



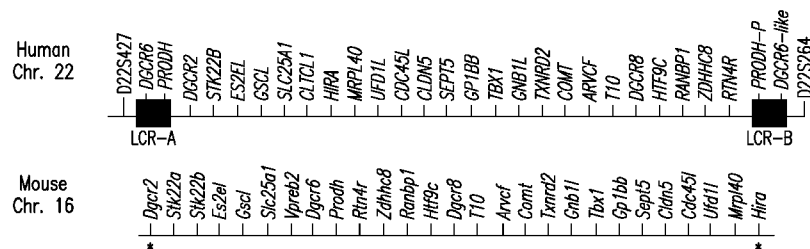
- (51) **International Patent Classification:**
A61K 48/00 (2006.01) *A61K 38/17* (2006.01)
- (21) **International Application Number:**
PCT/US2012/045093
- (22) **International Filing Date:**
29 June 2012 (29.06.2012)
- (25) **Filing Language:** English
- (26) **Publication Language:** English
- (30) **Priority Data:**
61/502,661 29 June 2011 (29.06.2011) US
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- (81) **Designated States (unless otherwise indicated, for every kind of national protection available):** AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.
- (84) **Designated States (unless otherwise indicated, for every kind of regional protection available):** ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

- without international search report and to be republished upon receipt of that report (Rule 48.2(g))

(54) **Title:** INHIBITOR OF NEURONAL CONNECTIVITY LINKED TO SCHIZOPHRENIA SUSCEPTIBILITY AND COGNITIVE DYSFUNCTION

**FIG. 1A**

(57) **Abstract:** The present invention relates to methods and compositions for enhancing neuronal connectivity. It is based, at least in part, on the discovery of a protein, termed "Mirta22," that inhibits the formation of structures which create connections between neurons. It is further based, in part, on the discovery that inhibiting Mirta22 activity by short hairpin RNA was able to restore these structures. Mirta22 was discovered in experiments relating to 22q 11 microdeletions, which have been linked to schizophrenia. Accordingly, the present invention provides for methods of treating schizophrenia comprising administering an agent that inhibits Mirta22 activity. It may also be used in the treatment of other disorders that would benefit from enhanced neural connectivity, including learning and memory disorders.

INHIBITOR OF NEURONAL CONNECTIVITY LINKED TO SCHIZOPHRENIA SUSCEPTIBILITY AND COGNITIVE DYSFUNCTION

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GRANT INFORMATION

This invention was made with government support under Grant Nos. MH067068 and MH077235 awarded by the National Institute of Mental Health. The government has certain rights in the invention.

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CROSS REFERENCE TO RELATED APPLICATIONS

This application claims priority to U.S. Provisional Application Serial No. 61/502,661, filed June 29, 2011, the contents of which are incorporated by reference in its entirety herein.

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1. INTRODUCTION

The present invention relates to methods and compositions for enhancing neuronal connectivity.

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2. BACKGROUND OF THE INVENTION

Association between recurrent de novo 22q11.2 microdeletions and schizophrenia represented a shift in our understanding of the genetic architecture of schizophrenia (SCZ), highlighting the role that rare and highly penetrant mutations play in the disease risk (1,2). This view has been strengthened recently by the
25 identification of a widespread role of chromosomal microdeletions and microduplications (copy-number variants or CNVs) in determining susceptibility to schizophrenia and other psychiatric disorders (3-5). Individuals with 22q11.2 microdeletions have specific behavioral impairments and exhibit a spectrum of deficits in cognitive abilities linked to activity in the hippocampus and prefrontal
30 cortex, such as measures of attention, working memory and executive function. Up to one third of children with the microdeletion develop schizophrenia or schizoaffective disorder in adolescence or early adulthood accounting for 1–2% of sporadic schizophrenia cases (2). Understanding how the genes disrupted by this microdeletion contribute to the emergence of the psychiatric and cognitive phenotypes
35 associated with this genomic imbalance would provide important clues for the

pathogenesis of SCZ and can guide future analysis of other CNVs that cause psychiatric disorders (6,7).

3. SUMMARY OF THE INVENTION

5 The present invention relates to methods and compositions for enhancing neuronal connectivity. It is based, at least in part, on the discovery of a protein, termed "Mirta22," that inhibits the formation of structures which create connections between neurons. It is further based, in part, on the discovery that inhibiting Mirta22 activity by short hairpin RNA was able to restore these structures.

10 Mirta22 was discovered in experiments relating to 22q11 microdeletions, which have been linked to schizophrenia. Accordingly, the present invention provides for methods of treating schizophrenia comprising administering an agent that inhibits Mirta22 activity. It may also be used in the treatment of other disorders that would benefit from enhanced neural connectivity, including learning and memory disorders.

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4. BRIEF DESCRIPTION OF THE FIGURES

FIGURE 1A-D. Generation of *Df(16)A^{+/-}* mice. (A) Schematic showing the human chromosome 22q11.2 region and the syntenic mouse region. Asterisks note *Dgcr2* and *Hira*, the two endpoints of the targeted deletion. (B) Wild-type chromosomal loci for *Dgcr2* and *Hira*, and their corresponding targeted loci after the introduction of a 5'HPRT and a 3'HPRT mini-cassette, respectively. (C) Cre-induced recombination between loxP sites in *cis* leads to a functional *HPRT* minigene and a 1.3 Mb deletion in the mice. After the loxP sites recombine to produce the deletion, an 829-bp PCR product can be detected. (D) FISH analysis of

20 Het deletion mice. Metaphase preps from splenocyte cultures demonstrate a Het deletion

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FIGURE 2A-E. Production, validation and analysis of miRNA biogenesis in *Dgcr8*-deficient mice. (A) Diagram showing the genomic structure of the *Dgcr8* gene and the gene trap insert located in the intron between exons 8 and 9 (diagram not drawn to scale). En2 intron 1:1.5 kb of mouse En2 Intron 1;SA: splice acceptor of mouse En2 exon 2; β -geo: fusion of β -galactosidase and neomycin transferase; pA: SV40 polyadenylation signal. Red arrowheads indicate the approximate genomic location of the PCR primers used for genotyping. (B) PCR

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genotyping designed to verify the gene trap insertion in HET *Dgcr8*^{+/-} mice. (C) Northern blot analysis indicating the reduction in levels of the WT 4.5-kb *Dgcr8* mRNA and the production of the expected 7.2-kb chimeric mRNA in *Dgcr8*^{+/-} mice (arrowhead). (D) qRT-PCR analysis confirming reduction in the levels of WT *Dgcr8* mRNA. (E) Western blot analysis indicating a reduction of the 120-kD WT *Dgcr8* protein and the production of the expected ~220 kD chimeric protein (arrow).

FIGURE 3A-J. Drastic reduction in mir-185 levels affects neuronal connectivity. (A) Expression of mir-185 mRNA in HPC and cortex as shown by *in situ* hybridization in coronal brain sections using an antisense mir-185 probe. An antisense U6 probe and a scramble probe were used as positive and negative controls, respectively. Images were taken at either × 4 (left panels) or × 10 (right panels) magnification. (B-D) mir-185 expression levels in HPC (B) or PFC (C) of *Df(16)A*^{+/-} (n = 7 for mutant, n = 9 for WT littermates) and in HPC (D) of *Dgcr8*^{+/-} (n = 10 for mutant and WT littermates), as assayed by qRT-PCR. Expression levels in mutant animals were normalized to their respective WT littermates. (E) Representative *Df(16)A*^{+/-} neurons at DIV9 transfected with pre-scramble or pre-mir-185 mimic and enhanced GFP for visualization. Scale Bar, 20 μm. (F,G) Reduction in the number of primary dendrites (F) and branch points (G) in *Df(16)A*^{+/-} neurons at DIV9 relative to WT neurons is reversed by the transfection of pre-mir-185, but not pre-scramble mimic (pre-scr) (n = 21 for WT + pre-scr; n = 21 for *Df(16)A*^{+/-} + pre-scr; n = 21 for *Df(16)A*^{+/-} + pre-mir-185). Values of *Df(16)A*^{+/-} neurons were normalized to WT+pre-scr. (H) Representative images of spines on *Df(16)A*^{+/-} neurons at DIV19, transfected with pre-scramble or pre-mir-185 mimic as well as enhanced GFP. Scale Bar, 5 μm. (I) Reduction in the density of mushroom spines (estimated over 75 μm of dendritic length) in *DIV19 Df(16)A*^{+/-} neurons relative to WT control neurons is reversed by the transfection of pre-mir-185, but not pre-scramble mimic, into *Df(16)A*^{+/-} neurons. (n = 23 for WT + pre-scr; n = 21 for *Df(16)A*^{+/-} + pre-scr; n = 23 for *Df(16)A*^{+/-} + pre-mir-185). Values of *Df(16)A*^{+/-} neurons were normalized to WT+pre-scr. (J) Transfection of pre-mir-185 mimic, but not pre-scramble control, significantly increased the width of mushroom spines of *Df(16)A*^{+/-} neurons at DIV19 (P < 0.001, Kolmogorov-Smirnov test) (n = 568 for WT + pre-scr; n = 339 for *Df(16)A*^{+/-} + pre-scr; n = 527 for *Df(16)A*^{+/-} + pre-mir-185). For all data, ***P < 0.001, **P < 0.01, *P < 0.05 (Student's t-test); Error bars indicate s.e.m.

FIGURE 4A-E. 2310044H10Rik (Mirta22) is robustly upregulated in the brain of *Df(16)A^{+/-}* mice. (A) Changes in gene expression in the PFC (upper panel) or HPC (lower panel) of *Df(16)A^{+/-}* and WT littermate control mice at E16, P6 and adulthood (n = 10 each group): Volcano plot of the P-values and the corresponding relative expression of each gene. Light blue dots, genes within *Df(16)A* deficiency; light green dots, upregulated miRNA-containing transcripts; red dots (identified by an arrow), indicate probe sets representing Mirta22. (B) Top 10 protein encoding genes that show significant upregulation in the PFC (upper panel) or HPC (lower panel) of *Df(16)A^{+/-}* and WT littermate mice at E16, P6 and adulthood. Mirta22 is highlighted in red (and identified with an “•”). (C,D) Temporal expression of 2310044H10Rik (Mirta22) in the PFC (c) and HPC (d) of *Df(16)A^{+/-}* and WT littermate mice as monitored by qRT-PCR. n = 9–10 for each group. (E) Increased expression of endogenous 2310044H10Rik (Mirta22) in DIV9 hippocampal neurons isolated from *Df(16)A^{+/-}* animals as assayed by qRT-PCR (n = 3 each genotype). Expression levels in mutant neurons were normalized to WT neurons. ***P < 0.001, **P < 0.01, *P < 0.05 (Student's t-test); Error bars indicate s.e.m.

FIGURE 5A-H. miR-185 directly targets and represses 2310044H10Rik (Mirta22). (A) Structure of the 3'UTR of 2310044H10Rik (Mirta22) showing miRNA binding sites predicted by TargetScan or mirDB. Blocks in mouse 2310044H10Rik (Mirta22) 3'UTR that are highly conserved in rat and human orthologues are shown below the mouse 3'UTR. Evolutionary conservation is also assessed by the “30-way multiz alignment and conservation analysis” in the USCS browser, with conserved blocks indicated by green peaks. *mir-185* and *mir-485* binding sites located within the conserved blocks are shown in red (and identified with an “•”). (B-C) qRT-PCR quantification of endogenous 2310044H10Rik (Mirta22) in DIV7 hippocampal neurons. Expression levels in *anti-mir-185*-treated and *pre-mir-185*-treated neurons were normalized to expression levels under respective controls. (B) Increased expression levels of *Mirta22* in neurons transfected with *anti-miR-185* at DIV5 (n = 5, each treatment). (C) Reduced expression levels of *Mirta22* in DIV9 hippocampal neurons transfected with *pre-miR-185* mimic at DIV7 (n = 3, each treatment). (D-E) qRT-PCR quantification of endogenous 2310044H10Rik (Mirta22) in N18 cells. Expression levels in *pre-mir-185*-treated and *anti-mir-185*-treated cells were normalized to expression levels under respective controls. (D) Reduced

expression levels of *Mirta22* in cells transfected with *pre-mir-185* mimic (n = 3, each treatment). (E) Up-regulation of *Mirta22* in cells transfected with an *anti-mir-185* LNA oligo (n = 3, each treatment). (F-H) Repression effects of *pre-mir-185*, *pre-mir-485* and *pre-mir-491* on *Mirta22* 3'UTR were examined by a dual-luciferase reporter assay. psiCHECK2 plasmids containing a *Renilla* luciferase gene under the control of either Wt or one of the mutant *2310044H10Rik (Mirta22)* 3'UTRs were co-transfected with pre-miRNA or pre-scramble mimic into the N18 neuroblastoma cell line (n = 3 for each condition). Firefly luciferase expressed from the same plasmid was used as internal control. Values are *Renilla* luciferase levels relative to firefly luciferase levels and normalized to the relative expression levels under pre-scramble treatment (F, H) or to the relative expression levels from plasmid with no 3'UTR (G). *Pre-mir-185* significantly decreases the *2310044H10Rik (Mirta22)* 3'UTR reporter expression over a concentration range of 10nM to 0.01nM (F). *pre-mir-185* mediated repression on *2310044H10Rik (Mirta22)* 3'UTR reporter expression depends on conserved miRNA binding sites (G). *2310044H10Rik (Mirta22)* 3'UTR luciferase reporters with mutations at Site 1 (Mut1) or Site 2 (Mut2) or both sites (Mut1&2) were analyzed. Mutated *2310044H10Rik (Mirta22)* 3'UTR reporters express significantly higher luciferase activities than Wt *Mirta22* 3'UTR reporters. *Pre-mir-485* and *pre-mir-491* significantly decreases the *2310044H10Rik (Mirta22)* 3'UTR reporter expression (H). For all data, ***P < 0.001, **P < 0.01, *P < 0.05 (Student's t-test); Error bars indicate s.e.m.

FIGURE 6A-D. Genomic structure, neuronal expression and subcellular localization of *2310044H10Rik (Mirta22)*. (A) *Top*: Structure of mRNA transcripts of *2310044H10Rik (Mirta22)* and its human orthologue, *C19orf63*. RefSeq reports a *2310044H10Rik (Mirta22)* transcript with 7 exons (blue rectangles), which is predicted to encode a signal peptide and a transmembrane domain (red rectangles). The peptide epitope used to generate a polyclonal antibody is marked by green rectangle. For *C19orf63*, RefSeq reports 2 alternatively spliced transcripts: one that encodes a predicted transmembrane protein and one with an additional exon that encodes a predicted secreted protein. *Bottom*: Protein sequence alignment of predicted transmembrane isoforms encoded by *2310044H10Rik (Mirta22)* and its human orthologue. Black blocks indicate completely conserved residues; grey blocks indicate similar residues (defined by Boxshade default similarities); white blocks indicate different residues. (B) *Upper*: Representative western blot assays of *2310044H10Rik*

(Mirta22) in PFC lysates prepared from *Df(16)A^{+/-}* animals and Wt littermates.

Alpha-tubulin is used as loading control. *Lower*: Quantification of 2310044H10Rik (Mirta22) protein level in PFC of Wt and *Df(16)A^{+/-}* animals (n = 9 each genotype).

Expression levels in mutant mice were normalized to Wt littermates. Results are

- 5 expressed as mean $\pm$ SEM. **p < 0.01 (Student's *t*-test). (C) Quantification of 2310044H10Rik (Mirta22) immunocytochemical signals in Wt and *Df(16)A^{+/-}* cultured neurons (n = 34 for Wt; n = 31 for *Df(16)A^{+/-}*). Expression levels in mutant neurons were normalized to Wt neurons. Results are expressed as mean $\pm$ SEM. *p < 0.05 (Student's *t*-test). (D) *Upper panel*: 2310044H10Rik (Mirta22) co-localizes with
- 10 neuron specific marker NeuN, but not with glia specific marker GFAP, in cultured hippocampal neurons at DIV20. *Lower panel*: 2310044H10Rik (Mirta22) (green) co-localizes with Golgi specific marker GM130 (red) in the soma. 2310044H10Rik (Mirta22) is also found in vesicles and tubular-like clusters in the dendrites, which are highlighted by the dendritic marker MAP2 (blue).

- 15 **FIGURE 7A-L. Elevated 2310044H10Rik (Mirta22) Levels Contribute to Structural Alterations of *Df(16)A^{+/-}* Neurons.** (A) qRT-PCR results showing that the *Mirta22* mRNA was significantly increased in DIV12 hippocampal cultured neurons transfected with a full length *Mirta22* cDNA plasmid. (B) Right: Representative western blot showing the reduction of the endogenous Mirta22 protein
- 20 in DIV12 hippocampal neurons transfected with *Mirta22* cDNA (lane 2) or empty vector (lane 1). Alpha-tubulin is the loading control. Left: Quantification of western blots showing a 58% increase (p < 10⁻⁴; n = 4 each condition) in Mirta22 signal in *Mirta22* cDNA-transfected cells, compared to empty vector-transfected cells. (C-D) Transfection of a 2310044H10Rik (*Mirta22*) expression construct into Wt neurons at
- 25 DIV7 results in decrease in the number of primary dendrites (C) and branch points (D) at DIV9 (n = 24 for empty vector transfected and n = 27 for *Mirta22* transfected cells). In (D), values of *Mirta22* overexpressing neurons were normalized to empty vector-transfected neurons. (E-F) Introduction of 2310044H10Rik (*Mirta22*) into Wt neurons at DIV17 results in decrease in the density of mushroom spines (E) and the
- 30 width of those spines (F) (P < 0.01, Kolmogorov-Smirnov test) at DIV19 [n = 16 for vector transfected and n = 17 for *Mirta22* transfected neurons (E); n = 278 for spines on vector transfected neurons and n = 231 for spines on *Mirta22* transfected neurons (F)]. Values of *Mirta22* overexpressing neurons were normalized to empty vector-transfected neurons.

(G) qRT-PCR results showing that the endogenous *Mirta22* mRNA was significantly reduced in DIV12 hippocampal cultured neurons transfected with *Mirta22* shRNA. (H) Right: Representative western blot showing the reduction of the endogenous *Mirta22* protein in DIV12 hippocampal neurons transfected with *Mirta22* shRNA (lane 2) or scramble shRNA (lane 1). Alpha-tubulin is the loading control. Left: Quantification of western blots showing a 48% decrease ($p < 0.01$; $n = 4$ each condition) in *Mirta22* levels in *Mirta22* shRNA-transfected cells, compared to scramble shRNA-transfected cells. (I-J) Reduction in the number of primary dendrites (I), but not reduction in the number branch points (J), in *Df(16)A^{+/-}* neurons at DIV9 relative to Wt neurons is reversed by the transfection of a construct that expresses *2310044H10Rik* (*Mirta22*) shRNA^{+/+} ($n = 24$ for Wt + scr shRNA; $n = 21$ for *Df(16)A^{+/-}* + scr shRNA; $n = 25$ for *Df(16)A^{+/-}* + *Mirta22* shRNA). Scr shRNA: scramble shRNA. N.S.: not significant. In (J), values of *Df(16)A^{+/-}* neurons were normalized to Wt + scr shRNA. (K) Reduction in the density of mushroom spines (estimated over 75 μ m of dendritic length) in *Df(16)A^{+/-}* neurons at DIV19 relative to Wt neurons is reversed by the introduction of *Mirta22* shRNA, but not scramble shRNA^{+/+} ($n = 22$ for Wt + scr shRNA; $n = 24$ for *Df(16)A^{+/-}* + scr shRNA; $n = 15$ for *Df(16)A^{+/-}* + *Mirta22* shRNA). Values of *Df(16)A^{+/-}* neurons were normalized to Wt + scr shRNA. (L) Transfection of *Mirta22* shRNA does not affect the width of mushroom spines of *Df(16)A^{+/-}* neurons at DIV19 ($p > 0.05$, Kolmogorov-Smirnov test). $n = 342$ for Wt + pre-scr; $n = 289$ for *Df(16)A^{+/-}* + pre-scr, $n = 177$ for *Df(16)A^{+/-}* + *pre-miR-185*. (A-E, G-K) Results are expressed as mean $\pm$ SEM. * $p < 0.05$, ** $p < 0.01$ (Student's *t*-test).

FIGURE 8A-B. *Dgcr8* levels in *Df(16)A^{+/-}* mice during brain development. (A,B) *Dgcr8* expression levels in HPC of E17 (a) and P6 (b) *Df(16)A^{+/-}* mice ($n = 10$) and their respective WT littermate controls ($n = 10$), as assayed by qRT-PCR. Expression levels in mutant animals were normalized to their respective WT littermates. *** $P < 0.001$, ** $P < 0.01$ (Student's *t*-test); Error bars indicate s.e.m.

FIGURE 9A-B. *mir-185* levels in *Df(16)A^{+/-}* mice during brain development. (A,B) *mir-185* expression levels in HPC and PFC of E17 (a) and P6 (b) *Df(16)A^{+/-}* mice ($n = 10$ for HPC or PFC) and their respective WT littermate control mice ($n = 10$ for HPC or PFC), as assayed by qRT-PCR. Expression levels in mutant animals were normalized to their respective WT littermates. *** $P < 0.001$, ** $P < 0.01$ (Student's *t*-test); Error bars indicate s.e.m.

FIGURE 10. 2310044H10Rik (Mirta22) expression in the brain of *Dgcr8*^{+/-} mice. Expression levels in HPC and PFC of adult *Dgcr8*^{+/-} mice (n = 7 for HPC; n = 10 for PFC) and their respective WT littermate controls (n = 8 for HPC; n = 10 for PFC), as assayed by qRT-PCR. Expression levels in mutant animals were
 5 normalized to their respective WT littermates. *P < 0.05 (Student's t-test); Error bars indicate s.e.m.

FIGURE 11A-B. mir-485 and mir-491 levels are reduced in *Df(16)A*^{+/-} mice. (A,B) Expression levels of mir-485 (a) and mir-491 (b) in HPC of adult *Df(16)A*^{+/-} mice (n = 7) and their respective WT littermate control mice (n = 9), as
 10 assayed by qRT-PCR. Expression levels in mutant animals were normalized to their respective WT littermates. **P < 0.01, *P < 0.05 (Student's t-test); Error bars indicate s.e.m.

FIGURE 12. Expression levels of a sample of putative *miR-185* targets predicted by both TargetScan and miRanda (*Nptx2*, *Coro2b*, *Epb4.1ll*, *Lpcat3*,
 15 *Gsk3b*, *Slc6a1*, *Grin2b* and *Vipr2*) in the HPC of adult *Df(16)A*^{+/-} mice (n = 10) and their respective Wt littermate mice (n = 10), as assayed by qRT-PCR. Expression levels in mutant animals were normalized to their respective Wt littermates. For all genes tested, expression levels were not significantly altered in mutant animals. Expression levels in mutant animals were normalized to their respective Wt
 20 littermates. Results are expressed as mean ± SEM. *p < 0.05, **p < 0.01, ***p < 0.001 (Student's t-test).

FIGURE 13A-G. Specificity of 2310044H10Rik (Mirta22) Antibody. (A) Anti-2310044H10Rik (anti-Mirta22) antibody, as well as polyclonal and
 25 monoclonal anti-FLAG antibodies, recognize a 28 kD band in western blots on lysates of 293T cells transfected with a plasmid expressing full length 2310044H10Rik (Mirta22) cDNA with a C-terminal FLAG tag ("Flag tagged" lanes). Lysates of cells transfected with empty vector were used as control ("control" lanes). Note that there is a 28 kD band recognized by anti-2310044H10Rik antibody, but not by anti-FLAG antibodies, in control lysates. This likely represents the endogenous human
 30 2310044H10Rik (Mirta22) orthologous protein (C19orf63). (B) *Upper panel:* Representative western blot showing the reduction of the endogenous 2310044H10Rik (Mirta22) protein in N18 cells transfected with 2310044H10Rik (*Mirta22*) shRNA (lane 2) compared to scramble shRNA-transfected cells (lane 1). Alpha-tubulin is the loading control. *Lower panel:* Quantification of western blots

showing a 32% reduction ($p < 0.01$; $n = 6$ each condition) in 2310044H10Rik (Mirta22) signal in 2310044H10Rik (Mirta22) shRNA-transfected cells, compared to scramble shRNA-transfected cells. Expression levels in 2310044H10Rik (Mirta22) shRNA-manipulated neurons were normalized to their respective controls. (C) *Upper panel*: Representative western blot showing the expected increase in 2310044H10Rik (Mirta22) protein levels in N18 cells transfected with 2310044H10Rik (Mirta22) cDNA (lane 2) compared to empty vector-transfected cells (lane 1). Alpha-tubulin is the loading control. *Lower panel*: Quantification of western blots showing a 57% increase ($p < 0.05$; $n = 6$ each condition) in 2310044H10Rik (Mirta22) signal in 2310044H10Rik (Mirta22) cDNA-transfected cells, compared to empty vector-transfected cells. Expression levels in 2310044H10Rik (Mirta22) cDNA-manipulated neurons were normalized to their respective controls. (D) Anti-2310044H10Rik (anti-Mirta22) antiserum, but not pre-immune serum, recognizes endogenous Mirta22 protein (green signal) in DIV20 Wt hippocampal neurons, as assayed by immunocytochemistry. (E) Representative immunocytochemistry images showing that 2310044H10Rik (Mirta22) signal (green) as compared to Golgi marker GM130 (blue), is largely reduced in 2310044H10Rik (Mirta22) shRNA-transfected (RFP+) DIV14 Wt hippocampal neurons (lower panel) but not in scramble shRNA-transfected (RFP+) neurons (upper panel). Note that in 2310044H10Rik (Mirta22) shRNA-treated culture (lower panel), 2310044H10Rik (Mirta22) signal (green) in un-transfected neurons (marked by white arrows) is not reduced, as compared to scramble shRNA-transfected (RFP+) neurons (shown in upper panel). Transfection was performed at DIV12 and neurons were fixed and immunostained 2 days later. (F) Quantification of Mirta 22 immunocytochemical signal shown in (E). Note that in 2310044H10Rik (Mirta22) shRNA-treated neuronal cultures, there is a 64% decrease ($p < 0.001$) in 2310044H10Rik (Mirta22) signal in transfected neurons (RFP+, $n = 20$), compared to un-transfected neurons in (RFP-, $n = 20$). In scramble shRNA-treated cultures, there is no difference in 2310044H10Rik (Mirta22) signal between transfected (RFP+, $n = 10$) and un-transfected (RFP-, $n = 10$) neurons. Mirta22 signal measurements in transfected (RFP+) and un-transfected (RFP-) neurons in scramble shRNA treated culture and 2310044H10Rik (Mirta22) shRNA-transfected neurons (RFP+) were normalized to un-transfected neurons (RFP-) in 2310044H10Rik (Mirta22) shRNA treated culture. (G) Anti-Mirta22 antibody serum, but not pre-immune serum, recognizes endogenous Mirta22 protein in western blot assays of hippocampal protein

extracts. Representative western blot images from two animals are shown in each case. Results are expressed as mean $\pm$ SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ (Student's *t*-test).

FIGURE 14A-C. 2310044H10Rik (Mirta22) distribution in the adult brain. (A-C) Mirta22 distribution as assayed by immunohistochemistry on brain slices using anti-Mirta22 antibody. Images were taken at 4X (A), 10X (B) and 20X (C) magnification. Red boxes in 4X and 10X images outline the area shown in 10X and 20X images, respectively. Note the widespread distribution of Mirta22 immunoreactivity in the brain, including in the cortex and hippocampus. Cortical neurons with processes can be seen clearly in the 20X image (C).

FIGURE 15A-B. Bidirectional manipulation of 2310044H10Rik (Mirta22) levels. (A) Mirta22 expression levels in N18 cells, a mouse neuroblastoma cell line, transfected with plasmids expressing Mirta22 shRNA or scramble shRNA ($n = 3$ for each treatment), as assayed by qRT-PCR. Expression levels in Mirta22 shRNA-treated cells were normalized to scramble shRNA-treated cells. (B) Mirta22 expression levels in N18 cells transfected with plasmids expressing Mirta22 cDNA or empty vector ($n = 3$ for each treatment), as assayed by qRT-PCR. Expression levels in Mirta22 cDNA-treated cells were normalized to vector-treated cells. * $P < 0.05$ (Student's *t*-test); Error bars indicate s.e.m.

FIGURE 16A-C. Mirta-encoding cDNA. (A) Human Isoform 1, Genbank Accession No. NM_175063 (SEQ ID NO:3); (B) Human Isoform 2, Genbank Accession No. NM_175063 (SEQ ID NO:5); (C) Murine mirta-encoding cDNA, GenBank Accession No. NM_197991 (SEQ ID NO:19).

FIGURE 17A-F. (A) Representative images of Wt neurons at DIV9 transfected with anti-*miR* control or *anti-miR-185* oligos and enhanced GFP to facilitate visualization of dendrites and spines. (B-C) Reduction in the number of primary dendrites (B) and branch points (C) in Wt neurons at DIV9, 2 days after transfected with *anti-miR-185* relative to Wt neurons transfected with anti-*miR* control ($n = 21$ for Wt + *anti-miR-185*; $n = 20$ for Wt + anti-*miR* control). In (C), values of Wt + *anti-miR-185* were normalized to Wt + anti-*miR* control. (D) Representative images of spines on Wt neurons at DIV19, transfected with anti-*miR* control or *anti-miR-185* as well as enhanced GFP. (E) Reduction in the density of mushroom spines (quantified over 75 μm of dendritic length) in neurons transfected with *anti-miR-185* relative to neurons transfected with anti-*miR* control ($n = 20$ for Wt

+ *anti-miR-185*; n = 20 for Wt + *anti-miR* control). Values of Wt + *anti-miR-185* were normalized to Wt + *anti-miR* control. (F) Transfection of *anti-miR-185* oligos significantly decreased the width of mushroom spines compared to that of the neurons transfected with *anti-miR* control at DIV19 (15%, $P < 0.001$, Kolmogorov-Smirnov test) (n = 232 for Wt + *anti-miR-185*; n = 293 for Wt + *anti-miR* control).
 5 (B, C, E) Results are expressed as mean $\pm$ SEM. * $p < 0.05$, ** $p < 0.01$ (Student's *t*-test).

FIGURE 18A-D. miR-185 Levels Affect Dendritic and Spine Development. (A) Sholl analysis of dendritic complexity using 10 μ m concentric
 10 circles around the soma. Wt neurons transfected with either *anti-miR-185* (n = 21) or *anti-miR* control (n = 20) at DIV7 and fixed at DIV9. Increase in branching is prevalent in the vicinity of the most proximal dendrites. (B) Distribution of spine morphotypes (other than mushroom spines, quantified over 75 μ m of dendritic length from soma), as well as total protrusions, in cultured Wt hippocampal neurons
 15 transfected with *anti-mir-185* relative to neurons transfected with *anti-mir* control (n = 20 for Wt + *anti-mir* control; n = 20 for Wt + *anti-mir-185*). (C) Increase in the number of primary dendrites and branch points in DIV9 (2 days following transfection) Wt neurons transfected with *pre-mir-185* relative to Wt neurons transfected with pre-scramble mimic (pre-scr) (n = 21 for Wt + *pre-mir-185*; n = 20
 20 for Wt + pre-scr). For the number of branch point, values of Wt + *pre-mir-185* were normalized to Wt + pre-scr. (D) Left: Increase in the density of mushroom spines (quantified over 75 μ m of dendritic length) in DIV19 Wt neurons transfected with *pre-mir-185* relative to Wt neurons transfected with pre-scr (n = 23 for Wt + pre-scr; n = 23 for Wt + *pre-mir-185*). Values of Wt neurons with *pre-mir-185* were
 25 normalized to Wt with pre-scr. Right: Transfection of *pre-mir-185* mimic, but not pre-scramble control, significantly increased the width of mushroom spines on Wt neurons at DIV19 (18%, $P < 0.001$, Kolmogorov-Smirnov test) (n = 568 for Wt + pre-scr; n = 527 for Wt + *pre-mir-185*). (E) Sholl analysis of dendritic complexity using 10 μ m concentric circles around the soma. *Dff(16)A^{+/-}* neurons transfected with either
 30 *pre-miR-185* mimic (n = 21) or pre-scramble mimic (n = 21) at DIV7 and fixed at DIV9. Reduction in branching in *Dff(16)A^{+/-}* neurons is reversed by *pre-miR-185* mimic throughout the dendritic tree, although it is more pronounced in the vicinity of the most proximal dendrites. (F) Distribution of spine morphotypes (other than mushroom spines, quantified over 75 μ m of dendritic length from soma), as well as

total protrusions, in cultured DIV19 *Df(16)A^{+/-}* hippocampal neurons transfected *pre-mir-185* or pre-scramble mimic relative to Wt neurons transfected with pre-scramble mimic (n = 23 for Wt + pre-scr; n = 21 for *Df(16)A^{+/-}* + pre-scr; n = 23 for *Df(16)A^{+/-}* + *pre-mir-185*).

- 5 Results are expressed as mean $\pm$ SEM. *p < 0.05, **p < 0.01, ***p < 0.001 (Student's *t*-test).

FIGURE 19A-D. Generation of *Df(16)B* and *Dp(16)B*. (A) *Dgcr14* and *Hira* loci and the corresponding targeted loci following the introduction of 5' *HPRT* and 3' *HPRT* mini-cassettes, respectively. (B) Upon exposure to Cre recombina-
 10 se, recombination between the Lox P sites of modified loci positioned in the trans orientation, led to the generation of a deletion between *Dgcr14* and *Hira*, as well as a duplication of the region. (C) Southern blot of 2 ES cell clones, using probes A and B (positions of probes, as well as expected sizes upon digestion with KpnI, indicated in panel B. E5 and E8 possess both the deletion (indicated by a 22.9
 15 kb band) as well as the duplication (indicated by a 16.6 kb band). Wt indicates lane with DNA from wild type ES cells. M indicates the marker lane, BstEII cut λ DNA. (D) FISH verification. Top panel: metaphase chromosome spread of MEFs possessing the duplication. A slightly brighter red signal (arrow) indicates the duplicated region. Middle panel: Interphase FISH of the duplication. The duplication is demonstrated by
 20 a third separate red signal. Bottom panel: Metaphase chromosome spread of MEFs possessing the deletion. The red signal indicates the deleted region, and the green signal is from the control probe, located outside of the deleted region. MEF: mouse embryonic fibroblast. FISH: fluorescent *in situ* hybridization. RP23-420H6: mouse BAC probe located within the deleted and duplicated regions. RP23-290E4: mouse
 25 BAC probe located outside of the deleted and duplicated regions.

FIGURE 20. List of transcripts outside the 22q11.2 syntenic region significantly misregulated in a reciprocal manner in both PFC and HPC.

FIGURE 21. Three-factor ANOVA of the impact of mir-185, mir-485 and mir-491 on luciferase activity, related to Figure 4.

30 FIGURE 22. 293T cells transfected with *pre-miR-185* mimic or pre-scramble (*pre-scr*) oligo. Expression levels of *C19orf63*, the human homolog of *2310044H10Rik (Mirta22)*, were assayed by qRT-PCR. Expression levels of *C19orf63* in cells transfected with *pre-miR-185* (n = 3) were normalized to the *pre-scr* controls (n = 3).

FIGURE 23A-B. Coordinated Mild Dysregulation of Golgi-related Putative miR-185 Targets in *Df(16)A^{+/-}* mice. (A) 2708 genes that were predicted to be *miR-185* targets by TargetScan Mouse v5.2 were imported into the DAVID bioinformatics resources 6.7 (<http://david.abcc.ncifcrf.gov>) and 2695 genes have

5 corresponding DAVID IDs. Functional annotation analysis using *Mus musculus* genes as background identified Gene Ontology (cellular component) term "Golgi apparatus" as the top enriched gene cluster (gene count = 159, Enrichment Score = 8.56, FDR-corrected $P = 2 \times 10^{-9}$) and term "Golgi apparatus part" as the second best hit with FDR-corrected $P = 4 \times 10^{-3}$ (left). The same gene list was also imported into Gene Set

10 Enrichment Analysis (GSEA v2.0). The Gene Ontology (cellular component) terms "Golgi apparatus part" (NES = 1.35, $P = 0.1$) and "Golgi apparatus" (NES = 1.3, $P = 0.1$) were again among the top enriched gene sets (right). (B) Expression heatmap plot of the potential miR-185 targets that serve Golgi apparatus related functions (GO term) and are differentially expressed ($p < 0.005$) between adult HPC of *Df(16)A^{+/-}*

15 mice and Wt littermates. ID is Affymetrix ID (see FIGURE 28) and Rank is the ranking position in the list of all differentially expressed genes according to significance level. Note that the majority (91%, 42 out of 46) of the genes are upregulated.

FIGURE 24. *miR-185* Reduction Results in Coordinated Mild and

20 Age-specific Dysregulation of Golgi-related Genes, Related to Figure 5

Expression heatmap plot of all potential *miR-185* targets that have Golgi apparatus related function (GO term) in E17 (left panel), P6 (middle panel) and adult (right panel) HPC of *Df(16)A^{+/-}* mice and Wt littermates.

FIGURE 25. Schematic outlining the proposed relationship among

25 22q11.2 microdeletions, miRNA dysregulation, 2310044H10Rik (*Mirta22*) and neuronal maturation. In normal postnatal development, miR-185, together with other miRNAs (such as mir-485 and mir-491) whose biogenesis is controlled by *Dgcr8*, repress *Mirta22* levels and promote the development of dendrites and spines. In 22q11.2 microdeletion carriers, miRNA biogenesis deficits due to hemizyosity of

30 *Dgcr8* along with a drastic reduction in the levels of miR-185 results in protracted elevation of *Mirta22* levels, which in turn may impair the growth of dendrites and spines and affect formation and maintenance of neural circuits.

FIGURE 26A-E. *2310044H10Rik (Mirta22)* Levels Affect Dendritic and Spine Development, Related to Figure 7. (A) Sholl analysis of dendritic

complexity (using 10- μ m concentric circles around the soma) of Wt neurons transfected with a plasmid carrying either full length *2310044H10Rik (Mirta22)* (n = 25) or no insert (n = 24) at DIV7 and fixed at DIV9. Increase in branching in Wt neurons transfected with full length *2310044H10Rik (Mirta22)*, is prevalent throughout the dendritic tree. (B) Sholl analysis of dendritic complexity (using 10- μ m concentric circles around the soma) of *Df(16)A^{+/-}* neurons transfected with either *2310044H10Rik (Mirta22)* shRNA (n = 29) or scramble shRNA (n = 26) at DIV7 and fixed at DIV9. Reduction in branching in *Df(16)A^{+/-}* neurons is partially reversed by *2310044H10Rik (Mirta22)* shRNA, especially in the vicinity of the most proximal dendrites. (C) Reduction in the number of primary dendrites in *Df(16)A^{+/-}* neurons at DIV9 relative to Wt neurons is reversed by the transfection of a construct that expresses an independent *2310044H10Rik (Mirta22)* shRNA (n = 16 for Wt + scr shRNA; n = 16 for *Df(16)A^{+/-}* + scr shRNA; n = 16 for *Df(16)A^{+/-}* + *Mirta22* shRNA). Scr shRNA: scramble shRNA. (D) Sholl analysis showing that reduction in branching in *Df(16)A^{+/-}* neurons is partially reversed by *2310044H10Rik (Mirta22)* shRNA, especially in the vicinity of the most proximal dendrites. (E) Reduction in the density of mushroom spines (estimated over 75 μ m of dendritic length) in *Df(16)A^{+/-}* neurons at DIV19 relative to Wt neurons is reversed by the introduction of *2310044H10Rik (Mirta22)* shRNA, but not scramble shRNA (n = 12 for Wt + scr shRNA; n = 12 for *Df(16)A^{+/-}* + scr shRNA; n = 12 for *Df(16)A^{+/-}* + *Mirta22* shRNA). Values of *Df(16)A^{+/-}* neurons were normalized to Wt + scr shRNA. Results are expressed as mean $\pm$ SEM. *p < 0.05, **p < 0.01, ***p < 0.001 (Student's t-test).

FIGURE 27. List of transcripts outside the 22q11.2 syntenic region misregulated in a reciprocal manner, related to Figure 4.

FIGURE 28. Gene Expression profile of predicted mir-185 targets that have Golgi related functions, related to Figure 23.

5. DETAILED DESCRIPTION OF THE INVENTION

As discussed in the working example below, it was discovered that a consequence of 22q11.2 hemizygous microdeletion is heterozygosity for expression of the microRNA, mir-185. It was further discovered that decreased levels of mir-185 consequent to heterozygosity lead to overexpression of *Mirta22*, and that overexpression of *Mirta22* is associated with changes in neuronal morphology,

including decreased numbers of primary dendrites, decreased spine density, and a small but significant reduction in mushroom spine width, all of which are, in turn, associated with decreased neuronal connectivity. According to the invention, neuronal connectivity may be increased by decreasing the activity of Mirta22 and/or
 5 increasing the activity of mir-185. Further, as both mir-485 and mir-491 also have binding sites in proximity to Mirta22, the present invention also provides for increasing neuronal connectivity by increasing the activity of mir-485 and/or mir-491.

For clarity of description, and not by way of limitation, the detailed description of the invention is divided into the following subsections:

- 10 (i) mir-185, mir-485 and mir-491;
 (ii) Mirta22; and
 (iii) methods of treatment.

5.1 MIR-185, MIR-485 AND MIR-491

15 The present invention provides for mir-185, mir-485 and mir-491 nucleic acid molecules. These nucleic acid molecules may be comprised of deoxyribonucleotides and/or ribonucleotides and may optionally comprise non-naturally occurring nucleotides, for example phosphorothioate residues, to promote stability. In non-limiting embodiments the mir-185, mir-485 or mir-491 nucleic acid
 20 molecules are between about 15 and 100, or between about 15 and 80, or between about 15 and 70, or between about 15 and 60, or between about 15 and 50, or between about 15 and 40, or between about 15 and 30, nucleotides in length or between about 22 and 100, or between about 22 and 80, or between about 22 and 70, or between about 22 and 60, or between about 22 and 50, or between about 22 and 40, or between
 25 about 22 and 30, nucleotides in length.

In certain non-limiting embodiments, the present invention provides for murine mir-185 nucleic acid molecules, including, as non-limiting examples, the mir-185 RNA molecule as well as precursors thereof. In certain non-limiting
 30 embodiments, the present invention provides for murine mir-185 nucleic acid molecules. Such molecules may comprise the sequence, GenBank Acc. No. NR_029571: agggattggagagaaaggcagttcctgatgtccctcccagg
 ggctggcttctctgtgtcctt (SEQ ID NO:6), or a sequence at least 95 percent homologous thereto or at least 99 percent homologous thereto (homology, as that term is used

herein, may be measured using standard software such as BLAST or FASTA). In related non-limiting embodiments, some or all of the thymidines in SEQ ID NO:6 may be substituted with uridine. For example, a murine mir-185 nucleic acid may comprise the sequence

- 5 agggauuggagagaaaggcaguuccugaugguccccucccaggggcuggcuuccucugguccuu (SEQ ID NO:7). or may comprise the subsequence (miRBase Acc. No. MIMAT0000214) uggagagaaaggcaguuccuga (SEQ ID NO: 8). In further embodiments, a murine mir-185 nucleic acid may comprise a 15-21 nucleotide fragment of SEQ ID NO:6 or SEQ ID NO:7 or SEQ ID NO:8.

- 10 In certain non-limiting embodiments, the present invention provides for human mir-185 nucleic acid molecules, including, as non-limiting examples, the mir-185 RNA molecule as well as precursors thereof. In certain non-limiting embodiments, the present invention provides for human mir-185 nucleic acid molecules. Such molecules may comprise the sequence, GenBank Acc No.

- 15 NR_029706: agggggcgagggattggagagaaaggcagttctgatgtccctcccca
ggggctggcttctcttgtt ccttcctccca (SEQ ID NO:9), or a sequence at least 95 percent homologous thereto or at least 99 percent homologous thereto. In related non-limiting embodiments, some or all of the thymidines in SEQ ID NO:9 may be substituted with uridine. For example, a human mir-185 nucleic acid may comprise the sequence:
- 20 agggggcgagggauuggagagaaaggcaguuccugaugguccccucccaggggcuggcuuccucuggu
ccuuccuccca (SEQ ID NO:10) or the sequence uggagagaaaggcaguuccuga (SEQ ID NO:8). In further embodiments, a human mir-185 nucleic acid may comprise a 15-21 nucleotide fragment of SEQ ID NO:8 or SEQ ID NO:9 or SEQ ID NO:10.

- In certain non-limiting embodiments, the present invention provides
25 for murine mir-485 nucleic acid molecules, including, as non-limiting examples, the mir-485 RNA molecule as well as precursors thereof. In certain non-limiting embodiments, the present invention provides for murine mir-485 nucleic acid molecules. Such molecules may comprise the sequence, MiRBase Acc. No.

- MI0003492: ACUUGGAGAGAGGCUGGCCGUGAUGAAU
30 UCGAUUCAUCUAAACGAGUCAUACACGGCUCUCCUCUCUUCUAGU (SEQ ID NO:11) or MiRBase Acc. No. MIMAT0003128:
AGAGGCUGGCCGUGAUGAAUUC

(SEQ ID NO:12), or a sequence at least 95 percent homologous thereto or at least 99 percent homologous thereto (homology, as that term is used herein, may be measured

using standard software such as BLAST or FASTA). In further embodiments, a murine mir-485 nucleic acid may comprise a 15-21 nucleotide fragment of SEQ ID NO:11 or SEQ ID NO:12.

In certain non-limiting embodiments, the present invention provides for human mir-485 nucleic acid molecules, including, as non-limiting examples, the mir-485 RNA molecule as well as precursors thereof. In certain non-limiting embodiments, the present invention provides for human mir-485 nucleic acid molecules. Such molecules may comprise the sequence, MiRBase Acc. No. MI0002469: ACUUGGAGAGAGGCUGGCCGUGAUGAAU
 5 UCGAUUCAUCAAAGCGAGUCAUACACGGCUCUCCUCUCUUUUAGU (SEQ ID NO:13) or MiRBase Acc. No. MIMAT0002175: AGAGGCUGGCCGUGAUGAAUUC
 10 (SEQ ID NO:14), or a sequence at least 95 percent homologous thereto or at least 99 percent homologous thereto. In further embodiments, a human mir-485 nucleic acid may comprise a 15-21 nucleotide fragment of SEQ ID NO:13 or SEQ ID NO:14.
 15

In certain non-limiting embodiments, the present invention provides for murine mir-491 nucleic acid molecules, including, as non-limiting examples, the mir-491 RNA molecule as well as precursors thereof. In certain non-limiting embodiments, the present invention provides for murine mir-491 nucleic acid molecules. Such molecules may comprise the sequence, MiRBase Acc. No. MI0004680: AAUUGACUUAGCUGGGAAGUGGGGAACCCU
 20 UCCAUGAGGAGUAGAACACUCCUUAUGCAAGAUUCCCUUCUACCUGACU GAGUUGA (SEQ ID NO:15) or MiRBase Acc. No. MIMAT0003486: AGUGGGGAACCCUCCAUGAGG (SEQ ID NO:16), or a sequence at least 95 percent homologous thereto or at least 99 percent homologous thereto (homology, as that term is used herein, may be measured using standard software such as BLAST or FASTA). In further embodiments, a murine mir-491 nucleic acid may comprise a 15-21 nucleotide fragment of SEQ ID NO:15 or SEQ ID NO:16.
 25

In certain non-limiting embodiments, the present invention provides for human mir-491 nucleic acid molecules, including, as non-limiting examples, the mir-491 RNA molecule as well as precursors thereof. In certain non-limiting embodiments, the present invention provides for human mir-491 nucleic acid molecules. Such molecules may comprise the sequence, MiRBase Acc. No. MI0003126: UUGACUUAGCUGGGUAGUGGGGAACCCU
 30

UCCAUGAGGAGUAGAACACUCCUUAUGCAAGAUUCCCUUCUACCUGGCU
GGGUUGG (SEQ ID NO:17), or MiRBase Acc. No. MIMAT0002807:

AGUGGGGAACCCUCCAUGAGG (SEQ ID NO:18), or a sequence at least 95
percent homologous thereto or at least 99 percent homologous thereto. In further
5 embodiments, a human mir-491 nucleic acid may comprise a 15-21 nucleotide
fragment of SEQ ID NO:17 or SEQ ID NO:18.

The present invention further provides for nucleic acid molecules
comprising nucleotide sequences that are the complements of any of the above-
described nucleotide sequences (SEQ ID NO:6-18).

10

5.2 MIRTA22

The present invention provides for Mirta22, also known in the
literature as 2310044H10Rik (murine) and C19orf63 (human).

In certain non-limiting embodiments, the present invention provides
15 for a murine Mirta22 protein comprising an amino acid sequence as depicted in
FIGURE 6A (SEQ ID NO:1) or a subsequence comprising amino acids 11-269
thereof, or a subsequence comprising amino acids 38-269 thereof, or a variant of the
complete protein or subsequence that is at least about 95 percent or at least about 99
percent homologous thereto.

20 In certain non-limiting embodiments, the present invention provides
for a nucleic acid encoding a murine Mirta22 protein, as described above. For
example, said nucleic acid may comprise the sequence set forth in GenBank
Accession No. NM_197991.2 and as depicted in FIGURE 16C (SEQ ID NO:19), or
may be at least about 95 percent or at least about 99 percent homologous thereto, or
25 may comprise a subsequence of SEQ ID NO:19 comprising the coding sequence from
nucleotide 115 through 921, or a sequence that is at least about 95 percent or at least
about 99 percent homologous thereto, or a nucleic acid that is complementary to any
of the foregoing sequences. The present invention further provides for an antisense
RNA or interfering RNA ("RNAi") or short hairpin RNA ("shRNA") that comprises a
30 nucleotide sequence complementary to a portion of SEQ ID NO:19 over a region of
between about 15 and 35 or between about 15 and 25 nucleotides, where there can be
no more than five or no more than four or no more than three or no more than two or
no more than one or zero mismatches in complementarity.

In certain non-limiting embodiments, the present invention provides for an antibody or fragment (e.g., Fab or F(ab')₂ thereof, or single chain antibody, or diabody, that specifically binds to murine Mirta22 protein, for example a protein having SEQ ID NO:1. Antibody or the source of antibody fragment may be

5 polyclonal or monoclonal antibody. In a specific, non-limiting embodiment of the invention, the antibody may be directed to the subfragment of murine Mirta22, EQAQKAKNPQEQKSFFAKY (SEQ ID NO:20).

In certain non-limiting embodiments, the present invention provides for a human Mirta22 protein (isotype 1) comprising an amino acid sequence as depicted in FIGURE 6A (SEQ ID NO:2), GenBank Accession No. NM_206538.2,

10 MAAASAGATRLLLLLLMAVAAPSRARGSGCRAGTGARGAGAEGREGEACG
TVGLLLEHSFEIDDSANFRKRGSLLWNQQDGTLSLSQRQLSEEERGRLRDVA
ALNGLYRVRIPRRPGALDGLEAGGYVSSFVPACSLVESHLSDQLTLHVDVAG
NVVGVSVVTHPGGCRGHEVEDVDLELFNTSVQLQPPTTAPGPETAAFIERLE

15 MEQAQKAKNPQEQKSFFAKYWMYIIPVVLFLMMSGAPDTGGQGGGGGGGGG
GGGSGR (SEQ ID NO:2) or a subsequence thereof comprising amino acids 28-263, or a variant of the complete protein or subsequence that is at least about 95 percent or at least about 99 percent homologous thereto.

In certain non-limiting embodiments, the present invention provides

20 for a nucleic acid encoding a human Mirta22 protein (isotype 1), as described above. For example, said nucleic acid may comprise the sequence as set forth in GenBank Accession No. NM_206538.2 and as depicted in FIGURE 16A (SEQ ID NO:3) or a sequence that is at least about 95 percent or at least about 99 percent homologous thereto, or may comprise a subsequence of SEQ ID NO:3 comprising the coding

25 sequence between nucleotides 67 and 855 (inclusive; all ranges recited in this document are inclusive of their stated limits unless provided otherwise) or a sequence that is at least about 95 percent or at least about 99 percent homologous thereto, or may comprise a subsequence of SEQ ID NO:3 comprising the coding sequence without the portion encoding the signal peptide between nucleotides 148 and 855, or a

30 sequence that is at least about 95 percent or at least about 99 percent homologous thereto, or a nucleic acid that is complementary to any of the foregoing sequences. The present invention further provides for an antisense RNA or RNAi or shRNA comprising a nucleotide sequence that is complementary to a portion of SEQ ID NO:3 over a region of between about 15 and 35 or between about 15 and 25 nucleotides,

where there can be no more than five or no more than four or no more than three or no more than two or no more than one or zero mismatches in complementarity.

In certain non-limiting embodiments, the present invention provides for an antibody or fragment (e.g., Fab or F(ab')₂ thereof, or single chain antibody, or diabody, that specifically binds to human Mirta22 protein, for example a protein having SEQ ID NO:2. Antibody or the source of antibody fragment may be polyclonal or monoclonal antibody. In a specific, non-limiting embodiment of the invention, the antibody may be directed to the subfragment of human Mirta22 sharing the sequence with the murine protein, EQAQKAKNPQEQKSFFAKY (SEQ ID NO: 20).

In certain non-limiting embodiments, the present invention provides for a human Mirta22 protein (isotype 2) comprising an amino acid sequence as depicted in GenBank Accession No. NM_175063.4,
 MAAASAGATRLLLLLLMAVAAPSRARGSGCRAGTGARGAGAEGR
 EGEACGTVGLLLEHSFEIDDSANFRKRGSLLWNQQDGTLSLSQRQLSEEERGR
 LRDVAALNGLYRVRIPRRPGALDGLEAGGYVSSFVPACSLVESHLSDQLTLH
 VDVAGNVVGVSVVTHPGGCRGHEVEDVDLELFNTSVQLQPPTTAPGPETAAF
 IERLEMEQAQKAKNPQE
 QKSFFAKYWHIILGGAVLLTALRPAAPGPAPPPQEA (SEQ ID NO:4), or a subsequence thereof comprising amino acids 28-263, or a variant of the complete protein or subsequence that is at least about 95 percent or at least about 99 percent homologous thereto.

In certain non-limiting embodiments, the present invention provides for a nucleic acid encoding a human Mirta22 protein (isotype 2), as described above. For example, said nucleic acid may comprise the sequence as set forth in GenBank Accession No. NM_175063.4 and as depicted in FIGURE 16B (SEQ ID NO:5) or a sequence that is at least about 95 percent or at least about 99 percent homologous thereto, or may comprise a subsequence of SEQ ID NO:5 comprising the coding sequence between nucleotides 67 and 831 or a sequence that is at least about 95 percent or at least about 99 percent homologous thereto, or may comprise a subsequence of SEQ ID NO:5 comprising the coding sequence without the portion encoding the signal peptide between nucleotides 148 and 831, or a sequence that is at least about 95 percent or at least about 99 percent homologous thereto, or a nucleic acid that is complementary to any of the foregoing sequences. The present invention

further provides for an antisense RNA or RNAi or shRNA comprising a nucleotide sequence that is complementary to a portion of SEQ ID NO:5 over a region of between about 15 and 35 or between about 15 and 25 nucleotides, where there can be no more than five or no more than four or no more than three or no more than two or
5 no more than one or zero mismatches in complementarity.

In certain non-limiting embodiments, the present invention provides for an antibody or fragment (e.g., Fab or F(ab')₂ thereof, or single chain antibody, or diabody, that specifically binds to human Mirta22 protein, for example a protein having SEQ ID NO:4. Antibody or the source of antibody fragment may be
10 polyclonal or monoclonal antibody. In a specific, non-limiting embodiment of the invention, the antibody may be directed to the subfragment of human Mirta22 sharing the sequence with the murine protein, EQAQKAKNPQE QKSFFAKY (SEQ ID NO:20).

15 5.3 METHODS OF TREATMENT

In certain non-limiting embodiments, the present invention provides for a method for increasing connectivity between neurons comprising administering to said neurons an effective amount of an agent that inhibits Mirta22 activity.

“Increasing connectivity between neurons” includes increasing the
20 number of primary dendrites and/or increasing the spine density and/or increasing the width of mushroom spines on neurons and/or increasing the number of synapses.

The method for increasing connectivity between neurons may be practiced in vitro or in vivo.

The neurons in said methods may be human or non-human neurons.

“Inhibits Mirta22 activity” includes reducing and/or antagonizing
25 Mirta22 activity. Agents that inhibit Mirta22 activity may act, for example, by reducing transcription or translation or processing of Mirta22 or by antagonizing the function of Mirta22, for example by binding to it such as to reduce or prevent it from performing its natural function.

30 Agents that inhibit Mirta22 activity include, but are not limited to, mir-185, mir-485, mir-491, antisense RNA comprising a sequence complementary to the Mirta22 mRNA; RNAi comprising a sequence complementary to the Mirta22 mRNA, shRNA comprising a sequence complementary to the Mirta22 mRNA, an anti-

Mirta22 antibody or fragment thereof, a single chain antibody that specifically binds to Mirta22, or a diabody that specifically binds to Mirta22.

In certain non-limiting embodiments, the present invention provides for a method for increasing connectivity between neurons comprising administering to
5 said neurons an effective amount of an agent that increases mir-185 activity.

“Increasing mir-185 activity” includes increasing the amount or functionality of mir-185. For example, but not by way of limitation, agents that increase mir-185 activity include a mir-185 nucleic acid, as well as agents that promote mir-185 functionality, for example agents that increase the activity of Dgcr8
10 including Dgcr8 protein or a nucleic acid encoding Dgcr8 protein. Agents that directly increase mir-185 activity include mir-185 nucleic acid. Agents that indirectly increase mir-185 include agents that increase the activity of Dgcr8.

In certain non-limiting embodiments, the present invention provides for a method for increasing connectivity between neurons comprising administering to
15 said neurons an effective amount of an agent that increases mir-485 activity.

“Increasing mir-485 activity” includes increasing the amount or functionality of mir-485. For example, but not by way of limitation, agents that increase mir-485 activity include a mir-485 nucleic acid, as well as agents that promote mir-485 functionality, for example agents that increase the activity of Dgcr8
20 including Dgcr8 protein or a nucleic acid encoding Dgcr8 protein. Agents that directly increase mir-485 activity include mir-485 nucleic acid. Agents that indirectly increase mir-485 include agents that increase the activity of Dgcr8.

In certain non-limiting embodiments, the present invention provides for a method for increasing connectivity between neurons comprising administering to
25 said neurons an effective amount of an agent that increases mir-491 activity.

“Increasing mir-491 activity” includes increasing the amount or functionality of mir-491. For example, but not by way of limitation, agents that increase mir-491 activity include a mir-491 nucleic acid, as well as agents that promote mir-491 functionality, for example agents that increase the activity of Dgcr8
30 including Dgcr8 protein or a nucleic acid encoding Dgcr8 protein. Agents that directly increase mir-491 activity include mir-491 nucleic acid. Agents that indirectly increase mir-491 include agents that increase the activity of Dgcr8.

In related non-limiting embodiments, the present invention provides for a method for increasing connectivity between neurons in a subject in need of such

treatment comprising administering to said subject an effective amount of an agent that inhibits Mirta22 activity.

A subject in need of such treatment is a subject who suffers from schizophrenia or a related disorder or a subject who has a disorder of memory or learning or a subject who would benefit from enhancement of neural connectivity or memory or learning capabilities. A subject may be a human or non-human subject.

Schizophrenia-related disorders include but are not limited to brief psychotic disorder, schizophreniform disorder, schizoaffective disorder, and delusional disorder.

Disorders of memory include, but are not limited to, Alzheimer's disease, Lewy body dementia, vascular dementia, corticobasal degeneration, Creutzfeld-Jacob disease, frontotemporal dementia, Huntington's disease, mild cognitive impairment, progressive supranuclear palsy, and Picks disease.

Disorders of learning include, but are not limited to, attention deficit hyperactivity disorder, dyslexia, dyscalcula, nonverbal learning disorder and mental retardation.

Additional subjects that may benefit from treatment according to the invention include, but are not limited to, subjects suffering from autism, Asperger's syndrome, depression, bipolar disorder, obsessive compulsive disorder, and subjects suffering from memory or learning deficits secondary to cerebral infarction, surgical, radiologic, or chemotherapeutic treatment of the brain, or trauma. The present invention provides for a method for increasing connectivity between neurons in such subjects comprising administering to said subject an effective amount of an agent that inhibits increases mir-185, mir-485 and/or mir-491 activity and/or inhibits Mirta22 activity.

In certain non-limiting embodiments, the present invention provides for a method of treating schizophrenia or a related disorder in a subject, comprising administering to said subject an effective amount of an agent that inhibits Mirta22 activity.

In certain non-limiting embodiments, the present invention provides for a method of treating schizophrenia or a related disorder in a subject, comprising administering to said subject an effective amount of an agent that increases mir-185, mir-485 and/or mir-491 activity.

In certain non-limiting embodiments, the present invention provides for a method of enhancing memory in a subject, comprising administering to said subject an effective amount of an agent that inhibits Mirta22 activity.

5 In certain non-limiting embodiments, the present invention provides for a method of enhancing memory in a subject, comprising administering to said subject an effective amount of an agent that increases mir-185, mir-485 and/or mir-491 activity.

Enhancement of memory may be determined using standard tests. In non-human animals, for example, memory may be tested using performance in a passing through a maze. In humans, memory may be tested, for example but not by way of limitation, using the Mini-Mental State Exam or the Wechsler Memory Scale.

In any of the foregoing, the species of origin of nucleic acids or antibodies or related molecules being utilized is preferably the same as or closely related to the species of the neurons or subject being treated.

15 According to the invention, the agent may be administered by any route, including but not limited to intrathecal, intravenous, nasal, intraarterial, oral, intramuscular, subcutaneous, pulmonary, intraperitoneal, etc.

The present invention provides for pharmaceutical compositions comprising an agent for use according to the invention in a suitable carrier, such as, but not limited to, normal saline or sterile water.

The present invention is exemplified but not limited by the following working example, which sets for additional specific non-limiting embodiments of the present invention. The contents of the working example are hereby incorporated by reference into the detailed description of the invention.

25

6. WORKING EXAMPLE

6.1 MATERIALS AND METHODS

Mice, qRT-PCR and expression profiling. Generation of *Df(16)A^{+/-}* and *Dgcr8^{+/-}* mice is described in (8). Total RNA was isolated by miRNeasy mini kit. qRT-PCR was performed as described in (8). For expression profiling, cDNA was generated and exposed to the Affymetrix Mouse genome 430 2.0 array, which includes 45,000 probe sets from > 34,000 well characterized mouse genes. Data were

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obtained using GeneChip Analysis Software Microarray Suite version 5 and analyzed with limma package in the bioconductor project (www.bioconductor.org)_ .

Analysis of dendritic complexity and spine morphology. Dissected E17 hippocampal neurons were plated at 2×10^5 cells/ml in 6-well plates and cultured
 5 for 9 – 19 days, depending on the experiments. Images of basal dendrites and dendritic spines were acquired as described previously (9). An experimenter blind to the genotype performed imaging and analysis. Semi-automatic tracing and quantification was performed using ImageJ and NeuroStudio (30,31). The number of primary dendrites and total branch points for each treatment condition were calculated
 10 by L-measure [version 4.0](32). Statistics were conducted using Student's t-test (number of primary dendrites and branch points, as well as spine density) and Kolmogorov-Smirnov test (width of mushroom spines).

Luciferase assay. Mirta22 3'UTR was cloned into psiCHECK2. Binding site mutant clones were generated by PCR-based mutagenesis. N18
 15 neuroblastoma cells were transfected with various psiCHECK2 reporter constructs together with pre-mir-185 mimic or pre-scramble control unless mentioned otherwise and luciferase assays were performed using the Promega Dual-Luciferase Reporter Assay System. All experiments were performed at least 2 times and all data presented is the average of 3 technical repeats.

Antibodies. A 20 amino acid peptide ([H]-
 20 CEQAQKAKNPQEQKSFFAKY -[NH₂]) was used to generate a rabbit polyclonal antibody. The specificity of the antibody was determined as described in FIGURE 11. Western blot, immunohistochemistry and immunocytochemistry assays were conducted as previously described (8,9).

In situ hybridization. Digoxigenin tail labeled anti-mir-185 locked nucleic acid (LNA) and scramble LNA oligonucleotides were obtained from Exiqon. In situ hybridization was conducted as described previously (33).

qRT-PCR and primers. Total RNA was isolated from brain or culture cells using the miRNeasy mini kit (QIAGEN) according to manufacturer's
 30 instructions. qRT-PCR was performed as described in detail previously (8). The sequences of primers used are as follows:

Mirta22: F: CTGCTGTCAATGGCCTCTAC (SEQ ID NO:1),

R: GTCCGAAAGGTGCGACTC (SEQ ID NO:2)

Hybridization probe: CATGGCCGCCAGCTTCTGA (SEQ ID NO:3)

mmu-miR-185: ABI Taqman assay ID 002271

Neuronal culture and transfection. Dissociated neurons were isolated from E17 mouse embryos and plated at 2×10^5 cells/ml in 6-well plates containing glass coverslips coated with poly-D-lysine. Neurons were cultured for 9 –
5 19 days, depending on the experiments. A pRFP-C-2310044H10Rik shRNA and pRFP-C-scramble shRNA control (Origene), a Mirta22 (2310044H10Rik) cDNA clone (Origene), as well as pre-mir-185 mimic and a pre-scramble control (Ambion) were used for high efficiency calcium-phosphate mediated transfections as described previously (34). For all experiments, 5 µg of total plasmids and/or 100 pmol of pre-
10 miRNA mimics were used per well.

Analysis of dendritic complexity. Images of basal dendrites were acquired as described previously (9). An experimenter blind to the genotype performed all imaging and analysis. The basal dendrite branches were semi-automatically traced from the somata of neurons using NeuronStudio(31). The .swc
15 files were imported into L-measure (32). The number of primary dendrites and total branch points for each treatment condition were calculated by L-measure (version 4.0). Statistical analysis was conducted using the Student's t-test as implemented in L-measure. Sholl analysis was conducted in ImageJ (<http://rsbweb.nih.gov/ij/>) using the “Sholl analysis” plugin
20 (<http://biology.ucsd.edu/labs/ghosh/software/ShollAnalysis.pdf>). The results were combined in the MS-Excel and a student t-test was conducted to determine the intersection number and significance at each step size between the experimental group and the controls.

Analysis of spine morphology. Images of dendritic spines were
25 acquired as described previously (9). For each experiment, images across all genotypes were acquired with similar optimal settings for laser power, detector gain, and amplifier offset. An experimenter blind to the genotype performed all imaging and analysis. Quantification of spine density, length, and width was performed using ImageJ (<http://rsbweb.nih.gov/ij/>) and NeuroStudio30. One basal dendrite, each at
30 least 75 µm in length from the first branch point, was analyzed per neuron. We analyzed at least 4 neurons from each animal. Spine width was measured as the distance of a straight line drawn across the widest part of the spine head. Changes in spine density between groups were assessed by Student's t-test. The distribution of width of mushroom spines was compared using the Kolmogorov-Smirnov test.

Expression profiling. A total of 48 PFC and 48 HPC from 12 *Df(16)A*^{+/-} mutants and 12 WT littermate control mice at E17 and P6 were dissected and processed using standard protocols recommended by Affymetrix. RNA quality was assessed with Bioanalyzer (Agilent Technologies) and all RNAs had a RIN > 7.0.

- 5 For hybridization, cRNA was exposed to the Affymetrix Mouse genome 430 2.0 array, which includes 45,000 probe sets from > 34,000 well characterized mouse genes, and then scanned (Agilent). Sequence clusters were created and refined from the UniGene and the Whitehead Institute Center for Genome Research databases.

- Microarray data analysis.** Initial intensity files (CEL files) were
10 obtained from microarray images using GeneChip (Affymetrix) Analysis Software Microarray Suite version 5 (Affymetrix) and analyzed with limma package in the bioconductor project (www.bioconductor.org). Data analysis of *Df(16)A*^{+/-} and Wt littermate expression profiling at E17 and P6 was performed as previously described (8). Briefly, the raw intensity data were first normalized by the Robust Multichip
15 Average (RMA) method, linear models were applied and differential expression was determined by limma package. The gene list was ranked according to their nominal P values and the significance level was estimated by adjusting P values using Benjamini and Hochberg FDR35 to control false positive rate due to multiple testing.

- Functional and geneset enrichment analysis of predicted *miR-185* targets.** *miR-185* target predictions were obtained using TargetScan Mouse v5.2. The
20 gene list was imported into DAVID gene functional annotation database. 92% (2708 out of 2932) predicted targets genes were mapped into the DAVID database. Functional annotation was conducted using the program's functional annotation clustering analysis with default settings. The gene list from the top cluster ("Golgi apparatus") was converted into Affymetrix IDs using the DAVID ID conversion tool
25 and further mapped to the corresponding IDs of the Affymetrix mouse 430 2.0 chip. This final probeset list was used as a user-defined geneset for downstream analysis. Geneset enrichment analysis was conducted using the ErmineJ software (38). Analysis was conducted using the receiver operator characteristic (ROC) analysis
30 based on p value rankings.

Luciferase Assays. Mirta22 3'UTR was cloned into XhoI and NotI sites of psiCHECK2 (Promega). Binding site mutant clones were generated by PCR-based mutagenesis. Site Mut1 sequence (starting from position 289 in 3'UTR): GGAgTtTGCCAAGCTCggTaaA (SEQ ID NO:4; lower case letters denote altered

nucleotide). Site Mut2 sequence (starting from position 350): AtTGTCACgCTaaA (SEQ ID NO:5). Mutations are predicted by RNAhybrid36 to disrupt the binding of mir-185 at the seeds and secondary binding sites. N18 neuroblastoma cells were transfected with various psiCHECK2 reporter constructs (100 ng per well of a 24-well plate) together with pre-mir-185 mimic or pre-scramble control (1 nM = 0.5 pmol), unless mentioned otherwise, and luciferase assays were performed 24 hrs post-transfection using the Dual-Luciferase Reporter Assay System (Promega) under the manufacturer's instructions. All experiments were performed at least 2 times and all data presented is the average of 3 technical repeats.

Antibody production. A 20 amino acid peptide ([H]-CEQAQKAKNPQEQKSFFAKY-[NH₂]) was designed, synthesized and conjugated to KLH (Keyhole Limpet Hemocyanin) and then injected into rabbits using Covance custom antibody production service (Covance). Antibody serum was then purified using Melon Gel IgG purification Kit (Thermo Scientific) and the purified antibody was used for western blot (1:50), immunohistochemistry and immunocytochemistry assays (1:25).

In western blot assays of 293T cell lysates, the purified antibody specifically recognized overexpressed full length Mirta22 tagged with C-terminal FLAG (~28 kD), which was independently verified using both polyclonal and monoclonal anti-FLAG antibodies (FIGURE 13A). The specificity of the antibody was tested in western blot assays of protein extracts from N18 cells transfected with a *2310044H10Rik* shRNA or a full length *2310044H10Rik* cDNA plasmid. The intensity of the band at ~28 kD, changed as predicted for each manipulation (Figure 13B, C). Finally, when used to stain cultured neurons the antibody generated a specific staining pattern whereas the pre-immune serum failed to show signal (Figure 13D).

Western blot assays. The western blot procedure has been described previously (8). Briefly, equal amounts of protein were separated and then transferred onto an ECF plus membrane. The membrane was probed with 1:50 purified Mirta22 antibody and then with 1:5000 horseradish peroxidase conjugated secondary antibody. The washed membrane was incubated with HRP substrate and chemiluminescence images were obtained using Alpha imaging system.

Immunohistochemistry assays. Immunohistochemistry assays were conducted according to (37). Briefly, fresh brains were dissected and immediately

frozen in OCT. Brains were then sliced at a thickness of 15–20 μm and brain sections were dried at room temperature and then fixed. Sections were incubated overnight with one or two primary antibodies and subsequently incubated for 1 hr with secondary antibody coupled to the Alexa Fluor fluorochromes (1:1,000, Invitrogen).

5 Images were examined under a fluorescence microscope (Nikon).

Immunocytochemistry assays. Cells were cultured on coverslips, fixed, permeabilized, and then exposed to primary antibodies and secondary antibodies as described previously (9). Images were examined under a fluorescence microscope (Nikon). For quantification of Mirta22 signals, images were acquired as
10 described previously (9). The raw LSM images were projected and exported using ImageJ. Mirta22 immunocytochemical signal from cell body of each individual neuron was calculated as total integrated density minus background of the cell body area.

15 6.2 RESULTS

Using chromosomal engineering, we generated a mouse model carrying a hemizygous 1.3-Mb chromosomal deficiency on mouse chromosome 16 [*Df(16)A*], which is syntenic to the 22q11.2 1.5-Mb microdeletion (FIGURE 1A-D)(8). Analysis of *Df(16)A*^{+/-} mice provided evidence for abnormalities in dendritic
20 morphogenesis and formation of dendritic spines of hippocampal pyramidal neurons both in culture and in vivo (8,9). Such changes may account, at least in part, for the regional decreases in grey matter volumes observed in some 22q11.2 deletion carriers (10). Even modest alterations in dendritic and spine formation may result in a suboptimal number of synaptic connections, formation of inappropriate connections
25 or changes in the integration of synaptic inputs, and may ultimately lead to considerable changes in information processing (11,12).

The *Df(16)A*^{+/-} strain also provided compelling evidence that the 22q11.2 deletion results in abnormal processing of brain microRNAs (miRNAs), a class of small, noncoding RNAs that regulate the stability and translation of mRNAs
30 (13-16). One gene disrupted by the 22q11.2 microdeletion is *Dgcr8*, a component of the 'microprocessor' complex that is essential for miRNA production (17). *Dgcr8*^{+/-} mice were generated (FIGURE 2A-E). *Dgcr8* haploinsufficiency results in the downregulation of a specific subset of mature miRNAs and contributes to a number of

cellular, synaptic and behavioral alterations found in *Df(16)A^{+/-}* mice (8,18). miRNA dysregulation likely accounts for a fraction of the transcript misexpression observed in the brains of *Df(16)A^{+/-}* mice but direct targets of the affected miRNAs have hitherto not been reported. The present experiments highlight an important component of this dysregulation and identify a previously uncharacterized gene as a major miRNA transcriptional target mediating the effects of the 22q11.2 microdeletions on neuronal maturation and connectivity.

A drastic reduction of *miR-185* levels in *Df(16)A^{+/-}* mice. Studies in the *Df(16)A^{+/-}* mouse strain have shown that the 22q11.2 microdeletion results in abnormal processing of a specific subset of brain miRNAs due to the removal of one copy of the *Dgcr8* gene and decrease in its expression in the adult brain (8) as well as earlier in the development (FIGURE 8). It is noteworthy that, in addition to *Dgcr8*, the 22q11.2 microdeletion and the equivalent mouse deficiency remove one miRNA gene (*mir-185*) located within the minimal 1.5-Mb 22q11.2 critical region (FIGURE 1A). In situ hybridization assays indicated that *mir-185* is expressed in several brain regions such as hippocampus (HPC) and cortex (FIGURE 3A), including frontal cortex. Quantitative real-time PCR (qRT-PCR) analysis showed that expression of mature *mir-185* is dramatically reduced by ~70-80% in both HPC ($P < 10^{-6}$) and prefrontal cortex (PFC, $P < 10^{-11}$) of adult *Df(16)A^{+/-}* mice (FIGURE 3B, C). This reduction was also observed at earlier developmental stages (E17 and P6) (FIGURE 9A, B). *mir-185* showed a more modest decrease in *Dgcr8^{+/-}* mice (~20% in HPC, $P < 0.05$; FIGURE 3D) suggesting that the severe reduction of mature *mir-185* expression in *Df(16)A^{+/-}* mice is due to a combined effect of hemizyosity of *mir-185* gene and impaired maturation of the pri-*mir-185* transcript produced from the remaining copy, due to the reduction in the *Dgcr8* levels. This represents a genuine gene X gene interaction within a pathogenic CNV that results in reduction of the expression of a resident gene considerably greater than expected by the 50% decrease in gene dosage. Such large reduction in relative expression is unique among genes affected by the microdeletion and may represent an important and previously unappreciated component of the 22q11.2-associated miRNA dysregulation. As such, drastically diminished *mir-185* activity may lead, either on its own or in combination with other miRNAs affected due to the *Dgcr8* deficiency, to altered developmental regulation of one or more target genes and impact a number of neural processes.

Altered *miR-185* levels contribute to structural alterations of *Df(16)A*^{+/-} neurons. miRNAs have been shown to affect structural indices of neuronal connectivity, such as dendritic tree and dendritic spine development (19-21). 22q11.2 microdeletion results in impaired dendritic tree and dendritic spine development in the brain and these deficits are recapitulated in primary neuronal cultures. Impaired dendritic tree and dendritic spine development could be partially accounted for by the 50% decrease in the levels of *Dgcr8* but it remained unknown whether reduction in the levels of *mir-185* also contributes (8,9,18). Localization of *Mirta22* within the Golgi apparatus and dendritic shafts suggests that diminishment of the repressive influence of *miR-185* on *Mirta22* levels may also contribute to these deficits.

To investigate this, we first asked whether reduction of *miR-185* levels results in deficits in dendritic and spine development similar to those observed in *Df(16)A*^{+/-} neurons (9). We introduced an anti-*miR-185* and a scramble control LNA oligonucleotide into Wt primary hippocampal neurons and measured dendritic and spine morphology two days post-transfection at DIV9 and DIV19, respectively. As mentioned above, we confirmed that introduction anti-*miR-185* LNA oligo resulted in a significant increase of *Mirta22* mRNA levels when compared to anti-*miR* control transfected primary neurons (FIGURE 5C). Analysis of dendritic architecture indicated that reduction of *miR-185* levels leads to deficits in dendritic complexity (FIGURE 17A), including a significant reduction in the number of primary dendrites (21%, $P < 0.05$; FIGURE 17B) and a significant reduction in total branch points in transfected neurons (16%, $P < 0.05$; FIGURE 17C). This finding was confirmed by a Sholl analysis, which compares branch point numbers at varying distances from the soma (FIGURE 18A). Moreover, reduction of *miR-185* levels in DIV19 neurons results in decreased mushroom spine density (21%, $P < 0.05$; FIGURES 17D and 17E and FIGURE 18B) and a significant reduction in their median width (15% decrease, $P < 0.001$, Kolmogorov-Smirnov test; FIGURE 17F). These structural deficits recapitulate those observed in *Df(16)A*^{+/-} neurons. Thus the neuronal deficits in *Df(16)A*^{+/-} mice are, at least in part, due to the aberrantly low level of *miR-185*. Consistently, introduction *pre-miR-185* mimic into Wt neurons increased the number of primary dendrites, the number of branch points, the density and head width of mushroom spines (FIGURES 18C and 18D).

We also examined whether elevation of *miR-185* levels could, at least partially, reverse cytoarchitectural alterations observed in *Df(16)A^{+/-}* neurons (9). We transfected primary hippocampal neurons from *Df(16)A^{+/-}* mice and their Wt littermates with a miRNA precursor mimic ("*pre-miR-185*," Ambion), which is processed into mature miRNA or a scramble precursor oligonucleotide ("pre-scramble") with no homology to the mouse genome, which serves as a control for nonspecific effects of small RNA expression. A co-transfected GFP reporter plasmid allowed us to analyze the dendritic architecture (FIGURE 3E) and spine morphology (FIGURE 3H) of GFP-positive pyramidal neurons at DIV9 (2 days following transfection) and DIV19 (2 days following transfection), respectively. In control experiments we confirmed that introduction of *pre-miR-185* resulted in significant decrease in the levels of *Mirta22* when compared to pre-scramble transfected neurons ($P < 0.01$; FIGURE 4B). Consistent with previous results, compared to WT neurons, *Df(16)A^{+/-}* neurons transfected with pre-scramble showed reduced dendritic complexity as manifested by a decrease in the number of primary dendrites (25%, $P < 10^{-10}$; FIGURE 3F) and the number of dendritic branchpoints (38%, $P < 10^{-4}$; FIGURE 3G). They also showed reduced spine density (38%, $P < 10^{-6}$, FIGURE 3I) as well as a small, but statistically significant decrease in the head-width (8% in median width, $P < 0.01$; FIGURE 3J) of mushroom spines. Increase in mir-185 activity largely reversed the deficits in dendritic complexity (FIGURE 3F, G, and FIGURE 18E) and the reduction in spine density (FIGURE 3I) and significantly increased the spine head-width of mushroom spines in *Df(16)A^{+/-}* hippocampal neurons (18% increase in median width, $P < 0.001$, Kolmogorov-Smirnov test; FIGURE 3J). Thus, introduction of pre-mir-185 into hippocampal neurons from *Df(16)A^{+/-}* mice reversed some key structural connectivity deficits.

A primary transcriptional consequence of 22q11.2 genomic losses.

Impaired production of miRNAs may have considerable impact on target transcript stability. Previous microarray analysis of adult *Df(16)A^{+/-}* mice revealed that reduction in dosage of genes in the 22q11.2 region results in genome-wide alterations of transcriptional programs in the HPC and PFC (8). We extended analysis of these two brain regions to two earlier developmental stages, embryonic day 17 (E17) and postnatal day 6 (P6), when mir-185 and possibly other miRNAs are already decreased in *Df(16)A^{+/-}* mice. Only one gene, 2310044H10Rik, was consistently found to be significantly upregulated in at least two of the three developmental stages examined

and in at least one of the two brain areas tested. Indeed, 2310044H10Rik was among the top upregulated genes in both postnatal stages examined (adult and P6) and the top upregulated transcript in the frontal cortex of P6 mutant mice (FIGURE 4A, B).

Notably, no significant difference in 2310044H10Rik expression was found in either
5 frontal cortex or hippocampus at E17 (FIGURE 4A, B).

In independent experiments, we attempted to distinguish primary versus secondary gene targets of the 22q11.2 microdeletion by looking for genes whose expression changes in opposite direction as a result of genomic losses or gains in this locus. Such genes are likely to represent primary targets and direct
10 transcriptional readouts of the underlying copy number imbalances. By contrast, expression changes specific to genomic losses or in the same direction independently of genomic dosage are more likely to be secondary reflections of affected physiological process or malfunctioning brain regions (39). We compared the PFC and HPC gene expression profiles in mice carrying a deletion or duplication at the
15 syntenic mouse locus using as reference compound heterozygous mice balanced for copy number (FIGURE 19). We identified a number of inversely altered transcripts in either PFC or HPC (P-value <0.001, FIGURE 27), in addition to the transcripts from the 22q11.2 region. The majority of the identified transcripts are pri-miRNA forms. Twelve transcripts were significantly misregulated in a reciprocal manner in both PFC
20 and HPC (FIGURE 20). Among them, 2310044H10Rik is the only gene with protein coding potential.

The pattern of upregulation in both PFC and HPC was confirmed by TaqMan qRT-PCR (PFC: E17, 20% P = 0.24; P6, 59%, P < 0.01; Adult, 76%, P < 10⁻⁶; HPC: E17, 20%, P = 0.16; P6, 50%, P < 0.05; Adult, 38%, P < 0.05; FIGURE 4C,
25 D). This analysis revealed a profile of temporal regulation where levels of 2310044H10Rik rapidly decline during the first week after birth (between E17 and P6) and remain constantly low thereafter, as well as a corresponding pattern of expression misregulation in *Df(16)A^{+/-}* mice where elevated expression of 2310044H10Rik persists throughout postnatal and adult life. Increased brain
30 expression of 2310044H10Rik is recapitulated in primary hippocampal neurons from *Df(16)A^{+/-}* mice (FIGURE 4E). Importantly, there is not any known miRNA within or surrounding the 2310044H10Rik genomic locus suggesting that the observed upregulation is not due to impaired processing of overlapping pri-miRNA transcripts.

2310044H10Rik is major downstream effector of miRNA

dysregulation. The gene expression profiling described above highlighted the impaired developmental regulation of 2310044H10Rik levels as a major consequence of the 22q11.2 deletion at the transcriptomic level. Notably, 2310044H10Rik mRNA levels were also elevated in *Dgcr8*^{+/-} mice (HPC: 30%, $P < 0.05$; PFC: 24%, $P < 0.05$; FIGURE 10), strongly suggesting that upregulation may be due to miRNA dysregulation. Indeed, two miRNA target site prediction programs, TargetScan(22) and mirDB (23), report that the 3'UTR of 2310044H10Rik contains binding sites of miRNAs shown both by microarray profiling (8) or qRT-PCR (FIGURE 11) to be affected in *Df(16)A*^{+/-} mice. Specifically, mirDB predicted 5 such miRNAs with binding sites in the 3'UTR of 2310044H10Rik including mir-185 and mir-485, whereas TargetScan predicted 13 miRNA sites, including sites for mir-185, mir-485, mir-491 and mir-224. Notably, both programs predicted that the 3'UTR of 2310044H10Rik contains sites for mir-185 and mir-485 (FIGURE 5A).

Because increased brain expression of 2310044H10Rik is recapitulated in primary neurons from *Df(16)A*^{+/-} mice (FIGURE 4E), we first used primary neurons to determine if endogenous 2310044H10Rik expression is actually under the control of *miR-185*. To examine the effect of *miR-185* overexpression on 2310044H10Rik level, we introduced into primary neuronal cultures a miRNA precursor mimic ("*pre-miR-185*"), which is processed into mature miRNA, or a scramble precursor ("*pre-scramble*") with no homology to the mouse genome, which serves as a control for nonspecific effects of small RNA expression. 24 hours post-transfection, there was a 54% decrease in the levels of 2310044H10Rik in pre-mir-185 transfected neurons when compared to pre-scramble transfected neurons ($P < 0.01$; FIGURE 5B). In a complementary experiment, introduction of an anti-*miR-185* LNA oligo or a scramble control oligo resulted in an increase of 2310044H10Rik mRNA levels in anti-*miR-185* transfected cells when compared to scramble transfected cells ($P < 0.05$; FIGURE 5C). Taken together, these results confirm that 2310044H10Rik expression in primary neurons is under the repressive control of *miR-185*. Essentially identical results were obtained when 2310044H10Rik expression was assayed in N18 cells (FIGURES 5D and 5E).

To further characterize the nature of the inhibition of mir-185 on 2310044H10Rik expression and test if it is 2310044H10Rik 3'UTR-dependent as predicted by TargetScan and mirDB (see above), 2310044H10Rik 3'UTR fused

luciferase reporter genes (see Methods and Figure legend for details) were cotransfected with either “pre-mir-185” mimic or a scramble precursor (“pre-scramble”), into N18 cells, a mouse neuroblastoma cell line. Pre-scramble did not affect the reporter activity. Introduction of pre-mir-185 mimic led to a dramatic decrease of luciferase activity as compared to the pre-scramble control ($P < 0.001$ for all pre-mir-185 concentrations used, compared to pre-scramble control; FIGURE 5F). Repression by mir-185 occurs over a 10^3 -fold range and more than 55% repression was still observed at a pre-mir-185 mimic concentration of 0.01 nM (FIGURE 5F).

To investigate if mir-185-mediated repression is specific and operates directly via the two binding sites predicted by TargetScan (FIGURE 5A), we engineered luciferase reporters carrying mutated versions of 2310044H10Rik 3'UTR with either individual or both mir-185 binding sites mutated (Mut1/Site 1 mutant; Mut2/Site 2 mutant; Mut1&2/Site 1 and 2 mutants). The pre-mir-185 mimic significantly reduced the luciferase activity of the WT reporter to ~25% relative to a control reporter without 3'UTR, while it reduced the luciferase activities of the Mut1 and Mut2 reporters to 80% ($P < 0.01$) and 33% ($P < 0.05$) respectively, compared to WT 2310044H10Rik 3'UTR fused reporter (FIGURE 5G). Notably, the pre-mir-185 mimic could not repress luciferase activity driven from a mutant reporter where both binding sites are simultaneously disrupted (FIGURE 5G). Both mir-185 cognate binding sites have an impact on the 3'UTR-mediated regulation of Mirta22 expression, although the site disrupted in the Mut1 reporter (Site 1) seems to be the major target site via which mir-185 directly exerts its repressive effect.

We further addressed the dependence of 2310044H10Rik 3'UTR reporter repression on the levels of mir-485 or mir-491, which are also predicted to target binding sites in the 3'UTR of 2310044H10Rik gene. None of these miRNAs is located within the 22q11.2 microdeletion, but both are modestly down-regulated in HPC of *Df(16)A^{+/-}* mice due to the *Dgcr8* hemizyosity (FIGURE 11). The pre-miRNA mimics of either miRNA modestly but significantly reduced the luciferase activity of the 3'UTR fused reporter compared to the pre-scramble control (pre-mir-485: 27%, $P < 0.05$; pre-mir-491: 35%, $P < 0.05$; FIGURE 5G). A three factor ANOVA analysis indicated that all three miRNAs (*miR-185*, *miR-485* and *miR-491*) and their interactions have significant impact on the luciferase activity with the exception of the interaction between *miR-485* and *miR-491* (FIGURE 21). Taken together, these findings suggest that the persistent elevation of 2310044H10Rik levels

observed in *Df(16)A*^{+/-} mice is likely the result of the combined hemizyosity at *mir-185* and *Dgcr8* loci. Although more than one miRNA contributes, the major effect is due to the dramatic downregulation of *mir-185*. Consistent with this notion and the less profound reduction of *mir-185* in *Dgcr8*^{+/-} mice (FIGURE 3D), *2310044H10Rik* is only modestly upregulated in this strain (FIGURE 10). Due to confirmed miRNA-mediated regulation, we renamed the gene *Mirta22* (miRNA target of the 22q11.2 deletion).

Interestingly, a comparison between the 3'UTR of human and mouse *Mirta22* orthologues as implemented by the "30-way multiz alignment and conservation analysis" in the USCS browser (FIGURE 5A) reveals that *mir-185* cognate Site 1, as well as one *mir-485* binding site are located within a highly conserved region, suggesting that these sites may also play a critical role in regulating the levels of the human orthologue (*C19orf63*). Consistent with this expectation, introduction of *pre-miR-185* into human 293T cells resulted in a significant decrease of endogenous *C19orf63* levels (FIGURE 22). In addition, similar to the pattern observed in the mouse brain, expression of *C19orf63* decreases in infant brain (40). It is also noteworthy that, inspection of our gene expression database as well as qRT-PCR analysis of a sample of eight high-likelihood *miR-185* targets identified by more than one prediction programs did not reveal any additional significant changes of transcript levels in the brains of *Df(16)A*^{+/-} mice (FIGURE 12). Furthermore, unlike *2310044H10Rik* no other of the top upregulated protein coding genes (shown in FIGURE 4B) are consistently altered in the both HPC and frontal cortex of E17, P6 and adult *Df(16)A*^{+/-} mice and only one of them (*B3gat1*, see below) was predicted to contain *miR-185* seed sites in its 3'UTR. Overall, although additional downstream targets of *miR-185* may exist, especially at the level of translation, our analysis suggests that *Mirta22* represents a major downstream effector of *miR-185* and a major hub target of the miRNA dysregulation due to the 22q11.2 microdeletion.

***Mirta22* encodes a novel neuronal protein residing in the Golgi apparatus.** *Mirta22* encodes a 28 kD protein without any known sequence homology or functional domain (<http://www.uniprot.org/uniprot/Q3TAS6>). The murine orthologue is located on mouse chromosome 7 and contains seven coding exons. The human orthologue (*C19orf63*) is located on chromosome 19q13.33 and encodes a protein with 92.3% identity to the murine *Mirta22* protein (FIGURE 6A). One mouse reference sequence (isoform 1) is reported in GeneBank, while two *C19orf63*

isoforms (isoform 1 and 2) are reported in GeneBank and in the literature (24). The protein encoded by isoform 1 is predicted to contain an N-terminal signal peptide, as well as a putative transmembrane segment (FIGURE 6A, red rectangles), which separates a long N-terminal region from a short C-terminal segment that contains a characteristic polyglycine tail with unknown function. Isoform 2 is generated by alternative splicing and differs from isoform 1 by an alternatively spliced exon located after exon 6. The protein encoded by isoform 2 is shorter by only 8 amino acids, contains the N-terminal signal peptide but not the transmembrane segment and it is predicted to be secreted (FIGURE 6A).

To investigate the distribution in the brain, as well as the subcellular localization of Mirta22, we raised a polyclonal antibody against a protein segment spanning both isoforms (amino acids 207–226, FIGURE 6A, green rectangle; see Materials and Methods and FIGURE 13A, B and G for antibody generation and specificity). Immunohistochemical analysis demonstrated that Mirta22 is widely distributed in the brain (FIGURE 14). The specificity of the antibody was validated by Western blot analysis on protein extracts from 293T cells transfected with a plasmid expressing full length *Mirta22* cDNA with a C-terminal FLAG tag (FIGURE 13A) or a mouse neuroblastoma cell line (N18), transfected with either a *Mirta22* shRNA (Figure 13B) or a full length cDNA plasmid (FIGURE 13C). Western blot assays of protein extracts from the brain of *Df(16)A^{+/-}* mice and their Wt littermates showed the expected increase (25%) in the levels of Mirta22 in mutant mice (FIGURE 6B). A similar in magnitude increase of the Mirta22 immunocytochemical signal was observed in *Df(16)A^{+/-}* cultured neurons, as compared to Wt neurons (FIGURE 6C). Analysis of cultured hippocampal neurons revealed that Mirta22 immunoreactivity is colocalized with neuron-specific marker NeuN, but not astrocyte-specific marker GFAP, indicating that Mirta22 is a neuronal protein (FIGURE 6D, upper panel). Mirta22 immunoreactivity is found both in the soma, where it colocalizes with the Golgi apparatus marker GM13025, as well as in vesicle-like clusters and tubular-like clusters within the dendritic shafts (FIGURE 6D, lower panel). Mirta22 immunoreactivity was not detected in cultures stained with preimmune serum (FIGURE 13D) and was diminished by 64% in *Mirta22* shRNA-transfected neurons (RFP+ neuron, FIGURE 13E, lower panel).

***miR-185* reduction results in coordinated mild dysregulation of Golgi-related genes.** Accumulating evidence suggests that miRNAs may target

functionally connected genes, often in a developmental stage-specific manner (41, 42). Consistent with this notion, functional annotation clustering analysis of 2708 predicted *miR-185* targets (TargetScan Mouse v5.2) included in the DAVID *Mus musculus* gene functional annotation database (<http://david.abcc.ncifcrf.gov>) identified as the top enriched gene cluster (gene count = 159, Enrichment Score = 8.56, FDR-corrected $P = 2 \times 10^{-9}$) the Gene Ontology (cellular component) term "Golgi apparatus" (FIGURE 23A). Gene set enrichment analysis (GSEA) on the 2708 predicted *miR-185* targets ranked based on the gene expression profile of *Df(16)A^{+/-}* mice also indicated that the Gene Ontology terms "Golgi apparatus part" and "Golgi apparatus" were among the top 20 genesets in the adult HPC (FIGURE 23A). A global perspective on the enrichment of this *miR-185* target gene set among the differentially expressed genes in the *Df(16)A^{+/-}* mice²⁹ showed a significant enrichment in the adult HPC expression profile ($P = 5 \times 10^{-4}$) where, as expected, most of top genes were upregulated (42 genes were upregulated and only 4 genes were downregulated at $P < 0.005$, FIGURE 23B and FIGURE 28). A considerably more modest enrichment was suggested for the E17 ($P = 0.02$) and P6 HPC ($P = 0.016$) profiles (FIGURE 24). Interestingly, there was no significant enrichment within the PFC profiles in all three ages tested (E17: $P = 0.6311$; P6: $P = 0.1326$; Adult: $P = 0.244$). Expression changes were modest, with only 4 out of 159 Golgi-related probe sets included among the top 100 probe sets in the adult HPC. Overall, in addition to the robust and pervasive upregulation of *Mirta22*, reduction in *miR-185* levels appears to affect expression of a group of Golgi apparatus-related genes in a milder, age and region-specific manner.

Elevation of Mirta22 levels inhibits dendritic and spine development in *Df(16)A^{+/-}* neurons.

The observation that reduction of *Mirta22* levels partially reverses the deficits in neuronal connectivity observed in *Df(16)A^{+/-}* mice strongly suggests that *Mirta22* may be an inhibitor of dendritic and spine development. To test this possibility and determine whether elevation of *Mirta22* levels phenocopies aspects of the *Df(16)A*, we introduced a *Mirta22* cDNA into WT primary hippocampal neurons and measured dendritic and spine morphology two days post-transfection at DIV9 and DIV19, respectively. Control experiments using qRT-PCR and western blot confirmed that the *Mirta22*-encoding plasmid drives increased expression of *Mirta22* at both mRNA and protein levels (FIGURE 7A, B). Analysis of dendritic architecture indicated that elevation of *Mirta22* levels results in a significant

reduction in the number of primary dendrites (18%, $P < 0.001$; FIGURE 7C) and total branch points in transfected neurons (41%, $P < 10^{-5}$; FIGURE 7D). This finding was confirmed by Sholl analysis (Figure 26A). Moreover, elevation of Mirta22 levels in DIV19 neurons results in decreased spine density (22%, $P < 0.05$; FIGURE 7E) and a small but significant reduction in the mushroom spine width (8% decrease in median width, $P < 0.001$, Kolmogorov-Smirnov test; FIGURE 7F). These structural deficits recapitulate those observed in *Df(16)A^{+/-}* neurons, suggesting the neuronal deficits in *Df(16)A^{+/-}* mice are, at least in part, due to the aberrantly high level of Mirta22.

We showed that the elevation of mir-185 levels in primary hippocampal neurons from *Df(16)A^{+/-}* mice reverses some key structural connectivity deficits that emerge as a result of the microdeletion. We also showed that Mirta22 is a major target of mir-185. Therefore, we considered the possibility that elevation of Mirta22 levels may contribute to at least some of the effects of the mir-185 deficiency on the neuronal morphology in *Df(16)A^{+/-}* mice. To test this hypothesis, we asked first whether reduction of Mirta22 levels could partially reverse the neuronal abnormalities observed in *Df(16)A^{+/-}* mice (9). We transfected primary hippocampal neurons isolated from *Df(16)A^{+/-}* embryos and their WT littermates with constructs that coexpress turbo RFP (tRFP) and either a shRNA engineered to knock down expression of endogenous mouse Mirta22 or a scramble control shRNA (scr shRNA). We confirmed that the Mirta22 shRNAs can effectively knockdown the expression of Mirta22 at both mRNA and protein levels (Figures 7G, 7H and 15A). We analyzed dendritic architecture and spine morphology two days following transfection, at DIV9 and DIV19 respectively. Consistent with previous reports, we observed significantly reduced dendritic complexity in *Df(16)A^{+/-}* neurons transfected with the control shRNA (FIGURE 7I, J). Introduction of Mirta22 shRNA restored to WT levels the number of primary dendrites of *Df(16)A^{+/-}* neurons at DIV9 (Mirta22 shRNA versus scr shRNA, 40% increase, $P < 10^{-5}$; FIGURE 7I). An increase in the total number of branch points in *Df(16)A^{+/-}* neurons was also observed but did not reach significance (25% increase, $P = 0.16$; FIGURE 7J). Sholl analysis confirmed that introduction of *Mirta22* shRNA in *Df(16)A^{+/-}* neurons increased branch point numbers mainly in the proximal dendritic segments from the soma (FIGURE 26B). Furthermore, while DIV19 *Df(16)A^{+/-}* neurons transfected with the control shRNA had fewer and thinner mushroom spines than WT neurons, introduction of Mirta22 shRNA into *Df(16)A^{+/-}* neurons reversed the deficit in spine density (Mirta22 shRNA

versus scr shRNA, 91% increase $P < 10^{-6}$; FIGURE 7K) while it had no impact on spine width (FIGURE 7L). The observation that reduction of *Mirta22* levels partially reverses the structural deficits observed in *Df(16)A^{+/-}* mice was confirmed by using an independent *Mirta22* shRNA (Figures 26C-E) and strongly suggests that *Mirta22* acts as an inhibitor mediating the effects of the structural mutation of dendritic and spine growth.

6.3 DISCUSSION

Elucidation of the biological processes affected by pathogenic CNVs may offer novel insights into the pathogenesis of psychiatric disorders as well as disorders of cognitive development (2). However, the transcriptional networks and signaling cascades that mediate the effects of CNVs on neuronal structure and function remain to a large extent uncharacterized. This study was designed to identify such downstream targets and processes disrupted by a *bona fide* pathogenic mutation that predisposes to schizophrenia and cognitive dysfunction. By applying an array of morphological, molecular and cellular assays to a mouse model of this mutation we provide a number of novel mechanistic insights.

First, taken together with previous results on the effects of *Dgcr8* hemizyosity (18, 8) we provide a comprehensive view of the pattern of miRNA dysregulation emerging due to 22q11.2 deletions, which is shaped by the combined (synergistic and additive) effect of *miR-185* and *Dgcr8* hemizyosity (see graphic summary). In this context, our results shows how a genuine gene X gene interaction within a pathogenic CNV can result in a considerably greater reduction of the expression of a resident gene than expected by the 50% decrease in gene dosage indicating that mechanisms other than simple haploinsufficiency could represent an important and previously unappreciated component of CNV pathogenicity. Along these lines, our results also raise the more general and intriguing possibility that 22q11.2 microdeletions, by partially disabling the miRNA machinery, create a sensitized genetic background, which promotes the effects of deleterious mutations that affect the expression or activity of a subset of miRNAs (43, 44).

Second, by comparing gene expression profiles over three developmental stages and variable levels of genomic dosage at the 22q11.2 locus we identified elevated levels of a previously uncharacterized gene, *Mirta22* as the most robust change in gene expression resulting from the 22q11.2 microdeletion, as well as

the major downstream transcriptional effects of the 22q11.2- associated miRNA dysregulation.

Third, using physiologically relevant cellular models we provide unequivocal evidence that *Mirta22* as well as its primary miRNA regulator (*miR-185*)
5 mediate, at least in part, the effects of the 22q11.2 microdeletions on dendrite and spine formation. Although results from acute manipulations of gene expression via transient transfections of primary neurons cannot be over-interpreted quantitatively, the observed convergence and remarkable consistency of data from a multitude of experimental manipulations and approaches convincingly identified *Mirta22* as a
10 miRNA-regulated inhibitory factor of neuronal maturation that does not belong to any previously described class of developmental modulators.

Localization of *Mirta22* in the Golgi apparatus and in vesicle and tubular-like extensions in dendrites is consistent with a role in membrane and protein trafficking and secretion, which is necessary for establishment and maintenance of
15 neuronal connections (28). *Mirta22* is likely to act in concert with other genes within the 22q11.2 deletion (2), including the *Zdhhc8* palmitoyl-transferase, which is also located in the Golgi apparatus and has been shown to modulate dendritic and spine development (9). A potential interaction between miRNA dysregulation and altered neuronal palmitoylation is supported by previous findings (45, 21) and contributions
20 from more than one gene to the morphological phenotypes described in this study are consistent with an oligogenic basis for the psychiatric and cognitive symptoms associated with 22q11.2 microdeletions (2). Moreover, although *Mirta22* represents a major downstream effector of *miR-185* dysregulation, our finding of a coordinated *miR-185* targeting of Golgi apparatus-related genes suggests that *Mirta22*
25 upregulation may act, in an age and brain region specific manner, in concert with other modestly altered *miR-185* targets to interfere with the Golgi-related processes required for neuronal maturation. Thus, our findings highlight a link between the Golgi apparatus and neuronal phenotypes associated with 22q11.2 microdeletions.

The reduction in the levels of *Mirta22* soon after birth during periods
30 of active neuronal maturation and synaptogenesis suggests that repression of this gene may play an important role in promoting neural circuit formation, especially in the postnatal brain, after embryonically generated neurons have migrated to their final destinations. Consistent with the notion that miRNAs function predominantly as fine-tuning regulators of the expression levels of their targets (26, 27), *miR-185* and to a

lesser extent other miRNAs affected by the 22q11.2 deletion appear to restrict and optimize *Mir22* expression, presumably to avoid excessive inhibition during this critical stage of synapse formation. Accordingly, sustained derepression of the gene due to genomic loss at the 22q11.2 locus may have an impact on the formation of

5 neural circuits in early development, as well on their maintenance during adulthood. Such structural changes may result in local and long-distance disruptions of neuronal communication that may contribute to the cognitive dysfunction, psychiatric phenotypes or both (18, 46). In agreement with this prediction expression of the human orthologue of *Mir22* (*C19orf63*) declines in infant brains (40) and displays a

10 spatio-temporal pattern that significantly overlaps with the one of *Neurologin-3*, consistent with participation in processes related to synapse and circuit formation and maturation (47). Moreover, it has been shown that during the transition between human fetal and early postnatal development a large number of the genes reverse their direction of expression from an increase *in utero* to a decrease in the months after

15 birth and that ~40% of them are predicted miRNA targets (40). In that respect, *Mir22* is one of the first examples of a disease-related gene representative of this prominent type of transcriptional trajectory indicative of a miRNA-imposed inhibitory control over postnatal brain development.

Accumulating evidence suggests that miRNAs play an important role

20 in the pathogenesis and pathophysiology of psychiatric disorders and cognitive dysfunction (29,14). Our work represents one of the first examples where a major downstream target of a disease-related miRNA dysregulation has been unequivocally identified and its function has been characterized. It will be of interest to determine whether elevation in *Mir22* levels affect additional aspects of neuronal structure and

25 function. Future studies, entailing normalization of *Mir22* levels in mice compound heterozygous for *Df(16)A* and *Mir22* loss-of-function alleles should also establish which of the various behavioral (8), cognitive (48, 8) and circuit alterations (18, 46) observed in *Df(16)A^{+/-}* mice can be attributed to the inhibitory influence of *Mir22* upregulation. In addition, since inhibition of *Mir22* function is predicted to promote

30 formation of neuronal connections in the 22q11.2 deletion carriers, drugs that target this molecule may be used to ameliorate associated cognitive and psychiatric phenotypes.

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25 Various references are cited herein, the contents of which are hereby
incorporated by reference in their entireties.

WHAT IS CLAIMED IS:

1. A method for increasing connectivity between neurons comprising administering to said neurons an effective amount of an agent that inhibits Mirta22 activity.
- 5 2. The method of claim 1, where the agent is mir-185.
3. The method of claim 1, where the agent is selected from the group consisting of antisense RNA comprising a sequence complementary to the Mirta22 mRNA; RNAi comprising a sequence complementary to the Mirta22 mRNA, and shRNA comprising a sequence complementary to the Mirta22 mRNA.
- 10 4. The method of claim 1, where the agent is selected from the group consisting of an anti-Mirta22 antibody or fragment thereof, a single chain antibody that specifically binds to Mirta22, or a diabody that specifically binds to Mirta22.
5. A method for increasing connectivity between neurons comprising administering to said neurons an effective amount of an agent that increases mir-185 activity.
- 15 6. The method of claim 5, where the agent is a mir-185 nucleic acid.
7. A method for increasing connectivity between neurons in a subject in need of such treatment comprising administering to said subject an effective amount of an agent that inhibits Mirta22 activity.
- 20 8. The method of claim 7, where the agent is a mir-185 nucleic acid.
9. The method of claim 7, where the agent is selected from the group consisting of antisense RNA comprising a sequence complementary to the Mirta22 mRNA; RNAi comprising a sequence complementary to the Mirta22 mRNA, and shRNA comprising a sequence complementary to the Mirta22 mRNA.
- 25 10. The method of claim 7, where the agent is selected from the group consisting of an anti-Mirta22 antibody or fragment thereof, a single chain antibody that specifically binds to Mirta22, or a diabody that specifically binds to Mirta22.
11. A method for increasing connectivity between neurons in a subject in need of such treatment comprising administering to said subject an effective amount of an agent that increases mir-185 activity.
- 30 12. The method of claim 5, where the agent is a mir-185 nucleic acid.

13. A method of treating schizophrenia or a related disorder in a subject, comprising administering to said subject an effective amount of an agent that inhibits Mirta22 activity.
14. The method of claim 13, where the agent is a mir-185 nucleic acid.
- 5 15. The method of claim 13, where the agent is selected from the group consisting of antisense RNA comprising a sequence complementary to the Mirta22 mRNA; RNAi comprising a sequence complementary to the Mirta22 mRNA, and shRNA comprising a sequence complementary to the Mirta22 mRNA.
- 10 16. A method of treating schizophrenia or a related disorder in a subject, comprising administering to said subject an effective amount of an agent that increases mir-185 activity.
17. The method of claim 16, where the agent is a mir-185 nucleic acid.
18. A method of enhancing memory in a subject, comprising administering to said subject an effective amount of an agent that inhibits Mirta22 activity.
- 15 19. The method of claim 18, where the agent is a mir-185 nucleic acid.
20. The method of claim 18, where the agent is selected from the group consisting of antisense RNA comprising a sequence complementary to the Mirta22 mRNA; RNAi comprising a sequence complementary to the Mirta22 mRNA, and shRNA comprising a sequence complementary to the Mirta22 mRNA.
- 20 21. A method of enhancing memory in a subject, comprising administering to said subject an effective amount of an agent that increases mir-185 activity.
22. The method of claim 21, where the agent is a mir-185 nucleic acid.

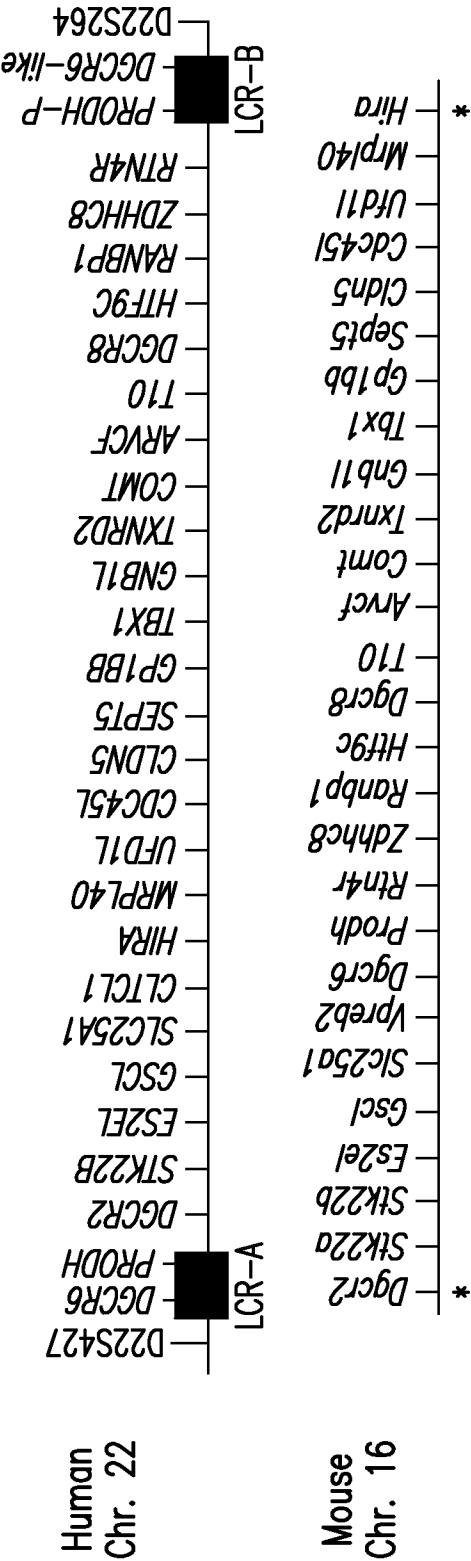


FIG. 1A

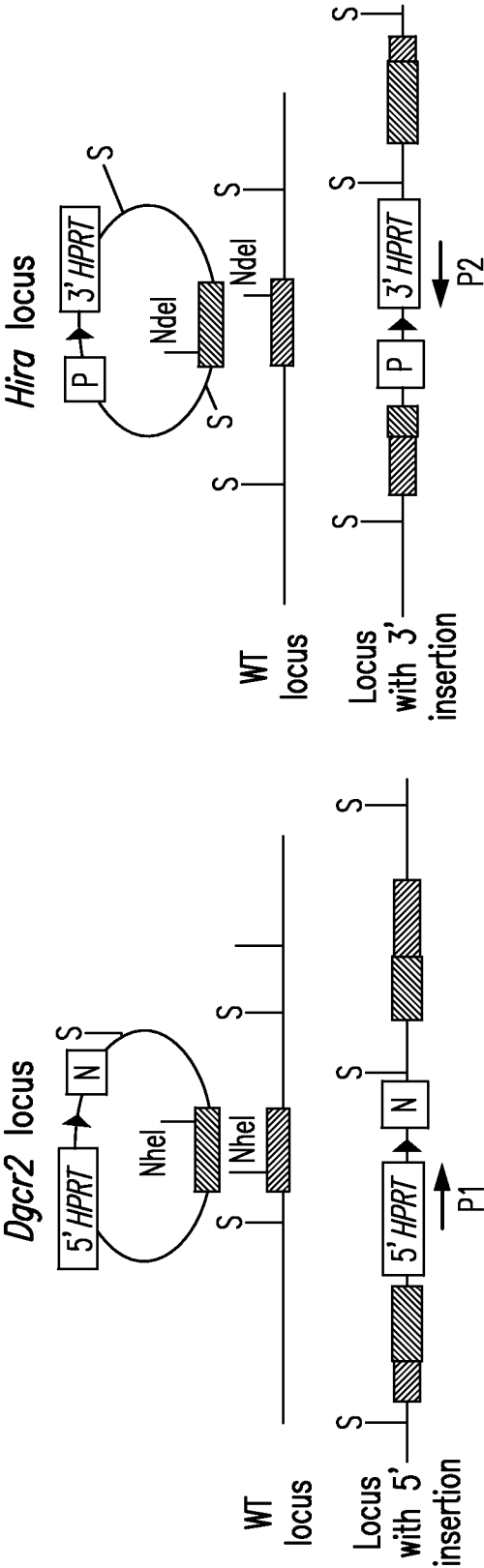


FIG. 1B

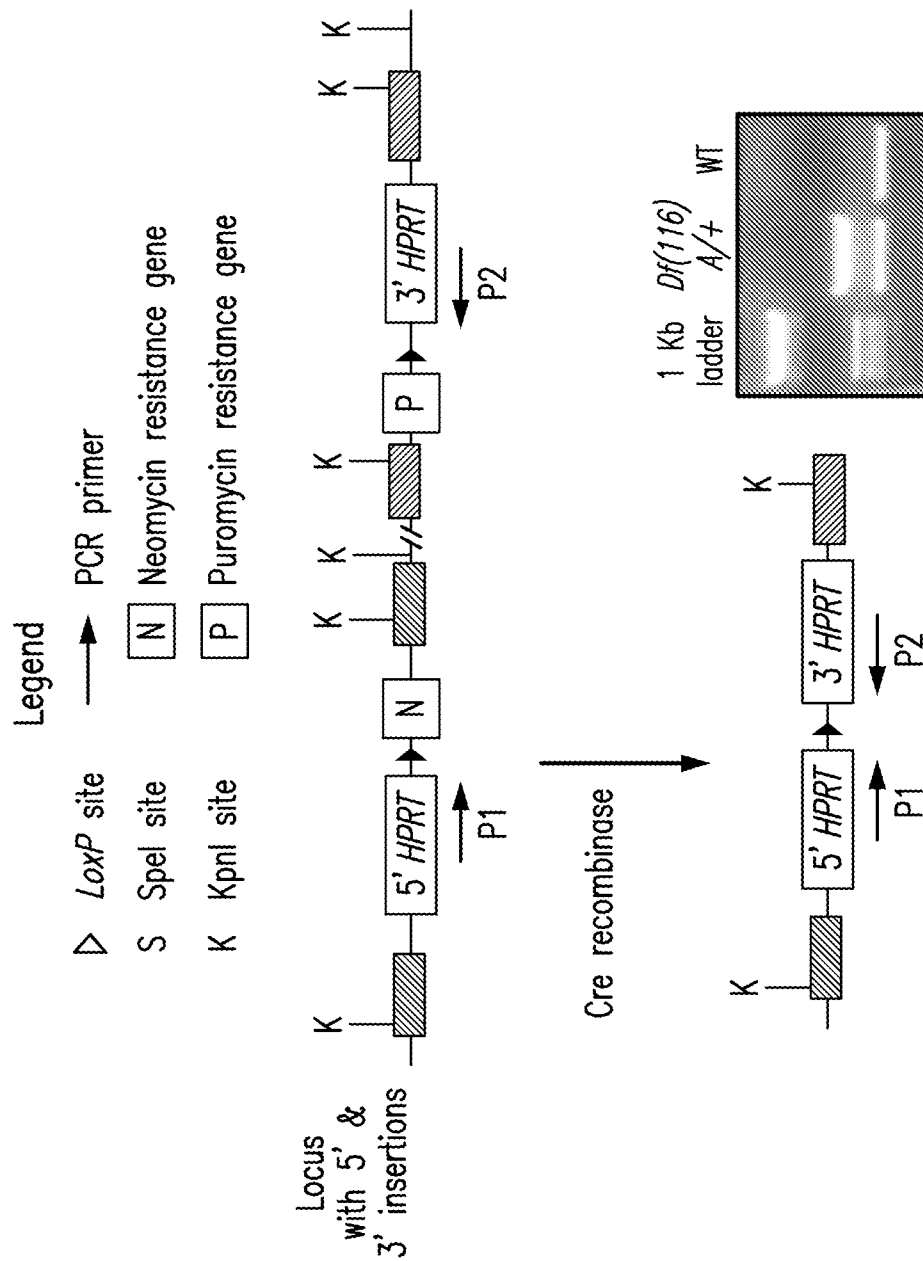
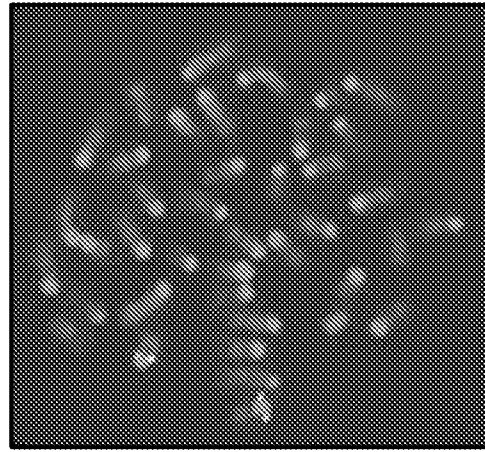


FIG. 1C

FIG. 1D



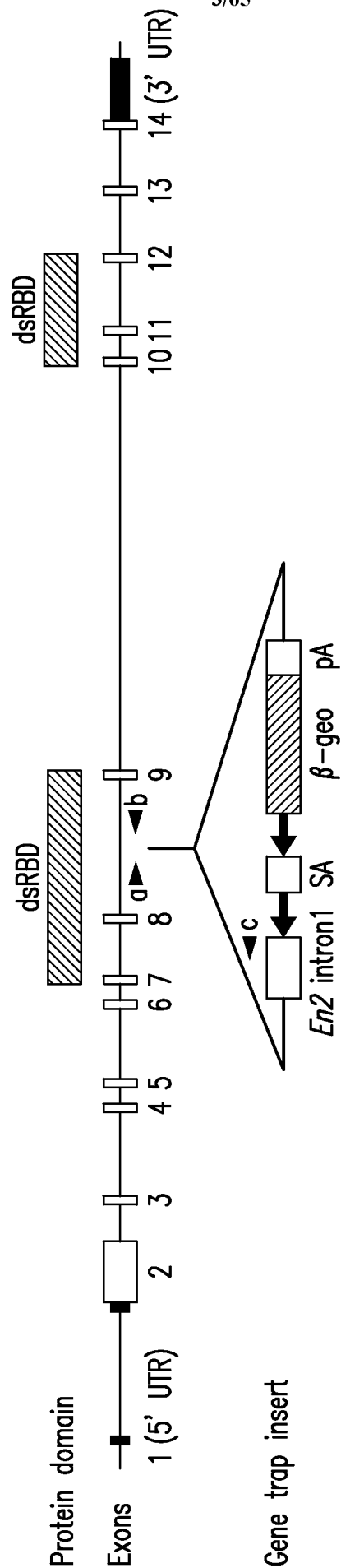


FIG. 2A

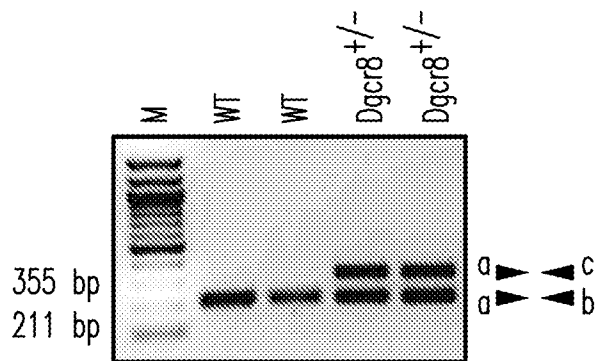


FIG. 2B

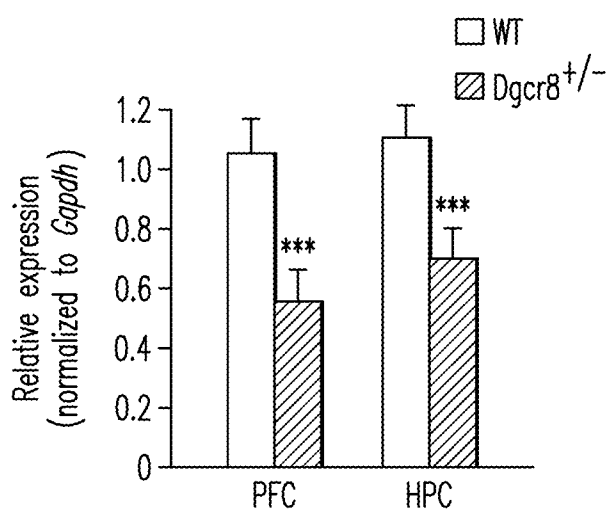


FIG. 2D

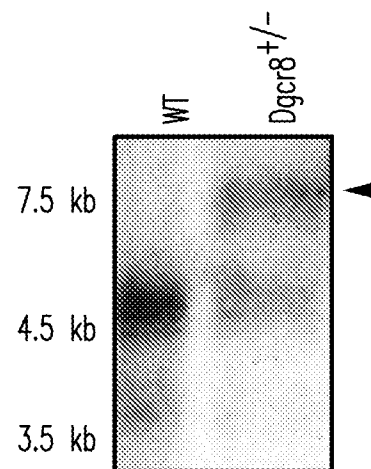


FIG. 2C

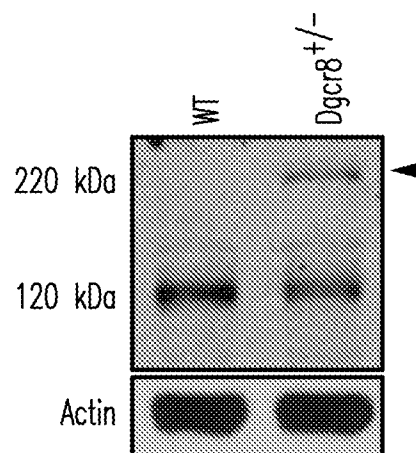


FIG. 2E

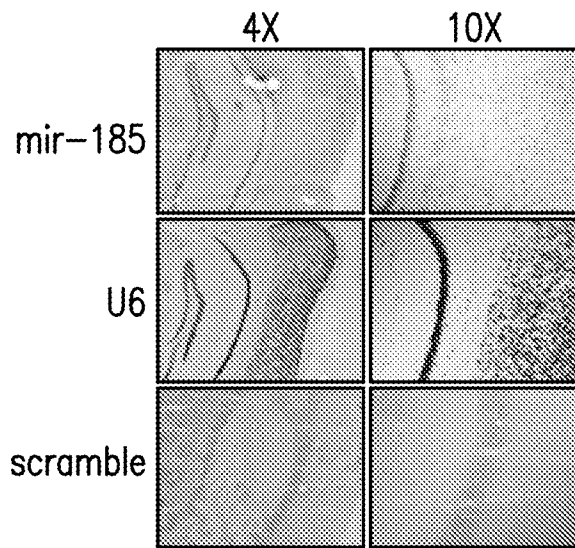


FIG. 3A

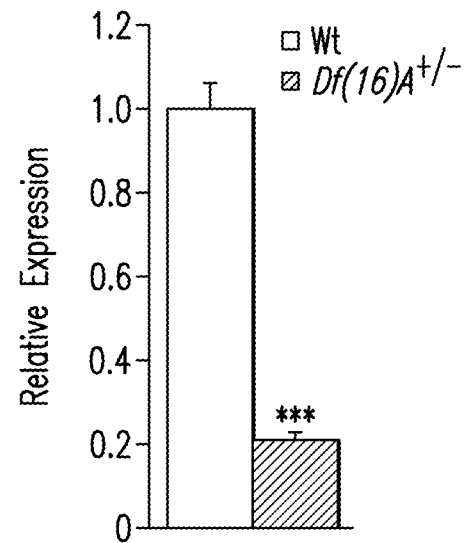


FIG. 3B

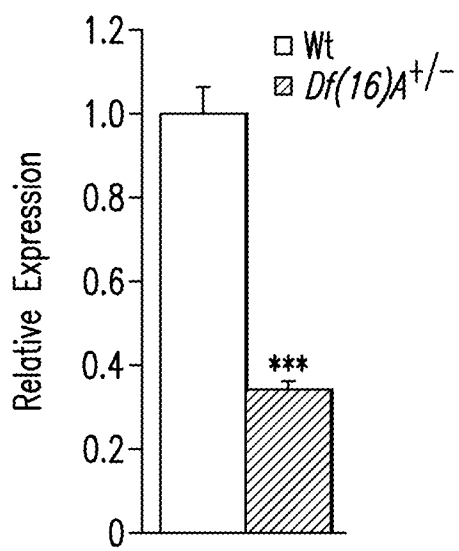


FIG. 3C

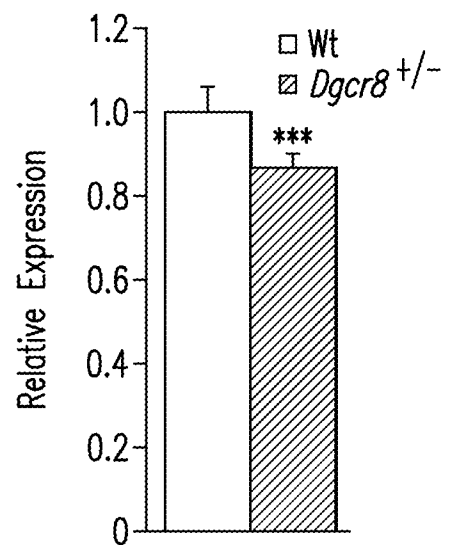


FIG. 3D

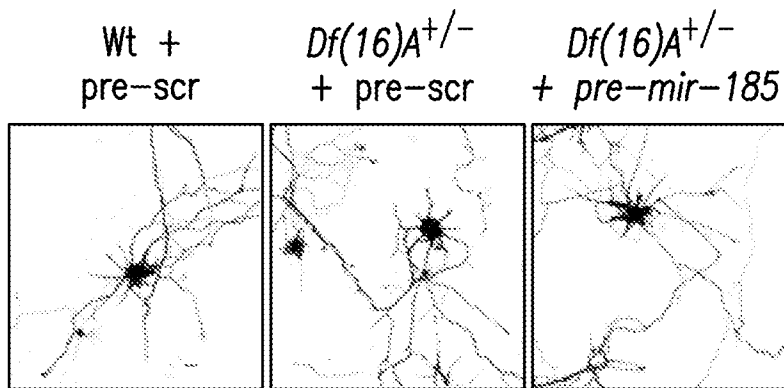


FIG. 3E

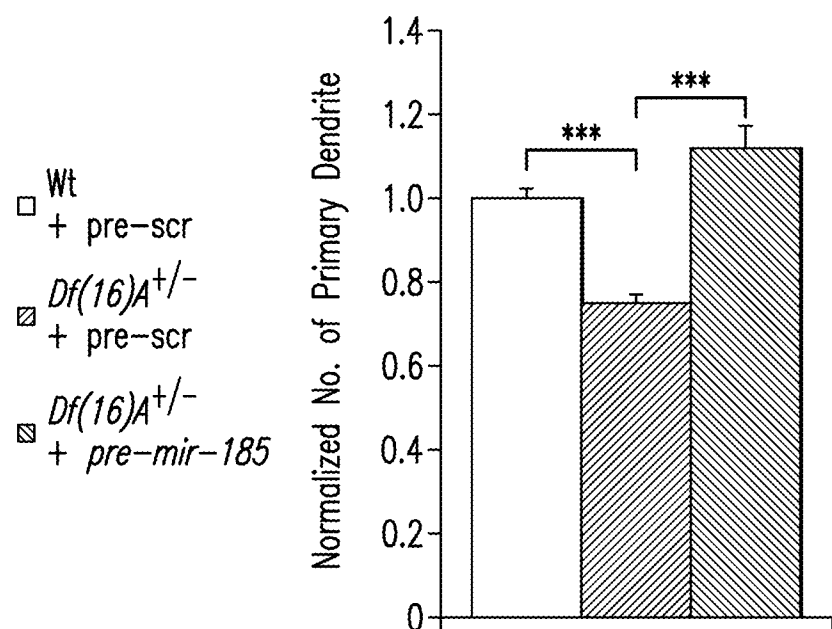


FIG. 3F

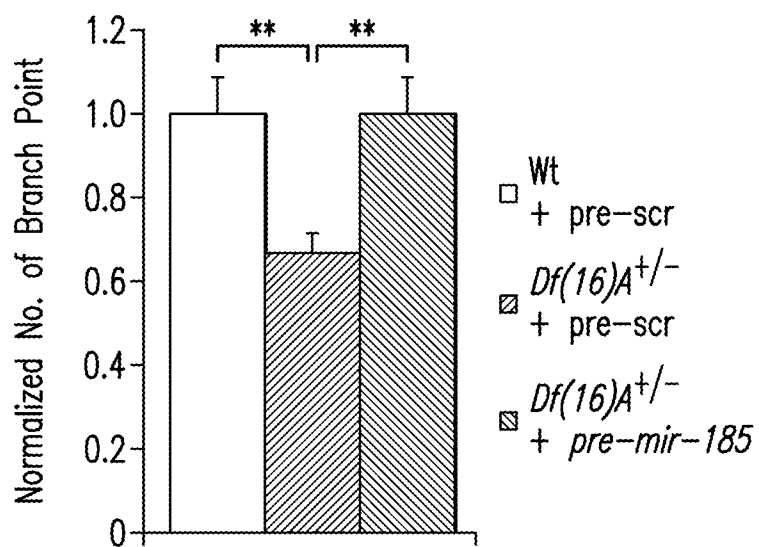


FIG. 3G

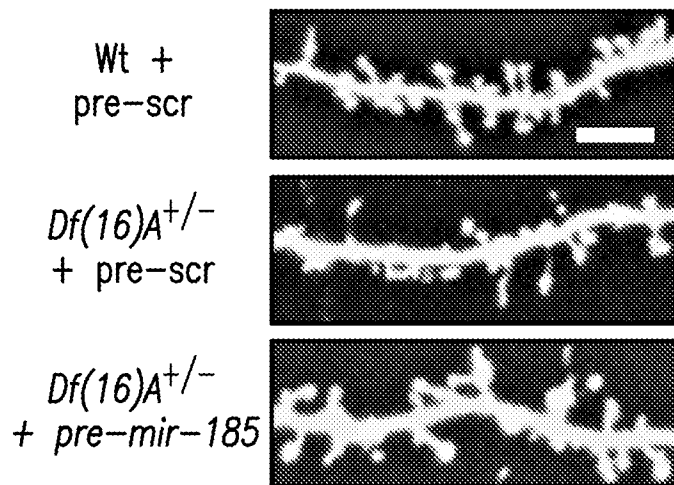


FIG. 3H

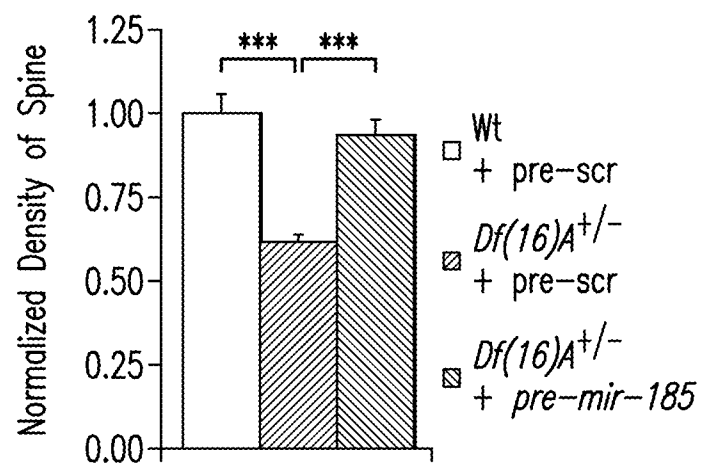


FIG. 3I

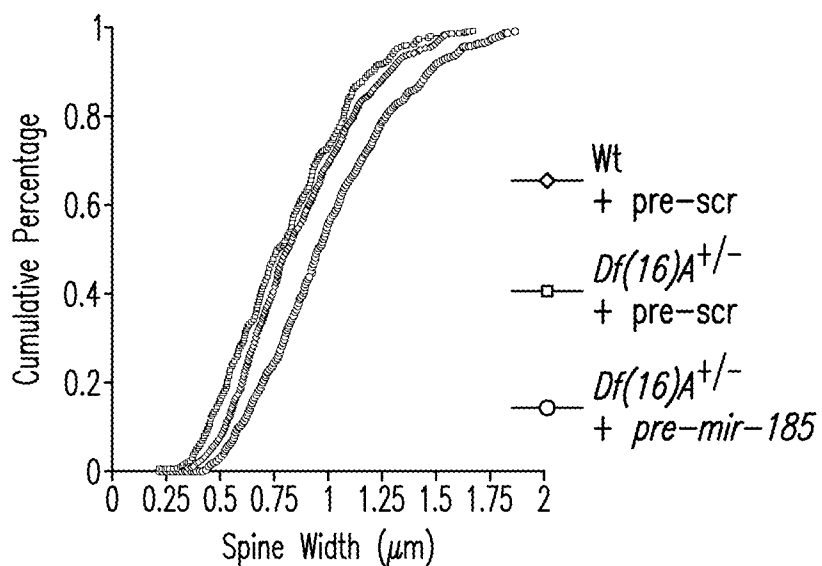


FIG. 3J

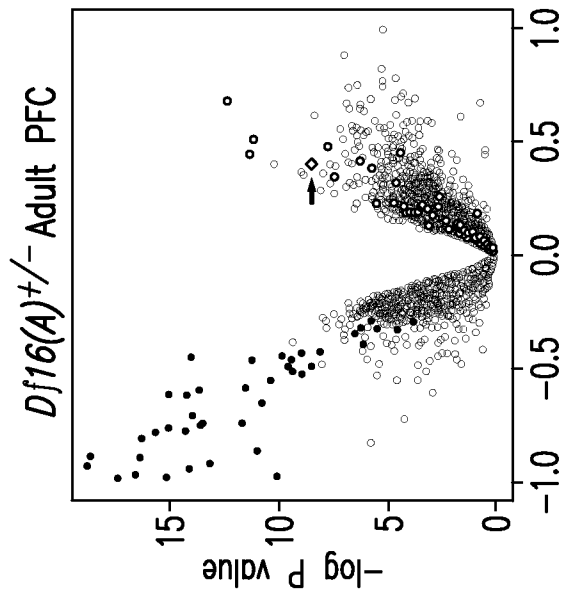


FIG. 4A-3

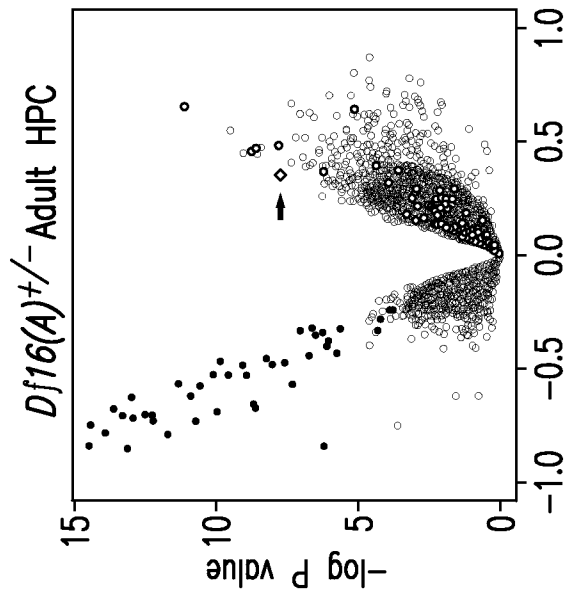


FIG. 4A-6

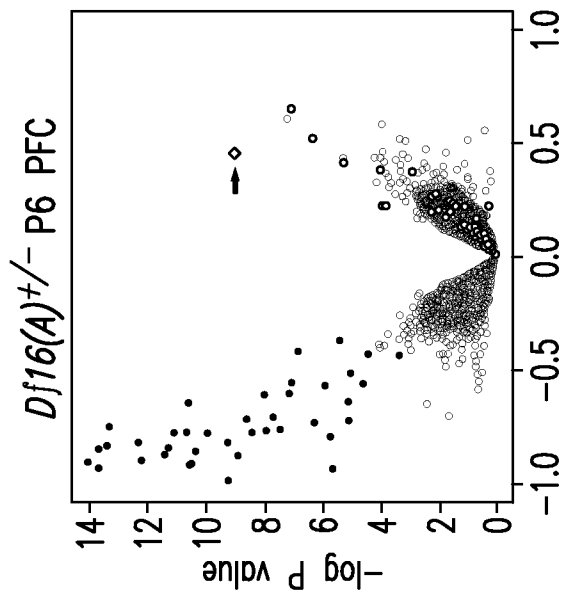


FIG. 4A-2

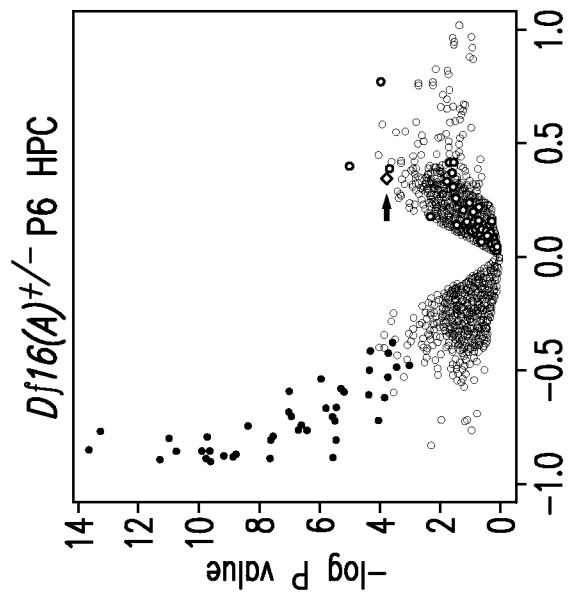


FIG. 4A-5

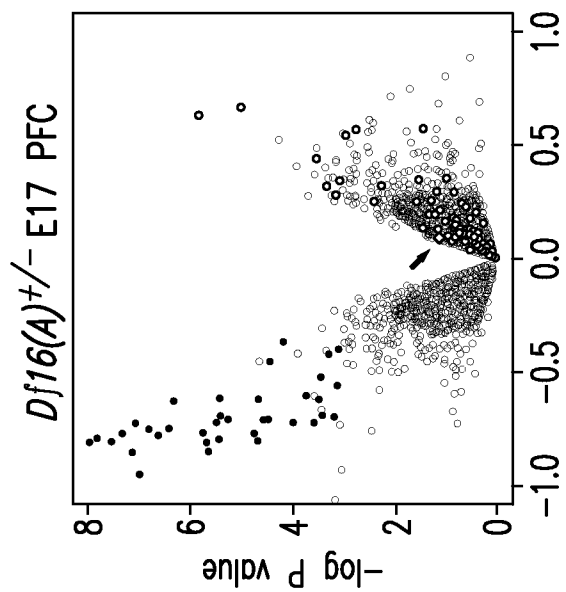


FIG. 4A-1

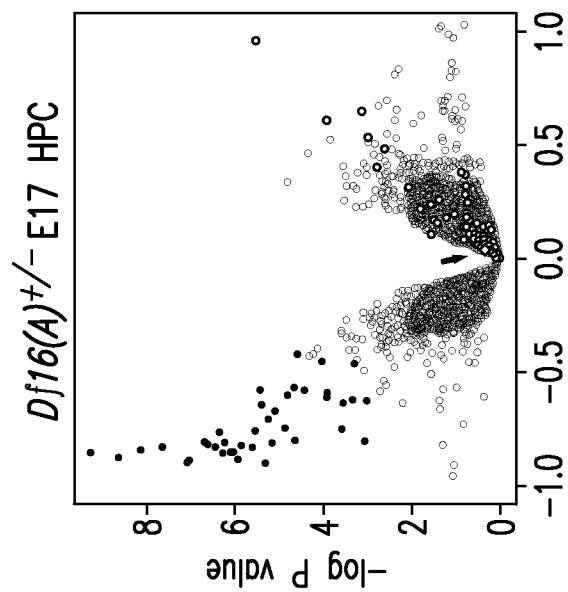


FIG. 4A-4

E17 PFC			P6 PFC			Adult PFC		
AffyID	Symbols	P.Value	AffyID	Symbols	P.Value	AffyID	Symbols	P.Value
1450625_at	Col5a2	1.15E-04	1424038_a_at	2310044H10Rik	9.08E-10	1455326_at	Clec16a	1.38E-09
1430644_at	Wbscr25	1.92E-04	1428021_at	Mccc2	5.82E-05	1424038_a_at	2310044H10Rik	3.40E-09
1419147_at	Rec8	4.30E-04	1430976_a_at	Mrpl9	6.26E-05	1456488_at	Wdr33	4.36E-09
1419028_at	Arpp21	4.49E-04	1438967_x_at	Amhr2	9.71E-05	1428208_at	Bc17a	1.06E-08
1427083_a_at	Map4k5	4.79E-04	1427962_at	Ccdc102a	1.03E-04	1437125_at	Carmk2a	3.73E-08
1440240_at	Npb	6.16E-04	1437236_a_at	Zfp110	1.49E-04	1439661_at	Slc16a14	3.76E-08
1421648_at	Nlgn1	6.30E-04	1451688_s_at	Cant1	1.76E-04	1426446_at	6430548M08Rik	6.03E-08
1424659_at	Slit2	8.03E-04	1453380_a_at	Xrcc6bp1	1.77E-04	1458870_x_at	Mycbp2	1.04E-07
1458119_at	Slc25a27	8.84E-04	1420117_at	Golph3	2.04E-04	1452322_a_at	Brwd1	1.32E-07
1449914_at	Ribc1	9.42E-04	1431678_at	4930556N08Rik	2.32E-04	1450661_x_at	Nfic	1.61E-07
E17 HPC			P6 HPC			Adult HPC		
AffyID	Symbols	P.Value	AffyID	Symbols	P.Value	AffyID	Symbols	P.Value
1450625_at	Col5a2	4.40E-05	1460613_x_at	Gh	8.98E-05	1455326_at	Clec16a	8.58E-10
1460245_at	Klrd1	2.84E-04	1424038_a_at	2310044H10Rik	1.68E-04	1423125_at	Dclk1	2.98E-09
1460451_at	Tmem52	2.96E-04	1418547_at	Tfpi2	2.42E-04	1425690_at	B3gat1	3.94E-09
1453816_at	Nsa2	4.81E-04	1460471_at	Ooep	3.33E-04	1424038_a_at	2310044H10Rik	1.81E-08
1420170_at	Myh9	5.38E-04	1418541_at	Cenpo	3.44E-04	1441312_at	Cnnm1	4.33E-08
1434423_at	Gulp1	5.72E-04	1427202_at	4833442J19Rik	4.12E-04	1449931_at	Cpeb4	4.44E-08
1442160_at	Fam19a3	5.73E-04	1438624_x_at	Hs3st2	4.66E-04	1421200_at	Dlg2	9.01E-08
1422263_at	Bdkrb2	8.09E-04	1421298_a_at	Hipk1	4.75E-04	1421339_at	Extl3	1.10E-07
1416529_at	Emp1	8.99E-04	1449191_at	Wfdc12	4.87E-04	1431254_at	Kbtbd11	1.40E-07
1418710_at	Cd59a	9.23E-04	1454576_at	A230102021Rik	5.02E-04	1452742_at	Trak1	1.45E-07

FIG. 4B

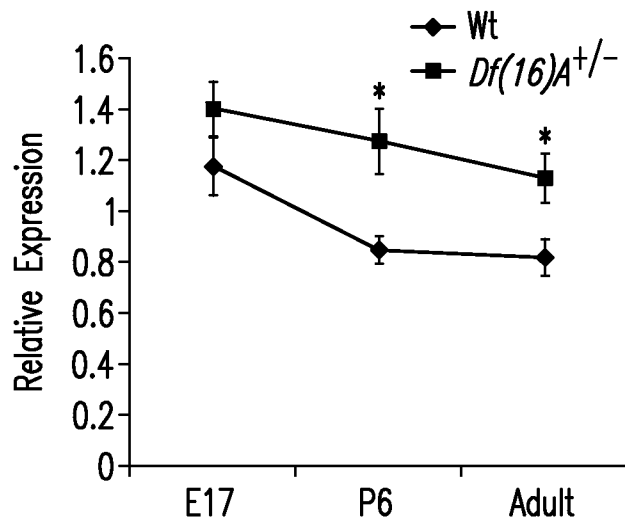


FIG. 4C

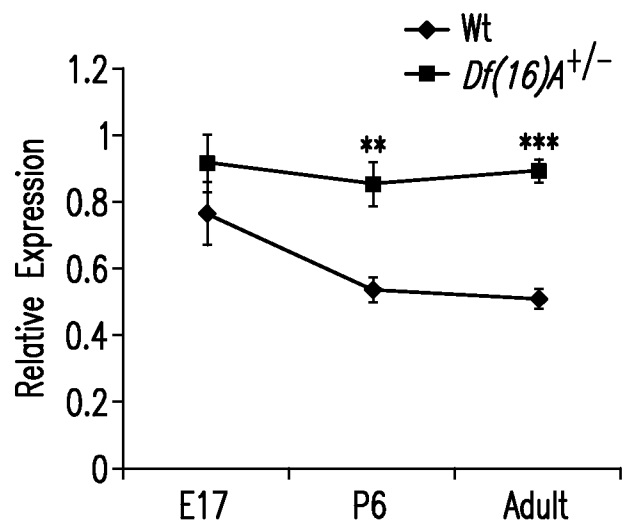


FIG. 4D

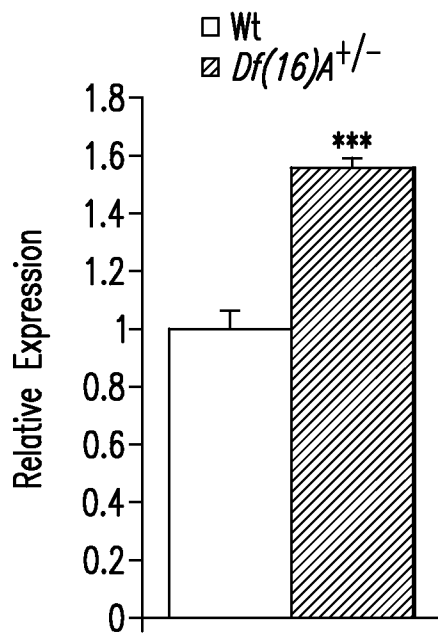


FIG. 4E

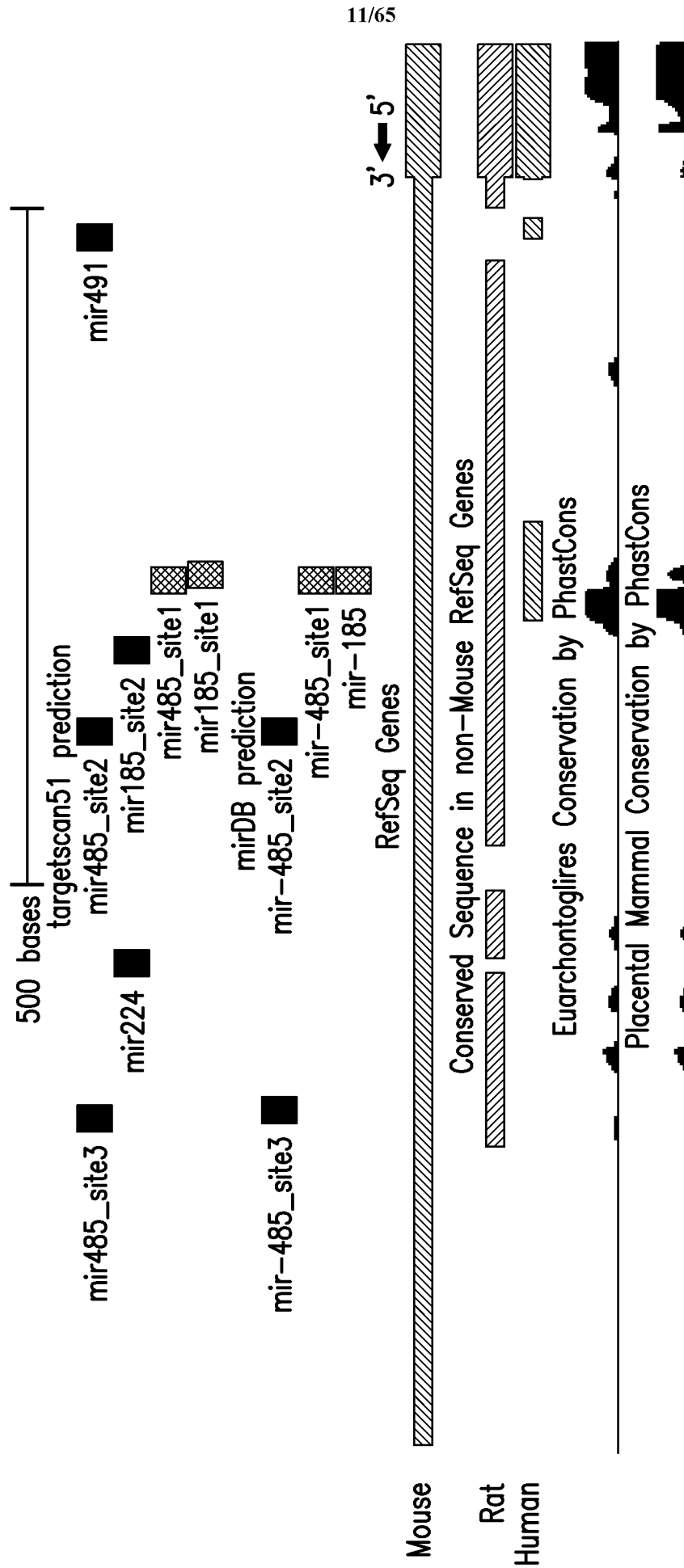


FIG. 5A

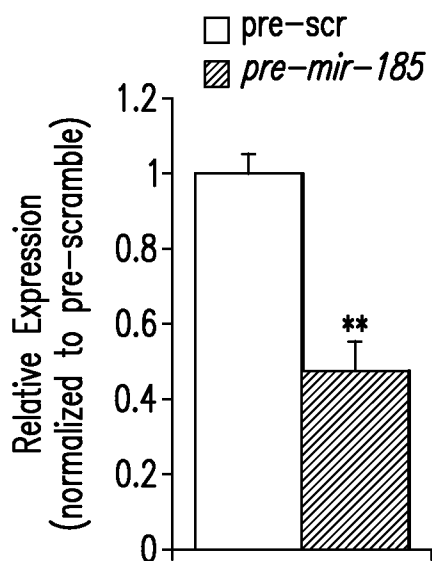


FIG. 5B

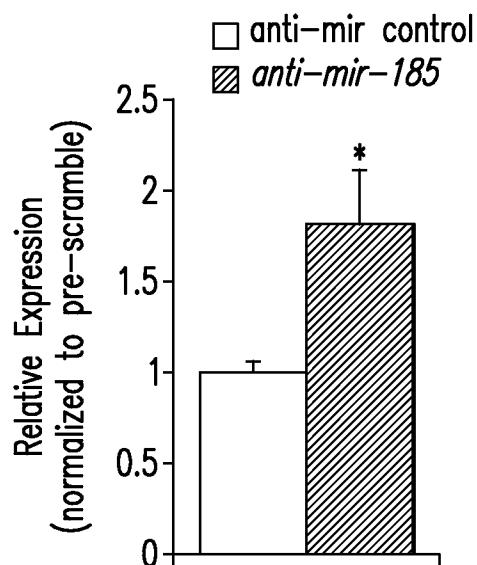


FIG. 5C

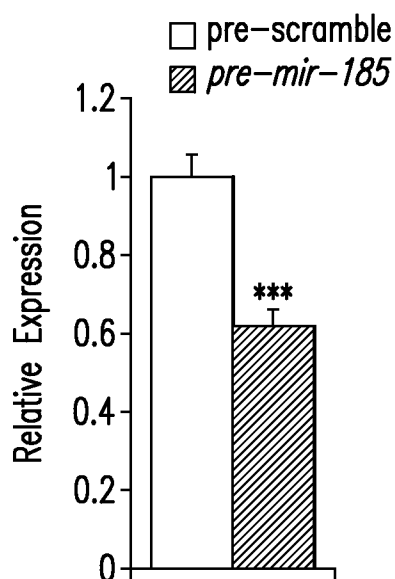


FIG. 5D

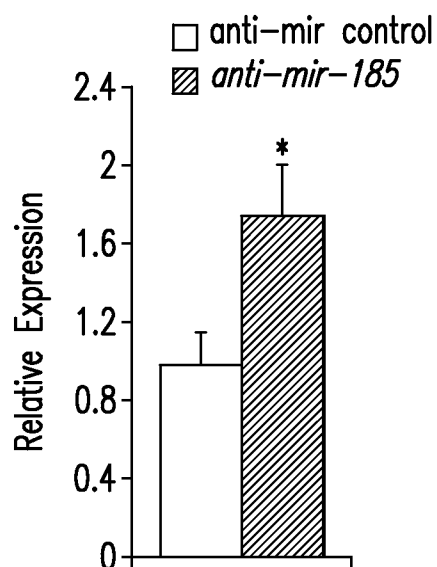


FIG. 5E

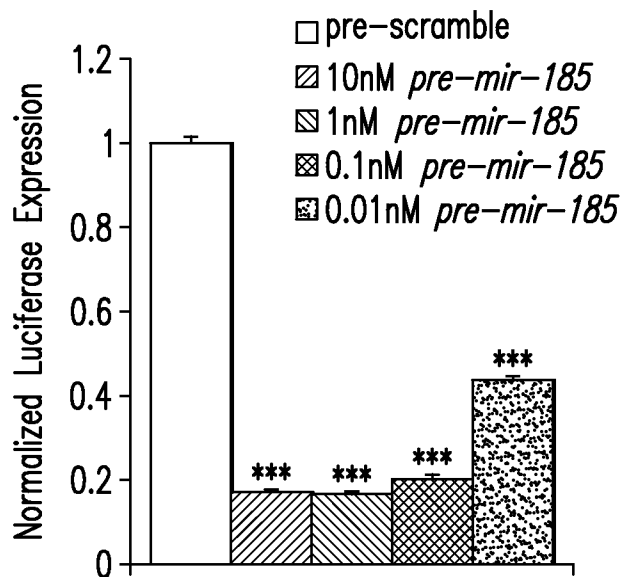


FIG. 5F

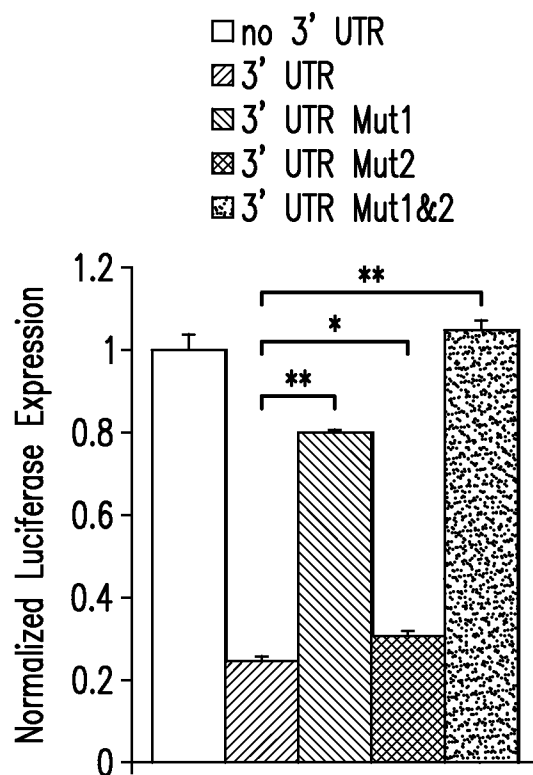


FIG. 5G

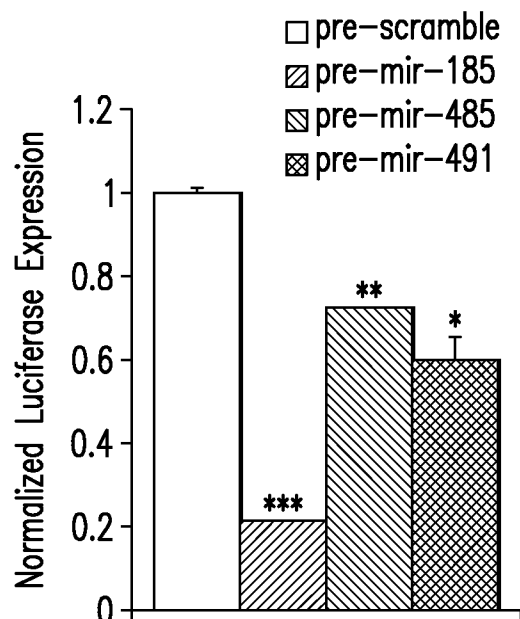


FIG. 5H

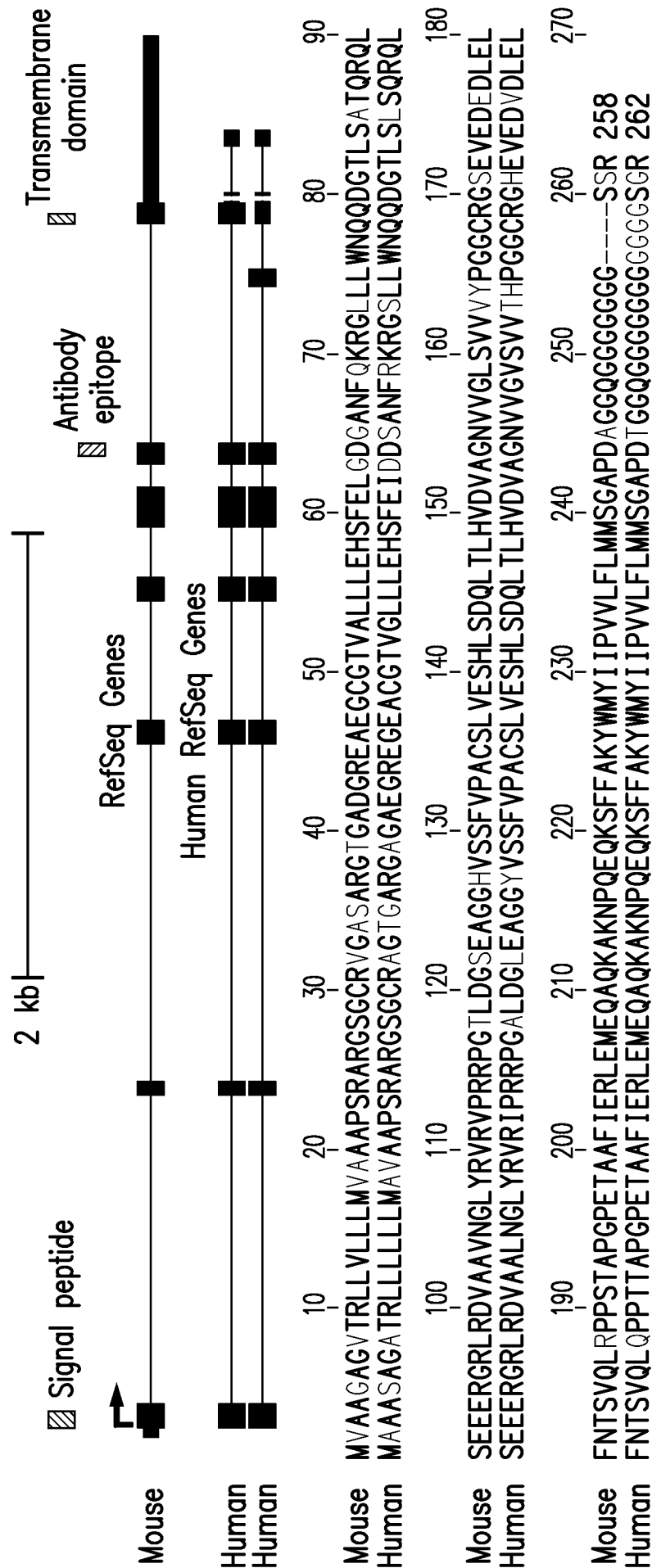


FIG. 6A

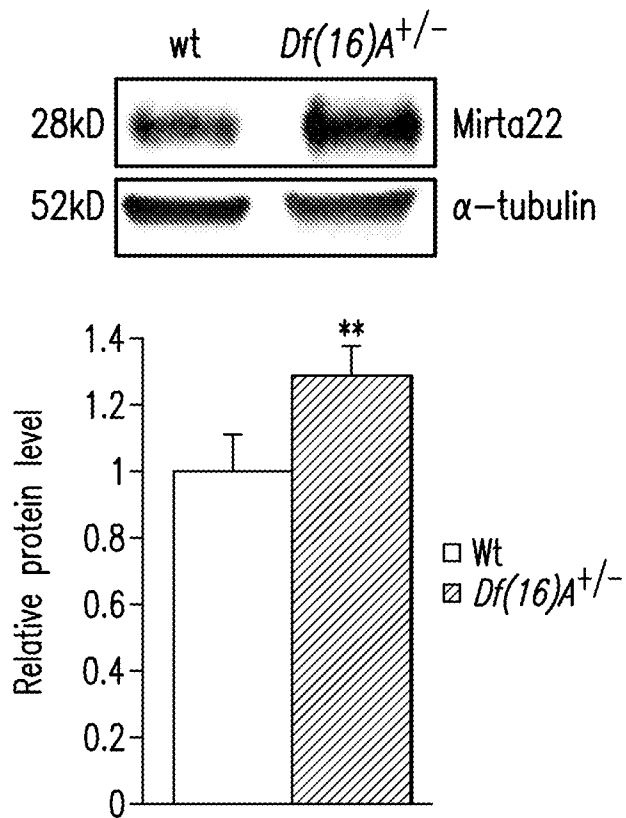


FIG. 6B

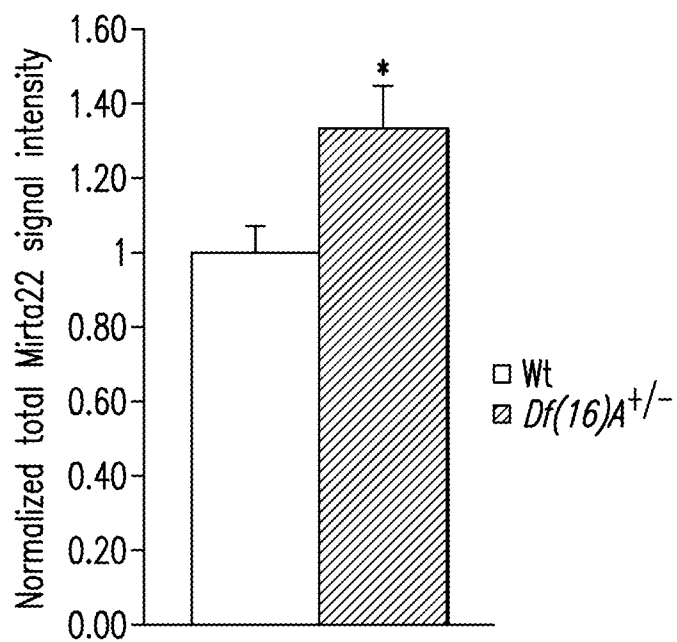


FIG. 6C

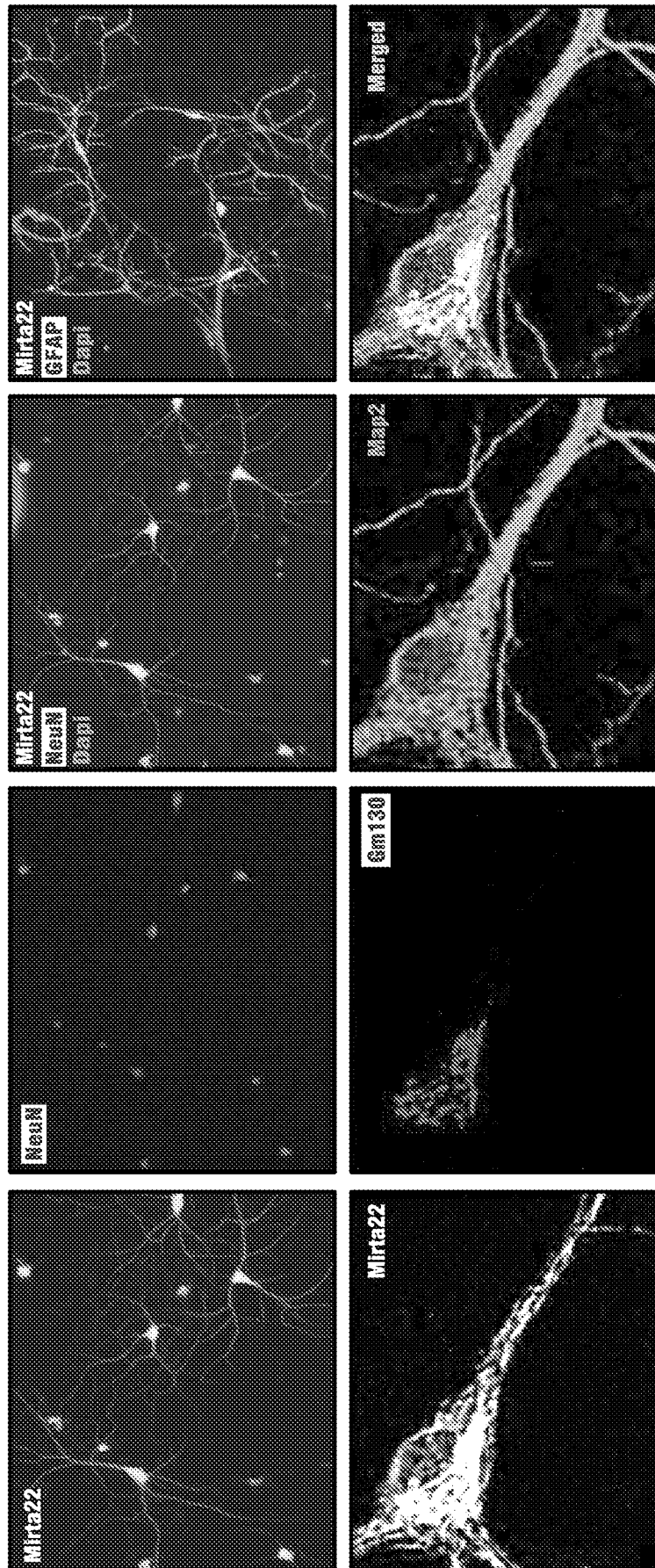


FIG. 6D

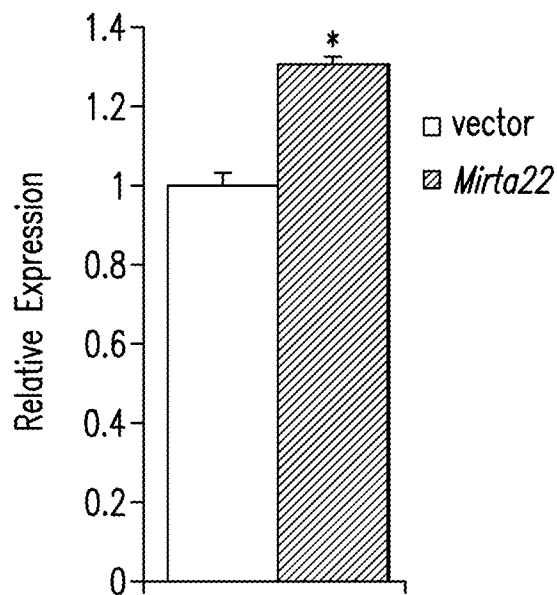


FIG. 7A

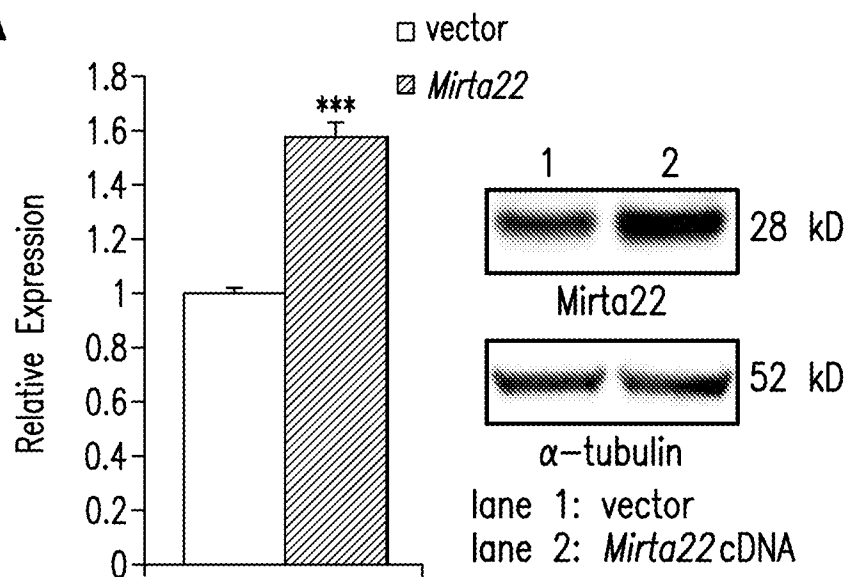


FIG. 7B

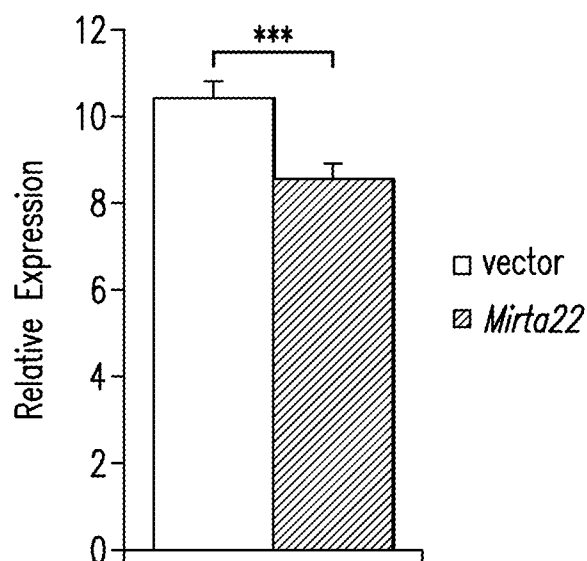


FIG. 7C

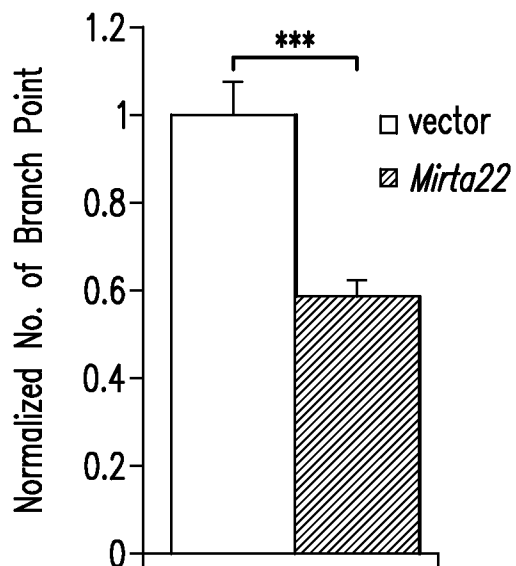


FIG. 7D

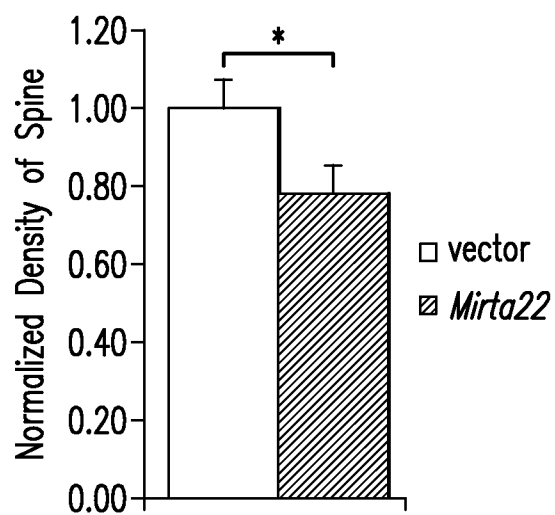


FIG. 7E

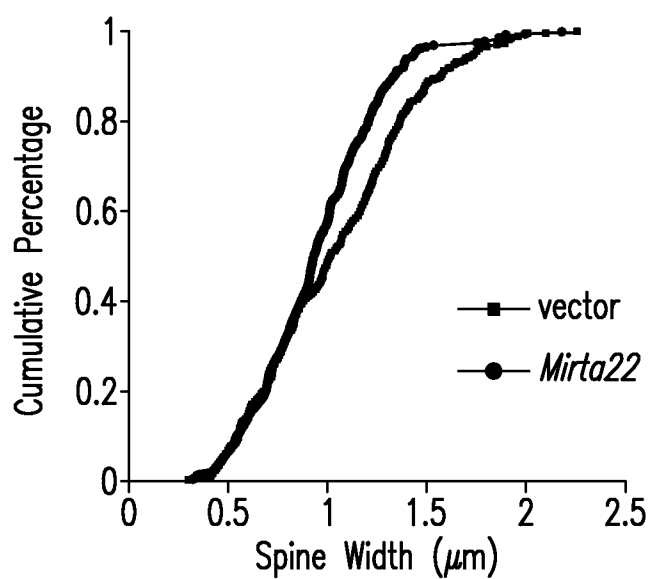


FIG. 7F

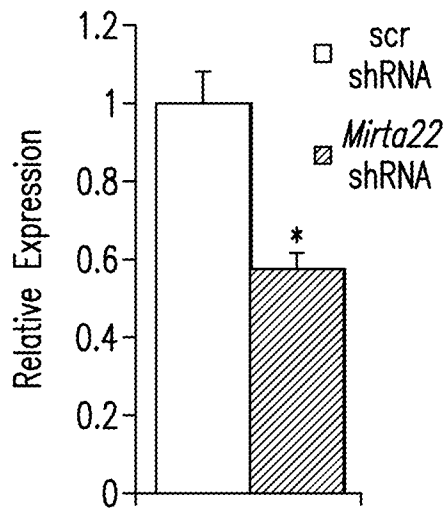


FIG. 7G

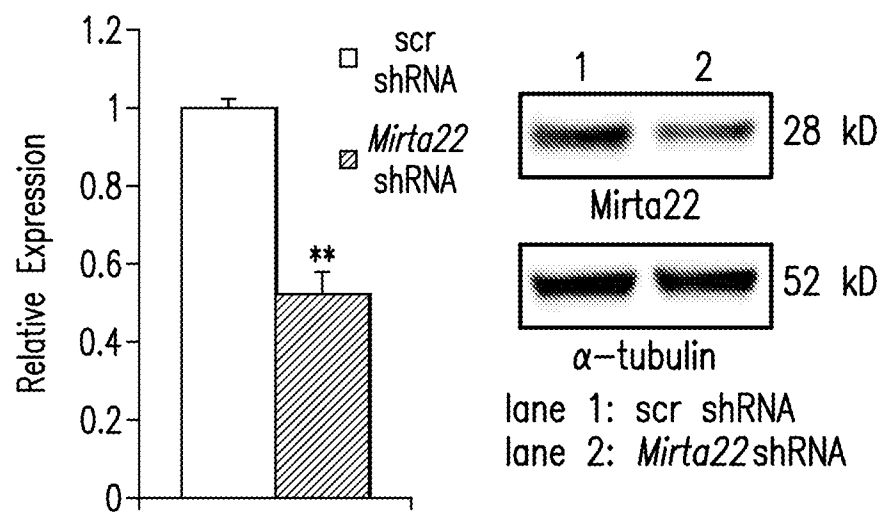


FIG. 7H

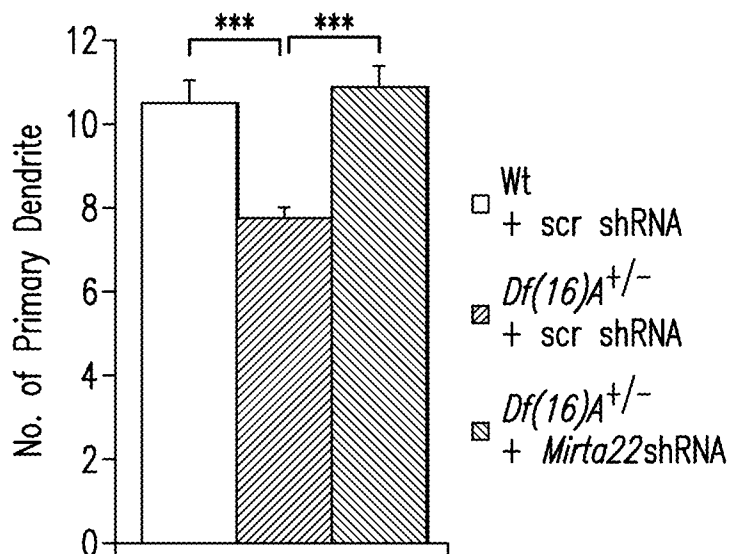


FIG. 7I

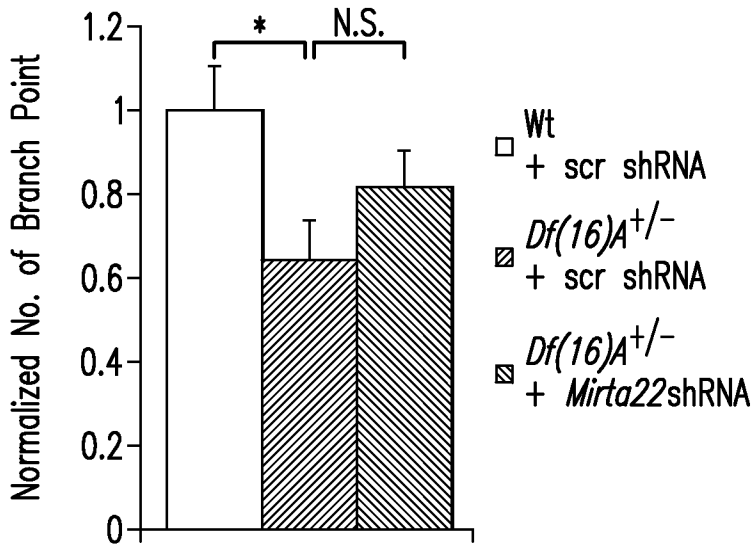


FIG. 7J

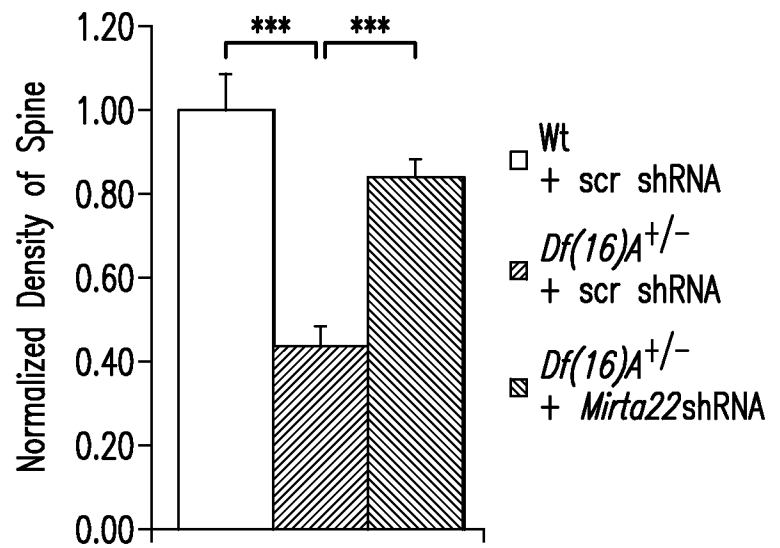


FIG. 7K

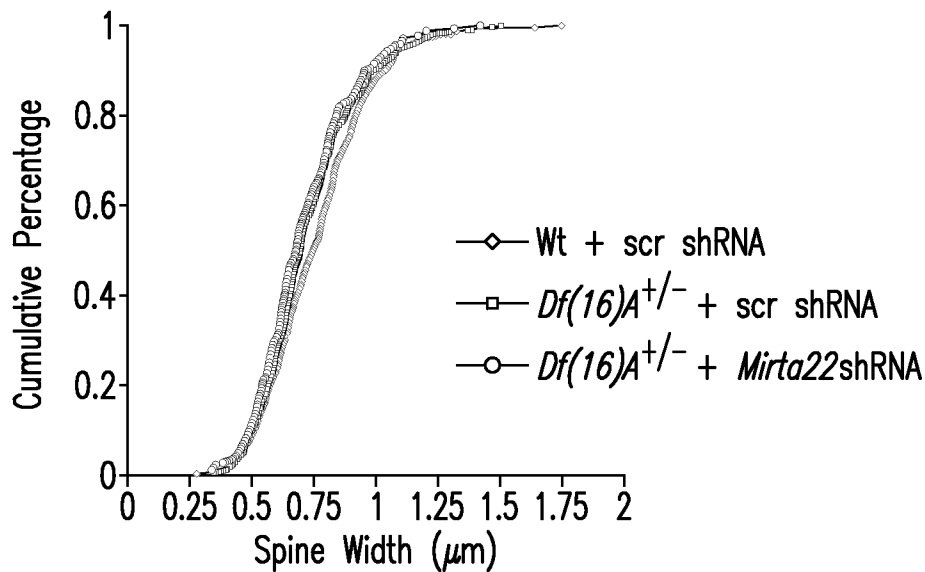


FIG. 7L

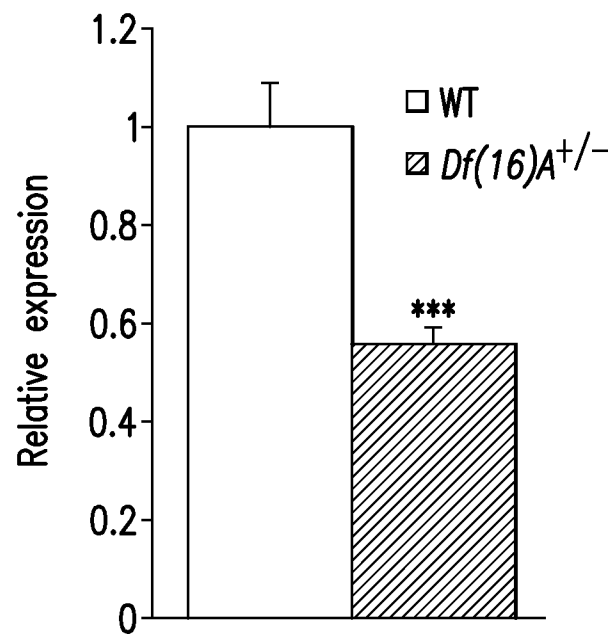


FIG. 8A

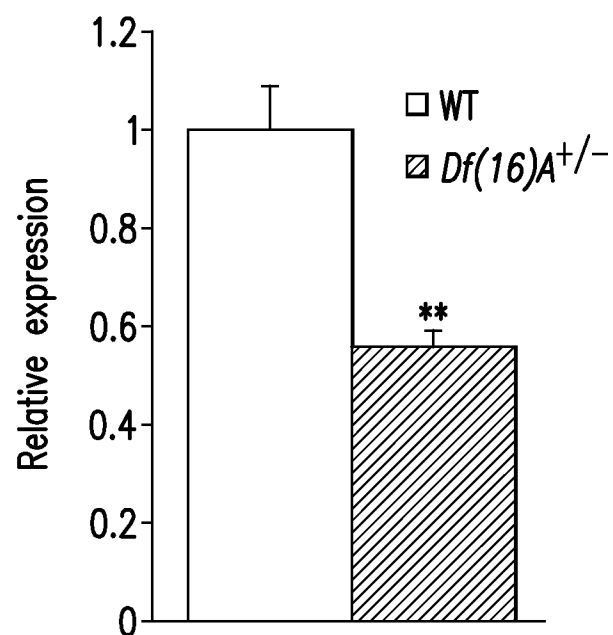


FIG. 8B

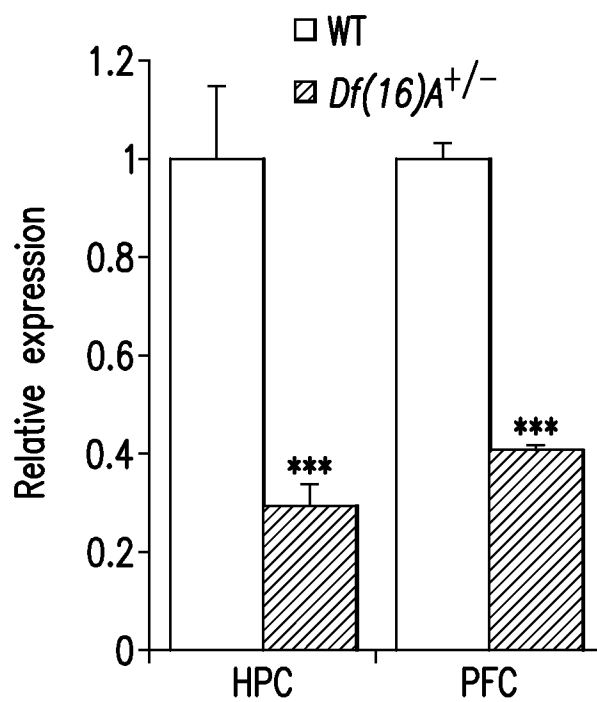


FIG. 9A

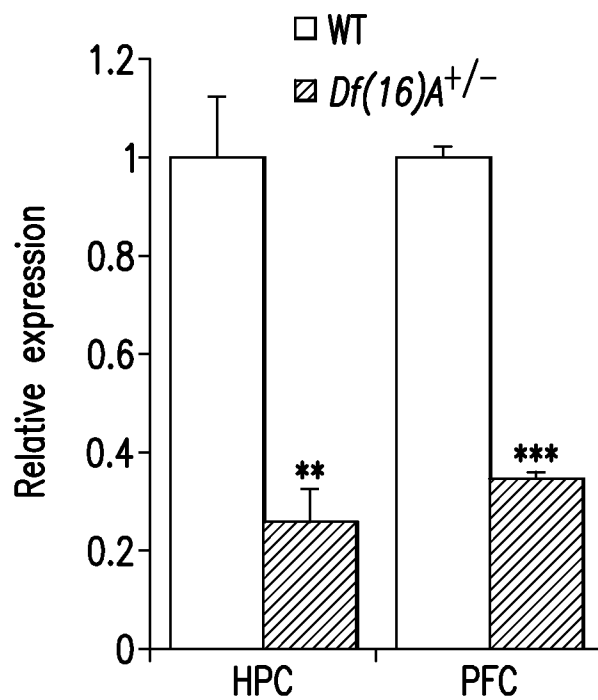


FIG. 9B

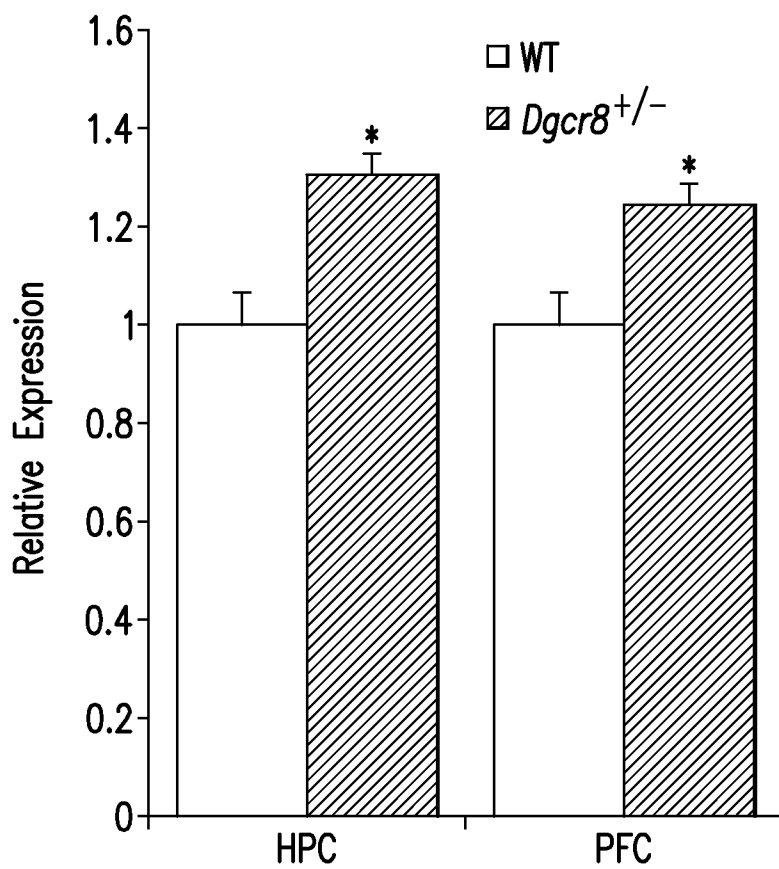


FIG. 10

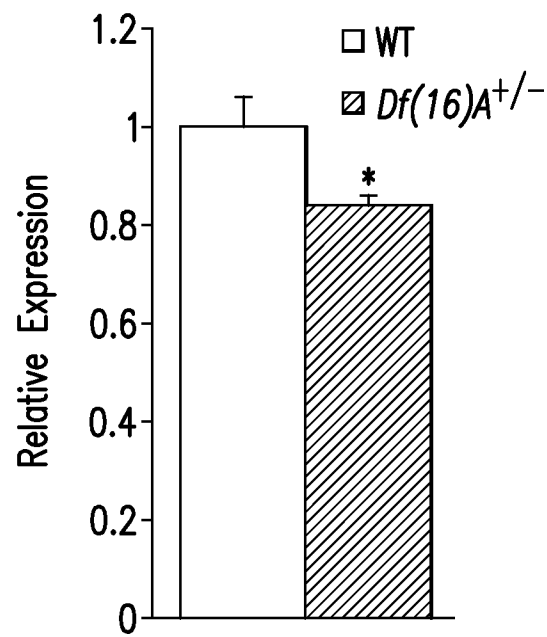


FIG. 11A

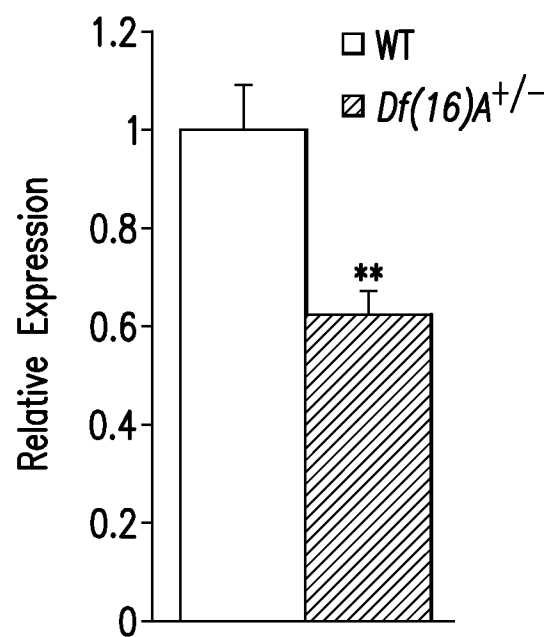


FIG. 11B

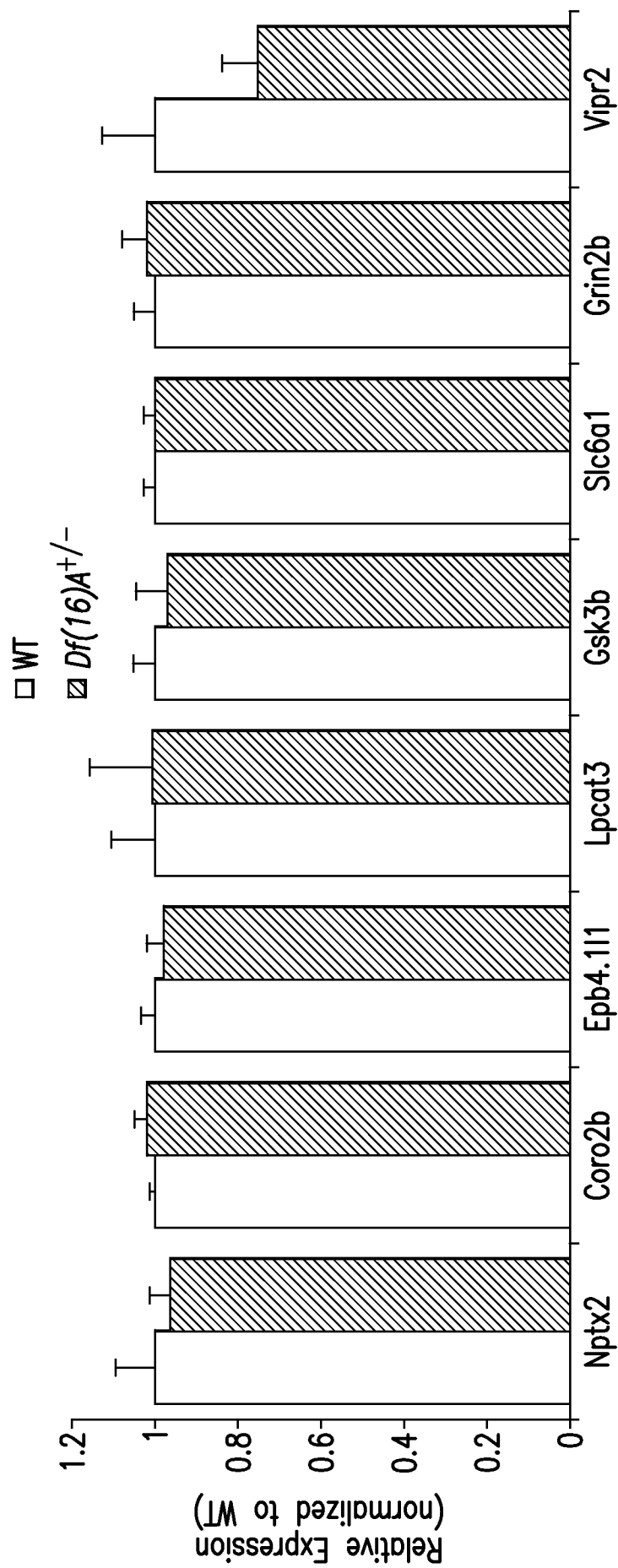


FIG. 12

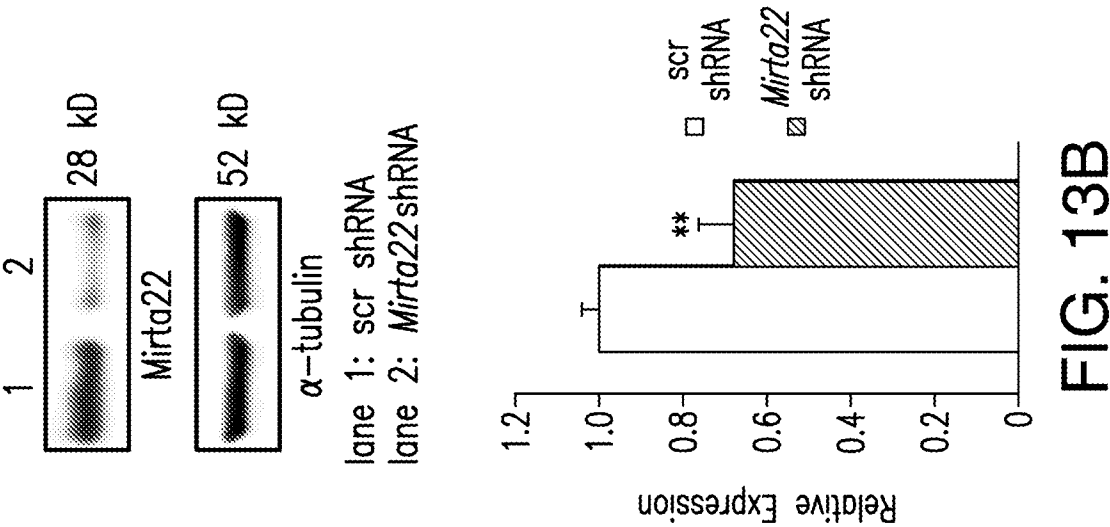


FIG. 13B

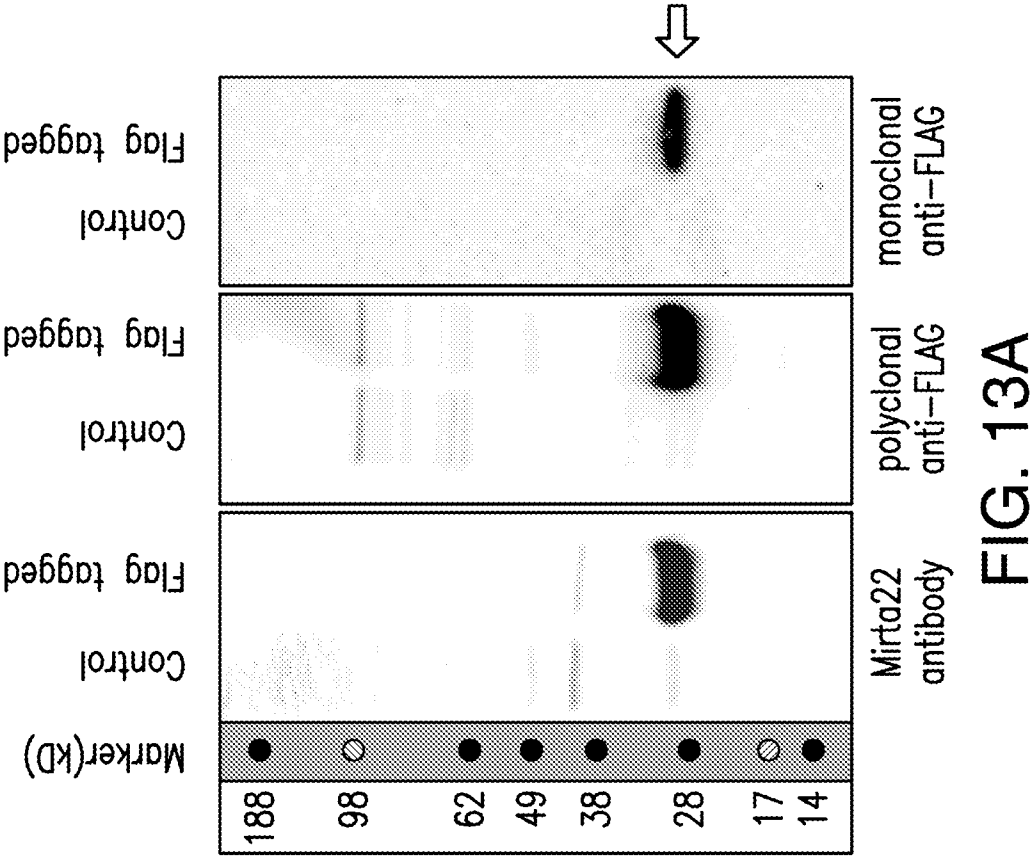


FIG. 13A

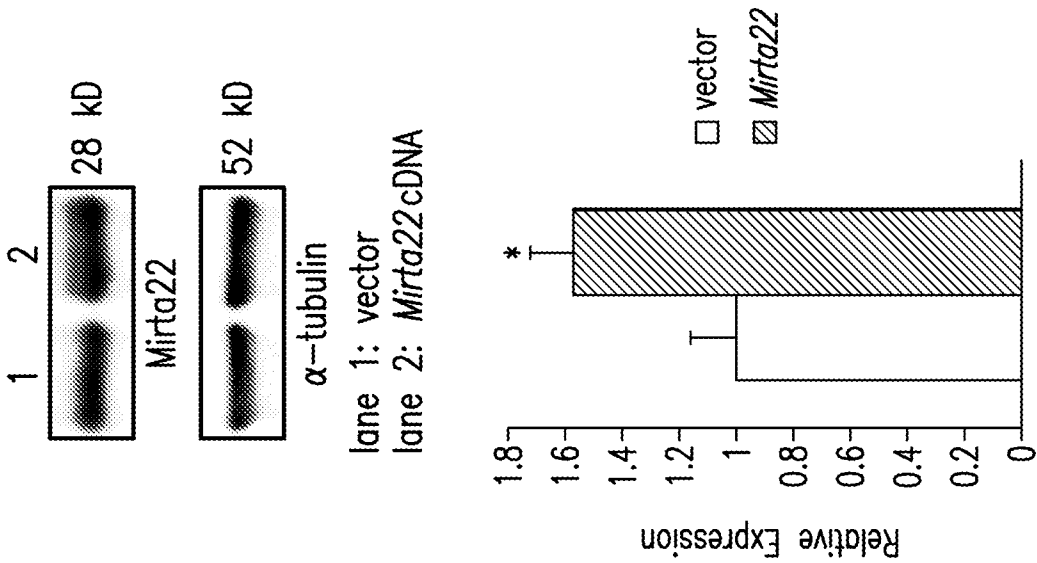


FIG. 13C

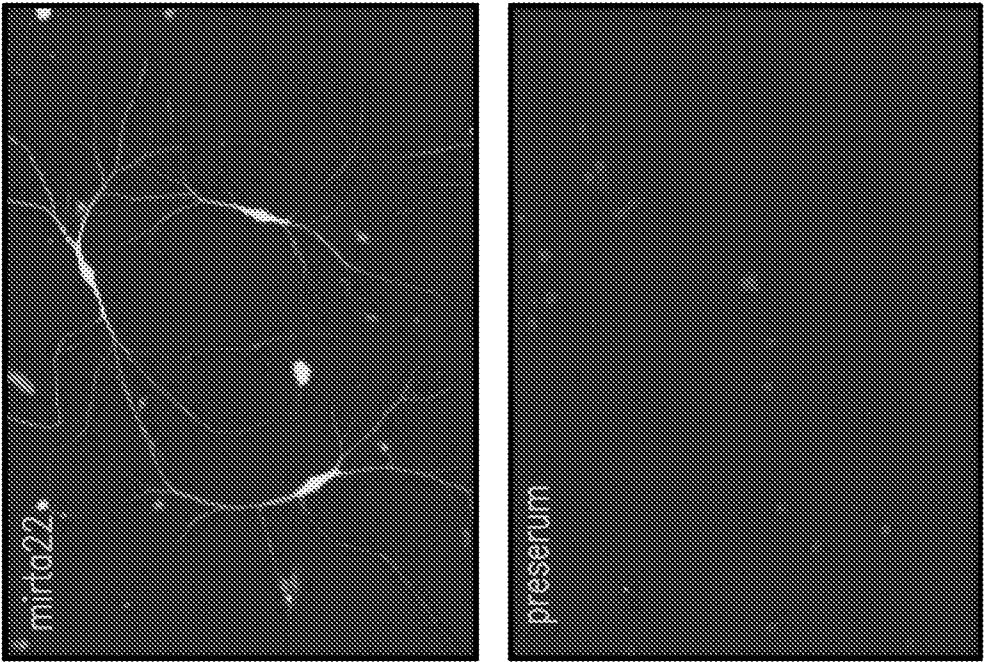


FIG. 13D

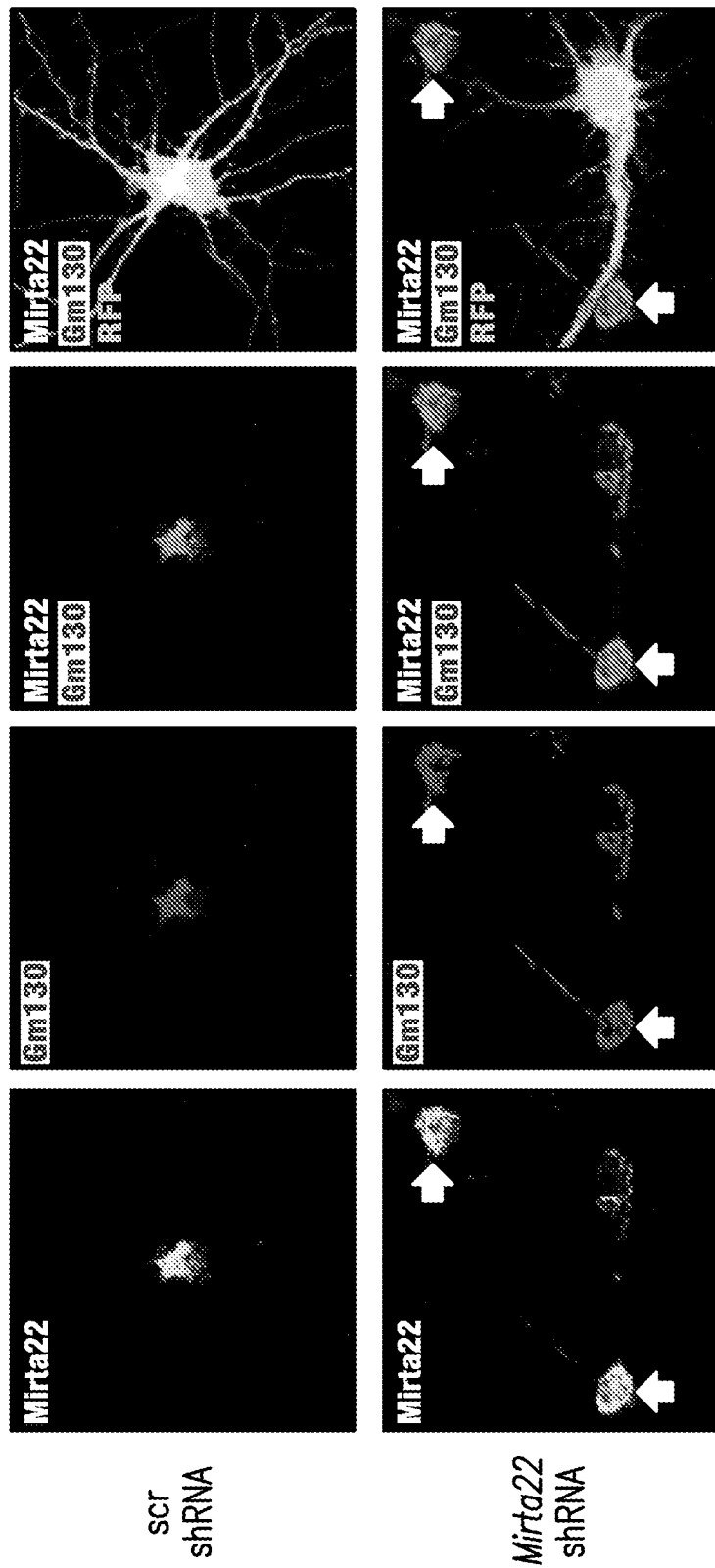


FIG. 13E

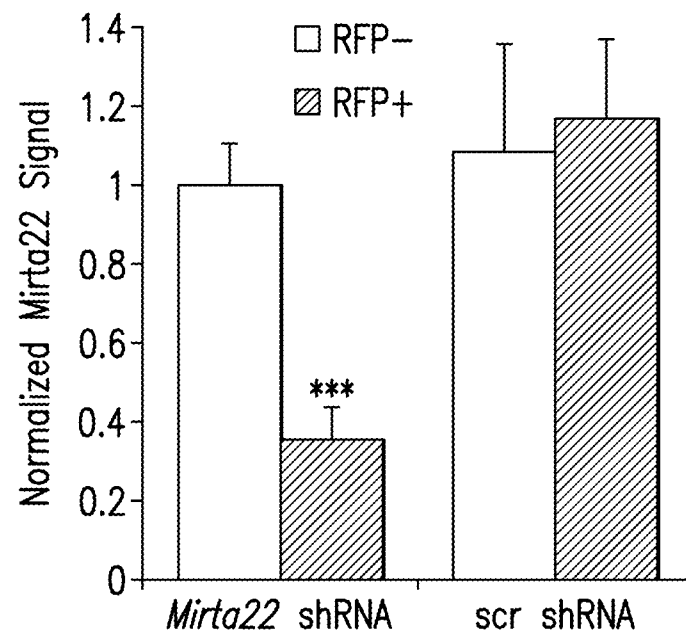


FIG. 13F

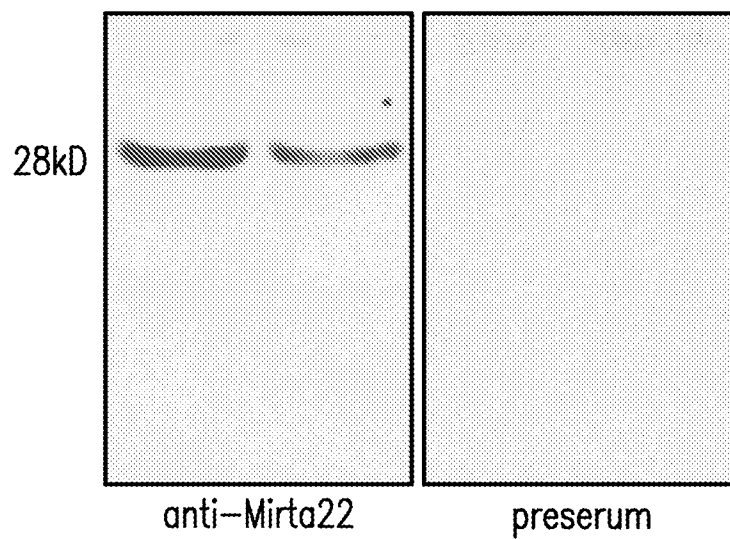


FIG. 13G

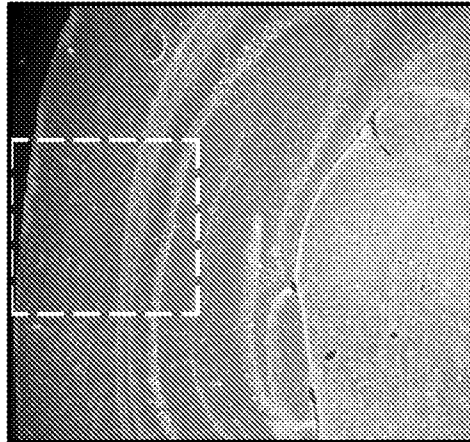


FIG. 14A

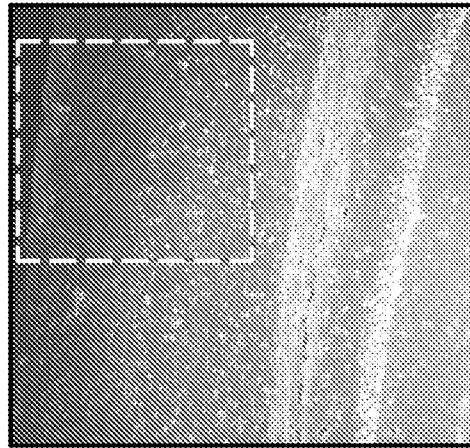


FIG. 14B

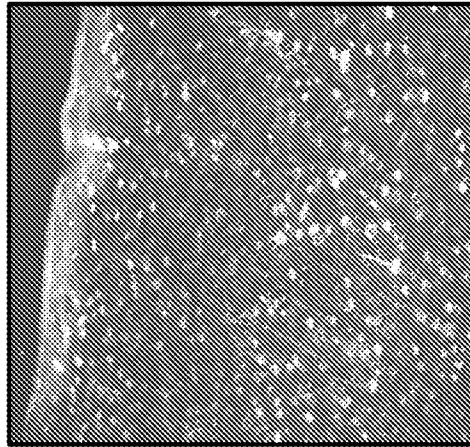


FIG. 14C

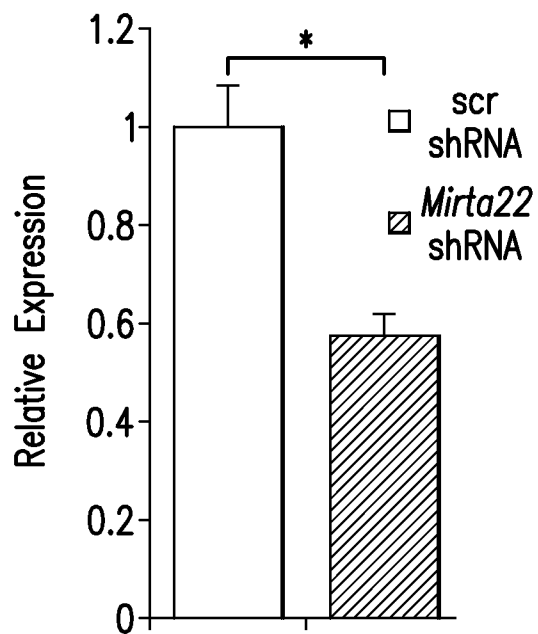


FIG. 15A

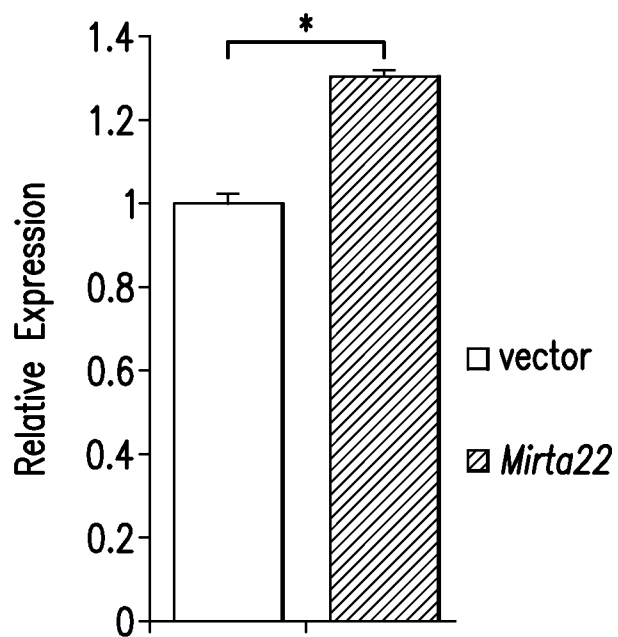


FIG. 15B

(SEQ ID NO:3)

```

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tttcccccg tcccacgagg ccacctgggc cagccccttg tcctctgcct tctgctggca 1021
gaggagcagc tggactgggg cctttggcac agcagccggt gtctcctgcg ccgcctccc 1081
ccatggcccc atgcagcccc aggggttcc cccctgccca tggagtagag ccgagatcc 1141
tgccactat gccagttctg acctgcac cccctaccc gagcccatgc agtctgggaa 1201
catgccgcct tctctccagc ctctgtgcct ttgttccagg tggctcacc ctctgtccc 1261
tggttgggt aggtggtcct gtccaggtc ctgcagcgc cccctcactt tgacactgga 1321
ctaggatgca gcctcccttc tgttccctc tgagggtacc ctgggtcccc tcatcagggg 1381
cagaggcatg aaagagtcgg ggctggatgg ccgggggctt ctgggcccga cgcctagtgc 1441
agcccctggg gtcgtggttt gacatttgc tgctgtgtc aaacaaggaa tccttgccct 1501
taaggtgaca ggccctccac aggttccag acttgaagga aaaggtttaa gaaagaaaac 1561
aaaaccaaca gttagtggag tcaaagccca gacactgtaa atagaacccc ctccaccacc 1621
ccccgccgcc cagcactcta cctggactgc ggtgctacga gggcctgcgg gcctttgctg 1681
tgtgccaccc tcctgttaag tctattttaa aacatcgacg atacattgaa atgtgtgaac 1741
gttttgaaaa gctacagctt ccagcagcca aaagcaactg ttgttttggc aagacggtcc 1801
tgatgtacaa gcttgattga aattcactgc tcacttgata cgttattcag aaacccaagg 1861
aatggctgtc cccatcctca tgttgctgtg tggagctcag ctgtgttgtg tggcagttta 1921
ttaaactgtc cccagatcg acacgcaaaa aaaaaaaaaa a

```

FIG. 16A

(SEQ ID NO:5)

```
1  ttccctcccg  cgtgctccgc  ggctcttggc  tcacagccgt  cccttcgctg  gtgggaagaa
61  gccgagatgg  cggcagccag  cgctggggca  acccggtgc  tcctgctctt  gctgatggcg
121  gtagcagcgc  ccagtcgagc  ccggggcagc  ggctgccggg  ccgggactgg  tgcgcgaggg
181  gctggggcgg  aaggtcgaga  gggcgaggcc  tgtggcacgg  tggggctgct  gctggagcac
241  tcatttgaga  tcgatgacag  tgccaacttc  cggaagcggg  gctcactgct  ctggaaccag
301  caggatggta  ccttgtecc  gtcacagcgg  cagctcagcg  aggaggagcg  gggccgactc
361  cgggatgtgg  cagccctgaa  tggcctgtac  cgggtccgga  tcccaaggcg  acccggggcc
421  ctggatggcc  tggaaactgg  tggctatgtc  tcctcctttg  tccttcgctg  ctccctgggtg
481  gagtgcaccc  tgtcggacca  gctgaccctg  cacgtggatg  tggccggcaa  cgtggtgggc
541  gtgtcggtag  tgacgcaccc  cgggggctgc  cggggccatg  aggtggagga  cgtggacctg
601  gagctgttca  acacctcgg  gcagctgcag  ccgcccacca  cagccccagg  cctgagacg
661  gcggccttca  ttgagcgcc  ggagatggaa  caggcccaga  aggccaagaa  cccccaggag
721  cagaagtcct  tcttcgcca  atactggcac  atcatcctgg  ggggggccc  gttgctcaca
781  gccctgcgtc  ctgctgcgcc  agggcccgcg  ccaccgccac  aggaggcctg  atggatgtac
841  atcattccc  tcgtcctgtt  cctcatgatg  tcaggagcgc  cagacaccgg  gggccagggt
901  gggggtgggg  gtgggggtgg  tgggtggggg  agtggccgg  gagggcccag  gctggtcagc
961  gtcccgtctt  gcacaccag  gggcctccct  ttctgctgga  gtcccctgtg  tcctcagcca
1021  tcccaagaag  ggtttgctgg  tcctccttt  cccccgtcc  cagaggcca  cctgggccag
1081  ccccttgctc  tctgccttct  gctggcagag  gagcagctgg  actggggcct  ttggcacagc
1141  agccggtgtc  tcctgcgccc  gcctccccc  tggccccatg  cagccccagg  ggcttcccc
1201  ctgcccattg  agtagagccc  gagatcctgg  ccactatgcc  agttctgacc  tcgcaccccc
1261  ctaccccag  cccatgcagt  ctgggaacat  gccgccttct  ctccagcctc  tgtgcctttg
1321  ttccaggtgg  tctcacccct  ctgtccctgg  ctgggctagg  tggctcctgtc  caggctcctg
1381  cagcgcctcc  ctcactttga  cactggacta  ggatgcagcc  tccttctgt  gtccccttga
1441  gggtagcctg  ggtccctca  tcaggggcag  aggcatgaaa  gagtcggggc  tggatggccg
1501  ggggcttctg  ggcccgcgc  ctagtgcagc  ccctggggtc  gtggtttgac  atttgtctgc
1561  ctggtgcaaa  caaggaatcc  ttgcctttaa  ggtgacaggc  cctccacagg  ctccagact
1621  tgaaggaaaa  ggtttaagaa  agaaaacaaa  accaacagtt  agtggagtca  aagcccagac
1681  actgtaata  gaacccctc  caccacccc  cgccgccag  catcctacct  ggactgcgtt
1741  gctacgagg  cctgcgggcc  tttgctgtgt  gccaccctcc  ctgtaagtct  atttaaaaac
1801  atcgacgata  cattgaaatg  tgtgaacgtt  ttgaaaagct  acagcttcca  gcagccaaaa
1861  gcaactgttg  ttttggaag  acggctcctga  tgtacaagct  tgattgaaat  tcactgctca
1921  cttgatacgt  tattcagaaa  cccaaggaa  ggctgtcccc  atcctcatgt  ggctgtgtgg
1981  agctcagctg  tgttgtgtgg  cagtttatta  aactgtcccc  cagatcgaca  cgcaaaaaaa
2041  aaaaaaaa
```

FIG. 16B

(SEQ ID NO:19)

```
1 gagaaatggc cgccctgact gcgggagcag gcggacgggc gctagtgcgc aggcgcggcg
61 tgcggcgcag gcgcgtgagc ctcaggatga accctgtgtt tcctagcggg ctgtatggct
121 ctcgggtttt ctcaacgctc ccgtatggtg gccgcgggtg ccggggtgac ccggctgcta
181 gtgctcttgc tgatggtagc cgcggctcct agcagagccc gaggcagcgg ctgccgggtc
241 ggggcctccg cgcgtgggac cggggccgat ggccgtgaag ctgagggctg tggcaccgtg
301 gctttgctgc tggagcattc atttgagctc ggtgatggag ccaacttcca gaagcgaggc
361 ttgctgctct ggaaccagca ggatggcacc ctgtcggcaa cacagcgaca gctcagttag
421 gaggagcgtg gccgactccg ggatgtggct gctgtcaatg gcctctacag ggtccgggtc
481 ccgaggcggc ctgggacact tgatggttca gaagctggcg gccatgtgtc ttccctcgtc
541 ccagcgtgct ccctgggtga gtcgcacctt tcggaccagc tgaccttgca cgtggatgtg
601 gctggcaacg tggtagggcct gtctgtggtg gtgtacctg ggggctgccg gggctccgag
661 gtggaagatg aggacctgga gctgttcaat acatctgtgc agctgcggcc tcccagcact
721 gctccaggcc ccgagactgc agccttcatt gagcgcctgg agatggagca ggcccagaag
781 gccaaagaacc cacaggagca gaagtcttcc ttgtccaaat actggatgta catcattcca
841 gttgtgctgt tcctcatgat gtcgggagcg ccggacgctg ggggccaggg cggcggtggg
901 ggcgggggca gcagccggtg agcagctgtg ccacctagag cccccccag agccagccca
961 agaaggagtt cctgtcccca catttcccta ttgcatgaat atggaaggct gtcccttcag
1021 tgagccctct ggcccttcctg taagcccctc ttctgttccc tgagccctct tctcatcctg
1081 ttgactgaga gcttgggtgg acctccctgt agccagctca ctgcaactgt gtcccaccat
1141 gtggcactgt gctcctctgt ctgctaaca cccaccagcc tgccccaccc caccaccaca
1201 tacactttgg gaacttgcca agctctctcc agcctctgtg cctttgccct gcaggccccg
1261 tgcgcccctc actgtcactc tccagccctt tgccaaggat ctgtggccca gaggcctctg
1321 ctcttagtgg ctaggtcagc ctccagccca ctgtccagggt ggcatgctgt cttctttgcc
1381 cccctctctg gtgcccaga ataccatggt gacctaccac tatectttct gcctttggat
1441 gtcatagcct ggatctgtca ccaggagagg attgtgggccc tccacgttag tctgtgaatg
1501 cacacttcga gtgacttggt tgcaggtttt gagagccggt tttgcactag ctgctcgaca
1561 gctgctggca tggccgtgct cttgcacatg cgccgctgtg ggcatgggga ttgctgtgca
1621 gcctcagctg tgttgtgtgg ctgctgatta aactgtcccc taaacagcca ctcttcagct
1681 cacttcctgc cttctgtgct tgtgaatagt cctgagttgc cgctgtggtt tgccctggtt
1741 atgtttgaat ggctttctta gggatatgta cagaggggtg cctgagcaga ttaaagttgc
1801 tgtgagcaag gacgccttcc gaactctggg aggaggctgg ttcttgacct tccta
```

FIG. 16C

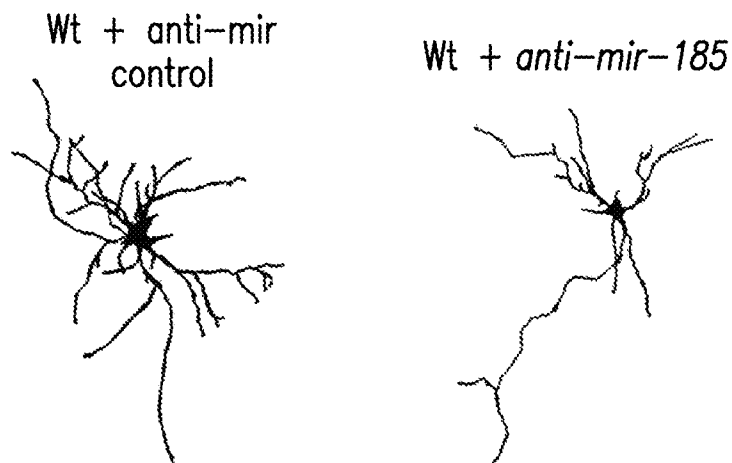


FIG. 17A

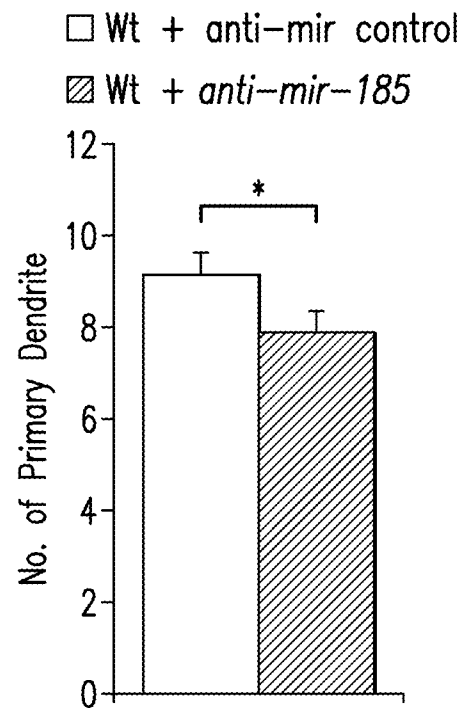


FIG. 17B

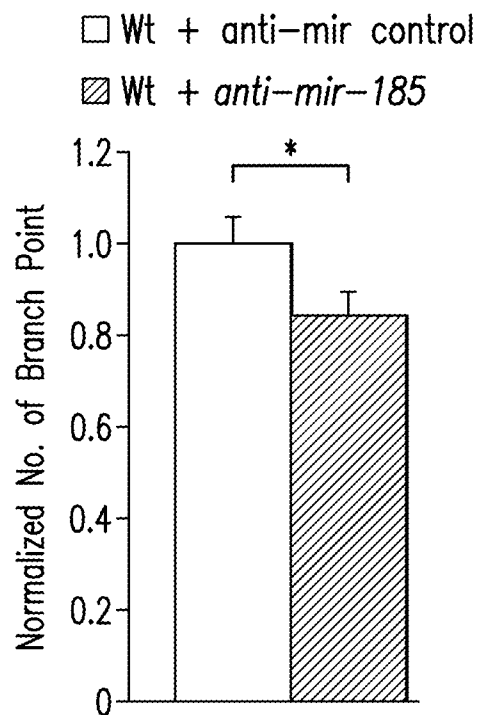


FIG. 17C

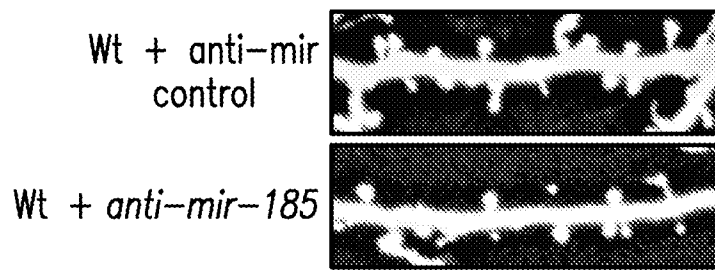


FIG. 17D

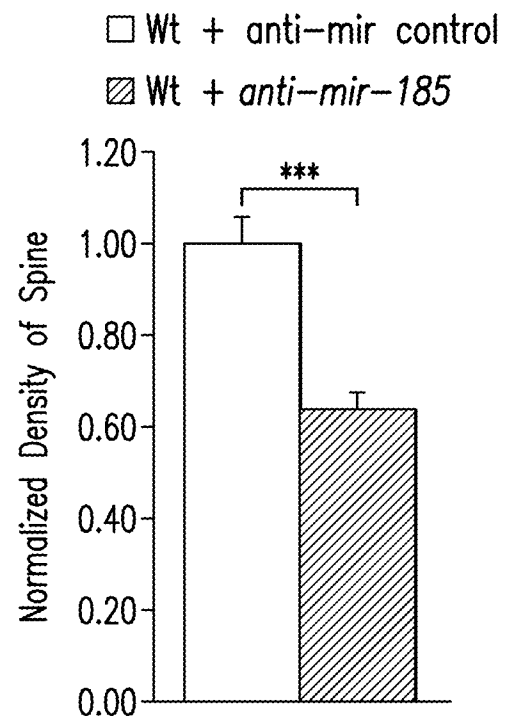


FIG. 17E

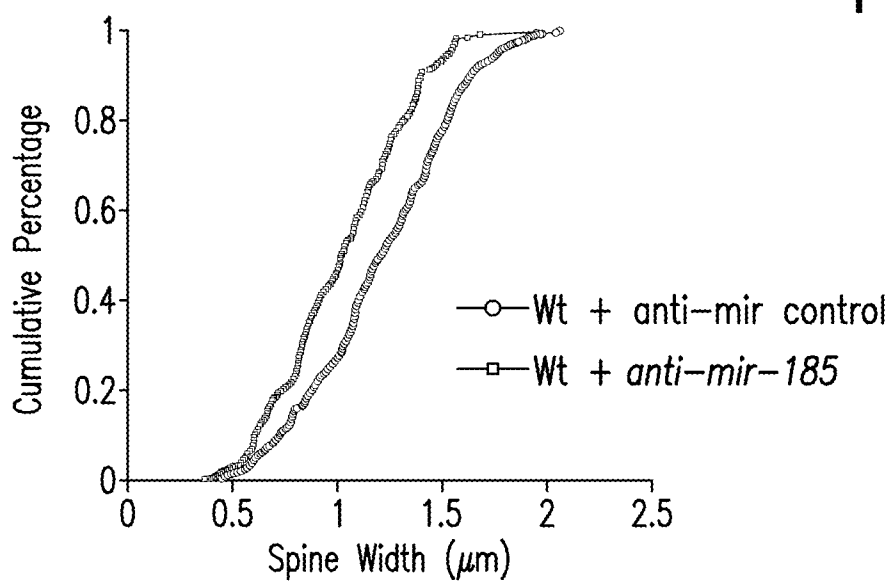


FIG. 17F

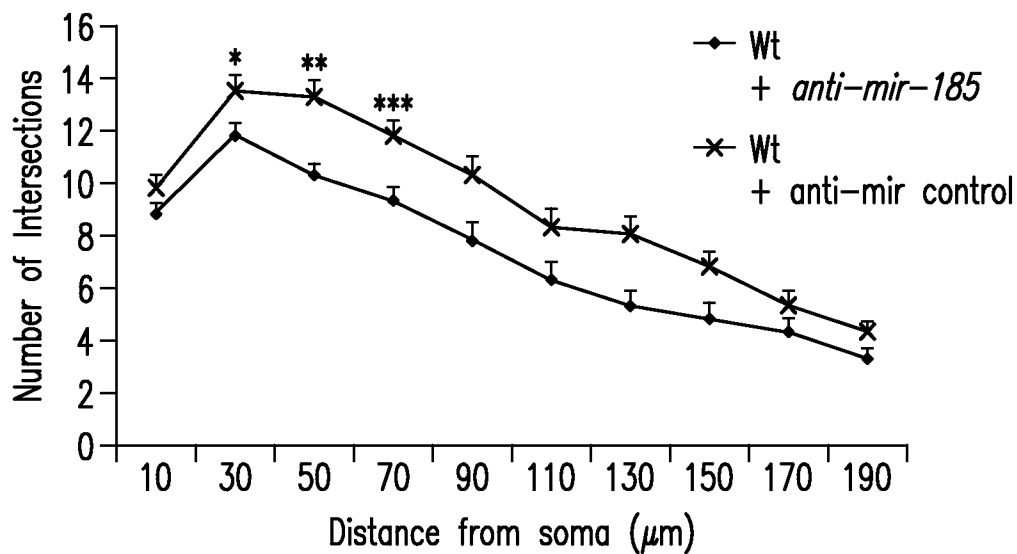


FIG. 18A

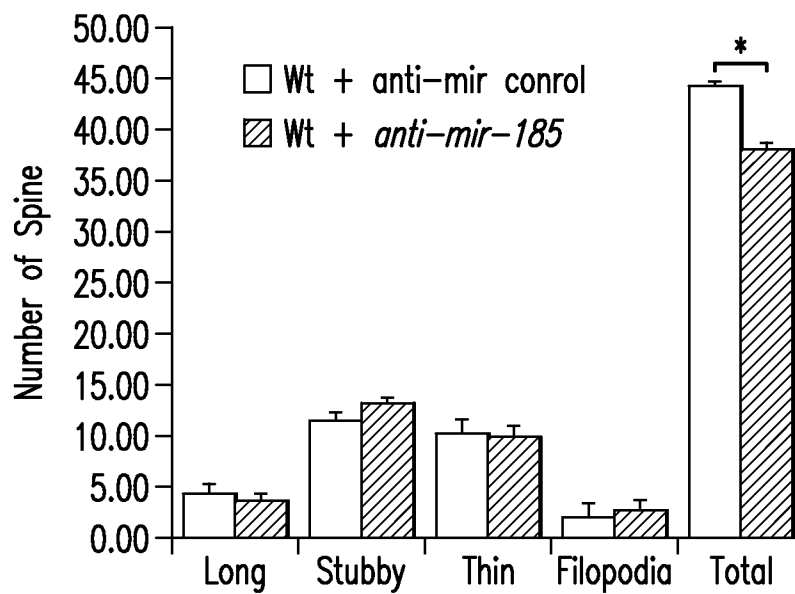
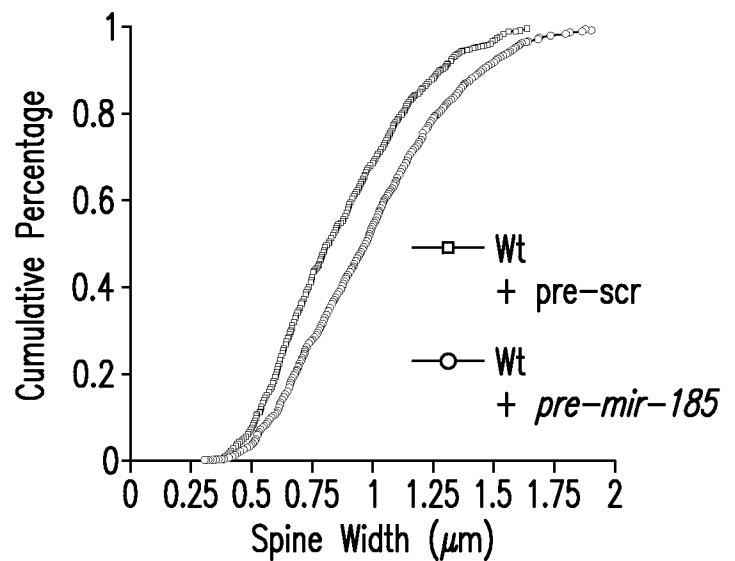
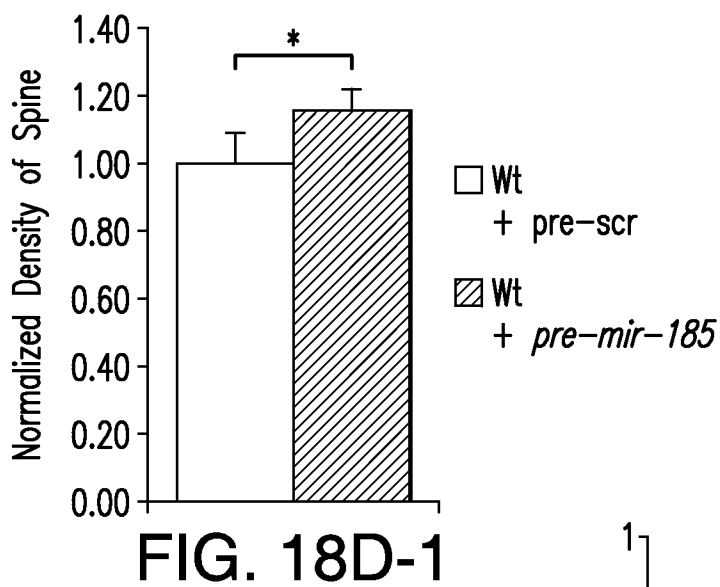
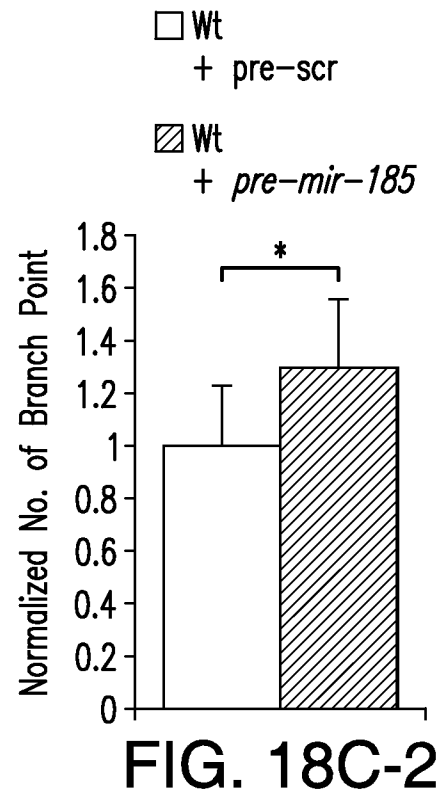
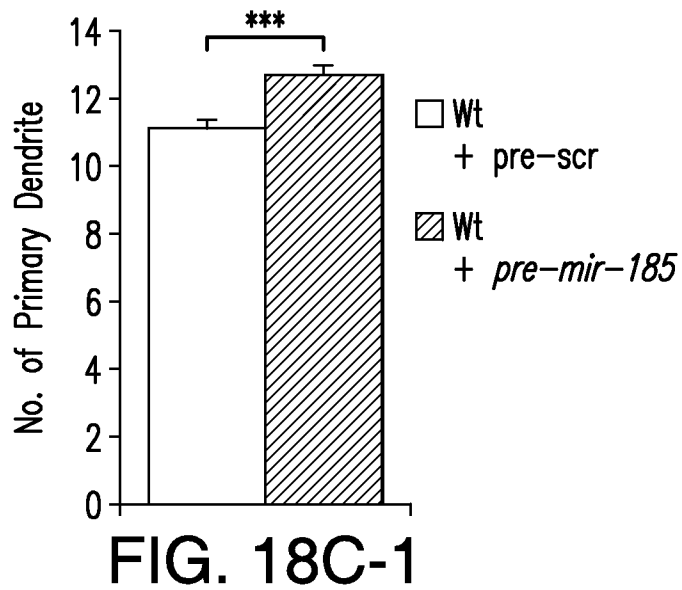


FIG. 18B



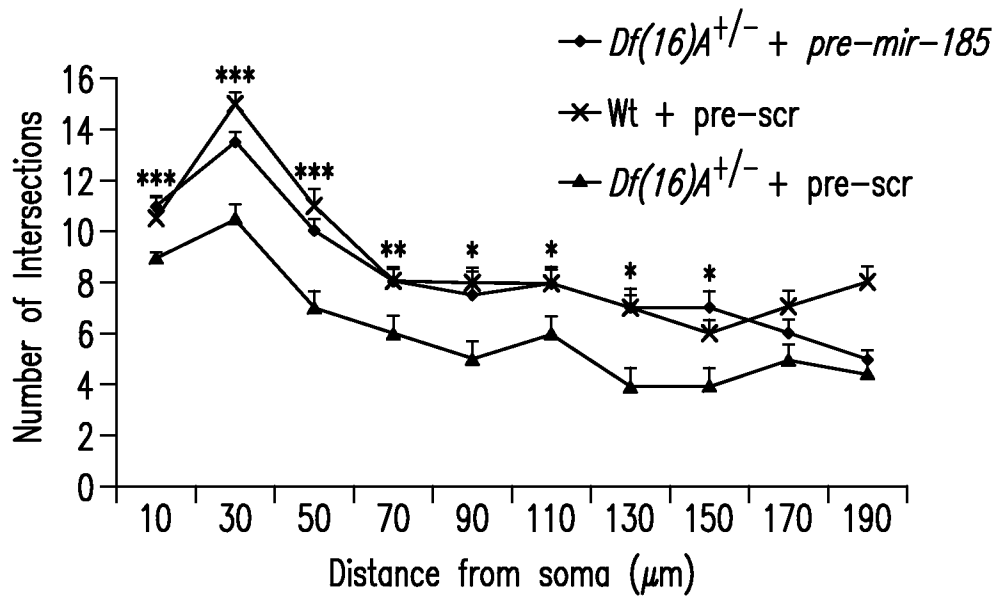


FIG. 18E

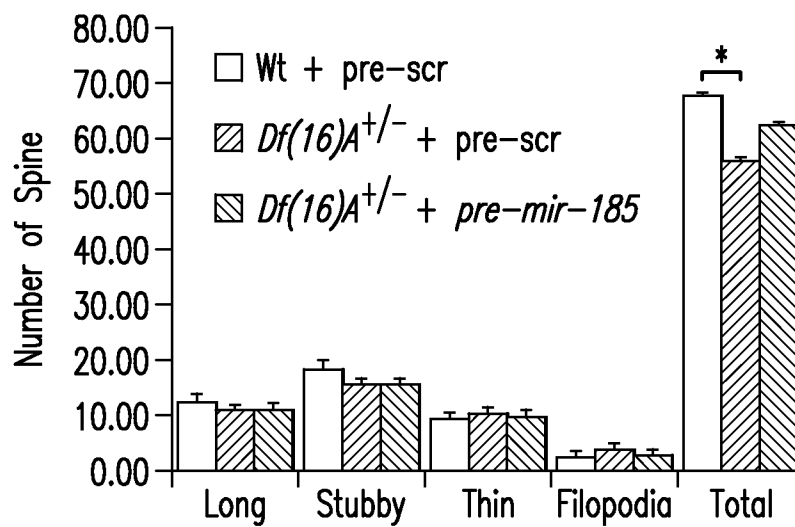
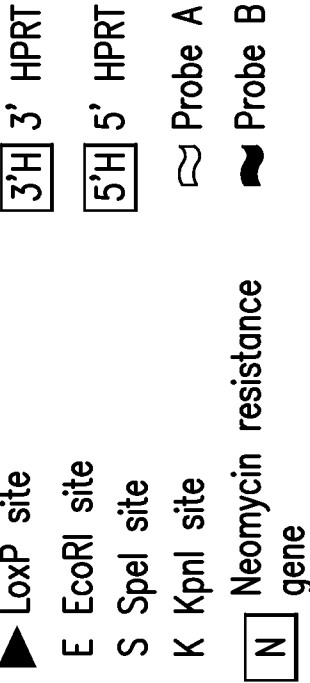


FIG. 18F



Dgcr14 locus

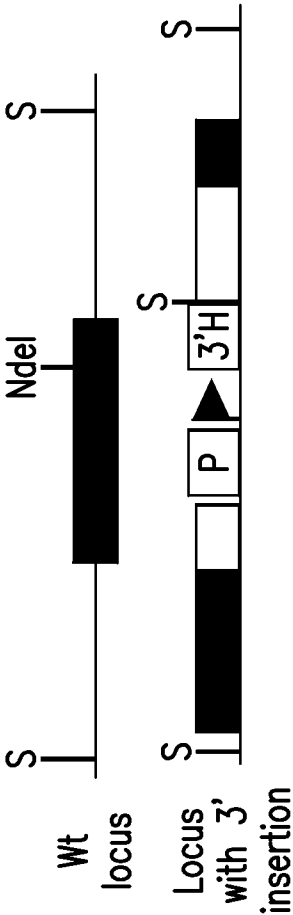
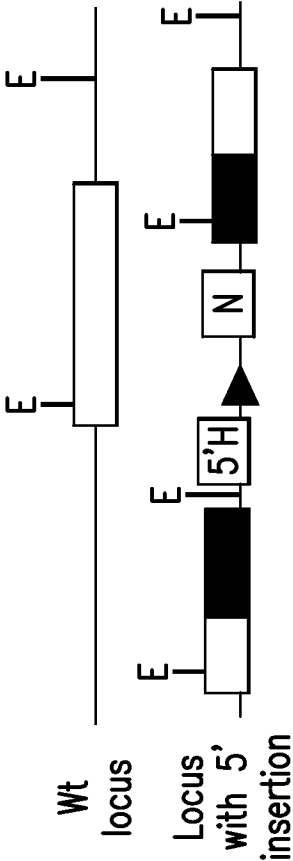
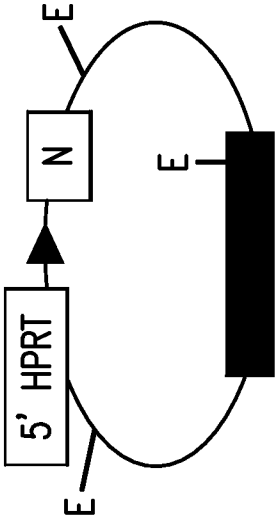


FIG. 19A

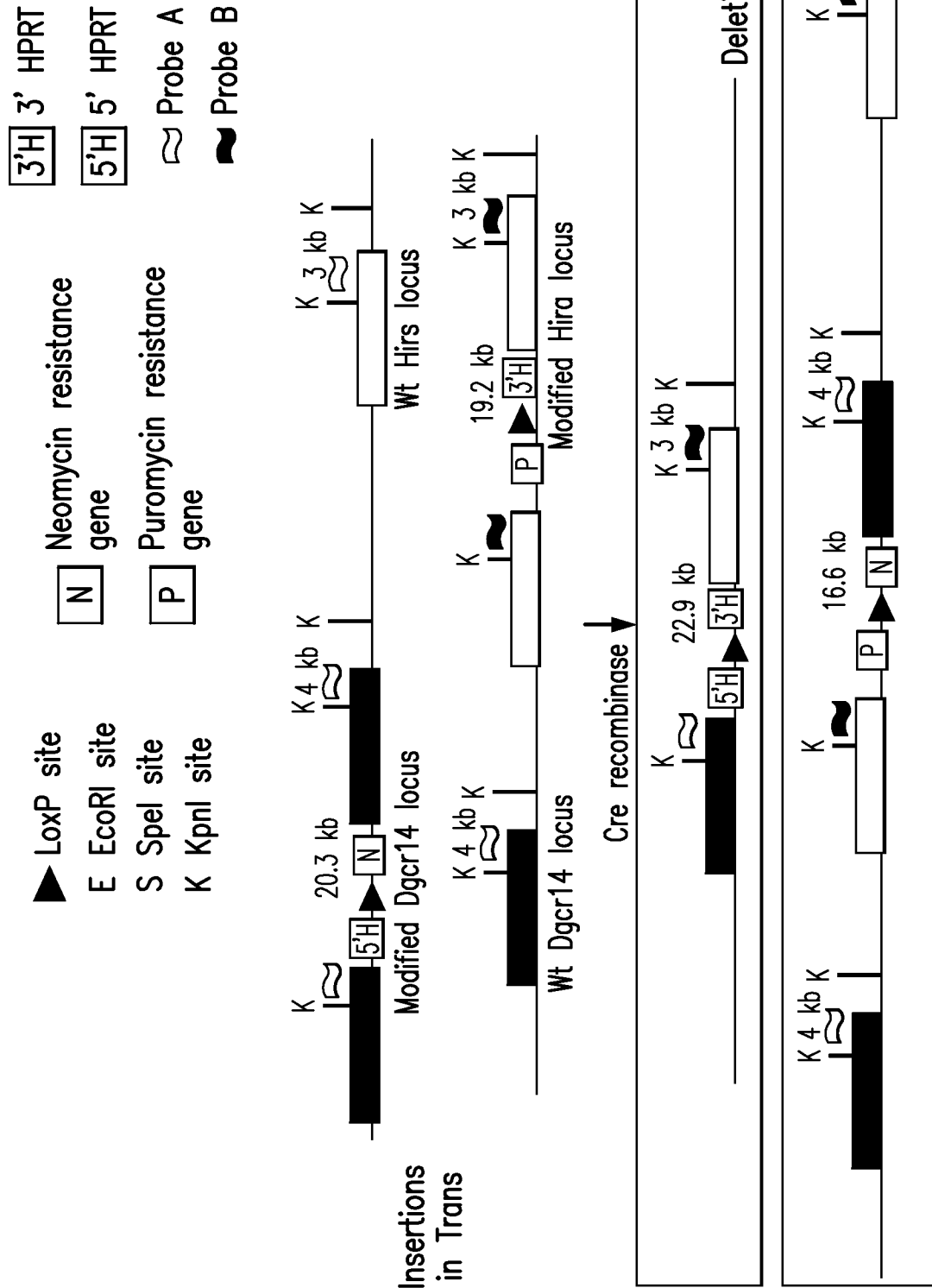


FIG. 19B

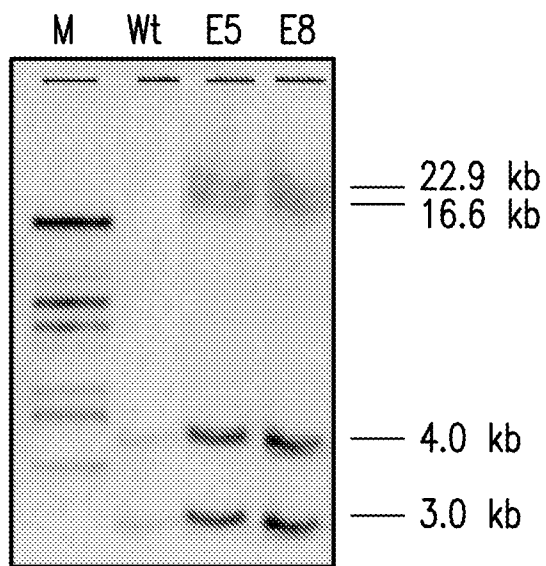


FIG. 19C

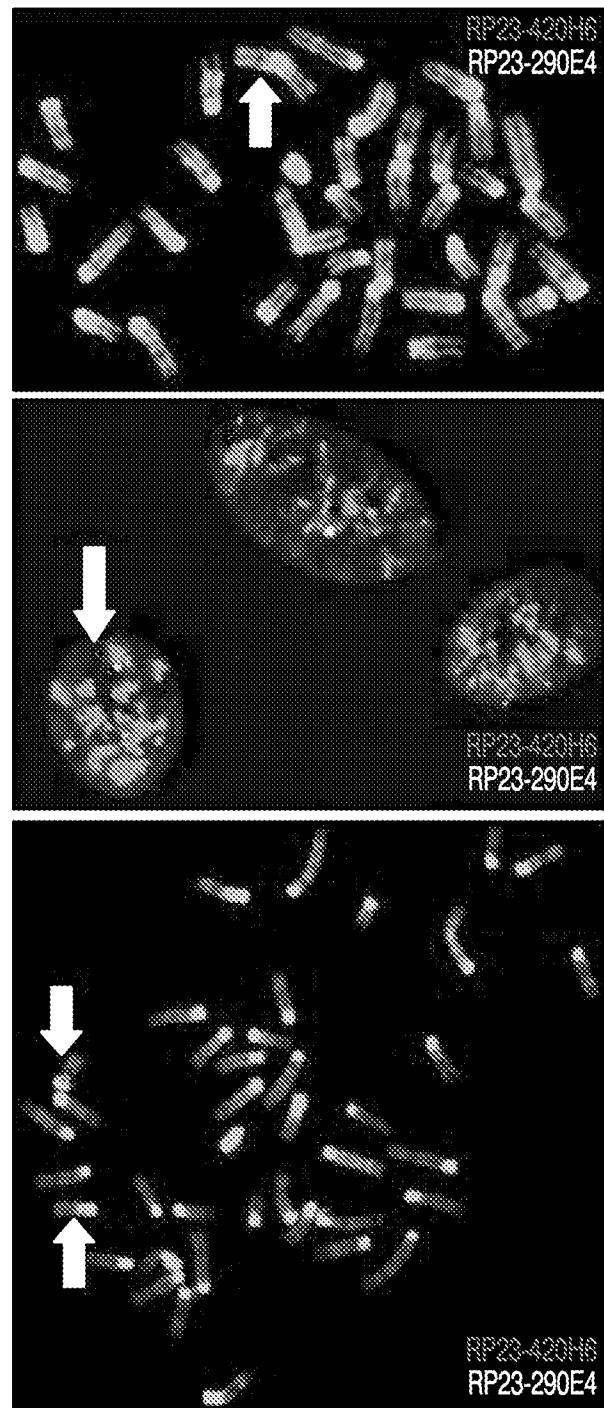


FIG. 19D

AffyID	Chr	Chr_start	Chr_end	PFC			HPC					
				Fold change (Del/Bal)*	Fold change (Dup/Bal)*	P Value	Fold change (Del/Bal)*	Fold change (Dup/Bal)*	P Value			
1424038_at	7	51745308	51751883	1.43	0.86	73.00	2.6E-11	1.29	0.88	68.49	1.1E-10	2310044H10Rik
1435179_at	13	83867710	83875274	1.46	0.89	52.60	8.5E-10	1.42	0.95	45.42	6.4E-09	C130071C03Rik (mir-9-2)
1460033_at	1	198850302	198864096	1.76	0.85	44.49	4.6E-09	1.71	0.86	42.71	1.1E-08	A330023F24Rik (mir-29c)
1435089_at	13	63116293	63400964	1.77	0.78	43.72	5.5E-09	1.41	0.82	43.86	8.9E-09	201011101Rik (mir-23b)
1428562_at	11	75275040	75280192	1.33	0.85	42.88	6.7E-09	1.33	0.92	30.51	2.4E-07	2210403K04Rik (mir-22)
1440357_at	15	85537748	85537832	2.07	0.87	32.51	9.4E-08	1.68	0.81	48.73	3.3E-09	(let-7b)
1438838_at	X	100764843	100819093	1.30	0.88	18.77	9.6E-06	1.30	0.93	19.90	7.7E-06	B230206F22Rik (mir-374)
1431094_at	12	110833610	110835408	1.20	0.91	17.13	1.9E-05	1.20	0.87	25.24	1.2E-06	1110006E14Rik (mir-136)
1457030_at	12	110973190	110987665	1.29	0.91	14.82	5.3E-05	1.22	0.80	9.73	7.9E-04	Mirg
1434730_at	7	86676773	86677477	1.42	0.96	14.75	5.5E-05	1.40	0.95	13.99	9.2E-05	A854517 (mir-9-3)
1427410_at	14	62221673	62301210	1.27	0.90	13.75	8.8E-05	1.49	0.95	33.16	1.2E-07	Dleu2 (mir-15a)
1442913_at	3	118242789	118243450	1.31	0.92	9.92	6.4E-04	1.39	0.84	22.06	3.5E-06	AK141880

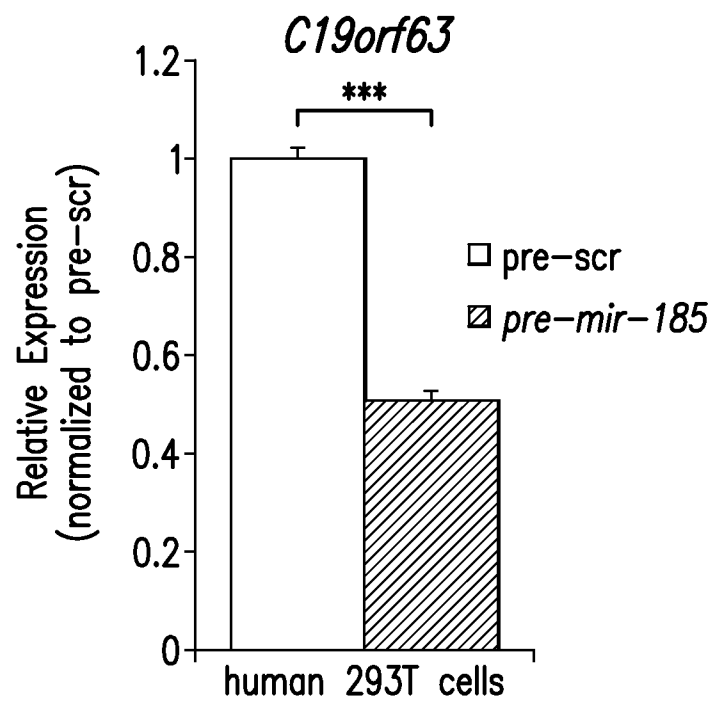
* Bal = mouse strain balanced for copy number; Del = mouse model of the 22q11.2 microdeletion; Dup = mouse model of the 22q11.2 microduplication

FIG. 20

Three factor ANOVA of the impact of *mir-185*, *mir-485* and *mir-491* on luciferase activity, related to Figure 4.

FACTOR	DF	SS	MS	F	Pr > F
pre_mir_185	1	0.582	0.582	6554	0
pre_mir_491	1	0.0307	0.0307	346.3	0
pre_mir_185*pre_mir_491	1	0.0062	0.0062	70.27	0
pre_mir_485	1	0.0011	0.0011	12.86	0.0059
pre_mir_185*pre_mir_485	1	0.0008	0.0008	9.069	0.0147
pre_mir_485*pre_mir_491	1	2.50E-05	2.50E-05	0.282	0.6084

FIG. 21

**FIG. 22**

David Functional Annotation Clustering

Current Background: Mus musculus

2695 DAVID IDs

Classification Stringency: Medium

Enrichment Score 8.56

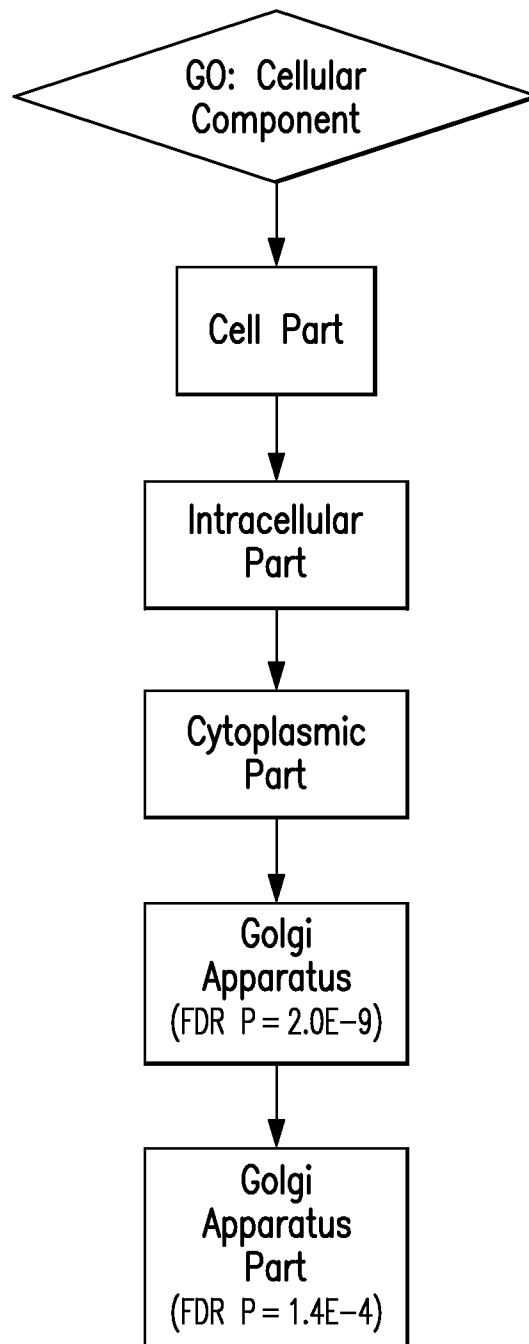


FIG. 23A-1

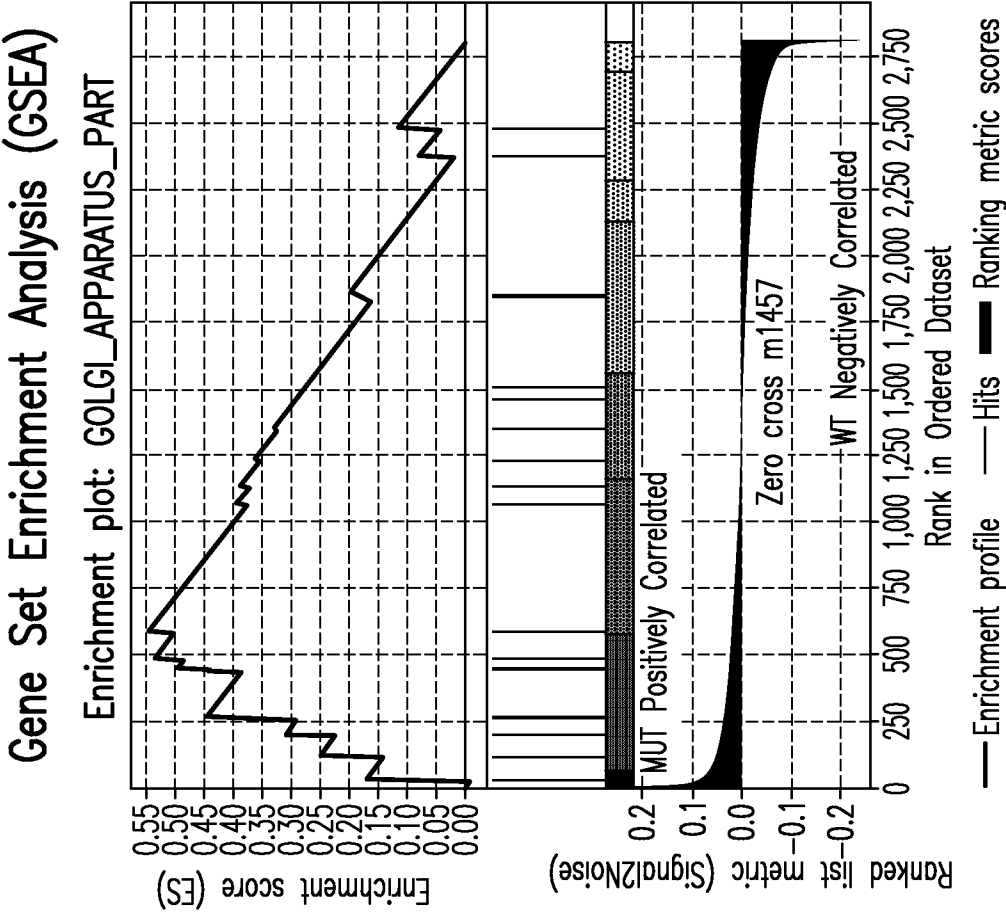


FIG. 23A-3

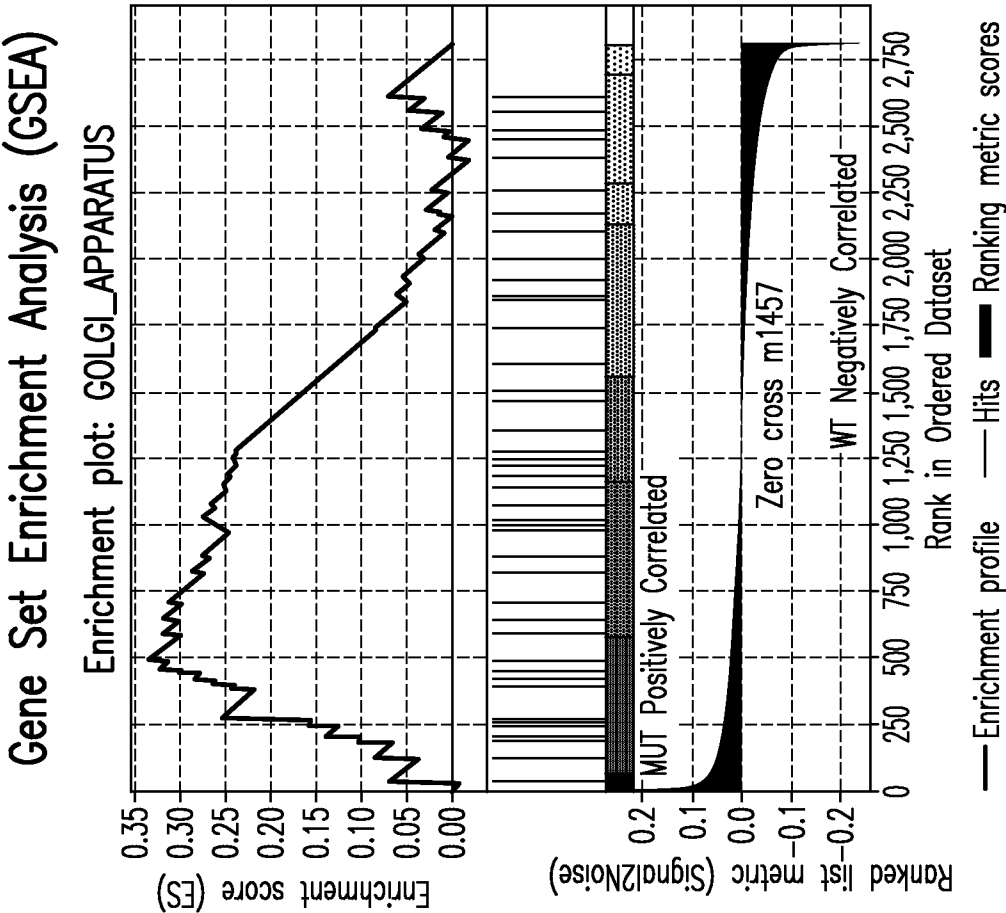


FIG. 23A-2

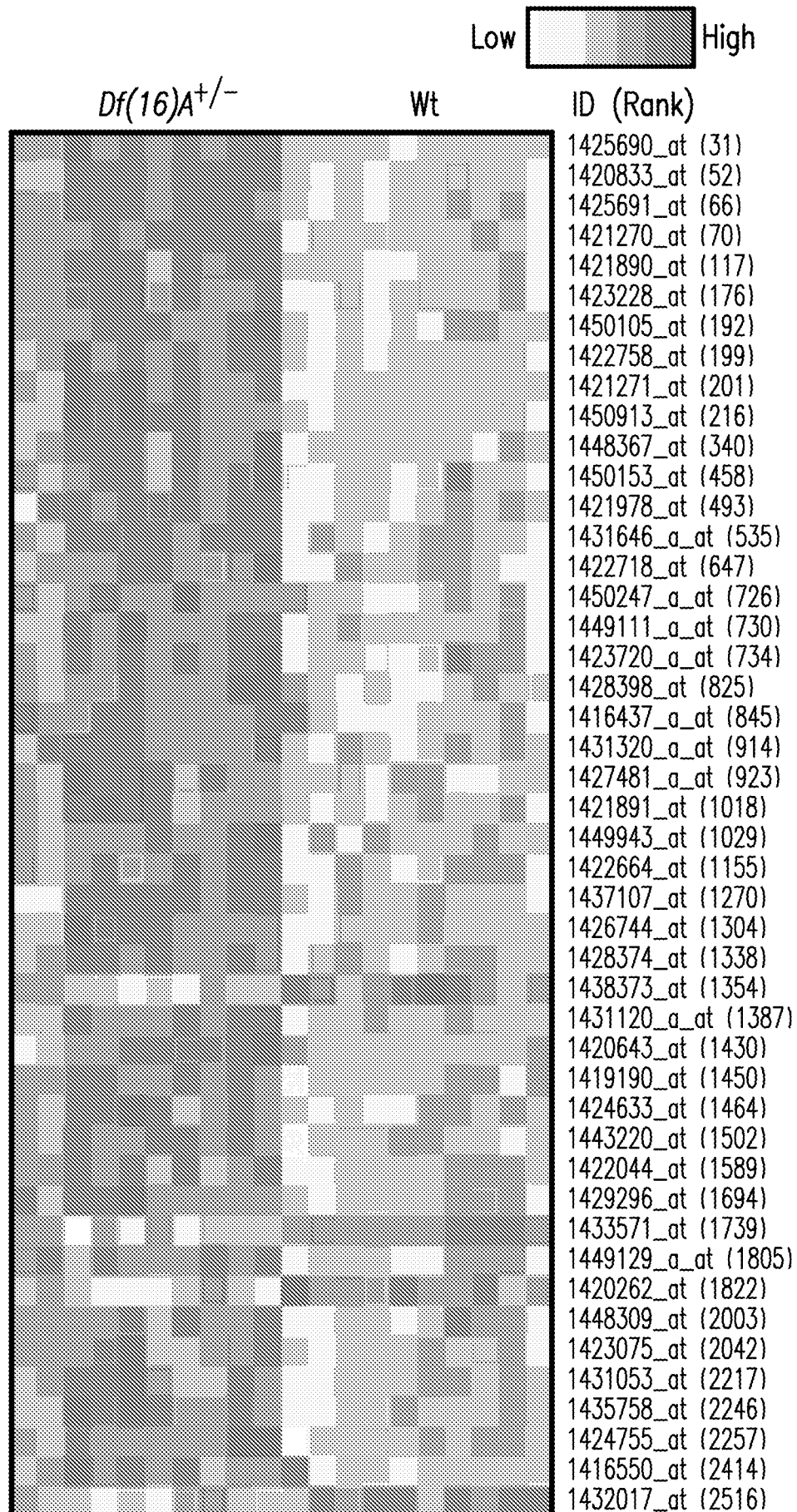


FIG. 23B

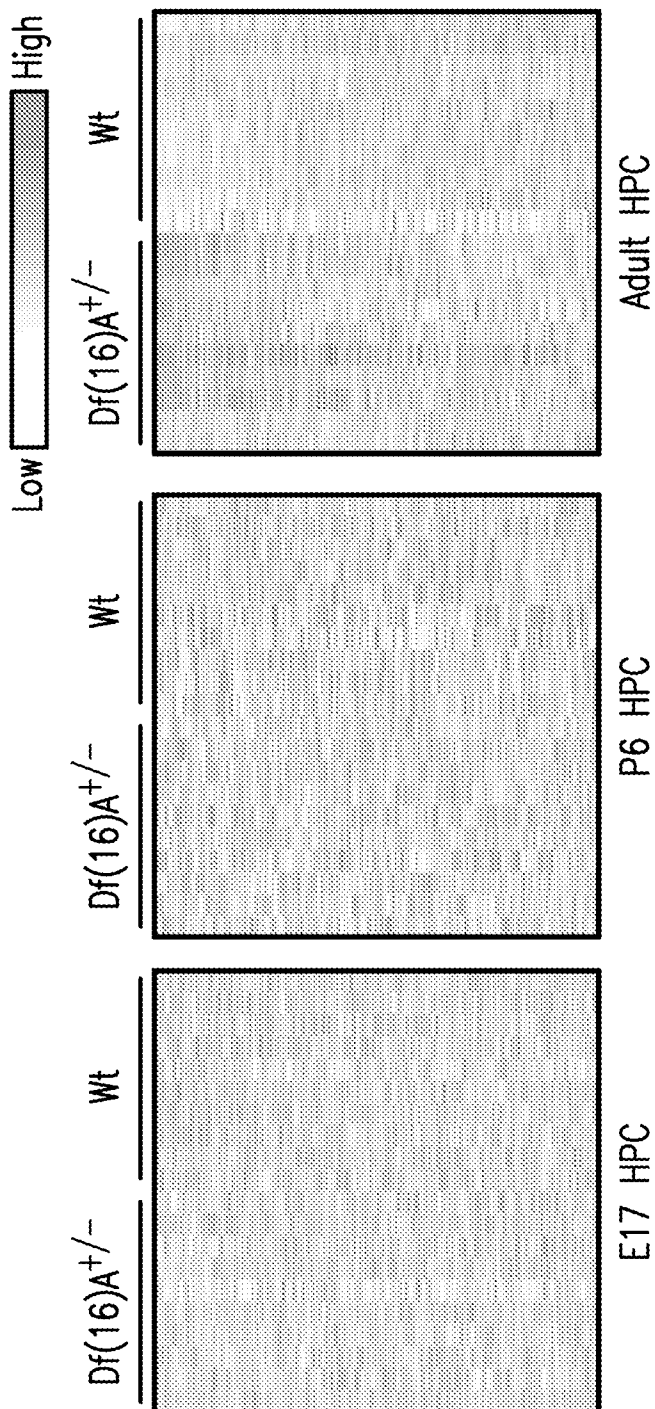


FIG. 24

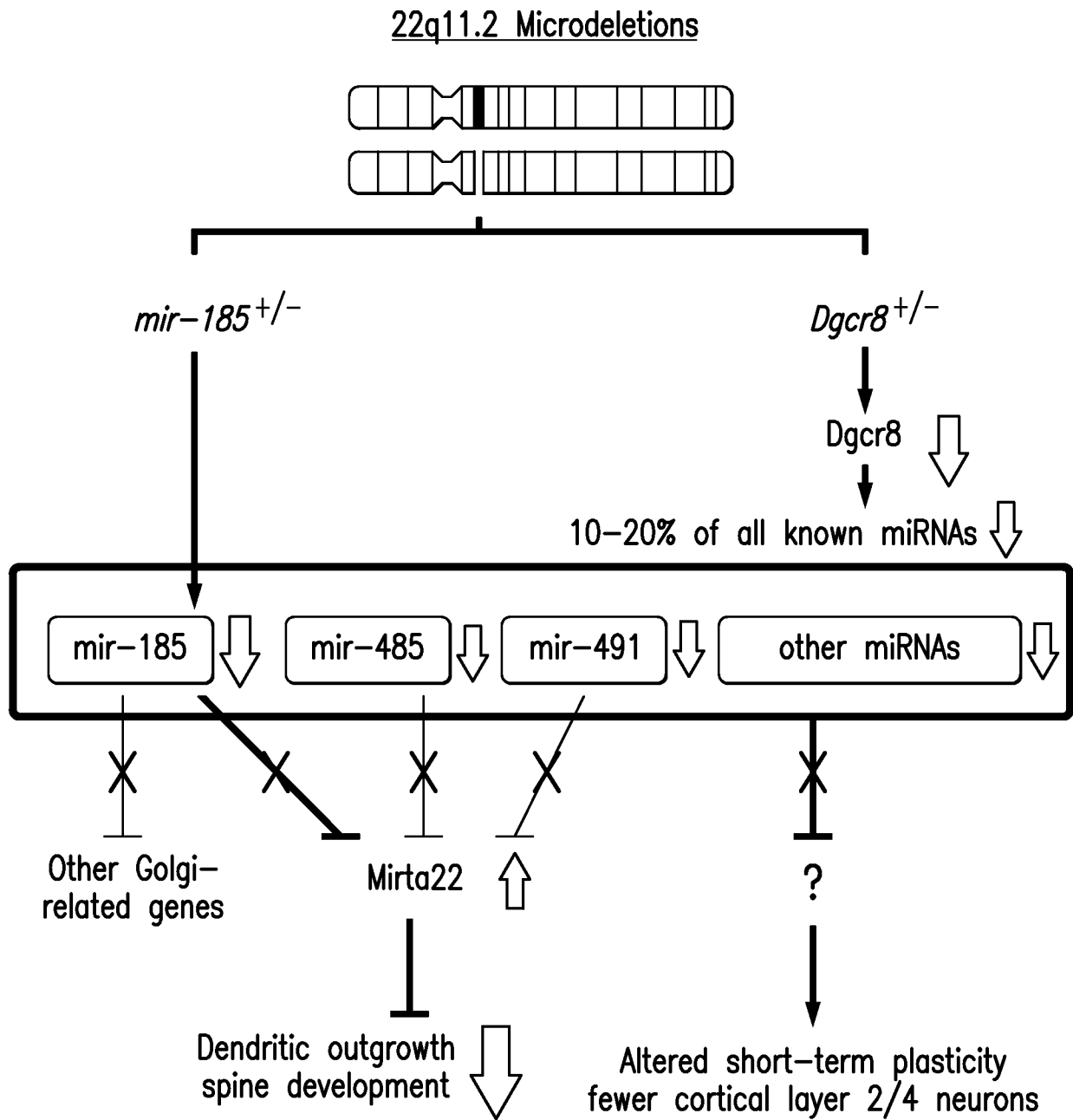


FIG. 25

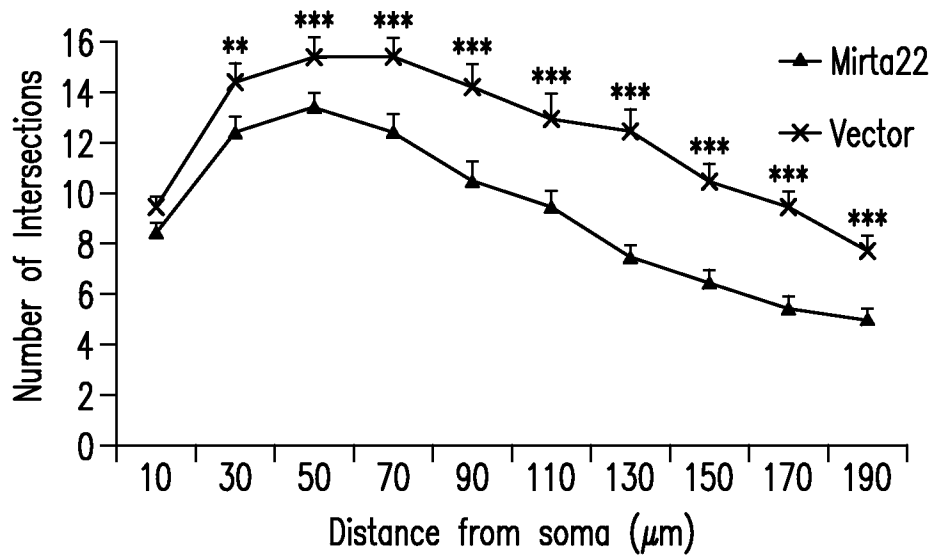


FIG. 26A

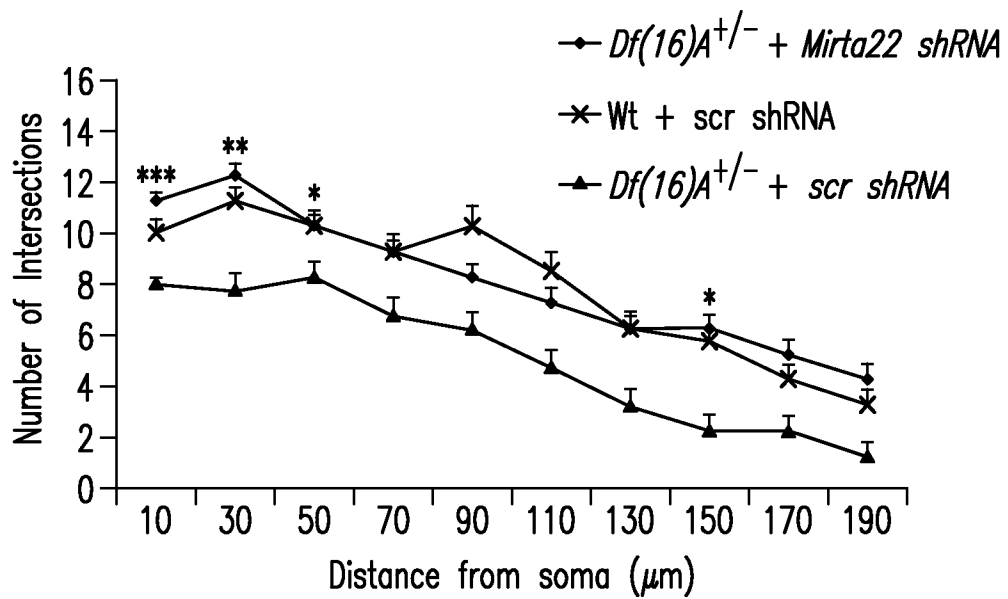


FIG. 26B

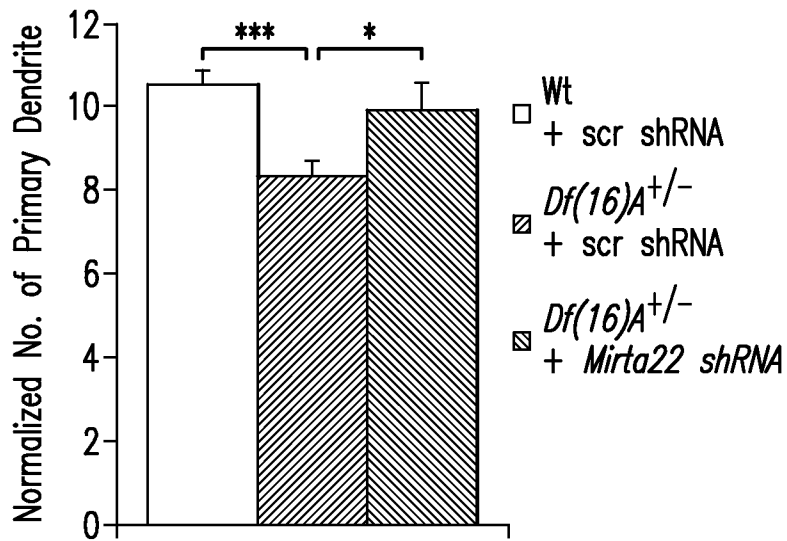


FIG. 26C

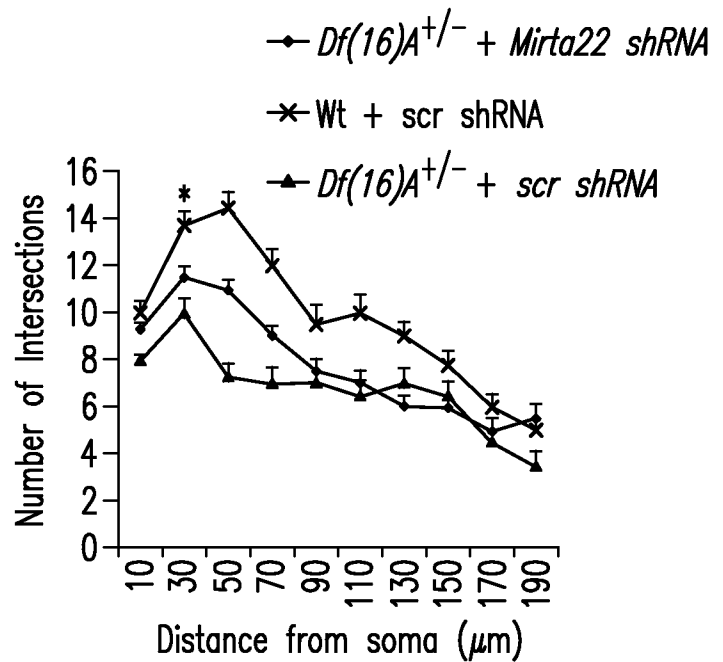


FIG. 26D

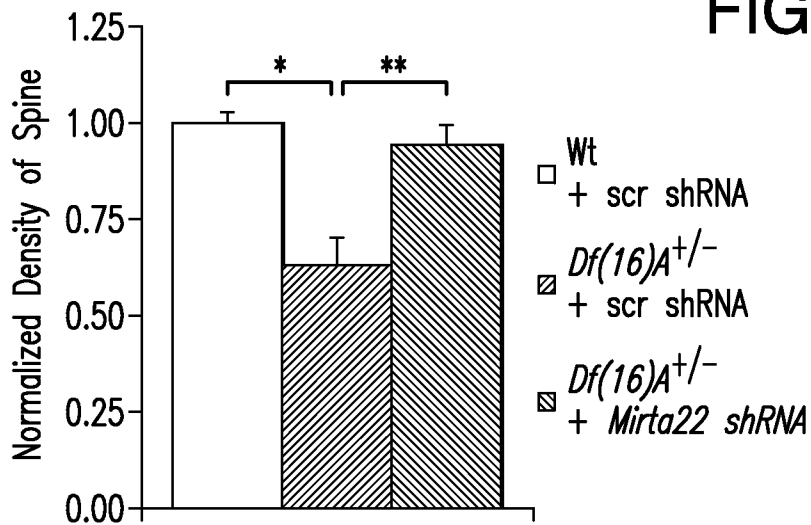


FIG. 26E

List of transcripts outside the 22q11.2 syntenic region misregulated in a reciprocal manner

Brain region	AffyID	Chr	Chr_start	Chr_end	Fold change (Del/Bal)*	Fold change (Dup/Bal)*	F	P Value	Symbols
PFC	1424038_a_at	7	51745308	51751883	1.43	0.86	73	2.60E-11	2310044H10Rik
PFC	1435179_at	13	83867710	83875274	1.46	0.89	52.6	8.50E-10	C130071C03Rik (miR-9-2)
PFC	1460033_at	1	196850302	196864096	1.76	0.85	44.49	4.60E-09	A330023F24Rik (miR-29c)
PFC	1435089_at	13	63116293	63400964	1.77	0.78	43.72	5.50E-09	2010111101Rik (miR-23b)
PFC	1428562_at	11	75275040	75280192	1.33	0.85	42.88	6.70E-09	2210403K04Rik (miR-22)
PFC	1440357_at	15	85537748	85537832	2.07	0.87	32.51	9.40E-08	(let-7b)
PFC	1438838_at	X	100764843	100819093	1.3	0.88	18.77	9.60E-06	B230206F22Rik (miR-374)
PFC	1431094_at	12	110833610	110835408	1.2	0.91	17.13	1.90E-05	1110006E14Rik (miR-136)
PFC	1439305_at	X	100772966	100773671	1.4	0.97	15.3	4.20E-05	(miR-374)
PFC	1430959_at	17	17967938	17976035	1.46	0.88	14.93	5.00E-05	Ncrna00085
PFC	1456904_at	11	97049276	97050652	0.82	1.08	14.92	5.10E-05	NA
PFC	1457030_at	12	110973190	110987665	1.29	0.91	14.82	5.30E-05	Mirg
PFC	1434730_at	7	86676773	86677477	1.42	0.96	14.75	5.50E-05	AI854517 (miR-9-3)
PFC	1427410_at	14	62221673	62301210	1.27	0.9	13.75	8.80E-05	Dleu2 (miR-15a)
PFC	1419161_a_at	7	94395304	94547116	1.17	0.97	13.66	9.20E-05	Nox4
PFC	1441666_at	1	148274740	148342171	0.81	1.02	13.48	1.00E-04	AK162963
PFC	1453713_s_at	17	17967938	17976035	1.8	0.89	13.18	1.20E-04	Ncrna00085
PFC	1457181_at	15	81822952	81846570	0.84	1.16	12.67	1.50E-04	Pppde2
PFC	1438053_at	16	56690444	56717467	0.71	1.06	11.63	2.60E-04	Tfg
PFC	1431343_at	14	8603700	8625033	1.16	0.91	11.59	2.60E-04	Gm10044
PFC	1430603_at	9	98462878	98464030	0.93	1.25	11.14	3.30E-04	4930579K19Rik
PFC	1459704_at	11	100447804	100448219	0.85	1.05	10.91	3.70E-04	Dnajc7
PFC	1457760_at	4	131435283	131436411	0.7	1.04	10.8	4.00E-04	A930004J17Rik
PFC	1425173_s_at	3	95392880	95423164	1.03	0.82	10.4	4.90E-04	Golph3l
PFC	1419586_at	X	19941733	19977475	0.78	1.03	10.36	5.00E-04	Rp2h

FIG. 27-1

PFC	1453087_at	5	139475528	139476533	0.91	1.08	10.35	5.10E-04	6330403L08Rik
PFC	1420483_at	1	36568720	36585082	1.23	0.94	10.07	5.90E-04	Cnnm3
PFC	1436503_at	6	128489839	128531624	1.1	0.87	9.98	6.20E-04	BC048546
PFC	1417272_at	5	65361313	65433140	0.82	1.1	9.97	6.30E-04	Fam114a1
PFC	1442913_at	3	118242789	118243450	1.31	0.92	9.92	6.40E-04	NA
PFC	1437686_x_at	5	136750114	137043301	1.2	0.95	9.79	6.90E-04	Cux1
PFC	1429599_a_at	5	91360221	91450395	1.26	0.98	9.79	6.90E-04	Mthfd2l
PFC	1424291_at	8	96738500	96838966	1.23	0.97	9.77	7.00E-04	Nup93
PFC	1427587_at	7	6336027	6349347	0.98	1.22	9.48	8.30E-04	Zfp28
PFC	1446835_at	17	6109658	6110258	0.96	1.31	9.48	8.30E-04	Tulp4
PFC	1429647_at	1	89083240	89084978	0.93	1.25	9.38	8.80E-04	1700027L20Rik
PFC	1452817_at	1	180885169	181448134	0.94	1.3	9.37	8.80E-04	Smyd3
PFC	1446932_at	4	108470038	108470615	0.98	1.34	9.2	9.70E-04	NA
HPC	1424038_a_at	7	51745308	51751883	1.29	0.88	68.49	1.10E-10	2310044H10Rik
HPC	1440357_at	15	85537748	85537832	1.68	0.81	48.73	3.30E-09	(let-7b)
HPC	1435179_at	13	83867710	83875274	1.42	0.95	45.42	6.40E-09	C130071C03Rik (miR-9-2)
HPC	1435089_at	13	63116293	63400964	1.41	0.82	43.86	8.90E-09	201011101Rik (miR-23b)
HPC	1460033_at	1	196850302	196864096	1.71	0.86	42.71	1.10E-08	A330023F24Rik (miR-29c)
HPC	1427410_at	14	62221673	62301210	1.49	0.95	33.16	1.20E-07	Dleu2 (miR-15a)
HPC	1428562_at	11	75275040	75280192	1.33	0.92	30.51	2.40E-07	2210403K04Rik (miR-22)
HPC	1431094_at	12	110833610	110835408	1.2	0.87	25.24	1.20E-06	1110006E14Rik (miR-136)
HPC	1442913_at	3	118242789	118243450	1.39	0.84	22.06	3.50E-06	AK141880
HPC	1438838_at	X	100764843	100819093	1.3	0.93	19.9	7.70E-06	B230206F22Rik (miR-374)
HPC	1447298_at	13	83879857	83880314	1.29	0.98	19.9	7.70E-06	(miR-9-2)
HPC	1436467_at	9	41422406	41423268	1.2	0.83	15.31	5.10E-05	D230004N17Rik (miR-125b-1)
HPC	1424525_at	18	66033147	66046233	1.06	0.83	14.34	7.80E-05	Grp
HPC	1434730_at	7	86676773	86677477	1.4	0.95	13.99	9.20E-05	AI854517 (miR-9-3)
HPC	1460695_a_at	13	63116293	63400964	1.11	0.89	13.92	9.40E-05	201011101Rik

FIG. 27-2

HPC	1455965_at	1	173180552	173190053	0.75	1.02	13.5	1.20E-04	<i>Adams4</i>
HPC	1428792_at	2	170172490	170253345	0.8	1.03	13.37	1.20E-04	<i>Bcas1</i>
HPC	1458783_at	11	22753661	227545585	1.19	0.97	13.05	1.40E-04	<i>B3gnt2</i>
HPC	1459340_at	3	94771437	94772099	1.25	1	12.96	1.50E-04	<i>NA</i>
HPC	1453184_at	11	61497911	61523452	1.22	0.97	11.96	2.40E-04	<i>Fam83g</i>
HPC	1419148_at	10	126437764	126458050	1.15	0.87	11.45	3.20E-04	<i>Avil</i>
HPC	1430600_at	14	55555306	55558114	0.81	1.02	11.27	3.50E-04	<i>Cnrm5</i>
HPC	1445039_at	1	120955632	120956425	0.9	1.09	11.21	3.60E-04	<i>NA</i>
HPC	1426852_x_at	15	54577482	54585316	1.11	0.92	11.15	3.70E-04	<i>Nov</i>
HPC	1440109_at	7	54261880	54262492	0.88	1.07	10.94	4.10E-04	<i>D7Etd413e</i>
HPC	1436796_at	18	35721811	35751699	0.88	1.06	10.91	4.20E-04	<i>Matr3</i>
HPC	1454212_x_at	15	63712281	63720628	0.86	1.01	10.71	4.60E-04	<i>Gsdmcl2</i>
HPC	1432862_at	2	146909611	146911081	0.89	1.15	10.37	5.60E-04	<i>Nkx2-4</i>
HPC	1439943_at	11	21138891	21221136	0.8	1.01	10.27	5.90E-04	<i>Vps54</i>
HPC	1426651_a_at	15	54577482	54585316	1.12	0.92	9.98	6.90E-04	<i>Nov</i>
HPC	1449465_at	5	21390271	21850523	1.14	0.93	9.95	7.00E-04	<i>Reln</i>
HPC	1415951_at	11	100277007	100286153	0.95	1.2	9.84	7.50E-04	<i>Fkbp10</i>
HPC	1459493_at	18	79083283	79083877	1.21	0.95	9.83	7.50E-04	<i>Setbp1</i>
HPC	1419411_at	10	127162447	127168824	1.2	0.84	9.76	7.80E-04	<i>Tac2</i>
HPC	1435047_at	13	110844396	111070414	1.03	0.9	9.74	7.90E-04	<i>Rab3c</i>
HPC	1457030_at	12	110973190	110987665	1.22	0.8	9.73	7.90E-04	<i>Mirg</i>
HPC	1438748_at	19	60600598	60656926	1.06	0.85	9.62	8.40E-04	<i>2700078E11Rik</i>
HPC	1422703_at	X	82947275	83022158	0.98	1.15	9.39	9.60E-04	<i>Gyk</i>

* Bal = mouse strain balanced for copy number; Del = mouse model of the 22q11.2 microdeletion; Dup = mouse model of the 22q11.2 microduplication

FIG. 27-3

Gene Expression profile of predicted mir-185 targets that have Golgi related functions

Rank	AffylID	Gene Symbols	P.Value	logFC
31	1425690_at	B3gat1	3.94E-09	0.478
52	1420833_at	Vamp2	2.92E-07	0.607
66	1425691_at	B3gat1	6.47E-07	0.319
70	1421270_at	Sh3rf1	7.53E-07	0.391
117	1421890_at	St3gal2	3.97E-06	0.292
176	1423228_at	B4galt6	1.23E-05	0.286
192	1450105_at	Adam10	1.48E-05	0.28
199	1422758_at	Chst2	1.57E-05	0.296
201	1421271_at	Sh3rf1	1.58E-05	0.276
216	1450913_at	B4galt6	1.80E-05	0.291
340	1448367_at	Sdf4	5.34E-05	0.267
458	1450153_at	Gopc	9.82E-05	0.233
493	1421978_at	Gad2	1.16E-04	0.437
535	1431646_a_at	Stx6	1.42E-04	0.215
647	1422718_at	Ap3s2	2.27E-04	0.236
726	1450247_a_at	Scamp5	3.09E-04	0.255
730	1449111_a_at	Grb2	3.13E-04	0.217
734	1423720_a_at	Sar1a	3.16E-04	0.292
825	1428398_at	B3galt5	4.25E-04	0.224
845	1416437_a_at	Mapk8ip3	4.48E-04	0.238
914	1431320_a_at	Myo5a	5.23E-04	0.179
923	1427481_a_at	Atp1a3	5.33E-04	0.198
1018	1421891_at	St3gal2	6.61E-04	0.194
1029	1449943_at	Lfng	6.74E-04	0.312
1155	1422664_at	Rab10	8.79E-04	0.183
1270	1437107_at	Rab6b	1.08E-03	0.395
1304	1426744_at	Srebf2	1.14E-03	0.295
1338	1428374_at	Glce	1.20E-03	0.164
1354	1438373_at	App	1.22E-03	-0.21
1387	1431120_a_at	Golga1	1.28E-03	0.259
1430	1420643_at	Lfng	1.37E-03	0.283
1450	1419190_at	Vti1a	1.41E-03	0.217
1464	1424633_at	Camk1g	1.44E-03	0.161
1502	1443220_at	NA	1.50E-03	0.217
1589	1422044_at	Ndst1	1.68E-03	0.352
1694	1429296_at	Rab10	1.93E-03	0.166
1739	1433571_at	Serinc5	2.05E-03	-0.245
1805	1449129_a_at	Kcnip3	2.18E-03	0.161
1822	1420262_at	NA	2.25E-03	-0.203
2003	1448309_at	Ap3m1	2.74E-03	0.196
2042	1423075_at	Lman2	2.85E-03	0.17
2217	1431053_at	Mphosph9	3.37E-03	0.257

FIG. 28-1

SUBSTITUTE SHEET (RULE 26)

2246	1435758_at	B4galt6	3.45E-03	0.158
2257	1424755_at	Hip1	3.51E-03	0.264
2414	1416550_at	Slc35b4	4.08E-03	0.135
2516	1432017_at	Hip1	4.47E-03	-0.146
2712	1436321_at	B3gnt7	5.02E-03	-0.185
2740	1432054_at	Golga1	5.13E-03	0.244
2995	1451484_a_at	Syn1	6.13E-03	0.13
3025	1452174_at	Srebf2	6.30E-03	0.132
3093	1439853_at	B4galnt2	6.58E-03	-0.181
3148	1455924_at	Rab6b	6.85E-03	0.145
3281	1431066_at	Fut11	7.49E-03	0.187
3345	1450730_at	Hs2st1	7.76E-03	0.177
3358	1450104_at	Adam10	7.83E-03	0.144
3427	1457356_at	NA	8.17E-03	-0.17
3683	1416459_at	Arf2	9.41E-03	0.119
3710	1428397_at	B3galt5	9.58E-03	0.156
3722	1460191_at	Ykt6	9.65E-03	0.144
3788	1436155_at	Nmnat2	1.01E-02	0.145
3822	1457045_at	Galnt13	1.03E-02	0.251
3826	1423743_at	Arcn1	1.03E-02	0.148
3969	1448477_at	Chst12	1.10E-02	0.113
3986	1415670_at	Copg	1.11E-02	0.124
3998	1419754_at	Myo5a	1.12E-02	0.134
4049	1444943_at	NA	1.14E-02	-0.184
4089	1423358_at	Ece2	1.16E-02	-0.207
4101	1444413_at	Ap3s2	1.17E-02	0.255
4459	1436525_at	Ap3s2	1.39E-02	0.135
4584	1455986_at	Zdhhc17	1.48E-02	0.245
4608	1423229_at	Inpp5e	1.49E-02	0.119
4656	1436193_at	Man1c1	1.52E-02	0.099
4708	1435762_at	Pacs1	1.55E-02	0.153
4795	1453095_at	Rab10	1.60E-02	0.094
4866	1434945_at	Lpcat2	1.66E-02	-0.1
4905	1443398_at	NA	1.68E-02	0.308
5038	1426575_at	Sgms1	1.77E-02	0.185
5075	1449063_at	Sec22b	1.80E-02	0.125
5154	1440684_at	Lpcat2	1.85E-02	-0.121
5157	1460329_at	B4galt6	1.85E-02	0.086
5225	1443842_at	Arfgef2	1.90E-02	0.133
5582	1430966_at	Cml3	2.16E-02	-0.158
5633	1460070_at	NA	2.20E-02	0.13
5663	1455735_at	Ap1s3	2.22E-02	0.129
5664	1418508_a_at	Grb2	2.22E-02	0.101
5671	1425363_at	B4galnt1	2.22E-02	-0.122
5681	1422468_at	Ppt1	2.23E-02	0.133

FIG. 28-2

5811	1416549_at	Slc35b4	2.33E-02	0.107
5837	1422129_at	Apc2	2.36E-02	0.114
5882	1425539_a_at	Rtn3	2.40E-02	0.108
6004	1447834_at	NA	2.51E-02	-0.162
6169	1423167_at	Mobkl3	2.63E-02	0.117
6247	1445103_at	NA	2.70E-02	-0.101
6461	1454865_at	Slc9a8	2.88E-02	0.11
6526	1424112_at	Igf2r	2.94E-02	0.126
6637	1451837_at	Ap3b2	3.04E-02	-0.111
6756	1459801_at	B3gal5	3.16E-02	-0.185
6868	1457316_at	Mtap6	3.24E-02	0.133
6882	1428150_at	Coro7	3.26E-02	0.105
6950	1426704_at	Gak	3.33E-02	-0.118
6971	1426576_at	Sgms1	3.35E-02	0.113
7015	1426274_at	Slc9a8	3.39E-02	0.194
7049	1453221_at	Gopc	3.42E-02	0.095
7088	1450729_at	Hs2st1	3.45E-02	0.116
7283	1458363_at	Zdhhc17	3.61E-02	0.114
7303	1423152_at	Vapb	3.63E-02	0.314
7426	1429661_at	Rhobtb3	3.74E-02	0.173
7435	1431136_at	Rab36	3.75E-02	0.141
7450	1447012_at	Gm10791	3.76E-02	-0.127
7669	1459144_at	NA	3.97E-02	-0.093
7928	1426703_at	Gak	4.23E-02	0.092
7956	1418582_at	Cbfa2t3	4.26E-02	0.102
8007	1458501_at	Vapb	4.31E-02	0.169
8158	1428149_at	Coro7	4.48E-02	0.162
8286	1460322_at	Chst3	4.60E-02	-0.114
8611	1448308_at	Ap3m1	4.93E-02	0.094
8746	1419189_at	Vti1a	5.07E-02	0.079
8831	1450384_at	Bace1	5.15E-02	0.086
8939	1423038_at	Stx6	5.28E-02	0.078
8968	1442028_at	B4galnt2	5.30E-02	-0.138
9185	1458189_at	Emid2	5.57E-02	-0.111
9251	1448464_at	Ykt6	5.63E-02	0.087
9285	1439610_at	Rab27b	5.68E-02	0.128
9324	1441216_at	St3gal1	5.73E-02	-0.124
9331	1421825_at	Bace1	5.74E-02	0.116
9491	1425876_a_at	Glce	5.92E-02	0.174
9552	1424856_at	Atp1a3	5.98E-02	-0.098
9608	1442234_at	Chst2	6.05E-02	0.127
9850	1416548_at	Slc35b4	6.36E-02	0.098
9907	1435718_at	Ap3s2	6.42E-02	0.069
9932	1416375_at	Ap3m1	6.45E-02	0.163
10029	1421892_at	St3gal2	6.58E-02	0.117

FIG. 28-3

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10410	1424708_at	Tmed10	7.12E-02	0.159
10733	1416374_at	Ap3m1	7.55E-02	0.087
10817	1450509_at	Chst11	7.66E-02	0.126
10868	1454821_at	B3gat1	7.75E-02	0.077
10928	1436676_at	Mapk8ip3	7.84E-02	-0.092
11304	1435199_at	Apc2	8.31E-02	0.182
11552	1415958_at	Slc2a4	8.65E-02	-0.131
11608	1452471_at	Il17rd	8.73E-02	-0.13
11746	1443857_at	Hook3	8.90E-02	0.138
11832	1436471_at	Rab36	9.03E-02	-0.073
11896	1445688_at	NA	9.12E-02	-0.093
11919	1419909_at	Mphosph9	9.17E-02	-0.094
12475	1433630_at	Map6d1	9.98E-02	0.068
12508	1428044_at	Ap3s2	1.00E-01	-0.102
12518	1428902_at	Chst11	1.00E-01	0.084
12643	1417215_at	Rab27b	1.02E-01	0.104
12745	1460083_at	Adam10	1.04E-01	-0.073
12789	1423759_a_at	Tmco1	1.04E-01	0.078
12811	1435801_at	Fktn	1.05E-01	0.101
12906	1427020_at	Scara3	1.06E-01	0.125
12922	1430549_at	Bet1l	1.07E-01	0.107
13027	1425316_at	B3gat1	1.08E-01	-0.106
13056	1446049_at	NA	1.09E-01	-0.096
13109	1418946_at	St3gal1	1.09E-01	0.091
13608	1453227_at	Rhobtb3	1.17E-01	0.105
13625	1437748_at	Fut11	1.17E-01	0.103
13687	1418929_at	Ift57	1.18E-01	0.084
13889	1419186_a_at	St8sia4	1.22E-01	-0.075
13935	1447656_at	Zdhhc17	1.22E-01	-0.109
13967	1418655_at	B4galnt1	1.23E-01	0.106
13968	1423647_a_at	Zdhhc3	1.23E-01	0.063
14000	1448404_at	Scamp2	1.23E-01	0.139
14073	1438008_at	Gga3	1.25E-01	0.094
14264	1440011_at	NA	1.27E-01	-0.176
14265	1445374_at	NA	1.28E-01	-0.07
14657	1451036_at	Spg21	1.34E-01	0.105
14708	1420928_at	St6gal1	1.35E-01	0.093
14715	1428367_at	Ndst1	1.35E-01	0.092
14960	1439094_at	Cltc	1.39E-01	-0.116
15016	1450528_at	B3galt5	1.40E-01	0.129
15062	1451017_at	Ergic3	1.40E-01	-0.064
15117	1421853_at	Psen1	1.41E-01	0.072
15173	1450844_at	Stx6	1.43E-01	0.06
15267	1452657_at	Ap1s2	1.44E-01	0.054
15301	1438063_at	Mphosph9	1.45E-01	0.107
15413	1458296_at	NA	1.47E-01	-0.147

FIG. 28-4

15447	1443881_at	Pofut1	1.48E-01	0.064
15609	1442463_at	NA	1.50E-01	0.101
15617	1456655_at	NA	1.50E-01	-0.09
15685	1424111_at	Igf2r	1.52E-01	0.062
15742	1442904_at	Chst2	1.53E-01	-0.144
16001	1415766_at	Sec22b	1.57E-01	0.075
16220	1439922_at	Prrc1	1.61E-01	0.117
16277	1421155_at	B3galt6	1.62E-01	0.067
16372	1424707_at	Tmed10	1.64E-01	0.063
16416	1435679_at	Optrn	1.65E-01	-0.075
16561	1418544_at	Kcnip3	1.67E-01	0.055
16664	1447347_at	NA	1.69E-01	-0.117
16694	1454982_at	Arfgef2	1.69E-01	0.064
16773	1435982_at	Stx12	1.71E-01	-0.065
17109	1434123_at	Fut11	1.77E-01	-0.052
17199	1440585_at	Stx6	1.78E-01	-0.086
17742	1429778_at	Optrn	1.90E-01	0.114
18074	1420621_a_at	App	1.96E-01	0.054
18150	1417327_at	Cav2	1.97E-01	0.093
18184	1440014_at	NA	1.98E-01	-0.102
18225	1455368_at	Zdhhc3	1.99E-01	0.074
18374	1440979_at	Igf2r	2.02E-01	-0.047
18470	1446737_a_at	Hook3	2.04E-01	0.095
18685	1436051_at	Myo5a	2.08E-01	0.057
18823	1442079_at	Sgms1	2.11E-01	-0.058
18878	1416086_at	Tpst2	2.12E-01	0.059
19009	1423168_at	Mobk13	2.14E-01	0.046
19055	1425549_at	Psen1	2.15E-01	0.07
19058	1420832_at	Qsox1	2.15E-01	-0.061
19193	1448936_at	Stx12	2.19E-01	0.054
19621	1440963_at	Cbfa2t3	2.28E-01	-0.069
19946	1445009_at	NA	2.34E-01	-0.072
20048	1420016_at	Ppt1	2.37E-01	-0.063
20091	1426393_a_at	Sdf4	2.37E-01	0.066
20302	1426886_at	Cln5	2.42E-01	-0.072
20382	1440915_at	Mphosph9	2.44E-01	0.108
20406	1422467_at	Ppt1	2.44E-01	0.057
20483	1438566_at	St8sia6	2.46E-01	-0.08
20589	1427442_a_at	App	2.49E-01	0.041
20638	1431325_at	Cml3	2.50E-01	0.083
20681	1420112_at	NA	2.51E-01	-0.054
20768	1444275_at	NA	2.52E-01	-0.07
21064	1452515_a_at	Xylt2	2.60E-01	-0.063
21300	1420927_at	St6gal1	2.65E-01	0.075
21507	1446617_at	NA	2.70E-01	-0.066

FIG. 28-5

SUBSTITUTE SHEET (RULE 26)

21565	1460436_at	Ndst1	2.72E-01	0.05
21605	1458920_at	NA	2.73E-01	0.116
21742	1415696_at	Sar1a	2.76E-01	0.057
21867	1418129_at	Dhcr24	2.80E-01	0.137
21870	1455584_at	Sdf4	2.80E-01	0.053
22017	1432230_at	Hip1	2.83E-01	-0.069
22036	1446696_at	Ift57	2.84E-01	-0.123
22131	1434397_at	Zdhhc17	2.86E-01	0.051
22165	1455741_a_at	Ece1	2.87E-01	0.05
22331	1436192_at	Arfgef2	2.91E-01	0.058
22341	1460431_at	Gcnt1	2.91E-01	0.095
22363	1440905_at	Hs2st1	2.91E-01	-0.077
22645	1436664_a_at	Slc35a2	2.98E-01	0.042
22797	1425031_at	Fktn	3.02E-01	0.051
22906	1451215_at	Prrc1	3.05E-01	0.048
23037	1451089_a_at	Arcn1	3.08E-01	0.035
23265	1439876_at	Vti1a	3.14E-01	-0.061
23269	1430904_at	Arfgap3	3.14E-01	-0.075
23324	1436062_at	Arcn1	3.15E-01	0.045
23558	1436956_at	NA	3.22E-01	-0.053
23566	1436101_at	NA	3.22E-01	0.044
23601	1453858_at	Slc35a2	3.23E-01	-0.044
23760	1424746_at	Kif1c	3.27E-01	0.069
23774	1439877_at	NA	3.27E-01	-0.098
23978	1449538_a_at	Gcnt1	3.32E-01	-0.06
24169	1458524_at	Fndc3a	3.37E-01	-0.073
24233	1457968_at	NA	3.38E-01	0.064
24321	1440457_at	NA	3.41E-01	-0.056
24330	1450137_at	Pofut1	3.41E-01	0.054
24445	1421609_a_at	Cml3	3.44E-01	0.048
24546	1422687_at	Nras	3.47E-01	0.041
24588	1418774_a_at	Atp7a	3.48E-01	-0.077
24690	1435157_at	Hook3	3.51E-01	0.034
24721	1432631_at	Prrc1	3.51E-01	-0.06
24887	1434316_at	NA	3.56E-01	-0.055
24932	1420821_at	Sgpp1	3.57E-01	-0.1
24999	1440340_at	B3galt6	3.59E-01	-0.062
25088	1434557_at	Hip1	3.61E-01	0.055
25240	1425128_at	B3gnt8	3.65E-01	-0.075
25460	1439433_a_at	Slc35a2	3.71E-01	0.074
25610	1422622_at	Nos3	3.75E-01	0.076
25660	1458342_at	Tmem90a	3.76E-01	0.078
25735	1443863_at	Fndc3a	3.78E-01	-0.039
25921	1424747_at	Kif1c	3.82E-01	-0.043
25932	1426903_at	Fndc3a	3.83E-01	0.038

FIG. 28-6

SUBSTITUTE SHEET (RULE 26)

26127	1455826_a_at	Bace1	3.88E-01	0.055
26230	1446314_at	Gcnt7	3.92E-01	-0.053
26239	1450399_at	Psen1	3.92E-01	0.035
26692	1423074_at	Lman2	4.05E-01	0.036
26788	1453393_a_at	Chst4	4.07E-01	-0.037
26884	1417730_at	Ext1	4.10E-01	0.046
27080	1425955_at	Cav2	4.15E-01	-0.053
27188	1434914_at	Rab6b	4.18E-01	0.033
27310	1424894_at	Rab13	4.20E-01	0.037
27493	1444873_at	NA	4.25E-01	-0.048
27548	1439899_at	Galnt13	4.27E-01	-0.06
27767	1458608_at	NA	4.33E-01	-0.046
27770	1423230_at	Inpp5e	4.33E-01	0.055
27794	1444705_at	NA	4.34E-01	-0.071
28091	1439417_at	Qsox1	4.43E-01	0.051
28164	1416828_at	Snap25	4.45E-01	-0.035
28323	1440386_at	Glce	4.50E-01	-0.038
28342	1459654_at	NA	4.50E-01	0.043
28620	1444563_at	NA	4.58E-01	-0.045
28728	1431761_at	Entpd4	4.61E-01	-0.048
28771	1441423_at	Ece1	4.62E-01	-0.048
28825	1426372_a_at	Bet1l	4.64E-01	0.026
29026	1451224_at	Scamp5	4.70E-01	0.031
29052	1444361_at	Ap1s2	4.71E-01	0.05
29079	1428147_at	Coro7	4.71E-01	0.038
29100	1420902_at	St6galnac3	4.72E-01	0.065
29292	1422550_a_at	Mtap6	4.78E-01	0.033
29339	1438661_a_at	Arf2	4.79E-01	0.054
29538	1439196_at	Hook3	4.85E-01	0.045
29727	1454626_at	Cltc	4.91E-01	0.025
29739	1458084_at	Zdhhc17	4.91E-01	-0.029
30054	1454077_at	Vti1a	5.00E-01	-0.03
30521	1418195_at	Galnt10	5.12E-01	0.056
30636	1458897_at	Ust	5.16E-01	0.036
30681	1422980_a_at	Bet1l	5.18E-01	0.038
31145	1459097_at	NA	5.32E-01	0.038
31455	1427617_at	Fut10	5.41E-01	-0.032
31508	1435027_at	Golga1	5.43E-01	0.022
31615	1459707_at	NA	5.46E-01	-0.027
31807	1426325_at	Kif1c	5.51E-01	0.034
32077	1444102_at	NA	5.60E-01	0.033
32304	1438355_at	Tmem90a	5.67E-01	0.029
32464	1446624_at	Fktn	5.72E-01	-0.037
32703	1421522_at	B4galnt2	5.79E-01	-0.038
32779	1439218_at	NA	5.81E-01	-0.049

FIG. 28-7

32833	1415959_at	Slc2a4	5.83E-01	-0.038
32882	1437388_at	Fut10	5.84E-01	0.035
32946	1416591_at	Rab34	5.86E-01	-0.028
32960	1416590_a_at	Rab34	5.87E-01	-0.024
33007	1416760_at	Galnt1	5.88E-01	0.023
33138	1432819_at	Prrc1	5.93E-01	-0.032
33158	1416611_at	Scamp2	5.94E-01	0.022
33175	1447235_at	NA	5.94E-01	0.033
33517	1455149_at	Sh3rf1	6.04E-01	0.035
33813	1432818_at	Prrc1	6.14E-01	0.029
34235	1421824_at	Bace1	6.27E-01	0.018
34412	1445505_at	Ndst1	6.32E-01	0.023
34607	1436865_at	Slc26a11	6.39E-01	-0.032
34680	1431549_at	NA	6.41E-01	-0.038
34744	1416017_at	Copg	6.44E-01	0.022
34779	1436499_at	Sgms1	6.45E-01	0.026
34819	1452365_at	Csgalnact1	6.47E-01	0.027
34832	1454924_at	Fut10	6.47E-01	0.027
34887	1451825_a_at	Copz1	6.49E-01	-0.017
35005	1441133_at	NA	6.52E-01	-0.023
35022	1429589_at	Gad2	6.52E-01	-0.019
35229	1429652_at	Prrc1	6.59E-01	0.026
35272	1430391_a_at	St8sia4	6.60E-01	0.029
35397	1453032_at	Mobkl3	6.64E-01	0.019
35670	1420333_at	Txndc8	6.74E-01	0.028
35685	1441993_at	Ap3s2	6.75E-01	-0.019
35814	1458616_at	NA	6.80E-01	0.019
35954	1416199_at	Kifc3	6.84E-01	-0.021
36001	1444884_at	Ppt1	6.86E-01	0.019
36267	1445966_at	NA	6.95E-01	-0.029
36278	1418014_a_at	B4galt1	6.95E-01	0.022
36301	1421967_at	B4galt5	6.96E-01	0.025
36355	1441148_at	NA	6.98E-01	0.027
36408	1459970_at	NA	7.00E-01	-0.021
36487	1437998_at	Ap1s2	7.03E-01	-0.02
36607	1458773_at	NA	7.07E-01	-0.021
36699	1453929_at	Rnf24	7.10E-01	0.032
36742	1438252_at	Qsox1	7.12E-01	-0.014
36795	1425310_a_at	Emid2	7.13E-01	-0.021
36894	1417214_at	Rab27b	7.16E-01	0.022
37011	1454060_a_at	Nras	7.20E-01	0.022
37023	1445178_at	Sh3rf1	7.20E-01	0.05
37039	1451895_a_at	Dhcr24	7.21E-01	-0.015
37096	1420261_at	Psen1	7.23E-01	-0.015
37194	1422739_at	Hs2st1	7.26E-01	0.022

FIG. 28-8

37503	1420831_at	Qsox1	7.35E-01	-0.026
37522	1427923_at	Zmpste24	7.36E-01	0.018
37524	1419910_at	Mphosph9	7.36E-01	-0.019
37638	1424058_at	Prrc1	7.41E-01	-0.021
37757	1458054_at	Ext1	7.45E-01	-0.022
37768	1456147_at	St8sia6	7.45E-01	0.035
37836	1429893_at	Il17rd	7.47E-01	0.019
38090	1446739_at	NA	7.56E-01	0.023
38187	1458258_at	NA	7.60E-01	0.012
38215	1447763_at	NA	7.61E-01	-0.017
38284	1436797_a_at	Surf4	7.63E-01	0.016
38293	1440493_at	Galnt10	7.63E-01	-0.02
38312	1460036_at	Ap1s2	7.64E-01	-0.052
38364	1440092_at	NA	7.66E-01	-0.028
38429	1444411_at	NA	7.68E-01	-0.025
38435	1419320_at	Chst5	7.69E-01	-0.02
38548	1418130_at	Dhcr24	7.73E-01	-0.015
38586	1436224_at	Kif1c	7.74E-01	0.019
38629	1423646_at	Zdhhc3	7.76E-01	-0.011
38767	1438705_at	Cbfa2t3	7.80E-01	0.017
38795	1442365_at	NA	7.81E-01	-0.019
38806	1440121_at	NA	7.82E-01	0.032
38913	1421517_at	St6galnac1	7.85E-01	-0.011
39243	1426927_at	Ap3b2	7.98E-01	-0.011
39548	1424561_at	Ece2	8.08E-01	-0.014
39772	1418101_a_at	Rtn3	8.15E-01	-0.008
40008	1424979_at	Aph1a	8.24E-01	0.014
40539	1423609_a_at	Mgat1	8.42E-01	0.007
40735	1415672_at	Golga7	8.48E-01	-0.008
40782	1456323_at	Pofut1	8.50E-01	-0.012
40839	1443082_at	NA	8.52E-01	-0.01
41035	1434862_at	Fut2	8.58E-01	-0.009
41074	1428103_at	Adam10	8.59E-01	-0.01
41243	1457009_at	Rhobtb3	8.65E-01	0.014
41365	1432533_a_at	Slc35a2	8.70E-01	0.009
41408	1436921_at	Atp7a	8.71E-01	0.016
41588	1446365_at	Vti1a	8.77E-01	-0.014
41613	1420834_at	Vamp2	8.78E-01	0.007
41628	1451853_at	Fktn	8.79E-01	0.007
41685	1452366_at	Csgalnact1	8.81E-01	0.008
41705	1435252_at	B3galt6	8.81E-01	0.008
41726	1438897_at	Zdhhc3	8.82E-01	-0.009
41824	1457583_at	NA	8.86E-01	-0.012
42111	1425975_a_at	Mapk8ip3	8.96E-01	-0.006
42167	1450246_at	Fut2	8.97E-01	0.006

FIG. 28-9

SUBSTITUTE SHEET (RULE 26)

42229	1448255_a_at	Surf4	9.00E-01	0.006
42245	1449432_a_at	Mmel1	9.00E-01	-0.006
42495	1443141_at	NA	9.08E-01	0.005
42617	1450497_at	Apc2	9.13E-01	-0.008
42710	1434177_at	Ece1	9.16E-01	-0.006
42987	1445466_at	Map6d1	9.26E-01	-0.007
43014	1445838_at	NA	9.26E-01	0.004
43082	1431941_at	Slc35b4	9.29E-01	0.005
43387	1431205_at	Slc9a8	9.40E-01	0.004
43455	1430932_at	Slc9a8	9.43E-01	0.004
43457	1416458_at	Arf2	9.43E-01	0.004
43643	1418194_at	Galnt10	9.50E-01	0.003
43797	1458179_at	NA	9.55E-01	0.003
43822	1422688_a_at	NA	9.56E-01	0.003
44117	1442402_at	Sh3rf1	9.66E-01	-0.003
44298	1455064_at	Rab36	9.72E-01	-0.002
44305	1445224_at	NA	9.73E-01	0.002
44314	1420903_at	St6galnac3	9.73E-01	-0.002
44475	1451554_a_at	Aph1a	9.78E-01	-0.004
44677	1424756_at	Hip1	9.85E-01	-0.001
44797	1449737_at	NA	9.89E-01	0.001
44798	1458525_at	NA	9.89E-01	-0.001
44883	1443581_at	St6gal1	9.92E-01	-0.001

FIG. 28-10