcgtcaa	aagcctctac	caatccgatt	1800
gtgtacaaat	cgggcagcag	aatgtggtat	cgcttctacg	atctgaacac	caatcgcgga	1860
ttctttagt	atcgcgatgg	cagcaaatc	tatgacatca	ctcaaatgtc	tgaagaacgt	1920
cgacccggct	acagctgggg	cggttcttac	ggtgaatcga	ttatcagctt	cgcgcaaaaa	1980
gtgggttata	tctaa					1995

<210> 68

<211> 664

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(32)

<221> BINDING

<222> (33)...(126)

<223> Carbohydrate binding module

<221> BINDING

<222> (184)...(307)

<223> Carbohydrate binding module

<221> DOMAIN

<222> (308)...(664)

<223> Catalytic domain

<400> 68

```
Met Lys Asn Ser Lys Thr Val Phe Thr Ala Gln Lys Lys Leu Met His
 1          5          10          15
Ser Cys Ile Ala Ala Ala Ile Gly Leu Ala Ile Ser Ser Gly Ala Trp
 20          25          30
Ser Ala Cys Thr Tyr Thr Val Thr Asn Asn Trp Gly Ser Gly Phe Thr
 35          40          45
Gly Glu Ile Lys Val Thr Asn Asn Thr Ser Ser Ala Val Asn Gly Trp
```

```

50      55      60
Ser Val Ser Trp Gln Glu Ser Gly Ala Ser Val Thr Asn Ser Trp Asn
65      70      75      80
Ala Thr Leu Ser Gly Ser Asn Pro Tyr Thr Ala Ala Ala Leu Gly Trp
85      90      95
Asn Ala Thr Leu Ala Pro Gly Ala Ser Ala Ser Phe Gly Phe Gln Ala
100     105     110
Asn Gly Thr Ala Ser Ala Pro Lys Val Asn Gly Thr Leu Cys Gly Thr
115     120     125
Ala Thr Ser Ser Thr Pro Ala Ser Ser Ser Val Ala Ser Ser Val
130     135     140
Lys Ser Ser Ala Pro Val Ser Ser Ser Ser Lys Ser Ser Ser Ile
145     150     155     160
Thr Val Ser Ser Ser Ile Ala Ser Ser Ser Ala Pro Ser Val Ser
165     170     175
Ser Leu Thr Ile Gln Glu Glu Gln Ala Gly Phe Cys Arg Val Asp Gly
180     185     190
Ile Ala Thr Glu Ser Thr Asn Thr Gly Phe Thr Gly Asn Gly Tyr Thr
195     200     205
Asn Ala Asn Asn Ala Gln Gly Ala Ala Ile Glu Trp Ala Val Asn Ala
210     215     220
Pro Ser Ser Gly Arg Tyr Thr Leu Thr Phe Arg Phe Ala Asn Gly Gly
225     230     235     240
Thr Ala Ala Arg Asn Gly Ser Leu Leu Ile Asn Gly Gly Ser Asn Gly
245     250     255
Asn Tyr Thr Val Asp Leu Pro Leu Thr Gly Ala Trp Ala Thr Trp Gln
260     265     270
Thr Ala Thr Val Glu Ile Asp Leu Val Gln Gly Thr Asn Thr Leu Lys
275     280     285
Leu Ser Ala Leu Thr Ala Asp Gly Leu Ala Asn Ile Asp Ser Leu Lys
290     295     300
Ile Asp Gly Asn Gln Pro Lys Ala Gly Thr Cys Ser Asn Thr Ser Ser
305     310     315     320
Ser Val Ala Ser Ser Ser Ser Val Lys Ser Ser Ser Ser Ser Ser
325     330     335
Ser Ser Ser Ser Thr Thr Ala Lys Met Leu Thr Leu Asp Gly Asn Pro
340     345     350
Ala Ala Ser Trp Phe Asn Lys Ser Arg Thr Lys Trp Asn Ser Ser Arg
355     360     365
Ala Asp Ile Val Leu Ser Tyr Gln Gln Ala Asn Gly Gly Trp Pro Lys
370     375     380
Asn Leu Asp Tyr Asn Ser Val Ser Ala Gly Asn Gly Gly Ser Asp Ser
385     390     395     400
Gly Thr Ile Asp Asn Gly Ala Thr Ile Thr Glu Met Val Tyr Leu Ala
405     410     415
Glu Val Tyr Lys Asn Gly Gly Asn Thr Lys Tyr Arg Asp Ala Val Arg
420     425     430
Lys Ala Ala Asn Phe Ile Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu
435     440     445
Pro Gln Phe Tyr Pro Leu Lys Gly Gly Tyr Ala Asp His Ala Thr Phe
450     455     460
Asn Asp Asn Gly Met Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala Val
465     470     475     480
Asn Lys Arg Ala Pro Phe Asp Asn Asp Val Phe Ser Asp Ala Asp Arg
485     490     495
Ala Lys Phe Lys Thr Ala Val Thr Lys Gly Ile Asp Tyr Ile Leu Lys
500     505     510
Ala Gln Trp Lys Gln Asn Gly Lys Leu Thr Ala Trp Cys Ala Gln His
515     520     525

Gly Ala Asn Asp Tyr Gln Pro Lys Ala Ala Arg Ala Tyr Glu Leu Val
530     535     540
Ser Leu Ser Gly Ser Glu Ser Val Gly Ile Ile Ala Phe Leu Met Thr
545     550     555     560
Gln Pro Gln Thr Ala Gln Ile Glu Ala Ala Val Lys Ala Gly Val Asn
565     570     575
Trp Phe Ala Ser Pro Asn Thr Tyr Leu Ala Asn Tyr Thr Tyr Asp Ser
580     585     590
Ser Lys Ala Ser Thr Asn Pro Ile Val Tyr Lys Ser Gly Ser Arg Met
595     600     605
Trp Tyr Arg Phe Tyr Asp Leu Asn Thr Asn Arg Gly Phe Phe Ser Asp
610     615     620
Arg Asp Gly Ser Lys Phe Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg
625     630     635     640
Arg Thr Gly Tyr Ser Trp Gly Gly Ser Tyr Gly Glu Ser Ile Ile Ser
645     650     655
Phe Ala Gln Lys Val Gly Tyr Leu
660

```

<210> 69

<211> 1035

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 69

```

atggcgcggtt tgttcgggtg cgtgtgtgcc agcctgggag gatggggccgc ggttctggcc 60
gccgcgggcg gcccggtatt gtcccgctgt ctccgcgaac cggacccttg gtttcgcagt 120
ccggcggggc aacaggcggt gacgaacgtt ttgtcctggc agagcgcgac aggcgcctgg 180
ccgaaaaaac tggacaccac ccgcgagcgc cgtcggcagg attccgcccc gcccgagggc 240
actttcgaca acggcgccac caccggcgag ttgcggttgc tggcgcgggc gtttcgggcc 300
accggcgatc cgcgctgcga agccggcggt ctccgggggc tggacggcat cctcgcgggc 360
cagcttccca gcggcggtgt gccgcagtgt catcctcgcg gcgcgcctta tcagcgccac 420
atcaccttca acgacggtgt catggtgcgc atcctggagc tgctgcgcga gatagaccgc 480
gcgcgggagt ttgcgtgggt ggacgaggcg cggcgcgcg gcggtgcgcg ggccttcaat 540
cgcggggtgt agtgctcctt gcgctgccag gtggtcgtcg agggcagact caccgtgtgg 600
tgtgcccagc atgacgcgga gaactttcaa ccgcgaccgc cagcgcccta cgaactggaa 660
tcgctcagcg gcgcggaag cgccggcctc ctggtgttcc tcatgagcct ggagcgcca 720
accgcggaga tcgcgcgcgc ggtcgaggcc gggcgcgccct ggttttcggc ggtaagctt 780
gaagggttcc gtctcgaacg aacggcgac gacgcgcggg ttgtggaaga gccggcgcg 840
ccgcgctctt gggcgcggtt ctacgagatc gggaccaatc gcccatctt tgccggtcgc 900
gacggtgtca agaagtacgc cctgagcgag atcgagcggg aacgcggggt cggctatgcg 960
tggtacggcg cctggggtga accggtcgcc cgccattatg cccagtggcg ggagcgttac 1020
gggacgcaga aatga 1035

```

<210> 70

<211> 344

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(22)

<221> DOMAIN

<222> (23)...(344)

<223> Catalytic domain

<400> 70

```

Met Ala Arg Leu Phe Arg Cys Val Cys Ala Ser Leu Gly Gly Trp Ala
1      5      10      15
Ala Val Leu Ala Ala Ala Ala Gly Pro Asp Trp Ser Arg Leu Leu Ala
20     25     30
Gln Pro Asp Pro Trp Phe Arg Ser Pro Ala Gly Gln Gln Ala Val Thr
35     40     45
Asn Val Leu Ser Trp Gln Ser Ala Thr Gly Ala Trp Pro Lys Asn Leu
50     55     60
Asp Thr Thr Arg Glu Pro Arg Arg Gln Asp Ser Ala Pro Pro Glu Gly
65     70     75     80
Thr Phe Asp Asn Gly Ala Thr Thr Gly Glu Leu Arg Phe Leu Ala Arg
85     90     95
Ala Phe Ala Ala Thr Gly Asp Pro Arg Cys Glu Ala Ala Val Leu Arg
100    105    110
Gly Leu Asp Gly Ile Leu Ala Ala Gln Leu Pro Ser Gly Gly Trp Pro
115    120    125
Gln Cys His Pro Pro Arg Ala Pro Tyr Gln Arg His Ile Thr Phe Asn
130    135    140
Asp Gly Val Met Val Arg Ile Leu Glu Leu Leu Arg Glu Ile Asp Arg
145    150    155    160
Ala Pro Glu Phe Arg Trp Val Asp Glu Ala Arg Arg Ala Arg Val Arg
165    170    175
Ala Ala Phe Thr Arg Gly Leu Glu Cys Leu Leu Arg Cys Gln Val Val
180    185    190
Val Glu Gly Arg Leu Thr Val Trp Cys Ala Gln His Asp Ala Glu Asn
195    200    205
Phe Gln Pro Arg Pro Ala Arg Ala Tyr Glu Leu Glu Ser Leu Ser Gly
210    215    220
Ala Glu Ser Ala Gly Ile Leu Val Phe Leu Met Ser Leu Glu Pro Pro
225    230    235    240
Thr Pro Glu Ile Ala Arg Ala Val Glu Ala Gly Ala Ala Trp Phe Ser
245    250    255
Ala Val Lys Leu Glu Gly Phe Arg Leu Glu Arg Thr Ala Asp Asp Ala
260    265    270
Arg Val Val Glu Glu Pro Gly Ala Pro Pro Leu Trp Ala Arg Phe Tyr
275    280    285
Glu Ile Gly Thr Asn Arg Pro Ile Phe Ala Gly Arg Asp Gly Val Lys
290    295    300
Lys Tyr Ala Leu Ser Glu Ile Glu Arg Glu Arg Val Gly Tyr Ala
305    310    315    320
Trp Tyr Gly Ala Trp Gly Glu Pro Val Ala Arg His Tyr Ala Gln Trp
325    330    335
Arg Glu Arg Tyr Gly Thr Gln Lys
340

```

<210> 71

<211> 1038

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 71

gtgactcgtg tcgcccttgc gatggggctt gttgcatggg ttccggcgct cgcttcagct 60

```

gggcccgtg catatttga gaagccggac gactggttcg ccagtccga ggcagggca 120
atcgcccgca acgtactcgc gcctcaggcc gatctcggcg ggtggccgaa gaacatcgac 180
acaacgaagc cgttcaccgg cgaccggacg caaatcaaac cgaccttcga taacagcgcg 240
acaaccgacg agtcccggtt tctggcgcg atccacaacg cgactcgcga cgagaagtac 300
cgaccccgct tcgagaagg gctcgattac atcttgaag cacaatacgc aaacggcggt 360
tggccgcagt cgacccggcc cggcaccggc taccaccggc acatcacctt caacgacaat 420
gccatggtcc gtttgatgga gctcgtgcgc gaagtccgga cctcgaatcg gtacgacttc 480
ctggacggcg accgcggcaa ggctcgccgc gccgctttcg atcgcgccat cgaatgcac 540
ctgaagtgcg agatcaaggc cgacagtaag ctgacggcat ggtcgcccca gcacgacgag 600
aaggacctcg ctcccgggcc ggccgggaac tacgagctcg tctcactcag cggctcggag 660
tcggtcggga tcgtccgcct actcatgagc ctgacgcgac caagcccgga ggtcgctcgg 720
gccatcgacg cgcgggtcgc gtggttcagg tcggcgaagc tcgaaggcac caaggtcgtt 780
gtcgagcgcg acccgaagta tccggggcgc cgggaacgcg tgggtgtgaa ggaaccaaag 840
gcaccggcac tctggggcgc tctctacgaa atcggcacga atcgcccat cttctccgac 900
cgcgacggca tcaagaagta cgcgctcgcc gagatcgccc ccgaacggcg gaatggctat 960
gcctggtatg gcacctggcc cgcgcacctg ctggagaagg aatacccagg gtggaaaaag 1020
aagctggccc ggccgtga 1038

```

<210> 72

<211> 345

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(20)

<221> DOMAIN

<222> (21)...(345)

<223> Catalytic domain

<400> 72

```

Met Thr Arg Val Ala Leu Ala Met Gly Leu Val Ala Trp Val Pro Ala
1      5      10      15
Leu Ala Ser Ala Gly Pro Ala Ala Tyr Leu Gln Lys Pro Asp Asp Trp
20     25     30
Phe Ala Ser Pro Glu Ala Arg Ala Ile Ala Ala Asn Val Leu Ala His
35     40     45
Gln Ala Asp Leu Gly Gly Trp Pro Lys Asn Ile Asp Thr Thr Lys Pro
50     55     60
Phe Thr Gly Asp Arg Thr Gln Ile Lys Pro Thr Phe Asp Asn Ser Ala
65     70     75     80
Thr Thr Asp Glu Leu Arg Phe Leu Ala Arg Ile His Asn Ala Thr Arg
85     90     95
Asp Glu Lys Tyr Arg Thr Ala Phe Glu Lys Gly Leu Asp Tyr Ile Leu
100    105    110
Lys Ala Gln Tyr Ala Asn Gly Gly Trp Pro Gln Ser His Pro Pro Gly
115    120    125
Thr Gly Tyr His Arg His Ile Thr Phe Asn Asp Asn Ala Met Val Arg
130    135    140
Leu Met Glu Leu Val Arg Glu Val Ala Thr Ser Asn Arg Tyr Asp Phe
145    150    155    160
Leu Asp Ala Asp Arg Lys Ala Cys Arg Ala Ala Phe Asp Arg Gly
165    170    175
Ile Glu Cys Ile Leu Lys Cys Gln Ile Lys Val Asp Ser Lys Leu Thr
180    185    190
Ala Trp Cys Ala Gln His Asp Glu Lys Asp Leu Ala Pro Arg Pro Ala

```

```

      195          200          205
Arg Thr Tyr Glu Leu Val Ser Leu Ser Gly Ser Glu Ser Val Gly Ile
210          215          220
Val Arg Leu Leu Met Ser Leu Asp Arg Pro Ser Pro Glu Val Ala Arg
225          230          235          240
Ala Ile Asp Gly Ala Val Ala Trp Phe Gln Ser Ala Lys Leu Glu Gly
245          250          255
Thr Lys Val Val Glu Arg Asp Pro Lys Tyr Pro Gly Gly Arg Glu
260          265          270
Arg Val Val Val Lys Asp Pro Lys Ala Pro Pro Leu Trp Ala Arg Phe
275          280          285
Tyr Glu Ile Gly Thr Asn Arg Pro Ile Phe Ser Asp Arg Asp Gly Ile
290          295          300
Lys Lys Tyr Ala Leu Ala Glu Ile Gly Pro Glu Arg Arg Asn Gly Tyr
305          310          315          320
Ala Trp Tyr Gly Thr Trp Pro Arg Asp Leu Leu Glu Lys Glu Tyr Pro
325          330          335
Gly Trp Lys Lys Lys Leu Ala Arg Pro
340          345

```

<210> 73

<211> 1221

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 73

```

atgctcacca aaacatcaact acttattgca ttgctaggca gttgttgtat cgcaccatta      60
catgctggaca caccagcaag caatgcacccg acaaccaatg catcaattcc gctacagcaa      120
actgctggagc atgctgcccgc ctggaaaaat tatctcgcca aatccaacga gttgctgcaaa      180
gcagaccagg cgagctcaa agccgagctg aaaaaactcg ggcaaaaac cgcgagtttg      240
cctgagtaca ccaagaatt tggttttgaa gtgaagcagt catctgagtg gtttaaaagc      300
actgaagcta aacgagtgat ggatattatc ctatcgtttc aaactccttc tggcggctgg      360
tcaaaacgca ctgacatgag caaagcgccg cgcaaacccg gccaggcatt tgggtgtgaa      420
aaaaattaca tccccacctt tgataatggc gcgaccagca cacaattaat gctactggca      480
caggcgcatc aagccactgg cgataaacgc tacagcgatg catttgccgc cgggcttgaa      540
tttatcatca ccgctcaata tcccaatggc ggctggccac aaaattttcc attggttggc      600
aagtatcacg atcacatcac ttacaacgat gccctgatgc gogatttaat ggtagtgcta      660
cacaaggttg ccatggccaa ggatgaattt gcccttgtat ccaaggcgca gcaacaggcc      720
gcacaagcga gcctcgaacg cgcgctggac tgcgttttga aaaccagggt gatggccaat      780
ggccaattaa ctatatgggg tgcgcagcac gatgccaaaa ccttaaaacc cgccaagcg      840
cgcgcctatg aaatgatttc actcaccagt tctgaaagcg tgtgatgct cgatttttta      900
atggatttgc aacagcccaag cgctgacatt attaaatccg tgcacgcggc tgcgcttgg      960
tatgagcaaa ataaaattat cggaaaaacc tggacccggg gcgacacagt tctgaaagac      1020
gataaggatg caccgccaat ctgggcgctg ttttatgaga taggtacgaa caaacccctg      1080
tttggcgacc cgcgatctc tgtccattac gatctggcaa aggtatcgga agagcgccgc      1140
acgggttatg cctggtacac aacctcacc aatcaggtat taaaaaagta cgcgcgctgg      1200
gctaaacaat atccgcaata a

```

<210> 74

<211> 406

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(22)

<221> DOMAIN

<222> (23)...(406)

<223> Catalytic domain

<400> 74

```

Met Leu Thr Lys Thr Ser Leu Leu Ile Ala Leu Leu Gly Ser Cys Cys
1      5      10      15
Ile Ala Pro Leu His Ala Asp Thr Pro Ala Ser Asn Ala Pro Thr Thr
20      25      30
Asn Ala Ser Ile Pro Leu Gln Gln Thr Ala Ser Asp Ala Ala Trp
35      40      45
Lys Asn Tyr Leu Ala Lys Ser Asn Glu Leu Arg Lys Ala Asp Gln Ala
50      55      60
Gln Leu Lys Ala Glu Leu Lys Lys Leu Gly Gln Lys Thr Ala Ser Leu
65      70      75      80
Pro Glu Tyr Thr Lys Glu Phe Gly Phe Glu Val Lys Gln Ser Ser Glu
85      90      95
Trp Phe Lys Ser Thr Glu Gly Lys Arg Val Met Asp Ile Ile Leu Ser
100     105     110
Phe Gln Thr Pro Ser Gly Gly Trp Ser Lys Arg Thr Asp Met Ser Lys
115     120     125
Ala Pro Arg Lys Pro Gly Gln Ala Phe Gly Val Glu Lys Asn Tyr Ile
130     135     140
Pro Thr Phe Asp Asn Gly Ala Thr Ser Thr Gln Leu Met Leu Leu Ala
145     150     155     160
Gln Ala His Gln Ala Thr Gly Asp Lys Arg Tyr Ser Asp Ala Phe Ala
165     170     175
Arg Gly Leu Glu Phe Ile Ile Thr Ala Gln Tyr Pro Asn Gly Gly Trp
180     185     190
Pro Gln Asn Phe Pro Leu Val Gly Lys Tyr His Asp His Ile Thr Tyr
195     200     205
Asn Asp Ala Leu Met Arg Asp Leu Met Val Val Leu His Lys Val Ala
210     215     220
Met Ala Lys Asp Glu Phe Ala Phe Val Ser Lys Ala Gln Gln Gln Ala
225     230     235     240
Ala Gln Ala Ser Leu Glu Arg Ala Leu Asp Cys Val Leu Lys Thr Gln
245     250     255
Val Met Ala Asn Gly Gln Leu Thr Ile Trp Gly Ala Gln His Asp Ala
260     265     270
Lys Thr Leu Lys Pro Ala Lys Ala Arg Ala Tyr Glu Met Ile Ser Leu
275     280     285
Thr Ser Ser Glu Ser Val Trp Met Leu Asp Phe Leu Met Asp Leu Gln
290     295     300
Gln Pro Ser Ala Asp Ile Ile Lys Ser Val His Ala Ala Ala Trp
305     310     315     320
Tyr Glu Gln Asn Lys Ile Ile Gly Lys Thr Trp Thr Arg Gly Asp Thr
325     330     335
Val Leu Lys Asp Asp Lys Asp Ala Pro Pro Ile Trp Ala Arg Phe Tyr
340     345     350
Glu Ile Gly Thr Asn Lys Pro Leu Phe Gly Asp Arg Asp Ser Val
355     360     365
His Tyr Asp Leu Ala Lys Val Ser Glu Glu Arg Arg Thr Gly Tyr Ala
370     375     380
Trp Tyr Thr Thr Ser Pro Asn Gln Val Leu Lys Lys Tyr Ala Arg Trp
385     390     395     400
Ala Lys Gln Tyr Pro Gln

```

405

<210> 75

<211> 1767

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 75

```

atgaccacaa cccgccgcac tatcctgaaa gccgccgcca gccgccggcg gatcgccagc      60
accggctggc ccgcttggc cgcgcacag gccgcgcaag ccgccgaccc gtgggcccgc      120
gccagcaga tcctcgaccg cttcgccaag ccgctcagct tcccgaacag ggacttcccg      180
atcaccagat tcggcgccaa accctgcaag ctggtcaaaag ccagggcctt ggtcgaagta      240
agagtcaaa gctgaactcga aacgccagca ccgcaagcgc cggacgccta cccggcaatc      300
aaagccgcca tcggcgcgac gagcaaggcc ggaggaggcg gcgtgctgat cccggccggc      360
aactggtaact ccaagggcc tctcgtgctg ctgtcgaacg tgcacgtgca ccttgccaag      420
ggcgcgcaag tctacttcag cgcacaagcc aaggacttcg cccgcgacgg cgactacgac      480
tgccggcgcca accgcaagct ggtgctctcg cgtgggcaag gcaacgattg cctgaacttc      540
tcgcccatgg tctacgcgcg cgggcaaaaag aatatcgcca ttaccggcga agactggacc      600
agcatcctga accgccaagg cggcgtggcg ttcgaagacg gcagcgccaa tggctgggtg      660
ggcatgaacc ccgcccggcg gccgcccgcc agcaccacgc accaggcgcg agccaatccg      720
aacaacgccg aggagccaat cccagactg cccacgcgcc acgcgaactg gagcgccgac      780
gacaagtacc tgccgctgct gtccgaagcc ggcgtgcccg ccgagcgccg cgtgttcggt      840
ctggggcact acctgcggcc gtcgatggtc gaattcgtcg actgcgggga tgtgtgatg      900
cagggtacc aggtcatcaa caccgcgttc tggattcctc acccggtcaa ctacgcgaac      960
attcaactct ccaaagtgcg catggaagc atcgcccgca attcgacgg tttcgatccc      1020
gagtcctgcg acaccatcct ggtggacggc tgcctgttca ataccggcg cgactgcac      1080
gccatcaaat ccggcaagaa ccgagactcg caatacggcc caacgcgcaa tatggtggtc      1140
cagaactgca tcatgaacgg cggccacggc ggcgttacgc tgggcagcga aatggcgggt      1200
ggcatcgagc atatctacgc gcagaaaatc gaattccgca acgcgttctg ggaccacgac      1260
ccgctgggca ccggcatccg aatgaagacg aacatgaacc cggcgggcta ccttcgtcat      1320
ttctacgtgc gcgacgtgac gctgccgaat ggcgtgcgta ccaagagcgg cttctacaag      1380
acgctgcggg gatctccgct ggagggcaag gtctccacca gcggcgggcg tgttatcact      1440
atcgactcgc attacgcgcc gaatgacgac agcgtgcggc tgcggccgcc gcagggtgctg      1500
gacgtgcata tctgaaacgt ccgctcagc aatgtgaaaa cggccgaagg ctggttctcc      1560
tgetaccagg ccatggtgct gctcggggcc gtggcgcca gcttcaacgg cgcgcctggc      1620
acggccatcc tgccgatcac gaatgtcacc gtcagcgatt cggacttcgg cagcccgcg      1680
aacagcgag agccctggtt cgcgttcaac gtcaggggac tcaagctcgg caacgtgcgc      1740
atcgatggca aggagtacaa cgtatga      1767

```

<210> 76

<211> 588

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(34)

<221> DOMAIN

<222> (110)...(555)

<223> Catalytic domain

<400> 76

Met Thr Thr Thr Arg Arg Thr Ile Leu Lys Ala Ala Ala Ser Ala Gly

```

1      5      10      15
Ala Ile Ala Ser Thr Gly Trp Pro Ala Leu Ala Ala Gln Ala Ala
20
Gln Ala Ala Asp Pro Trp Ala Arg Ala Gln Gln Ile Ile Asp Arg Phe
35      40      45
Ala Lys Pro Leu Ser Phe Pro Asn Arg Asp Phe Pro Ile Thr Glu Phe
50      55      60
Gly Ala Lys Pro Cys Lys Leu Val Lys Ala Gln Gly Leu Val Glu Val
65      70      75      80
Arg Val Lys Gly Glu Leu Glu Thr Pro Ala Pro Gln Ala Pro Asp Ala
85      90      95
Tyr Pro Ala Ile Lys Ala Ala Ile Ala Ala Ala Ser Lys Ala Gly Gly
100      105      110
Gly Arg Val Leu Ile Pro Ala Gly Asn Trp Tyr Cys Lys Gly Pro Ile
115      120      125
Val Leu Leu Ser Asn Val His Val His Leu Ala Lys Gly Ala Gln Val
130      135      140
Tyr Phe Ser Ala Asn Ala Lys Asp Phe Ala Arg Asp Gly Asp Tyr Asp
145      150      155      160
Cys Gly Ala Asn Gly Lys Leu Val Leu Ser Arg Trp Gln Gly Asn Asp
165      170      175
Cys Leu Asn Phe Ser Pro Met Val Tyr Ala Arg Gly Gln Lys Asn Ile
180      185      190
Ala Ile Thr Gly Glu Asp Trp Thr Ser Ile Leu Asn Gly Gln Ala Gly
195      200      205
Val Ala Phe Glu Asp Gly Ser Gly Asn Gly Trp Trp Gly Met Asn Pro
210      215      220
Ala Gly Ala Pro Pro Gly Ser Thr Thr His Gln Gly Ala Ala Asn Pro
225      230      235      240
Asn Asn Ala Glu Glu Pro Ile Ala Arg Leu Pro Thr Arg His Ala Asn
245      250      255
Trp Ser Ala Asp Asp Lys Tyr Leu Pro Leu Leu Ser Glu Ala Gly Val
260      265      270
Pro Ala Glu Arg Arg Val Phe Gly Leu Gly His Tyr Leu Arg Pro Ser
275      280      285
Met Val Glu Phe Val Asp Cys Gly Asp Val Leu Met Gln Gly Tyr Gln
290      295      300
Val Ile Asn Thr Pro Phe Trp Ile His His Pro Val Asn Ser Arg Asn
305      310      315      320
Ile His Phe Ser Lys Val Arg Met Glu Ser Ile Gly Pro Asn Ser Asp
325      330      335
Gly Phe Asp Pro Glu Ser Cys Asp Thr Ile Leu Val Asp Gly Cys Leu
340      345      350
Phe Asn Thr Gly Asp Asp Cys Ile Ala Ile Lys Ser Gly Lys Asn Arg
355      360      365
Asp Ser Gln Tyr Gly Pro Thr Arg Asn Met Val Val Gln Asn Cys Ile
370      375      380
Met Asn Arg Gly His Gly Gly Val Thr Leu Gly Ser Glu Met Ala Gly
385      390      395      400
Gly Ile Glu His Ile Tyr Ala Gln Lys Ile Glu Phe Arg Asn Ala Phe
405      410      415
Trp Asp His Asp Pro Leu Gly Thr Ala Ile Arg Met Lys Thr Asn Met
420      425      430
Asn Arg Gly Gly Tyr Leu Arg His Phe Tyr Val Arg Asp Val Thr Leu
435      440      445
Pro Asn Gly Val Arg Thr Lys Ser Gly Phe Tyr Lys Thr Leu Pro Gly
450      455      460
Ser Pro Leu Ala Gly Lys Val Ser Thr Ser Gly Gly Ala Val Ile Thr
465      470      475      480

Ile Asp Cys Asp Tyr Ala Pro Asn Asp Asp Ser Val Arg Val Arg Pro
485      490      495
Pro Gln Val Ser Asp Val His Ile Ser Asn Val Arg Val Ser Asn Val
500      505      510
Lys Thr Ala Glu Gly Ser Phe Ser Cys Tyr Gln Ala Met Val Leu Leu
515      520      525
Gly Pro Val Ala Ala Ser Phe Asn Gly Ala Pro Gly Thr Ala Ile Leu
530      535      540
Pro Ile Thr Asn Val Thr Val Ser Asp Ser Asp Phe Gly Thr Pro Arg
545      550      555      560
Asn Ser Ala Glu Pro Trp Phe Ala Phe Asn Val Gln Gly Leu Lys Leu
565      570      575
Arg Asn Val Arg Ile Asp Gly Lys Glu Tyr Asn Val
580      585

```

<210> 77

<211> 2043

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 77

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atgaaaaact ccagagcaat ttttactaca tcaacacttt tacacggcgc gcttatcgcg 60
gctagtgtca gcatggcaat gagttctgcc gcatgggcgg gttgtacctt taccgtcacc 120
aataattggg gctcaggatt tacccggcga atcaaatga ccaacaacac caccgccagc 180
gtgaacaatt ggtctgtgtc atggcaggaa tccggtgcgg ctatcaccaa cgcttggaat 240
gcaacgctca gtggctcaaa cccttacaca gccgtatccg ctggttggaa tggcacactt 300
gcccccattg catcgccac ttttggtttc caggcaaacg gttctgccgg tgcacctaaa 360
gtgaattggc gcttctgcgg caccacact tcatacaac cggcatccag cagtgttgcc 420
agctcggtta aatcaagcgc gccgtatcgc tccagcagca gatcatccag ttcaatcgct 480
atcactagca gctctttagc gagaagtctt attgcctcca gcagctcact agttagtagc 540
tccagagcga gcagttagtc gccaaagcgtt ttctctttta cgatccagga agagcaagcg 600
ggcttctgtc gtgttgatgg cattgcgaca gaaagcacca acaccgggtt taccggcaat 660
ggctacacca atgcgaacaa cgcgcaaggc gcagcgattg aatgggcagt cagcgcacct 720
agcagtgggc gttatacagt agccttcgcg ttcggcaatg gcggcacagc agcgcgaac 780
ggctcgttgt taatcaatgg cggtagcaat ggtaattaca ctgtggagtt acccctgacc 840
ggcgcatggg caacctggca aattgccagc gtggaattg atttagtgca aggcataat 900
attttaaaac tctcgcggtt aaccgctgac ggtttggcca atatcgactc attaaaaata 960
gacggcgccg aaaccaaagc aggtacttgc agcactacat caagcagcag cgttgccagc 1020
agctcgtcgt ccgttaaatc cagcgcaagt tcttcttcga gttcatccac cgctgcaaaa 1080
atactgacat tagacggtaa ccggcgccgc agctggttca acaaatccag gaccaagtgg 1140
aatagcagcg gcgcgcatat tgtgtgtct taccagcaat ccaacggcgg ttggcctaaa 1200
aacctggatt acaactcagt gacgcagggc aatggcggga gcgacagcgg caccatcgac 1260
aatggtgcaa ccattaccga aatggtttac ctgctgaaa tttataaaaa cggcggaac 1320
accaaatatc gcgatgcagt gcgcagagca gcaaaacttt tagtgagctc gcaatacagc 1380
acaggcgctt tgccacaatt ttatccgttg aaaggcggtc atgcggatca tgcgaccttt 1440
aacgataacg gcattggcgtg cgcgttgacg gtattggatt tcgcagtaaa caaacgcgca 1500
ccgtttgata acgacatctt ctctgattct gatcggcgca aattcaaaac cgctgttgcc 1560
aaaggtgtgg attacatctt aaaagcgagc tggaaacaaa atggaaaact cactgcatgg 1620
tgtgcacaac acggtgctac ggattaccaa cggaaaaaag cgcgcgctta tgaattggaa 1680
tcattgagtg gtagcagtcg ggtcggcatt ctgcctctct tgatgaccca accacaaacc 1740
gcgcataatg aagcgcggtt caaggcggtt gtcactggt tcgccagtcc aaatacttat 1800
ttggttaact acacttacga ttcatcaaaa gcgtctacca acccgattgt gtataaatcc 1860
ggaagcagaa tgtggtatcg ctctatgac ctgaacacca accgtggttt ctttagtgat 1920
cgcgatggca gcaaatctta tgatatcacc caaatgtcag aagagcgctg caccggttat 1980
agctggggtg gctcttacgg tgaatctatt atttccttcg cgcaaaaagt gggttatctg 2040
taa 2043

```

<210> 78

<211> 680

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(33)

<221> BINDING

<222> (34)...(126)

<223> Carbohydrate binding module

<221> BINDING

<222> (199)...(322)

<223> Carbohydrate binding module

<221> DOMAIN

<222> (323)...(680)

<223> Catalytic domain

<400> 78

```

Met Lys Thr Ser Arg Ala Ile Phe Thr Thr Ser Thr Leu Leu His Arg
1      5      10      15
Ala Leu Ile Ala Ala Ser Val Ser Met Ala Met Ser Ser Ala Ala Trp
20      25      30
Ala Gly Cys Thr Tyr Thr Val Thr Asn Asn Trp Gly Ser Gly Phe Thr
35      40      45
Gly Glu Ile Lys Val Thr Asn Asn Thr Thr Ala Ser Val Asn Asn Trp
50      55      60
Ser Val Ser Trp Gln Glu Ser Gly Ala Ala Ile Thr Asn Ala Trp Asn
65      70      75      80
Ala Thr Leu Ser Gly Ser Asn Pro Tyr Thr Ala Val Ser Ala Gly Trp
85      90      95
Asn Gly Thr Leu Ala Pro Asn Ala Ser Ala Thr Phe Gly Phe Gln Ala
100     105     110
Asn Gly Ser Ala Gly Ala Pro Lys Val Asn Gly Ser Leu Cys Gly Thr
115     120     125
Asn Thr Ser Ser Thr Pro Ala Ser Ser Ser Val Ala Ser Ser Val Lys
130     135     140
Ser Ser Ala Pro Val Ser Ser Ser Arg Ser Ser Ser Ser Ile Ala
145     150     155     160
Ile Thr Ser Ser Ser Leu Ala Arg Ser Ser Ile Ala Ser Ser Ser Ser
165     170     175
Leu Val Ser Ser Ser Arg Ala Ser Ser Ser Ala Pro Ser Val Phe Ser
180     185     190
Phe Thr Ile Gln Glu Glu Gln Ala Gly Phe Cys Arg Val Asp Gly Ile
195     200     205
Ala Thr Glu Ser Thr Asn Thr Gly Phe Thr Gly Asn Gly Tyr Thr Asn
210     215     220
Ala Asn Asn Ala Gln Gly Ala Ala Ile Glu Trp Ala Val Ser Ala Pro
225     230     235     240
Ser Ser Gly Arg Tyr Thr Val Ala Phe Arg Phe Ala Asn Gly Gly Thr
245     250     255
Ala Ala Arg Asn Gly Ser Leu Leu Ile Asn Gly Gly Ser Asn Gly Asn
260     265     270

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Tyr Thr Val Glu Leu Pro Leu Thr Gly Ala Trp Ala Thr Trp Gln Ile
275     280     285
Ala Ser Val Glu Ile Asp Leu Val Gln Gly Asn Asn Ile Leu Lys Leu
290     295     300
Ser Ala Leu Thr Ala Asp Gly Leu Ala Asn Ile Asp Ser Leu Lys Ile
305     310     315     320
Asp Gly Ala Gln Thr Lys Ala Gly Thr Cys Ser Thr Thr Ser Ser Ser
325     330     335
Ser Val Ala Ser Ser Ser Ser Val Lys Ser Ser Ala Ser Ser Ser
340     345     350
Ser Ser Ser Ser Thr Ala Ala Lys Ile Leu Thr Leu Asp Gly Asn Pro
355     360     365
Ala Ala Ser Trp Phe Asn Lys Ser Arg Thr Lys Trp Asn Ser Ser Arg
370     375     380
Ala Asp Ile Val Leu Ser Tyr Gln Gln Ser Asn Gly Gly Trp Pro Lys
385     390     395     400
Asn Leu Asp Tyr Asn Ser Val Ser Ala Gly Asn Gly Gly Ser Asp Ser
405     410     415
Gly Thr Ile Asp Asn Gly Ala Thr Ile Thr Glu Met Val Tyr Leu Ala
420     425     430
Glu Ile Tyr Lys Asn Gly Gly Asn Thr Lys Tyr Arg Asp Ala Val Arg
435     440     445
Arg Ala Ala Asn Phe Leu Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu
450     455     460
Pro Gln Phe Tyr Pro Leu Lys Gly Gly Tyr Ala Asp His Ala Thr Phe
465     470     475     480
Asn Asp Asn Gly Met Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala Val
485     490     495
Asn Lys Arg Ala Pro Phe Asp Asn Asp Ile Phe Ser Asp Ser Asp Arg
500     505     510
Ala Lys Phe Lys Thr Ala Val Ala Lys Gly Val Asp Tyr Ile Leu Lys
515     520     525
Ala Gln Trp Lys Gln Asn Gly Lys Leu Thr Ala Trp Cys Ala Gln His
530     535     540
Gly Ala Thr Asp Tyr Gln Pro Lys Lys Ala Arg Ala Tyr Glu Leu Glu
545     550     555     560
Ser Leu Ser Gly Ser Glu Ser Val Gly Ile Leu Ala Phe Leu Met Thr
565     570     575
Gln Pro Gln Thr Ala Gln Ile Glu Ala Ala Val Lys Ala Gly Val Asn
580     585     590
Trp Phe Ala Ser Pro Asn Thr Tyr Leu Ala Asn Tyr Thr Tyr Asp Ser
595     600     605
Ser Lys Ala Ser Thr Asn Pro Ile Val Tyr Lys Ser Gly Ser Arg Met
610     615     620
Trp Tyr Arg Phe Tyr Asp Leu Asn Thr Asn Arg Gly Phe Phe Ser Asp
625     630     635     640
Arg Asp Gly Ser Lys Phe Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg
645     650     655
Arg Thr Gly Tyr Ser Trp Gly Gly Ser Tyr Gly Glu Ser Ile Ile Ser
660     665     670
Phe Ala Gln Lys Val Gly Tyr Leu
675     680

```

<210> 79

<211> 1746

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 79

```

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gctccgcggt taattgcgtt cggcgcgag gcaagtcctt gggaacgat gatgccttcg 120
atcctcgcac gcatcacacc acctcgtttt cgaaacgca cttctatct caatcgattc 180
ggcgccaagg cggatggagt cacagactgc acccggtt ttcatcgcg gatcgatgaa 240
tgcaccaaa cggcggtgg gaaagtcgtc gtgcccggc gcacttatct caccggcgcg 300
attcatttga agagcaacgt caacctcgaa gtctcggaag gcgcgacgat caagttcagt 360
caggaccoga aacactacct gcctgtgtc ttctcggtt gggaagggt cgaagtcttc 420
aactactcgc ctttcattta cgcgttcgaa cagcgaaca tcgcatcac cggcaaaagg 480
acgctcgacg gacagagtga ttcggaacac tggtgccgt ggaaacggcg tccgcagtac 540
ggatggaatg aaggatgaa acagcagcgt cccgatcgca acgcgttgtt cacaatggcg 600
gagaaaggcg tgcggtgcg cgagcgcatc ttggcgaa gtcattattt gaggcccgag 660
ttcattcagc cgtaccgctg ccagaacgtg ctgatccagg gcgtgacgat tcggaactcg 720
ccgatgtggg agattcatcc ggtgtgtgc cgaacgtga ctattcacga cgtgcacatc 780
gatagtcatg gaccaaaca cgcggtctgc aatccgaat cgtgcagcga cgtgttgatt 840
aaggatagct acttcgatac cggcgacgac tgcacgcga tcaaatcggg acgcaacgcc 900
gacgggcggc ggcttaaagc gccgactgag aacatcatcg ttcaaggatg tcgcatgaaa 960
gacggccacg gtggaatcac ggtcggcagc gagatctcgg cggcggtgcg aaacctgttt 1020
gccgagaatt gccgggtcga cagtccaaac ctgatcacg cctgcgcgt gaagaacaat 1080
gccatgcgcg cgggattact cgagaacttc cacttcgta acatcgaagt cgggcaggtg 1140
gccatgcgcg tgattacgat cgacttcaac tacgaagagg gcgcgaaagg gtctgtcacg 1200
ccggtcgttc cgcattacac ggtcgatcgt ttgcgcagca cgaagagcaa gcacgcactc 1260
gacgtccagg gtctgccgg cgccgggtc atcaacctgc gattgacaaa ctgcacattc 1320
aacgatgtgc agcaaccgaa cattctcaag aacgtcgaa aatcaacct tgagaacgtc 1380
acgattaacg gaaagacgat cacacaaaca ggatccatct cagaagagc ggccacgaca 1440
gcaatgaccg cgctttgcg cgacgcgtc aggaagaaa acggttatcc cgcgaagtgg 1500
acctatgac atgggctggt cctgaaagga atcgagcgcg ttgggaacaa taccggcgat 1560
aagaagtatc tgaagttcat ccaggacagc atggaccact tcgtcaacga cgcgggtcc 1620
attcgacct acacgatoga cgagtacaac atcgatcacg ttcttccgg acgaaacctc 1680
ctgttccttt acaaaactac cgtcaggaa aagtatcgca aagccgcgcg gttcttgcg 1740
gaacaa 1746

```

<210> 80

<211> 582

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(28)

<221> DOMAIN

<222> (81)...(458)

<223> Catalytic domain

<400> 80

```

Met Thr Thr Arg Glu Phe Ile Lys Gly Phe Leu Leu Thr Gly Ala
1      5      10      15
Ala Val Ala Val Ala Pro Arg Leu Ile Ala Phe Gly Ala Glu Ala Ser
20     25     30
Pro Trp Glu Thr Met Met Pro Ser Ile Leu Ala Arg Ile Thr Pro Pro
35     40     45
Arg Phe Pro Lys Arg Thr Phe Tyr Leu Asn Arg Phe Gly Ala Lys Gly
50     55     60
Asp Gly Val Thr Asp Cys Thr Ala Ala Phe His Arg Ala Ile Asp Glu

```

```

65      70      75      80
Cys Thr Lys Ala Gly Gly Lys Val Val Val Pro Ala Gly Thr Tyr
      85      90      95
Leu Thr Gly Ala Ile His Leu Lys Ser Asn Val Asn Leu Glu Val Ser
      100      105      110
Glu Gly Ala Thr Ile Lys Phe Ser Gln Asp Pro Lys His Tyr Leu Pro
      115      120      125
Val Val Phe Ser Arg Trp Glu Gly Val Glu Val Phe Asn Tyr Ser Pro
      130      135      140
Phe Ile Tyr Ala Phe Glu Gln Arg Asn Ile Ala Ile Thr Gly Lys Gly
      145      150      155      160
Thr Leu Asp Gly Gln Ser Asp Ser Glu His Trp Trp Pro Trp Asn Gly
      165      170      175
Arg Pro Gln Tyr Gly Trp Asn Glu Gly Met Lys Gln Gln Arg Pro Asp
      180      185      190
Arg Asn Ala Leu Phe Thr Met Ala Glu Lys Gly Val Pro Val Arg Glu
      195      200      205
Arg Ile Phe Gly Glu Gly His Tyr Leu Arg Pro Gln Phe Ile Gln Pro
      210      215      220
Tyr Arg Cys Gln Asn Val Leu Ile Gln Gly Val Thr Ile Arg Asn Ser
      225      230      235      240
Pro Met Trp Glu Ile His Pro Val Leu Cys Arg Asn Val Thr Ile His
      245      250      255
Asp Val His Ile Asp Ser His Gly Pro Asn Asn Asp Gly Cys Asn Pro
      260      265      270
Glu Ser Cys Ser Asp Val Leu Ile Lys Asp Ser Tyr Phe Asp Thr Gly
      275      280      285
Asp Asp Cys Ile Ala Ile Lys Ser Gly Arg Asn Ala Asp Gly Arg Arg
      290      295      300
Leu Lys Ala Pro Thr Glu Asn Ile Ile Val Gln Gly Cys Arg Met Lys
      305      310      315      320
Asp Gly His Gly Gly Ile Thr Val Gly Ser Glu Ile Ser Gly Gly Val
      325      330      335
Arg Asn Leu Phe Ala Glu Asn Cys Arg Leu Asp Ser Pro Asn Leu Asp
      340      345      350
His Ala Leu Arg Val Lys Asn Asn Ala Met Arg Gly Gly Leu Leu Glu
      355      360      365
Asn Phe His Phe Arg Asn Ile Glu Val Gly Gln Val Ala His Ala Val
      370      375      380
Ile Thr Ile Asp Phe Asn Tyr Glu Glu Gly Ala Lys Gly Ser Phe Thr
      385      390      395      400
Pro Val Val Arg Asp Tyr Thr Val Asp Arg Leu Arg Ser Thr Lys Ser
      405      410      415
Lys His Ala Leu Asp Val Gln Gly Leu Pro Gly Ala Pro Val Ile Asn
      420      425      430
Leu Arg Leu Thr Asn Cys Thr Phe Asn Asp Val Gln Gln Pro Asn Ile
      435      440      445
Leu Lys Asn Val Glu Gln Ser Thr Phe Glu Asn Val Thr Ile Asn Gly
      450      455      460
Lys Thr Ile Thr Gln Thr Gly Ser Ile Ser Glu Arg Ala Ala Thr Thr
      465      470      475      480
Ala Met Thr Ala Leu Trp Arg Asp Ala Ser Arg Lys Glu Asn Gly Tyr
      485      490      495
Pro Ala Lys Trp Thr Tyr Asp His Gly Leu Val Leu Lys Gly Ile Glu
      500      505      510
Arg Val Trp Asn Asn Thr Gly Asp Lys Lys Tyr Leu Lys Phe Ile Gln
      515      520      525
Asp Ser Met Asp His Phe Val Asn Asp Asp Gly Ser Ile Arg Thr Tyr
      530      535      540

Thr Ile Asp Glu Tyr Asn Ile Asp His Val Leu Pro Gly Arg Asn Leu
545      550      555      560
Leu Phe Leu Tyr Lys Thr Thr Gly Gln Glu Lys Tyr Arg Lys Ala Ala
      565      570      575
Ala Phe Leu Arg Glu Gln
      580

```

<210> 81

<211> 1065

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 81

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ctgctgcaga ccgctccggg tggctggtcc aagcactacc gcgggaagaa ggtcgactac 180
acgcgcaatt acgacaccgc cagcgcgcgc cgcctgcgcg cgcgcgcgcg gcatgacgac 240
gcgacgacgc acaacaaggc caccgaccgc gagatcgctt acctggtgca ggcacatgcc 300
aggacgggca acccggcgta cctcgacggt gcccgccgcg gggtcgaata cctgctgccc 360
gcgcagttacc cgaatgggtg ctggccgcag ttctaccccg accactcgtc ctaccggcac 420
cagatcacgc tcaacgacga cgcgatggtg catgccatca ccgtgctgca ggacatcgcc 480
gccggcgccg acggcgtgca ggcgttgacg cccgagttcg gcgtccgcgc cgcgcgcgcc 540
gcgcagcgcg gcatcggaac cctgctcgag ttgcaggtgc gcatcgacgg cgagccgacc 600
atctgggcgg cgcagttacg cagcgtatgc ctgcagccgg ccaaggcccg cgcctatgaa 660
ctgcccctgc tggccgtggc cgaatcggtc ggcgtggtgc gcctgctgat gcgccagccg 720
aggccggatg cccggaccgt cgcgcgcatc gaatcgccgg cccgctggct ggaggcgcat 780
cgctcgatg acctggcgct cgaacgcgtc gacgcaccgg ccgaggaaac gggcaaggac 840
gtcggggtcg tgaccgggcc cggcgctcgc ctgtgggcgc gcttctacga cctggatgga 900
cagcagcctc tggctcgtga cgcgcgacgc aagcccgctc cgttcgccag cctgcccaac 960
gagcgcgcga ccgctatgc ctggtagcgc acctggccgg agaagctgct ggcgcaggaa 1020
ctccgcgctt ggcgcgaggt ccatgccgcc ggcgcgcgcg cctga 1065

```

<210> 82

<211> 354

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(30)

<221> DOMAIN

<222> (31)...(354)

<223> Catalytic domain

<400> 82

```

Met Thr Leu Pro Val Ser Leu Arg Val Leu Leu Ala Leu Leu Ala
 1          5          10          15
Thr Leu Pro Val Ala Cys Ala Gly Ala Ala Val Ser Ala Ala Ala Thr
 20          25          30
Asp Pro Val Ala Glu Asn Met Leu Leu Leu Gln Thr Ala Ser Gly Gly
 35          40          45
Trp Ser Lys His Tyr Arg Gly Lys Lys Val Asp Tyr Thr Arg Asn Tyr
 50          55          60

Asp Thr Ala Glu Arg Ala Ala Leu Arg Ala Pro Gly Arg His Asp Asp
 65          70          75          80
Ala Thr Ile Asp Asn Lys Ala Thr Thr Ser Glu Ile Ala Tyr Leu Val
 85          90          95
Gln Ala His Ala Arg Thr Gly Asn Pro Ala Tyr Leu Asp Gly Ala Arg
100          105          110
Arg Gly Val Glu Tyr Leu Leu Arg Ala Gln Tyr Pro Asn Gly Gly Trp
115          120          125
Pro Gln Phe Tyr Pro Asp His Ser Ser Tyr Arg His Gln Ile Thr Leu
130          135          140
Asn Asp Asp Ala Met Val His Ala Ile Thr Val Leu Gln Asp Ile Ala
145          150          155          160
Ala Gly Arg Asp Gly Met Gln Ala Leu Thr Pro Glu Phe Gly Val Arg
165          170          175
Ala Ala Ala Ala Ala Gln Arg Gly Ile Gly Asn Leu Leu Glu Leu Gln
180          185          190
Val Arg Ile Asp Gly Glu Pro Thr Ile Trp Ala Ala Gln Tyr Asp Glu
195          200          205
His Ser Leu Gln Pro Ala Lys Ala Arg Ala Tyr Glu Leu Pro Ser Leu
210          215          220
Ala Val Ala Glu Ser Val Gly Val Val Arg Leu Leu Met Arg Gln Pro
225          230          235          240
Arg Pro Asp Ala Arg Thr Val Ala Ala Ile Glu Ser Ala Ala Arg Trp
245          250          255
Leu Glu Ala His Arg Leu His Asp Leu Ala Leu Glu Arg Val Asp Ala
260          265          270
Pro Ala Glu Glu Thr Gly Lys Asp Val Arg Val Val Thr Arg Pro Gly
275          280          285
Ala Ser Leu Trp Ala Arg Phe Tyr Asp Leu Asp Gly Gln Gln Pro Leu
290          295          300
Phe Val Asp Arg Asp Ser Lys Pro Val Pro Phe Ala Ser Leu Pro Asn
305          310          315          320
Glu Arg Arg Thr Gly Tyr Ala Trp Tyr Gly Thr Trp Pro Glu Lys Leu
325          330          335
Leu Ala Gln Glu Leu Pro Arg Trp Arg Glu Val His Ala Ala Gly Ala
340          345          350
Ala Pro

```

<210> 83

<211> 3618

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 83

```

ttgactgctc tttcaagaaa cagtcagggt gatgtaagct gggaaccgag ttccgcccac 60
caggtaacct acaatctaaa acgcagtacc acgaaggagg gtcccttatca gaccattgct 120
gaaaaaatgg cagaaccoga ctccgggat acagggttag agaattggcca gaagtattac 180
tatgttgatc ctgccgaaac gactagcggt gagagtgcag attcacaagc tataacggct 240
gtgcctgtag cgccattgca agctccgacc ggcctttcag caagtcatgg caatggcggg 300
gtaaccattc attgggaatc cgtcaatggt gccgagtcct accaagtctt gcgcagtaaa 360
caaaagggca tcggctatga agtcatcaaa aacggtgtaa cggaaccag ttatacagat 420
accgggattc ccgatggcga gaagtactat tatgtcgtat ccgccaagaa cgatacagct 480
gaaagtgcaa attcccaacc gattaacgggt gctgctgtat cgacgagtgg tgtaccagcc 540
attccaaacg gtatgaacgc tactgccggt gatggcagag ctgccttaac ctggtccgct 600
gtatccggcg cagattccta tagcatcaag cgcggtgagt ttaacagtgg tcaatatgag 660

```

```

gtcattgcta aaaaatataca ctctaccggt tatcaagata taggccttac aaacgggtgat 720
acctatgatt atgtgatttc cgctgtcaat gagcaagggg aaagttttagg ctccgaacc 780
atcgccgtta ctctcgcgaa agtaacgggt gtacggaag aaggcggaga ctttaaacg 840
attcaagaag ccatttgatgc tgcacctgat aacagtacga aacggcatgt tatttttata 900
aaaaatggtc aatatcgtga aaagcttaac atccctaaga gcaaaaccaa tctgagtttt 960
gtaggggaaa gtaagggaag gaccgtgctt gtttttaatg ataatgcaaa tacgcctggg 1020
ccgacggca aaccattagg cacttccaat agttcaagta tctttatcta tggcaatgat 1080
tttattgccc aaaaatttaac catccagaac gactctggtc aaggaaacagg tcaagcagtt 1140
gccgcttatg taaggggccga tcgtctctac tttgaaaacg tgcagttttt aggataccag 1200
gatacattat atgcccatac ggggaagacag tattataaaa actgctacgt agaaggggat 1260
gtggatttca tttttggcgg agccacagcc ttgtttgata cctgtcacct tcatagcaag 1320
cgtaacggca gtaaggttaac ccagctagat acogataag tcacaccgta tgggtatggt 1380
tttttagatt caaaaatocac ctccagtgaa ggcgtgacca atgtgcactc cggcgacact 1440
tggcgtcctt attcagctgt caccctatc aacaccgaaa tggatgcac gattgttct 1500
gacggatggg ataactgggg gaaagtgtga aacgagaaaa cagccagata ttccagaatc 1560
aataacatgg ggccaggggc agacccgaaa aagcgggatc catggaccac acaattaacg 1620
ccggagggaag cgaatcaata cactgtgcaa aatgtgtaga aaggatctga cggctgggat 1680
cctgagagaa tcgggattat cccattatca ccactgtcag caccgattat ttcacttgat 1740
caacgagatt ccattgtcaa tacaccaagc tttacaatta caggtcaagt ggaataagaa 1800
gcagccgttt ctgtcaatgg gaaggaaatt tccctacaaa aggatggcag cttcagcacg 1860
acggtgtgtc tgaatgacgg tttgaacact attacagtgg gagctgtaga tgcagcaggg 1920
aatcaggcta ttccctgagt gttaaaaatt gtttatgac atgagaaacc tgcgttatcc 1980
atcgatgatc ttaaaggaga aaaaaacggg aatcactaca atgtaatcta caatccgctg 2040
ccgattacag ggaagctgaa caaagcagga acagttatgg tgaatgggtga gaaagtaaat 2100
gtatccgaaa agttgacggt tagtacaaaa gtcattttaa agccgggggtt aaataacatt 2160
acgattaccg ctgttgatca ggcagggaat gaagccgaat ctatcactat caatgtggtt 2220
ccaaaaggga atgtgttcc agacggtccc gtcaagatta tcaaaagtga aacaacaaat 2280
gcaaataccg tttaggtttac ttttaatagc aagctagaaa aatttgattc tagtgatatt 2340
gcattgcaaa cggctacgaa cgtttgggca gctctcaatc ctggtttgaa acaattgatg 2400
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acgaagggaag cctttcaaga atagtggaacc attacgctcc caaaagttga agatccgttt 2520
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atgtataaag aaacaggtga tgctogttat aaggattccg ttttaaaagg aattgatttt 2820
ttactagaga tgcaggttga ttccggcggc tggccgagc tctatcctgc aagaagcgg 2880
tactcagatt atgtgacctt taatgataat gcgatgggtt cgtttatgag tgtattaacg 2940
atggttaaa gaaaagaagta tccgtttaat tccaaactag gtgacgagca actttctgag 3000
cagattgatg atgcattggg ccgtgggctg gattatatgt taaaatcgca aattaaggta 3060
gacggtgaag taaccgcatg gtgtgctcag catgacctg tgacgtatga accgaaagg 3120
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tgggttgaag aggcataaatt ggccgggaacg aagtatgtat caggcagatcc aaatgggcaa 3300
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aacacaactg gctactacga aaacagagta tacgtagaag tctgtgggga tcagtcataa 3540
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```

<210> 84

<211> 1205

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (268)...(556)

<223> Pectin methyl esterase domain

<221> DOMAIN

<222> (782)...(1164)

<223> Catalytic domain

<400> 84

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Met Thr Ala Leu Ser Arg Asn Ser Gln Val Asp Val Ser Trp Glu Pro
 1      5      10      15
Ser Ser Ala Gln Gln Val Thr Tyr Asn Leu Lys Arg Ser Thr Thr Lys
 20      25      30
Glu Gly Pro Tyr Gln Thr Ile Ala Glu Lys Met Ala Glu Thr Asp Phe
 35      40      45
Arg Asp Thr Gly Leu Glu Asn Gly Gln Lys Tyr Tyr Val Val Ser
 50      55      60
Ala Glu Thr Ser Ser Gly Glu Ser Ala Asp Ser Gln Ala Ile Thr Ala
 65      70      75      80
Val Pro Val Ala Pro Leu Gln Ala Pro Thr Gly Leu Ser Ala Ser His
 85      90      95
Gly Asn Gly Gly Val Thr Ile His Trp Glu Ser Val Asn Gly Ala Glu
100      105      110
Ser Tyr Gln Val Leu Arg Ser Lys Gln Lys Gly Ile Gly Tyr Glu Val
115      120      125
Ile Lys Asn Gly Val Thr Glu Thr Ser Tyr Thr Asp Thr Gly Ile Pro
130      135      140
Asp Gly Glu Lys Tyr Tyr Val Val Ser Ala Lys Asn Asp Thr Ala
145      150      155      160
Glu Ser Ala Asn Ser Gln Pro Ile Asn Gly Ala Ala Val Ser Thr Ser
165      170      175
Gly Val Pro Ala Ile Pro Asn Gly Met Asn Ala Thr Ala Gly Asp Gly
180      185      190
Arg Ala Ala Leu Thr Trp Ser Ala Val Ser Gly Ala Asp Ser Tyr Ser
195      200      205
Ile Lys Arg Gly Glu Phe Asn Ser Gly Gln Tyr Glu Val Ile Ala Lys
210      215      220
Asn Ile His Ser Thr Gly Tyr Gln Asp Ile Gly Leu Thr Asn Gly Asp
225      230      235      240
Thr Tyr Asp Tyr Val Ile Ser Ala Val Asn Glu Gln Gly Glu Ser Leu
245      250      255
Gly Ser Glu Pro Ile Ala Val Thr Pro Ala Lys Val Thr Val Val Ala
260      265      270
Lys Glu Gly Gly Asp Phe Lys Thr Ile Gln Glu Ala Ile Asp Ala Ala
275      280      285
Pro Asp Asn Ser Thr Lys Arg His Val Ile Phe Ile Lys Asn Gly Gln
290      295      300
Tyr Arg Glu Lys Leu Thr Ile Pro Lys Ser Lys Thr Asn Leu Ser Phe
305      310      315      320
Val Gly Glu Ser Lys Glu Gly Thr Val Leu Val Phe Asn Asp Asn Ala
325      330      335
Asn Thr Pro Gly Pro Asp Gly Lys Pro Leu Gly Thr Ser Asn Ser Ser
340      345      350
Ser Ile Phe Ile Tyr Ala Asn Asp Phe Ile Ala Gln Asn Leu Thr Ile
355      360      365
Gln Asn Asp Ser Gly Gln Gly Thr Gly Gln Ala Val Ala Ala Tyr Val
370      375      380
Arg Ala Asp Arg Leu Tyr Phe Glu Asn Val Gln Phe Leu Gly Tyr Gln
385      390      395      400

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```

Asp Thr Leu Tyr Ala His Thr Gly Arg Gln Tyr Tyr Lys Asn Cys Tyr
405 410 415
Val Glu Gly Asp Val Asp Phe Ile Phe Gly Gly Ala Thr Ala Leu Phe
420 425 430
Asp Thr Cys His Leu His Ser Lys Arg Thr Gly Ser Lys Leu Thr Ala
435 440 445
Ala Ser Thr Asp Gln Val Thr Pro Tyr Gly Tyr Val Phe Leu Asp Ser
450 455 460
Lys Ile Thr Ser Asp Glu Gly Val Thr Asn Val His Leu Gly Arg Pro
465 470 475 480
Trp Arg Pro Tyr Ser Ala Val Thr Tyr Ile Asn Thr Glu Met Asp Ala
485 490 495
Ser Ile Val Pro Asp Gly Trp Asp Asn Trp Gly Lys Val Glu Asn Glu
500 505 510
Lys Thr Ala Arg Tyr Ser Glu Tyr Asn Asn Met Gly Pro Gly Ala Asp
515 520 525
Pro Lys Lys Arg Asp Pro Trp Thr Thr Gln Leu Thr Pro Glu Glu Ala
530 535 540
Asn Gln Tyr Thr Val Gln Asn Val Met Lys Gly Ser Asp Gly Trp Asp
545 550 555 560
Pro Glu Arg Ile Gly Ile Ile Pro Leu Ser Pro Leu Ser Ala Pro Ile
565 570 575
Ile Ser Leu Asp Gln Arg Asp Ser Ile Val Asn Thr Pro Ser Phe Thr
580 585 590
Ile Thr Gly Gln Val Asp Lys Glu Ala Ala Val Ser Val Asn Gly Lys
595 600 605
Glu Ile Ser Leu Gln Lys Asp Gly Ser Phe Ser Thr Thr Val Val Leu
610 615 620
Asn Asp Gly Leu Asn Thr Ile Thr Val Gly Ala Val Asp Ala Ala Gly
625 630 635 640
Asn Gln Ala Ile Pro Ala Val Leu Lys Ile Val Tyr Asp His Glu Lys
645 650 655
Pro Val Val Ser Ile Asp Asp Leu Lys Gly Glu Lys Asn Gly Asn His
660 665 670
Tyr Asn Val Ile Tyr Asn Pro Leu Pro Ile Thr Gly Lys Leu Asn Glu
675 680 685
Ala Gly Thr Val Met Val Asn Gly Glu Lys Val Asn Val Ser Glu Lys
690 695 700
Leu Thr Phe Ser Thr Lys Val Ile Leu Lys Pro Gly Leu Asn Asn Ile
705 710 715 720
Thr Ile Thr Ala Val Asp Gln Ala Gly Asn Glu Ala Glu Ser Ile Thr
725 730 735
Ile Asn Val Val Pro Lys Gly Asn Ala Val Pro Asp Gly Pro Val Lys
740 745 750
Ile Ile Lys Ser Glu Thr Thr Asn Ala Asn Thr Val Glu Val Thr Phe
755 760 765
Asn Ser Lys Leu Glu Lys Phe Asp Ser Ser Asp Ile Ala Leu Gln Thr
770 775 780
Ala Thr Asn Val Trp Ala Ala Leu Asn Pro Gly Leu Lys Gln Leu Met
785 790 795 800
Thr Val Glu Ser Ile Thr Thr Lys Val Asn Lys Asp Asn Gln Thr Val
805 810 815
Ala Val Ile Lys Thr Lys Glu Ala Phe Gln Glu Asp Gly Thr Ile Thr
820 825 830
Leu Pro Lys Val Glu Asp Pro Phe His Ile Gln Tyr Leu Asn Ala Asp
835 840 845
Tyr Tyr Thr Gly Asp Arg Thr Gln Asp Ile Lys His Ala Asp Ala Leu
850 855 860
Leu Thr Trp Gln Met Asp His Gly Gly Trp Phe Lys Asn Trp Val Glu

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865      870      875      880
Lys Tyr Lys Arg Pro Trp Asp Gly Lys Glu Pro Lys Ser Glu Trp Tyr
885      890      895
Ser Thr Asn His Gly Glu Leu Gly Thr Ile Asp Asn Asp Ala Thr Thr
900      905      910
Asn Glu Ile Leu Phe Leu Ala Leu Met Tyr Lys Glu Thr Gly Asp Ala
915      920      925
Arg Tyr Lys Asp Ser Val Leu Lys Gly Ile Asp Phe Leu Leu Glu Met
930      935      940
Gln Val Asp Ser Gly Glu Trp Pro Gln Val Tyr Pro Ala Arg Ser Gly
945      950      955      960
Tyr Ser Asp Tyr Val Thr Phe Asn Asp Asn Ala Met Val Arg Val Met
965      970      975
Ser Val Leu Thr Met Val Lys Glu Lys Lys Tyr Pro Phe Asn Ser Asn
980      985      990
Leu Gly Asp Glu Gln Leu Ser Glu Gln Ile Asp Asp Ala Leu Gly Arg
995      1000      1005
Gly Leu Asp Tyr Met Leu Lys Ser Gln Ile Lys Val Asp Gly Glu Val
1010      1015      1020
Thr Ala Trp Cys Ala Gln His Asp Pro Val Thr Tyr Glu Pro Lys Gly
1025      1030      1035      1040
Ala Arg Ala Tyr Glu His Pro Ser Ile Ser Gly Ser Glu Ser Val Gly
1045      1050      1055
Ile Val Gln Tyr Leu Met Ser Leu Pro Asn Pro Ser Thr Glu Val Gln
1060      1065      1070
Ala Ala Ile His Gly Ala Leu Asn Trp Phe Glu Glu Ala Lys Leu Ala
1075      1080      1085
Gly Thr Lys Tyr Val Ser Gly Asp Pro Asn Gly Gln Tyr Phe Tyr Pro
1090      1095      1100
Asp Ala Asn Ser Asn Thr Trp Tyr Arg Phe Tyr Glu Ile Gly Thr Asn
1105      1110      1115      1120
Arg Pro Ile Phe Ser Gly Arg Asp Gly Val Ile Lys His Asn Ile Leu
1125      1130      1135
Glu Ile Glu Lys Glu Arg Arg Asp Gly Tyr Arg Trp Ala Gly Glu Trp
1140      1145      1150
Pro Gln Lys Leu Leu Asn Ile Ala Asn Thr Thr Gly Tyr Tyr Glu Asn
1155      1160      1165
Arg Val Tyr Val Glu Val Val Gly Asp Gln Ser Lys Asn Ala Ala Gly
1170      1175      1180
Glu Ser Leu Glu Ile Gly Asn Leu Tyr Arg Ile Glu Ala Ser Ala Ser
1185      1190      1195      1200
Gly Ser Thr Ser Lys
1205

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<210> 85

<211> 1152

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 85

```

atgtcgggttg gaccaggtgc taatccgaaa gctcgtgttc catggtccaa acagttatcg      60
ggtgttgagg caaagctgtt tcagcgcgag cggttcttca gcctcgtgc ggagcacact      120
tctaagaaaa atgatacagga agtcggcgcg atcgcgtgga aagatgcaca tggaaagccg      180
gatgagtggt atgcgagtggt tgaggcactg cggatggccg ataacgtcgt tctctatcaa      240
cgcgactcag gtggttgccc caagaacatc gacatggcga aggcactcaa cgcctgtgag      300
caggctgcga tcctccgcca gaagaaaaag aacgactcca cgcgcgcaa tggtcgcgact      360

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```

cacacacagt taccctttct ggcgcgcgtc tatacagcac agcgtcagga ggcacatcgc      420
gagtcgtttt tcaaaggatt ggattactta ctgaatgcgc agtatccaaa tggaggctgg      480
ccgcagtttt atccgaaccc gacgggctat cacaagcaca ttacttacaa cgacggtgcg      540
atgattggtg tgatgaaggt gctgcgcgat atcgcctcgg cgaagccttt gtacgccttt      600
gtcgcgaaag ctgcgcgcgc gaaggcgacg agtcagttg aaaaagggat cgcgtgcatt      660
ttgaaaacgc aggtggtggt agatggcggt cgcactgtgt ggagtgcgca acatgatgaa      720
gtaacgttag gcgcagctcc tgcgcgaacc ttcgagttaa ctctgttgag cggcggtgag      780
agcgtagata tcgttcgatt tttaatgtcg atcaaggatc cgtcgcttaa agtagttgat      840
gcggttgaat cgcggtttaa gtggtttgag caatcggagt taaaaggcgt gaagtggggt      900
aagaaggcgg acgcttctaa acctggcggg ttgtattgag tcgtagttaa ggaatccggag      960
agctcgtttt ggcgcgcctt ttatgagatt ggcacgaacc ggcgatcttt ttccggcgcg      1020
gatggagtgg tcaaatacga cgtggcgag atcgaacacg agcggcggac gaattatgag      1080
tggtacgttg atgaagcagc caagctgtcg aagaaagagt atccggcctg gcggaaaaga      1140
acatctctgt ga
1152

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<210> 86

<211> 383

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(383)

<223> Catalytic domain

<400> 86

```

Met Ser Val Gly Pro Gly Ala Asn Pro Lys Ala Arg Val Pro Trp Ser
 1      5      10      15
Lys Gln Leu Ser Gly Val Glu Ala Lys Leu Phe Gln Arg Glu Arg Phe
 20      25      30
Phe Ser Leu Ala Ala Glu His Thr Ser Lys Lys Asn Asp Gln Glu Val
 35      40      45
Gly Ala Ile Ala Trp Lys Asp Ala His Gly Lys Pro Asp Glu Trp Tyr
 50      55      60
Ala Ser Val Glu Ala Leu Arg Met Ala Asp Asn Val Val Leu Tyr Gln
 65      70      75      80
Arg Asp Ser Gly Gly Trp Pro Lys Asn Ile Asp Met Ala Lys Ala Leu
 85      90      95
Asn Asp Arg Glu Gln Ala Ala Ile Leu Arg Gln Lys Lys Lys Asn Asp
100      105      110
Ser Thr Ile Asp Asn Gly Ala Thr His Thr Gln Leu Ser Phe Leu Ala
115      120      125
Arg Val Tyr Thr Ala Gln Arg Gln Glu Arg His Arg Glu Ser Phe Phe
130      135      140
Lys Gly Leu Asp Tyr Leu Leu Asn Ala Gln Tyr Pro Asn Gly Gly Trp
145      150      155      160
Pro Gln Phe Tyr Pro Asn Pro Thr Gly Tyr His Lys His Ile Thr Tyr
165      170      175
Asn Asp Gly Ala Met Ile Gly Val Met Lys Val Leu Arg Asp Ile Ala
180      185      190
Ala Ala Lys Pro Leu Tyr Ala Phe Val Asp Glu Ala Arg Arg Ala Lys
195      200      205
Ala Thr Ser Ala Val Glu Lys Gly Ile Glu Cys Ile Leu Lys Thr Gln
210      215      220
Val Val Val Asp Gly Arg Arg Thr Val Trp Ser Ala Gln His Asp Glu
225      230      235      240
Val Thr Leu Ala Pro Ala Pro Ala Arg Thr Phe Glu Leu Thr Ser Leu

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245      250      255
Ser Gly Gly Glu Ser Val Asp Ile Val Arg Phe Leu Met Ser Ile Lys
260      265      270
Asp Pro Ser Pro Lys Val Val Asp Ala Val Glu Ser Ala Val Lys Trp
275      280      285
Phe Glu Gln Ser Glu Leu Lys Gly Val Lys Trp Val Lys Lys Ala Asp
290      295      300
Ala Ser Lys Pro Gly Gly Phe Asp Cys Val Val Val Lys Asp Pro Glu
305      310      315      320
Ser Ser Val Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg Pro Ile
325      330      335
Phe Ser Gly Arg Asp Gly Val Val Lys Tyr Asp Val Ala Gln Ile Glu
340      345      350
His Glu Arg Arg Thr Asn Tyr Glu Trp Tyr Val Asp Glu Ala Ala Lys
355      360      365
Leu Leu Lys Lys Glu Tyr Pro Ala Trp Arg Lys Arg Thr Ser Leu
370      375      380

```

<210> 87

<211> 1698

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 87

```

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gcagtggcga tgtttgcagc ttcttcagta ttccgcagcg ctacagggtg tttttccacg 120
actgatggcg gtgcgggcaag cggctcgcaa tccttcacgg cggccaaact tgaccagctc 180
aacaccattg ttgccaatgc gaagagtggc ggttaccggg ttgtgattac ctataccggg 240
aatgaagaca ccttgattaa ccagatgatc aaagaccaca ccgtggatto ttcaggcaac 300
tgcccgaaac cagcgtggag tgaacacctac cgcaaggtag aaattaagga gatgacaaa 360
gggtgcacca tcactgggac caatgggtct tcggcaaaact tcgggtattgt ggtgaacaag 420
tccagcaatg tgattatccg caacatgaaa atcgggtcgc tggccgggtg cagcaacgac 480
gcggatatga ttcgatatga tagcggcact aacgtatggg ttgaccacaa cgaattgttc 540
gcgggtgaaca acgaatgtaa aggttcacgg gatggcgatt tgaccttoga aagcgccatc 600
gacatcaaga aagattcaca caacatcacc gtgtcttaca acctgattcg cgacagcaaa 660
aaagtgggac ttgatggttc cagcagcagc gatatcgccg gtggccgcga gatcactttc 720
caccacaaca ttacaaaaa cgtgaatgca cgcttgccgt tgcaacggcg tggctggacg 780
cacatgtata acaacctgta cgacggcatt acgggttccg gtatcaacgt acgtcaggcc 840
ggttatgcgt tgattgaaag caactggttc caaatgcgg ttaaccgggt gacttgccgt 900
tacgacagca gcaactgcgg ttctcgggat ctgcgcaata acaacgtgaa gtcgccagca 960
gatttcgcga cctataacat caccctggacc agcggcgga ctattgatgc aaccaactgg 1020
acgaccaccg ctccgttccc gatcagcatt ccttacagct actcgcgggt gtctccacag 1080
tgcgtagaag acaagttggc cagcgttgcg ggtgtgggta aaaacgggtc agttctgaac 1140
tcctcagtggt gtggtggaag cagctctggt ccatcatcaa gctcagtcgc tactacttcc 1200
aatcatcca gctcggtagc aaccagcaag tccagctccg tcgctacgac gtcacgtaag 1260
tcctccagct cggtagtgcc atcatcatca agctcaagtt cagtgggtta taacggcagc 1320
atcgcggtta ccgccactgc taccggcaat agcattgttc tgagctggtc gccgaacaac 1380
ctgacactgg gcacccagga ggtgtatcgc gataccgatt cagaccocaa tggccgtgtg 1440
cgtattgctg ccctgagttc cagcactcgc atgtacacgg atgccactgc atcgccggcg 1500
caaacgttct actactggat caaaaacacc accaacgggt taaccaccaa ttccaatgcg 1560
gcttcggcgg caattggcga tgcagctcgc gccattcgcg catgcgcagg aaaccgagga 1620
agtggcgctc gcaccagtcg cgcagtttcg actgggtcaa atcctcgtgg gcctgccggg 1680
agccatccca gagcttga

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<210> 88

<211> 565

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(32)

<221> DOMAIN

<222> (33)...(375)

<223> Catalytic domain

<400> 88

```

Met Ser Thr Thr Lys Cys Phe Asn Thr Ala Pro Gly Phe Thr Leu Lys
1      5      10      15
Ala Val Ala Ala Val Ala Met Phe Ala Gly Ser Ser Val Phe Ala
20      25      30
Ala Ala Thr Gly Gly Phe Ser Thr Thr Asp Gly Gly Ala Ala Ser Gly
35      40      45
Ser Gln Ser Phe Thr Ala Ala Asn Leu Asp Gln Leu Asn Thr Ile Val
50      55      60
Ala Asn Ala Lys Ser Gly Gly Tyr Pro Val Val Ile Thr Tyr Thr Gly
65      70      75      80
Asn Glu Asp Ser Leu Ile Asn Gln Met Ile Lys Asp His Thr Val Asp
85      90      95
Ser Ser Gly Asn Cys Pro Asn Pro Arg Trp Ser Glu Thr Tyr Arg Lys
100     105     110
Val Glu Ile Lys Glu Met Thr Lys Gly Val Thr Ile Ile Gly Ala Asn
115     120     125
Gly Ser Ser Ala Asn Phe Gly Ile Val Val Asn Lys Ser Ser Asn Val
130     135     140
Ile Ile Arg Asn Met Lys Ile Gly Ala Leu Ala Gly Ala Ser Asn Asp
145     150     155     160
Ala Asp Met Ile Arg Ile Asp Ser Gly Thr Asn Val Trp Val Asp His
165     170     175
Asn Glu Leu Phe Ala Val Asn Asn Glu Cys Lys Gly Ser Pro Asp Gly
180     185     190
Asp Leu Thr Phe Glu Ser Ala Ile Asp Ile Lys Lys Asp Ser His Asn
195     200     205
Ile Thr Val Ser Tyr Asn Leu Ile Arg Asp Ser Lys Lys Val Gly Leu
210     215     220
Asp Gly Ser Ser Ser Ser Asp Ile Ala Gly Gly Arg Glu Ile Thr Phe
225     230     235     240
His His Asn Ile Tyr Lys Asn Val Asn Ala Arg Leu Pro Leu Gln Arg
245     250     255
Gly Gly Trp Thr His Met Tyr Asn Asn Leu Tyr Asp Gly Ile Thr Gly
260     265     270
Ser Gly Ile Asn Val Arg Gln Ala Gly Tyr Ala Leu Ile Glu Ser Asn
275     280     285
Trp Phe Gln Asn Ala Val Asn Pro Val Thr Cys Arg Tyr Asp Ser Ser
290     295     300
Asn Cys Gly Phe Trp Asp Leu Arg Asn Asn Asn Val Lys Ser Pro Ala
305     310     315     320
Asp Phe Ala Thr Tyr Asn Ile Thr Trp Thr Ser Gly Gly Thr Ile Asp
325     330     335
Ala Thr Asn Trp Thr Thr Thr Ala Pro Phe Pro Ile Ser Ile Pro Tyr
340     345     350
Ser Tyr Ser Pro Val Ser Pro Gln Cys Val Lys Asp Lys Leu Ala Ser
355     360     365
Val Ala Gly Val Gly Lys Asn Gly Ala Val Leu Asn Ser Ser Val Cys
370     375     380
Gly Gly Ser Ser Ser Val Pro Ser Ser Ser Ser Val Ala Thr Thr Ser
385     390     395     400
Lys Ser Ser Ser Ser Val Ala Thr Ser Lys Ser Ser Ser Val Ala Thr
405     410     415
Thr Ser Ser Lys Ser Ser Ser Ser Val Val Pro Ser Ser Ser Ser
420     425     430
Ser Ser Val Val Asn Asn Gly Ser Ile Ala Leu Thr Ala Thr Thr
435     440     445
Gly Asn Ser Ile Val Leu Ser Trp Ser Pro Asn Asn Leu Thr Leu Gly
450     455     460
Thr Gln Glu Val Tyr Arg Asp Thr Asp Ser Asp Pro Ser Gly Arg Val
465     470     475     480
Arg Ile Ala Ala Leu Ser Ser Thr Arg Met Tyr Thr Asp Ala Thr
485     490     495
Ala Ser Ala Gly Gln Thr Phe Tyr Tyr Trp Ile Lys Asn Thr Thr Asn
500     505     510
Gly Val Thr Thr Asn Ser Asn Ala Ala Ser Ala Ala Ile Gly Asp Ala
515     520     525
Ala Arg Ala Ile Arg Ala Cys Ala Gly Asn Arg Gly Ser Gly Ala Arg
530     535     540
Thr Ser Arg Ala Val Ser Thr Gly Ser Asn Pro Arg Gly Pro Ala Gly
545     550     555     560
Ser His Pro Arg Ala
565

```

<210> 89

<211> 1377

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 89

```

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gcaccgcggt tcttggcggt ttcttcggtg gcgagtcctg gggaacggt gatgccttcg 120
atcctcgaac gcataagcc acccggtttt ccgaacgca cgtgctatct caaccggttt 180
ggagcaaaag cgcgcgggca aactgattgc acttcagctt ttgcagcgcg aatcgatcag 240
tggtcgaaag cggcggtgg caaagtgtgc gttccgcagg gaatgtatct caccggcgca 300
attcacttga agagcaactt caatctcgag atctccgaag gcgcgacgat caagttcagt 360
caaaaaccga aagactatct cccggtggtt ttttcgcgtt gggaaggcgt cgaagtattc 420
aactactcac ctttcatcta cgcatttgaa cagcagaaca tcgcgatcac gggcaagggc 480
acgctcgatg ggcagagtga taacgaacac tggtyggcat ggaacggacg cgcacgggtac 540
ggttggaag aagggatgag ccaccagcgt ccgcatcgaa acgcgctctt tgcgatggcg 600
gaaaaagggt tttcggttcg cgaacgtggt ttccggcagg gtcattactt aaggccgcag 660
ttcattcagc cgtatcgctg ccagaacgta ttgatcgacg gagttacgat acgaaactcg 720
ccgatgtggg aaattcatcc ggtgctgtgc cggaaatgta tcgtgcaaaa cgtgcacatt 780
aacagtcatg gaccaaaca cgtggctgac aatcccgagt cgtgcactga tgtgctgatt 840
aagaactggt acttcgacac tggcgacgac tgtatcgagg tcaaatcagg acgcaacgcg 900
gacggccgcg ggcttaaacg gccgacagag aacgtgatcg tgcaagactg tcaaatgaaa 960
gatggacacg cggggatcac tgtcggcagt gagatctcag gcggtgtgag aaatctgttt 1020
gcggagaact gccggcttga tagtccaaac ctggaccatg ctttgcgggt taagaacaac 1080
gcgatgcgtg gagggtgctc cgagaatttg cacttccgaa acatcgaaat tggtcagggtg 1140
gcgatgcag tgatcacgat cgattttaat tacgaggaag gcgcgaaaag atcgttcacg 1200
ccggtggttc gtgactcac tgcgatggtt ttgcgcagca cgcgaagcaa atacgcgctc 1260
gacgttcaag gtctgtcggg cgcgccgacg gtaaatctgc gtctgacgaa ttgcacgttc 1320

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gacaatgttg ccgaaggga cgtcgtgaag aatgttaagg acgcgacaat tcaaaaa 1377

<210> 90

<211> 459

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(31)

<221> DOMAIN

<222> (32)...(459)

<223> Catalytic domain

<400> 90

```

Met Thr Thr Arg Glu Phe Ile Arg Asp Leu Leu Val Gly Gly Val
1      5      10      15
Val Val Ala Val Ala Pro Arg Phe Leu Ala Phe Ser Ser Val Ala Ser
20     25     30
Pro Trp Glu Thr Val Met Pro Ser Ile Leu Glu Arg Ile Lys Pro Pro
35     40     45
Arg Phe Pro Lys Arg Thr Cys Tyr Leu Asn Arg Phe Gly Ala Lys Gly
50     55     60
Asp Gly Gln Thr Asp Cys Thr Ser Ala Phe Arg Arg Ala Ile Asp Gln
65     70     75     80
Cys Ser Lys Ala Gly Gly Gly Lys Val Ile Val Pro Gln Gly Met Tyr
85     90     95
Leu Thr Gly Ala Ile His Leu Lys Ser Asn Val Asn Leu Glu Ile Ser
100    105    110
Glu Gly Ala Thr Ile Lys Phe Ser Gln Asn Pro Lys Asp Tyr Leu Pro
115    120    125
Val Val Phe Ser Arg Trp Glu Gly Val Glu Val Phe Asn Tyr Ser Pro
130    135    140
Phe Ile Tyr Ala Phe Glu Gln Gln Asn Ile Ala Ile Thr Gly Lys Gly
145    150    155    160
Thr Leu Asp Gly Gln Ser Asp Asn Glu His Trp Trp Pro Trp Asn Gly
165    170    175
Arg Ala Arg Tyr Gly Trp Lys Glu Gly Met Ser His Gln Arg Pro Asp
180    185    190
Arg Asn Ala Leu Phe Ala Met Ala Glu Lys Gly Val Ser Val Arg Glu
195    200    205
Arg Val Phe Gly Glu Gly His Tyr Leu Arg Pro Gln Phe Ile Gln Pro
210    215    220
Tyr Arg Cys Gln Asn Val Leu Ile Asp Gly Val Thr Ile Arg Asn Ser
225    230    235    240
Pro Met Trp Glu Ile His Pro Val Leu Cys Arg Asn Val Ile Val Gln
245    250    255
Asn Val His Ile Asn Ser His Gly Pro Asn Asn Asp Gly Cys Asn Pro
260    265    270
Glu Ser Cys Thr Asp Val Leu Ile Lys Asn Cys Tyr Phe Asp Thr Gly
275    280    285
Asp Asp Cys Ile Ala Val Lys Ser Gly Arg Asn Ala Asp Gly Arg Arg
290    295    300
Leu Lys Ala Pro Thr Glu Asn Val Ile Val Gln Asp Cys Gln Met Lys
305    310    315    320
Asp Gly His Gly Gly Ile Thr Val Gly Ser Glu Ile Ser Gly Gly Val

```

```

          325          330          335
Arg Asn Leu Phe Ala Glu Asn Cys Arg Leu Asp Ser Pro Asn Leu Asp
          340          345          350
His Ala Leu Arg Val Lys Asn Asn Ala Met Arg Gly Gly Leu Leu Glu
          355          360          365
Asn Leu His Phe Arg Asn Ile Glu Val Gly Gln Val Ala His Ala Val
          370          375          380
Ile Thr Ile Asp Phe Asn Tyr Glu Glu Gly Ala Lys Gly Ser Phe Thr
          385          390          395          400
Pro Val Val Arg Asp Tyr Thr Val Asp Gly Leu Arg Ser Thr Arg Ser
          405          410          415
Lys Tyr Ala Leu Asp Val Gln Gly Leu Ser Gly Ala Pro Ile Val Asn
          420          425          430
Leu Arg Leu Thr Asn Cys Thr Phe Asp Asn Val Ala Glu Gly Asn Val
          435          440          445
Val Lys Asn Val Lys Asp Ala Thr Ile Gln Lys
          450          455

```

<210> 91

<211> 1125

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 91

```

gtggtcctag gtaataacgg cggcagcttg agttgcgtcc aatatattgt gattgtgaaa 60
ggaccgggtg gacctcgacc gccggtgaaa ccggccgtcc aggcgccgt tagggttacc 120
tggagcgcat ccctagtcca gcggccgcaa tggtagggga gtgacgaagc gatccgcctc 180
gcggacaacg tcctcctcta ccagcgcaac accggcgggt ggccgaagga catagatatg 240
gccgagccca tcccggaaca caggaagtcc ttttcctca ccgagaagga gccgaccgat 300
gactcgacca tcgacaacgg tgccaccgtg acccagctca agtatctcgc ccgcgtctac 360
aaggcgacca ggctggaacg gtccaaggag ggcttcctca aaggtctcga ctacctcttg 420
gcccccagct acccgaacgg cggctggccc cagtattatc ctaacttgag gggctactac 480
gccaaacatca cttataacga caatgccatg gtgaacgtgc tcaccctcct ccagagcatc 540
gccaaaaagg ccccgagta cgacttcgtc gaccggcgcc gccgggagaa ggccggccgg 600
gccgtggcga aagggatcga ctgcatcctc aagaccaga tccgtgtcaa tggaaaactt 660
accgcctggt gcgcccagca tgacgccaag acgctggcgc ccgcgccggc ccgttcgtat 720
gagcttgagt ccatcagcgg tttcgagagc gtccggatcg tccggttctt aatgagcctc 780
gagaatccga gcccgaaagg catcgaggcg gttagggccg ccgtgaaatg gttcgaggag 840
gtcaagctta ccgggatcaa ggtggtcgag aaacccgacc cgtcccttcc gggcggttac 900
gaccgcgtgg tggtcgaaga ccccaacgcg ccgcccctct gggcccggtt ctacagatc 960
ggcaccaccc gtcccttctt ctgcggccgc gatggtatca aaaaatacag cctggcggag 1020
atcgaacacg aacgccgggt cggttactcc tggtagacca atgccccggc ctacctcatc 1080
gagaaggagt atccgctctg gccgggcaaa caccctacca agtaa 1125

```

<210> 92

<211> 374

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(374)

<400> 92

```

Met Val Leu Gly Asn Asn Gly Gly Ser Leu Ser Cys Val Gln Tyr Ile
 1          5          10          15
Val Ile Val Lys Gly Pro Gly Gly Pro Arg Pro Pro Val Lys Pro Ala
 20          25          30
Val Gln Ala Pro Val Arg Val Thr Trp Ser Ala Ser Leu Val Gln Arg
 35          40          45
Pro Glu Trp Tyr Gly Ser Asp Glu Ala Ile Arg Ile Ala Asp Asn Val
 50          55          60
Leu Leu Tyr Gln Arg Asn Thr Gly Gly Trp Pro Lys Asp Ile Asp Met
 65          70          75          80
Ala Glu Pro Ile Pro Glu His Arg Lys Ser Phe Phe Leu Thr Glu Lys
 85          90          95
Glu Arg Thr Asp Asp Ser Thr Ile Asp Asn Gly Ala Thr Val Thr Gln
100          105          110
Leu Lys Tyr Leu Ala Arg Val Tyr Lys Ala Thr Arg Leu Glu Arg Phe
115          120          125
Lys Glu Gly Phe Leu Lys Gly Leu Asp Tyr Leu Leu Ala Ala Gln Tyr
130          135          140
Pro Asn Gly Gly Trp Pro Gln Tyr Tyr Pro Asn Leu Arg Gly Tyr Tyr
145          150          155          160
Ala Asn Ile Thr Tyr Asn Asp Asn Ala Met Val Asn Val Leu Thr Leu
165          170          175
Leu Gln Ser Ile Ala Lys Lys Ala Pro Glu Tyr Asp Phe Val Asp Pro
180          185          190
Ala Arg Arg Glu Lys Ala Ala Arg Ala Val Ala Lys Gly Ile Asp Cys
195          200          205
Ile Leu Lys Thr Gln Ile Arg Val Asn Gly Lys Leu Thr Ala Trp Cys
210          215          220
Ala Gln His Asp Ala Lys Thr Leu Ala Pro Ala Pro Ala Arg Ser Tyr
225          230          235          240
Glu Leu Glu Ser Ile Ser Gly Phe Glu Ser Val Gly Ile Val Arg Phe
245          250          255
Leu Met Ser Leu Glu Asn Pro Ser Pro Lys Val Ile Glu Ala Val Glu
260          265          270
Ala Ala Val Lys Trp Phe Glu Glu Val Lys Leu Thr Gly Ile Lys Val
275          280          285
Val Glu Lys Pro Asp Pro Ser Leu Pro Gly Gly Tyr Asp Arg Val Val
290          295          300
Val Glu Asp Pro Asn Ala Pro Pro Ile Trp Ala Arg Phe Tyr Glu Ile
305          310          315          320
Gly Thr Asn Arg Pro Phe Phe Cys Gly Arg Asp Gly Ile Lys Lys Tyr
325          330          335
Ser Leu Ala Glu Ile Glu His Glu Arg Arg Val Gly Tyr Ser Trp Tyr
340          345          350
Thr Asn Ala Pro Ala Tyr Leu Ile Glu Lys Glu Tyr Pro Leu Trp Arg
355          360          365
Ala Lys His Pro Thr Lys
370

```

<210> 93

<211> 1062

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 93

gtggatccaa agaattggaa cccgaaaaaa gccgacgatt catggctcga aaagacgaaa 60

```

cccgattacc ggctgggtctc ctggcgcgac gttttagatc aaactcagct ctgggtacgcg 120
gtcgacgaag cgacgcgcac cgccaaccag gttttgctct ttcagcgcgga taacggcggcg 180
tgggaaaaaaa acgtcgacat ggccggcgatg ctcaactcaag ccgaacgaga aaaactcgtc 240
aaagaaaaaat ctcacaccga tacgaccatc gacaacggcg cgacgaccac gcagctgcgt 300
tatctggcaa aagtcacac ggcgaaaaac atcgaagctc ataaacagtc gtttctcaag 360
ggattggatt tctgctcgc gatgcagtat gaaaacggag gatttcgcga atattatcct 420
ttgaaaaaacg attattcgcg cgagattact ttcaacgacg acgcatgat caatgttctt 480
aaattgctgc gcgacgtggc aaaaaagaag gaagattatt tattcgtcga cgaagaccgc 540
cgcgccagag cggaaggcgc ggtcgaaaaa ggctgccgct tgatcttgaa aacacaggtc 600
gccatcgacg gcaaaaaaac gatctgggcy gcgcagtacg acgaaaaacac ttgaaacgcg 660
gcaaatcgca gaaagtgtga gcccgctcgc ctgccttcgc gcgaatcggt cagcgtggtc 720
agatttttga tgctcgacgc caaaccgcac gaggaaaaaa tcggagcgat cgaatcggcg 780
atcgaatggt ttcaaaaaaa caaactgagc ggcatctcgt gggaatcgaa aagcggagaa 840
aacctggtcg tcaagacaa agcggcgccg ccgatctggg gaaggtttta tcaattcgaa 900
accatgcgcc ccatttttat cgggcgcgac gcggtgattc gctacgatgt catgcaaatc 960
gaagccgaac gccgcaacgg ctacggctgg tacacgaacg agccgaacga gcttttgac 1020
aaagattatc cgaaatggaa agagaaaatt aagaaaaatt ag 1062

```

<210> 94

<211> 353

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(353)

<400> 94

```

Met Asp Pro Lys Asn Trp Asn Pro Lys Lys Ala Asp Asp Ser Trp Leu
 1          5          10          15
Glu Lys Thr Lys Pro Asp Tyr Arg Leu Val Ser Trp Arg Asp Val Leu
 20          25          30
Asp Gln Thr Gln Leu Trp Tyr Ala Val Asp Glu Ala Thr Arg Ile Ala
 35          40          45
Asn Gln Val Leu Leu Phe Gln Arg Asp Asn Gly Gly Trp Glu Lys Asn
 50          55          60
Val Asp Met Ala Ala Met Leu Thr Gln Ala Glu Arg Glu Lys Leu Val
 65          70          75          80
Lys Glu Lys Ser His Thr Asp Thr Thr Ile Asp Asn Gly Ala Thr Thr
 85          90          95
Thr Gln Leu Arg Tyr Leu Ala Lys Val Ile Thr Ala Lys Asn Ile Glu
100          105          110
Ala His Lys Gln Ser Phe Leu Lys Gly Leu Asp Phe Leu Leu Ala Met
115          120          125
Gln Tyr Glu Asn Gly Gly Phe Pro Gln Tyr Tyr Pro Leu Lys Asn Asp
130          135          140
Tyr Ser Arg Glu Ile Thr Phe Asn Asp Asp Ala Met Ile Asn Val Leu
145          150          155          160
Lys Leu Leu Arg Asp Val Ala Lys Lys Lys Glu Asp Tyr Leu Phe Val
165          170          175
Asp Glu Asp Arg Arg Ala Arg Ala Glu Gly Ala Val Glu Lys Gly Val
180          185          190
Arg Leu Ile Leu Lys Thr Gln Val Ala Ile Asp Gly Lys Lys Thr Ile
195          200          205
Trp Ala Ala Gln Tyr Asp Glu Asn Thr Leu Lys Pro Ala Asn Ala Arg
210          215          220
Lys Phe Glu Pro Ala Ser Leu Ala Ser Arg Glu Ser Val Ser Val Val

```

```

225          230          235          240
Arg Phe Leu Met Leu Asp Ala Lys Pro Asp Glu Glu Lys Ile Gly Ala
245          250          255
Ile Glu Ser Ala Ile Glu Trp Phe Gln Lys Asn Lys Leu Ser Gly Ile
260          265          270
Arg Trp Glu Ser Lys Ser Gly Glu Asn Leu Val Val Lys Asp Lys Ala
275          280          285
Ala Pro Pro Ile Trp Gly Arg Phe Tyr Gln Phe Glu Thr Met Arg Pro
290          295          300
Ile Phe Ile Gly Arg Asp Ala Val Ile Arg Tyr Asp Val Met Gln Ile
305          310          315          320
Glu Ala Glu Arg Arg Asn Gly Tyr Gly Trp Tyr Thr Asn Glu Pro Asn
325          330          335
Glu Leu Leu Asp Lys Asp Tyr Pro Lys Trp Lys Glu Lys Ile Lys Lys
340          345          350
Asn

```

<210> 95

<211> 1074

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 95

```

atgacgctac cegttgtttc cctgcgcgta ctgctggcgc tgctggccac gtcgccggtc      60
gcctgcgcgg gcgcgcgcgc acccgcgact cgcaccgata cggtcgccga gaacatgctg      120
cttctgcaga cgcctcccg cggtgtgtcc aagcactacc gcgagaagaa ggtcgactac      180
gcgcgcgact acgacgcgcg cgagcgcgcc gcgctgcgcg cgcgcgaccg gcatgacgat      240
gcgacgatcg acaacaaggc caccaccacc gagatcgctt acctggtgca ggcacatgcc      300
aggacgggca atccggccta cctcgacggc gcgcgcgcgc gcgtcgagta cctgctgcgc      360
gcccagtaac cgaacggcgg ttgcgcgcag ttctaccccg accattcgtc ctaccggcac      420
cagatcacgc tcaacgcacga tgcgatgttg caccgatca cctgctgca ggacatcgcc      480
gcgggcccga acggcatgca ggtgctggcg ccggagttcg gcgtccgcgc cgcgcggccc      540
gcgcagcgcg gcatcgaaaa cctgctcgag ttgcaggtgc ggatcgccgg ggtgcgcagc      600
atatggggcg cgcagtacga cgagaccagc ctgcaaccgg ccaaggcccg cgcgtacgaa      660
ctgccttcgc tggcgtggcg cgaatcggtc ggctgtgtgc gcctgctgat gcccagccg      720
gcgcctgatg cgcgcacggc cgcgcgatac gaggcggcgg ccgactggct ggaggcgcac      780
cgctgcgcgg acctcgccct ggaacgcata gaagccccg ccgaggaaac cggcaaggac      840
gtccgcgtcg tggccagacc gggcgctgcg ttgtgggcgc gcttctacga cctcgagcgg      900
caggtgcgcg tgttctgcga tcgcaacagc cgtcgggtgc ccttcgccga gcttcccaac      960
gagcgtcgta ccggctatgg ctggtatggc acctggccgg aaaaagctgt ggcacaggaa     1020
ctcccgcgct ggcgcaaggt ccatgcggcc agcgcggggc ctcgggcccg ttga      1074

```

<210> 96

<211> 357

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(31)

<221> DOMAIN

<222> (32)...(357)

<223> Catalytic domain

<400> 96

```

Met Thr Leu Pro Val Val Ser Leu Arg Val Leu Leu Ala Leu Leu Ala
 1      5      10      15
Thr Ser Pro Val Ala Cys Ala Gly Ala Ala Ala Pro Ala Thr Ala Thr
 20      25      30
Asp Pro Val Ala Glu Asn Met Leu Leu Leu Gln Thr Ala Ser Gly Gly
 35      40      45
Trp Ser Lys His Tyr Arg Glu Lys Lys Val Asp Tyr Ala Arg Asp Tyr
 50      55      60
Asp Ala Ala Glu Arg Ala Ala Leu Arg Ala Pro Asp Arg His Asp Asp
 65      70      75      80
Ala Thr Ile Asp Asn Lys Ala Thr Thr Thr Glu Ile Ala Tyr Leu Val
 85      90      95
Gln Ala His Ala Arg Thr Gly Asn Pro Ala Tyr Leu Asp Gly Ala Arg
100      105      110
Arg Gly Val Glu Tyr Leu Leu Arg Ala Gln Tyr Pro Asn Gly Gly Trp
115      120      125
Pro Gln Phe Tyr Pro Asp His Ser Ser Tyr Arg His Gln Ile Thr Leu
130      135      140
Asn Asp Asp Ala Met Val His Ala Ile Thr Val Leu Gln Asp Ile Ala
145      150      155      160
Ala Gly Arg Asn Gly Met Gln Val Leu Ala Pro Glu Phe Gly Val Arg
165      170      175
Ala Ala Ala Ala Ala Gln Arg Gly Ile Gly Asn Leu Leu Glu Leu Gln
180      185      190
Val Arg Ile Ala Gly Val Pro Thr Ile Trp Ala Ala Gln Tyr Asp Glu
195      200      205
Thr Ser Leu Gln Pro Ala Lys Ala Arg Ala Tyr Glu Leu Pro Ser Leu
210      215      220
Ala Val Ala Glu Ser Val Gly Val Val Arg Leu Leu Met Arg Gln Pro
225      230      235      240
Ala Pro Asp Ala Arg Thr Val Ala Ala Ile Glu Ala Ala Ala Asp Trp
245      250      255
Leu Glu Ala His Arg Leu Pro Asp Leu Ala Leu Glu Arg Ile Glu Ala
260      265      270
Pro Ala Glu Glu Thr Gly Lys Asp Val Arg Val Val Ala Arg Pro Gly
275      280      285
Ala Ser Leu Trp Ala Arg Phe Tyr Asp Leu Glu Arg Gln Val Pro Leu
290      295      300
Phe Val Asp Arg Asn Ser Arg Pro Val Pro Phe Ala Glu Leu Pro Asn
305      310      315      320
Glu Arg Arg Thr Gly Tyr Gly Trp Tyr Gly Thr Trp Pro Glu Lys Leu
325      330      335
Leu Ala Gln Glu Leu Pro Arg Trp Arg Lys Val His Ala Ala Ser Ala
340      345      350
Gly Ala Pro Ala Arg
355

```

<210> 97

<211> 2097

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 97

```

ttgaacgccg cgggcagccg gcggttcgcg caactcgtcg tcgcgggatct gcggcggtcg 60
gtgcccgcgc tggcgccctt ctttcgcgac gagccgctgg cgggaggagt cgcgcgcgtc 120
cagcgcagcg tcgatcgcat cgtcgcgcgc gacggcaccc gacagtttgc gacggtgcag 180
gaggcgatca accgcgcgcc gcagaacacc agcacgacca gccgtggat catcctcgtc 240
aaaccagcca cgtatcgcca ggtcgtctac ttgcagcggt agaagcgctt cgtcacgctg 300
atcggcgaag acccggcacg gacgacgac acgtaccacc tcaaagcgctc tgacgtgggg 360
ctcgacggca agcccatcgg cacgtttcgc acgcccagca tgggtggtgga tgcgcagcat 420
ttcacgatcg agaacctcac catcgagaac ggggcagggc cggtcggtca agcgcgtggc 480
ttgcgagtg gacggcgatc cgtgacggtg aggaacagcc gccctcgtgg ctygcaggac 540
acgatctttc tcaaccgtgg gcgcactac ttcagggact cgttcacgcg cgggcacgctg 600
gatttcattt tcggcgcgcg gacggcggtg ttgcagcgat gccatcttcg cgcctggcgg 660
gacggctacc tcacggcgcc gtccacgccc ggggagcaac gattcggctt cgtgttcctg 720
aacagcatcg tcagtggaga agctggcgcc cgcacgtacc tcggtcgacc gtggcgggcg 780
ttcgcgacg tggccttcac caagacgacg atggcgaggg tggtcgcgcc ggtgggctgg 840
aacaactggg accggcgcca gcgtgagaag accgtgcgtt ttctcgaagc aggcaccagc 900
ggcgcgggcg gcagcgtcgc tgcgcgcgtc gccctggcgc gccctcgcac gccagccag 960
ctcgctgac tgacgaccca ggtggtgctt ggcggcaccc acggctggga cccgcgtcgc 1020
gtcgccccc acccgtcgcc cgttcgcgc aacggcgccg cgtgcgcgc gccgcggg 1080
cccgacgtcg ctggcccgca gagccccccc gccctgacgt gggaccaggt cgcgcgccag 1140
ccagcgtcgt ggtggccac acccgaagcg ctgcggattg ccgagaacgt gccctctat 1200
caacggcaca ctggcggtg gcccaaaaac ctgcacatgg cgcagccgtt gacggacgcc 1260
gatcgcgcg cgtctcaggg cgtatcgcg ctgcagact cgaccatcga caatggcgcc 1320
acgacgcggc agatcgagtt tctcgcccg atcgccgcgc ccaaccgcca cgagcgcgcg 1380
caggcgtcga tgctggctgg gatcgactac ctgctcgcg cccaglatcc aaacggcgcc 1440
tgcccgaggt atttccgct cgcgaacgac tactcgcgcc acatcacgtt caagacgac 1500
gcatgatcg cggccgcgac gatcctgcag tcggtcgcg tggcccgcc gccgttcgcc 1560
ggcgtcgacg cgaactcgcc cggcgggcg cgggagggcg tcgcgcgcg ccacgcgtg 1620
attctggcct cgcagattcg cgtcaacggc cagctcactg gctggtgcca gcagcacgat 1680
gcacgcacgc tggagccag cgcggggcg accacgagc atccatcgat cagtggccgc 1740
gaaaccgtga cgaactcga tttcctcgcg tcgatcgaac cgcgcgacgg ccagacccaa 1800
gccgccatcg atgcccgat ggagtggtc aaggccgtgc agatccgcg ctygcgcacg 1860
gagcgcgcg ccgatccctc aggaccggcg ggttacgacg tgggtgatgg ggaggacccc 1920
aacgcggcg cgccttgggc cgccttctac gagattgga ccaatcgcc gatctactcg 1980
ggcggggag cgcctcatca gtacgggctc gccagatcg aaattgaac gcggaccgcg 2040
tacagctggg tcggaccgta cgcgcaggcg ctgctcgatg aagagcgacg gaagtaa 2097

```

<210> 98

<211> 698

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (45)...(333)

<223> Pectin methyl esterase domain

<221> DOMAIN

<222> (336)...(698)

<223> Catalytic domain

<400> 98

```

Met Asn Ala Ala Gly Ser Arg Arg Phe Ala Gln Leu Val Val Ala Asp
1          5          10          15
Leu Arg Arg Leu Val Pro Ala Leu Ala Pro Phe Phe Arg Asp Glu Pro
20          25          30
Leu Ala Gly Gly Val Ala Ala Leu Gln Arg Ser Val Asp Ala Ile Val
35          40          45

```

Ala Ala Asp Gly Thr Gly Gln Phe Ala Thr Val Gln Glu Ala Ile Asn
 50 55 60
 Ala Ala Pro Gln Asn Thr Ser Thr Thr Ser Arg Trp Ile Ile Leu Val
 65 70 75 80
 Lys Pro Gly Thr Tyr Arg Glu Val Val Tyr Val Gln Arg Glu Lys Arg
 85 90 95
 Phe Val Thr Leu Ile Gly Glu Asp Pro Ala Arg Thr Thr Ile Thr Tyr
 100 105 110
 His Leu Lys Ala Ser Asp Val Gly Leu Asp Gly Lys Pro Ile Gly Thr
 115 120 125
 Phe Arg Thr Pro Thr Met Val Val Asp Ala Asp Asp Phe Thr Ile Glu
 130 135 140
 Asn Leu Thr Ile Glu Asn Gly Ala Gly Pro Val Gly Gln Ala Leu Ala
 145 150 155 160
 Leu Arg Val Asp Gly Asp Arg Val Thr Val Arg Asn Ser Arg Leu Leu
 165 170 175
 Gly Trp Gln Asp Thr Ile Phe Leu Asn Arg Gly Arg His Tyr Phe Glu
 180 185 190
 Asp Ser Phe Ile Gly Gly His Val Asp Phe Ile Phe Gly Gly Ala Thr
 195 200 205
 Ala Val Phe Glu Arg Cys His Leu Arg Ala Trp Arg Asp Gly Tyr Leu
 210 215 220
 Thr Ala Ala Ser Thr Pro Ala Glu Gln Arg Phe Gly Phe Val Phe Leu
 225 230 235 240
 Asn Ser Ile Val Ser Gly Glu Ala Gly Ala Arg Thr Tyr Leu Gly Arg
 245 250 255
 Pro Trp Arg Ala Phe Ala His Val Ala Phe Ile Lys Thr Thr Met Gly
 260 265 270
 Glu Val Val Arg Pro Val Gly Trp Asn Asn Trp Asp Arg Pro Glu Arg
 275 280 285
 Glu Lys Thr Val Arg Phe Leu Glu Ala Gly Thr Ser Gly Ala Gly Gly
 290 295 300
 Ser Val Ala Ala Arg Val Ala Trp Ala Arg Val Ala Thr Pro Ala Glu
 305 310 315 320
 Leu Ala Asp Leu Thr Thr Glu Val Val Leu Gly Gly Thr Asp Gly Trp
 325 330 335
 Asp Pro Arg Arg Val Ala Pro Tyr Pro Ser Ala Val Arg Ala Asn Ala
 340 345 350
 Ala Pro Leu Pro Arg Pro Pro Gly Pro Asp Val Ala Gly Pro Gln Ser
 355 360 365
 Pro Pro Ala Leu Thr Trp Asp Gln Val Ala Arg Gln Pro Ala Ser Trp
 370 375 380
 Leu Ala Thr Pro Glu Ala Leu Arg Ile Ala Glu Asn Val Arg Leu Tyr
 385 390 395 400
 Gln Arg His Thr Gly Gly Trp Pro Lys Asn Leu Asp Met Ala Gln Pro
 405 410 415
 Leu Thr Asp Ala Asp Arg Ala Arg Leu Thr Ala Asp Arg Ala Leu Asp
 420 425 430
 Asp Ser Thr Ile Asp Asn Gly Ala Thr Thr Arg Gln Ile Glu Phe Leu
 435 440 445
 Ala Arg Ile Ala Ala Ala Asn Arg Asp Glu Arg Ala Gln Ala Ser Met
 450 455 460
 Leu Ala Gly Ile Asp Tyr Leu Leu Ala Ala Gln Tyr Pro Asn Gly Gly
 465 470 475 480
 Trp Pro Gln Tyr Phe Pro Leu Arg Asn Asp Tyr Ser Arg His Ile Thr
 485 490 495
 Phe Asn Asp Asp Ala Met Ile Ala Ala Ala Thr Ile Leu Gln Ser Val
 500 505 510
 Ala Leu Ala Arg Pro Pro Phe Ala Gly Val Asp Ala Thr Arg Arg Arg

515 520 525
 Arg Ala Ala Glu Ala Val Ala Arg Ala His Arg Val Ile Leu Ala Ser
 530 535 540
 Gln Ile Arg Val Asn Gly Gln Leu Thr Gly Trp Cys Gln Gln His Asp
 545 550 555 560
 Ala Arg Thr Leu Glu Pro Ala Arg Gly Arg Thr Tyr Glu His Pro Ser
 565 570 575
 Ile Ser Gly Arg Glu Thr Val Thr Ile Val Asn Phe Leu Arg Ser Ile
 580 585 590
 Glu Pro Arg Asp Arg Gln Thr Gln Ala Ala Ile Asp Ala Ala Met Glu
 595 600 605
 Trp Leu Lys Ala Val Gln Ile Arg Gly Trp Arg Thr Glu Arg Arg Pro
 610 615 620
 Asp Pro Ser Gly Pro Gly Gly Tyr Asp Val Val Met Val Glu Asp Pro
 625 630 635 640
 Asn Ala Ala Pro Leu Trp Ala Arg Phe Tyr Glu Ile Gly Thr Asn Arg
 645 650 655
 Pro Ile Tyr Ser Gly Arg Asp Gly Val Ile Lys Tyr Arg Leu Ala Glu
 660 665 670
 Ile Glu Ile Glu Arg Arg Thr Gly Tyr Ser Trp Val Gly Pro Tyr Ala
 675 680 685
 Gln Ala Leu Leu Asp Glu Glu Arg Arg Lys
 690 695

<210> 99

<211> 1782

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 99

```

atgttcaacta ctaacagctc tatttgccgc cggaaatccg cgcgttttcc actgactgcc 60
atggctgctg cgggtggctat gatcgcgggc acctctgcct ttgcggcctc taccggtggc 120
ttttcgacca cggatggcgg caatgtgtca gggcctaaat cctttaccgc ctcaagccac 180
acccaaatcc agcaaatcct tgaggatgcc aaagatggca attatccggt ggtgatcacc 240
tacaccggca atgaggattc actgattaac caagtctgoc gggatcacac cgtcgattct 300
tcaggcaact gccctaaagc gcgttggaat gatgcctacc gcaaagtcga aatcaaagaa 360
atgaccaagg gtgtcaccat tcagggtgcc aatggttcgt cggcgaattt cggaatcgtg 420
gtgaataaat ccagcaacgt gattattcgc aacatgaaga ttggtgact gggcggcgct 480
aataacgatg cggatatgat cgtgtggac agcgggtga acgtctggat cgatcacaac 540
gaattattcg ccgtgaacaa cgagtgttaag ggttcacccg atggcgatct gacctttgaa 600
agcgcgattg atatcaaaaa agcctcgcaa gatctacccg tgcctacaa cgtgattcgc 660
gacagtaaaa aagtcggttt ggatggctcc agcagcagcg atatcgccgg cggccgcaaa 720
attactttcc accacaatat ctaccgcaac gtagggtcgc gottaccttt gcagcgcggc 780
ggttggaagc acatgtacaa caacctgtac gacggcatta ccagctcggg catcaacgtg 840
cgccaaaacg gttatgcgtt aattgaaagc aactggttcc aaaacgcggg taaccgggtc 900
acctgcgctt ttgacagcag caactgcggc aagtgggac tgcgcaacaa taacatccgc 960
aaccgggtg attttgcgac ttacaacatc acctggacca gtggcgccac catcgacgcc 1020
accaactgga ccaccactgc gcccttcctc atcagcattc cctacagcta ttcaccggtt 1080
actcgcgaat gtgtgaaaga tcgtctggcg agttacgcgg gtgtgggtaa aaacggcgcg 1140
cagctgactg cctcgccctg cgggtggtgc gcatcgcca cacctgcac gtccacacct 1200
gcaagtcca gctctgcggc aaacagttcc gctgcacag gcagtgtgag ttgggtggc 1260
agtgcggtg atgcactcgt tgacattaac tggaccgtga atgccaacat taatgcgctg 1320
gaaatttatc aggatacgga ttctgatccc gccggacgtg tgcgattgc gtcgtgcca 1380
accagcgcca ccaactacac cgcaacaggt ctgagcaacg gcactacctt ttactttgg 1440
gtgaatatgc gcaccaacaa taatgtgtgg agcaactcca atgtgttcag cgccaagcca 1500
agttcaggta caaccccgtc atcatccagc agcgcgctt catcaacgcc aagtgtgtga 1560

```

```

ccggtgttaa gtgttacagg tgattaccca agcggcttct ccaagtgtgc tgatctgggt 1620
ggcacctgct cagtcgcctc gggcgatggt tgggttcctt ttggtcgcaa aggcaagtgg 1680
gtcaccaaaa aagtgtcagt cggtagctct attgcctgta ccggttgccg gtttgatctt 1740
gatccacaag gcaatcccaa taagtgttct tataaaaagt aa 1782

```

<210> 100

<211> 593

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(35)

<221> DOMAIN

<222> (36)...(593)

<223> Catalytic domain

<400> 100

```

Met Phe Thr Thr Asn Ser Ser Ile Cys Ala Arg Lys Ser Ala Arg Phe
1      5      10      15
Ser Leu Thr Ala Met Ala Ala Val Ala Met Ile Ala Gly Thr Ser
20      25      30
Ala Phe Ala Ala Ser Thr Gly Gly Phe Ser Thr Thr Asp Gly Gly Asn
35      40      45
Val Ser Gly Ser Lys Ser Phe Thr Ala Ser Ser His Thr Gln Ile Gln
50      55      60
Gln Ile Leu Glu Asp Ala Lys Asp Gly Asn Tyr Pro Val Val Ile Thr
65      70      75      80
Tyr Thr Gly Asn Glu Asp Ser Leu Ile Asn Gln Val Val Arg Asp His
85      90      95
Thr Val Asp Ser Ser Gly Asn Cys Pro Lys Ala Arg Trp Asn Asp Ala
100     105     110
Tyr Arg Lys Val Glu Ile Lys Glu Met Thr Lys Gly Val Thr Ile Gln
115     120     125
Gly Ala Asn Gly Ser Ser Ala Asn Phe Gly Ile Val Val Asn Lys Ser
130     135     140
Ser Asn Val Ile Ile Arg Asn Met Lys Ile Gly Ala Leu Gly Gly Ala
145     150     155     160
Asn Asn Asp Ala Asp Met Ile Arg Val Asp Ser Gly Val Asn Val Trp
165     170     175
Ile Asp His Asn Glu Leu Phe Ala Val Asn Asn Glu Cys Lys Gly Ser
180     185     190
Pro Asp Gly Asp Leu Thr Phe Glu Ser Ala Ile Asp Ile Lys Lys Ala
195     200     205
Ser Gln Asp Ile Thr Val Ser Tyr Asn Val Ile Arg Asp Ser Lys Lys
210     215     220
Val Gly Leu Asp Gly Ser Ser Ser Asp Ile Ala Gly Gly Arg Lys
225     230     235     240
Ile Thr Phe His His Asn Ile Tyr Arg Asn Val Gly Ala Arg Leu Pro
245     250     255
Leu Gln Arg Gly Gly Trp Thr His Met Tyr Asn Asn Leu Tyr Asp Gly
260     265     270
Ile Thr Ser Ser Gly Ile Asn Val Arg Gln Asn Gly Tyr Ala Leu Ile
275     280     285
Glu Ser Asn Trp Phe Gln Asn Ala Val Asn Pro Val Thr Cys Arg Phe
290     295     300

```

```

Asp Ser Ser Asn Cys Gly Lys Trp Asp Leu Arg Asn Asn Asn Ile Arg
305     310     315     320
Asn Pro Gly Asp Phe Ala Thr Tyr Asn Ile Thr Trp Thr Ser Gly Gly
325     330     335
Thr Ile Asp Ala Thr Asn Trp Thr Thr Thr Ala Pro Phe Pro Ile Ser
340     345     350
Ile Pro Tyr Ser Tyr Ser Pro Val Thr Pro Gln Cys Val Lys Asp Arg
355     360     365
Leu Ala Ser Tyr Ala Gly Val Gly Lys Asn Gly Ala Gln Leu Thr Ala
370     375     380
Ser Ala Cys Gly Gly Ala Ala Ser Ser Thr Pro Ala Ser Ser Thr Pro
385     390     395     400
Ala Ser Ser Ser Ser Ala Ala Asn Ser Ser Ala Ala Ser Gly Ser Val
405     410     415
Ser Leu Gly Gly Ser Ala Gly Asn Ala Ser Val Ala Leu Asn Trp Thr
420     425     430
Val Asn Ala Asn Ile Asn Ala Leu Glu Ile Tyr Gln Asp Thr Asp Ser
435     440     445
Asp Pro Ala Gly Arg Val Arg Ile Ala Ser Leu Pro Thr Ser Ala Thr
450     455     460
Asn Tyr Thr Ala Thr Gly Leu Ser Asn Gly Thr Thr Tyr Tyr Phe Trp
465     470     475     480
Val Lys Tyr Arg Thr Thr Asn Asn Val Trp Ser Asn Ser Asn Val Phe
485     490     495
Ser Ala Lys Pro Ser Ser Gly Thr Thr Pro Ser Ser Ser Ser Ala
500     505     510
Ala Ser Ser Thr Pro Ser Gly Ala Pro Val Leu Ser Gly Thr Gly Asp
515     520     525
Tyr Pro Ser Gly Phe Ser Lys Cys Ala Asp Leu Gly Gly Thr Cys Ser
530     535     540
Val Ala Ser Gly Asp Gly Trp Val Ala Phe Gly Arg Lys Gly Lys Trp
545     550     555     560
Val Thr Lys Lys Val Ser Val Gly Ser Ser Ile Ala Cys Thr Val Ala
565     570     575
Ala Phe Gly Ser Asp Pro Gln Gly Asn Pro Asn Lys Cys Ser Tyr Lys
580     585     590
Lys

```

<210> 101

<211> 1404

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 101

```

atgactatag accgtcgaga attccttata gacctcatta tcggcacccg cggcttcgca    60
atcgccaccga gtgatgcggt cggccaagct gatccatgga aaaccgtcta tccgcaaatac    120
ctcgctcgca tacggccacc gaaatttccg aagcgagatt tcatcatcac tagattcggc    180
gcaaagccgg gaaccgacag cggcgctgcg atcgcaaaag ctatcacccg gtgcagcaag    240
gcaggcgagg gacgtgttct cgttcccgca ggagagtttc tcaccggagc gatccatctg    300
aaatcgaaag taaactttca cgtgtcaaaa ggcgcgacgc tgaatttctc gaccgaccog    360
aaggcatatc tcccgattgt acatacgca tgggaaggaa tggagctgat gcatctgtca    420
ccgttcctct acgcttatga gcagacgaac atcgctatca cgggtcaggg aacgctcgac    480
ggccagggaa aatcattctt ctggaaatgg catggcaatc cggcttatgg cggcgatccg    540
aacacgctca gccacggccc cgtctgtgog cggctttacg agatgatgga taagaatgtg    600
ccggtcgccg aacgtgtctt cgtctcgga cattatctgc ggccgcagtt tattcagccg    660

```

```

tacaaatgca ggaacgtttt gatcgaagat gtgacgatcg tcgattcgcc gatgtgggaa    720
gttcatccgg tgcttttgca gaacgtcacg gtccgaaatg ttcacatttc atcgcatggt    780
ccgaacaatg accgatgcga tccggagtgc tgcaaggacg tactgatcga caactgtttt    840
ttcgacaccg gcgacgattg catcgogac aagtcgggcc gcaacaatga cggctgctgg    900
atcaatgtcc cgaccgagaa catcatcgtc cgcaactgca caatgaaaga cggctatggc    960
ggcatcacgg tcggcagtga gatttcggga ggctgtcgaa atctttttgc gcacgattgt    1020
cgactcgaca gtgcggatct ctggaccgcg cttcgcgta agaacaatgc gtccgagggc    1080
ggcaagctcg agaattttta ttttcggaat ataacggtcg gccaggtcgc acgcgctgtg    1140
gtcgagatcg attttaatta cgagggaagg gcgaaaaggct cgtatattcc tctcgttcga    1200
aattatgttg ttgaaggact gacatcgccc acaggcaatc gcgcgctcga tctgcaaggga    1260
ttggccaacg cgccgatcta caatgtaacg ctgcgaaact gtacgtttgg tctgtccga    1320
aatcgtatgt ttgtgaaaaa cgttcgtgga cttcggtcgc agaattgtga gatcgcgccg    1380
aggatgtaa acgaactggt atga                                     1404

```

<210> 102

<211> 467

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(28)

<221> DOMAIN

<222> (78)...(459)

<223> Catalytic domain

<400> 102

```

Met Thr Ile Asp Arg Arg Glu Phe Leu Ile Asp Leu Ile Ile Gly Thr
 1          5          10          15
Ala Gly Phe Ala Ile Ala Pro Ser Asp Ala Phe Gly Gln Ala Asp Pro
 20          25          30
Trp Lys Thr Val Tyr Pro Gln Ile Leu Ala Arg Ile Arg Pro Pro Lys
 35          40          45
Phe Pro Lys Arg Asp Phe Ile Ile Thr Arg Phe Gly Ala Lys Pro Gly
 50          55          60
Thr Asp Ser Ala Ala Ala Ile Ala Lys Ala Ile Thr Ala Cys Ser Lys
 65          70          75          80
Ala Gly Gly Gly Arg Val Leu Val Pro Ala Gly Glu Phe Leu Thr Gly
 85          90          95
Ala Ile His Leu Lys Ser Asn Val Asn Phe His Val Ser Lys Gly Ala
100          105          110
Thr Leu Lys Phe Ser Thr Asp Pro Lys Ala Tyr Leu Pro Ile Val His
115          120          125
Thr Arg Trp Glu Gly Met Glu Leu Met His Leu Ser Pro Phe Ile Tyr
130          135          140
Ala Tyr Glu Gln Thr Asn Ile Ala Ile Thr Gly Gln Gly Thr Leu Asp
145          150          155          160
Gly Gln Gly Lys Ser Phe Phe Trp Lys Trp His Gly Asn Pro Ala Tyr
165          170          175
Gly Gly Asp Pro Asn Thr Leu Ser Gln Arg Pro Ala Arg Ala Arg Leu
180          185          190
Tyr Glu Met Met Asp Lys Asn Val Pro Val Ala Glu Arg Val Phe Gly
195          200          205
Leu Gly His Tyr Leu Arg Pro Gln Phe Ile Gln Pro Tyr Lys Cys Arg
210          215          220
Asn Val Leu Ile Glu Asp Val Thr Ile Val Asp Ser Pro Met Trp Glu

```

```

225          230          235          240
Val His Pro Val Leu Cys Glu Asn Val Thr Val Arg Asn Val His Ile
          245          250          255
Ser Ser His Gly Pro Asn Asn Asp Gly Cys Asp Pro Glu Ser Cys Lys
          260          265          270
Asp Val Leu Ile Asp Asn Cys Phe Phe Asp Thr Gly Asp Asp Cys Ile
          275          280          285
Ala Ile Lys Ser Gly Arg Asn Asn Asp Gly Arg Arg Ile Asn Val Pro
          290          295          300
Thr Glu Asn Ile Ile Val Arg Asn Cys Thr Met Lys Asp Gly His Gly
305          310          315          320
Gly Ile Thr Val Gly Ser Glu Ile Ser Gly Gly Val Arg Asn Leu Phe
          325          330          335
Ala His Asp Cys Arg Leu Asp Ser Ala Asp Leu Trp Thr Ala Leu Arg
          340          345          350
Val Lys Asn Asn Ala Ser Arg Gly Gly Lys Leu Glu Asn Phe Tyr Phe
          355          360          365
Arg Asn Ile Thr Val Gly Gln Val Ala Arg Ala Val Val Glu Ile Asp
          370          375          380
Phe Asn Tyr Glu Glu Gly Ala Lys Gly Ser Tyr Ile Pro Val Val Arg
385          390          395          400
Asn Tyr Val Val Glu Gly Leu Thr Cys Ala Thr Gly Asn Arg Ala Val
          405          410          415
Asp Leu Gln Gly Leu Asp Asn Ala Pro Ile Tyr Asn Val Thr Leu Arg
          420          425          430
Asn Cys Thr Phe Gly Ser Val Arg Asn Arg Ser Val Val Lys Asn Val
          435          440          445
Arg Gly Leu Arg Leu Glu Asn Val Lys Ile Gly Gly Arg Ile Val Asn
          450          455          460
Glu Leu Val
465

```

<210> 103

<211> 1101

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 103

```

atgaacacccg cactgcaccg cgtcatccgc ctgccgctgc tgctggcgct gtgcctgccc      60
gcgctgcagg cacaggccac gcagaccgag cccgtgcgag agaacatgct gctgctgcag      120
accgcgtccg gcgctgggtc caagcaccac cagggcaagg cgtcgacta cggccacacg      180
ttcaccgatg ccgaacgtgc ggcgctgcgc gcgcccagacc gcagggacga tgcgacgatc      240
gacaacaagg cgaccacgct tgagatcgtc gcgctgctgg aagcccacca gcgcaccggc      300
aatgccgcct atctggcggc tgccgagcgc ggcgtggact acotgctggc cgcgcagtac      360
ccgaacggcg gctggccgca gtactaccgc gaccgttcgc tgtaccggca ccaggtcacc      420
ttcaacgatg atgcgatgac ccgcgtgctg gagctgctgc aggacatcgt cgagggcaag      480
ggcgcgctgg cgcagctgac acccacgcat ggcgaacgcg ccaggggcgc gctcgacagg      540
ggcatcgctt cgtgtgctgc caccacggta cggatcgatg gcgagctcac gctctgggcc      600
gcgcagtacg acgaagccac gctgcagccg gcgaaggcgc gctcctaaga gctgccatcg      660
ctggcggtcg ccgaatcggt cggggtgatg cggctgctga tgcgccagcc acagccgtcg      720
ccgcaggctc tgacggcggc cgaggccggc gcacgctggc tggaggcgca ccgatgccc      780
gacctggccc ggcaaaagat cgacgcgccc ggcgaagaaa ccggccagga cgtggtgatc      840
gtcgccgagc ccggcgcgct gctgtgggca cgcttctacg acctgcagca ccagcagccg      900
atgttcgtga accgcgaagg cgagcaggtg gcccgcttcg ccgacatgcc caacgaacgc      960
cgctcggtc acgcctggtg tggcgtgtgg ccggagaagc tgctgcagca ggagctgccca      1020
cgctggtaca acacccatgc cgaggcattg cgcgcgatta cgctgcgca tgccgagcca      1080

```

aggccgccga agcggccctg a

1101

<210> 104

<211> 366

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(26)

<221> DOMAIN

<222> (27)...(366)

<223> Catalytic domain

<400> 104

```

Met Asn Thr Ala Leu His Arg Val Ile Arg Leu Pro Leu Leu Leu Ala
 1      5      10      15
Leu Cys Leu Pro Ala Leu Gln Ala Gln Ala Thr Gln Thr Glu Pro Val
 20      25      30
Ala Glu Asn Met Leu Leu Leu Gln Thr Ala Ser Gly Gly Trp Ser Lys
 35      40      45
His His Gln Gly Lys Ala Val Asp Tyr Gly His Thr Phe Thr Asp Ala
 50      55      60
Glu Arg Ala Ala Leu Arg Ala Pro Asp Arg Arg Asp Asp Ala Thr Ile
 65      70      75      80
Asp Asn Lys Ala Thr Thr Leu Glu Ile Val Ala Leu Leu Glu Ala His
 85      90      95
Gln Arg Thr Gly Asn Ala Ala Tyr Leu Ala Ala Ala Gln Arg Gly Val
100      105      110
Asp Tyr Leu Leu Ala Ala Gln Tyr Pro Asn Gly Gly Trp Pro Gln Tyr
115      120      125
Tyr Pro Asp Arg Ser Leu Tyr Arg His Gln Val Thr Phe Asn Asp Asp
130      135      140
Ala Met Thr Arg Val Leu Glu Leu Leu Gln Asp Ile Val Glu Gly Lys
145      150      155      160
Gly Ala Leu Ala Gln Leu Thr Pro Thr His Gly Glu Arg Ala Arg Ala
165      170      175
Ala Leu Asp Arg Gly Ile Ala Cys Val Leu Ala Thr Gln Val Arg Ile
180      185      190
Asp Gly Glu Leu Thr Leu Trp Ala Ala Gln Tyr Asp Glu Ala Thr Leu
195      200      205
Gln Pro Ala Lys Ala Arg Ser Tyr Glu Leu Pro Ser Leu Ala Val Ala
210      215      220
Glu Ser Val Gly Val Met Arg Leu Leu Met Arg Gln Pro Gln Pro Ser
225      230      235      240
Pro Gln Val Leu Thr Ala Val Glu Ala Gly Ala Arg Trp Leu Glu Ala
245      250      255
His Arg Met Arg Asp Leu Ala Arg Arg Lys Ile Asp Ala Pro Gly Glu
260      265      270
Glu Thr Gly Gln Asp Val Val Ile Val Ala Glu Pro Gly Ala Ser Leu
275      280      285
Trp Ala Arg Phe Tyr Asp Leu Gln His Gln Gln Pro Met Phe Val Asn
290      295      300
Arg Glu Gly Glu Gln Val Ala Arg Phe Ala Asp Met Pro Asn Glu Arg
305      310      315      320
Arg Val Gly Tyr Ala Trp Tyr Gly Val Trp Pro Glu Lys Leu Leu Gln
325      330      335
Gln Glu Leu Pro Arg Trp Tyr Asn Thr His Ala Glu Ala Leu Arg Ala
340      345      350
Ile Thr Pro Ala His Ala Glu Pro Arg Pro Pro Lys Arg Pro
355      360      365

```

<210> 105

<211> 1203

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 105

```

atgcaattca tcgaaacaca gcaattgggg accgcccga aaccctggc gggacgagga 60
ggcgacaggc gctttccgcg ggtcatgccc gccgtttgcg cgggccttgc cctcgccgtg 120
tcgtcggccg agccggtccg ggcgcagggc gcggatgcgg atgcggatgg cccactgccc 180
aggtggaaca ggagggctgt ggtcgcgcc gaggaactgt tcgcctccga cgagggaacg 240
cgcggttgcg ccaacgtcct ccgctaccaa tcggcggaag gagcctggcc caaaaacacc 300
aatctggccg ccactcccct tcgcccgcag gacattccct cctcgacctc cggggtggcc 360
aacacgatcg acaatgaagc caccaccgtg cccattcggt ttttggcccg tttcgcgcaa 420
atcaacgagg acacggccag cgcgagggcg gtccagcgcg gattggacta tctcctcaag 480
gcgcaatata cgaacggtg ctggccgcag tatttccgc tcgcgcgagg ctaccactcg 540
cacatcacct acaacgacga cgccatggtg aatgtgctcg acctgctgct ggacgtgtcg 600
ctggcgaggg agccgttcga ttttggggac gaggatcgcc gccagcgggc cgcgaccgcc 660
gtggagcggg ggtatgaatg catcctccgc acccaaatcc ggcaggagga ccaaccacc 720
ggctggtgct cgcagtatga ccccgaaacc ttggccccgg cgtggggacg ggcgtacgag 780
ccgcccgcga tttccggagc cgagaccgtc ggcgtggcgc ggtttctgat gcggtggag 840
tcgccatcgc cggagccgtg cgaagccatc gagggcgcca tcgcctggct cgacacggtg 900
ggcatcgagg aattgcgtct cgaatggttc accaacagcg agggcaagcg tgaccggcgc 960
gtggtcgagg acgcttccgt gggcaccctt tggcgcgct tttacgaact cgaacgaac 1020
cgccccttgt tcgtggaccg cgacggggtg ctccgctacg acttcgagg actgacggcg 1080
gagcgccgcc aaggttacag ctactacggc acttggccgg cgccattgct ggccacggaa 1140
tatccgcgtc ggcgcaggat gaacgagtcg gccctgctcg agtcgtcctt catctcgcat 1200
tga 1203

```

<210> 106

<211> 400

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(43)

<221> DOMAIN

<222> (44)...(400)

<223> Catalytic domain

<400> 106

```

Met Gln Phe Ile Glu Thr Gln Gln Leu Gly Thr Ala Ala Lys Pro Val
 1      5      10      15
Ala Gly Arg Gly Gly Asp Arg Arg Phe Pro Arg Val Met Pro Ala Val
 20      25      30
Cys Ala Gly Leu Ala Leu Ala Val Ser Ser Ala Glu Pro Val Arg Ala
 35      40      45

```

```

Gln Gly Ala Asp Ala Asp Ala Asp Gly Pro Leu Pro Arg Trp Asn Arg
 50      55      60
Arg Leu Val Asp Arg Pro Glu Asp Trp Phe Ala Ser Asp Glu Gly Gln
 65      70      75      80
Arg Val Ala Ala Asn Val Leu Arg Tyr Gln Ser Ala Glu Gly Ala Trp
 85      90      95
Pro Lys Asn Thr Asn Leu Ala Ala Thr Pro Leu Arg Pro Glu Asp Ile
100      105      110
Pro Ser Ser Thr Ser Gly Val Ala Asn Thr Ile Asp Asn Glu Ala Thr
115      120      125
Thr Val Pro Ile Arg Phe Leu Ala Arg Phe Ala Gln Ile Asn Glu Asp
130      135      140
Thr Ala Ser Arg Glu Ala Val Gln Arg Gly Leu Asp Tyr Leu Leu Lys
145      150      155      160
Ala Gln Tyr Pro Asn Gly Gly Trp Pro Gln Tyr Phe Pro Leu Arg Arg
165      170      175
Gly Tyr His Ser His Ile Thr Tyr Asn Asp Asp Ala Met Val Asn Val
180      185      190
Leu Asp Leu Leu Leu Asp Val Ser Leu Gly Glu Glu Pro Phe Asp Phe
195      200      205
Val Asp Glu Asp Arg Arg Gln Arg Ala Ala Thr Ala Val Glu Arg Gly
210      215      220
Ile Glu Cys Ile Leu Arg Thr Gln Ile Arg Gln Glu Asp Gln Pro Thr
225      230      235      240
Gly Trp Cys Ala Gln Tyr Asp Pro Glu Thr Leu Ala Pro Ala Trp Gly
245      250      255
Arg Ala Tyr Glu Pro Pro Ser Ile Ser Gly Ala Glu Thr Val Gly Val
260      265      270
Ala Arg Phe Leu Met Arg Leu Glu Ser Pro Ser Pro Glu Ala Val Glu
275      280      285
Ala Ile Glu Gly Ala Ile Ala Trp Leu Asp Thr Val Gly Ile Glu Glu
290      295      300
Leu Arg Leu Glu Trp Phe Thr Asn Ser Glu Gly Lys Arg Asp Arg Arg
305      310      315      320
Val Val Glu Asp Ala Ser Val Gly Thr Leu Trp Ala Arg Phe Tyr Glu
325      330      335
Leu Glu Thr Asn Arg Pro Leu Phe Val Asp Arg Asp Gly Val Leu Arg
340      345      350
Tyr Asp Phe Ala Glu Leu Thr Ala Glu Arg Arg Gln Gly Tyr Ser Tyr
355      360      365
Tyr Gly Thr Trp Pro Ala Pro Leu Leu Ala Thr Glu Tyr Pro Arg Trp
370      375      380
Arg Arg Met Asn Glu Ser Ala Leu Leu Glu Ser Ser Phe Ile Ser His
385      390      395      400

```

<210> 107

<211> 1074

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 107

```

atgacgctac cgtttgtttc cctgcgcgta ctgctggcgc tgctggccac gtcgcgggtc    60
gcctgcgcgg gcgtgcggc acccgcgact gcgaccgata cggctgcgca gaacatgctg    120
cttctgcaga ccgcctccgg cggtctgttc aagcactacc gcgagaagaa ggtcgactac    180
gcgcgcgact acgacgcgcg cgagcgcgcc gcgtgcgcgc cgcgcgacgc gcatgacgat    240
gccacgatcg acaacaaggc caccgaccacc gagatcgcat acctggtgca ggcacatgcc    300

```

```

aggacgggca atccggccta cctcgaaggc gcgcggccgc gcgtcgagta cctgctgcgc 360
gcgcagttacc cgaacggcgg ctggcccgag ttctaccccg accattcgtc ctaccggcac 420
cagatcacgc tcaacgacga tgcgatggtg cagcccatca ccgtgctgca ggacatcgcc 480
gcggggccgca acggcatgca ggtgctggcg ccggagttcg gcgtccgcgc cgcgcgggcc 540
gcgcagcgcg gcacgggaaa cctgctcgag ttgcaggtgc ggatcgacgg ggtgccgacg 600
atctggggcg cgagtagca cgagaccacc ctgcaaccgg ccaaggcccg tgcgtacgag 660
ttgcctctgc tggccgtggc cgaatcggtg ggcgtgatgc gcctgctgat gcgccagccg 720
gggcctgatg cgcgcacgat cgcgcgcatc gagcgcgcg cggactggct ggaggcgcac 780
cgctcgcccg acctcgccct ggaacgcata gaagccccc cggaggaaac cggcaaggac 840
gtccgcgtcg tggccagacc gggcgcgctg ttgtggcgcg gcttctacga cctcgagcgg 900
caggtgccgc tgttcgtcga tcgcaacagc cgtccggttc cattcgccga gcttcccaac 960
gagcgtcgta ccggctatgg ctggtatggc acctggccgg aaaagctgct ggcacaggaa 1020
ctcccgcgct ggcgcaaggt ccacgcggcc agcgcgggcg ctccggcccg ttga 1074

```

<210> 108

<211> 357

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(31)

<221> DOMAIN

<222> (32)...(357)

<223> Catalytic domain

<400> 108

```

Met Thr Leu Pro Val Val Ser Leu Arg Val Leu Leu Ala Leu Leu Ala
 1      5      10      15
Thr Ser Pro Val Ala Cys Ala Gly Ala Ala Ala Pro Ala Thr Ala Thr
 20      25      30
Asp Pro Val Ala Glu Asn Met Leu Leu Leu Gln Thr Ala Ser Gly Gly
 35      40      45
Trp Ser Lys His Tyr Arg Glu Lys Lys Val Asp Tyr Ala Arg Asp Tyr
 50      55      60
Asp Ala Ala Glu Arg Ala Ala Leu Arg Ala Pro Asp Arg His Asp Asp
 65      70      75      80
Ala Thr Ile Asp Asn Lys Ala Thr Thr Thr Glu Ile Ala Tyr Leu Val
 85      90      95
Gln Ala His Ala Arg Thr Gly Asn Pro Ala Tyr Leu Asp Gly Ala Arg
100      105      110
Arg Gly Val Glu Tyr Leu Leu Arg Ala Gln Tyr Pro Asn Gly Gly Trp
115      120      125
Pro Gln Phe Tyr Pro Asp His Ser Ser Tyr Arg His Gln Ile Thr Leu
130      135      140
Asn Asp Asp Ala Met Val His Ala Ile Thr Val Leu Gln Asp Ile Ala
145      150      155      160
Ala Gly Arg Asn Gly Met Gln Val Leu Ala Pro Glu Phe Gly Val Arg
165      170      175
Ala Ala Ala Ala Ala Gln Arg Gly Ile Gly Asn Leu Leu Glu Leu Gln
180      185      190
Val Arg Ile Asp Gly Val Pro Thr Ile Trp Ala Ala Gln Tyr Asp Glu
195      200      205
Thr Thr Leu Gln Pro Ala Lys Ala Arg Ala Tyr Glu Leu Pro Ser Leu
210      215      220
Ala Val Ala Glu Ser Val Gly Val Met Arg Leu Leu Met Arg Gln Pro

```

```

225      230      235      240
Gly Pro Asp Ala Arg Thr Ile Ala Ala Ile Glu Ala Ala Asp Trp
245      250      255
Leu Glu Ala His Arg Leu Pro Asp Leu Ala Leu Glu Arg Ile Glu Ala
260      265      270
Pro Ala Glu Glu Thr Gly Lys Asp Val Arg Val Val Arg Pro Gly
275      280      285
Ala Ser Leu Trp Ala Arg Phe Tyr Asp Leu Glu Arg Gln Val Pro Leu
290      295      300
Phe Val Asp Arg Asn Ser Arg Pro Val Pro Phe Ala Glu Leu Pro Asn
305      310      315      320
Glu Arg Arg Thr Gly Tyr Gly Trp Tyr Gly Thr Trp Pro Glu Lys Leu
325      330      335
Leu Ala Gln Glu Leu Pro Arg Trp Arg Lys Val His Ala Ala Ser Ala
340      345      350
Gly Ala Pro Ala Arg
355

```

<210> 109

<211> 1422

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 109

atgacgacac	gacgcgaatt	catcaaaggc	tttctactta	cgggagcagc	cgtggccgtc	60
gctccgcgtt	tgcttgcgtt	cgccgcggag	gcaagtcogt	gggaaacgat	gatgccttcg	120
atcctcgcac	gcacacagac	acctcgtttt	cggaaacgca	ccttctatct	caatcgattc	180
ggcgccaagg	gtgatggagt	cacagactgc	accgcggctt	ttcatcgcg	gatcgatgaa	240
tgacccaaag	ccggcgggtg	gaaagtcgtc	gtgcggcg	gcacttatct	caccggcgcg	300
attcatttga	agagcaacgt	caacotcgaa	gtctcggaag	gcgcgacgat	caagttcagt	360
caggaccoga	aacactacct	gcctgttgtc	ttctcgcgtt	gggaagggtg	cgaagtcttc	420
aactactcgc	ctttcattta	cgcgttcgaa	cagcgaacaa	tcgcgatcac	cggcaaaaggc	480
acgtcgcgac	gacagagtga	ttcggaaacac	tggtggccgt	ggaacggcgg	tcgcgagtac	540
ggatggaaag	aagggatgaa	acagcagcgt	cccgatcgca	acgcgttggt	cacaatggcg	600
gagaaaggcg	tgccggtgcg	cgagcgcac	tttggcgaag	gtcattatct	gaggccgcag	660
ttcattcagc	cgtaccgctg	ccagaacgtg	ctgatccagg	gcgtgacgat	tcggaaactcg	720
ccgatgtggg	agattcatcc	ggtgttggtc	cgtaacgtga	ctattcacga	cgtgcacatc	780
gatagtcatg	gaccaaacaa	cgacggctgc	aatcccgaat	cgtgcagcga	cgtgttgatt	840
aaggtagctc	acttcgatac	cgccgacgac	tgcatcgcca	tcaaatcggg	acgcaacgcc	900
gacggcgggc	ggcttaaagc	cgccgactgag	aacatcatcg	ttcaaggatg	tcgcatgaaa	960
gacggccacg	gtggaatcac	ggtcggcagc	gagatctcgg	gcggcgtgcg	aaacctgttt	1020
gccgagaatt	gccgggtcga	cagtcacaa	ctcgatcacg	ccctgcgcgt	gaagaacaat	1080
gccatgcgcg	gcggattact	cgagaacttc	cacttccgta	acatcgaagt	cgggcagggtg	1140
gcccatgccg	tgattacgat	cgacttcaac	tacgaagagg	gcgcgaaaag	gtcgttcacg	1200
ccggtcgttc	gcgattacac	ggtcgcgtcg	ttgcgcagca	cgaagagcaa	gcacgcactc	1260
gacgtccagg	gtctgccccg	cgccgcggtc	atcaacctgc	gattgacgaa	ctgcacattc	1320
aacgatgtgc	agcaaccgaa	cattctcaag	aacgtcgaac	aatcaacctt	tgaaaaacgtc	1380
acgattaaag	gaaagacgat	cacacaaaca	ggatccaaag	aa		1422

<210> 110

<211> 474

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(21)

<221> DOMAIN

<222> (28)...(308)

<223> Pectin methyl esterase domain

<221> DOMAIN

<222> (309)...(637)

<223> Catalytic domain

<400> 110

```

Met Thr Thr Arg Arg Glu Phe Ile Lys Gly Phe Leu Leu Thr Gly Ala
1      5      10      15
Ala Val Ala Val Ala Pro Arg Leu Leu Ala Phe Ala Ala Glu Ala Ser
20      25      30
Pro Trp Glu Thr Met Met Pro Ser Ile Leu Ala Arg Ile Arg Pro Pro
35      40      45
Arg Phe Pro Lys Arg Thr Phe Tyr Leu Asn Arg Phe Gly Ala Lys Gly
50      55      60
Asp Gly Val Thr Asp Cys Thr Ala Ala Phe His Arg Ala Ile Asp Glu
65      70      75      80
Cys Thr Lys Ala Gly Gly Gly Lys Val Val Val Pro Ala Gly Thr Tyr
85      90      95
Leu Thr Gly Ala Ile His Leu Lys Ser Asn Val Asn Leu Glu Val Ser
100      105      110
Glu Gly Ala Thr Ile Lys Phe Ser Gln Asp Pro Lys His Tyr Leu Pro
115      120      125
Val Val Phe Ser Arg Trp Glu Gly Val Glu Val Phe Asn Tyr Ser Pro
130      135      140
Phe Ile Tyr Ala Phe Glu Gln Arg Asn Ile Ala Ile Thr Gly Lys Gly
145      150      155      160
Thr Leu Asp Gly Gln Ser Asp Ser Glu His Trp Trp Pro Trp Asn Gly
165      170      175
Arg Pro Gln Tyr Gly Trp Lys Glu Gly Met Lys Gln Gln Arg Pro Asp
180      185      190
Arg Asn Ala Leu Phe Thr Met Ala Glu Lys Gly Val Pro Val Arg Glu
195      200      205
Arg Ile Phe Gly Glu Gly His Tyr Leu Arg Pro Gln Phe Ile Gln Pro
210      215      220
Tyr Arg Cys Gln Asn Val Leu Ile Gln Gly Val Thr Ile Arg Asn Ser
225      230      235      240
Pro Met Trp Glu Ile His Pro Val Leu Cys Arg Asn Val Thr Ile His
245      250      255
Asp Val His Ile Asp Ser His Gly Pro Asn Asn Asp Gly Cys Asn Pro
260      265      270
Glu Ser Cys Ser Asp Val Leu Ile Lys Asp Ser Tyr Phe Asp Thr Gly
275      280      285
Asp Asp Cys Ile Ala Ile Lys Ser Gly Arg Asn Ala Asp Gly Arg Arg
290      295      300
Leu Lys Ala Pro Thr Glu Asn Ile Ile Val Gln Gly Cys Arg Met Lys
305      310      315      320
Asp Gly His Gly Gly Ile Thr Val Gly Ser Glu Ile Ser Gly Gly Val
325      330      335
Arg Asn Leu Phe Ala Glu Asn Cys Arg Leu Asp Ser Pro Asn Leu Asp
340      345      350
His Ala Leu Arg Val Lys Asn Asn Ala Met Arg Gly Gly Leu Leu Glu
355      360      365

Asn Phe His Phe Arg Asn Ile Glu Val Gly Gln Val Ala His Ala Val
370      375      380
Ile Thr Ile Asp Phe Asn Tyr Glu Glu Gly Ala Lys Gly Ser Phe Thr
385      390      395      400
Pro Val Val Arg Asp Tyr Thr Val Asp Arg Leu Arg Ser Thr Lys Ser
405      410      415
Lys His Ala Leu Asp Val Gln Gly Leu Pro Gly Ala Pro Val Ile Asn
420      425      430
Leu Arg Leu Thr Asn Cys Thr Phe Asn Asp Val Gln Gln Pro Asn Ile
435      440      445
Leu Lys Asn Val Glu Gln Ser Thr Phe Glu Asn Val Thr Ile Asn Gly
450      455      460
Lys Thr Ile Thr Gln Thr Gly Ser Lys Glu
465      470

```

<210> 111

<211> 1440

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 111

```

atgcaaaatc gtcgagaatt tttacaactt ttatttgccg gtgcggtgc cggacttgtt 60
ttgccgcaga tttctttcgg gcagactaaa caagccgacg cctggacgac cgaagtatccg 120
aagatttttag ccagaatcaa accgccgaaa ttctgcacaaa aagattttcc gatcaccaaa 180
tatggagccg ttgcggacgg gaaaaccctg gcgacccgaaa gcatcaaaaa agccatcgaa 240
gcgtgcgcca aatcgggcgg cgggcgcgtc gtctgcccc agggagaatt tttgaccggc 300
gcgattcatt tgaatcaaa cgtcaatctg cacatcacga aaggcgcgac cgtcaaat 360
tccaccaacc cgaagatta tctgccgacg gttcacacgc gctgggaagg gatggaattg 420
atgcataatt cgctttaat ttatgcctac gagcaaacca acatcgccgt caccggcgag 480
ggaacgctcg acgggcaggg caaggctttt ttctggaat ggacacgaaa cccgcgctac 540
ggcggaatc cggtgtgat cagccagcgt ccgcccgcgg cccggctgta tgaatgatg 600
gaaaaagcgg tgcctgtggc ggagcggatt ttccgcaaaa ctgagtatct tcgcccgag 660
tttatccagc cctataaatg caaaaatgtt ttgatcgaag gcgttaaat catcgattcg 720
ccgatgtggg aagttcaccc cgttttgtgc gaaaacgtga cgtccgaaa acttcatt 780
tctaccacg gaccgaacaa cgacgggtgc gatccgaaa gctgcaagg cgttttgatc 840
gaagactgct atttcgacac cggcgacgat tgcattgcca tcaaggcggg gcgcaatgaa 900
gacggcgac gcatcaatgt tccgaccgaa aacgtcgtcg tgcggcgggtg cgtgatgaag 960
gacggtcacg gcggaatcac catcggaagc gagatttcgg gcggcgtgag aaatgttttc 1020
gcgaaaaaca accggctcga cagcgcggat ttgtggactg cgctgagagt gaaaaacaac 1080
gcttcgcgg gcggaaaact ggagaatttt tacttcgcg atatcacctg cgggcaggtc 1140
tcgcgcgcgg tcgtcgaaat agattttaat tacgaggaag gcgctaaagg aaaacacacg 1200
ccgctgcttc gcaattacgt ggtcgaatgt ctaacctgca ataaaggcaa tcgagcggtc 1260
gatctgcagg gcttggacaa cgccccgatt tacgacatca cgtgaaaaa ctgtacgttt 1320
aacgtggtcg aaaagccgag cgtcgtgaaa aacgtcaaa gcgtcaaaact gaaaaacgtg 1380
aagattaacg gcaaaagtcgt cgagagtcgt gaaaatgctg caacgacggc taaaaataa 1440

```

<210> 112

<211> 479

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(27)

<221> DOMAIN

<222> (82)...(461)

<223> Catalytic domain

<400> 112

```

Met Gln Asn Arg Arg Glu Phe Leu Gln Leu Leu Phe Ala Gly Ala Gly
1      5      10      15
Ala Gly Leu Val Leu Pro Gln Ile Ser Phe Gly Gln Thr Lys Gln Ala
20      25      30
Asp Ala Trp Thr Thr Glu Tyr Pro Lys Ile Leu Ala Arg Ile Lys Pro
35      40      45
Pro Lys Phe Arg Lys Lys Asp Phe Pro Ile Thr Lys Tyr Gly Ala Val
50      55      60
Ala Asp Gly Lys Thr Leu Ala Thr Glu Ser Ile Lys Lys Ala Ile Glu
65      70      75      80
Ala Cys Ala Lys Ser Gly Gly Gly Arg Val Val Val Pro Gln Gly Glu
85      90      95
Phe Leu Thr Gly Ala Ile His Leu Lys Ser Asn Val Asn Leu His Ile
100     105     110
Thr Lys Gly Ala Thr Val Lys Phe Ser Thr Asn Pro Lys Asp Tyr Leu
115     120     125
Pro Ile Val His Thr Arg Trp Glu Gly Met Glu Leu Met His Ile Ser
130     135     140
Pro Leu Ile Tyr Ala Tyr Glu Gln Thr Asn Ile Ala Val Thr Gly Glu
145     150     155     160
Gly Thr Leu Asp Gly Gln Gly Lys Ala Phe Phe Trp Lys Trp His Gly
165     170     175
Asn Pro Arg Tyr Gly Gly Asn Pro Asp Val Ile Ser Gln Arg Pro Ala
180     185     190
Arg Ala Arg Leu Tyr Glu Met Met Glu Lys Gly Val Pro Val Ala Glu
195     200     205
Arg Ile Phe Gly Glu Thr Gln Tyr Leu Arg Pro Gln Phe Ile Gln Pro
210     215     220
Tyr Lys Cys Lys Asn Val Leu Ile Glu Gly Val Lys Ile Ile Asp Ser
225     230     235     240
Pro Met Trp Glu Val His Pro Val Leu Cys Glu Asn Val Thr Ile Arg
245     250     255
Lys Leu His Ile Ser Thr His Gly Pro Asn Asn Asp Gly Cys Asp Pro
260     265     270
Glu Ser Cys Lys Asp Val Leu Ile Glu Asp Cys Tyr Phe Asp Thr Gly
275     280     285
Asp Asp Cys Ile Ala Ile Lys Ala Gly Arg Asn Glu Asp Gly Arg Arg
290     295     300
Ile Asn Val Pro Thr Glu Asn Val Val Val Arg Gly Cys Val Met Lys
305     310     315     320
Asp Gly His Gly Gly Ile Thr Ile Gly Ser Glu Ile Ser Gly Gly Val
325     330     335
Arg Asn Val Phe Ala Glu Asn Asn Arg Leu Asp Ser Ala Asp Leu Trp
340     345     350
Thr Ala Leu Arg Val Lys Asn Asn Ala Ser Arg Gly Gly Lys Leu Glu
355     360     365
Asn Phe Tyr Phe Arg Asp Ile Thr Val Gly Gln Val Ser Arg Ala Val
370     375     380
Val Glu Ile Asp Phe Asn Tyr Glu Glu Gly Ala Lys Gly Lys His Thr
385     390     395     400
Pro Val Val Arg Asn Tyr Val Val Glu Asn Leu Thr Cys Asn Lys Gly
405     410     415
Asn Arg Ala Val Asp Leu Gln Gly Leu Asp Asn Ala Pro Ile Tyr Asp

```

```

420      425      430
Ile Thr Met Lys Asn Cys Thr Phe Asn Val Val Glu Lys Pro Ser Val
435      440      445
Val Lys Asn Val Lys Gly Val Lys Leu Glu Asn Val Lys Ile Asn Gly
450      455      460
Lys Val Val Glu Ser Leu Glu Asn Ala Ala Thr Thr Ala Lys Lys
465      470      475

```

<210> 113

<211> 1017

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 113

```

atgaagatat ttttaacaat attgctctcg gcattattca gcattttcaa tgcacagggtg      60
ctatcggatc ctgttgcgga tegtatgacc agctaccaac ttaaaaacgg aggctggccg      120
aagcaattgg cggataaatc tgttgtaaac tattcaaaac ctctctcacc tgctttgcaa      180
aaagtcatcg atcaatcgac cgaaaagtct gcgacaattg ataataatgc aaccacacgt      240
gagataaacc atcttctcct cgcttattcc aaaaccaaca atgacaagta tcttcaagcg      300
gcgacaaaag gtgttgagta tatcctgagt gctcaaaatg acaaaggagg atggcctcaa      360
tattatccag acagtagctc atatcgtggt cagatcacct acaatgacgg cgcgatgatt      420
aatgtattgg aaattttact ttccatatca acaaaaacaag agccctatgc tgttctaacg      480
aataaattta acgaaagaat agaaagggcc ttaacacgag ggattcactg catcttacaa      540
acccagggtt aacaaggaga taaactaacc atctgggccc cacagtacga tcagaaaaa      600
atggaaacct ctcaagccag actgtttgaa cggtagcgt tagcgacagc ggaatcggcg      660
ggcattctcc gctttttaat gcgtcttgac catcctactc cggaaataaa aaatgcaatc      720
aaccacgctg tagaatggtt ttccctccat aaagaggtag gctatgatta cgttaaaacg      780
gaaaaaaacg gaaaaacttt gcgggatttg gttttctcgc cggcctctac cgtatgggca      840
agatttttat acatcaggag gaatcaacc atctttggtg atcgcgataa tacgataaag      900
tattcgtcga atgaataaag cgaggaacga caaatggct actcttggtg tggtaactgg      960
ccagaaaaag taattacaaa agaatatgaa aaatggctta agaaggtaaa tgaataa      1017

```

<210> 114

<211> 338

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> SIGNAL

<222> (1)...(18)

<221> DOMAIN

<222> (19)...(388)

<223> Catalytic domain

<400> 114

```

Met Lys Ile Phe Leu Thr Ile Leu Leu Ser Ala Leu Phe Ser Ile Ser
 1           5           10           15
Asn Ala Gln Val Leu Ser Asp Pro Val Ala Asp Arg Met Thr Ser Tyr
          20           25           30
Gln Leu Lys Asn Gly Gly Trp Pro Lys His Leu Ala Asp Lys Ser Val
 35           40           45
Val Asn Tyr Ser Lys Pro Leu Ser Pro Ala Leu Gln Lys Val Ile Asp
 50           55           60

```

```

Gln Ser Thr Glu Lys Ser Ala Thr Ile Asp Asn Asn Ala Thr Thr Arg
65           70           75           80
Glu Ile Asn His Leu Leu Leu Ala Tyr Ser Lys Thr Asn Asn Asp Lys
          85           90           95
Tyr Leu Gln Ala Ala Thr Lys Gly Val Glu Tyr Ile Leu Ser Ala Gln
100           105           110
Asn Asp Lys Gly Gly Trp Pro Gln Tyr Tyr Pro Asp Ser Ser Tyr
115           120           125
Arg Gly Gln Ile Thr Tyr Asn Asp Gly Ala Met Ile Asn Val Leu Glu
130           135           140
Ile Leu Leu Ser Ile Ser Thr Lys Gln Glu Pro Tyr Ala Val Leu Thr
145           150           155           160
Asn Lys Phe Asn Glu Arg Ile Glu Arg Ala Leu Thr Arg Gly Ile His
165           170           175
Cys Ile Leu Gln Thr Gln Val Lys Gln Gly Asp Lys Leu Thr Ile Trp
180           185           190
Ala Ala Gln Tyr Asp Gln Lys Thr Met Glu Pro Ala Gln Ala Arg Leu
195           200           205
Phe Glu Pro Val Ala Leu Ala Thr Ala Glu Ser Ala Gly Ile Leu Arg
210           215           220
Phe Leu Met Arg Leu Asp His Pro Thr Pro Glu Ile Lys Asn Ala Ile
225           230           235           240
Asn His Ala Val Glu Trp Phe Ser Ser His Lys Glu Val Gly Tyr Asp
245           250           255
Tyr Val Lys Thr Glu Lys Asn Gly Lys Leu Leu Arg Asp Leu Val Ser
260           265           270
Ser Pro Ala Ser Thr Val Trp Ala Arg Phe Tyr Asp Ile Arg Thr Asn
275           280           285
Gln Pro Ile Phe Gly Asp Arg Asp Asn Thr Ile Lys Tyr Ser Leu Asn
290           295           300
Glu Ile Ser Glu Glu Arg Gln Asn Gly Tyr Ser Trp Tyr Gly Asn Trp
305           310           315           320
Pro Glu Lys Ile Ile Thr Lys Glu Tyr Glu Lys Trp Leu Lys Lys Val
325           330           335
Asn Glu

```

<210> 115

<211> 996

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 115

```

gtggccaagg cgatcgccgg tccgttgccg ccggcaccag ggcagggatc gccggtaacg      60
tggcgacaga ttctccggca gccatcgccg tggtagcggt ccgcggaacg gaaggcggtt      120
gccgaaaccc tgcgcgcgag ccagagagcc accggcggtt ggccgaagaa cacggattgg      180
acggcgctcc agagcgacgc tgagcggcag gcgctgcgaa atgcccgccg cgagaccgat      240
tcgacgatcg acaatggcgc caccgtcacc gagcttcgct ttctcaaccg cgtgtatgtc      300
gccacgcgcg acgagctttt accgggaggcc gtgcttcgct gcctcgacta cctgttggcg      360
tcgcagtaca gcaacggcgg ctggccacaa tactttccgt tcgggaccga ttactcgcgg      420
gacatcacgt tcaacgacga cgcgatgacc ggcggtgtgc tgctgctgaa ggatgccgcg      480
gacgggtcag caggttttca attcgtcgac aaggcgagac gtgaccgcgc tgcgcgggcc      540
gtgacgcgcg ccacgcgcgt gatcctccgc acgcagattc gggtcaacgg tacgtgacc      600
ggctgtgtcc agcagtacga cgcgcacgcg ctgacgcggc cgcgcggggc ctcgtacgag      660
catccgtcga ttgcgagccg cgagacggtc gggatcgccg ggctgctgat gggcgtagcg      720
aatccgtcgc cagagatcgt ggctgcggtt gacgcggctg ccgcatgggt gggtaaatcg      780

```

```

gaactgaagg gtgtgcccg ggcgacggcg ccaggacttt gggcgcgctt ctacgacatc 840
gctacgaatc ggcgcatcta ttcggggccgc gacggcggtca tcaagtaccg gctcgcagag 900
atcgagctcg agcggcgcac aggtacagc tgggttgccc cgtacgcgcg gccattttctg 960
acgaccgaat atccgaaatg gcgggcggca cgatga 996

```

<210> 116

<211> 331

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<221> DOMAIN

<222> (1)...(331)

<223> Catalytic domain

<400> 116

```

Met Ala Lys Ala Ile Gly Gly Pro Leu Pro Pro Ala Pro Gly Gln Gly
1      5      10      15
Ser Pro Val Thr Trp Ala Thr Ile Leu Arg Gln Pro Ser Pro Trp Tyr
20     25     30
Ala Ser Ala Asp Ala Lys Ala Val Ala Glu Thr Val Arg Ala Ser Gln
35     40     45
Arg Ala Thr Gly Gly Trp Pro Lys Asn Thr Asp Trp Thr Ala Leu Gln
50     55     60
Ser Asp Ala Glu Arg Gln Ala Leu Arg Asn Ala Arg Ala Glu Thr Asp
65     70     75     80
Ser Thr Ile Asp Asn Gly Ala Thr Val Thr Glu Leu Arg Phe Leu Thr
85     90     95
Arg Val Tyr Val Ala Thr Arg Asp Glu Leu Leu Arg Glu Ala Val Leu
100    105    110
Arg Gly Leu Asp Tyr Leu Leu Ala Ser Gln Tyr Ser Asn Gly Gly Trp
115    120    125
Pro Gln Tyr Phe Pro Leu Arg Thr Asp Tyr Ser Arg Asp Ile Thr Phe
130    135    140
Asn Asp Asp Ala Met Thr Gly Val Val Leu Leu Leu Lys Asp Ala Ala
145    150    155    160
Asp Gly Ser Ala Gly Phe Glu Phe Val Asp Lys Ala Arg Arg Asp Arg
165    170    175
Ala Ala Ala Ala Val Thr Arg Ala Ile Ala Val Ile Leu Arg Thr Gln
180    185    190
Ile Arg Val Asn Gly Thr Leu Thr Gly Trp Cys Gln Gln Tyr Asp Ala
195    200    205
Asp Ala Leu Thr Pro Ala Arg Gly Arg Ser Tyr Glu His Pro Ser Ile
210    215    220
Ala Ser Arg Glu Thr Val Gly Ile Ala Arg Leu Leu Met Gly Val Pro
225    230    235    240
Asn Pro Ser Pro Glu Ile Val Ala Ala Val Asp Ala Ala Ala Ala Trp
245    250    255
Leu Gly Lys Ser Glu Leu Lys Gly Val Pro Glu Ala Thr Ala Pro Gly
260    265    270
Leu Trp Ala Arg Phe Tyr Asp Ile Ala Thr Asn Arg Pro Ile Tyr Ser
275    280    285
Gly Arg Asp Gly Val Ile Lys Tyr Arg Leu Asp Glu Ile Glu Leu Glu
290    295    300
Arg Arg Thr Gly Tyr Ser Trp Val Gly Pro Tyr Ala Ala Ala Phe Leu
305    310    315    320
Thr Thr Glu Tyr Pro Lys Trp Arg Ala Ala Arg

```

325

330

<210> 117

<211> 1725

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<400> 117

```

atgaagaatt ttgggtttgg taactacaag tttttttag cggaatgtc tgtcgcgtct 60
ttttcgtatg cggcaagcta tacaccccg tcaacagcag tttegaaat caacagctat 120
cgaggctatt cggagctgac ttccagctgca tccggcatgg atatcgacca gtacacctac 180
aacatgacca cttggcaaat ccaaacgggc ggtttttaca aagccatggc cgacaagtat 240
aaaagcgctt atggcgcgcg tcaaaaatcc gaatggcaag ctaaaggcgg tggcgacctc 300
ggcactatag acaacaacgc cccatccag gaaatgcgtt tgcctgcgct gcgttacaaa 360
gaaacgcgca acaacaatta caaatccgca tttaagacaa gtttcaacaa ggcgttcaat 420
ttttttttga ccatgcagcg ctccaaaggc ggactccac aagtttggcc caaacgcggc 480
aactattctg accaaatcac gctaatgac aacgccatga tccgcgcat ggtcacgatg 540
atggatatcg ccaacaagac gagtccattt gattcggata tcatcgacga cgccacccgc 600
agcaaatga aatcgctctc cgacaagcg gtcgattact tgcctcaaggc gcaaatcgtg 660
aacgcggaag aggtcacggt atggtgcgcc cagcacgaca ccaacagcct cgccccgta 720
ggcgacgag cctacgaact cccgagcaaa tccggcaacg aatccatggg cgttgtgtgg 780
tttttgatga actggccaga ccaaaacgaa gcaatccaga aggcggtcaa aggcgcaatc 840
gcttggtaca aaaagaataa actaaaagac aaggcgttta gcaagaccgc aggcgttgtg 900
gacaaggcgg gttcatcgct gtggttccgc ttttacgaag tcaacaacga caactacttt 960
ttctgcgacc gcgatggtgc tagcaccaag acgcaggact tcatgaaat cagcgaagaa 1020
cgtcgcaagg gctaccagtg ggcaggcgat tacggctctg caattctagg caccgaaaaa 1080
gcataccttg aagcactcgc caagatggac gacaactatg ttccacctcc gccagcacca 1140
gctatgtgcg gaaacgcacac ttgcaaaacg tacatcgatg gcgttgactt tattgacatt 1200
caaggcgctg aggaacaacac caacacggga ttcgttggcg aaggttacgc caacgttgac 1260
aactccaccg gaagctatgt gacctacggc gtcaccgcct tcaagggaag caaatacact 1320
ttgtcatca gctttgcaaa cggcggtggt tccgcacgcg gttacagcgt ttctgcagga 1380
gacaagacgt tacttgacga cggcagcatg gaatctacag ccgcattggac cacttggaag 1440
atgcaatcca tcgaaatcga attgccaatg ggctatagcg aactcaagtt cacaagcctt 1500
tcgaaagacg gtatggcgaa catcgattac atcggctgga tgaacgatga ttgaaagtt 1560
ggcgaagtgg aagtaccacg ctcatccatt gaagcaatac gcgccatccg caaagcccag 1620
caggacaacc gctactttgt ggactttggc ggcaacaata atagccagcg ggcctacttt 1680
aagcgtggca tcaacacgct ccgcgtgaat gggaagatga ggtaa 1725

```

<210> 118

<211> 574

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<221> SIGNAL

<222> (1)...(24)

<221> DOMAIN

<222> (25)...(574)

<223> Catalytic domain

<400> 118

Met Lys Asn Phe Gly Phe Gly Asn Tyr Lys Phe Phe Val Ala Ala Met
1 5 10 15

```

Ser Val Ala Ser Phe Ser Tyr Ala Ala Ser Tyr Thr Pro Pro Ser Thr
20 25 30
Ala Val Ser Lys Ile Asn Ser Tyr Arg Gly Tyr Ser Glu Leu Thr Ser
35 40 45
Ala Ala Ser Gly Met Asp Ile Asp Gln Tyr Thr Tyr Asn Met Thr Thr
50 55 60
Trp Gln Ile Ala Asn Gly Gly Phe Tyr Lys Ala Met Ala Asp Lys Tyr
65 70 75 80
Lys Ser Ala Tyr Gly Gly Gly Gln Lys Ser Glu Trp Gln Ala Lys Gly
85 90 95
Gly Gly Asp Leu Gly Thr Ile Asp Asn Asn Ala Thr Ile Gln Glu Met
100 105 110
Arg Leu Leu Ala Val Arg Tyr Lys Glu Thr Thr Asn Asn Asn Tyr Lys
115 120 125
Ser Ala Phe Lys Thr Ser Phe Asn Lys Ala Val Asn Phe Leu Leu Thr
130 135 140
Met Gln Arg Ser Lys Gly Gly Leu Pro Gln Val Trp Pro Lys Arg Gly
145 150 155 160
Asn Tyr Ser Asp Gln Ile Thr Leu Asn Asp Asn Ala Met Ile Arg Ala
165 170 175
Met Val Thr Met Met Asp Ile Ala Asn Lys Thr Ser Pro Phe Asp Ser
180 185 190
Asp Ile Ile Asp Asp Ala Thr Arg Ser Lys Met Lys Ser Ala Leu Asp
195 200 205
Lys Ala Val Asp Tyr Leu Leu Lys Ala Gln Ile Val Asn Asp Gly Lys
210 215 220
Val Thr Val Trp Cys Ala Gln His Asp Thr Asn Ser Leu Ala Pro Val
225 230 235 240
Gly Ala Arg Ala Tyr Glu Leu Pro Ser Lys Ser Gly Asn Glu Ser Met
245 250 255
Gly Val Val Trp Phe Leu Met Asn Trp Pro Asp Gln Asn Glu Ala Ile
260 265 270
Gln Lys Ala Val Lys Gly Ala Ile Ala Trp Tyr Lys Lys Asn Lys Leu
275 280 285
Lys Asp Lys Ala Phe Ser Lys Thr Ala Gly Val Val Asp Lys Ala Gly
290 295 300
Ser Ser Leu Trp Phe Arg Phe Tyr Glu Val Asn Asn Asp Asn Tyr Phe
305 310 315 320
Phe Cys Asp Arg Asp Gly Ala Ser Thr Lys Thr Gln Asp Phe Met Lys
325 330 335
Ile Ser Glu Glu Arg Arg Lys Gly Tyr Gln Trp Ala Gly Asp Tyr Gly
340 345 350
Ser Ala Ile Leu Gly Thr Glu Asn Ala Tyr Leu Glu Ala Leu Ala Lys
355 360 365
Met Asp Asp Asn Tyr Val Pro Pro Pro Ala Pro Ala Met Cys Gly
370 375 380
Asn Asp Thr Cys Lys Thr Tyr Ile Asp Gly Val Asp Phe Ile Asp Ile
385 390 395 400
Gln Gly Val Lys Glu Thr Thr Asn Thr Gly Phe Val Gly Glu Gly Tyr
405 410 415
Ala Asn Val Asp Asn Ser Thr Gly Ser Tyr Val Thr Tyr Gly Val Thr
420 425 430
Ala Phe Lys Glu Gly Lys Tyr Thr Leu Phe Ile Ser Phe Ala Asn Gly
435 440 445
Gly Gly Ser Ala Arg Gly Tyr Ser Val Ser Ala Gly Asp Lys Thr Leu
450 455 460
Leu Ala Asp Gly Ser Met Glu Ser Thr Ala Ala Trp Thr Thr Trp Lys
465 470 475 480
Met Gln Ser Ile Glu Ile Glu Leu Pro Met Gly Tyr Ser Glu Leu Lys
485 490 495
Phe Thr Ser Leu Ser Lys Asp Gly Met Ala Asn Ile Asp Tyr Ile Gly
500 505 510
Trp Met Asn Asp Asp Leu Lys Val Gly Glu Val Glu Val Pro Arg Ser
515 520 525
Ser Ile Glu Ala Ile Arg Ala Ile Arg Lys Ala Gln Gln Asp Asn Arg
530 535 540
Tyr Phe Val Asp Phe Gly Gly Asn Asn Asn Ser Ala Gly Ala Tyr Phe
545 550 555 560
Lys Arg Gly Ile Asn Thr Phe Arg Val Asn Gly Lys Met Arg
565 570

```

<210> 119

<211> 1848

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<400> 119

```

gtgtcatggc aggaatccgg tgcggctatc accaacgcct ggaatgcaac gctcagtggc 60
tcaaacccctt acacagccgt atccgctggt tggaaatggca cacttgcccc caatgcatcg 120
gccacttttg gtttccaggc aaacgggtctt gccggtgcac ctaaagttaa tggcagcttg 180
tgcggcacca acacttcac aacacccgca tccagcagtg ttgccagcto gggtaaatca 240
agcgcgcccg tatcgctccag cagcagatca tccagttcaa tcgctatcac tagcagctct 300
ttagcgagaa gtctctattgc ctccagcagc tcaactagtt gtagctccag agcagcagct 360
agtgcgcgcaa gcgtttttctc ttttacgac caggaagagc aagcgggctt ctgtcgtgtt 420
gatggcattg cgacagaaag caccacaccc ggttttaccg gcaatggcta caccaatgcg 480
aacaacgcgc aaggcgcagc gattgaatgg gcagtcagcg cacttagcag tggcggttat 540
acagtagcct tccgcttcgc caatggcggc acagcagcgc gcaacggctc gttgttaatc 600
aatggcggta gcaatggtaa ttacactgtg gagttacccc tgaccggcgc atgggcaacc 660
tggcaaatgg ccagcgtgga aattgattta gtgcaaggca ataattttt aaaactctcg 720
gcgttaaccg ctgacgggtt ggccaatata gactcattaa aaatagacgg cgcgcaaac 780
aaagcaggta cttgcagcac tacatcaagc agcagcgttg ccagcagctc gtcgtccgtt 840
aaatccagcg caagttcttc ttcgagttca tccaccgctg caaaaatact gacattagac 900
ggtaaccgcg ccgccagctg gttcaacaaa tccaggacca agtggaatag cagccgcgcc 960
gatatttgtt tgccttacc gcaatccaac gccggttggc caaaaaacct ggattacaac 1020
tcagtgagcg caggcaatgg cgggagcgac agcgcacca tcgacaatgg tgcaaccatt 1080
accgaaatgg ttacctcgc tgaaatttat aaaaacggcg gcaacaccaa atatcgcat 1140
gcagtgcgca gagcagcaaa ctttttagtg agctcgcaat acagcacagg cgccttgcca 1200
caattttatc cgttgaaagg cggctatgcg gatcatgca cctttaacga taacggcatg 1260
gcgtacgcgt tgacgggtatt ggatttcgca gtaaacaaac gcgcaccgtt tgataacgac 1320
atcttctctg attctgatcg gccgaaatc aaaaacgcgt ttgccaaagg tgtggattac 1380
attttaaaag cgcagtgga acaaaatgga aaactcactg catggtgtgc acaacacggg 1440
gctacggatt accaaccgaa aaaagcgcgc gcttatgaat tggaaatcatt gagtggtagc 1500
gagtcggtcg gcattctcgc cttcttgatg acccaaccac aaaccgcgca aatcgaagcg 1560
gggtcaagg cgggtgtcaa ctggttcgcc agtccaata cttatttggc taactacact 1620
tacgattcat caaaagcgtc taccaccccg atttgtata aatccggaag cagaatgtgg 1680
tatcgcttct atgacctgaa caccaaccgt ggtttcttta gtgatcgca tggcagcaaa 1740
ttctatgata tcacccaat gtcagaagag cgtcgaccg gttatagctg gggtggtctc 1800
tacggtgaat ctattatttc cttcgcgcaa aaagtgggtt atctgtaa 1848

```

<210> 120

<211> 615

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<221> BINDING

<222> (1)...(61)

<223> Carbohydrate binding module

<221> BINDING

<222> (139)...(257)

<223> Carbohydrate binding module

<221> DOMAIN

<222> (258)...(615)

<223> Catalytic domain

<400> 120

```

Met Ser Trp Gln Glu Ser Gly Ala Ala Ile Thr Asn Ala Trp Asn Ala
 1      5      10      15
Thr Leu Ser Gly Ser Asn Pro Tyr Thr Ala Val Ser Ala Gly Trp Asn
 20      25      30
Gly Thr Leu Ala Pro Asn Ala Ser Ala Thr Phe Gly Phe Gln Ala Asn
 35      40      45
Gly Ser Ala Gly Ala Pro Lys Val Asn Gly Ser Leu Cys Gly Thr Asn
 50      55      60
Thr Ser Ser Thr Pro Ala Ser Ser Ser Val Ala Ser Ser Val Lys Ser
 65      70      75      80
Ser Ala Pro Val Ser Ser Ser Ser Arg Ser Ser Ser Ile Ala Ile
 85      90      95
Thr Ser Ser Ser Leu Ala Arg Ser Ser Ile Ala Ser Ser Ser Ser Leu
100      105      110
Val Ser Ser Ser Arg Ala Ser Ser Ser Ala Pro Ser Val Phe Ser Phe
115      120      125
Thr Ile Gln Glu Glu Gln Ala Gly Phe Cys Arg Val Asp Gly Ile Ala
130      135      140
Thr Glu Ser Thr Asn Thr Gly Phe Thr Gly Asn Gly Tyr Thr Asn Ala
145      150      155      160
Asn Asn Ala Gln Gly Ala Ala Ile Glu Trp Ala Val Ser Ala Pro Ser
165      170      175
Ser Gly Arg Tyr Thr Val Ala Phe Arg Phe Ala Asn Gly Gly Thr Ala
180      185      190
Ala Arg Asn Gly Ser Leu Leu Ile Asn Gly Gly Ser Asn Gly Asn Tyr
195      200      205
Thr Val Glu Leu Pro Leu Thr Gly Ala Trp Ala Thr Trp Gln Ile Ala
210      215      220
Ser Val Glu Ile Asp Leu Val Gln Gly Asn Asn Ile Leu Lys Leu Ser
225      230      235      240
Ala Leu Thr Ala Asp Gly Leu Ala Asn Ile Asp Ser Leu Lys Ile Asp
245      250      255
Gly Ala Gln Thr Lys Ala Gly Thr Cys Ser Thr Thr Ser Ser Ser Ser
260      265      270
Val Ala Ser Ser Ser Ser Ser Val Lys Ser Ser Ala Ser Ser Ser Ser
275      280      285
Ser Ser Ser Thr Ala Ala Lys Ile Leu Thr Leu Asp Gly Asn Pro Ala
290      295      300
Ala Ser Trp Phe Asn Lys Ser Arg Thr Lys Trp Asn Ser Ser Arg Ala
305      310      315      320
Asp Ile Val Leu Ser Tyr Gln Gln Ser Asn Gly Gly Trp Pro Lys Asn
325      330      335
Leu Asp Tyr Asn Ser Val Ser Ala Gly Asn Gly Gly Ser Asp Ser Gly
340      345      350

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Thr Ile Asp Asn Gly Ala Thr Ile Thr Glu Met Val Tyr Leu Ala Glu
355      360      365
Ile Tyr Lys Asn Gly Gly Asn Thr Lys Tyr Arg Asp Ala Val Arg Arg
370      375      380
Ala Ala Asn Phe Leu Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu Pro
385      390      395      400
Gln Phe Tyr Pro Leu Lys Gly Gly Tyr Ala Asp His Ala Thr Phe Asn
405      410      415
Asp Asn Gly Met Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala Val Asn
420      425      430
Lys Arg Ala Pro Phe Asp Asn Asp Ile Phe Ser Asp Ser Asp Arg Ala
435      440      445
Lys Phe Lys Thr Ala Val Ala Lys Gly Val Asp Tyr Ile Leu Lys Ala
450      455      460
Gln Trp Lys Gln Asn Gly Lys Leu Thr Ala Trp Cys Ala Gln His Gly
465      470      475      480
Ala Thr Asp Tyr Gln Pro Lys Lys Ala Arg Ala Tyr Glu Leu Glu Ser
485      490      495
Leu Ser Gly Ser Glu Ser Val Gly Ile Leu Ala Phe Leu Met Thr Gln
500      505      510
Pro Gln Thr Ala Gln Ile Glu Ala Ala Val Lys Ala Gly Val Asn Trp
515      520      525
Phe Ala Ser Pro Asn Thr Tyr Leu Ala Asn Tyr Thr Tyr Asp Ser Ser
530      535      540
Lys Ala Ser Thr Asn Pro Ile Val Tyr Lys Ser Gly Ser Arg Met Trp
545      550      555      560
Tyr Arg Phe Tyr Asp Leu Asn Thr Asn Arg Gly Phe Phe Ser Asp Arg
565      570      575
Asp Gly Ser Lys Phe Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg Arg
580      585      590
Thr Gly Tyr Ser Trp Gly Gly Ser Tyr Gly Glu Ser Ile Ile Ser Phe
595      600      605
Ala Gln Lys Val Gly Tyr Leu
610      615

```

<210> 121

<211> 1047

<212> DNA

<213> Bacteria

<400> 121

```

atgatgagat caagcatcgt caagctagtt gctttcagtg ttgtgggttat gttatgggtc 60
gggtgatccct ttcaaacggc agaagcgaat acgccaaatt tcaacttaca aggctttggc 120
acgttaaatg gggaacaac ttggtggtgca ggtggagatg tagtgacggt tcgtacaggg 180
aatgaatata taaacgcttt gaagtccaaa aaccctaata gtcggttaac aatttatgta 240
aacggtagca taacacctaag taatacgtct gatagtaaga tcgatattaa ggatgtttcc 300
aatgtatcga ttttaggggt tgggtacaaat ggacgattaa atgggacggt tattaaagta 360
tggcgagcga ataataatcat cattcgcaac ttgacgatcc atgaagtcca tacagtgat 420
aaagatgcga ttacgattga agggccctct cggaacattt ggattgacca taacgagctt 480
tatgccagct tgaacgttca taaagaccac tatgacggct tgtttgacgt aaagcgcgat 540
gcttacaata ttaccttctc ttggaattat gtccatgatg gctggaagc gatgctcatg 600
gggaactctg atagtataaa ctacgaccga aacataacat tccaccataa ctacttcaaa 660
aacttaact ctccggtacc tgcgtaccgt tttggaaaagg cgcacttggt tagcaattac 720
.tttgagaaca ttttagaacc aggcattaat tcacggatgg gagcggaat gctcgttgaa 780
cataacgttt ttgagaatgc caccaaccog ttaggattct ggcatagcag tcgaacaggt 840
tattggaatg ttgccaataa ccgctatatc aatagcacgg gtagcatgcc gaccacttcc 900
acgaccaatt atcgacctcc ttatccctat acggtcacac cagttggtga tgtgaaatcg 960
gttgtcacac gttatggcgg agttggtgtc atccagccgt atgcaagaaa gccatccgag 1020
cgattgctct ggtggctttt tgcataa 1047

```

<210> 122

<211> 348

<212> PRT

<213> Bacteria

<220>

<221> SIGNAL

<222> (1)...(29)

<221> DOMAIN

<222> (30)...(348)

<223> Catalytic domain

<400> 122

```

Met Met Arg Ser Ser Ile Val Lys Leu Val Ala Phe Ser Val Val Val
1      5      10      15
Met Leu Trp Leu Gly Val Ser Phe Gln Thr Ala Glu Ala Asn Thr Pro
20      25      30
Asn Phe Asn Leu Gln Gly Phe Ala Thr Leu Asn Gly Gly Thr Thr Gly
35      40      45
Gly Ala Gly Gly Asp Val Val Thr Val Arg Thr Gly Asn Glu Leu Ile
50      55      60
Asn Ala Leu Lys Ser Lys Asn Pro Asn Arg Pro Leu Thr Ile Tyr Val
65      70      75      80
Asn Gly Thr Ile Thr Pro Ser Asn Thr Ser Asp Ser Lys Ile Asp Ile
85      90      95
Lys Asp Val Ser Asn Val Ser Ile Leu Gly Val Gly Thr Asn Gly Arg
100     105     110
Leu Asn Gly Ile Gly Ile Lys Val Trp Arg Ala Asn Asn Ile Ile Ile
115     120     125
Arg Asn Leu Thr Ile His Glu Val His Thr Gly Asp Lys Asp Ala Ile
130     135     140
Ser Ile Glu Gly Pro Ser Arg Asn Ile Trp Ile Asp His Asn Glu Leu
145     150     155     160
Tyr Ala Ser Leu Asn Val His Lys Asp His Tyr Asp Gly Leu Phe Asp
165     170     175
Val Lys Arg Asp Ala Tyr Asn Ile Thr Phe Ser Trp Asn Tyr Val His
180     185     190
Asp Gly Trp Lys Ala Met Leu Met Gly Asn Ser Asp Ser Asp Asn Tyr
195     200     205
Asp Arg Asn Ile Thr Phe His His Asn Tyr Phe Lys Asn Leu Asn Ser
210     215     220
Arg Val Pro Ala Tyr Arg Phe Gly Lys Ala His Leu Phe Ser Asn Tyr
225     230     235     240
Phe Glu Asn Ile Leu Glu Thr Gly Ile Asn Ser Arg Met Gly Ala Glu
245     250     255
Met Leu Val Glu His Asn Val Phe Glu Asn Ala Thr Asn Pro Leu Gly
260     265     270
Phe Trp His Ser Ser Arg Thr Gly Tyr Trp Asn Val Ala Asn Asn Arg
275     280     285
Tyr Ile Asn Ser Thr Gly Ser Met Pro Thr Thr Ser Thr Thr Asn Tyr
290     295     300
Arg Pro Pro Tyr Pro Tyr Thr Val Thr Pro Val Gly Asp Val Lys Ser
305     310     315     320
Val Val Thr Arg Tyr Ala Gly Val Gly Val Ile Gln Pro Tyr Ala Arg
325     330     335
Lys Pro Ser Glu Arg Leu Leu Trp Trp Leu Phe Ala

```

340

345

<210> 123

<211> 1830

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<400> 123

```

ttgagtctac ttagtgaat gacccttttg cctgtaatgg caagtaacaa cgtagctccc      60
tggggctggg ccacotgctc cgatgagtc ggcacagctt atactctgaa cggaggttgc      120
ttttctgatg catcttccgt tactctgaaa gctcttggca atgaacaaac agatgacaaa      180
caaatcaaac aggtctatcg tcagaaagac atcatttatct tagatgggtc caatggcgat      240
ttcatcctta atgaatacat caagatttctg accaaaaaca aaacctcat tggtatcaac      300
aacgcccggc tgtgtacaaa gttctaccta accgctgatg atattacgta ccttaaagca      360
caaggactgg agggactgag tagtacaat caacatacag gaactctgcc tgatggcaca      420
acagtgacct gtgacgagcg tgcctttttc accaagaaag ccacatgga actccaatat      480
cagaaaaacg gatcctatac cctacccaat aatcaggta tcttttattt agatgccgct      540
tctgagaata tcactcatccg aaatatctcg ctgataggcg caggagccgt agatatagac      600
ggagctgacc tgattaccaa tcagggttaag cactgtctgga ttgaccattg cactgtttgtg      660
gactctcaag atgggtgccct ggacagcaag gtatgcgact gggccacctt tacctataac      720
cacttctact atacagaccg cagttactca catgcctaca cttgcgggtg cggatgggtc      780
agcaatcatg aaatgggtgat tcacatgacc ttgtcatgta atatctgggg agcaaatgt      840
atgcgtcgtc tgccgcaagc agatgactgt ttcatacacc ttgtgaacaa ctatcacaa      900
tgtcctggca atagtctcg tatgaccatt aacagttaca gcaagcatt ggttgagggt      960
aactatgctg ctgcaggtgt caacaagcca ttagatggca gtggggccaa ccgtaatgta      1020
acagctaaag ataatagttt tgcaaaactca caagccggtt ctgttgtgtc tgtgccatac      1080
gactatacca agattgcagc cgcgcgactt ccagctacgc tgactggaac agaggggtgca      1140
ggcgccacat taggcaacga tgcaacatac attctgtcta ctattccaac tgcgaccga      1200
caagaaggcg aatcttact ctactatttc attgatggc ttgtgggaac taatagtga      1260
ggctattcca ttataggtt taatgatggc gcaacattgc tgcgaacaa taaagagaaa      1320
gcattgtcta atgttagtgc aattcaactt ggtgacgata attatacgag tattaactt      1380
tctaaggag cagaaaacat cttcacagca cctactggca aaaaagtaag ttgtattacc      1440
ttctattctt atatcaatat aaaagaagaa aaactcgact tcaccaaata tccagaatat      1500
ggtttcgca cctgtttctg gcagaaagtt gccaacctca cttattctgc gacttctgat      1560
gacgtacaaa tcttgaaatc tcgtgatcca cagaatactg acgtggcatc attccattc      1620
actccaacaa atgttgtaag ttcaaaaaat tcaggatgaac agctttgttt cttaatgaaa      1680
gtcacctata gtgatgaaag cacagggtatc tctgctatcc agaaaaaat gcctatcgat      1740
ggcgttacct ataaccttca aggtatcogt atagataatc ccaccaaggg aatctatatt      1800
cagaacggaa agaaaatcat tatcaataa      1830

```

<210> 124

<211> 609

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<221> SIGNAL

<222> (1)...(21)

<221> DOMAIN

<222> (22)...(390)

<223> Catalytic domain

<400> 124

Leu Ser Leu Leu Ser Val Met Thr Leu Leu Pro Val Met Ala Ser Asn
 1 5 10 15
 Asn Val Ala Pro Trp Gly Trp Ala Thr Cys Ser Asp Glu Ser Ala Thr
 20 25 30
 Ala Tyr Thr Leu Asn Gly Gly Cys Phe Ser Asp Ala Ser Ser Val Thr
 35 40 45
 Leu Lys Ala Leu Gly Asn Glu Gln Thr Asp Asp Lys Gln Ile Lys Gln
 50 55 60
 Ala Ile Ala Gln Lys Asp Ile Ile Ile Leu Asp Gly Ser Asn Gly Asp
 65 70 75 80
 Phe Ile Leu Asn Glu Tyr Ile Lys Ile Ser Thr Lys Asn Lys Thr Ile
 85 90 95
 Ile Gly Ile Asn Asn Ala Arg Leu Cys Thr Lys Phe Tyr Leu Thr Ala
 100 105 110
 Asp Asp Ile Thr Tyr Leu Lys Ala Gln Gly Leu Glu Gly Leu Ser Ser
 115 120 125
 Thr Asn Gln His Thr Gly Thr Leu Pro Asp Gly Thr Thr Val Thr Cys
 130 135 140
 Asp Glu Arg Ala Phe Phe Thr Lys Lys Ala Ile Met Glu Leu Gln Tyr
 145 150 155 160
 Gln Lys Thr Gly Ser Tyr Thr Leu Pro Asn Lys Ser Gly Ile Phe Tyr
 165 170 175
 Leu Asp Ala Ala Ser Glu Asn Ile Ile Ile Arg Asn Ile Ser Leu Ile
 180 185 190
 Gly Pro Gly Ala Val Asp Ile Asp Gly Ala Asp Leu Ile Thr Asn Gln
 195 200 205
 Gly Lys His Val Trp Ile Asp His Cys Thr Phe Val Asp Ser Gln Asp
 210 215 220
 Gly Ala Leu Asp Ser Lys Val Cys Asp Trp Ala Thr Tyr Thr Tyr Asn
 225 230 235 240
 His Phe Tyr Tyr Thr Asp Arg Ser Tyr Ser His Ala Tyr Thr Cys Gly
 245 250 255
 Cys Gly Trp Val Ser Asn His Glu Met Val Ile His Met Thr Phe Ala
 260 265 270
 Cys Asn Ile Trp Gly Ala Lys Cys Met Arg Arg Leu Pro Gln Ala Asp
 275 280 285
 Asp Cys Phe Ile His Leu Val Asn Asn Tyr His Asn Cys Pro Gly Asn
 290 295 300
 Ser Val Gly Met Thr Ile Asn Ser Tyr Ser Lys Ala Leu Val Glu Gly
 305 310 315 320
 Asn Tyr Ala Ala Ala Gly Val Asn Lys Pro Leu Asp Gly Ser Gly Ala
 325 330 335
 Asn Arg Asn Val Thr Ala Lys Asp Asn Ser Phe Ala Asn Ser Gln Ala
 340 345 350
 Gly Ser Val Val Ser Val Pro Tyr Asp Tyr Thr Lys Ile Ala Ala Ala
 355 360 365
 Asp Val Pro Ala Thr Leu Thr Gly Thr Glu Gly Ala Gly Ala Thr Leu
 370 375 380
 Gly Asn Asp Ala Thr Tyr Ile Leu Ser Thr Ile Pro Thr Val Asp Arg
 385 390 395 400
 Gln Glu Gly Glu Ser Ser Leu Tyr Tyr Phe Ile Asp Gly Leu Val Gly
 405 410 415
 Thr Asn Ser Glu Gly Tyr Ser Ile Ile Glu Phe Asn Asp Gly Ala Thr
 420 425 430
 Leu Leu Leu Asn Asn Lys Glu Lys Ala Trp Ser Asn Gly Ser Ala Ile
 435 440 445
 Gln Leu Gly Asp Asp Asn Tyr Thr Ser Ile Lys Leu Ser Asn Gly Ala
 450 455 460
 Glu Asn Ile Phe Thr Ala Pro Thr Gly Lys Lys Val Ser Gly Ile Thr

 465 470 475 480
 Phe Tyr Ser Tyr Ile Asn Ile Lys Glu Glu Lys Leu Asp Phe Thr Lys
 485 490 495
 Tyr Pro Glu Tyr Gly Phe Arg Thr Cys Phe Trp Gln Lys Val Ala Asn
 500 505 510
 Leu Thr Tyr Ser Ala Thr Ser Asp Asp Val Gln Ile Leu Lys Ser Arg
 515 520 525
 Asp Pro Gln Asn Thr Asp Val Ala Ser Phe His Phe Thr Pro Thr Asn
 530 535 540
 Val Val Ser Phe Lys Asn Ser Gly Glu Gln Leu Cys Phe Leu Met Lys
 545 550 555 560
 Val Thr Tyr Ser Asp Glu Ser Thr Gly Ile Ser Ala Ile Gln Lys Lys
 565 570 575
 Met Pro Ile Asp Gly Val Thr Tyr Asn Leu Gln Gly Ile Arg Ile Asp
 580 585 590
 Asn Pro Thr Lys Gly Ile Tyr Ile Gln Asn Gly Lys Lys Ile Ile Ile
 595 600 605
 Lys

<210> 125

<211> 1170

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<400> 125

```

atgagggtcta aaatcatcag cgccataaat aattatagtg ttattattct cgaagggtcgc 60
aatggcgattt tcaatattag tgctacaatg agtttcagta gcaaatcaaa caaaaccata 120
gttggtgttaa ataatgctcg cctatgcacc aagttctatc taaccgatga aataaagact 180
gcgcgtcgatg ctgctaattgt aaaatcagca agttcaacca gtggagggtgg tacactctca 240
aatgggaaat cagtggtcaga acaacgtgaa taccttactc gtcaacaat tatcgatcta 300
actggcgatg cttcggaaatc gtgtcagaaa gcgggcatct ttagcttcag tagttgtacc 360
aatatcatca tgcgaaacct cgttttgggt ggccctggcc catgcgatgt aggtggcaac 420
gatttgcctt cgctcaactg ttctaagcat ttttgggtcg atcactgtga gtttaaccgat 480
ggatagatag gcaatttcga tattaccaag agtagcgatt tcaatactgt tacttggtgt 540
atattcaatt ataccgatcg tgcatacgac cacatgaact ccaatcttat tggtagctcc 600
gatagcgaag atgctgccta tttgaacact actatggcat gcaatatttg gggctacaag 660
tgcaatcagc gaatgccaat ggctcgtgct ggtaatatcc accttgtgaa caacttttac 720
gattgcgctg gcaatagtg ggctgttaac cctcgtaaaa attctgagtt cttagtcgag 780
aactgcctact ttgcccaggg tgtgaagcca ttctgcgaga gtggtgcgtt gggatacaac 840
tttattgatt gctatacaga agattcatal acttttcagc agagtgttac agtgtctgtg 900
ccatacgttt actctaagtt tgaatgtcaa ttagtaccgc agcaactcaa taaatatgct 960
ggcgcaacgc ttacttctcc gcttgcata ggtcgggaag aggtgtgtgt tactcctatt 1020
agtgtgtgtc ctgttgatag cgaatgtgtg ttggtcgaat actatcgcct gactggtaat 1080
cgtgttaaca cgctcaatag aggcataat atcgttagaa ctatttaacg caacggcaaa 1140
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<210> 126

<211> 389

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<221> DOMAIN

<222> (24)...(325)

<223> Catalytic domain

<400> 126

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1      5      10      15
Leu Asp Gly Ser Asn Gly Asp Phe Thr Ile Ser Ala Thr Met Ser Phe
20     25     30
Ser Ser Lys Ser Asn Lys Thr Ile Val Gly Val Asn Asn Ala Arg Leu
35     40     45
Cys Thr Lys Phe Tyr Leu Thr Asp Glu Ile Lys Thr Ala Leu Asp Ala
50     55     60
Ala Asn Val Lys Ser Ala Ser Ser Thr Ser Gly Gly Gly Thr Leu Ser
65     70     75     80
Asn Gly Lys Ser Val Ser Glu Gln Arg Glu Tyr Leu Thr Arg Gln Thr
85     90     95
Ile Ile Asp Leu Thr Gly Asp Ala Ser Glu Ser Cys Gln Lys Ala Gly
100    105    110
Ile Phe Ser Phe Ser Ser Cys Thr Asn Ile Ile Met Arg Asn Leu Val
115    120    125
Leu Val Gly Pro Gly Pro Cys Asp Val Gly Gly Asn Asp Leu Leu Ser
130    135    140
Leu Thr Gly Ser Lys His Phe Trp Val Asp His Cys Glu Leu Thr Asp
145    150    155    160
Gly Ile Asp Gly Asn Phe Asp Ile Thr Lys Ser Ser Asp Phe Asn Thr
165    170    175
Val Thr Trp Cys Ile Phe Asn Tyr Thr Asp Arg Ala Tyr Asp His Met
180    185    190
Asn Ser Asn Leu Ile Gly Ser Ser Asp Ser Glu Asp Ala Ala Tyr Leu
195    200    205
Asn Thr Thr Met Ala Cys Asn Ile Trp Gly Tyr Lys Cys Asn Gln Arg
210    215    220
Met Pro Met Ala Arg Ala Gly Asn Ile His Leu Val Asn Asn Phe Tyr
225    230    235    240
Asp Cys Ala Gly Asn Ser Val Ala Val Asn Pro Arg Lys Asn Ser Glu
245    250    255
Phe Leu Val Glu Asn Cys Tyr Phe Ala Thr Gly Val Lys Pro Phe Ser
260    265    270
Gln Ser Gly Ala Leu Gly Tyr Asn Phe Ile Asp Cys Tyr Thr Glu Asp
275    280    285
Ser Tyr Thr Phe Gln Gln Ser Gly Thr Val Ser Val Pro Tyr Val Tyr
290    295    300
Ser Lys Phe Asp Val Gln Leu Val Pro Glu Gln Leu Asn Lys Tyr Ala
305    310    315    320
Gly Ala Thr Leu Thr Ser Pro Leu Val Ile Gly Arg Glu Glu Gly Val
325    330    335
Val Thr Pro Ile Ser Ala Val Ser Val Asp Ser Asp Val Val Leu Val
340    345    350
Glu Tyr Tyr Ser Leu Thr Gly Asn Arg Val Asn Thr Leu Asn Arg Gly
355    360    365
Ile Asn Ile Val Arg Thr Ile Tyr Ala Asn Gly Lys Val Thr Thr Gln
370    375    380
Lys Val Leu Val Lys
385

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<210> 127

<211> 1449

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<400> 127

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aaagccagtg gcagtagcta cggcgatgta aaagacgccc atctgcaaaag ctactggcaa 180
cggcctgcta ataacggcca aagagtgtcg gttaagtggg gcagcgctat cagcgtaa 240
caggtaatac tgcgtgaaca gggcagtaat gtaaccagct ggcggctggt aaataatgac 300
aacggcgccag tattggcaac cggcaccagc attggcagca acagaacggt taacttcagc 360
actgtaagca cgaaaaaact caatctggaa atactaactg ccagcggtgc cccgcgcatt 420
gctgagtttg aagtttattt aaataccaat ggcggcaacc cgccaaatcc tactgaccgg 480
gaaccaggcc cggtaacttc ttgcgcagcg tctccacagg gctatgcctc gcttaacgg 540
ggcactaccg gcggcagtg cagcaacgcg gtcacggtaa cggtaagcac cggcgctcaa 600
atggtatcgg cgctacaaaa ccgcgatcta aaccggccgc tcactatccg ggtaatggc 660
actatcacac cgggtaattc tggcggtgtc agtaagtgtg acattaaaga tatggataat 720
gtcagcatta ttggtgtagg caacaatgcg ttgtttgacg gtatcggtat taaaatctgg 780
cgggccaaata acgttattat ccgcaacctt acaatgcgtt atgttaacac cggcgataaa 840
gacgctatta ccattgaagg cccggcgcggt aatatctgga ttgaccacaa cgaatctat 900
aacagcctga atgtgggtaa agatttttac gacgagctta taagcggtaa aaaagacgta 960
gataacgtaa ctatctctta caactacctg cagcacagct ggaaaacctc gctgtggggc 1020
agcagtgatt ccgacaacta caaccgcggt attaccttcc accataacca ctggcataag 1080
gtaaatccac gcctgccact gttccgtttt ggccagggcc atatttacia taactattac 1140
aacgacattc aggcacccgg tattaacagc cggatgggtg cggtaattcg tattgaaac 1200
aatgtgtttg aaaacgcgaa aaaccgata gtgtcgtttt attccagcgg ctacggttac 1260
tgggacaccc gcggtaatag ctttagcaat attacctggc aggaataccc cagcgacggc 1320
attatcgccg ggccaaatgt acaaccaca gcggtgctaa acctgcccta cagctttaac 1380
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agtttctaa

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<210> 128

<211> 482

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample.

<221> SIGNAL

<222> (1)...(24)

<221> DOMAIN

<222> (5)...(482)

<223> Catalytic domain

<400> 128

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1      5      10      15
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20      25      30
Gly Ala Gly Ala Asp Gly Ser Ser Lys Ala Ser Gly Ser Ser Tyr Gly
35      40      45
Asp Val Lys Asp Ala Asp Leu Gln Ser Tyr Trp Gln Pro Pro Ala Asn
50      55      60
Asn Gly Gln Arg Val Ser Val Lys Trp Ser Ser Ala Ile Ser Val Asn
65      70      75      80
Gln Val Ile Leu Arg Glu Gln Gly Ser Asn Val Thr Ser Trp Arg Leu
85      90      95

```

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Val Asn Asn Asp Asn Gly Ala Val Leu Ala Thr Gly Thr Ser Ile Gly
100 105 110
Ser Asn Arg Thr Val Asn Phe Ser Thr Val Ser Thr Lys Lys Leu Asn
115 120 125
Leu Glu Ile Leu Thr Ala Ser Gly Ala Pro Arg Ile Ala Glu Phe Glu
130 135 140
Val Tyr Leu Asn Thr Asn Gly Gly Asn Pro Pro Asn Pro Thr Asp Pro
145 150 155 160
Glu Pro Gly Pro Val Thr Ser Cys Ala Ala Ser Pro Gln Gly Tyr Ala
165 170 175
Ser Leu Asn Gly Gly Thr Thr Gly Gly Ser Gly Ser Asn Ala Val Thr
180 185 190
Val Thr Val Ser Thr Gly Ala Gln Met Val Ser Ala Leu Gln Asn Arg
195 200 205
Asp Leu Asn Arg Pro Leu Thr Ile Arg Val Asn Gly Thr Ile Thr Pro
210 215 220
Gly Asn Ser Gly Gly Val Ser Lys Phe Asp Ile Lys Asp Met Asp Asn
225 230 235 240
Val Ser Ile Ile Gly Val Gly Asn Asn Ala Leu Phe Asp Gly Ile Gly
245 250 255
Ile Lys Ile Trp Arg Ala Asn Asn Val Ile Ile Arg Asn Leu Thr Met
260 265 270
Arg Tyr Val Asn Thr Gly Asp Lys Asp Ala Ile Thr Ile Glu Gly Pro
275 280 285
Ala Arg Asn Ile Trp Ile Asp His Asn Glu Ile Tyr Asn Ser Leu Asn
290 295 300
Val Gly Lys Asp Phe Tyr Asp Glu Leu Ile Ser Gly Lys Lys Asp Val
305 310 315 320
Asp Asn Val Thr Ile Ser Tyr Asn Tyr Leu His Asp Ser Trp Lys Thr
325 330 335
Ser Leu Trp Gly Ser Ser Asp Ser Asp Asn Tyr Asn Arg Arg Ile Thr
340 345 350
Phe His His Asn His Trp His Lys Val Asn Ser Arg Leu Pro Leu Phe
355 360 365
Arg Phe Gly Gln Gly His Ile Tyr Asn Asn Tyr Tyr Asn Asp Ile Gln
370 375 380
Asp Thr Gly Ile Asn Ser Arg Met Gly Ala Val Ile Arg Ile Glu Asn
385 390 395 400
Asn Val Phe Glu Asn Ala Lys Asn Pro Ile Val Ser Phe Tyr Ser Ser
405 410 415
Gly Tyr Gly Tyr Trp Asp Thr Arg Gly Asn Ser Phe Ser Asn Ile Thr
420 425 430
Trp Gln Glu Tyr Pro Ser Asp Gly Ile Ile Ala Gly Pro Asn Val Gln
435 440 445
Pro Thr Ala Val Leu Asn Leu Pro Tyr Ser Phe Asn Leu Leu Pro Thr
450 455 460
Asn Gln Val Lys Ala His Val Leu Ala Asn Ala Gly Val Asn Lys Cys
465 470 475 480
Ser Phe

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<210> 129

<211> 1173

<212> DNA

<213> Bacillus halodurans ATCC 27557

<220>

<400> 129

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gtgggtgtga acaatgtctg cctatgcacc aaattctaca ttacgcccga gataaaagaa 180
gccctcgatg ctgcccgatg gaaatctaag agctcaagta gtggcactgg tggaaactctt 240
tctaattggt a cgtcggtcag tgaggctcgc gaattggcta ctgcgtcaaac gttgattgat 300
tatctcgggc atagctcaga atcggtatcag aaagctggta tctttggctt tagcaactgc 360
actaatatta ttatgcgcaa cattgttttc gttggccctg gtccatgcga tgtagggtggc 420
aacgacttgc tttcgctcgt tggttcgaag catttctggg tcgaccactg cgagtttacc 480
gatggcatcg atggcaactt cgacatcacc aagagtagcg acttcaacac cgtttcgtgg 540
tgcactttca gctatacoga ccgcgcatac gaccacatga attccaactt tattggtagc 600
tccgattcag agaatgcggc ttaccttaat actactatgg ctccaacgt ctggggcaat 660
aagtgc aatc agcgtatgcc tatggctcgt gccggttaata ttcaactcgt aaataattat 720
tacaactgcc ctggcaatag cgtggctgtg aatcctcgca aaaactcaga atttttgggtg 780
gagaattgct atttcgcaag tggcgtaag cctttctcgc agagcggcgc tcttagctat 840
ctatttatcg attgctacac cgaagatact tacaccttcc agaaactcgg ctctactacg 900
gtgccataca catatagcaa attcgatgct cagctgttcc ccgagcaact cacc caattc 960
gctggcgcaa cattgacttc gccgcttgtt attggtaggg aatctgagaa tgttacacca 1020
gtctcagtc a ttgctgcaaa tagcgatgtc atatctgtag aatactattc gctcactggc 1080
aagcgc atca gcgaaccaac taaaggcatc aatatcgtaa gaactattta tactaacggc 1140
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<210> 130

<211> 390

<212> PRT

<213> Bacillus halodurans ATCC 27557

<220>

<222> DOMAIN

<222> (38)...(326)

<223> Catalytic domain

<400> 130

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 1          5          10          15
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 20          25          30
Ser Gly Lys Ser Asn Lys Thr Ile Val Gly Val Asn Asn Ala Arg Leu
 35          40          45
Cys Thr Lys Phe Tyr Ile Thr Pro Glu Ile Lys Glu Ala Leu Asp Ala
 50          55          60
Ala Asp Val Lys Ser Lys Ser Ser Ser Gly Thr Gly Gly Thr Leu
 65          70          75          80
Ser Asn Gly Thr Ser Val Ser Glu Ala Arg Glu Leu Ala Thr Arg Gln
 85          90          95
Thr Leu Ile Asp Tyr Leu Gly Asp Ser Ser Glu Ser Tyr Gln Lys Ala
100          105          110
Gly Ile Phe Gly Phe Ser Asn Cys Thr Asn Ile Ile Met Arg Asn Ile
115          120          125
Val Phe Val Gly Pro Gly Pro Cys Asp Val Gly Gly Asn Asp Leu Leu
130          135          140
Ser Leu Val Gly Ser Lys His Phe Trp Val Asp His Cys Glu Phe Thr
145          150          155          160
Asp Gly Ile Asp Gly Asn Phe Asp Ile Thr Lys Ser Ser Asp Phe Asn
165          170          175
Thr Val Ser Trp Cys Thr Phe Ser Tyr Thr Asp Arg Ala Tyr Asp His
180          185          190
Met Asn Ser Asn Leu Ile Gly Ser Ser Asp Ser Glu Asn Ala Ala Tyr
195          200          205

```

```

Leu Asn Thr Thr Met Ala Ser Asn Val Trp Gly Asn Lys Cys Asn Gln
210          215          220
Arg Met Pro Met Ala Arg Ala Gly Asn Ile His Leu Val Asn Asn Tyr
225          230          235          240
Tyr Asn Cys Pro Gly Asn Ser Val Ala Val Asn Pro Arg Lys Asn Ser
245          250          255
Glu Phe Leu Val Glu Asn Cys Tyr Phe Ala Ser Gly Val Lys Pro Phe
260          265          270
Ser Gln Ser Gly Ala Leu Ser Tyr Leu Phe Ile Asp Cys Tyr Thr Glu
275          280          285
Asp Thr Tyr Thr Phe Gln Lys Ser Gly Ser Thr Thr Val Pro Tyr Thr
290          295          300
Tyr Ser Lys Phe Asp Ala Gln Leu Val Pro Glu Gln Leu Thr Gln Phe
305          310          315          320
Ala Gly Ala Thr Leu Thr Ser Pro Leu Val Ile Gly Arg Glu Ser Glu
325          330          335
Asn Val Thr Pro Val Ser Val Ile Ala Ala Asn Ser Asp Val Ile Ser
340          345          350
Val Glu Tyr Tyr Ser Leu Thr Gly Lys Arg Ile Ser Glu Pro Thr Lys
355          360          365
Gly Ile Asn Ile Val Arg Thr Ile Tyr Thr Asn Gly Asn Val Thr Thr
370          375          380
Gln Lys Val Leu Val Lys
385          390

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<210> 131

<211> 972

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 131

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accatcgaca atggtgcaac cattaccgaa atggtttacc tcgctgaaat ttataaaaac 240
ggcggcaaca ccaaatatcg cgaatcagtg cgcagagcag caaacttttt agtgagctcg 300
caatacagca caggcgctt gccacaattt tatccgttga aaggcggtta tgcggatcat 360
gcgaccttta acgataacgg catggcgtag gogttgacgg tatggattt cgcagtaaac 420
aaacgcgcac cgtttgataa cgacattttc tctgattctg atcgggcgaa attcaaaacc 480
gctgttgcca aaggtgtgga ttacatttta aaagcgagtg ggaacaaaaa tggaaaactc 540
actgcatggt gtgcacaaca cgggtgctac gattaccaac cgaaaaaagc gcgcgcttat 600
gaattggaat cattgagtg tagcgagtcg gtcggcattc tcgccttctt gatgacccaa 660
ccacaaccg cgcaaatcga agcggcggtc aaggcgggtg tcaactggtt cgccagtcga 720
aatacttatt tggctaacta cacttacgat tcatcaaaag cgtctaccaa cccgattgtg 780
tataaatccg gaagcagaat gtggtatcgc ttctatgacc tgaacaccaa ccgtggtttc 840
tttagtgatc gcgatggcag caaattctat gatataccac aaatgtcaga agagcgtcgc 900
accggttata gctggggtgg ctcttacggt gaattctatta ttctctcgc gcaaaaagtg 960
ggttatctgt ag
972

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<210> 132

<211> 323

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 132

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Met Ala Lys Ile Leu Thr Leu Asp Gly Asn Pro Ala Ala Ser Trp Phe
 1          5          10          15
Asn Lys Ser Arg Thr Lys Trp Asn Ser Ser Arg Ala Asp Ile Val Leu
 20          25          30
Ser Tyr Gln Gln Ser Asn Gly Gly Trp Pro Lys Asn Leu Asp Tyr Asn
 35          40          45
Ser Val Ser Ala Gly Asn Gly Gly Ser Asp Ser Gly Thr Ile Asp Asn
 50          55          60
Gly Ala Thr Ile Thr Glu Met Val Tyr Leu Ala Glu Ile Tyr Lys Asn
 65          70          75          80
Gly Gly Asn Thr Lys Tyr Arg Asp Ala Val Arg Arg Ala Ala Asn Phe
 85          90          95
Leu Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu Pro Gln Phe Tyr Pro
100          105          110
Leu Lys Gly Gly Tyr Ala Asp His Ala Thr Phe Asn Asp Asn Gly Met
115          120          125
Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala Val Asn Lys Arg Ala Pro
130          135          140
Phe Asp Asn Asp Ile Phe Ser Asp Ser Asp Arg Ala Lys Phe Lys Thr
145          150          155          160
Ala Val Ala Lys Gly Val Asp Tyr Ile Leu Lys Ala Gln Trp Lys Gln
165          170          175
Asn Gly Lys Leu Thr Ala Trp Cys Ala Gln His Gly Ala Thr Asp Tyr
180          185          190
Gln Pro Lys Lys Ala Arg Ala Tyr Glu Leu Glu Ser Leu Ser Gly Ser
195          200          205
Glu Ser Val Gly Ile Leu Ala Phe Leu Met Thr Gln Pro Gln Thr Ala
210          215          220
Gln Ile Glu Ala Ala Val Lys Ala Gly Val Asn Trp Phe Ala Ser Pro
225          230          235          240
Asn Thr Tyr Leu Ala Asn Tyr Thr Tyr Asp Ser Ser Lys Ala Ser Thr
245          250          255
Asn Pro Ile Val Tyr Lys Ser Gly Ser Arg Met Trp Tyr Arg Phe Tyr
260          265          270
Asp Leu Asn Thr Asn Arg Gly Phe Ser Asp Arg Asp Gly Ser Lys
275          280          285
Phe Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg Arg Thr Gly Tyr Ser
290          295          300
Trp Gly Gly Ser Tyr Gly Glu Ser Ile Ile Ser Phe Ala Gln Lys Val
305          310          315          320
Gly Tyr Leu

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<210> 133

<211> 972

<212> DNA

<213> Artificial Sequence

<220>

<223> Synthetically generated polynucleotide

<400> 133

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tggccaaaaaa acctggatta caactcagtg agcgcaggca atggcgggag cgacagcggc    180
accatogaca atggtgcaac cattaccgaa atggtttacc tcgctgaaat ttataaaaaac    240
ggcggaaca ccaaatatcg cgatgcagtg cgcagagcag caaacttttt agtgagctcg    300

```

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caatacagca caggcgccctt gccacaattt tatccgttga aaggcggcta tcatgatcat      360
gcgaccttta acgataacgg catggcgtag gcgttgacgg tattggattt ccagtaaaac    420
aaacgcgcac cgtttgataa cgacattttc tctgattctg atcgggcgaa attcaaaacc    480
gctgttgcca aaggtgtgga ttacatttta aaagcgtagt ggaaacaaaa tggaaaactc    540
actgcatggt gtgcacaaca cggtgctttg gattaccaac cgaaaaaagg tcgcgcttat    600
gaattggaat cattgagtgg taaggagtgc gtccggcattc tcgccttctt gatgacccaa    660
ccacaacccg cgcaaatcga agcggcggtc aaggcgggtg tcaacttggt cgccagtcga    720
aatacttatt tggctaacta cacttacgat tcatcaaaag cgtctacca cccgattgtg    780
tataaaaagg gaagcagaat gtggtatcgc ttctatgacc tgtataccaa ccgtggtttc    840
tttagtgatc gcgatggcag caaattctat gatataccc aaatgtcaga agagcgtcgc    900
accggttata gctggggtgg ctcttggggt gaagttatta tttccttcgc gcaaaaagtg    960
ggttatctgt ag                                     972

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<210> 134

<211> 323

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetically generated polypeptide

<400> 134

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Met Ala Lys Ile Leu Thr Leu Asp Gly Asn Pro Ala Ala Ser Trp Phe
 1           5           10           15
Asn Lys Ser Arg Thr Lys Trp Asn Ser Ser Arg Ala Asp Ile Val Leu
 20           25           30
Ser Tyr Gln Gln Ser Asn Gly Gly Trp Pro Lys Asn Leu Asp Tyr Asn
 35           40           45
Ser Val Ser Ala Gly Asn Gly Gly Ser Asp Ser Gly Thr Ile Asp Asn
 50           55           60
Gly Ala Thr Ile Thr Glu Met Val Tyr Leu Ala Glu Ile Tyr Lys Asn
 65           70           75           80
Gly Gly Asn Thr Lys Tyr Arg Asp Ala Val Arg Arg Ala Ala Asn Phe
 85           90           95
Leu Val Ser Ser Gln Tyr Ser Thr Gly Ala Leu Pro Gln Phe Tyr Pro
100           105           110
Leu Lys Gly Gly Tyr His Asp His Ala Thr Phe Asn Asp Asn Gly Met
115           120           125
Ala Tyr Ala Leu Thr Val Leu Asp Phe Ala Val Asn Lys Arg Ala Pro
130           135           140
Phe Asp Asn Asp Ile Phe Ser Asp Ser Asp Arg Ala Lys Phe Lys Thr
145           150           155           160
Ala Val Ala Lys Gly Val Asp Tyr Ile Leu Lys Ala Gln Trp Lys Gln
165           170           175
Asn Gly Lys Leu Thr Ala Trp Cys Ala Gln His Gly Ala Leu Asp Tyr
180           185           190
Gln Pro Lys Lys Gly Arg Ala Tyr Glu Leu Glu Ser Leu Ser Gly Lys
195           200           205
Glu Ser Val Gly Ile Leu Ala Phe Leu Met Thr Gln Pro Gln Thr Ala
210           215           220
Gln Ile Glu Ala Ala Val Lys Ala Gly Val Asn Trp Phe Ala Ser Pro
225           230           235           240
Asn Thr Tyr Leu Ala Asn Tyr Thr Tyr Asp Ser Ser Lys Ala Ser Thr
245           250           255
Asn Pro Ile Val Tyr Lys Lys Gly Ser Arg Met Trp Tyr Arg Phe Tyr
260           265           270
Asp Leu Tyr Thr Asn Arg Gly Phe Phe Ser Asp Arg Asp Gly Ser Lys
275           280           285
Phe Tyr Asp Ile Thr Gln Met Ser Glu Glu Arg Arg Thr Gly Tyr Ser
290           295           300
Trp Gly Gly Ser Trp Gly Glu Val Ile Ile Ser Phe Ala Gln Lys Val
305           310           315           320
Gly Tyr Leu

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REFERENCES CITED IN THE DESCRIPTION

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Patentkrav

1. Isoleret, syntetisk eller rekombinant nukleinsyre, der omfatter

- 5 (a) en nukleinsyresekvens, der har mindst 90 %, 91 %, 92 %, 93 %, 94 %, 95 %, 96 %, 97 %, 98 %, 99 % eller mere sekvensidentitet eller er 100 % identisk med SEQ ID NO: 77 over et område på mindst 950, 1000, 1050, 1100, 1150 eller
10 flere rester eller den fulde længde af et gen eller et transkript, hvor nukleinsyren koder for mindst ét polypeptid med en pektatlyaseaktivitet;
- (b) en nukleinsyresekvens, der omfatter en sekvens ifølge SEQ ID NO: 77;
- (c) en nukleinsyresekvens, der koder for et polypeptid med en
15 sekvens ifølge SEQ ID NO: 78, SEQ ID NO: 132 eller SEQ ID NO: 134;
- (d) en sekvens, der under stringente betingelser hybridiserer til en nukleinsyre, der omfatter SEQ ID NO: 77, SEQ ID NO: 131 eller SEQ ID NO: 133, hvor nukleinsyren koder for et
20 polypeptid med en pektatlyaseaktivitet, hvor de stringente betingelser inkluderer et vasketrin, der omfatter en vask i 0,2X SSC ved en temperatur på ca. 65 °C i ca. 15 minutter;
- (e) nukleinsyren fra (a), (b), (c) eller (d), der koder for en
25 pektatlyase, men mangler en signalsekvens, et præprodomæne eller en kombination deraf;
- (f) nukleinsyren fra (a), (b), (c), (d) eller (e), der koder for en pektatlyase, men også omfatter en heterolog sekvens; eller
- 30 (g) en sekvens, der er komplementær til (a), (b), (c), (d), (e) eller (f).

2. Nukleinsyreprobe til identificering af en nukleinsyre, der koder for et polypeptid med en pektatlyaseaktivitet, hvor
35 proben omfatter ca. 60 til 100 eller ca. 60 til 150 fortløbende baser af en sekvens, der omfatter SEQ ID NO: 77, SEQ ID NO: 131 eller SEQ ID NO: 133, hvor proben identificerer nukleinsyren ved at binde eller hybridisere under stringente

betingelser,

hvor de stringente betingelser inkluderer et vasketrin, der omfatter en vask i 0,1X SSC, 0,5 % SDS ved en temperatur på 68 °C i ca. 15 minutter.

5

3. Isoleret, syntetisk eller rekombinant nukleinsyre

(A) med en sekvens, der omfatter en sekvensmodifikation af SEQ ID NO: 131, hvor modifikationen af SEQ ID NO: 131 omfatter en eller flere af følgende ændringer:

10 nukleotiderne i resterne 352 til 354 er CAT eller CAC,
nukleotiderne i resterne 544 til 546 er GTG, GTT, GTC eller GTA,

nukleotiderne i resterne 568 til 570 er TTG, TTA, CTT, CTC, CTA eller CTG,

15 nukleotiderne i resterne 589 til 591 er GGT, GGC, GGA eller GGG,

nukleotiderne i resterne 622 til 624 er AAG eller AAA,

nukleotiderne i resterne 655 til 657 er ATG,

nukleotiderne i resterne 667 til 669 er GAG eller GAA,

20 nukleotiderne i resterne 763 til 765 er CGG, CGT, CGC, CGA, AGA, AGG,

nukleotiderne i resterne 787 til 789 er AAG eller AAA,

nukleotiderne i resterne 823 til 825 er TAT eller TAC,

nukleotiderne i resterne 925 til 927 er TGG, eller

25 nukleotiderne i resterne 934 til 936 er GTT, GTG, GTC eller GTA; eller

(B) med sekvensen ifølge krav 1(a), der yderligere omfatter en sekvensmodifikation, hvor sekvensmodifikationen omfatter en eller flere af følgende ændringer:

30 nukleotiderne i ækvivalenten til resterne 352 til 354 i SEQ ID NO: 131 er ændret til CAT eller CAC,

nukleotiderne i ækvivalenten til resterne 544 til 546 i SEQ ID NO: 131 er ændret til GTG, GTT, GTC eller GTA,

nukleotiderne i ækvivalenten til resterne 568 til 570 i SEQ ID

35 NO: 131 er ændret til TTG, TTA, CTT, CTC, CTA eller CTG,

nukleotiderne i ækvivalenten til resterne 589 til 591 i SEQ ID NO: 131 er ændret til GGT, GGC, GGA eller GGG,

nukleotiderne i ækvivalenten til resterne 622 til 624 i SEQ ID

NO: 131 er ændret til AAG eller AAA,
nukleotiderne i ækvivalenten til resterne 655 til 657 i SEQ ID
NO: 131 er ændret til ATG,
nukleotiderne i ækvivalenten til resterne 667 til 669 i SEQ ID
5 NO: 131 er GAG eller GAA,
nukleotiderne i ækvivalenten til resterne 763 til 765 i SEQ ID
NO: 131 er ændret til CGG, CGT, CGC, CGA, AGA, AGG,
nukleotiderne i ækvivalenten til resterne 787 til 789 i SEQ ID
NO: 131 er ændret til AAG eller AAA,
10 nukleotiderne i ækvivalenten til resterne 823 til 825 i SEQ ID
NO: 131 er ændret til TAT eller TAC,
nukleotiderne i ækvivalenten til resterne 925 til 927 i SEQ ID
NO: 131 er ændret til TGG, eller
nukleotiderne i ækvivalenten til resterne 934 til 936 i SEQ ID
15 NO: 131 er ændret til GTT, GTG, GTC eller GTA.

4. Fremgangsmåde til frembringelse af en variant af en
nukleinsyre, der koder for et polypeptid med en
pektatlyaseaktivitet, der omfatter
20 (A)
(a) tilvejebringelse af en templatenukleinsyre, der omfatter
en sekvens ifølge SEQ ID NO: 77; og
(b) modificering, deletering eller tilføjelse eller en
kombination deraf af et eller flere nukleotider i
25 templatesekvensen til frembringelse af en variant af
templatenukleinsyren,
hvor varianten har mindst 90 % identitet med
nukleinsyresekvensen ifølge krav 1; eller
(B)
30 (a) tilvejebringelse af en nukleinsyre, der koder for et
polypeptid med en pektatlyaseaktivitet, der omfatter en
sekvens ifølge krav 1; og
(b) identificering af en kodon i nukleinsyren fra trin (a) og
udskiftning af den med en anden kodon, der koder for den samme
35 aminosyre som den udskiftede kodon, til derved modificering af
kodoner i en nukleinsyre, der koder for en pektatlyase.

5. Ekspressionskassette, vektor eller kloningsvehikel, der

omfatter en nukleinsyre, der omfatter en sekvens ifølge krav 1 eller 3.

6. Værtscelle, der omfatter en ekspressionskassette, en vektor eller et kloningsvehikel ifølge krav 5.

7. Transgent ikke-humant dyr eller transgen plante eller transgent frø, der omfatter en sekvens ifølge krav 1 eller 3 eller omfatter en ekspressionskassette, en vektor eller et kloningsvehikel ifølge krav 5.

8. Fremgangsmåde til frembringelse af et rekombinant polypeptid, der omfatter trinene: (a) tilvejebringelse af en nukleinsyre, hvor nukleinsyren omfatter en sekvens ifølge krav 1 eller 3; og (b) ekspression af nukleinsyren fra trin (a) under betingelser, der åbner mulighed for ekspression af polypeptidet, til derved frembringelse af et rekombinant polypeptid, hvor eventuelt fremgangsmåden er til overekspression af polypeptidet ved anvendelse af en højaktivitetspromotor, en dicistronisk vektor eller ved genamplifikation af vektoren.

9. Isoleret, syntetisk eller rekombinant polypeptid,
(a) der har mindst 90 %, 91 %, 92 %, 93 %, 94 %, 95 %, 96 %, 97 %, 98 %, 99 % eller mere sekvensidentitet eller er 100 % identisk med SEQ ID NO: 78 over et område på mindst 300, 350, 400, 450, 500, 550, 600, 650 eller flere rester eller den fulde længde, og hvor polypeptidet har pektatlyaseaktivitet,
(b) der kodes af en nukleinsyre med en sekvens ifølge krav 1 eller 3,
(c) som er polypeptidet ifølge (a) eller (b), men mangler en signalsekvens, et præprodomæne eller en kombination deraf; eller
(d) som er polypeptidet ifølge (a), (b) eller (c), men også omfatter en heterolog sekvens; hvor polypeptidet har pektatlyaseaktivitet; eller
(e) som er polypeptidet ifølge (a), (b), (c) eller (d), hvor polypeptidet omfatter mindst ét glycosyleringssted eller er

glycosyleret.

10. Isoleret, syntetisk eller rekombinant polypeptid,

(A) der har en sekvens, der omfatter en sekvensmodifikation af
5 SEQ ID NO: 132, hvor modifikationen af SEQ ID NO: 132 omfatter
en eller flere af følgende mutationer: alanin i
aminosyreposition 118 er histidin, alanin i aminosyreposition
182 er valin, threonin i aminosyreposition 190 er leucin,
alanin i aminosyreposition 197 er glycin, serin i
10 aminosyreposition 208 er lysin, threonin i aminosyreposition
219 er methionin, threonin i aminosyreposition 223 er
glutaminsyre, serin i aminosyreposition 255 er arginin, serin
i aminosyreposition 263 er lysin, asparagin i
aminosyreposition 275 er tyrosin, tyrosin i aminosyreposition
15 309 er tryptofan, eller serin i aminosyreposition 312 er
valin; eller

(B) der har sekvensen ifølge krav 9 og omfatter en
sekvensmodifikation, hvor sekvensmodifikationen omfatter en
eller flere af følgende ændringer:

20 aminosyren i ækvivalenten til alanin i rest 118 i SEQ ID NO:
132 er ændret til en histidin,
aminosyren i ækvivalenten til alanin i rest 182 i SEQ ID NO:
132 er ændret til en valin,
aminosyren i ækvivalenten til threonin i rest 190 i SEQ ID NO:
25 132 er ændret til en leucin,
aminosyren i ækvivalenten til alanin i rest 197 i SEQ ID
NO: 132 er ændret til en glycin,
aminosyren i ækvivalenten til serin i rest 208 i SEQ ID NO:
132 er ændret til en lysin,
30 aminosyren i ækvivalenten til threonin i rest 219 i SEQ ID
NO: 132 er ændret til en methionin,
aminosyren i ækvivalenten til threonin i rest 223 i SEQ ID NO:
132 er ændret til en glutaminsyre,
aminosyren i ækvivalenten til serin i rest 255 i SEQ ID
35 NO: 132 er ændret til en arginin,
aminosyren i ækvivalenten til serin i rest 263 i SEQ ID NO:
132 er ændret til en lysin,
aminosyren i ækvivalenten til asparagin i rest 275 i SEQ ID

NO: 132 er ændret til en tyrosin,
aminosyren i ækvivalenten til tyrosin i rest 309 i SEQ ID NO:
132 er ændret til en tryptofan, eller
aminosyren i ækvivalenten til serin i rest 312 i SEQ ID NO:
5 132 er ændret til en valin.

11. Heterodimer eller homodimer, der omfatter et polypeptid
ifølge et hvilket som helst af kravene 9 eller 10.

10 12. Immobiliseret polypeptid, hvor polypeptidet omfatter en
sekvens ifølge et hvilket som helst af kravene 9 eller 10.

13. Isoleret, syntetisk eller rekombinant antistof, der
binder specifikt til et polypeptid ifølge krav 9 eller 10.

15

14. Fremgangsmåde til hydrolysering, flydendegørelse,
fjernelse, opløsning eller opbrydning af en pektin-, pektat
(polygalacturonsyre)-, homogalacturonan-, rhamnogalacturonan-
eller plantecellevægsomfattende sammensætning, der omfatter
20 følgende trin:

(a) tilvejebringelse af et polypeptid med en
pektatlyaseaktivitet ifølge et hvilket som helst af kravene 9
eller 10;

(b) tilvejebringelse af en sammensætning, der omfatter en
25 pektin, en pektat, homogalacturonan, rhamnogalacturonan eller
plantecellevæg; og

(c) etablering af kontakt mellem polypeptidet fra trin (a) og
sammensætningen fra trin (b) under betingelser, hvor
polypeptidet hydrolyserer, flydendegør, fjerner, opløser eller
30 opbryder sammensætningen.

15. Sammensætning, der omfatter et polypeptid ifølge et
hvilket som helst af kravene 9 eller 10 eller en nukleinsyre
ifølge et hvilket som helst af kravene 1 eller 3.

35

16. Fremgangsmåde til vask af et emne, der omfatter følgende
trin:

(a) tilvejebringelse af en sammensætning, der omfatter et

polypeptid med en pektatlyaseaktivitet ifølge et hvilket som helst af kravene 9 eller 10;

(b) tilvejebringelse af et emne; og

(c) etablering af kontakt mellem polypeptidet fra trin (a) og emnet fra trin (b) under betingelser, hvor sammensætningen kan vaske emnet.

17. Fremgangsmåde til rensning af fiber, tråd, tekstil, stof eller cellulosemateriale, der omfatter følgende trin:

10 (a) tilvejebringelse af et polypeptid med en pektatlyaseaktivitet ifølge et hvilket som helst af kravene 9 eller 10;

(b) tilvejebringelse af en fiber, en tråd, et tekstil, et stof eller et cellulosemateriale; og

15 (c) etablering af kontakt mellem polypeptidet fra trin (a) og fiberen, tråden, tekstilet, stoffet eller cellulosematerialet fra trin (b) under betingelser, hvor pektatlyasen kan rense fiberen, tråden, tekstilet, stoffet eller cellulosematerialet.

20 18. Fremgangsmåde til forbedring af ekstraktion af olie fra et olierigt plantemateriale, der omfatter følgende trin:

(a) tilvejebringelse af et polypeptid med en pektatlyaseaktivitet ifølge et hvilket som helst af kravene 9 eller 10;

25 (b) tilvejebringelse af et olierigt plantemateriale; og

(c) etablering af kontakt mellem polypeptidet fra trin (a) og det olierige plantemateriale.

30 19. Fremgangsmåde til fremstilling af en frugt- eller grønsagsjuice, sirup, puré eller ekstrakt, der omfatter følgende trin:

(a) tilvejebringelse af et polypeptid med en pektatlyaseaktivitet ifølge et hvilket som helst af kravene 9 eller 10;

35 (b) tilvejebringelse af en sammensætning eller en væske, der omfatter et frugt- eller grønsagsmateriale; og

(c) etablering af kontakt mellem polypeptidet fra trin (a) og sammensætningen til derved fremstilling af frugt- eller

grønsagsjuicen, siruppen, puréen eller ekstraktet.

20. Fremgangsmåde til behandling af et papir, et papirprodukt eller en træmasse, der omfatter følgende trin:

- 5 (a) tilvejebringelse af et polypeptid med en pektatlyaseaktivitet ifølge et hvilket som helst af kravene 9 eller 10;
- (b) tilvejebringelse af en sammensætning, der omfatter et papir, et papirprodukt eller en træmasse; og
- 10 (c) etablering af kontakt mellem polypeptidet fra trin (a) og sammensætningen fra trin (b) under betingelser, hvor pektatlyasen kan behandle papiret, papirproduktet, papirmassen eller træmassen.

15 21. Fremgangsmåde til biologisk rensning, der omfatter følgende trin:

- (a) tilvejebringelse af en pektatlyase ifølge et hvilket som helst af kravene 9 eller 10;
- (b) tilvejebringelse af et pektin-, polygalacturonsyre- (pektat-), homogalacturonan-, rhamnogalacturonan- eller
- 20 plantecellevægsomfattende materiale;
- (c) etablering af kontakt mellem pektatlyasen fra (a) og materialet fra (b) under alkaliske betingelser i en bicarbonatbuffer eller tilsvarende.

25

22. Isoleret, syntetisk eller rekombinant nukleinsyre ifølge krav 1, hvor nukleinsyren omfatter SEQ ID NO: 133.

23. Isoleret, syntetisk eller rekombinant polypeptid ifølge

30 krav 9, hvor polypeptidet omfatter SEQ ID NO: 134.

24. Isoleret, syntetisk eller rekombinant polypeptid ifølge krav 9, hvor polypeptidet omfatter SEQ ID NO: 132.

35 25. Formulering til behandling af et materialt, hvilken formulering omfatter mindst ét polypeptid ifølge krav 9 eller 10 ved en dosis, der er udvalgt blandt:

i. mellem 1 og 100 g pr. ton materiale, der skal behandles;

- ii. mellem 1 µg og 100 µg pr. g material, der skal behandles;
- iii. mellem 0,5 mg og 50 mg pr. 0,45 kg materiale, der skal behandles;
- iv. en dosis, der omfatter en enzymstyrke på mellem 100 og 5 40.000 enheder/ml.

DRAWINGS

FIGURE 1

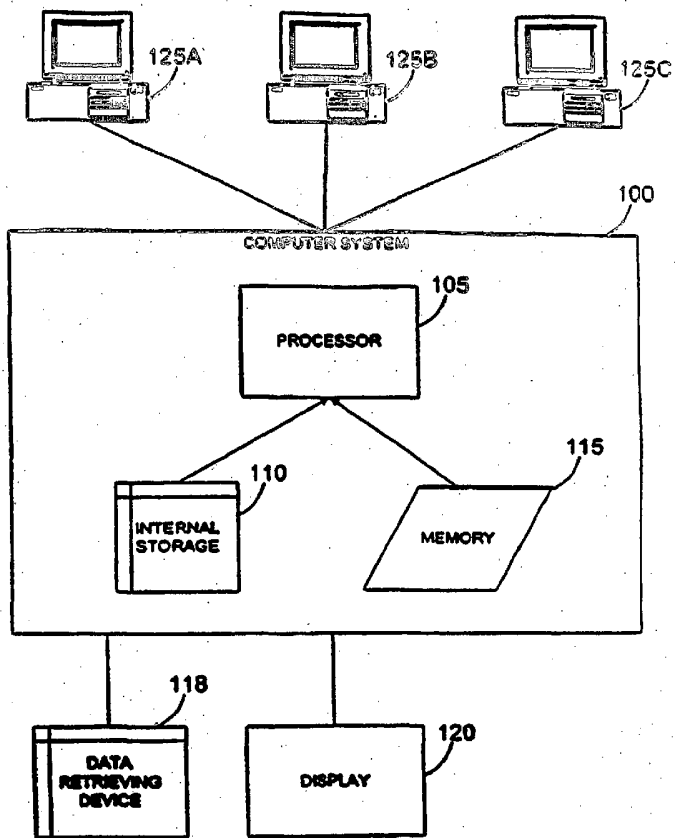


FIGURE 2

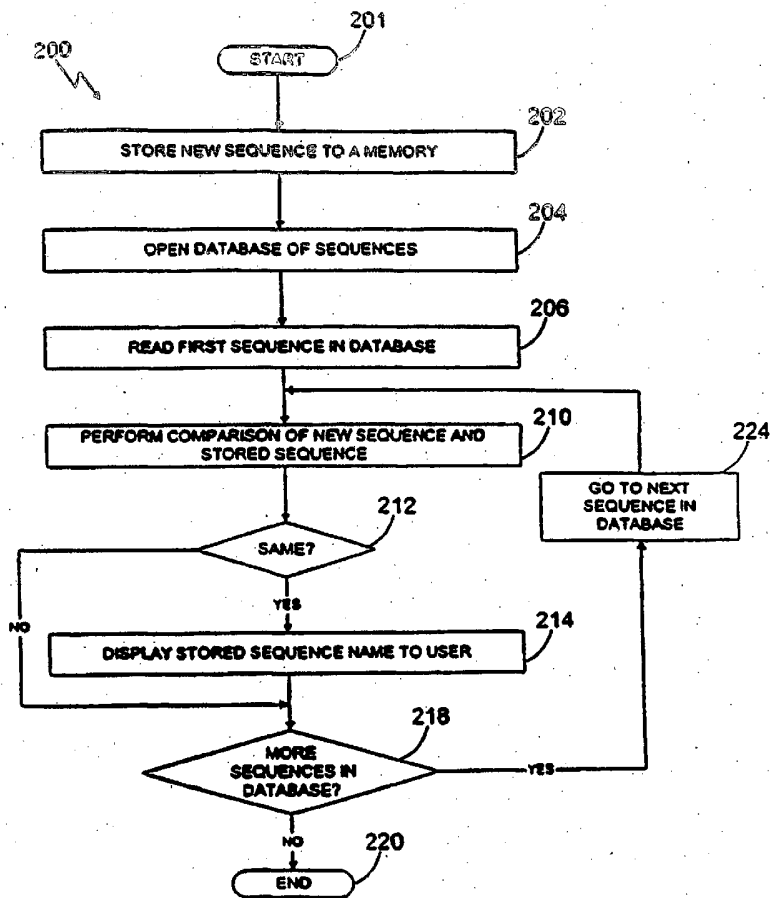


FIGURE 3

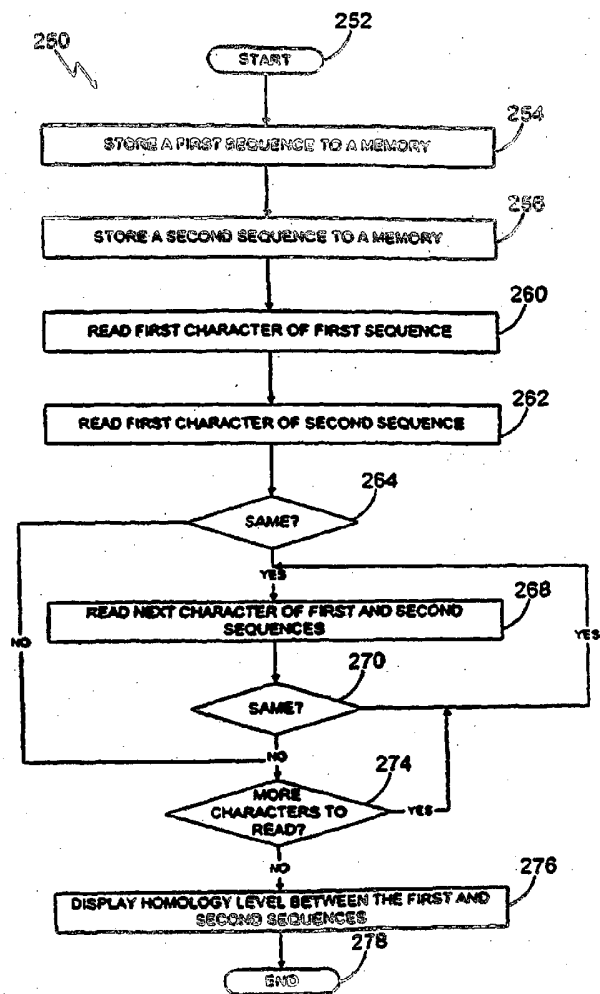


FIGURE 4

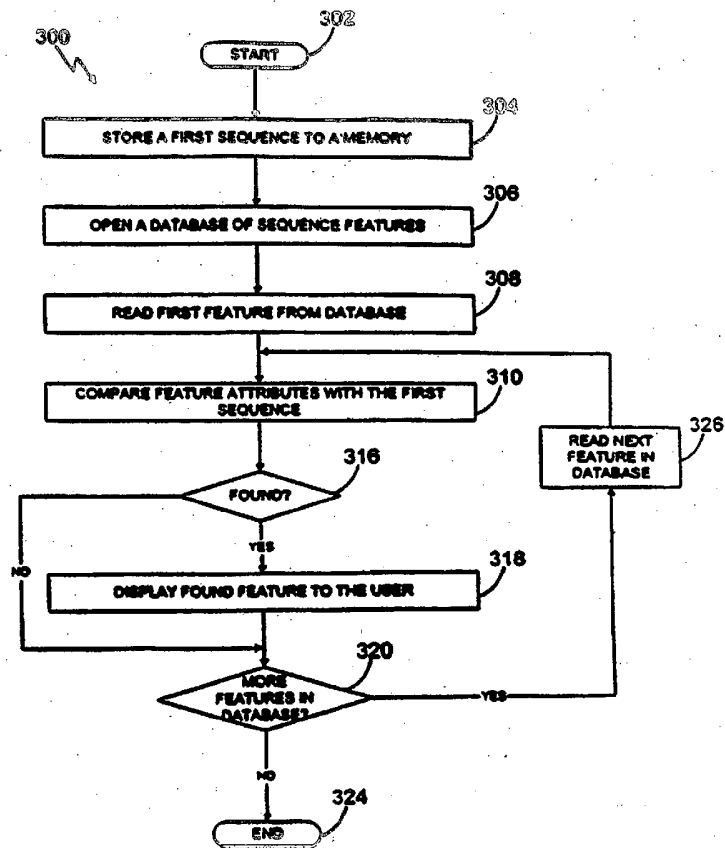


FIGURE 5A

SEQ ID NO. in provisional	Relative Substrate Specificity	Relative Substrate Specificity Value	Characterization Activity Temp	Characterization Activity pH	Enzyme	Characterization Description	Characterization Substrate
101, 102			room temp	9	Pectinase	pga assay	Polygalacturonic acid
103, 104					Pectate lyase		
105, 106	Cotton	4.8	40	9	Pectate lyase	Application Bioscoring	Cotton
107, 108	Cotton	1.5	40	9	Pectate lyase	Application Bioscoring	Cotton
109, 110			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
111, 112					Pectate lyase		
113, 114					Pectinase		
115, 116			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
117, 118	Cotton	3	40	9	Pectate lyase	Application Bioscoring	Cotton
119, 120			room temp	9	Pectate lyase		
121, 122	Cotton	7.8	50	9	Pectate lyase	Application Bioscoring	Cotton
123, 124	Cotton	10.3	40	9	Pectate lyase	Application Bioscoring	Cotton
125, 126	Cotton	3.6	40	9	Pectate lyase	Application Bioscoring	Cotton
127, 128		2.5	40	9	Pectate lyase	Application Bioscoring	Cotton
129, 130	Cotton	1.8	room temp	11	Pectate lyase	pga assay	Polygalacturonic acid
131, 132			room temp	9	Pectate lyase	Application Bioscoring	Cotton
133, 134			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
135, 136					Pectate lyase		
137, 138					Pectate lyase		
139, 140					Pectate lyase		
141, 142					Pectate lyase		
143, 144					Pectate lyase		
145, 146					Pectate lyase		
147, 148					Pectate lyase		
149, 150					Pectate lyase		
151, 152					Pectate lyase		
153, 154					Pectate lyase		
155, 156					Pectate lyase		
157, 158					Pectate lyase		
159, 160					Pectate lyase		
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309, 310					Pectate lyase		
311, 312					Pectate lyase		
313, 314					Pectate lyase		
315, 316					Pectate lyase		
317, 318					Pectate lyase		
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559, 560					Pectate lyase		</

FIGURE 5B

31, 32	Cotton	4.3	40	9	Pectate lyase	Application Bioscoring	Cotton
31, 32			room temp	8	Pectate lyase	pga assay	Polygalacturonic acid
33, 34			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
35, 36	Cotton	1.6	40	9	Pectate lyase	Application Bioscoring	Cotton
35, 36			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
37, 38	Cotton	11.2	50	9	Pectate lyase	Application Bioscoring	Cotton
39, 40			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
41, 42	Cotton	2	40	9	Pectate lyase	Application Bioscoring	Cotton
43, 44			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
45, 46			room temp	10	Pectate lyase	pga assay	Polygalacturonic acid
47, 48					Pectate lyase		
49, 50					Pectate lyase		
51, 52					Pectate lyase		
53, 54	Cotton	13.4	40	9	Pectate lyase	Application Bioscoring	Cotton
55, 56			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
57, 58			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
59, 60	Cotton	12.5	40	9	Pectate lyase	Application Bioscoring	Cotton
61, 62			room temp	10	Pectate lyase	pga assay	Polygalacturonic acid
63, 64					Pectate lyase		
65, 66					Pectate lyase		
67, 68	Cotton	9.8	40	9	Pectate lyase	Application Bioscoring	Cotton
69, 70			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
71, 72			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
73, 74	Cotton	0.8	40	9	Pectate lyase	Application Bioscoring	Cotton
75, 76			room temp	10	Pectate lyase	pga assay	Polygalacturonic acid
77, 78					Pectinase		
79, 80	Cotton	16.2	40	9	Pectate lyase	Application Bioscoring	Cotton
81, 82			room temp	9	Pectate lyase	pga assay	Polygalacturonic acid
83, 84	Cotton	3.2	40	9	Pectate lyase	Application Bioscoring	Cotton

FIGURE 5C

85, 86	Cotton	0.1	40	9	Pectate lyase	Application Bioscoring	Cotton
85, 86			room lamp	9	Pectate lyase	pja assay	Polygalacturonic acid
87, 88	Cotton	9.4	40	9	Pectate lyase	Application Bioscoring	Cotton
89, 90					Pectinase		
91, 92			room lamp	10	Pectate lyase	pja assay	Polygalacturonic acid
93, 94					Pectate lyase		
95, 96	Cotton	5.4	40	9	Pectate lyase	Application Bioscoring	Cotton
95, 96			room lamp	9	Pectate lyase	pja assay	Polygalacturonic acid
97, 98					Pectate lyase		
99, 100	Cotton	16	50	9	Pectate lyase	Application Bioscoring	Cotton
99, 100	Cotton	12.2	40	9	Pectate lyase	Application Bioscoring	Cotton

FIGURE 6

Pectate lyase GSSM™ upmutants														
Upmutant	Mutation													
	A118H	A182V	T190L	A197G	S208K	T219M	T223E	S255R	S263K	N275Y	Y309W	S312V		
A	x			x	x	x				x	x	x		
B	x		x		x	x				x	x	x		
C	x	x			x		x		x	x	x	x		
D	x	x	x		x			x	x	x	x	x		
E	x				x				x	x	x	x		
F	x	x			x		x	x	x	x	x	x		
G	x	x	x		x		x	x	x	x	x	x		
H	x	x	x	x	x		x		x	x	x	x		
I	x			x	x		x		x	x	x	x		
J	x			x	x				x	x	x	x		
K	x		x	x	x	x	x		x	x	x	x		
L	x	x	x		x			x	x	x	x	x		
M	x	x		x	x		x		x	x	x	x		
N	x		x	x	x				x	x	x	x		
O	x			x	x				x		x	x		
P	x	x		x	x	x		x		x	x	x		
Q	x		x		x				x		x	x		
R	x		x	x	x				x	x	x	x		
S	x		x	x	x	x	x	x	x	x	x	x		

FIGURE 7

Upmutant	Melting temperature (°C)	Specific Activity (SA)				
		at 30°C	SA at 40°C	SA at 50°C	SA at 60°C	SA at 70°C
AA	60.6	370.3	582.1	848.2	ND	ND
BB	58.3	366.8	542.4	648.6	ND	ND
CC	57	331.1	439.7	713.6	ND	ND
DD	56	468.6	595.8	714.6	ND	ND
EE	60	102.2	202.5	378.1	ND	ND
FF	58.5	585.5	744.5	955.8	ND	ND
GG	57.2	323.8	590.4	909.3	ND	ND
HH	59.7	267.6	425.5	706.3	ND	ND
II	57.8	357.1	527.3	875.1	ND	ND
JJ	57.5	372.2	537.6	834.9	ND	ND
KK	58.9	444.1	678.9	859.4	ND	ND
LL	58.4	375.2	557.6	1007.1	ND	ND
A	71	ND	ND	311.1	483.3	777.7
B	70.4	ND	ND	317.0	432.4	628.0
C	72.5	ND	ND	377.6	468.6	849.1
D	73.25	ND	ND	323.8	352.2	926.4
E	71	ND	ND	340.4	557.6	641.7
F	71.75	ND	ND	389.3	550.8	438.3
G	73	ND	ND	297.4	545.9	790.4
H	72.25	ND	ND	356.1	480.3	980.2
I	70.4	ND	ND	363.9	701.4	666.2
J	72.8	ND	ND	241.6	622.2	694.6
K	71.5	ND	ND	298.4	701.4	700.4
L	73	ND	ND	245.5	947.0	1003.7
M	73	ND	ND	304.2	547.8	309.1
N	73	ND	ND	685.8	1010.5	1284.5
O	71.3	ND	ND	268.0	517.5	330.7
P	72	ND	ND	398.2	542.9	689.7
Q	70.8	ND	ND	354.1	382.5	526.3
R	71.9	ND	ND	361.0	408.9	722.0
S	70.9	ND	ND	753.3	1186.6	579.1

FIGURE 8

Vmax													Kinetic	
1620													Time:	2:00
1	2	3	4	5	6	7	8	9	10	11	12	Interest:	0.11	
													Fluoresc:	11
A	21.001	49.326	NoFl	10.691	363.25	NoFl	-21.00	33.455	78.312	20.313	59.575	27.576		
B	397.30	13.539	0.759	39.177	NoFl	NoFl	-2.789	-2.740	5.672	-2.255	591.56	NoFl		
C	2.787	26.455	20.911	241.11	45.819	5.218	NoFl	75.481	71.450	134.55	NoFl	35.028	Auto: Once Calibrate: Once	
D	0.853	24.278	33.829	40.308	26.291	12.818	37.998	59.704	51.230	33.400	31.101	39.545	Lag Time: 0.00 End Time: 2.00 OD Mix: 0 CO Max: 0.5 Vmax Plz: 11/11	
E	19.889	3.173	11.375	NoFl	37.245	-19.36	5.922	NoFl	21.055	NoFl	4.384	NoFl		
F	10.759	13.045	NoFl	NoFl	8.385	25.478	7.558	4.158	715.891	1.097	0.252	-4.364		
G	0.478	4.149	-0.384	5.544	31.309	7.636	7.145	57.089	7.636	18.154	33.023	10.364	Plate Last Read: 3:50 PM 1/12/2002	
H	6.584	51.590	26.907	NoFl	1.185	3.019	48.200	0.000	9.872	39.536	356.78	9.818		

Wavelength Combination: Lmt
Data Mode: Absorbance