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(56) Related Art

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(54) Title: METHODS OF TREATING T CELL EXHAUSTION BY INHIBITING OR MODULATING T CELL RECEPTOR SIG-NALING

(57) Abstract: The present invention relates to T cell compositions and methods of using the same in the context of therapy and treatment. In particular, the invention provides chimeric antigen receptor (CAR) T cells that are modified to maintain functionality under conditions in which unmodified CAR T cells display exhaustion. Compositions and methods disclosed herein find use in inhibiting or reversing CAR T cell exhaustion (e.g., by modulating CAR surface expression) thereby enhancing CAR T cell function. Compositions and methods of the invention find use in both clinical and research settings, for example, within the fields of biology, immunology, medicine, and oncology.

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METHODS OF TREATING T CELL EXHAUSTION BY INHIBITING OR MODULATING T CELL RECEPTOR SIGNALING

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. provisional application Serial No. 62/479,930, filed March 31, 2017, which is incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

The present invention relates to methods of treating T cell exhaustion. In particular, the invention relates to methods of preventing or reversing T cell exhaustion by transiently inhibiting T cell receptor (TCR) signaling to restore T cell function.

BACKGROUND

T cells are immune cells that become activated via T cell receptor (TCR) signaling and co-stimulation following engagement with antigen. Physiologic activation through the T cell receptor renders T cells capable of mediating potent antitumor and/or anti-infective effects. During resolution of an acute inflammatory response, a subset of activated effector T cells differentiate into long-lived memory cells. By contrast, in patients with chronic infections or cancer, T cells may undergo pathologic differentiation toward a state of dysfunction, which has been termed T cell exhaustion. T cell exhaustion is characterized by marked changes in metabolic function, transcriptional programming, loss of effector function (e.g, cytokine secretion, killing capacity), and co-expression of multiple surface inhibitory receptors. The root cause of T cell exhaustion is persistent antigen exposure leading to continuous TCR signaling. Prevention or reversal of T cell exhaustion has been long sought as a means to enhance T cell effectiveness (e.g., in patients with cancer or chronic infections).

Any discussion of the prior art throughout the specification should in no way be considered as an admission that such prior art is widely known or forms part of the common general knowledge in the field.

Unless the context clearly requires otherwise, throughout the description and the claims, the words "comprise", "comprising", and the like are to be construed in an inclusive sense as opposed to an exclusive or exhaustive sense; that is to say, in the sense of "including, but not limited to".

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SUMMARY

According to a first aspect, the present invention provides a chimeric antigen receptor (CAR) comprising:

- a) an extracellular ligand-binding domain;
- b) a transmembrane domain;
- c) a cytoplasmic domain comprising one or more signaling domains; and
- d) a regulatable destabilization domain (RDD), wherein the RDD comprises a dihydrofolate reductase destabilization domain (DHFR DD) or an FK506 binding protein 12 (FKBP) destabilization domain (FKBP DD) comprising the following amino acid substitutions: E31G, F36V, R71G and K105E.

According to a second aspect, the present invention provides a genetically modified T cell comprising a nucleic acid sequence encoding the CAR of the invention.

According to a third aspect, the present invention provides a method of prolonging a T lymphocyte cell (T cell) effector function in a mammal comprising introducing into the mammal a T cell comprising a chimeric antigen receptor of the invention.

According to a fourth aspect, the present invention provides a method for treating a mammal suffering from cancer comprising introducing into the mammal the genetically modified T cell of the invention for treating cancer in the mammal.

According to a fifth aspect, the present invention provides a method for stimulating a T cell-mediated immune response to a target cell population or tissue in a mammal comprising administering to a mammal an effective amount of a T cell genetically modified to express a chimeric antigen receptor (CAR) of the invention.

According to a sixth aspect, the present invention provides a method of providing an anti-cancer immune response in a mammal, the method comprising administering to the mammal an effective amount of a T cell genetically modified to express a CAR of the invention.

According to a seventh aspect, the present invention provides a method of treating or delaying the progression of cancer in a patient comprising administering to the patient a therapeutically effective amount of T cells genetically modified to express a chimeric antigen receptor (CAR) of the invention.

According to an eighth aspect, the present invention provides a therapeutically effective amount of a composition comprising T cells genetically modified to express the CAR of the invention for use in treating or delaying the progression of cancer in a subject. 5

According to a ninth aspect, the present invention provides a use of the genetically modified T cell of the invention for the manufacture of a medicament for treating a mammal suffering from cancer.

According to a tenth aspect, the present invention provides a use of a T cell genetically modified to express a CAR of the invention for the manufacture of a medicament for providing an anti-cancer immune response in a mammal.

According to an eleventh aspect, the present invention provides a use of a therapeutically effective amount of T cells genetically modified to express a chimeric antigen receptor (CAR) of the invention for the manufacture of a medicament for treating or delaying the progression of cancer in a patient.

The invention relates to methods of treating T cell T cell exhaustion by transiently inhibiting T cell receptor (TCR) signaling. In one aspect, the present invention provides compositions and methods for use in preventing chimeric antigen receptor (CAR) T cell exhaustion. In particular, the invention provides CAR T cells modified to maintain functionality under conditions in which unmodified CAR T cells become dysfunctional (e.g., display exhaustion). CAR T cells modified according to the invention (e.g., via modulation of CAR surface expression), compositions containing same, and methods of using same

enhance T cell functionality including, for example, activity against cancer or infectious disease.

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In some embodiments, CAR T cell exhaustion is inhibited or reversed via modulation of CAR surface expression. In some embodiments, modulation of CAR surface expression is made possible via fusion of a regulatable destabilization domain (RDD) to a CAR T cell. In some embodiments, the RDD is a ligand-regulatable destabilization domain. For example, regulation of CAR expression is achieved, in some embodiments, by fusing the CAR with an RDD, which renders the resulting modified CAR protein unstable and prone to proteasomal degradation. By adding a ligand such as a small molecule or drug (e.g., Shield-1 or trimethoprim (TMP)), the modified CAR protein is shielded from degradation and the CAR construct comprising an RDD is stably expressed (see Banaszynski et al., (2006) Cell 126: 995–1004; Banaszynski et al., (2008) Nat Med 14: 1123–1127;. Iwamoto et al., (2010). Chem Biol 17: 981–988.)

In some embodiments, the RDD comprises a binding domain derived from dihydrofolate reductase (DHFR) RDD. In some embodiments, the DHFR-derived RDD comprises a binding domain from a bacterial DHFR. In some embodiments, the bacterial DHFR is an *Escherichia coli* DHFR. In some embodiments, the DHFR-derived RDD comprises a binding domain from human DHFR. In some embodiments, the RDD comprises a binding domain derived from an FK506 Binding Protein (FKBP). In some embodiments, the FKBP is FKBP12. In some embodiments, the nucleic acid sequence of a FKBP DD comprises the sequence set forth in SEQ ID NO: 1. In one embodiment, the FKBP DD comprise the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 2. In another embodiment, the FKBP DD portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO. 2.

In some embodiments, an ecDHFR destabilization domain is used. In some embodiments, the nucleic acid sequence of an ecDHFR DD comprises the sequence set forth in SEQ ID NO: 3. In one embodiment, the ecDHFR DD comprise the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 4. In another embodiment, the ecDHFR DD portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO. 4. In other embodiments, the nucleic acid sequence of an ecDHFR DD comprises the sequence set forth in SEQ ID NO: 5. In one embodiment, the ecDHFR DD comprise the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 6. In another embodiment, the ecDHFR DD portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO. 6.

The invention is not limited by the type of ligand. In some embodiments, Shield-1 or trimethoprim (TMP) is used to stabilize and promote stable surface expression a fusion protein containing a DHFR RDD or a FKBP RDD.

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In some embodiments, a T cell is provided that is genetically engineered to express a CAR comprising an RDD (e.g., fused to the CAR) that marks the CAR for degradation (e.g., ubiquitin-mediated and/or proteosomal degradation) such that the addition or presence of a stabilizing small molecule drug that directly interacts (e.g., binds) with the RDD prevents CAR degradation, and removal (e.g., by metabolic clearance) or absence of the stabilizing small molecule drug that directly interacts with the RDD allows CAR degradation to occur. While an understanding of a mechanism is not needed to practice the present invention, and while the present invention is not limited to any particular mechanism, in some embodiments, tonic CAR signaling is prevented by allowing degradation or destruction of CAR proteins such that surface expression levels of any CAR proteins are below the concentration necessary to permit tonic CAR signaling. In some embodiments, modulation (e.g., reduction) of CAR protein levels and CAR surface expression via use of a such a "drug on" RDD to the CAR reinvigorates exhausted T cells that have already developed the hallmarks of T cell exhaustion due to tonic CAR signaling. For example, CAR T cells that are exposed to the stabilizing small molecule demonstrate all of the phenotypic and functional hallmarks of T cell exhaustion. However, when the stabilizing small molecule is removed and CAR protein levels are reduced by protein degradation, the phenotypic and functional indicators of T cell exhaustion are reversed and effector T cell function is restored.

In another aspect, the invention provides a genetically modified T cell comprising an isolated nucleic acid sequence encoding a chimeric antigen receptor (CAR), wherein the isolated nucleic acid sequence comprises nucleic acid sequence (e.g., human, mouse, or humanized mouse nucleic acid sequence) of a tumor antigen-binding domain (e.g., a single chain variable fragment, or scFv), the nucleic acid sequence of a transmembrane domain, the nucleic acid sequence of an intracellular domain of one or more costimulatory molecules, the nucleic acid sequence of a CD3 zeta signaling domain, and the nucleic acid sequence of an RDD domain.

The invention additionally includes a vector comprising an isolated nucleic acid sequence encoding a chimeric antigen receptor (CAR), wherein the isolated nucleic acid sequence comprises nucleic acid sequence (e.g., human, mouse, or humanized mouse nucleic acid sequence) of a tumor antigen-binding domain (e.g., a single chain variable fragment, or scFv), the nucleic acid sequence of a transmembrane domain, the nucleic acid sequence of an

intracellular domain of one or more costimulatory molecule, the nucleic acid sequence of a CD3 zeta signaling domain, and the nucleic acid sequence of an RDD domain.

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In addition, the invention includes methods for providing anti-tumor immunity in a subject having cancer. The methods comprise administering to the subject an effective amount of a genetically modified T cell comprising an isolated nucleic acid sequence encoding a chimeric antigen receptor (CAR), wherein the isolated nucleic acid sequence comprises nucleic acid sequence (e.g., human, mouse, or humanized mouse nucleic acid sequence) of a tumor antigen-binding domain (e.g., a single chain variable fragment, or scFv), the nucleic acid sequence of a transmembrane domain, the nucleic acid sequence of an intracellular domain of one or more costimulatory molecule, the nucleic acid sequence of a CD3 zeta signaling domain, and the nucleic acid sequence of an RDD domain, thereby providing anti-tumor immunity in the subject. In some embodiments, the subject is a human.

Further included in the invention is a method for stimulating a beneficial and/or therapeutic T cell-mediated immune response to a cell population, tumor or tissue in a subject. The method comprises administering to the subject an effective amount of a genetically modified T cell comprising an isolated nucleic acid sequence encoding a chimeric antigen receptor (CAR), wherein the isolated nucleic acid sequence comprises nucleic acid sequence (e.g., human, mouse, or humanized mouse nucleic acid sequence) of a tumor antigen-binding domain (e.g., a single chain variable fragment, or scFv), the nucleic acid sequence of a transmembrane domain, the nucleic acid sequence of an intracellular domain of one or more costimulatory molecule, the nucleic acid sequence of a CD3 zeta signaling domain, and the nucleic acid sequence of an RDD domain domain, thereby stimulating a T cell-mediated immune response in the subject.

Also provided here are methods of treating cancer in a subject. The methods comprise administering to a subject having cancer a therapeutically effective amount of a genetically modified T cell comprising an isolated nucleic acid sequence encoding a chimeric antigen receptor (CAR), wherein the isolated nucleic acid sequence comprises nucleic acid sequence (e.g., human, mouse, or humanized mouse nucleic acid sequence) of a tumor antigen-binding domain (e.g., a single chain variable fragment, or scFv), the nucleic acid sequence of a transmembrane domain, the nucleic acid sequence of an intracellular domain of one or more costimulatory molecule, the nucleic acid sequence of a CD3 zeta signaling domain, and the nucleic acid sequence of an RDD domain, thereby treating cancer in the subject.

The invention further includes a method of generating a persisting population of genetically engineered T cells (e.g., memory T cells) in a subject diagnosed with cancer, the

method comprising: administering to the subject an effective amount of a genetically modified T cell comprising an isolated nucleic acid sequence encoding a chimeric antigen receptor (CAR), wherein the wherein the isolated nucleic acid sequence comprises nucleic acid sequence (e.g., human, mouse, or humanized mouse nucleic acid sequence) of a tumor antigen-binding domain (e.g., a single chain variable fragment, or scFv), the nucleic acid sequence of a transmembrane domain, the nucleic acid sequence of an intracellular domain of one or more costimulatory molecule, the nucleic acid sequence of a CD3 zeta signaling domain, and the nucleic acid sequence of an RDD domain, wherein the persisting population of genetically engineered T cells persists in the subject (e.g., for weeks, a month, months, etc.) after administration. In some embodiments, the persisting population of genetically engineered T cells persists in the human for at least three months after administration.

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The invention is not limited by the means of expressing a CAR containing an RDD. In some embodiments, a CAR is expressed constitutively. In other embodiments, a CAR is expressed in a regulated fashion (e.g., using a system to regulate expression via a small molecule or using an endogenously regulated system). A CAR, in another embodiment, is genetically integrated into the cellular DNA using a retroviral, lentiviral or other viral vector or via CRISPR/Cas9 based system. In yet another embodiment, a CAR is expressed via RNA or an oncolytic virus or other transient expression system known in the art. CARs can be delivered *ex vivo* into T cells for adoptive transfer, or delivered via *in vivo* genetic transfer.

The invention is not limited by the type of T cell genetically modified to express and/or contain a CAR of the invention. In some embodiments, the T cells are CD3+ T cells (e.g., CD4+ and/or CD8+ T cells). In certain embodiments, the T cells are CD8+ T cells. In other embodiments, the T cells are CD4+ T cells. In some embodiments, the T cells are natural killer (NK) T cells. In some embodiments, the T cells are alpha beta T cells. In some embodiments, the T cells are a combination of CD4+ and CD8 T+ cells (e.g., CD3+). In certain embodiments, the T cells are memory T cells. In certain embodiments, the memory T cells are central memory T cells. In other embodiments, the memory T cells are effector memory T cells. In some embodiments, the T cells are tumor-infiltrating lymphocytes (TILs). In certain embodiments, the T cells are a combination of CD8+ T cells, CD4+ T cells, NK T cells, memory T cells, and/or gamma delta T cells. In some embodiments, the T cells are cytokine-induced killer cells.

In some embodiments, CAR T cells include peripheral blood derived T cells genetically modified with a CAR that recognizes and responds to tumor antigens. Such

receptors are generally composed of extracellular domains comprising a single-chain antibody (scFy) specific for tumor antigen, a transmembrane domain, and an intracellular signaling domain (See, e.g., Westwood, J. A. et al, 2005, Proc. Natl. Acad. Sci., USA, 102(52):19051-19056). In other embodiments, the T cell is engineered to express a CAR of human or murine origin that recognizes a tumor antigen. The invention is not limited by the type of tumor antigen recognized. Indeed, any CAR that recognizes a tumor antigen finds use in the compositions and methods of the invention. Examples include, but are not limited to, a CAR that recognize an antigen selected from HER2, CD19, CD20, CD22, CD30, CD33/IL3Ra, CD123, CD38, receptor tyrosine kinase-like orphan receptor 1 (ROR1), ErbB3/4, Glycolipid F77, epidermal growth factor receptor (EGFR), EGFR variant III (EGFRVIII), melanoma antigen recognized by T cells 1 (MART-1), EphA2, FAP, human carcinoembryonic antigen (CEA), EGP2, EGP40, mesothelin, TAG72, prostate-specific membrane antigen (PSMA), NKG2D ligands, B7-H6, IL-13 receptor α2, IL-11 receptor α, MUC1, MUC16, CA9, disialoganglioside 2 (GD2), GD3, tyrosine protein kinase Met (c-Met) or hepatocyte growth factor receptor (HGFR), HMW-MAA, CD171, Lewis Y, G250/CAIX, melanoma antigen gene (MAGE) Family Member A3 (MAGE-A3), HLA-AI, MAGE A1, NY-ESO-1, PSC1, folate receptor-α, CD44v7/8, 8H9, NCAM, VEGF receptors, Fetal AchR, NKG2D ligands, CD44v6, TEM1, TEM8, GP1000, p53, Epstein-Barr Virus (EBV) protein or antigen, or other viral-associated antigens expressed by a tumor.

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The invention is not limited by the type CAR. Indeed, any CAR that binds with specificity to a desired antigen (e.g., tumor antigen or other type of antigen) may be modified as disclosed and described herein in order to modulate the CAR's surface expression. In certain embodiments, the CAR comprises an antigen-binding domain. In certain embodiments, the antigen-binding domain is a single-chain variable fragment (scFv) containing heavy and light chain variable regions that bind with specificity to a desired antigen. In some embodiments, the CAR comprises a transmembrane domain and a signaling domain comprising one or more immunoreceptor tyrosine-based activation motifs (ITAMs). In some embodiments, the transmembrane domain comprises a transmembrane domain of a protein selected from the group consisting of the T-cell receptor (TCR) alpha chain, the TCR beta chain, CD3 zeta, CD28, CD3 epsilon, CD45, CD4, CD5, CD8, CD9, CD16, CD22, CD33, CD37, CD64, CD80, CD86, CD134, CD137, and CD154. The intracellular signaling domain, in some embodiments, comprises a functional signaling domain of a 4-1BB polypeptide, a functional signaling domain of a CD3 zeta polypeptide, or both. In some

embodiments, the CAR comprises one or more co-stimulatory domains (e.g., domains that provide a second signal to stimulate T cell activation). The invention is not limited by the type of co-stimulatory domain. Indeed, any co-stimulatory domain known in the art may be used including, but not limited to, CD28, OX40/CD134, 4-1BB/CD137/TNFRSF9, the high affinity immunoglobulin E receptor-gamma subunit (Fc ϵ RI γ), ICOS/CD278, interleukin 2 subunit beta (ILR β) or CD122, cytokine receptor common subunit gamma (IL-2R γ) or CD132, and CD40. In one embodiment, the co-stimulatory domain is 4-1BB.

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The invention is not limited to any particular RDD or the domain within the CAR to which the RDD is fused. In some embodiments, the RDD is fused to the CAR antigenbinding domain. In other embodiments, the RDD is fused to a hinge or transmembrane CAR domain. In still further embodiments, the RDD is fused to a cytoplasmic CAR stimulatory domain such as, but not limited to, CD28, 4-1BB, OX-40. In yet other embodiments, the RDD is fused to a CAR CD3 zeta domain. Indeed, according to the present methods and compositions, cells comprising a CAR comprising an RDD may have the RDD attached to any portion of the CAR provided the CAR retains antigen-specific activity.

In another aspect, the invention provides a method of treating a disease or condition in a subject comprising administering to the subject (e.g., a patient) having a disease or condition an effective amount of CAR T cells comprising a CAR containing or fused to an RDD. The invention is not limited by the type of disease or condition treated. Indeed, any disease or condition that is treatable (e.g., for which signs or symptoms of the disease are ameliorated upon treatment) via administration of CAR T cells can be treated in an improved and more effective manner using compositions and methods of the invention. In one embodiment, the disease or condition is cancer. In another embodiment, the disease or condition is an infectious disease. The invention is not limited by the type of cancer or by the type of infectious disease. Indeed, any cancer or disease known in the art for which CAR T cell therapy is used for treatment may be treated with the compositions and methods of the invention. For example, compositions and methods of the invention can be used to modify any CAR T cell therapy known in the art via genetically introducing an RDD into the CAR of therapeutic CAR T cells. In some embodiments, administration to a subject (e.g., a patient) having a disease or condition of an effective amount of CAR T cells of the invention inhibits or reverses T cell exhaustion in the patient (e.g., compared to a subject receiving the same amount of CAR T cells lacking a DD).

In another aspect, the invention provides a method of maintaining, regaining, or enhancing functionality of CAR T cells (e.g., CAR T cells that would otherwise experience antigen induced tonic signaling and exhaustion in the context of treating a disease or condition) via selective modulation (e.g., reduction) of CAR cell surface expression. In some embodiments, incorporation of an RDD into the CAR allows modulation (e.g., reduction) of CAR expression during culture and expansion of T cells in vitro or ex vivo, which in turn enhances functionality (e.g., effector function) of the CAR T cells in the context of treating a disease or condition in a subject. The invention is not limited by the type of functionality maintained, regained or enhanced. In some embodiments, the functionality is antigen induced cytokine production. In other embodiments, the functionality is CAR T cell cytotoxicity (e.g., increased recognition of tumor targets). In still other embodiments, the functionality is increased memory cell formation and/or enhanced proliferation in response to antigen. In some embodiments, modulation (e.g., reduction) of CAR cell surface expression results in measurable reduction of markers indicative of exhaustion including, but not limited to, PD-1, TIM-3, and LAG-3 in the CAR T cells. In other embodiments, modulation (e.g., reduction) of CAR cell surface expression results in a reduction in the levels of CAR T cell programmed cell death. In still further embodiments, modulation (e.g., reduction) of CAR cell surface expression on CAR T cells significantly enhances clinical efficacy of CAR T cell therapy.

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In another aspect, the invention provides methods of treating or delaying the progression of cancer in a patient comprising administering to the patient a therapeutically effective amount of a composition comprising CAR T cells modified (e.g., genetically) to express a CAR containing or fused to an RDD. In certain embodiments, the therapeutically effective amount of the composition comprising modified CAR T cells reduces the number of cancer cells in the patient following such treatment. In certain embodiments, the therapeutically effective amount of the composition comprising modified CAR T cells reduces and/or eliminates the tumor burden in the patient following such treatment. In certain embodiments, the method further comprises administering radiation therapy to the patient. In certain embodiments, the radiation therapy is administered before, at the same time as, and/or after the patient receives the therapeutically effective amount of the composition comprising modified CAR T cells. In certain embodiments, the method further comprises administering to the patient one or more anticancer agents and/or one or more chemotherapeutic agents. In certain embodiments, the one or more anticancer agents and/or one or more chemotherapeutic agents are administered before, at the same time as, and/or after the patient receives the therapeutically effective amount of the composition comprising modified CAR T cells. In

certain embodiments, treatment of a patient with a therapeutically effective amount of modified CAR T cells and a course of an anticancer agent produces a greater tumor response and clinical benefit in such patient compared to those treated with the modified CAR T cells or anticancer drugs/radiation alone. Since the doses for all approved anticancer drugs and radiation treatments are known, the present invention contemplates the various combinations of them with the modified CAR T cells.

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In certain embodiments, the invention provides a therapeutically effective amount of a composition comprising CAR T cells modified according to the present disclosure (e.g., for use in treating or delaying the progression of cancer in a subject). As described herein, the composition may be administered before, during, or after other types of cancer treatment (e.g., chemotherapy, surgical resection of cancer, or radiation therapy). The invention also provides the use of the composition to induce cell cycle arrest and/or apoptosis. The invention also relates to the use of the compositions for sensitizing cells to additional agent(s), such as inducers of apoptosis and/or cell cycle arrest, and chemoprotection of normal cells through the induction of cell cycle arrest. Compositions of the invention are useful for the treatment, amelioration, or prevention of disorders, such as any type of cancer or infectious disease and additionally any cells responsive to induction of apoptotic cell death (e.g., disorders characterized by dysregulation of apoptosis, including hyperproliferative diseases such as cancer). In certain embodiments, the compositions can be used to treat, ameliorate, or prevent a cancer that additionally is characterized by resistance to cancer therapies (e.g., those cancer cells which are chemoresistant, radiation resistant, hormone resistant, and the like). The invention also provides pharmaceutical compositions comprising the composition (e.g., immunotherapeutic compositions) comprising modified CAR T cells of the invention in a pharmaceutically acceptable carrier.

In another embodiment, the invention provides a method of treating or delaying the progression of cancer in a patient comprising administering to the patient a therapeutically effective amount of a composition comprising CAR T cells modified (e.g., genetically) according to the present disclosure in combination with a therapeutically effective amount of an inhibitor of TCR signaling (e.g., in order to prevent T cell exhaustion). Multiple cycles of treatment may be administered to a subject. In certain embodiments, the inhibitor of TCR signaling is administered according to a standard dosing regimen (e.g., daily or intermittently). In another embodiment, the inhibitor of TCR signaling is administered for a period of time sufficient to restore at least partial T cell function, then discontinued.

These and other embodiments of the subject invention will readily occur to those of skill in the art in view of the disclosure herein.

DESCRIPTION OF THE DRAWINGS

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- FIG. 1: Characterization of the GD2.28z.FKBP CAR. T cells were transduced with lentivirus encoding the GD2.28z.FKBP CAR on day 1 after activation and subsequently cultured with various concentrations of shield-1 in the growth medium. On day 7, CAR expression was quantified via FACS.
- FIG. 2: Removal of S1 from culture medium results in reversal of T cell exhaustion marker surface expression.
 - FIG. 3: Removal of S1 from culture medium results in maintenance of CD62L expression and prevention of apoptosis.
 - FIG. 4: Removal of S1 from culture medium results in reversal of function T cell exhaustion.
- FIG. 5: Removal of surface CAR results in more effective prevention of T cell exhaustion compared PD-1/PDL-1 blockade.
 - FIG. 6: Removal of surface CAR rescues exhaustion in PD-1/TIM-3/LAG-3 triple positive CAR T cells after only 4 days.
- FIG. 7: Dasatinib inhibits cytokine secretion of CAR T cells in response to tumor antigen.
 - FIG. 8: Dasatinib reverses exhaustion marker expression and co-expression.
 - FIG. 9: Dasatinib treatment results in maintenance of CD62L expression.
 - FIG. 10: Dasatinib Treatment results in augmented IL-2 and IFNγ secretion in response to tumor antigen.
- FIG. 11 depicts a schematic of a chimeric antigen receptor (CAR) in one embodiment of the invention. For example, the CAR may contain an extracellular antigen-binding domain (e.g., a single chain variable fragment (scFv)) and extracellular hinge region, a transmembrane domain, an intracellular co-stimulatory domain (e.g., containing CD28 or 4-1BB domains), an intracellular CD3 zeta domain, and a regulatable destabilization domain (RDD).
 - FIG. 12 shows dose-dependent regulation of several different CARs of multiple sizes containing an RDD fused at a C-terminus position. A) CARs fused to an FKBP12 DD that can be regulated by the rapalog shield-1. B) CARs fused to an E. coli derived DHFR DD that

can be regulated by trimethoprim.

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FIG. 13 shows rapid CAR protein stabilization or destabilization in the presence or absence of stabilizing drug/small molecule, respectively. In general, IC50, that is, the time it takes for 50% of surface CAR to be degraded, was less than 2 hours.

FIG. 14 shows that stabilizing drug regulates RDD-CAR anti-tumor activity. The absence of stabilizing drug results in low CAR surface expression in vitro with accompanying attenuated cytokine secretion in the response to tumor and attenuated cytotoxicity compared to CAR T cells cultured in the presence of stabilizing drug and high CAR surface expression.

FIG. 15 shows that trimethoprim regulates CAR surface expression and activity of DHFR DD-CARs *in vivo*. FIG. 15A shows that CAR expression is regulatable in vivo, and that the dynamic range of CAR surface expression achieved in vitro is similar to that observed in vivo. FIG. 15B shows that mice that received injections of the stabilizing drug trimethoprim (TMP) not only exhibited CAR expression, but that the higher-expressing CAR T cells in this group also exhibit higher CD69 surface expression, indicating that they became activated in response to the tumor in vivo. FIG. 15C shows differences in the tumor burden of mice which received daily injections of vehicle versus mice that received TMP. Mice that received TMP, and thus expressed CAR on the surface, show a greater capacity to control tumor growth than mock and vehicle controls.

FIG. 16 shows that tonic signaling in HA-GD2.28z is prevented or reversed via a fused RDD. HA-GD2.28z FKBP CAR tonically signals in the absence of tumor antigen when the CAR is expressed on the T cell surface evidenced by western blotting in which the CAR CD3 zeta domain is phosphorylated at baseline. The tonic signaling leads to T cell exhaustion, which renders these CAR T cells less effective. Tonic signaling is prevented in RDD-CAR T cells by expanding them in the absence of stabilizing drug. When CAR expression was reduced or inhibited and not expressed on the T cell surface (in the absence of shield-1, CAR always OFF), or if the CAR was expressed at one point but then removed from the surface for 72 hours (CAR OFF D7), an attenuation in CD3 zeta phosphorylation was observed.

FIG. 17 shows that co-expression of canonical exhaustion markers on HA-GD2.28z.FKBP are rapidly downregulated upon removal of shield-1 from the culture medium and that prevention or reversal of tonic signaling improves the exhaustion phenotype of CAR T cells. The co-expression of PD-1/TIM-3/LAG-3 is correlated with a decrease in effector function. When the CAR is always expressed (cultured with shield-1, "CAR always

ON"), these markers are co-expressed in a majority of cells. However, when the CAR is not expressed on the surface (no shield-1, "CAR always OFF"), or cultured in shield-1 for 7 days and then removed, only a fraction of cells that co-express exhaustion markers are observed. FIG. 17B, shows a similar trend for cells that co-express Tbet and Blimp-1, which are transcription factors correlated with T cell exhaustion.

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FIG. 18 shows that inhibition or removal of tonic signaling rescues memory phenotype. T cell exhaustion inhibits T cells' ability to form memory. Exhausted (CAR always ON) cells have a high percentage of effector T cells (Teff) and a low percentage of stem cell memory (Tscm). Conversely, non-exhausted cells (CAR always OFF), or those cells in which exhaustion was reversed by removing stabilizing drug from culture (CAR OFF D7 or CAR OFF D10), exhibit an increased stem cell memory compartment.

FIG. 19 shows time-dependent regulation of CAR surface expression via a fused FKBP or *E.coli*-derived DHFR destabilization domain (DD). Exhausted CAR T cells fail to secrete effector cytokines IL-2 and IFNγ in response to tumor. However, CAR T cells in which exhaustion was prevented or reversed exhibit an enhanced capacity to secrete IL-2 and IFNγ.

FIG. 20 shows that exhausted CAR T cells have significantly reduced cytotoxicity. However, CAR T cells in which exhaustion was prevented or reversed exhibit enhanced cytotoxicity.

FIG. 21 shows mice that were exposed to iterative dosing with stabilizing drug. Four days of T cell "rest" were provided in which vehicle was injected instead of TMP. TMP injections were then resumed for an additional 3 days. The data show a profound rescue of function where tumor burden was lower after the "rest" period than it was before the "rest" period.

FIG. 22 shows dose response curves and EC₅₀ values of various regulatable DD-CARs. CAR Median Fluorescence Intensity (MFI) was calculated by first gating on CAR+ and CAR- cells, then subtracting the MFI of the CAR- population from that of the CAR+ population. Values were normalized to the max CAR MFI in the drug titration study, and a non-linear regression and least squares fit curve was generated. EC₅₀ values of each curve are plotted in the bar graphs. For GD2.28z.FKBP, HA-GD2.28z.FKBP, GD2.28z.ecDHFR, n=3 donors. For CD19.28z.FKBP, CD19.28z.ecDHFR, and HA-GD2.28z.ecDHFR, n=2 donors.

FIG. 23 shows that certain destabilization domains are more optimal for drugdependent regulation CAR expression. (A) T cells were transduced with lentivirus to express

GD2.28z.ecDHFR wherein the ecDHFR domain contained the following mutations: R12H, N18T, V19A, and G67S. T cells were removed from culture and stained with a fluorescent-conjugated anti-idiotype antibody, and CAR surface expression was subsequently assessed via FACS. Slight drug-dependent CAR regulation was observed due to high background expression in the absence of stabilizing drug. (B) T cells were transduced with lentivirus to express GD2.28z.ecDHFR wherein the ecDHFR domain contained the following mutations: R12Y, G67S, Y100I. T cells were again removed from culture, stained with anti-idiotype antibody and CAR expression was assessed via FACS. In this DD-CAR iteration incorporating a different ecDHFR domain, drug-dependent regulation of CAR surface expression was observed. The dotted lines in both subfigures were drawn to highlight the differences in CAR expression level between 0uM TMP and 10uM TMP.

FIG. 24 shows drug-dependent regulation of Her.28z CAR via a fused FKBP DD. The FKBP12 DD domain was fused to the far c-terminus of the Her2.28z CAR. T cells were transduced with lentivirus to express the Her2.28z.FKBP. On day 7 post-activation, CAR T cells were cultured in the presence or absence of shield-1. 24 hours after addition of drug, CAR T cells were stained with protein L and CAR expression was assessed via FACS. Both the percentage of CAR+ cells (quantified in the histograms) as well as CAR MFI are increased upon incubation with either 0.1uM or 1uM Shield-1 (S1).

FIG. 25 provides a table of nucleic acid and amino acid sequences useful in embodiments of the invention described herein.

DEFINITIONS

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For purposes of interpreting this specification, the following definitions will apply and whenever appropriate, terms used in the singular will also include the plural and vice versa. In the event that any definition set forth below conflicts with any document incorporated herein by reference, the definition set forth below shall control.

It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting.

The articles "a" and "an" are used herein to refer to one or to more than one (i.e., to at least one) of the grammatical object of the article. By way of example, "an element" means one element or more than one element.

"About" as used herein when referring to a measurable value such as an amount, a temporal duration, and the like, is meant to encompass variations of \pm 00% or \pm 10%, more preferably \pm 5%, even more preferably \pm 1%, and still more preferably \pm 10.1% from the

specified value, as such variations are appropriate to perform the disclosed methods.

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"T cell exhaustion" refers to decrease of T cell function, which may occur as a result of an infection (e.g., a chronic infection) or a disease. T cell exhaustion is associated with increased expression of PD-1, TIM-3, and LAG-3, apoptosis, and reduced cytokine secretion. Accordingly, the terms "ameliorate T cell exhaustion," "inhibit T cell exhaustion," "reduce T cell exhaustion" and the like refer to a condition of restored functionality of T cells characterized by one or more of the following: decreased expression and/or level of one or more of PD-1, TIM-3, and LAG-3; increased memory cell formation and/or maintenance of memory markers (e.g., CD62L); prevention of apoptosis; increased antigen-induced cytokine (e.g., IL-2) production and/or secretion; enhanced cytotoxicity/killing capacity; increased recognition of tumor targets with low surface antigen; enhanced proliferation in response to antigen.

As used herein, the terms "antigen-binding domain," as in "GD2-binding domain" may refer to any antigen-specific biding domain (e.g., GD2 specific binding domain) known to one of skilled in the art. In one example, a GD2 binding domain comprises a single-chain variable fragment (scFv) comprising the variable regions of the heavy (V_H) and light chains (V_L) of an antibody binding specifically to GD2. Anti-GD2 antibodies, antibody fragments, and their variants are well known in the art and include, for example, 14G2a, ch14.18, hu14.18K322A, m3F8, hu3F8-IgG1, hu3F8-IgG4, HM3F8, UNITUXIN, DMAb-20 or any other antibody that binds with specificity to GD2. In one embodiment, the GD2 binding domain is a homologue, a variant, an isomer, or a functional fragment of an anti-GD2 antibody. Each possibility represents a separate embodiment of the present invention.

"Activation," as used herein, refers to the state of a T cell that has been sufficiently stimulated to induce detectable cellular proliferation. Activation can also be associated with induced cytokine production, and detectable effector functions. The term "activated T cells" refers to, among other things, T cells that are undergoing cell division.

The term "antibody," as used herein, refers to an immunoglobulin molecule which specifically binds with an antigen. Antibodies can be intact immunoglobulins derived from natural sources or from recombinant sources and can be immunoreactive portions of intact immunoglobulins. Antibodies may be multimers of individual immunoglobulin molecules. The antibodies of the present invention may exist in a variety of forms including, for example, polyclonal antibodies, monoclonal antibodies, Fv, Fab and F(ab)₂, as well as single chain antibodies, human antibodies, and humanized antibodies (see Harlow et al., 1999, In: Using Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, NY; Harlow

et al., 1989, In: Antibodies: A Laboratory Manual, Cold Spring Harbor, N.Y.; Houston et al., 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; Bird et al., 1988, Science 242:423-426).

The term "antibody fragment" refers to a portion of an intact antibody and preferably refers to the antigenic determining variable regions of an intact antibody. Examples of antibody fragments include, but are not limited to, Fab, Fab', F(ab')₂, and Fv fragments, linear antibodies, scFv antibodies, and multispecific antibodies formed from antibody fragments.

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An "antibody heavy chain," as used herein, refers to the larger of the two types of polypeptide chains present in all antibody molecules in their naturally occurring conformations. An "antibody light chain," as used herein, refers to the smaller of the two types of polypeptide chains present in all antibody molecules in their naturally occurring conformations. Kappa and lamda light chains refer to the two major antibody light chain isotypes.

By the term "synthetic antibody" as used herein, is meant an antibody which is generated using recombinant DNA technology, such as, for example, an antibody expressed by a bacteriophage as described herein. The term should also be construed to mean an antibody which has been generated by the synthesis of a DNA molecule encoding the antibody and which DNA molecule expresses an antibody protein, or an amino acid sequence specifying the antibody, wherein the DNA or amino acid sequence has been obtained using synthetic DNA or amino acid sequence technology which is available and well known in the art.

The term "antigen" or "Ag" as used herein is defined as a molecule that provokes an immune response. This immune response may involve either antibody production, or the activation of specific immunologically-competent cells, or both. The skilled artisan will understand that any macromolecule, including virtually all proteins or peptides, can serve as an antigen. Furthermore, antigens can be derived from recombinant or genomic DNA. A skilled artisan will understand that any DNA, which comprises a nucleotide sequences or a partial nucleotide sequence encoding a protein that elicits an immune response therefore encodes an "antigen" as that term is used herein. Furthermore, one skilled in the art will understand that an antigen need not be encoded solely by a full length nucleotide sequence of a gene. Moreover, a skilled artisan will understand that an antigen need not be encoded by a "gene" at all. It is readily apparent that an antigen can be generated, synthesized or can be derived from a biological sample. Such a biological sample can include, but is not limited to a tissue sample, a tumor sample, a cell or a biological fluid.

The term "anti-tumor effect" as used herein, refers to a biological effect which can be

manifested by a decrease in tumor volume, a decrease in the number of tumor cells, a decrease in the number of metastases, an increase in life expectancy, or amelioration of various physiological symptoms associated with the cancerous condition. An "anti-tumor effect" can also be manifested by the ability of the peptides, polynucleotides, antibodies (or antigen-binding portions thereof), and CAR T cells of the invention in prevention of the occurrence of tumor in the first place.

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The term "autoimmune disease" as used herein is defined as a disorder that results from an autoimmune response. An autoimmune disease is the result of an inappropriate and excessive response to a self-antigen. Examples of autoimmune diseases include but are not limited to, Addision's disease, alopecia greata, ankylosing spondylitis, autoimmune hepatitis, autoimmune parotitis, Crohn's disease, diabetes (Type I), dystrophic epidermolysis bullosa, epididymitis, glomerulonephritis, Graves' disease, Guillain-Barr syndrome, Hashimoto's disease, hemolytic anemia, systemic lupus erythematosus, multiple sclerosis, myasthenia gravis, pemphigus vulgaris, psoriasis, rheumatic fever, rheumatoid arthritis, sarcoidosis, scleroderma, Sjogren's syndrome, spondyloarthropathies, thyroiditis, vasculitis, vitiligo, myxedema, pernicious anemia, ulcerative colitis, among others.

As used herein, the term "autologous" is meant to refer to any material derived from the same individual to which it is later to be re-introduced into the individual. "Allogeneic" refers to a graft derived from a different animal of the same species. "Xenogeneic" refers to a graft derived from an animal of a different species.

The term "cancer" as used herein is defined as disease characterized by the rapid and uncontrolled growth of aberrant cells. Cancer cells can spread locally or through the bloodstream and lymphatic system to other parts of the body. Examples of various cancers include but are not limited to, breast cancer, prostate cancer, ovarian cancer, cervical cancer, skin cancer, pancreatic cancer, colorectal cancer, renal cancer, liver cancer, brain cancer, lymphoma, leukemia, lung cancer and the like.

"Co-stimulatory ligand," as used herein, includes a molecule on an antigen presenting cell (APC) (e.g., dendritic cell, B cell, and the like) that specifically binds a cognate co-stimulatory molecule on a T cell, thereby providing a signal which, in addition to the primary signal provided by, for instance, binding of a TCR/CD3 complex with an MHC molecule loaded with peptide, mediates a T cell response, including, but not limited to, proliferation, activation, differentiation, and the like. A co-stimulatory ligand can include, but is not limited to, CD7, B7-1 (CD80), B7-2 (CD86), PD-L1, PD-L2, 4-1BBL, OX40L, inducible costimulatory ligand (ICOS-L), intercellular adhesion molecule (ICAM), CD30L, CD40,

CD70, CD83, HLA-G, MICA, MICB, HVEM, lymphotoxin beta receptor, 3/TR6, ILT3, ILT4, HVEM, an agonist or antibody that binds Toll ligand receptor and a ligand that specifically binds with B7-H3. A co-stimulatory ligand also encompasses, inter alia, an antibody that specifically binds with a co-stimulatory molecule present on a T cell, such as, but not limited to, CD27, CD28, 4-1BB, OX40, CD30, CD40, PD-1, ICOS, lymphocyte function-associated antigen-1 (LFA-1), CD2, CD7, LIGHT, NKG2C, B7-H3, and a ligand that specifically binds with CD83.

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A "co-stimulatory molecule" refers to the cognate binding partner on a T cell that specifically binds with a co-stimulatory ligand, thereby mediating a co-stimulatory response by the T cell, such as, but not limited to, proliferation. Co-stimulatory molecules include, but are not limited to an MHC class I molecule, BTLA and a Toll ligand receptor.

A "co-stimulatory signal", as used herein, refers to a signal, which in combination with a primary signal, such as TCR/CD3 ligation, leads to T cell proliferation and/or upregulation or downregulation of key molecules.

An "effective amount" as used herein, means an amount which provides a therapeutic or prophylactic benefit.

The term "therapeutic" as used herein means a treatment and/or prophylaxis. A therapeutic effect is obtained by suppression, remission, or eradication of a disease state.

The term "therapeutically effective amount" refers to the amount of the subject compound that will elicit the biological or medical response of a tissue, system, or subject that is being sought by the researcher, veterinarian, medical doctor or other clinician. The term "therapeutically effective amount" includes that amount of a compound that, when administered, is sufficient to prevent development of, or alleviate to some extent, one or more of the signs or symptoms of a disorder or disease being treated. The therapeutically effective amount will vary depending on the compound, the disease and its severity and the age, weight, etc., of the subject to be treated.

To "treat" a disease as the term is used herein, means to reduce the frequency or severity of at least one sign or symptom of a disease or disorder experienced by a subject.

"Encoding" refers to the inherent property of specific sequences of nucleotides in a polynucleotide, such as a gene, a cDNA, or an mRNA, to serve as templates for synthesis of other polymers and macromolecules in biological processes having either a defined sequence of nucleotides (e.g., rRNA, tRNA and mRNA) or a defined sequence of amino acids and the biological properties resulting therefrom. Thus, a gene encodes a protein if transcription and translation of mRNA corresponding to that gene produces the protein in a cell or other

biological system. Both the coding strand, the nucleotide sequence of which is identical to the mRNA sequence and is usually provided in sequence listings, and the non-coding strand, used as the template for transcription of a gene or cDNA, can be referred to as encoding the protein or other product of that gene or cDNA.

As used herein "endogenous" refers to any material from or produced inside an organism, cell, tissue or system. As used herein, the term "exogenous" refers to any material introduced from or produced outside an organism, cell, tissue or system.

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The term "expression" as used herein relates to the transcription and/or translation of a particular nucleotide sequence driven by its promoter. "Expression also refers to the presence of a protein on the surface of a cell (e.g., CAR expression on the surface of a T cell).

The term "transfected" or "transformed" or "transduced" as used herein refers to a process by which exogenous nucleic acid is transferred or introduced into a host cell. A "transfected" or "transformed" or "transduced" cell is one that has been transfected, transformed or transduced with exogenous nucleic acid. The cell includes the primary subject cell and its progeny.

A "vector" is a composition of matter which comprises an isolated nucleic acid and which can be used to deliver the isolated nucleic acid to the interior of a cell. Numerous vectors are known in the art including, but not limited to, linear polynucleotides, polynucleotides associated with ionic or amphiphilic compounds, plasmids, and viruses. Thus, the term "vector" includes an autonomously replicating plasmid or a virus. The term should also be construed to include non-plasmid and non-viral compounds which facilitate transfer of nucleic acid into cells, such as, for example, polylysine compounds, liposomes, and the like. Examples of viral vectors include, but are not limited to, adenoviral vectors, adeno-associated virus vectors, retroviral vectors, and the like.

"Expression vector" refers to a vector comprising a recombinant polynucleotide comprising expression control sequences operatively linked to a nucleotide sequence to be expressed. An expression vector comprises sufficient cis-acting elements for expression; other elements for expression can be supplied by the host cell or in an in vitro expression system. Expression vectors include all those known in the art, such as cosmids, plasmids (e.g., naked or contained in liposomes) and viruses (e.g., lentiviruses, retroviruses, adenoviruses, and adeno-associated viruses) that incorporate the recombinant polynucleotide.

A "lentivirus" as used herein refers to a genus of the Retroviridae family. Lentiviruses are unique among the retroviruses in being able to infect non-dividing cells. Lentivirus-derived vectors can deliver a significant amount of genetic information into the DNA of a

host cell, so they are one of the most efficient methods of a gene delivery vector. HIV, SIV, and FIV are all examples of lentiviruses.

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"Homologous" refers to the sequence similarity or sequence identity between two polypeptides or between two nucleic acid molecules. When a position in both of the two compared sequences is occupied by the same base or amino acid monomer subunit, e.g., if a position in each of two DNA molecules is occupied by adenine, then the molecules are homologous at that position. The percent of homology between two sequences is a function of the number of matching or homologous positions shared by the two sequences divided by the number of positions compared × 100. For example, if 6 of 10 of the positions in two sequences are matched or homologous then the two sequences are 60% homologous. By way of example, the DNA sequences ATTGCC and TATGGC share 50% homology. Generally, a comparison is made when two sequences are aligned to give maximum homology.

The term "immunoglobulin" or "Ig," as used herein is defined as a class of proteins, which function as antibodies. Antibodies expressed by B cells are sometimes referred to as the B cell receptor (BCR) or antigen receptor. The five members included in this class of proteins are IgA, IgG, IgM, IgD, and IgE. IgA is the primary antibody that is present in body secretions, such as saliva, tears, breast milk, gastrointestinal secretions and mucus secretions of the respiratory and genitourinary tracts. IgG is the most common circulating antibody. IgM is the main immunoglobulin produced in the primary immune response in most subjects. It is the most efficient immunoglobulin in agglutination, complement fixation, and other antibody responses, and is important in defense against bacteria and viruses. IgD is the immunoglobulin that has no known antibody function, but may serve as an antigen receptor. IgE is the immunoglobulin that mediates immediate hypersensitivity by causing release of mediators from mast cells and basophils upon exposure to allergen.

"Isolated" means altered or removed from the natural state. For example, a nucleic acid or a peptide naturally present in a living animal is not "isolated," but the same nucleic acid or peptide partially or completely separated from the coexisting materials of its natural state is "isolated." An isolated nucleic acid or protein can exist in substantially purified form, or can exist in a non-native environment such as, for example, a host cell.

As used herein, a "substantially purified" cell is a cell that is essentially free of other cell types. A substantially purified cell also refers to a cell which has been separated from other cell types with which it is normally associated in its naturally occurring state. In some instances, a population of substantially purified cells refers to a homogenous population of

cells. In other instances, this term refers simply to cell that have been separated from the cells with which they are naturally associated in their natural state. In some embodiments, the cells are cultured in vitro. In other embodiments, the cells are not cultured in vitro.

Unless otherwise specified, a "nucleotide sequence encoding an amino acid sequence" includes all nucleotide sequences that are degenerate versions of each other and that encode the same amino acid sequence. The phrase nucleotide sequence that encodes a protein or an RNA may also include introns to the extent that the nucleotide sequence encoding the protein may in some version contain an intron(s).

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The term "operably linked" refers to functional linkage between a regulatory sequence and a heterologous nucleic acid sequence resulting in expression of the latter. For example, a first nucleic acid sequence is operably linked with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Generally, operably linked DNA sequences are contiguous and, where necessary to join two protein coding regions, in the same reading frame.

The term "polynucleotide" as used herein is defined as a chain of nucleotides. Furthermore, nucleic acids are polymers of nucleotides. Thus, nucleic acids and polynucleotides as used herein are interchangeable. One skilled in the art has the general knowledge that nucleic acids are polynucleotides, which can be hydrolyzed into the monomeric "nucleotides." The monomeric nucleotides can be hydrolyzed into nucleosides. As used herein polynucleotides include, but are not limited to, all nucleic acid sequences which are obtained by any means available in the art, including, without limitation, recombinant means, i.e., the cloning of nucleic acid sequences from a recombinant library or a cell genome, using ordinary cloning technology and PCR, and the like, and by synthetic means.

As used herein, the terms "peptide," "polypeptide," and "protein" are used interchangeably, and refer to a compound comprised of amino acid residues covalently linked by peptide bonds. A protein or peptide must contain at least two amino acids, and no limitation is placed on the maximum number of amino acids that can comprise a protein's or peptide's sequence. Polypeptides include any peptide or protein comprising two or more amino acids joined to each other by peptide bonds. As used herein, the term refers to both short chains, which also commonly are referred to in the art as peptides, oligopeptides and oligomers, for example, and to longer chains, which generally are referred to in the art as

proteins, of which there are many types. "Polypeptides" include, for example, biologically active fragments, substantially homologous polypeptides, oligopeptides, homodimers, heterodimers, variants of polypeptides, modified polypeptides, derivatives, analogs, fusion proteins, among others. The polypeptides include natural peptides, recombinant peptides, synthetic peptides, or a combination thereof.

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The term "promoter" as used herein is defined as a DNA sequence recognized by the synthetic machinery of the cell, or introduced synthetic machinery, required to initiate the specific transcription of a polynucleotide sequence. As used herein, the term "promoter/regulatory sequence" means a nucleic acid sequence required for expression of a gene product operably linked to the promoter/regulatory sequence. In some instances, this sequence may be the core promoter sequence and in other instances, this sequence may also include an enhancer sequence and other regulatory elements required for expression of the gene product. The promoter/regulatory sequence may, for example, be one that expresses the gene product in a tissue specific manner. A "constitutive" promoter is a nucleotide sequence which, when operably linked with a polynucleotide encoding a gene product, causes the gene product to be produced in a cell under most or all physiological conditions of the cell. An "inducible" promoter is a nucleotide sequence which, when operably linked with a polynucleotide which encodes or specifies a gene product, causes the gene product to be produced in a cell substantially only when an inducer which corresponds to the promoter is present in the cell. A "tissue-specific" promoter is a nucleotide sequence which, when operably linked with a polynucleotide encoding a gene, causes the gene product to be produced in a cell substantially only if the cell is a cell of the tissue type corresponding to the promoter.

The term "overexpressed" tumor antigen or "overexpression" of the tumor antigen is intended to indicate an abnormal level of expression of the tumor antigen in a cell from a disease area like a solid tumor within a specific tissue or organ of the patient relative to the level of expression in a normal cell from that tissue or organ. Patients having solid tumors or a hematological malignancy characterized by overexpression of the tumor antigen can be determined by standard assays known in the art.

By the term "modulating," as used herein, refers to mediating a detectable increase or decrease in the level of an activity in a subject (e.g., a response in a subject or the expression of a protein in a cell). Modulating the level of activity can occur in the presence of a treatment or compound, or in the absence of the treatment or compound. By way of example, to modulate the level of chimeric antigen receptor (CAR) expression on the surface of a CAR

T cell genetically modified with a CAR containing a regulatable destabilization domain (RDD) means to increase CAR expression via exposure of the CAR T cell to a small molecule/drug that stabilizes the RDD compared with the level of CAR expression in the absence of the small molecule/drug. Similarly, it is possible to modulate the level of chimeric antigen receptor (CAR) expression on the surface of a CAR T cell genetically modified with a CAR containing a regulatable destabilization domain (RDD) by decreasing CAR expression via removal of the stabilizing small molecule/drug that stabilizes the RDD compared with the level of CAR expression in the presence of the small molecule/drug. The term encompasses perturbing and/or affecting a native signal or response or non-modulated expression thereby mediating a beneficial therapeutic response in a subject, preferably, a human.

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"Parenteral" administration of an immunogenic composition includes, e.g., subcutaneous (s.c.), intravenous (i.v.), intramuscular (i.m.), or intrasternal injection, or infusion techniques.

The terms "patient," "subject," "individual," and the like are used interchangeably herein and are intended to include living organisms in which an immune response can be elicited (e.g., mammals). Examples of subjects include humans, dogs, cats, mice, rats, and transgenic species thereof.

By the term "specifically binds," as used herein with respect to an antibody, is meant an antibody which recognizes a specific antigen, but does not substantially recognize or bind other molecules in a sample. For example, an antibody that specifically binds to an antigen from one species may also bind to that antigen from one or more species. But, such cross-species reactivity does not itself alter the classification of an antibody as specific. In another example, an antibody that specifically binds to an antigen may also bind to different allelic forms of the antigen. However, such cross reactivity does not itself alter the classification of an antibody as specific. In some instances, the terms "specific binding" or "specifically binding," can be used in reference to the interaction of an antibody, a protein, or a peptide with a second chemical species, to mean that the interaction is dependent upon the presence of a particular structure (e.g., an antigenic determinant or epitope) on the chemical species; for example, an antibody recognizes and binds to a specific protein structure rather than to proteins generally. If an antibody is specific for epitope "A", the presence of a molecule containing epitope A (or free, unlabeled A), in a reaction containing labeled "A" and the antibody, will reduce the amount of labeled A bound to the antibody.

By the term "stimulation," is meant a primary response induced by binding of a stimulatory molecule (e.g., a TCR/CD3 complex) with its cognate ligand thereby mediating a

signal transduction event, such as, but not limited to, signal transduction via the TCR/CD3 complex. Stimulation can mediate altered expression of certain molecules, such as downregulation of TGF- β , enhanced expression of IL-2 and/or IFN- γ , and/or reorganization of cytoskeletal structures, and the like.

A "stimulatory molecule," as the term is used herein, means a molecule on a T cell that specifically binds with a cognate stimulatory ligand present on an antigen presenting cell.

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A "stimulatory ligand," as used herein, means a ligand that when present on an antigen presenting cell (e.g., a dendritic cell, a B-cell, and the like) can specifically bind with a cognate binding partner (referred to herein as a "stimulatory molecule") on a T cell, thereby mediating a primary response by the T cell, including, but not limited to, activation, initiation of an immune response, proliferation, and the like. Stimulatory ligands are well-known in the art and encompass, inter alia, an MHC Class I molecule loaded with a peptide, an anti-CD3 antibody, a superagonist anti-CD28 antibody, and a superagonist anti-CD2 antibody.

The terms "sensitize" and "sensitizing," as used herein, refer to making, through the administration of a first agent, an animal or a cell within an animal more susceptible, or more responsive, to the biological effects (e.g., promotion or retardation of an aspect of cellular function including, but not limited to, cell division, cell growth, proliferation, invasion, angiogenesis, necrosis, or apoptosis) of a second agent. The sensitizing effect of a first agent on a target cell can be measured as the difference in the intended biological effect (e.g., promotion or retardation of an aspect of cellular function including, but not limited to, cell growth, proliferation, invasion, angiogenesis, or apoptosis) observed upon the administration of a second agent with and without administration of the first agent. The response of the sensitized cell can be increased by at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 150%, at least about 200%, at least about 400%, at least about 500% over the response in the absence of the first agent.

As used herein, the term "subject suspected of having cancer" refers to a subject that presents one or more symptoms indicative of a cancer (e.g., a noticeable lump or mass) or is being screened for a cancer (e.g., during a routine physical). A subject suspected of having cancer may also have one or more risk factors for developing cancer. A subject suspected of having cancer has generally not been tested for cancer. However, a "subject suspected of having cancer" encompasses an individual who has received a preliminary diagnosis (e.g., a

CT scan showing a mass) but for whom a confirmatory test (e.g., biopsy and/or histology) has not been done or for whom the type and/or stage of cancer is not known. The term further includes people who previously had cancer (e.g., an individual in remission). A "subject suspected of having cancer" is sometimes diagnosed with cancer and is sometimes found to not have cancer.

As used herein, the term "subject diagnosed with a cancer" refers to a subject who has been tested and found to have cancerous cells. The cancer may be diagnosed using any suitable method, including but not limited to, biopsy, x-ray, blood test, etc.

As used herein, the term "subject at risk for cancer" refers to a subject with one or more risk factors for developing a specific cancer. Risk factors include, but are not limited to, gender, age, genetic predisposition, environmental exposure, and previous incidents of cancer, preexisting non-cancer diseases, and lifestyle.

As used herein, the term "characterizing cancer in a subject" refers to the identification of one or more properties of a cancer sample in a subject, including but not limited to, the presence of benign, pre-cancerous or cancerous tissue and the stage of the cancer.

DETAILED DESCRIPTION

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The invention is based on the discovery that transient inhibition or modulation of TCR signaling, via modulation of chimeric antigen receptor (CAR) surface expression in CAR T cells, can prevent or reverse CAR T cell exhaustion and restore CAR T cell function. T cells that express CARs undergo tonic, antigen-independent signaling due to receptor clustering and replicate the fundamental biology of T cell exhaustion, as shown by high levels of PD-1, TIM-3, and LAG-3 expression, diminished antigen-induced cytokine production and excessive programmed cell death. A GD2.28z CAR fused to an FKBP12 destabilization domain (DD) (GD2.28z.FKBP) was generated and the regulatable DD (RDD) conferred instability to the CAR and induced rapid CAR protein degradation (see, Banaszynski et. al, Cell 2006). Surface expression was observed to be rapidly and dosedependently regulated by adding or subtracting trimethoprim or the stabilizing rapalog shield-1 (S1) in culture medium. Similar regulatability of CAR expression was also accomplished using an *E. coli* DHFR mutant fused to the CAR (GD2.28z.DHFR), which was regulated in a dose-dependent manner with trimethoprim (see Examples 1 and 2, FIGS. 2, 12-13, 22-24).

Because tonic signaling is highly dependent upon CAR receptor levels in CAR T cells, control of CAR expression levels can be used to regulate the level of tonic signaling

(e.g., in vitro or in vivo). Since tonic signaling is highly dependent upon CAR receptor levels, precise control of CAR expression levels also precisely regulated levels of tonic signaling. Drug regulated control of levels of CAR expression therefore allowed modulation of the duration and intensity of GD2.28z CAR tonic signaling. Using this system, phenotypic and functional changes associated with exhaustion were reversed upon cessation of CAR signaling (see Examples 1 and 2). Removal of the small molecule or drug (S1 or trimethoprim) from the culture medium and consequent removal of surface CAR on day 7 post-activation reversed canonical exhaustion marker expression to control levels by day 10 (see Example 1, FIG. 2). This was illustrated by measuring levels of PD-1/TIM-3/LAG-3 triple expressing cell which is highly specific for dysfunctional, exhausted T cells. By Day 10, increases in levels of triple expressing exhausted cells exist, but that removal of S1 on Day 7 resulted in normalization of these levels by Day 10. Similar results were obtained on day 14 (see Example 1).

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Thus, in some embodiments, the invention provides the ability to control, with precision, CAR T cell signaling and/or activity via modulation of surface expression of CAR. For example, as detailed herein (see Example 2), the invention provides the ability to titrate down the dose of stabilizing drug, resulting in a rapid decrease of CAR expression, which in turn inhibits and/or eliminates CAR T cell signaling and/or activity.

Thus, in some embodiments, regulatable DD-CAR technology can be used to address several important challenges currently facing the field of CAR T cell therapy. First, the rapid on/off switching of DD-CAR surface expression in response to drug allows for precise regulation of CAR activity, and thus provides an opportunity to mitigate CAR T cell toxicity (e.g., cytokine release syndrome (CRS) or on-target off-tumor toxicity) while preserving the option to continue therapy once the toxicity has resolved (See Example 2). Second, expansion of CAR T cells in the absence of stabilizing drug prevents CAR tonic signaling and in turn enhances the functional capacity of these cells (See Example 2). Last, toggling DD-CAR expression *in vivo* via iterative drug dosing may be one method by which CAR T cell exhaustion is prevented or reversed and/or memory could be induced (See Example 2).

Accordingly, the invention provides CAR T cells modified to express a CAR fused to an RDD) (e.g., a FKBP RDD or a DHFR RDD) that provides control, in a dose- and time-dependent manner, over CAR expression on the surface of CAR T cells. The ability to control CAR expression on the surface of CAR T cells in turn permits the ability to maintain functionality of the CAR T cells under conditions in which unmodified CAR T cells (that is, CAR T cells lacking a CAR fused to an RDD domain) display T cell exhaustion or lack of

functionality. In this way, compositions and methods of the invention find use in preventing exhaustion of CAR T cells and enhancement of CAR T cell functionality (e.g., activity against cancer or infectious disease).

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The invention is not limited by the type of CAR T cells functionality maintained, regained or enhanced (e.g., prolonged). Indeed, compositions comprising CAR T cells modified to express a CAR fused to an RDD domain and methods utilizing the same may be used to maintain, regain, or enhance CAR T cell functionality including, but not limited to, cytotoxic activities against tumor cells; promotion of CAR T cell survival and function; induction of cytokine expression such as expression of interleukin-2 (IL-2) to promote T cell survival, expression of Fas Ligand (FasL) and/or tumor necrosis factor-related apoptosis inducing ligand (TRAIL) to induce tumor cell apoptosis, and/or to induce interferon (IFN)-gamma to activate innate immune responses (e.g., against cancer); and/or potentiate the induction of cell cycle arrest and/or apoptosis. In some embodiments, CAR T cells of the invention sensitize cancer cells to induction of cell cycle arrest and/or apoptosis, including cells that are normally resistant to such inducing stimuli.

Modification of CAR T cells to express a CAR fused to an RDD domain (e.g., mutant FKBP or DHFR) significantly enhanced functionality of CAR T cells exposed to conditions that induce CAR T cell exhaustion (See, e.g., Examples 1 and 2). While an understanding of a mechanism is not needed to practice the present invention, and while the present invention is not limited to any particular mechanism of action, reduction of expression of CAR on the CAR T cell surface prevents dysfunction of the CAR T cells associated with CAR T cell exhaustion. In some embodiments, use of a T cell modified to express a CAR comprising an RDD of the invention prolongs the anti-tumor response of the CAR T cell compared to CAR T cell expressing the same CAR but without the RDD.

Thus, the invention provides compositions and methods for reducing CAR T cell exhaustion comprising CAR T cells modified to express a CAR fused to an RDD domain (e.g., FKBP12 or mutant DHFR) thereby providing the ability to strictly regulate the level of CAR expression on the CAR T cell surface. The invention is not limited by the disease or condition that can be treated using modified CAR T cells of the invention. Indeed, the compositions and methods provided herein may be useful in the treatment of any disease for which increased activity of CAR T cells may provide a therapeutic benefit.

Accordingly, the invention provides a composition comprising CAR T cells modified to express a CAR fused to an RDD domain. The invention is not limited by the type of modified CAR T cell. In some embodiments, the CAR T cells are CD3+ T cells (e.g., CD4+

and/or CD8+ T cells). In certain embodiments, the CAR T cells are CD8+ T cells. In other embodiments, the CAR T cells are CD4+ T cells. In some embodiments, the CAR T cells are natural killer (NK) T cells. In some embodiments, the CAR T cells are gamma delta T cells. In some embodiments, the CAR T cells are alpha beta T cells. In some embodiments, the CAR T cells are a combination of CD4+ and CD8 T+ cells (e.g., that are CD3+). In certain embodiments, the CAR T cells are memory T cells. In certain embodiments, the CAR T cells are a combination of CD8+ T cells, CD4+ T cells, NK T cells, memory T cells, alpha beta T cells and/or gamma delta T cells. In some embodiments, the CAR T cells are cytokine-induced killer T cells. In some embodiments, the CAR T cells are tumor infiltrating lymphocytes. In some embodiments, the CAR T cells are engineered to express a tumor antigen specific CAR. In another embodiment, the CAR T cells are engineered to express a CAR specific for an infectious disease antigen. In some embodiments, the CAR T cells are anti-tumor T cells. In some embodiments, the composition further comprises a pharmaceutically acceptable carrier (e.g., buffer).

Thus, in one aspect, the present invention provides compositions and methods for treating cancer, among other diseases. The cancer may be a hematological malignancy, a solid tumor, a primary or a metastasizing tumor. Other diseases treatable using the compositions and methods of the invention include viral, bacterial and parasitic infections as well as autoimmune diseases.

In some embodiments, the invention provides a T cell engineered to express a CAR wherein the CAR T cell exhibits an antitumor property. The CAR of the invention can be engineered to comprise an extracellular domain having an antigen-binding domain fused to an intracellular signaling domain of the CD3 zeta chain. The CAR of the invention when expressed in a T cell is able to redirect antigen recognition based on the antigen-binding specificity. An exemplary antigen is GD2 as this antigen is expressed on a variety of cancers including neuroblastoma, osteosarcomas and some sarcomas (see, e.g., Thomas et al., PLoS One, 2016. 11(3): p. e0152196; Long et al., Nature Medicine, 2015. 21(6): p. 581-590; Long et al., Cancer Immunology Research, 2016. 4(10): p. 869-880; Yu et al., N Engl J Med, 2010. 363(14): p. 1324-34; Perez Horta et al., Immunotherapy, 2016. 8(9): p. 1097-117; Heczey et al, Molecular Therapy). However, the invention is not limited to targeting GD2. Rather, the invention includes any antigen-binding moiety that when bound to its cognate antigen, affects a tumor cell so that the tumor cell fails to grow, is prompted to die, or otherwise is affected so that the tumor burden in a patient is diminished or eliminated. The antigen-binding moiety is

preferably fused with an intracellular domain from one or more of a costimulatory molecule, a CD3 zeta chain, and an RDD.

The present invention provides chimeric antigen receptor (CAR) comprising an extracellular and intracellular domain. In some embodiments, the CAR of the invention is fully human. The extracellular domain comprises a target-specific binding element otherwise referred to as an antigen-binding moiety. The intracellular domain or otherwise the cytoplasmic domain comprises a costimulatory signaling region and a zeta chain portion. The costimulatory signaling region refers to a portion of the CAR comprising the intracellular domain of a costimulatory molecule. Costimulatory molecules are cell surface molecules other than antigens receptors or their ligands that are required for an efficient response of lymphocytes to antigen.

Between the extracellular domain and the transmembrane domain of the CAR, or between the cytoplasmic domain and the transmembrane domain of the CAR, there may be incorporated a spacer domain. As used herein, the term "spacer domain" generally means any oligo- or polypeptide that functions to link domains (e.g., the transmembrane domain to either the extracellular domain or the cytoplasmic domain in the polypeptide chain). A spacer domain may comprise up to 300 amino acids, preferably 10 to 100 amino acids and most preferably 25 to 50 amino acids.

20 Chimeric Antigen Receptor Constructs

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As described herein, a CAR T cell of the invention genetically modified to express a CAR fused to an RDD may contain a CAR that comprises a target-specific binding element otherwise referred to as an antigen-binding moiety. The choice of moiety depends upon the type and number of ligands that define the surface of a target cell. For example, the antigen-binding domain may be chosen to recognize a ligand that acts as a cell surface marker on target cells associated with a particular disease state. Examples of cell surface markers that may act as ligands for the antigen moiety domain of the CAR include those associated with viral, bacterial and parasitic infections, autoimmune disease and cancer cells.

As used herein, the term "antigen" refers to a molecule capable of being bound by an antibody or T-cell receptor. An antigen is additionally capable of inducing a humoral immune response and/or cellular immune response leading to the production of B and/or T lymphocytes. Embodiments of the present invention involve nucleic acids, including nucleic acids encoding an antigen-specific chimeric antigen receptor (CAR) polypeptide, including a CAR that has been humanized to reduce immunogenicity (hCAR), a transmembrane domain,

one or more intracellular signaling domains (e.g., a costimulatory domain and/or CD3 zeta), and an RDD domain. It is contemplated that human CAR nucleic acids are human genes that enhance cellular immunotherapy for human patients. In certain embodiments, the CAR recognizes an epitope comprised of the shared space between one or more antigens. In some embodiments, the CAR is specific for a carbohydrate antigen.

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Non-human antigen-binding regions have typically been used in constructing chimeric antigen receptors. However, a potential problem with using non-human antigen-binding regions, such as murine monoclonal antibodies, is the lack of human effector functionality. Furthermore, non-human monoclonal antibodies can be recognized by the human host as a foreign protein, and therefore, repeated injections of such foreign antibodies can lead to the induction of immune responses leading to harmful hypersensitivity reactions. For murine-based monoclonal antibodies, this is often referred to as a Human Anti-Mouse Antibody (HAMA) response. Therefore, in some embodiments, antigen-binding regions of human antibodies are used (e.g., because they do not elicit strong a HAMA response). In some embodiments, the antigen-binding domains may be chimeric (i.e., human/mouse chimeras) or humanized to minimize such immunogenicity. However, in other embodiments, antigen-binding regions of murine antibodies are used.

In some embodiments, the CAR comprises: a) an extracellular domain comprising an antigen-binding region, b) a transmembrane domain, c) an intracellular signaling domain, and d) an RDD. For example, in some embodiments, the CAR comprises an anti-GD2 antigenbinding domain, a CD28 transmembrane domain, CD3-zeta, CD28 and/or 4-1BB stimulatory domain, and an RDD (e.g., a DHFR or FKBP DD). As described in detail herein, many different CARs containing an RDD and methods of using the same are made possible. In one embodiment, the CAR comprises a) an extracellular domain comprising an antigenbinding region comprising SEQ ID NO: 44, b) a transmembrane domain comprising SEQ ID NO: 48, c) an intracellular signaling domain comprising SEQ ID NO: 50 and/or SEQ ID NO: 52, and d) an RDD comprising SEQ ID NO: 54. In another embodiment, a CAR comprises a) an extracellular domain comprising an antigen-binding region comprising SEQ ID NO: 60, b) a transmembrane domain comprising SEQ ID NO: 64, c) an intracellular signaling domain comprising SEQ ID NO: 66 and/or SEQ ID NO: 68, and d) an RDD comprising SEQ ID NO: 70. In another embodiment, a CAR comprises a) an extracellular domain comprising an antigen-binding region comprising SEQ ID NO: 76, b) a transmembrane domain comprising SEQ ID NO: 80, c) an intracellular signaling domain comprising SEQ ID NO: 82 and/or SEQ ID NO: 84, and d) an RDD comprising SEQ ID NO: 86. In still another embodiment, a CAR

comprises a) an extracellular domain comprising an antigen-binding region comprising SEQ ID NO: 92, b) a transmembrane domain comprising SEQ ID NO: 96, c) an intracellular signaling domain comprising SEQ ID NO: 98 and/or SEQ ID NO: 100, and d) an RDD comprising SEQ ID NO: 102. In another embodiment, a CAR comprises a) an extracellular domain comprising an antigen-binding region comprising SEQ ID NO: 108, b) a transmembrane domain comprising SEQ ID NO: 112, c) an intracellular signaling domain comprising SEQ ID NO: 114 and/or SEQ ID NO: 116, and d) an RDD comprising SEQ ID NO: 118. In another embodiment, a CAR comprises a) an extracellular domain comprising an antigen-binding region comprising SEQ ID NO: 124, b) a transmembrane domain comprising SEQ ID NO: 130, c) an intracellular signaling domain comprising SEQ ID NO: 132 and/or SEQ ID NO: 134, and d) an RDD comprising SEQ ID NO: 136.

Antigen-binding Region

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In certain embodiments, the binding region comprises complementary-determining regions (CDRs) of a monoclonal antibody, variable regions of a monoclonal antibody, and/or antigen-binding fragments thereof. A CDR is a short amino acid sequence found in the variable domains of antigen receptor (e.g., immunoglobulin and T-cell receptor) proteins that complements an antigen and therefore provides the receptor with its specificity for that particular antigen. Each polypeptide chain of an antigen receptor contains three CDRs (CDR1, CDR2, and CDR3). Since the antigen receptors are typically composed of two polypeptide chains, there are six CDRs for each antigen receptor (e.g., antibody or TCR) that can come into contact with the antigen--each heavy and light chain contains three CDRs. Because most sequence variation associated with immunoglobulins and T-cell receptors are found in the CDRs, these regions are sometimes referred to as hypervariable domains.

A CAR may be engineered to target a tumor antigen of interest by way of engineering a desired antigen-binding moiety that specifically binds to an antigen on a tumor cell. As used herein, a "tumor antigen" or "hyperproliferative disorder antigen" or "antigen associated with a hyperproliferative disorder" or "cancer antigen," refers to antigens that are common to specific hyperproliferative disorders such as cancer. Exemplary antigens mentioned herein are included by way of example. The examples are not intended to be exclusive and further examples will be readily apparent to those of skill in the art.

Tumor antigens are moieties (e.g., proteins, carbohydrates, etc.) that are produced by tumor cells that elicit an immune response, particularly T-cell mediated immune responses. Thus, an antigen-binding moiety can be selected based on the particular type of cancer to be

treated. Tumor antigens are well known in the art and include, for example, a glioma-associated antigen, carcinoembryonic antigen (CEA), beta-human chorionic gonadotropin, alphafetoprotein (AFP), lectin-reactive AFP, thyroglobulin, RAGE-1, MN-CA IX, human telomerase reverse transcriptase, RU1, RU2 (AS), intestinal carboxyl esterase, mut hsp70-2, M-CSF, prostate-specific antigen (PSA), PAP, NY-ESO-1, LAGE-1a, p53, prostein, PSMA, Her2/neu, survivin and telomerase, prostate-carcinoma tumor antigen-1 (PCTA-1), MAGE, ELF2M, neutrophil elastase, ephrinB2, CD22, insulin growth factor (IGF)-I, IGF-II, IGF-I receptor, mesothelin, and other tumor antigens described herein or known in the art.

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A tumor antigen may comprise one or more antigenic cancer antigens/epitopes associated with a malignant tumor. Malignant tumors express a number of proteins that can serve as target antigens for an immune attack. These molecules include but are not limited to tissue-specific antigens such as melanoma antigen recognized by T cells 1 (MART-1), tyrosinase and GP100 in melanoma and prostatic acid phosphatase (PAP) and prostate-specific antigen (PSA) in prostate cancer. Other target molecules belong to the group of transformation-related molecules such as the oncogene HER-2/Neu/ErbB-2. Still another group of target antigens are onco-fetal antigens such as carcinoembryonic antigen (CEA). In B-cell lymphoma the tumor-specific idiotype immunoglobulin constitutes a truly tumor-specific immunoglobulin antigen that is unique to the individual tumor. B-cell differentiation antigens such as CD19, CD20 and CD37 are other candidates for target antigens in B-cell lymphoma.

The tumor antigen may also be a tumor-specific antigen (TSA) or a tumor-associated antigen (TAA). A TSA is unique to tumor cells and does not occur on other cells in the body. A TAA is not unique to a tumor cell and instead is also expressed on some normal cells under conditions that fail to induce a state of immunologic tolerance to the antigen. The expression of the antigen on the tumor may occur under conditions that enable the immune system to respond to the antigen. TAAs may be antigens that are expressed on normal cells during fetal development when the immune system is immature and unable to respond or they may be antigens that are normally present at extremely low levels on normal cells but which are expressed at much higher levels on tumor cells.

Examples of TSA or TAA include, but are not limited to, differentiation antigens such as MART-1/MelanA (MART-1), gp100 (Pmel 17), tyrosinase, TRP-1, TRP-2 and tumor-specific multilineage antigens such as MAGE-1, MAGE-3, BAGE, GAGE-1, GAGE-2, p15; overexpressed embryonic antigens such as CEA; overexpressed oncogenes and mutated tumor-suppressor genes such as p53, Ras, HER-2/neu; unique tumor antigens resulting from

chromosomal translocations; such as BCR-ABL, E2A-PRL, H4-RET, IGH-IGK, MYL-RAR; and viral antigens, such as the Epstein Barr virus antigens EBVA and the human papillomavirus (HPV) antigens E6 and E7. Other large, protein-based antigens include TSP-180, MAGE-4, MAGE-5, MAGE-6, RAGE, NY-ESO-1, p185erbB2, p180erbB-3, c-met, nm-23H1, PSA, TAG-72, CA 19-9, CA 72-4, CAM 17.1, NuMa, K-ras, beta-Catenin, CDK4, Mum-1, p 15, p 16, 43-9F, 5T4, 791Tgp72, alpha-fetoprotein, beta-HCG, BCA225, BTAA, CA 125, CA 15-3\CA 27.291\BCAA, CA 195, CA 242, CA-50, CAM43, CD68\P1, CO-029, FGF-5, G250, Ga733\EpCAM, HTgp-175, M344, MA-50, MG7-Ag, MOV18, NB/70K, NY-CO-1, RCAS1, SDCCAG16, TA-90\Mac-2 binding protein\cyclophilin C-associated protein, TAAL6, TAG72, TLP, and TPS.

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In some embodiments, intracellular tumor associated antigens are targeted, such as HA-1, survivin, WT1, and p53. This can be achieved by a CAR expressed on a T cell that recognizes the processed peptide derived from the intracellular tumor associated antigen in the context of HLA.

Depending on the desired antigen to be targeted, a CAR can be engineered to include the appropriate antigen-binding moiety specific to the desired antigen target. For example, if GD2 is the desired antigen that is to be targeted, an antibody for GD2 (e.g., 14G2a, ch14.18, hu14.18K322A, m3F8, hu3F8-IgG1, hu3F8-IgG4, HM3F8, UNITUXIN, DMAb-20 or any other antibody that binds with specificity to GD2) can be used as the antigen-binding moiety for incorporation into the CAR of the invention.

In some embodiments, the antigen is a pathogen-specific antigen. The invention is not limited by the type of pathogen-specific antigen. A pathogen-specific antigen may be from any pathogen including, but not limited to, a fungus, bacteria, or virus. Exemplary viral pathogens include those of the families of Adenoviridae, EpsteinBarr virus (EBV),

- 25 Cytomegalovirus (CMV), Respiratory Syncytial Virus (RSV), JC virus, BK virus, HSV, HHV family of viruses, Picornaviridae, Herpesviridae, Hepadnaviridae, Flaviviridae, Retroviridae, Orthomyxoviridae, Paramyxoviridae, Papovaviridae, Polyomavirus, Rhabdoviridae, and Togaviridae. Exemplary pathogenic viruses cause smallpox, influenza, mumps, measles, chickenpox, ebola, and rubella. Exemplary pathogenic fungi include
- Candida, Aspergillus, Cryptococcus, Histoplasma, Pneumocystis, and Stachybotrys. Exemplary pathogenic bacteria include Streptococcus, Pseudomonas, Shigella, Campylobacter, Staphylococcus, Helicobacter, E. coli, Rickettsia, Bacillus, Bordetella, Chlamydia, Spirochetes, and Salmonella. In one embodiment the pathogen receptor Dectin-1 is used to generate a CAR that recognizes the carbohydrate structure on the cell wall of fungi.

T cells genetically modified to express a CAR based on the specificity of Dectin-1 recognize Aspergillus and target (e.g., inhibit) hyphal growth. In another embodiment, CARs are made based on an antibody recognizing viral determinants (e.g., the glycoproteins from CMV or Ebola) to interrupt viral infection and pathology.

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In some embodiments, the invention includes a full length CAR cDNA or coding region. The antigen-binding regions or domain can comprise a fragment of the $V_{\rm H}$ and $V_{\rm L}$ chains of a single-chain variable fragment (scFv) derived from a particular human monoclonal antibody, such as those described in U.S. Pat. Nos. 7,109,304; 8,822,196; 9,868,774; 9,790,282; 9,765,342; 9,624,306; 9,522,915; 9,359,447; 9,845,362; 9,815,901; 9,777,061; 9,598,489; 9,394,368; 9,446,105; and 9,334,330, each of which is incorporated herein by reference. The fragment can also be any number of different antigen-binding domains of a human antigen-specific antibody. In a more specific embodiment, the fragment is an antigen-specific scFv encoded by a sequence that is optimized for human codon usage for expression in human cells.

The antibody can be multimeric, such as a diabody or multimers. The multimers can be formed by cross pairing of the variable portion of the light and heavy chains into what has been referred to as a diabody (see Winter et al., Protein Eng. 1996 Mar;9(3):299-305). The hinge portion of the construct can have multiple alternatives from being totally deleted, to having the first cysteine maintained, to a proline rather than a serine substitution, to being truncated up to the first cysteine. The Fc portion can be deleted. Any protein that is stable and/or dimerizes can serve this purpose (e.g., a single one of the Fc domains, e.g., either the CH2 or CH3 domain from human immunoglobulin, can be used). The hinge, CH2 and CH3 region of a human immunoglobulin that has been modified to improve dimerization can also be used.

The antigen-binding domain of a CAR can be any domain that binds to antigen including but not limited to monoclonal antibodies, polyclonal antibodies, synthetic antibodies, human antibodies, humanized antibodies, and fragments thereof. In some instances, it is beneficial for the antigen-binding domain to be derived from the same species in which the CAR will ultimately be used in. For example, for use in humans, it may be beneficial for the antigen-binding domain of the CAR to comprise a human antibody or fragment thereof. Thus, in one embodiment, the antigen biding domain portion comprises a human antibody or a fragment thereof.

For *in vivo* use of antibodies in humans, it may be preferable to use human antibodies. Completely human antibodies are particularly desirable for therapeutic treatment of human

subjects. Human antibodies can be made by a variety of methods known in the art including phage display methods using antibody libraries derived from human immunoglobulin sequences, including improvements to these techniques. See, also, U.S. Pat. Nos. 4,444,887 and 4,716,111; and PCT publications WO 98/46645, WO 98/50433, WO 98/24893, WO98/16654, WO 96/34096, WO 96/33735, and WO 91/10741; each of which is incorporated herein by reference in its entirety. A human antibody can also be an antibody wherein the heavy and light chains are encoded by a nucleotide sequence derived from one or more sources of human DNA.

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Human antibodies can also be produced using transgenic mice which are incapable of expressing functional endogenous immunoglobulins, but which can express human immunoglobulin genes. For example, the human heavy and light chain immunoglobulin gene complexes may be introduced randomly or by homologous recombination into mouse embryonic stem cells. Alternatively, the human variable region, constant region, and diversity region may be introduced into mouse embryonic stem cells in addition to the human heavy and light chain genes. The mouse heavy and light chain immunoglobulin genes may be rendered non-functional separately or simultaneously with the introduction of human immunoglobulin loci by homologous recombination. For example, it has been described that the homozygous deletion of the antibody heavy chain joining region (JH) gene in chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production. The modified embryonic stem cells are expanded and microinjected into blastocysts to produce chimeric mice. The chimeric mice are then bred to produce homozygous offspring which express human antibodies. The transgenic mice are immunized in the normal fashion with a selected antigen. Anti-tumor antigen antibodies directed against the tumor antigen can be obtained from the immunized, transgenic mice using conventional hybridoma technology. The human immunoglobulin transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and somatic mutation. For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, e.g., PCT Publication Nos. WO 98/24893, WO 96/34096, and WO 96/33735; and U.S. Pat. Nos. 5,413,923; 5,625,126; 5,633,425; 5,569,825; 5,661,016; 5,545,806; 5,814,318; and 5,939,598, each of which is incorporated by reference herein in their entirety. In addition, companies such as Abgenix, Inc. (Freemont, Calif.) and Genpharm (San Jose, Calif.) can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above. For a specific discussion of transfer of a human germ-line

immunoglobulin gene array in germ-line mutant mice that will result in the production of human antibodies upon antigen challenge see, e.g., Jakobovits et al., Proc. Natl. Acad. Sci. USA, 90:2551 (1993); Jakobovits et al., Nature, 362:255-258 (1993); Bruggermann et al., Year in Immunol., 7:33 (1993); and Duchosal et al., Nature, 355:258 (1992).

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Human antibodies can also be derived from phage-display libraries (Hoogenboom et al., J. Mol. Biol., 227:381 (1991); Marks et al., J. Mol. Biol., 222:581-597 (1991); Vaughan et al., Nature Biotech., 14:309 (1996)). Phage display technology (McCafferty et al., Nature, 348:552-553 (1990)) can be used to produce human antibodies and antibody fragments in vitro, from immunoglobulin variable (V) domain gene repertoires from unimmunized donors. According to this technique, antibody V domain genes are cloned in-frame into either a major or minor coat protein gene of a filamentous bacteriophage, such as M13 or fd, and displayed as functional antibody fragments on the surface of the phage particle. Because the filamentous particle contains a single-stranded DNA copy of the phage genome, selections based on the functional properties of the antibody also result in selection of the gene encoding the antibody exhibiting those properties. Thus, the phage mimics some of the properties of the B cell. Phage display can be performed in a variety of formats; for their review see, e.g., Johnson, Kevin S, and Chiswell, David J., Current Opinion in Structural Biology 3:564-571 (1993). Several sources of V-gene segments can be used for phage display. Clackson et al., Nature, 352:624-628 (1991) isolated a diverse array of antioxazolone antibodies from a small random combinatorial library of V genes derived from the spleens of unimmunized mice. A repertoire of V genes from unimmunized human donors can be constructed and antibodies to a diverse array of antigens (including self-antigens) can be isolated essentially following the techniques described by Marks et al., J. Mol. Biol., 222:581-597 (1991), or Griffith et al., EMBO J., 12:725-734 (1993). See, also, U.S. Pat. Nos. 5,565,332 and 5,573,905, each of which is incorporated herein by reference in its entirety.

Human antibodies may also be generated by in vitro activated B cells (see, U.S. Pat. Nos. 5,567,610 and 5,229,275, each of which is incorporated herein by reference in its entirety). Human antibodies may also be generated in vitro using hybridoma techniques such as, but not limited to, that described by Roder et al. (Methods Enzymol., 121:140-167 (1986)).

Alternatively, in some embodiments, a non-human antibody is humanized, where specific sequences or regions of the antibody are modified to increase similarity to an antibody naturally produced in a human. In one embodiment, the antigen-binding domain portion is humanized.

A humanized antibody can be produced using a variety of techniques known in the art, including but not limited to, CDR-grafting (see, e.g., European Patent No. EP 239,400; International Publication No. WO 91/09967; and U.S. Pat. Nos. 5,225,539, 5,530,101, and 5,585,089, each of which is incorporated herein in its entirety by reference), veneering or 5 resurfacing (see, e.g., European Patent Nos. EP 592,106 and EP 519,596; Padlan, 1991, Molecular Immunology, 28(4/5):489-498; Studnicka et al., 1994, Protein Engineering, 7(6):805-814; and Roguska et al., 1994, PNAS, 91:969-973, each of which is incorporated herein by its entirety by reference), chain shuffling (see, e.g., U.S. Pat. No. 5,565,332, which is incorporated herein in its entirety by reference), and techniques disclosed in, e.g., U.S. Patent Application Publication No. US2005/0042664, U.S. Patent Application Publication 10 No. US2005/0048617, U.S. Pat. No. 6,407,213, U.S. Pat. No. 5,766,886, International Publication No. WO 9317105, Tan et al., J. Immunol., 169:1119-25 (2002), Caldas et al., Protein Eng., 13(5):353-60 (2000), Morea et al., Methods, 20(3):267-79 (2000), Baca et al., J. Biol. Chem., 272(16):10678-84 (1997), Roguska et al., Protein Eng., 9(10):895-904 (1996), 15 Couto et al., Cancer Res., 55 (23 Supp):5973s-5977s (1995), Couto et al., Cancer Res., 55(8):1717-22 (1995), Sandhu J S, Gene, 150(2):409-10 (1994), and Pedersen et al., J. Mol. Biol., 235(3):959-73 (1994), each of which is incorporated herein in its entirety by reference. Often, framework residues in the framework regions will be substituted with the corresponding residue from the CDR donor antibody to alter, preferably improve, antigen-20 binding. These framework substitutions are identified by methods well-known in the art, e.g., by modeling of the interactions of the CDR and framework residues to identify framework residues important for antigen-binding and sequence comparison to identify unusual framework residues at particular positions. (See, e.g., Queen et al., U.S. Pat. No. 5,585,089; and Riechmann et al., 1988, Nature, 332:323, which are incorporated herein by reference in 25 their entireties).

A humanized antibody has one or more amino acid residues introduced into it from a source which is nonhuman. These nonhuman amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Thus, humanized antibodies comprise one or more CDRs from nonhuman immunoglobulin molecules and framework regions from human. Humanization of antibodies is well-known in the art and can essentially be performed following the method of Winter and co-workers (Jones et al., Nature, 321:522-525 (1986); Riechmann et al., Nature, 332:323-327 (1988); Verhoeyen et al., Science, 239:1534-1536 (1988)), by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody, i.e., CDR-grafting (EP

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239,400; PCT Publication No. WO 91/09967; and U.S. Pat. Nos. 4,816,567; 6,331,415; 5,225,539; 5,530,101; 5,585,089; 6,548,640, the contents of which are incorporated herein by reference herein in their entirety). In such humanized chimeric antibodies, substantially less than an intact human variable domain has been substituted by the corresponding sequence from a nonhuman species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some framework (FR) residues are substituted by residues from analogous sites in rodent antibodies. Humanization of antibodies can also be achieved by veneering or resurfacing (EP 592,106; EP 519,596; Padlan, 1991, Molecular Immunology, 28(4/5):489-498; Studnicka et al., Protein Engineering, 7(6):805-814 (1994); and Roguska et al., PNAS, 91:969-973 (1994)) or chain shuffling (U.S. Pat. No. 5,565,332), the contents of which are incorporated herein by reference herein in their entirety.

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In some instances, a human scFv may also be derived from a yeast display library.

The choice of human variable domains, both light and heavy, to be used in making the humanized antibodies is to reduce antigenicity. According to the so-called "best-fit" method, the sequence of the variable domain of a rodent antibody is screened against the entire library of known human variable-domain sequences. The human sequence which is closest to that of the rodent is then accepted as the human framework (FR) for the humanized antibody (Sims et al., J. Immunol., 151:2296 (1993); Chothia et al., J. Mol. Biol., 196:901 (1987), the contents of which are incorporated herein by reference herein in their entirety). Another method uses a particular framework derived from the consensus sequence of all human antibodies of a particular subgroup of light or heavy chains. The same framework may be used for several different humanized antibodies (Carter et al., Proc. Natl. Acad. Sci. USA, 89:4285 (1992); Presta et al., J. Immunol., 151:2623 (1993), the contents of which are incorporated herein by reference herein in their entirety).

Antibodies can be humanized with retention of high affinity for the target antigen and other favorable biological properties. According to one aspect of the invention, humanized antibodies are prepared by a process of analysis of the parental sequences and various conceptual humanized products using three-dimensional models of the parental and humanized sequences. Three-dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available which illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind the target antigen.

In this way, FR residues can be selected and combined from the recipient and import sequences so that the desired antibody characteristic, such as increased affinity for the target antigen, is achieved. In general, the CDR residues are directly and most substantially involved in influencing antigen-binding.

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A "humanized" antibody retains a similar antigenic specificity as the original antibody, i.e., in the present invention, the ability to bind human GD2. However, using certain methods of humanization, the affinity and/or specificity of binding of the antibody for human GD2 may be increased using methods of "directed evolution," as described by Wu et al., J. Mol. Biol., 294:151 (1999), the contents of which are incorporated herein by reference herein in their entirety.

In one embodiment, the antigen-binding moiety portion of the CAR of the invention targets GD2. In some embodiments, the antigen-binding moiety portion in the CAR of the invention is a fully human anti-GD2 scFV. In some embodiments, the nucleic acid sequence of the anti-GD2 scFV comprises the sequence set forth in SEQ ID NO: 43. In one embodiment, the anti-GD2 scFV comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 44. In another embodiment, the anti-GD2 scFV portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 44. In other embodiments, the nucleic acid sequence of the anti-GD2 scFV comprises the sequence set forth in SEQ ID NO: 87. In one embodiment, the anti-GD2 scFV comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 88. In another embodiment, the anti-GD2 scFV portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 88.

In other embodiments, the antigen-binding moiety portion of the CAR of the invention targets Her2. In some embodiments, the nucleic acid sequence of the anti-Her2 scFV comprises the sequence set forth in SEQ ID NO: 123. In one embodiment, the anti-Her2 scFV comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 124. In another embodiment, the anti-Her2 scFV portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 124.

In other embodiments, the antigen-binding moiety portion of the CAR of the invention targets CD19. In some embodiments, the nucleic acid sequence of the anti-CD19 scFV comprises the sequence set forth in SEQ ID NO: 11. In one embodiment, the anti-CD19 scFV comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 12. In another embodiment, the anti-CD19 scFV portion of the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 12.

Transmembrane Domain

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With respect to the transmembrane domain, the CAR can be designed to comprise a transmembrane domain that is fused to the extracellular domain of the CAR. In one embodiment, a transmembrane domain that naturally is associated with one of the domains in the CAR is used. In some embodiments, the transmembrane domain is selected or modified by amino acid substitution to avoid binding of such domains to the transmembrane domains of the same or different surface membrane proteins to minimize interactions with other members of the receptor complex.

The transmembrane domain may be derived either from a natural or from a synthetic source. Where the source is natural, the domain may be derived from any membrane-bound or transmembrane protein. Transmembrane regions that find particular use in this invention include, but are not limited to, a transmembrane region derived from (i.e. comprise at least the transmembrane region(s) of) the alpha or beta chain of the T-cell receptor, CD28, CD3 zeta, CD3 epsilon, CD45, CD4, CD5, CD8, CD9, CD16, CD22, CD33, CD37, CD64, CD80, CD86, CD134, CD137, or CD154. In some embodiments, a variety of human hinges can be employed as well including the human Ig (immunoglobulin) hinge.

In some embodiments, the transmembrane domain is synthetic, in which case it comprises predominantly hydrophobic residues such as leucine and valine. In some embodiments, triplet of phenylalanine, tryptophan and valine will be found at each end of a synthetic transmembrane domain. Optionally, a short oligo- or polypeptide linker, preferably between 2 and 10 amino acids in length forms the linkage between the transmembrane domain and the cytoplasmic signaling domain of the CAR.

In one embodiment, the transmembrane domain in the CAR of the invention is the CD28 transmembrane domain. In one embodiment, the CD28 transmembrane domain comprises the nucleic acid sequence of SEQ ID NO: 15. In one embodiment, the CD28 transmembrane domain comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 16. In another embodiment, the CD28 transmembrane domain comprises the amino acid sequence of SEQ ID NO: 16.

In some embodiments, a CD28 hinge region is attached to the CD28 transmembrane domain of the CAR. In one embodiment, the CD28 hinge domain comprises the nucleic acid sequence of SEQ ID NO: 13. In one embodiment, the CD28 hinge domain comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 14. In another embodiment, the CD28 hinge domain comprises the amino acid sequence of SEQ ID NO: 14.

Intracellular or Cytoplasmic Domains

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The intracellular signaling domain of a CAR of the invention is responsible for activation of at least one of the normal effector functions of the T cell in which the chimeric receptor has been placed. The term "effector function" refers to a specialized function of a differentiated cell. Effector function of a T cell, for example, may be cytolytic activity or helper activity including the secretion of cytokines. Effector function in a naive, memory, or memory-type T cell includes antigen-dependent proliferation. Thus, the term "intracellular signaling domain" refers to the portion of a protein that transduces the effector function signal and directs the cell to perform a specialized function. While usually the entire intracellular signaling domain is employed, in many cases it will not be necessary to use the entire intracellular polypeptide. To the extent that a truncated portion of the intracellular signaling domain may find use, such truncated portion may be used in place of the intact chain as long as it still transduces the effector function signal. The term intracellular signaling domain is thus meant to include any truncated portion of the intracellular signaling domain sufficient to transduce the effector function signal.

Examples of intracellular signaling domains include, but are not limited to, cytoplasmic sequences of the T cell receptor (TCR) and co-receptors that act in concert to initiate signal transduction following antigen receptor engagement, as well as any derivative or variant of these sequences and any synthetic sequence that has the same functional capability. It is known that signals generated through the TCR alone are insufficient for full activation of the T cell and that a secondary or co-stimulatory signal is also required. Thus, T cell activation can be mediated by two distinct classes of cytoplasmic signaling sequence: those that initiate antigen-dependent primary activation through the TCR (primary cytoplasmic signaling sequences) and those that act in an antigen-independent manner to provide a secondary or co-stimulatory signal (secondary cytoplasmic signaling sequences).

Primary cytoplasmic signaling sequences regulate primary activation of the TCR complex either in a stimulatory way, or in an inhibitory way. Primary cytoplasmic signaling sequences that act in a stimulatory manner may contain signaling motifs which are known as immunoreceptor tyrosine-based activation motifs or ITAMs.

Examples of ITAM containing primary cytoplasmic signaling sequences that are of particular use in the invention include those derived from CD3 zeta, FcR gamma, FcR beta, CD3 gamma, CD3 delta, CD3 epsilon, CD5, CD22, CD79a, CD79b, and CD66d. In some

embodiments, the cytoplasmic signaling molecule in the CAR of the invention comprises a cytoplasmic signaling sequence derived from CD3 zeta.

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The cytoplasmic domain of the CAR can be designed to comprise the CD3-zeta signaling domain by itself or combined with any other desired cytoplasmic domain(s) useful in the context of the CAR of the invention. For example, the cytoplasmic domain of the CAR can comprise a CD3-zeta chain portion and a costimulatory signaling region or domain. The costimulatory signaling region refers to a portion of the CAR comprising the intracellular domain of a costimulatory molecule. A costimulatory molecule is a cell surface molecule other than an antigen receptor or their ligands that is required for an efficient response of lymphocytes to an antigen. Examples of such molecules include CD27, CD28, 4-1BB (CD137), OX40, CD30, CD40, PD-1, ICOS, lymphocyte function-associated antigen-1 (LFA-1), CD2, CD7, LIGHT, NKG2C, B7-H3, a ligand that specifically binds with CD83, and the like. In some embodiments, the CARs described herein comprise an ITAM-containing CD3-zeta domain in combination with CD28 or 4-1BB as the co-stimulatory signaling element. In some embodiments, the CARs described herein comprise an ITAM-containing CD3-zeta domain in combination with at least one other costimulatory domain are within the scope of the invention.

An intracellular signaling domain may comprise CD3-zeta or any of its homologs (e.g., eta, delta, gamma, or epsilon), MB 1 chain, B29, Fc RIII, Fc RI, and combinations of signaling molecules, such as CD3-zeta and CD28, CD27, 4-1BB, DAP-10, OX40, and combinations thereof. Intracellular signaling portions of other members of the families of activating proteins can be used, such as FcyRIII and FcyRI (see, Gross et al. (1992), Stancovski et al. (1993), Moritz et al. (1994), Hwu et al. (1995), Weijtens et al. (1996), and Hekele et al. (1996) for disclosures of cTCR's using these alternative transmembrane and intracellular domains.

The cytoplasmic signaling sequences within the cytoplasmic signaling portion of the CAR of the invention may be linked to each other in any order. Optionally, a short oligo- or polypeptide linker, preferably between 2 and 10 amino acids in length may form the linkage. A glycine-serine doublet provides a particularly suitable linker. The antigen-specific extracellular domain and the intracellular signaling-domain may be linked by a transmembrane domain, such as the human IgG_{4Fc} hinge and Fc regions. Alternatives include the human CD4 transmembrane domain, the human CD28 transmembrane domain, the human CD3-zeta transmembrane domain, or a cysteine mutated human CD3-zeta

transmembrane, or other transmembrane domains from other human transmembrane signaling proteins, such as CD16 and CD8 and erythropoietin receptor.

In some embodiments, any part of the endogenous T cell receptor complex is utilized in the intracellular domain. One or multiple cytoplasmic domains may be employed, as so-called third generation CARs have at least two or three signaling domains fused together for additive or synergistic effect, for example.

In one embodiment, the cytoplasmic domain comprises the signaling domain of CD3-zeta and the signaling domain of CD28. In another embodiment, the cytoplasmic domain comprises the signaling domain of CD3-zeta and the signaling domain of 4-1BB and/or OX-40.

In one embodiment, the cytoplasmic domain in a CAR of the invention comprises the signaling domain of CD28 and the signaling domain of CD3-zeta, wherein the signaling domain of CD28 comprises the nucleic acid sequence set forth in SEQ ID NO: 17 and the signaling domain of CD3-zeta comprises the nucleic acid sequence set forth in SEQ ID NO: 19. In one embodiment, the signaling domain of CD28 comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 18. In another embodiment, the signaling domain of CD28 comprises the amino acid sequence of SEQ ID NO: 18. In one embodiment, the signaling domain of CD3-zeta comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 20. In another embodiment, the signaling domain of CD3-zeta comprises the amino acid sequence of SEQ ID NO: 20.

Regulatable Destabilization Domain

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The regulatable destabilization domain (RDD) of a CAR of the invention is utilized to modulate (e.g., reduce) in a time and/or dose dependent manner the expression of the CAR on the surface of a T cell. In this way, CAR T cell exhaustion can be prevented/inhibited or reversed leading to a maintained, regained or enhanced functionality (e.g., prolonged effector functions) of CAR T cells.

For example, in some embodiments, a CAR T cell genetically engineered to express a CAR containing an RDD (e.g., fused to the CAR) marks the CAR for ubiquitin mediated degradation, such that when in the presence of a stabilizing small molecule that binds the RDD, CAR degradation is prevented, and removal or absence of the stabilizing small molecule that binds the RDD allows CAR degradation to occur. In some embodiments, modulation of CAR protein levels and CAR surface expression via fusion of an RDD to the CAR reverses exhaustion of CAR T cells that have already developed the hallmarks of T cell

exhaustion due to tonic CAR signaling. For example, CAR T cells that are exposed to the stabilizing small molecule demonstrate all of the phenotypic and functional hallmarks of T cell exhaustion. However, when the stabilizing small molecule is removed and CAR protein levels are reduced by protein degradation, the phenotypic and functional indicators of T cell exhaustion are reversed and T cell function is restored.

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The invention is not limited to any particular RDD. Indeed, any RDD that confers CAR instability such that rapid CAR protein degradation occurs when the RDD is not stabilized by the presence of its cognate small molecule, yet whose structure or presence within the CAR does not disrupt CAR function may be used in the invention. A non-limiting example of an RDD is derived from the FK506 binding protein (FKBP), referred to herein as an FKBP DD (see Banaszynski et. al, Cell 2006). Surface expression of a CAR containing the FKBP RDD can be rapidly and dose-dependently regulated by adding or subtracting the stabilizing rapalog shield-1 (S1) to CAR T cells or to an organism treated with CAR T cells comprising an FKBP DD. Another non-limiting example of an RDD is derived from E. coli dihydrofolate reductase (DHFR), referred to herein as a DHFR DD, which can be rapidly and dose-dependently regulated with the stabilizing small molecule drug trimethoprim. As described herein, in some embodiments, CAR expression is achieved in vivo using a clinically established and well tolerated dose of trimethoprim (see Kremers et al., J Clin Pharmacol. 1974 Feb-Mar;14(2):112-7). For example, maximum expression of a CAR comprising the trimethoprim-controlled DHFR RDD occurred at about 500 nM of trimethoprim, well within the compound's EC50 of 50 nM. This is well within the mean steady-state plasma concentration of trimethoprim when taken 160 mg twice a day (1.72 μg/mL, about 6μM). In some embodiments, CAR expression is achieved in vivo using a dose that is below a clinically established dose of trimethoprim.

In some embodiments, the DHFR DD is derived from *E. coli* DHFR. In some embodiments, the DHFR DD is derived from human DHFR. Similarly, the invention is not limited by the type RDD used. For example, any DHFR that contains mutations that achieve destabilization of the DHFR domain and any protein attached thereto may be used. For example, any RDD (DHFR or FKBP DD) that confers instability/destabilization such that rapid protein degradation occurs when the RDD is not stabilized, using for example trimethoprim or S1, yet whose structure or presence within a CAR does not disrupt CAR effector function can be used. In some embodiments, an *E. coli* DHFR DD of SEQ ID NO. 6 that contains mutations at amino acids 12 and 100 is used.

The invention is not limited by the location of the RDD within the CAR. For example, in some embodiments, an RDD is fused (e.g., genetically linked) on the N- or the C- terminus of the CAR. A CAR can be expressed with an RDD fused to an intracellular component of the CAR, fused to an extracellular component of the CAR, or fused to a hinge or transmembrane region. Exemplary intracellular components of the CAR include, but are not limited to, a co-stimulatory domain or the CD3-zeta domain. In some embodiments, the intracellular domain is the CD3-zeta domain.

The intracellular component of the CAR (e.g., CD3-zeta) may be linked to an RDD in a random or specified order. Optionally, a short oligo- or polypeptide linker, preferably between 2 and 10 amino acids in length may form the linkage.

CAR Expression Vectors and Genetically Modified T cells

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The present invention encompasses a DNA construct comprising sequences of a CAR, wherein the sequence comprises the nucleic acid sequence of an antigen-binding moiety (e.g., a tumor antigen-binding domain or a pathogen antigen-binding domain) operably linked to the nucleic acid sequence of an intracellular domain which is operably linked to the nucleic acid sequence of an RDD. An exemplary intracellular domain that can be used in the CAR of the invention includes but is not limited to the intracellular domain of CD3-zeta, CD27, CD28 and the like. In some instances, the CAR can comprise any combination of CD3-zeta, CD28, 4-1BB, and the like. Exemplary DDs that can be used in the CAR of the invention include but are not limited to a DHFR DD and a FKBP DD.

In one embodiment, the CAR of the invention comprises anti-GD2 scFv, human CD28 hinge and transmembrane domain, and CD28 and CD3-zeta signaling domains. In one embodiment the anti-GD2 scFv is fully human. In one embodiment, the CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 7. In another embodiment, the CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 8. In another embodiment, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 8.

In some embodiments, the CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 23. In another embodiment, the CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 24. In another embodiment, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 24.

In other embodiments, a CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 39. In another embodiment, the CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 40. In another embodiment, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 40.

In still other embodiments, a CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 55. In other embodiments, a CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 56. In other embodiments, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 56.

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In still other embodiments, a CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 71. In other embodiments, a CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 72. In other embodiments, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 72.

In other embodiments, a CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 87. In other embodiments, a CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 88. In other embodiments, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 88.

In other embodiments, a CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 103. In other embodiments, a CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 104. In other embodiments, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 104.

In some embodiments, a CAR of the invention comprises the nucleic acid sequence set forth in SEQ ID NO: 119. In other embodiments, a CAR of the invention comprises the nucleic acid sequence that encodes the amino acid sequence of SEQ ID NO: 120. In other embodiments, the CAR of the invention comprises the amino acid sequence set forth in SEQ ID NO: 120.

In some embodiments, the invention provides isolated nucleic acid segments and expression cassettes incorporating DNA sequences that encode a CAR fused to an RDD. Vectors of the present invention are designed, primarily, to deliver desired genes to immune cells, preferably T cells under the control of regulated eukaryotic promoters, for example,

EFlα promoter MNDU3 promoter, CMV promoter, or Ubiquitin promoter. Also, the vectors may contain a selectable marker, if for no other reason, to facilitate their manipulation in vitro. In other embodiments, the CAR can be expressed from mRNA in vitro transcribed from a DNA template.

CAR molecules fused to an RDD are recombinant and are distinguished by their ability to both bind antigen and transduce activation signals via immunoreceptor activation motifs (ITAM's) present in their cytoplasmic tails. CAR constructs utilizing an antigenbinding moiety (for example, generated from single chain antibodies (scFv)) afford the additional advantage of being HLA-independent in that they do not require an antigen be presented by a major histocompatibility complex (MHC) protein, but rather bind native antigen on the target cell surface in an HLA-independent fashion.

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The nucleic acid sequences coding for the desired molecules can be obtained using recombinant methods known in the art, such as, for example by screening libraries from cells expressing the gene, by deriving the gene from a vector known to include the same, or by isolating directly from cells and tissues containing the same, using standard techniques. Alternatively, a gene of interest can be produced synthetically, rather than cloned.

The present invention also provides vectors in which a DNA of the present invention is inserted. Vectors derived from retroviruses such as the lentivirus are suitable tools to achieve long-term gene transfer since they allow long-term, stable integration of a transgene and its propagation in daughter cells. Lentiviral vectors have the added advantage over vectors derived from onco-retroviruses such as murine leukemia viruses in that they can transduce non-proliferating cells, such as hepatocytes. They also have the added advantage of low immunogenicity. In another embodiment, the desired CAR can be expressed in the cells by way of transponsons.

The expression of natural or synthetic nucleic acids encoding CARs is typically achieved by operably linking a nucleic acid encoding the CAR polypeptide or portions thereof to a promoter, and incorporating the construct into an expression vector. The vectors can be suitable for replication and integration into eukaryotes. Typical cloning vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the desired nucleic acid sequence.

Expression constructs of the invention may also be used for nucleic acid immunization and gene therapy, using standard gene delivery protocols. Methods for gene delivery are known in the art. See, e.g., U.S. Pat. Nos. 5,399,346, 5,580,859, 5,589,466,

incorporated by reference herein in their entireties. In another embodiment, the invention provides a gene therapy vector.

Nucleic acid can be cloned into a number of types of vectors. For example, the nucleic acid can be cloned into a vector including, but not limited to a plasmid, a phagemid, a phage derivative, an animal virus, and a cosmid. Vectors of particular interest include expression vectors, replication vectors, probe generation vectors, and sequencing vectors.

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Further, the expression vector may be provided to a cell in the form of a viral vector. Viral vector technology is well known in the art and is described, for example, in Sambrook et al. (2001, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York), and in other virology and molecular biology manuals. Viruses, which are useful as vectors include, but are not limited to, retroviruses, adenoviruses, adeno-associated viruses, herpes viruses, and lentiviruses. In general, a suitable vector contains an origin of replication functional in at least one organism, a promoter sequence, convenient restriction endonuclease sites, and one or more selectable markers, (e.g., WO 01/96584; WO 01/29058; and U.S. Pat. No. 6,326,193).

A number of viral based systems have been developed for gene transfer into mammalian cells. For example, retroviruses provide a convenient platform for gene delivery systems. A selected gene can be inserted into a vector and packaged in retroviral particles using techniques known in the art. The recombinant virus can then be isolated and delivered to cells of the subject either in vivo or ex vivo. A number of retroviral systems are known in the art. In some embodiments, lentivirus vectors are used. In other embodiments, adenovirus vectors are used. A number of adenovirus vectors are known in the art.

Additional promoter elements, e.g., enhancers, can be used to regulate the frequency of transcriptional initiation. Typically, these are located in the region 30-110 bp upstream of the start site, although a number of promoters have recently been shown to contain functional elements downstream of the start site as well. The spacing between promoter elements frequently is flexible, so that promoter function is preserved when elements are inverted or moved relative to one another. In the thymidine kinase (tk) promoter, the spacing between promoter elements can be increased to 50 bp apart before activity begins to decline. Individual elements can function either cooperatively or independently to activate transcription.

In some embodiments, the vector comprises a promoter. In some embodiments, the promoter is a constitutive promoter. In some embodiments, the constitutive promoter comprises the immediate early cytomegalovirus (CMV) promoter sequence. This promoter

sequence is a strong constitutive promoter sequence capable of driving high levels of expression of any polynucleotide sequence operatively linked thereto. In some embodiments, the constitutive promoter comprises the eukaryotic translation elongation factor-1 alpha promoter sequence. In some embodiments, the constitutive promoter is selected from the group consisting of the simian virus 40 (SV40) early promoter, mouse mammary tumor virus (MMTV), human immunodeficiency virus (HIV) long terminal repeat (LTR) promoter, MoMuLV promoter, an avian leukemia virus promoter, an Epstein-Barr virus immediate early promoter, and a Rous sarcoma virus promoter. In some embodiments, the constitutive promoter is selected from the group consisting of the actin I promoter, the myosin promoter, a hemoglobin promoter, and the creatine kinase promoter. In some embodiments, the promoter is an inducible promoter. The use of an inducible promoter provides a molecular switch capable of turning on expression of the polynucleotide sequence to which it is operatively linked when such expression is desired, or turning off the expression when expression is not desired. Examples of inducible promoters include, but are not limited to a metallothionein promoter, a glucocorticoid promoter, a progesterone promoter, and a tetracycline promoter.

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In order to assess the expression of a CAR polypeptide or portions thereof, the expression vector to be introduced into a cell can also contain either a selectable marker gene or a reporter gene or both to facilitate identification and selection of expressing cells from the population of cells sought to be transfected or infected through viral vectors. In other aspects, the selectable marker may be carried on a separate piece of DNA and used in a cotransfection procedure. Both selectable markers and reporter genes may be flanked with appropriate regulatory sequences to enable expression in the host cells. Useful selectable markers include, for example, antibiotic-resistance genes, such as neo and the like.

Reporter genes are used for identifying potentially transfected cells and for evaluating the functionality of regulatory sequences. In general, a reporter gene is a gene that is not present in or expressed by the recipient organism or tissue and that encodes a polypeptide whose expression is manifested by some easily detectable property, e.g., enzymatic activity. Expression of the reporter gene is assayed at a suitable time after the DNA has been introduced into the recipient cells. Suitable reporter genes may include genes encoding luciferase, beta-galactosidase, chloramphenicol acetyl transferase, secreted alkaline phosphatase, or the green fluorescent protein gene (e.g., Ui-Tei et al., 2000 FEBS Letters 479: 79-82). Suitable expression systems are well known and may be prepared using known techniques or obtained commercially. In general, the construct with the minimal 5' flanking

region showing the highest level of expression of reporter gene is identified as the promoter. Such promoter regions may be linked to a reporter gene and used to evaluate agents for the ability to modulate promoter-driven transcription.

Methods of introducing and expressing genes into a cell are known in the art. In the context of an expression vector, the vector can be readily introduced into a host cell, e.g., mammalian, bacterial, yeast, or insect cell by any method in the art. For example, the expression vector can be transferred into a host cell by physical, chemical, or biological means.

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Physical methods for introducing a polynucleotide into a host cell include calcium phosphate precipitation, lipofection, particle bombardment, microinjection, electroporation, and the like. Methods for producing cells comprising vectors and/or exogenous nucleic acids are well-known in the art. See, for example, Sambrook et al. (2001, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York). A preferred method for the introduction of a polynucleotide into a host cell is calcium phosphate transfection.

Chemical means for introducing a polynucleotide into a host cell include colloidal dispersion systems, such as macromolecule complexes, nanocapsules, microspheres, beads, and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. An exemplary colloidal system for use as a delivery vehicle in vitro and in vivo is a liposome (e.g., an artificial membrane vesicle).

In the case where a non-viral delivery system is utilized, an exemplary delivery vehicle is a liposome. The use of lipid formulations is contemplated for the introduction of the nucleic acids into a host cell (in vitro, ex vivo or in vivo). In another aspect, the nucleic acid may be associated with a lipid. The nucleic acid associated with a lipid may be encapsulated in the aqueous interior of a liposome, interspersed within the lipid bilayer of a liposome, attached to a liposome via a linking molecule that is associated with both the liposome and the oligonucleotide, entrapped in a liposome, complexed with a lipid, contained as a suspension in a lipid, mixed with a lipid, combined with a lipid, contained as a suspension in a lipid, contained or complexed with a micelle, or otherwise associated with a lipid. Lipid, lipid/DNA or lipid/expression vector associated compositions are not limited to any particular structure in solution. For example, they may be present in a bilayer structure, as micelles, or with a "collapsed" structure. They may also simply be interspersed in a solution, possibly forming aggregates that are not uniform in size or shape. Lipids are fatty substances which may be naturally occurring or synthetic lipids. For example, lipids include the fatty droplets that naturally occur in the cytoplasm as well as the class of

compounds which contain long-chain aliphatic hydrocarbons and their derivatives, such as fatty acids, alcohols, amines, amino alcohols, and aldehydes.

Lipids suitable for use can be obtained from commercial sources. For example, dimyristyl phosphatidylcholine ("DMPC") can be obtained from Sigma, St. Louis, Mo.; dicetyl phosphate ("DCP") can be obtained from K & K Laboratories (Plainview, N.Y.); cholesterol ("Choi") can be obtained from Calbiochem-Behring; dimyristyl phosphatidylglycerol ("DMPG") and other lipids may be obtained from Avanti Polar Lipids, Inc. (Birmingham, Ala.). "Liposome" is a generic term encompassing a variety of single and multilamellar lipid vehicles formed by the generation of enclosed lipid bilayers or aggregates. Liposomes can be characterized as having vesicular structures with a phospholipid bilayer membrane and an inner aqueous medium. Multilamellar liposomes have multiple lipid layers separated by aqueous medium. They form spontaneously when phospholipids are suspended in an excess of aqueous solution. The lipid components undergo self-rearrangement before the formation of closed structures and entrap water and dissolved solutes between the lipid bilayers (Ghosh et al., 1991 Glycobiology 5: 505-10). However, compositions that have different structures in solution than the normal vesicular structure are also encompassed. For example, the lipids may assume a micellar structure or merely exist as nonuniform aggregates of lipid molecules. Also contemplated are lipofectamine-nucleic acid complexes.

In some embodiments, nucleic acids encoding the CAR constructs are delivered into cells using a lentiviral or retroviral vector. CAR-expressing lentiviral or adenoviral vectors can be delivered into different types of eukaryotic cells as well as into tissues and whole organisms using transduced cells as carriers or cell-free local or systemic delivery of encapsulated, bound or naked vectors. The method used can be for any purpose where stable expression is required or sufficient.

In another embodiment, the desired CAR can be expressed in the cells by way of transponsons.

The disclosed methods can be applied to the modulation of T cell activity in basic research and therapy, in the fields of cancer, stem cells, acute and chronic infections, and autoimmune diseases, including the assessment of the ability of the genetically modified T cell to kill a target cancer cell.

Types and Sources of T Cells

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In one aspect, the CAR T cells provided herein are autologous CAR T cells. In some embodiments, the autologous CAR T cells are manufactured from T cells obtained from a

subject or patient to be treated. In some embodiments, the T cells obtained from a subject are isolated from samples comprising peripheral blood mononuclear cells, bone marrow, lymph node tissue, cord blood, thymus tissue, tissue from a site of infection, ascites, pleural effusion, spleen tissue, and tumors. In certain embodiments, any number of T cell lines available in the art, may be used. In certain embodiments of the present invention, T cells can be obtained from a unit of blood collected from a subject using any number of techniques known to the skilled artisan, such as FICOLLTM separation. In some embodiments, cells from the circulating blood of an individual are obtained by apheresis. The apheresis product typically contains lymphocytes, including T cells, monocytes, granulocytes, B cells, other nucleated white blood cells, red blood cells, and platelets. In one embodiment, the cells collected by apheresis may be washed to remove the plasma fraction and to place the cells in an appropriate buffer or media for subsequent processing steps. In one embodiment of the invention, the cells are washed with phosphate buffered saline (PBS). In an alternative embodiment, the wash solution lacks calcium, magnesium, and other divalent cations. After washing, the cells may be resuspended in a variety of biocompatible buffers, such as, for example, Ca²⁺ free, Mg²⁺-free PBS, PlasmaLyte A, or other saline solution with or without buffer. Alternatively, the undesirable components of the apheresis sample may be removed and the cells directly resuspended in culture medium.

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In another embodiment, T cells are isolated from peripheral blood lymphocytes by lysing the red blood cells and depleting the monocytes, for example, by centrifugation through a PERCOLLTM gradient or by counterflow centrifugal elutriation.

Specific subpopulations of T cells can be further isolated by positive or negative selection techniques. Exemplary subpopulations of T cells include CD3⁺, CD28⁺, CD4⁺, CD8⁺, CD45RA⁺, and CD45RO⁺T cells. For example, in one embodiment, T cells are isolated by incubation with anti-CD3/anti-CD28-conjugated beads, such as DYNABEADSTM M-450 CD3/CD28 T, for a time period sufficient for positive selection of the desired T cells. In one embodiment, the time period is about 30 minutes. In a further embodiment, the time period ranges from 30 minutes to 36 hours or longer and all integer values there between. In a further embodiment, the time period is at least 1, 2, 3, 4, 5, or 6 hours. In yet another preferred embodiment, the time period is 10 to 24 hours. In one preferred embodiment, the incubation time period is 24 hours. For isolation of T cells from patients with leukemia, use of longer incubation times, such as 24 hours, can increase cell yield. Longer incubation times may be used to isolate T cells in any situation where there are few T cells as compared to other cell types, such in isolating tumor infiltrating lymphocytes (TIL) from tumor tissue or

from immune-compromised individuals. Further, use of longer incubation times can increase the efficiency of capture of CD8⁺ T cells. Thus, by simply shortening or lengthening the time T cells are allowed to bind to the CD3/CD28 beads and/or by increasing or decreasing the ratio of beads to T cells, subpopulations of T cells can be preferentially selected for or against at culture initiation or at other time points during the process. Additionally, by increasing or decreasing the ratio of anti-CD3 and/or anti-CD28 antibodies on the beads or other surface, subpopulations of T cells can be preferentially selected for or against at culture initiation or at other desired time points. The skilled artisan recognizes that multiple rounds of selection can also be used in the context of this invention. In certain embodiments, it may be desirable to perform the selection procedure and use the "unselected" cells in the activation and expansion process. "Unselected" cells can also be subjected to further rounds of selection.

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Enrichment of a T cell population by negative selection can be accomplished with a combination of antibodies directed to surface markers unique to the negatively selected cells. One method is cell sorting and/or selection via negative magnetic immunoadherence or flow cytometry that uses a cocktail of monoclonal antibodies directed to cell surface markers present on the cells negatively selected. For example, to enrich for CD4⁺cells by negative selection, a monoclonal antibody cocktail typically includes antibodies to CD14, CD20, CD11b, CD16, HLA-DR, and CD8. In certain embodiments, it may be desirable to enrich for or positively select for regulatory T cells which typically express CD4⁺, CD25⁺, CD62L^{hi}, GITR⁺, and FoxP3⁺. Alternatively, in certain embodiments, T regulatory cells are depleted by anti-C25 conjugated beads or other similar method of selection.

T cells can also be frozen after a washing step. In some embodiments, the freeze and subsequent thaw step provides a more uniform product by removing granulocytes and to some extent monocytes in the cell population. After the washing step that removes plasma and platelets, the cells may be suspended in a freezing solution.

In certain embodiments, cryopreserved cells are thawed and washed as described herein and allowed to rest (e.g., for one hour or more) at room temperature prior to use according to the methods of the present invention.

Also contemplated in the context of the invention is the collection of blood samples or apheresis product from a subject at a time period prior to when the expanded cells as described herein might be needed. As such, the source of the cells to be expanded can be collected at any time point necessary, and desired cells, such as T cells, isolated and frozen for later use in T cell therapy for any number of diseases or conditions that would benefit from T cell therapy, such as those described herein. In one embodiment a blood sample or an

apheresis is taken from a generally healthy subject. In certain embodiments, a blood sample or an apheresis is taken from a generally healthy subject who is at risk of developing a disease, but who has not yet developed a disease, and the cells of interest are isolated and frozen for later use. In certain embodiments, the T cells may be expanded, frozen, and used at a later time. In certain embodiments, samples are collected from a patient shortly after diagnosis of a particular disease as described herein but prior to any treatments.

Activation and Expansion of T Cells

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Whether prior to or after genetic modification of the T cells to express a desirable CAR, the T cells can be activated and expanded generally using methods as described, for example, in U.S. Pat. Nos. 6,352,694; 6,534,055; 6,905,680; 6,692,964; 5,858,358; 6,887,466; 6,905,681; 7,144,575; 7,067,318; 7,172,869; 7,232,566; 7,175,843; 5,883,223; 6,905,874; 6,797,514; 6,867,041; and U.S. Patent Application Publication No. 20060121005, each of which is incorporated by reference.

Generally, the T cells of the invention are expanded by contact with a surface having attached thereto an agent that stimulates a CD3/TCR complex associated signal and a ligand that stimulates a co-stimulatory molecule on the surface of the T cells. In particular, T cell populations may be stimulated as described herein, such as by contact with an anti-CD3 antibody, or antigen-binding fragment thereof, or an anti-CD2 antibody immobilized on a surface, or by contact with a protein kinase C activator (e.g., bryostatin) in conjunction with a calcium ionophore. For co-stimulation of an accessory molecule on the surface of the T cells, a ligand that binds the accessory molecule is used. For example, a population of T cells can be contacted with an anti-CD3 antibody and an anti-CD28 antibody, under conditions appropriate for stimulating proliferation of the T cells. To stimulate proliferation of either CD4⁺T cells or CD8⁺ T cells, an anti-CD3 antibody and an anti-CD28 antibody. Examples of An anti-CD28 antibody such as B-T3, XR-CD28 (Diaclone, Besancon, France) can be used as can other methods commonly known in the art (see Berg et al., Transplant Proc. 30(8):3975-3977, 1998; Haanen et al., J. Exp. Med. 190(9):13191328, 1999; Garland et al., J. Immunol Meth. 227(1-2):53-63, 1999).

In certain embodiments, the primary stimulatory signal and the co-stimulatory signal for the T cell may be provided by different protocols. For example, the agents providing each signal may be in solution or coupled to a surface. When coupled to a surface, the agents may be coupled to the same surface (i.e., in "cis" formation) or to separate surfaces (i.e., in "trans" formation). Alternatively, one agent may be coupled to a surface and the other agent in

solution. In one embodiment, the agent providing the co-stimulatory signal is bound to a cell surface and the agent providing the primary activation signal is in solution or coupled to a surface. In certain embodiments, both agents can be in solution. In another embodiment, the agents may be in soluble form, and then cross-linked to a surface, such as a cell expressing Fc receptors or an antibody or other binding agent which will bind to the agents. In this regard, see for example, U.S. Patent Application Publication Nos. 20040101519 and 20060034810 for artificial antigen presenting cells (aAPCs) that are contemplated for use in activating and expanding T cells in the present invention.

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In one embodiment, the two agents are immobilized on beads, either on the same bead, i.e., "cis," or to separate beads, i.e., "trans." By way of example, the agent providing the primary activation signal is an anti-CD3 antibody or an antigen-binding fragment thereof and the agent providing the co-stimulatory signal is an anti-CD28 antibody or antigen-binding fragment thereof; and both agents are co-immobilized to the same bead in equivalent molecular amounts.

In further embodiments of the present invention, T cells are combined with agent-coated beads, the beads and the cells are subsequently separated, and then the cells are cultured. In an alternative embodiment, prior to culture, the agent-coated beads and cells are not separated but are cultured together. In a further embodiment, the beads and cells are first concentrated by application of a force, such as a magnetic force, resulting in increased ligation of cell surface markers, thereby inducing cell stimulation.

For example, cell surface proteins may be ligated by allowing paramagnetic beads to which anti-CD3 and anti-CD28 are attached to contact the T cells. In one embodiment the cells (for example, 10^4 to 10^9 T cells) and beads (for example, DYNABEADSTM M-450 CD3/CD28 T paramagnetic beads at a ratio of 1:1) are combined in a buffer, preferably PBS (without divalent cations such as, calcium and magnesium). Accordingly, any cell number is within the context of the present invention. In certain embodiments, it may be desirable to significantly decrease the volume in which particles and cells are mixed together (i.e., increase the concentration of cells), to ensure maximum contact of cells and particles. For example, in one embodiment, a concentration of about 2 billion cells/ml is used. In another embodiment, greater than 100 million cells/ml is used. In a further embodiment, a concentration of cells of 10, 15, 20, 25, 30, 35, 40, 45, or 50 million cells/ml is used. In yet another embodiment, a concentration of cells from 75, 80, 85, 90, 95, or 100 million cells/ml is used. In further embodiments, concentrations of 125 or 150 million cells/ml can be used.

In one embodiment of the present invention, the mixture may be cultured for several hours (about 3 hours) to about 14 days or any hourly integer value in between. In another embodiment, the mixture may be cultured for 21 days. In one embodiment of the invention the beads and the T cells are cultured together for about eight days. In another embodiment, the beads and T cells are cultured together for 2-3 days. Several cycles of stimulation may also be desired such that culture time of T cells can be 60 days or more. Conditions appropriate for T cell culture include an appropriate media (e.g., Minimal Essential Media or RPMI Media 1640 or, X-vivo 15, (Lonza)) that may contain factors necessary for proliferation and viability, including serum (e.g., fetal bovine or human serum), interleukin-2 (IL-2), insulin, IFN-γ, IL-4, IL-7, GM-CSF, IL-10, IL-12, IL-15, TGFβ, and TNF-α or any other additives for the growth of cells known to the skilled artisan. Other additives for the growth of cells include, but are not limited to, surfactant, plasmanate, and reducing agents such as N-acetyl-cysteine and 2-mercaptoethanol. Media can include RPMI 1640, AIM-V, DMEM, MEM, α-MEM, F-12, X-Vivo 15, and X-Vivo 20, Optimizer, with added amino acids, sodium pyruvate, and vitamins, either serum-free or supplemented with an appropriate amount of serum (or plasma) or a defined set of hormones, and/or an amount of cytokine(s) sufficient for the growth and expansion of T cells. Antibiotics, e.g., penicillin and streptomycin, may also be included. The target cells are maintained under conditions necessary to support growth, for example, an appropriate temperature (e.g., 37C.) and atmosphere (e.g., air plus 5% CO₂).

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As described herein, CAR T cells tonically signal in the absence of tumor antigen when the CAR is expressed on the T cell surface. Tonic signaling is demonstrated by Western blotting in which the CAR CD3-zeta domain is phosphorylated at baseline (see Example 2, FIG. 16). Tonic signaling leads to T cell exhaustion, rendering the CAR T cells ineffective. Tonic signaling was prevented in RDD-CAR T cells by expanding them in the absence of stabilizing drug. When CAR expression was reduced or inhibited and not expressed on the T cell surface (in the absence of the appropriate small molecule – in this case, shield-1 – the CAR is not expressed on the cell surface – "always OFF"), or if the CAR was expressed at one point but then removed from the surface for a period of time (e.g., 72 hours - CAR OFF D7), an attenuation in the CD3-zeta phosphorylation was observed. Furthermore, when the CAR is not expressed on the surface (no shield-1, "CAR always OFF"), or cultured in shield-1 for 7 days and then removed, only a fraction of cells that co-express exhaustion markers were observed (see Example 2, FIG. 17). Therefore, in some embodiments, the present

invention provides that inhibiting CAR expression using an RDD-CAR of the invention has a profound effect on how CAR T cells are regulated on a transcriptional level by preventing or reversing CAR tonic signaling.

Accordingly, in some embodiments, because CARs tonically signal in the absence of antigen (e.g., during culture), expanding RDD-CAR T cells without stabilizing drug so that CAR expression on the surface is inhibited augments CAR T cell effector function when used therapeutically to treat a patient compared to T cells in which the CAR is continuously expressed (e.g., when expanded *in vitro* or *ex vivo*).

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In some embodiments, suppression of CAR expression during expansion *in vitro* generates and/or leads to healthier, more functional CAR T cells (e.g., with normal effector function) compared to CAR T cells in which CAR expression during expansion in vitro is not suppressed. Thus, in some embodiments, CAR expression is modulated (e.g., reduced) in T cells containing a CAR of the invention comprising an RDD (e.g., DHFR DD regulated by trimethoprim) during culture and expansion by adding drug to increase expression or removing drug to decrease expression. In some embodiments, suppressing CAR expression during culture and/or expansion inhibits tonic signaling *in vitro* leading to or generating T cells with better effector function in the context in treating a condition or disease in a subject compared to T cells in which tonic signaling was not inhibited.

T cells that have been exposed to varied stimulation times may exhibit different characteristics. For example, typical blood or apheresed peripheral blood mononuclear cell products have a helper T cell population (T_H , CD4+) that is greater than the cytotoxic or suppressor T cell population (T_C , CD8+). Ex vivo expansion of T cells by stimulating CD3 and CD28 receptors produces a population of T cells that prior to about days 8-9 consists predominately of T_H cells, while after about days 8-9, the population of T cells comprises an increasingly greater population of T_C cells. Accordingly, depending on the purpose of treatment, infusing a subject with a T cell population comprising predominately of T_H cells may be advantageous. Similarly, if an antigen-specific subset of T_C cells has been isolated it may be beneficial to expand this subset to a greater degree.

Further, in addition to CD4 and CD8 markers, other phenotypic markers vary significantly, but in large part, reproducibly during the course of the cell expansion process. Thus, such reproducibility enables the ability to tailor an activated T cell product for specific purposes.

Therapeutic Applications

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The present invention provides, in some embodiments, a cell (e.g., T cell) modified to express a CAR that combines an antigen recognition domain of a specific antibody with an intracellular domain of CD3-zeta, CD28, 4-1BB, or any combinations thereof, and an RDD. Therefore, in the presence of its target antigen, the modified T cell elicits a CAR-mediated T-cell response.

The invention provides the use of a CAR to redirect the specificity of a primary T cell to a tumor antigen. Thus, the present invention also provides a method for stimulating a T cell-mediated immune response to a target cell population or tissue in a subject comprising the step of administering to the subject a T cell that expresses a CAR, wherein the CAR comprises a binding moiety that specifically interacts with a predetermined target, a zeta chain portion comprising for example the intracellular domain of human CD3-zeta, a costimulatory signaling region, and an RDD.

In some embodiments, the present invention provides a cellular therapy where T cells are genetically modified to express a CAR containing an RDD and the CAR T cell is infused to a recipient in need thereof. The infused cell is able to kill tumor cells expressing the CAR T cell's target antigen in the recipient. Unlike other biologic therapies, CAR T cells are able to replicate *in vivo* resulting in long-term persistence that can lead to sustained tumor control. In some embodiments, a CAR T cell comprising a CAR including an RDD displays a prolonged anti-tumor response (e.g., compared to CAR T cell comprising a CAR lacking an RDD). In some embodiments, inclusion of an RDD within the CAR serves as a type of safety switch to inhibit CAR T cell activity/functionality (e.g., to mitigate CAR toxicity). For example, in some embodiments, removal of a stabilizing small molecule or drug (e.g., trimethoprim) from the CAR T cells results in rapidly reduced or inhibited expression of the CAR on the surface of the CAR T cell, thereby effectively shutting down or preventing targeting and effector functions of the CAR T cell. In particular, as shown in Example 2, the IC50 (the time it takes for 50% of surface CAR to be degraded), was observed to be less than 2 hours in the absence of stabilizing drug/molecule (see Example 2, FIG. 13). Therefore, in some embodiments, removing stabilizing drugs from patients who receive DD-CARs serves as a rapid and reversible safety switch. Thus, the invention provides, in some embodiments, the ability to mitigate or eliminate toxicity events that may occur with CAR T cell therapies, such as, but not limited to, cytokine release syndrome (CRS) or on-target off-tumor toxicity. Further, in the context of on-target off-tumor toxicity, in some embodiments, the invention

makes possible the ability to titrate down the dose of stabilizing drug such that a CAR T cell therapy recognizes a desired target (e.g., tumor antigen) but not healthy tissue.

In one embodiment, the CAR T cells of the invention undergo robust in vivo T cell expansion and persist for an extended period of time. In another embodiment, the CAR T cells of the invention evolve into specific memory T cells that can be reactivated to inhibit any additional tumor formation or growth. For example, GD2-specific CAR T cells of the invention can undergo robust in vivo T cell expansion and persist at high levels for an extended amount of time in blood and bone marrow and form specific memory T cells.

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The anti-tumor immunity response elicited by the CAR-modified T cells may be an active or a passive immune response. In addition, the CAR mediated immune response may be part of an adoptive immunotherapy approach in which CAR-modified T cells induce an immune response specific to the antigen-binding moiety in the CAR. For example, GD2-specific CAR T cells elicit an immune response specific against cells expressing GD2.

While the data disclosed herein specifically describe lentiviral vectors comprising anti-GD2 scFv (e.g. 14G2a scFv), CD28 hinge and transmembrane domain, CD28 and CD3-zeta signaling domains, and RDD of FKBP or DHFR, the invention should be construed to include any number of variations for each of the components of the construct as described elsewhere herein. That is, the invention includes the use of any antigen-binding moiety in the CAR to generate a CAR-mediated T-cell response specific to the antigen-binding moiety. For example, the antigen-binding moiety in the CAR of the invention can target a tumor antigen for the purposes of treating cancer. Similarly, the invention includes use of any hinge, transmembrane domain, stimulation domain and/or regulatable destabilization domain in a CAR to generate a CAR-mediated T-cell response. Indeed, methods described herein provide the ability to characterize and identify CAR T cells that possess desired effector functions and that are regulatable via the incorporation of an RDD into the CAR.

In one embodiment, the antigen bind moiety portion of the CAR of the invention is designed to treat a particular cancer. GD2 is expressed on a variety of cancers including neuroblastoma, osteosarcomas and some sarcomas (see, e.g., Thomas et al., PLoS One, 2016. 11(3): p. e0152196; Long et al., Nature Medicine, 2015. 21(6): p. 581-590; Long et al., Cancer Immunology Research, 2016. 4(10): p. 869-880; Yu et al., N Engl J Med, 2010. 363(14): p. 1324-34; Perez Horta et al., Immunotherapy, 2016. 8(9): p. 1097-117; Heczey et al, Molecular Therapy). Thus, CARs designed to target GD2 can be used to treat any disease or disorder, including neuroblastoma, osteosarcoma, and other sarcomas, characterized by cells and/or tissues displaying or overexpressing GD2. However, the invention is not limited

to targeting GD2. Indeed, the invention includes any antigen-binding moiety that when bound to its cognate antigen, affects a tumor cell so that the tumor cell fails to grow, is prompted to die, or otherwise is affected so that the tumor burden in a patient is diminished or eliminated. For example, FR α is a glycosylphosphatidylinositol-anchored protein that is overexpressed on the surface of cancer cells in a variety of epithelial malignancies, but is limited in normal tissue. As such, CARs designed to target FR α can be used to treat any disease or disorders, including but not limited to epithelial cancers, characterized by cells and/or tissues displaying an overexpression of FR α . For example, the CAR designed to target FR α can be used to treat cancers and disorders including but are not limited to ovarian cancer, lung cancer, breast cancer, renal cancer, colorectal cancer, other solid cancers and the like.

The CAR-modified T cells of the invention may also serve as a type of vaccine for *ex vivo* immunization and/or in vivo therapy in a mammal. Preferably, the mammal is a human. With respect to *ex vivo* immunization, at least one of the following occurs *in vitro* prior to administering the cell into a mammal: i) expansion of the cells, ii) introducing a nucleic acid encoding a CAR to the cells, and/or iii) cryopreservation of the cells.

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Ex vivo procedures are well known in the art. Briefly, cells are isolated from a mammal (preferably a human) and genetically modified (i.e., transduced or transfected *in vitro*) with a vector expressing a CAR disclosed herein. The CAR-modified cell can be administered to a mammalian recipient to provide a therapeutic benefit. The mammalian recipient may be a human and the CAR-modified cell can be autologous with respect to the recipient. Alternatively, the cells can be allogeneic, syngeneic or xenogeneic with respect to the recipient.

A procedure for *ex vivo* expansion of hematopoietic stem and progenitor cells is described in U.S. Pat. No. 5,199,942, incorporated herein by reference and can be applied to the cells of the present invention. The present invention is not limited to any particular method of *ex vivo* expansion of the cells. Briefly, *ex vivo* culture and expansion of T cells comprises: (1) collecting CD34+hematopoietic stem and progenitor cells from a mammal from peripheral blood harvest or bone marrow explants; and (2) expanding such cells *ex vivo*. In addition to the cellular growth factors described in U.S. Pat. No. 5,199,942, other factors such as flt3-L, IL-1, IL-3 and c-kit ligand, can be used for culturing and expansion of the cells.

In addition to using a cell-based vaccine in terms of *ex vivo* immunization, the present invention also provides compositions and methods for *in vivo* immunization to elicit an immune response directed against an antigen in a patient.

In particular, the CAR-modified T cells of the invention are used in the treatment of cancer. In certain embodiments, the cells of the invention are used in the treatment of patients at risk for developing cancer. Thus, the present invention provides methods for the treatment or prevention of cancer comprising administering to a subject in need thereof, a therapeutically effective amount of the CAR-modified T cells of the invention.

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CAR-modified T cells of the present invention may be administered either alone, or as a pharmaceutical composition in combination with diluents and/or with other components such as IL-2 or other cytokines or cell populations. Briefly, pharmaceutical compositions of the present invention may comprise CAR T cells as described herein, in combination with one or more pharmaceutically or physiologically acceptable carriers, diluents or excipients. Such compositions may comprise buffers such as neutral buffered saline, phosphate buffered saline and the like; carbohydrates such as glucose, mannose, sucrose or dextrans, mannitol; proteins; polypeptides or amino acids such as glycine; antioxidants; chelating agents such as EDTA or glutathione; adjuvants (e.g., aluminum hydroxide); and preservatives. Compositions of the present invention are preferably formulated for intravenous administration.

Pharmaceutical compositions of the present invention may be administered in a manner appropriate to the disease to be treated (or prevented). The quantity and frequency of administration will be determined by such factors as the condition of the patient, and the type and severity of the patient's disease, although appropriate dosages may be determined by clinical trials.

When "an immunologically effective amount", "an anti-tumor effective amount", "an tumor-inhibiting effective amount", or "therapeutic amount" is indicated, the precise amount of the compositions of the present invention to be administered can be determined by a physician with consideration of individual differences in age, weight, tumor size, extent of infection or metastasis, and condition of the patient (subject). A pharmaceutical composition comprising the CAR T cells described herein may be administered at a dosage of 10⁴ to 10⁹ cells/kg body weight, preferably 10⁵ to 10⁶ cells/kg body weight, including all integer values within those ranges. T cell compositions may also be administered multiple times at these dosages. The cells can be administered by using infusion techniques that are commonly known in immunotherapy (see, e.g., Rosenberg et al., New Eng. J. of Med. 319:1676, 1988).

The optimal dosage and treatment regime for a particular patient can readily be determined by a person of ordinary skill in the art by monitoring the patient for signs of disease and adjusting the treatment accordingly.

In certain embodiments, it may be desired to administer genetically modified T cells to a subject and then subsequently redraw blood (or have an apheresis performed), activate T cells therefrom according to the present invention, and reinfuse the patient with these activated and expanded T cells. This process can be carried out multiple times every few weeks.

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The administration of a composition described herein may be carried out in any convenient manner. In some embodiments, the T cell compositions of the present invention are preferably administered by i.v. injection. However, the administration is not limited only to this route. The compositions of T cells may be injected directly into a tumor, lymph node, or site of infection. The compositions described herein may be administered to a patient subcutaneously, intradermally, intratumorally, intranodally, intramedullary, intramuscularly, by intravenous (i.v.) injection, or intraperitoneally. Compositions may also be administered, for example, by injection, transfusion, implantation, or transplantation.

In certain embodiments of the present invention, cells activated and expanded using the methods described herein, or other methods known in the art where T cells are expanded to therapeutic levels, are administered to a patient in conjunction with (e.g., before, simultaneously or following) any number of treatment modalities (e.g., for cancer or infectious disease). Exemplary treatment modalities include, but are not limited to, treatment with agents such as antiviral therapy, cidofovir and interleukin-2, Cytarabine (also known as ARA-C) or natalizumab treatment for MS patients or efalizumab treatment for psoriasis patients. In further embodiments, the T cells of the invention may be used in combination with chemotherapy, radiation, immunosuppressive agents, such as cyclosporin, azathioprine, methotrexate, mycophenolate, and FK506, antibodies, or other immunoablative agents such as CAM PATH, anti-CD3 antibodies or other antibody therapies, cytoxin, fludaribine, cyclosporin, FK506, rapamycin, mycophenolic acid, steroids, FR901228, cytokines, and irradiation. These drugs inhibit either the calcium dependent phosphatase calcineurin (cyclosporine and FK506) or inhibit the p70S6 kinase that is important for growth factor induced signaling (rapamycin) (Liu et al., Cell 66:807-815, 1991; Henderson et al., Immun. 73:316-321, 1991; Bierer et al., Curr. Opin. Immun. 5:763-773, 1993). In a further embodiment, the cell compositions of the present invention are administered to a patient in conjunction with (e.g., before, simultaneously or following) bone marrow transplantation, T

cell ablative therapy using either chemotherapy agents such as, fludarabine, external-beam radiation therapy (XRT), cyclophosphamide, or antibodies such as OKT3 or CAMPATH. In another embodiment, the cell compositions of the present invention are administered following B-cell ablative therapy such as agents that react with CD20, e.g., Rituxan. For example, in one embodiment, subjects may undergo standard treatment with high dose chemotherapy followed by peripheral blood stem cell transplantation. In certain embodiments, following the transplant, subjects receive an infusion of the expanded immune cells of the present invention. In an additional embodiment, expanded cells are administered before or following surgery.

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The efficacy of any of the methods described herein may be tested in various models known in the art, such as clinical or pre-clinical models. Suitable pre-clinical models are exemplified herein. For any exemplary model, after developing tumors, mice are randomly recruited into treatment groups receiving treatment or control treatment. Tumor size (e.g., tumor volume) is measured during the course of treatment, and overall survival rate is also monitored.

In some embodiments, a sample is obtained prior to treatment with T cells (e.g., alone or in combination with another therapy described herein) as a baseline for measuring response to treatment. In some embodiments, the sample is a tissue sample (e.g., formalinfixed and paraffin-embedded (FFPE), archival, fresh or frozen). In some embodiments, the sample is whole blood. In some embodiments, the whole blood comprises immune cells, circulating tumor cells and any combinations thereof.

Responsiveness to treatment may refer to any one or more of: extending survival (including overall survival and progression free survival); resulting in an objective response (including a complete response or a partial response); or improving signs or symptoms of cancer. In some embodiments, responsiveness may refer to improvement of one or more factors according to the published set of RECIST guidelines for determining the status of a tumor in a cancer patient, i.e., responding, stabilizing, or progressing. For a more detailed discussion of these guidelines, see Eisenhauer et al., Eur J Cancer 2009;45: 228-47; Topalian et al., N Engl J Med 2012;366:2443-54; Wolchok et al., Clin Can Res 2009;15:7412-20; and Therasse, P., et al. J. Natl. Cancer Inst. 92:205-16 (2000). A responsive subject may refer to a subject whose cancer(s) show improvement, e.g., according to one or more factors based on RECIST criteria. A non-responsive subject may refer to a subject whose cancer(s) do not show improvement, e.g., according to one or RECIST criteria.

Conventional response criteria may not be adequate to characterize the anti-tumor activity of immunotherapeutic agents, which can produce delayed responses that may be preceded by initial apparent radiological progression, including the appearance of new lesions. Therefore, modified response criteria have been developed that account for the possible appearance of new lesions and allow radiological progression to be confirmed at a subsequent assessment. Accordingly, in some embodiments, responsiveness may refer to improvement of one of more factors according to immune-related response criteria2 (irRC). See, e.g., Wolchok et al., Clin Can Res 2009; 15:7412-20. In some embodiments, new lesions are added into the defined tumor burden and followed, e.g., for radiological progression at a subsequent assessment. In some embodiments, presence of non-target lesions are included in assessment of complete response and not included in assessment of radiological progression. In some embodiments, radiological progression may be determined only on the basis of measurable disease and/or may be confirmed by a consecutive assessment >4 weeks from the date first documented.

The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The foregoing embodiments are therefore to be considered in all respects illustrative rather than limiting the invention described herein. Scope of the invention is thus indicated by the appended claims rather than by the foregoing description, and all changes that come within the meaning and range of equivalency of the claims are intended to be embraced therein. All publications, patents, and patent applications cited herein are hereby incorporated by reference in their entirety for all purposes.

EXAMPLES

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The following examples illustrate but do not limit the compounds, compositions, and methods of the present invention. Other suitable modifications and adaptations of the variety of conditions and parameters normally encountered in clinical therapy and which are obvious to those skilled in the art are within the spirit and scope of the invention.

Example 1

A Method of Preventing or Reversing T Cell Exhaustion by Inhibiting or Modulating TCR Signaling

Introduction

We previously reported that GD2-CAR expressing T cells develop functional

exhaustion within 10 days in culture and are characterized by co-expression of inhibitory receptors, failure to secrete cytokines in response to tumor antigen, and aberrant metabolic function (Long et. al, Nat Med 2015). Control cultures included untransduced T cells (mock) and those expressing CD19-CAR, which does not manifest tonic signaling or develop exhaustion *in vitro*. Previous work also demonstrated that the zeta chain was required for exhaustion in this system, with CD28 signaling enhancing the potency of the signaling stimulus in inducing exhaustion. Using this model system, we have now optimized a robust, manipulatable, and reproducible *in vitro* human model of T cell exhaustion to evaluate approaches to prevent or reverse T cell exhaustion.

Results

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We engineered a GD2.28z CAR fused to an FKBP12 mutant destabilization domain (Banaszynski et. al, Cell 2006) (GD2.28z.FKBP) which confers its instability to the CAR and induces rapid protein degradation. We observed that surface expression could be rapidly and dose-dependently regulated by adding or subtracting the stabilizing rapalog shield-1 (S1) in culture medium (Fig. 1). Similar regulatability of CAR expression was also accomplished using an E. coli DHFR mutant (GD2.28z.DHFR, not shown), which could be regulated by trimethoprim, an antibiotic that is commonly used clinically.

Since tonic signaling is highly dependent upon GD2-CAR receptor levels, precise control of CAR expression levels also precisely regulates levels of tonic signaling. Drug regulated control of levels of CAR expression therefore also allowed modulation of the duration and intensity of GD2.28z tonic signaling. Using this system, we demonstrated that phenotypic and functional changes associated with exhaustion were reversed upon cessation of CAR signaling. As shown in FIG. 2, removal of S1 drug from the culture medium and consequent removal of surface CAR on day 7 post-activation reverses canonical exhaustion marker expression to control levels by day 10 (Fig. 2, n=3). This is most well illustrated by measuring levels of PD-1/TIM-3/LAG-3 triple expressing cell which is highly specific for dysfunctional, exhausted T cells. We demonstrate that Day 10 clear induces increases in levels of triple expressing exhausted cells, but that removal of S1 on Day 7 results in normalization of these levels by Day 10. Similar results were obtained on day 14 for cells in which S1 was removed from culture medium on day 7 or day 10 (not shown).

Additionally, removal of S1 on day 7 or 10, allow transient degradation of CAR proteins results in maintenance of memory markers (ex. CD62L) and prevention of apoptosis (i.e., annexin V staining) by day 14 compared to T cells that received S1 for the entire duration of the culture (S1) (Fig. 3).

Because phenotypic markers may not be entirely predictive of T cell function, we also performed functional experiments on CAR T cells provided transient drug exposure in culture. CAR T cells were washed, resuspended in media containing S1, and mixed at a 1:1 ratio with Nalm6 leukemic cells stably expressing surface GD2. Culture supernatants were harvested approximately 24 hours later and cytokine levels were evaluated via ELISA.

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Similar to GD2.28z CAR that lacks a destabilization domain and therefore have persistent high levels of CAR signaling, cells expressing the GD2.28z.FKBP CAR that experienced continuous drug treatment (Fig. 4, grey bars) secreted minimal amounts of IL-2 on both day 10 and day 14 post-activation, consistent with T cell exhaustion. Alternatively, CAR T cells that were not exposed to drug during culture (black bars) and therefore did not experience tonic signaling demonstrated significant bioactivity as measured by IL-2 production. Finally, CAR T cells that were exposed to drug during the initial 7 or 10 days of culture and therefore acquired phenotypic and functional evidence of T cell exhaustion, but had drug removed from the culture medium on day 7 or day 10 (blue and red bars, respectively) displayed a restored capacity to secrete IL-2 in response to tumor antigen.

Remarkably, exhausted T cells on day 10 (grey bar, day 10 ELISA) could be reinvigorated by removing S1 from the culture medium and "rested" for only 4 days (red bar, day 14 ELISA). Similar, but less dramatic augmentation of IFNγ secretion in conditions in which S1 was removed from culture medium was also observed. These functional data cannot be attributed to differential CAR surface expression, as all groups exhibited similar levels of surface CAR at the conclusion of this co-culture assay (not shown).

We then compared whether prevention or reversal of T cell exhaustion by removal of surface CAR was more or less potent than treatment with well-characterized anti-PD-1 checkpoint inhibitor, nivolumab (Nivo). CAR T cells were either treated with continuous S1 (and thus exhibit continuous tonic signaling), continuous S1 + nivolumab, or no S1 until the time of the co-culture assay. Interestingly, nivolumab treatment resulted in only modest augmentation of IL-2 secretion at day 10, which was sustained until day 14, suggesting that nivolumab only partially prevented the onset of T cell exhaustion in this system (Fig. 5).

Conversely, culturing CAR T cells without S1, then adding it back to the medium just prior to the co-culture assay (left chart, blue bars), resulted in a far superior prevention of exhaustion, as IL-2 secretion was augmented 5-10 fold compared to CAR T cells that experienced continuous S1 (black bars). Further, removing tonic signaling on day 7 by removing S1 from the culture medium also resulted in superior IL-2 secretion compared to

CAR T cells that experienced continuous S1, and those that experienced continuous S1 and were simultaneously treated with S1. Collectively, these data demonstrate that modulating tonic signaling exhibits more potent effects on prevention or reversal of exhaustion compared to PD-1 blockade.

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Functional studies by several groups, including our lab have verified that co-expression of PD-1, TIM-3, and LAG-3 (triple positive, TP) denotes an exhausted cell subset that is highly dysfunctional. We thus sought to analyze whether cessation of tonic signaling in this cell subset could reverse their phenotype and restore their ability to secrete IL-2 in response to tumor antigen. A high affinity version of our GD2.28z CAR (HA-GD2.28z), which exhibits an even more dramatic exhausted phenotype, was fused to the FKBP12 mutant destabilization domain in order to control its surface expression. On day 10 post-activation, HA-GD2.28z.FKBP CAR T cells that had experienced continuous S1 treatment were sorted in order to isolate a pure PD-1/TIM-3/LAG-3 exhausted population. "Triple positive" exhausted cells were then re-cultured either with or without S1 to test whether removal of tonic signaling could restore their function. FACS and co-culture assays were conducted 4 days later.

Removal of S1 resulted in a dramatic reversal of the exhausted phenotype. After only 4 days without S1 in the medium, pre-sorted triple positive cells exhibited far less expression of exhaustion markers in both CD4+ and CD8+ CAR T cells (Fig. 6). Importantly, these phenotypic changes also conferred functional augmentation in IL-2 secretion, as removal of S1 resulted in a 2-fold increase in IL-2 secretion compared to triple positive cells that received continuous S1 treatment from days 10-14 (Fig. 6).

We hypothesized that we could recapitulate the effects of removing surface CAR, and thus tonic signaling, by simply inhibiting kinases in the TCR signaling pathway that are also integral to CAR signaling. One such kinase is Lck, which acts to phosphorylate CD3-zeta in response to TCR or CAR ligation. Dasatinib, a potent receptor tyrosine kinase inhibitor and BCR/ABL antagonist, has also been shown to inhibit T cell activation, proliferation, and cytokine secretion by binding to and inhibiting Lck at low concentrations (Schade et. al, Blood, 2008 and Lee et. al, Leukemia, 2010).

At 100 nM and 1μ M concentrations, dasatinib potently inhibits CD19.28z CAR T cell cytokine secretion in response to tumor antigen on day 14 post-activation (Fig. 7), proving that dasatinib disrupts CAR signaling.

We then asked whether transient dasatinib exposure could reverse T cell exhaustion

by treating HA-GD2.28z CAR T cells with dasatinib on days 10-14 post-activation. Cells were treated with dasatinib for 4 days, then drug was extensively washed from the media, and cells were re-cultured for an additional 24 hours before examining their phenotype and function via FACS and tumor co-culture assays. Interestingly, 4-day treatment with dasatinib reversed exhaustion marker expression and co-expression in a dose-dependent manner (Fig. 8).

Furthermore, dasatinib treatment resulted in preservation of T cell memory via maintenance of CD62L expression in a dose-dependent manner (Fig 9.).

Finally, similar to removal of surface CAR, dasatinib treatment reinvigorated exhausted T cells in a functionally significant manner, as dasatinib-treated CAR T cells secreted more IL-2 (and to a lesser extent, IFN γ) in response to tumor antigen compared to those that never received dasatinib (Fig. 10).

Collectively, these data demonstrate that selective inhibition or modulation of TCR signaling can substantially enhance the function of exhausted T cells that experience continuous antigen exposure in the context of cancer or chronic infection. In future studies, we will conduct *in vivo* studies to assess the feasibility of exhaustion reversal in this setting and whether such reversal can enhance antitumor effects in murine models.

Example 2

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20 Prevention of CAR T cell exhaustion and enhancement of CAR T cell effector function via introduction of a destabilization domain into the CAR

Experiments were conducted in order to test whether it would be possible to control CAR T cell activity by tuning CAR T cell surface expression, thus offering a method by which CAR T cells could be turned "off" in patients experiencing toxicity without completely eliminating the cells (in stark contrast to a suicide switch). Experiments were also conducted during development of embodiments of the invention in order to determine whether expansion of CAR T cells in the absence of surface CAR *in vitro* would allow mitigation of CAR tonic signaling, which may yield a healthier, more efficacious infusion product. Finally, experiments were conducted in order to test whether regulation of CAR surface expression using regulatable, drug-sensitive destabilization domains would allow for prevention or reversal of T cell exhaustion and/or maintenance/induction of T cell memory.

Materials and Methods

Cells and culture conditions

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NALM6-GL (acute lymphoblastic leukemia line, stably transfected with GFP and luciferase) and NALM6-GL-GD2 (stably transfected to overexpress GD2 synthetase) cell lines were cultured in RPMI-1640. 293T and 143B cell lines were cultured in DMEM (Life Technologies). DMEM and RPMI-1640 were supplemented with 10% heat-inactivated FBS (Gibco, Life Technologies), 10mM HEPES, 100U/mL penicillin, 100 µg/ml streptomycin and 2mM L-glutamine (Gibco, Life Technologies).

Primary human T cells were obtained from healthy donor buffy coats using a Pan T cell negative selection kit (Miltenyi Biotec). Donor T cells were then aliquoted and stored in Cryostor (StemCell Technologies) in liquid nitrogen. T cells were cultured in AimV (Gibco, Life Technologies) supplemented with 5% heat-inactivated FBS, 10mM HEPES, 1% glutamax (Gibco, Life Technologies), and 100u/uL recombinant human IL-2 (Peprotech). Shield-1 and trimethoprim lactate (Sigma Aldrich or Alfa Aesar) were cultured at 1μM unless otherwise specified.

Lentiviral production and T cell transduction

All DD-CAR lentiviral supernatants were produced via transient transfection of the 293T cell line. Briefly, 293T cells were transfected via Lipofectamine 2000 (Life Technologies) with the plasmids encoding the CARs and plasmids encoding packaging proteins Gag-Pol, REV, and envelope protein VSVG. Supernatants were collected at 24 and 48 hours post-transfection and either immediately frozen or first concentrated via ultracentrifuge spin at 30,000 RPM and then frozen and stored at -80C.

Upon thawing, T cells were activated at a 3:1 bead:cell ratio using anti-CD3/anti-CD28-coated magnetic beads (Dynabeads, Thermo Fisher) at a concentration of $1x10^6$ cells/mL. On day 1 post activation, lentivirus was added directly to the T cells. On day 4 post-activation, magnetic beads were removed from culture, and T cells were cultured at $0.5x10^6$ cells/mL every day thereafter. Media supplemented with IL-2 and stabilizing drug was changed every two days. Transduction efficiencies were routinely 70–90% for all CARs.

Flow Cytometry

All samples were analyzed with an LSR Fortessa (BD Bioscience) or a Cytoflex (Beckman Coulter) and data were analyzed using FlowJo. Cells were washed twice with PBS and labelled with stain at $1x10^6$ cells/mL in PBS, followed by two washes with FACS buffer (PBS supplemented with 2% FBS and 0.4% 0.5M EDTA). GD2 CARs were detected with the 14g2a anti-idiotype antibody (clone 1A7). CD19 CARs were detected with the FMC63

anti-idiotype antibody (clone 136.20.1). T cell phenotype was evaluated via: CD4 (OKT4, Biolegend), CD8 (SK1, Biolegend), PD-1 (eBioJ105, eBioscience), TIM-3 (F38-2E2, Biolegend), LAG-3 (3DS223H, eBioscience), CD45RO (UCHL1, eBioscience), CCR7 (150503, BD Biosciences), CD69 (FN50, Biolegend), IL-2 (MQ1-17H12, Biolegend), and IFNγ (4S.B3, Biolegend). For co-culture assays in which cytokine production was assessed, tumor cells and CAR T cells were co-cultured in the presence of 1:1000 monensin (eBioscience) for at least 6 hours. When assessing IL-2 and IFNy, cells were surface stained, then fixed and permeablized (eBioscience) prior to incubation with intracellular antibodies. All FACS plots displaying CAR T cell phenotype data were pre-gated on CAR+ cells. For mock-transduced T cells, whole T cell populations were used for analysis.

CyTOF

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T cells were removed from culture, fixed in 2% PFA for 10 minutes at room temperature, pelleted and frozen at -80C. Upon thawing, each sample was barcoded with a 20-plex Pd barcoding kit (Fluidigm). Cell samples were then pooled and stained for surface marker expression with heavy-metal conjugated antibodies for 30 minutes at room temperature. Cells were then permeablized in methanol and subsequently stained with heavy metal-conjugated anti-human T-bet (4B10, Biolegend) and anti-human Blimp-1 (ROS195G, Biolegend) for 30 minutes at room temperature. Samples were run on a Helios mass cytometer (Fluidigm) and analyses were complete using Cytobank online software.

Incucyte assay

50,000 NALM6-GL-GD2 tumor cells were co-cultured with T cells at a 1:2 or 1:8 E:T ratio in 200uL of complete AimV medium without IL-2 supplementation in each well of a 96-well plate. Plates were loaded into the incucyte and 488nm fluorescent images were acquired every 2 hours for 48-72 hours. GFP+ tumor cells were identified by size and fluorescence intensity masks, and the total integrated GFP intensity of all counted tumor cells was quantified for each individual well. Values were normalized to t=0, and replicate wells were averaged for data display.

For experiments in which HA-GD2.28z.FKBP T cells were expanded in the absence of shield-1, 1μ M shield-1was added to CAR T cells 18-24 hours prior to co-culture with tumor cells.

Cytokine release assay

50,000 NALM6-GL-GD2 tumor cells were co-cultured with T cells at a 1:1 E:T ratio in 200µL of complete AimV medium without IL-2 supplementation in each well of a 96-well

plate. After 24 hours, supernatants were removed and stored at -20C. IL-2 and IFNγ secretion was assessed via ELISA (Biolegend).

For experiments in which HA-GD2.28z.FKBP T cells were expanded in the absence of shield-1, 1μ M shield-1was added to CAR T cells 18-24 hours prior to co-culture with tumor cells.

Western Blot

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2x10⁶ CAR-T cells were removed from culture, pelleted, and resuspended in 100uL of RIPA lysis buffer (10mM Tris-Cl pH 8.0, 1mM EDTA, 1% Triton X-100, 0.1% sodium deoxycholate, 0.1% SDS, 140mM NaCl) supplemented with phosphatase and protease inhibitors (Thermo Fisher). After incubating for 30 minutes at 4C, supernatants were cleared by centrifugation at 14,000 RPM for 20 minutes at 4C. Protein concentration in the cleared lysates was measured by a colorimetric reaction (BioRad).

15μg of protein lysate was mixed with 6x loading buffer and loaded onto 10% SDS-PAGE gels assembled into a mini-protean electrophoresis systems (BioRad). Electrophoresis was performed in tris-glycine-SDS buffer (BioRad) at 100V for 20 minutes and later increased to 150V for 50 minutes. Protein transfer into Immobilion-FL PVDF membranes was performed at 100V for 1 hour in tris-glycine buffer (BioRad # 1610771). Primary antibodies targeting CD3-zeta (Cell signaling) and pY142-CD3-zeta (Cell Signaling) were used. The Odyssey (LI-COR) imaging system, LI-COR buffers, and LI-COR secondary antibodies (Goat Anti-Mouse IgG Antibody-800CW-Conjugated and Goat Anti-Rabbit IgG Antibody-680LT-Conjugated) were used for protein detection.

In vivo experiments

6-8 week old NSG mice were engrafted with 1x10⁶ NALM6-GL-GD2 via intravenous injection. At day 7 post-engraftment, 1-5x10⁶ GD2.28z.DHFR or HA-GD2.28z.DHFR

25 CAR+ T cells were infused intravenously. NALM6-GL-GD2 tumor burden was evaluated using the Xenogen IVIS Lumina (Caliper Life Sciences). Mice were first injected intraperitoneally with 3 mg D-luciferin (Caliper Life Sciences) and then imaged 4 minutes later with an exposure time of 30 seconds, or, in cases where 30 seconds resulted in signal saturation, "auto" exposure was selected. Luminescence images were analyzed using Living

30 Image software (Caliper Life Sciences).

Mice treated with trimethoprim were injected intraperitoneally at a concentration of 300mg/kg. Mice treated with vehicle were injected with an equivalent volume of water intraperitoneally.

Blood samples were taken via retro-orbital bleed and briefly stored in EDTA-coated microvettes (Kent Scientific). Spleens were mechanically disaggregated by passage through a 70-µm filter (BD Biosciences). Both blood and spleen were lysed in ACK lysis buffer (Fisher Scientific) for 5 minutes and subsequently stained with surface marker antibodies for FACS analysis.

Construction of CAR vectors

All CAR sequences were inserted into the pELNS lentiviral backbone under the control of an EF-1 alpha promotor. Destabilization domain sequence insertion downstream of the CD3z domain was accomplished by subcloning a custom gene fragment (IDT Technologies) which included a portion of the CD3z sequence (beginning with the intrinsic BmgBI restriction site) followed by the destabilization domain sequence, a stop codon, and a SalI restriction site. The additional restriction sites not present in either the CD3z or DD sequences was avoided.

Each CAR includes a signal peptide, single chain variable fragment (scFv), extracellular hinge region, transmembrane domain, intracellular co-stimulatory domain, and intracellular CD3-zeta domain.

Results

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First, the dose dependent regulation of DD-CAR surface expression was tested for 3 different CARs fused with two different RDDs. CD19.28z, GD2.28z, and a high-affinity GD2.28z (HA-GD2.28z) were each fused to a 12 kDa FK506 binding protein (FKBP) DD or an E. coli-derived dihydrofolate reductase (DHFR) DD. In separate experiments, regulation of a Her2.28z CAR fused to the FKBP DD (Figure 24) was also tested. The rapalog shield-1 (S1) was used to stabilize FKBP-fused CARs, while the FDA-approved antibiotic trimethoprim (TMP) was used to stabilize DHFR-fused CARs. Stabilizing drug was incubated with DD-CAR T cells for at least 48 hours prior to assessing surface CAR expression via FACS. The data demonstrate precise dose-dependent regulation of surface CAR for all 7 of the CARs tested (See FIGS. 12, 24). The differential EC₅₀ values of the DD-CARs (FIG. 22B) demonstrate heterogeneity in the sensitivity of a given DD-CAR protein to the stabilizing drug, regardless of the DD to which the CAR was fused. These studies also demonstrate intrinsic differences in DDs, as each DHFR-fused CAR demonstrated greater sensitivity to stabilizing drug compared to its FKBP-fused counterpart (see FIG. 22). Furthermore, it was observed that not all RDDs confer instability to the CAR protein, even if they do so for other proteins (see FIG. 23). Two previously published iterations of the DHFR DD (see Iwamoto et al., Chemistry & Biology 17, 981–988 (2010)) were tested on the

GD2.28z CAR, and one of them allowed for drug-dependent regulation (See FIGS. 12, 22, 23).

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Next, the kinetics of the drug-dependent regulation of DD-CARs was tested and characterized. DD-CAR T cells were cultured either in the presence or absence of 1uM stabilizing drug for at least 48 hours. For DD-CAR T cells cultured in the absence of drug, drug was introduced into the culture and samples removed over the course of 48 hours to assess the increase in CAR surface expression (See FIG. 13, left side of each plot). Conversely, for DD-CAR T cells cultured in the presence of drug, we removed drug from the culture to assess the decrease in CAR surface expression (See FIG. 13, right side of each plot). These studies indicated that DD-CAR surface expression increases gradually over the course of 24 hours subsequent to exposure to drug. However, removal of drug induced a rapid loss of approximately half of the DD-CAR surface expression within the first 2 hours, then a gradual loss of the remaining surface CAR over the next 46 hours. The rapid "off" kinetics of DD-CARs indicated that this method of CAR regulation provides a realistic therapeutic opportunity to mitigate CAR toxicity in a clinical setting.

It was next determined whether the differences in DD-CAR surface expression would elicit differential anti-tumor responses. Therefore, it was assessed whether drug-dependent regulation of DD-CAR surface expression would also allow for regulation of T cell activation. Indeed, when DD-CAR T cells were co-cultured with antigen-bearing tumor cells in the absence of drug, negligible levels of cytokine secretion (see FIG. 14A) and cytotoxicity were observed (see FIG. 14B) compared to DD-CAR T cells co-cultured in the presence of stabilizing drug. Experiments were also conducted in an *in vivo* setting.

Mice were engrafted with antigen-bearing tumor and subsequently infused with CAR T cells. Mice were then injected with either stabilizing drug or vehicle control every day thereafter. After only 1 dose of stabilizing drug, drug-dependent upregulation of CAR expression was observed similar to that observed *in vitro* (See FIG. 15A). Higher levels of CD69 in mice dosed with stabilizing drug were also observed, indicating that the DD-CAR T cells with higher surface expression became more activated in response to tumor. Finally, after 7 days post-CAR infusion, a dramatic reduction in tumor burden was observed in mice receiving stabilizing drug versus those receiving vehicle control. Collectively, these data provide that CAR surface expression is intimately linked to CAR T cell activity, and thus, RDD-CAR T cell activity can be precisely tuned via administration (or lack thereof) of stabilizing drug.

As described herein, many constitutively expressed CARs exhibit tonic signaling in

the absence of antigen, consequently driving them to become exhausted (see Long et al., Nature Medicine 21, 581–590 (2015)). It was determined whether expanding DD-CAR T cells in the absence of stabilizing drug would alleviate this tonic signaling and consequently yield a healthier, more potent CAR T cell infusion product. One readout for tonic signaling is the constitutive phosphorylation of the CAR CD3-zeta domain in the absence of antigen. HA-GD2.28z.FKBP CAR T cells exhibited this phenotype when cultured in the presence of S1 (see FIG. 16, CAR Always ON). However, CD3-zeta phosphorylation was absent in DD-CAR T cells cultured in the absence S1 (CAR Always OFF), indicating that low CAR surface expression mitigates tonic signaling. This phenotype was also reversible, as indicated by the lack of CD3-zeta phosphorylation in DD-CAR T cells that were initially cultured with S1, but then had drug removed on day 7 of the culture (FIG. 16, CAR OFF D7).

It was next determined whether the prevention or reversal of tonic signaling in HA-GD2.28z.FKBP-expressing CAR T cells altered their exhaustion phenotype. DD-CAR T cells expanded in the presence of drug (CAR Always ON) exhibited co-expression of multiple inhibitory receptors (see FIG. 17A) and transcription factors commonly associated with T cell exhaustion (FIG. 17B). However, DD-CAR T cells expanded in the absence of drug (CAR Always OFF) or those expanded in drug from days 1-7 but not thereafter (CAR OFF D7) exhibited significantly less exhaustion marker co-expression (FIG. 17). Similarly, exhausted DD-CAR T cells expanded in the presence of drug displayed a high frequency of effector T cells and concomitant low frequency of memory cells (FIG. 18), similar to that observed in the literature (see Wherry and Kurachi, Nature Reviews Immunology 15, nri3862 (2015). However, a rescue in DD-CAR T cell memory was observed simply by expanding the cells in the absence of stabilizing drug for at least 4 days, indicating that mitigation of tonic signaling in CAR T cells can simultaneously prevent exhaustion and facilitate memory formation.

Next, it was determined whether the dramatic phenotypic changes observed when tonic signaling was mitigated may also coincide with an augmentation in T cell function. To test this, DD-CAR T cells were expanded either in the presence (CAR Always ON) or absence (CAR Always OFF) of stabilizing drug. To interrogate the plasticity of exhausted CAR T cells to become reinvigorated, cells were expanded in the presence of drug for either 7 or 10 days, after which drug was removed and T cells were "rested" (CAR OFF D7 and CAR OFF D10, respectively). CAR T cells were then co-cultured with antigen-bearing tumor for 24 hours and cytokine secretion was assessed via ELISA (FIG. 19) or they were co-cultured for 72 hours in an incucyte during which cytotoxicity was assessed (FIG. 20). Importantly, regardless of the conditions in which DD-CAR T cells were cultured, drug was

added to each condition 18-24 hours prior to the co-culture assay to ensure normalization of CAR expression during the assay. CAR T cells expanded with drug (CAR Always ON) secreted low levels of IL-2 and IFNγ in response to tumor (FIG. 19) and exhibited impaired cytotoxicity (FIG. 20), consistent with the phenotype of exhausted T cells. Conversely, CAR T cells grown in the absence of drug secreted 2-10 fold more cytokine (FIG. 19) and were capable of limiting tumor growth (FIG. 20), indicating that mitigation of tonic signaling profoundly augmented the functional capacity of CAR T cells. Furthermore, CAR T cells that experienced tonic signaling for 7 or 10 days, but were provided 4-7 days of "rest" prior to the co-culture assay, were functionally reinvigorated (FIGS. 19 and 20, CAR OFF D7 and CAR OFF D10). These observations indicated that DD-CAR T cells benefit from periods of "rest", during which DD-CAR expression is low and cells are allowed to quiesce and form memory.

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Based on the *in vitro* functional studies, it was determined whether toggling DD-CAR T cell expression *in vivo* could provide an opportunity to prolong the CAR T cell anti-tumor response by limiting chronic antigen exposure and mitigating CAR T cell exhaustion. To test this, antigen-bearing tumor cells were engrafted into NSG mice and subsequently infused 2-5E6 HA-GD2.28z.DHFR-expressing T cells. Mice were dosed with TMP everyday thereafter.

A rapid anti-tumor response was observed upon infusion of the DD-CAR, but eventually the disease relapsed, suggesting that the CAR T cells may have become dysfunctional or exhausted (see FIG. 21). Mice were then dosed with vehicle for 4 days to provide a period during which DD-CAR T cells could recover (see FIG. 21, "rest"). During this time period, tumor burden increased at a more rapid rate compared to the previous period of TMP dosing, indicating that, in fact, the anti-tumor response was being tuned down. Upon reinfusion of TMP for 3 days (FIG. 21, "restim"), a profound reinvigoration of the anti-tumor response was observed in all mice tested (FIG. 21), similar to the observations *in vitro* (FIGS. 19, 20). Furthermore, 2 out of 3 mice exhibited a lower tumor burden after the reinfusion of TMP than before the period of rest, indicating that the benefits of iterative drug dosing may outweigh the temporary acceleration of tumor growth during vehicle infusion.

All publications and patents mentioned in the above specification are herein incorporated by reference. Various modifications and variations of the described method and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed

should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention that are obvious to those skilled in relevant fields are intended to be within the scope of the following claims.

CLAIMS

- 1. A chimeric antigen receptor (CAR) comprising:
 - a) an extracellular ligand-binding domain;
 - b) a transmembrane domain;
 - c) a cytoplasmic domain comprising one or more signaling domains; and
- d) a regulatable destabilization domain (RDD), wherein the RDD comprises a dihydrofolate reductase destabilization domain (DHFR DD) or an FK506 binding protein 12 (FKBP) destabilization domain (FKBP DD) comprising the following amino acid substitutions: E31G, F36V, R71G and K105E.
- 2. The CAR of claim 1, wherein the extracellular ligand-binding domain specifically binds a tumor antigen.
- 3. The CAR of claim 2, wherein the tumor antigen is disialoganglioside GD2, CD19, or Her2.
- 4. The CAR of claim 1, wherein the extracellular ligand-binding domain comprises a single chain variable fragment (scFv) domain.
- 5. The CAR of claim 4, wherein the scFv comprises the variable regions of the heavy (V_H) and light chains (V_L) of an antibody binding specifically to disialoganglioside GD2.
- 6. The CAR of claim 5, wherein the antibody binding specifically to disialoganglioside GD2 is selected from the group consisting of 14G2a, ch14.18, hu14.18K322A, m3F8, hu3F8-IgG1, hu3F8-IgG4, HM3F8, UNITUXIN, and DMAb-20.
- 7. The CAR of claim 1, wherein the transmembrane domain is a CD28 transmembrane domain.
- 8. The CAR of claim 1, wherein the cytoplasmic domain comprises a 4-1BB signaling domain.

- 9. The CAR of claim 1, wherein the cytoplasmic domain comprises a CD28 signaling domain.
- 10. The CAR of claim 1, wherein the cytoplasmic domain comprises a CD3-zeta signaling domain.
- 11. The CAR of claim 1, wherein the cytoplasmic domain comprises a 4-1BB signaling domain and a CD3-zeta signaling domain.
- 12. The CAR of claim 1, wherein the RDD comprises the dihydrofolate reductase (DHFR) destabilization domain (DHFR DD).
- 13. The CAR of claim 12, wherein the RDD comprises a human DHFR destabilization domain.
- 14. The CAR of claim 1, wherein the RDD comprises the FK506 binding protein 12 (FKBP) destabilization domain (FKBP DD).
- 15. The CAR of claim 1, wherein the RDD comprises a human FK506 binding protein 12 (FKBP) destabilization domain (FKBP DD).
- 16. The CAR of claim 1, wherein the extracellular ligand-binding domain comprises the amino acid sequence of SEQ ID NO: 76.
- 17. The CAR of claim 1, wherein the transmembrane domain comprises the amino acid sequence of SEQ ID NO: 48.
- 18. The CAR of claim 1, wherein the signaling domain comprises a CD28 domain comprising the amino acid sequence of SEQ ID NO: 50.

- 19. The CAR of claim 1, wherein the signaling domain comprises a CD28 domain comprising the amino acid sequence of SEQ ID NO: 50 and a CD3-zeta domain comprising the amino acid sequence of SEQ ID NO: 52.
- 20. The CAR of claim 10, wherein the CD3-zeta signaling domain comprises the amino acid sequence of SEQ ID NO: 52.
- 21. The CAR of claim 1, wherein the DHFR DD comprises the amino acid sequence of SEQ ID NO: 70.
- 22. The CAR of claim 1, wherein the chimeric antigen receptor comprises the amino acid sequence of SEQ ID NO: 56.
- 23. The CAR of claim 2, wherein the CAR exhibits antigen-independent tonic signaling.
- 24. A genetically modified T cell comprising a nucleic acid sequence encoding the CAR of any one of claims 1-23.
- 25. The genetically modified T cell of claim 24, wherein the nucleic acid sequence is human, mouse, or humanized mouse nucleic acid sequence.
- 26. The genetically modified T cell of claim 24, wherein the nucleic acid sequence comprises the nucleotide sequence of SEQ ID NO: 39 for encoding the extracellular ligand-binding domain.
- 27. The genetically modified T cell of claim 24, wherein the nucleic acid sequence comprises the nucleotide sequence of SEQ ID NO: 47 for encoding the transmembrane domain.
- 28. The genetically modified T cell of claim 24, wherein the nucleic acid sequence comprises the nucleotide sequence of SEQ ID NO: 49 for encoding the signaling domain.

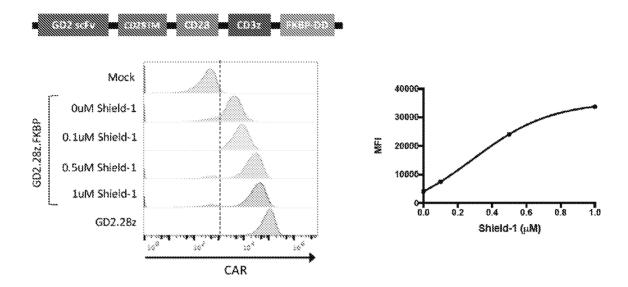
- 29. The genetically modified T cell of claim 24, wherein the nucleic acid sequence comprises the nucleotide sequence of SEQ ID NO: 49 for encoding the signaling domain comprising a CD28 domain and the nucleotide sequence of SEQ ID NO: 51 for encoding a CD3-zeta domain.
- 30. The genetically modified T cell of claim 24, wherein the nucleic acid sequence comprises the nucleotide sequence of SEQ ID NO: 51 for encoding a CD3-zeta domain.
- 31. The genetically modified T cell of claim 24, wherein the nucleic acid sequence comprises the nucleotide sequence of SEQ ID NO: 69 for encoding DHFR DD.
- 32. The genetically modified T cell of claim 24, wherein the chimeric antigen receptor comprises the amino acid sequence of SEQ ID NO: 55.
- 33. A method of prolonging a T lymphocyte cell (T cell) effector function in a mammal comprising introducing into the mammal a T cell comprising a chimeric antigen receptor of any one of claims 1 to 23.
- 34. A method for treating a mammal suffering from cancer comprising introducing into the mammal the genetically modified T cell of any one of claims 24 to 32 for treating cancer in the mammal.
- 35. A method for stimulating a T cell-mediated immune response to a target cell population or tissue in a mammal comprising administering to a mammal an effective amount of a T cell genetically modified to express a chimeric antigen receptor (CAR) of any one of claims 1 to 23.
- 36. A method of providing an anti-cancer immune response in a mammal, the method comprising administering to the mammal an effective amount of a T cell genetically modified to express a CAR of any one of claims 1 to 23.
- 37. The method of claim 36, wherein the cancer is selected from the group consisting of neuroblastoma, glioblastoma, midline glioma, osteosarcomas, sarcoma, B lineage acute

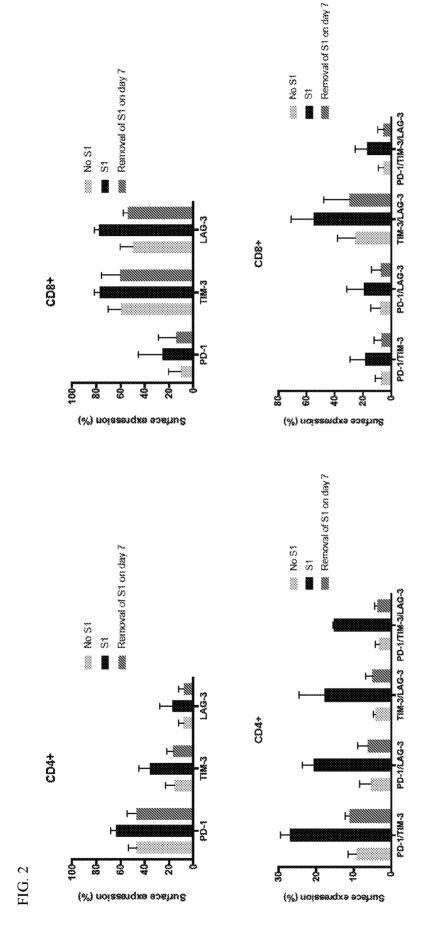
lymphoblastic leukemia, B-cell chronic lymphocytic leukemia, B-cell non-Hodgkin's lymphoma, leukemia and lymphoma, acute lymphoblastic leukemia, Hodgkin's lymphoma, and childhood acute lymphoblastic leukemia.

- 38. A method of treating or delaying the progression of cancer in a patient comprising administering to the patient a therapeutically effective amount of T cells genetically modified to express a chimeric antigen receptor (CAR) of any one of claims 1 to 23.
- 39. The method of claim 38, wherein the administering reduces the number of cancerous cells in the patient.
- 40. The method of claim 38, wherein the administering reduces the tumor burden and/or clears the tumor in the patient.
- 41. A therapeutically effective amount of a composition comprising T cells genetically modified to express the CAR of any one of claims 1 to 23 for use in treating or delaying the progression of cancer in a subject.
- 42. The composition for use of claim 41, wherein the composition provides one or more of the following effects:
 - reduces the number of cancer cells in the subject.
 - reduces the tumor burden and/or clears the tumor in the subject.
- 43. Use of the genetically modified T cell of any one of claims 24 to 32 for the manufacture of a medicament for treating a mammal suffering from cancer.
- 44. Use of a T cell genetically modified to express a CAR of any one of claims 1 to 23 for the manufacture of a medicament for providing an anti-cancer immune response in a mammal.
- 45. Use of a therapeutically effective amount of T cells genetically modified to express a

chimeric antigen receptor (CAR) of any one of claims 1 to 23 for the manufacture of a medicament for treating or delaying the progression of cancer in a patient.

FIG. 1





*BTLA, CD160, 284 and CD39 exhibit the same rescue pattern (not shown) *Data collected on day 10 post-activation

FIG. 3 *Data collected on day 14, gated on CD8+

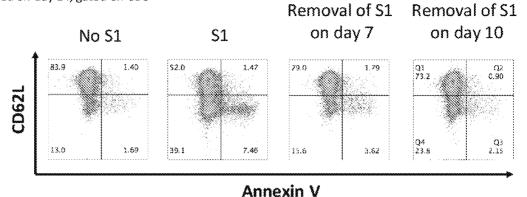
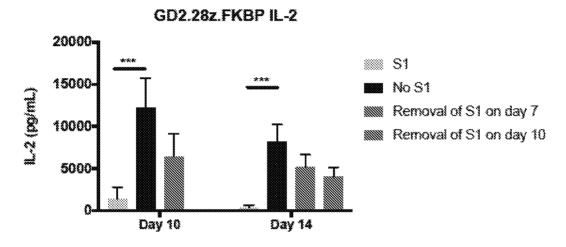


FIG. 4



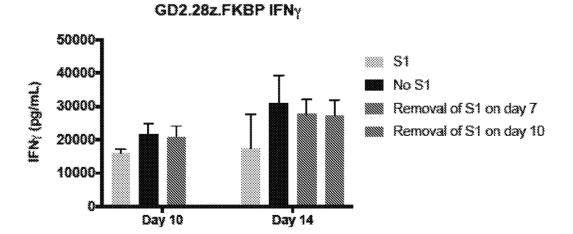
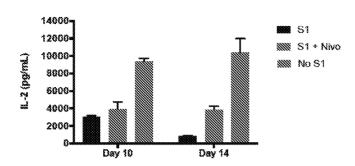
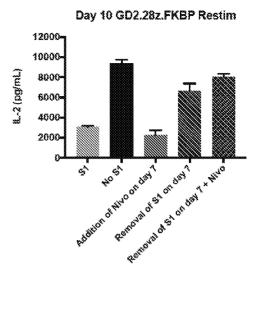
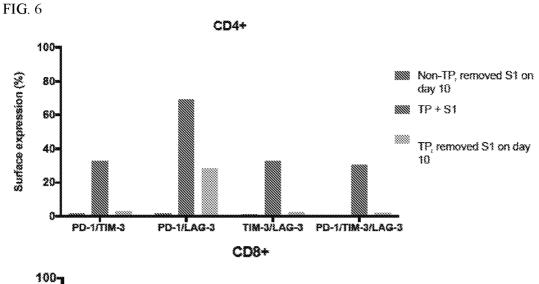
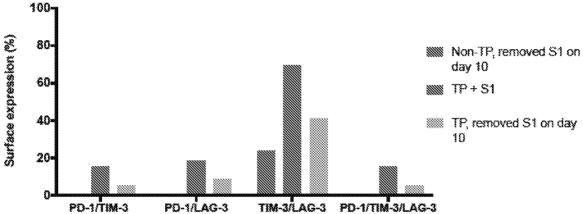


FIG. 5











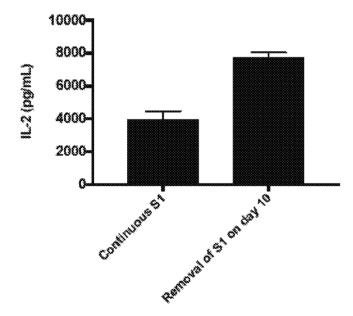
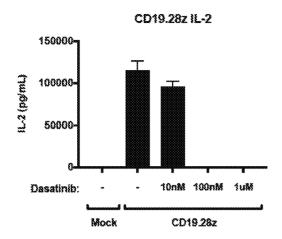
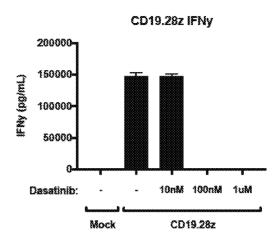


FIG. 7





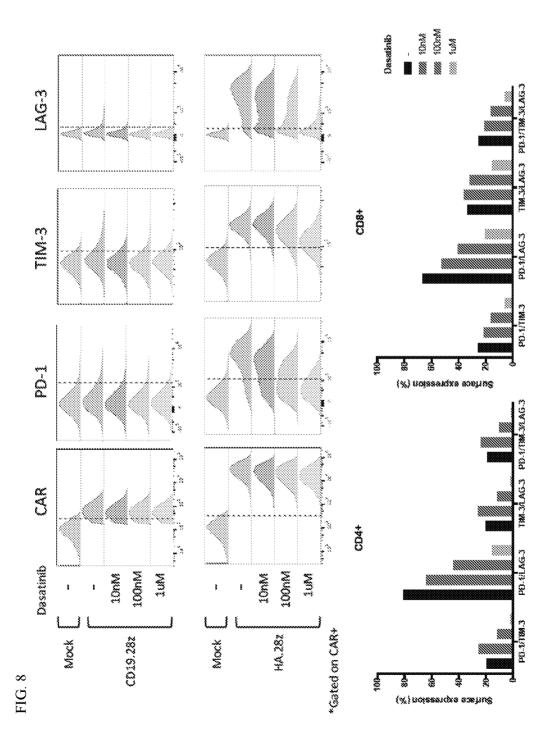
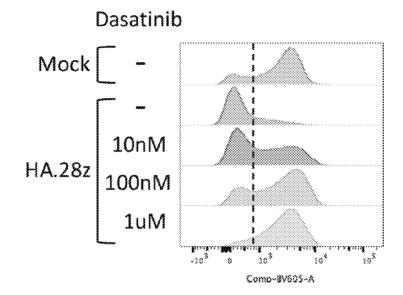
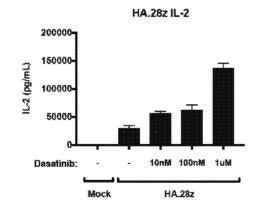


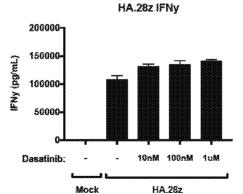
FIG. 9

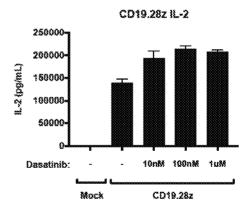


^{*}Gated on CAR+









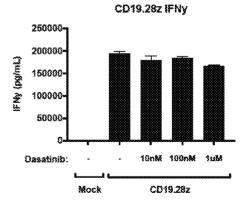
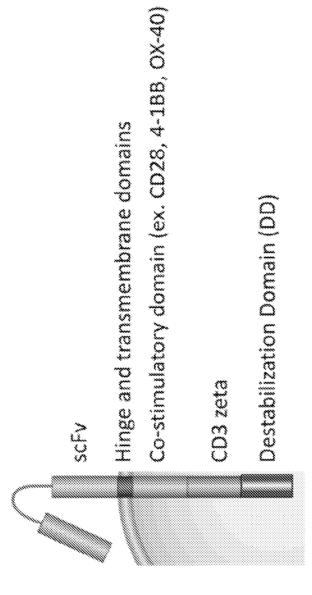
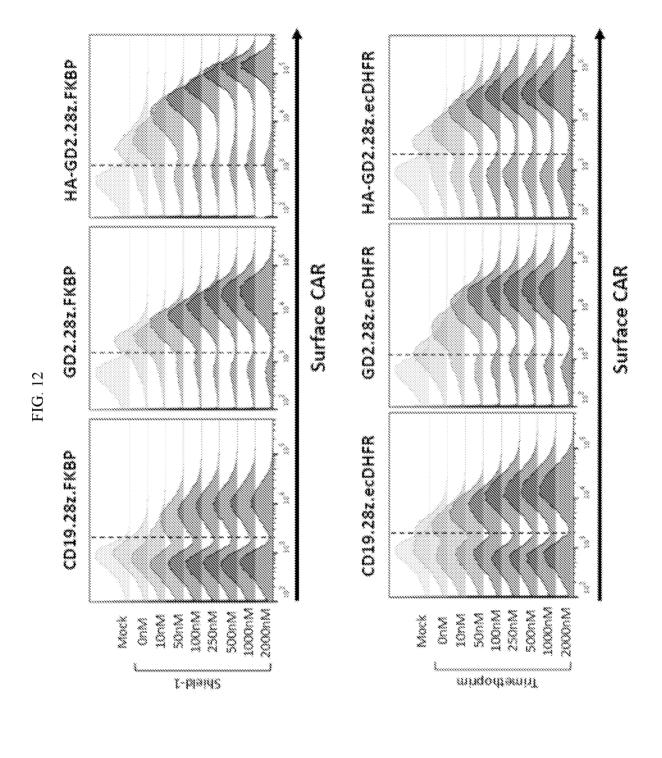


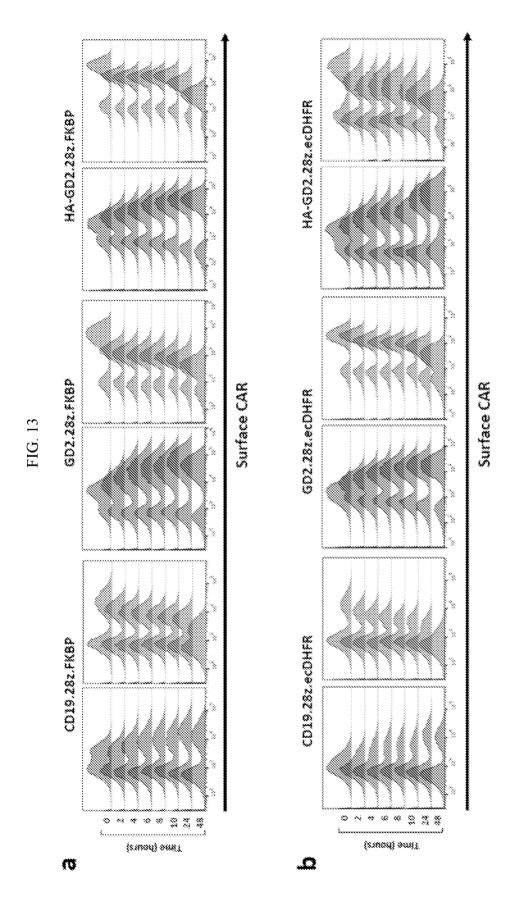
FIG. 11

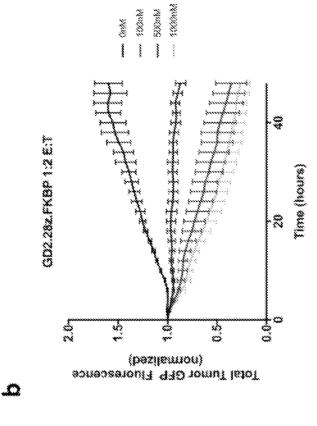




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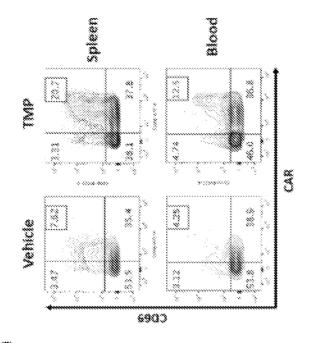




+ Shield-1 HA-GD2.282.FKBP - Shield-1 FNY Mock **22C**

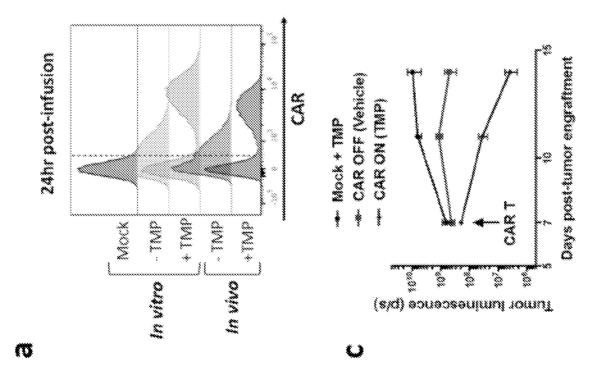
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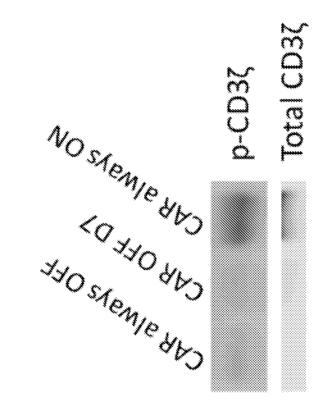
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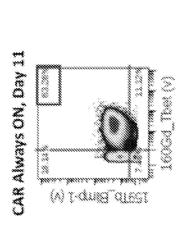


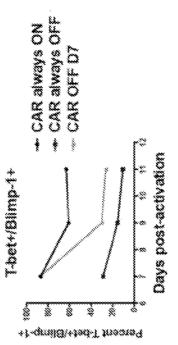
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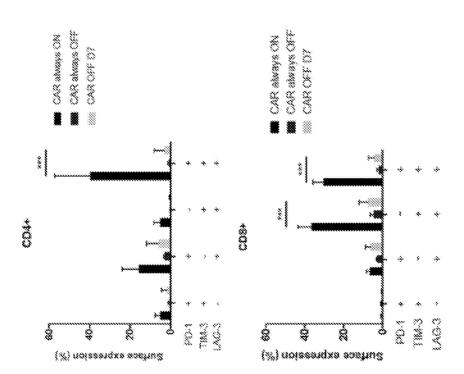


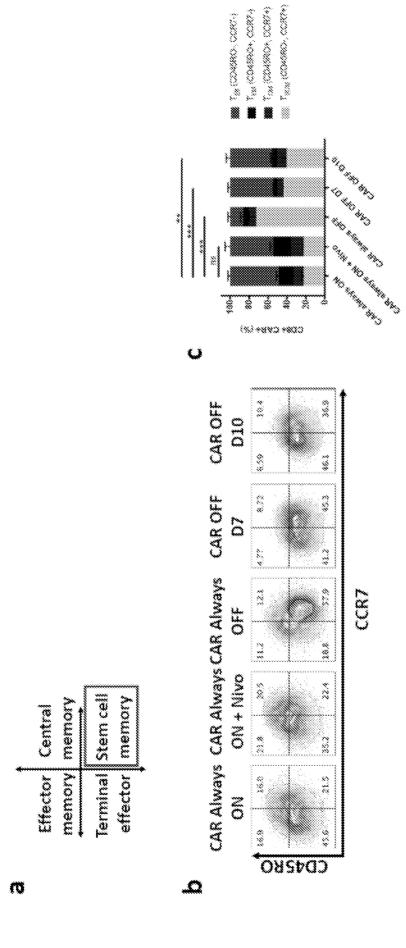


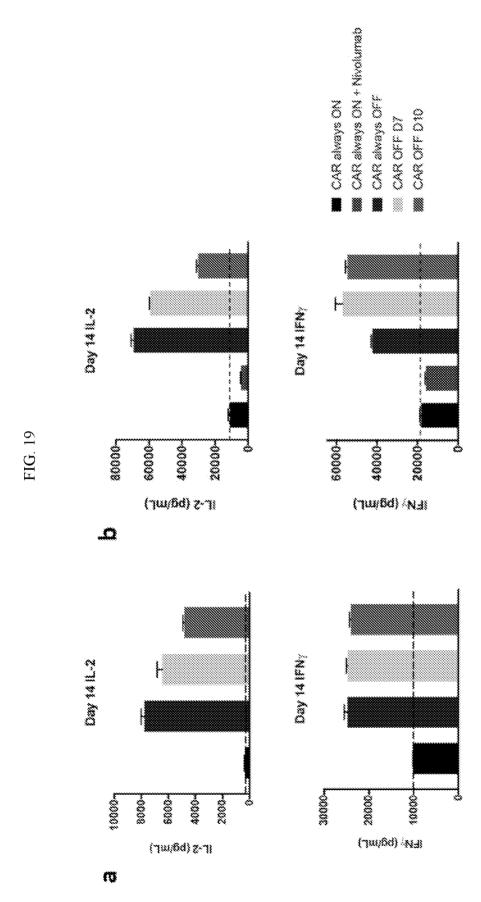


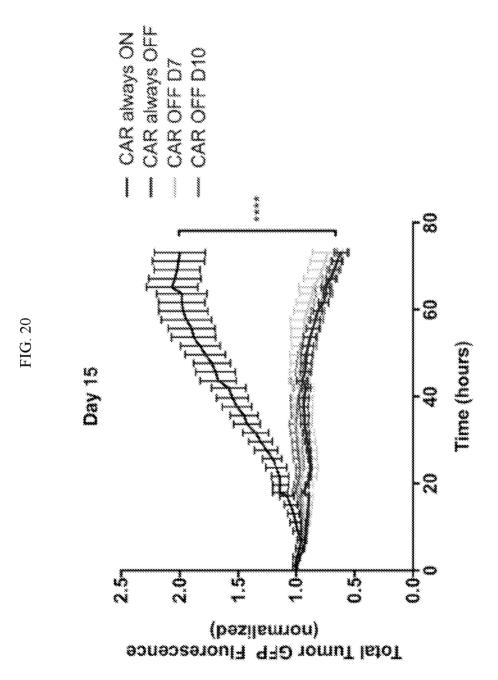
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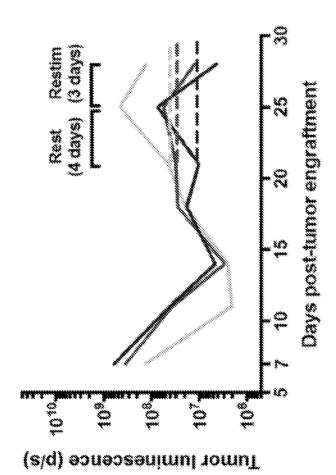
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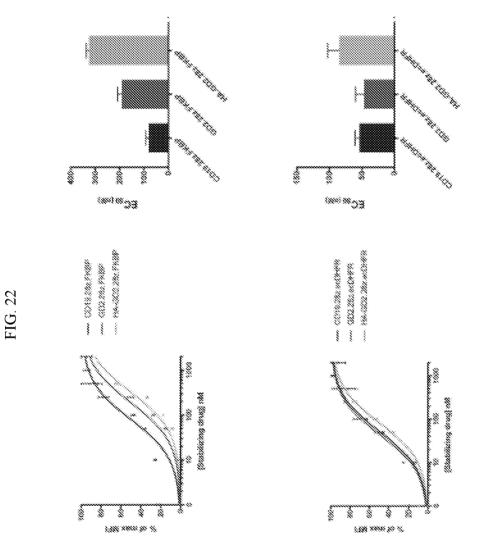






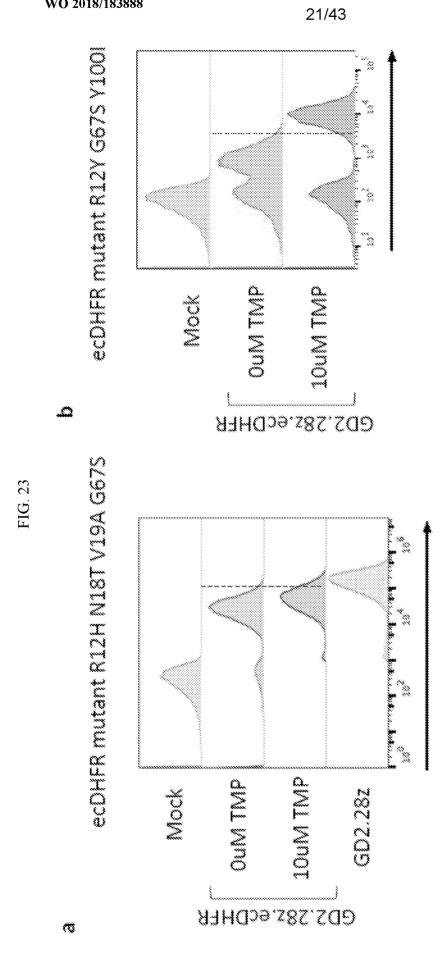






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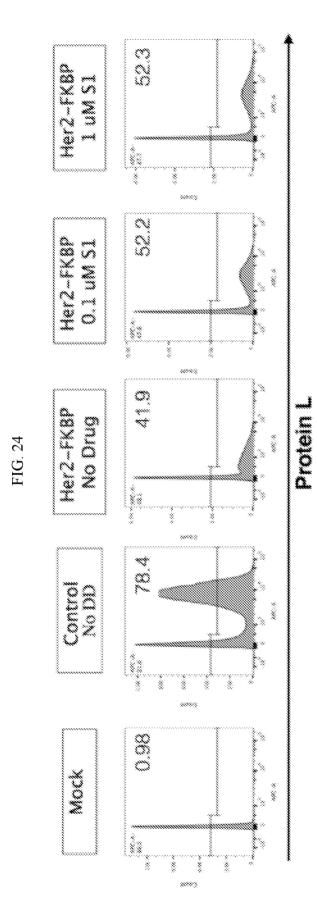


FIG. 25

SEQ		
ID NO:		
1	FKBP12 destabilization domain (E31G, F36V,	ATGGTGCAGGTGGAAACCATCTCCCCAGGAGACGGGCGCACC TTCCCCAAGCGCGGCCAGACCTGCGTGGTGCACTACACCGGG
	R71G, K105E)	ATGCTTGGAGATGGAAAGAAGTTGACTCCTCCCGGGACAGA
	nucleic acid sequence	AACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGATC
	nucleic acid sequence	CGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTCA
		GGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTGC
		CACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCGT
		CTTCGATGTGGAGCTTCTAGAACTGGAA
2	FKBP12 destabilization	MVQVETISPGDGRTFPKRGQTCVVHYTGMLGDGKKVDSSRDRN
	domain (E31G, F36V,	KPFKFMLGKQEVIRGWEEGVAQMSVGQGAKLTISPDYAYGATG
	R71G, K105E)	HPGIIPPHATLVFDVELLELE
	Amino acid sequence	
3	ecDHFR destabilization	ATGATCAGTCTGATTGCGGCGTTAGCGGTAGATCACGTTATC
	domain (R12H, N18T,	GGCATGGAAACCGTCATGCCGTGGAACCTGCCTGCCGATCTC
	V19A, G67S)	GCCTGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATG
	nucleic acid sequence	GGCCGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGA
		CGCAAAAATATTATCCTCAGCAGCAGCGAGTACGGACGAT
		CGCGTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCG TGTGGTGACGTACCAGAAATCATGGTTATTGGCGGCGGTCGC
		GTTTATGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTG
		ACGCATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCG
		GATTACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTC
		CACGATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAG
		ATTCTGGAGCGCGA
4	ecDHFR destabilization	MISLIAALAVDHVIGMETVMPWNLPADLAWFKRNTLNKPVIMG
	domain (R12H, N18T,	RHTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIAACGDV
	V19A, G67S)	PEIMVIGGGRVYEQFLPKAQKLYLTHIDAEVEGDTHFPDYEPDD
	Amino acid sequence	WESVFSEFHDADAQNSHSYCFEILERR
5	ecDHFR destabilization	ATGATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATC
	domain (R12Y, G67S,	GGCATGGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCTC
	Y100I) nucleic acid sequence	GCCTGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATG GGCCGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGA
	nucleic acid sequence	CGCAAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGAT
		CGCGTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCG
		TGTGGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGC
		GTTATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTG
		ACGCATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCG
		GATTACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTC
		CACGATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAG
		ATTCTGGAGCGCGA
6	ecDHFR destabilization	MISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMG
	domain (R12Y, G67S,	RHTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIAACGDV
	Y100I)	PEIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEGDTHFPDYEPDD
7	Amino acid sequence CD19.28z.FKBP	WESVFSEFHDADAQNSHSYCFEILERR ATGCTGCTGCTGGAGCTGCCCC
'	nucleic acid sequence	ACCCCGCCTTTCTGCTGATCCCCGATATCGACATCCAGATGAC
	nucleic acid sequence	ACAGACTACATCCTCCCTGTCTGCCTCTCTGGGAGACAGAGT
		CACCATCAGTTGCAGGGCAAGTCAGGACATTAGTAAATATTT
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FIG. 25 (cont'd)

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		CCTCAGCTAGCTTCGAAATTGAAGTTATGTATCCTCCTCA
		CCTAGACAATGAGAAGAGCAATGGAACCATTATCCATGTGAA
		AGGGAAACACCTTTGTCCAAGTCCCCTATTTCCCGGACCTTCT
		AAGCCCTTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCT
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		ACAAGCAGGCCCAGAACCAGCTCTATAACGAGCTCAATCTAG
		GACGAAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGC
		CGGGACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGAACCC
		TCAGGAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
		CGGAGGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGG
		AGGGGCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACA
		GCCACCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTG
		CCCCCTCGCGGAGTGCAGGTGGAAACCATCTCCCCAGGAGAC
		GGGCGCACCTTCCCCAAGCGCGGCCAGACCTGCGTGGTGCAC
		TACACCGGGATGCTTGGAGATGGAAAGAAAGTTGACTCCTCC
		CGGGACAGAAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAG
		GAGGTGATCCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAG
		TGTGGGTCAGGGAGCCAAACTGACTATATCTCCAGATTATGC
		CTATGGTGCCACTGGGCACCCAGGCATCATCCCACCACATGC
		CACTCTCGTCTTCGATGTGGAGCTTCTAGAACTGGAATGA
8	CD19.28z.FKBP	MLLLVTSLLLCELPHPAFLLIPDIDIQMTQTTSSLSASLGDRVTISC
	Amino acid sequence	RASQDISKYLNWYQQKPDGTVKLLIYHTSRLHSGVPSRFSGSGS
		GTDYSLTISNLEQEDIATYFCQQGNTLPYTFGGGTKLEITGSTSGS
		GKPGSGEGSTKGEVKLQESGPGLVAPSQSLSVTCTVSGVSLPDY
		GVSWIRQPPRKGLEWLGVIWGSETTYYNSALKSRLTIIKDNSKS
		QVFLKMNSLQTDDTAIYYCAKHYYYGGSYAMDYWGQGTSVT
		VSSASFEIEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
		FWVLVVVGGVLACYSLLVTVAFIIFWVRSKRSRLLHSDYMNMT
		PRRPGPTRKHYQPYAPPRDFAAYRSHMRVKFSRSADAPAYKQG
		QNQLYNELNLGRREEYDVLDKRRGRDPEMGGKPRRKNPQEGL
		YNELQKDKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDT
		YDALHMQALPPRGVQVETISPGDGRTFPKRGQTCVVHYTGMLG
		DGKKVDSSRDRNKPFKFMLGKQEVIRGWEEGVAQMSVGQGAK
0	CD 40 40 ELVDD	LTISPDYAYGATGHPGIIPPHATLVFDVELLELE
9	CD19.28z,FKBP	ATGCTGCTGCTGACATCTCTGCTGCTGCTGCGAGCTGCCCC
	Leader sequence	ACCCCGCCTTTCTGCTGATCCCC
10	nucleic acid sequence	MILLATCLI CELDIDAELLID
10	CD19.28z.FKBP	MLLLVTSLLLCELPHPAFLLIP
	Leader sequence	
11	Amino acid sequence FMC63 scFv (CD19	GACATCCAGATGACACAGACTACATCCTCCCTGTCTGCCTCTC
11	binding domain)	TGGGAGACAGAGTCACCATCAGTTGCAGGGCAAGTCAGGAC
	nucleic acid sequence	ATTAGTAAATATTTAAATTGGTATCAGCAGAAACCAGATGGA
	nucleic acid sequence	ACTGTTAAACTCCTGATCTACCATACATCAAGATTACACTCAG
		GAGTCCCATCAAGGTTCAGTGGCAGTGGGTCTGGAACAGATT
		ATTCTCTCACCATTAGCAACCTGGAGCAAGAAGATATTGCCA
		CTTACTTTTGCCAACAGGGTAATACGCTTCCGTACACGTTCGG
		AGGGGGACTAAGTTGGAAATAACAGGCTCCACCTCTGGATC
		CGGCAAGCCCGGATCTGGCGAGGGATCCACCAAGGGCGAGG
		TGAAACTGCAGGAGTCAGGACCTGGCCTGGTGGCGCCCTCAC
		AGAGCCTGTCCGTCACATGTACTGTCTCAGGGGTCTCATTACC
	1	11011000101000101101110111011010101010

lG. 25	(cont'd)	
		CGACTATGGTGTAAGCTGGATTCGCCAGCCTCCACGAAAGGG
		TCTGGAGTGGCTGGGAGTAATATGGGGTAGTGAAACCACATA
		CTATAATTCAGCTCTCAAATCCAGACTGACCATCATCAAGGA
		CAACTCCAAGAGCCAAGTTTTCTTAAAAATGAACAGTCTGCA
l		AACTGATGACACAGCCATTTACTACTGTGCCAAACATTATTA
		CTACGGTGGTAGCTATGCTATGGACTACTGGGGTCAAGGAAC
		CTCAGTCACCGTCTCCTCA
10	EMCCC E (CD10	DIOMETER CLICACIONE CONTROL CO
12	FMC63 scFv (CD19	DIQMTQTTSSLSASLGDRVTISCRASQDISKYLNWYQQKPDGTV
	binding domain)	KLLIYHTSRLHSGVPSRFSGSGSGTDYSLTISNLEQEDIATYFCQQ
	Amino acid sequence	GNTLPYTFGGGTKLEITGSTSGSGKPGSGEGSTKGEVKLQESGPG
		LVAPSQSLSVTCTVSGVSLPDYGVSWIRQPPRKGLEWLGVIWGS
		ETTYYNSALKSRLTIIKDNSKSQVFLKMNSLQTDDTAIYYCAKH
		YYYGGSYAMDYWGQGTSVTVSS
13	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
	nucleic acid sequence	AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT
	nuciese uesa sequence	CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC
14	CD28 hinge region	IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
14		IE VIVITERE I L'UNEASNOTTIN VAGANL CESPLEFOFSAF
1.5	Amino acid sequence	
15	CD28 transmembrane	TTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCTTGCTATA
	region nucleic acid	GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG
	sequence	
16	Amino acid sequence	FWVLVVVGGVLACYSLLVTVAFIIFWV
17	CD28 intracellular	AGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAA
	domain nucleic acid	CATGACTCCCGCCGCCCCGGGCCCACCCGCAAGCATTACCA
	sequence	GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
	sequence	
18	CD28 transmembrane	RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
10	region Amino acid	RSKRSRLEIISD I WINWITT ING OF TRIGITY OF TATTROT AATRO
10	sequence	
19	CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA
	domain nucleic acid	GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
	sequence	AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
		ACCCTGAGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
		GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
		GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG
		GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA
		CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC
		CTCGC
20	CD3 zeta intracellular	RVKFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP
20		
	domain Amino acid	EMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG HDGLYQGLSTATKDTYDALHMQALPPR
2.1	sequence	
21	FKBP12 destabilization	GGAGTGCAGGTGGAAACCATCTCCCCAGGAGACGGGCGCAC
	domain (E31G, F36V,	CTTCCCCAAGCGCGGCCAGACCTGCGTGGTGCACTACACCGG
	R71G, K105E)	GATGCTTGGAGATGGAAAGAAAGTTGACTCCTCCCGGGACAG
	nucleic acid sequence	AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT
		CCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTC
		AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG
		CCACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCG
		TCTTCGATGTGGAGCTTCTAGAACTGGAA
22	FKBP12 destabilization	GVQVETISPGDGRTFPKRGQTCVVHYTGMLGDGKKVDSSRDRN
_	domain (E31G, F36V,	KPFKFMLGKQEVIRGWEEGVAQMSVGQGAKLTISPDYAYGATG
	R71G, K105E)	HPGIIPPHATLVFDVELLELE
	Amino acid sequence	III GIII I IIA I E VI D V LEELELE
22		A TOOTOOTOOTOOTO A CATOTOTOOTOOTOOO A COTOCOCO
23	CD19.28z.ecDHFR	ATGCTGCTGCTGACATCTCTGCTGCTGCTGCGAGCTGCCCC
	nucleic acid sequence	ACCCCGCCTTTCTGCTGATCCCCGATATCGACATCCAGATGAC
		ACAGACTACATCCTCCCTGTCTGCCTCTCTGGGAGACAGAGT
		CACCATCAGTTGCAGGGCAAGTCAGGACATTAGTAAATATTT
		AAATTGGTATCAGCAGAAACCAGATGGAACTGTTAAACTCCT
		GATCTACCATACATCAAGATTACACTCAGGAGTCCCATCAAG
	-	

I <u>G. 25 (</u>	cont'd)	
		GTTCAGTGGCAGTGGGTCTGGAACAGATTATTCTCTCACCATT
		AGCAACCTGGAGCAAGAAGATATTGCCACTTACTTTTGCCAA
		CAGGGTAATACGCTTCCGTACACGTTCGGAGGGGGGGACTAAG
		TTGGAAATAACAGGCTCCACCTCTGGATCCGGCAAGCCCGGA
		TCTGGCGAGGGATCCACCAAGGGCGAGGTGAAACTGCAGGA
		GTCAGGACCTGGCCTGGTGGCGCCCTCACAGAGCCTGTCCGT
		CACATGTACTGTCTCAGGGGTCTCATTACCCGACTATGGTGTA
		AGCTGGATTCGCCAGCCTCCACGAAAGGGTCTGGAGTGGCTG
		GGAGTAATATGGGGTAGTGAAACCACATACTATAATTCAGCT
		CTCAAATCCAGACTGACCATCATCAAGGACAACTCCAAGAGC
		CAAGTTTTCTTAAAAATGAACAGTCTGCAAACTGATGACACA
		GCCATTTACTACTGTGCCAAACATTATTACTACGGTGGTAGCT
		ATGCTATGGACTACTGGGGTCAAGGAACCTCAGTCACCGTCT
		CCTCAGCTAGCTTCGAAATTGAAGTTATGTATCCTCCTCCTTA
		CCTAGACAATGAGAAGAGCAATGGAACCATTATCCATGTGAA
		AGGGAAACACCTTTGTCCAAGTCCCCTATTTCCCGGACCTTCT
		AAGCCCTTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCT
		TGCTATAGCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGG
		TGAGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATG
		AACATGACTCCCCGCCCCCGGGCCCACCCGCAAGCATTAC
		CAGCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCCC
		ATATGAGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGT
		ACAAGCAGGCCAGAACCAGCTCTATAACGAGCTCAATCTAG
		GACGAAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGC
		CGGGACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGAACCC
		TCAGGAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
		CGGAGGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGG
		AGGGGCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACA
		GCCACCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTG
		CCCCTCGCATCAGTCTGATTGCGGCGTTAGCGGTAGATTAC
		GTTATCGGCATGGAAAACGCCATGCCGTGGAACCTGCCTG
		GATCTCGCCTGGTTTAAACGCAACACCTTAAATAAACCCGTG
		ATTATGGGCCGCCATACCTGGGAATCAATCGGTCGTCCGTTG
		CCAGGACGCAAAAATATTATCCTCAGCAGTCAACCGAGTACG
		GACGATCGCGTAACGTGGGTGAAGTCGGTGGATGAAGCCATC
		GCGCGTGTGGTGACGTACCAGAAATCATGGTGATTGGCGGC
		GGTCGCGTTATTGAACAGTTCTTGCCAAAAAGCGCAAAAACTG
		TATCTGACGCATATCGACGCAGAAGTGGAAGGCGACACCCAT
		TTCCCGGATTACGAGCCGGATGACTGGGAATCGGTATTCAGC
		GAATTCCACGATGCTGATGCGCAGAACTCTCACAGCTATTGC
		TTTGAGATTCTGGAGCGGCGATGA
24	CD19.28z.ecDHFR	MLLLVTSLLLCELPHPAFLLIPDIDIQMTQTTSSLSASLGDRVTISC
	Amino acid sequence	RASQDISKYLNWYQQKPDGTVKLLIYHTSRLHSGVPSRFSGSGS
	Tamas was sequence	GTDYSLTISNLEQEDIATYFCQQGNTLPYTFGGGTKLEITGSTSGS
		GKPGSGEGSTKGEVKLQESGPGLVAPSQSLSVTCTVSGVSLPDY
		GVSWIRQPPRKGLEWLGVIWGSETTYYNSALKSRLTIIKDNSKS
		QVFLKMNSLQTDDTAIYYCAKHYYYGGSYAMDYWGQGTSVT
		VSSASFEIEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
		FWVLVVVGGVLACYSLLVTVAFIIFWVRSKRSRLLHSDYMNMT
		PRRPGPTRKHYQPYAPPRDFAAYRSHMRVKFSRSADAPAYKQG
		QNQLYNELNLGRREEYDVLDKRRGRDPEMGGKPRRKNPQEGL
		YNELQKDKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDT
		YDALHMQALPPRISLIAALAVDYVIGMENAMPWNLPADLAWFK
		RNTLNKPVIMGRHTWESIGRPLPGRKNIILSSQPSTDDRVTWVKS
		VDEAIAACGDVPEIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEG
		DTHFPDYEPDDWESVFSEFHDADAQNSHSYCFEILERR
25	CD19.28z.ecDHFR	ATGCTGCTGCTGACATCTCTGCTGCTGCGAGCTGCCCC
~~	nucleic acid sequence	ACCCCGCCTTTCTGCTGATCCCC
26	CD19.28z.ecDHFR	MLLLVTSLLLCELPHPAFLLIP
	Amino acid sequence	

FIG. 25 (cont'd)

	(cont'd)	
27	FMC63 scFv (CD19	GACATCCAGATGACACAGACTACATCCTCCCTGTCTGCCTCTC
	binding domain)	TGGGAGACAGAGTCACCATCAGTTGCAGGGCAAGTCAGGAC
	nucleic acid sequence	ATTAGTAAATATTTAAATTGGTATCAGCAGAAACCAGATGGA
		ACTGTTAAACTCCTGATCTACCATACATCAAGATTACACTCAG
		GAGTCCCATCAAGGTTCAGTGGCAGTGGGTCTGGAACAGATT
		ATTCTCTCACCATTAGCAACCTGGAGCAAGAAGATATTGCCA
		CTTACTTTTGCCAACAGGGTAATACGCTTCCGTACACGTTCGG
		AGGGGGACTAAGTTGGAAATAACAGGCTCCACCTCTGGATC
		CGGCAAGCCCGGATCTGGCGAGGGATCCACCAAGGGCGAGG
		TGAAACTGCAGGAGTCAGGACCTGGCCTGGTGGCGCCCTCAC
		AGAGCCTGTCCGTCACATGTACTGTCTCAGGGGTCTCATTACC
		CGACTATGGTGTAAGCTGGATTCGCCAGCCTCCACGAAAGGG
		TCTGGAGTGGCTGGGAGTAATATGGGGTAGTGAAACCACATA
		CTATAATTCAGCTCTCAAATCCAGACTGACCATCATCAAGGA
		CAACTCCAAGAGCCAAGTTTTCTTAAAAATGAACAGTCTGCA
		AACTGATGACACAGCCATTTACTACTGTGCCAAACATTATTA
		CTACGGTGGTAGCTATGCTATGGACTACTGGGGTCAAGGAAC
		CTCAGTCACCGTCTCCTCA
28	FMC63 scFv (CD19	DIQMTQTTSSLSASLGDRVTISCRASQDISKYLNWYQQKPDGTV
	binding domain)	KLLIYHTSRLHSGVPSRFSGSGSGTDYSLTISNLEQEDIATYFCQQ
	Amino acid sequence	GNTLPYTFGGGTKLEITGSTSGSGKPGSGEGSTKGEVKLQESGPG
		LVAPSQSLSVTCTVSGVSLPDYGVSWIRQPPRKGLEWLGVIWGS
		ETTYYNSALKSRLTIIKDNSKSQVFLKMNSLQTDDTAIYYCAKH
		YYYGGSYAMDYWGQGTSVTVSS
29	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
	nucleic acid sequence	AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT
	•	CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC
30	CD28 hinge region	IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
	Amino acid sequence	
31	CD28 transmembrane	TTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCTTGCTATA
	region nucleic acid	GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG
	sequence	
32	CD28 transmembrane	FWVLVVVGGVLACYSLLVTVAFIIFWV
	region Amino acid	
	sequence	
33	CD28 intracellular	AGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAA
33	domain	CATGACTCCCGCCGCCCCGGGCCCACCCGCAAGCATTACCA
	nucleic acid sequence	GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
34	CD28 intracellular	RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
34	I	RSARSKLLIISD I MINWITERRE OF IRATI QF I AFFRDEAA I RS
	domain	
35	Amino acid sequence	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA
33	CD3 zeta intracellular	
	domain	GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
	nucleic acid sequence	AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
		ACCCTGAGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
		GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
		GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG
		GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA
		CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC
		CTCGC
36	CD3 zeta intracellular	RVKFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP
	domain	EMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG
	Amino acid sequence	HDGLYQGLSTATKDTYDALHMQALPPR
37	ecDHFR destabilization	ATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGC
	domain (R12Y, G67S,	ATGGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCTCGCC
	Y100I)	TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC
	nucleic acid sequence	CGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGACGC
	and the state of t	AAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC
		GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT
	L	

FIG. 25 (cont'd)

1		SFEIEVMYPPPYLDNEKSNGTIHVKGKHLCPSPLFPGPSKPFWVL
		FTGYNMNWVRQNIGKSLEWIGAIDPYYGGTSYNQKFKGRATLT VDKSSSTAYMHLKSLTSEDSAVYYCVSGMEYWGQGTSVTVSSA
		STSGSGKPGSGEGSTKGEVKLQQSGPSLVEPGASVMISCKASGSS
	Amino acid sequence	SGSGTDFTLKISRVEAEDLGVYFCSQSTHVPPLTFGAGTKLELKG
40	GD2.28z.FKBP	MLLLVTSLLLCELPHPAFLLIPDILLTQTPLSLPVSLGDQASISCRS SQSLVHRNGNTYLHWYLQKPGQSPKLLIHKVSNRFSGVPDRFSG
40	CIDA AO ELEPP	TTCGATGTGGAGCTTCTAGAACTGGAATGA
		ACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCGTC
		GGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTGCC
		ACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGATCC GAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTCAG
		TGCTTGGAGATGGAAAGAAAGTTGACCCCCGGGACAGAA
		TCCCCAAGCGCGGCCAGACCTGCGTGGTGCACTACACCGGGA
		GAGTGCAGGTGGAAACCATCTCCCCAGGAGACGGGCGCACCT
		GGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCACCAAGG ACACCTACGACGCCCTTCACATGCAGGCCCTGCCCCTCGCG
		CAGTGAGATTGGGATGAAAGGCGAGGGGCCAGG
		CTGTACAATGAACTGCAGAAAGATAAGATGGCGGAGGCCTA
		AGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAGGAAGGC
		GCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACGAAGAG AGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGGACCCTG
		GAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAAGCAGG
		CTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCCAGAGT
		GACTCCCCGCCCCCGGGCCCACCCGCAAGCATTACCAGCC
		ATAGCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTGAG GAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAACAT
		CCCTTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCTGAG
		GAAACACCTTTGTCCAAGTCCCCTATTTCCCGGACCTTCTAAG
		GACAATGAGAAGAGCAATGGAACCATTATCCATGTGAAAGG
		TGCTAGCTTCGAAATTGAAGTTATGTATCCTCCTCCTTACCTA
		CCTGACCAGCGAGGACAGCGCCGTGTACTACTGCGTGTCCGG CATGGAATACTGGGGCCAGGGCACAAGCGTGACCGTGTCCTC
		CGTGACCAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGG
		CACCAGCTACAACCAGAAGTTCAAGGGCAGAGCCACCCTGAC
		GAGCCTGGAATGGATCGGCGCCATCGACCCCTACTACGGCGG
		ACCGCTACAACATGAACTGGGTGCGCCAGAACATCGGCAA
		GTGAAGCTGCAGCAGAGCGCCCCTCTCTGGTGGAACCTGGC GCCTCTGTGATGATCTCCTGCAAGGCCAGCGGCAGCTCCTTC
		GCGGCAAGCCTGGATCTGGCGAGGGAAGCACCAAGGGCGAA
		CGCCGGAACAAAGCTGGAACTGAAGGGCAGCACAAGCGGCA
		ACTTCTGCAGCCAGTCCACCCACGTGCCCCCCTGACATTTGG
		GCCCGACAGATTTTCTGGCAGCGGCTCCGGCACCGACTTCAC CCTGAAGATCAGCCGGGTGGAAGCCGAGGACCTGGGCGTGT
		CAAGCTGCTGATTCACAAGGTGTCCAACCGGTTCAGCGCACTTCAG
		CACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGAGCCC
		AGCTGCAGATCCAGCCAGAGCCTGGTGCACCGGAACGCAA
	nacicie aciu sequence	CCCTCTGAGCCTGCTGTGTCTCTGGGCGATCAGGCCAGCATC
39	GD2.28z.FKBP nucleic acid sequence	ATGCTGCTGCTGACATCTCTGCTGCTGCGAGCTGCCCC ACCCGCCTTTCTGCTGATCCCCGATATCCTGCTGACCCAGAC
20	Amino acid sequence	ESVFSEFHDADAQNSHSYCFEILERR
	Y100I)	EIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEGDTHFPDYEPDDW
	domain (R12Y, G67S,	HTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIAACGDVP
38	ecDHFR destabilization	ISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMGR
		TGGAGCGCGA
		TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC
		CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT
		ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG
		GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGCGTT

FIG. 25 (cont'd)

<u>IG. 25</u>	(cont'd)	
		VVVGGVLACYSLLVTVAFIIFWVRSKRSRLLHSDYMNMTPRRP
		GPTRKHYQPYAPPRDFAAYRSRVKFSRSADAPAYKQGQNQLYN
		ELNLGRREEYDVLDKRRGRDPEMGGKPRRKNPQEGLYNELQK
		DKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDTYDALHM
		QALPPRGVQVETISPGDGRTFPKRGQTCVVHYTGMLGDGKKVD
		SSRDRNKPFKFMLGKQEVIRGWEEGVAQMSVGQGAKLTISPDY
		AYGATGHPGIIPPHATLVFDVELLELE
41	GD2.28z.FKBP	ATGCTGCTGCTGACATCTCTGCTGCTGTGCGAGCTGCCCC
	Leader sequence	ACCCCGCCTTTCTGCTGATCCCC
	nucleic acid sequence	
42	GD2,28z,FKBP	MLLLVTSLLLCELPHPAFLLIP
	Leader sequence	
	Amino acid sequence	
43	14G2a scFv (GD2	GATATCCTGCTGACCCAGACCCCTCTGAGCCTGCCTGTGTCTC
43		
	binding domain)	TGGGCGATCAGGCCAGCATCAGCTGCAGATCCAGCCAGAGCC
	nucleic acid sequence	TGGTGCACCGGAACGGCAACACCTACCTGCACTGGTATCTGC
		AGAAGCCCGGCCAGAGCCCCAAGCTGCTGATTCACAAGGTGT
		CCAACCGGTTCAGCGGCGTGCCCGACAGATTTTCTGGCAGCG
		GCTCCGGCACCGACTTCACCCTGAAGATCAGCCGGGTGGAAG
		CCGAGGACCTGGGCGTGTACTTCTGCAGCCAGTCCACCCAC
		TGCCCCCCTGACATTTGGCGCCGGAACAAAGCTGGAACTGA
		AGGGCAGCACAAGCGGCAGCGGCAAGCCTGGATCTGGCGAG
		GGAAGCACCAAGGGCGAAGTGAAGCTGCAGCAGAGCGGCCC
		CTCTCTGGTGGAACCTGGCGCCTCTGTGATGATCTCCTGCAAG
		GCCAGCGCAGCTCCTTCACCGGCTACAACATGAACTGGGTG
		CGCCAGAACATCGGCAAGAGCCTGGAATGGATCGGCGCCATC
		GACCCCTACTACGGCGCCACCAGCTACAACCAGAAGTTCAAG
		GGCAGAGCCACCCTGACCGTGGACAAGAGCAGCTCCACCGCC
		TACATGCACCTGAAGTCCCTGACCAGCGAGGACAGCGCCGTG
		TACTACTGCGTGTCCGGCATGGAATACTGGGGCCAGGGCACA
		AGCGTGACCGTGTCCTCT
4.4	14C2 E (CD2	
44	14G2a scFv (GD2	DILLTQTPLSLPVSLGDQASISCRSSQSLVHRNGNTYLHWYLQKP
	binding domain)	GQSPKLLIHKVSNRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGV
	Amino acid sequence	YFCSQSTHVPPLTFGAGTKLELKGSTSGSGKPGSGEGSTKGEVK
		LQQSGPSLVEPGASVMISCKASGSSFTGYNMNWVRQNIGKSLE
		WIGAIDPYYGGTSYNQKFKGRATLTVDKSSSTAYMHLKSLTSED
		SAVYYCVSGMEYWGQGTSVTVSS
45	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
	nucleic acid sequence	AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT
	nucleic uciu sequence	CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC
46	CD28 hinge region	IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
40		IE VIVITEET TEDNEKSINGTIIII VKOKIILCESTLIT OFSKE
47	Amino acid sequence	TTTTCCCTCCTCCTCCTTCCCCCCACTCCTTCCCTATA
47	CD28 transmembrane	TTTTGGGTGCTGGTGGTGGTGGGGGAGTCCTGGCTTGCTATA
	region	GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG
	nucleic acid sequence	
48	CD28 transmembrane	FWVLVVVGGVLACYSLLVTVAFIIFWV
	region	
	Amino acid sequence	
49	CD28 intracellular	AGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAA
'	domain	CATGACTCCCGCCGCCCCGGGCCCACCCGCAAGCATTACCA
	nucleic acid sequence	
50		GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
50	CD28 intracellular	RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
	domain	
	Amino acid sequence	
51	CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA
	domain	GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
	nucleic acid sequence	AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
	nucicie aciu scquence	ACCCTGAGATGGGGGGAAAGCCGAGAAGAACCCTCAG
		GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
		GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG

FIG. 25 (cont'd)

U. 23	(cont d)	
		GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC
		CTCGC
52	CD3 zeta intracellular	RVKFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP
32	domain	EMGGKPRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG
	Amino acid sequence	HDGLYQGLSTATKDTYDALHMQALPPR
53	FKBP12 destabilization	GGAGTGCAGGTGGAAACCATCTCCCCAGGAGACGGCGCAC
	domain (E31G, F36V,	CTTCCCCAAGCGCGGCCAGACCTGCGTGGTGCACTACACCGG
	R71G, K105E)	GATGCTTGGAGATGGAAAGAAAGTTGACTCCTCCCGGGACAG
	nucleic acid sequence	AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT
		CCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTC
		AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG
		CCACTGGGCACCCAGGCATCTACAACTCCAACATGCCACTCTCG
54	FKBP12 destabilization	TCTTCGATGTGGAGCTTCTAGAACTGGAA GVQVETISPGDGRTFPKRGQTCVVHYTGMLGDGKKVDSSRDRN
34	domain (E31G, F36V,	KPFKFMLGKQEVIRGWEEGVAQMSVGQGAKLTISPDYAYGATG
	R71G, K105E)	HPGIIPPHATLVFDVELLELE
	Amino acid sequence	III GIII I IIA LE VI D V ELELELE
55	GD2.28z.ecDHFR (R12Y,	ATGCTGCTGCTGACATCTCTGCTGCTGCGAGCTGCCCC
-	G67S, Y100I)	ACCCCGCCTTTCTGCTGATCCCCGATATCCTGCTGACCCAGAC
	nucleic acid sequence	CCCTCTGAGCCTGCCTGTGTCTCTGGGCGATCAGGCCAGCATC
	_	AGCTGCAGATCCAGCCAGAGCCTGGTGCACCGGAACGGCAA
		CACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGAGCCC
		CAAGCTGCTGATTCACAAGGTGTCCAACCGGTTCAGCGGCGT
		GCCCGACAGATTTTCTGGCAGCGGCTCCGGCACCGACTTCAC
		CCTGAAGATCAGCCGGGTGGAAGCCGAGGACCTGGGCGTGT
		ACTTCTGCAGCCAGTCCACCCACGTGCCCCCCTGACATTTGG
		CGCCGAACGCTGGAACTGAAGGGCAGCAAGCGGCA
		GCGGCAAGCCTGGATCTGGCGAGGGAAGCACCAAGGGCGAA GTGAAGCTGCAGCAGAGCGGCCCCTCTCTGGTGGAACCTGGC
		GCCTCTGTGATGATCTCCTGCAAGGCCAGCGGCAGCTCCTTC
		ACCGCTACAACATGAACTGGGTGCGCCAGAACATCGGCAA
		GAGCCTGGAATGGATCGGCGCCATCGACCCCTACTACGGCGG
		CACCAGCTACAACCAGAAGTTCAAGGGCAGAGCCACCCTGAC
		CGTGGACAAGAGCAGCTCCACCGCCTACATGCACCTGAAGTC
		CCTGACCAGCGAGGACAGCGCCGTGTACTACTGCGTGTCCGG
		CATGGAATACTGGGGCCAGGGCACAAGCGTGACCGTGTCCTC
		TGCTAGCTTCGAAATTGAAGTTATGTATCCTCCTCCTTACCTA
		GACAATGAGAAGAGCAATGGAACCATTATCCATGTGAAAGG
		GAAACACCTTTGTCCAAGTCCCCTATTTCCCGGACCTTCTAAG
		CCCTTTTGGGTGCTGGTGGTGGTGGGGGAGTCCTGGCTTGCT
		ATAGCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTGAG GAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAACAT
		GACTCCCCGCCCCCGGGCCCACCCGCAAGCATTACCAGCC
		CTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCCAGAGT
		GAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAAGCAGG
		GCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACGAAGAG
		AGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGGACCCTG
		AGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAGGAAGGC
		CTGTACAATGAACTGCAGAAAGATAAGATGGCGGAGGCCTA
		CAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGGGCAAGG
		GGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCACCAAGG
		ACACCTACGACGCCCTTCACATGCAGGCCCTGCCCCCTCGCA
		TCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGCAT
		GGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCTCGCCTG
		GTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGCCG
		CCATACCTGGGAATCAATCGGTCGTCGTTGCCAGGACGCAA
		AAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGCGT
		AACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGTGGGTTAT
		TGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGCGTTAT

FIG. 25 (cont'd)

U. 23 (t	Zoni u)	
56	GD2.28z.ecDHFR (R12Y,	TGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACGCA TATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGATTA CGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCACGA TGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTCTG GAGCGGCGATGA MLLLVTSLLLCELPHPAFLLIPDILLTQTPLSLPVSLGDQASISCRS SOSI VALIDAGCTTYL HWYL OV DCOSDRALL HAVYSDRES CANDRES CONDRESS CON
	G67S, Y100I) Amino acid sequence	SQSLVHRNGNTYLHWYLQKPGQSPKLLIHKVSNRFSGVPDRFSG SGSGTDFTLKISRVEAEDLGVYFCSQSTHVPPLTFGAGTKLELKG STSGSGKPGSGEGSTKGEVKLQQSGPSLVEPGASVMISCKASGSS FTGYNMNWVRQNIGKSLEWIGAIDPYYGGTSYNQKFKGRATLT VDKSSSTAYMHLKSLTSEDSAVYYCVSGMEYWGQGTSVTVSSA SFEIEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKPFWVL VVVGGVLACYSLLVTVAFIIFWVRSKRSRLLHSDYMNMTPRRP GPTRKHYQPYAPPRDFAAYRSRVKFSRSADAPAYKQGQNQLYN ELNLGRREEYDVLDKRRGRDPEMGGKPRRKNPQEGLYNELQK DKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDTYDALHM QALPPRISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNK PVIMGRHTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIA ACGDVPEIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEGDTHFPD YEPDDWESVFSEFHDADAQNSHSYCFEILERR
57	GD2.28z.ecDHFR (R12Y, G67S, Y100I) Leader sequence nucleic acid sequence	ATGCTGCTGCTGACATCTCTGCTGCTGCGAGCTGCCCC ACCCCGCCTTTCTGCTGATCCCC
58	GD2.28z.ecDHFR (R12Y, G67S, Y100I) Leader sequence Amino acid sequence	MLLLVTSLLLCELPHPAFLLIP
59	14G2a scFv (GD2 binding domain) nucleic acid sequence	GATATCCTGCTGACCCAGACCCCTCTGAGCCTGCCTGTGTCTC TGGGCGATCAGGCCAGCATCAGCTGCAGATCCAGCCAGAGCC TGGTGCACCGGAACGGCAACACCTACCTGCACTGGTATCTGC AGAAGCCCGGCCAGAGCCCCAAGCTGCTGATTCACAAGGTGT CCAACCGGTTCAGCGGCGTGCCCGACAGATTTTCTGGCAGCG GCTCCGGCACCGACTTCACCCTGAAGATCAGCCGGGTGGAAG CCGAGGACCTGGGCGTGTACTTCTGCAGCCAGTCCACCCAC
60	14G2a scFv (GD2 binding domain) Amino acid sequence	DILLTQTPLSLPVSLGDQASISCRSSQSLVHRNGNTYLHWYLQKP GQSPKLLIHKVSNRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGV YFCSQSTHVPPLTFGAGTKLELKGSTSGSGKPGSGEGSTKGEVK LQQSGPSLVEPGASVMISCKASGSSFTGYNMNWVRQNIGKSLE WIGAIDPYYGGTSYNQKFKGRATLTVDKSSSTAYMHLKSLTSED SAVYYCVSGMEYWGQGTSVTVSS
61	CD28 hinge region nucleic acid sequence	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC
62	CD28 hinge region Amino acid sequence	IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
	Amino goid compand	

FIG. 25 (cont'd)

U. 23 ((cont a)	
64	CD28 transmembrane	FWVLVVVGGVLACYSLLVTVAFIIFWV
	region	
	Amino acid sequence	
65	CD28 intracellular	AGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAA
	domain	CATGACTCCCCGCCCCCCGGGCCCACCCGCAAGCATTACCA
	nucleic acid sequence	GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
66	CD28 intracellular	RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
	domain	
	Amino acid sequence	
67	CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA
	domain	GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
	nucleic acid sequence	AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
		ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
		GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
		GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG
		GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA
		CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC
60	CD2 mate feature at 11 1 1	CTCGC DAWESDS ADADAYWOCONOL VAIEL NIL CRREEVENT DERRORDE
68	CD3 zeta intracellular	RVKFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP
	domain	EMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG HDGLYQGLSTATKDTYDALHMQALPPR
69	Amino acid sequence ecDHFR destabilization	ATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGC
09	domain (R12Y, G67S,	ATGGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCTCGCC
	Y100I)	TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC
	nucleic acid sequence	CGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGACGC
	nucleic acid sequence	AAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC
		GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT
		GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGCGTT
		ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG
		CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT
		TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC
		GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC
		TGGAGCGGCGA
70	ecDHFR destabilization	ISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMGR
	domain (R12Y, G67S,	HTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIAACGDVP
	Y100I)	EIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEGDTHFPDYEPDDW
	Amino acid sequence	ESVFSEFHDADAQNSHSYCFEILERR
71	GD2.28z.ecDHFR (R12H,	ATGCTGCTGCTGACATCTCTGCTGCTGTGCGAGCTGCCCC
	N18T, V19A, G67S)	ACCCCGCCTTTCTGCTGATCCCCGATATCCTGCTGACCCAGAC
	nucleic acid sequence	CCCTCTGAGCCTGCCTGTGTCTCTGGGCGATCAGGCCAGCATC
		AGCTGCAGATCCAGCAGAGCCTGGTGCACCAGGACGCCAGAGAGGCCAG
		CACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGAGCCC
		CAAGCTGCTGATTCACAAGGTGTCCAACCGGTTCAGCGGCGT GCCCGACAGATTTTCTGGCAGCGGCTCCGGCACCGACTTCAC
		CCTGAAGATCAGCCGGGTGGAAGCCGACCTGAC
		ACTTCTGCAGCCAGTCCACCCACGTGCCCCCCTGACATTTGG
		CGCCGGAACAAAGCTGGAACTGAAGGGCAGCACAAGCGGCA
		GCGGCAAGCCTGGATCTGGCGAGGGAAGCACCAAGGGCGAA
		GTGAAGCTGCAGCAGAGCGGCCCCTCTCTGGTGGAACCTGGC
		GCCTCTGTGATGATCTCCTGCAAGGCCAGCGGCAGCTCCTTC
		ACCGCTACAACATGAACTGGGTGCGCCAGAACATCGGCAA
		GAGCCTGGAATGGATCGGCGCCATCGACCCCTACTACGGCGG
		CACCAGCTACAACCAGAAGTTCAAGGGCAGAGCCACCCTGAC
		CGTGGACAAGAGCAGCTCCACCGCCTACATGCACCTGAAGTC
		COTOGRESAGAGETCEACCOCCTACATOCACCTGAAGTC
		CCTGACCAGCGAGGACAGCGCCGTGTACTACTGCGTGTCCGG
		CCTGACCAGCGAGGACAGCGCCGTGTACTACTGCGTGTCCGG
		CCTGACCAGCGAGGACAGCGCCGTGTACTACTGCGTGTCCGG CATGGAATACTGGGGCCAGGGCACAAGCGTGACCGTGTCCTC

FIG. 25 (cont'd)

<u> IG. 25 (c</u>	ont'd)	
		CCCTTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCTTGCT
		ATAGCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTGAG
		GAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAACAT
		GACTCCCCGCCCCCGGGCCCACCCGCAAGCATTACCAGCC
		CTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCCCATATG
		AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAA
		GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
		AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
		ACCCTGAGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
		GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
		GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG
		GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA
		CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC
		CTCGCATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTAT
		CGGCATGGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCT
		CGCCTGGTTTAAACGCAACACCTTAAATAAACCCGTGATTAT
		GGGCCGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGG
		ACGCAAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGA
		TCGCGTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGC
		GTGTGGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCG
		CGTTATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCT
		GACGCATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCC
		GGATTACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATT
		CCACGATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGA
		GATTCTGGAGCGGCGATGA
72	ecDHFR destabilization	MLLLVTSLLLCELPHPAFLLIPDILLTQTPLSLPVSLGDQASISCRS
	domain (R12Y, G67S,	SQSLVHRNGNTYLHWYLQKPGQSPKLLIHKVSNRFSGVPDRFSG
	Y100I)	SGSGTDFTLKISRVEAEDLGVYFCSQSTHVPPLTFGAGTKLELKG
	Amino acid sequence	STSGSGKPGSGEGSTKGEVKLQQSGPSLVEPGASVMISCKASGSS
	Times acid sequence	FTGYNMNWVRQNIGKSLEWIGAIDPYYGGTSYNQKFKGRATLT
		VDKSSSTAYMHLKSLTSEDSAVYYCVSGMEYWGQGTSVTVSSA
		SFEIEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKPFWVL
		VVVGGVLACYSLLVTVAFIIFWVRSKRSRLLHSDYMNMTPRRP
		GPTRKHYQPYAPPRDFAAYRSHMRVKFSRSADAPAYKQGQNQL
		YNELNLGRREEYDVLDKRRGRDPEMGGKPRRKNPQEGLYNEL
		QKDKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDTYDAL
		HMQALPPRISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTL
		NKPVIMGRHTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEA
		IAACGDVPEIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEGDTHFP
		DYEPDDWESVFSEFHDADAQNSHSYCFEILERR
73	GD2.28z.ecDHFR (R12H,	ATGCTGCTGCTGACATCTCTGCTGCTGTGCGAGCTGCCCC
	N18T, V19A, G67S)	ACCCCGCCTTTCTGCTGATCCCC
	Leader sequence	
	nucleic acid sequence	
74	GD2.28z.ecDHFR (R12H,	MLLLVTSLLLCELPHPAFLLIP
1 .	N18T, V19A, G67S)	
	Leader sequence	
	Amino acid sequence	
75	14G2a scFv (GD2	GATATCCTGCTGACCCAGACCCCTCTGAGCCTGCCTGTGTCTC
'3		
	binding domain)	TGGGCGATCAGGCAGCATCAGCTGCAGTCCAGCCAGAGCC
	nucleic acid sequence	TGGTGCACCGGAACGCCAACACCTACCTGCACTGTATCTGC
		AGAAGCCCGGCCAGAGCCCCAAGCTGCTGATTCACAAGGTGT
		CCAACCGGTTCAGCGGCGTGCCCGACAGATTTTCTGGCAGCG
		GCTCCGGCACCGACTTCACCCTGAAGATCAGCCGGGTGGAAG
		CCGAGGACCTGGGCGTGTACTTCTGCAGCCAGTCCACCCAC
		TGCCCCCCTGACATTTGGCGCCGGAACAAAGCTGGAACTGA
		AGGGCAGCACAAGCGGCAGCGGCAAGCCTGGATCTGGCGAG
		GGAAGCACCAAGGGCGAAGTGAAGCTGCAGCAGAGCGGCCC
		CTCTCTGGTGGAACCTGGCGCCTCTGTGATGATCTCCTGCAAG
		GCCAGCGCAGCTCCTTCACCGGCTACAACATGAACTGGGTG
L	I.	,

<u>IG. 25</u>	(cont'd)	
		CGCCAGAACATCGGCAAGAGCCTGGAATGGATCGGCGCCATC
		GACCCCTACTACGGCGGCACCAGCTACAACCAGAAGTTCAAG
		GGCAGAGCCACCCTGACCGTGGACAAGAGCAGCTCCACCGCC
		TACATGCACCTGAAGTCCCTGACCAGCGAGGACAGCGCCGTG
		TACTACTGCGTGTCCGGCATGGAATACTGGGGCCAGGGCACA
		AGCGTGACCGTGTCCTCT
76	14G2a scFv (GD2	DILLTQTPLSLPVSLGDQASISCRSSQSLVHRNGNTYLHWYLQKP
'	binding domain)	GQSPKLLIHKVSNRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGV
	Amino acid sequence	YFCSQSTHVPPLTFGAGTKLELKGSTSGSGKPGSGEGSTKGEVK
	rimino acia sequence	LQQSGPSLVEPGASVMISCKASGSSFTGYNMNWVRQNIGKSLE
		WIGAIDPYYGGTSYNQKFKGRATLTVDKSSSTAYMHLKSLTSED
		SAVYYCVSGMEYWGQGTSVTVSS
77	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
' '	nucleic acid sequence	AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT
	nucieic acid sequence	
70	CD20 his second second	CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
78	CD28 hinge region	IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
5 0	Amino acid sequence	
79	CD28 transmembrane	TTTTGGGTGCTGGTGGTGGTGGGGGGAGTCCTGCTATA
	region	GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG
	nucleic acid sequence	
80	CD28 transmembrane	FWVLVVVGGVLACYSLLVTVAFIIFWV
	region	
	Amino acid sequence	
81	CD28 intracellular	AGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAA
	domain	CATGACTCCCGCCCCCGGGCCCACCCGCAAGCATTACCA
	nucleic acid sequence	GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
82	CD28 intracellular	RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
	domain	
	Amino acid sequence	
83	CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA
	domain	GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
	nucleic acid sequence	AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
		ACCCTGAGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
		GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
		GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG
		GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA
		CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC
		CTCGC
84	CD3 zeta intracellular	RVKFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP
	domain Amino acid	EMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG
	sequence	HDGLYQGLSTATKDTYDALHMQALPPR
85	ecDHFR destabilization	ATGATCAGTCTGATTGCGGCGTTAGCGGTAGATCACGTTATC
	domain (R12H, N18T,	GGCATGGAAACCGTCATGCCGTGGAACCTGCCTGCCGATCTC
	V19A, G67S)	GCCTGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATG
	nucleic acid sequence	GGCCGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGA
	_	CGCAAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGAT
		CGCGTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCG
		TGTGGTGACGTACCAGAAATCATGGTTATTGGCGGCGGTCGC
		GTTTATGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTG
		ACGCATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCG
		GATTACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTC
		CACGATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAG
		ATTCTGGAGCGGCGA
86	ecDHFR destabilization	ISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMGR
	domain (R12H, N18T,	HTWESIGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIAACGDVP
	V19A, G67S)	EIMVIGGGRVIEQFLPKAQKLYLTHIDAEVEGDTHFPDYEPDDW
	Amino acid sequence	ESVFSEFHDADAQNSHSYCFEILERR
87	HA-GD2.28z.FKBP	ATGGAATTCGGCCTGAGCTGGCTGTTTCTGGTGGCCATTCTGA
	nucleic acid sequence	AGGGCGTGCAGTGCTCCAGAGACATCCTGCTGACACAGACAC
	sequence	CTCTGAGCCTGCCTGTGTCTCTGGGAGATCAGGCCAGCATCA
L		, state and control of the control o

88

HA-GD2.28z.FKBP Amino acid sequence

COTTOTAL CALACIDA CONTACTOR CONTACTAL CALALACICA AND CONTACTAL CALACACTOR CALACACTOR CONTACTAL CALACACTOR CA
GCTGTAGAAGCAGCCAGAGCCTGGTGCACAGAAACGGCAAT
ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT
AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG
CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACCC
TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT
TCTGTAGCCAGAGCACACATGTGCCTCCACTGACCTTTGGCG
CTGGCACCAAACTGGAACTTAAAGGCGGCGGAGGATCTGGTG
GTGGTGGATCTGGCGGAGGCGGTTCTGAAGTGAAACTGCAGC
AGTCTGGCCCCTCTCTGGTTGAACCTGGCGCCTCTGTGATGAT
CTCTTGCAAGGCCAGCGGCAGCAGCTTCACCGGCTACAACAT
GAACTGGGTCCGACAGAACATCGGCAAGAGCCTGGAATGGA
TCGGCGCCATCGATCCTTACTACGGCGGCACCAGCTACAACC
AGAAGTTCAAGGGCAGAGCCACACTGACCGTGGACAAGAGC
AGCAGCACAGCCTACATGCACCTGAAGTCCCTGACAAGCGAG
GACAGCGCCGTGTACTACTGTGTGTCCGGCATGAAGTATTGG
GGCCAGGGCACAAGCGTGACCGTGTCTAGCGCTAAGACCACA
CCTCCTAGCGTGTACGGCAGAGTGACAGTGTCCAGCGCCGAG
CCTAAGAGCTGCGACAAGACACACCTGTCCTCCATGTCCA
GCTCCAGAACTGCTCGGCGGACCCTCCGTTTTCCTGTTTCCAC
CTAAGCCAAAGGACACCCTCATGATCAGCAGAACCCCTGAAG
TGACCTGCGTGGTCGATGTCTCCCACGAGGATCCCGAAG
TGAAGTTCAATTGGTACGTGGACGGCGTGGAAGTGCACAACG
CCAAGACCAAGCCTAGAGAGGAACAGTACAACAGCACCTAC
AGAGTGGTGCCGTGCTGACCGTGCTGCATCAGGACTGGCTG
AACGCAAAGAGTACAAGTGCAAAGTCTCCAACAAGGCCCT
GCCTGCTCCTATCGAGAAAACCATCAGCAAGGCCAAGGGCCA
GCCAAGAGAACCCCAGGTTTACACACTGCCTCCAAGCAGGGA
CGAGCTGACCAAGAATCAGGTGTCCCTGACCTGCCTGGTCAA
GGGCTTCTACCCTTCCGATATCGCCGTGGAATGGGAGAGCAA
TGGCCAGCCTGAGAACAACTACAAGACAACCCCTCCTGTGCT
GGACAGCGACGGCTCATTCTTCCTGTACAGCAAGCTGACAGT
GGATAAGTCCCGGTGGCAGCAGGGCAATGTGTTCAGCTGTTC
TGTGATGCACGAGGCCCTGCACAACCACTACACCCAGAAAAG
CCTGTCTCTGAGCCCCGGCAAGAAGGACCCTAAAGCTAGCT
CGAAATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAG
AAGAGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTT
TGTCCAAGTCCCCTATTTCCCGGACCTTCTAAGCCCTTTTGGG
TGCTGGTGGTGGTTGGGGGAGTCCTGGCTTGCTATAGCTTGCT
AGTAACAGTGGCCTTTATTATTTTCTGGGTGAGGAGTAAGAG
GAGCAGGCTCCTGCACAGTGACTACATGAACATGACTCCCCG
CCGCCCGGGCCCACCCGCAAGCATTACCAGCCCTATGCCCC
ACCACGCGACTTCGCAGCCTATCGCTCCAGAGTGAAGTTCAG
CAGGAGCGCAGACGCCCCCGCGTACAAGCAGGGCCAGAACC
AGCTCTATAACGAGCTCAATCTAGGACGAAGAGAGAGAGTAC
GATGTTTTGGACAAGAGACGTGGCCGGGACCCTGAGATGGGG
GGAAAGCCGAGAAGGAAGAACCCTCAGGAAGGCCTGTACAA
TGAACTGCAGAAAGATAAGATGGCGGAGGCCTACAGTGAGA
TTGGGATGAAAGGCGAGCGCCGGAGGGGCAAGGGGCACGAT
GGCCTTTACCAGGGTCTCAGTACAGCCACCAAGGACACCTAC
GACGCCCTTCACATGCAGGCCCTGCCCCCTCGCGGAGTGCAG
GTGGAAACCATCTCCCCAGGAGACGGCGCACCTTCCCCAAG
CGCGGCCAGACCTGCGTGGTGCACTACACCGGGATGCTTGGA
GATGGAAAGAAGTTGACTCCTCCCGGGACAGAAACAAGCC
CTTTAAGTTTATGCTAGGCAAGCAGGAGGTGATCCGAGGCTG
GGAAGAAGGGGTTGCCCAGATGAGTGTGGGTCAGGGAGCCA
AACTGACTATATCTCCAGATTATGCCTATGGTGCCACTGGGC
ACCCAGGCATCATCCCACCACATGCCACTCTCGTCTTCGATGT
GGAGCTTCTAGAACTGGAATGA
MEFGLSWLFLVAILKGVQCSRDILLTQTPLSLPVSLGDQASISCRS
SQSLVHRNGNTYLHWYLQKPGQSPKLLIHKVSNRFSGVPDRFSG

FIG. 25 (cont'd)

		KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPI EKTISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDI
		A VEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQG
		NVFSCSVMHEALHNHYTQKSLSLSPGKKDPKASFEIEVMYPPPY LDNEKSNGTIIHVKGKHLCPSPLFPGPSKPFWVLVVVGGVLACY
		SLLVTVAFIIFWVRSKRSRLLHSDYMNMTPRRPGPTRKHYQPYA
		PPRDFAAYRSRVKFSRSADAPAYKQGQNQLYNELNLGRREEYD
		VLDKRRGRDPEMGGKPRRKNPQEGLYNELQKDKMAEAYSEIG
		MKGERRRGKGHDGLYQGLSTATKDTYDALHMQALPPRGVQVE
		TISPGDGRTFPKRGQTCVVHYTGMLGDGKKVDSSRDRNKPFKF
		MLGKQEVIRGWEEGVAQMSVGQGAKLTISPDYAYGATGHPGII PPHATLVFDVELLELE
89	HA-GD2,28z,FKBP	ATGGAATTCGGCCTGAGCTGGCTGTTTCTGGTGGCCATTCTGA
	nucleic acid sequence	AGGGCGTGCAGTGCTCCAGA
90	HA-GD2.28z.FKBP	MEFGLSWLFLVAILKGVQCSR
	Amino acid sequence	
91	High affinity 14G2a scFv (GD2 binding domain)	GACATCCTGCTGACACAGACACCTCTGAGCCTGCCTGTGTCTC TGGGAGATCAGGCCAGCATCAGCTGTAGAAGCAGCCAGAGC
	nucleic acid sequence	CTGGTGCACAGAAACGGCAATACCTACCTGCACTGGTATCTG
	nucleic acid sequence	CAGAAGCCCGGCCAGTCTCCTAAGCTGCTGATCCACAAGGTG
		TCCAACAGATTCAGCGGCGTGCCCGATAGATTTTCTGGCTCTG
		GCAGCGGCACCGACTTCACCCTGAAGATCTCTAGAGTGGAAG
		CCGAGGACCTGGGCGTGTACTTCTGTAGCCAGAGCACACATG
		TGCCTCCACTGACCTTTGGCGCTGGCACCAAACTGGAACTTA
		AAGGCGGCGGAGGATCTGGTGGTGGTGGATCTGGCGGAGGC GGTTCTGAAGTGAAACTGCAGCAGTCTGGCCCCTCTCTGGTT
		GAACCTGGCGCCTCTGTGATGATCTCTTGCAAGGCCAGCGGC
		AGCAGCTTCACCGGCTACAACATGAACTGGGTCCGACAGAAC
		ATCGGCAAGAGCCTGGAATGGATCGGCGCCATCGATCCTTAC
		TACGGCGCACCAGCTACAACCAGAAGTTCAAGGGCAGAGC
		CACACTGACCGTGGACAAGAGCAGCAGCACAGCCTACATGC
		ACCTGAAGTCCCTGACAAGCGAGGACAGCGCCGTGTACTACT
		GTGTGTCCGGCATGAAGTATTGGGGCCAGGGCACAAGCGTGA
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTCG
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAACTTCAATTGGTACG TGGACGGCGTGGAAGTGCACAACGCCAAGACCAAGCCTAGA GAGGAACAGTACAACAGCACCTACAGAGTGTGTCCGTGCTG ACCGTGCTGCATCAGGACTGCTGAACGGCAAAGAGTACAA GTGCAAAGTCTCCAACAAGGCCTGCCTGCTCCTATCGAGAA AACCATCAGCAAGGCCAAGGGCCAAGAGAACCCAGG TTTACACACTGCCTCCAAGCAGGGACGAGCTGACCAAGAATC AGGTGTCCCTGACCTGCTTGTCAAGGGCTTCTACCCTTCCGA TATCGCCGTGGAATGGGAGAGCAATGGCCAGCCTGAGAACA ACTACAAGACAACCCCTCCTGTGCTGGACAGCGACGCTCAT TCTTCCTGTACAGCAAGCTGACAGTGGCCAGCGCTCAT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACACTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT
92	High affinity 14G2a scFv (GD2 binding domain)	CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG ACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG ATGTGTCCCACGAGGATCCCGAAGTGAAGT

FIG. 25 (cont'd)

Amino acid sequence VFCSQSTINPPLATEGACTRLELKGGGGSGGGGGSGGGSEVKLQ QSGSSLVEPGASVMSCKAGSGSFTGYNMSWYRONIGSKELSDAV YYCVSGMKYWGGGTSVTVSSAFSTGYNMSWYRONIGSKELSDAV YYCVSGMKYWGGGTSVTVSSAFSTGYNMSWYRONIGSKELSDAV YYCVSGMKYWGGGTSVTVSSAFSTGYNMSWYRONIGSKELSDAV YYCVSGMKYWGGGTSVTVSSAFSTGYNMSWYRONIGSKELSDAV YYCVSGMKYWGGGTSVTVSSAFSTGYNMSWYRONIGSKELSDAV YYCVSGMKYWGGGTSVTVSSAFTTPSVTGATTVTVSAFPRSCAFT KTHTCPPCPAPELLGGPSVFLFPPRFKD LLMISRTPEVTCVWDV SHEDPEVKFNWYDYDGVEVINAKTEREREQYNSYTRVNSLTTL HODWLNGKEYKCKVSNKALPAPIEKTISKAKGGPREPOVYTLPP SRDELTKNOVSLTCLVKGFYPSDLAVEWESNGGPENVYKTTPPV LDSDGSFFLYSKLTVDSKSRWQGGNVFSCSVMHEALHINHTYTLPV LDSDGSFFLYSKLTVDSKSRWQGGNVFSCSVMHEALHINHTYTLPV LDSDGSFFLYSKLTVDSKSRWQGGNVFSCSVMHEALHINHTYTLPV LDSDGSFFLYSKLTVDSKSRWQGGGCCTTTCAGAGCAATGAGAA AGCAATCGCCTATTTCCCTGCCTTCACCTAGACAATGAGAAG AGCAATCGAACCATTATCCATGTCAAAAGGGAAACACCTTTGT CAAGTTCCCCATTTCCCGGCCCTTTCAACACCCCTTCCATA GCTTGCAACTAACACACTTCATCCATGACACTTCATA CCTTGCTAGTACACAGTGGCTTTCTTATTATTTTTCTGGGTG PWLVVVGGVLACYSLLVTVAFIIFWV TTTTGGGTGGTGGTGGTGGGGGGAGTCCTGCACAGTGACTACATGAA CATGACTCCCCGCCCCCCCGCGCCCACCCCAAGCATTACCA GCTGCATTGCACAGTGACTTCGAAGCAGTTCCAAGAACAGACATTACCA GCTGCATTAGCCCCACCCCGAGCCCACCCCAAGCAATTACCA GCTGCATTAGCCCCACCCCGAGCCCACCCCAAGCAATTACCA GCTGCATTAGCCCCACCCCGAGCCCACCCCCAAGCAATTACCA GCTGCATTGCAGACCATTCCAAGAACACACTTTCCAAGACAACACACTTTCCAAGACACACTTTCCAAGACCACCCAC	U. 23	(cont a)	
SHEPPEKFNWYDYDGWEWINAKTREREGYNSTYRVSWJLL		Amino acid sequence	QSGPSLVEPGASVMISCKASGSSFTGYNMNWVRQNIGKSLEWIG AIDPYYGGTSYNQKFKGRATLTVDKSSSTAYMHLKSLTSEDSAV
SRDELTKNOVSLTCLVKGFYPSDIAVEWESNGQPINNYKTTPPY LDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS			KTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDV SHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVL
SLSEPGKDPK			SRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPV
CD28 hinge region nucleic acid sequence CAGATGGAACCATTATCCATGTGAAAGGAAACCACTTTGT			
CD28 hinge region	93	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
CD28 transmembrane region Amino acid sequence		nucleic acid sequence	
Amino acid sequence CD28 transmembrane region nucleic acid sequence FWVLVVVGGVLACYSLLVTVAFIIFWV			
CD28 transmembrane region CD28 transmembrane region CD28 transmembrane region Amino acid sequence CD28 transcription CD28 t	94		IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
region mucleic acid sequence 96	05		TTTTCCCTCCTCCTCCTCCTTCCCCCACTCCTCCTATA
mucleic acid sequence FWVLVVVGGVLACYSLLVTVAFIIFWV	93		
CD28 transmembrane region Amino acid sequence			derideradiaacadiddeerriairarrificidddid
Pegion	96		FWVLVVVGGVLACYSLLVTVAFIIFWV
Amino acid sequence CD28 intracellular domain CATGACTCCCGCGCCCCGGGCCCACCCCAAGCATTACCA		I	
domain nucleic acid sequence GCCCTATGCCCGCGCCCACCCGGAGCATTACCA GCCCTATGCCCCACCACGGGACTTCGCAGCCTATCGCTCC GCCCTATGCCCCACCACGGGACTTCGCAGCCTATCGCTCC GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS		Amino acid sequence	
Proceedings CCCTATGCCCACCACGCGACTTCGCACCCCCCCCCCCCC	97		
Section			
domain			
Amino acid sequence AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAA domain	98		RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
99 CD3 zeta intracellular domain nucleic acid sequence			
domain GCAGGGCCAGAACCAGCTCTATAAACGAGCTCAATCTAGGACG AAGAGAGGAGTACCATGTTTTGGACAAGAGACCTGGCCGGG ACCCTGAGATGGGGGGAAAGCCAGAAAGAACCCTACAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGCGGGA GGCCTACAGTGAGATGGGGGGAAAGCCAGAGAGGCCCGGAGGG GCAAGGGCCTGCACATGACACCCACAAGACACTACCAGCCCCTCCCC CTCGC 100	99		A GA GTGA A GTTCA GCA GGA GCGCA GA CGCCCCCGCGTA CA A
nucleic acid sequence			
ACCCTGAGATGGGGGAAAGCCGAGAAGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAAGATAAGAT			
GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCTTCACATGCAGGCCCTCCCC CCTCGC		•	
CCAAGGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA			GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC CTCGC			
CTCGC			
CD3 zeta intracellular domain			
domain	100	CD3 zeta intracellular	
Amino acid sequence	100	I	
FKBP12 destabilization GGAGTGCAGGTGGAAACCATCTCCCCAGGAGACGGGCGCAC CTTCCCCAAGCGCGGCCAGACCTGCGTGGTGCACTACACCGG GATGCTTGGAGATGGAAAGAAAGTTGACTCCTCCCGGGACAG AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT CCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTC AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG CCACTGGGCACCCAGGCATCATCCCACCACTCTCG TCTTCGATGTGGAGCTTCTAGAACTGGAA GVQVETISPGDGRTFPKRGQTCVVHYTGMLGDGKKVDSSRDRN KPFKFMLGKQEVIRGWEEGVAQMSVGQGAKLTISPDYAYGATG HPGIIPPHATLVFDVELLELE 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGGTGTCTCTGGAGACACACACCCTCTGGAGCTGCCTGTTCTGTGACACAGAACAC CTCTGAGCCTGCTGTCTCTGGAGAACACCCTCTGCACAGAAACGCCAGCATCA ACCTACCTGCACTGGTATCTGCAGAAACCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGACCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGATCCACAGAAACCCCTTGCACAAGAACCCCTGCACAGAAACCGCAGTGCTCCTGAAGATTCTGCAGAAGACTCCACAGAAACCCCTGCACAGAAACCCCTGCACACAAGATTCAGCGGCCGTGCCCGATTCACCC TGAAGATTTTCTGGCTCTGGCAGCAGCACCACCCCTTCACCC TGAAGATTTTCTGGCTCTGGCAGCAGCACCCCCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGGCCAGTCTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGACTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGACTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGGCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGACTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGGCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGACTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGGCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGACTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGCCTGTACT		Amino acid sequence	
R71G, K105E) nucleic acid sequence AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT CCGAGGCTGGGAAGAAGAAGTTGACTCCTCCCGGGACAG AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT CCGAGGCTGGGAAGAAGAGGGGTTGCCCAGATTATGCCTATGGTG CCACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCG TCTTCGATGTGGAGCTTCTAGAACTGGAA BY FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGCTGTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCCAGAGACACTCCTGCTGACACAGACAC CTCTGAGCCTGCCTGTTCTCTGGAGAAACCGCAAT ACCTACCTGCACTGGTTCTCTGGAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCTGTTCCT TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCTC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGTTTCCCCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGCGCTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGCGCGTTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTTTACT	101	FKBP12 destabilization	GGAGTGCAGGTGGAAACCATCTCCCCAGGAGACGGCGCAC
nucleic acid sequence AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT CCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTC AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG CCACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCG TCTTCGATGTGGAGCTTCTAGAACTGGAA 102 FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGCTGTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCCAGAGACACTCTGCTGACACAGACAC CTCTGAGCCTGCTGTGTCTCTGGGAGATCAGGCCAGCATCA GCTGTAGAAGCAGCCAGAGCCTGGTGCAACAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTTACT			
CCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTC AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG CCACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCG TCTTCGATGTGGAGCTTCTAGAACTGGAA 102 FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGCTGTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCCAGAGACACTCTGCTGACACAGACAC CTCTGAGCCTGCTGTTCTTGGGAGATCAGGCCAGCATCA GCTGTAGAAGCAGCCAGAGACCTGGTGCCAGAAACCGCAAT ACCTACCTGCACTGGTATCTGCAGAAACCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACC TGAAGATCTCTAGAGTGGAAGCCCGAGCACTGCCC TGAAGATCTCTAGAGTGGAAGCCCGAGCACTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGAGCACTTCACCC			
AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG CCACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCG TCTTCGATGTGGAGCTTCTAGAACTGGAA 102 FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCCAGAGAACACC CTCTGAGCCTGCTGTTCTCTGGAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAACCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCTG CCCGATAGATTTTCTGGCTCTGGCAGCGCACCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGCCTGTTCCT TGAAGATCTCTAGAGTGGAAGCCCGGCCACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGGCACCGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGGCCACTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGACCTGGTGTCTCT		nucleic acid sequence	
CCACTGGGCACCCAGGCATCATCCCACCACATGCCACTCTCG TCTTCGATGTGGAGCTTCTAGAACTGGAA 102 FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCCAGAGAACACCCTCTGACACAGACAC CTCTGAGCCTGCTGTTCTCTGGAGAAACAGCACAC GCTGTAGAAGCAGCCAGAGCCTGGCTGTCCAGAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGCACCTCACCC TGAAGATCTCTAGAGTGGAAGCCCGAGCACTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGAGCCTGTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGAGCCTGTTCACCC			
TCTTCGATGTGGAGCTTCTAGAACTGGAA 102 FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCTCTGAGAACACACCCTCTGAGAACACCAGACACCCTCTGAGAACACCAGACACACAC			
FKBP12 destabilization domain (E31G, F36V, R71G, K105E) Amino acid sequence HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGTGTTTCTGGTGGCCATTCTGA AGGGCGTGCTGTCTCTGGAGAAACGGCAAT ACCTACCTGCAGAGACACCCGGCCAGTCTCCT AAGCTGCTGATCCAGAGAGCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGCCACACCCCGCCTGTTCTCACACAAGATCCTGCTGAAGAACCCCCCGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGCCAACAGATCCCCCTGAAGATTCAGCGGCGTGCCCGAAGAACCCCCCGAAGACCCCCCCC			
domain (E31G, F36V, R71G, K105E) Amino acid sequence HA-GD2.28z.ecDHFR nucleic acid sequence AGGCGTGCAGTGCTCTGGAGATCTGGCCAGACACACACAC	102	FKBP12 destabilization	
Amino acid sequence 103 HA-GD2.28z.ecDHFR nucleic acid sequence ATGGAATTCGGCCTGAGCTGGTTTTCTGGTGGCCATTCTGA AGGGCGTGCAGTGCTCCAGAGACATCCTGCTGACACAGACAC CTCTGAGCCTGCTGTGTCTCTGGGAGATCAGGCCAGCATCA GCTGTAGAAGCAGCCAGAGCCTGGTGCACAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT		domain (E31G, F36V,	
103 HA-GD2.28z.ecDHFR nucleic acid sequence AGGGCGTGCCTGAGCTGGTCTCTGGACACAGACAC CTCTGAGCCTGCTGTCTCTGGAGATCAGGCCAGCATCA GCTGTAGAAGCAGCCAGAGACCTGGTGCACAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCTTCACCC TGAAGATCTCTAGAGTGGAAGCCCGAGGACCTGGCGTGTACT			HPGIIPPHATLVFDVELLELE
nucleic acid sequence AGGGCGTGCAGTGCTCCAGAGACATCCTGCTGACACAGACAC CTCTGAGCCTGCCTGTGTCTCTGGGAGATCAGGCCAGCATCA GCTGTAGAAGCAGCCAGAGCCTGGTGCACAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCTGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT			
CTCTGAGCCTGCCTGTGTCTCTGGGAGATCAGGCCAGCATCA GCTGTAGAAGCAGCCAGAGCCTGGTGCACAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT	103		
GCTGTAGAAGCAGCCAGAGCCTGGTGCACAGAAACGGCAAT ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT		nucleic acid sequence	
ACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCTCCT AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGCACCGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT			
AAGCTGCTGATCCACAAGGTGTCCAACAGATTCAGCGGCGTG CCCGATAGATTTTCTGGCTCTGGCAGCGCACCGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT			
CCCGATAGATTTTCTGGCTCTGGCAGCGCACCGACTTCACCC TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT			
TGAAGATCTCTAGAGTGGAAGCCGAGGACCTGGGCGTGTACT			
TCTGTAGCCAGAGCACATGTGCCTCCACTGACCTTTGGCG			TCTGTAGCCAGAGCACACATGTGCCTCCACTGACCTTTGGCG

HA-GD2.28z.ecDHFR Amino acid sequence

104

CTGGCACCAAACTGGAACTTAAAGGCGGCGGAGGATCTGGTG
GTGGTGGATCTGGCGGAGGCGGTTCTGAAGTGAAACTGCAGC
AGTCTGGCCCCTCTCTGGTTGAACCTGGCGCCTCTGTGATGAT
CTCTTGCAAGGCCAGCGGCAGCAGCTTCACCGGCTACAACAT
GAACTGGGTCCGACAGAACATCGGCAAGAGCCTGGAATGGA
TCGGCGCCATCGATCCTTACTACGGCGGCACCAGCTACAACC
AGAAGTTCAAGGGCAGAGCCACACTGACCGTGGACAAGAGC
AGCAGCACAGCCTACATGCACCTGAAGTCCCTGACAAGCGAG
GACAGCGCCGTGTACTACTGTGTGTCCGGCATGAAGTATTGG
GGCCAGGGCACAAGCGTGACCGTGTCTAGCGCTAAGACCACA
CCTCCTAGCGTGTACGGCAGAGTGACAGTGTCCAGCGCCGAG
CCTAAGAGCTGCGACAAGACACACCTGTCCTCCATGTCCA
GCTCCAGAACTGCTCGGCGGACCCTCCGTTTTCCTGTTTCCAC
CTAAGCCAAAGGACACCCTCATGATCAGCAGAACCCCTGAAG
TGACCTGCGTGGTCGATGTGTCCCACGAGGATCCCGAAG
TGAAGTTCAATTGGTACGTGGACGGCGTGGAAGTGCACAACG
CCAAGACCAAGCCTAGAGAGGAACAGTACAACAGCACCTAC
AGAGTGGTGCCGTGCTGACCGTGCTGCATCAGGACTGGCTG
AACGGCAAAGAGTACAAGTGCAAAGTCTCCAACAAGGCCCT
GCCTGCTCCTATCGAGAAAACCATCAGCAAGGCCAAGGGCCA
GCCAAGAGAACCCCAGGTTTACACACTGCCTCCAAGCAGGGA
CGAGCTGACCAAGAATCAGGTGTCCCTGACCTGCCTGGTCAA
GGGCTTCTACCCTTCCGATATCGCCGTGGAATGGGAGAGCAA
TGGCCAGCCTGAGAACAACTACAAGACAACCCCTCCTGTGCT
GGACAGCGACGCCTCATTCTTCCTGTACAGCAAGCTGACAGT
GGATAAGTCCCGGTGGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCA
TGTGATGCACGAGGCCCTGCACAACCACTACACCCAGAAAAG
CCTGTCTCTGAGCCCCGGCAAGAAGGACCCTAAAGCTAGCT
AAGAGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTT
TGTCCAAGTCCCCTATTTCCCGGACCTTCTAAGCCCTTTTGGG
TGCTGGTGGTTGGGGGAGTCCTGGCTTGCTATAGCTTGCT
AGTAACAGTGGCCTTTATTATTTTCTGGGTGAGGAGTAAGAG
GAGCAGGCTCCTGCACAGTGACTACATGAACATGACTCCCCG
CCGCCCGGGCCCACCCGCAAGCATTACCAGCCCTATGCCCC
ACCACGCGACTTCGCAGCCTATCGCTCCAGAGTGAAGTTCAG
CAGGAGCGCAGACGCCCCCGCGTACAAGCAGGGCCAGAACC
AGCTCTATAACGAGCTCAATCTAGGACGAAGAGAGAGAGTAC
GATGTTTTGGACAAGAGACGTGGCCGGGACCCTGAGATGGGG
GGAAAGCCGAGAAGGAAGAACCCTCAGGAAGGCCTGTACAA
TGAACTGCAGAAAGATAAGATGGCGGAGGCCTACAGTGAGA
TTGGGATGAAAGGCGAGCGCCGGAGGGCAAGGGGCACGAT
GGCCTTTACCAGGGTCTCAGTACAGCCACCAAGGACACCTAC
GACGCCCTTCACATGCAGGCCCTGCCCCCTCGCATCAGTCTG
ATTGCGGCGTTAGCGGTAGATTACGTTATCGGCATGGAAAAC
GCCATGCCGTGGAACCTGCCTGCCGATCTCGCCTGGTTTAAA
CGCAACACCTTAAATAAACCCGTGATTATGGGCCGCCATACC
TGGGAATCAATCGGTCGTCCGTTGCCAGGACGCAAAAATATT
ATCCTCAGCAGTCAACCGAGTACGGACGATCGCGTAACGTGG
GTGAAGTCGGTGGATGAAGCCATCGCGGCGTGTGGTGACGTA
CCAGAAATCATGGTGATTGGCGGCGGTCGCGTTATTGAACAG
TTCTTGCCAAAAGCGCAAAAACTGTATCTGACGCATATCGAC
GCAGAAGTGGAAGGCGACACCCATTTCCCGGATTACGAGCCG
GATGACTGGGAATCGGTATTCAGCGAATTCCACGATGCTGAT
GCGCAGAACTCTCACAGCTATTGCTTTGAGATTCTGGAGCGG
CGATGA MEFGLSWLFLVAILKGVQCSRDILLTQTPLSLPVSLGDQASISCRS
SQSLVHRNGNTYLHWYLQKPGQSPKLLIHKVSNRFSGVPDRFSG
SGSGTDFTLKISRVEAEDLGVYFCSQSTHVPPLTFGAGTKLELKG
GGGSGGGSGGGSEVKLQQSGPSLVEPGASVMISCKASGSSFT

'IG. 25 (c	cont'd)	
		GYNMNWVRQNIGKSLEWIGAIDPYYGGTSYNQKFKGRATLTVD
		KSSSTAYMHLKSLTSEDSAVYYCVSGMKYWGQGTSVTVSSAKT
		TPPSVYGRVTVSSAEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKP
		KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKT
		KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPI
		EKTISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDI
		AVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQG
		NVFSCSVMHEALHNHYTQKSLSLSPGKKDPKASFEIEVMYPPPY
		LDNEKSNGTIIHVKGKHLCPSPLFPGPSKPFWVLVVVGGVLACY
		SLLVTVAFIIFWVRSKRSRLLHSDYMNMTPRRPGPTRKHYQPYA
		PPRDFAAYRSRVKFSRSADAPAYKQGQNQLYNELNLGRREEYD
		VLDKRRGRDPEMGGKPRRKNPQEGLYNELQKDKMAEAYSEIG
		MKGERRRGKGHDGLYQGLSTATKDTYDALHMQALPPRISLIAA
		LAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMGRHTWES
		IGRPLPGRKNIILSSQPSTDDRVTWVKSVDEAIAACGDVPEIMVIG
		GGRVIEQFLPKAQKLYLTHIDAEVEGDTHFPDYEPDDWESVFSE
		FHDADAQNSHSYCFEILERR
105	HA-GD2.28z.ecDHFR	ATGGAATTCGGCCTGAGCTGGCTGTTTCTGGTGGCCATTCTGA
103	Leader sequence	AGGGCGTGCAGTGCTCGGAGCTGGCTGTTTCTGGTGGCCATTCTGA
		AUUUCUTUCAUTUCTCCAUA
100	nucleic acid sequence	MEECI CUII EL VAIL V CVO COD
106	HA-GD2.28z.ecDHFR	MEFGLSWLFLVAILKGVQCSR
	Leader sequence	
	Amino acid sequence	
107	High affinity 14G2a scFv	GACATCCTGCTGACACAGACACCTCTGAGCCTGCCTGTGTCTC
	(GD2 binding domain)	TGGGAGATCAGGCCAGCATCAGCTGTAGAAGCAGCCAGAGC
	nucleic acid sequence	CTGGTGCACAGAAACGGCAATACCTACCTGCACTGGTATCTG
		CAGAAGCCCGGCCAGTCTCCTAAGCTGCTGATCCACAAGGTG
		TCCAACAGATTCAGCGGCGTGCCCGATAGATTTTCTGGCTCTG
		GCAGCGGCACCGACTTCACCCTGAAGATCTCTAGAGTGGAAG
		CCGAGGACCTGGGCGTGTACTTCTGTAGCCAGAGCACACATG
		TGCCTCCACTGACCTTTGGCGCTGGCACCAAACTGGAACTTA
		AAGGCGGCGGAGGATCTGGTGGTGGTGGATCTGGCGGAGGC
		GGTTCTGAAGTGAAACTGCAGCAGTCTGGCCCCTCTCTGGTT
		GAACCTGGCGCCTCTGTGATGATCTCTTGCAAGGCCAGCGGC
		AGCAGCTTCACCGGCTACAACATGAACTGGGTCCGACAGAAC
		ATCGGCAAGAGCCTGGAATGGATCGGCCCATCGATCCTTAC
		TACGGCGCACCAGCTACAACCAGAAGTTCAAGGGCAGAGC
		CACACTGACCGTGGACAAGAGCAGCAGCACAGCCTACATGC
		ACCTGAAGTCCCTGACAAGCGAGGACAGCGCCGTGTACTACT
		GTGTGTCCGGCATGAAGTATTGGGGCCAGGGCACAAGCGTGA
		CCGTGTCTAGCGCTAAGACCACACCTCCTAGCGTGTACGGCA
		GAGTGACAGTGTCCAGCGCCGAGCCTAAGAGCTGCGACAAG
		ACACACACCTGTCCTCCATGTCCAGCTCCAGAACTGCTCGGC
		GGACCCTCCGTTTTCCTGTTTCCACCTAAGCCAAAGGACACCC
		TCATGATCAGCAGAACCCCTGAAGTGACCTGCGTGGTGGTCG
		ATGTGTCCCACGAGGATCCCGAAGTGAAGTTCAATTGGTACG
		TGGACGCCTGGAAGTGCACAACGCCAAGACCAAGCCTAGA
		GAGGAACAGTACAACAGCACCTACAGAGTGGTGTCCGTGCTG
		ACCGTGCTGCATCAGGACTGGCTGAACGGCAAAGAGTACAA
		GTGCAAAGTCTCCAACAAGGCCCTGCTCCTATCGAGAA
		AACCATCAGCAAGGCCAAGGCCAAGAGAAACCCCAGG
		TTTACACACTGCCTCCAAGCAGGACGAGCTGACCAAGAATC
		AGGTGTCCCTGACCTGCCTGGTCAAGGGCTTCTACCCTTCCGA
		TATCGCCGTGGAATGGGAGAGCAATGGCCAGCCTGAGAACA
		ACTACAAGACAACCCCTCCTGTGCTGGACAGCGACGGCTCAT
		TCTTCCTGTACAGCAAGCTGACAGTGGATAAGTCCCGGTGGC
		AGCAGGGCAATGTGTTCAGCTGTTCTGTGATGCACGAGGCCC
		AGCAGGGCAATGTGTTCAGCTGTTCTGTGATGCACGAGGCCC TGCACAACCACTACACCCAGAAAAGCCTGTCTCTGAGCCCCG
		AGCAGGGCAATGTGTTCAGCTGTTCTGTGATGCACGAGGCCC

FIG. 25 (cont'd)

Amino acid sequence YFCSQSTHVPPLTFGAGTKLELKGGGSGGGGGGGGSGKSUKLQ QSGSSLVEGASAVMSCKAGGSSFTGYNNMVRQNIGKSLENG AIDPYYCGTSYNOKFKGRATLTVDKSSSTAYMHLKSLTSEDSAV YYCVSGMK YWGGGTSYNVSKATTPSPYVGRYVTVSSAPPKSC KTHTCPPCPAPELLGGPSVFLFPPKKDTLMISRTPEVTCVVDV SHEDPEVKKNNVALPPEKKTSKAKGQRPEPQVYTLVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQRPEPQVYTLVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQRPEPQVYTLVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQRPEPQVYTLVL DSDGSFFLYSKLTVDKSRWQGGNVFSCSVMHEALHNBHYTQDK LSLSPGKKDPK ATTGAGGTTATGTATCCTTCCTCTCACTAGACAATGAGAAG AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT CCAAGTCCCCTATTTCCCGGCCTTCTAAGCCC CCAAGTCCCCTATTTCCCGGACCATGGCCC CCAAGTCCCCCTATTTCCCGGCCTTCTAAGCCC IEVMYPPPYLDNEKSNGTIHVKGKHLCPSPLFPGSKP ATTGAGGTGCTGTGAAAGGGGAACACCTTTGT CCAAGTCACCATTTCCCGGCCTTCATAGTCCCTCCTTAATTTTTTTCTGGGTG CCAAGTCACCATGACACGTCCCTATAGTCCCTCCTTAATTTTTTTT	U. 23 ((cont a)	
Amino acid sequence YFCSQSTHVPPLTFGAGTKLELKGGGSGGGGGGGGSGKSUKLQ QSGSSLVEGASAVMSCKAGGSSFTGYNNMVRQNIGKSLENG AIDPYYCGTSYNOKFKGRATLTVDKSSSTAYMHLKSLTSEDSAV YYCVSGMK YWGGGTSYNVSKATTPSPYVGRYVTVSSAPPKSC KTHTCPPCPAPELLGGPSVFLFPPKKDTLMISRTPEVTCVVDV SHEDPEVKKNNVALPPEKKTSKAKGQRPEPQVYTLVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQRPEPQVYTLVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQRPEPQVYTLVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQRPEPQVYTLVL DSDGSFFLYSKLTVDKSRWQGGNVFSCSVMHEALHNBHYTQDK LSLSPGKKDPK ATTGAGGTTATGTATCCTTCCTCTCACTAGACAATGAGAAG AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT CCAAGTCCCCTATTTCCCGGCCTTCTAAGCCC CCAAGTCCCCTATTTCCCGGACCATGGCCC CCAAGTCCCCCTATTTCCCGGCCTTCTAAGCCC IEVMYPPPYLDNEKSNGTIHVKGKHLCPSPLFPGSKP ATTGAGGTGCTGTGAAAGGGGAACACCTTTGT CCAAGTCACCATTTCCCGGCCTTCATAGTCCCTCCTTAATTTTTTTCTGGGTG CCAAGTCACCATGACACGTCCCTATAGTCCCTCCTTAATTTTTTTT		(GD2 binding domain	GQSPKLLIHKVSNRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGV
OSGPSLVPGGASVMISCKASGSSFIGYNMNWRQNIGKSLEWIG ADPYYGGTSYNQKFKGRATLTVDKSSSTAYMHLKSLTSEDS YYCVSGMK YWGGGTSVTVSSAKTTPPSVYGRVTVSSAEPKSCD KTHTCPPCPAPELIGGPSVFLPPKPKDTLMISK FTEPVTCVVVSUSHEDPEVKENWYDGVEVFINAKTK/PREEDYNSTYRVVSVLTVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPP SRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPV LDSDGSFFLYSKLTVDKSSRWQGGNVFSCSVMHEALHNHYTQKS LSLSPGKKDPK ATTGAAGTTATGTATCTCTCCTTACCTAGACAATGAAGAAG AGAAGACACATTATCCATTGAAAAGGGAAACACCTTTGT CCAAGTCCCCAATTATCCATGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGTGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGTG CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGCATAA GCTTGCTGTTGCTAAGCC CCAAGTCCCCCAATTATCCATGAAAGGGAAACACCTTGCATAA GCTTGCTGCTGTGTGTGGTGGTGGTGGTGGTGGTGGTGGT		Amino acid sequence	YFCSOSTHVPPLTFGAGTKLELKGGGGSGGGGGGGGGSEVKLO
ADPYYGGTSYNOKEKGRATLTVDKSSSTAYMHLKSLTSEDSAV YYCVSGMK YWGGGTSVTVSSAKTTPSPVYGRVTVSSAEPSKA YYCVSGMK YWGGGTSVTVSSAKTTPSPVYGRVTVSSAEPSKA HTTCPPCPAPELLGGPSVFLFPPKRDTLMISRTPEVTCVVDV SHEDPEVKFNWYDGVEVHNAKTKPREEQYNSTYR VVSVLTVL HQDWLMGKETKCKVSNKALPAPIEKTISKAKGQREPGVYTLPP SRDELTKNQVSLTCLVKGFYPSDIAVEWESNGGPENVYTLTPPV LDSDGSFFLYSKLTVDKSRWQGGNVFSCSVMHEALHNHYTQDK LSLSPGKRDPK ATTGAGGTTATGTATCCTCTCCTCACCTAGCAATGAGAAG AGCAATGGAACCATTATCCATGGAAAGGGAAACACCTTTGT CCAAGTCCCCTATTTCCCGGACCTTCAAGCCC CCAB hinge region Amino acid sequence CD28 transmembrane region Amino acid sequence CD28 transmembrane region Amino acid sequence CD28 intracellular domain mucleic acid sequence AGGAGTAGAGGAGCTCCTGCACAGTGACTACATGAA Amino acid sequence CD28 intracellular domain Amino acid sequence AGGAGTAGAGGGAGCCCCCGGGGCCCACCCGCAAGCATTACCA CATGACTCCCCGCCGCCCCCGGGCCCACCCGCAAGCATTACCA CATGACTCCCCGCGCCCCCGGGCCCACCCGCAAGCATTACCA CATGACTCCCCGCGCCCCCGGGCCCACCCGCAAGCACTACCA CACGAGGCCAGAACCACACGCCTCTACAACGAGACCACCACGGGG GCCATAGCCCCACAGGGGGGAACACACCACAC			
PYCVSGMKYWGGGTSVTVSSAKTTPPSVYGRVTVSSAEPKSCD SHEDPEVKFNWYVDGVGVFNRATTKPREGYNSTTPRVVSVLTVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPP SRDELTKNQVSLTCLVKGFVPSDIAVEWESSGQPENNYKTTPPV LDSDGSFFLYSKLTVDKSSRWQGGNFSCSVMHEALHNHYTOK LSLSPGKKDPK ATTGAAGTTATGTATCCTCTCCTTACCTAGACAATGAGAAG ATTGAAGTTATGTATCCTCCCTAGACAATGAGAAG ATTGAAGTTATGTATCCTCCTCAGACAATGAGAACCCTTTGT CCAAGTCCCCCAATTATCCAGGAACCACTTAGGCACA CCAAGTCCCCCAATTATCCAGGAACACCCC CCAAGTCCCCCTATTTCCGGGACCTTCAGCACC CCAAGTCCCCCTATTTCCGGGACCTTCTAGCGCC CCTCCGC CCCCACGCAAGCATTATCCATTATATTTTTCTGGGTG CCTTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTC			
KTHICPPCPAPELLIGGESVELPPRENDTIMISRIPEVTCVVVDV SHEDPEVKRIWVDGVEVINAKTKPREEGEVNSTTYR VVSUTVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGOPREPQVYTLPP SRDELTKNOVSLITLLVKGFYPSDLAVEWESNGOPENNYKTIPPV LDSDGSFFLYSKLTVDKSRWQGGNYFSCSWHEALINHYTQKS LSLSPGKKDPK ATTGAAGTTATGTATCCTCTCTCTACCTAGACAATGAGAAG AGCAATGGACCATTATCCATGTGAAAGGGAAACACCTTTGT CD28 hinge region ATTGAAGTTATGTATCCTCCTCTACCTAGACAATGAGAAG AGCAATGGACCATTATCCATGTGAAAGGGAAACACCTTTGT CD28 transmembrane region IEVMYPPPYLDNEKSNGTIHVKGKHLCPSPLFPGPSKP CD28 transmembrane region Amino acid sequence CD28 intracellular domain Anino acid sequence CD28 intracellular domain Anino acid sequence CD28 intracellular domain Anino acid sequence CD3 zeta intracellular domain Anino acid sequence CD3 zeta intracellular domain AGGAGGACCACCCACCAGCCAGCCCCCCGCGAGCCCCCCGCGAGCCGGCCCCCC			
SHEDPEVKINWYDGVEVHNAKTKPREEQYNSTYR VVSVLTVL HODWLNGKGKYEKCKVSNKAL APPIEKTISK AKGOPREPQVTLPP SRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPV LDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALINHIYTQKS LSLSFGKSDPK			
HQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPP SRDELTKNQVSLTCLVKGFYPSDLAVEWESSGGPENNYKTTPPV LSLSPGKKDPK			KTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDV
SRDELTKNOVSLTCLVKGFYPSDIAVEWESNGOPENNYKTTPPV LDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS LSLSPGKKDPK			SHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVL
SRDELTKNOVSLTCLVKGFYPSDIAVEWESNGOPENNYKTTPPV LDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS LSLSPGKKDPK			HODWLNGKEYKCKVSNKALPAPIEKTISKAKGOPREPOVYTLPP
LDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS LSLSPGKKDPK			
LSLSPGKEDPK			
CD28 hinge region nucleic acid sequence			
nucleic acid sequence AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT			
CD28 hinge region	109	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
CD28 hinge region		nucleic acid sequence	AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT
CD28 tinge region		_	CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC
Amino acid sequence CD2 transmembrane region nucleic acid sequence FWVLVVVGGVLACYSLLVTVAFIIFWV	110	CD28 hinge region	
CD28 transmembrane region	110		IE VIVITITI TEDNEKSNOTIIII VKOKIIECI SI EIT OI SKI
region GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG mucleic acid sequence region Amino acid sequence 113			
Italian	111	I	
Total		region	GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG
Total		nucleic acid sequence	
Pregion Amino acid sequence	112		FWVLVVVGGVLACYSLLVTVAFIIFWV
Amino acid sequence CD28 intracellular domain AGGAGTAAGAGGAGCAGGCCCACCGCAAGCATTACCA			
113 CD28 intracellular domain			
domain			4 GG 4 GT 4 4 G 4 GG 4 GG 6 GT GT GT GT G 4
Description CCCTATGCCCACCACGCGACTTCGCACCTCCCCCCCCCC	113	I	
CD28 intracellular domain			
Its CD3 zeta intracellular domain Amino acid sequence AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAGAGAGAGAGACGTGGCCGGG AAGAGGAGTACGATGTTTTGGACAAGAGAGAGAGACGTCGGGG AAGAGGAGTACGATGGCCTTAACAGAGAGAGAGAGACGTCGGGG AAGAGGAGTACGAATGGAAAGACAGAAGAGAGAGAGGGGGAAGGGCCTTACAATGGAAGGCAGAGGGGGAAGGGCCTTACAATGAACTGAGAAGAGAAGATAAGACTGCGGGA GGCCTACAGTGGAATTGGGATTGACAAGGCAGCCCCCCCC		nucleic acid sequence	GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
Its CD3 zeta intracellular domain Amino acid sequence AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAGAGAGAGAGACGTGGCCGGG AAGAGGAGTACGATGTTTTGGACAAGAGAGAGAGACGTCGGGG AAGAGGAGTACGATGGCCTTAACAGAGAGAGAGAGACGTCGGGG AAGAGGAGTACGAATGGAAAGACAGAAGAGAGAGAGGGGGAAGGGCCTTACAATGGAAGGCAGAGGGGGAAGGGCCTTACAATGAACTGAGAAGAGAAGATAAGACTGCGGGA GGCCTACAGTGGAGTTGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCCC CTCGC CTCGC CTCGC RVKFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP EMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG HDGLYQGLSTATKDTYDALHMQALPPR ATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGC AGAAAATATTATCCTCAGCAGACCCTTAAATAAACCCGTGCTGCCGATCTCGCC AGAAAATATTATCCTCAGCAGACCCTTAAATAAACCCATGCGGTGCGATCTCGCC GTGACGTGGAACTCAACCGGGTGATCGCGTGGAACACCTTTCCCCGATTACGGC CATATCGACGAAATCAATCGGTCGTCCGTTGCCAGACGC CATATCGACGAAATCAATCGGTGATTAGCGGCGTTACGGTGGTGAACACCACTATCCGCGCTT ATTGAACAGTTCTCCCCAAAAGCCCAAAAACTCTTCACCGGCCGTT ATTGAACAGTTCTTCCCAAAAGCCCAAAAACTTCACCGGCCGTT ATTGAACAGTTCTTCCCAAAAGCCCAAAAACTTCACCGGCCGTT ATTGAACAGTTCTTCCCAAAAGCCCAAAAACTTCACCGGCCGTT ATTGACCGCGAACACCTTCACAGCCAAAAACTTCACCGGCCGTT ATTGACCGCGAACTCGCGAAACCCATTTCCCCGAT TACGACCCGAAAACTCGTGAATCCACCAAAACCTCTTCCCCGAT TACGACCCGAAAACTCTCACAGCTATTCCACGATTCCACGCCCTTTCCCCGATTTCCCCCAACATTCCACACTTTCCCCGATTTCCCCCAACACCTCTCCTCTCTCT	114	CD28 intracellular	RSKRSRLLHSDYMNMTPRRPGPTRKHYOPYAPPRDFAAYRS
Amino acid sequence			
CD3 zeta intracellular domain AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATTTTTGGACAAGAAGAGACGTGGCCGGG AACCCTGAGATGGGGGGAAAGCCGAGAAGAAGAACCCTCAG GAAGGCCTACAATGAACTGCAGAAGAAGAAGACCCTCAG GAAGGCCTACAATGAACTGCAGAAGAAGAAGACCCTCAG GCACTACAGTGGACTGCCCGGAGGG GCAAGGGGCACGACCCGGAGGG GCAAGGGCACCACTACAATGAACTGCAGAAAGATAAGAT			
domain GCAGGGCCAGAACCAGCTCTATAAACGAGCTCAATCTAGGACG AAGAGAGGAGTACCATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGAACACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA GCCAAGGGCACCTACAGTGAGATTGGCAGTACAGCCC CCAGGGCCCTTCACATGCAGGCCCTTCACATGCAGGCCCTTCCCC CTCGC 116	115		
AGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGAAAACCCTCAG GAAGGCTGTACAATGAACTGCAGAAAGATAAAGTGGCGGGA GGCCTACAGTGAGATTGGGATGAAAGGCGAGGCGCGGAGGG GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTCCCCC CTCGC	113	I	
ACCCTGAGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAAGATAAGAT			
GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA GGCCTACAGTGAGATTGGGATGAAAGGCGCCCGGAGGG GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCCC CTCGC		nucleic acid sequence	AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCTTCACATGCAGGCCCTCCCC CTCGC			ACCCTGAGATGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG GCAAGGGCACGATGGCCTTTACCAGGGTCTCAGTACAGCCA CCAAGGACACCTACGACGCCTTCACATGCAGGCCCTCCCC CTCGC			GAAGGCCTGTACAATGAACTGCAGAAAGATAAGATGGCGGA
CD3 zeta intracellular domain RVFSRSADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDP EMGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG HDGLYQGLSTATKDTYDALHMQALPPR			GGCCTACAGTGAGATTGGGATGAAAGGCGAGCGCCGGAGGG
CCAAGGACACCTACGACGCCCTTCACATGCAGGCCCTGCCC CTCGC			
CTCGC			
CD3 zeta intracellular domain Amino acid sequence EMGGKPRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG HDGLYQGLSTATKDTYDALHMQALPPR			
domain			
Amino acid sequence	116	CD3 zeta intracellular	
ATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGC ATGGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCTCGCC TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC CGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGACGC AAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTGT ATTGAACAGTTCTTGCCAAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGCCAACCCATTTCCCGGAT TACGAGCCGGATGACTGGAAGCCAACACCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCGTATTCACGGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118		domain	EMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG
ATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGC ATGGAAAACGCCATGCCGTGGAACCTGCCTGCCGATCTCGCC TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC CGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGACGC AAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTGT ATTGAACAGTTCTTGCCAAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGCCAACCCATTTCCCGGAT TACGAGCCGGATGACTGGAAGCCAACACCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCGTATTCACGGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118		Amino acid sequence	HDGLYQGLSTATKDTYDALHMQALPPR
domain (R12Y, G67S, Y100I) nucleic acid sequence	117		ATCAGTCTGATTGCGGCGTTAGCGGTAGATTACGTTATCGGC
TGGTTTAAACGCAACACCTTAAATAAACCCGTGATTATGGGC CGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGACGC AAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC GTAACGTGGGTGAAGTCGGTGGATTGGCGGCGTGT GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTGT ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGCGAACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence SVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ATGCTGCTGCTCGTGACATCCCGGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCTCTTGTGGGCGATAGGCTCACAT CCCCGAGCTCCCTGTTCCGCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTTTTGGGCCGATATCCACAT			
nucleic acid sequence CGCCATACCTGGGAATCAATCGGTCGTCCGTTGCCAGGACGC AAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGCGTT ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ACCCCGCCTTTCTGCTGATCCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTGTGGGCGATAGCCCAGTC CCCGAGCTCCCTGTCCGCCTCTTGTGGGCGATAGGCTCACCAT		` ' '	
AAAAATATTATCCTCAGCAGTCAACCGAGTACGGACGATCGC GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTGT ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence AAAAATATTATCCTCAGCAGTCACCCGATCCCCC ACCCCGCCTTTCTGCTGACATCTGGTGCGAGCTGCCCC ACCCCGCCTTTCTGCTGATCCCCGATATCCAGGTCACCATC CCCGAGCTCCCTGTCCGCCTCTTGGGGCGATAGCCCAGTC CCCGAGCTCCCTGTCCGCCTCTTTTGGGGCGATAGGGTCACCAT			
GTAACGTGGGTGAAGTCGGTGATGAAGCCATCGCGGCGTGT GGTGACGTACCAGAAATCATGGTGATTGGCGGCGTGTT ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ACCCCGCCTTTCTGCTGATCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTTTTGGGCGATATCCAGATGACCCATC CCCGAGCTCCCTGTCCGCCTCTTTGGGGCGATAGCCCATC		nucieic acia sequence	
GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGCGTT ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ACCCCGCCTTTCTGCTGATCCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTTGGGGCGATAGCCCAGTC CCCGAGCTCCCTGTCCGCCTCTTTGGGGCGATAGGGTCACCAT			
ATTGAACAGTTCTTGCCAAAAAGCGCAAAAACTGTATCTGACG CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ATGCTGCTGCTGCTGCTGCTGCTGCTGCGAGCTGCCCC ACCCCGAGCTCCCTGTCCGCCTCTTTGGGGCGATAGCCCATC CCCGAGCTCCCTGTCCGCCTCTTTGGGGCGATAGGCTCACCAT			GTAACGTGGGTGAAGTCGGTGGATGAAGCCATCGCGGCGTGT
CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ACCCCGCCTTTCTGCTGACATCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTTGGGGCGATAGGGTCACCAT			GGTGACGTACCAGAAATCATGGTGATTGGCGGCGGTCGCGTT
CATATCGACGCAGAAGTGGAAGGCGACACCCATTTCCCGGAT TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP nucleic acid sequence ACCCCGCCTTTCTGCTGACATCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTTGGGGCGATAGGGTCACCAT			ATTGAACAGTTCTTGCCAAAAGCGCAAAAACTGTATCTGACG
TACGAGCCGGATGACTGGGAATCGGTATTCAGCGAATTCCAC GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA 118 ecDHFR destabilization domain (R12Y, G67S, Y100I) Amino acid sequence 119 Her2.28z.FKBP nucleic acid sequence ACCCCGCCTTTCTGCTGAGACTCTGTGGGAGATTCCAC CCCGAGCTCCCTGTCCGCCTCTTGTGGGCGATATCCAGATGACCCATC CCCGAGCTCCCTGTCCGCCTCTTTGGGGCGATATCCAGATGACCCATC			
GATGCTGATGCGCAGAACTCTCACAGCTATTGCTTTGAGATTC TGGAGCGGCGA			
TGGAGCGGCGA			
ISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMGR domain (R12Y, G67S, Y100I)			
domain (R12Y, G67S, Y100I)			
Y100I)	118	ecDHFR destabilization	ISLIAALAVDYVIGMENAMPWNLPADLAWFKRNTLNKPVIMGR
Y100I)		domain (R12Y. G67S.	HTWESIGRPLPGRKNIILSSOPSTDDRVTWVKSVDEAIAACGDVP
Amino acid sequence ESVFSEFHDADAQNSHSYCFEILERR 119 Her2.28z.FKBP ATGCTGCTGCTGACATCTCTGCTGCTGCGAGCTGCCCC			· ·
119 Her2.28z.FKBP ATGCTGCTGCTGACATCTCTGCTGCTGCGAGCTGCCCC nucleic acid sequence ACCCCGCCTTTCTGCTGATCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTGTGGGCGATAGGGTCACCAT			
nucleic acid sequence ACCCCGCCTTTCTGCTGATCCCCGATATCCAGATGACCCAGTC CCCGAGCTCCCTGTCCGCCTCTGTGGGCGATAGGGTCACCAT	110		
CCCGAGCTCCCTGTCCGCCTCTGTGGGCGATAGGGTCACCAT	119		
		nucleic acid sequence	
CACCTGCCGTGCCAGTCAGGATGTGAATACTGCTGTAGCCTG			CCCGAGCTCCCTGTCCGCCTCTGTGGGCGATAGGGTCACCAT
			CACCTGCCGTGCCAGTCAGGATGTGAATACTGCTGTAGCCTG

<u>IG. 25 (</u>	cont u)	T =
		GTATCAACAGAAACCAGGAAAAGCTCCGAAACTACTGATTTA
		CTCGGCATCCTTCATTCTGGAGTCCCTTCTCGCTTCTCTG
		GATCTAGATCTGGGACGGATTTCACTCTGACCATCAGCAGTC
		TGCAGCCGGAAGACTTCGCAACTTATTACTGTCAGCAACATT
		ATACTACTCCTCCCACGTTCGGACAGGGTACCAAGGTGGAGA
		TCAAAGGGTCTACATCTGGATCTGGGAAGCCGGGTTCTGGTG
		AGGGTTCTGGTGAGGTTCAGCTGGTGGAGTCTGGCGGTGGCC
		TGGTGCAGCCAGGGGGCTCACTCCGTTTGTCCTGTGCAGCTTC
		TGGCTTCAACATTAAAGACACCTATATACACTGGGTGCGTCA
		GGCCCCGGGTAAGGGCCTGGAATGGGTTGCAAGGATTTATCC
		TACGAATGGTTATACTAGATATGCCGATAGCGTCAAGGGCCG
		TTTCACTATAAGCGCAGACACATCCAAAAACACAGCCTACCT
		GCAGATGAACAGCCTGCGTGCTGAGGACACTGCCGTCTATTA
		TTGTTCTAGATGGGGAGGGGACGGCTTCTATGCTATGGACGT
		GTGGGGTCAAGGAACCCTGGTCACCGTCTCCTCGGCTAGCGA
		ACAAAAACTCATCTCAGAAGAGGATCTGTTCGAAATTGAAGT
		TATGTATCCTCCTCCTTACCTAGACAATGAGAAGAGCAATGG
		AACCATTATCCATGTGAAAGGGAAACACCTTTGTCCAAGTCC
		CCTATTTCCCGGACCTTCTAAGCCCTTTTGGGTGCTGGTGGTG
		GTTGGGGGAGTCCTGGCTTGCTATAGCTTGCTAGTAACAGTG
		GCCTTTATTATTTTCTGGGTGAGGAGTAAGAGGAGCAGGCTC
		CTGCACAGTGACTACATGAACATGACTCCCCGCCGCCCCCGGG
		CCCACCGCAAGCATTACCAGCCCTATGCCCCACCACGCGAC
		TTCGCAGCCTATCGCTCCAGAGTGAAGTTCAGCAGGAGCGCA
		GACGCCCCGCGTACAAGCAGGGCCAGAACCAGCTCTATAAC
		GAGCTCAATCTAGGACGAAGAGAGGAGTACGATGTTTTGGAC
		AAGAGACGTGGCCGGGACCCTGAGATGGGGGGGAAAGCCGAG
		AAGGAAGACCTCAGGAAGGCCTGTACAATGAACTGCAGA
		AAGATAAGATGGCGGAGGCCTACAGTGAGATTGGGATGAAA
		GGCGAGCGCCGGAGGGCCAAGGGGCACGATGGCCTTTACCA
		GGGTCTCAGTACAGCCACCAAGGACACCTACGACGCCCTTCA
		CATGCAGGCCCTGCCCCTCGCGGAGTGCAGGTGGAAACCAT
		CTCCCCAGGAGACGGCGCACCTTCCCCAAGCGCGGCCAGAC
		CTGCGTGGTGCACTACACCGGGATGCTTGGAGATGGAAAGAA
		AGTTGACTCCTCCCGGGACAGAAACAAGCCCTTTAAGTTTAT
		GCTAGGCAAGCAGGAGGTGATCCGAGGCTGGGAAGAAGGGG
		TTGCCCAGATGAGTGTGGGTCAGGGAGCCAAACTGACTATAT
		CTCCAGATTATGCCTATGGTGCCACTGGGCACCCAGGCATCA
		TCCCACCACATGCCACTCTCGTCTTCGATGTGGAGCTTCTAGA
120	H. 2.20 EV/DD	ACTGGAATGA
120	Her2.28z.FKBP	MLLLVTSLLLCELPHPAFLLIPDIQMTQSPSSLSASVGDRVTITCR
	Amino acid sequence	ASQDVNTAVAWYQQKPGKAPKLLIYSASFLYSGVPSRFSGSRSG
		TDFTLTISSLQPEDFATYYCQQHYTTPPTFGQGTKVEIKGSTSGS GKPGSGEGSGEVQLVESGGGLVQPGGSLRLSCAASGFNIKDTYI
		HWVRQAPGKGLEWVARIYPTNGYTRYADSVKGRFTISADTSKN
		TAYLQMNSLRAEDTAVYYCSRWGGDGFYAMDVWGQGTLVTV
		SSASEQKLISEEDLFEIEVMYPPPYLDNEKSNGTIIHVKGKHLCPS
		PLFPGPSKPFWVLVVVGGVLACYSLLVTVAFIIFWVRSKRSRLLH
		SDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRSRVKFSRSADAP
		AYKQGQNQLYNELNLGRREEYDVLDKRRGRDPEMGGKPRRKN
		PQEGLYNELQKDKMAEAYSEIGMKGERRRGKGHDGLYQGLST
		ATKDTYDALHMQALPPRGVQVETISPGDGRTFPKRGQTCVVHY
		TGMLGDGKKVDSSRDRNKPFKFMLGKQEVIRGWEEGVAQMSV
12:	T	GQGAKLTISPDYAYGATGHPGIIPPHATLVFDVELLELE
121	Her2.28z.FKBP	ATGCTGCTGCTGACATCTCTGCTGCTGCTGCGAGCTGCCCC
	Leader sequence	ACCCCGCCTTTCTGCTGATCCCC
	nucleic acid sequence	
122	Her2.28z.FKBP	MLLLVTSLLLCELPHPAFLLIP
	Leader sequence	
	Amino acid sequence	

FIG. 25 (cont'd)

<u>10. 23 (</u>		,
123	4D5 scFv (Her2 binding	CAGATGACCCAGTCCCCGAGCTCCCTGTCCGCCTCTGTGGGC
	domain)	GATAGGGTCACCATCACCTGCCGTGCCAGTCAGGATGTGAAT
	nucleic acid sequence	ACTGCTGTAGCCTGGTATCAACAGAAACCAGGAAAAGCTCCG
	_	AAACTACTGATTTACTCGGCATCCTTCCTTTATTCTGGAGTCC
		CTTCTCGCTTCTCTGGATCTAGATCTGGGACGGATTTCACTCT
		GACCATCAGCAGTCTGCAGCCGGAAGACTTCGCAACTTATTA
		CTGTCAGCAACATTATACTACTCCTCCCACGTTCGGACAGGGT
		ACCAAGGTGGAGATCAAAGGGTCTACATCTGGATCTGGGAAG
		CCGGGTTCTGGTGAGGTTCTGGTGAGGTTCAGCTGGTGAG
		TCTGGCGGTGGCCTGGTGCAGCCAGGGGGCTCACTCCGTTTG
		TCCTGTGCAGCTTCTGGCTTCAACATTAAAGACACCTATATAC
		ACTGGGTGCGTCAGGCCCCGGGTAAGGGCCTGGAATGGGTTG
		CAAGGATTTATCCTACGAATGGTTATACTAGATATGCCGATA
		GCGTCAAGGGCCGTTTCACTATAAGCGCAGACACATCCAAAA
		ACACAGCCTACCTGCAGATGAACAGCCTGCGTGCTGAGGACA
		CTGCCGTCTATTATTGTTCTAGATGGGGAGGGGACGGCTTCTA
		TGCTATGGACGTGTGGGGTCAAGGAACCCTGGTCACCGTCTC
		CTCGGCTAGC
124	ADS as For (III a 2 hi a 11	
124	4D5 scFv (Her2 binding	QMTQSPSSLSASVGDRVTITCRASQDVNTAVAWYQQKPGKAPK
	domain)	LLIYSASFLYSGVPSRFSGSRSGTDFTLTISSLQPEDFATYYCQQH
	Amino acid sequence	YTTPPTFGQGTKVEIKGSTSGSGKPGSGEGSGEVQLVESGGGLV
		QPGGSLRLSCAASGFNIKDTYIHWVRQAPGKGLEWVARIYPTNG
		YTRYADSVKGRFTISADTSKNTAYLQMNSLRAEDTAVYYCSRW
		GGDGFYAMDVWGQGTLVTVSSAS
125	MYC tag	GAACAAAACTCATCTCAGAAGAGGATCTG
	nucleic acid sequence	
126	MYC tag	EQKLISEEDL
	Amino acid sequence	_
127	CD28 hinge region	ATTGAAGTTATGTATCCTCCTCCTTACCTAGACAATGAGAAG
127	nucleic acid sequence	AGCAATGGAACCATTATCCATGTGAAAGGGAAACACCTTTGT
	nucleic acid sequence	
120	CD2011	CCAAGTCCCCTATTTCCCGGACCTTCTAAGCCC
128	CD28 hinge region	IEVMYPPPYLDNEKSNGTIIHVKGKHLCPSPLFPGPSKP
	Amino acid sequence	
129	CD28 transmembrane	TTTTGGGTGCTGGTGGTGGTTGGGGGAGTCCTGGCTTGCTATA
	region	GCTTGCTAGTAACAGTGGCCTTTATTATTTTCTGGGTG
	nucleic acid sequence	
130	CD28 transmembrane	FWVLVVVGGVLACYSLLVTVAFIIFWV
	region	
	Amino acid sequence	
131	CD28 intracellular	AGGAGTAAGAGGAGCAGGCTCCTGCACAGTGACTACATGAA
	domain	CATGACTCCCGCCGCCCCGGGCCCACCCGCAAGCATTACCA
	nucleic acid sequence	GCCCTATGCCCCACCACGCGACTTCGCAGCCTATCGCTCC
132	CD28 intracellular	RSKRSRLLHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRS
132		
	1	RSRRSRELISD I WINWITI RIG OF TRAITING TAFFRED FACTORS
	domain	RSRASRELISD I WINWITT RRI OF TRRITTQI TATTRDI AATRO
122	domain Amino acid sequence	·
133	domain Amino acid sequence CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG
133	domain Amino acid sequence CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGAACCCTCAG
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
133	domain Amino acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
133	domain Amino acid sequence CD3 zeta intracellular domain nucleic acid sequence	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
	domain Amino acid sequence CD3 zeta intracellular domain nucleic acid sequence CD3 zeta intracellular	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
	domain Amino acid sequence CD3 zeta intracellular domain nucleic acid sequence CD3 zeta intracellular domain	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
134	domain Amino acid sequence CD3 zeta intracellular domain nucleic acid sequence CD3 zeta intracellular domain Amino acid sequence	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
	domain Amino acid sequence CD3 zeta intracellular domain nucleic acid sequence CD3 zeta intracellular domain Amino acid sequence FKBP12 destabilization	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGAACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT
134	domain Amino acid sequence CD3 zeta intracellular domain nucleic acid sequence CD3 zeta intracellular domain Amino acid sequence	AGAGTGAAGTTCAGCAGGAGCGCAGACGCCCCCGCGTACAA GCAGGGCCAGAACCAGCTCTATAACGAGCTCAATCTAGGACG AAGAGAGAGTACGATGTTTTGGACAAGAGACGTGGCCGGG ACCCTGAGATGGGGGGAAAGCCGAGAAGGAAGACCCTCAG GAAGGCCTGTACAATGAACTGCAGAAAGATAAGAT

	(
	nucleic acid sequence	AAACAAGCCCTTTAAGTTTATGCTAGGCAAGCAGGAGGTGAT
		CCGAGGCTGGGAAGAAGGGGTTGCCCAGATGAGTGTGGGTC
		AGGGAGCCAAACTGACTATATCTCCAGATTATGCCTATGGTG
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Val Ala Pro Ser Gln Ser Leu Ser Val Thr Cys Thr Val Ser Gly Val 165 170 175

Ser Leu Pro Asp Tyr Gly Val Ser Trp Ile Arg Gln Pro Pro Arg Lys 180 185 190

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Asn Ser Ala Leu Lys Ser Arg Leu Thr Ile Ile Lys Asp Asn Ser Lys Page 7

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Gln Val Glu Thr Ile Ser Pro Gly Asp Gly Arg Thr Phe Pro Lys Arg

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Ser Gly Ser Gly Thr Asp Tyr Ser Leu Thr Ile Ser Asn Leu Glu Gln 65 70 75 80

Glu Asp Ile Ala Thr Tyr Phe Cys Gln Gln Gly Asn Thr Leu Pro Tyr 85 90 95

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Ser Ser Leu Ser Ala Ser Leu Gly Asp Arg Val Thr Ile Ser Cys Arg 35 40 45

Ala Ser Gln Asp Ile Ser Lys Tyr Leu Asn Trp Tyr Gln Gln Lys Pro 50 55 60

Asp Gly Thr Val Lys Leu Leu Ile Tyr His Thr Ser Arg Leu His Ser 65 70 75 80

Gly	Val	Pro	Ser	Arg 85	Phe	Ser	Gly	Ser	Gly 90	Ser	Gly	Thr	Asp	Tyr 95	Ser
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Gln	Gln	Gly 115	Asn	Thr	Leu	Pro	Tyr 120	Thr	Phe	Gly	Gly	Gly 125	Thr	Lys	Leu
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Gly 145	Ser	Thr	Lys	Gly	Glu 150	Val	Lys	Leu	Gln	Glu 155	Ser	Gly	Pro	Gly	Leu 160
Val	Ala	Pro	Ser	Gln 165	Ser	Leu	Ser	Val	Thr 170	Cys	Thr	Val	Ser	Gly 175	Val
Ser	Leu	Pro	Asp 180	Tyr	Gly	Val	Ser	Trp 185	Ile	Arg	Gln	Pro	Pro 190	Arg	Lys
Gly	Leu	Glu 195	Trp	Leu	Gly	Val	Ile 200	Trp	Gly	Ser	Glu	Thr 205	Thr	Tyr	Tyr
Asn	Ser 210	Ala	Leu	Lys	Ser	Arg 215	Leu	Thr	Ile	Ile	Lys 220	Asp	Asn	Ser	Lys
Ser 225	Gln	Val	Phe	Leu	Lys 230	Met	Asn	Ser	Leu	Gln 235	Thr	Asp	Asp	Thr	Ala 240
Ile	Tyr	Tyr	Cys	Ala 245	Lys	His	Tyr	Tyr	Tyr 250	Gly	Gly	Ser	Tyr	Ala 255	Met
Asp	Tyr	Trp	Gly 260	Gln	Gly	Thr	Ser	Val 265	Thr	Val	Ser	Ser	Ala 270	Ser	Phe
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Tyr A	Ala 370	Pro	Pro	Arg	Asp	Phe 375	Ala	Ala	Tyr	Arg	Ser 380	His	Met	Arg	Val
Lys P 385	Phe	Ser	Arg	Ser	Ala 390	Asp	Ala	Pro	Ala	Tyr 395	Lys	Gln	Gly	Gln	Asn 400
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Leu A	Asp	Lys	Arg 420	Arg	Gly	Arg	Asp	Pro 425	Glu	Met	Gly	Gly	Lys 430	Pro	Arg
Arg L	_ys	Asn 435	Pro	Gln	Glu	Gly	Leu 440	Tyr	Asn	Glu	Leu	Gln 445	Lys	Asp	Lys
Met A	\la 150	Glu	Ala	Tyr	Ser	Glu 455	Ile	Gly	Met	Lys	Gly 460	Glu	Arg	Arg	Arg
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Arg 545	Pro	Leu	Pro	Gly	Arg 550	Lys	Asn	Ile	Ile	Leu 555	Ser	Ser	Gln	Pro	Ser 560
Thr	Asp	Asp	Arg	Val 565	Thr	Trp	Val	Lys	Ser 570	Val	Asp	Glu	Ala	Ile 575	Ala
Ala	Cys	Gly	Asp 580	Val	Pro	Glu	Ile	Met 585	Val	Ile	Gly	Gly	Gly 590	Arg	Val
Ile	Glu	Gln 595	Phe	Leu	Pro	Lys	Ala 600	Gln	Lys	Leu	Tyr	Leu 605	Thr	His	Ile
Asp	Ala 610	Glu	Val	Glu	Gly	Asp 615	Thr	His	Phe	Pro	Asp 620	Tyr	Glu	Pro	Asp
Asp 625	Trp	Glu	Ser	Val	Phe 630	Ser	Glu	Phe	His	Asp 635	Ala	Asp	Ala	Gln	Asn 640
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120 180 aggttcagtg gcagtgggtc tggaacagat tattctctca ccattagcaa cctggagcaa 240 gaagatattg ccacttactt ttgccaacag ggtaatacgc ttccgtacac gttcggaggg 300 360 gggactaagt tggaaataac aggctccacc tctggatccg gcaagcccgg atctggcgag ggatccacca agggcgaggt gaaactgcag gagtcaggac ctggcctggt ggcgccctca 420 480 cagagectgt cegteacatg tactgtetea ggggteteat taccegaeta tggtgtaage 540 tggattcgcc agcctccacg aaagggtctg gagtggctgg gagtaatatg gggtagtgaa accacatact ataattcagc tctcaaatcc agactgacca tcatcaagga caactccaag 600

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Asp Arg Val Thr Ile Ser Cys Arg Ala Ser Gln Asp Ile Ser Lys Tyr 20 25 30

Leu Asn Trp Tyr Gln Gln Lys Pro Asp Gly Thr Val Lys Leu Leu Ile 35 40 45

Tyr His Thr Ser Arg Leu His Ser Gly Val Pro Ser Arg Phe Ser Gly 50 55 60

Ser Gly Ser Gly Thr Asp Tyr Ser Leu Thr Ile Ser Asn Leu Glu Gln 65 70 75 80

Glu Asp Ile Ala Thr Tyr Phe Cys Gln Gln Gly Asn Thr Leu Pro Tyr 85 90 95

Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Thr Gly Ser Thr Ser Gly 100 105 110

Ser Gly Lys Pro Gly Ser Gly Glu Gly Ser Thr Lys Gly Glu Val Lys 115 120 125

Leu Gln Glu Ser Gly Pro Gly Leu Val Ala Pro Ser Gln Ser Leu Ser 130 135 140

Val Thr Cys Thr Val Ser Gly Val Ser Leu Pro Asp Tyr Gly Val Ser Page 23 Trp Gly Ser Glu Thr Thr Tyr Tyr Asn Ser Ala Leu Lys Ser Arg Leu

185

170

175

60

165

180

Thr Ile Ile Lys Asp Asn Ser Lys Ser Gln Val Phe Leu Lys Met Asn 195 200 205

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Leu Val Thr Val Ala Phe Ile Ile Phe Trp Val
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Pro Arg Arg Pro Gly Pro Thr Arg Lys His Tyr Gln Pro Tyr Ala Pro

Pro Arg Asp Phe Ala Ala Tyr Arg Ser 35

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Gln As	n Gln	Leu 20	Tyr	Asn	Glu	Leu	Asn 25	Leu	Gly	Arg	Arg	Glu 30	Glu	Tyr	
Asp Va	1 Leu 35	Asp	Lys	Arg	Arg	Gly 40	Arg	Asp	Pro	Glu	Met 45	Gly	Gly	Lys	
Pro Ar 50		Lys	Asn	Pro	Gln 55	Glu	Gly	Leu	Tyr	Asn 60	Glu	Leu	Gln	Lys	
Asp Ly 65	s Met	Ala	Glu	Ala 70	Tyr	Ser	Glu	Ile	Gly 75	Met	Lys	Gly	Glu	Arg 80	
Arg Ar	g Gly	Lys	Gly 85	His	Asp	Gly	Leu	Tyr 90	Gln	Gly	Leu	Ser	Thr 95	Ala	
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gacacccatt	tcccggatta	cgagccggat	gactgggaat	cggtattcag	cgaattccac	420
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Asn Thr Leu Asn Lys Pro Val Ile Met Gly Arg His Thr Trp Glu Ser 35 40 45

Ile Gly Arg Pro Leu Pro Gly Arg Lys Asn Ile Ile Leu Ser Ser Gln 50 55 60

Pro Ser Thr Asp Asp Arg Val Thr Trp Val Lys Ser Val Asp Glu Ala 65 70 75 80

Ile Ala Ala Cys Gly Asp Val Pro Glu Ile Met Val Ile Gly Gly 85 90 95

Arg Val Ile Glu Gln Phe Leu Pro Lys Ala Gln Lys Leu Tyr Leu Thr 100 105 110

His Ile Asp Ala Glu Val Glu Gly Asp Thr His Phe Pro Asp Tyr Glu 115 120 125

Pro Asp Asp Trp Glu Ser Val Phe Ser Glu Phe His Asp Ala Asp Ala 130 135 140

Gln Asn Ser His Ser Tyr Cys Phe Glu Ile Leu Glu Arg Arg 145 150 155

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<211> 1791

<212> DNA

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<212> PRT

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<223> synthetic

<400> 40

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Leu Pro Val Ser Leu Gly Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser 35 40 45

Gln Ser Leu Val His Arg Asn Gly Asn Thr Tyr Leu His Trp Tyr Leu 50 55 60

Gln Lys Pro 65	Gly Gln	Ser Pro) Lys	Leu	Leu	Ile 75	His	Lys	Val	Ser	Asn 80
Arg Phe Ser	Gly Val 85	Pro Asp) Arg	Phe	Ser 90	Gly	Ser	Gly	Ser	Gly 95	Thr
Asp Phe Thr	Leu Lys 100	Ile Ser	` Arg	Val 105	Glu	Ala	Glu	Asp	Leu 110	Gly	Val
Tyr Phe Cys 115	Ser Gln	Ser Thi	His 120	Val	Pro	Pro	Leu	Thr 125	Phe	Gly	Ala
Gly Thr Lys 130	Leu Glu	Leu Lys	-	Ser	Thr	Ser	Gly 140	Ser	Gly	Lys	Pro
Gly Ser Gly 145	Glu Gly	Ser Thi	` Lys	Gly	Glu	Val 155	Lys	Leu	Gln	Gln	Ser 160
Gly Pro Ser	Leu Val 165	Glu Pro	Gly	Ala	Ser 170	Val	Met	Ile	Ser	Cys 175	Lys
Ala Ser Gly	Ser Ser 180	Phe Thi	Gly	Tyr 185	Asn	Met	Asn	Trp	Val 190	Arg	Gln
Asn Ile Gly 195	Lys Ser	Leu Glu	ı Trp 200	Ile	Gly	Ala	Ile	Asp 205	Pro	Tyr	Tyr
Gly Gly Thr 210	Ser Tyr	Asn Glr 21	-	Phe	Lys	Gly	Arg 220	Ala	Thr	Leu	Thr
Val Asp Lys 225	Ser Ser	Ser Thr 230	` Ala	Tyr	Met	His 235	Leu	Lys	Ser	Leu	Thr 240
Ser Glu Asp	Ser Ala 245	Val Tyr	Tyr	Cys	Val 250	Ser	Gly	Met	Glu	Tyr 255	Trp
Gly Gln Gly	Thr Ser 260	Val Thr	`Val	Ser 265	Ser	Ala	Ser	Phe	Glu 270	Ile	Glu

Val	Met	Tyr 275	Pro	Pro	Pro	Tyr	Leu 280	Asp	Asn	Glu	Lys	Ser 285	Asn	Gly	Thr
Ile	Ile 290	His	Val	Lys	Gly	Lys 295	His	Leu	Cys	Pro	Ser 300	Pro	Leu	Phe	Pro
Gly 305	Pro	Ser	Lys	Pro	Phe 310	Trp	Val	Leu	Val	Val 315	Val	Gly	Gly	Val	Leu 320
Ala	Cys	Tyr	Ser	Leu 325	Leu	Val	Thr	Val	Ala 330	Phe	Ile	Ile	Phe	Trp 335	Val
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				405	Arg				410					415	
			420		Met			425					430		
	·	435			Glu		440					445			-
	450				Lys	455					460				•
G1y 465	Leu	ıyr	GIN	σ1У	Leu 470	ser	ınr	ΑТА	ınr	Lys 475	ASP	ınr	ıyr	ASP	A1a 480

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Ser Pr	o Gly	Asp 500	Gly	Arg	Thr	Phe	Pro 505	Lys	Arg	Gly	Gln	Thr 510	Cys	Val	
Val Hi	s Tyr 515		Gly	Met	Leu	Gly 520	Asp	Gly	Lys	Lys	Val 525	Asp	Ser	Ser	
Arg As 53	_	Asn	Lys	Pro	Phe 535	Lys	Phe	Met	Leu	Gly 540	Lys	Gln	Glu	Val	
Ile Ar 545	g Gly	Trp	Glu	Glu 550	Gly	Val	Ala	Gln	Met 555	Ser	Val	Gly	Gln	Gly 560	
Ala Ly	s Leu	Thr	Ile 565	Ser	Pro	Asp	Tyr	Ala 570	Tyr	Gly	Ala	Thr	Gly 575	His	
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Ala Phe Leu Leu Ile Pro 20

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Pro Lys Leu Leu Ile His Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 55 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile
70 75 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95

Thr His Val Pro Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu 100 105 110

Lys Gly Ser Thr Ser Gly Ser Gly Lys Pro Gly Ser Gly Glu Gly Ser 115 120 125

Thr Lys Gly Glu Val Lys Leu Gln Gln Ser Gly Pro Ser Leu Val Glu 130 135 140

Pro Gly Ala Ser Val Met Ile Ser Cys Lys Ala Ser Gly Ser Ser Phe 145 150 155 160

Thr Gly Tyr Asn Met Asn Trp Val Arg Gln Asn Ile Gly Lys Ser Leu 165 170 175

Glu Trp Ile Gly Ala Ile Asp Pro Tyr Tyr Gly Gly Thr Ser Tyr Asn Page 35

Gln Lys Phe Lys Gly Arg Ala Thr Leu Thr Val Asp Lys Ser Ser Ser 195 200 Thr Ala Tyr Met His Leu Lys Ser Leu Thr Ser Glu Asp Ser Ala Val 215 220 Tyr Tyr Cys Val Ser Gly Met Glu Tyr Trp Gly Gln Gly Thr Ser Val 230 235 Thr Val Ser Ser <210> 45 <211> 117 <212> DNA <213> Artificial Sequence <220> <223> synthetic <400> 45 attgaagtta tgtatcctcc tccttaccta gacaatgaga agagcaatgg aaccattatc 60 catgtgaaag ggaaacacct ttgtccaagt cccctatttc ccggaccttc taagccc 117 <210> 46 <211> 39 <212> PRT <213> Artificial Sequence <220> <223> synthetic <400> 46 Ile Glu Val Met Tyr Pro Pro Pro Tyr Leu Asp Asn Glu Lys Ser Asn 5 15 1 10

Phe Pro Gly Pro Ser Lys Pro

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Pro Arg Asp Phe Ala Ala Tyr Arg Ser 35 40

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Asp	Val	Leu 35	Asp	Lys	Arg	Arg	Gly 40	Arg	Asp	Pro	Glu	Met 45	Gly	Gly	Lys		
Pro	Arg 50	Arg	Lys	Asn	Pro	G1n 55	Glu	Gly	Leu	Tyr	Asn 60	Glu	Leu	Gln	Lys		
Asp 65	Lys	Met	Ala	Glu	Ala 70	Tyr	Ser	Glu	Ile	Gly 75	Met	Lys	Gly	Glu	Arg 80		
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gaca	agaaa	aca a	agcco	ettta	aa gt	tttat	gcta	a ggo	caago	agg	aggt	gato	cg a	aggct	gggaa	;	180
gaag	ggggt	ttg	ccag	gatga	ag tg	gtggg	gtcag	g gga	agcca	aaac	tgad	tata	atc 1	ccag	gattat		240
gcct	atgg	gtg (ccact	ggg	ca co	cagg	gcato	ato	cca	cac	atgo	cact	ct o	gtct	tcgat		300
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Gly Ly	s Lys 35	Val	Asp	Ser	Ser	Arg 40	Asp	Arg	Asn	Lys	Pro 45	Phe	Lys	Phe	
Met Le 50	-	Lys	Gln	Glu	Val 55	Ile	Arg	Gly	Trp	Glu 60	Glu	Gly	Val	Ala	
Gln Me 65	t Ser	Val	Gly	Gln 70	Gly	Ala	Lys	Leu	Thr 75	Ile	Ser	Pro	Asp	Tyr 80	
Ala Ty	r Gly	Ala	Thr 85	Gly	His	Pro	Gly	Ile 90	Ile	Pro	Pro	His	Ala 95	Thr	
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180

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Gln Ser Leu Val His Arg Asn Gly Asn Thr Tyr Leu His Trp Tyr Leu 50 55 60

Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile His Lys Val Ser Asn 65 70 75 80

Arg Phe Ser Gly Val Pro Asp Arg Phe Ser Gly Ser Gly Thr 85 90 95

Asp Phe Thr Leu Lys Ile Ser Arg Val Glu Ala Glu Asp Leu Gly Val 100 105 110

Tyr Phe Cys Ser Gln Ser Thr His Val Pro Pro Leu Thr Phe Gly Ala 115 120 125

Gly Thr Lys Leu Glu Leu Lys Gly Ser Thr Ser Gly Ser Gly Lys Pro Page 42 135

Gly 145	Ser	Gly	Glu	Gly	Ser 150	Thr	Lys	Gly	Glu	Val 155	Lys	Leu	Gln	Gln	Ser 160
Gly	Pro	Ser	Leu	Val 165	Glu	Pro	Gly	Ala	Ser 170	Val	Met	Ile	Ser	Cys 175	Lys
Ala	Ser	Gly	Ser 180	Ser	Phe	Thr	Gly	Tyr 185	Asn	Met	Asn	Trp	Val 190	Arg	Gln
Asn	Ile	Gly 195	Lys	Ser	Leu	Glu	Trp 200	Ile	Gly	Ala	Ile	Asp 205	Pro	Tyr	Tyr
Gly	Gly 210	Thr	Ser	Tyr	Asn	Gln 215	Lys	Phe	Lys	Gly	Arg 220	Ala	Thr	Leu	Thr
Val 225	Asp	Lys	Ser	Ser	Ser 230	Thr	Ala	Tyr	Met	His 235	Leu	Lys	Ser	Leu	Thr 240
Ser	Glu	Asp	Ser	Ala 245	Val	Tyr	Tyr	Cys	Val 250	Ser	Gly	Met	Glu	Tyr 255	Trp
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Val	Met	Tyr 275		Pro	Pro		Leu 280		Asn	Glu	Lys	Ser 285		Gly	Thr
Ile	Ile 290	His	Val	Lys	Gly	Lys 295	His	Leu	Cys	Pro	Ser 300	Pro	Leu	Phe	Pro
Gly 305	Pro	Ser	Lys	Pro	Phe 310	Trp	Val	Leu	Val	Val 315	Val	Gly	Gly	Val	Leu 320
Ala	Cys	Tyr	Ser	Leu 325	Leu	Val	Thr	Val	Ala 330	Phe	Ile	Ile	Phe	Trp 335	Val
Arg	Ser	Lys	Arg	Ser	Arg	Leu	Leu	His	Ser	Asp	Tyr	Met	Asn	Met	Thr

Page 43

35233-W0-10RD_ST25.txt 345 350

Pro Arg Arg Pro	Gly Pro Thr	Arg Lys His	Tyr Gln Pro	Tyr Ala Pro
355		360	365	

340

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Leu Asn Leu Gly Arg Arg Glu Glu Tyr Asp Val Leu Asp Lys Arg Arg 405 410 415

Gly Arg Asp Pro Glu Met Gly Gly Lys Pro Arg Arg Lys Asn Pro Gln
420 425 430

Glu Gly Leu Tyr Asn Glu Leu Gln Lys Asp Lys Met Ala Glu Ala Tyr 435 440 445

Ser Glu Ile Gly Met Lys Gly Glu Arg Arg Gly Lys Gly His Asp 450 455 460

Gly Leu Tyr Gln Gly Leu Ser Thr Ala Thr Lys Asp Thr Tyr Asp Ala 465 470 475 480

Leu His Met Gln Ala Leu Pro Pro Arg Ile Ser Leu Ile Ala Ala Leu 485 490 495

Ala Val Asp Tyr Val Ile Gly Met Glu Asn Ala Met Pro Trp Asn Leu 500 505 510

Pro Ala Asp Leu Ala Trp Phe Lys Arg Asn Thr Leu Asn Lys Pro Val 515 520 525

Ile Met Gly Arg His Thr Trp Glu Ser Ile Gly Arg Pro Leu Pro Gly 530 540

Arg Lys Asn Ile Ile Leu Ser Ser Gln Pro Ser Thr Asp Asp Arg Val Page 44

545 550 560 Thr Trp Val Lys Ser Val Asp Glu Ala Ile Ala Ala Cys Gly Asp Val 565 570 575 Pro Glu Ile Met Val Ile Gly Gly Gly Arg Val Ile Glu Gln Phe Leu 580 585 Pro Lys Ala Gln Lys Leu Tyr Leu Thr His Ile Asp Ala Glu Val Glu 595 600 605 Gly Asp Thr His Phe Pro Asp Tyr Glu Pro Asp Asp Trp Glu Ser Val 610 615 620 Phe Ser Glu Phe His Asp Ala Asp Ala Gln Asn Ser His Ser Tyr Cys 625 630 635 640 Phe Glu Ile Leu Glu Arg Arg 645 <210> 57 <211> 66 <212> DNA <213> Artificial Sequence <220> <223> synthetic <400> 57 atgctgctgc tcgtgacatc tctgctgctg tgcgagctgc cccaccccgc ctttctgctg 60 atcccc 66 <210> 58 <211> 22 <212> PRT <213> Artificial Sequence <220> <223> synthetic <400> 58 Met Leu Leu Val Thr Ser Leu Leu Cys Glu Leu Pro His Pro

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Asn	Gly	Asn 35	Thr	Tyr	Leu	His	Trp 40	Tyr	Leu	Gln	Lys	Pro 45	Gly	Gln	Ser
Pro	Lys 50	Leu	Leu	Ile	His	Lys 55	Val	Ser	Asn	Arg	Phe 60	Ser	Gly	Val	Pro
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Ser	Arg	Val	Glu	Ala 85	Glu	Asp	Leu	Gly	Val 90	Tyr	Phe	Cys	Ser	G1n 95	Ser
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Pro 145	Gly	Ala	Ser	Val	Met 150	Ile	Ser	Cys	Lys	Ala 155	Ser	Gly	Ser	Ser	Phe 160
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Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg Arg Glu Glu Tyr Page 50

20

Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu Met Gly Gly Lys 35 40 45

Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu Leu Gln Lys 50 55 60

Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys Gly Glu Arg 65 70 75 80

Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu Ser Thr Ala 85 90 95

Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala Leu Pro Pro Arg 100 105 110

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Asn Thr Leu Asn Lys Pro Val Ile Met Gly Arg His Thr Trp Glu Ser 35 40 45

Ile Gly Arg Pro Leu Pro Gly Arg Lys Asn Ile Ile Leu Ser Ser Gln 50 55 60

Pro Ser Thr Asp Asp Arg Val Thr Trp Val Lys Ser Val Asp Glu Ala 65 70 75 80

Ile Ala Ala Cys Gly Asp Val Pro Glu Ile Met Val Ile Gly Gly 85 90 95

Arg Val Ile Glu Gln Phe Leu Pro Lys Ala Gln Lys Leu Tyr Leu Thr 100 105 110

His Ile Asp Ala Glu Val Glu Gly Asp Thr His Phe Pro Asp Tyr Glu 115 120 125

Pro Asp Asp Trp Glu Ser Val Phe Ser Glu Phe His Asp Ala Asp Ala 130 135 140

Gln Asn Ser His Ser Tyr Cys Phe Glu Ile Leu Glu Arg Arg 145 150 155

<210> 71

<211> 1950

<212> DNA

<213> Artificial Sequence

<220>

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1440

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<211> 649

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<400> 72

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Leu Pro Val Ser Leu Gly Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser 35 40 45

Gln Ser Leu Val His Arg Asn Gly Asn Thr Tyr Leu His Trp Tyr Leu 50 55 60

Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile His Lys Val Ser Asn 70 75 80

Arg Phe Ser Gly Val Pro Asp Arg Phe Ser Gly Ser Gly Thr 85 90 95

Asp Phe Ti	nr Leu L 100	ys Ile	Ser	Arg	Val 105	Glu	Ala	Glu	Asp	Leu 110	Gly	Val
Tyr Phe C	/s Ser G L5	Gln Ser	Thr	His 120	Val	Pro	Pro	Leu	Thr 125	Phe	Gly	Ala
Gly Thr Ly 130	/s Leu G	Glu Leu	Lys 135	Gly	Ser	Thr	Ser	Gly 140	Ser	Gly	Lys	Pro
Gly Ser G	ly Glu G	Gly Ser 150	Thr	Lys	Gly	Glu	Val 155	Lys	Leu	Gln	Gln	Ser 160
Gly Pro S		/al Glu 165	Pro	Gly	Ala	Ser 170	Val	Met	Ile	Ser	Cys 175	Lys
Ala Ser G	ly Ser S 180	Ser Phe	Thr	Gly	Tyr 185	Asn	Met	Asn	Trp	Val 190	Arg	Gln
Asn Ile G	ly Lys S 95	Ser Leu	Glu	Trp 200	Ile	Gly	Ala	Ile	Asp 205	Pro	Tyr	Tyr
Gly Gly TI 210	nr Ser T	Гуr Asn	Gln 215	Lys	Phe	Lys	Gly	Arg 220	Ala	Thr	Leu	Thr
Val Asp Ly 225	s Ser S	Ser Ser 230	Thr	Ala	Tyr	Met	His 235	Leu	Lys	Ser	Leu	Thr 240
Ser Glu A	•	Ala Val 245	Tyr	Tyr	Cys	Val 250	Ser	Gly	Met	Glu	Tyr 255	Trp
Gly Gln G	ly Thr S 260	Ser Val	Thr	Val	Ser 265	Ser	Ala	Ser	Phe	Glu 270	Ile	Glu
Val Met Ty 2	r Pro P 75	Pro Pro	Tyr	Leu 280	Asp	Asn	Glu	Lys	Ser 285	Asn	Gly	Thr
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Ala	Cys	Tyr	Ser	Leu 325	Leu	Val	Thr	Val	Ala 330	Phe	Ile	Ile	Phe	Trp 335	Val
Arg	Ser	Lys	Arg 340	Ser	Arg	Leu	Leu	His 345	Ser	Asp	Tyr	Met	Asn 350	Met	Thr
Pro	Arg	Arg 355	Pro	Gly	Pro	Thr	Arg 360	Lys	His	Tyr	Gln	Pro 365	Tyr	Ala	Pro
Pro	Arg 370	Asp	Phe	Ala	Ala	Tyr 375	Arg	Ser	His	Met	Arg 380	Val	Lys	Phe	Ser
Arg 385	Ser	Ala	Asp	Ala	Pro 390	Ala	Tyr	Lys	Gln	Gly 395	Gln	Asn	Gln	Leu	Tyr 400
Asn	Glu	Leu	Asn	Leu 405	Gly	Arg	Arg	Glu	Glu 410	Tyr	Asp	Val	Leu	Asp 415	Lys
Arg	Arg	Gly	Arg 420	Asp	Pro	Glu	Met	Gly 425	Gly	Lys	Pro	Arg	Arg 430	Lys	Asn
Pro	Gln	Glu 435	Gly	Leu	Tyr	Asn	Glu 440	Leu	Gln	Lys	Asp	Lys 445	Met	Ala	Glu
Ala	Tyr 450	Ser	Glu	Ile	Gly	Met 455	Lys	Gly	Glu	Arg	Arg 460	Arg	Gly	Lys	Gly
His 465	Asp	Gly	Leu	Tyr	Gln 470	Gly	Leu	Ser	Thr	Ala 475	Thr	Lys	Asp	Thr	Tyr 480
Asp	Ala	Leu	His	Met 485	Gln	Ala	Leu	Pro	Pro 490	Arg	Ile	Ser	Leu	Ile 495	Ala
Ala	Leu	Ala	Val 500	Asp	Tyr	Val	Ile	Gly 505	Met	Glu	Asn	Ala	Met 510	Pro	Trp

Asn L		Pro 515	Ala	Asp	Leu	Ala	Trp 520	Phe	Lys	Arg	Asn	Thr 525	Leu	Asn	Lys	
Pro V 5	al I 30	Ile	Met	Gly	Arg	His 535	Thr	Trp	Glu	Ser	Ile 540	Gly	Arg	Pro	Leu	
Pro G 545	ly A	۱rg	Lys	Asn	Ile 550	Ile	Leu	Ser	Ser	Gln 555	Pro	Ser	Thr	Asp	Asp 560	
Arg V	al T	ſhr	Trp	Val 565	Lys	Ser	Val	Asp	Glu 570	Ala	Ile	Ala	Ala	Cys 575	Gly	
Asp V	al P	Pro	Glu 580	Ile	Met	Val	Ile	Gly 585	Gly	Gly	Arg	Val	Ile 590	Glu	Gln	
Phe L		Pro 595	Lys	Ala	Gln	Lys	Leu 600	Tyr	Leu	Thr	His	Ile 605	Asp	Ala	Glu	
Val G 6	lu 0 10	Sly	Asp	Thr	His	Phe 615	Pro	Asp	Tyr	Glu	Pro 620	Asp	Asp	Trp	Glu	
Ser V 625	al P	he	Ser	Glu	Phe 630	His	Asp	Ala	Asp	Ala 635	Gln	Asn	Ser	His	Ser 640	
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<211> 244

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<223> synthetic

<400> 76

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Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Leu Val His Arg 20 25 30

Asn Gly Asn Thr Tyr Leu His Trp Tyr Leu Gln Lys Pro Gly Gln Ser 35 40 45

Pro Lys Leu Leu Ile His Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 55 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 70 75 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95

Thr His Val Pro Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu 100 105 110

Lys Gly Ser Thr Ser Gly Ser Gly Lys Pro Gly Ser Gly Glu Gly Ser 115 120 125

Thr Lys Gly Glu Val Lys Leu Gln Gln Ser Gly Pro Ser Leu Val Glu 130 135 140

Pro Gly Ala Ser Val Met Ile Ser Cys Lys Ala Ser Gly Ser Ser Phe 145 150 155 160

Thr Gly Tyr Asn Met Asn Trp Val Arg Gln Asn Ile Gly Lys Ser Leu Page 59

165

Glu Trp Ile Gly Ala Ile Asp Pro Tyr Tyr Gly Gly Thr Ser Tyr Asn 180 185 190 Gln Lys Phe Lys Gly Arg Ala Thr Leu Thr Val Asp Lys Ser Ser Ser 200 Thr Ala Tyr Met His Leu Lys Ser Leu Thr Ser Glu Asp Ser Ala Val 210 215 220 Tyr Tyr Cys Val Ser Gly Met Glu Tyr Trp Gly Gln Gly Thr Ser Val 225 230 235 240 Thr Val Ser Ser <210> 77 <211> 117 <212> DNA <213> Artificial Sequence <220> <223> synthetic <400> 77 attgaagtta tgtatcctcc tccttaccta gacaatgaga agagcaatgg aaccattatc 60 catgtgaaag ggaaacacct ttgtccaagt cccctatttc ccggaccttc taagccc 117 <210> 78 <211> 39 <212> PRT <213> Artificial Sequence <220> <223> synthetic

Gly Thr Ile Ile His Val Lys Gly Lys His Leu Cys Pro Ser Pro Leu Page 60

Ile Glu Val Met Tyr Pro Pro Pro Tyr Leu Asp Asn Glu Lys Ser Asn

10

<400> 78

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123

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Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg Arg Glu Glu Tyr 20 25 30										
Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu Met Gly Gly Lys 35 40 45										
Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu Leu Gln Lys 50 55 60										
Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys Gly Glu Arg 65 70 75 80										
Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu Ser Thr Ala 85 90 95										
Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala Leu Pro Pro Arg 100 105 110										
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300

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<210> 86

<211> 158

<212> PRT

<213> Artificial Sequence

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<223> synthetic

<400> 86

Ile Ser Leu Ile Ala Ala Leu Ala Val Asp Tyr Val Ile Gly Met Glu
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Asn Ala Met Pro Trp Asn Leu Pro Ala Asp Leu Ala Trp Phe Lys Arg 20 25 30

Asn Thr Leu Asn Lys Pro Val Ile Met Gly Arg His Thr Trp Glu Ser 35 40 45

Ile Gly Arg Pro Leu Pro Gly Arg Lys Asn Ile Ile Leu Ser Ser Gln 50 55 60

Pro Ser Thr Asp Asp Arg Val Thr Trp Val Lys Ser Val Asp Glu Ala 65 70 75 80

Ile Ala Ala Cys Gly Asp Val Pro Glu Ile Met Val Ile Gly Gly Gly 85 90 95

Arg Val Ile Glu Gln Phe Leu Pro Lys Ala Gln Lys Leu Tyr Leu Thr 100 105 110

His Ile Asp Ala Glu Val Glu Gly Asp Thr His Phe Pro Asp Tyr Glu 115 120 125

Pro Asp Asp Trp Glu Ser Val Phe Ser Glu Phe His Asp Ala Asp Ala 130 135 140

Gln Asn Ser His Ser Tyr Cys Phe Glu Ile Leu Glu Arg Arg 145 150 155

<210> 87

<211> 2538

<212> DNA

<213> Artificial Sequence

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<223> synthetic

<400> 87

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180	ctacctgcac	acggcaatac	gtgcacagaa	ccagagcctg	gtagaagcag	agcatcagct
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<211> 845 <212> PRT

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<223> synthetic

<400> 88

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Pro Val Ser Leu Gly Asp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln 35 40 45

Ser Leu Val His Arg Asn Gly Asn Thr Tyr Leu His Trp Tyr Leu Gln 50 55 60

Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile His Lys Val Ser Asn Arg 70 75 80

Phe Ser Gly Val Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp 85 90 95

Phe Thr Leu Lys Ile Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr 100 105 110

Phe Cys Ser Gln Ser Thr His Val Pro Pro Leu Thr Phe Gly Ala Gly 115 120 125

Thr Lys Leu Glu Leu Lys Gly Gly Gly Gly Ser Gly Gly Gly Ser 130 135 140

Gly Gly Gly Ser Glu Val Lys Leu Gln Gln Ser Gly Pro Ser Leu 145 150 155 160

Val Glu Pro Gly Ala Ser Val Met Ile Ser Cys Lys Ala Ser Gly Ser 165 170 175

Ser Phe Thr Gly Tyr Asn Met Asn Trp Val Arg Gln Asn Ile Gly Lys Page 67

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Ser Ser Thr 225	Ala Tyr	Met His 230	Leu	Lys	Ser	Leu 235	Thr	Ser	Glu	Asp	Ser 240
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Val Lys Phe	Asn Trp 340	Tyr Val	Asp	Gly 345	Val	Glu	Val	His	Asn 350	Ala	Lys
Thr Lys Pro 355	Arg Glu	Glu Gln	Tyr 360	Asn	Ser	Thr	Tyr	Arg 365	Val	Val	Ser
Val Leu Thr 370	Val Leu	His Gln 375	-	Trp	Leu	Asn	Gly 380	Lys	Glu	Tyr	Lys

Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Page 68

202					390					222					400
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Thr	Val	Ala	Phe 580	Ile	Ile	Phe	Trp	Val 585	Arg	Ser	Lys	Arg	Ser 590	Arg	Leu

Leu His Ser Asp Tyr Met Asn Met Thr Pro Arg Arg Pro Gly Pro Thr

Page 69

Arg Lys His Tyr Gln Pro Tyr Ala Pro Pro Arg Asp Phe Ala Ala Tyr 610 615 620

Arg Ser Arg Val Lys Phe Ser Arg Ser Ala Asp Ala Pro Ala Tyr Lys 625 630 635 640

Gln Gly Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg Arg Glu 645 650 655

Glu Tyr Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu Met Gly 660 665 670

Gly Lys Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu Leu 675 680 685

Gln Lys Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys Gly 690 695 700

Glu Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu Ser 705 710 715 720

Thr Ala Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala Leu Pro
725 730 735

Pro Arg Gly Val Gln Val Glu Thr Ile Ser Pro Gly Asp Gly Arg Thr 740 745 750

Phe Pro Lys Arg Gly Gln Thr Cys Val Val His Tyr Thr Gly Met Leu 755 760 765

Gly Asp Gly Lys Lys Val Asp Ser Ser Arg Asp Arg Asn Lys Pro Phe 770 780

Lys Phe Met Leu Gly Lys Gln Glu Val Ile Arg Gly Trp Glu Glu Gly 785 790 795 800

Val Ala Gln Met Ser Val Gly Gln Gly Ala Lys Leu Thr Ile Ser Pro Page 70

805

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Pro Lys Leu Leu Ile His Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 55 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 70 75 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95

Thr His Val Pro Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu 100 105 110

Lys Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser 115 120 125

Glu Val Lys Leu Gln Gln Ser Gly Pro Ser Leu Val Glu Pro Gly Ala 130 135 140

Ser Val Met Ile Ser Cys Lys Ala Ser Gly Ser Ser Phe Thr Gly Tyr 145 150 155 160

Asn Met Asn Trp Val Arg Gln Asn Ile Gly Lys Ser Leu Glu Trp Ile 165 170 175

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Met	His 210	Leu	Lys	Ser	Leu	Thr 215	Ser	Glu	Asp	Ser	Ala 220	Val	Tyr	Tyr	Cys
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Ser	Ala	Lys	Thr	Thr 245	Pro	Pro	Ser	Val	Tyr 250	Gly	Arg	Val	Thr	Val 255	Ser
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Pro	Ala	Pro 275	Glu	Leu	Leu	Gly	Gly 280	Pro	Ser	Val	Phe	Leu 285	Phe	Pro	Pro
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Pro Se	r Asp	Ile 420	Ala	Val	Glu	Trp	Glu 425	Ser	Asn	Gly	Gln	Pro 430	Glu	Asn		
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Pro Ar 50		Lys	Asn	Pro	G1n 55	Glu	Gly	Leu	Tyr	Asn 60	Glu	Leu	Gln	Lys	
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Met Leu Gly Lys Gln Glu Val Ile Arg Gly Trp Glu Glu Gly Val Ala 50 55 60

Gln Met Ser Val Gly Gln Gly Ala Lys Leu Thr Ile Ser Pro Asp Tyr 65 70 75 80

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Pro Glu Val Thr Cys Val	Val Val Asp	Val Ser His	Glu Asp Pro Glu
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Thr Lys Pro Arg Glu Glu	Gln Tyr Asn	Ser Thr Tyr	Arg Val Val Ser
355	360		365
Val Leu Thr Val Leu His	Gln Asp Trp	Leu Asn Gly	Lys Glu Tyr Lys
370	375	380	
Cys Lys Val Ser Asn Lys		Ala Pro Ile	Glu Lys Thr Ile
385 390		395	400
Ser Lys Ala Lys Gly Gln	Pro Arg Glu	Pro Gln Val	Tyr Thr Leu Pro
405		410	415

Pro	Ser	Arg	Asp 420	Glu	Leu	Thr	Lys	Asn 425	Gln	Val	Ser	Leu	Thr 430	Cys	Leu
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Gly	Gln 450	Pro	Glu	Asn	Asn	Tyr 455	Lys	Thr	Thr	Pro	Pro 460	Val	Leu	Asp	Ser
Asp 465	Gly	Ser	Phe	Phe	Leu 470	Tyr	Ser	Lys	Leu	Thr 475	Val	Asp	Lys	Ser	Arg 480
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Asp	Pro	Lys 515	Ala	Ser	Phe	Glu	Ile 520	Glu	Val	Met	Tyr	Pro 525	Pro	Pro	Tyr
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Thr	Val	Ala	Phe 580	Ile	Ile	Phe	Trp	Val 585	Arg	Ser	Lys	Arg	Ser 590	Arg	Leu
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Arg Ser Arg 625	Val Lys	Phe Sei 630	r Arg	Ser	Ala	Asp 635	Ala	Pro	Ala	Tyr	Lys 640
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Gly Lys Pro 675	Arg Arg	Lys Ası	n Pro 680	Gln	Glu	Gly	Leu	Tyr 685	Asn	Glu	Leu
Gln Lys Asp 690	Lys Met	Ala Gl		Tyr	Ser	Glu	Ile 700	Gly	Met	Lys	Gly
Glu Arg Arg 705	Arg Gly	Lys Gl ₂ 710	/ His	Asp	Gly	Leu 715	Tyr	Gln	Gly	Leu	Ser 720
Thr Ala Thr	Lys Asp 725	Thr Ty	n Asp	Ala	Leu 730	His	Met	Gln	Ala	Leu 735	Pro
Pro Arg Ile	Ser Leu 740	Ile Ala	a Ala	Leu 745	Ala	Val	Asp	Tyr	Val 750	Ile	Gly
Met Glu Asn 755	Ala Met	Pro Tr	760	Leu	Pro	Ala	Asp	Leu 765	Ala	Trp	Phe
Lys Arg Asn 770	Thr Leu	Asn Ly: 77		Val	Ile	Met	Gly 780	Arg	His	Thr	Trp
Glu Ser Ile 785	Gly Arg	Pro Lei 790	ı Pro	Gly	Arg	Lys 795	Asn	Ile	Ile	Leu	Ser 800
Ser Gln Pro	Ser Thr 805	Asp As _l	o Arg	Val	Thr 810	Trp	Val	Lys	Ser	Val 815	Asp
Glu Ala Ile	Ala Ala 820	Cys Gl	/ Asp	Val 825	Pro	Glu	Ile	Met	Val 830	Ile	Gly

Gly Gly Arg Val Ile Glu Gln Phe Leu Pro Lys Ala Gln Lys Leu Tyr 835 840 845 Leu Thr His Ile Asp Ala Glu Val Glu Gly Asp Thr His Phe Pro Asp 850 Tyr Glu Pro Asp Asp Trp Glu Ser Val Phe Ser Glu Phe His Asp Ala 865 870 875 Asp Ala Gln Asn Ser His Ser Tyr Cys Phe Glu Ile Leu Glu Arg Arg 885 890 <210> 105 <211> 63 <212> DNA <213> Artificial Sequence <220> <223> synthetic <400> 105 atggaattcg gcctgagctg gctgtttctg gtggccattc tgaagggcgt gcagtgctcc 60 63 aga <210> 106 <211> 21 <212> PRT <213> Artificial Sequence <220> <223> synthetic <400> 106 Met Glu Phe Gly Leu Ser Trp Leu Phe Leu Val Ala Ile Leu Lys Gly 1 5 10 15 Val Gln Cys Ser Arg 20 <210> 107 <211> 1482 <212> DNA

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Pro Lys Leu Leu Ile His Lys Val Ser Asn Arg Phe Ser Gly Val Pro 50 55 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile 65 70 75 80

Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Phe Cys Ser Gln Ser 85 90 95

Thr His Val Pro Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu 100 105 110

Lys Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser 115 120 125

Glu Val Lys Leu Gln Gln Ser Gly Pro Ser Leu Val Glu Pro Gly Ala 130 135 140

Ser Val Met Ile Ser Cys Lys Ala Ser Gly Ser Ser Phe Thr Gly Tyr
Page 88

155

Asn Met Asn Trp Val Arg Gln Asn Ile Gly Lys Ser Leu Glu Trp Ile 165 170 175

Gly Ala Ile Asp Pro Tyr Tyr Gly Gly Thr Ser Tyr Asn Gln Lys Phe 180 185 190

Lys Gly Arg Ala Thr Leu Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr 195 200 205

Met His Leu Lys Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 210 215 220

Val Ser Gly Met Lys Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser 225 230 235 240

Ser Ala Lys Thr Thr Pro Pro Ser Val Tyr Gly Arg Val Thr Val Ser 245 250 255

Ser Ala Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys 260 265 270

Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro 275 280 285

Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys 290 295 300

Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp 305 310 315 320

Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu 325 330 335

Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu 340 345 350

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Page 89

Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly 370 375 Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu 390 395 Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr 405 410 Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn 420 425 430 Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe 435 445 440 Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn 450 455 460 Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr 465 470 475 480 Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys Lys Asp Pro Lys 485 <210> 109 <211> 117 <212> DNA <213> Artificial Sequence <220> <223> synthetic

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Phe Pro Gly Pro Ser Lys Pro
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cgggaccctg agatgggggg aaagccgaga aggaagaacc ctcaggaagg cctgtacaat

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Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu Leu Gln Lys 50 55 60

Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys Gly Glu Arg 65 70 75 80

Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu Ser Thr Ala 85 90 95

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Ile Gly Arg Pro Leu Pro Gly Arg Lys Asn Ile Ile Leu Ser Ser Gln 50 55 60

Pro Ser Thr Asp Asp Arg Val Thr Trp Val Lys Ser Val Asp Glu Ala 65 70 75 80

Ile Ala Ala Cys Gly Asp Val Pro Glu Ile Met Val Ile Gly Gly 85 90 95

Arg Val Ile Glu Gln Phe Leu Pro Lys Ala Gln Lys Leu Tyr Leu Thr 100 105 110

His Ile Asp Ala Glu Val Glu Gly Asp Thr His Phe Pro Asp Tyr Glu 115 120 125

Pro Asp Asp Trp Glu Ser Val Phe Ser Glu Phe His Asp Ala Asp Ala 130 135 140

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Page 96

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Gln Asp Val Asn Thr Ala Val Ala Trp Tyr Gln Gln Lys Pro Gly Lys

Ala Pro Lys Leu Leu Ile Tyr Ser Ala Ser Phe Leu Tyr Ser Gly Val

Pro Ser Arg Phe Ser Gly Ser Arg Ser Gly Thr Asp Phe Thr Leu Thr

Ile Ser Ser Leu Gln Pro Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln

His Tyr Thr Thr Pro Pro Thr Phe Gly Gln Gly Thr Lys Val Glu Ile

Lys Gly Ser Thr Ser Gly Ser Gly Lys Pro Gly Ser Gly Glu Gly Ser

Gly Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly

Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Asn Ile Lys Asp

Thr Tyr Ile His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp

Val Ala Arg Ile Tyr Pro Thr Asn Gly Tyr Thr Arg Tyr Ala Asp Ser

Val Lys Gly Arg Phe Thr Ile Ser Ala Asp Thr Ser Lys Asn Thr Ala

Tyr Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Page 97

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Gly Lys Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu Leu Page 98

Glu Tyr Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu Met Gly

Gln Lys Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys Gly 450 455 460

Glu Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu Ser 465 470 475 480

Thr Ala Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala Leu Pro
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Pro Arg Gly Val Gln Val Glu Thr Ile Ser Pro Gly Asp Gly Arg Thr 500 505 510

Phe Pro Lys Arg Gly Gln Thr Cys Val Val His Tyr Thr Gly Met Leu 515 520 525

Gly Asp Gly Lys Lys Val Asp Ser Ser Arg Asp Arg Asn Lys Pro Phe 530 540

Lys Phe Met Leu Gly Lys Gln Glu Val Ile Arg Gly Trp Glu Glu Gly 545 550 555 560

Val Ala Gln Met Ser Val Gly Gln Gly Ala Lys Leu Thr Ile Ser Pro 565 570 575

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