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(54) Title: COMPOSITIONS USEFUL IN THE PREVENTION OR TREATMENT OF SKIN CANCER

(57) **Abrégé/Abstract:**

The invention relates to compounds of general formula (I): $\text{CH}_3-(\text{CH}=\text{CH})_n-\text{R}$ (I) wherein $n = 3, 5, 7$; R is selected from $-\text{CO}-\text{OR}'$, $-\text{CO}-\text{O}^{(t)}$, or $-\text{CH}_2-\text{O}-\text{CO}-\text{R}'$, R' being selected from H, C_1-C_{22} alkyl or alkenyl, aryl or aralkyl, or sugars; with the proviso that when R is $-\text{CH}_2-\text{O}-\text{CO}-\text{R}'$ and R' alkyl, said alkyl being selected from C_1 to C_{11} ; and pharmaceutically acceptable salts thereof, preferably such as sodium, potassium, lysine salts, each compound of general formula (I) being used as such or in admixture with one or more of the other ones, for use in the prevention or treatment of skin cancer resulting from DNA damage produced by UV radiation.

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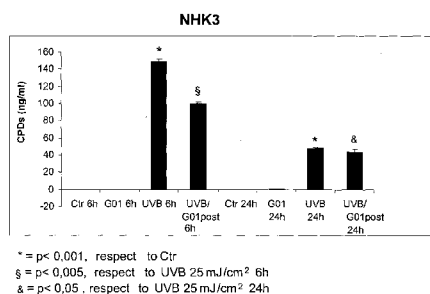
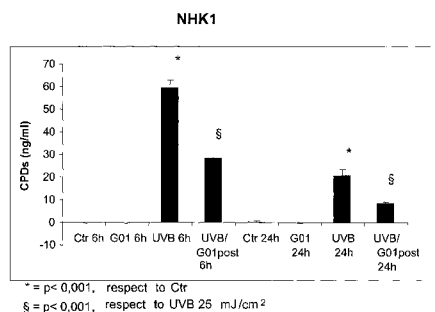


Fig. 7

(57) Abstract: The invention relates to compounds of general formula (I): CH₃-(CH=CH)_n-R (I) wherein n = 3, 5, 7; R is selected from -CO-OR', -CO-O⁽⁻⁾, or -CH₂-O-CO-R', R' being selected from H, C₁-C₂₂ alkyl or alkenyl, aryl or aralkyl, or sugars; with the proviso that when R is -CH₂-O-CO-R' and R' alkyl, said alkyl being selected from C₁ to C₁₁; and pharmaceutically acceptable salts thereof, preferably such as sodium, potassium, lysine salts, each compound of general formula (I) being used as such or in admixture with one or more of the other ones, for use in the prevention or treatment of skin cancer resulting from DNA damage produced by UV radiation.

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COMPOSITIONS USEFUL IN THE PREVENTION OR TREATMENT OF SKIN CANCER

Field of the Invention

The object of the present invention is a novel anticancer use of known
5 compounds, and compositions thereof which contain said compounds as active ingredient.

Background of the Invention

In each cell, the DNA is the repository of the genetic information; its stability and integrity are essential to life. The DNA is subject to environmental insults and any
10 consequent damage, if not repaired, leads to a mutation and, possibly, to the onset of a disease state.

Skin cancer from damage caused by excessive exposure to ultraviolet radiation is an example of the link between environmental damage induced to the DNA and the onset of a disease.

15 Another example is the DNA damage caused by tobacco smoke, which may thus lead to mutations in the lung cells and subsequent lung cancer.

In addition to the attack of environmental agents, the DNA is also subject to damage caused by metabolism by-products.

Whatever the origin of the harmful attack, it is estimated that a single cell can
20 suffer up to a million DNA changes in a day. In addition to the damage caused by environmental insults or metabolites, the process of DNA replication during cell division is subject to errors itself. The speed at which the DNA polymerase adds not correct nucleotides during the DNA replication is an important factor in determining the rate of spontaneous mutation in an organism.

25 Although the cell is equipped with enzymatic systems that normally recognize and correct many of these errors, some mutations remain. For a given gene, estimates of the frequency with which the human DNA undergoes errors that are not corrected are roughly between 1×10^{-4} to 1×10^{-6} mutations per gamete. A rate of 1×10^{-6} expresses the possibility of finding a mutation in a specific locus for a
30 million gametes.

The DNA repair processes are present in both prokaryote and eukaryote organisms. Many proteins involved in these processes are highly conserved during

evolution because of the importance of this repair function. The cells have developed several mechanisms to detect and correct the various types of damage that can occur to DNA, regardless of the source of damage. Because the DNA is a molecule that plays an active and critical role in cell division, the DNA repair is

5 closely linked to the regulation of the cell cycle.

During the cell cycle, in fact, control mechanisms (checkpoints) ensure that the DNA of a cell is intact before allowing that the DNA replication and the cell division happen. A defect in the checkpoints may lead to an accumulation of damages, which in turn lead to mutations.

10 Among the harmful environmental factors, for example, it is particular well-known that the ultraviolet radiation (UVA and UVB) is the main etiologic agent in the development of many skin cancers. It causes DNA damage through the formation of pyrimidine cyclobutane dimers (CPD) and pyrimidine(6-4)pyrimidone adducts, or 6-4 photoproducts (6-4 PP). Both compounds are able to modify the DNA

15 structure, thus preventing transcription and replication.

The phosphorylated form of histone γ H2AX, the p53 protein and the protein GADD45a (growth arrest and DNA damage response gene), are also cited as known markers of DNA damage induced by UV irradiation.

In addition, 8-oxo-2'-deoxyguanosine (8-oxo-dG or 8-OHdG), an oxidized

20 derivative of deoxyguanosine, is one of the major products of the DNA oxidation and, therefore, another typical marker of damage against it, particularly under exposure to UVA ultraviolet radiation.

The DNA repair is a physiological process that continuously operates in cells to protect the genome from damage and harmful mutations. The damage from CPD dimers and 6-4 PP photoproducts is counteracted through a process known as

25 nucleotide excision repair (NER). In eukaryotes this complex process may repair a damage that involves up to 30 nucleotides.

Defects to some of these genes, if not repaired, can lead for example to genodermatoses, such as xeroderma pigmentosum, as well as other serious pre-

30 cancerous dermatological disorders, *i.e.* characterized by a very high risk of developing skin cancer (about one thousand times greater than normal).

With reference to other technical problems and, therefore, in view of different uses, the same Applicant has in the past studied a class of dienes characterized by a general formula $\text{CH}_3(-\text{CH}=\text{CH})_n-\text{R}$ (wherein $n=2-7$) and, in this respect, is the holder of various publications.

5 WO2010/052328 suggests for these compounds, investigated with reference to retinol as a positive control, a pharmaceutical or cosmetic use to improve the human epidermis cell repair activity by means of the production of keratin mediated by cytokeratins cK-19. The transcriptional study on the expression of cK-19 showed an increase (up-regulation) of cK-19 in the final differentiation of the
10 human epidermis reconstructed *in vitro*.

WO2010/052329 describes for these compounds, through the definition of an inhibitory action of the enzyme 5α -reductase, a cosmetic use directed to promote the integrity of the skin through the effect of increasing the strength of the collagen and elastin.

15 WO2011/132177 suggests the use of some of these compounds to counteract the oxidizing action of the free radicals ROS and preserve the physiological conditions of the human epidermis.

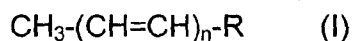
WO2012/007572 describes an activity of such compounds in the induction of melanogenesis through a non-receptor route. In particular, experiments performed
20 on primary melanocytes cultures showed the ability to induce the expression and activity of tyrosinase, the main melanogenic enzyme, and to increase the intracellular content of melanin.

None of these prior art documents relates to the behavior of these diene compounds with reference to the problem of human DNA damage caused by
25 environmental agents, such as those caused by excessive UV radiation may be. In fact, none of the experiments on which the evidence of activity in the uses as described in these prior art is based, contemplate, for example, UV irradiation, and therefore none of them measure the effects of UV radiation as a damage factor to human DNA, and thus as a carcinogen in case of skin cancers that develop
30 through an initial stage of DNA damage caused by UV radiation.

Summary of the Invention

Experimental studies, reported in the following paragraphs of the present description, have now surprisingly shown that for some of said compounds it is possible to obtain a significant activity of inhibition of the DNA damage produced by UV radiation, thereby envisaging a use for them as anticancer, both as
5 prevention and therapy, in the case of skin cancer induced by this kind of DNA damage.

Object of the invention are therefore compounds of general formula (I):



10 wherein $n = 3, 5, 7$; R is selected from $-\text{CO}-\text{OR}'$, $-\text{CO}-\text{O}^{(-)}$, or $-\text{CH}_2-\text{O}-\text{CO}-\text{R}'$, R' being selected from H, C_1 - C_{22} alkyl or alkenyl, aryl or aralkyl, or sugars; with the proviso that when R is $-\text{CH}_2-\text{O}-\text{CO}-\text{R}'$ and R' alkyl, said alkyl being selected from C_1 to C_{11} ;

and pharmaceutically acceptable salts thereof, preferably such as sodium,
15 potassium, lysine salts, each compound of general formula (I) being used as such or in admixture with one or more of the other ones, for use in the prevention or treatment of skin cancer resulting from DNA damage produced by UV radiation.

20 Among the dermatological disorders to be treated according to such use, for example, xeroderma pigmentosum and photocarcinogenesis are cited as non-limiting examples.

A secondary use, dependent on the anticancer one, particularly in the case of prevention of DNA damage caused by UV radiation, is envisaged as active
25 ingredient in compositions directed to the photoprotection of the skin from UVA and UVB, specifically to prevent DNA damage.

Therefore, a further object of the present invention is such photoprotective use dependent on the above described use.

The present invention also relates to any composition for the uses described
30 above in both the therapeutic and the prevention field, comprising as active ingredient a compound of general formula (I) as identified above, with any suitable excipient, particularly for topical application on the skin, including the scalp.

Detailed Description of the Invention

Some preferred compounds corresponding to the formula (I) for the use envisaged in the present invention are:

2,4,6 octatrienoic acid

5 2,4,6 octatrienoic acid sodium salt, or potassium salt

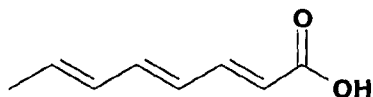
2,4,6-octatrienoic acid ethyl ester

2,4,6-octatrienoic acid L-lysine salt

2,4,6-octatrien-1-ol acetate

10 The following examples illustrate the invention without limiting its scope in any way.

The formula and the molecular weight for some of the compounds of general formula (I) are shown below.

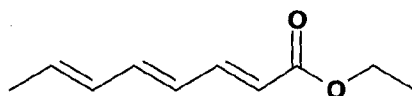


$C_8H_{10}O_2$ M.W. 138.17

15 2,4,6 octatrienoic acid

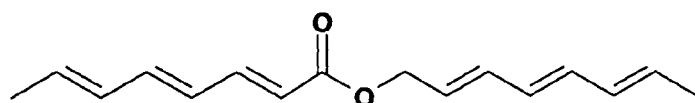
Sodium salt: $C_8H_9O_2Na$

Lysin salt: $C_8H_9O_2 \cdot C_6H_{15}N_2O_2$



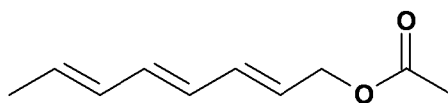
20 $C_{10}H_{14}O_2$ M.W. 166.22

2,4,6-octatrienoic acid ethyl ester



25 $C_{16}H_{20}O_2$ M.W. 244.34

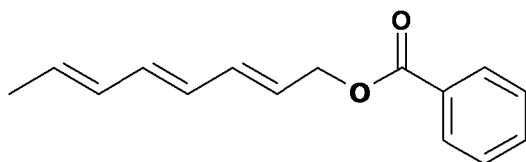
2,4,6-octatrienyl ester of 2,4,6-octatrienoic acid



$C_{10}H_{14}O_2$ M.W. 166.22

2,4,6-octatrien-1-ol acetate

5



$C_{15}H_{16}O_2$ M.W.228.29

2,4,6-octatrienyl ester of benzoic acid; 2,4,6-octatrien-1-ol, benzoate

In a composition according to the invention, said compound is present as active
10 ingredient in an amount preferably in the range from 10^{-6} to 3×10^{-2} mol/100g.

Description of the Drawings

Figs.1, 3, 6 show Western Blot immunofixation patterns, as described in more detail below.

15 Figs. 2, 4, 5, 7, 8, 9, 10, 11, 12 show diagrams relating to activity tests, as described in more detail below.

In the figures, the following meanings have to be understood:

Ctr = control

UVA or UVB = treatment with radiation only

20 G01 = treatment with compound of the invention only

G01/UVA = pre-treatment with compound of the invention, and subsequent UVA radiation

G01/UVB = pre-treatment with compound of the invention, and subsequent UVB radiation

25 UVA/G01post = UVA radiation and subsequent treatment with compound of the invention

UVB/G01post = UVB radiation and subsequent treatment with compound of the invention

Non-limiting examples of compositions particularly suitable for the above-stated use are given below.

The quantities of the components are expressed as percentages by weight:

5 Example 1

ANTICANCER DERMATOLOGICAL PREPARATION

	Component.....	Amount (% w/w)
	2-Octyldodecanol	5-20
	Cetostearyl alcohol.....	5-15
10	Cetyl esters wax	1-5
	Sorbitan monostearate	1-5
	Polisorbate 60	1-5
	Benzilic alcohol.....	0.1-1
	2,4,6-Octatrienoic acid	0.1-0.5
15	Purified water	q.s. to 100 mL

Example 2

DERMATOLOGICAL PREPARATION HIGH PROTECTION AGAINST UV DAMAGE

	Component.....	Amount (% w/w)
20	C ₂₀ -C ₂₂ alkyl phosphate.....	0.50-5.00
	C ₂₀ -C ₂₂ alcohols	0.50-5.00
	Cetearyl glucoside.....	0.10-2.00
	Tromethamine	0.10-0.80
25	Dibutyl adipate.....	1.00-7.00
	Ethylhexyl methoxycinnamate	1.00-10.00
	Isostearyl isostearate	2.00-8.00
	Ethylhexyl salicylate	1.00-5.00
	Butylene glycol cocoate.....	1.00-5.00
30	Butyl methoxydibenzoylmethane	1.00-5.00
	Diethylamino Hydroxybenzoyl Hexyl Benzoate	1.00-5.00
	Octocrylene	1.00-5.00

	Bis-Ethylhexyloxyphenol Methoxyphenyl Triazine	1.00-4.00
	Diethylhexyl syringylidene malonate	0.10-1.00
	2,4,6-Octatrienoic acid	0.05-0.50
	Lysine Octatrienoate	0.05-0.50
5	Phenoxyethanol.....	0.80-1.00
	Glycerin	1.00-5.00
	Disodium EDTA.....	0.01-0.09
	Tocopheryl acetate.....	0.05-1.00
	Potassium sorbate.....	0.005-0.10
10	Citric acid.....	q.s. to pH 5.5
	Purified water	q.s. to 100 g

Example 3

DERMATOLOGICAL PREPARATION FOR FACE PROTECTION

15	Component.....	Amount (% w/w)
	Xanthan gum	0.10-0.20
	Sodium hyaluronate	0.05-1.00
	Inositol	0.05-1.00
	Xylitol.....	0.05-1.00
20	Taurine	0.05-1.00
	Betaine	0.05-1.00
	C ₁₄ -C ₂₂ Alcohols	0.05-2.00
	C ₁₂ -C ₂₀ Alkyl glucoside	0.05-1.00
	Octyldodecanol.....	1.00-3.00
25	Ethylhexyl methoxycinnamate	0.50-3.00
	Octocrylene	0.10-3.00
	Butylene glycol	0.10-5.00
	Glycerin	1.00-4.00
	2,4,6-Octatrienoic acid	0.05-0.50
30	Phenoxyethanol.....	0.80-1.00
	Sodium hydroxide.....	0.001-0.20
	Citric acid.....	0.001-0.30

	Hydroxyethyl acrylate/Sodium acryloyldimethyl.....	0.01-2.00
	Taurate copolymer	
	Polyisobutene.....	0.01-1.50
	PEG-7 Trimethylolpropane Coconut Ether	0.01-1.00
5	Purified water	q.s. to100

Example 4

PROTECTIVE AND ANTICANCER DERMATOLOGICAL PREPARATION FOR THE SCALP

10	Component.....	Amount (% w/w)
	Ethanol	10.00-30.00
	Pentylene glycol	0.025-0.20
	Potassium Octatrienoate	0.05-0.50
	PEG-6 Caprylic/Capric Glycerides	0.10-1.00
15	Purified water	q.s. to100.00

Example 5

DERMATOLOGICAL PREPARATION WITHSUNSCREEN FOR SKIN PROTECTION

20	Component.....	Amount (% w/w)
	Bis-Ethylhexyloxyphenol Methoxyphenyl Triazine.....	1.00-4.00
	Octyldodecanol.....	1.00-6.00
	Denat. Alcohol Type D	1.00-15.00
	C ₁₂ -C ₁₅ Alkyl benzoate	1.00-15.00
25	Diethylamino Hydroxybenzoyl Hexyl Benzoate.....	1.00-5.00
	Caprylic/capric Triglyceride	1.00-60.00
	Ethylhexyl methoxycinnamate	0.50-10.00
	Octocrylene	0.10-5.00
	Simmondsia Chinensis Seed Oil	0.10-10.00
30	2,4,6-Octatrienoic acid	0.01-1.00

Example 6

DERMATOLOGICAL PREPARATION FOR THE TREATMENT OF THE FACE

Component.....	Amount (% w/w)
Propanediol	1.00-7.00
Xylitol.....	0.30-1.00
5 Cetearyl glucoside.....	0.10-2.00
Polyglyceryl-3 Rice Branate	0.10-3.00
Cetearyl alcohol.....	0.05-1.00
Disodium EDTA.....	0.01-0.10
2,4,6-Octatrienoic acidpotassium salt.....	0.05-0.50
10 C ₁₂ -C ₁₅ alkyl benzoate.....	1.00-5.00
Ethylhexyl methoxycinnamate	0.50-10.00
Octocrylene	0.50-5.00
Butyrospermum parkii butter	0.50-3.00
Citric acid.....	0.001-0.30
15 Simmondsia Chinensis seed oil.....	0.10-0.30
Hydrogenated Evening Primrose Oil	0.50-3.00
Octyldodecanol.....	0.50-3.00
Caprylic/Capric triglyceride.....	1.00-5.00
Isosterayl isostearate	0.10-5.00
20 Beta sitosterol.....	0.10-0.50
Delta tocoferol	0.05-0.20
Caprylyl glycol	0.05-0.50
1,2 Hexanediol	0.10-0.70
Sodium hydroxide.....	0.001-0.20
25 Phenoxyethanol.....	0.50-0.99
Perfume.....	0.05-0.50
Purified water	q.s. to 100.00

Example 7

30 DERMATOLOGICAL PREPARATION FOR SCALP SKIN

Component.....	Amount (% w/w)
Cyclopentasiloxane	1.00-50.00

	Denat. Alcohol Type C	1.00-15.00
	C ₁₂ -C ₁₅ alkyl benzoate	1.00-10.00
	Ethylhexyl methoxycinnamate	1.00-5.00
	Octocrylene	0.10-0.50
5	Disiloxane	1.00-49.00
	Perfume	0.05-0.30
	Oleyl erucate	0.50-3.00
	Dimethiconol	0.10-10.00
	2,4,6-Octatrienoic acid	0.01-1.00
10	Octyldodecanol	0.01-1.00

Example 8

PROTECTIVE TREATMENT CREAM

	Component	Amount (% w/w)
15	Propylene glycol	0.50-7.00
	Pentylene glycol	1.00-3.00
	Steareth-21	0.10-3.00
	Steareth-2	0.10-3.00
	Caprylic/capric triglyceride	1.00-10.00
20	Cyclopentasiloxane	0.50-20.00
	Cetearyl alcohol	0.01-2.00
	Octyldodecanol	0.10-5.00
	Disodium EDTA	0.01-0.10
	2,4,6-Octatrienoic acidL-lysine salt	0.05-1.00
25	Phenoxyethanol	0.80-1.00
	Sodium hydroxide	0.001-0.20
	Citric acid	0.001-0.30

An experimental study on cell cultures was conducted as described below, with reference to the figures in the attached drawings, to demonstrate the activity of the compounds of the present invention.

EXPERIMENTAL STUDY

The aim of the study is to evaluate the repair activity of UV induced DNA damage exercised by a compound according to said general formula (I), namely 2,4,6-trans-octatrienoic acid, identified in the description of the study hereinafter with the initials G01.

5 The study is performed on primary cultures of human keratinocytes, NHKs.

This activity has been evaluated in several respects, as follows.

Cell viability after treatment with G01

The cells (three different cultures of keratinocytes NHKs) were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the
10 plate. The cells were treated with eight different concentrations of the compound of the invention G01 under evaluation, diluted in the culture medium, for 48 ± 2 h. The values of cell viability for each concentration tested, resulting from the spectrophotometric reading, were compared with the value obtained from the control. The results obtained showed the absence of cellular toxicity in all eight
15 different concentrations.

1) Assessment of the activity in the pre-treatment with the compound of the invention

Cell viability after pre-treatment with G01 and subsequent UV irradiation

The cells were plated in the specific culture medium and maintained in culture for
20 24 ± 2 h to allow adhesion to the plate. The cells were treated for 24 ± 2 h with the concentrations of the compound G01 under evaluation, selected based on the previous results. Subsequently, the cells in the plates identified as +UVA and +UVB were respectively exposed, in the irradiation medium, to doses of UVA of 10 J/cm^2 and UVB of 25 mJ/cm^2 , while the control plate was maintained in the dark
25 and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium. The cell viability was measured (incorporation of NR) 24 ± 2 h and 48 ± 2 h after the irradiation, following examination of the morphology of the cells under a microscope, in the absence and the presence of treatment with G01 and/or UVA/UVB irradiation. The
30 cell viability values for each condition tested were compared with the value obtained from the control and plotted on a graph to highlight any differences between the various experimental conditions.

The results obtained showed a reduction in cell viability following irradiation with UVA and UVB, and a general protective effect of cell viability, particularly 48h after irradiation, if a pre-treatment with a compound of the invention G01 was present, particularly significant at the highest concentration tested (90 μ M).

5 Western Blot assessment of the ability to repair a DNA damage after pre-treatment with G01 and subsequent UV irradiation

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 \pm 2h to allow adhesion to the plate. Then, the keratinocytes were treated for 24 \pm 2h with the compound of the invention G01, at concentrations of 10 60 μ M and 90 μ M. Subsequently, the plates identified as +UVA, +G01/UVA, +UVB, and +G01/UVB were respectively exposed, in the irradiation medium, to doses of UVA of 10 J/cm² and UVB of 25 mJ/cm², while the control plates (Ctr) and those treated only with G01 were maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then 15 replaced with the culture medium. Based on the preliminary results obtained on a broader time range, a post-treatment time of 6h was selected, after which the plates were processed for Western Blot protein determination.

For each condition tested (treatment with G01 and/or UVA/UVB irradiation), the protein expression of the phosphorylated histone γ H2AX, p53 and GADD45a 20 (growth arrest and DNA damage response gene), known markers of DNA damage induced by UV radiation, was compared with the expression in the control cells (Ctr).

The results obtained by averaging the responses of three different keratinocytes cultures have shown the protective activity of the compound G01 under 25 examination, especially at the highest dose, as the cells pre-treated with G01 induced the protein expression of p53 and Gadd45a to a lesser extent. In confirmation of these results, the pre-treated keratinocytes show, after UVB irradiation, a reduced fraction of phosphorylated histone γ H2AX (fosfo γ H2AX), whose expression is also associated with the amount of DNA damage.

30 Fig.1 shows the Western Blot patterns drawn from experiments performed on one (NHK2) of the three different keratinocytes cultures described above, in which

GAPDH is the constitutively expressed housekeeping, used to normalize the values of the samples.

Flow cytometric assessment of UV induced apoptosis after a pre-treatment with G01 and subsequent UV irradiation

- 5 The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Each experimental condition was plated in duplicate. The cells were treated for 24 ± 2 h with G01, in a concentration of $90 \mu\text{M}$. Subsequently, the plates identified as +UVA, +G01/UVA, +UVB, and +G01/UVB were respectively exposed, in the irradiation medium, to doses of UVA of 25 J/cm^2 and UVB of 80 mJ/cm^2 , while the control plates (Ctr) and those treated only with the compound under examination (G01) were maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium. Based on the preliminary results obtained on a broader time range, a post-treatment time of 24h was selected, after which the plates were processed according to the procedure given in the user manual of the specific kit for annexin V flow cytometric analysis (Annexin V), a typical assay to detect the early stages of apoptosis. The positivity for annexin V in each condition tested (treatment with G01 and/or UVA/UVB irradiation) was compared with the positivity in the control cells.
- 10 of 25 J/cm^2 and UVB of 80 mJ/cm^2 , while the control plates (Ctr) and those treated only with the compound under examination (G01) were maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium. Based on the preliminary results obtained on a broader time range, a post-treatment time of 24h was selected, after which the plates were processed according to the procedure given in the user manual of the specific kit for annexin V flow cytometric analysis (Annexin V), a typical assay to detect the early stages of apoptosis. The positivity for annexin V in each condition tested (treatment with G01 and/or UVA/UVB irradiation) was compared with the positivity in the control cells.
- 15 was selected, after which the plates were processed according to the procedure given in the user manual of the specific kit for annexin V flow cytometric analysis (Annexin V), a typical assay to detect the early stages of apoptosis. The positivity for annexin V in each condition tested (treatment with G01 and/or UVA/UVB irradiation) was compared with the positivity in the control cells.
- 20 The results obtained have shown the ability of G01 to reduce the apoptosis induced by with UVA and UVB radiation.

Fig. 2 shows the corresponding diagrams drawn from the experiments performed on three different keratinocytes cultures, NHK1, NHK2 and NHK3.

Western Blot assessment of UV induced apoptosis after a pre-treatment with G01 and subsequent UV irradiation

- 25 The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Each experimental condition was plated in duplicate. The cells were treated for 24 ± 2 h with the compound G01, in a concentration of $90 \mu\text{M}$. Subsequently, the cells identified as +UVA, +G01/UVA, +UVB, and +G01/UVB were respectively exposed, in the irradiation medium, to doses of UVA of 25 J/cm^2 and UVB of 80 mJ/cm^2 , while the control plates and those treated only with G01 were maintained in the dark and at room temperature for a
- 30 +UVA, +G01/UVA, +UVB, and +G01/UVB were respectively exposed, in the irradiation medium, to doses of UVA of 25 J/cm^2 and UVB of 80 mJ/cm^2 , while the control plates and those treated only with G01 were maintained in the dark and at room temperature for a

period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium. Based on the preliminary results obtained on a broader time range, a post-treatment time of 24h was selected, after which the plates were processed for Western Blot protein determination. The protein expression of non-cleaved caspase 3 in each condition tested (treatment with G01 and/or UVA/UVB irradiation) was compared with the expression in the control cells (Ctr).

The results obtained have shown the ability of G01 to reduce caspase 3 activation (corresponding to a reduction of the protein expression of the zymogen form of the protein), one of the key mediators of the apoptosis process induced by UVA and UVB.

Fig. 3 shows the Western Blot patterns drawn from the experiments performed on three different keratinocytes cultures, NHK1, NHK2 and NHK3.

ELISA assessment of DNA damage after pre-treatment with G01 and subsequent UVB irradiation: CPDs determination

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Then, the keratinocytes were treated for 24 ± 2 h with the compound G01, in a concentration of $90 \mu\text{M}$. Subsequently, the plates identified as +UVB and +G01/UVB were exposed, in the irradiation medium, to a UVB dose of 80 mJ/cm^2 , while the control plates (Ctr) and those treated only with the compound under examination (G01) were maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium. Based on the preliminary results obtained on a broader time range, post-treatment times of 6h and 24h were selected, after which the plates were processed according to the procedure described in the work instruction specific for the DNA extraction. The determination of the pyrimidine cyclobutane dimers, CPDs, was performed using a specific ELISA kit following the procedure described in the user manual.

The results obtained have shown the ability of the compound G01 to promote the removal of CPDs adducts, known signal of DNA damage induced by UVB.

Fig. 4 shows the corresponding diagrams drawn from the experiments performed on three different keratinocytes cultures, NHK1, NHK2 and NHK3.

2) Assessment of the activity in the post-treatment with the compound of the invention

- 5 The following experiments were performed in order to assess the efficacy of compound G01 applied to NHKs cell cultures after irradiation with UVA and UVB.

Cell viability after UV irradiation and subsequent treatment with G01

The cells were plated in the specific culture medium and maintained in culture for 24 ± 2h to allow adhesion to the plate. The cells in the plates identified as +UVA and +UVB were respectively exposed, in the irradiation medium, to doses of UVA of 10 J/cm² and UVB of 25 mJ/cm², while the control plate (Ctr) was maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium, containing the compound G01 in concentrations selected based on the cell viability results obtained after treatment with the same compound (see above). The cell viability was measured (incorporation of NR) at a time of 24 ± 2h and 48 ± 2h after the irradiation, in the absence and the presence of UVA/UVB irradiation and/or treatment with the compound of the invention G01. The cell viability values for each condition tested were compared with the value obtained from the control and plotted on a graph to highlight any differences between the various experimental conditions.

The results obtained showed a reduction in cell viability following irradiation with UVA and UVB, and a significant protective action, particularly 48h after irradiation, of the highest concentration tested (90 μM) of the compound G01.

- 25 Flow cytometric assessment of apoptosis induced by UV irradiation and subsequent treatment with G01

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2h to allow adhesion to the plate. Each experimental condition was plated in duplicate. Subsequently, the plates identified as +UVA, +G01/UVA, +UVB, and +G01/UVB were respectively exposed, in the irradiation medium, to doses of UVA of 25 J/cm² and UVB of 80 mJ/cm², while the control plates (Ctr) and those treated only with the compound under examination (G01) were maintained

in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium containing the compound G01 in a concentration of 90 μ M. Based on the preliminary results obtained on a broader time range, a post-treatment time of 24h
5 was selected, after which the plates were processed according to the procedure given in the user manual of the specific kit for annexin V flow cytometric analysis. The positivity for annexin V in each condition tested (treatment with the compound of the invention under examination and/or UVA/UVB irradiation) was compared with the positivity in the control cells.

10 The results obtained have shown the ability of the compound G01 to reduce the apoptosis induced by irradiation with UVA and UVB.

Fig. 5 shows the corresponding diagrams drawn from the experiments performed on two different keratinocytes cultures, NHK1 and NHK2.

Western Blot assessment of apoptosis induced by UV irradiation and subsequent
15 treatment with G01

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 \pm 2h to allow adhesion to the plate. Subsequently, the plates identified as +UVA, +G01/UVA, +UVB, and +G01/UVB were respectively exposed, in the irradiation medium, to doses of UVA of 25 J/cm² and UVB of 80 mJ/cm², while
20 the control plates (Ctr) and those treated only with the compound under examination (G01) were maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium containing the compound G01, in a concentration of 90 μ M. Based on the preliminary results obtained on a broader time range, a
25 post-treatment time of 24h was selected, after which the plates were processed for Western Blot protein determination. The protein expression of non-cleaved caspase 3 in each condition tested (treatment with the compound G01 under examination and/or UVA/UVB irradiation) was compared with the expression in the control cells.

30 The results obtained have shown the ability of the compound of the invention to reduce caspase 3 activation (corresponding to a reduction of the protein

expression of the zymogen form of the protein), one of the key mediators of the apoptosis process induced by UVA and UVB.

Fig. 6 shows the Western Blot patterns drawn from the experiments performed on two different keratinocytes cultures, NHK1 and NHK3.

5 ELISA assessment of the damage induced by UVB irradiation and subsequent treatment with G01: CPDs determination

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Then, the plates identified as +UVB and +G01/UVB were respectively exposed, in the irradiation medium, to a UVB dose of 25 mJ/cm^2 , while the control plates (Ctr) and those treated only with the compound under examination (G01) were maintained in the dark and at room temperature for a period of time equal to the irradiation time. The irradiation medium was then replaced with the culture medium containing the compound G01, in a concentration of $90 \mu\text{M}$. Based on the preliminary results obtained on a broader time range, post-treatment times of 6h and 24h were selected, after which the plates were processed according to the procedure described in the work instruction specific for the DNA extraction. The CPDs determination was performed using a specific ELISA kit following the procedure described in the user manual.

20 The results obtained have shown the ability of the compound G01 under examination to promote the removal of CPDs products, known signal of DNA damage induced by UVB.

Fig. 7 shows the corresponding diagrams drawn from the experiments performed on two different keratinocytes cultures, NHK1 and NHK3.

25 ELISA assessment of the damage induced by UVA irradiation and subsequent treatment with G01: 8-OHdG determination

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Then, the plates identified as +UVA and +UVA/G01 post were respectively exposed, in the irradiation medium, to a UVA dose of 10 J/cm^2 . The irradiation medium was then replaced with the culture medium containing the compound of the invention G01, in a concentration of $90 \mu\text{M}$. Based on preliminary results obtained on a broader time range, post-treatment

times of 6h and 24h were selected, after which the plates were processed according to the procedure described in the work instruction specific for the DNA extraction. The determination of 8-OHdG, marker of DNA oxidative damage induced by UVA, was performed using a specific ELISA kit following the procedure described in the user manual.

The results obtained are summarized in the graph of Fig. 8. They show, in the cells irradiated and post-treated with the compound of the invention, a significant reduction in the amount of 8-OHdG produced by UVA radiation at both post-treatment times compared to the cells only irradiated and not treated with the compound.

Immunofluorescence assessment of the ability to repair a DNA damage following UVA or UVB irradiation and subsequent treatment with G01

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Subsequently, the plates identified as +UVA, +UVA/G01post, UVB, and +UVB/G01post were respectively exposed, in irradiation medium, to doses of UVA of 10 J/cm^2 or UVB of 25 mJ/cm^2 . The irradiation medium was then replaced with the culture medium containing the compound of the invention G01, in a concentration of $90 \mu\text{M}$. Based on preliminary results obtained on a broader time range, a post-treatment time of 24h was selected, after which the plates were processed for the immunofluorescence determination of the protein expression of the phosphorylated histone γH2AX , whose expression is associated to the amount of DNA damage (double strand breaks). The protein expression of the phosphorylated histone γH2AX after treatment with the compound of the invention G01, or only after irradiation with UVA/UVB, was compared with the expression in the control cells.

The results obtained are summarized in the graph of Fig. 9. They show, in the cells irradiated and post-treated with the compound of the invention, a significant reduction in the fraction of phosphorylated histone γH2AX compared to the cells only irradiated and not treated with the compound.

Western Blot assessment of the ability to repair a DNA damage following UVB irradiation and subsequent treatment with G01

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Subsequently, the cells identified as +UVB and +G01/UVB post were exposed, in the irradiation medium, to a UVB dose of 25 mJ/cm^2 . The irradiation medium was then replaced with the culture medium containing the compound of the invention G01, in a concentration of $90 \mu\text{M}$. Based on preliminary results obtained on a broader time range, a post-treatment time of 24 h was selected, after which the plates were processed for Western Blot protein determination. The protein expression of p53, phospho-p53, GADD45a, and phosphorylated histone γH2AX was measured in the condition of UVB irradiation.

Western Blot analysis at pre-set post-treatment times "time course" of the expression levels of p53, phospho-p53 and GADD45a after UVB irradiation and subsequent treatment with G01

The NHKs cells were plated in the specific culture medium and maintained in culture for 24 ± 2 h to allow adhesion to the plate. Subsequently, the plates identified as +UVB and +G01/UVB post were exposed, in the irradiation medium, to a UVB dose of 25 mJ/cm^2 . The irradiation medium was then replaced with the culture medium containing the compound of the invention G01, in a concentration of $90 \mu\text{M}$. Post-treatment times of 1-3-6-24 h were selected, after which the plates were processed for Western Blot protein determination. The protein expression of p53, phospho-p53, and GADD45a after UVB irradiation and post-treatment with the compound of the invention G01 was compared with the expression in the control cells (Ctr).

The results obtained are summarized in the diagrams of Figures 10 to 12, which represent the densitometries of the Western Blot electrophoretic patterns of immunofixation obtained at the above-specified times. They show that the post-treatment with the compound of the invention of cells exposed to UVB causes an acceleration of the increase in the levels of p53, phospho-p53, and GADD45a in the shorter times, resulting in a rapid activation of the DNA damage repair mechanisms. As final general result, this action causes a decrease in the expression of the markers of DNA damage at 24 h after UVB irradiation and, thus, a cytoprotective effect.

Overall, the experimental results obtained in the post-treatment of UV irradiation show, for the compound of the invention, an effective activity against DNA damage, similar to that demonstrated above in the pre-treatment before UV irradiation, particularly for what concerns key parameters such as preservation of cell viability, positivity to annexin V, inhibition of caspase 3 protein expression, CPDs and 8-OHdG inhibition, markers of DNA damage.

Moreover, in cells exposed to UVB irradiation, the post-treatment with compounds of the invention results in a timely activation of the DNA damage repair mechanisms. This results in a decrease of the expression of the markers of damage at 24h after the irradiation, and a cytoprotective effect.

Taken together, the experimental data therefore envisage an anticancer use of the compounds of general formula (I), both in the prevention of a cancer that may develop and in the treatment of an already developed cancer, as a result of a human DNA damaged induced by UV radiation.

As secondary use, dependent on the anticancer one, particularly in the case of prevention of DNA damage caused by UV radiation, a use as active ingredient is envisaged in compositions directed to the photoprotection of the skin, including the scalp skin, from UVA and UVB, to prevent DNA damage potentially developable in a cancer.

For this reason, a further object of the present invention is also such photoprotective use dependent on the one described above.

CLAIMS

1. Compound of general formula (I):



wherein $n = 3, 5$ or 7 ; R is $-\text{CO}-\text{OR}'$ or $-\text{CO}-\text{O}^{(-)}$, R' is H , C_1 - C_{22} alkyl, alkenyl, aryl, aralkyl or sugars,

or pharmaceutically acceptable salts thereof,

for use in the treatment of skin cancer resulting from DNA damage produced by UV radiation, wherein the compound is for topical administration to the skin in an amount sufficient to produce at least one of: (i) preservation of cell viability, (ii) positivity to annexin V, (iii) apoptosis inhibition, (iv) inhibition of caspase-3 protein expression and (v) inhibition of pyrimidine cyclobutane dimers and 8-oxo-2'-deoxyguanosine in human keratinocytes.

2. The compound for use according to claim 1, wherein the compound is for further use for photoprotection of the scalp from DNA damage produced by UV radiation.

3. The compound for use according to claim 1 or 2, wherein the pharmaceutically acceptable salts are selected from sodium, potassium and lysine salts.

4. A composition comprising a compound of general formula (I):



wherein $n = 3, 5$ or 7 ; R is $-\text{CO}-\text{OR}'$, $-\text{CO}-\text{O}^{(-)}$ or $-\text{CH}_2-\text{O}-\text{CO}-\text{R}'$, R' is H , C_1 - C_{22} alkyl, alkenyl, aryl, aralkyl or sugars,

or pharmaceutically acceptable salts thereof,

and an excipient,

for use in the treatment of skin cancer resulting from DNA damage produced by UV radiation, wherein the composition is for topical administration to the skin in an

amount sufficient to produce at least one of: (i) preservation of cell viability, (ii) positivity to annexin V, (iii) apoptosis inhibition, (iv) inhibition of caspase-3 protein expression and (v) inhibition of pyrimidine cyclobutane dimers and 8-oxo-2'-deoxyguanosine in human keratinocytes.

5. The composition for use according to claim 4, wherein the composition is for further use for photoprotection of the scalp from DNA damage produced by UV radiation.

6. The composition for use according to claim 4 or 5, wherein the pharmaceutically acceptable salts are selected from sodium, potassium and lysine salts.

7. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrienoic acid.

8. The composition for use according to any one of claims 4 to 6, wherein said compound is a 2,4,6-octatrienoic acid salt.

9. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrienoic acid sodium salt.

10. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrienoic acid potassium salt.

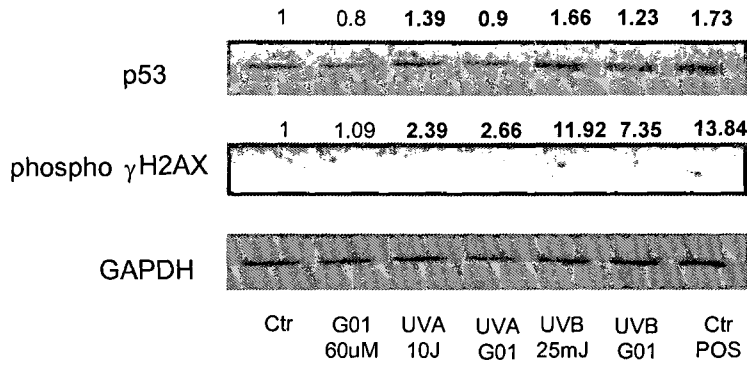
11. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrienoic acid L-lysine salt.

12. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrienoic acid ethyl ester.

13. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrienyl ester of 2,4,6-octatrienoic acid.

14. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrien-1-ol acetate.
15. The composition for use according to any one of claims 4 to 6, wherein said compound is 2,4,6-octatrien-1-ol benzoate.
16. The composition for use according to any one of claims 4 to 6, comprising a mixture of two or more compounds of formula (I).
17. The composition for use according to any one of claims 4 to 15, comprising said compound in an amount from 10^{-6} to 3×10^{-2} mol/100g.
18. The composition for use according to any one of claims 4 to 15, comprising said compound in an amount of 90 μ M.
19. The composition for use according to any one of claims 4 to 18, wherein the excipient is formulated for topical administration on the human skin or scalp.

NHK2



NHK2

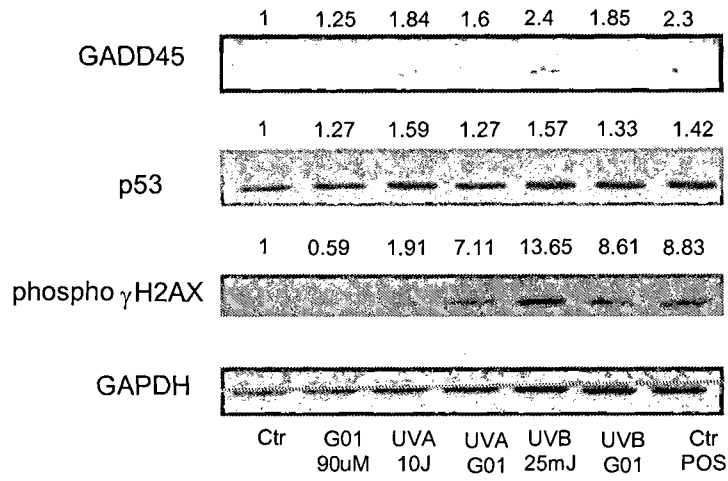
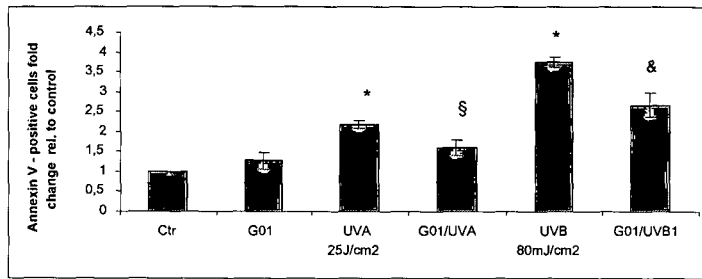


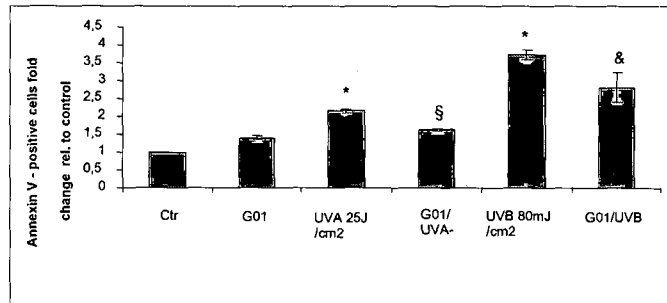
Fig. 1

2/10
NHK1



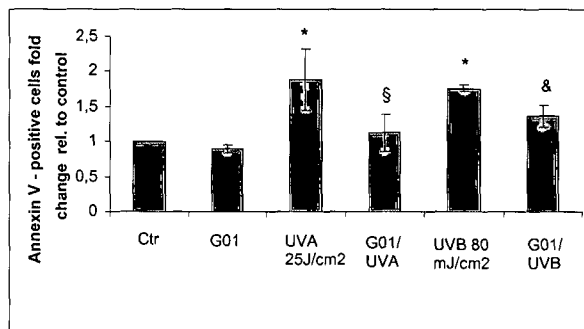
* = p < 0,01, respect to Ctr
 § = p < 0,05, respect to UVA 25 J/cm²
 & = p < 0,05, respect to UVB 80 mJ/cm²

NHK2



* = p < 0,01, respect to Ctr
 § = p < 0,05, respect to UVA 25 J/cm²
 & = p < 0,05, respect to UVB 80 mJ/cm²

NHK3



* = p < 0,01, respect to Ctr
 § = p < 0,05, respect to UVA 25 J/cm²
 & = p < 0,05, respect to UVB 80 mJ/cm²

Fig.2

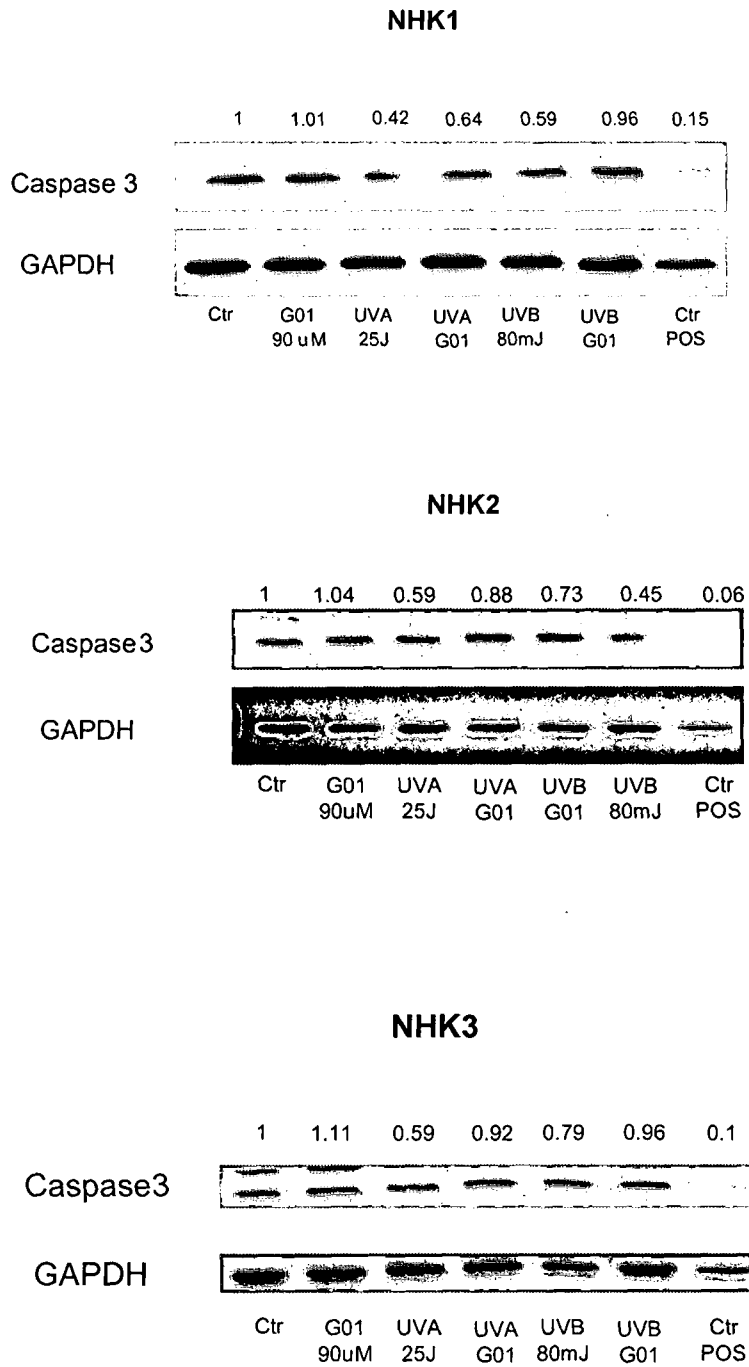
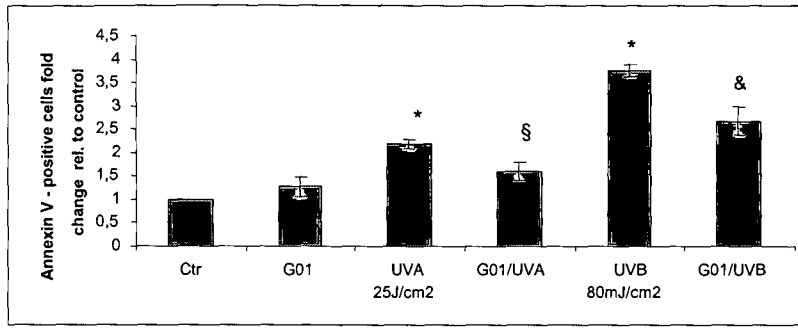


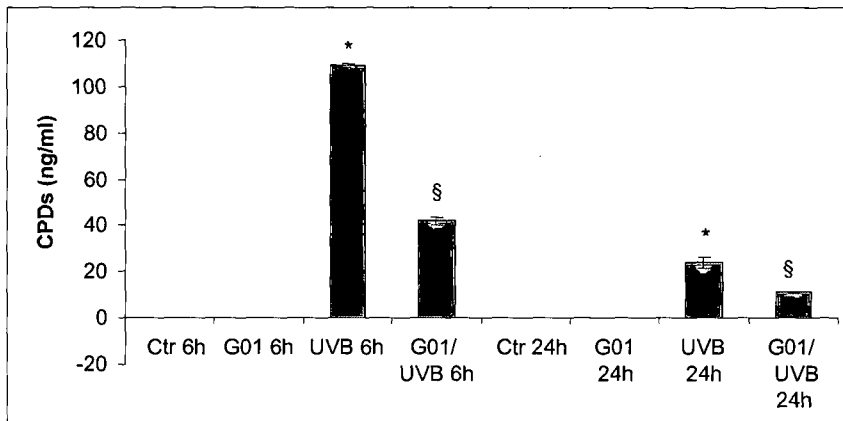
Fig. 3

4/10
NHK1



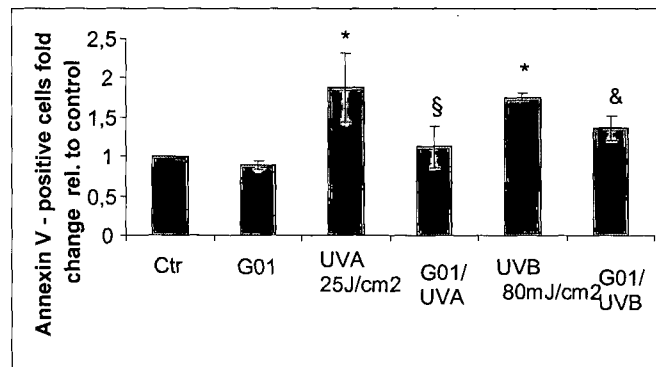
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 § = p < 0,05, respect to UVA 25 J/cm²
 & = p < 0,05, respect to UVB 80 mJ/cm²

NHK2



* = p < 0,001, respect to Ctrl
 § = p < 0,001, respect to UVB 25 mJ/cm²

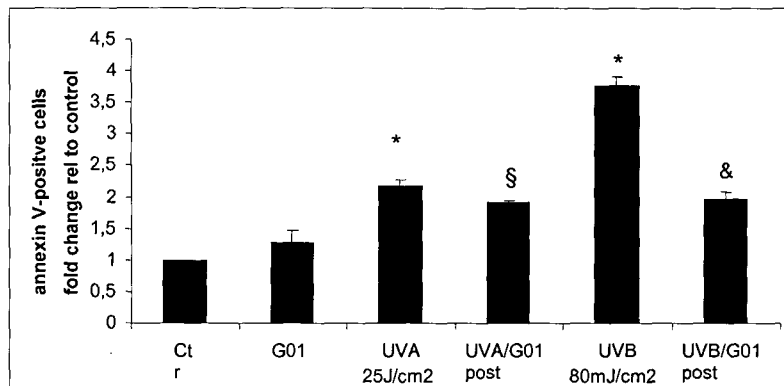
NHK3



* = p < 0,01, respect to Ctrl
 § = p < 0,05, respect to UVA 25 J/cm²
 & = p < 0,05, respect to UVB 80 mJ/cm²

Fig. 4

NHK1

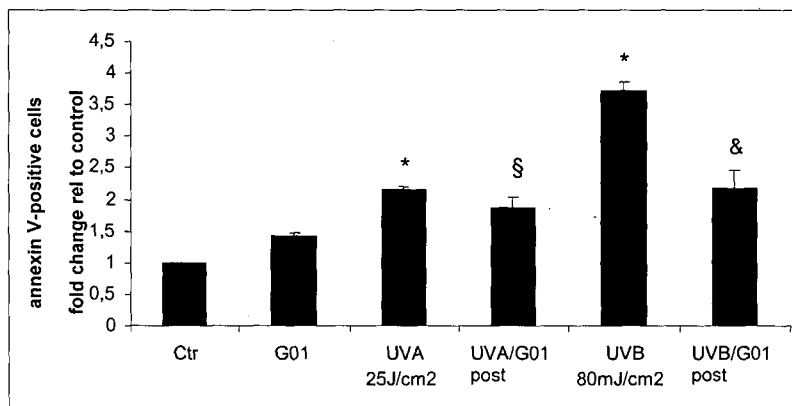


* = $p < 0,01$, respect to Ctr

§ = $p < 0,05$, respect to UVA 25 J/cm²

& = $p < 0,01$, respect to UVB 80 mJ/cm²

NHK2



* = $p < 0,01$, respect to Ctr

§ = $p < 0,05$, respect to UVA 25 J/cm²

& = $p < 0,01$, respect to UVB 80 mJ/cm²

Fig. 5

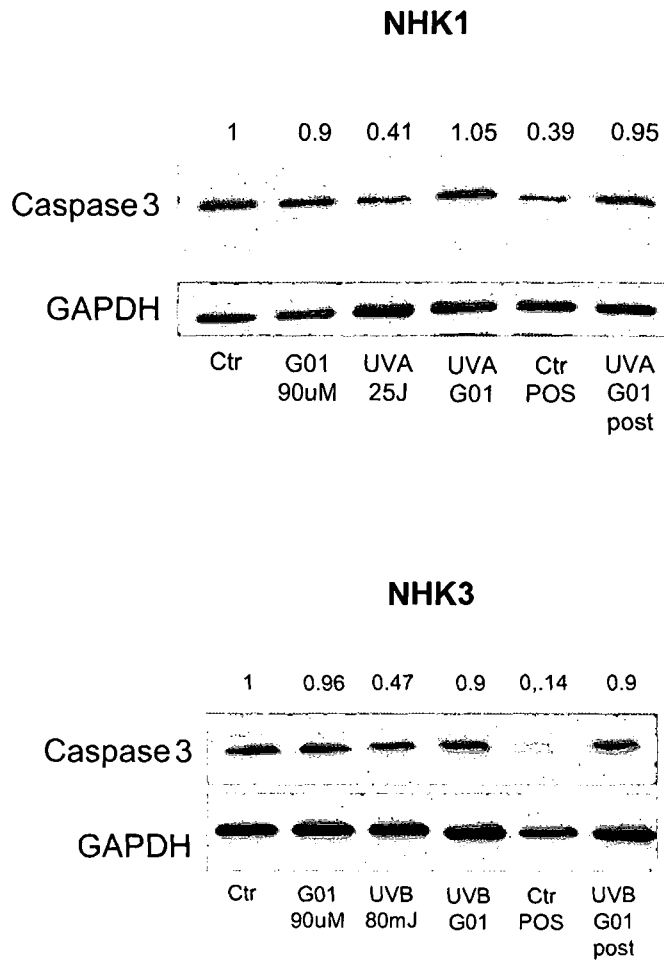
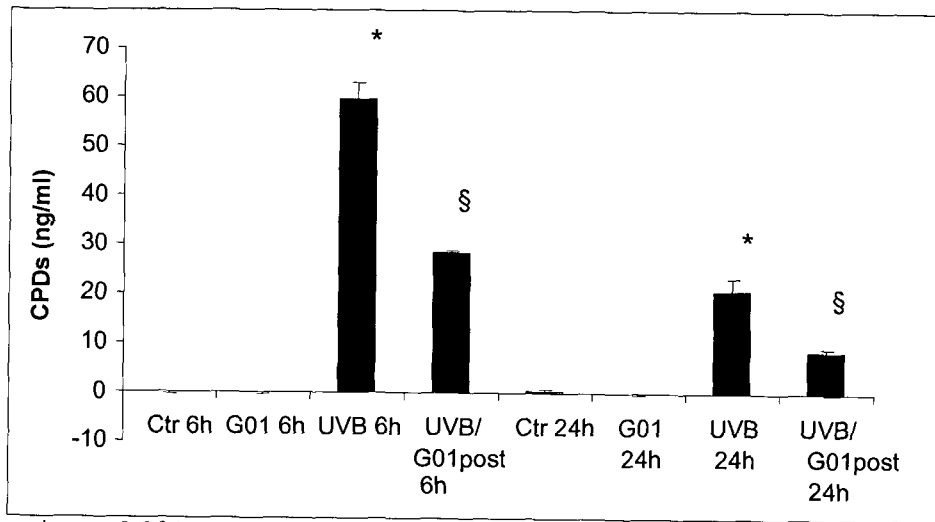
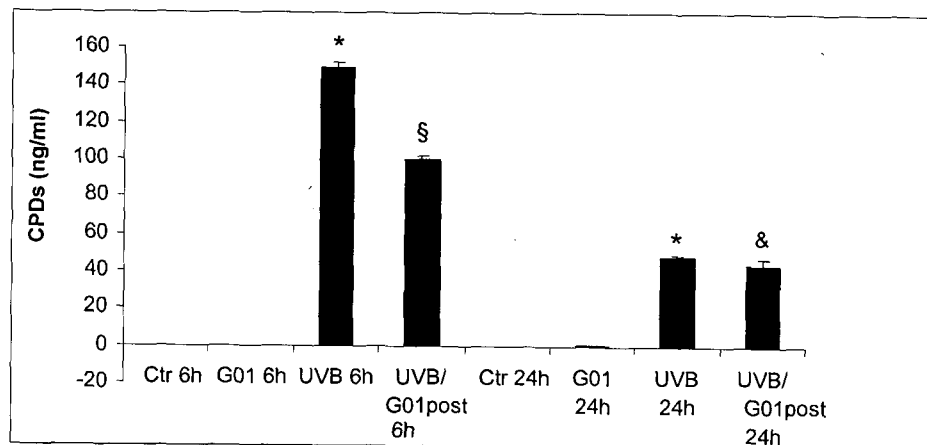


Fig. 6

7/10

NHK1* = $p < 0,001$, respect to Ctr§ = $p < 0,001$, respect to UVB 25 mJ/cm^2 **NHK3*** = $p < 0,001$, respect to Ctr§ = $p < 0,005$, respect to UVB 25 mJ/cm^2 6h& = $p < 0,05$, respect to UVB 25 mJ/cm^2 24h**Fig. 7**

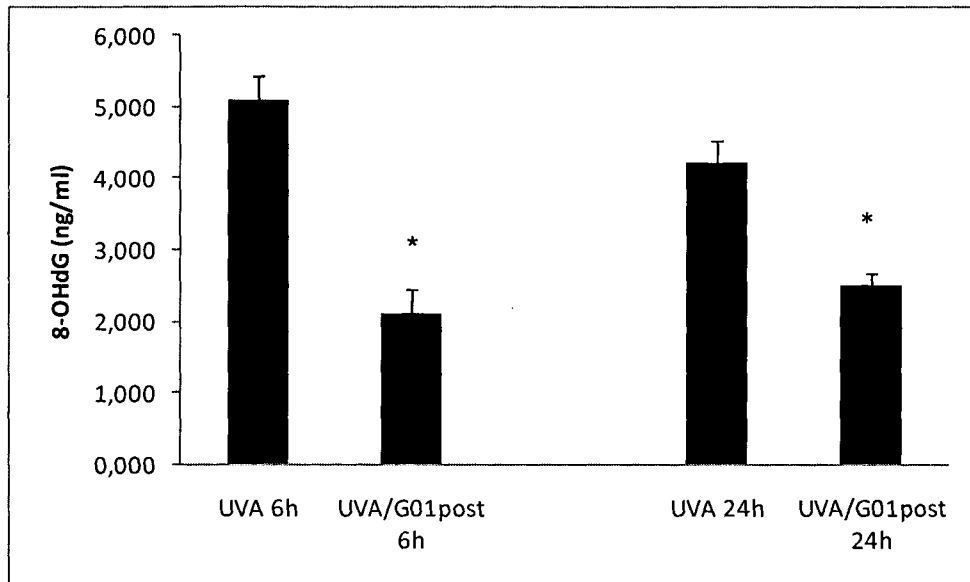


Fig. 8

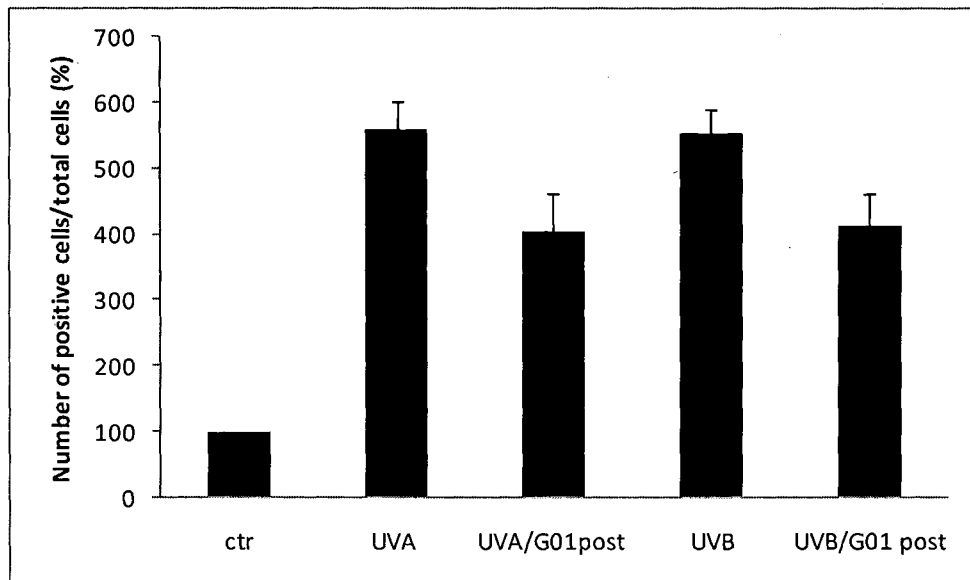


Fig. 9

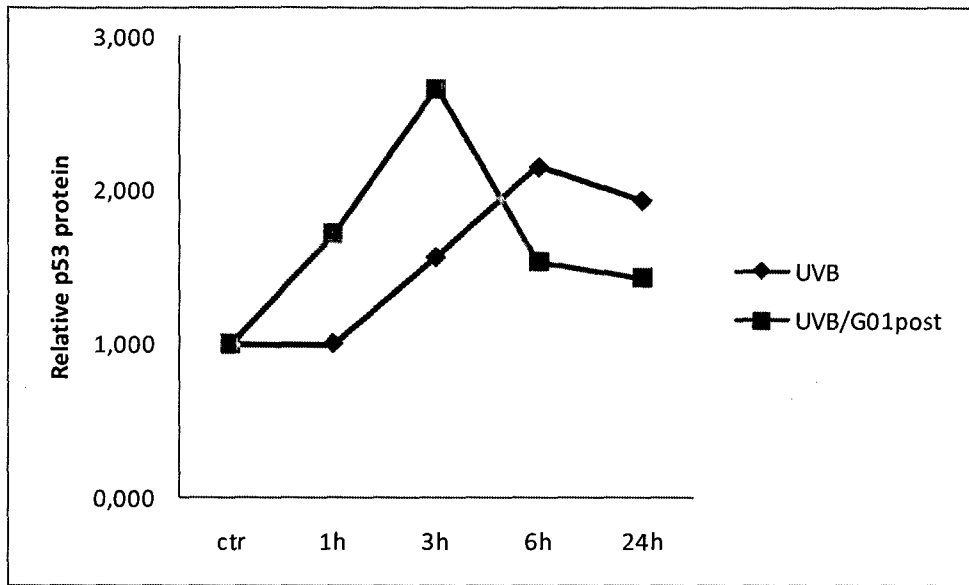


Fig. 10

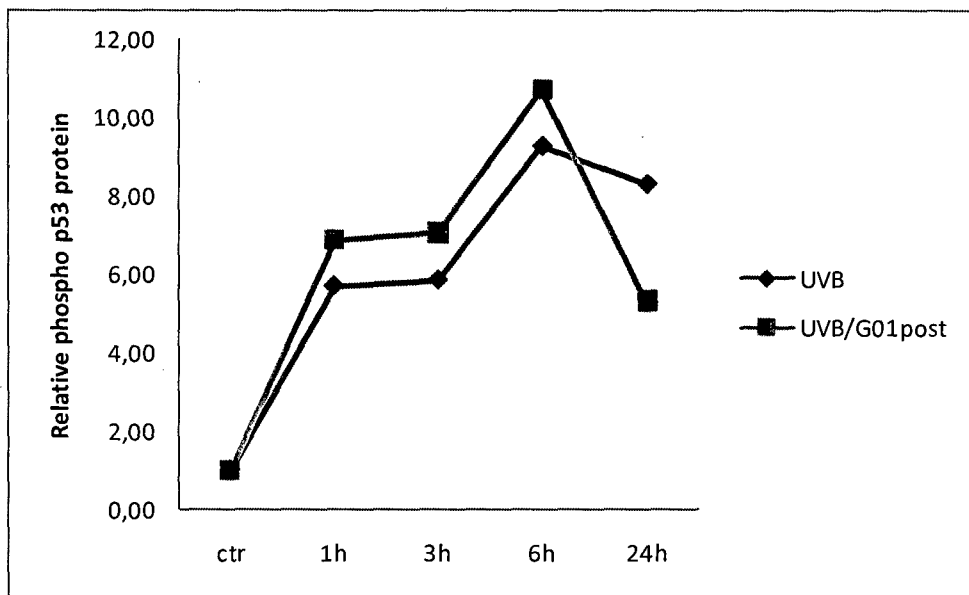


Fig. 11

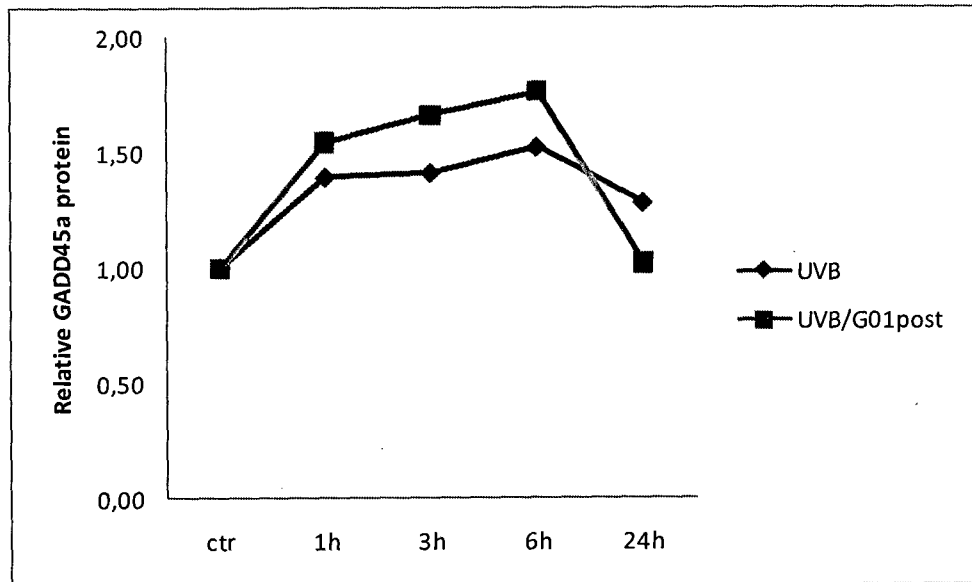


Fig. 12