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# DESCRIPTION

**[0001]** The present invention relates to compositions for use in improving the success rate of embryo implantation and the success rate of pregnancy in females, by providing an immunopermissive uterine environment prior to insemination or implantation of embryos. The methods of the present invention are used to make the uterus more receptive or less hostile to, for example, transferred embryos, sperm.

## BACKGROUND

**[0002]** The uterine environment, which, if hostile/non-receptive, can be responsible for poor implantation rates of good quality embryos in human and animals alike. It is believed that an inadequately primed uterine environment may also be responsible for many cases of reproductive failure in terms of failed implantation and spontaneous abortion. Similarly, a failure in uterine priming is recognised in humans as being causative to pregnancy complications such as pre-eclampsia and foetal growth restriction by preventing appropriate placental development.

**[0003]** Genetically altered or modified animals provide valuable models for testing novel gene and drug therapies *in vivo* and are the main reason the numbers of animal experiments have been rising in the last decade. In the UK, over four times as many scientific procedures using genetically modified animals were carried out in 2011 as compared to 1995. The use of genetically modified animals now represents over 50% of all scientific procedures on animals. The largest category of use is breeding (to produce genetically modified animals), with rodents accounting for almost 1.8 million procedures in the UK in 2012 alone, on a background trend for this number to increase annually. Embryo transfer in rodents underpins the development of transgenic approaches, re-derivation of specific strains and facilitates the transport of animal-lines across large distances. Typically, embryo transfer requires induction of pseudopregnancy in recipient females. This phenomenon prepares the uterus for implanting embryos, however, the success rate of transferring genetically modified embryos, despite induction of pseudopregnancy, remains relatively low.

**[0004]** Mice are spontaneous ovulators and can become pseudopregnant following an estrus in which the female is mated with a genetically sterile male such as the T145H-Re strain (which is sterile due to a chromosomal translocation) obtainable from Harlan Laboratories Inc or a vasectomised male. Both sets of males ejaculate seminal plasma devoid of functional sperm. However, both genetically sterile and vasectomised mice are relatively costly. In the instance of vasectomised males, sterility cannot be guaranteed to be 100% effective and needs testing for each male, while the production of genetically sterile males generates unwanted surplus females.

**[0005]** Alternatively, pseudopregnancy can be induced by simulating the normal vaginal stimuli

attained by mating with artificial mechanical stimulation, for example by a vibrating engraving tool (Kenney et al; J Reprod. Fert. 1977, 49, 305-309). It was found that the number and rate of intromissions were crucial influences on reproductive success (Diamond; Science, 1970, 169, 4, 995-997). Whilst this approach has seen some success in rats and mice mechanical stimulation had no effect on the induction of pseudopregnancy in the Golden Hamster (Diamond et al J. Reprod. Fert. 1968, 17, 165-168). When the female is mated with an infertile male or mechanically stimulated, the corpus luteum persists without an embryo, leading to pseudopregnancy. The female will develop mammary glands, lactate and build nests in the pseudopregnant state. There is a need to improve the methods of inducing a pseudopregnant state in laboratory test animals.

**[0006]** Although the protocols for embryo transfer in an array of rodent species are relatively well-established, their poor optimization means that there is a significant wastage of animals, raising a number of financial and ethical issues in animal units worldwide. The prior art standard approach currently relies on mating recipient females with vasectomised males to induce pseudopregnancy rather than mechanical stimulation, where copulatory activity and seminal exposure of the maternal reproductive tract triggers a neuroendocrine and localised (to the uterus, principally) inflammatory response involving a complex cascade of cytokine and prostaglandin-mediated events geared towards creating an immunopermissive environment in the uterus, thereby favouring pre-implantation embryo development and/or blastocyst implantation and the establishment of pregnancy. Even in the absence of fertilisation, luteal development and progesterone production are supported, and the maternal physiology is orchestrated to render the uterus receptive to transferred embryos for up to 10-13 days. This technique is routinely used to support the development of normal (cryopreserved strain regeneration/re-derivation) or genetically modified (transgenic/chimaeric/cloned) embryos.

**[0007]** However, the efficacy of this approach is limited. Typically, four times as many females are prepared for the procedure compared to those becoming pregnant. When implanting fresh or frozen embryos this represents a considerable wastage of valuable biological material and effort. Moreover, numbers of young vasectomised males also need to be maintained alongside the prospective recipients: these can only mate 2-3 times a week and are typically replaced every 6-9 months in order to maintain performance.

**[0008]** Mating predominantly occurs when the recipient female is in estrus. The estrus cycle lasts 4-5 days in the mouse and rat (equivalent to a woman's average 28 day menstrual cycle), which leads to the need to rely on a large pool of potential recipient females to take part in potential matings with vasectomised or otherwise sterile males. Typically, 75% of recipients are not in estrus in randomly cycling populations, leading to large numbers of females and vasectomised or otherwise sterile males being kept and, in the case of the former, often not used as surrogates in order to guarantee adequate numbers of recipients for use in timed transfers. This is particularly evident in instances where the embryos to be transferred are particularly valuable. Improvements to this approach have relied on timed estrus induction via the Whitten effect in recipient females. This strategy relies on pheromonal stimulation of recipient females, which typically brings them into estrus 3 days after exposure to stud male

urine-soiled bedding. However, the cycling stage of females at the time of pheromonal exposure, proximity to stud cages and the age of recipients can all have adverse effects on the reliability of this approach, making it relatively inefficient.

**[0009]** The chances of females being in estrus (sexually receptive) at the right time is 1:4 due to the length of their cycle (4 days). Thus, if 4 recipients are required, 16 females will be mated to 16 males, which translates to a 25% success rate. This figure can be even lower as some females will refuse to mate with their partner. The key point is that although most breeders either select females in estrus, or induce estrus before mating with sterile males, still only a relatively low percentage (often about 50% - but as low as 15% in some facilities) of oestrus females will become 'plugged' and so assumed to be pseudopregnant. Furthermore, females also have a very limited functional lifespan of a few months of age as embryo transfer recipients. Females rapidly accumulate abdominal fat as they mature, making laparotomic embryo transfers (the most common and successful method) technically too challenging.

**[0010]** By the compositions and methods of the present invention it is envisaged that the need for vasectomised or otherwise sterile male mice can be dramatically reduced along with a significant reduction of female mice usage.

**[0011]** The present invention aims to improve the pregnancy rates in mammalian females in terms of positive pregnancies and/or increased litter number following artificial or natural insemination or following transplantation of fresh or frozen or otherwise preserved embryos.

#### **BRIEF SUMMARY OF THE DISCLOSURE**

**[0012]** The present invention is defined by the claims. In accordance with the present inventions there is provided a composition of matter comprising eotaxin and RANTES for use in improving pregnancy rates and/or for reducing maternal alloreactivity against seminal fluid/sperm or embryos and/or for providing an immunopermissive uterine environment in females prior to implantation of an embryo or prior to insemination.

**[0013]** Preferably, the IL-12 is either IL-12 p40 or IL-12p70.

**[0014]** Preferably, the MIP is either MIP-1a or MIP-1b.

**[0015]** Preferably, the composition further comprises any one, two, three, four five or six additional cytokines selected from the group comprising IL-12, MCP-1, MIP, IL-17, IL-9, and GM-CSF. Thus, the composition of the present invention may be, as an illustrative example, eotaxin and RANTES in addition to one or more of TNF- $\alpha$ , TGF- $\beta$ , IL-12, MCP-1, MIP, IL-17, IL-9 and GM-CSF. It is within the scope of the invention to provide a number of specific combinations of the specified cytokines for use in inducing a uterus to be more receptive or less hostile to transferred embryo, sperm or other allografted tissue.

[0016] Accordingly, the compositions of the present invention may include a variety of multiple cytokines as it is recognised that high concentrations of a specific cytokine in seminal fluid do not necessarily reflect their biological significance.

[0017] The compositions of the present invention are selected from the following cytokines:

1. (i) eotaxin and RANTES; and optionally one or more additional cytokine selected from the group comprising;
2. (ii) TNF- $\alpha$ , TGF $\beta$ , IL-12, MCP-1, MIP IL-17, IL-9, and GM-CSF and optionally a further additional cytokine selected from the group comprising;
3. (iii) IL-1 $\alpha$ , IL-1 $\beta$ , IL-1ra, IL-2ra, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL10, IL-13, IL-15, IL-16, IL-18, FGF, G-CSF, IFN- $\alpha$ 2, IFN- $\gamma$ , IP-10, PDGF, VEGF, CTACK, KC, GRO $\alpha$ , HGF, ICAM-1, Leptin, LIF, MCP-3, M-CSF, MIF, MIG,  $\beta$ -NGF, SCF, SCGF- $\beta$ , SDF-1 $\alpha$ , TNF- $\beta$ , TRAIL VEGF and VCAM-1.

[0018] Table 1 below lists the acronyms for cytokines referred to in the present invention:

| <b>Table 1: Cytokines analysed using bio-plex assays</b> |                                   |
|----------------------------------------------------------|-----------------------------------|
| <b>IL-1<math>\alpha</math></b>                           | Interleukin-1a                    |
| <b>IL-1<math>\beta</math></b>                            | Interleukin-1 $\beta$             |
| <b>IL-1ra</b>                                            | Interleukin-1 receptor antagonist |
| <b>IL-2ra</b>                                            | Interleukin-2 receptor antagonist |
| <b>IL-2</b>                                              | Interleukin-2                     |
| <b>IL-3</b>                                              | Interleukin-3                     |
| <b>IL-4</b>                                              | Interleukin-4                     |
| <b>IL-5</b>                                              | Interleukin-5                     |
| <b>IL-6</b>                                              | Interleukin-6                     |
| <b>IL-7</b>                                              | Interleukin-7                     |
| <b>IL-8</b>                                              | Interleukin-8                     |
| <b>IL-9</b>                                              | Interleukin-9                     |
| <b>IL-10</b>                                             | Interleukin-10                    |
| <b>IL12 (p40)</b>                                        | Interleukin-12 (p40)              |
| <b>IL-12 (p70)</b>                                       | Interleukin-12 (p70)              |
| <b>IL-13</b>                                             | Interleukin-13                    |
| <b>IL-15</b>                                             | Interleukin-15                    |
| <b>IL-16</b>                                             | Interleukin-16                    |
| <b>IL-17</b>                                             | Interleukin-17                    |
| <b>IL-18</b>                                             | Interleukin-18                    |

| <b>Table 1: Cytokines analysed using bio-plex assays</b> |                                                                |
|----------------------------------------------------------|----------------------------------------------------------------|
| <b>Eotaxin</b>                                           | Eotaxin                                                        |
| <b>FGF</b>                                               | Basic fibroblast growth factor                                 |
| <b>G-CSF</b>                                             | Granulocyte-colony stimulating factor                          |
| <b>GM-CSF</b>                                            | Granulocyte macrophage-colony stimulating factor               |
| <b>IFN-<math>\alpha</math> 2</b>                         | Interferon- $\alpha$ 2                                         |
| <b>IFN-<math>\gamma</math></b>                           | Interferon- $\gamma$                                           |
| <b>IP-10</b>                                             | IFN- $\gamma$ inducible protein-10                             |
| <b>LEPTIN</b>                                            | Hormone associated with weight control                         |
| <b>MCP-1</b>                                             | Macrophage chemotactic protein-1                               |
| <b>MIP-1<math>\alpha</math></b>                          | Macrophage inflammatory protein-1 $\alpha$                     |
| <b>MIP-1<math>\beta</math></b>                           | Macrophage inflammatory protein-1 $\beta$                      |
| <b>PDGF</b>                                              | Platelet derived growth factor                                 |
| <b>RANTES</b>                                            | Regulated upon activation normal T cell expressed and secreted |
| <b>TNF-<math>\alpha</math></b>                           | Tumour necrosis factor                                         |
| <b>VEGF</b>                                              | Vascular endothelial growth factor                             |
| <b>CTACK</b>                                             | Cutaneous T cell attracting chemokine                          |
| <b>KC</b>                                                | Ketatinocyte derived cytokine                                  |
| <b>GRO<math>\alpha</math></b>                            | Growth regulated oncogene- $\alpha$                            |
| <b>HGF</b>                                               | Hepatocyte growth factor                                       |
| <b>ICAM 1</b>                                            | Intercellular cell adhesion molecule                           |
| <b>LIF</b>                                               | Leukaemia inhibitory factor                                    |
| <b>MCP3</b>                                              | Monocyte chemoattractant protein-3                             |
| <b>M-CSF</b>                                             | Macrophage-colony stimulating factor                           |
| <b>MIF</b>                                               | Macrophage migration inhibitory factor                         |
| <b>MIG</b>                                               | Monokine induced by IFN- $\gamma$                              |
| <b><math>\beta</math>-NGF</b>                            | Basic-nerve growth factor                                      |
| <b>SCF</b>                                               | Stem cell factor                                               |
| <b>SCGF-<math>\beta</math></b>                           | Stem cell growth factor- $\beta$                               |
| <b>SDF-1<math>\alpha</math></b>                          | Stromal cell derived factor-1 $\alpha$                         |
| <b>TGF-<math>\beta</math>1</b>                           | Transforming growth factor $\beta$ 1                           |
| <b>TNF-<math>\beta</math></b>                            | Tumour necrosis factor- $\beta$                                |
| <b>TRAIL</b>                                             | Tumour necrosis factor related apoptosis inducing ligand       |
| <b>VCAM-1</b>                                            | Vascular cell adhesion molecule-1                              |

**[0019]** The present invention resides in harnessing the properties of seminal agents which promote uterine receptivity and in providing a chemicophysical alternative to vasectomised or otherwise sterile males, preferably in the form of a vaginal insert such as a pessary, gel, spray or allied to any other dissolvable carrier.

**[0020]** The present invention, advantageously, given that the demand for transgenic non-human animal models is set to rise further, provides an alternative to vasectomised or otherwise sterile males inducing pseudopregnancy and also advantageously reduces the number of females required by improving uterine receptivity to transferred embryos.

**[0021]** The present invention advantageously obviates the need for vasectomised or otherwise sterile males given that their contribution solely relates to triggering the neuroendocrine and uterine inflammatory responses required to induce pseudopregnancy.

**[0022]** According to a further aspect of the invention there is provided a pharmaceutical composition as herein before described in the form of an intra-uterine device for improving pregnancy rates and/or for reducing maternal alloreactivity against seminal fluid/sperm or embryos and/or for providing an immunopermissive uterine environment in females prior to implantation of an embryo or prior to insemination.

**[0023]** Preferably, the female is mammalian and more preferably is human.

**[0024]** Preferably, the mammalian female is selected from the group comprising mouse, rat, rabbit, gerbil, guinea pig, hamster, primate (monkey, ape), canine, feline, porcine or any other laboratory animal or endangered species into which embryos are placed.

**[0025]** Preferably, the female is a mammal and more preferably still is a rare breed/species or a breed/species that is endangered.

**[0026]** Preferably, the female may be selected from the group comprising animals the orders of Artiodactyla, Carnivora, Cetacea, Chiroptera, Dermoptera, Edentata, Hyracoidae, Insectivora, Lagomorpha, Marsupialia, Perissodactyla, Pholidata, Pinnipedia, Primates, Proboscidea, Rodentia, Sirenia and Tubulidentata.

**[0027]** Preferably, the amount of any one of the cytokines present in the composition is released, from any one of its deliverable forms as described herein after, *in situ* either above or within their approximate physiological range found in seminal fluid.

**[0028]** Cytokines are measured as pg/ml as the standardised recognised values in the art.

**[0029]** The present invention resides in harnessing the properties of seminal agents, especially cytokines (which are protein modulators of the immune response and which promote uterine receptivity), by providing a chemicophysical formulation preferably in a form suitable for vaginal

delivery for insertion into the female prior to mating/insemination or prior to implantation of embryos. The introduced formulation releases agents which enhance the receptivity of the uterine environment.

**[0030]** The approach used in the present invention is to mimic the biochemical signalling mediated by seminal plasma by using a pessary-based, gel-based, solution-based, emulsion-based, powder-based or aerosol-based delivery system. Pessaries are already routinely used in an array of large domestic species (e.g. cattle) for the synchronisation of estrous cyclicity for embryo transfer/artificial insemination. However, to date, no pessary has been used with the compositions of the present composition or for the specified function of promoting uterine receptivity and/or inducing a pseudopregnant state.

**[0031]** Preferably, the cytokines are recombinant. That is to say that they are made by genetically engineering a bacterium or other cell type using recombinant technology.

**[0032]** The present invention provides compositions for females comprising recombinant cytokine preparations, typically in the form of a pessary placed in the vagina or an aerosol foam released in the vagina prior to insemination/embryo transfer in order to reduce maternal immune alloreactivity against sperm/embryos, thereby improving pregnancy rate/outcome. The use of this mode of delivery as a strategy for improving endometrial receptivity is novel.

**[0033]** Preferably, the composition further includes adjuvants such as preservatives, antioxidants, wetting agents, emulsifying agents and dispersing agents. Prevention of the action of microorganisms may be ensured by the inclusion of various antibacterial and antifungal agents, for example, paraben, chlorobutanol or phenol sorbic acid. It may also be desirable to include isotonic agents such as sugars or sodium chloride, for example. It may also be beneficial to include waxes, water and co-solvents.

**[0034]** Preferably the composition is in a form suitable for vaginal delivery such as a vaginal capsule, vaginal gel, vaginal tablet, vaginal powder, vaginal solution, vaginal pessary, vaginal cup, vaginal sponge or vaginal foam or spray. Most preferably the composition is in the form of a vaginal pessary.

**[0035]** Preferably, the vaginal formulation is dissolving or non-dissolving, degradable or non-degradable.

**[0036]** Preferably, the compositions of the present invention are prepared as a vaginal suppository, tablet, powder, bioadhesive tablet, capsule, microparticle, bioadhesive microparticle, microcapsule, microsphere, liposome, cream, lotion, foam, spray, film, ointment, solution, gel, or a sustained release gel, tablet or capsule, or a sustained release suppository administered to the vagina or incorporated into a vaginal device.

**[0037]** Preferably, the compositions of the present invention are prepared as multiwalled, multicored, microencapsulated preparations. More preferably, the active components of the

composition are when used as dried material encapsulated in a shell/coat like a gelatin capsule.

**[0038]** Compositions for vaginal administration are preferably prepared by mixing the compositions of the present invention with suitable pharmaceutical ingredients or non-irritating excipients or carriers such as cocoa butter, polyethylene glycol or a suppository wax which are solid at room temperature but liquid at body temperature and therefore melt in the rectum or vaginal cavity and release the active compound. A typical example of a vaginal pessary would include the active ingredients and the following excipients: medium chain tryglycerides and hard fat.

**[0039]** Alternatively, the composition can be incorporated into an intravaginal device or a coating of such device, for example, a tampon or tampon-like device coating, or incorporated into a sponge, foam, strip, powder, pessary, or other material. Absorbent material or matrix of such devices may be impregnated with a composition of the present invention as a liquid solution, suspension lotion, powder, cream, microemulsions or suspension of liposomes, bioadhesive nanoparticles, or bioadhesive microparticles. Preferably, the vaginal device is dissolving or non-dissolving, degradable or non-degradable.

**[0040]** Preferably, the compositions further include a mucoadhesive agent, sorption promoter or penetration enhancer. The compositions of the present invention are delivered by transmucosal vaginal delivery and comprise contacting the vaginal mucosa with the compositions of the present invention.

**[0041]** Preferably, the compositions for vaginal delivery are for rapid delivery, controlled delivery, continuous delivery or pulsed delivery.

**[0042]** It is envisaged that the composition of the present invention will be formulated in one embodiment as a pessary with a slow wax melt or a fast wax melt to achieve continuous or rapid delivery respectively. In an alternative embodiment, the composition of the invention will be formulated into a foam or gel with appropriate additives to permit controlled release.

**[0043]** Preferably, in the instance of providing the composition of the present invention as a vaginal pessary, tablet, bioadhesive tablet, capsule, powder, microparticle, bioadhesive microparticle, that the coating will be abrasive. This is an appropriate modification to the deliverable composition, especially if the penis of the male of the species to be prepared for uterine receptivity has a rough epidermis, keratinous spines, etc. In this instance it is desirable for the inserted vaginal delivery vehicle to have a rough outer coating similar to that as on the penis in order to elicit a maternal inflammatory response to improve penetration of the preparation into the mucosa, elicit an initial inflammatory response and aid generic neuroendocrine stimulation.

**[0044]** In a particularly preferred embodiment of the invention the composition is in the form of a pessary. Preferably the pessary is suitably sized and shaped so as to be inserted into the

vagina of the female. The pessary may have a solid core and an outer porous layer. Typical dimensions of a pessary for mice are overall diameter of 4 mm and a length of 7mm. The diameter and length of the pessary is dependent upon the size of the vagina of the species into which it is inserted, it is desirable that the pessary be sized and shaped so that when inserted into the vagina it is retained without discomfort.

**[0045]** According to a further aspect of the invention there is provided the composition of the first aspect of the invention for the manufacture of a medicament for improving pregnancy rates and/or reducing maternal alloreactivity against seminal fluid/sperm or embryos and/or for providing an immunopermissive uterine environment in females prior to implantation of an embryo or prior to insemination.

**[0046]** Described but not claimed is a method of reducing maternal alloreactivity in a female against seminal fluid/sperm, embryos or other allograft (including existing microflora) comprising exposing the vaginal/gastrointestinal tract mucosa to a composition as hereinbefore described, the method comprising:

1. (i) introducing at least one vaginal delivery vehicle comprising the composition of the present invention into the vagina of the female;
2. (ii) optionally inserting further vaginal delivery vehicle(s);
3. (iii) allowing a sufficient period of time to elapse to allow the active components of the vaginal delivery vehicle to be released and to penetrate into the vagina and be absorbed transmucosally and/or diffuse and/or be transported into the uterus; and
4. (iv) inseminating the female by either mating with a male or by artificial insemination, donor gametes or introducing an embryo or other allograft into the uterus for implantation.

**[0047]** Described but not claimed is a method of improving pregnancy rate or outcome in a female prior to insemination or implantation of an embryo comprising exposing the vaginal/gastrointestinal tract mucosa to a composition as hereinbefore described, the method comprising:

1. (i) introducing at least one vaginal delivery vehicle comprising the compositions of the present invention into the vagina of the female ;
2. (ii) optionally inserting further vaginal delivery vehicle(s);
3. (iii) allowing a sufficient period of time to elapse to allow the active components of the vaginal delivery vehicle to be released and to penetrate into the vagina and be absorbed transmucosally and/or diffuse and/or be transported into the uterus; and
4. (iv) inseminating the female by either mating with a male or by artificial insemination, donor gametes or introducing an embryo or other allograft into the uterus for implantation.

**[0048]** In a further embodiment of the invention, steps (iii) and (iv) are performed simultaneously/close chronological sequence in the case of artificial or natural insemination. The methods of the invention uses a composition comprising recombinant cytokine-containing pessaries, gel, spray or foam placed in the vagina at the time of insemination/embryo transfer to reduce maternal immune alloreactivity against sperm/embryos/gametes and endogenous or exogenous microflora, thereby improving pregnancy rate.

**[0049]** The number of doses and period between doses can be varied according to requirements and may vary depending on species or breed, super/ovulation status, seasonal effect, lactational status and number of previous failed pregnancies or previous inseminations or IVF treatments. It may also vary depending on maternal age and whether the female is *primigravida*. Preferably, the methods of the invention when used for laboratory non-human animals further includes the step of synchronizing estrus in recipient females.

**[0050]** According to a yet further aspect of the invention there is provided a kit of parts and optionally a set of written instructions therefore, the kit comprising a number of vaginal delivery vehicles containing the compositions of the invention and an apparatus for inserting said vaginal delivery vehicles into the vagina of the recipient female.

**[0051]** It will be appreciated that features described for the first aspect of the invention are equally applicable to each and all aspects of the invention and apply *mutatis mutandis*.

**[0052]** The invention will be described by way of example only with reference to the following figures wherein:

## **BRIEF DESCRIPTION OF THE DRAWINGS**

**[0053]** Embodiments of the invention are further described hereinafter with reference to the accompanying drawings, in which:

Figure 1 shows the Bayesian mathematical modelling of cytokine networks in mouse seminal plasma. The nodes (cytokines) are colour-coded according to the conditional probability of corresponding mediator relative concentrations being high (green), low (red) or medium (white) concentration given the state(s) of their parent nodes (the bar charts adjacent to each node reflect underlying conditional probabilities of categorization into one of the three concentration bins from low on the left to high on the right) . The normalized concentration (low or high) determines the intensity of the node colour. Edges (causal connecting lines between nodes) represent causal directed interactions between nodes. These cytokines will interact with the maternal reproductive tract to induce immunopermissiveness to paternal antigens.

Figure 2 shows the Bayesian mathematical modelling of cytokine networks in rat seminal plasma (details discussed above). Very high confidence level edges are coloured in red, based upon the confidence analysis of the Bayesian result (based on occurrence in >90%

bootstrapping iterations).

Figure 3 shows a bar chart of the percentage of successful deliveries following induction of pseudopregnancy by mating with a vasectomised male or a pessary (n=14 per group) of full mix, mix 1, mix 2 and mix 3.

## DETAILED DESCRIPTION

**[0054]** Reference herein to "embryo" is intended to include a blastula, blastocyst, fertilized ovum or an organism in its early stages of development, especially before it has reached a distinctively recognizable form that is to be implanted into a female recipient. The terms are used interchangeably.

**[0055]** Reference herein to an "improved pregnancy rate" is intended to include a positive pregnancy outcome or improved perinatal survival or general viability following artificial insemination with processed semen or natural insemination or following transplantation of fresh or frozen or otherwise preserved embryos. The term pregnancy as used hereinafter is to be interpreted as encompassing a pregnancy resulting from natural or artificial insemination or following transplantation of a fresh or frozen or otherwise preserved embryo(s) and gametes.

**[0056]** Reference herein to an "intra-uterine device" is intended to include any pessary-based, gel-based, solution-based, emulsion-based, powder-based or aerosol-based delivery system that is capable of delivering the compositions of the present invention into the vagina so as to permit the compositions of the present invention to have a pharmacological effect on the uterine environment.

**[0057]** Reference herein to a "pessary" is intended as a means of delivery of the pharmaceutical substances of the present invention so that they are easily absorbed through the mucosal surfaces of the vagina, or intended to have action in the locality, for example against inflammation, or on the uterus.

**[0058]** "Pharmaceutical ingredient" or "excipient" means a pharmacologically inactive pharmaceutically acceptable compound added to a mucoadhesive composition of the invention. The ingredient or excipient does not have any pharmacological properties.

**[0059]** "Rapid delivery" means initial immediate rapid release and delivery of the components from the composition. The rapid delivery is typically followed by a time-dependent reduction in release of the components from the composition or device and delivery of the drug to the plasma/uterine wall tissues (or gastrointestinal tract, where appropriate).

**[0060]** "Controlled delivery" means a release wherein the active agent is released from the

material in a predesigned manner. The release of the active agent may be constant over a long period, it may be cyclic over a long period, or it may be triggered by the environment or other external events.

**[0061]** "Continuous delivery" means continuous and uninterrupted release of the components from the formulation or device and delivering such components in a continuous manner. Continuous delivery may be preceded by the rapid delivery.

**[0062]** "Pulsed delivery" means a release and delivery of the components in intermittent intervals. Such pulsed delivery may be provided, for example, by formulating the composition in individual layers interspaced with inactive layers of dissolvable coatings or by using different pharmaceutical ingredients.

### ***Seminal Fluid cytokine analysis***

**[0063]** Sexually mature CD1 male mice (n = 20) and Wistar rats (n = 20) were sacrificed and seminal fluid collected from the seminal glands *post mortem*, a *post mortem* approach was chosen to avoid collecting samples by electroejaculation since semen quality is variable by this method, and because the samples coagulate rapidly, making analysis problematic. Seminal vesicle sampling is ideal as the fluid (rather than that of the accessory glands) contains the maternal tract immunomodulatory factors investigated and because coagulating gland secretions can more easily be avoided.

**[0064]** Seminal fluid samples were weighed individually, suspended in phosphate buffered saline (PBS) supplemented with 0.5% bovine serum albumin (BSA), and weighed again. By inference to standard weight:volume ratio of murine seminal fluid, it was possible to determine the original volume isolated and the dilution factor introduced by the PBS. This step was necessary because the fluid is too viscous to be pipetted accurately. Samples were spun and the supernatant frozen at -80°C until analysed simultaneously for the following 23 cytokines: interleukin (IL)-1 $\alpha$ , IL-1 $\beta$ , IL-2, IL-3, IL-4, IL-5, IL-6, IL-9, IL-10, IL-12 (p40), IL-12 (p70), IL-13, IL-17, eotaxin, granulocyte-colony stimulating factor (G-CSF), granulocyte macrophage-colony stimulating factor (GM-CSF), interferon (IFN)- $\gamma$ , keratinocyte-derived chemokine (KC), monocytes chemotactic protein (MCP)-1, macrophage inhibitory protein (MIP)-1 $\alpha$ , MIP-1 $\beta$ , regulated upon activation normal T cell expressed and secreted (RANTES) and tumour necrosis factor (TNF)- $\alpha$ . This was achieved by custom 23-plex fluid-phase immunoassay kits run on a Luminex-100™ equipped with StarStation™ software. Serum diluent was used in all cases to avoid false positive/negatives and dilution adjusted to 1:1 in order to maximise sensitivity to baseline levels. Similar analysis was performed on rat seminal fluid.

### **EXAMPLE 1**

**[0065]** Table 2 below shows a variety of cytokines analyzed and measured in mouse seminal fluid. Eotaxin and RANTES appear to be the predominant cytokines present, with levels above 500pg/ml. IL-9, TNF- $\alpha$  and MIP-1a had levels above 100 pg/ml whereas several cytokines such as G-CSF and IFN- $\gamma$  had levels between 50-100 pg/ml and several others such as IL-13 and TGF- $\beta$  had levels below 50 pg/ml.

Table 2

| Mouse          | Mean   | SEM   |
|----------------|--------|-------|
| IL-1 $\alpha$  | 8.19   | 1.96  |
| IL-1 $\beta$   | 87.48  | 9.04  |
| IL-2           | 3.03   | 0.49  |
| IL-3           | 0.35   | 0.04  |
| IL-4           | 0.11   | 0.01  |
| IL-5           | 0.56   | 0.07  |
| IL-6           | 3.63   | 0.44  |
| IL-9           | 135.14 | 33.47 |
| IL-10          | 19.95  | 3.36  |
| IL-12 p40      | 5.25   | 0.53  |
| IL-12 p70      | 10.91  | 1.08  |
| IL-13          | 20.64  | 1.86  |
| IL-17          | 5.10   | 0.90  |
| Eotaxin        | 857.22 | 73.85 |
| G-CSF          | 45.03  | 3.33  |
| GM-CSF         | 4.16   | 0.39  |
| IFN- $\gamma$  | 46.38  | 3.95  |
| KC             | 37.17  | 3.56  |
| MCP-1          | 30.23  | 2.65  |
| MIP-1 $\alpha$ | 114.32 | 8.31  |
| MIP-1 $\beta$  | 6.68   | 1.36  |
| RANTES         | 618.62 | 84.17 |
| TNF- $\alpha$  | 102.27 | 9.11  |
| TGF- $\beta$   | 27.63  | 6.54  |

**[0066]** Table 3 below shows a variety of cytokines analyzed and measured in rat seminal fluid. RANTES appears to be the predominant cytokine present. Of the cytokines analyzed only RANTES and GRO/KC had levels above 200pg/ml. IL-10 and IL-6 had levels above 100 pg/ml whereas several cytokines such as MCP-1 had levels between 50-100 pg/ml and several others such as IL-17 had levels below 50 pg/ml.

TABLE 3

| Rat            | Mean   | SEM  |
|----------------|--------|------|
| IL-1 $\alpha$  | 3.28   | 0.97 |
| IL-1 $\beta$   | 20.41  | 0.84 |
| IL-2           | 29.11  | 3.40 |
| IL-4           | 20.17  | 1.13 |
| IL-5           | 9.58   | 0.95 |
| IL-6           | 149.17 | 1.13 |
| IL-9           | 54.56  | 0.84 |
| IL-10          | 114.89 | 1.45 |
| IL-12 p70      | 55.14  | 4.31 |
| IL-13          | 8.29   | 1.35 |
| IL-17          | 15.80  | 1.11 |
| IL-18          | 6.66   | 0.89 |
| TNF- $\alpha$  | 2.27   | 0.16 |
| IFN- $\gamma$  | 2.93   | 0.39 |
| Eotaxin        | 34.84  | 1.45 |
| GCSF           | 1.51   | 0.07 |
| GMCSF          | 40.53  | 2.10 |
| MCP-1          | 61.56  | 2.21 |
| LEPTIN         | 43.69  | 2.61 |
| MIP-1 $\alpha$ | 0.13   | 0.02 |
| IP-10          | 4.24   | 0.34 |
| GRO/KC         | 228.00 | 2.10 |
| RANTES         | 287.31 | 2.21 |
| VEGF           | 0.00   | 0.00 |
| TGF- $\beta$   | 0.00   | 0.00 |

**EXAMPLE 2**

**[0067]** Eotaxin and RANTES appear to be the predominant cytokines each being present in an amount of more than 500pg/ml (see Tables 1 and 2). The cytokines IL-1 $\alpha$ , IL-6, IL-10, IL-12 (p40), IL-12 (p70), GM-CSF and MIP-1 $\beta$  were present at levels below 20pg/ml and cytokines IL-1 $\beta$ , IL-9, IL-13, G-CSF, TNF- $\alpha$ , MCP-1, KC, MIP-1 $\alpha$  and IFN- $\gamma$  were present at levels above 20 and below 150 pg/ml.

[0068] Based on these analyses, a solution of cell culture-tested recombinant mouse cytokines was made up in PBS using recombinant cytokines at the concentrations found in seminal fluid (Table 3). This was stored at -80°C until required for imbining the pessary.

TABLE 4 Cytokine Concentrations *in utero* in a Mouse Pessary Preparation once solubilised.

| Cytokine       | Pessary solution concentration (pg/ml) |
|----------------|----------------------------------------|
| IL-1 $\alpha$  | 8.19                                   |
| IL-1 $\beta$   | 87.48                                  |
| IL-6           | 3.63                                   |
| IL-9           | 135.14                                 |
| IL-10          | 19.95                                  |
| IL-12 (p40)    | 5.25                                   |
| IL-12 (p70)    | 10.91                                  |
| IL-13          | 20.64                                  |
| G-CSF          | 45.03                                  |
| GM-CSF         | 4.16                                   |
| TNF- $\alpha$  | 102.27                                 |
| MCP-1          | 30.23                                  |
| RANTES         | 618.62                                 |
| Eotaxin        | 857.22                                 |
| KC             | 37.17                                  |
| MIP-1 $\alpha$ | 114.32                                 |
| MIP-1 $\beta$  | 6.68                                   |
| IFN- $\gamma$  | 46.38                                  |

[0069] Pessaries also include an amount of TGF- $\beta$ 1 as this is important in eliciting uterine receptivity.

### EXAMPLE 3

[0070] Additional formulation components of pessaries for laboratory animals was dictated principally by toxicity (in case of accidental ingestion), palatability (to dissuade ingestion) and impact on luminal pH (the bioactivity of certain cytokines is promoted by vaginal pH). The size and shape of the pessaries is largely determined by the species for which their use is intended. For example, pessaries of approximately 4mm in diameter are particularly suitable for mice since the size has been determined as appropriate for insertion without undue discomfort and is also of a suitable size to be retained in the vaginal vestibule. Larger laboratory animals or indeed larger breeds of mice may necessitate larger pessaries. Pessaries were made from

laser-etched nylon at a setting of between 5-10 Watts, use of this technique makes it possible to manipulate porosity (which facilitates 'loading') and overall shape and dimensions. It is envisaged that pessaries will be provided in a range of sizes and that the stalks may be snapped off from a central holding unit for use as desired and that a range of different sizes of pessaries may be provided to the user. Pessaries are prepared for use by soaking them overnight in 500µl of 100 times the concentration of cytokine solution so as to load the pessaries with the necessary active agents to guarantee a seminal fluid like final concentration in the maternal reproductive tract. A pessary head is then removed from the stalk and inserted by means a suitable device directly into the mouse vagina. The pessary is then left in the mouse vagina for a period of time and then it is either removed at the time of embryo transfer when the animals are anaesthetised, or it self-dissolves or the pessary self ejects once the active ingredients have been absorbed.

**[0071]** It will be appreciated that the above embodiment is only one example of a means of delivering the compositions of the present invention and that the pessary may be in the form of a slow or fast melt wax type formulation and that the method of delivering the compositions of the present invention may vary from species to species. The delivery means may also be in the form of a biodegradable product and for example, in humans a vaginal sponge may be a more convenient method of delivering the compositions.

#### **EXAMPLE 4**

**[0072]** A full mix and three different combinations of reduced cytokine mixes, at physiological concentrations, were prepared as shown in the Table 5. The full mix comprised IL-12 (P40), RANTES, Eotaxin, MIP-1 $\beta$ , TNF- $\alpha$ , 1L13, IFN- $\gamma$ , TGF- $\beta$ , IL-1 $\beta$ , IL-6, 1L-12 (p70), MCP-1, IL-10, IL-1a, IL-9, GM-CSF, KC, MIP-1 and GCSF.

**[0073]** Mix 1 contained IL-12 (P40), RANTES, MIP-1 $\beta$ , IL-13 and TGF- $\beta$ . IL-12 (P40) was included due to its role as a major network ancestral parent (given the marked number of outgoing edges) and its putative causal control of multiple downstream mediators. RANTES and MIP-1 $\beta$  were included as these were hub nodes (likely involved in multiple signalling integration based on the high number of incoming edges). The importance of MIP-1 signalling highlighted by the conservation of this hub node function across species where its function is likely analogous to that highlighted in the rat network (Figure 2) IL-13 was included following a similar rationale and because it appeared to be the principal node fulfilling this function in the right hand side of the network main branch illustrated in Figure 1. TGF- $\beta$  was included due to its purported potent immunomodulatory properties which temper the mating-induced inflammatory response.

**[0074]** Mix 2 contained IL-12 (P40), RANTES, MIP-1 $\beta$ , IL-13 and IFN- $\gamma$ . The choice of mediators for this mix is given above with the exception of IFN- $\gamma$  which, despite being a recognised abortifacient likely has a physiological role to play as a hub node at low levels as presented herein.

**[0075]** Mix 3 contained RANTES, TNF- $\alpha$  and TGF- $\beta$  for the reasons highlighted above - TNF- $\alpha$  was added given that it was the only terminal node of the network. Its likely key role is underscored by the fact that its terminal node position was consistent across both the mouse and rat, despite the difference in species and the change in some of the network nodes analysed. Its role in reproductive immunity is hotly debated but it is increasingly being recognised as a likely key player in triggering the early inflammatory response associated with this process. This selection was based on the mathematical modelling of cytokine networks in seminal plasma (see Figures 1 and 2). The full mix composition is based on the most extensive murine multiplex immunoassay-based analysis available for seminal plasma analysis.

**Table 5:** The cytokine composition of the four pessaries used in this study (shaded boxes indicate inclusion in the mix)

| Cytokine    | Full Mix | Mix 1 | Mix 2 | Mix 3 |
|-------------|----------|-------|-------|-------|
| IL-12 (p40) |          |       |       |       |
| RANTES      |          |       |       |       |

|                |  |  |  |  |
|----------------|--|--|--|--|
| Eotaxin        |  |  |  |  |
| MIP-1 $\beta$  |  |  |  |  |
| TNF- $\alpha$  |  |  |  |  |
| IL-13          |  |  |  |  |
| IFN- $\gamma$  |  |  |  |  |
| TGF- $\beta$   |  |  |  |  |
| IL-1 $\beta$   |  |  |  |  |
| IL-6           |  |  |  |  |
| IL-12 (p70)    |  |  |  |  |
| MCP-1          |  |  |  |  |
| IL-10          |  |  |  |  |
| IL-1 $\alpha$  |  |  |  |  |
| IL-9           |  |  |  |  |
| GM-CSF         |  |  |  |  |
| KC             |  |  |  |  |
| MIP-1 $\alpha$ |  |  |  |  |
| G-CSF          |  |  |  |  |

**[0076]** The data shown in Figure 3, confirms that the current nylon prototype pessary, 'loaded' with either a full complement or reduced numbers of cytokines, can successfully induce pseudopregnancy in CaCBAXC57Bl/6 F1s. We found that our three mixes of reduced cytokines gave different rates of success but that one combination in particular, Mix 3, induced pseudopregnancy with a similar success rate as purchased vasectomised males (n=14).

**[0077]** Throughout the description and claims of this specification, the words "comprise" and

"contain" and variations of them mean "including but not limited to", and they are not intended to (and do not) exclude other moieties, additives, components, integers or steps. Throughout the description and claims of this specification, the singular encompasses the plural unless the context otherwise requires. In particular, where the indefinite article is used, the specification is to be understood as contemplating plurality as well as singularity, unless the context requires otherwise.

**[0078]** Features, integers, characteristics, compounds, chemical moieties or groups described in conjunction with a particular aspect, embodiment or example of the invention are to be understood to be applicable to any other aspect, embodiment or example described herein unless incompatible therewith. All of the features disclosed in this specification (including any accompanying claims, abstract and drawings), and/or all of the steps of any method or process so disclosed, may be combined in any combination, except combinations where at least some of such features and/or steps are mutually exclusive.

## REFERENCES CITED IN THE DESCRIPTION

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

### Non-patent literature cited in the description

- **KENNEY et al.**J Reprod. Fert., 1977, vol. 49, 305-309 [\[0005\]](#)
- **DIAMOND**Science, 1970, vol. 169, 4995-997 [\[0005\]](#)
- **DIAMOND et al.**J. Reprod. Fert., 1968, vol. 17, 165-168 [\[0005\]](#)

## PATENTKRAV

1. Stofsammensætning, der omfatter eotaxin og RANTES, til anvendelse til forbedring af svangerskabsrater og/eller til reduktion af maternel alloreaktivitet mod sædfluid/sperm eller embryoer og/eller til tilvejebringelse af et immunpermissivt uterint miljø hos hunner før  
5 implantering af et embryo eller før insemination.
2. Sammensætning til anvendelse ifølge krav 1, der endvidere omfatter et hvilket som helst eller flere af de supplerende otte cytokiner valgt fra gruppen omfattende TNF- $\alpha$ , TGF- $\beta$ , IL-12, MCP-1, MIP, IL-17, IL-9 og GM-CSF.
3. Sammensætning til anvendelse ifølge krav 2, hvor IL-12 enten er IL-12 p40 eller IL-  
10 12p70 og hvor MIP enten er MIP-1a eller MIP-1b.
4. Farmaceutisk sammensætning til anvendelse ifølge et hvilket som helst foregående krav i form af en intrauterin anordning til anvendelse til forbedring af svangerskabsrater og/eller til reduktion maternel alloreaktivitet mod sædfluid/sperm eller embryoer og/eller til tilvejebringelse af et immunpermissivt uterint miljø hos hunner før implantering af et embryo  
15 eller før insemination.
5. Farmaceutisk sammensætning til anvendelse ifølge krav 4, hvor aktive ingredienser frigives på stedet enten over eller inden for det omtrentlige fysiologiske interval som de, der findes i sædfluid.
6. Farmaceutisk sammensætning til anvendelse ifølge enten krav 4 eller 5, hvor den  
20 intrauterine anordning er vaginalkapsel, vaginalgel, vaginaltablet, vaginalpulver, vaginalopløsning, vaginalpessar, vaginalkop, vaginalsvamp, vaginalaerosol eller vaginalskum eller spray.
7. Farmaceutisk sammensætning til anvendelse ifølge enten krav 5 eller 6, der endvidere indbefatter en adjuvans, excipiens eller bærer.
- 25 8. Farmaceutisk sammensætning til anvendelse ifølge et hvilket som helst krav 5 til 7 til hurtig indgivelse, styret indgivelse, kontinuerlig indgivelse eller pulseret indgivelse.
9. Pessar til transmukosal vaginal indgivelse, der omfatter sammensætningen ifølge et hvilket som helst krav 1 til 3, til anvendelse til fremme af uterin modtagelighed.
10. Pessar ifølge krav 9 til anvendelse ved fremme af uterin modtagelighed hos en rotte,  
30 eventuelt hvor gnaveren er en mus eller rotte.
11. Kit af dele, der omfatter et antal vaginale indgivelsesvehikler, der indeholder sammensætningerne ifølge et hvilket som helst af kravene 1 til 8 eller pessarerne ifølge krav 9 til 10, hvilket kit er beregnet til anvendelse til forbedring af svangerskabsrater og/eller til reduktion af maternel alloreaktivitet mod sædfluid/sperm eller embryoer og/eller til

tilvejebringelse af et immunpermissivt uterint miljø hos hunner før implantering af et embryo eller før insemination, og en anordning til indføring af de vaginale indgivelsesvehikler i modtagerhunnens vagina og eventuelt et sæt skriftlige instruktioner dertil.

# DRAWINGS

FIGURE 1

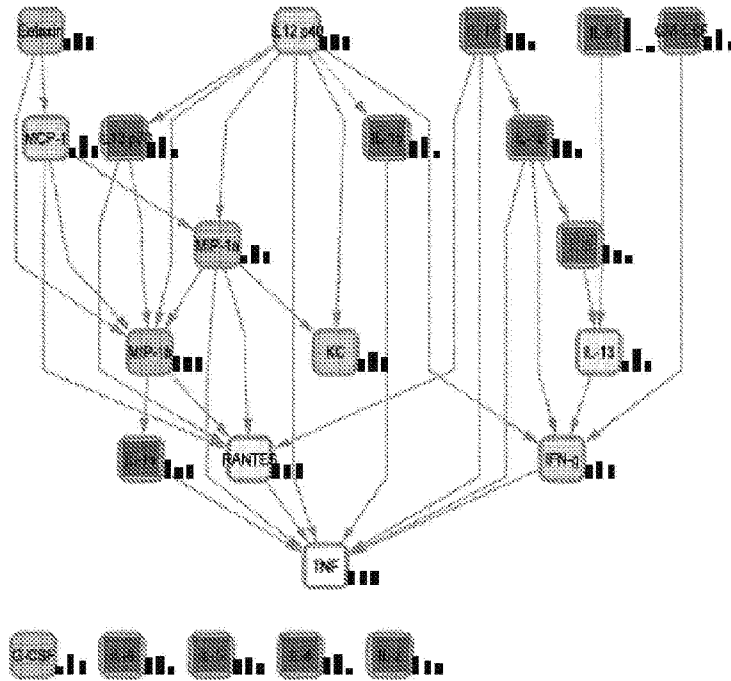


FIGURE 2

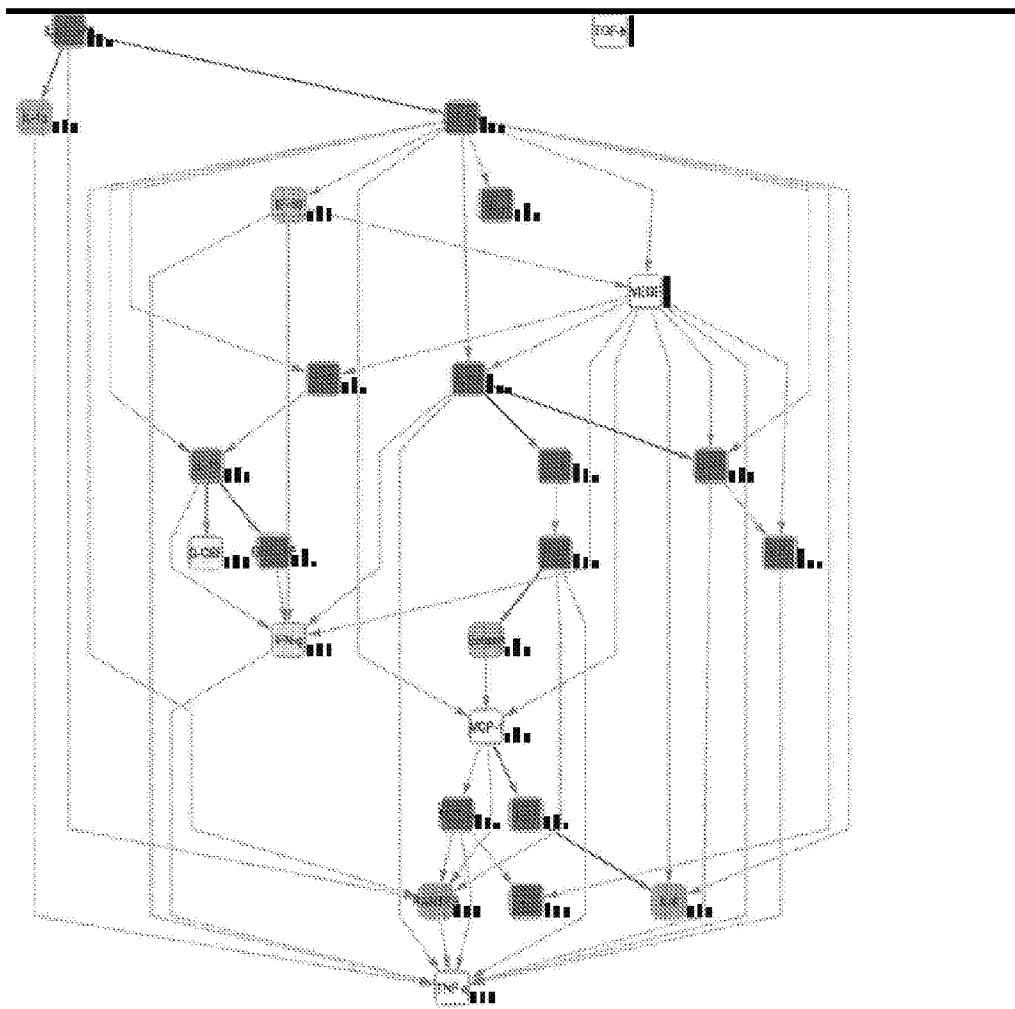


FIGURE 3

