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(71) Demandeur/Applicant:
ZEUS SCIENTIFIC, INC., US

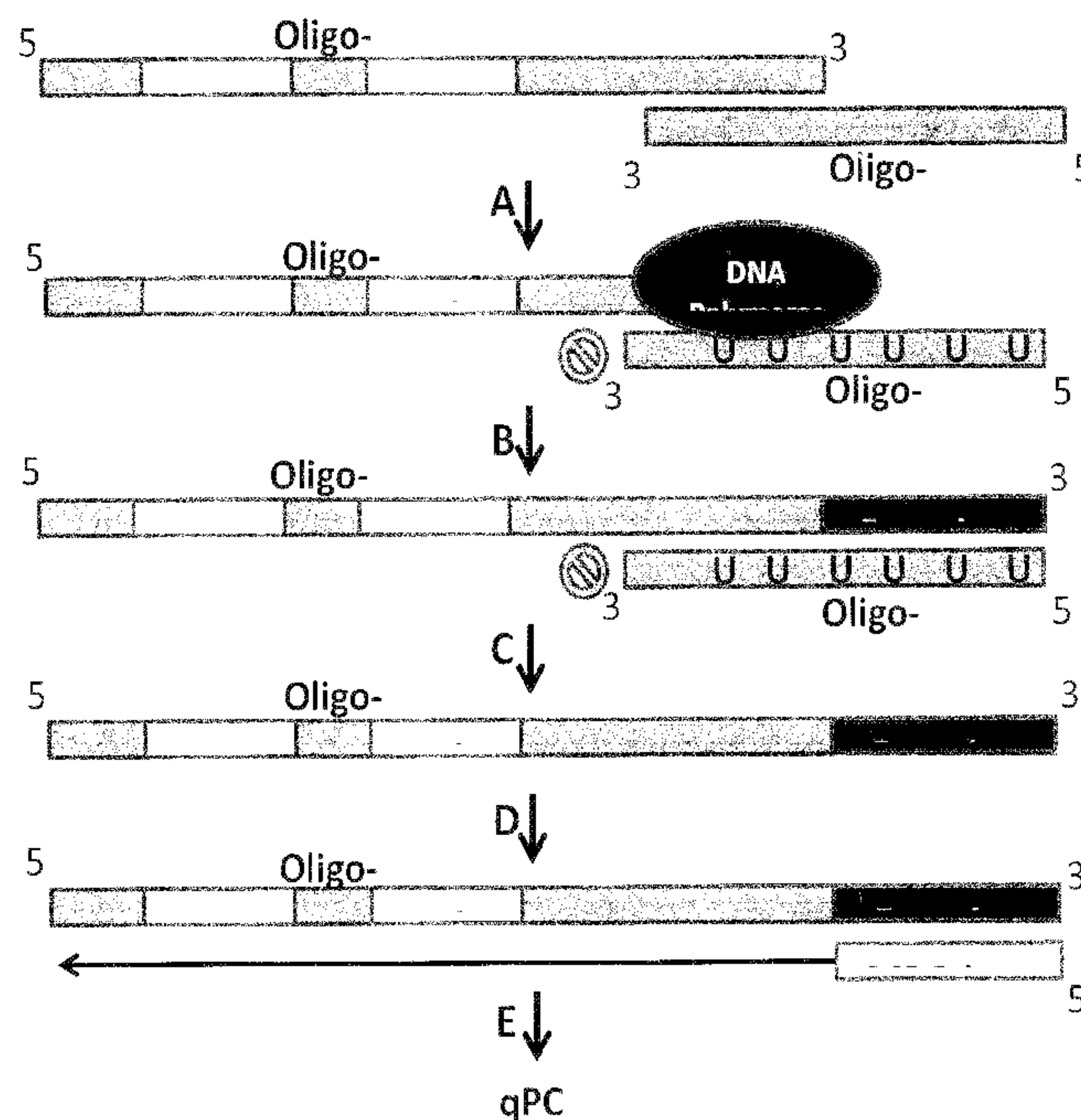
(72) Inventeurs/Inventors:
O'HARA, SHAWN MARK, US;
SWEITZIG, DANIEL, US

(74) Agent: THERIAULT, MARIO D., P. ENG.

(54) Titre : TESTS D'ACTIVITE AMELIOREE D'ADN POLYMERASE ET PROCEDES PERMETTANT LA DETECTION DE MICROBES VIABLES

(54) Title: IMPROVED DNA POLYMERASE ACTIVITY ASSAYS AND METHODS ENABLING DETECTION OF VIABLE MICROBES

Figure 1



(57) Abrégé/Abstract:

A method for performing a diagnostic assay for the detection of the presence or amount of a microorganism within a sample matrix containing active DNA polymerase, is disclosed. The method utilizes the measurement of DNA polymerase extension activity, wherein the assay comprises the steps of incubating DNA polymerase in the sample matrix with a selected suitable substrate, and performing PCR cycling and detection via the use of a selected suitable nucleic acid probe, thereby to detect endogenous DNA polymerase extension activity in the sample matrix as an indication of the presence or amount of said microorganism.



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(71) Applicant: ZEUS SCIENTIFIC, INC. [US/US]; P.O.Box 38, Raritan, NJ 08869-0038 (US).

(72) Inventors: O'HARA, Shawn, Mark; P.O.Box 38, Raritan, NJ 08869-0038 (US). SWEITZIG, Daniel; P.O.Box 38, Raritan, NJ 08869-0038 (US).

(74) Agent: WILCOX, James; 1767 Route 313, Perkasi, PA 18944 (US).

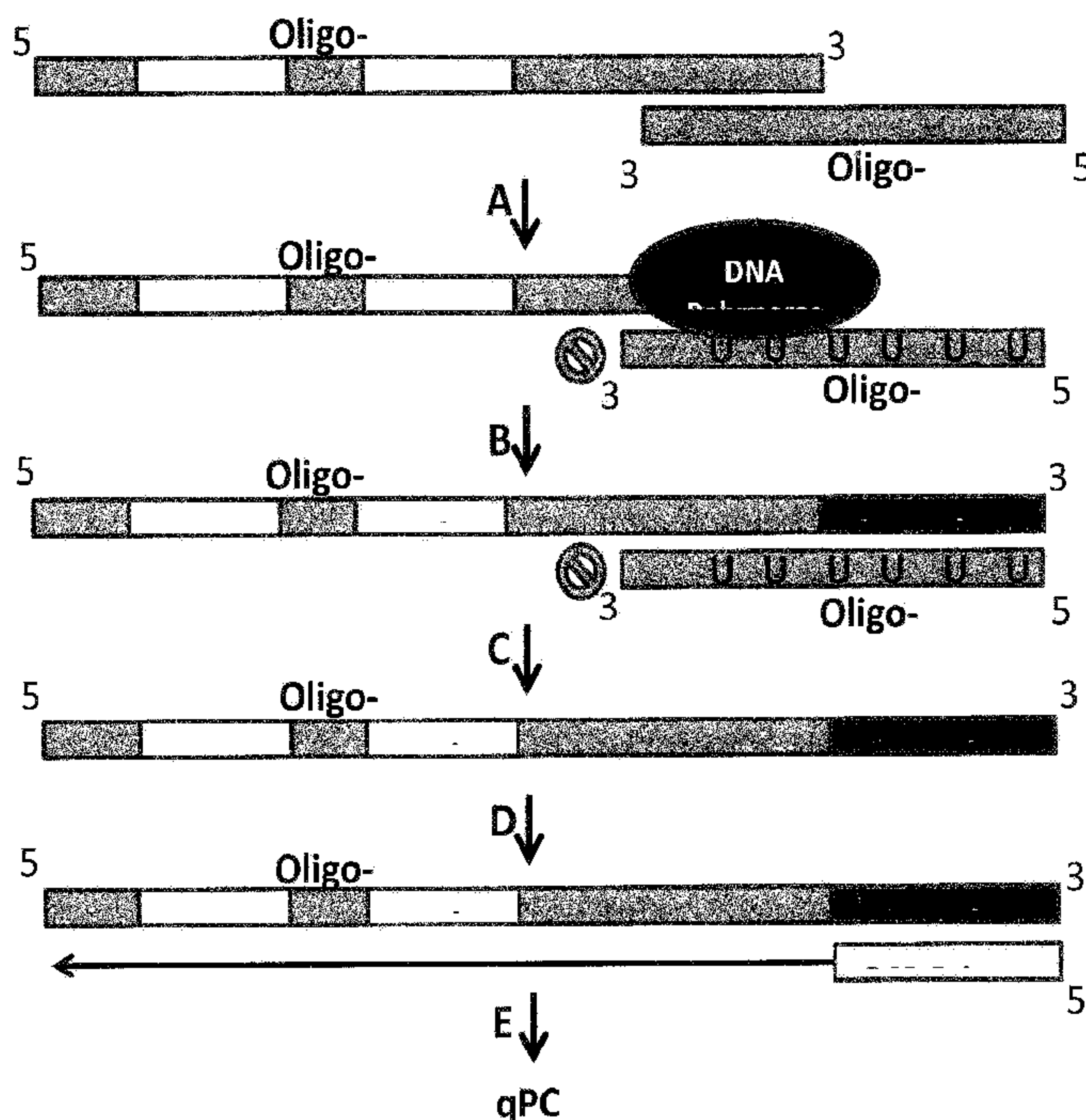
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(54) Title: IMPROVED DNA POLYMERASE ACTIVITY ASSAYS AND METHODS ENABLING DETECTION OF VIABLE MICROBES

Figure 1



(57) Abstract: A method for performing a diagnostic assay for the detection of the presence or amount of a microorganism within a sample matrix containing active DNA polymerase, is disclosed. The method utilizes the measurement of DNA polymerase extension activity, wherein the assay comprises the steps of incubating DNA polymerase in the sample matrix with a selected suitable substrate, and performing PCR cycling and detection via the use of a selected suitable nucleic acid probe, thereby to detect endogenous DNA polymerase extension activity in the sample matrix as an indication of the presence or amount of said microorganism.

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5 **Improved DNA Polymerase Activity Assays and Methods Enabling Detection of Viable
Microbes**

CROSS REFERENCE TO RELATED APPLICATION

10 This application is a non-provisional application, which incorporates by reference herein
and claims priority of US Provisional Application No. 61/583,568, filed January 5, 2012.

BACKGROUND OF THE INVENTION

The reference numbers used in this section and throughout this disclosure refer to the documents set forth in the “References” section herein.

15 DNA polymerase activity is indispensable for genome replication and organism
propagation across all biological domains (1-3). Since its initial characterization (4), the ability
to harness DNA polymerase activity *in vitro* has become a fundamental tool in the field of
molecular biology research (5). Above and beyond its established importance in research, *in*
vitro measurement of DNA polymerase activity potentially offers numerous useful applications
within the pharmaceutical and clinical setting. For instance, since bacterial DNA polymerase is
20 actively being targeted for development of novel antimicrobial agents (6, 7), a rapid and sensitive
assay capable of measuring DNA polymerase activity is desirable. Also, loss or gain of DNA
polymerase activity is intimately involved in human disease. For example, emerging links
between DNA polymerase activity and genetic aberrations are designating the enzyme as a target
for anti-cancer therapies (8, 9). Deficiencies in DNA polymerase activity have also been linked
25 to mitochondrial disorders (10). Furthermore, measurement of DNA polymerase activity has the
potential to be used as a rapid and sensitive diagnostic tool, capable of detecting virtually any
organism harboring active DNA polymerase within a given environmental or biological matrix
where sterility is expected.

30 The most common method used to measure DNA polymerase activity *in vitro* depends
upon incorporation of radiolabeled nucleotides (11). However, routine use of such DNA
polymerase assays is undesirable due to the inherent risks and restrictions associated with
radioisotopes. Consequently, over the past few decades numerous non-radioactive *in vitro*

5 polymerase assays have been developed. Some rely upon the measurement of fluorescence generated by DNA polymerase-mediated release of single stranded binding protein (12) or binding of PicoGreenTM to double stranded DNA (13,14). Other methods rely on microplate coupling and detection of fluorescently-labeled nucleotides (15). More recently, molecular beacon-based (16) and electrochemical-based (17) DNA polymerase assays have been
10 developed. Despite successfully averting the use of radioactivity, the above assays are limited by such factors as poor sensitivity, a small linear dynamic range of measurement, or the use of purified polymerase.

As will be apparent to those skilled in the relevant art, the measurement of DNA polymerase extension activity in accordance with the present invention as described herein
15 represents a useful tool with far reaching applications such as, but not limited to, screening candidate-polymerase inhibitors *in vitro*, or detecting the presence any microbe (harboring active DNA polymerases) within a diverse range of sample types. This is a substantial improvement over the state of the present art, because if intended for these purposes, routine use of traditional polymerase assays that incorporate radiolabeled nucleotides is unattractive. Consequently,
20 numerous non-radioactive DNA polymerase extension assays have been developed in recent decades. Despite successfully averting the use of radioactivity, current fluorescence-based DNA polymerase assays also suffer from various deficiencies. For example, detection of DNA polymerase activity via several existing non-radioactive assays is dependent upon the binding of PicoGreenTM to newly-generated double stranded DNA (13,14). If intended to analyze DNA
25 polymerase activity from freshly lysed organisms, PicoGreenTM-based assays would likely be hampered by background fluorescence via binding of PicoGreenTM to genomic DNA. Microplate-based DNA polymerase assays have also been developed (15). Decreased sensitivity of microplate-based assays can be expected for numerous reasons, including dependence upon intermediate binding of either product or substrate to a microplate and/or
30 inefficient incorporation of modified dNTPs by DNA polymerase. More recently, real-time measurement of DNA polymerase activity via molecular beacons has been described (16). Despite improved sensitivity, direct measurement of molecular beacon fluorescence could also potentially be hindered by exposure to crude cellular lysates.

5 SUMMARY OF THE INVENTION

The present invention improves upon the technology of the background art as described above, and provides a rapid, highly sensitive and quantitative assay, capable of measuring DNA polymerase extension activity from purified enzymes or directly from microbial lysates, including crude microbial lysates. The invention as described herein provides a significant and
10 unexpected advancement toward sensitive detection of potentially any microorganism containing active DNA polymerase within a given sample matrix. The present invention involves methodology for enzymatic template generation and amplification (ETGA). Accordingly herein is described the first characterization of a novel ETGA methodology based upon the measurement of DNA polymerase extension activity coupled to a quantitative PCR readout. For
15 the remainder of the disclosure herein, this type of diagnostic assay provided by the invention is referred to as DPE-PCR. The DPE-PCR assay of this invention can be used to measure low levels of purified enzyme and is capable of detecting endogenous DNA polymerase extension activity directly from microbial cell lysates.

BRIEF DESCRIPTION OF THE DRAWING FIGURES

20 Figure 1 is a Schematic overview of a preferred DPE-PCR diagnostic assay in accordance with the present invention, and wherein: **(Step A)** DNA polymerase is incubated with a substrate consisting of pre-annealed Oligo-1 and Oligo-2. **(Step B)** During a 20 minute incubation at 37 ° C, DNA polymerase extends only the 3' end of Oligo-1. **(Step C)** 3 µL of the reaction mixture is subsequently placed into a hot start qPCR reaction containing uracil DNA glycosylase (UDG).
25 Prior to activation of Taq, UDG degrades the deoxyuridine within Oligo-2, leaving only a single stranded product derived from polymerase-mediated extension of Oligo-1. **(Step D)** After activation of Taq, amplification is initiated via primer binding to the Oligo-1 extension product. **(Step E)** PCR cycling and detection via Taqman probe .

Figure 2 is a schematic representation of the sensitive detection of purified DNA
30 polymerase using DPE-PCR in accordance with a preferred embodiment of the present invention, and wherein: **(A)** A commercial source of DNA polymerase I was assayed in duplicate at 10 fold increments starting at 2×10^{-5} Units (U) down to 2×10^{-11} U per reaction. A representative DPE-PCR curve is shown for each polymerase input level and No Input Control

5 (NIC). **(B)** A plot was constructed from $n = 4$ data points per polymerase input level, taken from two independent experiments and linear regression analysis was performed **(C)** Triplicate reactions containing 2×10^{-7} U of DNA polymerase I, Klenow, Klenow (exo-) and *E. coli* DNA Ligase were assayed in comparison to a NIC. A representative DPE-PCR curve is presented for each of the assayed enzymes and NIC **(D)** DPE-PCR signal was compared in reactions
 10 containing a dNTP mixture with either dCTP or ddCTP, a schematic representing the available sites for dCTP or ddCTP incorporation within the DNA substrate is presented adjacent to the DPE-PCR curves.

Figure 3 is a schematic overview of coupling bead lysis to DPE-PCR in accordance with a preferred embodiment of the present invention.

15 Figure 4 is a graphical representation of how the performance of DPE-PCR in accordance with the present invention enables sensitive and quantitative detection of gram negative and gram positive bacteria via measurement of DNA polymerase extension activity in crude lysates, and wherein: **(A)** Decreasing amounts of *E. coli cfu* were spiked into bead lysis-coupled DPE-PCR. No Input Controls (NIC) were also included to monitor reagent background levels. All *cfu*
 20 spikes and NICs were performed in triplicate. A representative DPE-PCR curve is shown below for each level of bacterial input. Colony count plating and gsPCR were performed in an effort to obtain a better estimate of the actual *cfu* placed into each reaction and is presented in Supplemental Figure 3 **(B)** A plot of *E. coli* DNA polymerase activity and linear regression analysis is presented. Graphs were generated using the average Ct values obtained from
 25 triplicate reactions of bacterial spikes ranging from 1×10^5 - 1×10^1 input *cfu*. **(C and D)** *cfu* titration experiments were performed for *S. aureus* exactly as described above for *E. coli*. Colony count plating and gsPCR were performed in an effort to obtain a better estimate of the actual *cfu* placed into each reaction.

Figure 5 shows a graphical representation of the detection of bacteria by DPE-PCR in
 30 accordance with another preferred embodiment of the present invention, and wherein: **(A)** 5 μ L of *E. coli* suspension were added to bead lysis-coupled DNA polymerase assays comprised of a dNTP mix containing either 50 μ M dCTP or 50 μ M ddCTP. DPE-PCR curves representing *E. coli*-derived DNA polymerase activity is presented. Approximate *cfu* input as determined by plating is presented in the upper left region of the qPCR graph **(B)** 5 μ L of *E. coli* suspension

5 were added to bead lysis tubes containing 50 μ L reaction buffer comprised of a dNTP mix with either 50 μ M dCTP or 50 μ M ddCTP. Prior to lysis, 1 μ L of dCTP [2.5 mM, 0.25 mM 0.025 mM 0.0025 mM] was added to selected ddCTP-containing reactions. Reactions containing dCTP alone or ddCTP alone were run in parallel as “non-terminated” and “terminated” comparators. The resultant DPE-PCR curves representing *E. coli*-derived DNA polymerase activity is presented. Approximate *cfu* input as determined by plating is presented in the lower left region of the qPCR graph (C) *E. coli* gene specific PCR was also performed on the same lysates used for DNA polymerase detection presented in Figure 2B. Linear plots of dCTP-dependent rescue of bacterial DNA polymerase detection vs. gsPCR of genomic DNA are shown. Plots were generated using the average qPCR Ct values from triplicate reactions at the indicated conditions (D-F) ddCTP termination and dCTP rescue experiments were performed for *S. aureus* exactly as described above for *E. coli*.

Figure 6 is a graphical illustration of another embodiment of the present invention in which DPE-PCR is an indicator of *E. coli* viability in response to heat treatment, and wherein: (A) 200 μ L aliquots of an *E. coli* suspension (~ 2000 *cfu*/ μ L) were incubated at 25° C, 45° C, 65° C, 85° C and 105° C for 20 minutes. After heating, each bacterial stock was cooled to room temperature and 5 μ L were transferred to the bead lysis-coupled DPE-PCR assay. DPE-PCR curves representing *E. coli*-derived DNA polymerase activity following each of the indicated temperature treatments are presented. (B) Plots were generated from triplicate DPE-PCR reactions and gsPCR of genomic DNA (from the same lysates) after the indicated temperature treatments of *E. coli* suspensions. Parallel plating was also performed in triplicate for each of the treated *E. coli* suspensions. Representative *cfu* monitoring plates are presented below the graph, revealing bacterial viability status after treatment at each temperature. (C) DPE-PCR is compared to gsPCR of genomic DNA in response to the various temperature treatments. “Fold Reduction of qPCR Signal” was calculated using the indicated equation and the values obtained were used to generate comparative bar graphs.

Figure 7 is a graphical illustration of another embodiment of the present invention, in which DPE-PCR is an indicator of *S. aureus* viability in response to heat treatment, and wherein: (A) 200 μ L aliquots of an *S. aureus* suspension (~ 2000 *cfu*/ μ L) were incubated at 25° C, 45° C, 65° C, 85° C and 105° C for 20 minutes. After heating, each bacterial stock was cooled to room

5 temperature and 5 μ L were transferred to the bead lysis-coupled DPE-PCR assay. DPE-PCR curves representing *S.aureus*-derived DNA polymerase activity following each of the indicated temperature treatments are presented. **(B)** Plots were generated from triplicate DPE-PCR reactions and gsPCR of genomic DNA (from the same lysates) after the indicated temperature treatments of *S. aureus* suspensions. Parallel plating was also performed in triplicate for each of
10 the treated *S. aureus* suspensions. Representative *cfu* monitoring plates are presented below the graph, revealing bacterial viability status after treatment at each temperature. **(C)** DPE-PCR is compared to gsPCR of genomic DNA in response to the various temperature treatments. “Fold Reduction of qPCR Signal” was calculated using the indicated equation and the values obtained were used to generate comparative bar graphs.

15 Figure 8 sets forth Table 1, as referred to herein, in which results are set forth showing the sensitive and linear detection of 17 additional clinically relevant microbial species in accordance with the teachings of the present invention.

DETAILED DESCRIPTION OF THE INVENTION

During the past fifty years, *in vitro* measurement of DNA polymerase activity has
20 become an essential molecular biology tool. Traditional methods used to measure DNA polymerase activity *in vitro* are undesirable due to the usage of radionucleotides. Fluorescence-based DNA polymerase assays have been developed; however, they also suffer from various limitations. Herein is disclosed a rapid, highly sensitive and quantitative assay capable of measuring DNA polymerase extension activity from purified enzymes or directly from microbial
25 lysates. When tested with purified DNA polymerase, the assay has been found to detect as little as 2×10^{-11} U of enzyme (≈ 50 molecules), while demonstrating excellent linearity ($R^2 = 0.992$). The assay was also able to detect endogenous DNA polymerase extension activity down to at least 10 colony forming units of input gram-positive or gram-negative bacteria when coupled to bead mill lysis while maintaining an $R^2 = 0.999$. Furthermore, experimental evidence presented
30 herein suggests that DNA polymerase extension activity is an indicator of bacterial viability, as demonstrated by the reproducibly strong concordance between assay signal and bacterial colony formation. Together, the novel methodology of the invention described herein represents a significant advancement toward sensitive detection of potentially any microorganism containing active DNA polymerase within a given sample matrix.

5 To further illustrate the foregoing concepts and advantages of the invention, the following examples are provided as illustrative of this invention, but are in no way to be construed as limitative thereof.

Example

MATERIALS AND METHODS:

10 DNA substrate preparation

 The sequence of the DNA substrate (and qPCR primers presented below) was adapted from DNA oligos previously used to measure bacterial-derived ATP via T4 DNA ligase (18). Briefly, Oligo 1 and Oligo 2 (see Figure 1) were pre-annealed and diluted to a working concentration of 0.01 μ M.

15 DNA polymerase activity reaction using commercial polymerase

 DNA Pol I (NEB cat# M0209L), Klenow (NEB cat# M0210S) and Klenow exo(-) (NEB cat# M0212S) were diluted to the indicated U/ μ L stock in Tris EDTA (T.E.) pH 8.0. To begin, 2 μ L of DNA polymerase stock at each concentration were placed into a 50 μ L polymerase assay mixture containing the following components: 50 μ M dNTP, 20 mM Tris pH 8.0, 10 mM
20 ammonium sulfate, 10 mM potassium chloride, 2 mM magnesium sulfate, 1% BSA, 0.1% Triton, 0.1% Tween, and 0.001 μ M pre-annealed DNA substrate. Reactions were vortexed briefly and placed at 37° C for 20 minutes. After 20 minutes, 3 μ L of each reaction were immediately placed into a quantitative PCR (qPCR) reaction.

Detection by qPCR

25 The qPCR reaction master mix was prepared using the following components: LightCycler 480 Master Mix (Roche cat# 04707494001), 333 nM of each primer, 166 nM Target probe (FAM), 166 nM internal control probe (TxRed), and 1.2 U of UDG (Bioline cat# BIO-20744). As a tool to monitor PCR inhibition, each qPCR reaction also included 40 copies of competitive internal control DNA. For each qPCR reaction, 3 μ L of DNA polymerase reaction
30 were added to 27 μ L of master mix and a two-step qPCR was run on a SmartCycler (Cepheid, Sunnyvale CA) as follows: Initial incubation of 40° C for 10 minutes and 50° C for 10 minutes and at 95° C for 5 minutes (to activate Taq), followed by 45 cycles of 5s denaturation at 95° C and 20s annealing/extension at 65° C. Cycle threshold (Ct) values were generated automatically by the SmartCycler software using 2nd derivative analysis of the emerging qPCR curves.

5 Bacterial strains and media

Staphylococcus aureus (ATCC 25923) and *Escherichia coli* (ATCC 25922) were used in this study. Cultures were grown in/on Brain-Heart Infusion liquid media/agar (Teknova.) The ATCC reference numbers and growth media for the additional 17 microorganisms tested are listed in Figure 5.

10 Detection of bacterial DNA polymerase activity following bead mill lysis

S. aureus and *E. coli* cultures were grown to an OD₆₀₀ of 1.0 ± 0.2 (approximately 1×10^9 cfu/mL.) For each organism, 1 mL of culture was pelleted and washed three times in T.E. Bacterial suspensions were serially diluted in T.E., and 5 μ L of each stock were added to bead lysis-reactions containing 50 μ L of lysis-reaction buffer. A titration curve of 1×10^5 to 1×10^0 cfu/reaction was performed in triplicate for each organism, including triplicate reactions without bacterial suspension (No Input Control). After the addition of 5 μ L bacterial stock (or No Input Control), lysis/reaction tubes were bead milled for 6 min. at 2800 rpm, followed by incubation at 37° C for 20min. After a 20 minute incubation, samples were heated to 95° C for 5 min. and removed to cool at room temperature. Samples were then spun at 12k x g for 30 seconds and 3 μ L of each reaction were placed into qPCR. Five micro-liters of each bacterial stock was plated to obtain more accurate cfu input levels. Organism-specific PCR was also performed on the same lysates used for DNA polymerase detection. Primer and probe sequences for *S. aureus* and *E. coli* gene specific PCR are listed in Figure 2.

Dideoxy termination experiments

25 Termination of purified DNA polymerase extension activity with ddCTP:

DNA polymerase assay reactions were prepared as described above with a dNTP mix containing either 50 μ M dCTP or 50 μ M ddCTP (Affymetrix #77332.) Reactions containing either dNTP mix were spiked with 2×10^{-9} U of DNA polymerase I (New England Biolabs # M0209). Reactions were incubated at 37° C for 20 minutes and 3 μ L of each reaction were subsequently placed into qPCR.

Elimination of microbial detection via ddCTP:

S. aureus and *E. coli* cultures were grown, washed and diluted as described above. To demonstrate ddCTP-dependent termination of microbial DNA polymerase, 5 μ L of bacterial

5 stock were added to bead lysis tubes containing 50 μ L of reaction buffer with either 50 μ M dCTP or 50 μ M ddCTP. Lysis, incubation and qPCR were performed as described above. Five micro-liters of each bacterial stock were plated to determine more accurate *cfu* input levels. Gene specific PCR of genomic DNA was also performed on the same lysates used for DNA polymerase detection.

10 dCTP rescue of microbial detection:

S. aureus and *E. coli* cultures were grown, washed and diluted as described above. Five micro-liters of bacterial stock were added to bead lysis tubes containing 50 μ L of reaction buffer with 50 μ M ddCTP. Prior to lysis, 1 μ L of dCTP at 2.5 mM, 0.25 mM 0.025 mM 0.0025 mM was added to ddCTP-containing reactions. Reactions containing 50 μ M dCTP alone and ddCTP
15 alone were run in parallel as “non-terminated” and “terminated” comparators. Lysis, incubation and qPCR were performed as described above. Five micro-liters of each bacterial stock were plated to determine more accurate *cfu* input levels. Gene-specific PCR was also performed on the same lysates used for DNA polymerase detection.

Viability Assessment Experiments

20 *S. aureus* and *E. coli* cultures were grown, washed and diluted as described above. Two hundred micro-liters of bacterial stocks at approximately 2000 *cfu*/ μ L (in T.E.) were incubated at 25° C, 45° C, 65° C, 85° C and 105° C for 20 minutes. After heating, samples were cooled to room temperature and 5 μ L of each bacterial stock were added to bead lysis tubes containing 50 μ L of reaction buffer. Lysis, incubation and qPCR were performed as described above. Five
25 micro-liters of each bacterial stock (treated at various temperatures) were also added to 1 ml of T.E. and 50 μ L were plated for colony count determination. Gene specific PCR was also performed on the same lysates used for DNA polymerase detection.

RESULTS AND DISCUSSION

In the development of the present invention, it was set out to develop a rapid, simple,
30 highly sensitive and quantitative assay capable of measuring DNA polymerase extension activity derived from purified commercial sources or freshly lysed cells, which would improve upon and overcome the disadvantages of the foregoing described methodologies of the know art. Figure 1

5 shows a schematic overview of the mechanisms involved in coupling DNA polymerase extension activity to qPCR. Notably, Oligo 2 (see Figure 1, step C) is removed by uracil DNA glycosylase (UDG) prior to Taq activation, thus preventing non-specific extension of the substrate just prior to PCR cycling. A microbial detection method linking T4 DNA ligase activity to PCR amplification has been previously reported (18), which contains similarities to
10 our DPE-PCR assay and is another example of an ETGA methodology. However, during the development of the present invention a modified version of this method, aimed at detecting NAD-dependent DNA ligase activity, suffered from various limitations (unpublished data), leading to the development of the novel DNA polymerase-based approach of the invention as described herein.

15 **Sensitive and linear detection of purified DNA polymerase extension activity**

An experiment was performed to determine the approximate analytical sensitivity of the DPE-PCR assay using commercially available DNA polymerase I. As shown in Figure 2A, detection of DNA polymerase I was achieved over a wide range of input enzyme. In fact, measurement of DNA polymerase I extension activity is achieved down to as little as 2×10^{-11}
20 units (U) of enzyme (equivalent to approximately 50 molecules of polymerase). To our knowledge, detection of DNA polymerase extension activity at this level is unrivaled in existing DNA polymerase assays. In theory, this level of sensitivity could enable single microbe detection as *E. coli* has been reported to contain approximately 400 DNA polymerase I molecules per cell (11). Regression analysis also showed a strong positive linear correlation ($R^2 = 0.992$) between the DPE-PCR cycle threshold (Ct) values and units of input commercial DNA
25 polymerase I after graphing data from two independent limit of detection experiments (Figure 2B). After sensitivity and linearity experiments were performed, it was important to determine if the DPE-PCR assay signal was independent of intrinsic exonuclease activity. To this end, we subsequently compared signals generated by 2×10^{-7} U of DNA polymerase I to those generated
30 from DNA polymerase I lacking 5' $\rightarrow$ 3' exonuclease activity (Klenow) and another version of the enzyme lacking all exonuclease activity (Klenow exo -). For additional specificity and background signal determination, *E. coli* DNA ligase at 2×10^{-7} U and a No Input Control (NIC) were tested in parallel. As shown in Figure 2C, both Klenow and Klenow exo - were detected at similar levels when compared to wild type DNA polymerase I, providing evidence that the

5 DPE-PCR assay signal is derived from DNA polymerase-dependent extension and not intrinsic exonuclease activity. In addition to using exonuclease free polymerases, we set out to further demonstrate that DPE-PCR assay signal is derived from DNA polymerase-dependent extension of the DNA substrate prior to qPCR. Since incorporation of dideoxy nucleotides is a well-established method used for termination of DNA polymerase chain extension activities (19,20),
10 we chose to substitute dCTP with dideoxyCTP (ddCTP) within our reaction mix. The schematic shown in Figure 2D reveals the first possible position within the substrate that ddCTP can be incorporated by DNA polymerase. If ddCTP is incorporated into this position, the extension product of Oligo 1 would be insufficient in length for subsequent detection by qPCR primer 1 (Figure 1). As shown in Figure 2D, substitution of dCTP with ddCTP eliminates signal
15 generated by DNA polymerase I, thus demonstrating that the DPE-PCR assay signal is dependent upon DNA polymerase extension of the substrate prior to qPCR. The presence of a low copy internal amplification control confirms that qPCR was not inhibited by the presence of low amounts of ddCTP that are carried over from the DNA polymerase assay reagents (Supplemental Figure 1C). In addition, we feel it is important to note that we have sporadically
20 observed a weak, but detectable signal in the absence of input-DNA polymerase (No Input Control). Due to the exquisite sensitivity of the DPE-PCR assay, we have demonstrated that weak background noise signals can be derived from several potential sources such as, but not limited to, DNA polymerase contamination present in the reagents prior to reaction assembly, DNA polymerase introduced by the operator during experimental setup and/or incomplete
25 degradation of Oligo 2 (Figure 1) prior to activation of Taq (unpublished data). Notably, these irregular sources of background noise are controllable by instituting stricter reagent preparation procedures and good aseptic technique.

Sensitive universal detection of microbes via measurement of endogenous DNA polymerase extension activity directly from cell lysates

30 In addition to detecting purified polymerase activity, a simple universal method that measures microbial-derived DNA polymerase activity would be highly desirable. If achieved, such a method could enable the screening of candidate antimicrobial agents in actively growing cultures, thus allowing comparison of DNA polymerase extension activity to organism proliferation. Additionally, measurement of DNA polymerase extension activity could be used

5 to screen environmental or biological samples for the presence of any microorganism harboring active DNA polymerase. To this end, we developed a simple method that couples microbial lysis to a DPE-PCR assay provided by the invention. As shown in Figure 3, a liquid sample known to contain, or suspected of containing, microbes is added to a bead mill lysis tube, disrupted and immediately transitioned into the DPE-PCR assay. We chose one gram negative
10 bacteria (*E. coli*) and one gram positive bacteria (*S. aureus*) to demonstrate the ability of our assay to measure microbial-derived DNA polymerase extension activity in crude cellular lysates. As shown in Figure 4A, when linked with bead mill lysis, the DPE-PCR assay is capable of detecting a wide dynamic range of input *E. coli*, down to and below 10 colony forming units (*cfu*) per lysis tube. Linear regression analysis of *E. coli* detection was also performed down to
15 10 *cfu* of input bacteria and showed a strong positive linear correlation between input *cfu* and DNA polymerase extension activity signal as indicated by an R^2 value of 0.999 (Figure 4B). Colony count plating and *E. coli*-gene specific qPCR (gsPCR) were run in parallel, confirming both the input level of *cfu* per reaction and the ability to monitor intact genomic DNA from the exact same lysates. DNA polymerase extension activity from *S. aureus* lysates was detected to a
20 similar input level (Figure 4C). *S. aureus* detection was plotted down to 10 *cfu* of input bacteria and also showed a strong linear correlation between input *cfu* and DNA polymerase extension activity signal ($R^2 = 0.999$, Figure 4D). Colony count plating and gsPCR were performed in parallel to confirm the amount of *S. aureus* present in each bead lysis tube, as well as the presence of directly analyzable genomic DNA. Complete tables of plating, gsPCR and DNA
25 polymerase activity results for both *E. coli* and *S. aureus* can be found in Figures 3 and 4. We subsequently tested the ability of the DPE-PCR assay to measure DNA polymerase activity from seventeen additional clinically relevant microorganisms. As shown in Table 1, we were able to detect DNA polymerase activity from all seventeen additional organisms including six gram-negative bacteria, six gram-positive bacteria and five *Candida* species. Detection of the
30 seventeen additional microbes exhibited a strong positive linear correlation to input *cfu* with impressive low limits of detection. To date, and without failure we have similarly tested and detected a total of 31 different microbial species (data not shown). The upper linear dynamic ranges have yet to be fully characterized. More results containing parallel plating data and DPE-PCR results for each of 17 additional microbes are presented in Figure 8. Together, these data
35 support the notion that the performance of DPE-PCR in accordance with the teachings of the

5 present invention has the potential to be useful as a universal “pan” test for the sensitive detection of any microbe in a normally sterile environment.

As shown in Figure 2D, substitution of dCTP with ddCTP in the reaction mix represents a powerful tool for blocking the detection of DNA polymerase-dependent extension activity within our assay. To demonstrate that the signal derived from bacterial spikes was dependent upon their DNA polymerase extension activity, and not the other endogenous bacterial enzyme activities present in the lysates, we set up an experiment to compare DPE-PCR signals obtained from *E. coli* and *S. aureus* using a standard DNA polymerase reaction mix containing (dATP,dTTP,dGTP, dCTP) versus a reaction mix containing (dATP,dTTP,dGTP, ddCTP). As shown in Figure 5A, when compared to the standard reaction mix, substitution of ddCTP blocked the generation of signal derived from *E. coli cfu* spikes (Figure 5A). A dCTP rescue experiment was subsequently performed by comparing DNA polymerase extension activity from bacteria lysed in a reaction mix containing 100% ddCTP (50 μ M), to those containing 50 μ M ddCTP spiked with increasing amounts of dCTP (see materials and methods for a detailed description of rescue experiments). Figure 5B demonstrates the rescue effect that increasing amounts of dCTP has on quantifiable DNA polymerase extension activity derived from *E. coli* lysates. In addition to measuring microbial DNA polymerase extension activity, gsPCR was run in parallel to verify that equivalent amounts of *E. coli* were present in each of the assayed lysates. A graphical comparison of DNA polymerase activity versus presence of genomic DNA is presented in Figure 5C. Signal termination (via ddCTP) and dCTP rescue experiments were subsequently repeated with *S. aureus* and similar results were obtained (Figure 5D-F). Tables containing DPE-PCR and gsPCR data for both *E. coli* and *S. aureus* can be found in Figures 6 and 7. qPCR internal control values are provided to demonstrate that low levels of ddCTP carried over into qPCR are not inhibitory, and thus are not responsible for the disappearance of DNA polymerase activity signal (Figures 6A and 7A). Together, the data presented in Figure 5 strongly support the claim that the DPE-PCR assay is specifically detecting microbial DNA polymerase extension activity and signal is not derived from substrate modification via enzymatic activities other than DNA polymerase.

5 Measurement of DNA polymerase extension activity as an indicator of bacterial viability

Traditional methods for determining bacterial viability are dependent upon growth and visualization of a particular microbe on solid medium (21). Although bacterial growth and visualization is the current industry gold standard, the traditional *cfu* viability determination methods are undesirable due to the length of time required for *cfu* formation. Furthermore, the ability to grow on solid media or in liquid culture can vary dramatically from one microbe to another, thus potentially limiting the detection of certain fastidious organisms (22). Due to the aforementioned limitations of traditional methods, there is a growing need in a wide variety of pharmaceutical (23), environmental, food processing and clinical testing arenas for the rapid assessment of microbial viability. Consequently, numerous molecular methods have been developed in an effort to quickly assess microbial viability status within a given matrix (24). Despite being rapid and sensitive, molecular methods that detect the presence of nucleic acid often fall short of representing an accurate measurement of cell viability. For example, amplification of endogenous DNA or RNA is a poor indicator of bacterial viability, due to the persistence of nucleic acid after cell death (25, 26). We set out to determine the feasibility of using DNA polymerase extension activity as an indicator of bacterial viability. To this end, an experiment was designed to compare detection of DNA polymerase extension activity and PCR-mediated detection of genomic DNA as indicators of bacterial viability following various amounts of heat treatment. To begin, *E. coli* suspensions were treated at increasing temperatures for a fixed period of time. After heat treatment, bacteria were subsequently assayed for the presence of both DNA polymerase extension activity and genomic DNA. Heat treated and non-heat treated bacterial stocks were also plated in parallel to monitor bacterial viability via the presence of visible *cfu*. Figure 6A represents the levels of *E. coli* DNA polymerase extension activity measured after the indicated amounts of heat treatment. Notably, a significant drop in *E. coli* DNA polymerase extension activity was observed after incubation of bacterial suspensions between 45° C and 65° C (Figure 6A). In contrast, gsPCR signal obtained from the same lysates remained relatively constant at all temperatures and is graphically compared to DNA polymerase activity in Figure 6B. Plating results presented below the graph further demonstrate that increasing levels of heat treatment are sufficient to prevent *cfu* formation and are paralleled by a dramatic loss of DNA polymerase activity; however, dead cells still contribute genomic DNA levels very close to their original input levels confirming that gsPCR is a poor indicator of the

5 presence of viable cells (Figure 6B). In Figure 6C, the bar graphs further highlight the relative
abilities of DPE-PCR and gsPCR to monitor the disappearance of *cfu* in response to lethal
amounts of heat treatment. Subsequently, we wanted to test whether measurement of DNA
polymerase extension activity could be used to indicate the viability status of a gram positive
organism as well. The previous *E. coli* experiments were repeated with *S. aureus* under the same
10 conditions.

Figure 7A-C show similar results obtained from heat treatment experiments repeated with
S. aureus. Collectively, the strong concordance between the presence of *cfu* and DNA
polymerase extension activity shown in Figures 4, 6, 7, and Table 1 of Figure 8, demonstrates
that DPE-PCR performed according to this invention has potential to be used as a general
15 indicator of cell viability, and may additionally present the possibility of measuring DNA
polymerase extension activity from microbes exposed to other clinically or pharmaceutically
relevant agents (bacteriostatic and bactericidal) aimed at reducing cell proliferation or viability.

In summary, in accordance with the present invention there has been developed a novel,
highly sensitive, quantitative and rapid DPE-PCR assay. In addition to quantitative detection of
20 extremely low levels of purified enzyme, we have demonstrated the ability of DPE-PCR to
reproducibly measure DNA polymerase extension activity from less than 10 *cfu* of bacteria via
coupling to bead lysis. We have also demonstrated the potential for DPE-PCR to universally
detect microbes by testing a panel of microorganisms comprised of seven gram-negative
bacteria, seven gram-positive bacteria and five *Candida* species. Furthermore, preliminary
25 evidence that the DPE-PCR assay can be used to assess bacterial viability was provided via the
reproducibly strong correlation between DNA polymerase extension activity and proliferation as
indicated by the presence of *cfu*. Considering the data disclosed herein, it is presently believed
that the novel methods and techniques of the invention such as the preferred DPE-PCR assay as
disclosed herein, has the potential to become a useful tool for a wide range of testing applications
30 within pharmaceutical, environmental, food and clinical settings.

The contents of all references, patents and published patent applications cited throughout
this application, are incorporated herein by reference to the same extent as if each individual

5 publication, patent or patent application was specifically and individually indicated to be incorporated by reference.

The foregoing detailed description has been given for clearness of understanding only and no unnecessary limitations should be inferred therefrom as modifications will be obvious to those skilled in the art. It is not an admission that any of the information provided herein is prior
10 art or relevant to the presently claimed inventions, or that any publication specifically or implicitly referenced is prior art.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs.

15 While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications and this application is intended to cover any variations, uses, or adaptations of the invention following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the
20 essential features hereinbefore set forth.

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5 What is claimed is:

1. A method for performing a diagnostic assay for the detection of the presence or amount of a microorganism within a sample matrix containing active DNA polymerase by the measurement of DNA polymerase extension activity, which assay comprises the steps of incubating DNA polymerase in the sample matrix with a selected suitable substrate, and
10 performing PCR cycling and detection via the use of a selected suitable nucleic acid probe, thereby to detect endogenous DNA polymerase extension activity in the sample matrix as an indication of the presence or amount of said microorganism.
2. The method of claim 1, wherein measurement of DNA polymerase extension activity is an indicator of bacterial viability in said sample matrix.
- 15 3. The method of claim 1, wherein the sample matrix is serum.
4. The method of claim 1, wherein the sample matrix is plasma.
5. The method of claim 1, wherein the sample matrix is wherein the sample matrix is selected from the group consisting of purified enzymes, microbial lysates and crude microbial lysates.
- 20 6. The method of claim 1, wherein the assay specifically detects microbial DNA polymerase extension activity and signal is not derived from modification of said substrate via enzymatic activities other than DNA polymerase.
7. The method of claim 5, wherein prior to the performance of the assay, the method comprises the additional step of adding a microbial lysate or crude microbial lysate
25 known to contain, or suspected of containing, a microorganism to a bead mill lysis tube, disrupting the microorganism cells and transferring the disrupted cells into the incubation step of the assay.
8. The method of claim 6, comprising the further step of adding a blocking agent to the sample mixture at a suitable point in the performance of the method to ensure that the
30 assay specifically detects microbial DNA polymerase extension activity and assay signal is not derived from modification of said substrate via enzymatic activities other than DNA polymerase.
9. The method of claim 1, wherein the sample matrix is a biological sample.
10. The method of claim 1, wherein the sample matrix is an environmental sample.

Figure 1

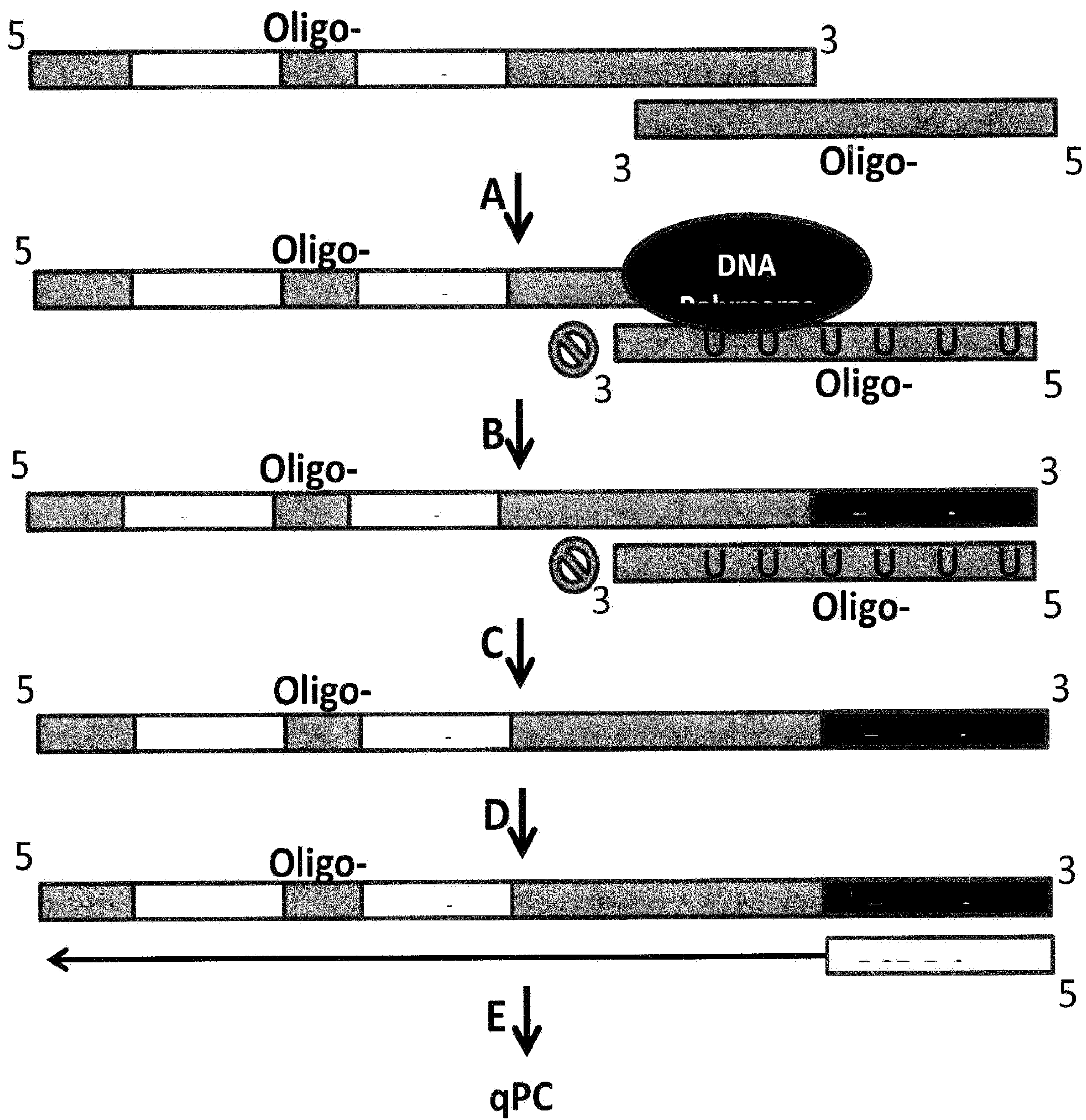
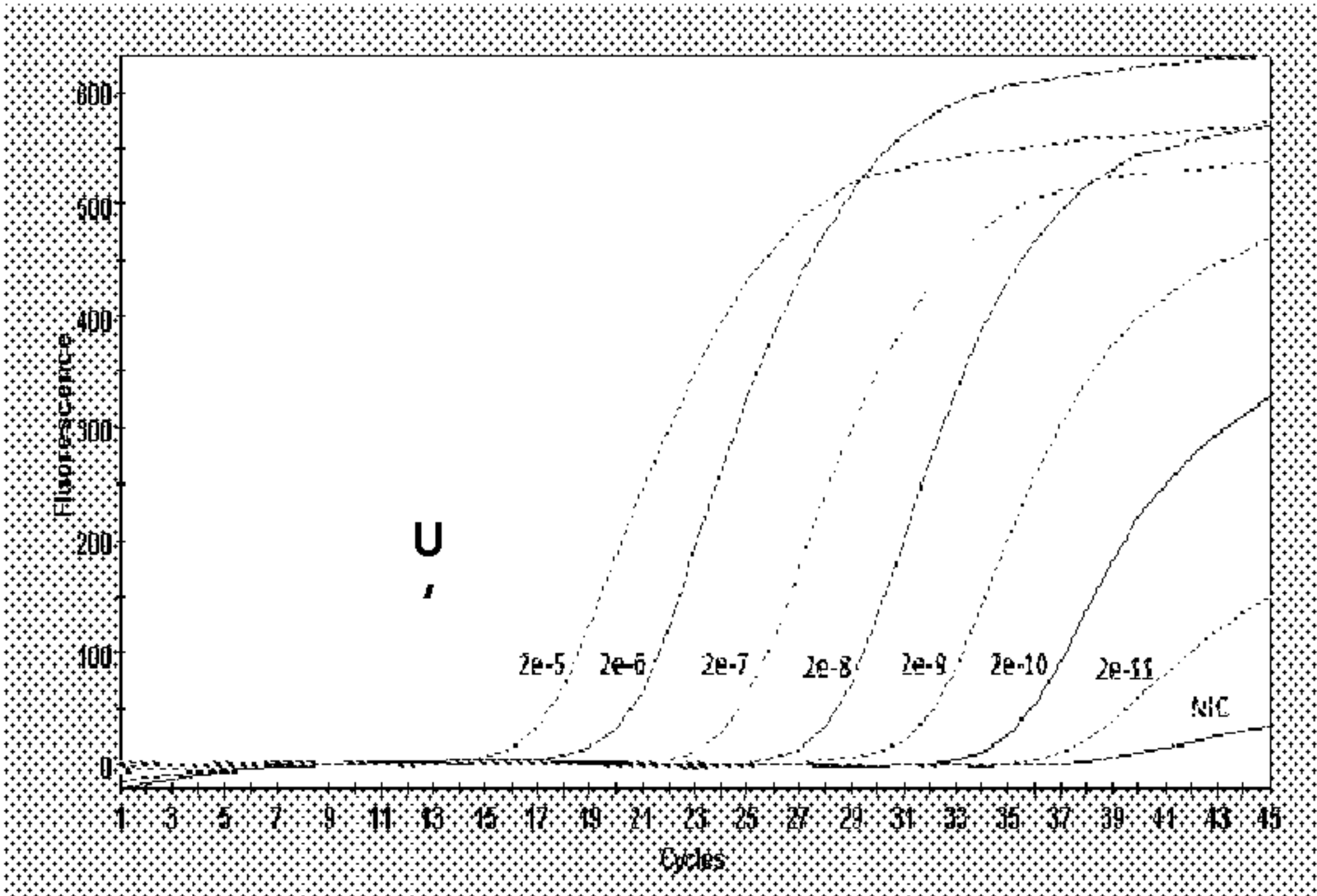
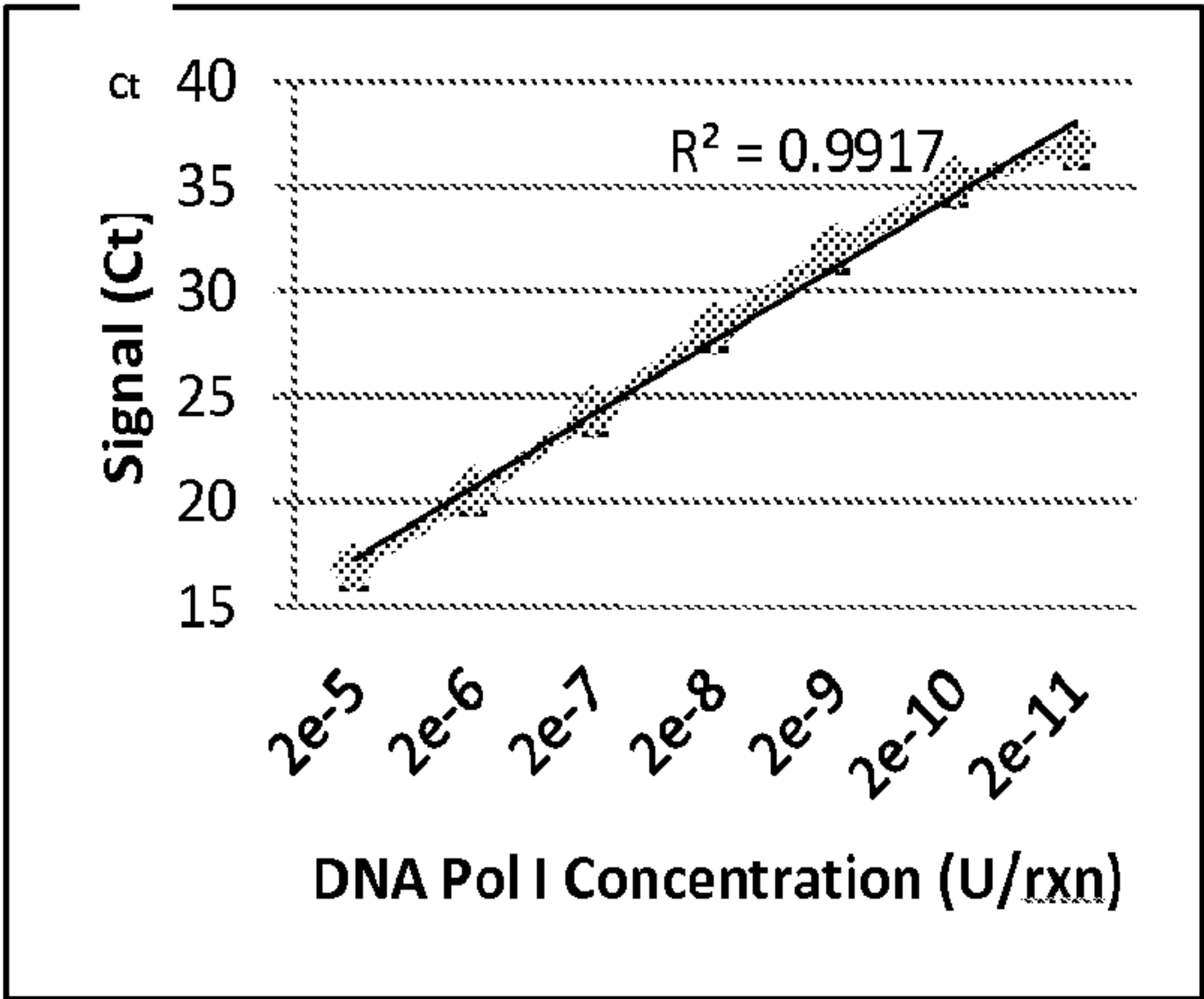


Figure 2

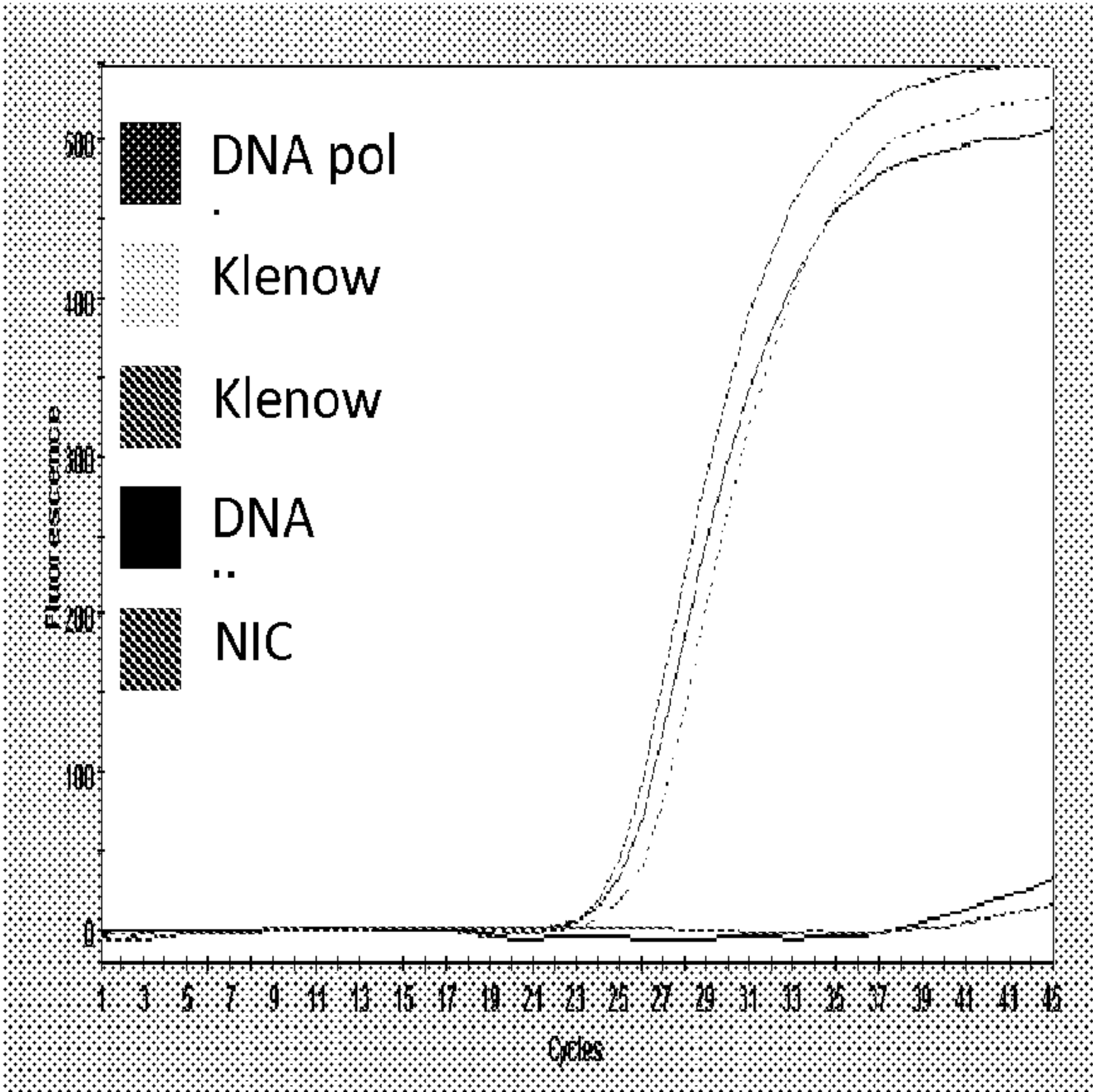
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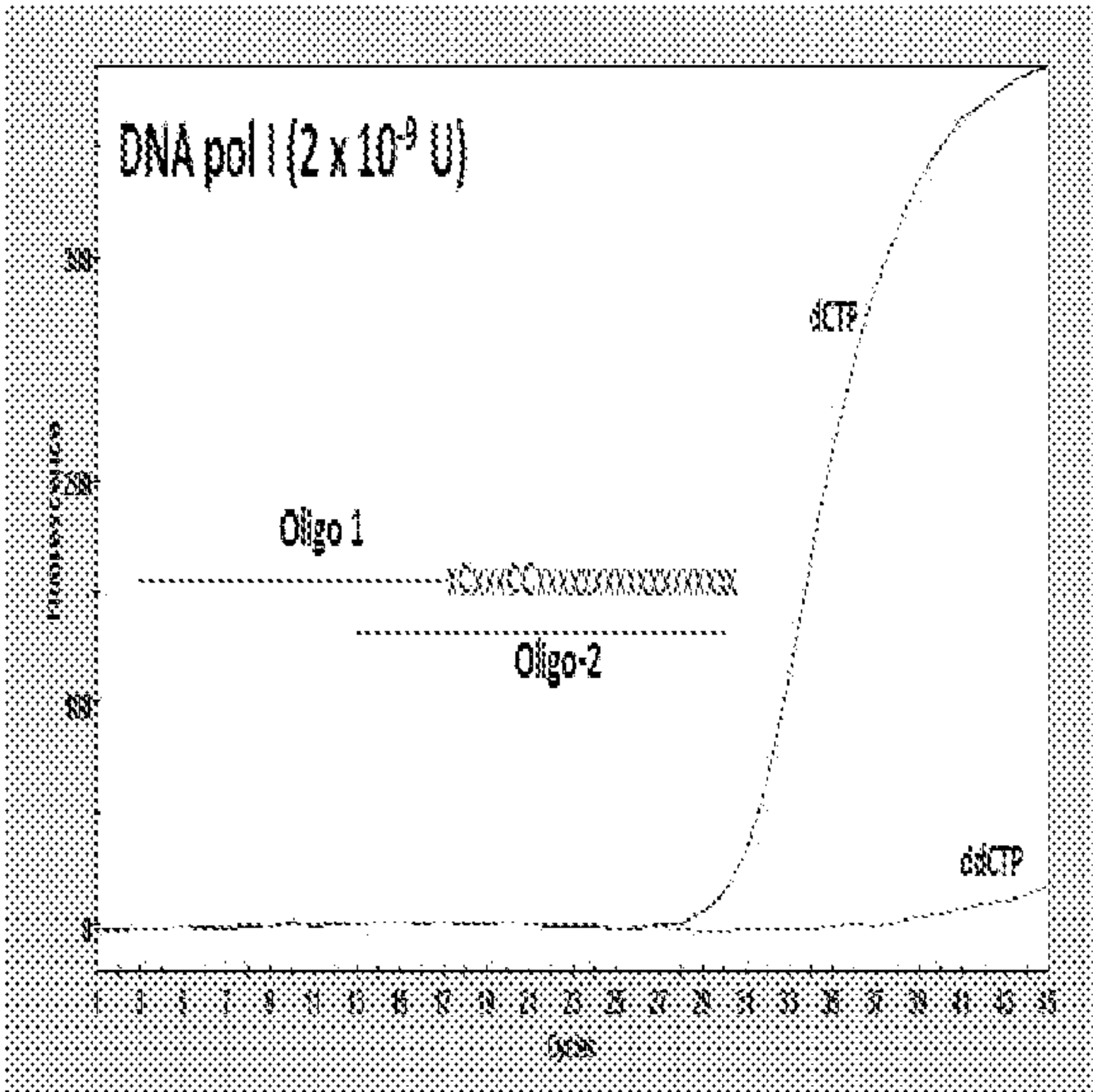


Figure 3

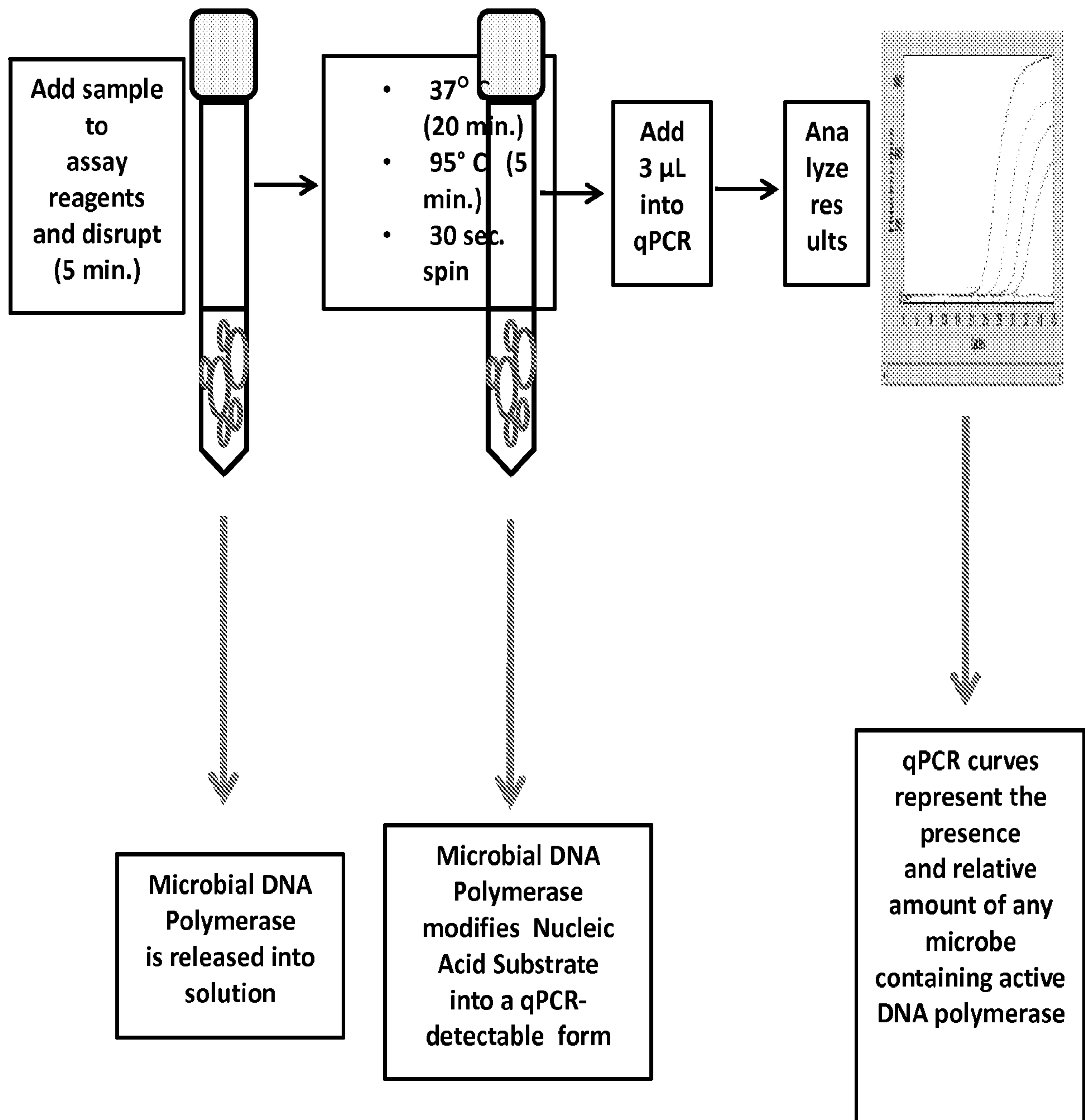
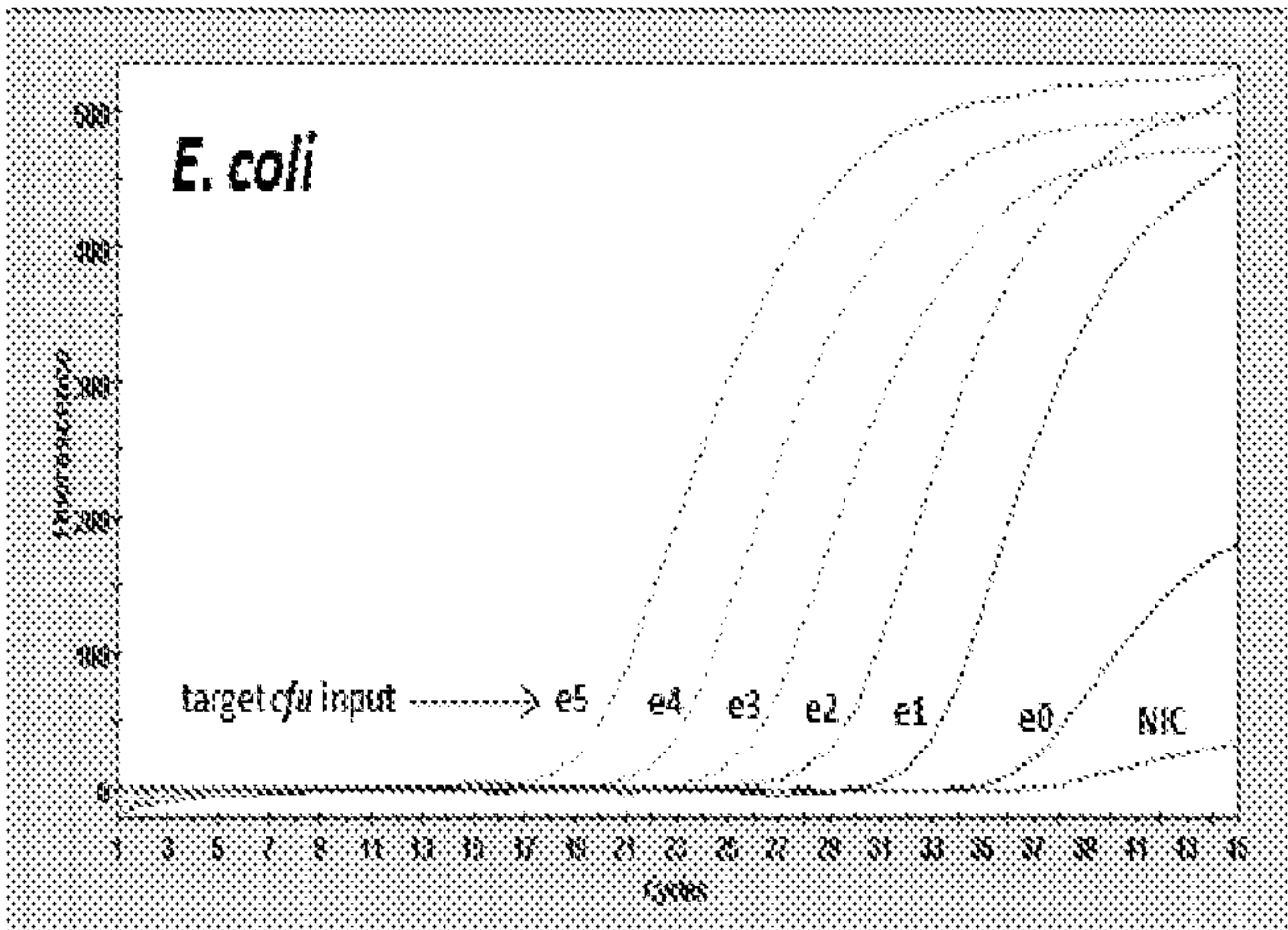
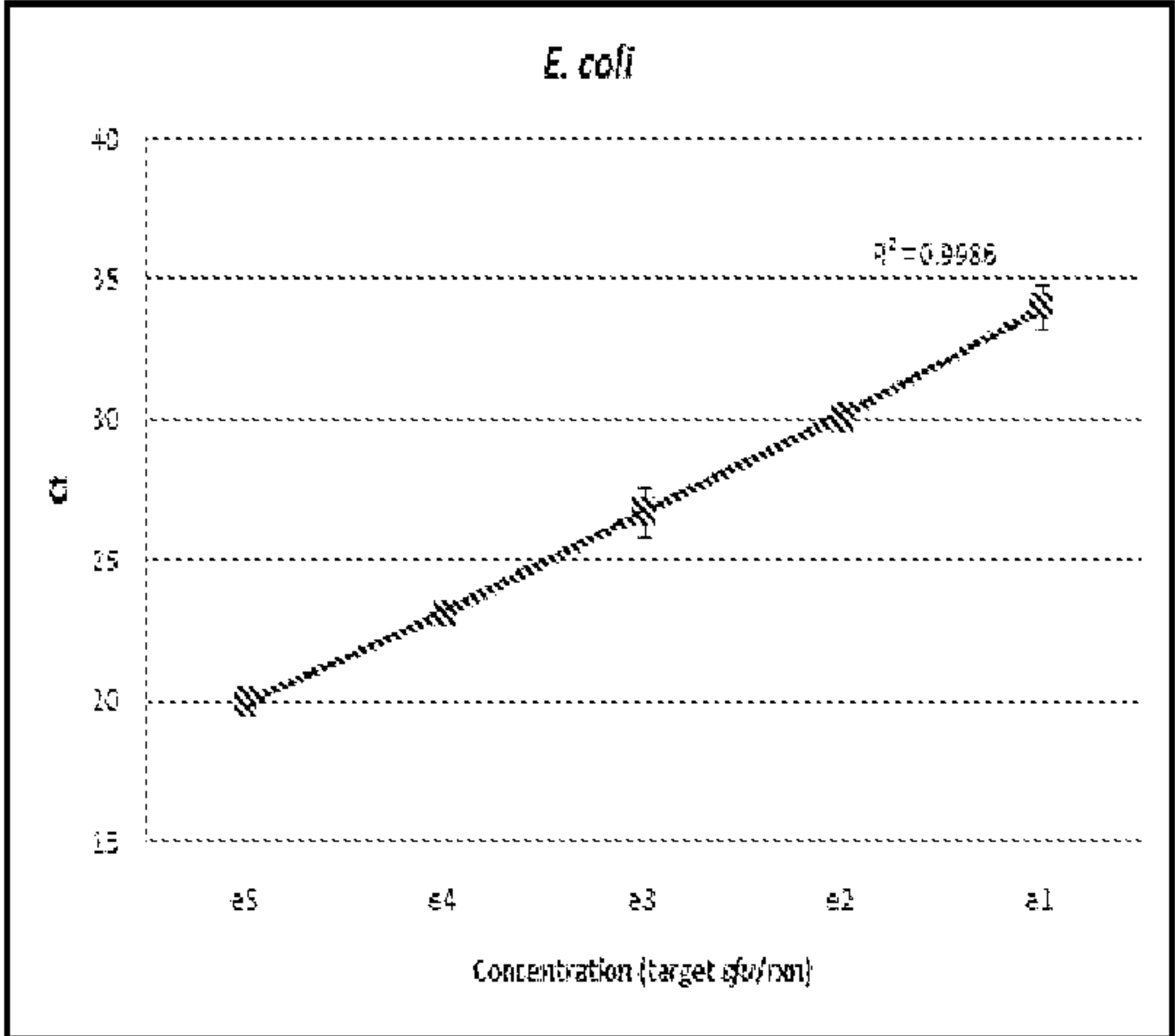


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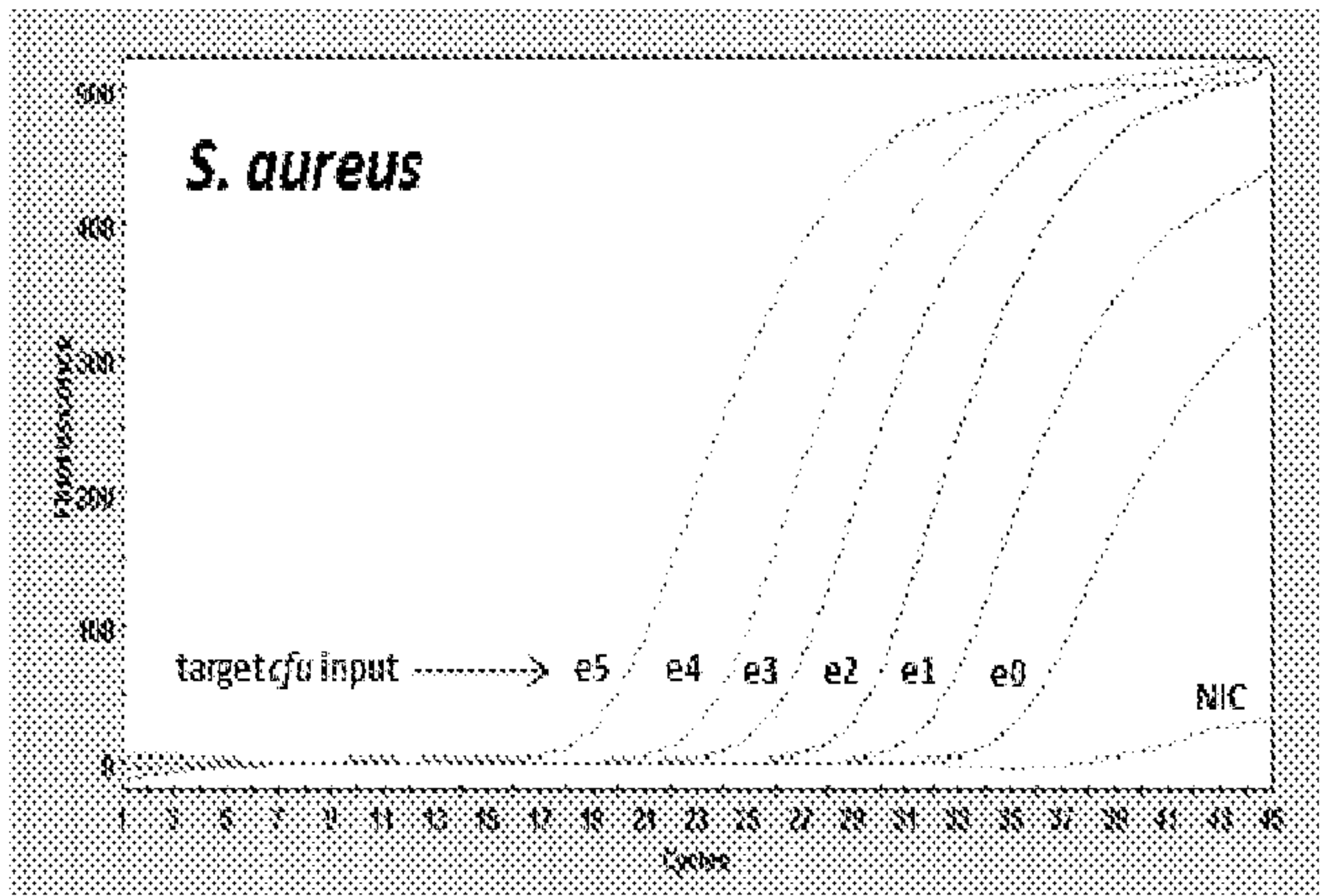
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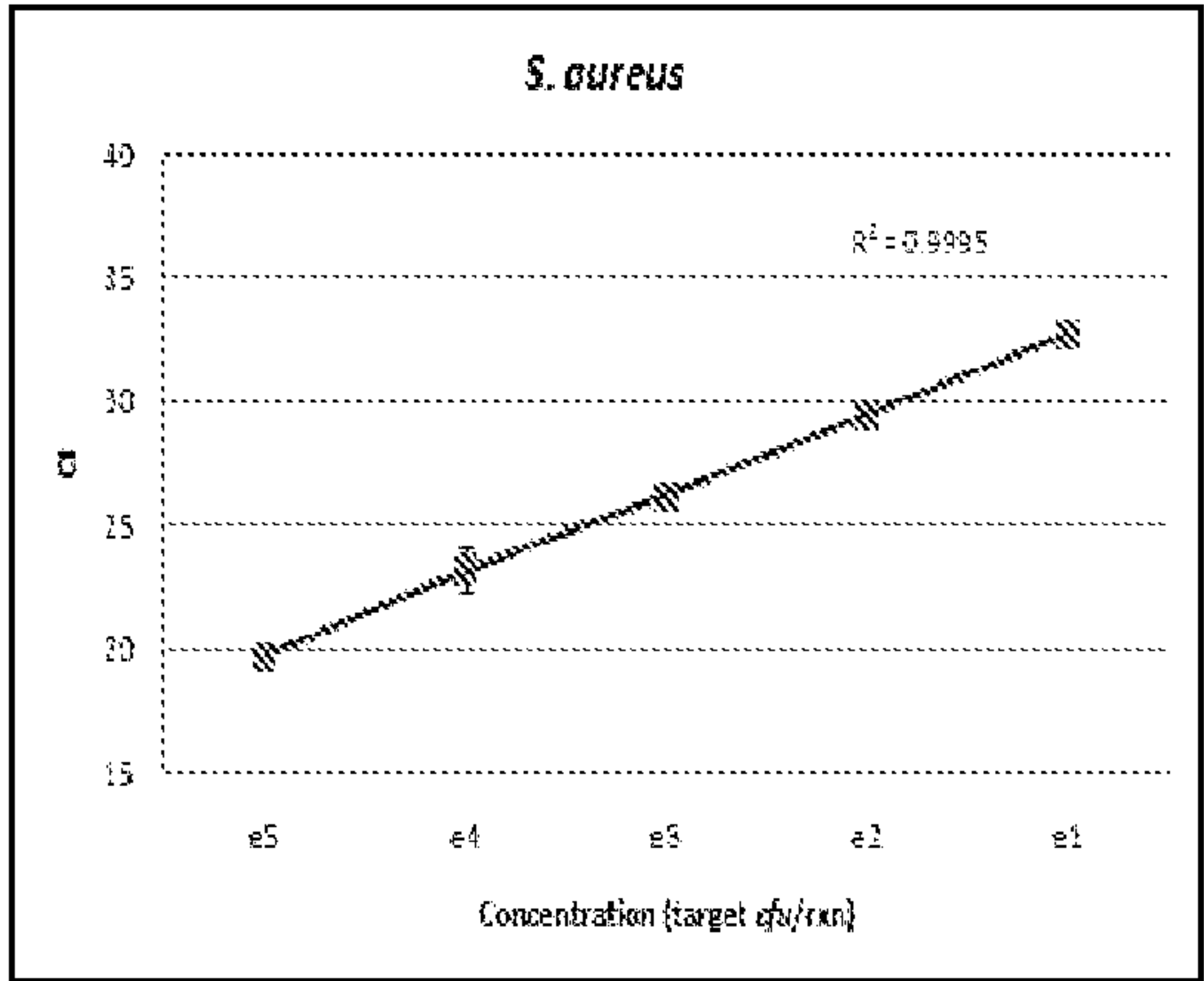
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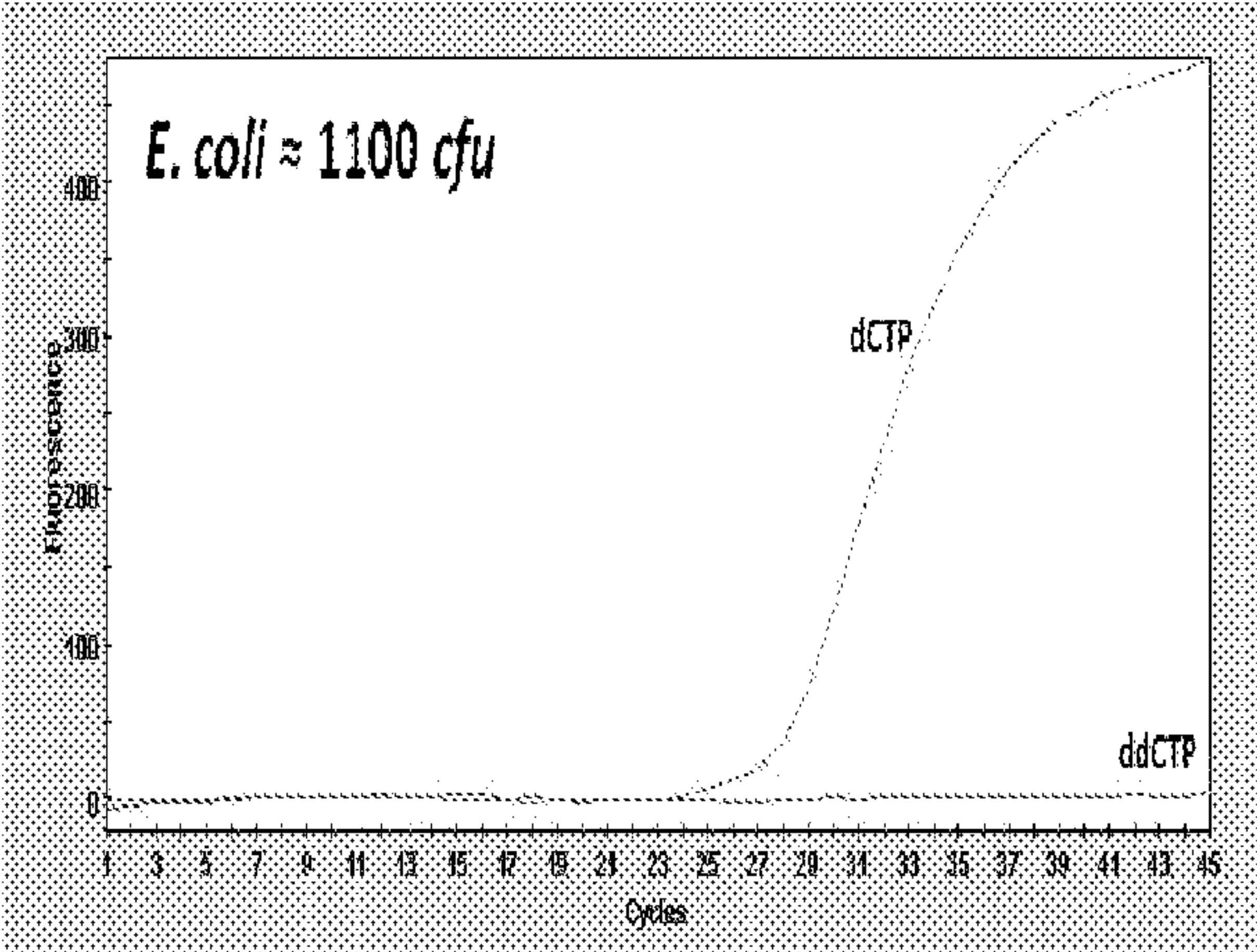


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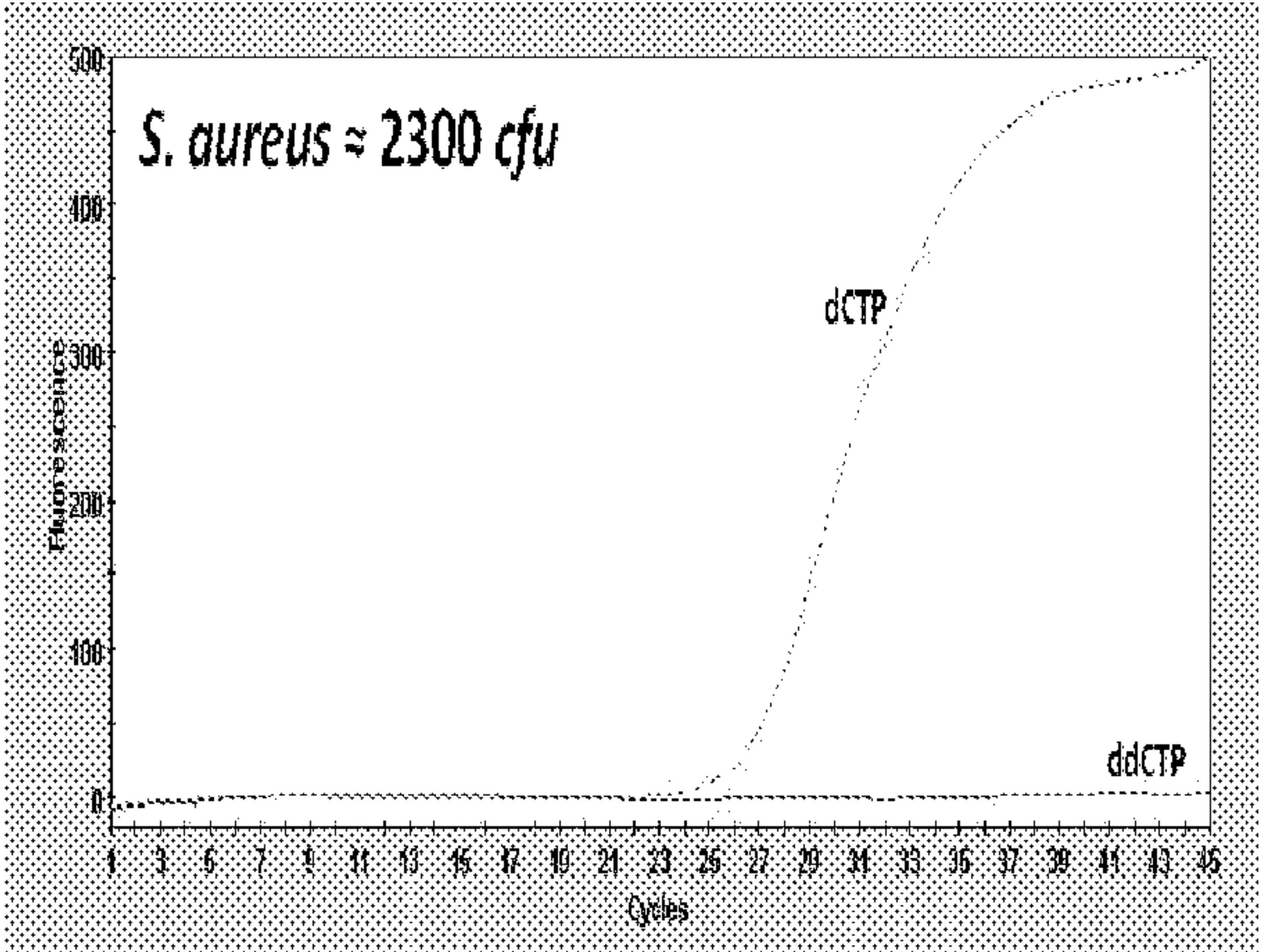
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Figure 5

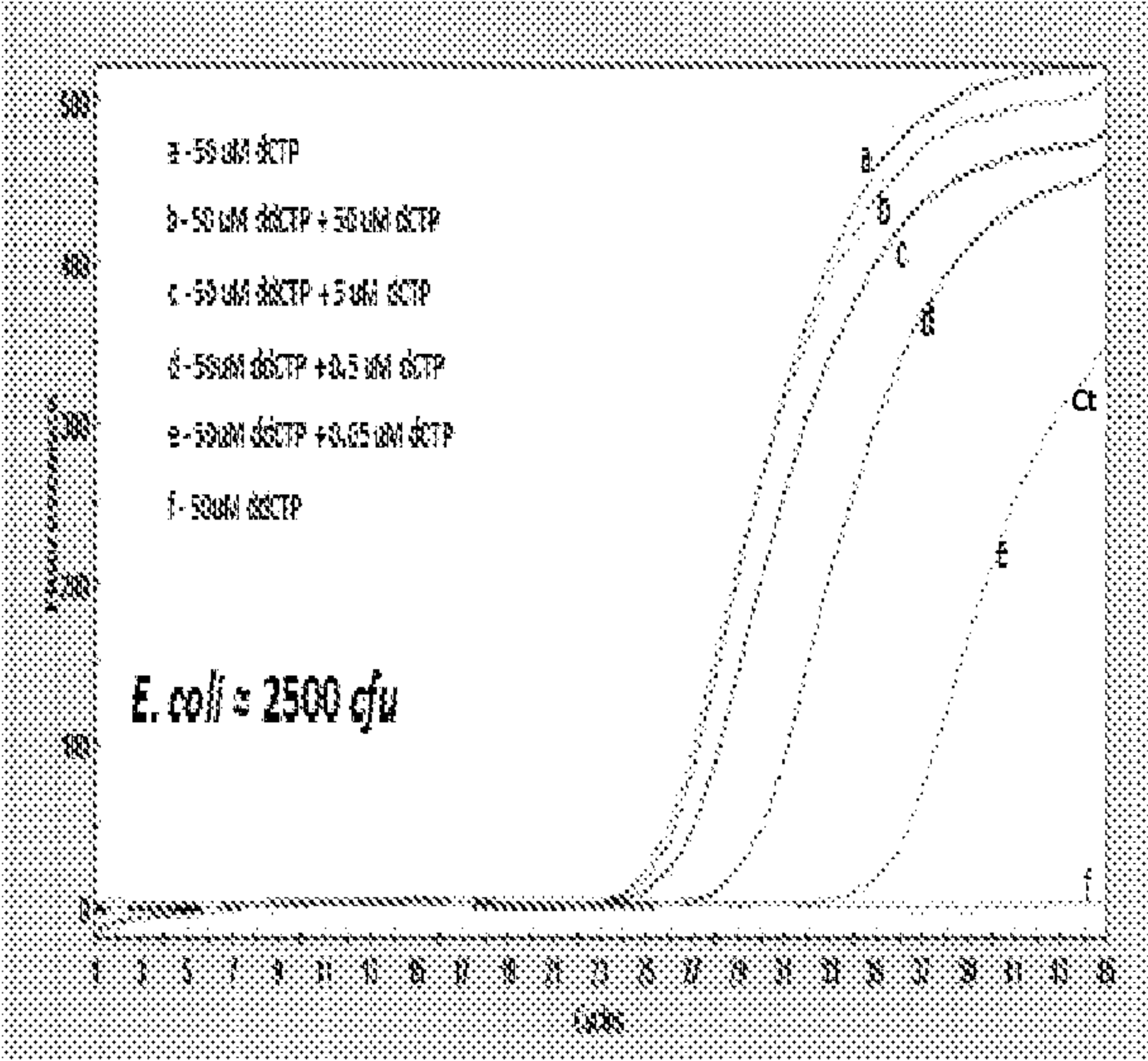
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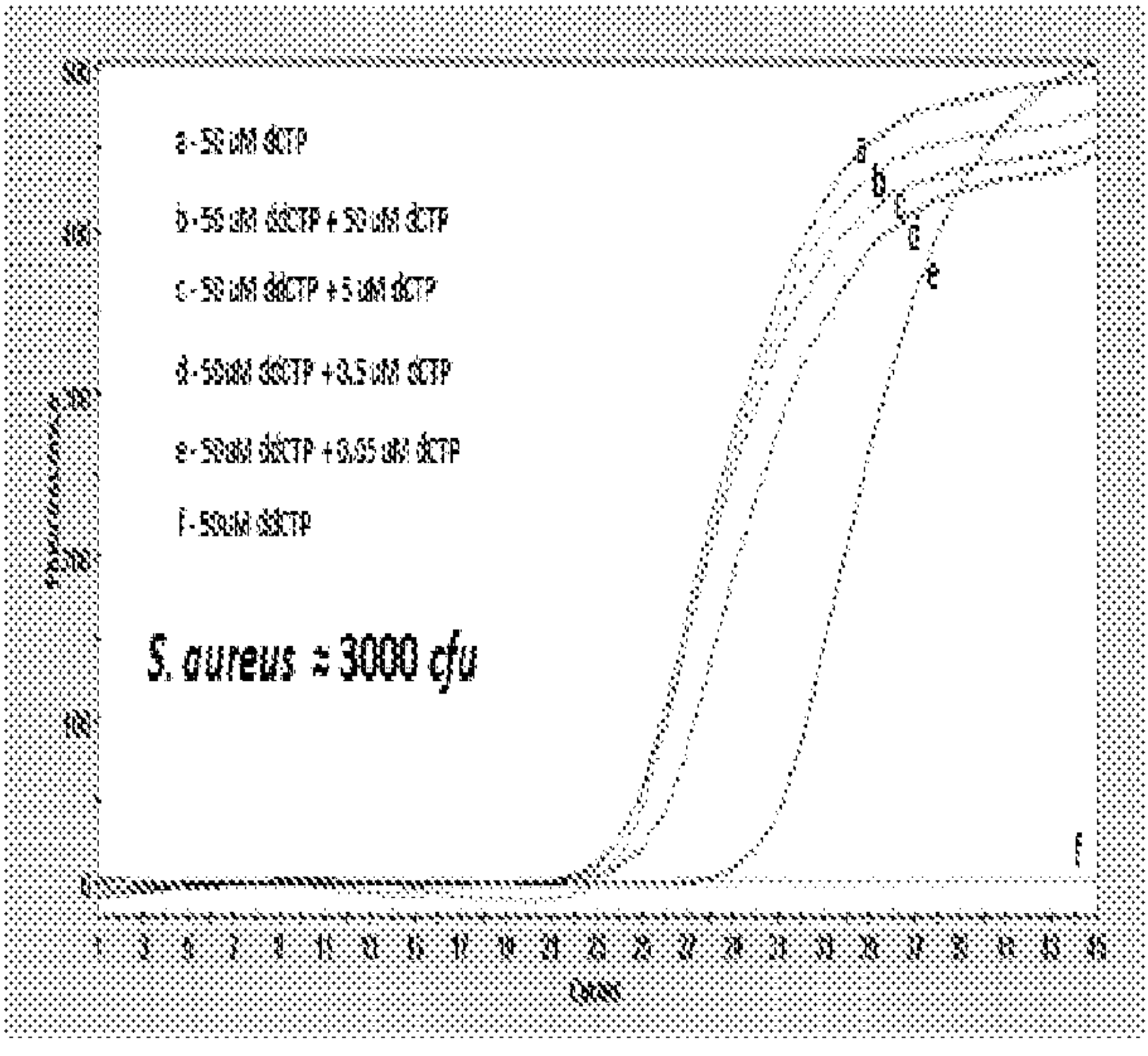
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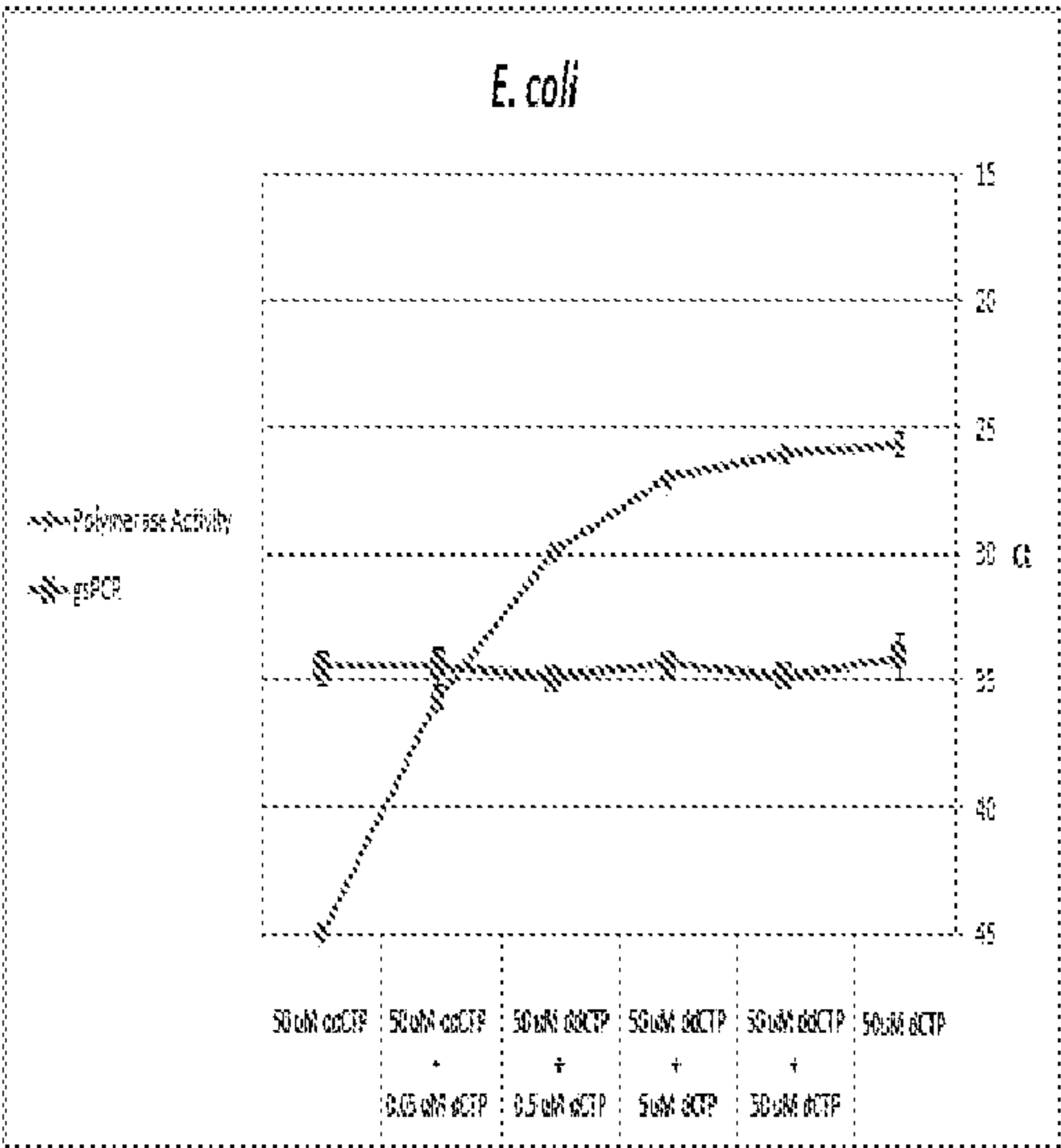
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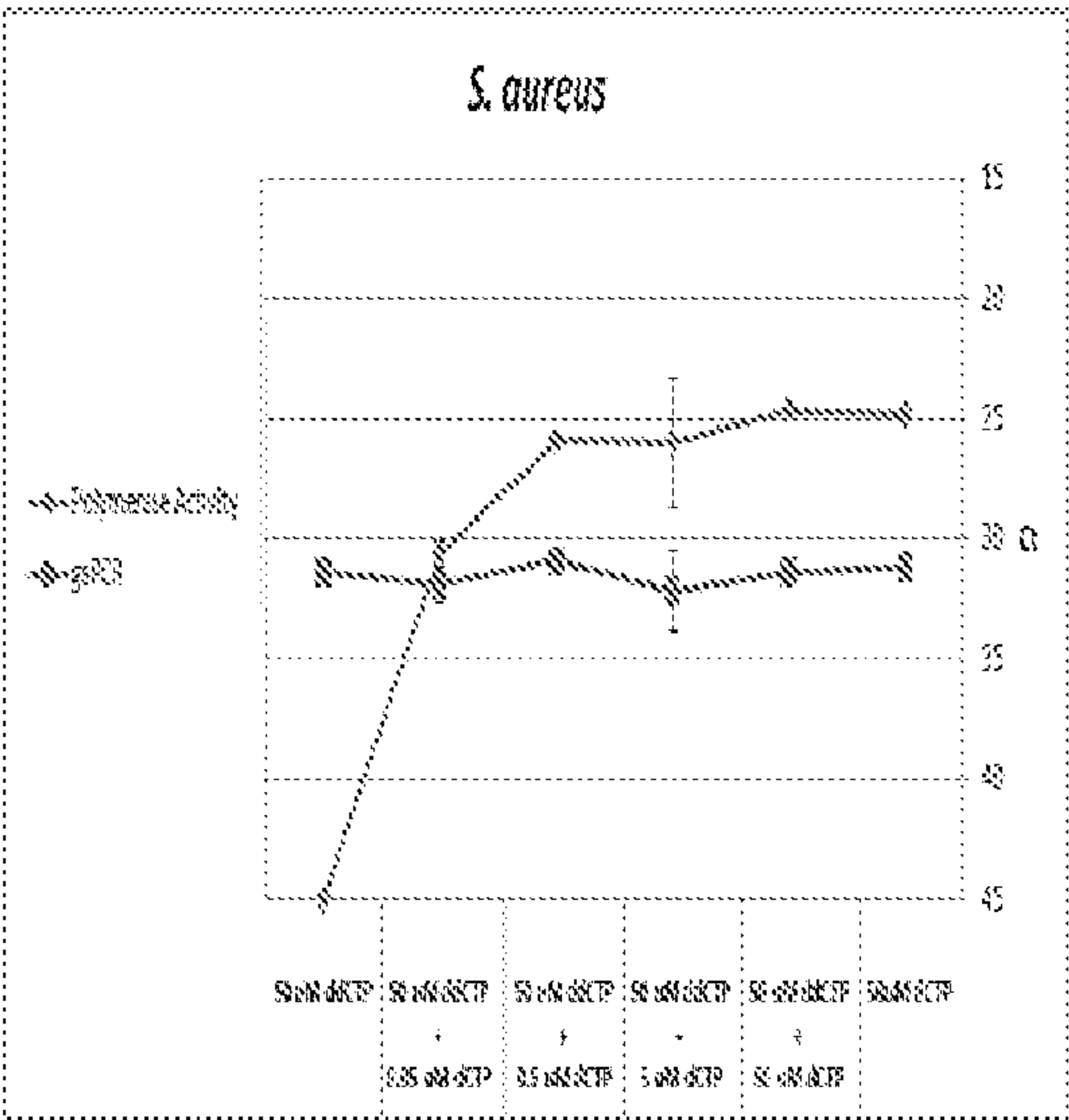


Figure 6

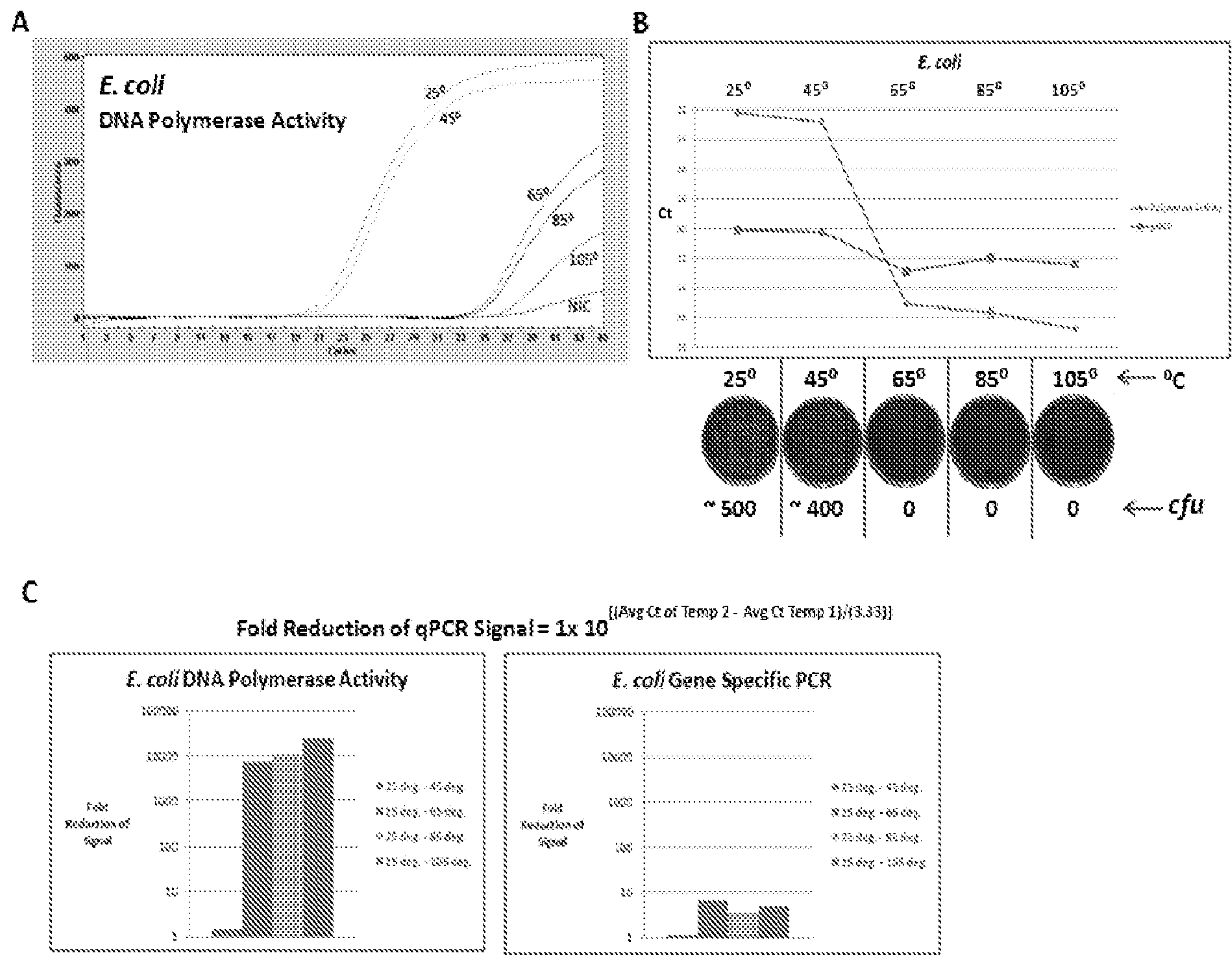


Figure 7

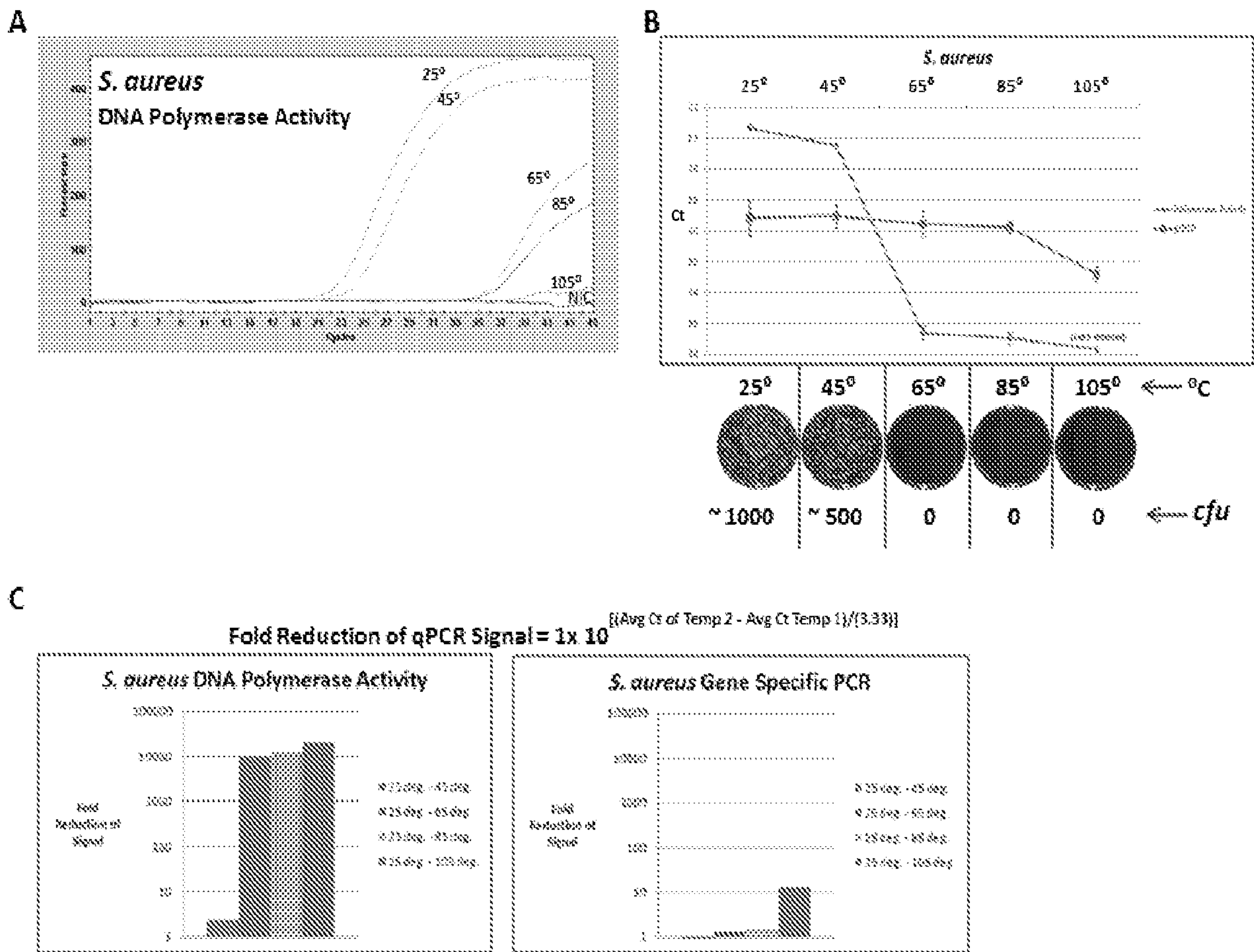


Figure 8

Table 1

Bacterial panel	Lower Limit Detected by DPE-PCR	R ² (1e4-1e1 cfu)
<i>Klebsiella pneumoniae</i>	<10	0.9957
<i>Pseudomonas aeruginosa</i>	<10	0.9860
<i>Enterobacter cloacae</i>	<10	0.9995
<i>Acinetobacter baumannii</i>	<10	0.9980
<i>Haemophilus influenzae</i>	<10	0.9996
<i>Serratia marcescens</i>	<10	0.9956
<i>Enterococcus faecalis</i>	<10	0.9963
<i>Enterococcus faecium</i>	<10	0.9899
<i>Streptococcus pyogenes</i>	<10	0.9945
<i>Streptococcus agalactiae</i>	<10	0.9969
<i>Streptococcus pneumoniae</i>	<10	0.9999
<i>Staphylococcus epidermidis</i>	<10	0.9990

Candida panel	Lower Limit Detected by DPE-PCR	R ² (1e5-1e3 cfu)
<i>Candida albicans</i>	≈ 20	0.9945
<i>Candida tropicalis</i>	≈ 20	0.9969
<i>Candida glabrata</i>	≈ 40	0.9111
<i>Candida parapsilosis</i>	≈ 20	0.9950
<i>Candida krusei</i>	≈ 15	0.9868

Figure 1

