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ANALETTE I. LOPEZ ET AL: "Biofunctionalization of silicone polymers using poly(amidoamine) dendrimers and a mannose derivative for prolonged interference against pathogen colonization", BIOMATERIALS., vol. 32, no. 19, 1 July 2011 (2011-07-01), pages 4336-4346, XP055416118, GB ISSN: 0142-9612, DOI: 10.1016/j.biomaterials.2011.02.056

Fortsættes ...

DESCRIPTION

TECHNICAL FIELD

[0001] The present disclosure relates to catheters including a hydrophilic coating that include mannose as an anti-infective agent and methods of forming such catheters.

BACKGROUND

[0002] It is desirable for medical devices that are inserted into the body to have a lubricated or lubricious outer surface to increase comfort of the patient during insertion into and/or removal from the body. Such devices may include, for example, urinary catheters, endoscopes, cardiovascular catheters, syringes, vascular stents, etc. One method for rendering the surface of a medical device lubricious is to coat at least the insertable portion of the medical device with a lubricious hydrophilic coating. An advantage associated with hydrophilic coatings is that the coatings become highly slippery or lubricous when wetted with a wetting fluid, such as saline or water. The high lubricity minimizes soft tissue damage and reduces overall discomfort during use of the medical device.

[0003] Although medical devices are typically prepared in sterile environments and are provided with safe handling instructions, the risk of bacterial contamination exists and is present whenever a medical device is inserted into a patient's body. For example, insertion of a urinary catheter into the urethra of a patient for draining a bladder inherently carries the risk of introducing bacteria and other contaminants into the urethra and bladder of the patient. The introduction of such contaminants into the urinary tract can lead to urinary tract and bladder infections. Such infections may result from the use of both intermittent and indwelling catheters. Additionally, it has been shown that some users who contract a urinary tract infection become more susceptible to recurrent urinary tract infections. ANALETTE I. LOPEZ ET AL: "Biofunctionalization of silicone polymers using poly(amidoamine) dendrimers and a mannose derivative for prolonged interference against pathogen colonization", BIOMATERIALS., vol. 32, no. 19, 1 July 2011 (2011-07-01), pgs. 4336-4346 discloses using mild CO₂ plasma to activate a PDMS surface followed by simple attachment of generation 5 (G5) poly(amidoamine) (PAMAM) dendrimers to generate an amino-terminated surface. Then, carboxy-terminated mannose are covalently attached to the PAMAM surface. Benign E.coli bacteria is then attached to the mannose.

[0004] The use of intermittent catheters comprising anti-infective agents and/or agents that support bladder health can help prevent urinary tract infections and reduce the frequency of recurrent infection. Therefore, there is a need for medical devices which reduce the risk of associated infections.

SUMMARY

[0005] In one aspect, a catheter includes a surface and a coating disposed on the surface, wherein the coating includes a hydrophilic polymer matrix and mannose located within the polymer matrix.

[0006] In yet another aspect, a method of forming a catheter includes forming a hydrophilic coating on a medical device, and immersing catheter including the coating in a solution including mannose, wherein in the solution carries mannose into a polymer matrix of the hydrophilic coating.

Brief Description of Drawings**[0007]**

Figs. 1 and 2 are photographs of catheter tubes showing the visual results of the dye uptake tests of Top Coat 1;

Figs. 3 and 4 are photographs of catheter tubes showing the visual results of the dye uptake tests of Top Coat 2;

Figs. 5 and 6 are photographs of catheter tubes showing the visual results of the dye uptake tests of Top Coat 3;

Figs. 7 and 8 are photographs of catheter tubes showing the visual results of the dye uptake tests of Top Coat 4; and

Fig. 9 is a photograph of catheter tubes showing the visual results of the dye uptake tests of Top Coat 5.

Detailed Description

[0008] The present disclosure relates to formulations for forming hygienic hydrophilic coatings, hygienic lubricious coatings, catheters including the aforementioned coatings and methods of forming such coatings. The present disclosure also discloses anti-infective additives and wetting fluids that impart anti-infective properties to hydrophilic coatings. The coatings, catheters including such coatings, anti-infective additives and wetting fluids are used in several different applications. For example, the coatings are used to ease insertion of the medical device into various types of body cavities and lumens, while at the same time providing an anti-infective effect.

[0009] Urinary catheters typically include an elongated tube or catheter shaft having an insertable portion that is inserted through the urethra and into the bladder to drain urine therefrom. The catheter tube includes a proximal insertion end portion that is inserted through the urethra and into the bladder. The proximal end portion has drainage eyes or holes that allow urine to drain from the bladder and through the catheter tube. The catheter tube also includes a distal end portion that has a drainage element, such as a funnel, associated therewith to drain the urine into a collection container, such as a toilet or waste collection bag.

[0010] The elongated catheter tube is made of any suitable materials, which are well known in the field. Such materials include for example polyvinyl chloride, polyurethane, latex rubbers, silicone, thermoplastic elastomers (TPE), thermoplastic olefin (TPO), PEBAX etc. and copolymers and mixtures thereof.

[0011] A hygienic anti-infective hydrophilic coating is disposed on a surface of the catheter shaft. For example, the hydrophilic coating is disposed on the insertable portion of the catheter shaft. The hydrophilic coating includes a hydrophilic polymer, anti-infective agent(s) and, optionally, other additives. The hydrophilic polymer is any suitable hydrophilic polymer capable of providing hydrophilicity to the coating and/or lubriciousness when the coating is hydrated. The polymer is synthetic or bio-derived and can be blends or copolymers of both. Suitable hydrophilic polymers include but are not limited to poly(lactams), for example polyvinylpyrrolidone (PVP), polyethylene oxide, polyurethanes, homo and copolymers of acrylic and methacrylic acid, polyvinyl alcohol, polyvinyl ethers, maleic anhydride based copolymers, polyesters, vinyl amines, polyethylenimines, polyethylene oxides, poly(carboxylic acids), polyamides, polyanhydrides, polyphosphazenes, cellulose, for example methyl cellulose, carboxymethyl cellulose, hydroxymethyl cellulose, and hydroxypropyl cellulose, heparin, dextran, polysaccharides, for example chitosan, hyaluronic acid, alginates, gelatin, and chitin, polyesters, for example polylactides, polyglycolides, and polycaprolactones, polypeptides, for example collagen and fibrins.

[0012] The anti-infective agents of the hydrophilic coating are agents that are capable of inhibiting the growth or migration of an infectious agent and/or are capable of killing an infectious agent. Such infectious agents may include bacteria, viruses, fungi and the like. The anti-infective agents of the hydrophilic coatings include, for example, mannose, or honey (natural or synthetic). Beneficially, such agents also provide favorable osmolality without the need for additional tonicity agents.

[0013] Hydrophilic Coatings and Wetting Fluids Containing Mannose

[0014] Hygienic hydrophilic coatings of the present disclosure include a hydrophilic polymer, mannose and, optionally, other additives and agents. As used herein, the term "mannose" includes D-mannose, L-mannose, oligomannose, polymannose and derivatives thereof.

[0015] The hygienic hydrophilic coating (in the dry state) includes

hydrophilic polymer(s) in an amount between about 80 wt% and about 99.9 wt% of the coating;
and

mannose in an amount between about 0.1 wt% and about 20 wt% of the coating.

In one embodiment, the hygienic hydrophilic coating (in the dry state) includes

hydrophilic polymer(s) in an amount between about 90 wt% and about 99 wt% of the coating;
and

mannose in an amount of about 1 wt% and about 10 wt%.

[0016] The hygienic hydrophilic coating may optionally include other agents or additives, such as organic acids, salts of organic acids, antioxidants and/or peroxide generating enzymes.

[0017] The organic acids may be, for example, citric acid, ascorbic acid (also an anti-oxidant), glycolic acid, acetic acid, butyric acid, formic acid, gluconic acid, lactic acid, malic acid, pyroglutamic acid, tartaric acid and succinic acid etc., glutaric acid, benzoic acid, polycarboxylic acid and mixtures thereof. The acid may be used to aid in maintaining a slightly acid pH in the coating and the surrounding environment, e.g., inside the package in which the device having the coating is stored. As explained in more detail below, the mannose may be catalyzed to produce peroxide, and a slightly acidic pH may help support peroxide stability in the coating composition and the surrounding environment. Depending on the application, it may be desirable to maintain a pH of approximately 4 to 8 in the coating and/or surrounding environment, preferably from approximately 5 to 7, and more preferably approximately 5.5 to 6.5. The organic acids, or mixtures of organic acids, may be in an amount between about 0.01 wt% and about 8.0 wt% of the coating.

[0018] The antioxidants may be, for example, ascorbic acid, α -tocopherol, BHT-Alcohol and mixtures thereof. The antioxidants may be in an amount between about 0.01 wt% and about 1 wt% of the coating.

[0019] The peroxide generating enzyme may be any suitable enzyme that interacts with mannose to generate peroxide. Such enzymes include, for example, an oxidoreductase such as glucose oxidase or peroxidase. The peroxidase may be, for example, lactoperoxidase. The peroxide generating enzyme may be in an amount that interacts with mannose to generate an effective amount of peroxide. For example, the enzyme may be in an amount sufficient to catalyze the mannose so as to produce an anti-infective amount of peroxide over the shelf life of the medical product. In one embodiment, the enzyme may be in an amount between about 0.0005 wt% and about 0.5 wt% of the coating. In another embodiment, the enzyme may be below 0.01 wt%. In other embodiments, the enzyme could be between about 10 U active units per 100 grams dry weight of the other components of the coating and about 4000 U active units per 100 grams dry weight.

[0020] When a peroxide generating enzyme, such as glucose oxidase, is included in the coating, such enzyme may catalyse a suitable substrate (e.g. mannose) to generate hydrogen peroxide. The amount of peroxide generating enzyme may be sufficient so that the catalyzation may generate peroxide steadily over a relatively long period of time, resulting in a continuous release of peroxide that can provide anti-infective protection during storage and use of the catheter.

[0021] The hydrophilic coating containing mannose may be formed from a hydrophilic coating formulation, such as a solution or dispersion, containing the hydrophilic polymer(s), mannose and, optionally, other agents/additives. The hydrophilic coating formulation may be applied to the catheter shaft by, for example, dip coating, spraying, painting or any other suitable coating procedure. Depending on the desired coating, the coating formulation may then be dried and/or cured to form the hydrophilic coating on the catheter. The hydrophilic coating may be formed directly on the surface of the catheter shaft. Alternatively, a base/tie coating layer may first be formed on the surface of catheter shaft, and then the hydrophilic coating may be formed on the base layer. In one embodiment, the base coat layer and formulation include a hydrophilic polymer(s). The base coat layer may have good adhesion to both the surface of the catheter shaft and hydrophilic coating.

[0022] In one embodiment, the hydrophilic coating formulation (in the liquid state) includes

hydrophilic polymer(s) in an amount between about 0.9 wt% and about 20 wt% of the coating formulation;

mannose in an amount between about 0.1 wt% and about 5.0 wt% of the coating formulation; and

solvent in an amount between about 75 wt% and about 99 wt% of the coating formulation.

The solvent may be, for example, water, ethanol, methanol or any combination thereof or any other suitable solvent.

[0023] The hydrophilic coating formulation may optionally also include an effective amount of a peroxide generating enzyme that is effective to catalyse the mannose in the coating to generate peroxide during storage and use. In one embodiment, the peroxide generating enzyme may be in an amount between about 0.0005 wt% and about 0.25 wt% of the coating formulation. In another embodiment, the enzyme may be below 0.01 wt%. In another embodiment, the enzyme could be between about 10 U active units per 100 grams dry weight and about 1400 U active units per 100 grams dry weight. The amount of enzyme may be sufficient to yield between about 10 mg/100 g background hydrogen peroxide in the coating over time and about 25 mg/100 g background hydrogen peroxide in the coating over time. In yet another embodiment, the peroxide generating enzyme may be in an amount between about 0.075 wt% and about 0.5 wt% of the coating formulation. In yet another embodiment, the amount of peroxide could be between about 15 U active units per 100 grams dry weight and about 2800 U active units per 100 grams dry weight.

[0024] The coating may also optionally include a curing agent, such as benzophenone. The curing agent may be in an amount between about 0.01 wt% and about 1.0 wt%. The coating formulation may also optionally include an organic acid in an amount between about 0.1 wt% and about 8.0 wt%, and/or an antioxidant in an amount between about 0.05 wt% and about 1.0 wt%. The coating formulation may also optionally include any other suitable agents/additives, such as plasticizers, humectants, compounds that aid in attachment of the coating to a surface, and/or polyelectrolytes.

[0025] In one embodiment, the coating formulation includes

hydrophilic polymer(s), such as PVP, in an amount between about 0.9 wt% and about 20 wt% of the coating formulation;

mannose, such as a mixture of d-mannose and oligomannose, in an amount between about 0.1 wt% and about 5.0 wt% of the coating formulation;

a curing agent in an amount between about 0.1 wt% and about 1.0 wt% of the coating formulation;

a peroxide generating enzyme, such as glucose oxidase, in an amount between about 0.01 wt% and about 0.07 wt% of the coating formulation; or the enzyme may be below 0.01 wt%; or the enzyme between about 10 U active units per 100 ml solution) and about 1400 U active units per 100 ml solution; and

a solvent in an amount between about 73 wt% and about 98.8 wt% of the coating formulation.

[0026] Once the coating is formed on the catheter, the catheter may be packaged in a catheter assembly for storage, distribution and use by the end-user. The catheter assembly includes a package that contains the catheter and a wetting fluid that wets the hydrophilic coating to render it lubricious. The wetting fluid includes any suitable wetting agent that wets the hydrophilic coating. The wetting agent may be for example, water or saline. The hydrophilic coated catheter may be packaged in contact with the wetting fluid or the package includes separate compartments for the catheter and the wetting fluid wherein the compartments are brought into communication during the manufacturing process or just prior to use by the user.

[0027] In one embodiment, the catheter assembly includes a package having therein a catheter including a hydrophilic coating that has a hydrophilic polymer(s), mannose and, optionally, any of the above-mentioned additives/agents. In another embodiment, the package includes therein a catheter having a hydrophilic coating that has a hydrophilic polymer, mannose, a peroxide generating enzyme and, optionally, any other of the above-mentioned additives/agents.

[0028] In yet another embodiment, the catheter within the package includes a hydrophilic

coating that has a hydrophilic polymer and mannose and, optionally, any of the above-mentioned additives/agents. The package also includes a wetting fluid that includes a wetting agent, such as water or saline, and a peroxide generating enzyme. In this embodiment, when the wetting fluid wets the hydrophilic coating and enters the matrix of the hydrophilic coating, the peroxide generating enzyme will enter the matrix and catalyze the mannose to generate peroxide while the catheter is stored within the package and during use by the user.

[0029] In yet another embodiment, the catheter includes a hydrophilic coating that does not include any mannose when the coating is formed. Mannose may be introduced into the coating by contacting the coating with a fluid that includes mannose. Optionally, the fluid may also include a peroxide generating enzyme. For instance, after the hydrophilic coating is formed on the catheter and prior to placement in contact with the wetting fluid, the hydrophilic coating may be placed into contact with a solution that includes mannose. For example, the catheter including the hydrophilic coating formed thereon may be immersed in a carrier solution of mannose wherein the solution enters the polymer matrix of the hydrophilic coating, thereby carrying the mannose into the matrix to introduce the mannose into the matrix. In other embodiments, the carrier solution may also include the peroxide generating enzyme so that both the mannose and the peroxide generating are carried into and introduced into the matrix of the hydrophilic coating. After a sufficient amount of mannose and, when included peroxide generating enzyme, have been introduced into the coating, the catheter may be removed from the carrier solution and placed in a package. The package may optionally include a wetting solution.

[0030] The mannose may also be introduced into the polymer matrix of the hydrophilic coating on a catheter by contacting the coating with a wetting fluid that includes mannose. Optionally, the wetting fluid may also include a peroxide generating enzyme. The catheter may be placed in the package with a wetting fluid that includes mannose. When the wetting fluid wets the hydrophilic coating, it enters the matrix of the hydrophilic coating, carrying mannose with it. When the user removes the catheter for use, the hydrophilic catheter includes mannose therein, and peroxide generating enzyme when one is used, which provides an anti-infective effect during storage and use.

[0031] Thus, the present disclosure also relates to a wetting fluid including mannose, a wetting fluid including mannose and a peroxide generating enzyme, and a wetting fluid including a peroxide generating enzyme.

[0032] Additionally, the above described catheter assemblies may be sterilized in a suitable manner. For example, the catheter assemblies may be radiation sterilized with gamma or E-beam radiation. Furthermore, any of the packages disclosed above or discussed herein may be gas and liquid impermeable. Alternatively and depending on the application, the packages may be gas permeable.

[0033] Mannose and/or Sugar Alcohol Additives for Hydrophilic Formulations and Wetting Fluids

[0034] The present disclosure also relates to a mannose additive that may be added to a hydrophilic formulation or a wetting fluid. The mannose containing additive includes

mannose in an amount between about 75 wt% and about 85 wt% of the additive; and

solvent, such as water, in an amount between about 15 wt% and about 25 wt%.

The additive may also include other agents such as any of the above mentioned organic acids, antioxidants and/or peroxide generating enzyme. When included, the organic acids may be in an amount between about 0.1 wt% and about 8.0 wt% of the additive, the antioxidant may be in an amount between about 0.01 wt% and about 1.0 wt% of the additive, and the peroxide generating enzyme may be in an amount between about 0.0005 wt% and about 0.5 wt% of the coating formulation. In another embodiment, the enzyme may be below 0.01 wt%. In another embodiment, the enzyme could be between about 10 U active units per 100 grams dry weight and about 1400 U active units per 100 grams dry weight. The amount of enzyme may be sufficient to yield between about 10 mg/100 g background hydrogen peroxide in the coating over time and about 25mg/100 g background hydrogen peroxide in the coating over time. The additive may also include a thixotropic agent, such as one or more of pectin, agar, alginate or the like.

[0035] In addition to mannose, the additive includes a sugar alcohol, such as 3-carbon sugar alcohols (Glycerol); 4-carbon sugar alcohols (Erythritol, Threitol); 5-carbon sugar alcohols (Arabitol, Xylitol, Ribitol) sugar alcohols, 6-carbon sugar alcohols (Mannitol, Sorbitol, Galactitol, Fucitol, Iditol, Inositol), 7-carbon sugar alcohols (Volemitol), 12-carbon sugar alcohols (Isomalt, Maltitol, Lactitol), 18-carbon sugar alcohols (Maltotriitol), 24-carbon sugar alcohols (Maltotetraitol) and Polyglycitol.

[0036] The mannose or sugar alcohol containing additive may be added to a hydrophilic coating formulation that forms a hydrophilic coating. It may be added to a wetting fluid, or may be added to a carrier solution for introducing mannose or sugar alcohol into a hydrophilic coating. Thus, the present disclosure also relates to hydrophilic coatings and wetting fluids containing sugar alcohol or sugar alcohol and an enzyme. In one embodiment, an additive includes a hydrophilic polymer, mannose, and at least one sugar alcohol, such as xylitol for example. Optionally, this additive also includes one or more of inulins (such as, for example, inulin extracted from asteraceae plants or synthetic inulin), at least one peroxide generating enzyme and/or at least one organic acid or salts thereof. Furthermore, a hydrophilic coating or a wetting fluid also includes a hydrophilic polymer, mannose, and at least one sugar alcohols, such as xylitol for example. Optionally, this additive may also include one or more of inulins, at least one peroxide generating enzyme and/or at least one organic acid or salts thereof. Similar to as discussed above relative to mannose, the sugar alcohol or additives, such as those mentioned above, can be included in the coating by addition to the coating formulation that forms the coating or by infiltration into an already formed coating.

Hydrophilic Coatings and Wetting Fluids Containing Honey

[0037] Hygienic hydrophilic coatings of the present disclosure include a hydrophilic polymer, honey (natural or synthetic) and, optionally, other additives and agents. As used herein natural honey refers to honey made naturally by bees (whether considered active or not) and synthetic honey refers to substances that include substantially the same components of natural honeys but are man-made.

[0038] The hygienic hydrophilic coating includes

hydrophilic polymer(s) in an amount between about 70 wt% and about 99 wt% of the coating;
and

honey in an amount between about 1 wt% and about 30 wt% of the coating.

In one embodiment, the hygienic hydrophilic coating includes

hydrophilic polymer(s) in an amount between about 80 wt% and about 98 wt% of the coating;
and

honey in an amount of about 2 wt% and about 20 wt%.

The hygienic hydrophilic coating may optionally include other agents or additives, such as any of the above-mentioned organic acids, antioxidants and/or peroxide generating enzyme. The organic acids may be in an amount between about 0.01 wt% and about 8.0 wt% of the coating. The antioxidants may be in an amount between about 0.01 wt% and about 1.0 wt% of the coating. The peroxide generating enzyme may be in an amount between about 0.0005 wt% and about 0.5 wt% of the coating. In another embodiment, the enzyme may be below 0.01 wt%. In another embodiment, the enzyme could be between about 10 U active units per 100 grams dry weight and about 1400 U active units per 100 grams dry weight. The amount of enzyme may be sufficient to yield between about 10 mg/100 g background hydrogen peroxide in the coating over time and about 25 mg/100 g background hydrogen peroxide in the coating overtime.

[0039] The honey may be an inherently active honey that generates a background level of hydrogen peroxide. Alternatively, when a peroxide generating enzyme is included in the coating, such enzyme may catalyse the saccharides of the honey to generate peroxide, such as hydrogen peroxide. The amount of peroxide generating enzyme may be sufficient so that the catalyzation may generate peroxide over a relatively long period of time, resulting in a continuous release of peroxide that can provide anti-infective protection during storage and use of the catheter.

[0040] The hydrophilic coating containing honey may be formed from a hydrophilic coating formulation, such as a solution or dispersion, containing the hydrophilic polymer(s), honey and, optionally, other agents/additives. The hydrophilic coating formulation may be applied to the catheter shaft by any of the above discussed procedures. Depending on the desired coating,

the coating formulation may then be dried and/or cured to form the hydrophilic coating on the catheter. The hydrophilic coating may be formed directly on the surface of the catheter shaft or over a base/tie coating layer as described above.

[0041] In one embodiment, the hydrophilic coating formulation includes

hydrophilic polymer(s) in an amount between about 80 wt% and about 97 wt% of the coating formulation;

honey in an amount between about 1 wt% and about 20 wt% of the coating formulation; and

a solvent in an amount between about 80 wt% and about 99 wt% of the coating formulation.

The solvent may be, for example, water, ethanol, methanol or any combination thereof or any other suitable solvent. The curing agent may be, for example, benzophenone.

[0042] The hydrophilic coating formulation may, optionally, also include an additional effective amount of a peroxide generating enzyme that is effective to catalyse the saccharides of the honey to generate peroxide. In one embodiment, the peroxide generating enzyme may be in an amount between about 0.05 wt% and about 0.5 wt% of the coating formulation. In another embodiment, the enzyme may be below 0.01 wt%. In other embodiments, the enzyme could be between about 5 U active units per 100 grams dry weight of the other components of the coating and about 4000 U active units per 100 grams dry weight. The amount of enzyme may be sufficient to yield between about 10 mg/100 g background hydrogen peroxide in the coating over time and about 25 mg/100 g background hydrogen peroxide in the coating over time. In another embodiment, the enzyme could be between about 10 U active units per 100 grams dry weight and about 1400 U active units per 100 grams dry weight. The coating may also optionally include any of the above mentioned curing agents. The curing agent may be in an amount between about 0.1 wt% and about 1.0 wt%. The coating formulation may also optionally include an organic acid in an amount between about 0.01 wt% and about 8.0 wt%, and/or an antioxidant in an amount between about 0.05 wt% and about 1.0 wt%. The coating formulation may also optionally include any other suitable agents/additives, such as plasticizers, humectants, compounds that aid in attachment of the coating to a surface, and/or polyelectrolytes.

[0043] In one embodiment, the coating formulation includes

hydrophilic polymer(s), such as PVP, in an amount between about 0.9 wt% and about 20 wt% of the coating formulation;

honey in an amount between about 0.5 wt% and about 5.0 wt% of the coating formulation;

curing agent in an amount between about 0.1 wt% and about 1.0 wt% of the coating formulation;

peroxide generating enzyme, such as glucose oxidase, in an amount between 0.0005 wt% and about 0.5 wt% of the coating formulation; or the enzyme may be below 0.01 wt%; or the

enzyme could be between about 5 U active units per 100 grams dry weight of the other components of the coating and about 1400 U active units per 100 grams dry weight. The amount of enzyme may be sufficient to yield between about 10 mg/100 g background hydrogen peroxide in the coating over time and about 25 mg/100 g background hydrogen peroxide in the coating over time; and

solvent in an amount between about 73 wt% and about 98.5 wt% of the coating formulation.

[0044] Once the coating is formed on the catheter, the catheter may be packaged in a catheter assembly for storage, distribution and use by the end-user. The catheter assembly includes a package that contains the catheter and a wetting fluid that wets the hydrophilic coating to render it lubricious. The wetting fluid includes any suitable wetting agent that wets the hydrophilic coating. The wetting agent may be for example, water or saline. The hydrophilic coated catheter may be packaged in contact with the wetting fluid or the package includes separate compartments for the catheter and the wetting fluid wherein the compartments are brought into communication during the manufacturing process or just prior to use by the user.

[0045] In one embodiment, the catheter assembly includes a package having therein a catheter including a hydrophilic coating that has a hydrophilic polymer, honey and, optionally, any of the above-mentioned additives/agents. In another embodiment, the package includes therein a catheter having a hydrophilic coating that has a hydrophilic polymer, honey, a peroxide generating enzyme and, optionally, any other of the above-mentioned additives/agents.

[0046] In yet another embodiment the catheter within the package includes a hydrophilic coating that has a hydrophilic polymer and honey. The package also includes a wetting fluid that includes a wetting agent, such as water or saline, and a peroxide generating enzyme. In this embodiment, when the wetting fluid wets the hydrophilic coating and enters the matrix of the hydrophilic coating, the peroxide generating enzyme will enter the matrix and catalyze the saccharides of the honey to generate peroxide while the catheter is stored within the package and during use by the user.

[0047] In yet another embodiment, the catheter includes a hydrophilic coating that does not include any honey when the coating is formed. Honey may be introduced into the coating by contacting the coating with a fluid that includes honey. Optionally, the fluid may also include a peroxide generating enzyme. For instance, after the hydrophilic coating is formed and prior to placement in contact with the wetting fluid, the hydrophilic coating may be placed into contact with a solution that includes honey. For example, the catheter including the hydrophilic coating formed thereon may be immersed in a carrier solution including honey wherein the solution enters the polymer matrix of the hydrophilic coating, thereby carrying the components of the honey into the matrix to introduce the honey components into the matrix. In other embodiments, the carrier solution may also include the peroxide generating enzyme so that both the honey and the peroxide generating are carried into and introduced into the hydrophilic

coating. After a sufficient amount of honey and, when included peroxide generating enzyme, have been introduced into the coating, the catheter may be removed from the carrier solution and placed in a package. The package may optionally include a wetting solution.

[0048] The honey may also be introduced into the polymer matrix of the hydrophilic coating on a catheter by contact with a wetting fluid that includes honey. Optionally, the wetting fluid may also include a peroxide generating enzyme. The catheter may be placed in the package with a wetting fluid that includes honey. When the wetting fluid wets the hydrophilic coating, it enters the matrix of the hydrophilic coating, carrying the honey components with it. When the user removes the catheter for use, the hydrophilic catheter includes honey therein, and peroxide generating enzyme when one is used, which provides an anti-infective effect during storage and use.

[0049] Thus, the present disclosure also relates to a wetting fluid including honey and a wetting fluid including honey and a peroxide generating enzyme.

[0050] Additionally, the above described catheter assemblies may be sterilized in a suitable manner. For example, the catheter assembly may be radiation sterilized with gamma or E-beam radiation.

EXAMPLES

[0051] Hydrophilic coatings were formed on the outer surfaces of catheters made from polyvinyl chloride (PVC catheters). The catheters had a size of CH14 and a shore hardness of 82A. The hydrophilic coatings included a base coat layer formed on the outer surface of the catheter and a top coat layer formed over the base coat layer. The base coat composition was prepared as indicated below. The base coat composition was applied to the outer surface of each of the catheters and then cured and dried to form a base coat layer thereon.

[0052] The base coat composition included the following components:

Base Coat Composition

Component	Amount
Methanol	97.98% (w/w)
Polyvinylpyrrolidone K90 (PVP) (Ashland)	1.61% (w/w)
Irgacure 2959 (BASF)	0.01% (w/w)
Polyethylene glycol diacrylate (PEG400DA) (SR344, Sartomer, inhibitor removed)	0.40% (w/w)

[0053] The base coat composition was prepared by slowly adding the PVP to methanol while mixing until the PVP was dissolved. PEG400DA and Irgacure 2959 were then added and

allowed to fully dissolve while the solution was stirred.

[0054] Top coat compositions were prepared with the components as shown in the table below. Each of the catheters had one of the top coat compositions applied over the base coat layer. The top coat compositions were then cured and dried to form a hydrophilic coating on the outer surface of the catheter.

[0055] The formulations of the top coat compositions were as follows:

Top Coats

	Top Coat 1	Top Coat 2	Top Coat 3	Top Coat 4	Top Coat 5
Ethanol (Absolute)	62.41 g	62.41 g	62.41 g	62.41 g	62.41 g
De-ionized water	14.00 g	14.00 g	14.00 g	14.00 g	14.00 g
PVP K90 (Ashland)	5.95 g	5.95 g	5.95 g	5.95 g	5.95 g
BHT-A (Sigma Aldrich)	0.01 g	0.01 g	0.01 g	0.01 g	0.01 g
PEG400DA (SR344, Sartomer)	0.30 g	0.30 g	0.30 g	0.30 g	0.30 g
Glycerol (Sigma Aldrich)	0.73 g	0.73 g	0.73 g	0.73 g	0.73 g
Benzophenone (Sigma Aldrich)	0.01 g	0.01 g	0.01 g	0.01 g	0.01 g
d-Mannose (Sigma Aldrich)	0.20 g	2.00 g	---	---	---
Xylitol (Sigma Aldrich)	---	---	0.20 g	2.00 g	---
Natural Honey (Boyne Valley)	---	---	---	---	5.0g

[0056] Each of the top coat compositions were prepared by adding PVP to the ethanol and water and mixing until dissolved. The BHT-A, PEG400DA, glycerol, and benzophenone were then added and allowed to fully dissolve under stirring. Finally, d-mannose, Xylitol and Natural Honey, were added to the respective coating formulation and allowed to fully dissolve under stirring.

[0057] To form the hydrophilic coating on the outer surfaces of each of the catheters, the catheters were immersed in the base coat composition for a period of 10 seconds and then withdrawn at a rate of 0.7 cm/sec using a Harland PCX coating machine containing UV lamps. The base coat composition was then cured and dried under UV lamps for 45 seconds to form a base coat layer covering the outer surface of the catheter. The catheters were then immersed into one of the five top coat compositions for 10 seconds and withdrawn at a rate of 0.5 cm/sec. The top coat composition was then UV cured and dried under UV lamps for 10 minutes to form the top coat layer resulting in the formation of a hydrophilic coating on the catheter.

[0058] Catheters coated with each of the formulations were divided into two groups. The first group was tested in an unsterilized state and the second group was packaged in aluminium foil pouches, sealed and sterilized by gamma irradiation in the package at 30-35 kGy.

[0059] The unsterilized and sterilized initial and abraded coefficients of friction (CoFs) of catheter including Top Coats 1-4 were measured with the hydrophilic coating in a hydrated state. The sterilized initial, abraded and ten minute dry-out coefficients of friction (CoFs) of catheters including Top Coat 5 were also tested. When testing the CoFs, the catheters were immersed in water for 30 seconds to achieve a hydrated state. Additionally, a ten minute dry-out time CoF was measured for Top Coat 5.

[0060] CoF measurements are an indicator of lubricity and were measured using a Harland Friction Tester Model FTS5500. The CoFs of the catheters were determined by inserting a mandrel into 127 mm section of the coated catheter tube. The tube was then clamped between two pieces of silicone rubber at 100g load wherein the silicone rubber had a shore hardness of 60A. The catheter tube with the mandrel inserted therein was pulled through the two pieces of silicone rubber at a speed of 10 mm/s. The force required to pull about 80 mm of the catheter tube through the two pieces of silicone rubber was measured. The CoF value was calculated from the ratio of recorded to applied loads (i.e., the recorded load divided by the applied load) when steady state was reached. The CoF of each type of catheter was measured immediately after hydration ("initial"), immediately after being abraded ("abraded") and immediately after a ten-minute dry-out time ("dry-out").

[0061] In measuring the abraded CoFs, the catheter, with the hydrophilic coating in a hydrated state, was cycled back and forth 25 times through a hole in a 1 mm thick, silicone pad having a shore hardness of 60A. The hole was just smaller than the outer diameter of the catheter tube and the abrasion took place under water. Abrading the catheter in this fashion is designed to remove any portions of the coating that is not well adhered to the tubes. After abrasion, the CoF was measured as described above.

[0062] In measuring the ten minute dry-out time CoF, the catheter, immediately after hydration for "dry sterilized" catheter or immediately after removal from the package for "wet sterilized" catheters, was placed in an atmosphere having a temperature of 23°C and a relative humidity of 50% for 10 minutes before measuring the CoF.

Results

Coefficient of Friction Measurements

[0063] Table 1 shows the average CoFs for the initial and abraded CoFs for unsterilized and sterilized catheters coated with Top Coats 1-4.

Table 1

	Unsterilized		Sterilized	
	Initial CoF Avg.	Abraded CoF Avg.	Initial CoF Avg.	Abraded CoF Avg.
Top Coat 1	0.0253	0.0276	0.0166	0.0175
Top Coat 2	0.0215	0.0259	0.0135	0.0187
Top Coat 3	0.0227	0.0200	0.0190	0.0192
Top Coat 4	0.0301	0.0248	0.0127	0.0143

[0064] Table 2 shows the average CoFs for the initial, abraded and ten minute dry out CoFs for sterilized catheters coated with Top Coats 5.

Table 2

	Sterilized		
	Initial CoF Avg.	Abraded CoF Avg.	10 Min Dry-out Avg.
Top Coat 5	0.0122	0.0132	0.0143

Dye Uptake Tests

[0065] Dye uptake tests were conducted on the catheters post abrasion to assess the coating integrity and the level of adhesion/non-adhesion between the hydrophilic coatings and catheters. After the CoFs of the abraded catheters were measured, the catheters were dried-out (dehydrated). The dried-out catheters were then immersed in a water soluble red dye for 2 minutes. The catheters were then visually inspected to determine if the dye had been uniformly taken up throughout the coating or if sections of the coated portion of the catheter were dye-free. A uniform dye uptake throughout the coated portion of the catheter indicates that the hydrophilic coating has good integrity and good adhesion to the catheter. If the coated portion of the catheter has undyed sections, this is an indication that the hydrophilic coating or sections thereof have significantly thinned and/or separated from the catheter due to lack of adhesion to the catheter.

[0066] Referring to Figs. 1-9, visual inspection of catheters including hydrophilic coatings formed from Top Coats 1 through Top Coat 5, sterilized and unsterilized, show that the hydrophilic coatings exhibited uniform dye uptake which is an indication that the hydrophilic coating had strong adhesion to the catheter and good coating cohesion.

REFERENCES CITED IN THE DESCRIPTION

Cited references

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Non-patent literature cited in the description

- **ANALETTE I. LOPEZ et al.** Biofunctionalization of silicone polymers using poly(amidoamine) dendrimers and a mannose derivative for prolonged interference against pathogen colonization *BIOMATERIALS*, 2011, vol. 32, 194336-4346 [0003]

Patentkrav

1. Kateter, der omfatter:

en overflade; og

en befugtet smørende hydrofil belægning, der er anbragt på overfladen, hvor belægningen omfatter en hydrofil polymermatrix, og der er placeret mannose inden i den hydrofile polymermatrix.

2. Kateter ifølge krav 1, hvor den hydrofile polymer omfatter polyvinylpyrrolidon.

3. Kateter ifølge et hvilket som helst af de foregående krav, hvor belægningen yderligere omfatter et peroxidgenererende enzym i polymermatrixen.

4. Kateter ifølge krav 3, hvor enzymet omfatter en oxidoreduktase.

5. Kateter ifølge et hvilket som helst af kravene 3 og 4, hvor enzymet omfatter glucoseoxidase.

6. Kateter ifølge et hvilket som helst af kravene 3 og 4, hvor enzymet omfatter peroxidase.

7. Kateter ifølge krav 6, hvor peroxidasen omfatter lactoperoxidase.

8. Kateter ifølge et hvilket som helst af de foregående krav, hvor belægningen yderligere omfatter et tiksotropisk middel.

9. Kateter ifølge krav 8, hvor det tiksotropiske middel er én eller flere af pektin, agar og/eller alginat.

10. Kateter ifølge et hvilket som helst af de foregående krav, hvor belægningen yderligere indbefatter sukkeralkohol.

11. Kateter ifølge et hvilket som helst af de foregående krav, hvor belægningen yderligere omfatter en organisk syre.

12. Kateter ifølge et hvilket som helst af de foregående krav, hvor belægningen yderligere omfatter en antioxidant.

13. Kateter ifølge krav 12, hvor antioxidant omfatter ascorbinsyre, α -tocopherol, BHT-alkohol og blandinger deraf.

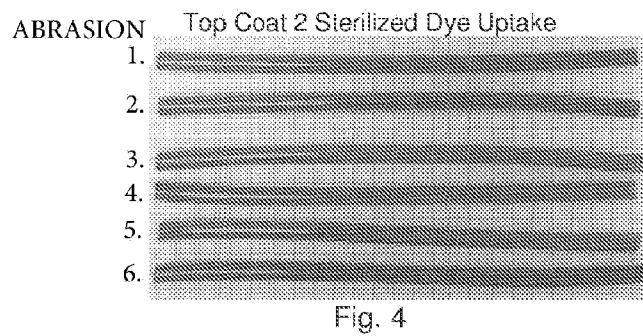
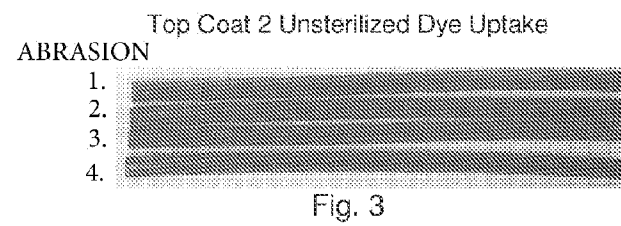
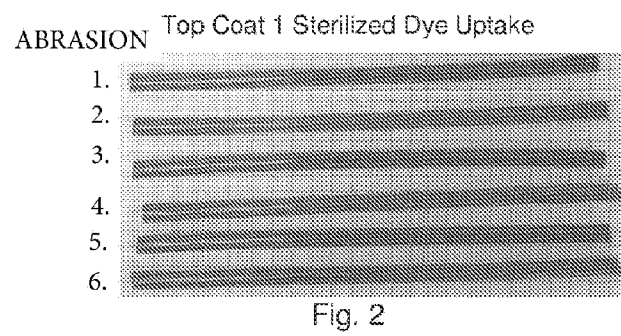
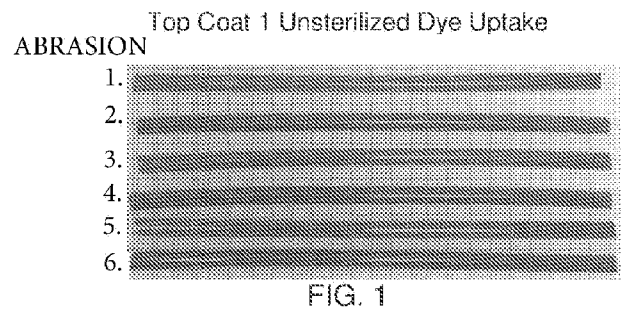
14. Fremgangsmåde til dannelse af et kateter ifølge krav 1, hvilken fremgangsmåde omfatter:

dannelse af en hydrofil belægning på en overflade af katetret;

og nedsænkning af katetret, der indbefatter den hydrofile belægning, i en opløsning, der indeholder mannose, hvor opløsningen bærer mannose ind i en polymermatrix af den hydrofile belægning.

15. Fremgangsmåde ifølge krav 14, hvor den hydrofile belægning yderligere indbefatter et peroxidgenererende enzym i polymermatrixen.

DRAWINGS



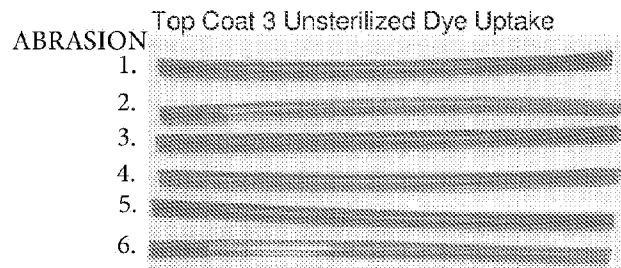


Fig. 5

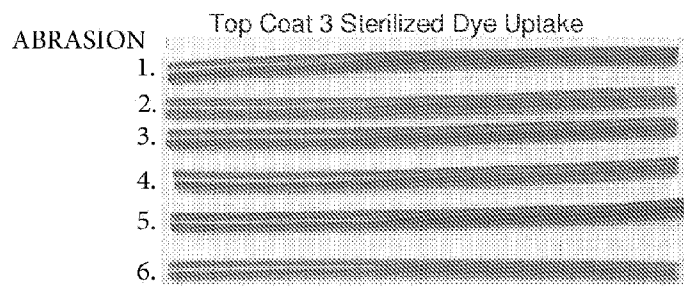


Fig. 6

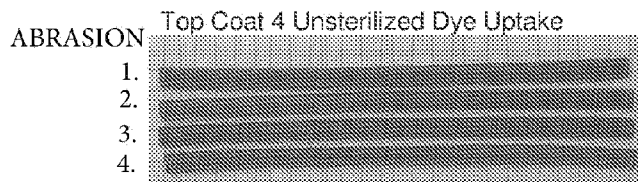


Fig. 7

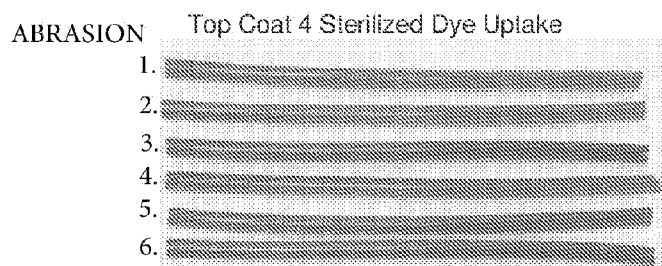


Fig. 8

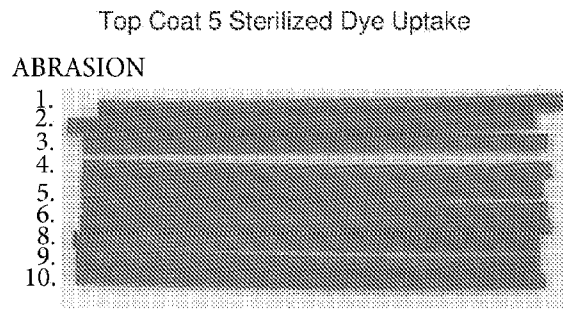


Fig. 9