



(51) International Patent Classification:

C12P 19/18 (2006.01) *C12P 19/44* (2006.01)
C07H 3/06 (2006.01) *C12N 1/21* (2006.01)
C07H 15/08 (2006.01)

(21) International Application Number:

PCT/DK2013/050306

(22) International Filing Date:

25 September 2013 (25.09.2013)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

PA 2012 70582 25 September 2012 (25.09.2012) DK
PA 2013 70311 7 June 2013 (07.06.2013) DK

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(81) Designated States (unless otherwise indicated, for every
kind of national protection available): AE, AG, AL, AM,
AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY,
BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM,
DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT,
HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KN, KP, KR,
KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME,
MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ,
OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA,
SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM,
TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM,
ZW.

(84) Designated States (unless otherwise indicated, for every
kind of regional protection available): ARIPO (BW, GH,
GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, SZ, TZ,
UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ,
TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK,
EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV,
MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM,
TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW,
KM, ML, MR, NE, SN, TD, TG).

Declarations under Rule 4.17:

— of inventorship (Rule 4.17(iv))

Published:

— with international search report (Art. 21(3))

(54) Title: GLYCOCONJUGATE SYNTHESIS

(57) Abstract: The invention relates to a method for producing a glycoconjugate comprising an oligosaccharide part covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, using a genetically modified cell.



WO 2014/048439 A1

GLYCOCONJUGATE SYNTHESIS

FIELD OF THE INVENTION

The present invention relates to a method of making glycoconjugates using genetically modified cells.

5 BACKGROUND OF THE INVENTION

In general, glycoconjugates are carbohydrate derivatives in which the carbohydrate part is linked to another chemical moiety by covalent linkage. Glycoconjugates like glycoproteins, glycopeptides, peptidoglycans, glycolipids, lipopolysaccharides play important roles in living systems, particularly in cell-cell or cell-matrix interactions.

- 10 Glycoproteins are proteins that contain sugar moiety/moieties, particularly oligosaccharide chain(s), covalently attached to the amino acid unit(s) of the polypeptide chain of a protein, preferably via the OH-group of an amino acid (serine, threonine, hydroxyproline) or the amide nitrogen of asparagine or glutamine (O- vs. N-glycosides). Glycoproteins are often parts of cell membranes. Glycopeptides are similar to glycoproteins in that the glycosyl
- 15 moiety is attached to an amino acid, but that particular amino acid can be a single aglycon or the part of an oligopeptide.

- Glycolipids are lipid (fatty acid) containing carbohydrate conjugates, generally classified in the groups of glyceroglycolipids, glycosphingolipids and glycosylphosphatidylinositols, with the role providing energy for the cell. Lipopolysaccharides, also lipid containing
- 20 glycoconjugates, but having antennary-like polysaccharide chains, are components of the outer membrane of bacteria and involved in inducing immune response.

- Preparing complex molecules such as glycoconjugates has always been a challenge for chemists and biologists. Structurally altered analogues of glycopeptides can be important targets in studying and understanding basic physiological processes taking place in cell
- 25 systems. Glycoconjugates and analogues thereof can be synthesized by both chemical and enzymatic ways. In chemical synthesis, the carbohydrate part has been reacted with the aglycon under glycosylation condition, in which both the participating educts shall be present in protected form, that is all functional groups that are not involved in forming the desired linkage and can be affected under glycosylation conditions shall be masked. This chemical
- 30 approach, in many cases, can suffer from low stereoselectivity with regard to the newly formed glycosidic bond, low overall yield due to the high number of protection/deprotection

steps, the use of sophisticated and often expensive purification methodologies such as column chromatography, and sometimes the use of toxic reagents. Alternatively, enzyme mediated glycosylation reactions can require fewer or even just a few elementary steps and/or take place with remarkable stereo- and/or regioselectivity, but the narrow substrate specificity of the enzymes towards the donors and the acceptors often requires the use of different enzymes depending on the structure of the target, which makes it practically impossible to develop a general methodology.

The production of some oligosaccharides having lactose, galactose or GlcNAc at the reducing terminus, and optionally their allyl glycosides, has been reported by using genetically modified *E. coli* cells which have been able to internalize lactose, allyl galactoside, allyl lactoside or GlcNAc-OAll added as exogenous precursors to the fermentation broth (WO 01/04341, Fort et al. *J. Chem. Soc., Chem. Comm.* 2558 (2005), WO 2007/101862, EP-A-1911850, WO 2010/070104, US 2012/208181).

SUMMARY OF THE INVENTION

The invention relates to a method for producing a glycoconjugate comprising an oligosaccharide part covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the method comprising:

(i) providing a genetically modified cell comprising a gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an acceptor molecule intracellularly,

(ii) culturing said genetically modified cell in the presence of an exogenous acceptor comprising a mono- or disaccharide part covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, so that

(a) said exogenous acceptor molecule is internalized by said genetically modified cell, and

(b) said glycoconjugate is formed from said internalized acceptor molecule by a glycosyl transfer mediated by said glycosyl transferase enzyme expressed by said cell, and then

(iii) separating said glycoconjugate from the culture medium.

Preferably, the mono- or disaccharide part of said exogenous acceptor is selected from galactose, glucose, N-acetyl-glucosamine and lactose, preferably lactose.

Also preferably, the recombinant glycosyl transferase enzyme is selected from the group consisting of N-acetyl-glucosaminyl transferases, galactosyl transferases, N-acetyl-

5 galactosaminyl transferases, glucuronosyl transferases, sialyl transferases and fucosyl transferases, more preferably β -1,3-N-acetyl-glucosaminyl transferase, β -1,6-N-acetyl-glucosaminyl transferase, β -1,3-galactosyl transferase, β -1,4-galactosyl transferase, β -1,3-N-acetyl-galactosaminyl transferase, β -1,3-glucuronosyl transferase, α -2,3-sialyl transferase, α -2,6-sialyl transferase, α -2,8-sialyl transferase, α -1,2-fucosyl transferase, α -1,3-fucosyl
10 transferase and α -1,4-fucosyl transferase.

Particularly, in the method of the invention the genetically modified cell is *E. coli* of LacZ^- , LacY^+ genotype, the exogenous acceptor consists of lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the
15 glycosyl transferase is a fucosyl transferase, and the glycoconjugate produced by the method is a fucosylated lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, preferably the fucosylated lactose moiety is selected from 2'-fucosyllactose, 3-fucosyllactose and difucosyllactose.

20 Also particularly, in the method of the invention the genetically modified cell is *E. coli* of LacZ^- , LacY^+ genotype, the exogenous acceptor consists of lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the glycosyl transferase is a sialyl transferase, and the glycoconjugate produced by
25 the method is a sialylated lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, preferably the sialylated lactose moiety is selected from 3'-sialyllactose and 6'-sialyllactose.

Yet particularly, in the method of the invention the genetically modified cell is *E. coli* of
30 LacZ^- , LacY^+ genotype, the exogenous acceptor consists of lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the glycosyl transferase is a sialyl transferase and a fucosyl transferase, and the glycoconjugate produced by the method is a sialylated and fucosylated lactose covalently
35 linked to a non-sugar moiety selected from the group consisting of amino acids, peptides,

proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, preferably the sialylated fucosylated lactose moiety is 3'-sialyl-3-fucosyllactose.

Yet particularly, in the method of the invention the genetically modified cell is *E. coli* of

- 5 LacZ⁻, LacY⁺ genotype, the exogenous acceptor consists of lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the glycosyl transferase is a N-acetylglucosaminyl transferase and optionally a galactosyl transferase, and the glycoconjugate produced by the method is a N-
- 10 acetylglucosaminylated lactose optionally substituted by galactosyl residue and covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, preferably the N-acetylglucosaminylated lactose moiety optionally substituted by galactosyl residue is 3'-N-acetylglucosaminyl lactose, LNT or LNnT.

- 15 The second aspect of the invention is to provide a glycoconjugate consisting of an oligosaccharide, preferably a human milk oligosaccharide, attached to an amino acid, preferably serine, threonine or hydroxyproline, more preferably via an O-glycosidic linkage.

The third aspect of the invention relates to providing a glycoconjugate consisting of an oligosaccharide, preferably a human milk oligosaccharide, attached to a polyethylene glycol

20 moiety via an O-glycosidic linkage.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with this invention, an "oligosaccharide part or residue of a glycoconjugate" preferably means an oligosaccharide residue having at least two monosaccharide units. The oligosaccharide residue can have a linear or branched structure containing monosaccharide

25 units that are linked to each other by interglycosidic linkages. The monosaccharides units can be any 5-9 carbon atom sugars that are aldoses (e.g. D-glucose, D-galactose, D-mannose, D-ribose, D-arabinose, L-arabinose, D-xylose, etc.), ketoses (e.g. D-fructose, D-sorbose, D-tagatose, etc.), deoxysugars (e.g. L-rhamnose, L-fucose, etc.), deoxy-aminosugars (e.g. N-acetylglucosamine, N-acetylmannosamine, N-acetylgalactosamine, etc.), uronic acids,

30 ketoaldonic acids (e.g. sialic acid) or equivalents. The glycoconjugate has an oligosaccharide part covalently attached to a non-sugar moiety (aglycon) by either a covalent bond, which is a linkage between any atom of the oligosaccharide residue and any atom of the non-sugar moiety, or by a linker, which consists of one, two, three or four atoms such as -O-, -C-, -NH-

, -N(OH)-, -S-, -C(=O)-, -C(=S)-, -C(=NH)-, -C(=N-OH)-, -C(=O)-O-, -O-C(=O)-, -C(=O)-S-, -S-C(=O)-, -C(=S)-O-, -O-C(=S)-, -C(=S)-S-, -S-C(=S)-, -C(=O)-NH-, -NH-C(=O)-, -C(=NH)-O-, -O-C(=NH)-, -C(=S)-NH-, -NH-C(=S)-, -C(=NH)-S- and -S-C(=NH).. Preferably, the C-1 (in case of aldoses) or C-2 (in the case of ketoses) anomeric carbon atom of the reducing end of the oligosaccharide residue is linked to the non-sugar moiety by a covalent bond or a linker, thus forming O-, N-, S- or C-glycosides. Similarly, an "exogenous acceptor having a mono- or disaccharide part covalently attached to a non-sugar moiety" means that the mono- or disaccharide moiety is linked to the non-sugar moiety (aglycon) by either a covalent bond or by a linker in the above defined manner. The monosaccharide residue or the monosaccharide units of the disaccharide moiety can be those defined above.

Also in this invention, a "glycoconjugate having an oligosaccharide part covalently attached to an amino acid" preferably means an oligosaccharide as defined above that is linked to an amino acid by a covalent bond as defined above. The amino acid can be any natural or non-natural amino acid, that is an alkanolic acid derivative having at least one amino group as a substituent. Preferably, the amino acid is selected from the group consisting of α -amino acids and β -amino acids such as Ala, Arg, Asn, Asp, Cys, Glu, Gln, Gly, His, Ile, Leu, Lys, Met, Phe, Pro, Ser, Thr, Trp, Tyr, Val, hydroxyproline, α -methylserine, β -alanine, etc. The above-mentioned amino acids can be either directly bound to the carbohydrate or a mono- or disaccharide part, or via a linker defined above (e.g. for urea-linked glycopeptides see WO 2009/040363). Also preferably, the C-1 (in case of aldoses) or C-2 (in the case of ketoses) anomeric carbon atom of the reducing end of the carbohydrate or a mono- or disaccharide part is linked to the amino acid thus forming O-, N-, S- or C-glycosides. O-Glycosides (O-glycans) can be formed involving OH-containing amino acids such as serine, threonine, hydroxyproline, etc., N-glycosides (N-glycans) can be made using the α -, β -, etc. amino group of any amino acid or the additional amino group of the side chain of e.g. lysine, asparagine or glutamine, while C-glycosides (C-glycans) contain a C-C bond coupling a C-atom of the amino acid to a C-atom of the oligosaccharide part, preferably to the anomeric carbon atom of the non-reducing end. Correspondingly, an "exogenous acceptor having a mono- or disaccharide part covalently attached to an amino acid" means a mono- or disaccharide defined above that is linked to an amino acid by a covalent bond as defined above.

Herein, a "glycoconjugate having an oligosaccharide part covalently attached to a peptide" or an "exogenous acceptor having a mono- or disaccharide part covalently attached to a peptide" preferably means an oligosaccharide or a mono- or disaccharide, respectively, defined above that is linked to an amino acid defined above by a covalent bond, provided that the amino acid is the part of an oligopeptide (also referred to as glycopeptides). The type of linkages can be similar to those mentioned above.

Also herein, a "glycoconjugate having an oligosaccharide part covalently attached to a protein" or an "exogenous acceptor having a mono- or disaccharide part covalently attached to a protein" preferably means a glycopeptide-like derivative wherein the amino acid attached to the carbohydrate moiety is a monomer unit of a protein; these compounds can be referred to as glycoproteins.

Also herein, a "glycoconjugate having an oligosaccharide part covalently attached to a lipid" preferably means an oligosaccharide defined above that is linked to a lipid by a covalent bond as defined above (also referred to as saccharolipids or glycolipids). "Lipid" denotes any organic compound, including fats, waxes, oils, sterols, fat-soluble vitamins (such as vitamins A, D, E and K), mono-, di- and triglycerides, sphingolipids and others, that are hydrophobic but soluble in alcohols, ether and most of other organic solvents; together with carbohydrates and protein they constitute basic and principal structural building blocks of living cells. Fatty acids are made of hydrocarbon chain having typically 4 to 28 carbon atoms, but preferably an even number of carbon atoms with a carboxyl group at the terminus (alkanoic acids), which carbon chain can be saturated or unsaturated one or more times and can be substituted by groups containing oxygen, halogen, nitrogen and/or sulfur. The double bond(s), if present, can be both cis or trans. Exemplary saturated fatty acids are caprylic acid, capric acid, lauric acid, myristic acid, palmitic acid, stearic acid, arachidic acid, behenic acid, lignoceric acid, cerotic acid, etc., and exemplary unsaturated fatty acids are miristoleic acid, palmitoleic acid, sapienic acid, oleic acid, elaidic acid, vaccenic acid, linoleic acid, linoelaidic acid, α -linolenic acid, arachidonic acid, eicosapentaenoic acid, erucic acid, docosahexaenoic acid, etc. Fats and oils are mostly mixture of fatty acids and/or their esters with glycerol, whereas waxes are generally esters of fatty acids with long chain alcohols. Mono-, di- and triglycerides are mono-, di- and triesters of fatty acids, respectively, with glycerol. Sphingolipids are lipid-like compounds containing a sphingoid long-chain hydroxyl amine base (e.g. sphingosine) that is coupled to a fatty acid molecule through an amide bond, thereby forming the ceramide unit. All of these lipids can be attached to a saccharidic moiety, especially via the anomeric carbon atom of the non-reducing end of the saccharidic moiety. The most important glycolipids are glyceroglycolipids, glycosphingolipids (including cerebrosides, gangliosides, globosides and glycoposphosphingolipids) and lipopolysaccharides. Likewise, an "exogenous acceptor having a mono- or disaccharide part covalently attached to a lipid" means a mono- or disaccharide as defined above that is linked to a lipid by a covalent bond as defined above.

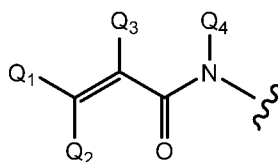
Also in this invention, a "glycoconjugate having an oligosaccharide part covalently attached to a polyethylene glycol" preferably means an oligosaccharide as defined above that is linked to the polyethylene glycol by a covalent bond as defined above. The polyethylene glycol (PEG) is a water soluble polyether of molecular formula $C_{2n}H_{4n+2}O_{n+1}$, having oxyethylene

(-CH₂-O-CH₂- or CH₂-CH₂-O-) repeating units and wherein n is 2 to 100, preferably 2 to 50, particularly 2 to 25, more particularly 2 to 10. Lower molecular weight PEGs are available in a purified form and are referred to as a "monodisperse PEG", and are also available as mixtures of PEGs and are referred to as a "polydisperse PEG". With regard to their geometry, PEGs can be in a linear, branched, star or comb configuration. Linear PEGs are preferably lower molecular weight PEGs (i.e., n is 2 to 10, preferably 3 to 6). Branched PEGs preferably have 3 to 10 linear, preferably lower molecular weight, PEG chains emanating from a central core group. Star PEGs preferably have 10 to 100 linear or branched, preferably lower molecular weight, PEG chains emanating from a central core group. Comb PEGs have multiple linear, branched and/or star, preferably lower molecular weight, PEG chains bonded to a polymer backbone. Terminal primary hydroxy group of PEGs can be bonded by an ether bond with an alkyl group, preferably methyl. In addition, their terminal hydroxy group can be replaced by amino, alkyl amino, dialkyl amino, acylamino, thiol or alkyl thio groups or their terminal hydroxymethyl group can be oxidized to a carboxyl, which can be esterified or be present in amide form with ammonia or a primary or secondary amine. PEGs can be attached to a saccharide unit at another one of its terminal ends. The attachment is preferably a glycosidic-like bond. Similarly, an "exogenous acceptor having a mono- or disaccharide part covalently attached to a PEG" means a mono- or disaccharide as defined above that is linked to a PEG by a covalent bond as defined above. Preferred PEGs are linear and of 2-10 repeating units.

Also herein, a "glycoconjugate having an oligosaccharide part covalently attached to polyvinyl alcohol" preferably means an oligosaccharide as defined above that is linked to a polyvinyl alcohol residue by a covalent bond as defined above. Polyvinyl alcohol (PVA) is a water-soluble polymer of molecular formula (C₂H₄O)_x having -CH₂-CH(OH)- monomer units. When attached to carbohydrate, either ends of the polymer chain can be glycosylated. Similarly, an "exogenous acceptor having a mono- or disaccharide part covalently attached to PVA" means a mono- or disaccharide as defined above that is linked to PVA by a covalent bond as defined above.

Also herein, a "longer alkyl chain" preferably means a linear or branched hydrocarbon chain having at least 6 carbon atoms, preferably 6-24 carbon atoms, that can be saturated or non-saturated. A "glycoconjugate having an oligosaccharide part covalently attached to a longer alkyl chain" or an "exogenous acceptor having a mono- or disaccharide part covalently attached to a longer alkyl chain" means that an above-defined hydrocarbon moiety is linked to an oligosaccharide defined above or a mono- or disaccharide defined above, respectively, by a covalent bond, preferably to the anomeric carbon atom of the carbohydrate in the form of an O-, N- or S-glycoside.

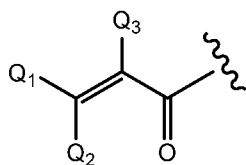
Also herein, an "α,β-unsaturated amido" preferably means a residue of formula **A**

**A**

wherein Q_1 , Q_2 , Q_3 and Q_4 are, independently, H and C_1 - C_6 -alkyl, which alkyl optionally can be substituted with halogen, OH, nitro or phenyl groups. A "glycoconjugate having an oligosaccharide part covalently attached to an α,β -unsaturated amido group" or an

- 5 "exogenous acceptor having a mono- or disaccharide part covalently attached to an α,β -unsaturated amido group" means that an above-defined residue of formula **A**, via its N atom, is linked to an oligosaccharide defined above or a mono- or disaccharide defined above, respectively, by a covalent bond, preferably to the anomeric carbon atom of the carbohydrate in the form of an N-glycoside.

- 10 Also herein, an " α,β -unsaturated carbonyl group" preferably means a residue of formula **B**

**B**

wherein Q_1 , Q_2 and Q_3 are as defined at residue of formula **A**.

Preferably, in the above defined glycoconjugates and mono- or disaccharide derivatives the non-sugar (or non-carbohydrate) moiety is directly attached to the anomeric carbon atom.

- 15 Also herein, the term "human milk oligosaccharides" means tri- and higher oligosaccharides found in human milk (see Urashima et al.: *Milk oligosaccharides* Nova Medical Books, NY, 2011).

- Also herein, a "genetically modified cell" preferably means a microorganism in which at least one alteration in its DNA sequence has been introduced into its genome in order that it has a particular phenotype. The alteration can result in a change in the original characteristics of the wild type cell, e.g., the modified cell is able to perform additional chemical transformation due to the introduced new genetic material that encodes the expression of an enzymes not being in the wild type cell, or is not able to carry out transformation like degradation due to
- 20

removal of gene/genes (knockout). A genetically modified cell can be produced in a conventional manner by genetic engineering techniques that are well-known to those skilled in the art.

In accordance with this invention, it has been surprisingly discovered that mono- and
5 disaccharide conjugates, namely mono- and disaccharides covalently attached to a non-sugar moiety, which non-sugar moiety is selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, preferably mono- and disaccharide conjugates containing a galactose residue, more preferably lactose conjugates can be internalized by a transport
10 mechanism involving permeases, allowing thus this carbohydrate precursors to be glycosylated in a genetically modified cell able to act so.

Also in accordance with this invention, a glycoconjugate having an oligosaccharide part covalently linked to a non-sugar moiety, which is selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated
15 amido group and polyvinyl alcohols, can be produced using a genetically modified cell, the method comprises:

(i) providing a genetically modified cell comprising one or more recombinant genes encoding one or more glycosyl transferase enzymes that are able to transfer the glycosyl residue of activated glycosyl nucleotides to an internalized acceptor molecule,

20 (ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently attached to a non-sugar moiety, which non-sugar moiety is selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, inducing

25 (a) internalization of the exogenous acceptor molecule by the genetically modified cell, and

(b) formation of the glycoconjugate of interest from the internalized acceptor molecule or glycosylated derivative thereof by one or more glycosyl transfers mediated by the one or more glycosyl transferase enzymes, and

(iii) collecting the glycoconjugate from the fermentation broth.

30 The genetically modified cell used in the method according to the present invention can be selected from the group of bacteria and yeasts, preferably the genetically modified cell stems

from a bacterium. Bacteria suitable for genetic modification preferably are selected from the group comprising *Escherichia coli*, *Bacillus* spp. (e.g. *Bacillus subtilis*), *Campylobacter pylori*, *Helicobacter pylori*, *Agrobacterium tumefaciens*, *Staphylococcus aureus*, *Thermophilus aquaticus*, *Azorhizobium caulinodans*, *Rhizobium leguminosarum*, *Neisseria gonorrhoeae*,
 5 *Neisseria meningitis*, *Lactobacillus* spp., *Lactococcus* spp., *Enterococcus* spp., *Bifidobacterium* spp., *Sporolactobacillus* spp., *Micromonospora* spp., *Micrococcus* spp., *Rhodococcus* spp., *Pseudomonas*, among which *E. coli* is of preferable choice. The genetically modified cell used in connection to the present invention preferably lacks of enzyme activity directed to degrade the acceptor molecule, glycosylated derivative thereof and the glycoconjugate of interest as
 10 product of the method.

The genetically modified cell used in the method according to the present invention comprises one or more endogenous or recombinant genes encoding one or more glycosyl transferase enzymes that are able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule. The gene or an equivalent DNA sequence
 15 thereof, if it is recombinant, is introduced into the cell by known techniques, using an expression vector. The origin of the heterologous nucleic acid sequence can be any animal (including human) or plant, eukaryotic cells such as those from *Saccharomyces cerevisiae*, *Saccharomyces pombe*, *Candida albicans* and the like, prokaryotic cells such as those originated from *E. coli*, *Bacillus subtilis*, *Campylobacter pylori*, *Helicobacter pylori*,
 20 *Agrobacterium tumefaciens*, *Staphylococcus aureus*, *Thermophilus aquaticus*, *Azorhizobium caulinodans*, *Rhizobium leguminosarum*, *Rhizobium meliloti*, *Neisseria gonorrhoeae* and *Neisseria meningitis*, or virus. The glycosyl transferase enzyme/enzymes expressed by the protein(s) encoded by the gene(s) or equivalent DNA sequence(s) are preferably glucosyl transferases, galactosyl transferases, N-acetylglucosaminyl transferases, N-
 25 acetylgalactosaminyl transferases, glucuronosyl transferases, xylosyl transferases, mannosyl transferases, fucosyl transferases, sialyl transferases and the like. In a preferred embodiment, the glycosyl transferases are selected from the group consisting of β -1,3-N-acetylglucosaminyl-transferase, β -1,6-N-acetylglucosaminyl-transferase, β -1,3-galactosyl-transferase, β -1,4-galactosyl-transferase, β -1,3-N-acetylgalactosaminyl-transferase, β -1,3-
 30 glucuronosyl-transferase, α -2,3-sialyl-transferase, α -2,6-sialyl-transferase, α -2,8-sialyl-transferase, α -1,2-fucosyl-transferase, α -1,3-fucosyl-transferase and α -1,4-fucosyl-transferase. More preferably, the glycosyl transferases are selected from β -1,3-N-acetylglucosaminyl transferase, β -1,6-N-acetylglucosaminyl transferase, β -1,3-galactosyl transferase, β -1,4-galactosyl transferase, α -2,3-sialyl transferase, α -2,6-sialyl transferase, α -
 35 1,2-fucosyl transferase, α -1,3-fucosyl transferase and α -1,4 fucosyl transferase. that is from those involved in the construction of HMO core structures as well as fucosylated and/or sialylated HMOs that are linked to a non-sugar moiety, which non-sugar moiety is selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups,

polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols. The genes encoding the above-mentioned transferases have been described in the literature.

In glycosyl transferase mediated glycosylation reactions activated sugar nucleotides serve as donors. The activated sugar nucleotides generally comprising a phosphorylated glycosyl residue attached to a nucleoside, the specific glycosyl transferase enzyme accept only the specific sugar nucleotide. Thus, preferably the following activated sugar nucleotides are involved in the glycosyl transfer: UDP-Glc, UDP-Gal, UDP-GlcNAc, UDP-GalNAc, UDP-glucuronic acid, UDP-Xyl, GDP-Man, GDP-Fuc and CMP-sialic acid, particularly those selected from the group consisting of UDP-Gal, UDP-GlcNAc, GDP-Fuc and CMP-sialic acid.

- 10 In one embodiment of the method, the genetically modified cell used in the method according to the present invention has one or more sets of genes encoding one or more enzymes responsible for the synthesis of one or more activated glycosyl nucleotide mentioned above by a *de novo* pathway. The sets of genes are either naturally present in the cell or introduced into the cell by means of gene technology or recombinant DNA techniques, all of them are parts of the general knowledge of the skilled person. Also in this embodiment, the genetically modified cell having one or more sets of genes encoding one or more enzymes responsible for the synthesis of one or more activated glycosyl nucleotide by a *de novo* pathway produces one or more activated glycosyl nucleotide, ready for glycosylation in glycosyl transferase mediated reaction in the cell, when cultured. The production of the activated glycosyl nucleotides by the cell takes place under the action of enzymes involved in the *de novo* biosynthetic pathway of that respective sugar nucleotide stepwise reaction sequence starting from a simple carbon source like glycerol, fructose or glucose (for a review for monosaccharide metabolism see e.g. H. H. Freeze and A. D. Elbein: *Chapter 4: Glycosylation precursors*, in: *Essentials of Glycobiology*, 2nd edition (Eds. A. Varki et al.), Cold Spring Harbour Laboratory Press (2009)).

According to the preferred embodiment disclosed above, the genetically modified cell is cultured in the presence of a carbon-based substrate such as glycerol, glucose, glycogen, fructose, maltose, starch, cellulose, pectin, chitin, etc. Preferably, the cell culturing is performed on glycerol and/or glucose and/or fructose.

- 30 In another embodiment, the genetically modified cell can utilize salvaged monosaccharide for producing activated sugar nucleotide. In the salvage pathway, monosaccharides derived from degraded oligosaccharides phosphorylated by kinases, and converted to nucleotide sugars by pyrophosphorylases. The enzymes involved in the procedure can be heterologous ones, or native ones of the cell used for genetic modification. Preferably, the synthesis of UDP-

arabinose, GDP-fucose or CMP-sialic acid can be accomplished using the salvage pathway when exogenous arabinose, fucose or sialic acid is also added to the culture.

It should be emphasized, that whatever way, either the *de novo*, or the salvage pathway taken for producing activated sugar nucleotides by the genetically modified cell is
5 advantageous compared to *in vitro* versions of transfer glycosylation, as it avoids using the very expensive sugar nucleotide type donors added exogenously, hence the donors are formed by the cell *in situ* and the phosphatidyl nucleoside leaving groups are recycled in the cell.

The method of this invention is also based on the way transporting an exogenous acceptor
10 molecule into the genetically modified cell for glycosylation thus producing glycoconjugates of interest. The internalization cannot affect the basic and vital functions or destroy the integrity of the cell. In one embodiment the internalization takes place via a passive transport mechanism during which the exogenous acceptor molecule diffuses passively across the plasma membrane of the cell. The flow is directed by the concentration difference in the
15 extra- and intracellular space with respect to the acceptor molecule to be internalized, which acceptor is supposed to pass from the place of higher concentration to the zone of lower concentration tending towards an equilibrium. In other embodiment the exogenous precursor is internalized with the aid of an active transport. In this case the genetically modified cell used in the method claimed comprises transporter proteins, called permeases, with which the
20 cell is able to admit exogenous substances and to concentrate them in the cytoplasm. Permeases act as enzymes, mention should be made of lactose permease (LacY) and sucrose permease (CscB). LacY has specificity towards galactose and simple galactosyl disaccharides, CscB is supposed to transport fructose and fructosyl disaccharides (see e.g. J. Sugihara et al. *Biochem.* **50**, 11009 (2011)). The specificity towards the sugar moiety of the substrate to be
25 internalized can be altered by mutation by means of known recombinant DNA techniques. In a preferred embodiment the internalization of the exogenous lactose derivative acceptor takes place via an active transport mechanism mediated by lactose permease.

During fermentation the exogenous substrate is internalized to and accumulated in the cell. The internalized substrate, acting as acceptor, participates in a glycosyl transferase induced
30 glycosylation reaction, in which a glycosyl residue of an activated nucleotide donor is transferred so that the acceptor is glycosylated. Optionally, when more than one glycosyl transferase is expressed by the cell, additional glycosylation reactions can occur resulting in the formation of the target glycoconjugate. Thus the expression "formation of the glycoconjugate of interest from the glycosylated derivative of the internalized acceptor
35 molecule" in describing the method to be claimed refers to the case when at least two, identical or different monosaccharide units are transferred to the acceptor added exogenously

and internalized by an active and/or passive mechanism by the cell, and the so glycosylated acceptor derivative is not yet intended to be the target glycoconjugate of interest, instead it is involved in further glycosylation reaction(s) in order to make the glycoconjugate of interest to be isolated. Of course, the cell preferably lacks any enzyme activity which would degrade the acceptor or the oligosaccharide derivatives produced in the cell.

At the end of incubation the glycoconjugate of interest as product of the method can be accumulated both in the intra- and the extracellular matrix. The product can be transported to the supernatant in a passive way, that is it diffuses outside across the cell membrane. The transport can be facilitated by sugar efflux transporters, proteins that promote the effluence of sugar derivatives from the cell to the supernatant. The sugar efflux transporter can be present exogenously or endogenously and is overexpressed under the conditions of the fermentation to enhance the export of the oligosaccharide derivative produced. The specificity towards the sugar moiety of the product to be secreted can be altered by mutation by means of known recombinant DNA techniques.

According to a preferred embodiment, the method also comprises the addition of an inducer to the culture medium. The role of the inducer is to promote the expression of enzymes involved in the *de novo* or salvage pathway and/or of permeases involved in the active transport and/or of sugar efflux transporters of the cell. Preferably, the inducer is isopropyl- β -D-thiogalactoside (IPTG).

In step (iii) of the method according to the present invention the glycoconjugate of interest is collected from the fermentation broth. According to a general manipulation, the supernatant containing the glycoconjugate of interest is separated from the cells by centrifugation. The separated cells are resuspended in water and subjected to heat and/or acid treatment in order to be permeabilized for releasing the glycoconjugates of interest piled up intracellularly.

The product is separated from the treated cell by centrifugation. The two supernatants containing the extra- and intracellular products, respectively, are combined and the products are purified and isolated by means of standard separation, purification and isolation techniques like gel and/or cationic ion exchange resin (H^+ form) chromatography. Preferably, the oligosaccharide derivative is collected only from the supernatant.

When carrying out a fermentation, a genetically modified cell, particularly a Lac Z^-Y^+ *E. coli* cell, is preferably cultured in the following way:

(a) a first phase of exponential cell growth ensured by a carbon-based substrate, and

(b) a second phase of cell growth limited by a carbon-based substrate which is added continuously.

Preferably, said carbon-based substrate is selected from the group consisting of glycerol and glucose. More preferably, the carbon-based substrate added during the second phase
5 glycerol.

Also preferably, said culturing is performed under conditions allowing the production of a culture with a high cell density.

Also preferably, said culturing further comprises a third phase of slowed cell growth obtained by continuously adding to the culture an amount of said carbon-based substrate that is less
10 than the amount of the carbon-based substrate added in said second phase so as to increase the content of the product produced in the high cell density culture.

Also preferably, the amount of the carbon-based substrate added continuously to the cell culture during said third phase is at least 30% less than the amount of the carbon-based substrate added continuously during said second phase.

15 Also preferably, the method further comprises the addition of an inducer to said culture medium to induce the expression in said cell of said enzyme and/or of a protein involved in said transport. The inducer is preferably isopropyl β -D-thiogalactoside (IPTG) and the protein is lactose permease.

The exogenous acceptor having a mono- or disaccharide part covalently attached to a non-sugar moiety, which non-sugar moiety is selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, to be internalized by and glycosylated in the fermented cell can be added to the culture medium at once or continuously. If added, it is done. The pure acceptor as solid/liquid or a concentrated aqueous solution of the acceptor can be added at
20 once at the end of the first phase of exponential cell growth, then the fermentation is continued by addition of the carbon-based substrate as described above. Alternatively, the continuous addition is beneficial when higher amount exogenous acceptor is intended to be used at a given volume. To avoid overflow metabolism and other side processes during the fermentation, the exogenous acceptor is dissolved in the feeding solution to be added during
25 the second (and optionally the third) phase, therefore a continuous addition of the acceptor (with the carbon-based substrate) is realized.
30

According to a first particular embodiment, it is provided a method for producing a glycoconjugate of interest having an oligosaccharide part covalently linked to an amino acid using a genetically modified cell, the method comprises:

- 5 (i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,
- (ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently linked to an amino acid inducing
 - 10 (a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and
 - (b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,
 - (iii) collecting the glycoconjugate from the culture medium.

15 The amino acid moiety is preferably selected from serine, threonine and hydroxyproline, more preferably serine.

The genetically modified cell can have more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the
20 internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred embodiment the exogenous acceptor is a mono- or disaccharide covalently linked, as defined above, to an amino acid, preferably serine, threonine or hydroxyproline, more preferably serine, and the glycoconjugate of interest is an
25 oligosaccharide covalently linked, as defined above, to an amino acid, preferably serine, threonine or hydroxyproline, more preferably serine, and they form an O-glycoside.

Also preferably with regard to the first particular embodiment, the method comprises:

- (i) providing a genetically modified cell comprising

a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor, which is selected from serine, threonine and hydroxyproline O-glycosides of a mono- or disaccharide, particularly a serine O-glycoside of a mono- or disaccharide, inducing

(a) internalization of said exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell,

(b) formation of the activated sugar nucleotide by a *de novo* pathway, and

(c) formation of a glycoconjugate selected from serine, threonine and hydroxyproline O-glycans, preferably a serine O-glycan, from said internalized acceptor molecule by a glycosyl transfers mediated by the glycosyl transferase enzyme,

(iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of genes encoding set of enzymes responsible for the synthesis of more than one activated sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

Also more preferably, the mono- or disaccharide part of said exogenous serine, threonine or hydroxyproline O-glycoside acceptor is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, particularly serine O-glycoside of lactose, the internalization of the above defined exogenous acceptor takes place via an active

transport mechanism mediated by a permease, preferably a lactose permease, and the product of the fermentation is selected from a serine, threonine and hydroxyproline O-glycosides of glycosylated galactose, glucose, N-acetyl-glucosamine and lactose, more preferably of glycosylated lactose, particularly serine O-glycoside of glycosylated lactose.

- 5 According to an even more preferable way to perform this first particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or at least one
- 10 galactosyl-transferase, and the glycoconjugate of interest produced by the method is a serine, threonine or hydroxyproline, preferably serine, O-glycoside of fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose, preferably of a human milk oligosaccharide.

Accordingly, the method even more preferably comprises:

- 15 (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
- a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and
- a set of genes encoding enzymes responsible for the synthesis of the activated fucose
- 20 nucleotide, preferably GDP-fucose, by a *de novo* pathway,
- (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor which is serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, inducing
- (a) internalization of the exogenous serine, threonine or hydroxyproline, preferably serine, O-
- 25 glycoside of lactose via an active transport mechanism by a lactose permease of the genetically modified cell,
- (b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

formation of a glycoconjugate from the internalized serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose by a fucosyl transfer mediated by the fucosyl transferase enzyme,

- 5 (iii) collecting the glycoconjugate that is a fucosylated serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, preferably serine, threonine or hydroxyproline, preferably serine, O-glycoside of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, from the culture medium.

Yet even more preferably, the method comprises:

- (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
- 10 a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and
- a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,
- 15 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, inducing
- (a) internalization of the exogenous serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose via an active transport mechanism by a lactose permease of the
- 20 genetically modified cell,
- (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and
- (c) formation of a glycoconjugate from the internalized serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose by a sialyl transfer mediated by the sialyl
- 25 transferase enzyme,
- (iii) collecting the glycoconjugate that is a sialylated serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, preferably serine, threonine or hydroxyproline, preferably serine, O-glycoside of 3'-sialyllactose or 6'-sialyllactose, from the culture medium.

Yet even more preferably, the method comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, inducing

(a) internalization of the exogenous serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

(c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the glycoconjugate that is a sialylated and fucosylated serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, preferably serine, threonine or

hydroxyproline, preferably serine, O-glycoside of 3'-sialyl-3-fucosyllactose, from the culture medium.

Yet even more preferably, the method comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

- 5 a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized
10 acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

optionally a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

- 15 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, inducing

(a) internalization of the exogenous serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose via an active transport mechanism by a lactose permease of the
20 genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

(c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

- 25 (d) formation of the glycoconjugate of interest from the internalized serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose by a N-acetylglucosaminyl transfer

mediated by the N-acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfer mediated by the galactosyl transferase enzyme,

(iii) collecting the glycoconjugate that is an N-acetylglucosaminylated serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose optionally substituted by galactose, preferably serine, threonine or hydroxyproline, preferably serine, O-glycoside of 3'-N-acetylglucosaminyl lactose, LNT or LNnT, from the culture medium.

The most preferred way to carry out the first particular embodiment of the invention comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

10 a recombinant gene encoding an α -1,2-fucosyl transferase, and

a set of genes encoding enzymes responsible for the synthesis of GDP-fucose by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, inducing

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(a) internalization of the exogenous serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of GDP-fucose by a *de novo* pathway, and

20 (c) formation of a glycoconjugate from the internalized serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose by a fucosyl transfer mediated by the α -1,2-fucosyl transferase,

(iii) collecting the glycoconjugate that is a serine, threonine or hydroxyproline, preferably serine, O-glycoside of 2'-fucosyllactose, from the culture medium.

25 According to a second particular embodiment, it is provided a method for producing a glycoconjugate of interest having an oligosaccharide part covalently linked to a peptide using a genetically modified cell, the method comprises:

(i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,

5 (ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently linked to a peptide inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and

(b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,

10 (iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the
15 internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred embodiment the exogenous acceptor that is a mono- or disaccharide part covalently linked to a peptide as defined above, and the glycoconjugate of interest that is an oligosaccharide part covalently linked to a peptide as defined above.

20 Also preferably with regard to the second particular embodiment, the method comprises:

(i) providing a genetically modified cell comprising

a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

25 a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor that is a mono- or disaccharide part covalently linked to a peptide inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell,

5 (b) formation of the activated sugar nucleotide by a *de novo* pathway, and

(c) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfers mediated by the glycosyl transferase enzyme,

(iii) collecting the glycoconjugate from the culture medium.

10 The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of genes encoding set of enzymes responsible for the synthesis of more than one activated sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the
15 internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

20 Also more preferably, the mono- or disaccharide part of the exogenous acceptor molecule is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, the internalization of the above defined exogenous acceptor takes place via an active transport mechanism mediated by a permease, preferably a lactose permease, and the product of the fermentation is selected from a glycosylated galactose, glucose, N-acetyl-glucosamine and lactose covalently linked to a peptide, more preferably glycosylated lactose
25 covalently linked to a peptide.

According to an even more preferable way performing this second particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is lactose covalently linked to a peptide, the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or at least one galactosyl-transferase, and the
30 glycoconjugate of interest produced by the method is a fucosylated and/or sialylated and/or

N-acetylglucosaminylated and/or galactosylated lactose, preferably a human milk oligosaccharide, covalently linked to a peptide.

Accordingly, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

- 5 a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

- 10 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a peptide inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

- 15 (b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the glycoconjugate that is a fucosylated lactose, preferably 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, covalently linked to a peptide from the culture medium.

- 20 Moreover, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a peptide inducing

5 (a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

10 (c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme,

(iii) collecting the glycoconjugate that is a sialylated lactose, preferably 3'-sialyllactose or 6'-sialyllactose, covalently linked to a peptide from the culture medium.

In addition, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

15 a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

20 a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

25 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a peptide inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

5 (c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

10 (iii) collecting the glycoconjugate that is a sialylated fucosylated lactose, preferably 3'-sialyl-3-fucosyllactose, covalently linked to a peptide from the culture medium.

Furthermore, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

15 a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

20 a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

25 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a peptide inducing

(a) internalization of the exogenous lactose derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

5 (c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

(d) formation of the glycoconjugate of interest from the internalized lactose derivative by a N-acetylglucosaminyl transfer mediated by the N-acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfers mediated by the galactosyl transferase enzyme,

10 (iii) collecting the glycoconjugate that is an N-acetylglucosaminylated lactose optionally substituted by galactose and covalently linked to a peptide, preferably 3'-N-acetylglucosaminyl lactose, LNT or LNnT covalently linked to a peptide from the culture medium.

According to a third particular embodiment, it is provided a method for producing a
15 glycoconjugate of interest having an oligosaccharide part covalently linked to a protein using a genetically modified cell, the method comprises:

(i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,

20 (ii) culturing the genetically modified cell in the presence of an exogenous acceptor comprising a mono- or disaccharide part covalently linked to a protein inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and

(b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl
25 transfer mediated by the glycosyl transferase enzyme expressed by the cell,

(iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred embodiment the exogenous acceptor that is a mono- or disaccharide part covalently linked to a protein as defined above, and the glycoconjugate of interest that is an oligosaccharide part covalently linked to a protein as defined above.

Also preferably with regard to the third particular embodiment, the method comprises:

(i) providing a genetically modified cell comprising

a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor that is a mono- or disaccharide part covalently linked to a protein inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell,

(b) formation of the activated sugar nucleotide by a *de novo* pathway, and

(c) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfers mediated by the glycosyl transferase enzyme,

(iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor

molecule or glycosylated derivative thereof, as well as it can comprise more than one set of genes encoding set of enzymes responsible for the synthesis of more than one activated sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

Also more preferably, the mono- or disaccharide part of said exogenous acceptor molecule is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, the internalization of the above defined exogenous acceptor takes place via an active transport mechanism mediated by a permease, preferably a lactose permease, and the product of the fermentation is selected from a glycosylated galactose, glucose, N-acetyl-glucosamine and lactose covalently linked to a protein, more preferably glycosylated lactose covalently linked to a protein.

According to an even more preferable way performing this third particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is lactose covalently linked to a protein, the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or at least one galactosyl-transferase, and the glycoconjugate of interest produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose, preferably a human milk oligosaccharide, covalently linked to a protein.

Accordingly, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a protein inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

- 5 (c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the glycoconjugate that is a fucosylated lactose, preferably 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, covalently linked to a protein from the culture medium.

Moreover, the method most preferably comprises:

- 10 (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and

- 15 a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a protein inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

- 20 (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

(c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme,

- 25 (iii) collecting the glycoconjugate that is a sialylated lactose, preferably 3'-sialyllactose or 6'-sialyllactose, covalently linked to a protein from the culture medium.

In addition, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of said activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing said genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a protein inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

(c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the glycoconjugate that is a sialylated fucosylated lactose, preferably 3'-sialyl-3-fucosyllactose, covalently linked to a protein from the culture medium.

Furthermore, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

- 5 optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

- 10 a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a protein inducing

- 15 (a) internalization of the exogenous lactose derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

(c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

- 20 (d) formation of the glycoconjugate of interest from the internalized lactose derivative by a N-acetylglucosaminyl transfer mediated by the N-acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfers mediated by the galactosyl transferase enzyme,

(iii) collecting the glycoconjugate that is an N-acetylglucosaminylated lactose optionally substituted by galactose and covalently linked to a protein, preferably 3'-N-

- 25 acetylglucosaminyl lactose, LNT or LNnT covalently linked to a protein from the culture medium.

According to a fourth particular embodiment, it is provided a method for producing a glycoconjugate of interest having an oligosaccharide part covalently linked to a lipid using a genetically modified cell, the method comprises:

- 5 (i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,
- (ii) culturing the genetically modified cell in the presence of an exogenous acceptor comprising a mono- or disaccharide part covalently linked to a lipid inducing
 - 10 (a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and
 - (b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,
- (iii) collecting the glycoconjugate from the culture medium.

- 15 The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

- 20 In a more preferred embodiment the exogenous acceptor is a mono- or disaccharide part covalently linked to a lipid as defined above, and the glycoconjugate of interest is an oligosaccharide part covalently linked to a lipid as defined above.

Also preferably with regard to the fourth particular embodiment, the method comprises:

- (i) providing a genetically modified cell comprising
 - 25 a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor that is a mono- or disaccharide part covalently linked to a lipid inducing

5 (a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell,

(b) formation of the activated sugar nucleotide by a *de novo* pathway, and

(c) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfers mediated by the glycosyl transferase enzyme,

10 (iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of

15 genes encoding set of enzymes responsible for the synthesis of more than one activated sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

20 In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

Also more preferably, the mono- or disaccharide part of the exogenous acceptor molecule is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, the internalization of the above defined exogenous acceptor takes place via an active transport mechanism mediated by a permease, preferably a lactose permease, and the

25 product of the fermentation is selected from a glycosylated galactose, glucose, N-acetyl-glucosamine and lactose covalently linked to a lipid, more preferably glycosylated lactose covalently linked to a lipid.

According to an even more preferable way performing this fourth particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the

exogenous acceptor is lactose covalently linked to a lipid, the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or at least one galactosyl-transferase, and the glycoconjugate of interest produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose, preferably a human milk oligosaccharide, covalently linked to a lipid.

Accordingly, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a lipid inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the glycoconjugate that is a fucosylated lactose, preferably 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, covalently linked to a lipid from the culture medium.

Moreover, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and

5 a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a lipid inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

10 (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

(c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme,

15 (iii) collecting the glycoconjugate that is a sialylated lactose, preferably 3'-sialyllactose or 6'-sialyllactose, covalently linked to a lipid from the culture medium.

In addition, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

20 a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

25 a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a lipid inducing

5 (a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

10 (c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

15 (iii) collecting the glycoconjugate that is a sialylated fucosylated lactose, preferably 3'-sialyl-3-fucosyllactose, covalently linked to a lipid from the culture medium.

Furthermore, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

20 a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

25 a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to a lipid inducing

5 (a) internalization of the exogenous lactose derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

10 (c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

(d) formation of the glycoconjugate of interest from the internalized lactose derivative by a N-acetylglucosaminyl transfer mediated by the N-acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfers mediated by the galactosyl transferase enzyme,

15 (iii) collecting the glycoconjugate that is an N-acetylglucosaminylated lactose optionally substituted by galactose and covalently linked to a lipid, preferably 3'-N-acetylglucosaminyl lactose, LNT or LNnT covalently linked to a lipid from the culture medium.

According to a fifth particular embodiment, it is provided a method for producing a glycoconjugate of interest having an oligosaccharide part covalently linked to a longer alkyl group using a genetically modified cell, the method comprises:

20 (i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently linked to a longer alkyl group inducing

25 (a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and

(b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,

(iii) collecting the glycoconjugate from the culture medium.

5 The longer alkyl group preferably consists of 6-24 carbon atoms, is preferably linear and saturated, and forms with the carbohydrate part of the acceptor and the glycoconjugate, to which it is attached, an O-glycoside. More preferably, the longer alkyl group consists of 6-20, even more preferably 6-14, particularly 6-10 carbon atoms.

10 The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

15 In a more preferred embodiment the exogenous acceptor is a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a mono- or disaccharide, and the glycoconjugate produced is a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of an oligosaccharide.

Also preferably with regard to the fifth particular embodiment, the method comprises:

(i) providing a genetically modified cell comprising

20 a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

25 (ii) culturing the genetically modified cell in the presence of an exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a mono- or disaccharide, inducing

(a) internalization of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a mono- or disaccharide via an active and/or passive transport mechanism by the genetically modified cell,

(b) formation of the activated sugar nucleotide by a *de novo* pathway, and

5 (c) formation of the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate from the internalized C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a mono- or disaccharide by a glycosyl transfers mediated by the glycosyl transferase enzyme,

10 (iii) collecting the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of
15 genes encoding set of enzymes responsible for the synthesis of more than one activated sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

20 In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

Also more preferably, the mono- or disaccharide part of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside acceptor is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, the internalization of the above defined exogenous acceptor takes place via an active transport mechanism mediated
25 by a permease, preferably a lactose permease, and the product of the fermentation is selected from a glycosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of galactose, glucose, N-acetyl-glucosamine and lactose, more preferably glycosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose.

30 According to an even more preferable way to perform this fifth particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside

of lactose, the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or at least one galactosyl-transferase, and the glycoconjugate produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, preferably of a human milk oligosaccharide.

Accordingly, the method even more preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor which is C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, inducing

(a) internalization of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(c) formation of a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate from the internalized C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate which is a fucosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, particularly C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-

alkyl O-glycoside of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, from the culture medium.

Yet even more preferably, the method comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

- 5 a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

- 10 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor which is a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, inducing

- (a) internalization of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose via an active transport mechanism by a lactose permease of the
15 genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

- (c) formation of a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate from the internalized C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose by a sialyl transfer mediated by the sialyl transferase enzyme,
20

(iii) collecting the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate which is a sialylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, particularly C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of 3'-sialyllactose or 6'-sialyllactose, from the culture medium.

- 25 Yet even more preferably, the method comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

5 a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

10 a set of genes encoding enzymes responsible for the synthesis of said activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, inducing

15 (a) internalization of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

20 (c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate from the internalized C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

25 (iii) collecting the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate which is a sialylated and fucosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, particularly C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of 3'-sialyl-3-fucosyllactose, from the culture medium.

Yet even more preferably, the method comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

optionally a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, inducing

(a) internalization of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

(c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

(d) formation of the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate from the internalized C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose by a N-acetylglucosaminyl transfer mediated by the N-acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfers mediated by the galactosyl transferase enzyme,

- (iii) collecting the C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of a glycoconjugate which is an N-acetylglucosaminylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose optionally substituted by galactose, particularly C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of 3'-N-acetylglucosaminyl lactose, LNT or LNnT, from the culture medium.

The most preferred way to carry out the fifth particular embodiment of the invention comprises:

- (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
- a recombinant gene encoding an α-1,2-fucosyl transferase, and
- 10 a set of genes encoding enzymes responsible for the synthesis of GDP-fucose by a *de novo* pathway,
- (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose, inducing
- 15 (a) internalization of the exogenous C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose via an active transport mechanism by a lactose permease of the genetically modified cell,
- (b) formation of GDP-fucose by a *de novo* pathway, and
- (c) formation of a glycoconjugate from the internalized C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of lactose by a fucosyl transfer mediated by the α-1,2-fucosyl transferase,
- 20 (iii) collecting the glycoconjugate which is a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀-alkyl O-glycoside of 2'-fucosyllactose, from the culture medium.

- According to a sixth particular embodiment, it is provided a method for producing a
- 25 glycoconjugate of interest having an oligosaccharide part covalently linked to polyethylene glycol using a genetically modified cell, the method comprises:

(i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,

5 (ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently linked to polyethylene glycol inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and

(b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,

10 (iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the
15 internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred embodiment the exogenous acceptor is a mono- or disaccharide part covalently linked to polyethylene glycol as defined above, and the glycoconjugate of interest is of an oligosaccharide part covalently linked to polyethylene glycol as defined above.

20 Also preferably with regard to the sixth particular embodiment, the method comprises:

(i) providing a genetically modified cell comprising

a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

25 a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor that is a mono- or disaccharide part covalently linked to polyethylene glycol inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell,

5 (b) formation of the activated sugar nucleotide by a *de novo* pathway, and

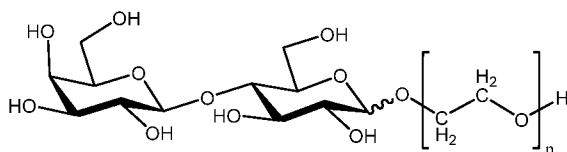
(c) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfers mediated by the glycosyl transferase enzyme,

(iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one
 10 recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of genes encoding set of enzymes responsible for the synthesis of more than one activated
 15 sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

Also more preferably, the mono- or disaccharide part of the exogenous acceptor molecule is
 20 selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose. In this regard, a preferred exogenous acceptor can be characterized by formula **7**

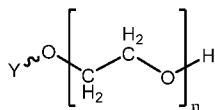


7

wherein n is an integer from 1 to 10, preferably from 2 to 6.

According to an even more preferable way to perform this sixth particular embodiment of the
 25 present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the

- exogenous acceptor is a compound of formula **7**, the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase, and/or at least one galactosyl-transferase and the glycoconjugate produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated compound of formula **7**, preferably polyethylene glycol O-glycoside of a human milk oligosaccharide of formula **7a**

**7a**

wherein Y is a human milk oligosaccharide glycosyl residue, and

n is as defined above.

- Accordingly, the method even more preferably comprises:
- (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
 - a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and
 - a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,
 - (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **7**, inducing
 - (a) internalization of the exogenous lactose derivative of formula **7** via an active transport mechanism by a lactose permease of the genetically modified cell,
 - (b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and
 - (c) formation of a glycoconjugate from the internalized lactose derivative of formula **7** by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting a fucosylated lactose derivative of formula **7**, particularly a corresponding polyethylene glycol O-glycoside of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, from the culture medium.

Yet even more preferably, the method comprises:

- 5 (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
- a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and
- a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid
10 nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,
- (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **7**, inducing
- (a) internalization of the exogenous lactose derivative of formula **7** via an active transport mechanism by a lactose permease of the genetically modified cell,
- 15 (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and
- (c) formation of a glycoconjugate from the internalized lactose derivative of formula **7** by a sialyl transfer mediated by the sialyl transferase enzyme,
- (iii) collecting the sialylated lactose derivative of formula **7**, particularly a corresponding
20 polyethylene glycol O-glycoside of 3'-sialyllactose or 6'-sialyllactose, from the culture medium.

Yet even more preferably, the method comprises:

- (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
- a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to
25 transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

5 a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of said activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **7**, inducing

10 (a) internalization of the exogenous lactose derivative of formula **7** via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

15 (c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized lactose derivative of formula **7** by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

20 (iii) collecting the sialylated and fucosylated lactose derivative of formula **7**, particularly the corresponding polyethylene glycol O-glycoside of 3'-sialyl-3-fucosyllactose, from the culture medium.

Yet even more preferably, the method comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

25 a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

optionally a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **7**, inducing

(a) internalization of the exogenous lactose derivative of formula **7** via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

(c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized lactose derivative of formula **7** by a N-acetylglucosaminyl transfer mediated by the N-acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfers mediated by the galactosyl transferase enzyme,

(iii) collecting the N-acetylglucosaminylated lactose derivative of formula **7** optionally substituted by galactose, particularly the corresponding polyethylene glycol O-glycoside of 3'-N-acetylglucosaminyl lactose, LNT or LNT, from the culture medium.

The most preferred way to carry out the sixth particular embodiment of the invention comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a recombinant gene encoding an α -1,2-fucosyl transferase, and

a set of genes encoding enzymes responsible for the synthesis of GDP-fucose by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **7**, inducing

5 (a) internalization of the exogenous lactose derivative of formula **7** via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of GDP-fucose by a *de novo* pathway, and

(c) formation of a glycoconjugate from the lactose derivative of formula **7** by a fucosyl transfer mediated by the α -1,2-fucosyl transferase,

10 (iii) collecting the corresponding polyethylene glycol O-glycoside of 2'-fucosyllactose, from the culture medium.

According to a seventh particular embodiment, it is provided a method for producing a glycoconjugate of interest having an oligosaccharide part covalently linked to an α,β -unsaturated amido group using a genetically modified cell, the method comprises:

15 (i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently linked to an α,β -unsaturated amido group inducing

20 (a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and

(b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,

(iii) collecting the glycoconjugate from the culture medium.

The α,β -unsaturated amido group is preferably a group of formula **A** defined above, more preferably Q_1 , Q_2 , Q_3 and Q_4 in group **A** are, independently, H and methyl, even more preferably H (that is group **A** is acrylamido).

- 5 The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.
- 10 In a more preferred embodiment the exogenous acceptor is a mono- or disaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q_1 , Q_2 , Q_3 and Q_4 in group **A** are, independently, H and methyl, even more preferably H, and the glycoconjugate produced is an oligosaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q_1 , Q_2 , Q_3 and Q_4 in group **A** are, independently, H and methyl, even more preferably H.
- 15 Also preferably with regard to the seventh particular embodiment, the method comprises:
- (i) providing a genetically modified cell comprising
- a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and
- 20 a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,
- (ii) culturing the genetically modified cell in the presence of an exogenous mono- or disaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q_1 , Q_2 , Q_3 and Q_4 in group **A** are, independently, H and methyl, even more preferably H, inducing
- 25 (a) internalization of the exogenous mono- or disaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q_1 , Q_2 , Q_3 and Q_4 in group **A** are, independently, H and methyl, even more preferably H via an active and/or passive transport mechanism by the genetically modified cell,
- (b) formation of the activated sugar nucleotide by a *de novo* pathway, and

(c) formation of an oligosaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q₁, Q₂, Q₃ and Q₄ in group **A** are, independently, H and methyl, even more preferably H, from the internalized mono- or disaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q₁, Q₂, Q₃ and Q₄ in group **A** are, independently, H and methyl, even more preferably H, by a glycosyl transfers mediated by the glycosyl transferase enzyme,

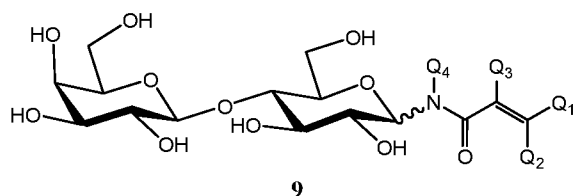
(iii) collecting the oligosaccharide attached to moiety **A** with its anomeric carbon atom, preferably Q₁, Q₂, Q₃ and Q₄ in group **A** are, independently, H and methyl, even more preferably H, from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of genes encoding set of enzymes responsible for the synthesis of more than one activated sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

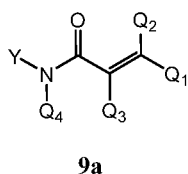
Also more preferably, the mono- or disaccharide part of the exogenous acceptor defined above is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, the internalization of the above defined exogenous acceptor takes place via an active transport mechanism mediated by a permease, preferably a lactose permease, and the product of the fermentation is a glycosylated galactose, glucose, N-acetyl-glucosamine or lactose, more preferably lactose, attached to moiety **A** with its anomeric carbon atom, preferably Q₁, Q₂, Q₃ and Q₄ in group **A** are, independently, H and methyl, even more preferably H.

According to an even more preferable way to perform this seventh particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is a compound of formula **9**



wherein Q_1 , Q_2 , Q_3 and Q_4 are, independently, H and methyl, even more preferably H,

the glycosyl transferase is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or one galactosyl-transferase, and the glycoconjugate produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated compound of formula **9**, preferably a compound of formula **9a**



wherein Y is a human milk oligosaccharide glycosyl residue, and

Q_1 , Q_2 , Q_3 and Q_4 are as defined above.

Accordingly, the method even more preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ^- , LacY^+ genotype comprising

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ^- , LacY^+ genotype in the presence of an exogenous lactose derivative of formula **9**, inducing

- (a) internalization of the exogenous lactose derivative of formula **9**, via an active transport mechanism by a lactose permease of the genetically modified cell,
- (b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and
- 5 (c) formation of a glycoconjugate from the internalized lactose derivative of formula **9** by a fucosyl transfer mediated by the fucosyl transferase enzyme,
- (iii) collecting the fucosylated lactose lactose derivative of formula **9**, particularly a compound of formula **9a**, wherein Y is a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, and Q₁, Q₂, Q₃ and Q₄ are as defined above, from the culture medium.
- 10 Yet even more preferably, the method comprises:
- (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
- a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and
- 15 a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,
- (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **9**, inducing
- (a) internalization of the exogenous lactose derivative of formula **9** via an active transport
- 20 mechanism by a lactose permease of the genetically modified cell,
- (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and
- (c) formation of a glycoconjugate from the internalized lactose derivative of formula **9** by a sialyl transfer mediated by the sialyl transferase enzyme,

(iii) collecting the sialylated lactose derivative of formula **9**, particularly a compound of formula **9a** wherein Y is a glycosyl residue of 3'-sialyllactose or 6'-sialyllactose, and Q₁, Q₂, Q₃ and Q₄ are as defined above, from the culture medium.

Yet even more preferably, the method comprises:

- 5 (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising
 - a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,
 - a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to
10 transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,
 - a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and
 - a set of genes encoding enzymes responsible for the synthesis of said activated fucose
15 nucleotide, preferably GDP-fucose, by a *de novo* pathway,
- (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **9**, inducing
 - (a) internalization of the exogenous lactose derivative of formula **9** via an active transport mechanism by a lactose permease of the genetically modified cell,
 - 20 (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,
 - (c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and
 - (d) formation of a glycoconjugate from the internalized lactose derivative of formula **9** by a
25 sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the sialylated and fucosylated lactose derivative of formula **9**, particularly a compound of formula **9a** wherein Y is a glycosyl residue of 3'-sialyl-3-fucosyllactose, and Q₁, Q₂, Q₃ and Q₄ are as defined above, from the culture medium.

Yet even more preferably, the method comprises:

- 5 (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme
10 that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

optionally a set of genes encoding enzymes responsible for the synthesis of the activated Gal
15 nucleotide, preferably UDP-Gal, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **9**, inducing

(a) internalization of the exogenous lactose derivative of formula **9** via an active transport mechanism by a lactose permease of the genetically modified cell,

20 (b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

(c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

(d) formation of a glycoconjugate by a N-acetylglucosaminyl transfer mediated by the N-
25 acetylglucosaminyl transferase enzyme optionally followed by a galactosyl transfer mediated by the galactosyl transferase enzyme,

(iii) collecting the N-acetylglucosaminylated lactose derivative of formula **9** optionally substituted by a galactose, particularly a compound of formula **9a** wherein Y is a glycosyl residue of LNT or LNnT, and Q₁, Q₂, Q₃ and Q₄ are as defined above, from the culture medium.

5 The most preferred way to carry out the seventh particular embodiment of the invention comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a recombinant gene encoding an α-1,2-fucosyl transferase, and

10 a set of genes encoding enzymes responsible for the synthesis of GDP-fucose by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous lactose derivative of formula **9**, inducing

(a) internalization of the exogenous lactose derivative of formula **9** by a lactose permease of the genetically modified cell,

15 (b) formation of GDP-fucose by a *de novo* pathway, and

(c) formation of a glycoconjugate from the internalized lactose derivative of formula **9** by a fucosyl transfer mediated by the α-1,2-fucosyl transferase,

(iii) collecting the compound of formula **9a** wherein Y is a glycosyl residue of 2'-fucosyllactose, and Q₁, Q₂, Q₃ and Q₄ are as defined above, from the culture medium.

20 According to a eighth particular embodiment, it is provided a method for producing a glycoconjugate of interest having an oligosaccharide part covalently linked to polyvinyl alcohol using a genetically modified cell, the method comprises:

(i) providing a genetically modified cell comprising a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an
25 activated sugar nucleotide to an internalized acceptor molecule,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor having a mono- or disaccharide part covalently linked to polyvinyl alcohol inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell, and

5 (b) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfer mediated by the glycosyl transferase enzyme expressed by the cell,

(iii) collecting the glycoconjugate from the culture medium.

The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to
10 transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred embodiment the exogenous acceptor is a mono- or disaccharide part
15 covalently linked to polyvinyl alcohol as defined above, and the glycoconjugate of interest is an oligosaccharide part covalently linked to polyvinyl alcohol as defined above.

Also preferably with regard to the eighth particular embodiment, the method comprises:

(i) providing a genetically modified cell comprising

a gene, preferably a recombinant gene encoding a glycosyl transferase enzyme that is able to
20 transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sugar nucleotide by a *de novo* pathway,

(ii) culturing the genetically modified cell in the presence of an exogenous acceptor that is a
25 mono- or disaccharide part covalently linked to polyvinyl alcohol inducing

(a) internalization of the exogenous acceptor molecule via an active and/or passive transport mechanism by the genetically modified cell,

(b) formation of the activated sugar nucleotide by a *de novo* pathway, and

(c) formation of the glycoconjugate from the internalized acceptor molecule by a glycosyl transfers mediated by the glycosyl transferase enzyme,

(iii) collecting the glycoconjugate from the culture medium.

- 5 The genetically modified cell can comprise more than one gene, preferably more than one recombinant gene, encoding more than one glycosyl transferase enzyme that are able to transfer the glycosyl residue of activated sugar nucleotides to an internalized acceptor molecule or glycosylated derivative thereof, as well as it can comprise more than one set of genes encoding set of enzymes responsible for the synthesis of more than one activated
- 10 sugar nucleotide by a *de novo* pathway, and the glycoconjugate is formed from the internalized acceptor by multiple glycosyl transfer mediated by multiple glycosyl transferases expressed by the cell.

In a more preferred realization the genetically modified cell lacks enzymes able to degrade the internalized acceptor molecule or glycosylated derivative thereof.

- 15 Also more preferably, the mono- or disaccharide part of the exogenous acceptor molecule is selected from galactose, glucose, N-acetyl-glucosamine and lactose, more preferably lactose, the internalization of the above defined exogenous acceptor takes place via an active transport mechanism mediated by a permease, preferably a lactose permease, and the product of the fermentation is selected from a glycosylated galactose, glucose, N-acetyl-
- 20 glucosamine and lactose covalently linked to polyvinyl alcohol, more preferably glycosylated lactose covalently linked to polyvinyl alcohol.

- According to an even more preferable way performing this eighth particular embodiment of the present invention, the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is lactose covalently linked to polyvinyl alcohol, the glycosyl transferase
- 25 is at least one fucosyl-transferase and/or at least one sialyl-transferase and/or at least one N-acetylglucosaminyl-transferase and/or one galactosyl-transferase, and the glycoconjugate of interest produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose, preferably a human milk oligosaccharide, covalently linked to polyvinyl alcohol.

- 30 Accordingly, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the fucosyl residue of an activated fucose nucleotide to an internalized acceptor molecule, and

5 a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to polyvinyl alcohol inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

10 (b) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a fucosyl transfer mediated by the fucosyl transferase enzyme,

15 (iii) collecting the glycoconjugate that is a fucosylated lactose, preferably 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, covalently linked to polyvinyl alcohol from the culture medium.

Moreover, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

20 a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule, and

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

25 (ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to polyvinyl alcohol inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

5 (c) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme,

(iii) collecting the glycoconjugate that is a sialylated lactose, preferably 3'-sialyllactose or 6'-sialyllactose, covalently linked to polyvinyl alcohol from the culture medium.

In addition, the method most preferably comprises:

10 (i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a sialyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

15 a gene, preferably a recombinant gene encoding a fucosyl transferase enzyme that is able to transfer the sialyl residue of an activated sialic acid nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway, and

20 a set of genes encoding enzymes responsible for the synthesis of the activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to polyvinyl alcohol inducing

(a) internalization of the exogenous lactose acceptor derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

25 (b) formation of an activated sialic acid nucleotide, preferably CMP-sialic acid, by a *de novo* pathway,

(c) formation of an activated fucose nucleotide, preferably GDP-fucose, by a *de novo* pathway, and

(d) formation of a glycoconjugate from the internalized lactose acceptor derivative by a sialyl transfer mediated by the sialyl transferase enzyme, and by a fucosyl transfer mediated by the fucosyl transferase enzyme,

(iii) collecting the glycoconjugate that is a sialylated fucosylated lactose, preferably 3'-sialyl-3-fucosyllactose, covalently linked to polyvinyl alcohol from the culture medium.

Furthermore, the method most preferably comprises:

(i) providing a genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype comprising

a gene, preferably a recombinant gene encoding a N-acetylglucosaminyl transferase enzyme that is able to transfer the N-acetylglucosaminyl residue of an activated GlcNAc nucleotide to an internalized acceptor molecule,

optionally a gene, preferably a recombinant gene encoding a galactosyl transferase enzyme that is able to transfer the galactosyl residue of an activated Gal nucleotide to an internalized acceptor molecule,

a set of genes encoding enzymes responsible for the synthesis of the activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway, and

a set of genes encoding enzymes responsible for the synthesis of the activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway,

(ii) culturing the genetically modified *E. coli* cell of LacZ⁻, LacY⁺ genotype in the presence of an exogenous acceptor that is lactose covalently linked to polyvinyl alcohol inducing

(a) internalization of the exogenous lactose derivative via an active transport mechanism by a lactose permease of the genetically modified cell,

(b) formation of an activated GlcNAc nucleotide, preferably UDP-GlcNAc, by a *de novo* pathway,

(c) optional formation of an activated Gal nucleotide, preferably UDP-Gal, by a *de novo* pathway, and

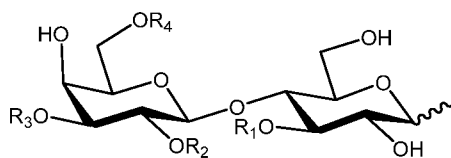
(d) formation of the glycoconjugate of interest from the internalized lactose derivative by a N-acetylglucosaminyl transfer mediated by the N-acetylglucosaminyl transferase enzyme

5 optionally followed by a galactosyl transfers mediated by the galactosyl transferase enzyme,

(iii) collecting the glycoconjugate that is an N-acetylglucosaminylated lactose optionally substituted by galactose and covalently linked to polyvinyl alcohol, preferably 3'-N-acetylglucosaminyl lactose, LNT or LNnT covalently linked to polyvinyl alcohol from the culture medium.

10 The present invention also relates to providing glycoconjugates having an oligosaccharide part covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols. Preferably, the above-mentioned glycoconjugates are O-, N- or S-glycosides. More preferably, the oligosaccharide moiety comprises

15 monosaccharide units selected from the group consisting of glucose, galactose, N-acetylglucosamine, fucose and sialic acid. Even more preferably, the oligosaccharide residue is a lactose moiety substituted by fucose and/or N-acetylglucosamine and/or galactose and/or sialic acid. Accordingly, the oligosaccharide moiety of the above defined glycoconjugates are characterized by formula **1**



1

wherein R_1 is fucosyl or H,

R_2 is fucosyl or H,

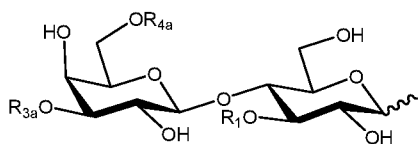
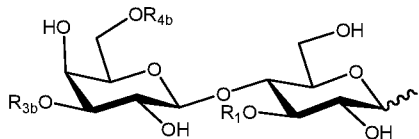
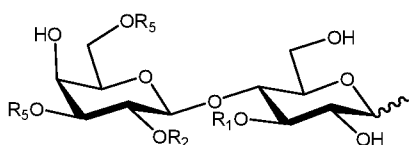
R_3 is selected from H, sialyl, N-acetyl-lactosaminyl and lacto-N-biosyl groups, wherein the N-acetyl lactosaminyl group may carry a glycosyl residue comprising one or more N-acetyl-lactosaminyl and/or one or more lacto-N-biosyl groups; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue,

R_4 is selected from H, sialyl and N-acetyl-lactosaminyl groups optionally substituted with a glycosyl residue comprising one or more N-acetyl-lactosaminyl and/or one or more lacto-N-biosyl groups; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue, and

5 provided that at least one of the R_1 , R_2 , R_3 and R_4 groups is different from H,

by culturing, as disclosed above, lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols.

Preferably, the moiety of formula **1** can be characterized by formula **1a**, **1b** or **1c**

**1a****1b****1c**

10

wherein R_1 and R_2 are as defined above,

R_{3a} is an N-acetyl-lactosaminyl group optionally substituted with a glycosyl residue comprising one N-acetyl-lactosaminyl and/or one lacto-N-biosyl group; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or

15 fucosyl residue,

R_{4a} is H or an N-acetyl-lactosaminyl group optionally substituted with a lacto-N-biosyl group; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue,

5 R_{3b} is a lacto-N-biosyl group optionally substituted with one or more sialyl and/or fucosyl residue(s),

R_{4b} is H or an N-acetyl-lactosaminyl group optionally substituted with one or two N-acetyl-lactosaminyl and/or one lacto-N-biosyl groups; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residues,

R₅ is, independently, H or sialyl,

10 and wherein at least one of R₁, R₂ or R₅ is not H.

More preferably, the moieties according to formulae **1a** or **1b** are characterized in that:

- the N-acetyl-lactosaminyl group in the glycosyl residue of R_{3a} is attached to another N-acetyl-lactosaminyl group with a 1-3 interglycosidic linkage,
- 15 - the lacto-N-biosyl group in the glycosyl residue of R_{3a} is attached to the N-acetyl-lactosaminyl group with a 1-3 interglycosidic linkage,
- the lacto-N-biosyl group in the glycosyl residue of R_{4a} is attached to the N-acetyl-lactosaminyl group with a 1-3 interglycosidic linkage,
- the N-acetyl-lactosaminyl group in the glycosyl residue of R_{4b} is attached to another N-acetyl-lactosaminyl group with a 1-3 or a 1-6 interglycosidic linkage,
- 20 - the lacto-N-biosyl group in the glycosyl residue of R_{4b} is attached to the N-acetyl-lactosaminyl group with a 1-3 interglycosidic linkage.

More preferably, the moieties according to formulae **1a**, **1b** and **1c** are human milk oligosaccharide (HMO) glycosyl groups.

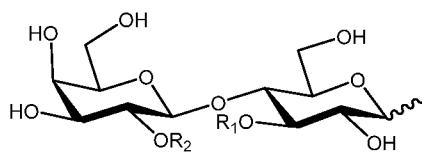
25 Also more preferably, the moiety of formula **1a** is a glycosyl residue of lacto-N-neotetraose, para-lacto-N-hexaose, para-lacto-N-neohexaose, lacto-N-neohexaose, para-lacto-N-octaose or lacto-N-neooctaose optionally substituted with one or more sialyl and/or fucosyl residue,

and the moiety of formula **1b** is a glycosyl residue of lacto-N-tetraose, lacto-N-hexaose, lacto-N-octaose, iso-lacto-N-octaose, lacto-N-decaose or lacto-N-neodecaose optionally substituted with one or more sialyl and/or fucosyl residue.

Preferably, the moieties of formula **1a** or **1b** are characterized in that:

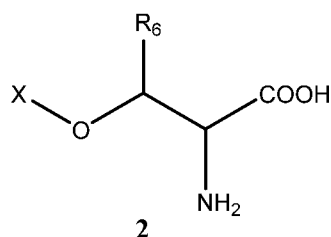
- 5 - the fucosyl residue attached to the N-acetyl-lactosaminyl and/or the lacto-N-biosyl group is linked to
 - the galactose of the lacto-N-biosyl group with 1-2 interglycosidic linkage and/or
 - the N-acetyl-glucosamine of the lacto-N-biosyl group with 1-4 interglycosidic linkage and/or
- 10 ○ the N-acetyl-glucosamine of the N-acetyl-lactosaminyl group with 1-3 interglycosidic linkage,
- the sialyl residue attached to the N-acetyl-lactosaminyl and/or the lacto-N-biosyl group is linked to
 - the galactose of the lacto-N-biosyl group with 2-3 interglycosidic linkage and/or
- 15 ○ the N-acetyl-glucosamine of the lacto-N-biosyl group with 2-6 interglycosidic linkage and/or
- the galactose of the N-acetyl-lactosaminyl group with 2-6 interglycosidic linkage.

According to a further preferred aspect, the moieties according to subformulae **1a**, **1b** or **1c** may be selected from the group of: a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 20 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose, lacto-N-neotetraose, LNFP-I, LNFP-II, LNFP-III, LNFP-V, LST-a, LST-b, LST-c, FLST-a, FLST-b, FLST-c, LNDFH-I, LNDFH-II, LNDFH-III, DS-LNT, FDS-LNT I and FDS-LNT II, or salts thereof. The glycosides may be alpha or beta-anomers, but preferably beta-anomers. Especially is preferred a residue of formula **1d**

**1d**

wherein R_1 is fucosyl or H, and R_2 is fucosyl or H, provided that at least one of the R_1 and R_2 groups is fucosyl.

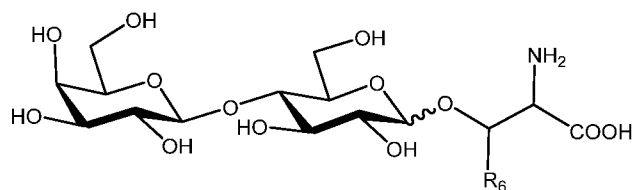
According to a preferred realization of the invention, the glycoconjugate obtainable by the method is an oligosaccharide covalently linked to an amino acid as described by the first particular embodiment above. Preferably, the oligosaccharide is linked to serine, threonine or hydroxyproline by an O-glycosidic linkage. More preferably, the glycoconjugate covalently linked to serine or threonine, and salts thereof, obtainable by the first particular embodiment, is characterized by formula **2**

**2**

wherein X is the moiety of formula **1** as defined above, and

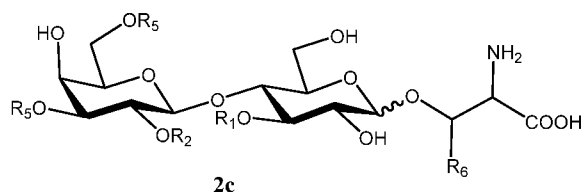
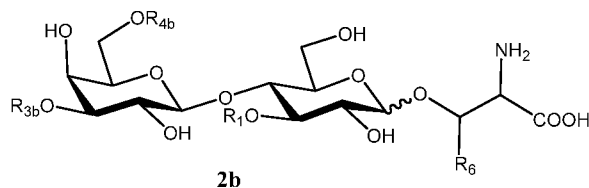
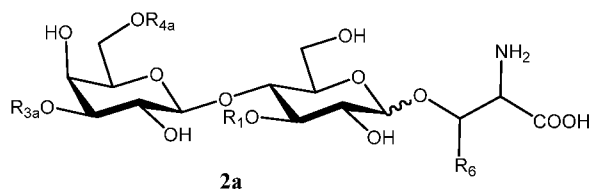
R_6 is H or methyl,

by culturing, as disclosed in the first particular embodiment, a lactose derivative of formula **3**

**3**

wherein R_6 is as defined above.

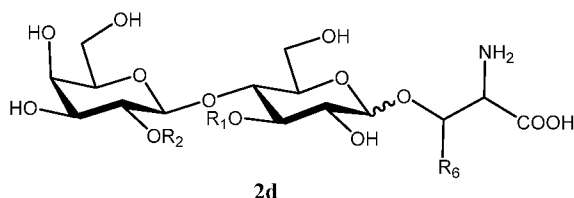
Preferably, in a compound of formula **2** moiety X is a group of formula **1a**, **1b** or **1c** as defined above, thus to form a compound of formula **2a**, **2b** or **2c**, respectively,



wherein R_1 , R_2 , R_{3a} , R_{3b} , R_{4a} , R_{4b} , R_5 and R_6 are as defined above.

More preferably, the compounds according to formulae **2a**, **2b** and **2c** are human milk oligosaccharide (HMO) glycosyl groups.

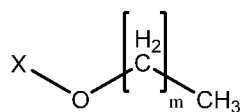
- 5 Also more preferably, the compound of formula **2a** has a glycosyl residue of lacto-N-neotetraose, para-lacto-N-hexaose, para-lacto-N-neohexaose, lacto-N-neohexaose, para-lacto-N-octaose or lacto-N-neooctaose optionally substituted with one or more sialyl and/or fucosyl residue, and the compound of formula **2b** has a glycosyl residue of lacto-N-tetraose, lacto-N-hexaose, lacto-N-octaose, iso-lacto-N-octaose, lacto-N-decaose or lacto-N-
- 10 neodecaose optionally substituted with one or more sialyl and/or fucosyl residue. Even more preferably, the glycosyl residue of compounds according to subformulae **2a**, **2b** or **2c** may be selected from the group of a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose, lacto-N-neotetraose, LNFP-I, LNFP-II, LNFP-III, LNFP-V, LST-a, LST-b, LST-c, FLST-a, FLST-
- 15 b, FLST-c, LNDFH-I, LNDFH-II, LNDFH-III, DS-LNT, FDS-LNT I and FDS-LNT II, more preferably from the group of a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose and lacto-N-neotetraose, or salts thereof. The glycosides may be alpha or beta-anomers, but preferably beta-anomers. Especially is preferred a compound of formula **2d**



wherein R_1 is fucosyl or H, R_2 is fucosyl or H, and R_6 is H or methyl, provided that at least one of the R_1 and R_2 groups is fucosyl.

According to another preferred realization of the invention, the glycoconjugate obtainable by the method is an oligosaccharide covalently linked to a longer alkyl group as described by the fifth particular embodiment above. Preferably, the oligosaccharide is linked to an alkyl group preferably consisting of 6-24, preferably 6-20, more preferably 6-14, particularly 6-10 carbon atoms, by an O-glycosidic linkage. More preferably, the glycoconjugate covalently linked to a longer alkyl group, obtainable by the fifth particular embodiment, is characterized by formula

4

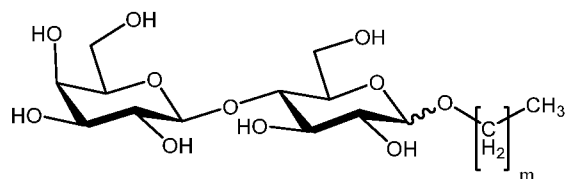


4

wherein X is the moiety of formula **1** as defined above, and

m is an integer from 5-23, preferably 5-19, more preferably 5-13, particularly 5-9,

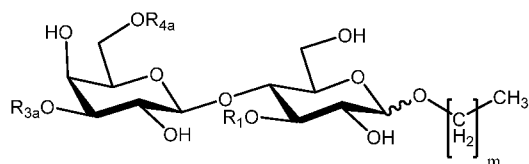
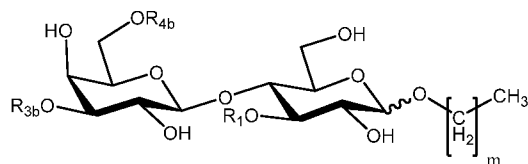
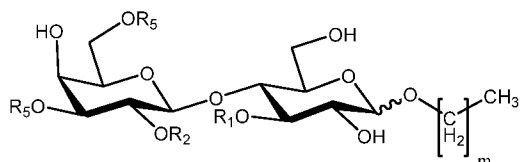
by culturing, as disclosed in the fifth particular embodiment, a lactose derivative of formula **5**



5

wherein m is as defined above.

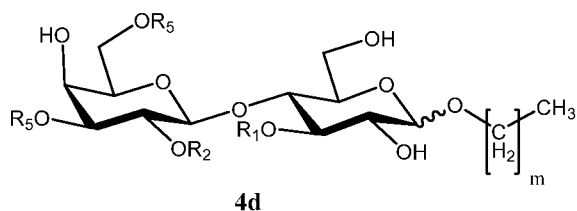
Preferably, in a compound of formula **4** moiety X is a group of formula **1a**, **1b** or **1c** as defined above, thus to form a compound of formula **4a**, **4b** or **4c**, respectively,

**4a****4b****4c**

wherein R_1 , R_2 , R_{3a} , R_{3b} , R_{4a} , R_{4b} , R_5 and m are as defined above.

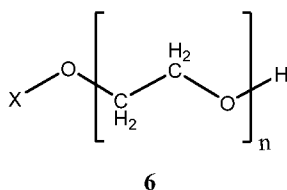
More preferably, the compounds according to formulae **4a**, **4b** and **4c** are human milk oligosaccharide (HMO) glycosyl groups.

- 5 Also more preferably, the compound of formula **4a** has a glycosyl residue of lacto-N-neotetraose, para-lacto-N-hexaose, para-lacto-N-neohexaose, lacto-N-neohexaose, para-lacto-N-octaose or lacto-N-neooctaose optionally substituted with one or more sialyl and/or fucosyl residue, and the compound of formula **4b** has a glycosyl residue of lacto-N-tetraose, lacto-N-hexaose, lacto-N-octaose, iso-lacto-N-octaose, lacto-N-decaose or lacto-N-
- 10 neodecaose optionally substituted with one or more sialyl and/or fucosyl residue. Even more preferably, the glycosyl residue of compounds according to subformulae **4a**, **4b** or **4c** may be selected from the group of: a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose, lacto-N-neotetraose, LNFP-I, LNFP-II, LNFP-III, LNFP-V, LST-a, LST-b, LST-c, FLST-a, FLST-
- 15 b, FLST-c, LNDFH-I, LNDFH-II, LNDFH-III, DS-LNT, FDS-LNT I and FDS-LNT II, more preferably from the group of a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose and lacto-N-neotetraose, or salts thereof. The glycosides may be alpha or beta-anomers, but preferably beta-anomers. Especially is preferred a compound of formula **4d**



wherein R_1 is fucosyl or H, R_2 is fucosyl or H, and m is 6-10, provided that at least one of the R_1 and R_2 groups is fucosyl.

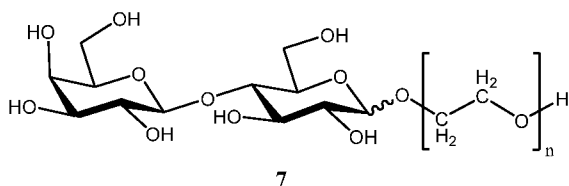
- 5 According to another preferred realization of the invention, the glycoconjugate obtainable by the method is an oligosaccharide covalently linked to a polyethylene glycol as described by the sixth particular embodiment above. Preferably, the glycoconjugate covalently linked to a polyethylene glycol, obtainable by the sixth particular embodiment, is characterized by formula **6**



- 10 wherein X is the moiety of formula **1** as defined above, and

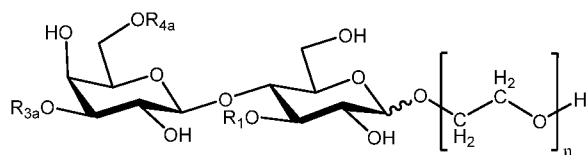
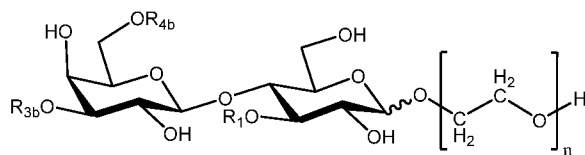
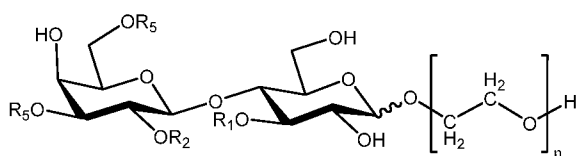
n is an integer from 1 to 10, preferably 2 to 6,

by culturing, as disclosed in the sixth particular embodiment, a lactose derivative of formula **7**



- 15 wherein n is as defined above.

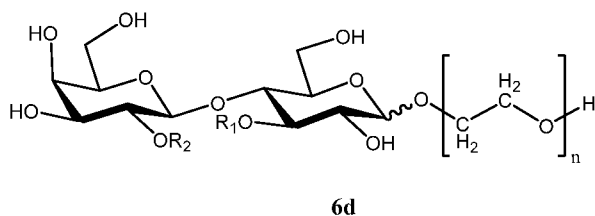
Preferably, in a compound of formula **6** moiety X is a group of formula **1a**, **1b** or **1c** as defined above, thus to form a compound of formula **6a**, **6b** or **6c**, respectively,

**6a****6b****6c**

wherein R_1 , R_2 , R_{3a} , R_{3b} , R_{4a} , R_{4b} , R_5 and n are as defined above.

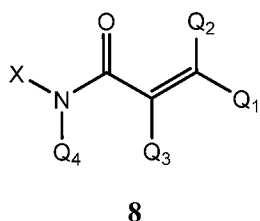
More preferably, the compounds according to formulae **6a**, **6b** and **6c** are human milk oligosaccharide (HMO) glycosyl groups.

- 5 Also more preferably, the compound of formula **6a** has a glycosyl residue of lacto-N-neotetraose, para-lacto-N-hexaose, para-lacto-N-neohexaose, lacto-N-neohexaose, para-lacto-N-octaose or lacto-N-neooctaose optionally substituted with one or more sialyl and/or fucosyl residue, and the compound of formula **6b** has a glycosyl residue of lacto-N-tetraose, lacto-N-hexaose, lacto-N-octaose, iso-lacto-N-octaose, lacto-N-decaose or lacto-N-
- 10 neodecaose optionally substituted with one or more sialyl and/or fucosyl residue. Even more preferably, the glycosyl residue of compounds according to subformulae **6a**, **6b** or **6c** may be selected from the group of: a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose, lacto-N-neotetraose, LNFP-I, LNFP-II, LNFP-III, LNFP-V, LST-a, LST-b, LST-c, FLST-a, FLST-
- 15 b, FLST-c, LNDFH-I, LNDFH-II, LNDFH-III, DS-LNT, FDS-LNT I and FDS-LNT II, more preferably from the group of a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose and lacto-N-neotetraose, or salts thereof. The glycosides may be alpha or beta-anomers, but preferably beta-anomers. Especially is preferred a compound of formula **6d**



wherein R_1 is fucosyl or H, R_2 is fucosyl or H, and n is 2-6, provided that at least one of the R_1 and R_2 groups is fucosyl.

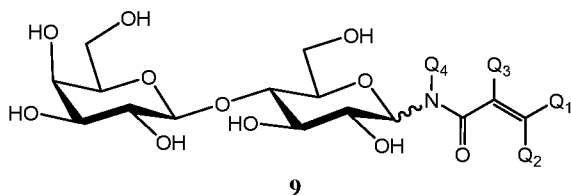
- According to another preferred realization of the invention, the glycoconjugate obtainable by the method is an oligosaccharide covalently linked to to an α,β -unsaturated amido group as described by the seventh particular embodiment above. Preferably, the glycoconjugate covalently linked to to an α,β -unsaturated amido group, obtainable by the seventh particular embodiment, is characterized by formula **8**



- wherein X is the moiety of formula **1** as defined above, and

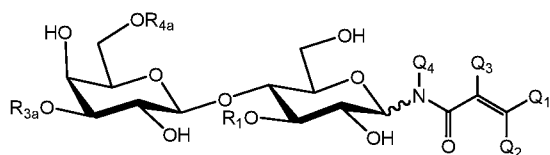
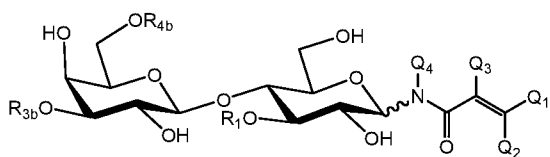
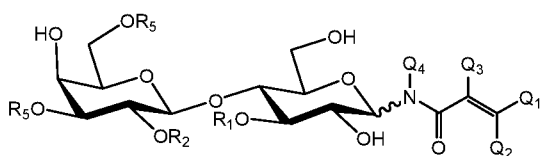
Q_1 , Q_2 , Q_3 and Q_4 are as defined above,

by culturing, as disclosed in the seventh particular embodiment, a lactose derivative of formula **9**



- wherein Q_1 , Q_2 , Q_3 and Q_4 are as defined above.

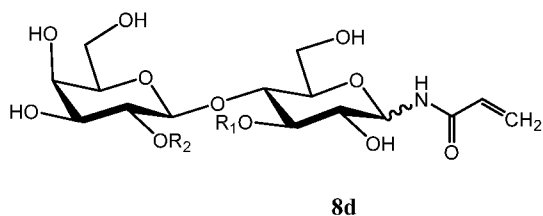
Preferably, in a compound of formula **8** moiety X is a group of formula **1a**, **1b** or **1c** as defined above, thus to form a compound of formula **8a**, **8b** or **8c**, respectively,

**8a****8b****8c**

wherein R_1 , R_2 , R_{3a} , R_{3b} , R_{4a} , R_{4b} , R_5 , Q_1 , Q_2 , Q_3 and Q_4 are as defined above.

More preferably, the compounds according to formulae **8a**, **8b** and **8c** are human milk oligosaccharide (HMO) glycosyl groups.

- 5 Also more preferably, the compound of formula **8a** has a glycosyl residue of lacto-N-neotetraose, para-lacto-N-hexaose, para-lacto-N-neohexaose, lacto-N-neohexaose, para-lacto-N-octaose or lacto-N-neooctaose optionally substituted with one or more sialyl and/or fucosyl residue, and the compound of formula **8b** has a glycosyl residue of lacto-N-tetraose, lacto-N-hexaose, lacto-N-octaose, iso-lacto-N-octaose, lacto-N-decaose or lacto-N-
- 10 neodecaose optionally substituted with one or more sialyl and/or fucosyl residue. Even more preferably, the glycosyl residue of compounds according to subformulae **8a**, **8b** or **8c** may be selected from the group of: a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose, lacto-N-neotetraose, LNFP-I, LNFP-II, LNFP-III, LNFP-V, LST-a, LST-b, LST-c, FLST-a, FLST-
- 15 b, FLST-c, LNDFH-I, LNDFH-II, LNDFH-III, DS-LNT, FDS-LNT I and FDS-LNT II, more preferably from the group of a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose and lacto-N-neotetraose, or salts thereof. The glycosides may be alpha or beta-anomers, but preferably beta-anomers. Especially is preferred a compound of formula **8d**



wherein R_1 is fucosyl or H, R_2 is fucosyl or H, provided that at least one of the R_1 and R_2 groups is fucosyl.

EXAMPLES

5 Example 1 - Tetraethylene glycol 2,3,6,2',3',4',6'-hepta-O-acetyl- β -lactoside:

A) To a solution of 2,3,6,2',3',4',6'-hepta-O-acetyl- α -lactosyl bromide (15.0 g) and tetraethylene glycol (13.23 ml) in CH_2Cl_2 (200 ml) Ag_2CO_3 (5.90 g) was added at 0 °C under N_2 . The reaction mixture was stirred at rt overnight then filtered through a pad of celite, the filtrate was washed with water and brine, dried and evaporated. The residue was purified by flash chromatography on silica gel to afford the title compound (12.66 g, 72.5 %) as a white foam. ^1H NMR (300 MHz, CDCl_3) δ 5.31 (d, J = 3.4 Hz, 1H), 5.17 (t, J = 9.3 Hz, 1H), 5.08 (dd, J = 7.8, 10.4 Hz, 1H), 4.94-4.83 (m, 2H), 4.56 (d, J = 7.9, 1H), 4.48-4.44 (m, 2H), 4.14-4.01 (m, 3H), 3.91-3.82 (m, 2H), 3.80-3.73 (m, 1H), 3.72-3.66 (m, 3H), 3.65-3.57 (m, 14H), 2.12 (s, 3H), 2.09 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 2.01 (s, 3-H), 2.01 (s, 3H), 1.94 (s, 3H).

B) To a solution of lactose peracetate (67.8 g) and tetraethylene glycol (69 ml) in CH_2Cl_2 (500 ml) $\text{BF}_3 \cdot \text{OEt}_2$ (64 ml) was added dropwise at 0 °C under Ar. The reaction mixture was stirred at rt overnight. The mixture was neutralised with triethyl amine, diluted with CH_2Cl_2 , washed with saturated NaHCO_3 solution and brine, dried and evaporated. The residue was chromatographed in silica gel to give the title compound (50 g, 62 %).

Example 2 - Tetraethylene glycol β -lactoside

To a solution of tetraethylene glycol 2,3,6,2',3',4',6'-hepta-O-acetyl- β -lactoside (12.66 g) in MeOH (60 ml) MeONa (25% in MeOH, 0.355 ml) was added at 0 °C. The reaction mixture was stirred at 0 °C for 3 h then neutralized with acidic resin, filtered and concentrated to afford the title compound (7.88 g, 97.5 %) as a white foam. ^1H NMR (300 MHz, MeOD) δ 4.36 (d, J = 7.3 Hz, 1H), 4.35 (d, J = 7.8 Hz, 1H), 4.00 (ddd, J = 2.9, 4.9, 9.6 Hz, 1H), 3.87 (dd,

$J = 3.3, 9.5 \text{ Hz, 1H}$), 3.82–3.81 (m, 1H), 3.78 (t, $J = 3.6 \text{ Hz, 1H}$), 3.75 – 3.74 (m, 1H), 3.72 – 3.68 (m, 4H), 3.67 – 3.64 (m, 10H), 3.61–3.55 (m, 5H), 3.53–3.50 (m, 2H), 3.44–3.89 (m, 1H), 3.30–3.24 (m, 1H); ^{13}C NMR (75 MHz, MeOD) δ 105.0, 104.3, 80.5, 77.0, 76.4, 76.2, 74.8, 74.7, 73.6, 72.5, 71.5, 71.4, 71.4, 71.4, 71.3, 70.3, 69.7, 62.5, 62.2, 61.9.

5 *Example 3 - Tetraethylene glycol β -glycoside of 2'-FL*

Plasmids and strain, suitable for performing the fermentation, were constructed, and the techniques with regard to cloning were performed in accordance with WO 01/04341 and Drouillard et al. *Angew. Chem. Int. Ed. Eng.* **45**, 1778 (2006), where the general culturing conditions have also been described. Engineered *E. coli* used in this example was constructed
10 from *E. coli* K12 strain by deleting genes that are liable to degrade the acceptor, the oligosaccharide product and its metabolic intermediates, inter alia the lacZ, lacA and wcaJ genes, maintaining manB, manC, gmd and wcaG genes involved in the GDP-fucose biosynthesis, and inserting *H. pylori* futC gene for α -1,2-fucosyl transferase.

The culture was carried out in a 1 l fermenter containing 0.75 l of mineral culture medium
15 (Samain et al. *J. Biotechnol.* **72**, 33 (1999)). The temperature was kept at around 33 °C and the pH regulated at 6.8 with aqueous NH_4OH solution. The inoculum (1 % of the volume of the basal medium) consisted in a LB medium and the culture of the producing strain. The exponential growth phase started with the inoculation and stopped until exhaustion of the carbon source (glucose 17.5 g/l) initially added to the medium. Tetraethylene glycol β -
20 lactoside (5.95 g dissolved in 50 ml of water) and the inducer (IPTG, 1-2 ml of a 50 mg/ml solution) were added at the end of the exponential phase. Then a fed-batch was realized, using a 500 g/l aqueous glycerol solution, with a high substrate feeding rate of $\approx 4.5 \text{ g/h}$ of glycerol for 1 l of culture for 5-6 hours followed by a lower glycerol feeding rate of $\approx 3 \text{ g/h}$ for 1 l culture for 20 hours.

25 At the end of the fermentation, the culture was centrifuged for 20-30 min at 6000 rpm at 25 °C. The supernatant was kept. The cells were acidified to pH ≈ 3 using a H^+ form resin or treated in water at 80-100 °C for 20-30 min. This resulted in the precipitation of the proteins. The resin was recovered by decantation and precipitated proteins were removed by centrifugation for 20-30 min at 6000 rpm at 25 °C. The combined supernatants were
30 subjected to standard chromatographic techniques to give the title compound (4.8 g).

LC-MS (instrument: Bruker microQToF II MS coupled with Dionex Ultimate 3000 UHPLC; ionization: ESI negative, dry temperature: 200 °C; mode: LC-MS, 1:1 split of flow; calibration: with Na-format cluster solution): 663.2744 Da $[\text{M-H}]^-$. ^1H NMR (300 MHz, D_2O) δ

5.29 (brs, 1H, H-1''), 4.48 (ddd, $J = 11.7, 7.8, 1.9$ Hz, 2H), 4.21 (td, $J = 7.5, 5.7$ Hz, 1H), 4.10–3.48 (m, 29H), 3.48–3.40 (m, 1H), 3.33 (ddd, $J = 9.5, 7.9, 1.9$ Hz, 1H), 1.21 (dd, $J = 6.6, 1.9$ Hz, 3H, H-6''). ^{13}C NMR (75 MHz, D_2O) δ 102.4, 100.4 and 99.5 (C-1, C-1' and C-1''), 76.4, 76.0, 75.4, 75.3, 74.3, 73.7, 73.0, 71.8, 69.8, 69.7, 69.6, 69.5, 69.2, 68.9, 68.3, 67.0, 61.2, 60.4, 60.3, 15.4 (C-6'').

Example 4 - Octyl β -glycoside of 2'-FL

The culture was carried out as described in Example 3 with the following differences:

- the starting volume of the mineral culture medium was 0.5 l,
- the acceptor was octyl β -lactoside (5 g, ref.) which was added as neat material to the medium at the end of the exponential phase,
- glycerol feeding lasted 93 hours.

The course of the fermentation was followed by LC-MS. A new compound was detected at 599.2937 Da $[\text{M-H}]^-$, corresponding to the title product (for LC-MS conditions see Example 3). The acceptor/product ratio measured is given in the following table:

	20 hours	93 hours
supernatant	66/34	34/66
intracellular	48/52	7/93

The culture was centrifuged for 20-30 min at 6000 rpm at 25 °C. The supernatant was acidified to $\text{pH} \approx 3$ using a H^+ form resin or treated in water at 80-100 °C for 20-30 min. The resin was recovered by decantation and precipitated proteins were removed by centrifugation for 20-30 min at 6000 rpm at 25 °C. A part of the supernatant was subjected to standard chromatographic techniques to give the title compound, the ^1H -NMR spectrum of which was in good agreement with that reported in Meloncelli et al. *Aust. J. Chem.* **62**, 558 (2009).

Example 5 - O-L-serine- β -glycoside of 2'-FL

The culture was carried out as described in Example 4 with the following difference:

- the acceptor was O- β -lactosyl-L-serine (5 g, Rude et al. *Carbohydr. Res.* **8**, 219 (1968)) which was dissolved in water and added to the medium at the end of the exponential phase,
- glycerol feeding lasted 48 hours.

The course of the fermentation was followed by LC-MS. A new compound was detected at 574.1916 Da $[M-H]^-$, corresponding to the title product (for LC-MS conditions see Example 3). After 22 hours from its addition practically no acceptor could be detected in the intracellular matrix and the supernatant.

5 *Example 6 - N-acryl- β -N-glycoside of 2'-FL*

The culture was carried out as described in Example 4 with the following difference:

- the acceptor was N-acryl- β -lactosyl-amine (5 g, Kallin et al. *J. Carbohydr. Chem.* **8**, 597 (1989)) which was added as neat material to the medium at the end of the exponential phase,
- 10 - glycerol feeding lasted 94 hours.

The course of the fermentation was followed by LC-MS. A new compound was detected at 540.1933 Da $[M-H]^-$, corresponding to the title product (for LC-MS conditions see Example 3). The acceptor/product ratio measured is given in the following table:

	21 hours	94 hours
supernatant	52/48	24/76
intracellular	34/66	22/78

CLAIMS

1. A method for producing a glycoconjugate comprising an oligosaccharide part covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the method comprising:

(i) providing a genetically modified cell comprising a gene encoding a glycosyl transferase enzyme that is able to transfer the glycosyl residue of an activated sugar nucleotide to an internalized acceptor,

(ii) culturing said genetically modified cell in the presence of an exogenous acceptor comprising a mono- or disaccharide part covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, inducing

(a) internalization of the exogenous acceptor by the genetically modified cell, and

(b) formation of said glycoconjugate from said internalized acceptor molecule by a glycosyl transfer mediated by said glycosyl transferase enzyme expressed by said cell, and then

(iii) collecting said glycoconjugate from the fermentation broth.

2. The method according claim 1, wherein said genetically modified cell provided in step (i) further comprises a set of genes encoding enzymes responsible for the synthesis of said activated sugar nucleotide by a *de novo* pathway, and wherein said culturing step (ii) further induces producing said activated sugar nucleotide by a *de novo* pathway.

3. The method according to any one of the claims 1 to 2, wherein said glycosyl transferase enzyme is selected from the group consisting of N-acetyl-glucosaminyl transferases, galactosyl transferases, N-acetyl-galactosaminyl transferases, glucuronosyl transferases, sialyl transferases and fucosyl transferases.

4. The method according to claim 3, wherein said glycosyl transferase enzyme is selected from the group consisting of β -1,3-N-acetyl-glucosaminyl transferase, β -1,6-N-acetyl-glucosaminyl transferase, β -1,3-galactosyl transferase, β -1,4-galactosyl transferase, α -2,3-sialyl transferase, α -2,6-sialyl transferase, α -1,2-fucosyl transferase, α -1,3-fucosyl transferase and α -1,4-fucosyl transferase.

5. The method according to any one of the claims 1 to 4, wherein the genetically modified cell is a bacterium or yeast, preferably a bacterium, more preferably of *E. coli* type.

6. The method according to any one of the claims 1 to 5, wherein the internalization of the exogenous acceptor takes place via an active transport mechanism mediated by a permease, preferably a lactose permease.

7. The method according to any one of the claims 1 to 6, wherein the genetically modified cell is *E. coli* of LacZ⁻, LacY⁺ genotype, the exogenous acceptor is a lactose covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols, the glycosyl transferase is a fucosyl transferase, and/or a sialyl transferase, and/or an N-acetylglucosaminyl transferase and/or a galactosyl transferase, and the glycoconjugate produced by the method is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose, preferably a human milk oligosaccharide, more preferably 2'-fucosyllactose, 3-fucosyllactose, difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, LNT or LNnT, covalently linked to a non-sugar moiety selected from the group consisting of amino acids, peptides, proteins, lipids, longer alkyl groups, polyethylene glycols, α,β -unsaturated amido group and polyvinyl alcohols.

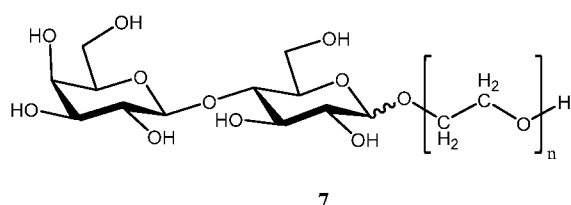
8. The method according to claim 7, wherein the exogenous acceptor is a serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, and the glycoconjugate is a serine, threonine or hydroxyproline, preferably serine, O-glycoside of a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose, preferably of a human milk oligosaccharide.

9. The method according to claim 8, wherein the glycoconjugate is a fucosylated serine, threonine or hydroxyproline, preferably serine, O-glycoside of lactose, preferably serine, threonine or hydroxyproline, preferably serine, O-glycoside of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose.

10. The method according to claim 7, wherein the exogenous acceptor is a C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀ alkyl O-glycoside of lactose, and the glycoconjugate is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀ alkyl O-glycoside of lactose, preferably of a human milk oligosaccharide.

11. The method according to claim 10, wherein the glycoconjugate is a fucosylated C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀ alkyl O-glycoside of lactose, particularly C₆-C₂₀, preferably a C₆-C₁₄, more preferably a C₆-C₁₀ alkyl O-glycoside of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose.

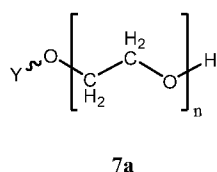
5 12. The method according to claim 7, wherein the exogenous acceptor consists of lactose covalently linked to polyethylene glycol, preferably a compound of formula **7**



wherein n is an integer from 1 to 10, preferably from 2 to 6,

and the glycoconjugate is a fucosylated and/or sialylated and/or N-acetylglucosaminylated

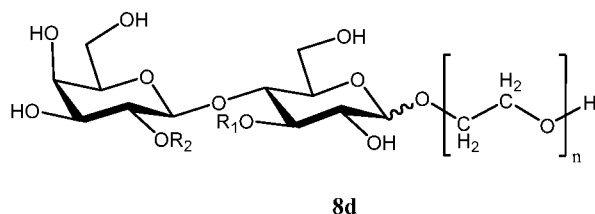
10 and/or galactosylated lactose covalently linked to polyethylene glycol, preferably a compound of formula **7a**



wherein Y is a human milk oligosaccharide glycosyl residue, and

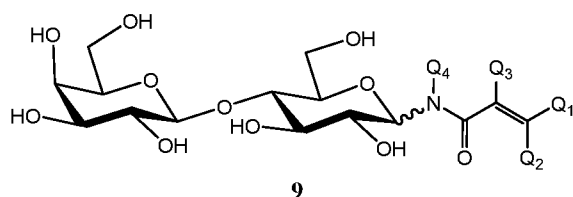
n is as defined above.

15 13. The method according to claim 12, wherein the compound of formula **7a** is of formula **8d**



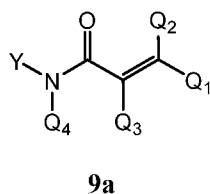
wherein R_1 is fucosyl or H, R_2 is fucosyl or H, n is an integer from 1 to 10, preferably 2 to 6, provided that at least one of the R_1 and R_2 groups is fucosyl.

14. The method according to claim 7, wherein the exogenous acceptor is a compound of formula **9**



wherein Q_1 , Q_2 , Q_3 and Q_4 are, independently, H and methyl, preferably H,

and the glycoconjugate is a fucosylated and/or sialylated and/or N-acetylglucosaminylated and/or galactosylated lactose derivative of formula **9**, preferably a compound of formula **9a**

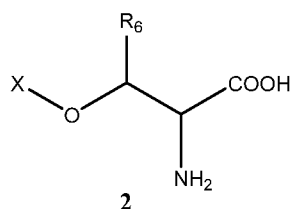


10 wherein Y is a human milk oligosaccharide glycosyl residue, and

Q_1 , Q_2 , Q_3 and Q_4 are as defined above.

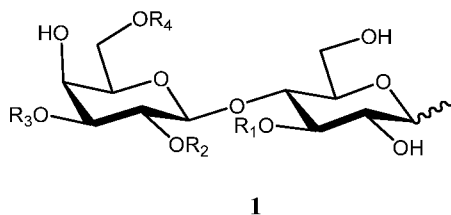
15. The method according to claim 14, wherein in compound of formula **9a** Y is a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose or difucosyllactose, and Q_1 , Q_2 , Q_3 and Q_4 are as defined above.

15 16. A compound of formula **2**



wherein R_6 is H or methyl, and

X is a moiety of formula **1**



wherein R_1 is fucosyl or H,

5 R_2 is fucosyl or H,

R_3 is selected from H, sialyl, N-acetyl-lactosaminyl and lacto-N-biosyl groups, wherein the N-acetyl lactosaminyl group may carry a glycosyl residue comprising one or more N-acetyl-lactosaminyl and/or one or more lacto-N-biosyl groups; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue,

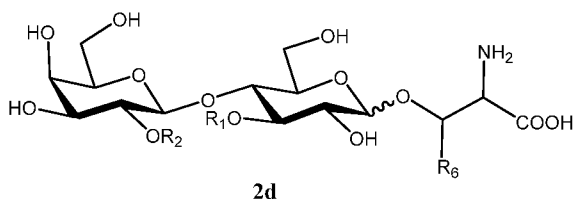
10 R_4 is selected from H, sialyl and N-acetyl-lactosaminyl groups optionally substituted with a glycosyl residue comprising one or more N-acetyl-lactosaminyl and/or one or more lacto-N-biosyl groups; each of the N-acetyl-lactosaminyl and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue, and

provided that at least one of the R_1 , R_2 , R_3 and R_4 groups is different from H,

15 preferably X is a human milk oligosaccharide glycosyl residue, more preferably a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose or lacto-N-neotetraose,

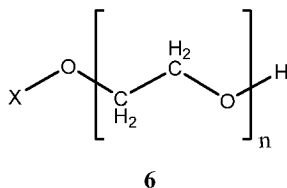
or a salt thereof.

17. The compound according to claim 16 characterized by formula **2d**



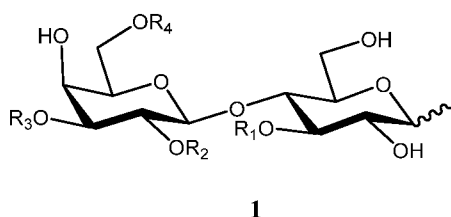
wherein R_1 is fucosyl or H, R_2 is fucosyl or H, and R_6 is H or methyl, provided that at least one of the R_1 and R_2 groups is fucosyl.

18. A compound of formula **6**



5 wherein n is an integer from 1 to 10, preferably 2 to 6, and

X is the moiety of formula **1**



wherein R_1 is fucosyl or H,

R_2 is fucosyl or H,

10 R_3 is selected from H, sialyl, N-acetyl-lactosaminy and lacto-N-biosyl groups, wherein the N-acetyl lactosaminy group may carry a glycosyl residue comprising one or more N-acetyl-lactosaminy and/or one or more lacto-N-biosyl groups; each of the N-acetyl-lactosaminy and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue,

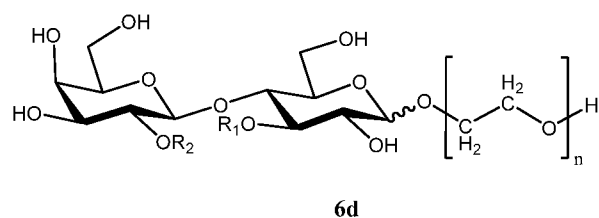
15 R_4 is selected from H, sialyl and N-acetyl-lactosaminy groups optionally substituted with a glycosyl residue comprising one or more N-acetyl-lactosaminy and/or one or more lacto-N-biosyl groups; each of the N-acetyl-lactosaminy and lacto-N-biosyl groups can be substituted with one or more sialyl and/or fucosyl residue, and

provided that at least one of the R_1 , R_2 , R_3 and R_4 groups is different from H,

preferably X is a human milk oligosaccharide glycosyl residue, more preferably a glycosyl residue of 2'-fucosyllactose, 3-fucosyllactose, 2',3-difucosyllactose, 3'-sialyllactose, 6'-sialyllactose, 3'-sialyl-3-fucosyllactose, lacto-N-tetraose or lacto-N-neotetraose,

or a salt thereof.

- 5 19. The compound according to claim 18 characterized by formula **6d**



wherein R_1 is fucosyl or H, R_2 is fucosyl or H, n is an integer from 1 to 10, preferably 2 to 6, provided that at least one of the R_1 and R_2 groups is fucosyl.

INTERNATIONAL SEARCH REPORT

 International application No.
 PCT/DK2013/050306

A. CLASSIFICATION OF SUBJECT MATTER

IPC: see extra sheet

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC: C07H, C12N, C12P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE, DK, FI, NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, PAJ, WPI data, BIOSIS, CHEM ABS Data, EMBASE, INSPEC, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2008002230 A1 (NILSSON KURT G I), 3 January 2008 (2008-01-03); page 1, line 4 - line 6; page 1, line 20; page 2, line 6 - line 8	1-11, 14-17
A	--	12-13, 18-19
X	EP 1911850 A1 (CENTRE NAT RECH SCIENT ET AL), 16 April 2008 (2008-04-16); claims 20, 22	1-11, 14-17
A	--	12-13, 18-19

☒ Further documents are listed in the continuation of Box C.☒ See patent family annex.

* Special categories of cited documents:

“A” document defining the general state of the art which is not considered to be of particular relevance

“E” earlier application or patent but published on or after the international filing date

“L” document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

“O” document referring to an oral disclosure, use, exhibition or other means

“P” document published prior to the international filing date but later than the priority date claimed

“T” later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

“X” document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

“Y” document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

“&” document member of the same patent family

Date of the actual completion of the international search

20-12-2013

Date of mailing of the international search report

27-12-2013

Name and mailing address of the ISA/SE

Patent- och registreringsverket

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/DK2013/050306

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 0104341 A1 (CENTRE NAT RECH SCIENT ET AL), 18 January 2001 (2001-01-18); whole document; page 12, line 13 - line 22; page 13, line 28 - line 2; page 13, line 17 - line 22	1-11, 14-17
A	--	12-13, 18-19
X	WO 2007101862 A1 (CENTRE NAT RECH SCIENT ET AL), 13 September 2007 (2007-09-13); whole document; page 29, line 21 - line 32	1-11, 14-17
A	--	12-13, 18-19
X	FORT S, et al. "Biosynthesis of conjugatable saccharidic moieties of GM2 and GM3 gangliosides by engineered E. coli", Chem Commun, 2005, pages 2558-2560; whole document	1-11, 14-17
A	--	
A	DROUILLARD S et al., "Large-Scale Synthesis of H-AntigenOligosaccharides by Expressing Helicobacterpylori alpha 1,2-Fycosyltransgerase in Metabolically Engineered Escheri coli Cells", Angewandte Chemie Intl. ed. 2006, vol 45, page1778-1780; whole document	1-19
A	--	
A	US 20120208181 A1 (MERIGHI MASSIMO ET AL), 16 August 2012 (2012-08-16); whole document; figures 1-3	1-19
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/DK2013/050306

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

- 1: Claims 8-9, and 16-17 as well as parts of claims 1-7, relating to glycoconjugates being amino acid O-glycosides of lactose.
- 2: Claims 10-11 as well as parts of claims 1-7, relating to glycoconjugates being alkyl O-glycosides of lactose.
- 3: Claims 12-13, 18-19 as well as parts of claims 1-7, relating to glycoconjugates being polyethylene glycol O-glycosides of lactose.
4. Claims 14-15 as well as parts of claims 1-7, relating to glycoconjugates being alpha-, beta-, unsaturated amido group O-glycosides of lactose.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

Continuation of: second sheet**International Patent Classification (IPC)*****C12P 19/18*** (2006.01)***C07H 3/06*** (2006.01)***C07H 15/08*** (2006.01)***C12P 19/44*** (2006.01)***C12N 1/21*** (2006.01)

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/DK2013/050306

WO	2008002230	A1	03/01/2008	NONE		
EP	1911850	A1	16/04/2008	NONE		
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				AT	318324 T	15/03/2006
				AU	6296100 A	30/01/2001
				AU	780290 B2	17/03/2005
				CA	2378562 A1	18/01/2001
				DE	60026142 D1	27/04/2006
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				EP	1991690 A1	19/11/2008
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				US	8507227 B2	13/08/2013
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				WO	2012112777 A3	08/11/2012