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Process for purifying (-)-delta9-trans-tetrahydrocannabinol

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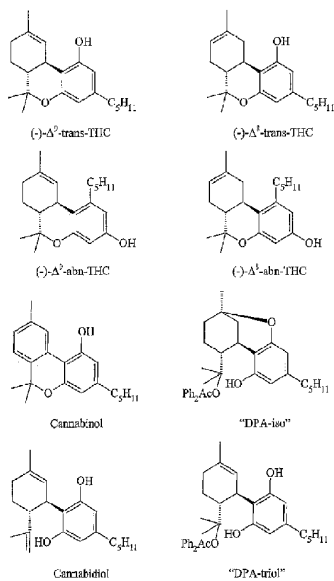
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(54) Title: PROCESS FOR PURIFYING (-)- Δ^5 -TRANS-TETRAHYDROCANNABINOL



(57) Abstract: A preparative separation process wherein (-)- Δ^5 -trans-tetrahydrocannabinol is separated from a mixture of cannabinoids is disclosed. The process comprises at least one chromatographic step wherein a mobile phase passes through a stationary phase. The stationary phase comprises a derivatised polysaccharide and the mobile phase comprises carbon dioxide.

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PROCESS FOR PURIFYING (-)- Δ^9 -TRANS-TETRAHYDROCANNABINOL

5 The present invention relates to a process for purifying (-)- Δ^9 -trans-tetrahydrocannabinol. The compound is separated from a mixture of cannabinoids using a chromatographic technique.

10 (-)- Δ^9 -trans-tetrahydrocannabinol is the active ingredient in marijuana. It is used therapeutically as an inhalant or an oral drug for stimulation of appetite among AIDS and cancer chemotherapy patients. Tetrahydrocannabinols (THCs) can be isolated from marijuana (a mixture of leaves and flowering heads of the plant *Cannabis Sativa*). Alternatively, THCs can be obtained by synthetic routes, e.g. as described in WO 02/096899. Enantiomerically pure THCs are required for formulation into drug
15 products, but the purification of THCs, whether produced by isolation or synthesis, is challenging. The present inventors have sought to provide a process for providing enantiomerically pure (-)- Δ^9 -trans-tetrahydrocannabinol ((-)- Δ^9 -THC).

20 Chromatographic techniques have been used to separate (-)- Δ^9 -THC from other cannabinoid compounds. The identification of cannabis products in drug samples has been achieved using Supercritical Fluid Chromatography. Such methods are described by Bäckström et al (Science & Justice, 1997, 37(2), 91-97), Cole ("Analysis of Cannabis by Supercritical Fluid Chromatography with Ultraviolet Detection", pages 145-148 in "Supercritical Fluid Methods and Protocols" ed. by Williams and Clifford), Veress
25 (Journal of Chromatography A, 668 (1994), 285-291) and Later et al (Journal of Chromatographic Science, 1986, 24, 249-253). In these methods, very small samples (typically μg amounts) are analysed and the (-)- Δ^9 -THC is often destroyed during the detection step (e.g. by flame ionisation detection or by chemical ionisation mass spectrometry). These chromatographic methods achieve separation of (-)- Δ^9 -THC from
30 other cannabinoid compounds, but are completely unsuitable for preparing sufficient quantities of enantiomerically pure (-)- Δ^9 -THC for incorporation into pharmaceutical products.

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Levin et al (Journal of Chromatography A, 654 (1993), 53-64) have developed an analytical procedure for separating enantiomeric mixtures of cannabinoid compounds. The chromatographic method uses a Daicel Chiralpak ® AD column, which is based on amylose tris(3,5-dimethylcarbamate) supported on macroporous silica gel. The mobile
5 phase is n-hexane with ethanol or propanol. The enantioselective analysis determines the optical purity of samples but does not provide useful quantities of separated enantiomers.

Although chromatographic procedures have been used to analyse samples of cannabinoid compounds, an effective preparative separation of enantiomerically pure (-)-
10 Δ^9 -THC has not been demonstrated. The present inventors have devised a chromatographic process that can be used to prepare quantities of enantiomerically pure (-)- Δ^9 -THC for incorporation into pharmaceutical products.

Accordingly, the present invention provides a preparative separation process
15 wherein (-)- Δ^9 -trans-tetrahydrocannabinol is separated from a mixture of cannabinoids, wherein the process comprises at least one chromatographic step wherein a mobile phase passes through a stationary phase, characterised in that the stationary phase comprises a derivatised polysaccharide and the mobile phase comprises carbon dioxide.

The inventors have found that a chromatographic process combining a
20 derivatised polysaccharide stationary phase and a carbon dioxide-containing mobile phase provides an effective preparative separation of (-)- Δ^9 -THC. By "preparative separation process" we mean a process that is capable of providing at least 0.1g of purified product, preferably at least 1g of purified product in a reasonable timeframe,
25 i.e. less than a day.

Preferably the mobile phase in the present invention is a mixture of carbon dioxide and one or more modifiers. The modifier can be any liquid solvent such as an

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alcohol, ethyl acetate, acetonitrile or methylene chloride. The modifier should be compatible with the stationary phase, e.g. ethyl acetate and methylene chloride cannot be used with a Chiralpak AD column as they will destroy the column. The modifier is suitably a C₁-C₅ alcohol, most preferably ethanol. A carbon dioxide and ethanol mobile phase has been found to be particularly advantageous. When (-)- Δ^9 -THC is prepared according to the synthetic route outlined in WO 02/096899, one of the impurities is "DPA-iso" (see figure 1 for the chemical structure). When the mobile phase is carbon dioxide/ ethanol, the DPA-iso elutes before the (-)- Δ^9 -THC. A minor impurity eluting in front of a major component usually focuses due to the effects of displacement chromatography, so it was possible to remove all of the DPA-iso. Using an alternative heptane/ethanol mobile phase, the DPA-iso eluted after the (-)- Δ^9 -THC. It is considerably more difficult to resolve minor components eluting in the tail of a major component, so the carbon dioxide/ethanol mobile phase provides a significantly improved process compared to heptane/ethanol.

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Carbon dioxide is easily removed, so the (-)- Δ^9 -THC product can be provided as a solution with the modifier as a solvent. It may therefore be desirable to choose a modifier in which the (-)- Δ^9 -THC is stable.

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The ratio of carbon dioxide to modifier, as a weight (g) to volume (cm³) is suitably in the range 100:1 to 50:50, preferably in the range 95:5 to 75:25, most preferably in the range 85:15 and 75:25. The ratio of carbon dioxide to modifier can be varied during the chromatographic process.

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The stationary phase comprises a derivatised polysaccharide and is a solid chiral stationary phase. The derivatised polysaccharide is suitably immobilised on a substrate such as silica gel, zirconium, alumina, ceramics or other silicas, and is preferably immobilised on silica gel. Examples of derivatised polysaccharides include amylosic, cellulosic, chitosan, xylan, curdlan, dextran and inulan classes of polysaccharides.

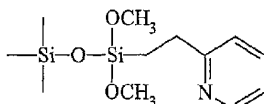
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The amylosic polysaccharides are preferred. A particularly preferred stationary phase is amylose tris(3,5-dimethylphenylcarbamate) supported on macroporous silica gel, which is available as Chiralpak ® AD, manufactured by Daicel Chemical Co. Another

preferred stationary phase is Chiralpak ® IA, which is similar to Chiralpak ® AD but has an immobilised chiral selector so that a wider variety of solvents can be used.

The stationary phase is preferably an encapsulated derivatised polysaccharide; the polysaccharide groups are not bonded to a substrate. It is thought that the encapsulated stationary phase may prevent decomposition of the (-)- Δ^9 -THC to (-)- Δ^8 -THC.

In a preferred embodiment of the invention the process comprises a further chromatographic step wherein a mobile phase passes through a stationary phase, wherein the stationary phase is an achiral stationary phase and is suitably selected from silica gel and derivatised silica gels, wherein the silica is derivatised with aminopropylsiloxane, diol-substituted propylsiloxane or 2-ethylpyridine siloxane groups. The 2-ethylpyridine siloxane immobilised on a silica support (shown below) is a preferred achiral stationary phase because the (-)- Δ^9 -THC does not degrade to form (-)- Δ^8 -THC, as was observed with some achiral stationary phases.



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The further chromatographic step ensures removal of the impurity (-)- Δ^9 -abn-THC (see figure 1 for the chemical structure) from a mixture of cannabinoids.

In the further chromatographic step, the mobile phase suitably comprises carbon dioxide and is preferably a mixture of carbon dioxide and one or more modifiers. The modifier can be any liquid solvent but is suitably a C₁-C₃ alcohol, most preferably ethanol. The ratio of carbon dioxide to modifier, as a weight (g) to volume (cm³) is suitably in the range 100:1 to 50:50, preferably in the range 100:1 to 75:25, most preferably in the range 95:5 and 90:10. The ratio of carbon dioxide to modifier can be varied during the chromatographic process.

Suitably a first chromatographic step uses the achiral stationary phase, preferably 2-ethylpyridine siloxane immobilised on a silica support and a second chromatographic step uses the derivatised polysaccharide stationary phase, and most preferably amylose

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tris(3,5-dimethylphenylcarbamate) supported on macroporous silica gel. It is preferred to use an amylosic stationary phase after a 2-ethylpyridine siloxane phase because it has been found that the amylosic phase can be destroyed by solvent impurities that might be present in the crude cannabinoid feed, whereas the 2-ethylpyridine siloxane is more robust. However, pure (-)- Δ^9 -THC can also be achieved by reversing the two steps, i.e. using the 2-ethylpyridine siloxane phase after the amylosic phase.

Suitable chromatographic apparatus is well known to the skilled person. It is preferred to use apparatus that is suitable for Supercritical Fluid chromatography such as the Novasep Supersep 10 SFC or the Novasep Supersep 100 SFC. The crude feed containing the mixture of cannabinoids is periodically injected into the apparatus wherein the mobile phase flows through the stationary phase which is located in a column. After detection at the column outlet, the purified fractions of the feed are directed to different traps. The carbon dioxide is removed from the purified fractions and is preferably recycled. Detection at the column outlet can be conducted by measuring UV absorption at an appropriate wavelength.

The column diameter is suitably from 0.5cm to 50cm and the column length is suitably from 5cm to 50cm. The particle size of the stationary phase is typically from 5 to 50 μ m.

The process is suitably carried out at temperatures from 5 to 45°C and at elevated pressures, e.g. from 80bar to 300bar. Typical flow rates depend upon the diameter of the column, and may vary from, e.g. 10g to 4kg/min.

In a further aspect the present invention provides a process for preparing a pharmaceutical product comprising (-)- Δ^9 -THC, wherein the process comprises a first step wherein (-)- Δ^9 -THC is separated from a mixture of cannabinoids by a preparative separation process according to the invention, and a further step wherein the (-)- Δ^9 -THC is combined with pharmaceutical carriers to form the pharmaceutical product. Suitable pharmaceutical carriers are known to the skilled person.

The following examples are illustrative but not limiting of the invention.

Crude feed

(-)- Δ^9 -trans-tetrahydrocannabinol was prepared as described in WO 02//096899. The crude reaction product included a variety of cannabinoid impurities, which are shown with (-)- Δ^9 -trans-THC in figure 1. The crude reaction product was dissolved in ethanol to provide the crude feed.

EXAMPLE 1:**Two step purification using a 2-ethylpyridine siloxane column and a Chiralpak AD column****Chromatographic apparatus**

A Novasep Supersep 10 SFC was used in both chromatographic steps. Two stationary phases were used: a 2-ethylpyridine siloxane stationary phase, manufactured by Princeton Chromatography Inc, with a particle diameter of 10 μ m, and a Chiralpak AD stationary phase (amylose tris(3,5-dimethylphenylcarbamate) supported on macroporous silica gel), manufactured by Daicel Chemical Co., with a particle diameter of 20 μ m. The chromatographic steps were carried out at 25°C and at a pressure of 100bar.

Step 1: Chromatographic separation using a 2-ethylpyridine siloxane column

The crude feed was filtered through 0.2 micrometer filter (Whatman PTFE w/GMF) and injected onto a chromatography column (length 25cm, inner diameter 2.1cm) containing 2-ethylpyridine siloxane stationary phase at a column flow rate of 40g/min using a mobile phase of 92% carbon dioxide and 8% ethanol. Column injections on the column were 0.85ml of crude feed injected for 5 seconds. After 128 column injections over 48 hours, an ethanolic solution was recovered with a (-)- Δ^9 -trans-THC concentration of about 25g/l. After removal of ethanol by rotary evaporation at 30°C under vacuum, about 22.5g of semi-purified (-)- Δ^9 -trans-THC was recovered exhibiting greater than 95% purity.

Step 2: Chromatographic separation using a Chiralpak AD column

The semi-purified (-)- Δ^9 -trans-THC was re-dissolved in absolute ethanol to a concentration of about 300g/l to produce a feed for injection onto Chiralpak AD column

(length 25cm, inner diameter 2.1cm) at a column flow rate of 40g/min using a mobile phase of 80% carbon dioxide and 20% ethanol. After 60 column injections of 0.85ml (over 5 seconds) of the semi-purified feed, an ethanolic solution was recovered (about 2.2 liters) and stored in a freezer for solvent evaporation at a later date. The estimated recovery of purified (-)- Δ^9 -trans-THC (>99.5% purity) was ~15 grams.

EXAMPLE 2:

**Two step purification using a Chiralpak AD column
and a 2-ethylpyridine siloxane column**

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Chromatographic apparatus

The apparatus was the same as the apparatus used in Example 1.

Step 1: Chromatographic separation using a Chiralpak AD column

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The crude feed was filtered through 0.2 micrometer filter (Whatman PTFE w/GMF) and injected onto a chromatography column (length 25cm, inner diameter 2.1cm) containing Chiralpak AD stationary phase at a column flow rate of 40g/min using a mobile phase of 80% carbon dioxide and 20% ethanol. After 35 column injections of 0.85ml, 6g of semi-purified (-)- Δ^9 -trans-THC (97.4% purity) were collected.

20

The majority of the remaining impurity was found to be (-)- Δ^9 -abn-THC (2.5% AUC).

Step 2: Chromatographic separation using a 2-ethylpyridine siloxane column

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The semi-purified (-)- Δ^9 -trans-THC was injected onto Chiralpak AD column (length 25cm, inner diameter 1cm) at a column flow rate of 20g/min using a mobile phase of 92% carbon dioxide and 8% ethanol. After 3 second column injections at 5ml/min of the semi-purified feed, (-)- Δ^9 -abn-THC impurity was reduced to less than 0.05% AUC.

COMPARATIVE EXAMPLE 1:
One step purification using a Chiralpak AD column
and a hexane/ethanol mobile phase

5 The chromatographic apparatus was the same as the apparatus used in Example 1.
The crude feed was filtered through 0.2 micrometer filter (Whatman PTFE w/GMF) and
injected onto a chromatography column (length 25cm, inner diameter 1cm) containing
Chiralpak AD stationary phase at a column flow rate of 4.8ml/min using a mobile phase
of 95% hexane and 5% ethanol. Preparative purification of (-)- Δ^9 -trans-THC from the
10 DPA-iso impurity was not achieved.

EXAMPLE 3:
Scaled-up two step purification using a 2-ethylpyridine siloxane column
and a Chiralpak AD column

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Chromatographic apparatus

A Novasep Supersep 100 SFC was used in both chromatographic steps.
Two stationary phases were used: a 2-ethylpyridine siloxane stationary phase,
manufactured by Princeton Chromatography Inc, with a particle diameter of 10 μ m, and a
20 Chiralpak AD stationary phase (amylose tris(3,5-dimethylphenylcarbamate) supported
on macroporous silica gel), manufactured by Daicel Chemical Co., with a particle
diameter of 20 μ m.

Step 1: Chromatographic separation using a 2-ethylpyridine siloxane column

25 The Novasep Supersep 100 SFC was fitted with a 100mm inner diameter
dynamic axial compression (DAC) column packed to a bed length of 250mm length with
2-ethylpyridine bonded silica. A mixture of liquid carbon dioxide (Airgas, Instrument
Grade) and Absolute Ethanol (Warner Graham, USP Grade) in a ratio of about
96:4 wt/wt were used as the mobile phase. The operating conditions were:

30 Column Temperature: 30°C
Column Pressure: 125 Bar
Liquid CO₂ flow rate: 1770 g/min
Ethanol flow rate: 80 g/min

Detection: UV 245nm

The product was concentrated in ethanol eluent by evaporation resulting in an amber solution. (-)- Δ^9 -trans-THC was isolated with a purity >96%.

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5 **Step 2: Chromatographic separation using a Chiralpak AD column**

The Novasep Supersep 100 SFC was fitted with a 100mm inner diameter dynamic axial compression (DAC) column packed to a bed length of 250mm length with Chiralpak AD. A mixture of liquid Carbon Dioxide (Airgas, Instrument Grade) and Absolute Ethanol (Warner Graham, USP Grade) in a ratio of about 86:14 wt/wt were used as the mobile phase. The SFC operating conditions were:

10 Column Temperature: 25°C

Column Pressure: 125 Bar

Liquid CO₂ flow rate: 858 g/min

Ethanol flow rate: 142 g/min

15 Detection: UV 245nm

The product was concentrated in ethanol eluent by evaporation resulting in a colourless solution. (-)- Δ^9 -trans-THC was isolated with a purity >99%.

20 In the claims which follow and in the preceding description of the invention, except where the context requires otherwise due to express language or necessary implication, the word "comprise" or variations such as "comprises" or "comprising" is used in an inclusive sense, i.e. to specify the presence of the stated features but not to preclude the presence or addition of further features in various embodiments of the invention.

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It is to be understood that, if any prior art publication is referred to herein, such reference does not constitute an admission that the publication forms a part of the common general knowledge in the art, in Australia or any other country.

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CLAIMS

1. A preparative separation process wherein (-)- Δ^9 -trans-tetrahydrocannabinol is separated from a mixture of cannabinoids, wherein the process comprises at least one
5 chromatographic step wherein a mobile phase passes through a stationary phase, characterised in that the stationary phase comprises a derivatised polysaccharide and the mobile phase comprises carbon dioxide.
2. A process according to claim 1, wherein the mobile phase is a mixture of carbon
10 dioxide and one or more modifiers.
3. A process according to claim 2, wherein the mobile phase is a mixture of carbon dioxide and ethanol.
- 15 4. A process according to claim 2 or claim 3, wherein the ratio of carbon dioxide to liquid modifier is in the range 95:5 to 75:25.
5. A process according to any preceding claim, wherein the derivatised polysaccharide is immobilised on a substrate chosen from silica gel, zirconium, alumina,
20 ceramics and other silicas.
6. A process according to any preceding claim, wherein the stationary phase comprises an amylosic polysaccharide.
- 25 7. A process according to claim 6, wherein the stationary phase is amylose tris(3,5-dimethylphenylcarbamate) supported on macroporous silica gel.
8. A process according to any preceding claim, wherein the process comprises a further chromatographic step wherein a mobile phase passes through a stationary phase,
30 wherein the stationary phase is an achiral stationary phase.
9. A process according to claim 8, wherein the achiral stationary phase is 2-ethylpyridine siloxane immobilised on a silica support.

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10. A process according to claim 8 or claim 9, wherein a first chromatographic step uses the achiral stationary phase and a second chromatographic step uses the stationary phase comprising a derivatised polysaccharide.
- 5 11. A process according to claim 8 or claim 9, wherein a first chromatographic step uses the stationary phase comprising a derivatised polysaccharide and a second chromatographic step uses the achiral stationary phase.
- 10 12. A process for preparing a pharmaceutical product comprising (-)- Δ^9 -THC, wherein the process comprises a first step wherein (-)- Δ^9 -THC is separated from a mixture of cannabinoids by a process according to any preceding claim, and a further step wherein the (-)- Δ^9 -THC is combined with pharmaceutical carriers to form the pharmaceutical product.
- 15 13. A preparative separation process substantially as herein described with reference to the examples.

Figure 1

