

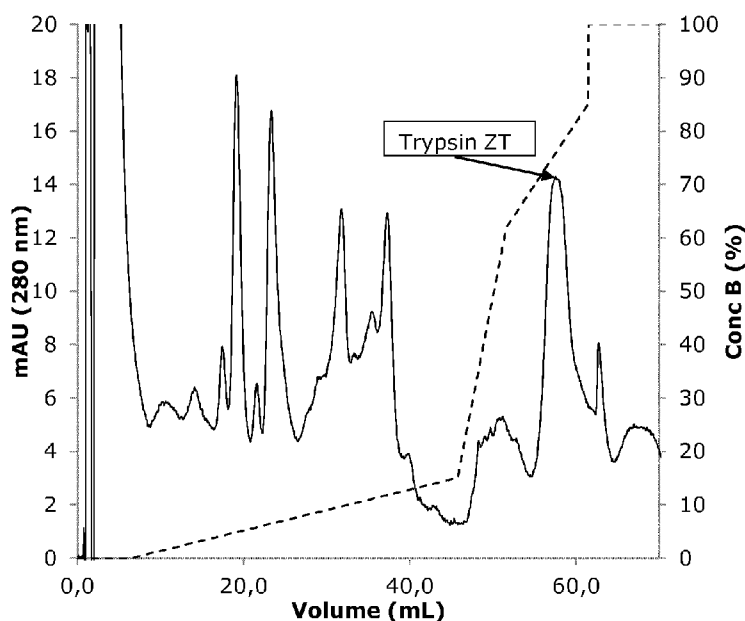


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[Continued on next page]

(54) Title: NOVEL TRYPSIN ISOFORMS AND THEIR USE

Figure 1



(57) Abstract: The present invention relates to the novel trypsin ZT isoforms. In particular, the invention relates to the use of trypsin ZT isoforms in medical devices, pharmaceuticals and cosmetics.



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NOVEL TRYPSIN ISOFORMS AND THEIR USE

Field of the invention

The present invention relates to the novel trypsin ZT isoforms. In particular, the
5 invention relates to the use of trypsin ZT isoforms in medical devices, pharmaceuticals and cosmetics.

Background of the invention

Proteinases are enzymes that are defined by their ability to cleave proteins.
10 Proteinases can cleave substrates at different amino residues within a protein sequence demonstrating a difference in substrate specificity. Trypsins are proteinases and are partly specified by their preference in cleaving C-terminal to arginine or lysine residues (Olsen, Ong et al., 2004, *Mol Cell Proteomics* **3**: 608-614). The Multiplex Substrate Profiling method reveals cleavage sites of proteinases and discloses the critical importance of residues
15 surrounding their cleavage site for their specificity (O'Donoghue, Eroy-Reveles et al., 2012, *Nat Methods* **9**: 1095-1100).

U.S. Patent Nos. 4,801,451 and 4,963,491 disclose a mixture of exo- and endopeptidases isolated from Antarctic krill (*Euphasia superba*) and the use of this mixture as a cleaning solution. 4,801,451 discloses the use of such enzymes to remove foreign
20 matter and dead tissue from wounds. WO 85/04809 discloses the use of krill enzymes as a digestion-promoting agent. EP-A1-0170115 discloses the use of krill enzymes to dissolve blood clots. However, all of these references disclose impure or poorly characterized materials. A purified peptidase or mixture of purified peptidases is desirable to provide a pharmaceutically useful product.

25 WO 96/24371 discloses the use of a krill-derived multifunctional proteolytic enzyme and a family of crustacean and fish derived proteolytic enzymes having structural similarity to the multifunctional enzyme derived from Antarctic krill. The document also relates to methods

of purifying the multifunctional enzyme and to pharmaceutical, cosmetic and other uses of the multifunctional enzyme. Structural similarity to the multifunctional enzyme derived from Antarctic krill is defined in that document as at least 70% homology with the krill derived multifunctional hydrolase.

5 WO 2000078332 discloses the use of cod derived trypsins and chymotrypsins in pharmaceutical compositions or medicaments for local and topical application to treat internal diseases and disorders and cosmetic use of such enzymes. The topical use for treating local, internal disorders, without the enzyme having to penetrate through open wounds or mucous tissue, was found to be effective. The serine proteinases disclosed in WO 2000078332 are
10 proteinases that have at least 90% amino acid sequence homology with trypsin I, trypsin II, trypsin III, trypsin IV derived from Atlantic cod and proteinases that are chymotrypsin having at least 90% amino acid sequence homology with any of chymotrypsin A and chymotrypsin B isolated from Atlantic cod.

A complementary DNA (cDNA) encoding trypsin Y was isolated from an Atlantic cod
15 cDNA library (Spilliaert and Gudmundsdottir, 1999, *Mar Biotechnol (NY)* **1**: 598-607). Cod trypsin Y has approximately 45% identity to the two Atlantic cod trypsin I and X (WO 2000078332). The native trypsin Y and recombinant forms of trypsin Y have previously briefly been described (Palsdottir and Gudmundsdottir, 2007, *Protein Expr Purif* **51**: 243-252, Palsdottir and Gudmundsdottir, 2008, *Food Chemistry* **111**: 408-414).

20 Summary of the invention

The present invention provides novel trypsin isoforms, called trypsin ZT isoforms. Such isoforms are derivable from fish, such as the Atlantic cod, or derivable in recombinant form using protein expression systems. The present invention further provides compositions
25 comprising at least one isolated cod trypsin ZT isoform according to the invention, together with suitable excipients and carriers.

The inventors have identified unexpected, beneficial and unique characteristics of the novel trypsin ZT isoforms over trypsins known previously. These characteristics provide an advantage in using trypsin ZT isoforms for various applications in medical devices, pharmaceuticals and cosmetics. The present invention provides said trypsin ZT isoforms *per se* and for use in treating or preventing diseases caused by pathogenic organisms. This is particularly relevant for use in treating or preventing a disease in the upper respiratory tract caused by pathogenic microorganisms. For that purpose, trypsin ZT isoforms can for example be used in medical devices or as active ingredient(s) in a pharmaceutical. Also, the present invention provides said trypsin ZT isoforms for use in the treatment of wound infections and wounds from burns, for use in removing dead or peeling skin from otherwise healthy skin for example in a medical device, as a pharmaceutical or as a cosmetic. The present invention further provides methods of preparing cod trypsin ZT isoforms of the present invention.

- 15 • Based on multiplex substrate profiling, trypsin ZT isoforms prefer cleavage at Arg residues compared to trypsin I. Trypsin I prefers cleavage at Lys residues compared to trypsin ZT isoforms.
- Multiplex substrate profiling shows that different amino acid residues in the substrate surrounding the cleavage site have a large effect on cleavage (about four amino acids residues on the N terminal side and about four amino acid residues on the C terminal side). Trypsin ZT isoforms prefer certain amino acid residues surrounding the cleavage site over trypsin I.
- Arginine and lysine rich amino acid sequences are frequently found in viral-, bacterial- and parasite sequences that are associated with infection.
- 25 • Numerical values based on standard scores (Z-score) ratio (multiplex substrate profiling data) show that trypsin ZT isoforms are much better adapted to cleave peptides containing several consecutive basic amino acid residues (Arg and Lys) compared to

trypsin I. Based on the data, trypsin ZT isoforms can act as antimicrobial agent against viruses and bacteria as they can cleave at clustered basic amino acid residues such as lysine and arginine.

5 Detailed description of the invention

The present invention provides novel trypsin isoforms, called trypsin ZT isoforms. Such isoforms are derivable from fish, such as the Atlantic cod, or derivable in recombinant form using protein expression systems. The present invention relates to using such trypsin ZT isoforms for treatment and prevention of disease and to the cosmetic field. In particular, the invention relates to novel Atlantic cod trypsin ZT isoforms, useful as pharmaceuticals, in
10 medical devices, and cosmetics.

Brief description of the drawings

FIG1 shows isolation of trypsin ZT isoforms with anion exchange chromatography.
15 Chromatogram from a MonoQ separation of benzamidine purified cod trypsin isoforms showing absorbance at 280 nm vs. elution volume. Cod trypsin isoforms were loaded on a MonoQ column and proteins eluted with a salt gradient (dotted line). Arrow shows the peak containing trypsin ZT isoforms. The column was equilibrated with a buffer (20 mM Tris, 10 mM CaCl₂, pH 8.0), and the enzymes were eluted with a linear 0-150 mM NaCl gradient in 40
20 column volumes, linear 150-620 mM NaCl in 6 column volumes, 620-850 mM NaCl in 10 column volumes at a flow rate of 1 mL/min.

FIG 2 shows SDS-PAGE of cod trypsin ZT isoforms. SDS-PAGE analysis of anion exchange chromatography peak labeled trypsin ZT from Figure 1. Trypsin 1, trypsin 5 and trypsin 7 are purified and isolated cod trypsin X isoforms. Cod trypsin X is closely related to
25 cod trypsin I, differ by about eight amino acid residues. The proteins were resolved by SDS-

PAGE and the gel silver stained. The bars and numbers on the left show the migration and molecular weight in kDa of standard proteins (Dalton Mark VII-L) separated on the gel.

FIG 3 shows Western blot analysis of cod trypsin X (trypsin 1, trypsin 5 and trypsin 7) and trypsin ZT isoforms. Cod trypsin X and trypsin ZT isoforms were subjected to SDS-

PAGE. Following transfer, samples were immunoblotted with trypsin I/X (top panel) and trypsin ZT isoform (bottom panel) reacting antibodies. This shows that trypsin ZT isoforms were separated from trypsin I/X using anion exchange. As can be seen in FIG 3, the polyclonal antibody that reacts against trypsin ZT isoforms does not cross-react with trypsin I/X and the peptide antibody raised against trypsin I/X does not cross-react with trypsin ZT isoforms.

FIG 4 shows a Venn diagram illustrating shared and unique cleavages generated by trypsin ZT and trypsin I at three different time points in a multiplex substrate profiling analysis. Number of shared (black) and unique cleavage sites by trypsin ZT (white on the left) and trypsin I (white on the right) after incubation for 5 min. (A), 15 min. (B) and 60 min (C). The figure shows that at all time points there are shared cleavage sites and unique cleavage sites for trypsin ZT and trypsin I in the 124 defined peptides (each containing 14 amino acid residues) within the library. The data demonstrates that the substrate specificity of trypsin ZT is different from that of trypsin I.

FIG 5 shows antiviral activity of trypsin ZT against enterovirus (Coxsackievirus B2).

The Y axis shows the percentage of wells infected compared to control and the X axis shows control (left bar) and treatment with trypsin ZT isoforms (right bar).

The inventors have identified novel trypsin isoforms termed trypsin ZT isoforms.

Trypsin ZT includes at least the following isoforms: Atlantic cod trypsin ZT-1 isoform, Atlantic cod trypsin ZT-2 isoform, Atlantic cod trypsin ZT-3 isoform and Atlantic cod trypsin ZT-4 isoform. All four Atlantic cod trypsin ZT isoforms have a similar molecular mass of about 25 kDa. The Atlantic cod trypsin ZT isoforms of the present invention are represented by the following amino acid sequences:

SEQ ID NO:1

IX₁GGX₂X₃CEPX₄SRPFMASLNYGYHFCGGVLINDQWVLSVAHCWYNPYMQVMLGEHDL
 RVFEGTEQLVKTNITFWHEX₅YDYQTLDYDMMMIKLYHPVEVTQSVAPISLPTGPPDGGMLC
 5 SVSGWGNMAMGEEVNLPTRLQCLDVPIVEX₆VX₇CX₈AX₉YPGMISPRMX₁₀CX₁₁GX₁₂MDGG
 RDX₁₃CNGDSGSPLVCEGVLTGLVSWGX₁₄GCAX₁₅PNX₁₆PGVYVKVYEX₁₇LSWIQTTLNANP

wherein

- X₁ is selected from I and V;
- 10 X₂ is selected from Q and H;
- X₃ is selected from D and E;
- X₄ is selected from R and N;
- X₅ is L;
- X₆ is selected from T and P;
- 15 X₇ is selected from D and A;
- X₈ is selected from E and Q;
- X₉ is selected from A and S;
- X₁₀ is selected from V and M;
- X₁₁ is selected from A and V;
- 20 X₁₂ is selected from Y and F;
- X₁₃ is selected from A and V;
- X₁₄ is selected from Q and R;
- X₁₅ is selected from L and E;
- X₁₆ is selected from Y and S; and
- 25 X₁₇ is selected from Y and F.

SEQ ID NO:2 The Atlantic cod trypsin ZT-1 isoform

IVGGHECEPNSRPFMASLNYGYHFCGGVLINDQWVLSVAHCWYNPYMQVMLGEHDLRV
FEGTEQLVKTNNTIFWHELYDYQTLDDYDMMMIKLYHPVEVTQSVAPISLPTGPPDGGMLCSV
SGWGNMAMGEEVNLPTRLQCLDVPIVEPVACQASYPGMISPRMMCVGFMDGGRDVCNG
DS GSPLVCEGVLTGLVSWGRGC AEPNSPGVYVKVYEFLSWIQTTLNANP

5

SEQ ID NO:3 The Atlantic cod trypsin ZT-2 isoform

IVGGHECEPNSRPFMASLNYGYHFCGGVLINDQWVLSVAHCWYNPYMQVMLGEHDLRV
FEGTEQLVKTNNTIFWHELYDYQTLDDYDMMMIKLYHPVEVTQSVAPISLPTGPPDGGMLCSV
SGWGNMAMGEEVNLPTRLQCLDVPIVETVDCEAAYPGMISPRMVCAGYMDGGRDACNGD

10 S GSPLVCEGVLTGLVSWGQGC ALPNYPGVYVKVYEYLSWIQTTLNANP

SEQ ID NO:4 The Atlantic cod trypsin ZT-3 isoform

IIGGQDCEPRSRPFMASLNYGYHFCGGVLINDQWVLSVAHCWYNPYMQVMLGEHDLRVF
EGTEQLVKTNNTIFWHELYDYQTLDDYDMMMIKLYHPVEVTQSVAPISLPTGPPDGGMLCSV
15 GWGNMAMGEEVNLPTRLQCLDVPIVEPVACQASYPGMISPRMMCVGFMDGGRDVCNGD
S GSPLVCEGVLTGLVSWGRGCAEPNSPGVYV KVYEFLSWIQTTLNANP

SEQ ID NO:5 The Atlantic cod trypsin ZT-4 isoform

IIGGQDCEPRSRPFMASLNYGYHFCGGVLINDQWVLSVAHCWYNPYMQVMLGEHDLRVF
20 EGTEQLVKTNNTIFWHELYDYQTLDDYDMMMIKLYHPVEVTQSVAPISLPTGPPDGGMLCSV
GWGNMAMGEEVNLPTRLQCLDVPIVETVDCEAAYPGMISPRMVCAGYMDGGRDACNGDS
GSPLVCEGVLTGLVSWGQGC ALPNYPGVYVKVYEYLSWIQTTLNANP

Trypsin ZT isoforms of the present invention have less than 50% amino acid identity
25 with the tryptins disclosed in WO 2000078332 and less than 45% homology with the
chymotrypsins disclosed in WO 2000078332.

The above trypsin ZT isoforms represents the active variants of these trypsins, i.e. variants that have been activated when the N terminus of the trypsins have been cleaved off. These trypsins are proteins expressed in the pyloric caeca (pancreatic tissue in fish) with a number of amino acids on the N terminal end that are important for secretion out of the cells and for keeping the enzyme inactive. The full length trypsin ZT isoforms are also disclosed herein as

SEQ ID NO:19 Uncleaved Atlantic cod trypsin ZT-1 isoform

10 MIGLALLMLLGAAAAVPRDVGKIVGGHECEPNSRPFMASLNYGYHFCGGVLINDQWVLSV
AHCWYNPYMVMQVMLGEHDLRVFEGTEQLVKTNIFWHELYDYQTLDYDMMMIKLYHPVEV
TQSVAPISLPTGPPDGGMLCSVSGWGNMAMGEEVNLPTLRLQCLDVPIVEPVACQASYPGM
ISPRMMCVGFMDGGRDVCNGDSGSPLVCEGVLTGLVSWGRGCAEPNSPGVYVKVYEFLS
WIQTTL DANP

15 SEQ ID NO:20 Uncleaved Atlantic cod trypsin ZT-2 isoform

MIGLALLMLLGAAAAVPRDVGKIVGGHECEPNSRPFMASLNYGYHFCGGVLINDQWVLSV
AHCWYNPYMVMQVMLGEHDLRVFEGTEQLVKTNIFWHELYDYQTLDYDMMMIKLYHPVEV
TQSVAPISLPTGPPDGGMLCSVSGWGNMAMGEEVNLPTLRLQCLDVPIVETVDCEAAYPGMI
SPRMVCAGYMDGGRDACNGDSGSPLVCEGVLTGLVSWGQGCALPNYPGVYVKVYEYLS
20 WIQTTL DANP

SEQ ID NO:21 Uncleaved Atlantic cod trypsin ZT-3 isoform

MIGLALLMLLGAAAAVPREDGRIIGGQDCEPRSRPFMASLNYGYHFCGGVLINDQWVLSVA
HCWYNPYMVMQVMLGEHDLRVFEGTEQLVKTNIFWHELYDYQTLDYDMMMIKLYHPVEVT
25 QSVAPISLPTGPPDGGMLCSVSGWGNMAMGEEVNLPTLRLQCLDVPIVEPVACQASYPGMI
SPRMMCVGFMDGGRDVCNGDSGSPLVCEGVLTGLVSWGRGCAEPNSPGVYVKVYEFLS
WIQTTL DANP

SEQ ID NO:22 Uncleaved Atlantic cod trypsin ZT-4 isoform

30 MIGLALLMLLGAAAAVPREDGRIIGGQDCEPRSRPFMASLNYGYHFCGGVLINDQWVLSVA
HCWYNPYMVMQVMLGEHDLRVFEGTEQLVKTNIFWHELYDYQTLDYDMMMIKLYHPVEVT

QSVAPISLPTGPPDGGMLCSVSGWGNMAMGEEVNLPTRLQCLDVPIVETVDCEAAYPGMI
SPRMVCAGYMDGGRDACNGDSGSPLVCEGLTGLVSWGQGCALPNYPGVYVKVYEYLS
WIQTTL DANP

5 Multiplex substrate profiling is used to analyze the substrate specificity of enzymes (proteinases) (O'Donoghue, Eroy-Reveles et al., 2012, *Nat Methods* **9**: 1095-1100). Proteinases can cleave substrates at different amino residues within a peptide sequence demonstrating a difference in substrate specificity. Trypsins are specified by their preference in cleaving C-terminal to arginine or lysine residues. Unexpectedly, multiplex substrate
10 profiling on trypsin ZT isoforms revealed that amino acids surrounding arginine or lysine in a substrate have a different effect on its cleavage compared to trypsin I. In short, trypsin ZT isoforms or trypsin I were incubated (5, 15 and 60 min.) with a library of peptides (124 defined peptides, 14-mer sequences, total of 1612 peptide bonds) with an extensive physiochemical diversity (O'Donoghue, Eroy-Reveles et al., 2012, *Nat Methods* **9**: 1095-
15 1100). After incubation, cleavage sites were identified by mass spectrometry analysis. The results are presented in the following examples and show that there are numerous cleavage sites unique to trypsin ZT compared trypsin I. Also, cleavage sites that showed preferential cleavage by trypsin ZT isoforms compared to trypsin I were identified. Further, the novel trypsin ZT isoforms have a different substrate specificity compared to a trypsin I, disclosed in
20 WO 2000078332. These findings were unexpected. Based on the results from the multiplex substrate profiling analysis, a table containing numerical values based on standard scores (Z-score) ratio at the P4-P4' positions in the substrate was generated where the substrate specificity of trypsin ZT was compared to trypsin I. Positive values (Z-score) reflect a preference for the amino acid residue in a certain P position in favor of trypsin ZT isoforms
25 compared to trypsin I.

Trypsin ZT isoforms have specific antimicrobial properties. Based on the values generated and presented in the examples, it can be seen that trypsin ZT isoforms are much

better adapted in cleaving peptides containing several consecutive positively charged residues (arginine or lysine) compared to trypsin I. Those types of sequences (lysine and arginine rich amino acid sequence) are frequently found in viral proteins (Suzuki, Orba et al., 2010, *PLoS Pathog* **6**: e1000801, Jiang, Cun et al., 2012, *J Virol* **86**: 7256-7267, Gallaher and Garry, 2015, *Viruses* **7**: 285-305).

Viroporins is a group of proteins that interact with plasma membranes modifying permeability and can promote the release of viral particles. Clustered lysine and/or arginine residues are important for the activity of viroporins. Based on the multiplex substrate profiling analysis, trypsin ZT isoforms can act as antiviral agents against RNA viruses as it can cleave these clustered basic amino acid residues . Viroporins have been described as present in a number of virus families, including the *Flaviviridae*, *Picornaviridae*, *Retroviridae*, *Coronaviridae*, *Reoviridae* and *Paramyxoviridae* (Royle, Dobson et al., 2015, *Viruses* **7**: 5375-5387). Studies have shown that basic residues, lysine or arginine, in viroporins are a critical requirement for their function. The majority of viroporins have been identified in RNA viruses. Viruses that contain lysine and arginine rich viroporins that would be a target for trypsin ZT isoforms include human immunodeficiency virus (HIV), rotavirus, Ebola, Reston virus (RESTV), Lloviu virus (LLOV), Sudan virus (SUDV), Bundibugyo (BDBV), Tai Forest virus (TAFV) (Suzuki, Orba et al., 2010, *PLoS Pathog* **6**: e1000801, Gallaher and Garry, 2015, *Viruses* **7**: 285-305).

The viroporins are found in a wide variety of DNA and RNA viruses such as *Paramecium bursaria chlorella virus 1*, simian virus 40 (SV40), human polyoma JC (JCV), severe acute respiratory syndrome coronavirus (SARS-CoV) (Kang, Moroni et al., 2004, *Proc Natl Acad Sci U S A* **101**: 5318-5324, Liao, Tam et al., 2006, *Adv Exp Med Biol* **581**: 199-202, Daniels, Sadowicz et al., 2007, *PLoS Pathog* **3**: e98).

Highly basic viral proteins, termed agnoproteins, have recently have been identified from DNA viruses such as from the *Polyomaviridae* and *Papillomaviridae* family (Royle, Dobson et al., 2015, *Viruses* **7**: 5375-5387). These proteins show a number of viroporin

characteristics. Conserved sequences have been identified in agnoproteins that contain a number of clustered basic amino acid residues (Royle, Dobson et al., 2015, *Viruses* 7: 5375-5387). Based on the multiplex substrate profiling analysis, trypsin ZT isoforms can act as antiviral agents against DNA viruses as it can cleave these clustered basic amino acid residues (Royle, Dobson et al., 2015, *Viruses* 7: 5375-5387).

Many viruses and bacteria have developed mechanisms to gain cell entry using cell penetrating peptides or proteins (Milletti, 2012, *Drug Discov Today* 17: 850-860). The viruses include pestiviruses of the *Flaviviridae* family, herpes virus (HSV-1), human immunodeficiency virus (HIV-1, hepatitis B and human respiratory syncytial virus (RSV) (Elliott and O'Hare, 1997, *Cell* 88: 223-233, Oess and Hildt, 2000, *Gene Ther* 7: 750-758, Langedijk, 2002, *J Biol Chem* 277: 5308-5314, Langedijk, Olijhoek et al., 2005, *International Congress Series* 1277: 95-107, Lu, Tager et al., 2006, *Anal Biochem* 353: 7-14, Godet, Guernon et al., 2010, *PLoS One* 5: e13760). Furthermore, these types of proteins have been found in bacteria such as *Mycobacterium tuberculosis* (Lu, Tager et al., 2006, *Anal Biochem* 353: 7-14). The cell penetrating peptides (CPP) have in common that they are rich in basic amino acid residues especially arginines (Milletti, 2012, *Drug Discov Today* 17: 850-860). Porins are outer membrane proteins that play a crucial role in bacterial pathogenicity and for protection and are for that reason useful targets for therapeutics (Galdiero, Falanga et al., 2012, *Curr Protein Pept Sci* 13: 843-854). These proteins often contain consecutive arginine amino acid residues. Based on the multiplex substrate profiling analysis trypsin ZT isoforms can act as antimicrobial agents against viruses and bacteria as it can cleave clustered basic amino acid residues.

Parasites depend on virulence determinants such as proteins containing arginine rich domains for example *Toxoplasma gondii* (Fentress, Steinfeldt et al., 2012, *Cell Microbiol* 14: 1921-1933). Multiplex substrate profiling analysis showed that trypsin ZT isoforms would be beneficial against parasites by acting against such proteins.

The Basic Local Alignment Search Tool (BLAST) was used to search for some of the unique and preferential amino acid sequences cleaved by trypsin ZT isoforms. The search results demonstrated that these types of sequences are found in many pathogens including viruses (e.g. from the *Paramyxoviridae* and *Coronaviridae* family of viruses), bacteria (e.g. *Haemophilus* genus) and parasites (e.g. *Plasmodium* genus). The *Paramyxoviridae* family includes viruses such as the respiratory syncytial virus (RSV) and human parainfluenza viruses and viruses that cause mumps and measles. The *Coronaviridae* family includes viruses that cause Severe Acute Respiratory Syndrome (SARS). The *Haemophilus* genus of bacteria includes *Haemophilus parainfluenzae* that is associated with endocarditis, meningitis, and bacteremia. The *Plasmodium* genus includes *Plasmodium falciparum* that causes malaria.

By the phrase “medical device” is meant, but not limited to, any instrument, apparatus, appliance, software, material or other article for the purpose of diagnosis, prevention, monitoring, treatment, or alleviation of disease, such as diagnosis, monitoring, treatment, alleviation of or compensation; for an injury or handicap, such as investigation, replacement or modification of the anatomy or of a physiological process; control of conception; including devices that do not achieve their principal intended action in or on the human body by pharmacological, immunological or metabolic means – but may be assisted in their function by such means.

The present invention relates to the novel trypsin ZT isoforms. The invention describes unpredicted and unique protein cleaving properties of trypsin ZT isoforms that make it valuable in medical applications. In particular, the invention relates to the novel Atlantic cod trypsin ZT isoforms, useful as pharmaceuticals, in medical devices, and cosmetics.

In one aspect of the invention, there is provided an isolated cod trypsin ZT isoform comprising an amino acid sequence according to SEQ ID NO:1, or an amino acid sequence with 80% sequence homology or more, thereof. Typically, such sequence homology is 85, 90, 95 or 99% homologous to the amino acid sequence of SEQ ID NO:1.

In one embodiment of this aspect there is provided an isolated cod trypsin ZT isoform, selected from trypsin ZT-1, comprising an amino acid sequence according to SEQ ID NO:2; trypsin ZT-2 comprising an amino acid sequence according to SEQ ID NO:3; trypsin ZT-3 comprising an amino acid sequence according to SEQ ID NO:4; and trypsin ZT-4, comprising
5 an amino acid sequence according to SEQ ID NO:5, or an amino acid sequence with 80% sequence homology or more, thereof. Included are also amino acid sequences with 80% sequence homology or more, to amino acid sequences set out in SEQ ID NOS:2-5. Typically, such sequence homology is 85, 90, 95 or 99% homologous to the amino acid sequence of SEQ ID NOS:2-5.

10 In one aspect of the invention, there is provided a composition comprising at least one cod trypsin ZT isoform according to the invention, together with suitable excipients and carriers. Said cod trypsin ZT isoform may be selected from cod trypsin ZT-1, comprising an amino acid sequence according to SEQ ID NO:2; cod trypsin ZT-2 comprising an amino acid sequence according to SEQ ID NO:3; cod trypsin ZT-3 comprising an amino acid sequence
15 according to SEQ ID NO:4; and cod trypsin ZT-4 comprising an amino acid sequence according to SEQ ID NO:5. Included are also amino acid sequences with 80% sequence homology or more, to amino acid sequences set out in SEQ ID NOS:2-5. Typically, such sequence homology is 85, 90, 95 or 99% homologous to the amino acid sequence of SEQ ID NOS:2-5.

20 Typically, said composition comprises said cod trypsin ZT isoforms present in admixture with other cod trypsins. Preferably, said cod trypsin ZT isoforms comprises at least 5% (w/w) of the total content of cod trypsins in said composition, such as at least 10, 15, 20, 25, 30, 35, 40, 45 or 50% or more (w/w) of the total content of cod trypsins in said composition.

25 Said composition is typically administered topically, as a lotion, a hydrogel, a mouth spray or a nasal spray. Said composition may further comprise a polyvalent alcohol, such as glycerol.

In one embodiment of this aspect, there is provided said composition for use in therapy.

In one embodiment of this aspect, there is provided said composition for use in treating or preventing a disease caused by a pathogenic organism selected from the group consisting
5 of a virus, a bacterium, a fungus, a parasite and a protozoan.

In one embodiment of this aspect, there is provided said composition for use in treating or preventing a disease in the upper respiratory tract caused by pathogenic organisms such as viruses, bacteria and fungus.

In one embodiment of this aspect, there is provided said composition for use in treating
10 or preventing pain, acute inflammation, chronic inflammation, arthritis, inflamed joints, bursitis, osteoarthritis, rheumatoid arthritis, juvenile rheumatoid arthritis, septic arthritis, fibromyalgia, systemic lupus erythematosus, phlebitis, tendinitis, rash, psoriasis, acne, eczema, facial seborrheic eczema, eczema of the hands, face or neck, foreskin infections, athlete's foot, fistulae infections, infected topical ulcers, navel infections in newborns,
15 wrinkles, scars, keloids, boils, warts and allergic itch, hemorrhoids, a fungal infection and an immunological disorder including an autoimmune disease.

In one embodiment of this aspect, there is provided said composition for use in the treatment of wound infections and wounds from burns.

In one embodiment of this aspect, there is provided said composition for use in
20 removing dead or peeling skin from otherwise healthy skin.

In one embodiment of this aspect, there is provided said composition for cosmetic use. Said composition may further comprise an additional cosmetically active compound.

In one aspect of the invention, there is provided a method of treating disease, comprising administering a therapeutically effective amount of a composition according to the
25 present invention, to a patient in need thereof.

In one embodiment of this aspect, said disease is caused by a pathogenic organism selected from the group consisting of a virus, a bacterium, a fungus, a parasite and a protozoan

5 In one embodiment of this aspect, said disease is a disease in the upper respiratory tract caused by pathogenic organisms such as viruses and bacteria.

In one embodiment of this aspect, said disease is selected from pain, acute inflammation, chronic inflammation, arthritis, inflamed joints, bursitis, osteoarthritis, rheumatoid arthritis, juvenile rheumatoid arthritis, septic arthritis, fibromyalgia, systemic lupus erythematosus, phlebitis, tendinitis, rash, psoriasis, acne, eczema, facial seborrheic eczema, 10 eczema of the hands, face or neck, foreskin infections, athlete's foot, fistulae infections, infected topical ulcers, navel infections in newborns, wrinkles, scars, keloids, boils, warts and allergic itch, hemorrhoids, a fungal infection and an immunological disorder including an autoimmune disease.

15 In one embodiment of this aspect, said disease is wound infections or wounds from burns.

In one aspect of the invention, there is provided an isolated cod trypsin ZT isoform, selected from full length trypsin ZT-1, comprising an amino acid sequence according to SEQ ID NO:19; full length trypsin ZT-2 comprising an amino acid sequence according to SEQ ID NO:20; full length trypsin ZT-3 comprising an amino acid sequence according to SEQ ID NO:21; and full length trypsin ZT-4, comprising an amino acid sequence according to SEQ ID NO:22. 20

In one aspect of the invention, there is provided a method of preparing cod trypsin ZT isoforms comprising an amino acid sequence according to any one of SEQ ID NO:2 to SEQ ID NO:5, comprising

- 25 (i) extracting trypsin ZT from cod viscera,
- (ii) applying the extract to at least one or more chromatography steps, including an affinity chromatography step using a p-aminobenzamidine affinity ligand, and

(iii) desorbing and eluting cod trypsin ZT isoforms bound to the p-aminobenzamidine affinity ligand.

In one embodiment of this aspect, said method further comprises at least one step using an anionic exchange resin after said affinity chromatography step (ii).

5 In one embodiment of this aspect, said cod trypsin ZT isoforms are selected from trypsin ZT-1, comprising an amino acid sequence according to SEQ ID NO:2; trypsin ZT-2 comprising an amino acid sequence according to SEQ ID NO:3; trypsin ZT-3 comprising an amino acid sequence according to SEQ ID NO:4; and trypsin ZT-4 comprising an amino acid sequence according to SEQ ID NO:5, or a mixture thereof.

10 Cod viscera is typically used to purify and isolate trypsin ZT isoforms. The inventors have identified novel methods described below, useful for a commercial scale production of trypsin ZT isoforms. Extraction is conducted at a pH of 6-8 at a temperature below 10°C. Cod viscera is mixed with water at a ratio (w/w) of 1:6 to 1:20. The extract is separated from the leftover offal and further clarified by sedimentation and filtration. This is followed by
15 microfiltration and ultrafiltration resulting a solution containing trypsin ZT isoforms that is used for chromatography separation that may include several steps.

An affinity chromatography is conducted, preferably by amino benzamidine affinity chromatography, to purify trypsin ZT isoforms. Trypsin ZT isoforms can then be further purified with anion exchange chromatography.

20 One method of application of trypsin ZT isoforms or mixtures of purified cod tryptins containing trypsin ZT isoforms, is in a preparation of hydrogel or lotion and water containing 0 to 90% (vol/vol) of a polyvalent alcohol (polyol) such as glycerol. A suitable concentration of trypsin ZT isoforms constitutes at least 1 to 100 protein concentration ratio to other trypsin forms, preferably 5%, or higher (w/w) of the total content of cod tryptins. The trypsin activity
25 is 0.1 to 10,000 enzyme units of activity for CBZ-Gly-Pro-Arg-pNA (carbobenzoxymethyl Gly-Pro-Arg-para nitroanilide) per 100 milliliters of the final hydrogel/lotion preparation.

The invention further provides (a) methods relating to certain conditions using effective amounts of the purified trypsin ZT isoforms described above, (b) compositions or substances for use in such methods, (c) pharmaceutical or medical device compositions containing effective amounts of trypsin ZT isoforms for use in such methods, and (d) uses of the enzyme or enzyme composition for manufacturing a medicament (pharmaceutical and medical devices) for use in such methods.

The methods are *inter alia* for:

treating or preventing pathogenic disease caused by viruses, bacteria and fungi that for example occur in the upper respiratory tract, lower respiratory tract and in lungs;

treating or prophylactically preventing dermatological conditions such as e.g. acne, rash, psoriasis or eczema, including facial seborrheic eczema or eczema of the hands, face, scalp or neck, hemorrhoids and the like, where preferably the amount of the trypsin ZT isoforms and other trypsins administered is a dermatological condition treating or preventing effective amount;

treating lung diseases where trypsin ZT isoforms with or without other trypsins are used for example in pressurized metered dose high efficiency inhaler in an effective dose for treatment;

treating or prophylactically preventing wound infection and debriding wounds (by applying to the wound a microbial infection-preventing effective amount of trypsin ZT

isoforms with or without other trypsins or by enhancing the healing of wounds by administering a microbe inhibiting effective amount of trypsin ZT isoforms with or without other trypsins, when treated the wound can be substantially freed of necrotic tissue;

treating burns, where preferably the amount of trypsin ZT isoforms with or without other trypsins administered in a sufficient amount to promote healing;

treating sunburns, where preferably the amount of trypsin ZT isoforms with or without other trypsins administered in a sufficient amount to promote healing;

treating radiation burns, where preferably the amount of trypsin ZT isoforms with or without other trypsins administered in a sufficient amount to promote healing;

removing dead or peeling skin from otherwise healthy skin to improve the skin's appearance, where preferably the amount of trypsin ZT isoforms with or without other

5 trypsins is administered in a dead skin removing effective amount;

treating cracked heels, where preferably the amount of trypsin ZT isoforms with or without other trypsins is administered in a sufficient amount for effective treatment;

for management of irritated skin, such as dry patches, itching, redness and blemishes, where preferably the amount of trypsin ZT isoforms with or without other trypsins is

10 administered in a sufficient amount for effective treatment;

treating or prophylactically preventing cystic fibrosis, cancer, e.g. by administering a tumor treating effective amount or a tumor metastasis preventing or inhibiting amount of trypsin ZT isoforms, atherosclerosis, asthma, septic shock, toxic shock syndrome, tissue adhesions such as tendon-sheath, abdominal post-surgical or joint adhesions, reperfusion

15 injury, malaria, immune disorder such as an autoimmune disease, apoptosis, colitis and enteritis, such as Crohn's disease, where preferably the amount of trypsin ZT isoforms and related peptidases administered are effective;

treating or prophylactically preventing a microbial infection, e.g. a viral infection such as a rhinovirus (RV), respiratory syncytial virus (RSV), influenza, herpes virus infection (e.g.

20 HSV-1 , HSV-2, herpes zoster or genital herpes infection), HIV, hepatitis, coronavirus, cytomegalovirus or papilloma virus infection; an infection causing a gastrointestinal disease such as ulcer or diarrhoea; a fungal infection such as systemic, skin, oral, vaginal or esophageal fungal infection, including e.g. yeast infection, including a fungal nail infection

and Candida infections; microbial infections of the eye, preferably treated with ocular

25 administrations; bacterial infections including infection by *Helicobacter pylori*, *Staphylococcus* spp., Methicillin-resistant *Staphylococcus aureus* (MRSA), *Streptococcus pneumonia*, *Haemophilus influenza*, *Streptococcus mitis*, *Streptococcus* spp., *Klebsiella*

spp., *Pseudomonas spp.*, *Neisseria gonorrhoeae*, *Haemophilus spp.*, *Chlamydia spp.*, syphilis and *E. coli* infections and bacterial infections causing chancroid; opportunistic microbial infections in immunocompromised patients where preferably the administered amount of the cod trypsins is a microbial infection-treating or -preventing effective amount or has inhibitory
5 activity against cell-cell or cell-virus adhesion;

treating or prophylactically preventing an indication selected from the group consisting of pain, inflammation, acute or chronic inflammation, arthritis, inflamed joints, bursitis, osteoarthritis, rheumatoid arthritis, juvenile rheumatoid arthritis, septic arthritis, fibromyalgia, systemic lupus erythematosus and phlebitis where preferably the amount is treating or
10 preventing effective amount;

removing dental plaque, where preferably the amount of trypsin ZT isoforms with or without other trypsins is administered in a dental plaque removing effective amount; and lysing blood clots, where preferably the amount of trypsin ZT isoforms is a clot lysing effective amount.

15 The method comprises administering a composition comprising the trypsin ZT isoforms as described above.

The invention provides topical cosmetic and medical compositions comprising trypsin ZT isoforms described above; and gel, cream or suppository composition.

The invention further provides a method of inhibiting or prophylactically preventing the
20 transmission of a pathogenic microbe by administering trypsin ZT isoforms with or without other trypsins. Preferably, trypsin ZT with or without other trypsins is applied to the portion of the body that comprises the primary transmission entryway for the microbe in question. In one embodiment, a spray, ointment or wash is applied to a body orifice involved in sexual activity, for instance, to prevent HIV or hepatitis transmission. In another embodiment, trypsin
25 ZT isoforms or related peptidases is applied to the upper airways, for example, via an aerosol, to inhibit or prevent the transmission of a common cold virus, such as a rhinovirus or a coronavirus.

The method of extra-corporeally treating a tissue, body fluid or composition of cells to remove cell adhesion components reduces the immune rejection of tissue, body fluid or composition of cells that is transplanted from one individual to another. In another aspect, such treatments remove or inactivate the cell adhesion components found in the treated
5 tissue, body fluid or composition of cells involved in a microbial infection.

In treating or prophylactically preventing septic shock or toxic shock syndrome by administering trypsin ZT isoforms or related peptidases, appropriate routes of administration include systemic administration. For vaginal infections associated with shock, vaginal flushes, creams, gels or suppositories may be used as a method of administration.

10 In treating or prophylactically preventing pain, inflammation, acute or chronic inflammation, arthritis, inflamed joints, bursitis, osteoarthritis, rheumatoid arthritis, juvenile rheumatoid arthritis, septic arthritis, fibromyalgia, systemic lupus erythematosus, phlebitis by administering trypsin ZT isoforms or related peptidases, appropriate routes of administration would include without limitation creams, gels or suppositories, in particular but
15 not limited to hydrogels containing glycerol or other polyols.

In treating or prophylactically preventing rash, psoriasis, acne, eczema, including facial seborrheic eczema or eczema of the hands, face, scalp or neck, foreskin infections, athlete's foot, fistulae infections, infected topical ulcers, navel infections in newborns, wrinkles, scars and keloids, boils, warts and allergic itch, hemorrhoids and the like, wounds, wound
20 infections, wounds from burns, removing dead or peeling skin from otherwise healthy skin to improve the skin's appearance, a fungal infection such as systemic, skin, oral, vaginal or esophageal fungal, including for example, yeast infection, including a fungal nail infection and Candida infections and immune disorders including autoimmune diseases by administering trypsin ZT isoforms with or without other trypsins, appropriate routes of
25 administration would include creams, gels or suppositories, in particular but not limited to hydrogels containing glycerol or other polyols.

The above objectives are achieved by using a composition containing an effective amount of trypsin ZT isoforms with or without other trypsins which is capable of relieving the pain, inflammation, arthritis, swelling, edema, psoriasis eczema, dermatitis, rash and/or other symptoms of the diseases mentioned in the introductory part. Trypsin ZT isoforms can be
5 obtained at a high yield and in a relatively simple manner from cod viscera. It has been found that trypsin ZT isoforms can be purified on a commercial scale in a relatively straightforward manner with reasonable yields. Trypsin ZT isoforms without or along with other Atlantic serine proteases can be used for the purpose of the invention.

Trypsin ZT isoforms presented in the present disclosure can be desorbed from the
10 affinity matrix by applying conditions that will destabilize the interaction between the enzyme and the affinity ligand. Such conditions include high salt followed by low pH, preferably in 30% or more glycerol. The column eluent is allowed to flow into a neutralizing buffer to stabilize the cod trypsin after the acid elution step. Trypsin ZT isoforms can then be further purified using anion exchange and eluted with increasing salt concentration. By these
15 methods trypsin ZT isoforms with a purity in excess of about 90% can be isolated. The cod trypsin fraction thus obtained contains 2-3 major isoforms as manifested by SDS electrophoresis, which yields two bands and MALDI-TOF mass spectrometry analysis. The molecular mass of trypsin ZT isoforms is about 25 kDa, whereas the calculated isoelectric point is 4.35 – 4.49 depending on the isoform.

Trypsin ZT isoforms with or without other trypsins in specific formulations can be used
20 as a pharmaceutical, in medical devices (such as chemical formulations) or in cosmetics depending on intended use. Trypsin ZT isoforms (with or without other trypsins) of the invention are administered topically, orally, rectally, vaginally, by instillation (for instance into the urinary tract or into fistulas), by the pulmonary route e.g. by use of an aerosol (for
25 example by using a pressurized metered-dose inhaler (pMDI)), by application of drops to the eye, or systemically, such as parenterally, including, for example, intramuscularly, subcutaneously, intraperitoneally, intraarterially or intravenously. Trypsin ZT

isoforms are administered in solution or combined with a pharmaceutically acceptable carrier or excipient according to standard pharmaceutical practice. For the oral mode of administration, trypsin ZT isoforms are used in the form of tablets, capsules, lozenges, chewing gum, troches, powders, syrups, elixirs, aqueous solutions and suspensions, and the like. For parenteral administration, sterile solutions of trypsin ZT isoforms are usually prepared, and the pH values of the solutions are suitably adjusted and buffered. For intravenous use, the total concentration of solutes should be controlled to render the preparation isotonic. For ocular administration, ointments or droppable liquids may be delivered by ocular delivery systems known to the art such as applicators or eye droppers.

For pulmonary administration, diluents and/or carriers will be selected to be appropriate to allow the formation of an aerosol. For topical administrations, trypsin ZT isoforms are typically administered in a hydrogel, containing 0 to 85% glycerol, such as about 20% to 30% glycerol and possibly up to 85 % glycerol.

For topical treatments, a suitable dose of trypsin ZT isoforms with or without other trypsins per application ranges from about $0.01 \mu\text{g}/\text{cm}^2$ to about $1 \text{ mg}/\text{cm}^2$, preferably from about $0.1 \mu\text{g}/\text{cm}^2$ to about $0.01 \text{ mg}/\text{cm}^2$ (using e.g. about $0.01 \text{ mg}/\text{ml}$ enzyme gel). For systematic treatments, dosages will generally be selected to maintain a serum level of trypsin ZT isoforms with or without other trypsins between about $0.1 \text{ mg}/100 \text{ ml}$ and about $100 \text{ mg}/100\text{ml}$, preferably between about $0.5 \text{ mg}/100 \text{ ml}$ and about $2.0 \text{ mg}/100 \text{ ml}$. In an alternative measure of preferred systematic administration amounts, preferably from about $0.1 \text{ mg}/\text{kg}$ to about $10 \text{ mg}/\text{kg}$, more preferably about $1 \text{ mg}/\text{kg}$, will be used. For vaginal and urinary tract treatments, suitable flushing/instillation solutions of trypsin ZT isoforms will generally have concentrations from about $1 \mu\text{g}/\text{ml}$ to about $15 \text{ mg}/\text{ml}$, preferably from about $100 \mu\text{g}/\text{ml}$ to about $3 \text{ mg}/\text{ml}$. For all treatments, the enzyme composition will generally be applied from about 1 to about 10 times per day, preferably from about 2 to about 5 times per day. These values, of course, will vary with a number of factors including the type and severity of the disease, and the age, weight and medical condition of the patient, as will be

recognized by those of ordinary skill in the medical arts. It is believed that substantially higher doses can be used without substantial adverse effect.

For wound healing, trypsin ZT isoforms with or without other trypsins are preferably applied more often than simply at the time at which the wound is first dressed. Preferably, 5 trypsin ZT isoforms are applied at least about every time the wound dressing is changed. Trypsin ZT isoforms can also be applied at least about every other day, more preferably, every day, or few times per day. For dermatological situations such as eczema, psoriasis and the like, a trypsin ZT isoform (with or without other trypsins) hydrogel is preferably applied every day, more preferably twice per day. For acute or chronic inflammation, arthritis, 10 inflamed joints, bursitis, osteoarthritis, and the like, trypsin ZT isoforms are preferably applied every day more preferably twice every day.

Numerous methods for determining percent homology of proteins are known in the art. Percent Identity was calculated using ClustalW2 (version Clustal2.1) when making sequence comparisons (EMBL-EBI website <http://www.ebi.ac.uk/Tools/msa/clustalw2/> date 4th of May 15 2015).

The invention will now be described by a number of non-limiting examples.

EXAMPLE 1.

Preparation of a mixture of proteases from cod

20 About 100 kg of frozen Atlantic cod viscera were thawed and added to a four-fold volume of cold potable water in an extraction tank and the pH adjusted to around pH 6 with sodium hydroxide solution. The mixture was stirred for about 10 hours at 0 to 5°C. After a brief period of crude sedimentation (about 30 minutes) the aqueous extract was run off the remaining insoluble viscera with a pump and collected in a sedimentation tank. The aqueous 25 extract was allowed to stand in the cooled sedimentation tank to sediment for about 40 to 80 hours. The supernatant was decanted from the supernatant tank to a holding tank using a pump. The supernatant was concentrated 10 to 20-fold by ultrafiltration and diafiltered to an

acceptable level of ionic strength with conductivity below about 2.5 mS/cm. About 10-15 liters of ultrafiltered and diafiltered protein concentrate was obtained.

EXAMPLE 2.

5 Purification of cod trypsin ZT isoforms from concentrated cod viscera extract

About 12 liters of ultrafiltered and diafiltered concentrate as obtained in Example 1 was applied to a continuous, connected series of about 1 liter packed chromatography columns, the first containing a carboxymethyl (CM) fast flow cation exchange resin (GE healthcare) and the second one a p-aminobenzamidinium affinity ligand coupled to a sepharose resin (GE
10 healthcare). The CM and p-aminobenzamidinium columns were pre-equilibrated with about 10 column volumes of 25 mM Tris buffer of pH 7.8, containing 2.5 mM calcium chloride (buffer A). The concentrate was pumped onto the CM and p-aminobenzamidinium columns at a flow rate of about 100 ml per minute. When the application of the concentrated solution onto the columns was completed, residual material was washed off the continuous column system
15 with about 8 liters of buffer A. After this wash was completed, the p-aminobenzamidinium affinity column was disconnected from the CM column and washed with about 5 column volumes of a high salt solution of 25 mM Tris buffer pH 7.5 containing 0.5 M NaCl and 2.5 mM calcium chloride. The cod tryptins were then desorbed from the affinity ligand and eluted off the column with an acid solution of 25 mM acetic acid pH 3.2 containing 10 mM calcium
20 chloride and 30% glycerol.

The purified cod trypsin ZT isoform-containing preparation was homogeneous by SDS PAGE electrophoresis. The purified preparation was filter-sterilized through a 0.22 micron filter and stored frozen at about -20°C. The cod trypsin ZT isoform-containing preparation was filter-sterilized through a 0.22 micron filter and stored frozen at about -20°C. The cod
25 trypsin ZT isoform-containing preparation was applied to an anion exchange chromatography column (MonoQ) and proteins eluted with a salt gradient (FIG 1). The column was equilibrated with a buffer (20 mM Tris, 10 mM CaCl₂, pH 8.0), and the enzymes were eluted

with a linear 0-150 mM NaCl gradient in 40 column volumes, linear 150-620 mM NaCl in 6 column volumes, 620-850 mM NaCl in 10 column volumes at a flow rate of 1 mL/min. The proteins in the peak labeled trypsin ZT from FIG 1 were subjected to SDS-PAGE analysis (FIG 2). The protein band on the gel from the trypsin ZT peak was excised from the gel and
5 analyzed using Matrix-Assisted Laser Desorption/Ionization Time-of-Flight Mass Spectrometry (MALDI-TOF MS). The mass spectrometry (MS) analysis revealed identity to trypsin ZT-1, trypsin ZT-2, trypsin ZT-3 and trypsin ZT-4 (Table 1 and Table 2). Antibodies against trypsin I and trypsin X were prepared in rabbits against a peptide ((NH₂-) CVLSGWVRDTMA (-COOH)) (SEQ ID NO: 23) corresponding to residues 228-239 at the
10 extreme C terminus of cod trypsin I and cod trypsin X. The antibodies were affinity purified from rabbit serum using the peptide coupled to a gel bead support. Polyclonal antibodies against trypsin ZT isoforms were made in mouse ascitic fluid as described in Overkamp et al. (Overkamp, Mohammed-Ali et al., 1988, *J Immunoassay* **9**: 51-68). A purified form of recombinant trypsin ZT-4 was injected intraperitoneally (i.p.) into Balb/C2 mice. The mice
15 were sacrificed on day 34 and ascitic fluid collected. The respective antibodies were used in Western blot analysis. Western blot analysis was performed on the proteins in the peak labeled trypsin ZT from FIG 1 (FIG 3). Trypsin ZT isoform antibody reacted against the proteins from the peak labeled trypsin ZT whereas an antibody against trypsin I and trypsin X did not (FIG 3).

20 Table 1 shows peptide masses obtained from MS analysis on protein band from SDS-PAGE gel in Figure 2 (non guanidilated sample), see column 1 Mr (expt). Mr stands for relative molecular mass in Da. Columns 2, 3 and 4 show the calculated values (for masses matching to masses in column 1) from an *in silico* trypsin digest of the amino acid sequences trypsin ZT-1, trypsin ZT-2, trypsin ZT-3 and trypsin ZT-4. Column 5 shows the sequence of
25 the peptide that give the mass value from the *in silico* digest. Column 6 shows modifications of the different peptides based on the *in silico* digest.

Table 1.

Mr (expt)	Trypsin ZT-4	Trypsin ZT-3	Trypsin ZT-2	Trypsin ZT-1	Sequence	Modifications
842,4077						
1045,4976						
1082,5603						
1144,4826	1144,5415	1144,5415			(R)IIGGQDCEPR(S)	Carbamidomethyl (C)
1149,5283	1149,6150	1149,6150	1149,6150	1149,6150	(R)VFEGTEQLVK(T)	
1212,4511						
1216,4408	1216,4908		1216,4908		(R)MVCAGYMDGGR(D)	Carbamidomethyl (C)
1260,4492		1260,4992		1260,4992	(R)MMCVGFMDGGR(D)	Carbamidomethyl (C)
1262,4221	1262,4962		1262,4962		(R)MVCAGYMDGGR(D)	2 Oxidation (M) Propionamide (C)
1276,4272		1276,4941		1276,4941	(R)MMCVGFMDGGR(D)	Carbamidomethyl (C) Oxidation (M)
1320,5165		1320,6253		1320,6253	(R)GCAEPNSPGVYVK(V)	
1354,5907						
1377,5624		1377,6467		1377,6467	(R)GCAEPNSPGVYVK(V)	Carbamidomethyl (C)
1493,5870						
1638,7873						
1707,6807						
1839,8099						
1994,9057						
2800,2263		2800,3831		2800,3831	(R)LQCLDVPIVEPVAC QASYPGMISPR(M)	2 Carbamidomethyl (C)
2814,2398		2814,3987		2814,3987	(R)LQCLDVPIVEPVAC QASYPGMISPR(M)	Carbamidomethyl (C) Propionamide (C)
2830,2763		2830,3936		2830,3936	(R)LQCLDVPIVEPVAC QASYPGMISPR(M)	Carboxymethyl (C) Oxidation (M) Propionamide (C)
2847,3361	2847,3725		2847,3725		(R)LQCLDVPIVETVDC EAAYPGMISPR(M)	Carbamidomethyl (C) Propionamide (C)

The sequences in Table 1 are as follows:

Seq ID NO	Amino acid sequence
SEQ ID NO 6	IIGGQDCEPR
SEQ ID NO 7	VFEGTEQLVK
SEQ ID NO 8	LQCLDVPIVEPVACQASYPGMISPR
SEQ ID NO 8	LQCLDVPIVEPVACQASYPGMISPR
SEQ ID NO 8	LQCLDVPIVEPVACQASYPGMISPR
SEQ ID NO 9	LQCLDVPIVETVDCEAAYPGMISPR
SEQ ID NO 10	MVCAGYMDGGR
SEQ ID NO 11	MMCVGFMDGGR
SEQ ID NO 10	MVCAGYMDGGR
SEQ ID NO 11	MMCVGFMDGGR
SEQ ID NO 12	GCAEPNSPGVYVK
SEQ ID NO 12	GCAEPNSPGVYVK

Table 2 shows peptide masses obtained from MS analysis on protein band from SDS-PAGE gel in Figure 2 (guanidilated sample), see column 1 Mr (expt). Mr stands for relative molecular mass in Da. Columns 2, 3 and 4 show the calculated values (for masses matching to masses in column 1) from an *in silico* trypsin digest of the amino acid sequences of trypsin ZT-1, trypsin ZT-2, trypsin ZT-3 and trypsin ZT-4. Column 5 shows the sequence of the peptide that give the mass value from the *in silico* digest. Column 6 shows modifications of the different peptides based on the *in silico* digest.

Table 2.

Mr (expt)	Trypsin ZT-4	Trypsin ZT-3	Trypsin ZT-2	Trypsin ZT-1	Sequence	Modifications
781,5661						
842,4931						
1144,5651	1144,5415	1144,5415			(R)IIGGQDCEPR(S)	Carbamidomethyl (C)
1173,6305						
1191,6567	1191,6368	1191,6368	1191,6368	1191,6368	(R)VFEGTEQLVK(T)	GuanidinyI (K)
1210,5861						
1212,5237						
1216,5105	1216,4908		1216,4908		(R)MVCAGYMDGGR(D)	Carbamidomethyl (C)
1222,6291						
1260,5201		1260,4992		1260,4992	(R)MMCVGFMDGGR(D)	Carbamidomethyl (C)
1270,6331						
1276,5112		1276,4941		1276,4941	(R)MMCVGFMDGGR(D)	Carbamidomethyl (C) Oxidation (M)
1638,8623						
1993,9786						
2635,2760		2635,2127		2635,2127	(R)DVCNGDSGSPLVC EGVLTGLVSWGR(G)	Carbamidomethyl (C) Carboxymethyl (C)
2649,2755		2649,2283		2649,2283	(R)DVCNGDSGSPLVC EGVLTGLVSWGR(G)	Carboxymethyl (C) Propionamide (C)
2752,4779						
2762,6478						
2766,5037						
2776,6052	2776,3354		2776,3354		(R)LQCLDVPIVETVD CEAAYPGMISPR(M)	
2800,4952		2800,3831		2800,3831	(R)LQCLDVPIVEPVA CQASYPGMISPR(M)	2 Carbamidomethyl (C)
2814,5098		2814,3987		2814,3987	(R)LQCLDVPIVEPVA CQASYPGMISPR(M)	Carbamidomethyl (C) Propionamide (C)
2830,5112		2830,3936		2830,3936	(R)LQCLDVPIVEPV ACQASYPGMISPR(M)	Carboxymethyl Oxidation Propionamide
2833,4913	2833,3569		2833,3569		(R)LQCLDVPIVETV DCEAAYPGMISPR(M)	2 Carbamidomethyl
2847,5160	2847,3725		2847,3725		(R)LQCLDVPIVETV DCEAAYPGMISPR(M)	Carbamidomethyl (C) Propionamide

The sequences in Table 2 are as follows:

Seq ID No	Amino acid sequence
SEQ ID NO 6	IIGGQDCEPR
SEQ ID NO 7	VFEGTEQLVK
SEQ ID NO 9	LQCLDVPIVETVDCEAAYPGMISPR
SEQ ID NO 8	LQCLDVPIVEPVACQASYPGMISPR
SEQ ID NO 8	LQCLDVPIVEPVACQASYPGMISPR
SEQ ID NO 8	LQCLDVPIVEPVACQASYPGMISPR
SEQ ID NO 9	LQCLDVPIVETVDCEAAYPGMISPR
SEQ ID NO 9	LQCLDVPIVETVDCEAAYPGMISPR
SEQ ID NO 10	MVCAGYMDGGR
SEQ ID NO 11	MMCVGFMDGGR
SEQ ID NO 11	MMCVGFMDGGR
SEQ ID NO 13	DVCNGDSGSPLVCEGVLTLGLVSWGR
SEQ ID NO 13	DVCNGDSGSPLVCEGVLTLGLVSWGR

5 **EXAMPLE 3.**

Multiplex substrate profiling on trypsin ZT isoforms

Multiplex substrate profiling was used to analyze the substrate specificity of enzymes (proteinases) (O'Donoghue, Eroy-Reveles et al., 2012, *Nat Methods* **9**: 1095-1100).

Proteinases can cleave substrates at different amino residues within a peptide sequence

10 demonstrating a difference in substrate specificity. Trypsins are specified by their preference in cleaving C-terminal to arginine or lysine residues.

Enzyme isoforms

15 The cod trypsin isoenzymes were separated by applying benzamidine purified cod trypsin to a MonoQ HR 5/5 ion exchange column. For isolation of trypsin I, the column was equilibrated with 20 mM Tris, 5 mM ethanolamine, 10 mM CaCl₂, pH 9.1, and the enzymes were eluted with a linear 0-200 mM NaCl gradient in 80 column volumes, linear 200-1000

mM NaCl gradient in 10 column volumes at a flow rate of 1 mL/min. The sample pH was adjusted to pH 9.1 with 1 M NaOH before application.

For isolation of trypsin ZT isoforms, the column was equilibrated with 25 mM Tris, 25 mM CaCl₂, 15% (vol/vol) glycerol, pH 7.5, and the enzymes were eluted with a linear 0-150 mM NaCl gradient in 5 column volumes, linear 150-550 mM NaCl gradient in 20 column volumes, linear 500-1000 mM NaCl gradient in 1 column volume at a flow rate of 1 mL/min.

The composition of both enzyme samples (trypsin ZT and trypsin I) was adjusted so the final concentration of Tris was 60 mM, CaCl₂ 30 mM and glycerol 15% (vol/vol) with a pH of 8.5.

Multiplex substrate profiling analysis

The purified trypsin ZT isoforms or trypsin I were incubated (5, 15 and 60 min.) with a library of peptides (124 defined peptides, 14-mer sequences, total of 1612 peptide bonds) with an extensive physiochemical diversity as described in O'Donoghue, Eroy-Reveles et al. (O'Donoghue, Eroy-Reveles et al., 2012, *Nat Methods* **9**: 1095-1100). After incubation, cleavage sites were identified by mass spectrometry analysis (O'Donoghue, Eroy-Reveles et al., 2012, *Nat Methods* **9**: 1095-1100).

Results

The multiplex substrate profiling analysis unexpectedly showed that there are numerous cleavage sites unique to trypsin ZT isoforms compared trypsin I, disclosed in WO 2000078332 (FIG 4 and Table 4). Also, cleavage sites that showed preferential cleavage by trypsin ZT isoforms compared to trypsin I were identified (Table 4). Based on the results from the multiplex substrate profiling analysis, a table containing numerical values based on standard scores (Z-score) ratio at the P4-P4' positions in the substrate was generated where the substrate specificity of trypsin ZT was compared to trypsin I (Table 3). Positive values (Z-score) reflect a preference for the amino acid residue in a certain P position in favor of trypsin

ZT isoforms compared to trypsin I. Unexpectedly, multiplex substrate profiling on trypsin ZT isoforms revealed that amino acids surrounding arginine or lysine in a substrate have a different effect on its cleavage compared to trypsin I.

Table 3 shows numerical values based on standard scores (Z-score) ratio at the P4-P4' positions in the substrate comparing the substrate specificity of trypsin ZT and trypsin I after incubation for 5, 15 and 60 min. Amino acid residues in a substrate undergoing cleavage were designated P1, P2, P3, P4 in the N-terminal direction from the cleaved bond (Schechter and Berger, 1967, *Biochem Biophys Res Commun* **27**: 157-162, Schechter and Berger, 1968, *Biochem Biophys Res Commun* **32**: 898-902). The residues in C-terminal direction are designated P1', P2', P3', P4'. The P positions are provided in the columns and the rows show amino acids in the different P positions. Positive values (Z-score) in the table reflect a preference for the amino acid residue in a certain P position in favor of trypsin ZT isoforms compared to trypsin I. Negative values (Z-score) reflect a preference in favor of trypsin I compared trypsin ZT. For example, based on the values from Table 3 it can be seen that trypsin ZT is much better adapted in cleaving peptides containing several consecutive positively charged residues (arginine or lysine) compared to trypsin I. Again, those types of sequences, that is lysine- and arginine-rich amino acid sequences, are frequently found in protein toxins, proteins and peptides involved in pathogenesis (Suzuki, Orba et al., 2010, *PLoS Pathog* **6**: e1000801, Fentress, Steinfeldt et al., 2012, *Cell Microbiol* **14**: 1921-1933, Jiang, Cun et al., 2012, *J Virol* **86**: 7256-7267, Milletti, 2012, *Drug Discov Today* **17**: 850-860, Gallaher and Garry, 2015, *Viruses* **7**: 285-305). As previously mentioned, trypsin ZT isoforms have therefore properties beneficial against proteins responsible for the pathogenicity of many microorganisms.

Table 3.

5 min.	Amino acid							
Position	P4	P3	P2	P1	P1'	P2'	P3'	P4'
G	-0,60	0,88	0,48	0,26	-1,21	0,11	0,97	0,08
A	0,01	0,48	1,92	-0,57	2,31	1,01	-0,62	-0,14
S	-0,06	-0,04	0,43	0,23	0,25	-0,52	-1,47	0,32
P	-1,04	-0,59	0,39	-0,16	0,25	1,86	-0,30	-0,50
V	0,99	-1,14	-1,88	0,23	0,43	0,54	0,01	0,38
T	0,05	-1,40	0,43	0,72	0,48	-0,02	-1,39	-0,06
L	-0,50	0,48	-2,26	0,25	-0,75	-0,43	0,88	0,91
I	1,42	-0,51	-0,31	0,72	-0,67	-0,36	-0,04	-0,62
N	0,95	0,93	-0,47	-0,15	-1,33	-0,79	-0,56	0,02
D	0,33	0,43	0,24	-0,48	-0,02	-0,53	-0,52	-0,69
Q	-1,38	-0,89	-0,87	0,70	-0,01	-0,55	-0,01	1,10
K	-0,12	0,38	1,88	-6,09	1,02	-1,11	-0,14	-0,27
E	0,38	-0,66	-0,96	0,30	-0,53	-1,06	0,48	-0,09
M	-1,91	-1,27	-0,28	0,25	-0,79	-0,86	0,33	0,38
H	-0,09	0,38	0,92	0,25	-0,39	0,58	0,38	1,49
F	0,49	-0,41	-0,26	0,72	-0,01	-0,42	-0,58	0,10
R	-0,69	-0,09	-0,07	2,69	1,95	0,99	0,12	0,27
Y	-0,60	0,75	1,04	0,23	-0,15	-0,42	0,95	-0,12
W	1,00	0,64	-0,84	-0,15	-0,59	1,00	1,36	-0,53

Table 3 continued.

15 min.	Amino acid							
Position	P4	P3	P2	P1	P1'	P2'	P3'	P4'
G	0,33	1,65	-0,27	0,48	-0,64	0,60	1,79	-0,36
A	-0,34	-0,66	1,33	-0,52	1,88	-0,60	-1,05	0,17
S	0,30	-0,41	0,73	0,44	0,51	0,03	-1,16	0,55
P	0,26	-0,62	0,23	0,12	0,54	1,70	0,66	-0,79
V	0,55	-0,08	-0,75	0,44	0,36	0,97	0,97	0,81
T	0,04	-0,25	-0,50	0,89	0,44	0,81	-1,21	-0,63
L	0,49	0,11	-1,57	0,18	0,18	0,44	0,73	0,73
I	-1,07	-0,89	-0,97	0,44	-0,54	0,12	0,32	0,04
N	-0,86	0,81	0,96	0,89	-1,50	0,67	-1,74	-0,89
D	-1,45	-0,43	0,07	-0,03	0,11	-2,02	0,42	-0,76
Q	-0,32	-0,71	-1,07	0,89	-0,28	-0,49	0,43	0,73
K	-0,72	-0,16	1,35	-6,74	0,96	-1,11	0,25	0,48
E	0,25	-0,72	-0,01	0,54	-0,39	-1,35	-1,49	0,25
M	-0,89	-0,59	-0,16	0,47	0,25	-2,23	0,19	1,55
H	0,33	0,26	1,26	0,89	-0,67	0,29	-0,19	0,40
F	1,34	0,15	-0,82	0,89	-0,33	-0,36	-0,26	0,20
R	-0,67	-0,17	-0,82	-1,11	2,17	1,35	0,65	-0,78
Y	0,30	0,40	0,58	0,44	-1,00	0,57	0,89	-1,07
W	0,49	-0,26	-1,38	0,13	-1,83	-0,57	0,36	0,50

Table 3 continued.

60 min.	Amino acid							
Position	P4	P3	P2	P1	P1'	P2'	P3'	P4'
G	0,45	0,80	-1,05	0,26	-0,27	0,71	0,65	-0,93
A	0,05	-0,74	1,65	-0,30	2,33	0,39	-0,43	-1,56
S	-0,30	0,57	0,52	0,52	0,60	-1,19	-0,08	0,59
P	-0,72	0,42	-1,24	-0,05	0,90	2,28	-0,28	-0,57
V	-0,25	0,51	-0,09	0,84	-0,55	1,03	-0,19	0,91
T	0,49	-0,07	0,91	0,52	-0,54	0,80	-1,21	0,13
L	-0,23	-0,47	-0,73	-0,06	0,66	0,22	0,41	0,38
I	-0,73	-0,53	0,04	0,20	0,44	0,20	0,85	0,61
N	-0,90	0,15	0,28	-0,05	-1,49	-0,43	-1,18	-0,18
D	-1,33	-0,03	-0,16	1,32	-0,70	-1,42	-0,26	0,67
Q	0,63	-0,44	-1,36	0,85	0,33	-0,17	-1,15	1,09
K	0,43	-0,02	1,24	-7,02	0,97	-1,46	0,73	0,90
E	-0,01	-1,45	-0,21	0,54	-1,36	-1,64	-1,11	-0,36
M	-1,39	-0,36	0,01	0,24	-0,24	-0,93	-0,37	0,79
H	0,79	-1,38	1,23	0,85	-1,06	-0,30	0,47	0,57
F	0,48	0,15	-1,67	0,52	-0,06	0,19	0,85	-0,07
R	-0,46	-0,02	-0,91	-0,91	1,67	0,91	1,50	-0,11
Y	0,73	0,41	0,25	0,52	-1,36	0,58	1,60	-2,25
W	0,85	1,02	0,09	0,90	-0,21	0,03	-0,49	-0,42

Table 4 shows precursor ion intensities of selected peptides from multiplex profiling after incubation with trypsin ZT or with trypsin I. The precursor ion intensities are peptide peak intensities in the MS before fragmentation that represent a measure of relative abundance for a given species. X in peptide sequence denotes a placeholder for the absence of an amino acid in a peptide sequence. The data shows that trypsin ZT isoforms are exclusively able to cleave three peptide sequences (QGKKAPXX, WGNRSPLE and QAVRPNGM) compared to trypsin I. Furthermore, the data show that trypsin ZT isoforms preferentially cleave two peptides compared to trypsin I. Trypsin ZT isoforms were able to cleave XIARQPWN within 5 minutes and FDNRVGKW within 15 minutes whereas cleavage within these two peptides was only detected with trypsin I after 60 minute incubation.

Table 4.

Sequence	SEQ ID NO.	Ion Precursor Intensities					
		Trypsin ZT			Trypsin I		
		5'	15'	60'	5'	15'	60'
QGKKAPXX	SEQ ID NO 14	175343	278749	256859	0	0	0
WGNRSPLE	SEQ ID NO 15	21152	42866	47692	0	0	0
QAVRPNGM	SEQ ID NO 16	0	12772	29668	0	0	0
FDNRVGKW	SEQ ID NO 17	0	187219	937228	0	0	164917
XIARQPWN	SEQ ID NO 18	130793	0	350390	0	0	188625

EXAMPLE 4.

Purification of cod trypsin ZT isoforms from concentrated cod viscera extract

About 12 liters of ultrafiltered and diafiltered concentrate as obtained in Example 1 was applied to a continuous connected series of about 1 liter packed chromatography columns, the first containing a CM fast flow cation exchange resin (GE healthcare), the second one a p-aminobenzamidinium affinity ligand coupled to a sepharose resin (GE healthcare) and the third one DEAE fast flow anion exchange resin (GE healthcare). The CM and p-aminobenzamidinium columns were pre-equilibrated with about 10 column volumes of 25 mM

Tris buffer of pH 7.8, containing 2.5 mM calcium chloride. The concentrate was pumped onto the CM and p-aminobenzamidine columns at a flow rate of about 100 ml per minute. When the application of the concentrated solution onto the columns was completed, residual material was washed off the continuous column system with about 8 liters of buffer A. After this wash was completed, the p-aminobenzamidine affinity column was disconnected from the CM column and washed with about 5 column volumes of a high salt solution of 25 mM Tris buffer pH 7.5 containing 0.5 M NaCl and 2.5 mM calcium chloride. The cod trypsins were then desorbed from the affinity ligand and eluted off the column with an acid solution of 25 mM acetic acid pH 3.2 containing 10 mM calcium chloride and 30% glycerol. The cod trypsin fraction was collected into a neutralizing buffer of 200 mM Tris pH 8.5 containing 30% glycerol. The DEAE anion exchange column was pre-equilibrated with about 10 column volumes of 25 mM Tris buffer of pH 7.8, containing 2.5 mM calcium chloride. The cod trypsin fraction was applied to the DEAE anion exchange resin and washed with about 5 column volumes of the equilibration buffer. Trypsin ZT isoforms were then desorbed from the column using up to a 0.9 M NaCl gradient salt solution. The purified cod trypsin ZT isoform preparation was homogeneous by SDS PAGE electrophoresis and FPLC Mono Q chromatography. The purified preparation was filter-sterilized through a 0.22 micron filter and stored frozen at about -20°C.

EXAMPLE 5.

Preparation of a hydrogel preparation of cod trypsins, including cod trypsin ZT isoforms

The purified cod trypsin ZT isoform containing preparation of Example 2 was mixed with hydrocolloid gel comprising an aqueous gel containing 0.8% w/v Carbomer 940 and 40% glycerol. The cod trypsin preparation was mixed in a 1:1 ratio with the hydrogel to give a final concentration of 4 enzyme unit per mg (U/mg) of the final gel-enzyme mixture (the enzyme hydrogel ointment), the enzyme unit being determined using Cbz- GPR-pNA as

substrate as previously described. Thus, the resulting enzyme hydrogel ointment contained about 0.01 mg/ml, or 1 U/ml of the cod trypsin enzymes, 0.4% Carbomer 940, 20% glycerol and 0.04% paraoxybenzoate.

5 **EXAMPLE 6.**

Antiviral activity of trypsin ZT isoforms against enterovirus as a model for rhinovirus

Picornaviruses are small single-stranded positive sense RNA viruses that lack a lipid envelope. Enterovirus and rhinovirus are two genera of *Picornaviridae*. Enterovirus replicates initially in the oropharynx and survives the acidic environment of the stomach. The small
10 intestine is the major invasion site of enterovirus. Rhinovirus replicates in the nasal passages but is destroyed in the acidic environment of the stomach. Rhinovirus and enterovirus have an identical morphology but can be distinguished based on clinical, biophysical and epidemiological studies. Rhinovirus is problematic to handle in cell cultures (Racaniello, 2007, *Fields virology*, 5th ed: 795–838). Therefore, enterovirus was used as a model for
15 rhinovirus to analyze the antiviral properties of trypsin ZT isoforms.

Materials and Methods

Enterovirus stock solution

Enterovirus (Coxsackievirus B2) was isolated from a patient at the University Hospital
20 of Iceland, department of Virology and grown in MA-104 cells (African green monkey kidney cells) or Vero cells (African green monkey kidney cells). The Coxsackievirus B2 was titrated (TCID₅₀ = 8,23) according to Reed-Muench (REED and MUENCH, 1938, *American Journal of Epidemiology* **27**: 493-497).

25 Enzyme isoforms

Cod trypsin isoenzymes were separated by applying benzamidine purified cod trypsin to a MonoQ HR 5/5 ion exchange column. For isolation of trypsin ZT isoforms, the column was

equilibrated with 25 mM Tris, 25 mM CaCl₂, pH 7.5, and the enzymes were eluted with a linear 0-400 mM NaCl gradient in 5 column volumes, linear 400-1000 mM NaCl in 20 column volumes at a flow rate of 1 mL/min.

5 Treatment

The MA-104 cells were grown in a 96-well microtiter plate to 95% confluency. The enterovirus was diluted from a stock solution to 10⁻⁶ using minimum essential medium (MEM) and 1.5 mL of the diluted solution was placed in two different vials. The vials were treated with 1.5 mL of trypsin ZT isoforms and 1.5 mL MEM as positive control. Negative control was
10 MEM (3 mL) without virus. All vials were incubated at 34 °C and 5% CO₂ for 90 min before adding 100 µL of benzamidine agarose solution to remove residual trypsin and incubated for another 30 min. The solution was centrifuged at 5500 rpm for 3 min before placing 2 mL of the supernatant into an Eppendorf vial. 200 µL of the solution was placed in each well, total of 10 wells in the microtiter plate. The plate was incubated for 30 min. After incubation, the
15 media was thrown off and MEM with 1% newborn calf serum (NCS) added to all the wells. A microscope was used to monitor for the presence of infection.

Results

Trypsin ZT isoforms were very effective as antiviral agents against enterovirus
20 (Coxsackievirus B2) that was used as a model for rhinovirus. The results are presented in FIG 5.

Claims

1. An isolated cod trypsin ZT isoform comprising an amino acid sequence according to
SEQ ID NO:1, or an amino acid sequence with 80% sequence homology or more,
5 thereof.
2. The isolated cod trypsin ZT isoform according to claim 1, selected from cod trypsin
ZT-1, comprising an amino acid sequence according to SEQ ID NO:2; cod trypsin ZT-
2 comprising an amino acid sequence according to SEQ ID NO:3; cod trypsin ZT-3
10 comprising an amino acid sequence according to SEQ ID NO:4; and cod trypsin ZT-4
comprising an amino acid sequence according to SEQ ID NO:5, or an amino acid
sequence with 80% sequence homology or more, thereof.
3. A composition comprising at least one cod trypsin ZT isoform according to claim 1 or
15 2, together with suitable excipients and carriers.
4. The composition according to claim 3, wherein said cod trypsin ZT isoform(s) are
present in admixture with other cod trypsins.
- 20 5. The composition according to claim 4, wherein said cod trypsin ZT isoform(s)
comprises at least 5% (w/w) of the total content of cod trypsins in said composition.
6. The composition according to any one of claims 3 to 5, which is administered
topically, as a lotion, a hydrogel, a mouth spray or a nasal spray.
- 25 7. The composition according to any one of claims 3 to 6, further comprising a
polyvalent alcohol, such as glycerol.

8. A composition according to any one of claims 3 to 7, for use in therapy.
9. A composition according to any one of claims 3 to 7, for use in treating or preventing
5 a disease caused by a pathogenic organism selected from the group consisting of a virus, a bacterium, a fungus, a parasite and a protozoan.
10. A composition according to any one of claims 3 to 7, for use in treating or preventing
10 a disease in the upper respiratory tract caused by pathogenic organisms such as viruses, bacteria and fungus.
11. A composition according to any one of claims 3 to 7, for use in treating or preventing
pain, acute inflammation, chronic inflammation, arthritis, inflamed joints, bursitis,
osteoarthritis, rheumatoid arthritis, juvenile rheumatoid arthritis, septic arthritis,
15 fibromyalgia, systemic lupus erythematosus, phlebitis, tendinitis, rash, psoriasis, acne, eczema, facial seborrheic eczema, eczema of the hands, face or neck, foreskin infections, athlete's foot, fistulae infections, infected topical ulcers, navel infections in newborns, wrinkles, scars, keloids, boils, warts and allergic itch, hemorrhoids, a fungal infection and an immunological disorder including an autoimmune disease.
20
12. A composition according to any one of claims 3 to 7, for use in the treatment of wound infections and wounds from burns.
13. A composition according to any one of claims 3 to 7, for use in removing dead or
25 peeling skin from otherwise healthy skin.
14. A composition according to any one of claims 3 to 7, for cosmetic use.

15. The composition for cosmetic use, according claim 14, said composition further comprising an additional cosmetically active compound.

5 16. A method of treating disease, comprising administering a therapeutically effective amount of a composition according to any one of claims 3 to 7, to a patient in need thereof.

10 17. The method according claim 16, wherein said disease is caused by a pathogenic organism selected from the group consisting of a virus, a bacterium, a fungus, a parasite and a protozoan.

18. The method according claim 16, wherein said disease is a disease in the upper respiratory tract caused by pathogenic organisms such as viruses and bacteria.

15

19. The method according claim 16, wherein said disease is selected from pain, acute inflammation, chronic inflammation, arthritis, inflamed joints, bursitis, osteoarthritis, rheumatoid arthritis, juvenile rheumatoid arthritis, septic arthritis, fibromyalgia, systemic lupus erythematosus, phlebitis, tendinitis, rash, psoriasis, acne, eczema, 20 facial seborrheic eczema, eczema of the hands, face or neck, foreskin infections, athlete's foot, fistulae infections, infected topical ulcers, navel infections in newborns, wrinkles, scars, keloids, boils, warts and allergic itch, hemorrhoids, a fungal infection and an immunological disorder including an autoimmune disease.

25 20. The method according claim 16, wherein said disease is wound infections or wounds from burns.

21. A method of preparing cod trypsin ZT isoforms comprising an amino acid sequence according to any one of SEQ ID NO:2 to SEQ ID NO:5, comprising

- (i) preparing an aqueous extract of cod viscera,
- (ii) subjecting the aqueous extract to at least one or chromatography step,
5 including an affinity chromatography step using a p-aminobenzamidine affinity ligand, and
- (iii) desorbing and eluting cod trypsin ZT isoforms bound to the p-aminobenzamidine affinity ligand.

10 22. The method according to claim 21, further comprising at least one step using an anionic exchange resin after said affinity chromatography step (ii).

23. The method according to claim 21, further comprising at least one step using a cation exchange resin prior to the affinity chromatography step (ii).

15

24. The method according to any one of claims 21 to 23, wherein said desorbed and eluted cod trypsin ZT isoforms are sterile filtered using a filter with 0.45 µm or smaller pore size.

20 25. The method according to any one of claims 21 to 24, wherein said cod trypsin ZT isoforms are selected from trypsin ZT-1, comprising an amino acid sequence according to SEQ ID NO:2; trypsin ZT-2 comprising an amino acid sequence according to SEQ ID NO:3; trypsin ZT-3 comprising an amino acid sequence according to SEQ ID NO:4; and trypsin ZT-4, comprising an amino acid sequence
25 according to SEQ ID NO:5, or a mixture thereof.

Figure 1.

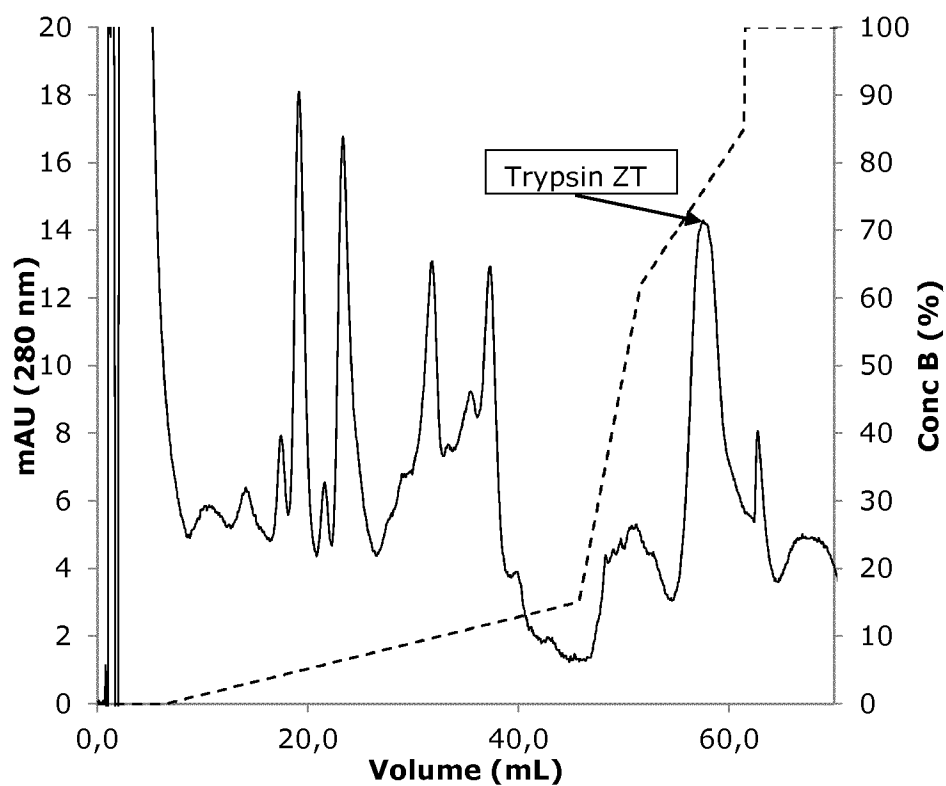


Figure 2.

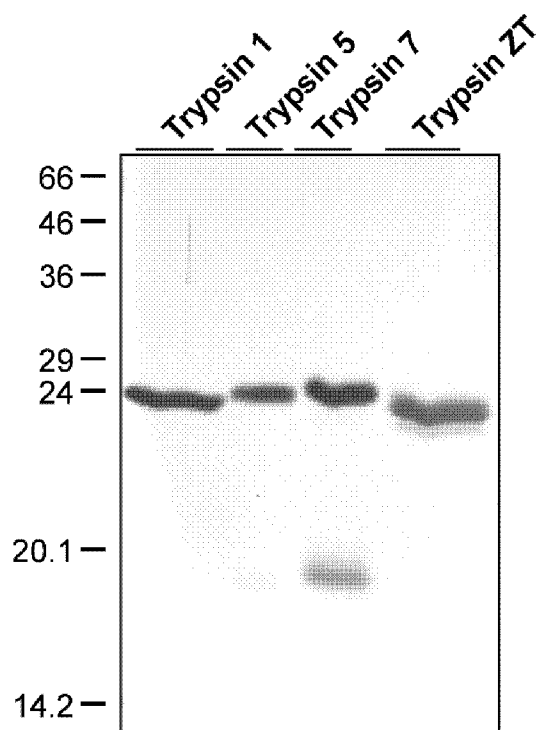


Figure 3.

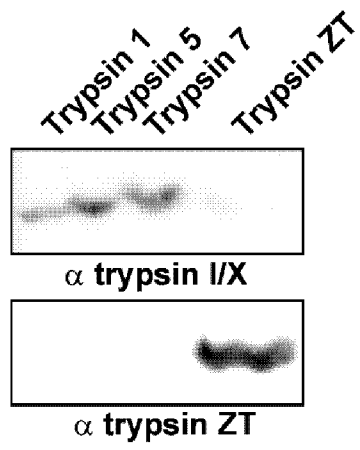


Figure 4.

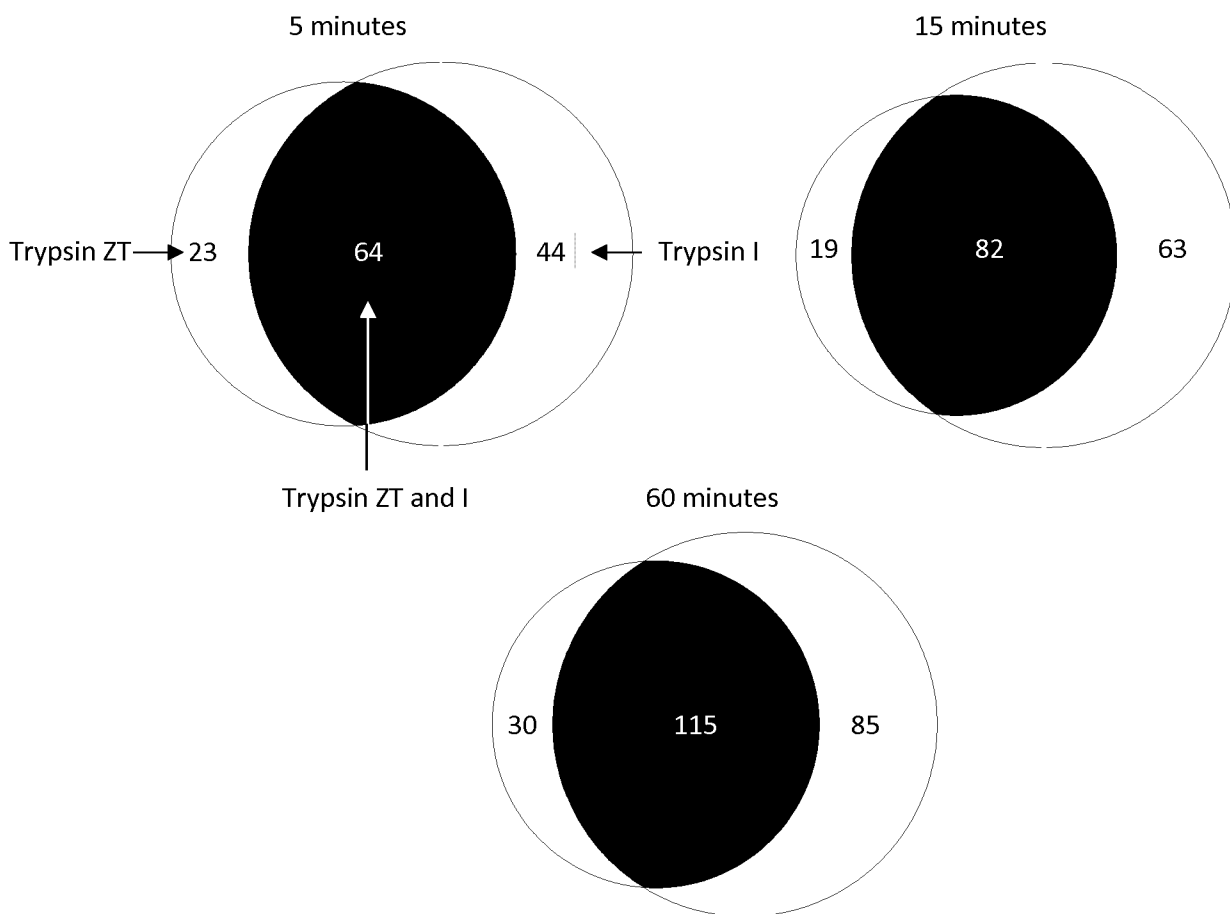
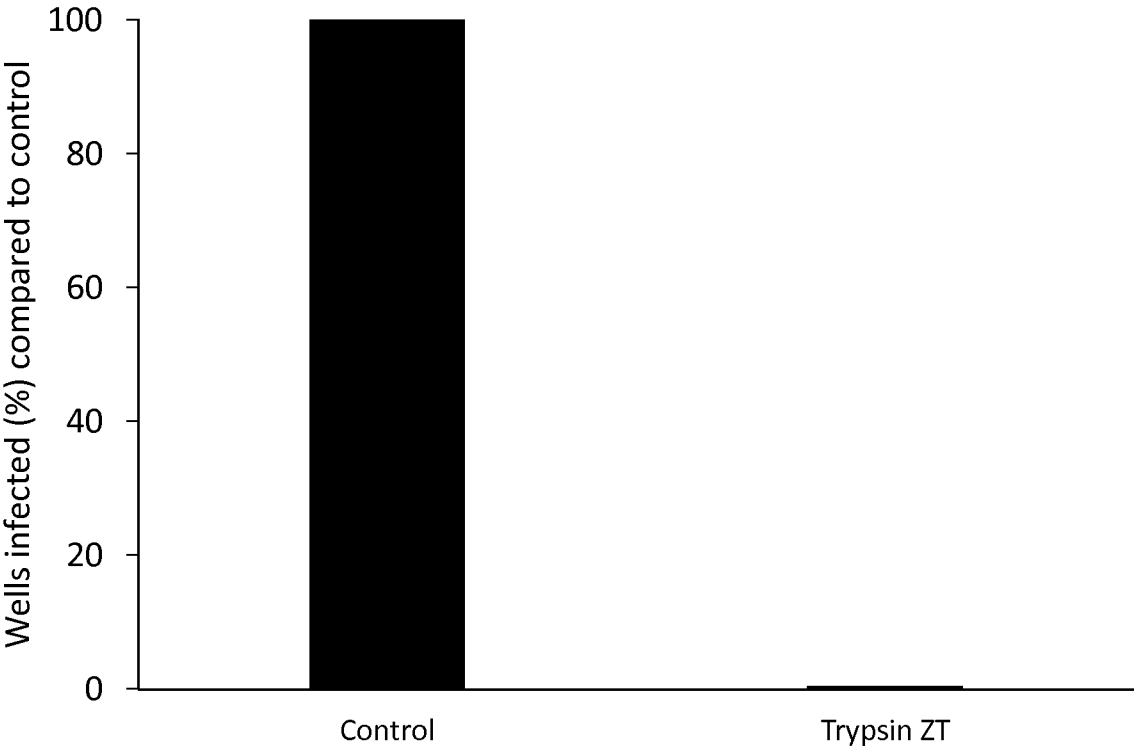


Figure 5.



INTERNATIONAL SEARCH REPORT

International application No

PCT/EP2016/067531

A. CLASSIFICATION OF SUBJECT MATTER
 INV. C12N9/76
 ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, BIOSIS, EMBASE, WPI Data, EMBL

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	SPILLIAERT R ET AL: "Atlantic cod trypsin Y-member of a novel trypsin group", 1 January 1999 (1999-01-01), MARINE BIOTECHNOLOGY, SPRINGER VERLAG, NEW YORK, NY, US, PAGE(S) 598 - 607, XP009187291, ISSN: 1436-2228 figures 1, 2 -& DATABASE UniProt [Online] 24 June 2015 (2015-06-24), XP002751139, Database accession no. Q8JFQ7	1-25
Y	WO 00/78332 A2 (BJARNASON JON BRAGI [IS]) 28 December 2000 (2000-12-28) cited in the application page 2, paragraph 2 page 4, paragraph 4 page 10, paragraphs 1, 2	1-25



Further documents are listed in the continuation of Box C.



See patent family annex.

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Date of the actual completion of the international search

19 September 2016

Date of mailing of the international search report

27/09/2016

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INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2016/067531

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