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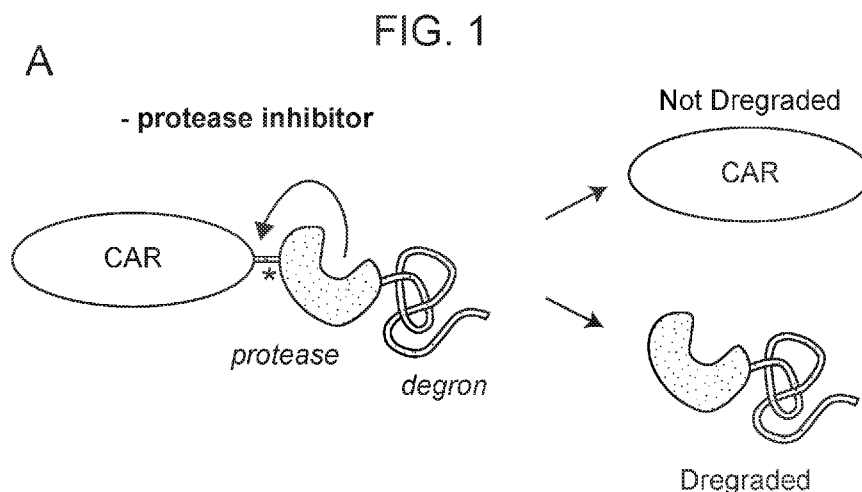
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(57) Abstract: Provided are polypeptides that include, from N-terminus to C-terminus, a chimeric antigen receptor (CAR), a protease, and a degranon, where the polypeptide further includes a cleavage site for the protease disposed between the CAR and the degranon. Also provided are cells that include such polypeptides (e.g., where the cells express the CAR on their surface) and pharmaceutical compositions including such cells. Nucleic acids that encode the polypeptides, cells including such nucleic acids, and pharmaceutical compositions including such cells, are also provided. Also provided are methods for controlling the expression of a CAR on the surface of a cell, and methods of using the cells of the present disclosure, including methods of using such cells to administer a regulatable CAR cell-based therapy (e.g., a regulatable CAR T cell therapy) to an individual.



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CHIMERIC ANTIGEN RECEPTOR POLYPEPTIDES AND METHODS OF USING SAME

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Patent Application No. 62/694,830, filed July 6, 2018, which application is incorporated herein by reference in its entirety.

INTRODUCTION

Engineering cells such as T cells, stem cells or other cellular components of tissues provides the potential to cure numerous diseases. Engineering T cells with chimeric antigen receptors (CARs) has already demonstrated impressive clinical efficacy and additional applications using engineered CARs in the context of cell therapy holds great promise. There are, however, substantial risks associated with unregulated CAR expression.

The constant antigenic burden during chronic viral infections, cancer, and the like, causes CD8⁺ T cells to develop a state of dysfunction often referred to as T cell exhaustion, characterized by a loss of some effector functions and up-regulation of multiple inhibitory receptors. The result is a population of antigen-specific CD8⁺ T cells that are unable to successfully clear or stabilize an infection or tumor. However, exhausted CD8⁺ T cells (T_{EX}) do retain some functionality. Blockade of inhibitory receptors such as programmed death 1 (PD-1), cytotoxic T-lymphocyte associated protein 4 (CTLA-4), Tim-3, and T-cell immunoreceptor with Ig and ITIM domains (TIGIT) has been used to reinvigorate T_{EX} in order to combat cancer and viral infections *in vitro* and in clinical studies. In mice, the use of anti-PD-L1 blocking antibodies has been shown to restore CD8⁺ T cell function against multiple epitopes during chronic lymphocytic choriomeningitis virus (LCMV) clone 13 infection. Barber et al. (2006) *Nature* 439:682-7. Clinical studies with therapeutic blockade of inhibitory receptor pathways (e.g., PD-1/PD-L1) in the field of human cancer research have rapidly resulted in several drugs approved by the FDA. Engineered T cells, such as T cells expressing chimeric antigen receptors (CARs), are another approach to combat malignancies. However, CAR-T cells also develop exhaustion, e.g., from tonic signaling or persistent antigen stimulation. Although advances have been made, patients do not respond equally to inhibitory receptor blockade therapy and the durability of the responses vary widely.

SUMMARY

Provided are polypeptides that include, from N-terminus to C-terminus, a chimeric antigen receptor (CAR), a protease, and a degron, where the polypeptide further includes a cleavage site for the protease disposed between the CAR and the degron. Also provided are cells that include such polypeptides (e.g., where the cells express the CAR on their surface) and pharmaceutical compositions including such cells. Nucleic acids that encode the polypeptides, cells including such nucleic acids, and pharmaceutical compositions including such cells, are also provided. Also provided are methods for controlling the expression of a CAR on the surface of a cell, and methods of using the cells of the present disclosure, including methods of using such cells to administer a regulatable CAR cell-based therapy (e.g., a regulatable CAR T cell therapy) to an individual.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 Schematic illustration of a polypeptide according to an example embodiment of the present disclosure. Shown is a polypeptide that includes a CAR, a protease, a degron, and a cleavage site (indicated by an asterisk) for the protease, where the cleavage site is disposed between the CAR and the degron. The cleavage site, protease, and degron may be collectively referred to herein as a Small Molecule-Assisted Shutoff (SMASh) tag. Panel A: In the absence of a protease inhibitor, the protease cleaves the cleavage site such that the CAR is separated from the degron and therefore not degraded. When expressed in a cell in the absence of the protease inhibitor, the CAR construct is expressed on the surface of the cell. Panel B: In the presence of an inhibitor of the protease, the CAR remains linked to the degron and, accordingly, is degraded.

FIG. 2 Panel A: Schematic illustration of a CAR that includes an antibody ("Ab") that binds to an antigen of interest, a transmembrane domain ("Tm"), and an intracellular signaling domain ("IS"). Panel B: Schematic illustration of a polypeptide according to one embodiment of the present disclosure. In this example, the CAR includes an anti-B7-H3 (CD276) scFv as the antibody ("B7H3 scFv"), a CD 8 transmembrane domain ("CD 8 Tm"), and a 4-1BB and CD 3 zeta signaling endodomain ("4-1BB" and "CD 3ζ").

FIG. 3 Flow cytometry plot showing CAR expression of B7H3 CAR constructs with a SMASh tag integrated at various positions in the CAR. All constructs with N-terminal SMASh domains were not expressed on the T cell surface, while constructs with a C-terminal SMASh domain were expressed at high levels.

FIG. 4 Panel A: Flow cytometry plot showing the decrease in CAR expression after incubation with 3 μM protease inhibitor (asunaprevir) for various amounts of time. Panels B

and C: Graphs showing the mean fluorescence intensities ("MFIs") of the flow cytometry data.

FIG. 5 Panel A: flow cytometry plot showing the recovery of CAR expression after removal of the protease inhibitor (asunaprevir) by washing. "No SMASh" control is a B7H3 CAR without the SMASh tag. Panel B: Graph showing the MFIs of the flow cytometry data.

FIG. 6 A series of flow cytometry plots of CAR expression and activation/exhaustion-associated markers. The data demonstrate that incubating CAR-SMASh T cells with the protease inhibitor (3 μ M asunaprevir) not only reduces CAR expression but also reduces expression of activation/exhaustion markers.

FIG. 7 Panel A: flow cytometry plot showing the decrease in CAR expression after incubation with the protease inhibitor (3 μ M simeprevir) for various amounts of time. Panel B: Graph showing the MFIs of the flow cytometry data.

FIG. 8 Panel A: Flow cytometry plot showing the recovery of CAR expression after removal of the protease inhibitor (simeprevir) by washing. "No SMASh" control is a B7H3 CAR without the SMASh tag. Panel B: Graph showing the MFIs of the flow cytometry data.

FIG. 9 A series of flow cytometry plots showing a dose-dependent relationship between the protease inhibitor (asunaprevir) and expression of CAR molecules and activation/exhaustion-associated markers.

FIG. 10 Microscope images demonstrating that incubation with the protease inhibitor (asunaprevir) renders T cells resistant to aggregation during *in vitro* culture, whereas T cells having control CAR constructs lacking the SMASh tag are not.

FIG. 11 Graph showing that B7H3 CAR-SMASh T cells which have been incubated with 3 μ M asunaprevir for various amounts of time prior to co-culture with B7H3-expressing Nalm6 GFP tumor cells have reduced cytotoxicity. Control B7H3-CAR T cells (red) which lack a SMASh tag have a strong cytotoxic effect against B7H3-expressing Nalm6 GFP tumor cells, as determined by tumor GFP fluorescence.

FIG. 12 Graph showing that B7H3 CAR-SMASh T cells which have had drug/protease inhibitor removed for various amounts of time prior to co-culture with B7H3-expressing Nalm6 GFP tumor cells retain their cytotoxic effects, as determined by tumor GFP fluorescence.

FIG. 13 Graph showing that B7H3 CAR-SMASh T cells which have been incubated with 3 μ M asunaprevir for various amounts of time prior to co-culture with B7H3 (low)-expressing Nalm6 GFP tumor cells have reduced cytotoxicity.

FIG. 14 Graph showing that B7H3 CAR-SMASh T cells which have had drug/protease inhibitor removed for various amounts of time prior to co-culture with B7H3

(low)-expressing Nalm6 GFP tumor cells retain their cytotoxic effects, as determined by tumor GFP fluorescence.

FIG. 15 Panel A: Graph showing interferon gamma (IFN γ) levels in co-culture supernatant of B7H3 CAR-SMASH T cells which have been incubated in 3 μ M asunaprevir time prior to co-culture with B7H3-expressing Nalm6 GFP. Panel B: Graph showing quantification of IFN γ levels in co-culture supernatant of B7H3 CAR-SMASH T cells which have had drug removed for various amounts of time prior to co-culture with B7H3-expressing Nalm6 GFP.

FIG. 16 Panel A: Graph showing interleukin-2 (IL-2) levels in co-culture supernatant of B7H3 CAR-SMASH T cells which have been incubated in 3 μ M asunaprevir time prior to co-culture with B7H3-expressing Nalm6 GFP. Panel B: Graph showing quantification of IL-2 levels in co-culture supernatant of B7H3 CAR-SMASH T cells which have had drug removed for various amounts of time prior to co-culture with B7H3-expressing Nalm6 GFP.

FIG. 17 Panel A: Flow cytometry plot showing the decrease in GD2 CAR-SMASH expression after incubation with 3 μ M asunaprevir for various amounts of time. Panel B: Graph showing the MFIs of the flow cytometry data.

FIG. 18 Panel A: Flow cytometry plot showing the decrease in CD19 CAR-SMASH expression after incubation with 3 μ M asunaprevir for various amounts of time. Panel B: Graph showing the MFIs of the flow cytometry data.

FIG. 19 Panel A: Flow cytometry plot showing the decrease in CD22 CAR-SMASH expression after incubation with 3 μ M asunaprevir for various amounts of time. Panel B: Graph showing the MFIs of the flow cytometry data.

FIG. 20 Graph showing tumor GFP for CD19 CAR-SMASH, GD2 CAR-SMASH, CD22 CAR-SMASH, and B7H3 CAR-SMASH T cells co-cultured with GFP-labeled tumor cells expressing cognate antigen in the presence of 3 μ M asunaprevir. The residual expression of GD2 and CD19 CAR-SMASH constructs in the “off” state results in significant cytotoxicity against tumor cells. However, B7H3 CAR-SMASH T cells exhibit minimal cytotoxicity against target cells in the presence of drug, as determined by tumor GFP fluorescence. Mock untransduced T cells served as a control.

FIG. 21 Graph showing the dose response of B7H3 CAR-SMASH expression to either of three different protease inhibitors: asunaprevir (ASV), danoprevir (DPV), or grazoprevir (GPV).

FIG. 22 Graph showing the expression of B7H3 CAR-nanoluciferase-SMASH on T cells in the presence of 3 μ M grazoprevir for various amounts of time, as determined by flow cytometry or bioluminescence. Nanoluciferase bioluminescence serves as a reporter for CAR expression, which can facilitate CAR tracking *in vivo*.

FIG. 23 Panel A: Graph showing MFIs of T cells expressing B7H3 CAR-SMASH vs B7H3 CAR-nLuc-SMASH in the presence or absence of 3 μ M grazoprevir. Inclusion of a nanoluciferase domain reduces the residual expression of CAR in the presence of drug. Panel B: Graph showing the MFIs of T cells expressing these constructs after incubation with 3 μ M grazoprevir for various amounts of time. The reduced half-life of CAR-nLuc-SMASH results in a faster drug response time relative to CAR-SMASH.

FIG. 24 *In vivo* mouse imaging data demonstrating drug-regulatable cytotoxic capacity of CAR T cells *in vivo*.

FIG. 25 Real-time expression of a C-terminal nanoluciferase (nLuc) domain-containing CAR as monitored *in vivo* by nanoluciferase-based imaging. As shown, infusion of a HCV NS3 protease inhibitor diminished CAR expression on the T cells *in vivo*.

FIG. 26 Data demonstrating *in vivo* control of B7H3 CAR-nanoluciferase-SMASH CAR-T cell activity against Nalm6 leukemia cells. NSG mice were infused with 10^6 Nalm6 leukemia cells expressing B7H3 antigen and firefly luciferase by tail vein injection. Four days later, mice were infused with 8×10^6 B7H3 CAR-nanoluciferase-SMASH CAR-T cells or mock untransduced control T cells by tail vein injection. Mice in the “CAR OFF” group were administered 50 mg/kg grazoprevir + 25 mg/kg ritonavir two to three times per day by oral gavage. Bioluminescence imaging was used to follow the expansion of the firefly luciferase-expressing Nalm6 cells. Left panel: a series of images showing the bioluminescence of the Nalm6 leukemia cells throughout the time course of the experiment. Right panel: a graph showing the average bioluminescence flux values of the various experimental groups throughout the time course of the experiment. Error bars represent standard error of mean. As seen in the data, B7H3 CAR-nanoluciferase-SMASH CAR-T cells display robust clearance of leukemia cells in the absence of drug but have diminished cytotoxic capacity in the presence of drug, as determined by bioluminescence imaging.

FIG. 27 Data demonstrating *in vivo* control of B7H3 CAR-nanoluciferase-SMASH CAR-T cell activity against a patient-derived medulloblastoma line (MED8A). NSG mice were subcutaneously implanted with 0.5×10^6 MED8A cells that were engineered to express firefly luciferase. Five days later, mice were infused with 5×10^6 B7H3 CAR-nanoluciferase-SMASH CAR-T cells or mock untransduced control T cells by tail vein injection. Mice in the “SMASH OFF” group were administered 50 mg/kg grazoprevir + 25mg/kg ritonavir two to three times per day by oral gavage. Bioluminescence imaging was used to follow the expansion of the firefly luciferase-expressing MED8A cells. Left panel: a series of images showing the bioluminescence of the MED8A cells throughout the time course of the experiment. Right panel: a graph showing the average bioluminescence flux values of the various experimental groups throughout the time course of the experiment.

Error bars represent standard error of mean. As seen in the data, B7H3 CAR-nanoluciferase-SMASH CAR-T cells display robust clearance of MED8A cells in the absence of drug but have diminished cytotoxic capacity in the presence of drug, as determined by bioluminescence imaging.

5 **FIG. 28** *In vivo* CAR expression of mice described in FIG. 27 was assessed by nanoluciferase imaging. Mice were injected with 200 µL of Nano-Glo substrate that was diluted 40x in PBS. After a 6 min incubation period, the mice were imaged on an IVIS Spectrum imager. As seen in the data, *in vivo* CAR expression was diminished in mice that received 50 mg/kg grazoprevir + 25 mg/kg ritonavir two to three times per day by oral
10 gavage.

DETAILED DESCRIPTION

Provided are polypeptides that include, from N-terminus to C-terminus, a chimeric antigen receptor (CAR), a protease, and a degron, where the polypeptide further includes
15 a cleavage site for the protease disposed between the CAR and the degron. Also provided are cells that include such polypeptides (e.g., where the cells express the CAR on their surface) and pharmaceutical compositions including such cells. Nucleic acids that encode the polypeptides, cells including such nucleic acids, and pharmaceutical compositions including such cells, are also provided. Also provided are methods for controlling the
20 expression of a CAR on the surface of a cell, and methods of using the cells of the present disclosure, including methods of using such cells to administer a regulatable CAR cell-based therapy (e.g., a regulatable CAR T cell therapy) to an individual.

Before the polypeptides, nucleic acids, cells, compositions and methods of the present disclosure are described in greater detail, it is to be understood that the
25 polypeptides, nucleic acids, cells, compositions and methods are not limited to particular embodiments described, as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the polypeptides, nucleic acids, cells, compositions and methods will be limited only by the appended claims.

30 Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range, is encompassed within the polypeptides, nucleic acids, cells, compositions and methods. The upper and lower limits of these smaller ranges may independently be
35 included in the smaller ranges, subject to any specifically excluded limit in the stated range.

Where the stated range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the polypeptides, nucleic acids, cells, compositions and methods.

Certain ranges are presented herein with numerical values being preceded by the term “about.” The term “about” is used herein to provide literal support for the exact number that it precedes, as well as a number that is near to or approximately the number that the term precedes. In determining whether a number is near to or approximately a specifically recited number, the near or approximating unrecited number may be a number which, in the context in which it is presented, provides the substantial equivalent of the specifically recited number.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the polypeptides, nucleic acids, cells, compositions and methods belong. Although any polypeptides, nucleic acids, cells, compositions and methods similar or equivalent to those described herein can also be used in the practice or testing of the polypeptides, nucleic acids, cells, compositions and methods, representative illustrative polypeptides, nucleic acids, cells, compositions and methods are now described.

All publications and patents cited in this specification are herein incorporated by reference as if each individual publication or patent were specifically and individually indicated to be incorporated by reference and are incorporated herein by reference to disclose and describe the materials and/or methods in connection with which the publications are cited. The citation of any publication is for its disclosure prior to the filing date and should not be construed as an admission that the present polypeptides, nucleic acids, cells, compositions and methods are not entitled to antedate such publication, as the date of publication provided may be different from the actual publication date which may need to be independently confirmed.

It is noted that, as used herein and in the appended claims, the singular forms “a”, “an”, and “the” include plural referents unless the context clearly dictates otherwise. It is further noted that the claims may be drafted to exclude any optional element. As such, this statement is intended to serve as antecedent basis for use of such exclusive terminology as “solely,” “only” and the like in connection with the recitation of claim elements, or use of a “negative” limitation.

It is appreciated that certain features of the polypeptides, nucleic acids, cells, compositions and methods, which are, for clarity, described in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the polypeptides, nucleic acids, cells, compositions and methods, which

are, for brevity, described in the context of a single embodiment, may also be provided separately or in any suitable sub-combination. All combinations of the embodiments are specifically embraced by the present disclosure and are disclosed herein just as if each and every combination was individually and explicitly disclosed, to the extent that such combinations embrace operable processes and/or compositions. In addition, all sub-combinations listed in the embodiments describing such variables are also specifically embraced by the present polypeptides, nucleic acids, cells, compositions and methods and are disclosed herein just as if each and every such sub-combination was individually and explicitly disclosed herein.

As will be apparent to those of skill in the art upon reading this disclosure, each of the individual embodiments described and illustrated herein has discrete components and features which may be readily separated from or combined with the features of any of the other several embodiments without departing from the scope or spirit of the present methods. Any recited method can be carried out in the order of events recited or in any other order that is logically possible.

POLYPEPTIDES AND NUCLEIC ACIDS

As summarized above, aspects of the present disclosure include polypeptides. The polypeptides include, from N-terminus to C-terminus, a chimeric antigen receptor (CAR), a protease, and a degron, where the polypeptide further includes a cleavage site for the protease disposed between the CAR and the degron. The SMASh concept in the context of the present disclosure is schematically illustrated in FIG. 1. The CAR is linked to a protease (e.g., a protease derived from hepatitis C virus (HCV) nonstructural protein 3 (NS3)) via a linker including a cleavage site for the protease (e.g., a HCV NS3 protease recognition site). In this example, the polypeptide includes a degron on the opposite side of the protease and cleavage site relative to the CAR. As shown in FIG. 1, panel A, upon expression and protein folding, the SMASh tag is removed from the CAR by its internal protease activity in the absence of an inhibitor of the protease, and is trafficked to the cell's protein degradation machinery by the degron. In the absence of an inhibitor of the protease, therefore, the CAR is expressed on the surface of the cell expressing the polypeptide. As shown in FIG. 1, panel B, addition of an inhibitor of the protease induces the rapid degradation of subsequently synthesized copies of the CAR, effectively shutting off further production of the CAR. Details regarding the subject polypeptides and components thereof will now be described in detail.

A "polypeptide" of the present disclosure is a single continuous chain of amino acids that includes a selected CAR of interest, a protease, a degron, and a cleavage site for the protease, which chain does not occur in nature. The degron is connected to the CAR of

interest through a cleavage site (e.g., a cleavable linker including a cleavage site) capable of being recognized by the protease of the polypeptide to allow self-removal of the degron (and optionally, the protease depending on the position of the cleavage site) from the CAR of interest. The position of the cleavage site in the polypeptide may be chosen to allow
5 release of the CAR of interest from the polypeptide essentially unmodified or with little modification, e.g., 20 or fewer, 15 or fewer, 10 or fewer, or 5 or fewer extra amino acids. The CAR is N-terminal to the degron. In some embodiments, the degron is located at the C-terminus of the polypeptide. In some embodiments, the polypeptide may be represented by the formula $\text{NH}_2\text{-X-C-P-D-COOH}$, where: X is an amino acid sequence of the CAR of
10 interest; C is an amino acid sequence of the cleavage site for the protease; P is an amino acid sequence of the protease; and D is an amino acid sequence of the degron. The inventors surprisingly found that only when the SMASh tag is C-terminal to the CAR could the CAR be expressed at acceptable levels on the cell surface (FIG. 3). In some embodiments, the cleavage site is between the CAR and the protease. In certain aspects,
15 the cleavage site is between the protease and the degron.

The CAR, cleavage site, protease and degron constitute domains of the polypeptide which are operably linked to one another. The domains may be contiguous (that is, immediately adjacent one another), or two or more domains may be linked by one or more linkers, spacers, and/or the like.

20 The polypeptide may include domains in addition to the CAR, cleavage site, protease, and degron. For example, the polypeptide may include a spacer domain between the CAR and the cleavage site. The inventors surprisingly found that such a spacer domain may decrease the response time of the polypeptide to an inhibitor of the protease. See, e.g., FIG. 22 and FIG. 23. A spacer domain may be positioned between the CAR and the
25 cleavage site, where the protease is on the opposite side of the cleavage site from the CAR (e.g., $\text{NH}_2\text{-X-S-C-P-D-COOH}$, where X is an amino acid sequence of the CAR of interest; S is an amino acid sequence of a spacer; C is an amino acid sequence of the cleavage site for the protease; P is an amino acid sequence of the protease; and D is an amino acid sequence of the degron). In such a configuration, upon cleavage at the cleavage site by
30 the protease, the spacer domain remains attached to the C-terminus of the CAR. In other aspects, a spacer domain is positioned between the CAR and the cleavage site, where the protease is on the same side of the cleavage site as the CAR (e.g., $\text{NH}_2\text{-X-S-P-C-D-COOH}$, where X is an amino acid sequence of the CAR of interest; S is the amino acid sequence of a spacer; P is an amino acid sequence of the protease; C is an amino acid sequence of
35 the cleavage site for the protease; and D is an amino acid sequence of the degron). In such a configuration, upon cleavage at the cleavage site by the protease, the spacer domain and

protease remain attached to the C-terminus of the CAR. In some embodiments, the spacer domain includes a linker, a reporter, or a combination thereof. Non-limiting examples of reporters are bioluminescence reporters. For example, the reporter may be a luciferase, e.g., a nanoluciferase. A polypeptide of the present disclosure may include one or more domains useful for purification (e.g., a purification tag, such as a FLAG tag, HIS tag, and/or the like), a localization sequence, a domain useful for detecting/imaging the CAR (e.g., a luciferase or other proteinaceous element that enables detection/imaging of the CAR directly or indirectly), and/or the like. The various domains of the subject polypeptides are operably linked to one another, meaning that such domains are linked to one another and retain their respective functions, e.g., protease function of the protease, etc.

The antigen-binding (e.g., antibody) portion of the CAR may specifically bind to the antigen (e.g., a cell surface antigen, such as an antigen on the surface of a cancer cell). The antigen-binding (e.g., antibody) portion of the CAR “specifically binds” to the antigen if it binds to or associates with the antigen with an affinity or K_a (that is, an equilibrium association constant of a particular binding interaction with units of $1/M$) of, for example, greater than or equal to about $10^5 M^{-1}$. In certain embodiments, the antigen-binding (e.g., antibody) portion of the CAR binds to an antigen with a K_a greater than or equal to about $10^6 M^{-1}$, $10^7 M^{-1}$, $10^8 M^{-1}$, $10^9 M^{-1}$, $10^{10} M^{-1}$, $10^{11} M^{-1}$, $10^{12} M^{-1}$, or $10^{13} M^{-1}$. “High affinity” binding refers to binding with a K_a of at least $10^7 M^{-1}$, at least $10^8 M^{-1}$, at least $10^9 M^{-1}$, at least $10^{10} M^{-1}$, at least $10^{11} M^{-1}$, at least $10^{12} M^{-1}$, at least $10^{13} M^{-1}$, or greater. Alternatively, affinity may be defined as an equilibrium dissociation constant (K_D) of a particular binding interaction with units of M (e.g., $10^{-5} M$ to $10^{-13} M$, or less). In certain aspects, specific binding means the antigen-binding (e.g., antibody) portion of the CAR binds to the target molecule with a K_D of less than or equal to about $10^{-5} M$, less than or equal to about $10^{-6} M$, less than or equal to about $10^{-7} M$, less than or equal to about $10^{-8} M$, or less than or equal to about $10^{-9} M$, $10^{-10} M$, $10^{-11} M$, or $10^{-12} M$ or less. The binding affinity of the antigen-binding (e.g., antibody) portion of the CAR for the target antigen can be readily determined using conventional techniques, e.g., by competitive ELISA (enzyme-linked immunosorbent assay), equilibrium dialysis, by using surface plasmon resonance (SPR) technology (e.g., the BIAcore 2000 instrument, using general procedures outlined by the manufacturer); by radioimmunoassay; or the like.

The CAR includes an antigen-binding portion that binds to a target antigen of interest, e.g., a particular antigen on the surface of a target cell. Non-limiting examples of an antigen-binding portion that may be included in the CAR include an antibody, a receptor (e.g., a variable lymphocyte receptor), a receptor fragment (e.g., an Fc receptor fragment), a ligand, a cytokine, a DARPIn, an adnectin, a nanobody, and a peptide. Suitable antigen-

binding portions include those described in Labanieh et al. (2018) *Nature Biomedical Engineering* 2:377-391, the disclosure of which is incorporated herein by reference in its entirety for all purposes.

In certain aspects, the antibody portion of the CAR is an antibody. In some
5 embodiments, the antibody is a single-chain antibody. According to certain embodiments, the single-chain antibody is a single-chain variable fragment (scFv). The single-chain antibody may be a monoclonal antibody, a chimeric antibody, a humanized antibody, a fully human antibody, and/or the like. In certain aspects, the antibody portion of the CAR is an antibody approved by the United States Food and Drug Administration and/or the European
10 Medicines Agency (EMA) for use as a therapeutic antibody (e.g., for antibody-dependent cellular cytotoxicity (ADCC), etc.), or a fragment thereof (e.g., a single-chain version of such an antibody, such as an scFv version of the antibody) that retains the ability to bind the target molecule.

In certain aspects, the antigen-binding portion of the CAR specifically binds a
15 molecule on the surface of a target cell. The target cell may be any cell type of interest. For example, the target cell may be a genetically and/or phenotypically normal cell. In other aspects, the target cell is a genetically and/or phenotypically abnormal cell. Abnormal cells of interest include, but are not limited to, cancer cells. By “cancer cell” is meant a cell exhibiting a neoplastic cellular phenotype, which may be characterized by one or more of,
20 for example, abnormal cell growth, abnormal cellular proliferation, loss of density dependent growth inhibition, anchorage-independent growth potential, ability to promote tumor growth and/or development in an immunocompromised non-human animal model, and/or any appropriate indicator of cellular transformation. “Cancer cell” may be used interchangeably herein with “tumor cell”, “malignant cell” or “cancerous cell”, and encompasses cancer cells
25 of a solid tumor, a semi-solid tumor, a hematological malignancy (e.g., a leukemia cell, a lymphoma cell, a myeloma cell, etc.), a primary tumor, a metastatic tumor, and the like.

In some embodiments, when the target cell is a cancer cell, the molecule on the surface of the cancer cell to which the antigen-binding portion of the CAR binds is a tumor-associated cell surface molecule or a tumor-specific cell surface molecule. By “tumor-associated cell surface molecule” is meant a cell surface molecule expressed on malignant
30 cells with limited expression on cells of normal tissues, a cell surface molecule expressed at much higher density on malignant versus normal cells, or a cell surface molecule that is developmentally expressed.

When the target cell is a cancer cell, the cancer cell may express on its surface a
35 tumor-associated molecule or tumor-specific molecule to which the antigen-binding portion of the CAR binds. In certain embodiments, such a tumor-associated molecule or tumor-

specific molecule is selected from HER2, B7-H3 (CD276), CD19, CD20, GD2, CD22, CD30, CD33, CD56, CD66/CEACAM5, CD70, CD74, CD79b, CD123, CD133, CD138, CD171, Nectin-4, Mesothelin, Transmembrane glycoprotein NMB (GPNMB), Prostate-Specific Membrane Antigen (PSMA), SLC44A4, CA6, tyrosine-protein kinase Met (c-Met),
 5 epidermal growth factor receptor variant III (EGFRvIII), mucin 1 (MUC1), ephrin type-A receptor 2 (EphA2), glypican 2 (GPC2), glypican 3 (GPC3), fms-like tyrosine kinase 3 (FLT3), folate receptor alpha (FR α), IL-13 receptor alpha 2 (IL13R α 2), fibroblast activation protein (FAP), receptor tyrosine kinase-like orphan receptor 1 (ROR1), B-cell maturation antigen (BCMA), delta-like 3 (DLL3), κ light chain, vascular endothelial growth factor
 10 receptor 2 (VEGFR2), Trophoblast glycoprotein (TPBG), anaplastic lymphoma kinase (ALK), CA-IX, an integrin, C-X-C chemokine receptor type 4 (CXCR4), neuropilin-1 (NRP1), matriptase, and any other tumor-associated or tumor-specific molecules of interest.

Non-limiting examples of antibodies which may be employed in the CAR of a polypeptide of the present disclosure include Adecatumumab, Ascrinvacumab,
 15 Cixutumumab, Conatumumab, Daratumumab, Drozitumab, Duligotumab, Durvalumab, Dusigitumab, Enfortumab, Enoticumab, Figitumumab, Ganitumab, Glembatumumab, Intetumumab, Ipilimumab, Iratumumab, Icrucumab, Lexatumumab, Lucatumumab, Mapatumumab, Narnatumab, Necitumumab, Nesvacumab, Ofatumumab, Olaratumab, Panitumumab, Patritumab, Pritumumab, Radretumab, Ramucirumab, Rilotumumab,
 20 Robatumumab, Seribantumab, Tarextumab, Teprotumumab, Tovetumab, Vantictumab, Vesencumab, Votumumab, Zalutumumab, Flanvotumab, Altumomab, Anatumomab, Arcitumomab, Bectumomab, Blinatumomab, Detumomab, Ibritumomab, Minretumomab, Mitumomab, Moxetumomab, Naptumomab, Nofetumomab, Pentumomab, Pintumomab, Racotumomab, Satumomab, Solitumab, Taplitumomab, Tenatumomab, Tositumomab,
 25 Tremelimumab, Abagovomab, Igovomab, Oregovomab, Capromab, Edrecolomab, Nacolomab, Amatuximab, Bavituximab, Brentuximab, Cetuximab, Derlotuximab, Dinutuximab, Ensituximab, Futuximab, Girentuximab, Indatuximab, Isatuximab, Margetuximab, Rituximab, Siltuximab, Ublituximab, Ecomeximab, Abituzumab, Alemtuzumab, Bevacizumab, Bivatuzumab, Brontictuzumab, Cantuzumab, Cantuzumab,
 30 Citatuzumab, Clivatuzumab, Dacetuzumab, Demcizumab, Dalotuzumab, Denintuzumab, Elotuzumab, Emactuzumab, Emibetuzumab, Enoblituzumab, Etaracizumab, Farletuzumab, Ficlatuzumab, Gemtuzumab, Imgatuzumab, Inotuzumab, Labetuzumab, Lifestuzumab, Lintuzumab, Lorvotuzumab, Lumretuzumab, Matuzumab, Milatuzumab, Nimotuzumab, Obinutuzumab, Ocaratuzumab, Otlertuzumab, Onartuzumab,
 35 Oportuzumab, Parsatuzumab, Pertuzumab, Pinatuzumab, Polatuzumab, Sibrotuzumab, Simtuzumab, Tacatuzumab, Tigatuzumab, Trastuzumab, Tucotuzumab, Vandortuzumab,

Vanucizumab, Veltuzumab, Vorsetuzumab, Sofituzumab, Catumaxomab, Ertumaxomab, Depatuxizumab, Ontuxizumab, Blontuvetmab, Tamtuvetmab, or an antigen-binding variant thereof. As used herein, the term “variant” refers to an antibody that binds to a particular cognate antigen (e.g., HER2 for trastuzumab) but has fewer or more amino acids than the parental antibody, has one or more amino acid substitutions relative to the parental antibody, is a single-chain variant (such as an scFv variant) of the parental antibody, or any combination thereof.

The CAR may include the antigen-binding (e.g., an antibody, such as an scFv), a transmembrane domain, and an intracellular signaling domain. A polypeptide that includes such a CAR is schematically illustrated in FIG. 2, panel A. As shown, the polypeptide includes a CAR that includes an antibody (Ab), transmembrane domain (Tm), and intracellular signaling domain (IS), where the polypeptide further includes a cleavage site for a protease, the protease, and a degron. A specific example of such a polypeptide, employed in the Experimental section herein, is shown in FIG. 2, panel B. In this example, the CAR includes an anti-B7-H3 (anti-CD276) scFv as the antibody (“B7H3 scFv”), a CD 8 transmembrane domain (“CD 8 Tm”), and a signaling endodomain including 4-1BB and CD3 zeta (“4-1BB” and “CD 3ζ”).

In some embodiments, the CAR includes one or more linker sequences between the various domains. A “variable region linking sequence” is an amino acid sequence that connects a heavy chain variable region to a light chain variable region and provides a spacer function compatible with interaction of the two sub-binding domains so that the resulting polypeptide retains a specific binding affinity to the same target molecule as an antibody that includes the same light and heavy chain variable regions. A non-limiting example of a variable region linking sequence is a serine-glycine linker, such as a serine-glycine linker that includes the amino acid sequence GGGSGGGSGGGGS (G₄S)₃ (SEQ ID NO:3). In certain aspects, a linker separates one or more heavy or light chain variable domains, hinge domains, transmembrane domains, co-stimulatory domains, and/or primary signaling domains. In particular embodiments, the CAR includes one, two, three, four, or five or more linkers. In particular embodiments, the length of a linker is about 1 to about 25 amino acids, about 5 to about 20 amino acids, or about 10 to about 20 amino acids, or any intervening length of amino acids. In some embodiments, the linker is 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or more amino acids long.

In some embodiments, the binding domain of the CAR is followed by one or more spacer domains that moves the antigen binding domain away from the effector cell surface (e.g., the surface of a T cell expressing the CAR) to enable proper cell/cell contact, antigen binding and/or activation. The spacer domain (and any other spacer domains, linkers,

and/or the like described herein) may be derived either from a natural, synthetic, semi-synthetic, or recombinant source. In certain embodiments, a spacer domain is a portion of an immunoglobulin, including, but not limited to, one or more heavy chain constant regions, e.g., CH2 and CH3. The spacer domain may include the amino acid sequence of a naturally occurring immunoglobulin hinge region or an altered immunoglobulin hinge region. In one embodiment, the spacer domain includes the CH2 and/or CH3 of IgG1, IgG4, or IgD. Illustrative spacer domains suitable for use in the CARs described herein include the hinge region derived from the extracellular regions of type 1 membrane proteins such as CD8 α and CD4, which may be wild-type hinge regions from these molecules or variants thereof. In certain aspects, the hinge domain includes a CD8 α hinge region. In some embodiments, the hinge is a PD-1 hinge or CD152 hinge.

The "transmembrane domain" (TM domain) is the portion of the CAR that fuses the extracellular binding portion and intracellular signaling domain and anchors the CAR to the plasma membrane of the cell (e.g., immune effector cell). The Tm domain may be derived either from a natural, synthetic, semi-synthetic, or recombinant source. In some embodiments, the Tm domain is derived from (e.g., includes at least the transmembrane region(s) or a functional portion thereof) of the alpha or beta chain of the T-cell receptor, CD35, CD3 ζ , CD3 γ , CD3 δ , CD4, CD5, CD8 α , CD9, CD16, CD22, CD27, CD28, CD33, CD37, CD45, CD64, CD80, CD86, CD134, CD137, CD152, CD154, and PD-1.

In one embodiment, a CAR includes a Tm domain derived from CD8 α . In certain aspects, a CAR includes a Tm domain derived from CD8 α and a short oligo- or polypeptide linker, e.g., between 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acids in length, that links the Tm domain and the intracellular signaling domain of the CAR. A glycine-serine linker may be employed as such a linker, for example.

The "intracellular signaling" domain of a CAR refers to the part of a CAR that participates in transducing the signal from CAR binding to a target molecule/antigen into the interior of the immune effector cell to elicit effector cell function, e.g., activation, cytokine production, proliferation and/or cytotoxic activity, including the release of cytotoxic factors to the CAR-bound target cell, or other cellular responses elicited with target molecule/antigen binding to the extracellular CAR domain. Accordingly, the term "intracellular signaling domain" refers to the portion of a protein which transduces the effector function signal and that directs the cell to perform a specialized function. To the extent that a truncated portion of an intracellular signaling domain is used, such truncated portion may be used in place of a full-length intracellular signaling domain as long as it transduces the effector function signal. The term intracellular signaling domain is meant to

include any truncated portion of an intracellular signaling domain sufficient for transducing effector function signal.

Signals generated through the T cell receptor (TCR) alone are insufficient for full activation of the T cell and a secondary or costimulatory signal is also required. Thus, T cell activation is mediated by two distinct classes of intracellular signaling domains: primary signaling domains that initiate antigen-dependent primary activation through the TCR (e.g., a TCR/CD3 complex) and costimulatory signaling domains that act in an antigen-independent manner to provide a secondary or costimulatory signal. As such, a CAR of the present disclosure may include an intracellular signaling domain that includes one or more “costimulatory signaling domains” and a “primary signaling domain.”

Primary signaling domains regulate primary activation of the TCR complex either in a stimulatory manner, or in an inhibitory manner. Primary signaling domains that act in a stimulatory manner may contain signaling motifs which are known as immunoreceptor tyrosine-based activation motifs (or “ITAMs”). Non-limiting examples of ITAM-containing primary signaling domains suitable for use in a CAR of the present disclosure include those derived from FcR γ , FcR β , CD3 γ , CD3 δ , CD3 ϵ , CD3 ζ , CD22, CD79 α , CD79 β , and CD66 δ . In certain embodiments, a CAR includes a CD3 ζ primary signaling domain and one or more costimulatory signaling domains. The intracellular primary signaling and costimulatory signaling domains are operably linked to the carboxyl terminus of the transmembrane domain.

In some embodiments, the CAR includes one or more costimulatory signaling domains to enhance the efficacy and expansion of T cells expressing the CAR. As used herein, the term “costimulatory signaling domain” or “costimulatory domain” refers to an intracellular signaling domain of a costimulatory molecule or an active fragment thereof. Example costimulatory molecules suitable for use in CARs contemplated in particular embodiments include TLR1, TLR2, TLR3, TLR4, TLR5, TLR6, TLR7, TLR8, TLR9, TLR10, CARD11, CD2, CD7, CD27, CD28, CD30, CD40, CD54 (ICAM), CD83, CD134 (OX40), CD137 (4-1BB), CD278 (ICOS), DAP10, LAT, KD2C, SLP76, TRIM, and ZAP70. In some embodiments, the CAR includes one or more costimulatory signaling domains selected from the group consisting of 4-1BB, CD28, CD137, and CD134, and a CD3 ζ primary signaling domain.

In certain aspects, a CAR of a polypeptide of the present disclosure includes an antigen-binding portion (e.g., a single chain antibody, such as an scFv) that binds to an antigen of interest; a transmembrane domain from a polypeptide selected from the group consisting of: CD4, CD8 α , CD154, and PD-1; one or more intracellular costimulatory signaling domains from a polypeptide selected from the group consisting of: 4-1BB, CD28,

CD134, and CD137; and an intracellular signaling domain from a polypeptide selected from the group consisting of: FcR γ , FcR β , CD3 γ , CD3 δ , CD3 ϵ , CD3 ζ , CD22, CD79 α , CD79 β , and CD66 δ . Such a CAR may further include a spacer domain between the antigen-binding portion and the transmembrane domain, e.g., a CD8 alpha hinge.

5 As summarized above, the polypeptides of the present disclosure include a degron. As used herein, a “degron” is a sequence of amino acids which provides a degradation signal that directs a polypeptide to intracellular pathways for proteolytic degradation. The degron may promote degradation of an attached polypeptide through either the proteasome or autophagy-lysosome pathways. In the polypeptide, the degron is operably linked to the
 10 CAR but need not be contiguous with it so long as the degron still functions to direct degradation of the CAR when the degron and CAR are linked (e.g., in the presence of an inhibitor of the protease such that the protease is not able to cleave the cleavage site and, therefore, the degron and CAR remain linked). In some embodiments, the degron induces rapid degradation of the polypeptide of interest. For a discussion of degrons and their
 15 function in protein degradation, see, e.g., Kanemaki et al. (2013) *Pflugers Arch.* 465(3):419-425, Eralles et al. (2014) *Biochim Biophys Acta* 1843(1):216-221, Schrader et al. (2009) *Nat. Chem. Biol.* 5(11):815-822, Ravid et al. (2008) *Nat. Rev. Mol. Cell. Biol.* 9(9):679-690, Tasaki et al. (2007) *Trends Biochem Sci.* 32(11):520-528, Meinnel et al. (2006) *Biol. Chem.* 387(7):839-851, Kim et al. (2013) *Autophagy* 9(7): 1100-1103, Varshavsky (2012) *Methods*
 20 *Mol. Biol.* 832: 1-11, and Fayadat et al. (2003) *Mol Biol Cell.* 14(3): 1268-1278; the disclosures of which are incorporated herein by reference in their entireties for all purposes.

In certain aspects, the degron is one found in p53, HIF1alpha, ubiquitin, or a functional variant thereof. In some embodiments, the degron includes portions of the HCV nonstructural proteins NS3 and NS4A. In certain aspects, the degron includes the amino
 25 acid sequence of SEQ ID NO:1 (the amino acid sequence of a degron from HCV genotype 1a), or a functional variant thereof having 70% or greater, 75% or greater, 80% or greater, 85% or greater, 90% or greater, 95% or greater, or 99% or greater amino acid sequence identity to the amino acid sequence of SEQ ID NO:1, or a fragment thereof, such as a
 30 fragment having a length of from 30 to 41 amino acids, 32 to 41 amino acids, 34 to 41 amino acids, 36 to 41 amino acids, or 38 to 41 amino acids, wherein a functional variant of the degron of SEQ ID NO:1 is capable of promoting degradation of the CAR when the CAR and degron are linked. It is to be understood that degrons including the residues corresponding to the reference sequence of SEQ ID NO:1 in HCV nonstructural proteins NS3 and NS4A obtained from other strains of HCV are also encompassed by the present disclosure.

Example Degron Amino Acid Sequence

PITKIDTKYIMTCMSADLEVVTSTWVLVGGVLAALAAYCLST (SEQ ID NO:1)

In certain aspects, a “variant” of any of the polypeptides or domains thereof of the present disclosure contains one or more conservative substitutions. A “conservative substitution” is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydrophobic nature of the polypeptide to be substantially unchanged. Modifications may be made in the structure of the polynucleotides and polypeptides contemplated in particular embodiments, polypeptides include polypeptides having at least about and still obtain a functional molecule that encodes a variant or derivative polypeptide with desirable characteristics. When it is desired to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, variant polypeptide, one skilled in the art, for example, can change one or more of the codons of the encoding DNA sequence.

As summarized above, the polypeptides of the present disclosure include a protease. The protease chosen for use in the polypeptide may be highly selective for the cleavage site in the polypeptide. Additionally, protease activity is preferably inhibitable with inhibitors that are cell-permeable and not toxic to the cell or individual under study or treatment. For a discussion of proteases, see, e.g., V. Y. H. Hook, *Proteolytic and cellular mechanisms in prohormone and proprotein processing*, RG Landes Company, Austin, Tex., USA (1998); N. M. Hooper et al., *Biochem. J.* 321: 265-279 (1997); Z. Werb, *Cell* 91: 439-442 (1997); T. G. Wolfsberg et al., *J. Cell Biol.* 131: 275-278 (1995); K. Murakami and J. D. Etlinger, *Biochem. Biophys. Res. Comm.* 146: 1249-1259 (1987); T. Berg et al., *Biochem. J.* 307: 313-326 (1995); M. J. Smyth and J. A. Trapani, *Immunology Today* 16: 202-206 (1995); R. V. Talanian et al., *J. Biol. Chem.* 272: 9677-9682 (1997); and N. A. Thornberry et al., *J. Biol. Chem.* 272: 17907-17911 (1997), the disclosures of which are incorporated herein by reference in their entireties for all purposes.

In some embodiments, the protease employed in the polypeptide is a sequence-specific non-human protease for which pharmacological inhibitors are available. In certain aspects, the protease employed in the polypeptide is a viral protease. Non-limiting examples of viral proteases that may be employed in the polypeptides of the present disclosure include a hepatitis C virus (HCV) protease, a rhinovirus protease, a coxsackie virus protease, a dengue virus protease, and a tev protease. As just one example, the viral protease may be a HCV protease. In certain embodiments, the viral protease is derived from HCV nonstructural protein 3 (NS3). NS3 consists of an N-terminal serine protease

domain and a C-terminal helicase domain. By “derived from HCV NS3” is meant the protease is the serine protease domain of HCV NS3 or a proteolytically active variant thereof capable of cleaving a cleavage site for the serine protease domain of HCV NS3. The protease domain of NS3 forms a heterodimer with the HCV nonstructural protein 4A (NS4A), which activates proteolytic activity. A protease derived from HCV NS3 may include the entire NS3 protein or a proteolytically active fragment thereof, and may further include a cofactor polypeptide, such as a cofactor polypeptide derived from HCV nonstructural protein 4A (NS4A), e.g., an activating NS4A region. NS3 protease is highly selective and can be well-inhibited by a number of non-toxic, cell-permeable drugs, which are currently clinically available. NS3 protease inhibitors that may be employed include, but are not limited to, simeprevir, danoprevir, asunaprevir, ciluprevir, boceprevir, sovalprevir, paritaprevir and telaprevir. A non-limiting example of a protease derived from HCV NS3 is provided below.

Example Protease Derived from HCV NS3

APITAYAQQTRGLLGCIITSLTGRDKNQVEGEVQIVSTATQTFLATCINGVCWAVYHG
AGTRTIASPKGPVIQMYTNVDQDLVGWPAPQGSRLTPCTCGSSDLYLVTRHADVIP
VRRRGDSRGSLLSPRPISYLGSSGGPLLCPAGHAVGLFRAAVCTRGVAKAVDFIPV
ENLETTMRSPVFTD (SEQ ID NO:2)

In some embodiments, the protease includes the sequence set forth in SEQ ID NO:2 or is a functional (proteolytic) variant thereof having 70% or greater, 75% or greater, 80% or greater, 85% or greater, 90% or greater, 95% or greater, or 99% or greater amino acid sequence identity to SEQ ID NO:2, and/or a functional (proteolytic) fragment thereof such as a fragment having a length of from 100 to 185, 120 to 185, 140 to 185, 160 to 185, 170 to 185, from 180 to 185, from 182 to 185, or from 184 to 185 amino acids.

When a protease derived from HCV NS3 is employed in a polypeptide of the present disclosure, the cleavage site should include a NS3 protease cleavage site. An NS3 protease cleavage site may include the four junctions between nonstructural (NS) proteins of the HCV polyprotein normally cleaved by the NS3 protease during HCV infection, including the NS3/NS4A, NS4A/NS4B, NS4B/NS5A, and NS5A/NS5B junction cleavage sites. For a description of NS3 protease and representative sequences of its cleavage sites for various strains of HCV, see, e.g., *Hepatitis C Viruses: Genomes and Molecular Biology* (S.L. Tan ed., Taylor & Francis, 2006), Chapter 6, pp. 163-206; the disclosure of which is incorporated herein by reference in its entirety.

In some embodiments, the protease is derived from HCV NS3 and engineered to include one or more amino acid substitutions relative to the amino acid sequence set forth

in SEQ ID NO:2. For example, the protease may include a substitution at the position corresponding to position 54 of the amino acid sequence set forth in SEQ ID NO:2. In certain aspects, such a substitution is a threonine to alanine substitution.

NS3 nucleic acid and protein sequences may be derived from HCV, including any isolate of HCV having any genotype (e.g., seven genotypes 1–7) or subtype. A number of NS3 nucleic acid and protein sequences are known and described, e.g., in USSN 15/737,712, the disclosure of which is incorporated herein by reference in their entirety for all purposes. Additional representative NS3 sequences are listed in the National Center for Biotechnology Information (NCBI) database. See, for example, NCBI entries: Accession Nos. YP_001491553, YP_001469631, YP_001469632, NP_803144, NP_671491, YP_001469634, YP_001469630, YP_001469633, ADA68311, ADA68307, AFP99000, AFP98987, ADA68322, AFP99033, ADA68330, AFP99056, AFP99041, CBF60982, CBF60817, AHH29575, AIZ00747, AIZ00744, ABI36969, ABN05226, KF516075, KF516074, KF516056, AB826684, AB826683, JX171009, JX171008, JX171000, EU847455, EF154714, GU085487, JX171065, and JX171063; all of which sequences are herein incorporated by reference. Any of these sequences or functional variants thereof having 70% or greater, 75% or greater, 80% or greater, 85% or greater, 90% or greater, 95% or greater, or 99% or greater amino acid sequence identity to any one of these sequences, or proteolytic fragments thereof, may be employed.

NS4A nucleic acid and protein sequences may be derived from HCV, including any isolate of HCV having any genotype (e.g., seven genotypes 1-7) or subtype. A number of NS4A nucleic acid and protein sequences are known. Representative NS4A sequences are listed in the National Center for Biotechnology Information (NCBI) database. See, for example, NCBI entries: Accession Nos. NP_751925, YP_001491554, GU945462, HQ822054, FJ932208, FJ932207, FJ932205, and FJ932199; all of which sequences (as entered by the date of filing of this application) are herein incorporated by reference. Any of these sequences or functional variants thereof having 70% or greater, 75% or greater, 80% or greater, 85% or greater, 90% or greater, 95% or greater, or 99% or greater amino acid sequence identity to any one of these sequences, or proteolytic fragments thereof, may be employed.

HCV polyprotein nucleic acid and protein sequences may be derived from HCV, including any isolate of HCV having any genotype (e.g., seven genotypes 1–7) or subtype. A number of HCV polyprotein nucleic acid and protein sequences are known. Representative HCV polyprotein sequences are listed in the National Center for Biotechnology Information (NCBI) database. See, for example, NCBI entries: Accession Nos. YP_001469631, NP_671491, YP_001469633, YP_001469630, YP_001469634,

YP_001469632, NC_009824, NC_004102, NC_009825, NC_009827, NC_009823, NC_009826, and EF108306; all of which sequences (as entered by the date of filing of this application) are herein incorporated by reference. Any of these sequences or functional variants thereof having 70% or greater, 75% or greater, 80% or greater, 85% or greater, 90% or greater, 95% or greater, or 99% or greater amino acid sequence identity to any one of these sequences, or proteolytic fragments thereof, may be employed.

As summarized above, the polypeptides of the present disclosure include a cleavage site for the protease, where the cleavage site is disposed between the cell surface molecule and the degron. The term "cleavage site" refers to the bond (e.g., a scissile bond) cleaved by an agent. A cleavage site for a protease includes the specific amino acid sequence recognized by the protease during proteolytic cleavage and may include surrounding amino acids (e.g., from one to six amino acids) on either side of the scissile bond, which bind to the active site of the protease and are needed for recognition as a substrate. In some embodiments, the cleavage site is provided as a cleavable linker, where "cleavable linker" refers to a linker including the cleavage site. A cleavable linker may include a cleavage site specific for an enzyme, such as a protease or other cleavage agent. A cleavable linker is typically cleavable under physiological conditions.

In some embodiments, the protease of the polypeptide is derived from HCV NS3 and the cleavage site includes an NS3 protease cleavage site. An NS3 protease cleavage site may include the HCV polyprotein NS3/NS4A, NS4A/NS4B, NS4B/NS5A, and NS5A/NS5B junction cleavage sites. A representative HCV NS4A/4B protease cleavage site is DEMEECSQHL (SEQ ID NO:4). A representative HCV NS5A/5B protease cleavage site is EDVVPCSMG (SEQ ID NO:5).

In certain aspects, a polypeptide of the present disclosure includes a SMASh tag that includes the amino acid sequence set forth in SEQ ID NO:6 (C-terminal SMASh tag with NS4A/4B protease cleavage site), or a functional variant thereof having 70% or greater, 75% or greater, 80% or greater, 85% or greater, 90% or greater, 95% or greater, or 99% or greater amino acid sequence identity to such a SMASh tag, where the polypeptide includes a degron operably linked to the cell surface molecule of interest, and/or a functional fragment thereof (e.g., having from 290 to 305, 295 to 305, or 300 to 305 amino acids), which is capable of promoting degradation of the cell surface molecule of interest and includes a protease capable of cleaving the polypeptide at a cleavage site in the absence of an inhibitor of the protease.

Example C-terminal SMASh tag with NS4A/4B Protease Cleavage Site

PGDEMEEC SQHLP GAGSSG DIMDYK DDDDKG SSGT GSGSGT SAPITAYAQQTRGL
 LGCIITSLTGRDKNQVEGEVQIVSTATQTFLATCINGVCWAVYHGAGTRRTIASPKGPVI
 QMYTNVDQDLVGWPAPQGSRS LTPCTCGSSDLYLVTRHADVIPVRRRGDSRGSLLS
 5 PRPISYLK GSSGG PLLCPAGHAVGLFRAAVCTRGVAKAVDFIPVENLETTMRSPVFT
 DNSSPPAVTLTHPITKIDTKYIMTCMSADLEVVTSTWVLVGGVLAALAAAYCLSTGCVVI
 VGRIVLSGKPAIIPDREVLV (SEQ ID NO:6)

Also provided by the present disclosure are nucleic acids encoding any of the polypeptides described herein, including any of the polypeptides having any of the features (e.g., domains, etc.) and combinations thereof described hereinabove. As a result of the degeneracy of the genetic code, there are many nucleotide sequences that may encode the polypeptides of the present disclosure. Some of these polynucleotides may bear minimal homology to the nucleotide sequence of any native gene. Polynucleotides that vary due to differences in codon usage are specifically contemplated in particular embodiments, for example polynucleotides that are optimized for human and/or primate codon selection.

Shown in Table 1 below are amino acid sequences of example polypeptides of the present disclosure (from N- to C-terminus), and nucleotide sequences of nucleic acids encoding the polypeptides. The polypeptides and nucleic acids are those employed in the Experimental section below. Not shown are signal sequences initially present at the N-termini of the polypeptides. For each polypeptide presented in Table 1 below, the CAR portion is shown in bold. Segments/domains of the polypeptides are indicated by alternating underlining, and the identities of the segments/domains are provided in the left column.

25 **Table 1 – Example Polypeptides**

B7H3-BBz-SMASh amino acid sequence (SEQ ID NO:7) CAR • B7H3 scFv (MGA271) heavy chain • linker • B7H3 scFv (MGA271) light chain • CD8 alpha hinge	<u>EVQLVESGGGLVQPGGSLRLSCAASGFTFSSFGMHWVRQ</u> <u>APGKGLEWVAYISSDSSAIYYADTVKGRFTISRDNKNSLY</u> <u>LQMNSLRDEDTAVYYCGRGRENIYYGSRLDYWGQGT</u> <u>VSSGGGGSGGGSGGGGSDIQLTQSPSFLSASVGDRVTIT</u> <u>CKASQNVDTNVAWYQQKPKGKAPKALIYSASYRYS</u> <u>SGVPSR</u> <u>FSGSGSGTDFTLTISSLQPEDFATYYCQQYNYPFTFGQGT</u> <u>KLEIKAAATTPAPRPPTPAPTIASQPLSLRPEACRPAAGG</u> <u>AVHTRGLDFACDIYIWAPLAGTCGVLLLSLVITLYCKRGRK</u> <u>KLLYIFKQPFMRPVQTTQEEDGCSCRFPEEEEGGCELRVK</u> <u>FSRSADAPAYKQQQNQLYNELNLGRREEYDVLDKRRGRD</u> <u>PEMGGKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERR</u>
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• CD8 alpha transmembrane domain • 4-1BB costimulatory domain • CD3 zeta domain SMASh tag • cleavage site (<i>italicized</i>)-containing domain • NS3 protease domain • degron domain • NS4A cofactor domain	<u>RGKGHDGLYQGLSTATKDTYDALHMQALPPR</u><i>PGDEMEEC</i> <i>SQHLP</i>GAGSSGDIMDYKDDDDKGSSGTGSGSGTS<i>APITAY</i> <i>AQQTRGLLGCIITSLTGRDKNQVEGEVQIVSTATQTFLATCIN</i> <i>GVCWAVYHGAGTRT</i>IASPKGPVIQMYTNVDQDLVGWPAPQ <i>GSRSLTPCTCGSSDLYLVTRHADVIPVRRRGDSRGSLLSPR</i> <i>PISYLGSSSGGPLLC</i>PAGHAVGLFRAAVCTRGVAKAVDFIPV <i>ENLETTMRSPVFTDNSSPPAVTLTHPITKIDTKYIMTCMSADL</i> <i>EVVTSTWVLVGGVLAALAAAYCLSTG</i><i>CVVIVGRIVLSGKPAIIP</i> <i>DREVL</i><i>Y</i>
B7H3-BBz-SMASh nucleotide sequence (SEQ ID NO:8)	GAGGTGCAGCTGGTGAATCTGGCGGCGGACTGGTGCA GCCTGGCGGATCTCTGAGACTGAGCTGTGCCGCCAGCG GCTTCACCTTCAGCAGCTTCGGAATGCACTGGGTGCGCC AGGCCCTGGCAAAGGACTGGAATGGGTGGCCTACATCA GCAGCGACAGCAGCGCCATCTACTACGCCGACACCGTG AAGGGCCGGTTCACCATCTCCCGGGACAACGCCAAGAAC AGCCTGTACCTGCAGATGAACTCCCTGCGGGACGAGGAC ACCGCCGTGTACTATTGCGGCAGAGGCAGAGAGAACATC TATTACGGCAGCAGACTGGACTACTGGGGCCAGGGCACA ACCGTGACAGTGTCTAGCGGAGGCGGAGGATCAGGCGG CGGAGGAAGTGGCGGAGGGGGATCTGATATCCAGCTGA CCCAGAGCCCCAGCTTCCTGAGCGCCTCTGTGGGCGAC AGAGTGACCATCACATGCAAGGCCAGCCAGAACGTGGAC ACCAACGTGGCCTGGTATCAGCAGAACCCCGGCAAGGC CCCTAAGGCCCTGATCTACAGCGCCAGCTACCGGTACAG CGGCGTGCCAGCAGATTTTCTGGCAGCGGCTCCGGCA CCGACTTCACCCTGACAATCAGCAGCCTGCAGCCCGAGG ACTTCGCCACCTACTACTGCCAGCAGTACAACAACCTACCC TTTACCTTCGGCCAGGGGACCAAGCTGGAAATCAAAGC GGCCGCAACAACAACACCAGCTCCTAGACCTCCAACCTCC TGCTCCTACAATCGCCAGCCAGCCTCTGTCTCTGAGGCC TGAAGCTTGTAGACCTGCTGCTGGCGGAGCCGTGCATAC CAGAGGACTGGATTTGCGCTGCGACATCTACATTTGGGC CCCTCTGGCTGGAACATGTGGCGTGCTGCTGTCTCT GGTCATCACCTGTACTGCAAGCGGGGCAGAAAGAAGCT GCTGTACATCTTCAAGCAGCCCTTCATGCGGCCCGTGCA GACCACACAAGAGGAAGATGGCTGCTCCTGCAGATTCCC CGAGGAAGAAGAAGGCGGCTGCGAGCTGAGAGTGAAGT TCTCTAGATCTGCCGACGCTCCCGCTACAAGCAGGGCC AGAATCAGCTGTACAACGAGCTGAACCTGGGGAGAAGAG AAGAGTACGACGTGCTGGATAAGCGGAGAGGCAGAGAT

	CCTGAGATGGGCGGCAAGCCCAGACGGAAGAATCCTCA AGAGGGCCTGTATAATGAGCTGCAGAAAGACAAGATGGC CGAGGCCTACAGCGAGATCGGAATGAAGGGCGAGCGCA GAAGAGGCAAGGGCCACGATGGACTGTATCAGGGCCTG AGCACAGCCACCAAGGATACCTATGATGCCCTGCACATG CAGGCCCTTCCACCTAGACCTGGCGACGAAATGGAAGAG TGTAGCCAGCATCTGCCTGGCGCCGGATCTAGCGGCGA CATCATGGACTACAAGGACGACGACGATAAGGGCAGCTC TGGCACAGGCTCTGGCTCTGGAACATCTGCCCCATCAC AGCCTACGCTCAGCAGACAAGAGGCCTGCTGGGCTGTAT CATCACCAGCCTGACCGGCAGAGACAAGAACCAGGTTGA GGGCGAAGTGCAGATCGTGTCTACCGCCACACAGACCTT TCTGGCCACCTGTATCAATGGCGTGTGCTGGGCCGTTTA TCACGGCGCTGGAACCAGAACAAATCGCCTCTCCTAAGGG CCCCGTGATCCAGATGTACACCAACGTGGACCAGGATCT CGTTGGCTGGCCTGCTCCTCAGGGAAGCAGAAGCCTGA CACCTTGATCCTGTGGCAGCAGCGATCTGTACCTGGTCA CAAGACACGCCGATGTGATCCCTGTGCGGAGAAGAGGC GATAGCAGAGGTTCTCTGCTGTCCCCTAGACCTATCAGC TACCTGAAGGGATCTTCTGGCGGCCCTCTGCTGTGTCT GCTGGACATGCTGTGGGACTGTTTAGAGCCGCGTGTGT ACAAGAGGCGTGGCCAAGGCTGTGGACTTCATCCCCGTG GAAAACCTGGAACCACCATGAGAAGCCCCGTGTTACCC GACAATAGCAGCCCTCCAGCCGTGACACTGACACACCCC ATCACCAAGATCGACACCAAGTACATCATGACCTGCATGA GCGCCGACCTGGAAGTGGTCACATCTACCTGGGTTCTCG TTGGAGGCGTTCTGGCTGCCCTGGCCGCTTATTGTCTGT CTACCGGCTGTGTGGTCATCGTGGGCAGAATCGTGCTGT CCGGCAAGCCTGCCATCATTCCCGATAGAGAGGTGCTGT ACTGA
CD19-BBz-SMASH amino acid sequence (SEQ ID NO:9) CAR • CD19 scFv (FMC63) light chain • linker • CD19 scFv (FMC63) heavy chain • CD8 alpha hinge • CD8 alpha transmembrane domain • 4-1BB costimulatory domain	<u>IQMTQTSSLSASLGDRVTISCRASQDISKYLNWYQQKPDG</u> <u>TVKLLIYHTSRLHSGVPSRFSGSGSGTDYSLTISNLEQEDIA</u> <u>TYFCQQGNLTPYTFGGGTKLEITGSTSGSGKPGSGEGSTK</u> <u>GEVKLQESGPGLVAPSQSLSVTCTVSGVSLPDYGVSWIRQ</u> <u>PPRKGLEWLGVWGSETTYYNALKSRLTIKDNSKSQVFL</u> <u>KMNSLQDDTAIYYCAKHYYYGGSYAMDYWGQGSVTVS</u> <u>SAAATTPAPRPPTPAPTIASQPLSLRPEACRPAAGGAVHT</u> <u>RGLDFACDIYWAPLAGTCGVLLLSLVITLYCKRGRKLLYI</u> <u>FKQPFMRPVQTTQEEDGCSCRFPEEEEGGCFLRVKFSRS</u> <u>ADAPAYKQGQNQLYNELNLGRREEYDVLDKRRGRDPENMG</u> <u>GKPRRKNPQEGLYNELQKDKMAEAYSEIGMKGERRRGKG</u> <u>HDGLYQGLSTATKDTYDALHMQALPPR</u>PGDEMEECsqHL

• CD3 zeta domain SMASh tag • cleavage site (<i>italicized</i>)-containing domain • NS3 protease domain • degron domain • NS4A cofactor domain	<u>PGAGSSGDIMDYKDDDDKGSSGTGSGSGTSAPITAYAQQT</u> <u>RGLLGCIITSLTGRDKNQVEGEVQIVSTATQTFLATCINGVC</u> <u>WAVYHGAGTRTIASPKGPVIQMYTNVDQDLVGWPAPQGSR</u> <u>SLTPCTCGSSDLYLVTRHADVIPVRRRGDSRGSLLSPRPISY</u> <u>LKGSSGGPLLCPAGHAVGLFRAAVCTRGVAKAVDFIPVENL</u> <u>ETTMRSPVFETDNSSPPAVTLTHPITKIDTKYIMTCMSADLEVY</u> <u>TSTWVLVGGVLAALAAAYCLSTGCVVIVGRIVLSGKPAIIPDRE</u> <u>VLV</u>
CD19-BBz-SMASh nucleotide sequence (SEQ ID NO:10)	ATCCAGATGACCCAGACCACCAGCAGCCTGAGCGCCAG CCTGGGCGATAGAGTGACCATCAGCTGCAGAGCCAGCC AGGACATCAGCAAGTACCTGAACTGGTATCAGCAGAAAC CCGACGGCACCGTGAAGCTGCTGATCTACCACACCAGCA GACTGCACAGCGGCGTGCCCAGCAGATTTTCTGGCAGC GGCTCCGGCACCGACTACAGCCTGACCATCTCCAACCTG GAACAGGAAGATATCGCTACCTACTTCTGTGTCAGCAAGGC AACACCCTGCCCTACACCTTCGGCGGAGGCACCAAGCTG GAAATCACCGGCAGCACAAGCGGCAGCGGCAAGCCTGG ATCTGGCGAGGGAAGCACCAGGGGCAAGTGAACTGC AGGAAAGCGGCCCTGGACTGGTGGCCCCAAGCCAGTCT CTGAGCGTGACCTGTACCGTGTCCGGCGTGTCCCTGCCT GACTATGGCGTGTCTGGATCAGACAGCCCCCAGAAAG GGCCTGGAATGGCTGGGAGTGATCTGGGGCAGCGAGAC AACCTACTACAACAGCGCCCTGAAGTCCCGGCTGACCAT CATCAAGGACAACCTCCAAGAGCCAGGTGTTCTGAAGAT GAACAGCCTGCAGACCGACGACACCGCCATCTACTACTG CGCCAAGCACTACTACTACGGCGGCAGCTACGCTATGGA CTACTGGGGCCAGGGCACCAGCGTGACCGTGTCTCTG CGGCCGCAACAACAACACCAGCTCCTAGACCTCCAATC CTGCTCCTACAATCGCCAGCCAGCCTCTGTCTCTGAGGC CTGAAGCTTGTAGACCTGCTGCTGGCGGAGCCGTGCATA CCAGAGGACTGGATTTTCGCCTGCGACATCTACATTTGGG CCCCTCTGGCTGGAACATGTGGCGTGCTGCTGTCTCTC TGGTCATCACCTGTACTGCAAGCGGGGCAGAAAGAAGC TGCTGTACATCTTCAAGCAGCCCTTCATGCGGCCCGTGC AGACCACACAAGAGGAAGATGGCTGCTCCTGCAGATTCC CCGAGGAAGAAGAAGGCGGCTGCGAGCTGAGAGTGAAG TTCTCTAGATCTGCCGACGCTCCCGCCTACAAGCAGGGC CAGAATCAGCTGTACAACGAGCTGAACCTGGGGAGAAGA GAAGAGTACGACGTGCTGGATAAGCGGAGAGGCAGAGA TCCTGAGATGGGCGGCAAGCCCAGACGGAAGAATCCTCA AGAGGGCCTGTATAATGAGCTGCAGAAAGACAAGATGGC CGAGGCCTACAGCGAGATCGGAATGAAGGGCGAGCGCA

	GAAGAGGCAAGGGCCACGATGGACTGTATCAGGGCCTG AGCACAGCCACCAAGGATACCTATGATGCCCTGCACATG CAGGCCCTTCCACCTAGACCTGGCGACGAAATGGAAGAG TG TAGCCAGCATCTGCCTGGCGCCGGATCTAGCGGCGA CATCATGGACTACAAGGACGACGACGATAAGGGCAGCTC TGGCACAGGCTCTGGCTCTGGAACATCTGCCCCATCAC AGCCTACGCTCAGCAGACAAGAGGCCTGCTGGGCTGTAT CATCACCAGCCTGACCGGCAGAGACAAGAACCAGGTTGA GGGCGAAGTGCAGATCGTGTCTACCGCCACACAGACCTT TCTGGCCACCTGTATCAATGGCGTGTGCTGGGCCGTTTA TCACGGCGCTGGAACCAGAACAAATCGCCTCTCCTAAGGG CCCCGTGATCCAGATGTACACCAACGTGGACCAGGATCT CGTTGGCTGGCCTGCTCCTCAGGGAAGCAGAAGCCTGA CACCTTGATCCTGTGGCAGCAGCGATCTGTACCTGGTCA CAAGACACGCCGATGTGATCCCTGTGCGGAGAAGAGGC GATAGCAGAGGTTCTCTGCTGTCCCCTAGACCTATCAGC TACCTGAAGGGATCTTCTGGCGGCCCTCTGCTGTGTCT GCTGGACATGCTGTGGGACTGTTTAGAGCCGCGTGTGT ACAAGAGGCGTGGCCAAGGCTGTGGACTTCATCCCCGTG GAAAACCTGGAAACCACCATGAGAAGCCCCGTGTTACCC GACAATAGCAGCCCTCCAGCCGTGACACTGACACACCCC ATCACCAAGATCGACACCAAGTACATCATGACCTGCATGA GCGCCGACCTGGAAGTGGTCACATCTACCTGGGTTCTCG TTGGAGGCGTTCTGGCTGCCCTGGCCGCTTATTGTCTGT CTACCGGCTGTGTGGTCATCGTGGGCAGAATCGTGCTGT CCGGCAAGCCTGCCATCATTCCCGATAGAGAGGTGCTGT ACTGA
GD2-BBz-SMASH amino acid sequence (SEQ ID NO:11) CAR • GD2 scFv (14G2A) • CD8 alpha hinge • CD8 alpha transmembrane domain • 4-1BB costimulatory domain • CD3 zeta domain SMASH tag • cleavage site (italicized)-containing domain • NS3 protease domain	<u>ILLTQTPLSLPVSLGDQASISCRSSQSLVHRNGNTYLHWYL</u> <u>QKPGQSPKLLIHKVSNRFSQVDFSGSGSGTDFTLKISR</u> <u>EAEDLG VYFCSQSTHPPLTFGAGTKLELKGSTSGSGKPG</u> <u>SGEGSTKGEVKLQQSGPSLVEPGASVMISCKASGSSFTGY</u> <u>NMNWVRQNIKGSLEWIG AIDPYYGGTSYNQKFKGRATLTV</u> <u>DKSSSTAYMHLKSLTSEDSAVYYCVSGMEYWGQGSVTV</u> <u>SSAAATTPAPRPPTPAPTIASQPLSLRPEACRPAAGGAV</u> <u>HTRGLDFACDIYIWAPLAGTCGVLLLSLVITLYCKRGRKKLL</u> <u>YIFKQPFMRPVQTTQEEDGCSCRFP EEEEEGGCELRVKFSR</u> <u>SADAPAYKQGQNQLYNELNLGRREEYDVL DKRRGRDPEM</u> <u>GGKPRRKNPQEGLYNELQDKMAEAYSEIGMKGERRRGK</u> <u>GHDGLYQGLSTATKDTYDALHMQALPPR</u> <i>PGDEMEEC</i> <u>SQH</u> <u>LPGAGSSGDIMDYKDDDDKSSSGTSGSGSTS</u> <i>APITAYAQQ</i> <u>TRGLLGCIITSLTGRDNQVEGEVQIVSTATQTFLATCINGVC</u> <u>WAVYHGAGTRTIA SPKGPVIQMYTNVDQDLVGWPAPQGSR</u>

• degron domain • NS4A cofactor domain	<u>SLTPCTCGSSDLYLVTRHADVIPVRRRGDSRGSLLSPRPISY</u> <u>LKGSSGGPLLCPAGHAVGLFRAAVCTRGVAKAVDFIPVENL</u> <u>ETMRSPVFETDNSSPPAVTLTHPITKIDTKYIMTCMSADLEV</u> <u>TSTWVLVGGVLAALAAAYCLSTGCVVIVGRIVLSGKPAIIPDRE</u> <u>VLV</u>
GD2-BBz-SMASH nucleotide sequence (SEQ ID NO:12)	ATCCTGCTGACCCAGACCCCTCTGAGCCTGCCTGTGTCT CTGGGCGATCAGGCCAGCATCAGCTGCAGATCCAGCCA GAGCCTGGTGCACCGGAACGGCAACACCTACCTGCACT GGTATCTGCAGAAGCCCGGCCAGAGCCCCAAGCTGCTG ATTCACAAGGTGTCCAACCGGTTTCAGCGGCGTGCCCGAC AGATTTTCTGGCAGCGGCTCCGGCACCGACTTCACCCCTG AAGATCAGCCGGGTGGAAGCCGAGGACCTGGGCGTGTA CTTCTGCAGCCAGTCCACCCACGTGCCCCCCTGACATT TGGCGCCGGAACAAAGCTGGAAGTGAAGGGCAGCACAA GCGGCAGCGGCAAGCCTGGATCTGGCGAGGGAAGCACC AAGGGCGAAGTGAAGCTGCAGCAGAGCGGCCCTCTCT GGTGGAACCTGGCGCCTCTGTGATGATCTCCTGCAAGGC CAGCGGCAGCTCCTTCACCGGCTACAACATGAACTGGGT GCGCCAGAACATCGGCAAGAGCCTGGAATGGATCGGCG CCATCGACCCCTACTACGGCGGCACCAGCTACAACCAGA AGTTCAAGGGCAGAGCCACCCTGACCGTGGACAAGAGC AGCTCCACCGCCTACATGCACCTGAAGTCCCTGACCAGC GAGGACAGCGCCGTGTACTACTGCGTGTCCGGCATGGA ATACTGGGGCCAGGGCACAAGCGTGACCGTGTCTCTG CGGCCGCAACAACAACACCAGCTCCTAGACCTCCAATC CTGCTCCTACAATCGCCAGCCAGCCTCTGTCTCTGAGGC CTGAAGCTTGTAGACCTGCTGCTGGCGGAGCCGTGCATA CCAGAGGACTGGATTTGCGCTGCGACATCTACATTTGGG CCCCTCTGGCTGGAACATGTGGCGTGTGCTGCTGTCTC TGGTCATCACCTGTACTGCAAGCGGGGCAGAAAGAAGC TGCTGTACATCTTCAAGCAGCCCTTCATGCGGCCCGTGC AGACCACACAAGAGGAAGATGGCTGCTCCTGCAGATTCC CCGAGGAAGAAGAAGGCGGCTGCGAGCTGAGAGTGAAG TTCTCTAGATCTGCCGACGCTCCCGCCTACAAGCAGGGC CAGAATCAGCTGTACAACGAGCTGAACCTGGGGAGAAGA GAAGAGTACGACGTGCTGGATAAGCGGAGAGGCAGAGA TCCTGAGATGGGCGGCAAGCCCAGACGGAAGAATCCTCA AGAGGGCCTGTATAATGAGCTGCAGAAAGACAAGATGGC CGAGGCCTACAGCGAGATCGGAATGAAGGGCGAGCGCA GAAGAGGCAAGGGCCACGATGGACTGTATCAGGGCCTG AGCACAGCCACCAAGGATACCTATGATGCCCTGCACATG CAGGCCCTTCCACCTAGACCTGGCGACGAAATGGAAGAG

	TGTAGCCAGCATCTGCCTGGCGCCGGATCTAGCGGCGA CATCATGGACTACAAGGACGACGACGATAAGGGCAGCTC TGGCACAGGCTCTGGCTCTGGAACATCTGCCCCATCAC AGCCTACGCTCAGCAGACAAGAGGCCTGCTGGGCTGTAT CATCACCAGCCTGACCGGCAGAGACAAGAACCAGGTTGA GGGCGAAGTGCAGATCGTGTCTACCGCCACACAGACCTT TCTGGCCACCTGTATCAATGGCGTGTGCTGGGCCGTTTA TCACGGCGCTGGAACCAGAACAAATCGCCTCTCCTAAGGG CCCCGTGATCCAGATGTACACCAACGTGGACCAGGATCT CGTTGGCTGGCCTGCTCCTCAGGGAAGCAGAAGCCTGA CACCTTGATCCTGTGGCAGCAGCGATCTGTACCTGGTCA CAAGACACGCCGATGTGATCCCTGTGCGGAGAAGAGGC GATAGCAGAGGTTCTCTGCTGTCCCTAGACCTATCAGC TACCTGAAGGGATCTTCTGGCGGCCCTCTGCTGTGTCCT GCTGGACATGCTGTGGGACTGTTTAGAGCCGCCGTGTGT ACAAGAGGCGTGGCCAAGGCTGTGGACTTCATCCCCGTG GAAAACCTGGAAACCACCATGAGAAGCCCCGTGTTACCC GACAATAGCAGCCCTCCAGCCGTGACACTGACACACCCC ATCACCAAGATCGACACCAAGTACATCATGACCTGCATGA GCGCCGACCTGGAAGTGGTCACATCTACCTGGGTTCTCG TTGGAGGCGTTCTGGCTGCCCTGGCCGCTTATTGTCTGT CTACCGGCTGTGTGGTCATCGTGGGCAGAATCGTGCTGT CCGGCAAGCCTGCCATCATTCCCGATAGAGAGGTGCTGT ACTGA
CD22-BBz-SMASH amino acid sequence (SEQ ID NO:13) CAR • CD22 scFv (M971) • CD8 alpha hinge • CD8 alpha transmembrane domain • 4-1BB costimulatory domain • CD3 zeta SMASH tag • cleavage site (italicized)-containing domain • NS3 protease domain • degron domain • NS4A cofactor domain	<u>QVQLQQSGPGLVKPSQTL</u><u>SLTCAISGDSVSSNSAAWNWIR</u> <u>QSPSRGLEWLGRYYRSKWYNDYAVSVKSRITINPDTSKN</u> <u>QFSLQLNSVTPEDTAVYYCAREVTGDLEDAFDIWGQGTMV</u> <u>TVSSGGGSDIQMTQSPSSLSASVGDRVTITCRASQTIWSY</u> <u>LNWYQQRPGKAPNLLIYAASSLQSGVPSRFSGRSGTDF</u> <u>LTISSLQAEDFATYYCQSYSIPQTFGQGTKLEIKAAATTP</u> <u>APRPPTPAPTIASQPLSLRPEACRPAAGGAVHTRGLDFAC</u> <u>DIYIWAPLAGTCGVLLLSLVITLYCKRGRKKLLYIFKQPFMR</u> <u>PVQTTQEEDGCSCRFPEEEEGGCELRVKFSRSADAPAYK</u> <u>QGQNQLYNELNLGRREEYDVLDKRRGRDPEMGGKPRRK</u> <u>NPQEGLYNELQKDKMAEAYSEIGMKGERRRGKGDGLYQ</u> <u>GLSTATKDTYDALHMQALPPR</u><i>PGDEMEEC</i><u>SQHLP</u><u>PGAGSS</u> <u>GDIMDYKDDDDKGSSGTGSGSGTSAPITAYAQQTRGLLGCI</u> <u>ITSLTGRDKNQVEGEVQIVSTATQTFLATCINGVCWAVYHGA</u> <u>GTRTIASPKGPVIQMYTNVDQDLVGWPA PQGSRSLTPCTC</u> <u>GSSDLYLVTRHADVIPVRRRGDSRGSLLSPRPISYLKSSG</u> <u>GPLLCPAGHAVGLFRAAVCTRGVAKAVDFIPVENLETTMRS</u>

	<u>PVFTDNSSPPAVTLTHPITKIDTKYIMTCMSADLEVVTSTWVL</u> <u>VGGVLAALAAAYCLSTGCVVIVGRIVLSGKPAIIPDREVLY</u>
CD22-BBz-SMASH nucleotide sequence (SEQ ID NO:14)	CAGGTGCAGCTGCAGCAGTCTGGCCCTGGCCTCGTGAA GCCTAGCCAGACCCTGAGCCTGACCTGTGCCATCAGCG GCGATAGCGTGTCCAGCAATAGCGCCGCTGGAAGTGG ATCAGACAGAGCCCTAGCAGAGGCCTGGAATGGCTGGG CCGGACCTACTACCGGTCCAAGTGGTACAACGACTACGC CGTGTCCGTGAAGTCCCGGATCACCATCAACCCGACAC CAGCAAGAACCAGTTCTCCCTGCAGCTGAACAGCGTGAC CCCCGAGGATACCGCCGTGTACTACTGCGCCAGAGAAGT GACCGGCGACCTGGAAGATGCCTTCGACATCTGGGGCC AGGGCACAAATGGTCACCGTGTCTAGCGGAGGCGGCGGA AGCGACATCCAGATGACACAGAGCCCCAGCTCCCTGAGC GCCAGCGTGGGAGACAGAGTGACCATCACCTGTGCGGC CAGCCAGACCATCTGGTCCTACCTGAACTGGTATCAGCA GCGGCCTGGCAAGGCCCCCAACCTGCTGATCTATGCCG CCAGCTCACTGCAGAGCGGCGTGCCAGCAGATTTTCCG GCAGAGGCAGCGGCACCGACTTCACCCTGACAATCAGTT CCCTGCAGGCCGAGGACTTCGCCACCTACTACTGCCAGC AGAGCTACAGCATCCCCAGACCTTCGGCCAGGGGACC AAGCTGGAATCAAAGCGGCCGCAACAACAACACCAGCT CCTAGACCTCCAACCTCTGCTCCTACAATCGCCAGCCAG CCTCTGTCTCTGAGGCCTGAAGCTTGTAGACCTGCTGCT GGCGGAGCCGTGCATACCAGAGGACTGGATTTCGCTG CGACATCTACATTTGGGCCCTCTGGCTGGAACATGTGG CGTGCTGCTGCTGTCTCTGGTCATCACCTGTACTGCAA GCGGGGCAGAAAGAAGCTGCTGTACATCTTCAAGCAGCC CTTCATGCGGCCCGTGACAGCACACAAGAGGAAGATGG CTGCTCCTGCAGATTCCCCGAGGAAGAAGAAGGCGGCT GCGAG CTGAGAGTGAAGTTCTCTAGATCTGCCGACGCTCCCGCC TACAAGCAGGGCCAGAATCAGCTGTACAACGAGCTGAAC CTGGGGAGAAGAGAAGAGTACGACGTGCTGGATAAGCG GAGAGGCAGAGATCCTGAGATGGGCGGCAAGCCCAGAC GGAAGAATCCTCAAGAGGGCCTGTATAATGAGCTGCAGA AAGACAAGATGGCCGAGGCCTACAGCGAGATCGGAATGA AGGGCGAGCGCAGAAGAGGCAAGGGCCACGATGGACTG TATCAGGGCCTGAGCACAGCCACCAAGGATACCTATGAT GCCCTGCACATGCAGGCCCTTCCACCTAGACCTGGCGAC GAAATGGAAGAGTGTAGCCAGCATCTG CCTGGCGCCGATCTAGCGGCGACATCATGGACTACAAG GACGACGACGATAAGGGCAGCTCTGGCACAGGCTCTGG

	CTCTGGAACATCTGCCCCTATCACAGCCTACGCTCAGCA GACAAGAGGCCTGCTGGGCTGTATCATCACCAGCCTGAC CGGCAGAGACAAGAACCAGGTTGAGGGCGAAGTGCAGA TCGTGTCTACCGCCACACAGACCTTTCTGGCCACCTGTAT CAATGGCGTGTGCTGGGCGGTTTATCACGGCGCTGGAAC CAGAACAATCGCCTCTCCTAAGGGCCCCGTGATCCAGAT GTACACCAACGTGGACCAGGATCTCGTTGGCTGGCCTGC TCCTCAGGGAAGCAGAAGCCTGACACCTTGACCTGTGG CAGCAGCGATCTGTACCTGGTCACAAGACACGCCGATGT GATCCCTGTGCGGAGAAGAGGCGATAGCAGAGTTCTCT GCTGTCCCCTAGACCTATCAGCTACCTGAAGGGATCTTCT GGCGGCCCTCTGCTGTGTCCTGCTGGACATGCTGTGGG ACTGTTTAGAGCCGCGGTGTGTACAAGAGGCGTGGCCAA GGCTGTGGACTTCATCCCCGTGGAAAACCTGGAAACCAC CATGAGAAGCCCCGTGTTACCGACAATAGCAGCCCTCC AGCCGTGACACTGACACACCCCATCACCAGATCGACAC CAAGTACATCATGACCTGCATGAGCGCCGACCTGGAAGT GGTCACATCTACCTGGGTTCTCGTTGGAGGCGTTCTGGC TGCCCTGGCCGCTTATTGTCTGTCTACCGGCTGTGTGGT CATCGTGGGCAGAATCGTGCTGTCCGGCAAGCCTGCCAT CATTCCCGATAGAGAGGTGCTGTACTGA
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Also provided are expression vectors that include any of the nucleic acids of the present disclosure. A “vector” is capable of transferring nucleic acid sequences to target cells (e.g., viral vectors, non-viral vectors, particulate carriers, and liposomes). Typically, “vector construct,” “expression vector,” and “gene transfer vector,” mean any nucleic acid construct capable of directing the expression of a nucleic acid of interest and which can transfer nucleic acid sequences to target cells. Thus, the term includes cloning and expression vehicles, as well as viral vectors.

In order to express a desired polypeptide, a nucleotide sequence encoding the polypeptide can be inserted into appropriate vector, e.g., using recombinant DNA techniques known in the art. Illustrative examples of viruses useful as vectors include, without limitation, retrovirus (including lentivirus), adenovirus, adeno-associated virus, herpesvirus (e.g., herpes simplex virus), poxvirus, baculovirus, papillomavirus, and papovavirus (e.g., SV40). Illustrative examples of expression vectors include, but are not limited to pCIneo vectors (Promega) for expression in mammalian cells; pLenti4/V 5-DEST™, pLenti6/V 5- DEST™, murine stem cell virus (MSCV), MSGV, moloney murine leukemia virus (MMLV), and pLenti6.2/V5-GW/lacZ (Invitrogen) for lentivirus-mediated gene transfer and expression in mammalian cells. In particular embodiments, a nucleic acid

sequence encoding a polypeptide of the present disclosure may be ligated into such expression vectors for the expression of the polypeptides in mammalian cells.

Expression control sequences, control elements, or regulatory sequences present in an expression vector are those non-translated regions of the vector – origin of replication, selection cassettes, promoters, enhancers, translation initiation signals (Shine Dalgarno sequence or Kozak sequence), introns, a polyadenylation sequence, 5' and 3' untranslated regions, and/or the like – which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including ubiquitous promoters and inducible promoters may be used.

Components of the expression vector are operably linked such that they are in a relationship permitting them to function in their intended manner. In some embodiments, the term refers to a functional linkage between a nucleic acid expression control sequence (such as a promoter, and/or enhancer) and a second polynucleotide sequence, e.g., a nucleic acid encoding the polypeptide, where the expression control sequence directs transcription of the nucleic acid encoding the polypeptide.

In certain aspects, the expression vector is an episomal vector or a vector that is maintained extrachromosomally. As used herein, the term “episomal” refers to a vector that is able to replicate without integration into the host cell's chromosomal DNA and without gradual loss from a dividing host cell also meaning that said vector replicates extrachromosomally or episomally. Such a vector may be engineered to harbor the sequence coding for the origin of DNA replication or “ori” from an alpha, beta, or gamma herpesvirus, an adenovirus, SV40, a bovine papilloma virus, a yeast, or the like. The host cell may include a viral replication transactivator protein that activates the replication. Alpha herpes viruses have a relatively short reproductive cycle, variable host range, efficiently destroy infected cells and establish latent infections primarily in sensory ganglia. Illustrative examples of alpha herpes viruses include HSV 1, HSV 2, and VZV. Beta herpesviruses have long reproductive cycles and a restricted host range. Infected cells often enlarge. Non-limiting examples of beta herpes viruses include CMV, HHV-6 and HHV-7. Gamma-herpesviruses are specific for either T or B lymphocytes, and latency is often demonstrated in lymphoid tissue. Illustrative examples of gamma herpes viruses include EBV and HHV-8.

CELLS

Also provided are cells that include any of the polypeptides, nucleic acids, and/or expression vectors of the present disclosure. In certain aspects, provided are cells where the CAR is expressed on the surface of the cell. By “expressed on the surface of the cell”

is meant the CAR has been trafficked to the cell membrane such that the antigen-binding portion of the CAR is displayed on the cell surface, the transmembrane portion passes through the cell membrane, and the intracellular signaling domain is disposed adjacent to the intracellular side of the cell membrane. Upon binding of the antigen-binding portion of the CAR to the target antigen, the intracellular signaling domain of the CAR participates in transducing the signal from the binding into the interior of the effector cell (e.g., T cell) to elicit effector cell function.

In some embodiments, the cells are eukaryotic cells. Eukaryotic cells of interest include, but are not limited to, yeast cells, insect cells, mammalian cells, and the like. Mammalian cells of interest include, e.g., murine cells, non-human primate cells, human cells, and the like.

"Recombinant host cells," "host cells," "cells," "cell lines," "cell cultures," and other such terms denoting microorganisms or higher eukaryotic cell lines, refer to cells which can be, or have been, used as recipients for a recombinant vector or other transferred DNA, and include the progeny of the cell which has been transfected. Host cells may be cultured as unicellular or multicellular entities (e.g., tissue, organs, or organoids) including an expression vector of the present disclosure.

In certain aspects, a cell of the present disclosure is an immune cell. Non-limiting examples of immune cells which may include any of the polypeptides, nucleic acids, and/or expression vectors of the present disclosure include T cells, B cells, natural killer (NK) cells, a macrophages, monocytes, neutrophils, dendritic cells, mast cells, basophils, and eosinophils. In some embodiments, a cell of the present disclosure is a T cell. Non-limiting examples of T cells include naive T cells (TN), cytotoxic T cells (T_C), memory T cells (TMEM), T memory stem cells (TSCM), central memory T cells (TCM), effector memory T cells (TEM), tissue resident memory T cells (TRM), effector T cells (TEFF), regulatory T cells (Tregs), CD4 T cells, CD8 T cells, virus-specific T cells, and gamma delta T cells ($\gamma\delta$). In certain aspects, a cell of the present disclosure is a stem cell, e.g., an embryonic stem cell or an adult stem cell.

Also provided are methods of making the cells of the present disclosure. In some embodiments, such methods include transfecting or transducing cells with a nucleic acid or expression vector of the present disclosure. The term "transfection" is used to refer to the uptake of foreign DNA by a cell. A cell has been "transfected" when exogenous DNA has been introduced inside the cell membrane. A number of transfection techniques are generally known in the art. See, e.g., Sambrook et al. (2001) Molecular Cloning, a laboratory manual, 3rd edition, Cold Spring Harbor Laboratories, New York, Davis et al. (1995) Basic Methods in Molecular Biology, 2nd edition, McGraw- Hill, and Chu et al. (1981) Gene

13:197. Such techniques can be used to introduce one or more exogenous DNA moieties into suitable host cells. The term refers to both stable and transient uptake of the genetic material.

5 In some embodiments, a cell of the present disclosure is produced by transducing the cell with a viral vector encoding the polypeptide. In certain aspects, the polypeptide includes a CAR and the cell is a T cell, such that provided are methods of producing a CAR T cell. In some embodiments, such methods include activating a population of T cells (e.g., T cells obtained from an individual to which a CAR T cell therapy will be administered), stimulating the population of T cells to proliferate, and transducing the T cell with a viral
10 vector encoding the polypeptide including the CAR. In some embodiments, the T cells are transduced with a retroviral vector, e.g., a gamma retroviral vector or lentiviral vector, encoding the polypeptide. In certain aspects, the T cells are transduced with a lentiviral vector encoding the polypeptide.

Cells of the present disclosure may be autologous/autogeneic ("self") or non-
15 autologous ("non-self," e.g., allogeneic, syngeneic or xenogeneic). "Autologous" as used herein, refers to cells from the same individual. "Allogeneic" as used herein refers to cells of the same species that differ genetically from the cell in comparison. "Syngeneic," as used herein, refers to cells of a different individual that are genetically identical to the cell in comparison. In some embodiments, the cells are T cells obtained from a mammalian
20 individual. In certain aspects, the mammalian individual is a primate. In some embodiments, the T cells are obtained from a human.

T cells may be obtained from a number of sources including, but not limited to, peripheral blood, peripheral blood mononuclear cells, bone marrow, lymph node tissue, cord blood, thymus tissue, tissue from a site of infection, ascites, pleural effusion, spleen
25 tissue, and tumors. In certain embodiments, T cells can be obtained from a unit of blood collected from an individual using any number of known techniques such as sedimentation, e.g., FICOLL™ separation.

In some embodiments, an isolated or purified population of T cells is used. In some
30 embodiments, after isolation of PBMC, both cytotoxic and helper T lymphocytes can be sorted into naive, memory, and effector T cell subpopulations either before or after activation, expansion, and/or genetic modification. Suitable approaches for such sorting are known and include, e.g., magnetic-activated cell sorting (MACS), where TN are CD45RA⁺ CD62L⁺ CD95⁻; TSCM are CD45RA⁺ CD62L⁺ CD95⁺; TCM are CD45RO⁺ CD62L⁺ CD95⁺; and TEM are CD45RO⁺ CD62L⁻ CD95⁺. An example approach for such
35 sorting is described in Wang et al. (2016) *Blood* 127(24):2980-90.

A specific subpopulation of T cells expressing one or more of the following markers: CD3, CD4, CD8, CD28, CD45RA, CD45RO, CD62, CD127, and HLA-DR can be further isolated by positive or negative selection techniques. In one embodiment, a specific subpopulation of T cells, expressing one or more of the markers selected from the group consisting of CD62L, CCR7, CD28, CD27, CD122, CD127, CD197; or CD38 or CD62L, CD127, CD197, and CD38, is further isolated by positive or negative selection techniques. In various embodiments, the manufactured T cell compositions do not express or do not substantially express one or more of the following markers: CD57, CD244, CD 160, PD-1, CTLA4, TIM3, and LAG3.

In order to achieve sufficient therapeutic doses of T cell compositions, the T cells may be subjected to one or more rounds of stimulation, activation and/or expansion. T cells can be activated and expanded generally using methods as described, for example, in U.S. Patents 6,352,694; 6,534,055; 6,905,680; 6,692,964; 5,858,358; 6,887,466; 6,905,681; 7,144,575; 7,067,318; 7,172,869; 7,232,566; 7,175,843; 5,883,223; 6,905,874; 6,797,514; and 6,867,041, each of which is incorporated herein by reference in its entirety for all purposes. In particular embodiments, T cells are activated and expanded for about 1 to 21 days, e.g., about 5 to 21 days. In some embodiments, T cells are activated and expanded for about 1 day to about 4 days, about 1 day to about 3 days, about 1 day to about 2 days, about 2 days to about 3 days, about 2 days to about 4 days, about 3 days to about 4 days, or about 1 day, about 2 days, about 3 days, or about 4 days prior to introduction of a nucleic acid (e.g., expression vector) encoding the polypeptide into the T cells.

In particular embodiments, T cells are activated and expanded for about 6 hours, about 12 hours, about 18 hours or about 24 hours prior to introduction of the genome editing compositions into the T cells. In certain aspects, T cells are activated at the same time that a nucleic acid (e.g., expression vector) encoding the polypeptide is introduced into the T cells.

In some embodiments, conditions appropriate for T cell culture include an appropriate media (e.g., Minimal Essential Media or RPMI Media 1640 or, X-vivo 15, (Lonza)) and one or more factors necessary for proliferation and viability including, but not limited to serum (e.g., fetal bovine or human serum), interleukin-2 (IL-2), insulin, IFN- γ , IL-4, IL-7, IL-21, GM-CSF, IL-10, IL-12, IL-15, TGF β , and TNF- α or any other additives suitable for the growth of cells known to the skilled artisan. Further illustrative examples of cell culture media include, but are not limited to RPMI 1640, Clicks, AEVI-V, DMEM, MEM, a-MEM, F-12, X-Vivo 15, and X-Vivo 20, Optimizer, with added amino acids, sodium pyruvate, and vitamins, either serum-free or supplemented with an appropriate amount of serum (or

plasma) or a defined set of hormones, and/or an amount of cytokine(s) sufficient for the growth and expansion of T cells.

In some embodiments, the nucleic acid (e.g., expression vector) encoding the polypeptide is introduced into the cell (e.g., a T cell) by microinjection, transfection, lipofection, heat-shock, electroporation, transduction, gene gun, microinjection, DEAE-dextran-mediated transfer, and the like. In certain aspects, the nucleic acid (e.g., expression vector) encoding the polypeptide is introduced into the cell (e.g., a T cell) by AAV transduction. The AAV vector may comprise ITRs from AAV2, and a serotype from any one of AAV1, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, or AAV 10. In preferred embodiments, the AAV vector may comprise ITRs from AAV2 and a serotype from AAV6. In some embodiments, the nucleic acid (e.g., expression vector) encoding the polypeptide is introduced into the cell (e.g., a T cell) by lentiviral transduction. The lentiviral vector backbone may be derived from HIV-1, HIV-2, visna-maedi virus (VMV) virus, caprine arthritis-encephalitis virus (CAEV), equine infectious anemia virus (EIAV), feline immunodeficiency virus (FIV), bovine immune deficiency virus (BIV), or simian immunodeficiency virus (SIV). The lentiviral vector may be integration competent or an integrase deficient lentiviral vector (IDLV). In one embodiment, IDLV vectors including an HIV-based vector backbone (i.e., HIV cis-acting sequence elements) are employed.

Also provided are viruses that include any of the polypeptides, nucleic acids, and/or expression vectors of the present disclosure.

COMPOSITIONS

Also provided by the present disclosure are compositions. A composition of the present disclosure may include any of the polypeptides, nucleic acids, expression vectors, and/or cells of the present disclosure.

In certain aspects, the compositions include any of the polypeptides, nucleic acids, expression vectors, and/or cells of the present disclosure present in a liquid medium. The liquid medium may be an aqueous liquid medium, such as water, a buffered solution, or the like. One or more additives such as a salt (e.g., NaCl, MgCl₂, KCl, MgSO₄), a buffering agent (a Tris buffer, N-(2-Hydroxyethyl)piperazine-N'-(2-ethanesulfonic acid) (HEPES), 2-(N-Morpholino)ethanesulfonic acid (MES), 2-(N-Morpholino)ethanesulfonic acid sodium salt (MES), 3-(N-Morpholino)propanesulfonic acid (MOPS), N-tris[Hydroxymethyl]methyl-3-aminopropanesulfonic acid (TAPS), etc.), a solubilizing agent, a detergent (e.g., a non-ionic detergent such as Tween-20, etc.), a nuclease inhibitor, glycerol, a chelating agent, and the like may be present in such compositions.

Pharmaceutical compositions are also provided. The pharmaceutical compositions may include any of the cells of the present disclosure, and a pharmaceutically acceptable

carrier. The pharmaceutical compositions generally include a therapeutically effective amount of the cells. By “therapeutically effective amount” is meant a number of cells sufficient to produce a desired result, e.g., an amount sufficient to effect beneficial or desired therapeutic (including preventative) results, such as a reduction in a symptom of a disease or disorder associated, e.g., with the target cell or a population thereof, as compared to a control. An effective amount can be administered in one or more administrations.

The cells of the present disclosure can be incorporated into a variety of formulations for therapeutic administration. More particularly, the cells of the present disclosure can be formulated into pharmaceutical compositions by combination with appropriate, pharmaceutically acceptable excipients or diluents.

Formulations of the cells suitable for administration to a patient (e.g., suitable for human administration) are generally sterile and may further be free of detectable pyrogens or other contaminants contraindicated for administration to a patient according to a selected route of administration.

The cells may be formulated for parenteral (e.g., intravenous, intra-arterial, intraosseous, intramuscular, intracerebral, intracerebroventricular, intrathecal, subcutaneous, etc.) administration, or any other suitable route of administration.

Pharmaceutical compositions that include the cells of the present disclosure may be prepared by mixing the cells having the desired degree of purity with optional physiologically acceptable carriers, excipients, stabilizers, surfactants, buffers and/or tonicity agents. Acceptable carriers, excipients and/or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid, glutathione, cysteine, methionine and citric acid; preservatives (such as ethanol, benzyl alcohol, phenol, m-cresol, p-chlor-m-cresol, methyl or propyl parabens, benzalkonium chloride, or combinations thereof); amino acids such as arginine, glycine, ornithine, lysine, histidine, glutamic acid, aspartic acid, isoleucine, leucine, alanine, phenylalanine, tyrosine, tryptophan, methionine, serine, proline and combinations thereof; monosaccharides, disaccharides and other carbohydrates; low molecular weight (less than about 10 residues) polypeptides; proteins, such as gelatin or serum albumin; chelating agents such as EDTA; sugars such as trehalose, sucrose, lactose, glucose, mannose, maltose, galactose, fructose, sorbose, raffinose, glucosamine, N-methylglucosamine, galactosamine, and neuraminic acid; and/or non-ionic surfactants such as Tween, Brij Pluronics, Triton-X, or polyethylene glycol (PEG).

An aqueous formulation of the polypeptides, nucleic acids, expression vectors, and/or cells may be prepared in a pH-buffered solution, e.g., at pH ranging from about 4.0

to about 7.0, or from about 5.0 to about 6.0, or alternatively about 5.5. Examples of buffers that are suitable for a pH within this range include phosphate-, histidine-, citrate-, succinate-, acetate-buffers and other organic acid buffers. The buffer concentration can be from about 1 mM to about 100 mM, or from about 5 mM to about 50 mM, depending, e.g., on the buffer and the desired tonicity of the formulation.

A tonicity agent may be included in the formulation to modulate the tonicity of the formulation. Example tonicity agents include sodium chloride, potassium chloride, glycerin and any component from the group of amino acids, sugars as well as combinations thereof. In some embodiments, the aqueous formulation is isotonic, although hypertonic or hypotonic solutions may be suitable. The term "isotonic" denotes a solution having the same tonicity as some other solution with which it is compared, such as physiological salt solution or serum. Tonicity agents may be used in an amount of about 5 mM to about 350 mM, e.g., in an amount of 100 mM to 350 mM.

A surfactant may also be added to the formulation to reduce aggregation and/or minimize the formation of particulates in the formulation and/or reduce adsorption. Example surfactants include polyoxyethylensorbitan fatty acid esters (Tween), polyoxyethylene alkyl ethers (Brij), alkylphenylpolyoxyethylene ethers (Triton-X), polyoxyethylene-polyoxypropylene copolymer (Poloxamer, Pluronic), and sodium dodecyl sulfate (SDS). Examples of suitable polyoxyethylenesorbitan-fatty acid esters are polysorbate 20, (sold under the trademark Tween 20™) and polysorbate 80 (sold under the trademark Tween 80™). Examples of suitable polyethylene-polypropylene copolymers are those sold under the names Pluronic® F68 or Poloxamer 188™. Examples of suitable Polyoxyethylene alkyl ethers are those sold under the trademark Brij™. Example concentrations of surfactant may range from about 0.001% to about 1% w/v.

In some embodiments, the pharmaceutical composition includes cells of the present disclosure, and one or more of the above-identified agents (e.g., a surfactant, a buffer, a stabilizer, a tonicity agent) and is essentially free of one or more preservatives, such as ethanol, benzyl alcohol, phenol, m-cresol, p-chlor-m-cresol, methyl or propyl parabens, benzalkonium chloride, and combinations thereof. In other embodiments, a preservative is included in the formulation, e.g., at concentrations ranging from about 0.001 to about 2% (w/v).

In certain aspects, provided is a pharmaceutical composition that includes a therapeutically effective amount of cells (e.g., T cells, such as CAR T cells) of the present disclosure. A "therapeutically effective amount" of such cells may vary according to factors such as the disease state, age, sex, and weight of the individual, and the ability of the cells to elicit a desired response in the individual. A therapeutically effective amount is also one

in which any toxic or detrimental effects of the cells are outweighed by the therapeutically beneficial effects. The term “therapeutically effective amount” includes an amount that is effective to “treat” an individual (e.g., a patient). When a therapeutic amount is indicated, the precise amount of the compositions contemplated in particular embodiments, to be administered, can be determined by a physician in view of the specification and with consideration of individual differences in age, weight, tumor size, extent of infection or metastasis, and condition of the patient (individual). In certain aspects, a pharmaceutical composition of the present disclosure includes from 1×10^6 to 5×10^{10} of the cells of the present disclosure.

10 METHODS OF USE

Also provided are methods that employ the polypeptides, nucleic acids, expression vectors, and/or cells of the present disclosure.

In certain aspects, a method for controlling the expression of a CAR on the surface of a cell is provided. Such methods include contacting a cell of the present disclosure with an inhibitor of the protease when expression of the CAR on the surface of the cell is not desired. In some embodiments, prior to the contacting, the method includes allowing the cell to express the CAR on the surface of the cell. In certain aspects the methods further include ceasing the contacting to express the CAR on the surface of the cell. In some embodiments, the protease is derived from HCV NS3, and the inhibitor of the protease is selected from the group consisting of asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), and grazoprevir (GPV). The methods may be carried out *in vitro* or *ex vivo* (e.g., in cultured cells), or *in vivo*, e.g., in an individual in a therapeutic context (e.g., an individual receiving a regulatable CAR cell-based therapy of the present disclosure).

In some embodiments, provided are methods of administering a regulatable CAR cell-based therapy to an individual in need thereof. In certain aspects, the individual in need thereof has cancer, and the antigen-binding portion of the CAR binds to a molecule on the surface of cells of the cancer. The methods of administering a regulatable CAR cell-based therapy to the individual include administering to the individual a pharmaceutical composition that includes a cell that includes any of the nucleic acids or expression vectors of the present disclosure. The pharmaceutical composition typically includes a therapeutically effective amount of such cells as described above. The cells may be any cells capable of effecting the desired therapy. In some embodiments, the cells are immune cells. Non-limiting examples of immune cells which may be administered include T cells, B cells, natural killer (NK) cells, macrophages, monocytes, neutrophils, dendritic cells, mast cells, basophils, and eosinophils. In some embodiments, the cells are T cells. In some embodiments, the cells are T cells such that the cells are CAR T cells. In certain aspects,

the cells are stem cells, e.g., embryonic stem cells or adult stem cells. In some embodiments, the pharmaceutical composition is produced by a method including removing cells from the individual and introducing into the removed cells or progeny thereof the desired nucleic acid or expression vector.

5 The methods of administering a regulatable cell-based therapy to an individual may further include contacting the administered cells or progeny thereof with an inhibitor of the protease when expression of the CAR on the surface of the administered cells or progeny thereof is not desired, where the contacting includes administering the inhibitor of the protease to the individual. Contacting the administered cells or progeny thereof with the
10 inhibitor may include administering to the individual an amount of the inhibitor effective to inhibit the protease of the polypeptides. As just one example, when the protease is derived from HCV NS3 as described elsewhere herein, the contacting may include administering to the individual by a suitable route of administration simeprevir, danoprevir, asunaprevir, ciluprevir, boceprevir, sovalprevir, paritaprevir, telaprevir, or any combination thereof, in an
15 amount effective to inhibit the protease of the polypeptides in the administered cells or progeny thereof. According to the methods of administering a regulatable cell-based therapy to an individual, the inhibitor of the protease may be administered to the individual prior to, concurrently with (that is, co-administered), and/or subsequent to administration of the pharmaceutical composition to the individual. In the methods of administering a
20 regulatable cell-based therapy to an individual, expression of the CAR on the surface of the administered cells or progeny thereof may not be desired for one or more reasons. For example, such expression may not be desired in order to prevent or delay the onset of cell exhaustion (e.g., T cell exhaustion) resulting from CAR activity. As such, the inhibitor of the protease may be administered to the individual to prevent or delay the onset of cell
25 exhaustion resulting from CAR activity. As another example, such expression may not be desired in order to reduce adverse side effects caused by the cells or progeny thereof, e.g., side effects relating to activity of the CAR expressed on the surface of the cells or progeny thereof.

 According to the methods of administering a regulatable CAR cell-based therapy
30 which include contacting the administered cells or progeny thereof with an inhibitor of the protease when expression of the CAR on the surface of the administered cells or progeny thereof is not desired, the methods may include administering the pharmaceutical composition to the individual prior to the contacting to allow expression of the CAR on the surface of the cells or progeny thereof in the individual, and subsequently administering the
35 inhibitor of the protease to the individual when expression of the CAR on the surface of the cells or progeny thereof is no longer desired. Expression of the CAR on the surface of the

cells or progeny thereof may no longer be desired for one or more reasons. For example, expression of the CAR on the surface of the cells or progeny thereof may no longer be desired in order to delay or prevent cell exhaustion resulting from CAR activity. Accordingly, the methods may include administering the protease to the individual to delay or prevent cell exhaustion resulting from CAR activity. Alternatively, or additionally, expression of the CAR on the surface of the cells or progeny thereof may no longer be desired because of adverse side effects caused by the cells or progeny thereof, such that the methods may include administering the protease to the individual to reduce adverse side effects caused by the cells or progeny thereof. Adverse side effects may include, but are not limited to, toxicity resulting from, e.g., unrestricted antigen-driven proliferation of the cells. Such toxicity may include cytokine release syndrome and/or neurotoxicity. Accordingly, the methods may include administering the protease to the individual to reduce adverse side effects caused by the cells or progeny thereof.

The methods of administering a regulatable cell-based therapy to an individual may further include administering to the individual a pharmacokinetic enhancer of the inhibitor of the protease. In certain embodiments, the pharmacokinetic enhancer increases the level of the inhibitor of the protease in the individual compared to the level of the inhibitor of the protease in the individual in the absence of administering the pharmacokinetic enhancer. The pharmacokinetic enhancer may vary depending upon the particular protease inhibitor employed. According to some embodiments, the pharmacokinetic enhancer is an inhibitor of an enzyme that metabolizes the inhibitor of the protease. For example, in certain embodiments, the inhibitor of the protease (e.g., grazoprevir and the like) is metabolized by a cytochrome P450 (CYP) enzyme, and the pharmacokinetic enhancer is an inhibitor of the CYP enzyme. A non-limiting example of such a pharmacokinetic enhancer is ritonavir.

The methods of administering a regulatable cell-based therapy to an individual may further include, subsequent to contacting the administered cells or progeny thereof with an inhibitor of the protease (e.g., to delay or prevent cell exhaustion, address adverse side effects, and/or the like), ceasing the contacting to express the CAR on the surface of the cells or progeny thereof.

When the protease of the polypeptide is derived from HCV NS3, the methods of administering a regulatable cell-based therapy to an individual may involve administering a protease inhibitor selected from the group consisting of asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), and grazoprevir (GPV).

The methods of administering a regulatable cell-based therapy to an individual may further include producing the pharmaceutical composition. Producing the pharmaceutical composition may include introducing an expression vector of the present disclosure into

cells or progeny thereof obtained from the individual. The introducing may include contacting the cells or progeny thereof obtained from the individual with an inhibitor of the protease prior to administering the pharmaceutical composition to the individual.

Methods that employ the cells of the present disclosure may be used in a variety of applications. For example, SMASh-mediated regulation of CARs could be used to enhance T cell function of engineered populations (cytotoxicity, proliferative capacity, cytokine secretion) during *ex vivo* expansion for adoptive transfer since diminished CAR expression would inhibit both tonic signaling (which can lead to exhaustion) and inhibit antigen induced signaling which can also lead to exhaustion *in vivo*. *In vivo* modulation of CAR signaling pathways in endogenous and/or transferred T cells in the context of chronic infection or cancer could lead to enhanced function.

SMASh-mediated regulation of CARs could potentially be used for localized regulation of receptor expression. For instance, if the small molecule protease inhibitor utilized to reduce receptor expression does not cross the blood:brain barrier, then one could achieve selective expression of a receptor within the CNS, which could be desirable in some circumstances. A further application is to enhance safety of genetically engineered populations since administration of an inhibitor of the protease can rapidly and potently diminish expression of a SMASh-regulated CAR on the surface of cells. A drug off system such as this is useful for enhancing the safety of engineered T cells including CAR T cells, where unrestricted antigen driven proliferation can lead to significant toxicity including cytokine release syndrome and neurotoxicity. Use of the SMASh-CARs allows regulated cell surface expression of the CAR for enhanced safety.

KITS

Also provided by the present disclosure are kits. In some embodiments, provided are kits that include any of the nucleic acids and/or expression vectors of the present disclosure, and instructions for introducing the nucleic acid or expression vector into a cell. The kits find use in a variety of *in vitro*, *ex vivo*, and *in vivo* applications. The instructions of such kits may further include instructions for regulatable expression of the CAR on the surface of the cell or progeny thereof. The instructions of such kits may further include instructions for contacting the cell or progeny thereof with an inhibitor of the protease when expression of the CAR on the surface of the cell or progeny thereof is not, or no longer, desired.

The kits of the present disclosure may further include any other reagents useful for regulatable expression of the CAR on the surface of the cell or progeny thereof, such as transfection/transduction reagents useful for introducing the nucleic acid or expression vector into a cell of interest, e.g., a T cell or other cell of interest.

In some embodiments, the kits further include an inhibitor of the protease of the polypeptide. For example, when a protease derived from HCV NS3 as described elsewhere herein is employed, the kit may include a suitable inhibitor of the protease, including but not limited to, imeprevir, danoprevir, asunaprevir, grazoprevir, simeprevir, 5 ciluprevir, boceprevir, sovalprevir, paritaprevir, telaprevir, or any combination thereof. Kits that include an inhibitor of the protease may further include instructions for contacting the cell or progeny thereof with the inhibitor when cell surface expression of the CAR is no longer desired.

Components of the kits may be present in separate containers, or multiple 10 components may be present in a single container. A suitable container includes a single tube (e.g., vial), one or more wells of a plate (e.g., a 96-well plate, a 384-well plate, etc.), or the like.

The instructions of the kits may be recorded on a suitable recording medium. For example, the instructions may be printed on a substrate, such as paper or plastic, etc. As 15 such, the instructions may be present in the kits as a package insert, in the labeling of the container of the kit or components thereof (i.e., associated with the packaging or sub-packaging), etc. In other embodiments, the instructions are present as an electronic storage data file present on a suitable computer readable storage medium, e.g., portable flash drive, DVD, CD-ROM, diskette, etc. In yet other embodiments, the actual 20 instructions are not present in the kit, but means for obtaining the instructions from a remote source, e.g. via the internet, are provided. An example of this embodiment is a kit that includes a web address where the instructions can be viewed and/or from which the instructions can be downloaded. As with the instructions, the means for obtaining the instructions is recorded on a suitable substrate.

25 Notwithstanding the appended claims, the present disclosure is also defined by the following clauses:

1. A polypeptide comprising from N-terminus to C-terminus:
a chimeric antigen receptor (CAR);
a protease; and
30 a degon,
wherein the polypeptide further comprises a cleavage site for the protease
disposed between the CAR and the degon.
2. The polypeptide of clause 1, wherein the cleavage site is between the CAR and
35 the protease.

3. The polypeptide of clause 1, wherein the cleavage site is between the protease and the degron.

5 4. The polypeptide of clause 2, further comprising a spacer domain between the CAR and the cleavage site.

5. The polypeptide of clause 3, further comprising a spacer domain between the CAR and the cleavage site.

10

6. The polypeptide of clause 4 or clause 5, wherein the spacer domain comprises a linker.

7. The polypeptide of any one of clauses 4 to 6, wherein the spacer domain
15 comprises a reporter.

8. The polypeptide of clause 7, wherein the reporter is a bioluminescence reporter.

9. The polypeptide of clause 8, wherein the bioluminescence reporter is a luciferase.

20

10. The polypeptide of any one of clauses 4 to 9, wherein the spacer domain decreases the response time of the polypeptide to an inhibitor of the protease.

25 11. The polypeptide of any one of clauses 1 to 10, wherein the antigen-binding portion of the CAR is an antibody.

12. The polypeptide of clause 11, wherein the antibody is a single chain variable fragment (scFv).

30

13. The polypeptide of any one of clauses 1 to 12, wherein the antigen-binding portion of the CAR specifically binds an antigen on the surface of a cancer cell.

14. The polypeptide of clause 13, wherein the antigen on the surface of the cancer cell
35 is a tumor-associated antigen or a tumor-specific antigen.

15. The polypeptide of clause 13, wherein the antigen on the surface of the cancer cell is selected from the group consisting of: B7-H3 (CD276), CD19, GD2, CD22, and HER2.

5 16. The polypeptide of any one of clauses 1 to 15, wherein the protease is a viral protease.

17. The polypeptide of clause 16, wherein the protease is derived from hepatitis C virus (HCV) nonstructural protein 3 (NS3).

10 18. The polypeptide of clause 17, wherein the protease further comprises a cofactor polypeptide derived from HCV nonstructural protein 4A (NS4A).

15 19. The polypeptide of clause 17 or clause 18, wherein the protease is engineered to include one or more amino acid substitutions relative to a wild-type HCV NS3 protease.

20 20. The polypeptide of clause 19, wherein the protease comprises an amino acid sequence that is at least 80% identical, at least 85% identical, at least 90% identical, at least 95% identical, at least 99% identical, or 100% identical, to the amino acid sequence set forth in SEQ ID NO:2, which protease comprises an alanine at position 54, and wherein numbering is as in SEQ ID NO:2.

25 21. The polypeptide of any one of clauses 1 to 20, wherein the cleavage site for the protease is selected from the group consisting of: an NS4A/4B junction cleavage site, an NS3/NS4A junction cleavage site, an NS4A/NS4B junction cleavage site, an NS4B/NS5A junction cleavage site, and an NS5A/NS5B junction cleavage site.

22. The polypeptide of any one of clauses 1 to 21, wherein the degron comprises portions of the HCV nonstructural proteins NS3 and NS4A.

30 23. The polypeptide of clause 22, wherein the degron comprises an amino acid sequence that is at least 80% identical, at least 85% identical, at least 90% identical, at least 95% identical, or 100% identical, to the amino acid sequence set forth in SEQ ID NO:1.

35 24. A cell comprising the polypeptide of any one of clauses 1 to 23.

25. The cell of clause 24, wherein the cell is a mammalian cell.

26. The cell of clause 25, wherein the cell is a human cell.

5 27. The cell of any one of clauses 24 to 26, wherein the cell is an immune cell.

28. The cell of clause 27, wherein the cell is a T cell.

10 29. The cell of any one of clauses 24 to 28, wherein the CAR is expressed on the surface of the cell.

30. A pharmaceutical composition, comprising:
the cell of any one of clauses 24 to 29; and
a pharmaceutically-acceptable carrier.

15

31. A nucleic acid encoding the polypeptide of any one of clauses 1 to 23.

32. An expression vector comprising the nucleic acid of clause 31.

20 33. A cell comprising the nucleic acid of clause 31 or the expression vector of clause 32.

34. The cell of clause 33, wherein the cell is a mammalian cell.

25 35. The cell of clause 34, wherein the cell is a human cell.

36. The cell of any one of clauses 33 to 35, wherein the cell is an immune cell.

37. The cell of clause 36, wherein the cell is a T cell.

30

38. A method of making the cell of any one of clauses 33 to 37, comprising introducing the nucleic acid of clause 31 or the expression vector of clause 32 into the cell.

35 39. A method for controlling the expression of a CAR on the surface of a cell, comprising:

contacting the cell of any one of clauses 33 to 37 with an inhibitor of the protease when expression of the CAR on the surface of the cell is not desired.

40. The method according to clause 39, wherein the protease is derived from HCV NS3, and wherein the inhibitor of the protease is selected from the group consisting of: asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), grazoprevir (GPV), and any combination thereof.

41. The method according to clause 39 or clause 40, wherein prior to the contacting, the cell is allowed to express the CAR on the surface of the cell.

42. The method according to any one of clauses 39 to 41, further comprising ceasing the contacting to express the CAR on the surface of the cell.

43. The method according to any one of clauses 39 to 42, wherein the method is performed *in vitro*.

44. The method according to any one of clauses 39 to 42, wherein the method is performed *ex vivo*.

45. The method according to any one of clauses 39 to 42, wherein the method is performed *in vivo*.

46. A pharmaceutical composition, comprising:
the cell of any one of clauses 33 to 37; and
a pharmaceutically-acceptable carrier.

47. A method of making the pharmaceutical composition of clause 46, comprising introducing the expression vector of clause 32 into cells obtained from an individual.

48. A method of administering a regulatable CAR cell-based therapy to an individual in need thereof, comprising administering to the individual the pharmaceutical composition of clause 46.

49. The method according to clause 48, further comprising contacting the administered cells or progeny thereof with an inhibitor of the protease when expression of the CAR on

the surface of the administered cells or progeny thereof is not desired, wherein the contacting comprises administering the inhibitor of the protease to the individual.

50. The method according to clause 49, wherein the inhibitor of the protease is
5 administered to the individual prior to administration of the pharmaceutical composition.

51. The method according to clause 49 or clause 50, wherein the pharmaceutical composition and the inhibitor of the protease are co-administered to the individual.

10 52. The method according to any one of clauses 49 to 51, wherein the inhibitor of the protease is administered to the individual to prevent or delay the onset of cell exhaustion resulting from CAR activity.

53. The method according to clause 49, comprising:
15 administering the pharmaceutical composition to the individual prior to the contacting to allow expression of the CAR on the surface of the cells or progeny thereof in the individual; and administering the inhibitor of the protease to the individual when expression of the CAR on the surface of the cells or progeny thereof is no longer desired.

20

54. The method according to clause 53, wherein the inhibitor of the protease is administered to the individual to delay or prevent cell exhaustion resulting from CAR activity.

25 55. The method according to clause 53 or clause 54, wherein the inhibitor of the protease is administered to the individual to reduce adverse side effects caused by the cells or progeny thereof.

56. The method according to any one of clauses 49 to 55, further comprising ceasing
30 the contacting to express the CAR on the surface of the cells or progeny thereof.

57. The method according to any one of clauses 49 to 56, wherein the protease is derived from HCV NS3, and wherein the inhibitor of the protease is selected from the group consisting of: asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), and
35 grazoprevir (GPV).

58. The method according to any one of clauses 48 to 57, further comprising administering to the individual a pharmacokinetic enhancer of the inhibitor of the protease.

59. The method according to clause 58, wherein the pharmacokinetic enhancer is an inhibitor of an enzyme that metabolizes the inhibitor of the protease.

60. The method according to clause 59, wherein the pharmacokinetic enhancer is an inhibitor of a cytochrome P450 (CYP) enzyme that metabolizes the inhibitor of the protease.

61. The method according to clause 60, wherein the pharmacokinetic enhancer is ritonavir.

62. The method according to clause 61, wherein the inhibitor of the protease is grazoprevir.

63. The method according to any one of clauses 48 to 62, further comprising producing the pharmaceutical composition.

64. The method according to clause 63, wherein producing the pharmaceutical composition comprises introducing the expression vector of clause 32 into cells or progeny thereof obtained from the individual.

65. The method according to clause 64, comprising contacting the cells obtained from the individual, or progeny thereof, with an inhibitor of the protease prior to administering the pharmaceutical composition to the individual.

66. A kit, comprising:
the nucleic acid of clause 31 or the expression vector of clause 32; and
instructions for introducing the nucleic acid or expression vector into a cell.

67. The kit of clause 66, wherein the instructions further comprise instructions for regulatable expression of the CAR on the surface of the cell or progeny thereof.

68. The kit of clause 67, wherein the instructions comprise instructions for contacting the cell or progeny thereof with an inhibitor of the protease when expression of the CAR on the surface of the cell or progeny thereof is not desired.

- 5 69. The kit of any one of clauses 66 to 68, further comprising an inhibitor of the protease.

The following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

EXAMPLE 1 - SMASH CARs: ENGINEERED PROTEIN EXPRESSION IN HUMAN CELLS USING 10 DRUG REGULATABLE PROTEASES

Materials and Methods

Materials

Asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), and grazoprevir (GPV) were purchased from ApexBio. Cell surface markers were stained with mouse monoclonal
15 antibodies targeting PD-1 (PE-Cy7, eBioscience), LAG3 (PE, eBioscience), Tim-3 (BV 510, BioLegend), and B7-H3 (PE, R&D Systems). CAR molecules were stained with recombinant B7-H3 Fc (R&D Systems), protein L (Pierce Protein Biology), 1A7 anti-idiotypic antibody, or FMC63 anti-idiotypic antibody 136.20.1 conjugated in house using DyLight 650 Antibody Labeling Kit (Thermo Fisher). T cells were cultured in AIM V (Thermo Fisher)
20 supplemented with 5% fetal bovine serum (Gibco), 100 U/mL rhIL-2 (PeproTech), 2 mM L-glutamine, 10 mM HEPES, 100 U/mL penicillin, and 100 mg/mL streptomycin (Invitrogen). Cancer cell lines were cultured in RPMI (Thermo Fisher) supplemented with 10% fetal bovine serum, 2 mM L-glutamine, 10 mM HEPES, 100 U/mL penicillin, and 100 mg/mL streptomycin.

Generation of CAR T Cells

SMASH-tagged CAR constructs were synthesized by custom gene synthesis (GeneArt) or assembled with the In-Fusion Cloning Kit (Takara Bio) into a retroviral vector. Viral supernatant was prepared by transfecting GP2-293 cells (Clontech) with CAR-SMASH plasmids complexed with Lipofectamine 2000 (Thermo Fisher). Viral supernatant was
30 harvested 48 and 72 hours post-transfection, centrifuged and filtered to remove cell debris, and stored at -80°C until use. CAR T cells were generated by retrovirally transducing activated T cells on Retronectin (Takara Bio) coated plates.

Flow Cytometry

Flow cytometry of samples was performed on an LSR Fortessa (BD Bioscience) and data were analyzed using FlowJo software. B7H3 CARs were detected using B7H3 Fc. GD2 and CD19 CARs were detected by 1A7 and FMC63 anti-idiotypic antibody, respectively. CD 22 CARs were detected using biotinylated protein L and APC conjugated streptavidin. FACS plots were gated on live single T cells. Untransduced T cells ("Mock") were used as controls.

CAR-SMASH Drug Response Assay

CAR T cells were incubated in the indicated concentration of drug in T cell culture media (above) until analysis by FACS. Media was changed every 48-72 hours. Removal of drug was achieved by washing CAR T cells in T cell culture media 3 times.

Co-culture Killing Assays

CAR T cells were co-cultured with indicated cancer cell lines, engineered to constitutively express green fluorescent protein, on day 10 post activation in 96-well plates at a ratio of 1:1 (50,000 cells each). Cancer cell fluorescence was monitored by time-lapse microscopy. Total integrated fluorescence intensity was calculated and plotted as a function of time. Co-culture supernatant was harvested after 24 hours of co-culture and analyzed for the presence of cytokine. Cytokine in culture supernatant was quantified using the Human IL-2 ELISA MAX™ and Human IFN-γ ELISA MAX™ Deluxe kits (BioLegend).

Results

A chimeric antigen receptor (CAR) incorporating the three critical elements for regulatory control in the SMASH-Tag system (a protease susceptible cleavable linker, the protease itself, and a degron domain) as an addition to the C-terminal domain of the CAR, was generated (termed "SMASH-CAR").

First, it was demonstrated that in the absence of exogenous protease inhibitors, expression of a B7-H3 CAR incorporating a 4-1BB and CD3 signaling endodomain as well as the trio of SMASH components (termed "B7H3 SMASH-CAR" – **FIG. 2, panel B**) shows high level surface expression on engineered human T cells (**FIG. 3**). The CAR was expressed using a retroviral construct and demonstrated expression that was approximately 2 logs above background levels.

When asunaprevir, a small molecule that specifically inhibits HCV NS3 protease activity was added to the system *ex vivo*, CAR expression was reduced by greater than 90% (**FIG. 4**). Reduction in CAR expression occurred within 24 hours and was maximal at 96h.

Following drug induced CAR removal, asunaprevir was subsequently removed, resulting in approximately 1.5 log re-expression within 24h and full re-expression within 72h (**FIG. 5**). Thus, we demonstrate drug-induced SMASh-CAR regulation.

It was further demonstrated that drug-induced loss of expression leads to diminished expression of markers of T cell exhaustion (**FIG. 6** and **FIG. 9**). Maximal effect was observed with 1 μ M of asunaprevir, but near maximal effect was observed at 0.3 μ M.

The level of regulation is biologically significant since drug-induced inhibition of CAR expression beginning at 24h following drug application demonstrates significant reduction in killing of B7H3+ tumor targets (**FIG. 11**). Upon re-expression of the CAR following asunaprevir removal, potent killing of tumor targets indistinguishable from constitutively expressed, non-regulated CAR T cells was observed (**FIG. 12**). Similarly drug-induced inhibition of CAR expression leads to very low levels of interferon production in response to antigen expressing targets, which is restored with 24h upon removal of the CAR from the media. Similar levels of regulation using multiple CAR constructs was observed, including those targeting GD2, CD19 and CD22 (**FIGs. 17-20**).

Example 2 – Drug-Regulatable Control of CAR Expression and CAR T Cell Cytotoxicity *In Vivo*

This example demonstrates the regulatable expression of a CAR on T cells *in vivo*. NSG mice were infused with 10^6 Nalm6 leukemia cells expressing B7H3 and firefly luciferase. Four days later, mice were infused with 8×10^6 B7H3-BBz-SMASh CAR T cells or mock untransduced control cells. Mice that received B7H3-BBz-SMASh CAR T cells were either administered drug (50 mg/kg grazoprevir + 25mg/kg ritonavir) or vehicle (50% polyethylene glycol 300) three times per day by oral gavage. Bioluminescence imaging was used to follow the expansion of the firefly luciferase-expressing Nalm6 cells. As shown in FIG. 24, the presence of drug dramatically reduces the cytotoxic capacity of the CAR T cells *in vivo*.

In a further study, real-time expression of a C-terminal nanoluciferase (nLuc) domain-containing CAR was monitored *in vivo* by nanoluciferase-based imaging. As shown in FIG. 25, infusion of a HCV NS3 protease inhibitor diminished CAR expression on T cells *in vivo*.

Example 3 – Regulatable CAR T Cell-Based Therapy

The following is a protocol for regulatable cell-based therapy using drug-regulatable CAR T cells.

Peripheral blood mononuclear cells (PBMCs) are harvested by leukapheresis and enriched for T cells. The T cells are activated and expanded by magnetic beads decorated with anti-CD3/CD28 antibodies. The CAR-SMASH transgene is delivered to the T cells via an appropriate gene delivery method (lentivirus, retrovirus, AAV, transposase, CRISPR-CAS9, TALENs, nanoparticle, mRNA, electroporation, etc.). The CAR T cells are co-cultured with beads for a period of 2-3 days prior to separation by exposure to a magnetic field. The CAR T cells are expanded for 5 to 21 days in the presence of a cytokine such as interleukin-2 that supports proliferation. The CAR T cells can optionally be cultured in the presence of HCV NS3 protease inhibitor such as asunaprevir (1 pM to 50 μ M) during part of or the entire *ex vivo* culture period to turn off CAR expression. Turning off CAR expression during the *ex vivo* culture period reduces premature T cell activation due to basal CAR activity or engagement of the CAR with cognate antigen present in the cell preparation. This reduction in premature CAR activity minimizes T cell exhaustion and preserves CAR T cell capacity for mounting an immune response upon engagement of target antigen. The CAR T cells are cryopreserved in cryopreservation media for later use and/or analysis.

Patients are infused with 1×10^6 to 5×10^{10} CAR T cells. When the SMASH tag includes a protease derived from HCV NS3, CAR expression can be controlled *in vivo* by administering an HCV NS3 protease inhibitor given one to six times per day at a dose of 0.1 to 300 mg/kg. The drug may be administered as needed to control adverse effects. Alternatively, or additionally, the drug is administered in order to minimize T cell exhaustion and/or dysfunction, and/or enhance T cell therapeutic capacity. The drug may be administered in such a manner to confer periods of high CAR expression (activity) and periods of low CAR expression (rest). The dosing schedule may include periods of rest lasting from 1 to 15 days, periods of CAR expression ranging between 1 to 30 days, and any combination thereof.

Example 4 – *In vivo* control of B7H3 CAR-nanoluciferase-SMASH CAR-T cell activity against Nalm6 leukemia cells

Assessed in this example was whether B7H3 CAR-nanoluciferase-SMASH CAR-T cell activity could be controlled *in vivo* against Nalm6 leukemia cells. NSG mice were infused with 10^6 Nalm6 leukemia cells expressing B7H3 antigen and firefly luciferase by tail vein injection. Four days later, mice were infused with 8×10^6 B7H3 CAR-nanoluciferase-SMASH CAR-T cells or mock un-transduced control T cells by tail vein injection. Mice in the “CAR OFF” group were administered 50 mg/kg grazoprevir + 25 mg/kg ritonavir (a pharmacokinetic enhancer of grazoprevir) two to three times per day by oral gavage.

Bioluminescence imaging was used to follow the expansion of the firefly luciferase-expressing Nalm6 cells.

Provided in FIG. 26 (left panel) is a series of images showing the bioluminescence of the Nalm6 leukemia cells throughout the time course of the experiment. Provided in FIG. 26 (right panel) is a graph showing the average bioluminescence flux values of the various experimental groups throughout the time course of the experiment. Error bars represent standard error of mean. As seen in the data, B7H3 CAR-nanoluciferase-SMASH CAR-T cells display robust clearance of leukemia cells in the absence of drug but have diminished cytotoxic capacity in the presence of drug, as determined by bioluminescence imaging.

Example 5 – *In vivo* control of B7H3 CAR-nanoluciferase-SMASH CAR-T cell activity against a patient-derived medulloblastoma cell line

Assessed in this example was whether B7H3 CAR-nanoluciferase-SMASH CAR-T cell activity could be controlled *in vivo* against the patient-derived medulloblastoma cell line MED8A. NSG mice were subcutaneously implanted with 0.5×10^6 MED8A cells that were engineered to express firefly luciferase. Five days later, mice were infused with 5×10^6 B7H3 CAR-nanoluciferase-SMASH CAR-T cells or mock untransduced control T cells by tail vein injection. Mice in the “SMASH OFF” group were administered 50 mg/kg grazoprevir + 25mg/kg ritonavir (a pharmacokinetic enhancer of grazoprevir) two to three times per day by oral gavage. Bioluminescence imaging was used to follow the expansion of the firefly luciferase-expressing MED8A cells.

Provided in FIG. 27 (left panel) is a series of images showing the bioluminescence of the MED8A cells throughout the time course of the experiment. Provided in FIG. 27 (right panel) is a graph showing the average bioluminescence flux values of the various experimental groups throughout the time course of the experiment. Error bars represent standard error of mean. As seen in the data, B7H3 CAR-nanoluciferase-SMASH CAR-T cells display robust clearance of MED8A cells in the absence of drug but have diminished cytotoxic capacity in the presence of drug, as determined by bioluminescence imaging.

Next, *in vivo* CAR expression in the mice described in FIG. 27 was assessed by nanoluciferase imaging. Mice were injected with 200 μ L of Nano-Glo substrate that was diluted 40x in PBS. After a 6-minute incubation period, the mice were imaged on an IVIS Spectrum imager. As seen in the data, *in vivo* CAR expression was diminished in mice that received 50 mg/kg grazoprevir + 25 mg/kg ritonavir two to three times per day by oral gavage.

Accordingly, the preceding merely illustrates the principles of the present disclosure. It will be appreciated that those skilled in the art will be able to devise various arrangements which, although not explicitly described or shown herein, embody the principles of the invention and are included within its spirit and scope. Furthermore, all examples and conditional language recited herein are principally intended to aid the reader in understanding the principles of the invention and the concepts contributed by the inventors to furthering the art, and are to be construed as being without limitation to such specifically recited examples and conditions. Moreover, all statements herein reciting principles, aspects, and embodiments of the invention as well as specific examples thereof, are intended to encompass both structural and functional equivalents thereof. Additionally, it is intended that such equivalents include both currently known equivalents and equivalents developed in the future, i.e., any elements developed that perform the same function, regardless of structure. The scope of the present invention, therefore, is not intended to be limited to the example embodiments shown and described herein.

WHAT IS CLAIMED IS:

1. A polypeptide comprising from N-terminus to C-terminus:
a chimeric antigen receptor (CAR);
a protease; and
a degron,
wherein the polypeptide further comprises a cleavage site for the protease
disposed between the CAR and the degron.
2. The polypeptide of claim 1, wherein the cleavage site is between the CAR and the protease.
3. The polypeptide of claim 1, wherein the cleavage site is between the protease and the degron.
4. The polypeptide of claim 2, further comprising a spacer domain between the CAR and the cleavage site.
5. The polypeptide of claim 3, further comprising a spacer domain between the CAR and the cleavage site.
6. The polypeptide of claim 4 or claim 5, wherein the spacer domain comprises a linker.
7. The polypeptide of any one of claims 4 to 6, wherein the spacer domain comprises a reporter.
8. The polypeptide of claim 7, wherein the reporter is a bioluminescence reporter.
9. The polypeptide of claim 8, wherein the bioluminescence reporter is a luciferase.
10. The polypeptide of any one of claims 4 to 9, wherein the spacer domain decreases the response time of the polypeptide to an inhibitor of the protease.

11. The polypeptide of any one of claims 1 to 10, wherein the antigen-binding portion of the CAR is an antibody.
12. The polypeptide of claim 11, wherein the antibody is a single chain variable fragment (scFv).
13. The polypeptide of any one of claims 1 to 12, wherein the antigen-binding portion of the CAR specifically binds an antigen on the surface of a cancer cell.
14. The polypeptide of claim 13, wherein the antigen on the surface of the cancer cell is a tumor-associated antigen or a tumor-specific antigen.
15. The polypeptide of claim 13, wherein the antigen on the surface of the cancer cell is selected from the group consisting of: B7-H3 (CD276), CD19, GD2, CD22, and HER2.
16. The polypeptide of any one of claims 1 to 15, wherein the protease is a viral protease.
17. The polypeptide of claim 16, wherein the protease is derived from hepatitis C virus (HCV) nonstructural protein 3 (NS3).
18. The polypeptide of claim 17, wherein the protease further comprises a cofactor polypeptide derived from HCV nonstructural protein 4A (NS4A).
19. The polypeptide of claim 17 or claim 18, wherein the protease is engineered to include one or more amino acid substitutions relative to a wild-type HCV NS3 protease.
20. The polypeptide of claim 19, wherein the protease comprises an amino acid sequence that is at least 80% identical, at least 85% identical, at least 90% identical, at least 95% identical, at least 99% identical, or 100% identical, to the amino acid

sequence set forth in SEQ ID NO:2, which protease comprises an alanine at position 54, and wherein numbering is as in SEQ ID NO:2.

21. The polypeptide of any one of claims 1 to 20, wherein the cleavage site for the protease is selected from the group consisting of: an NS4A/4B junction cleavage site, an NS3/NS4A junction cleavage site, an NS4A/NS4B junction cleavage site, an NS4B/NS5A junction cleavage site, and an NS5A/NS5B junction cleavage site.

22. The polypeptide of any one of claims 1 to 21, wherein the degron comprises portions of the HCV nonstructural proteins NS3 and NS4A.

23. The polypeptide of claim 22, wherein the degron comprises an amino acid sequence that is at least 80% identical, at least 85% identical, at least 90% identical, at least 95% identical, or 100% identical, to the amino acid sequence set forth in SEQ ID NO:1.

24. A cell comprising the polypeptide of any one of claims 1 to 23.

25. The cell of claim 24, wherein the cell is a mammalian cell.

26. The cell of claim 25, wherein the cell is a human cell.

27. The cell of any one of claims 24 to 26, wherein the cell is an immune cell.

28. The cell of claim 27, wherein the cell is a T cell.

29. The cell of any one of claims 24 to 28, wherein the CAR is expressed on the surface of the cell.

30. A pharmaceutical composition, comprising:
the cell of any one of claims 24 to 29; and
a pharmaceutically-acceptable carrier.

31. A nucleic acid encoding the polypeptide of any one of claims 1 to 23.
32. An expression vector comprising the nucleic acid of claim 31.
33. A cell comprising the nucleic acid of claim 31 or the expression vector of claim 32.
34. The cell of claim 33, wherein the cell is a mammalian cell.
35. The cell of claim 34, wherein the cell is a human cell.
36. The cell of any one of claims 33 to 35, wherein the cell is an immune cell.
37. The cell of claim 36, wherein the cell is a T cell.
38. A method of making the cell of any one of claims 33 to 37, comprising introducing the nucleic acid of claim 31 or the expression vector of claim 32 into the cell.
39. A method for controlling the expression of a CAR on the surface of a cell, comprising:
 - contacting the cell of any one of claims 33 to 37 with an inhibitor of the protease when expression of the CAR on the surface of the cell is not desired.
40. The method according to claim 39, wherein the protease is derived from HCV NS3, and wherein the inhibitor of the protease is selected from the group consisting of: asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), grazoprevir (GPV), and any combination thereof.
41. The method according to claim 39 or claim 40, wherein prior to the contacting, the cell is allowed to express the CAR on the surface of the cell.
42. The method according to any one of claims 39 to 41, further comprising ceasing the contacting to express the CAR on the surface of the cell.

43. The method according to any one of claims 39 to 42, wherein the method is performed *in vitro*.
44. The method according to any one of claims 39 to 42, wherein the method is performed *ex vivo*.
45. The method according to any one of claims 39 to 42, wherein the method is performed *in vivo*.
46. A pharmaceutical composition, comprising:
the cell of any one of claims 33 to 37; and
a pharmaceutically-acceptable carrier.
47. A method of making the pharmaceutical composition of claim 46, comprising introducing the expression vector of claim 32 into cells obtained from an individual.
48. A method of administering a regulatable CAR cell-based therapy to an individual in need thereof, comprising administering to the individual the pharmaceutical composition of claim 46.
49. The method according to claim 48, further comprising contacting the administered cells or progeny thereof with an inhibitor of the protease when expression of the CAR on the surface of the administered cells or progeny thereof is not desired, wherein the contacting comprises administering the inhibitor of the protease to the individual.
50. The method according to claim 49, wherein the inhibitor of the protease is administered to the individual prior to administration of the pharmaceutical composition.
51. The method according to claim 49 or claim 50, wherein the pharmaceutical composition and the inhibitor of the protease are co-administered to the individual.

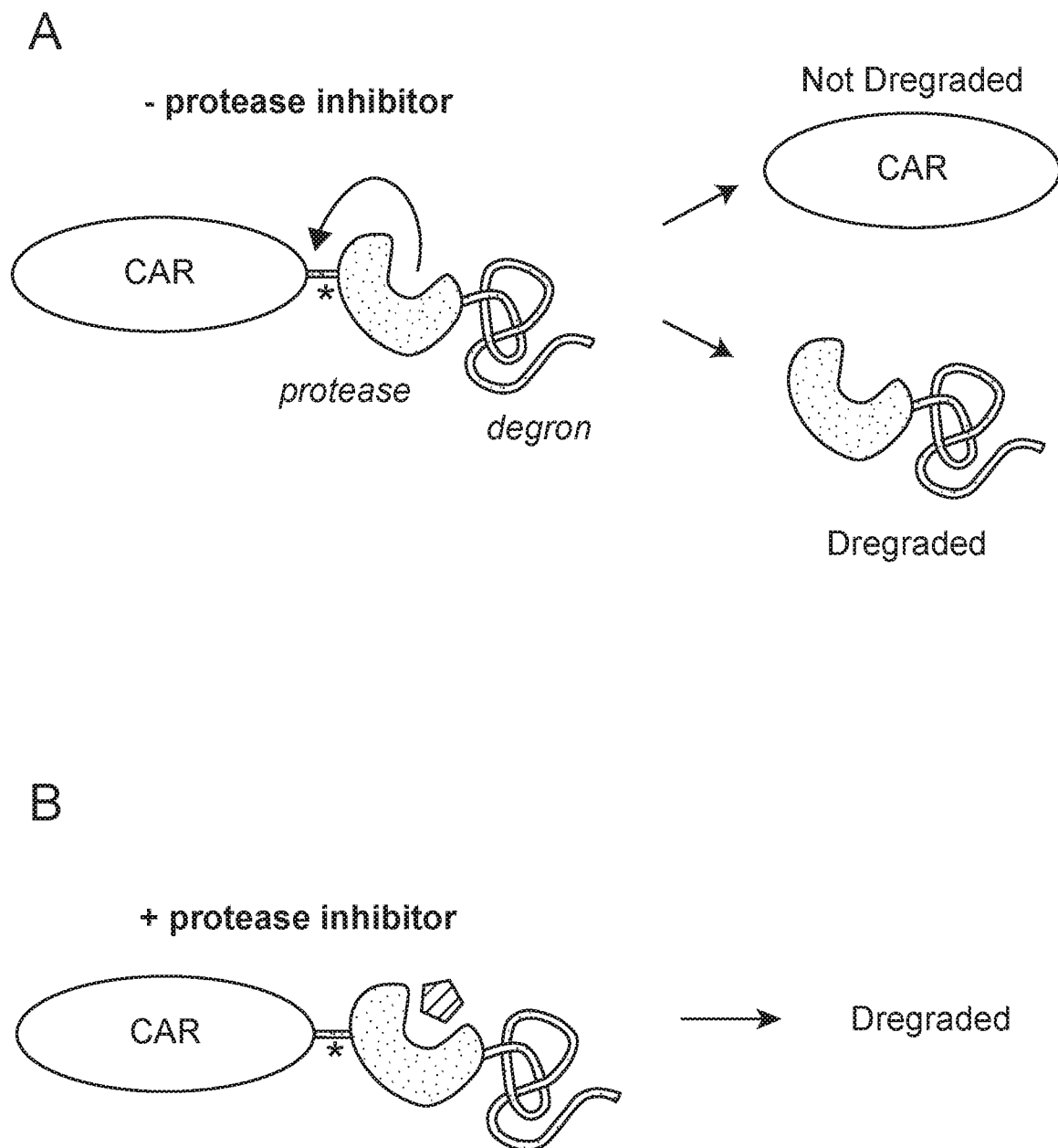
52. The method according to any one of claims 49 to 51, wherein the inhibitor of the protease is administered to the individual to prevent or delay the onset of cell exhaustion resulting from CAR activity.
53. The method according to claim 49, comprising:
administering the pharmaceutical composition to the individual prior to the
contacting to allow expression of the CAR on the surface of the cells or
progeny thereof in the individual; and
administering the inhibitor of the protease to the individual when expression of
the CAR on the surface of the cells or progeny thereof is no longer desired.
54. The method according to claim 53, wherein the inhibitor of the protease is administered to the individual to delay or prevent cell exhaustion resulting from CAR activity.
55. The method according to claim 53 or claim 54, wherein the inhibitor of the protease is administered to the individual to reduce adverse side effects caused by the cells or progeny thereof.
56. The method according to any one of claims 49 to 55, further comprising ceasing the contacting to express the CAR on the surface of the cells or progeny thereof.
57. The method according to any one of claims 49 to 56, wherein the protease is derived from HCV NS3, and wherein the inhibitor of the protease is selected from the group consisting of: asunaprevir (ASV), danoprevir (DPV), simeprevir (SPV), and grazoprevir (GPV).
58. The method according to any one of claims 48 to 57, further comprising administering to the individual a pharmacokinetic enhancer of the inhibitor of the protease.
59. The method according to claim 58, wherein the pharmacokinetic enhancer is an inhibitor of an enzyme that metabolizes the inhibitor of the protease.

60. The method according to claim 59, wherein the pharmacokinetic enhancer is an inhibitor of a cytochrome P450 (CYP) enzyme that metabolizes the inhibitor of the protease.
61. The method according to claim 60, wherein the pharmacokinetic enhancer is ritonavir.
62. The method according to claim 61, wherein the inhibitor of the protease is grazoprevir.
63. The method according to any one of claims 48 to 62, further comprising producing the pharmaceutical composition.
64. The method according to claim 63, wherein producing the pharmaceutical composition comprises introducing the expression vector of claim 32 into cells or progeny thereof obtained from the individual.
65. The method according to claim 64, comprising contacting the cells obtained from the individual, or progeny thereof, with an inhibitor of the protease prior to administering the pharmaceutical composition to the individual.
66. A kit, comprising:
the nucleic acid of claim 31 or the expression vector of claim 32; and
instructions for introducing the nucleic acid or expression vector into a cell.
67. The kit of claim 66, wherein the instructions further comprise instructions for regulatable expression of the CAR on the surface of the cell or progeny thereof.
68. The kit of claim 67, wherein the instructions comprise instructions for contacting the cell or progeny thereof with an inhibitor of the protease when expression of the CAR on the surface of the cell or progeny thereof is not desired.

69. The kit of any one of claims 66 to 68, further comprising an inhibitor of the protease.

1/33

FIG. 1



2/33

FIG. 2

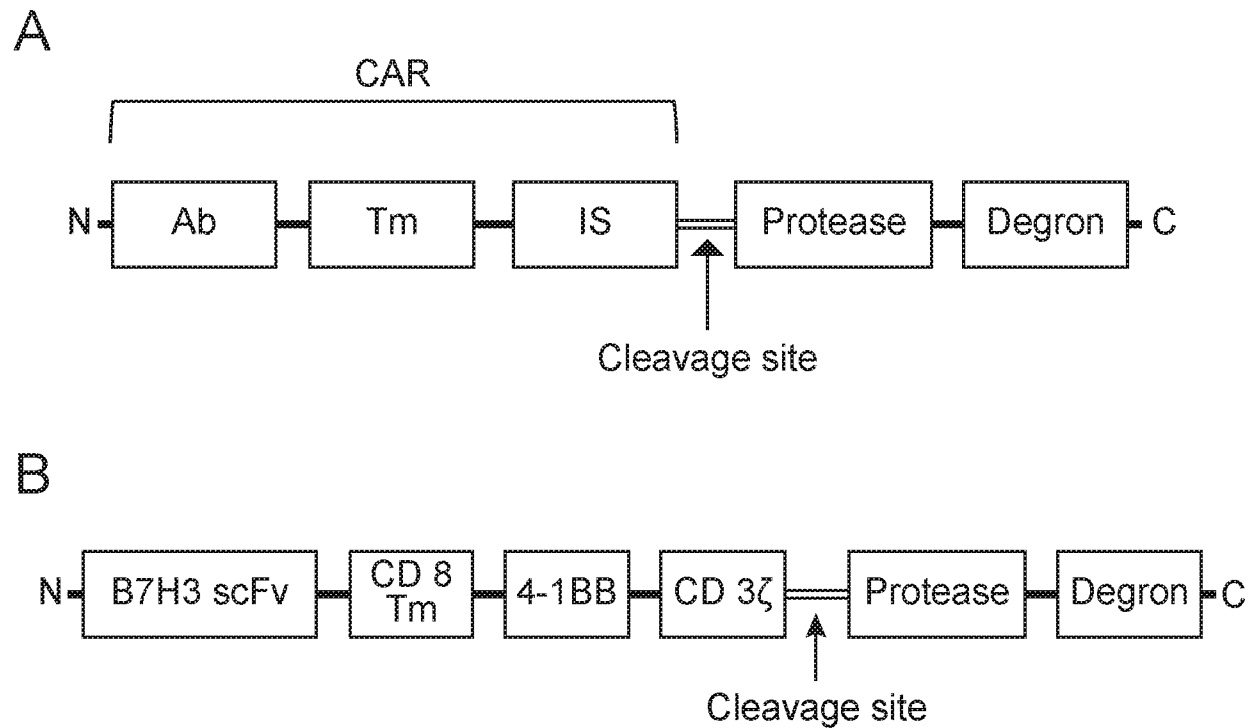
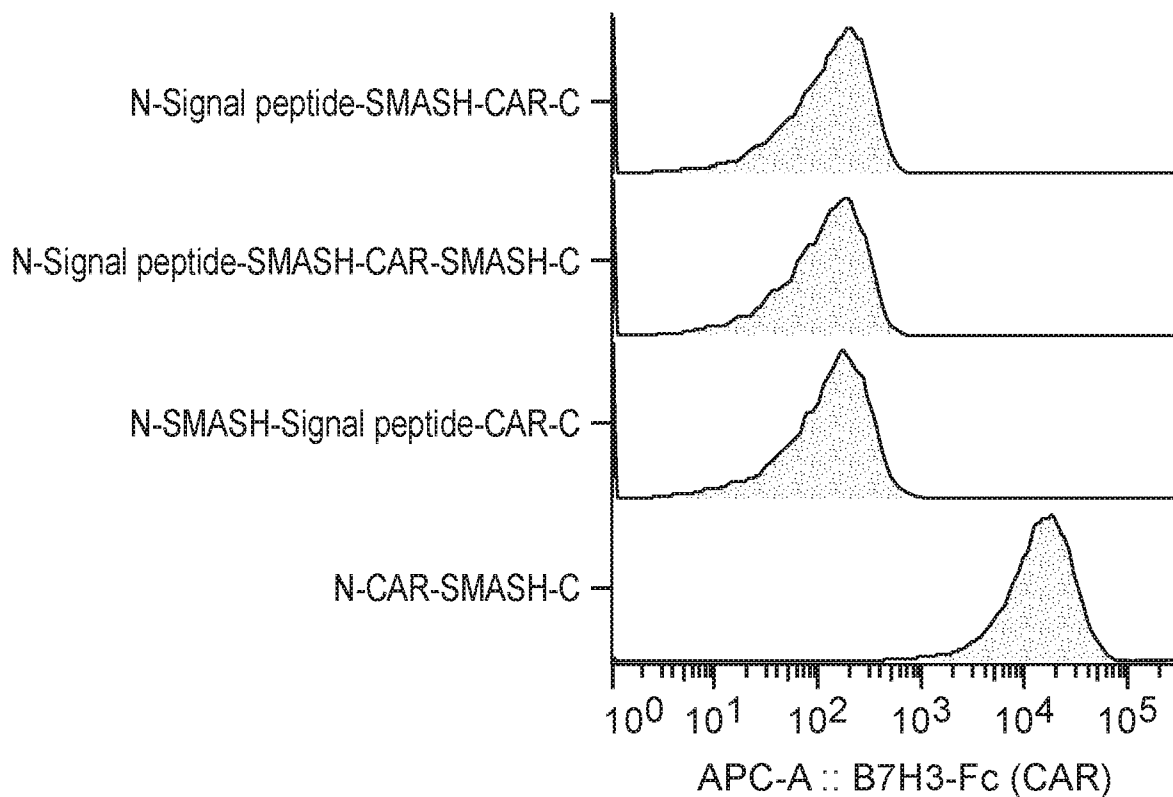
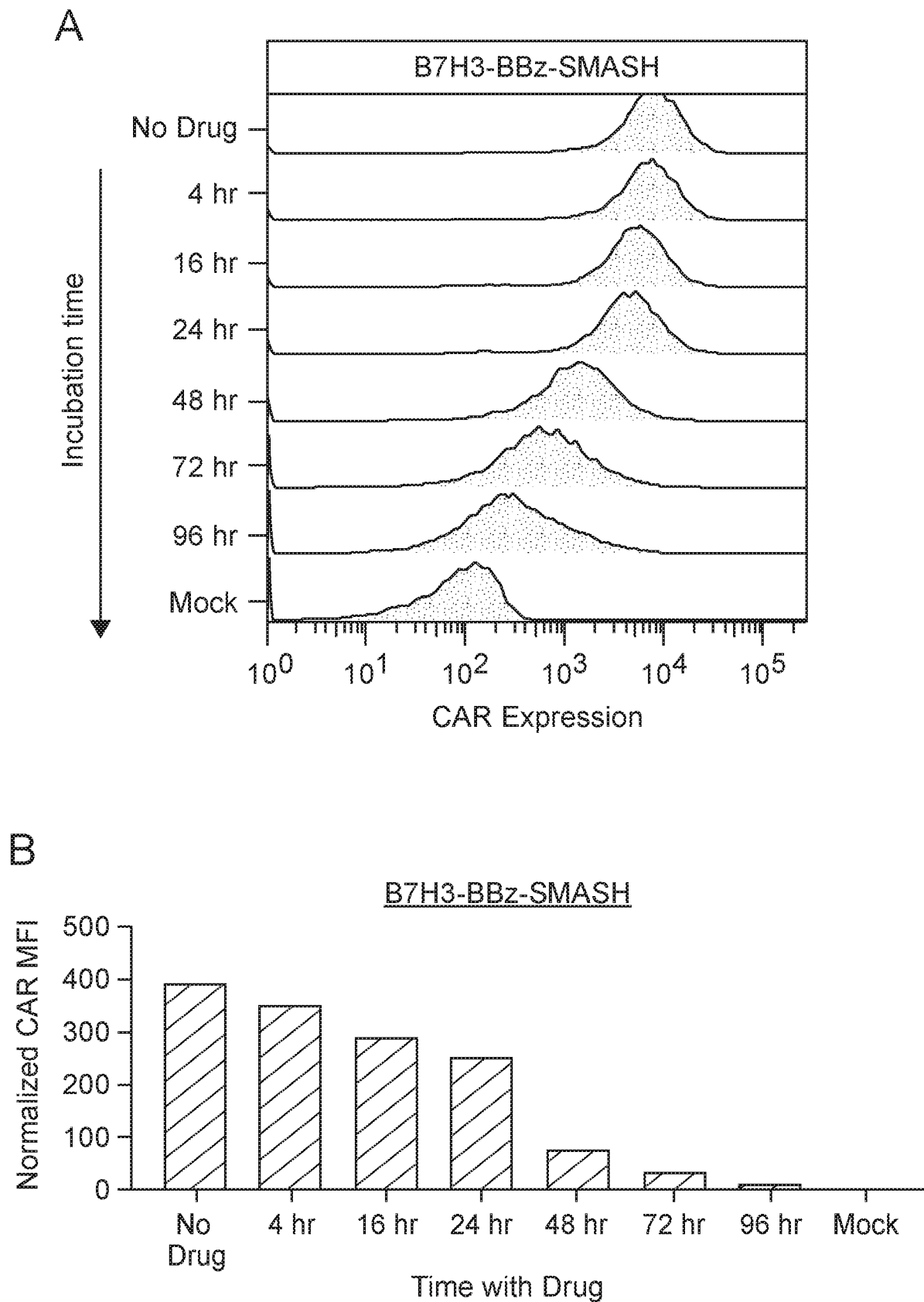


FIG. 3



3/33

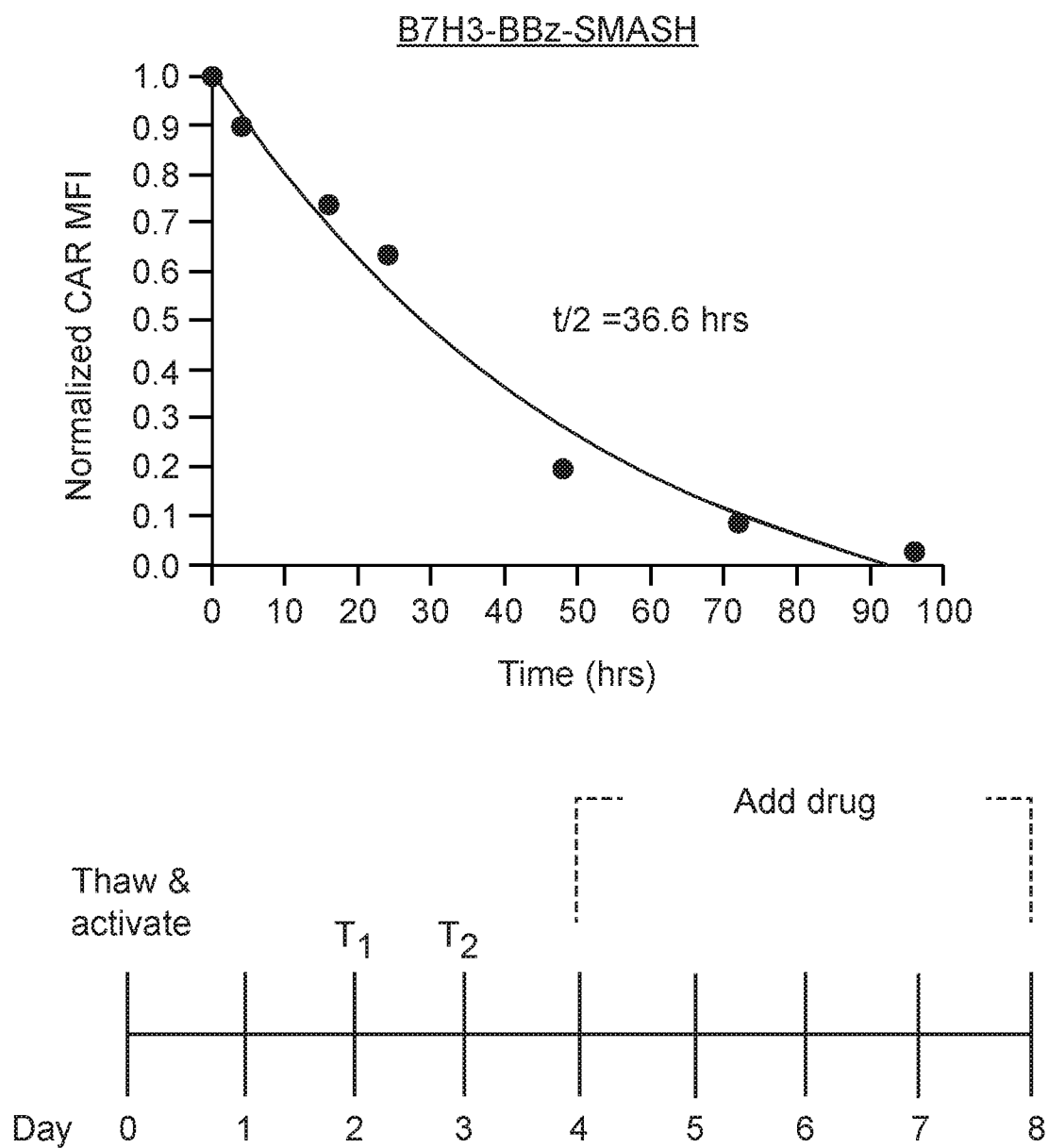
FIG. 4



4/33

FIG. 4 (Cont.)

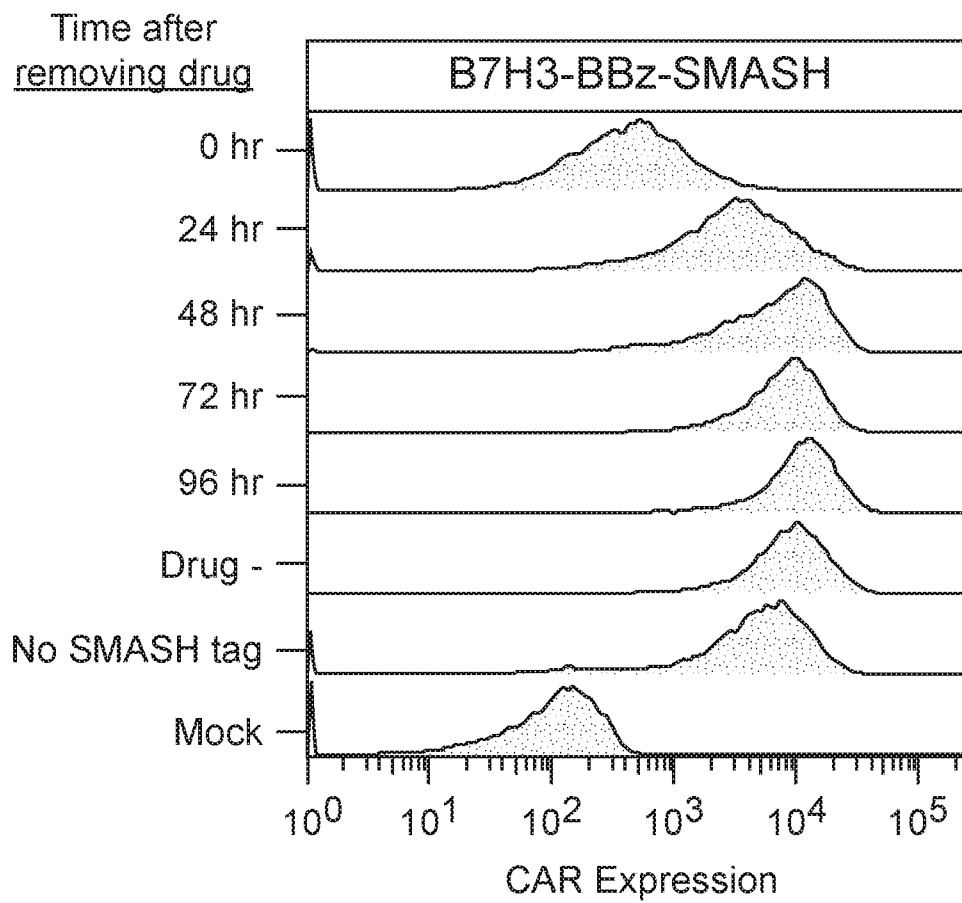
C



5/33

FIG. 5

A



B

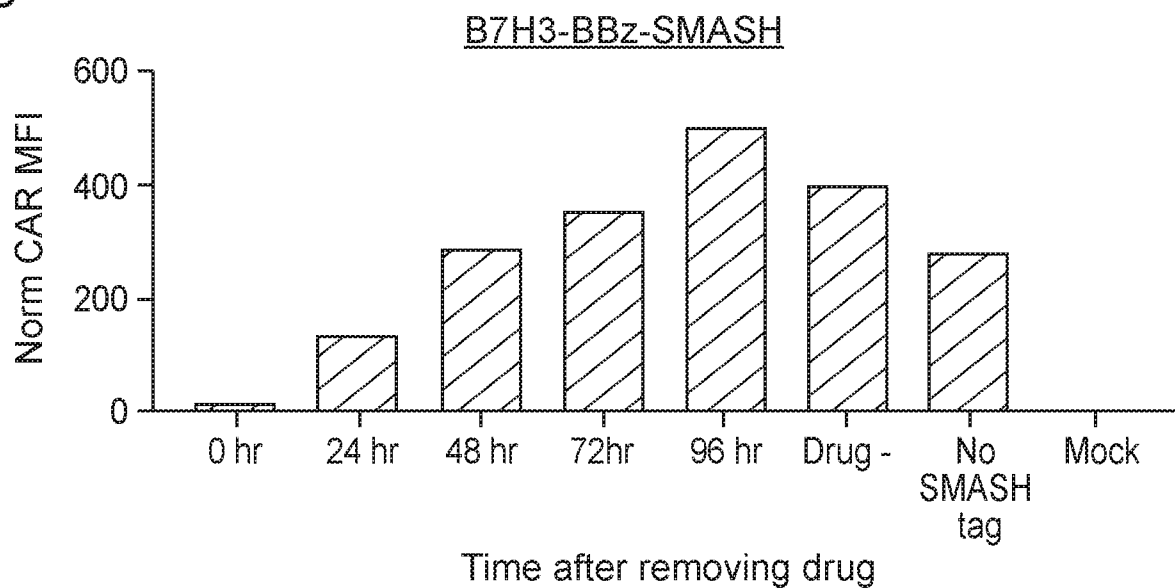


FIG. 6

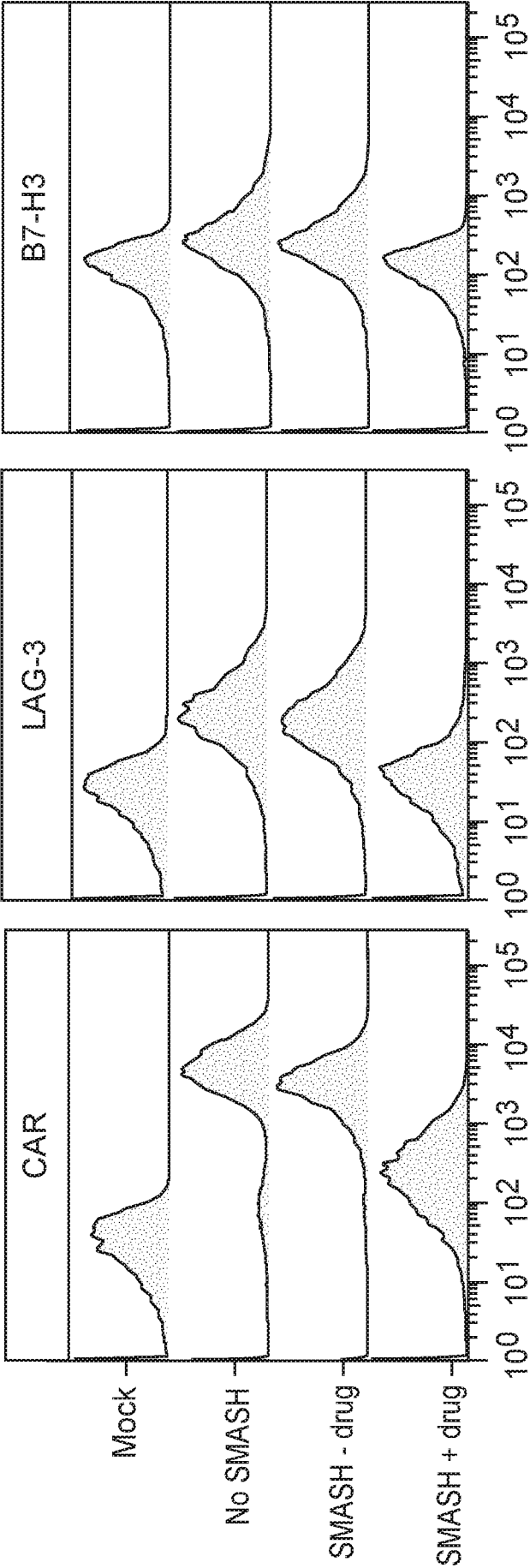
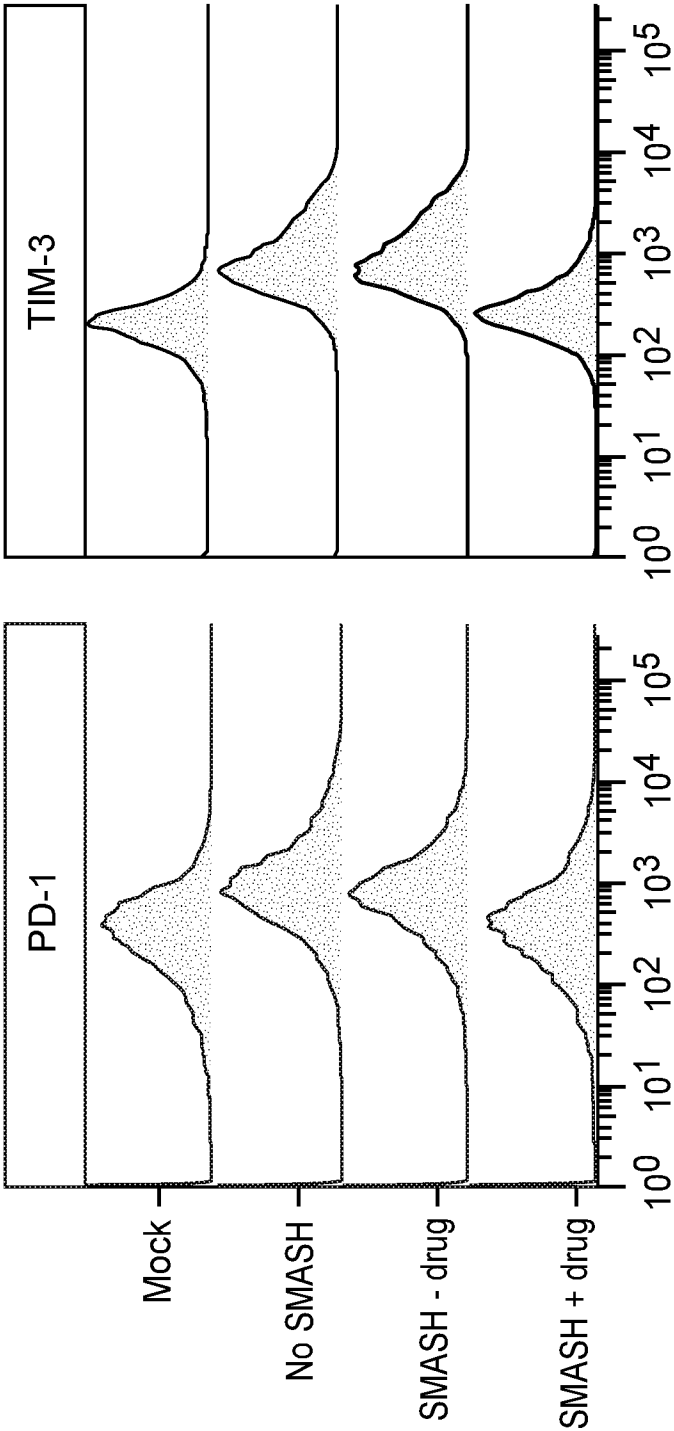


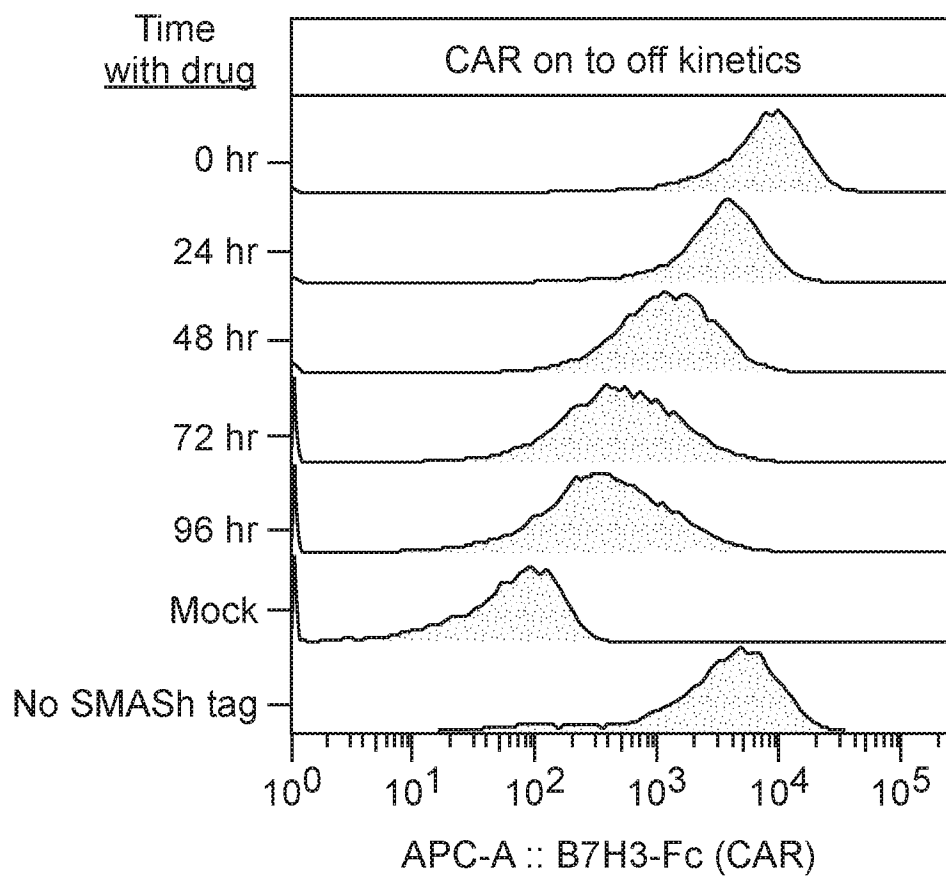
FIG. 6 (Cont.)



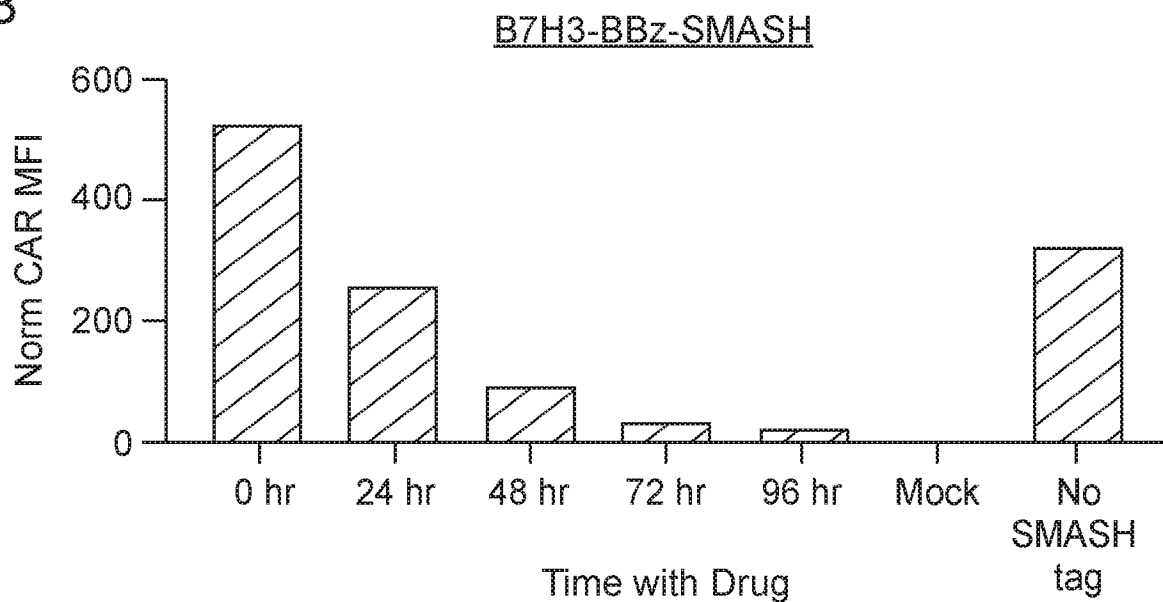
8/33

FIG. 7

A



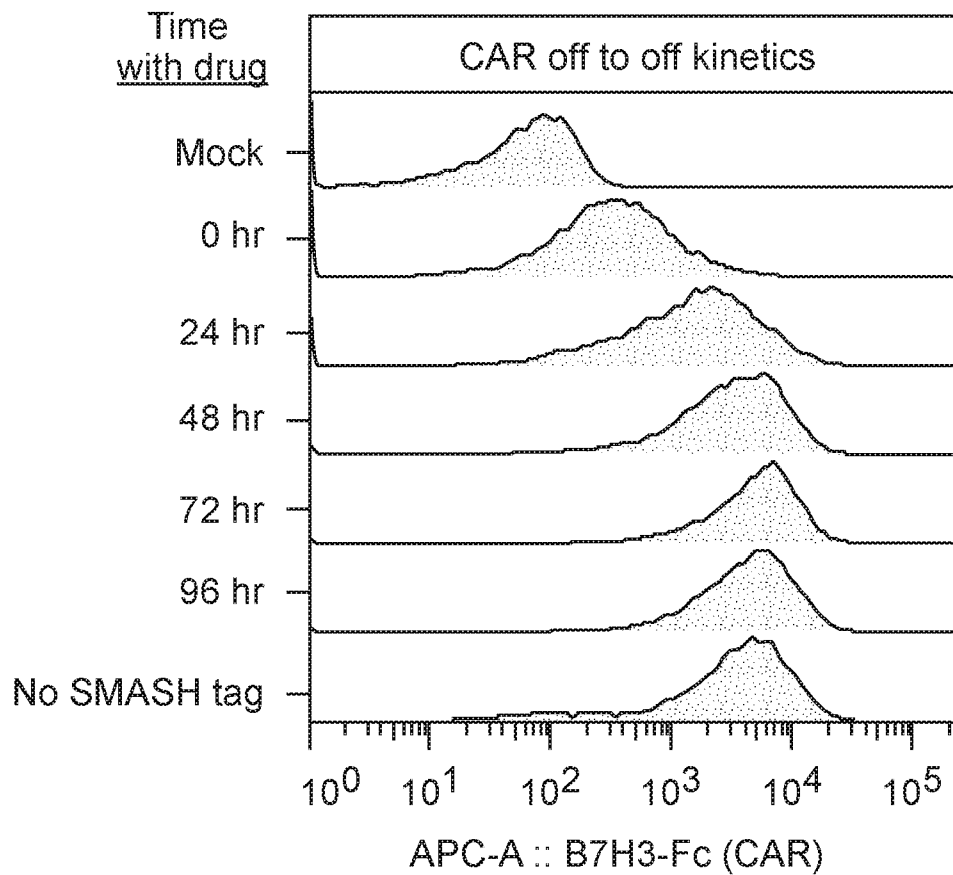
B



9/33

FIG. 8

A



B

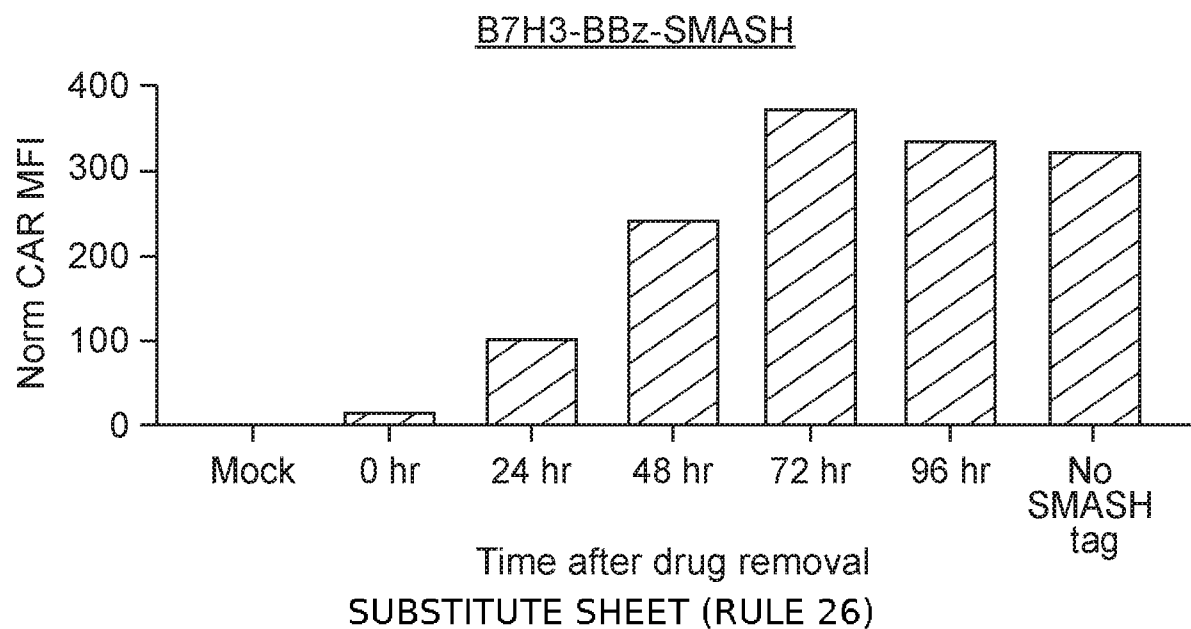


FIG. 9

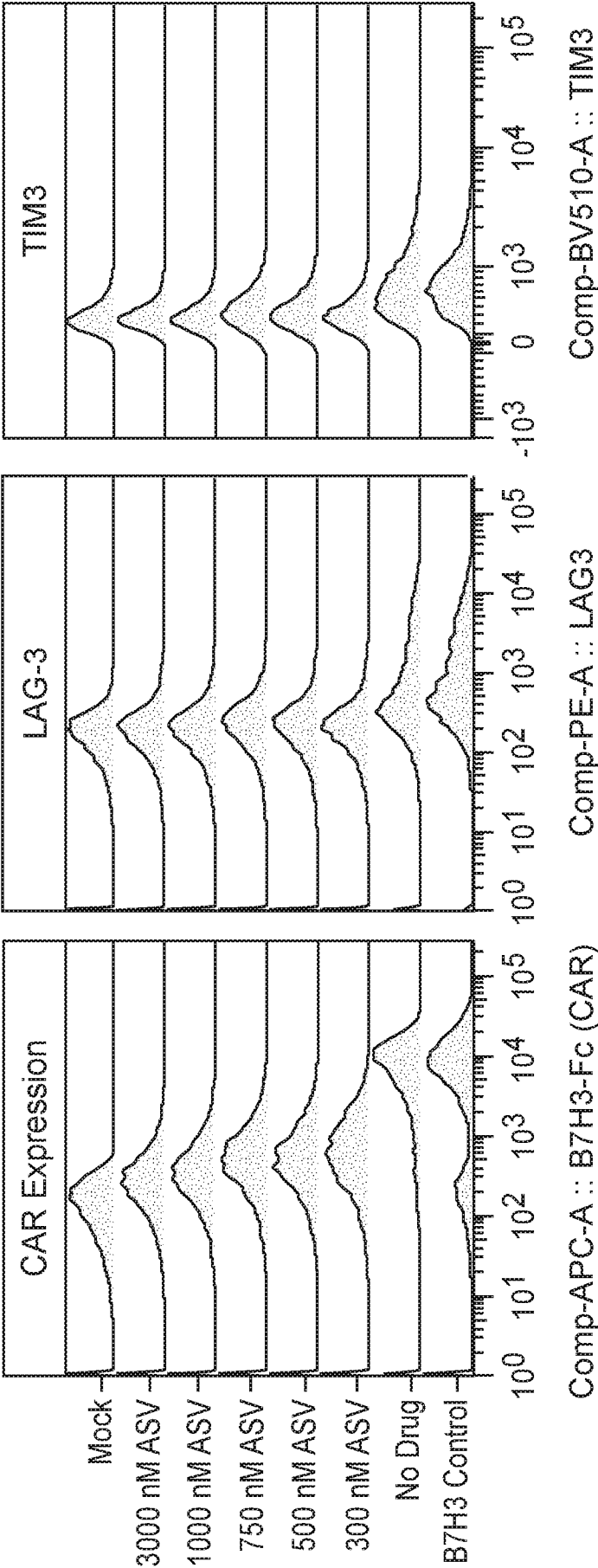
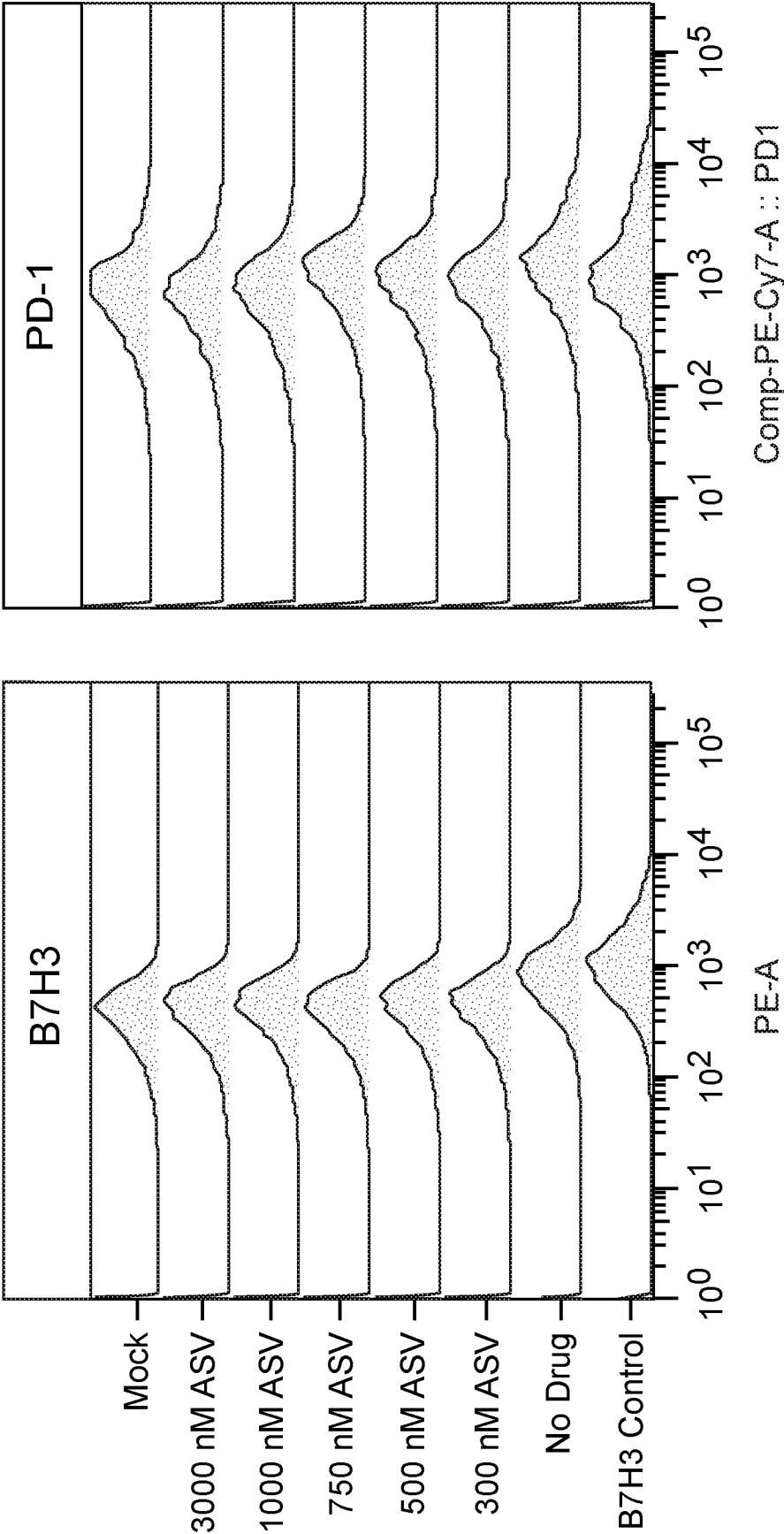
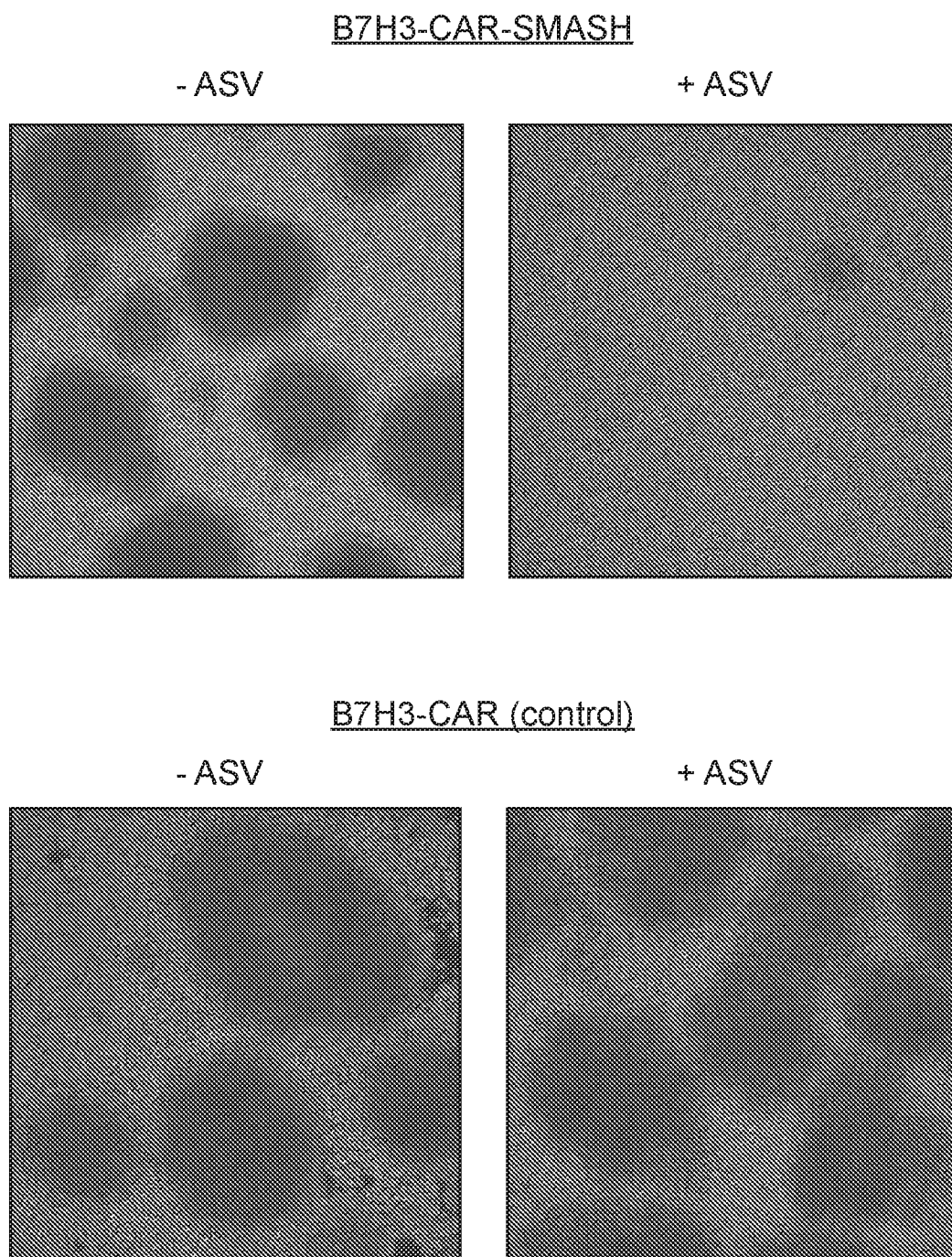


FIG. 9 (Cont.)



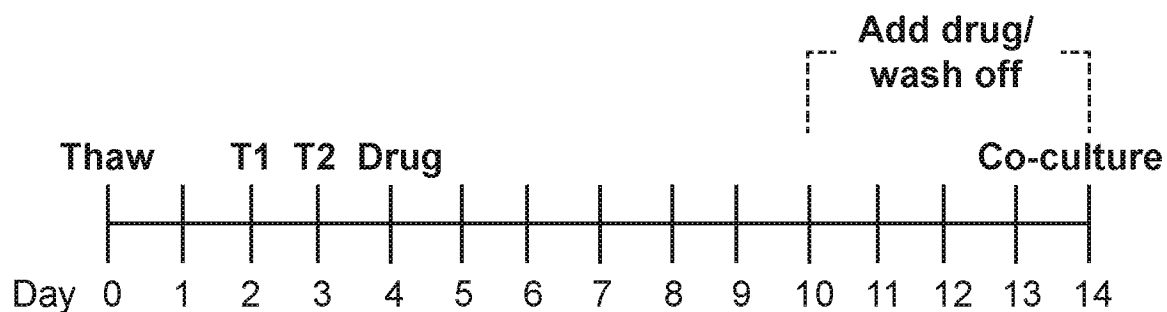
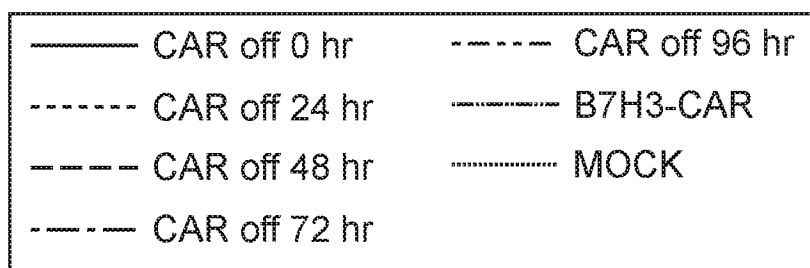
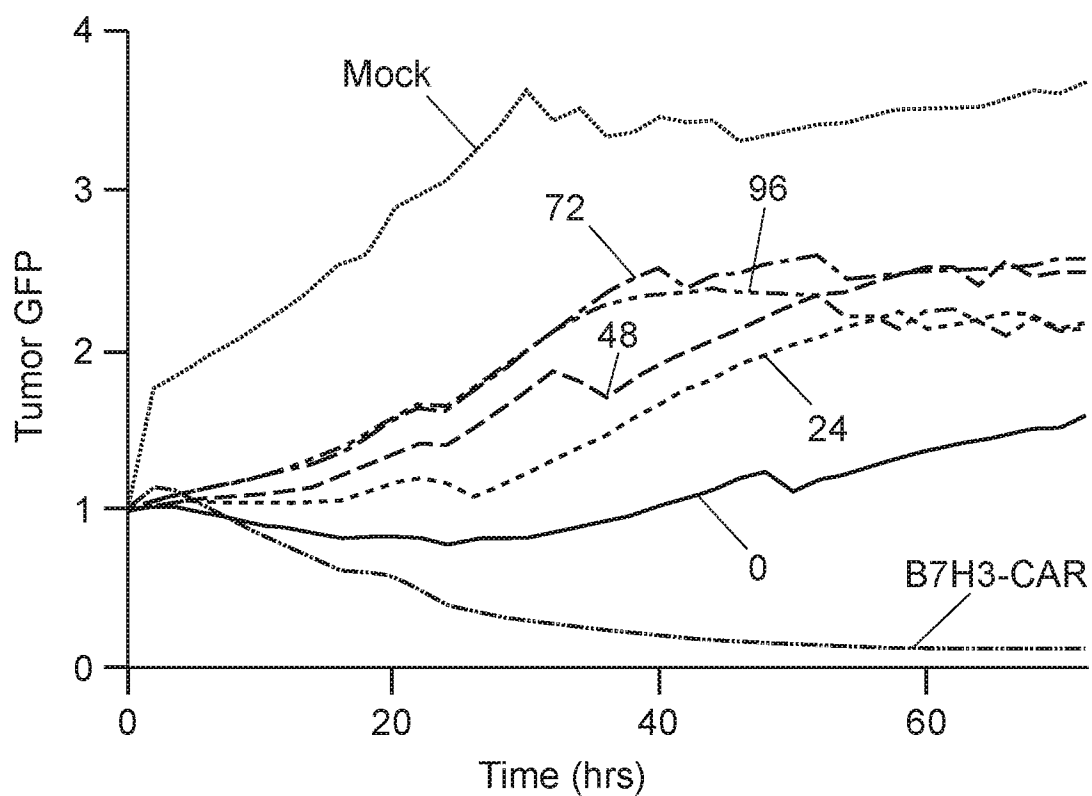
12/33

FIG. 10



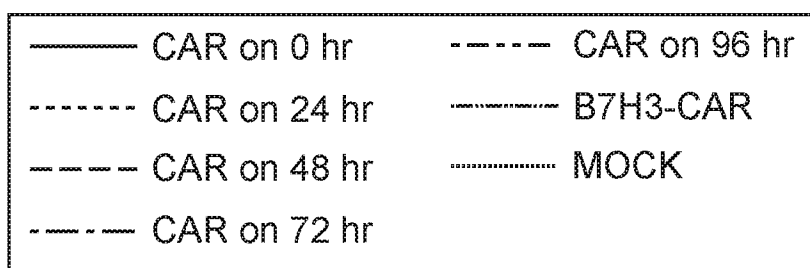
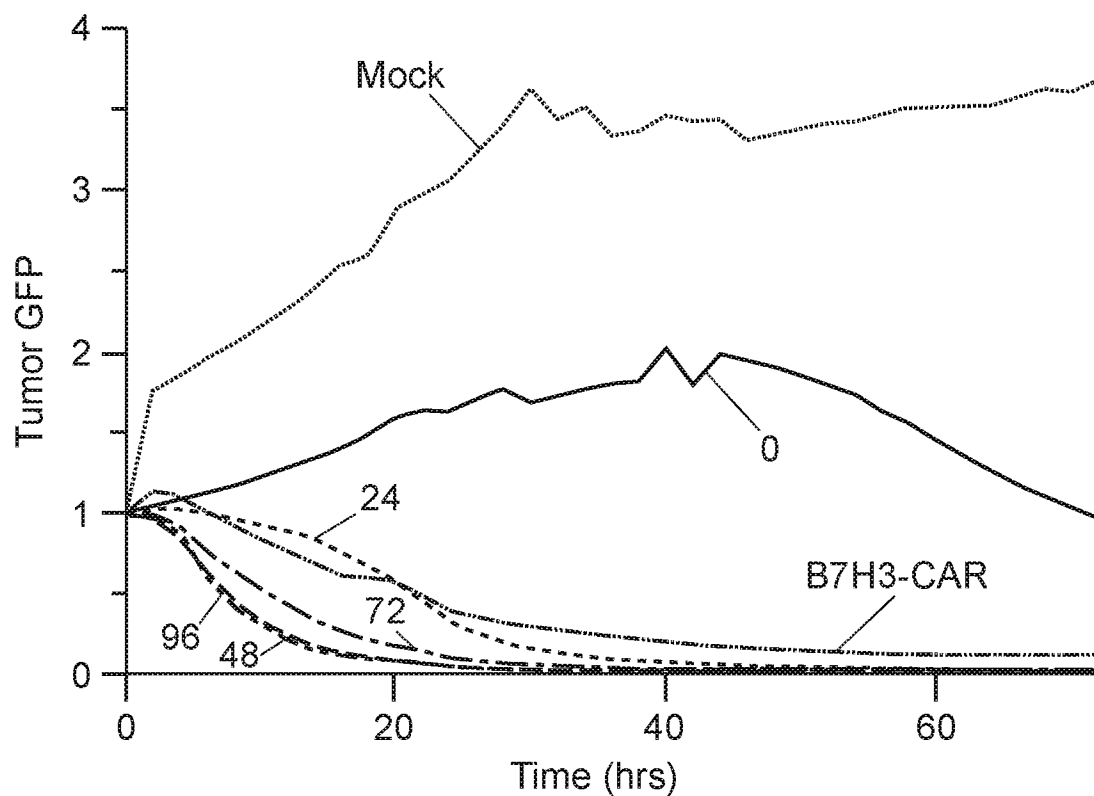
13/33

FIG. 11



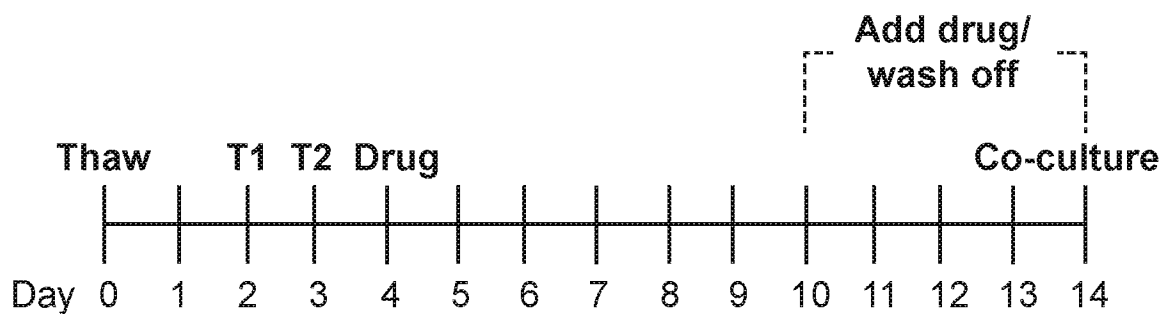
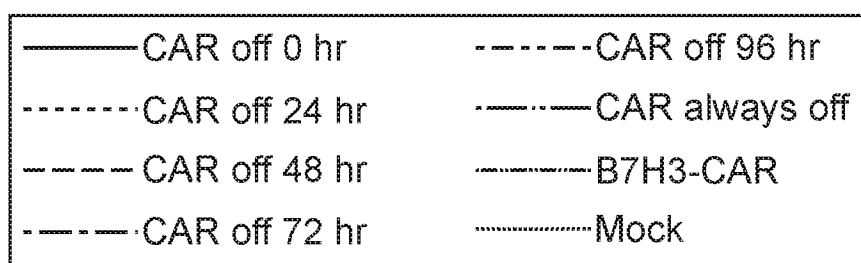
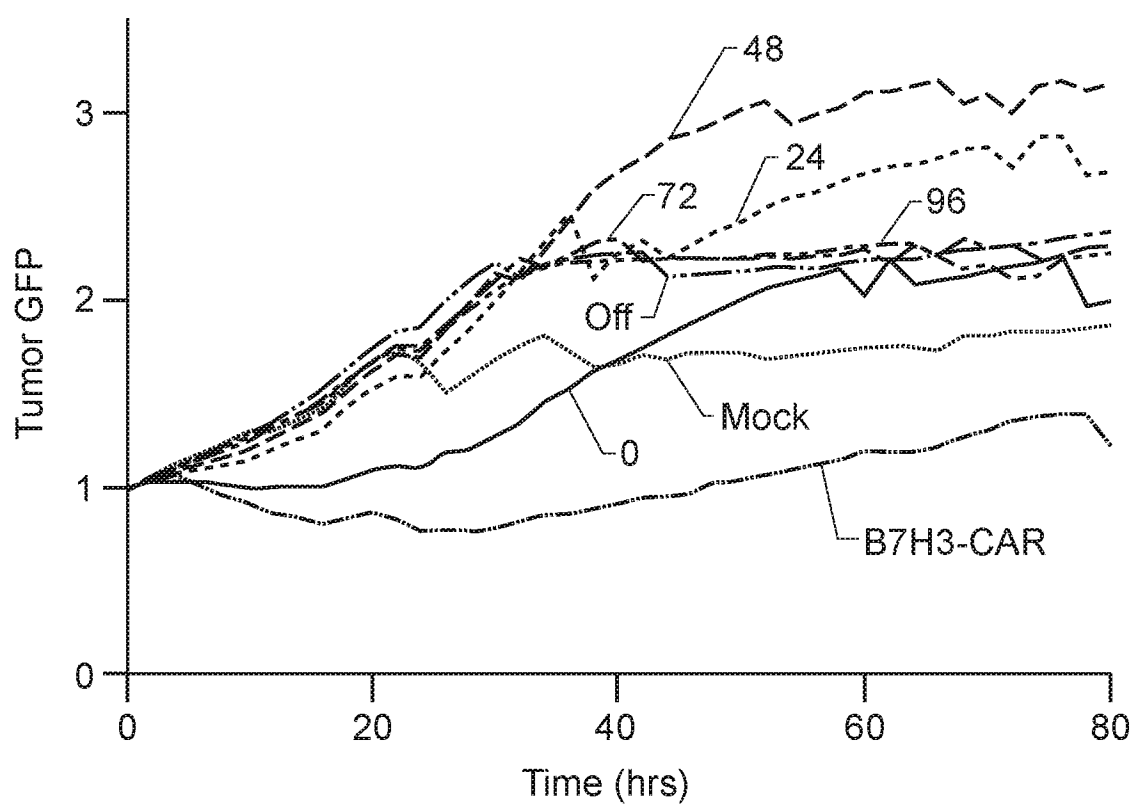
14/33

FIG. 12



15/33

FIG. 13

CAR OFF vs. Low B7H3

16/33

FIG. 14

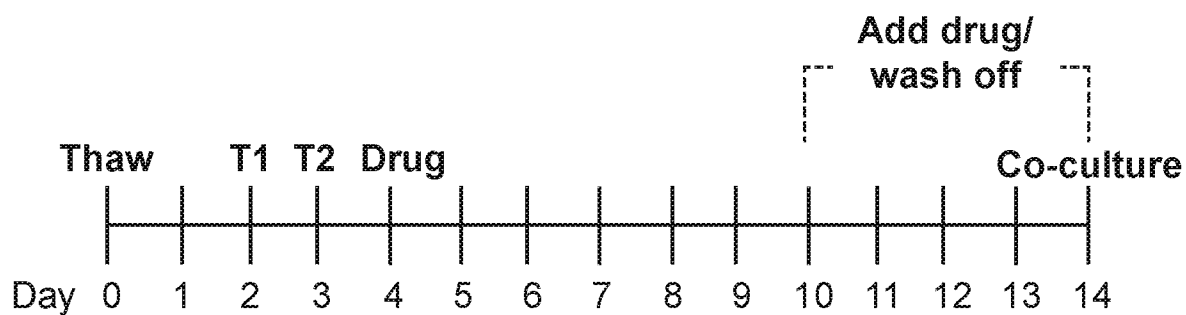
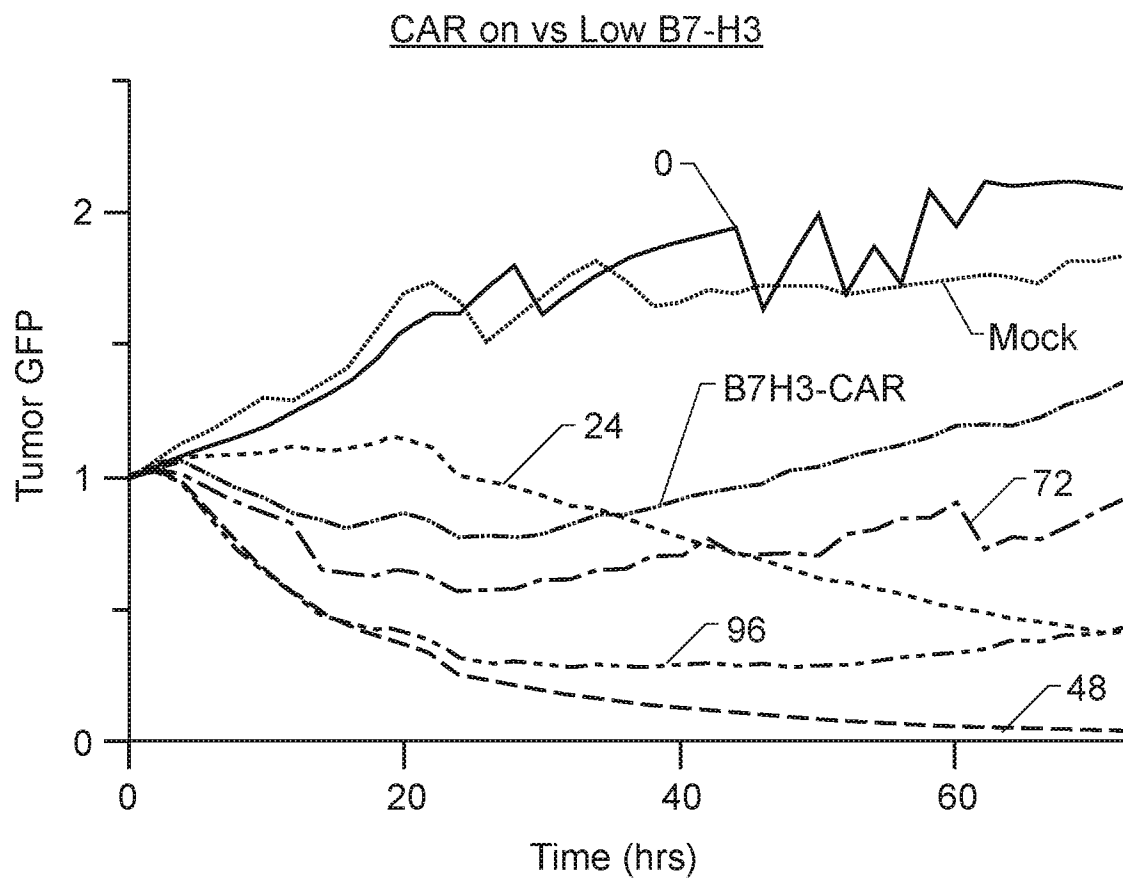


FIG. 15

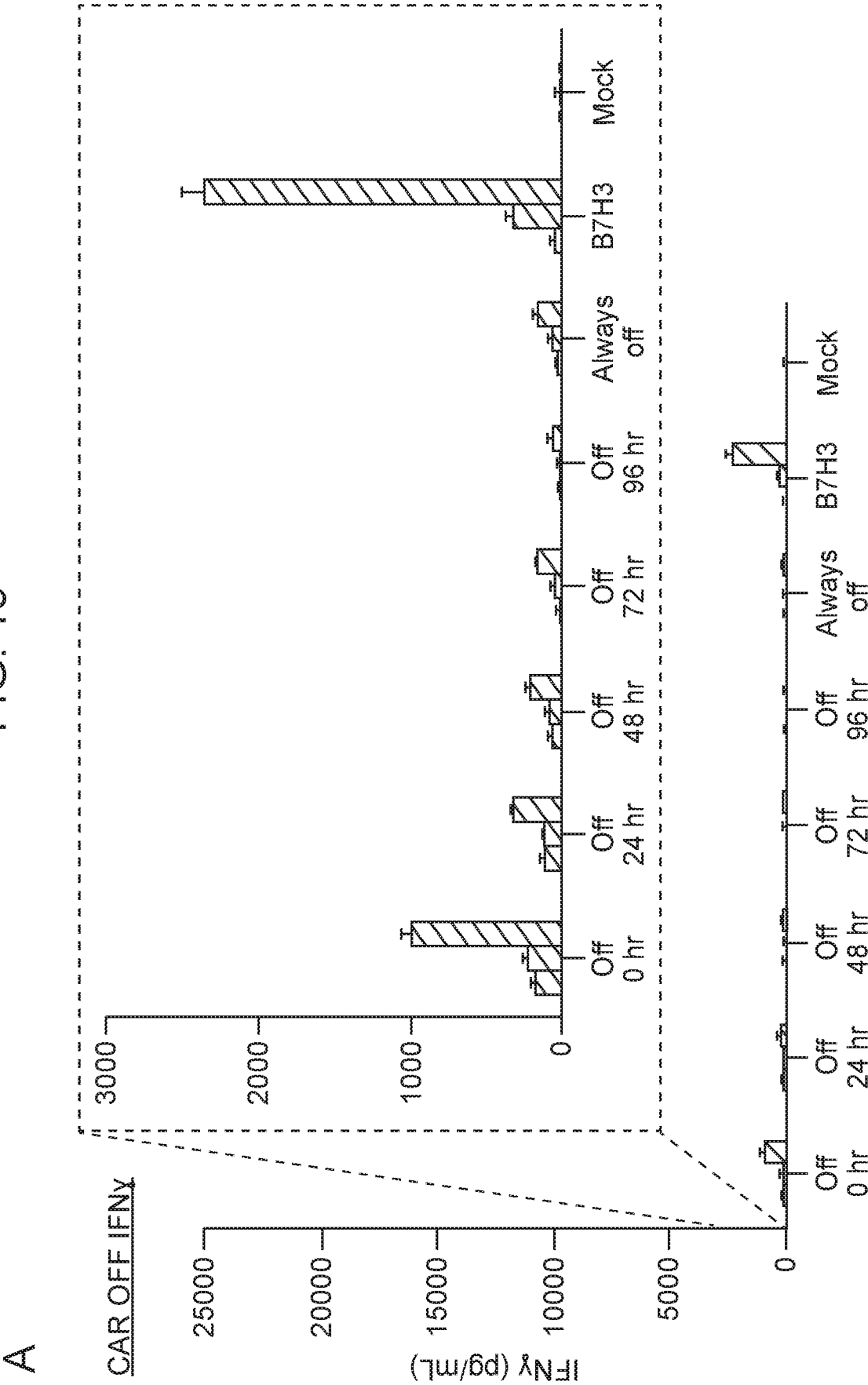


FIG. 15 (Cont.)

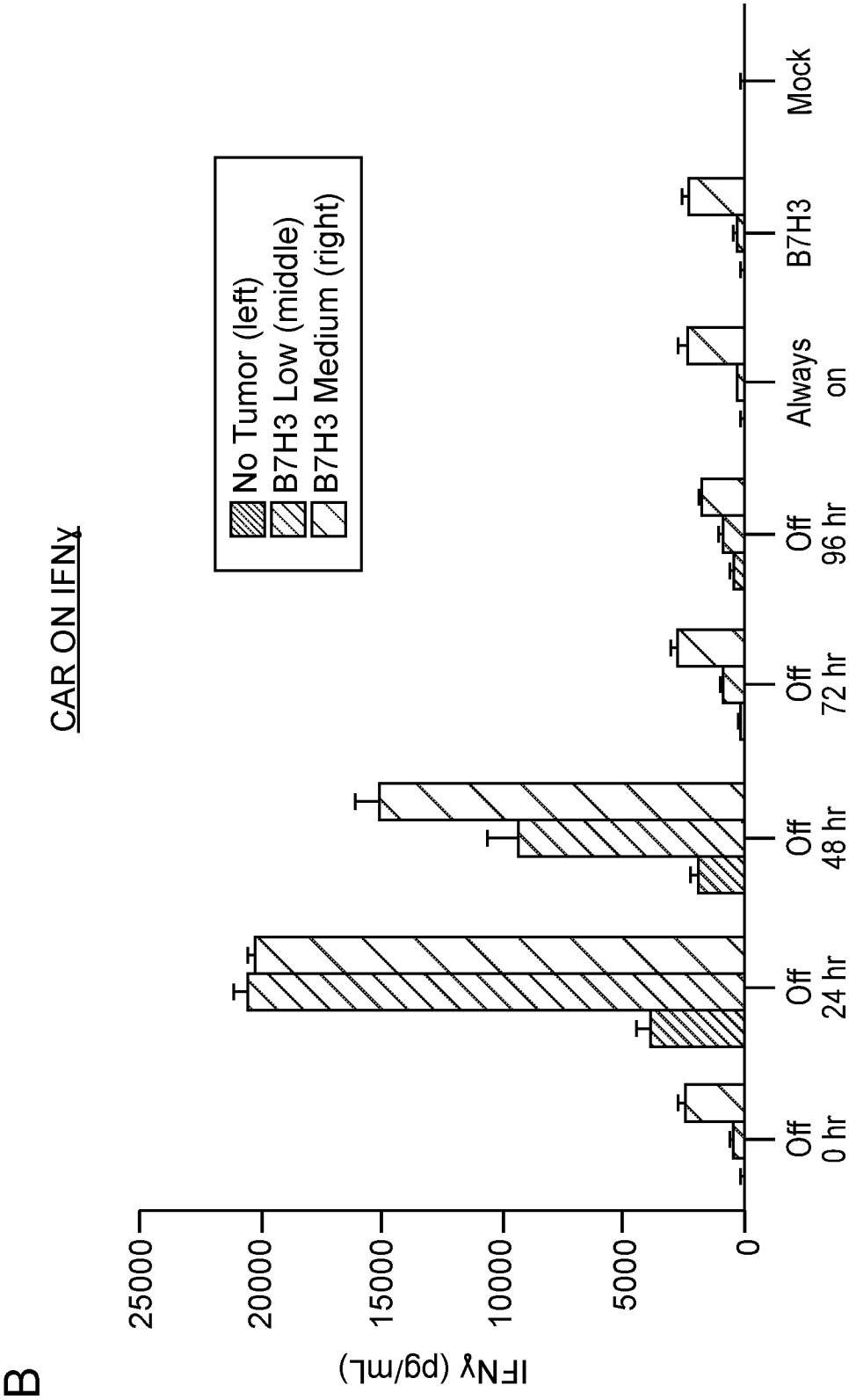


FIG. 16

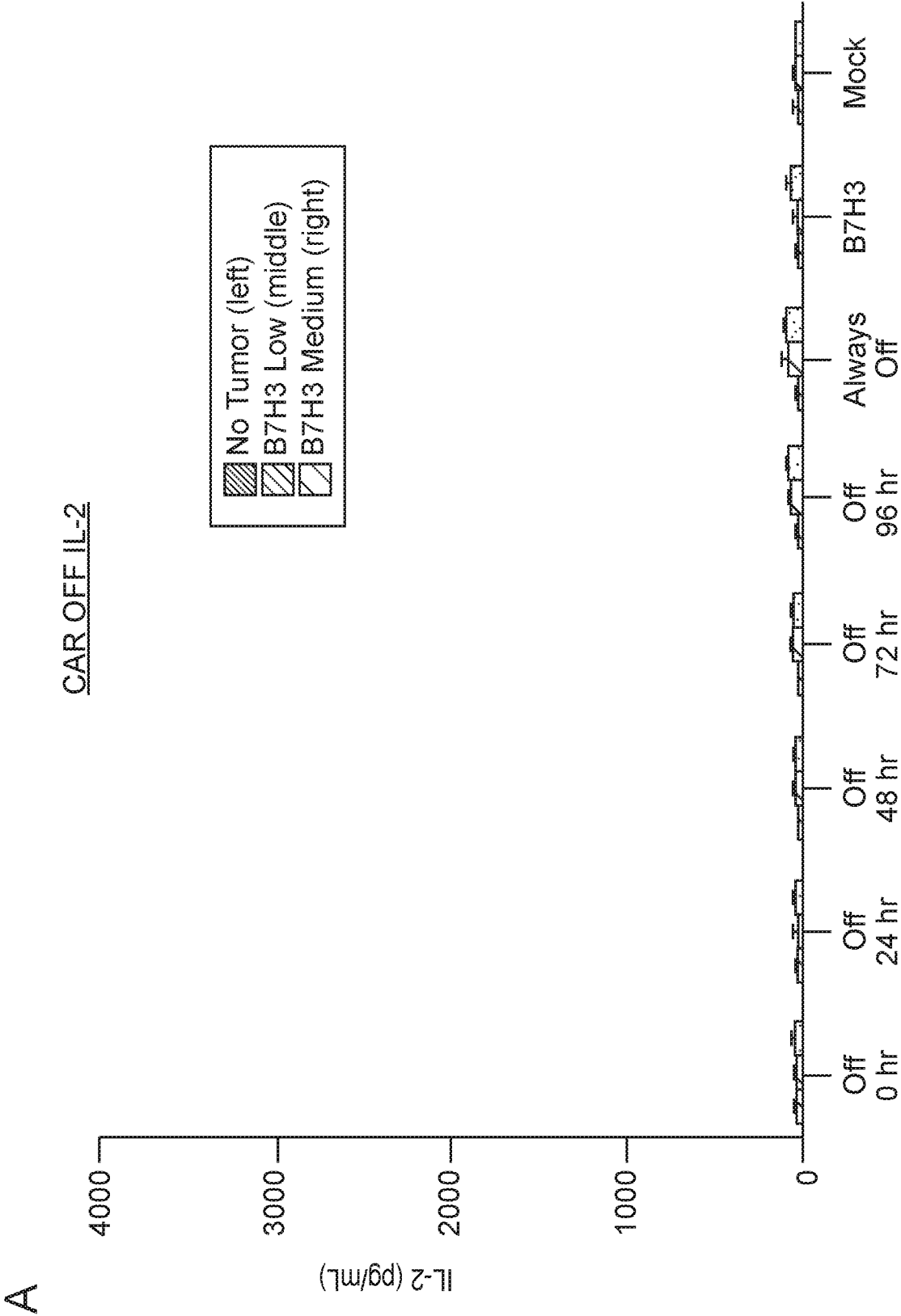
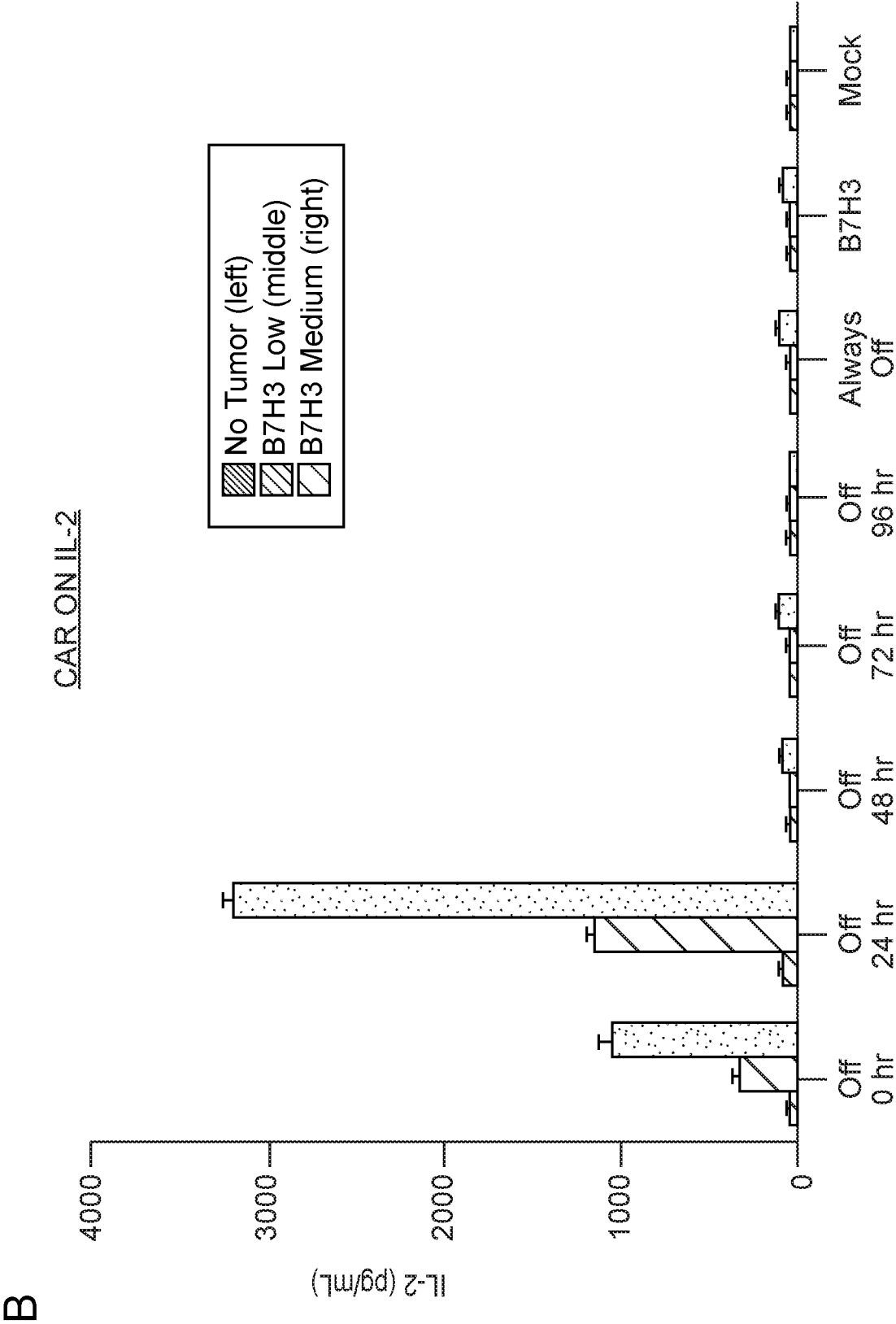


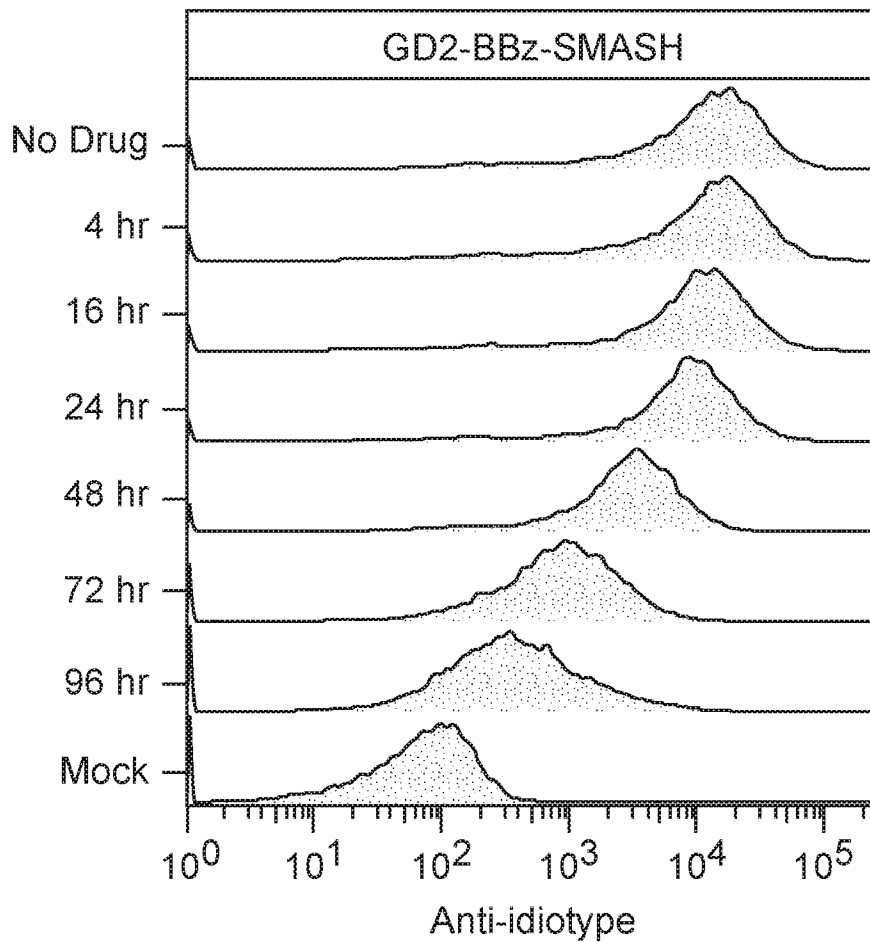
FIG. 16 (Cont.)



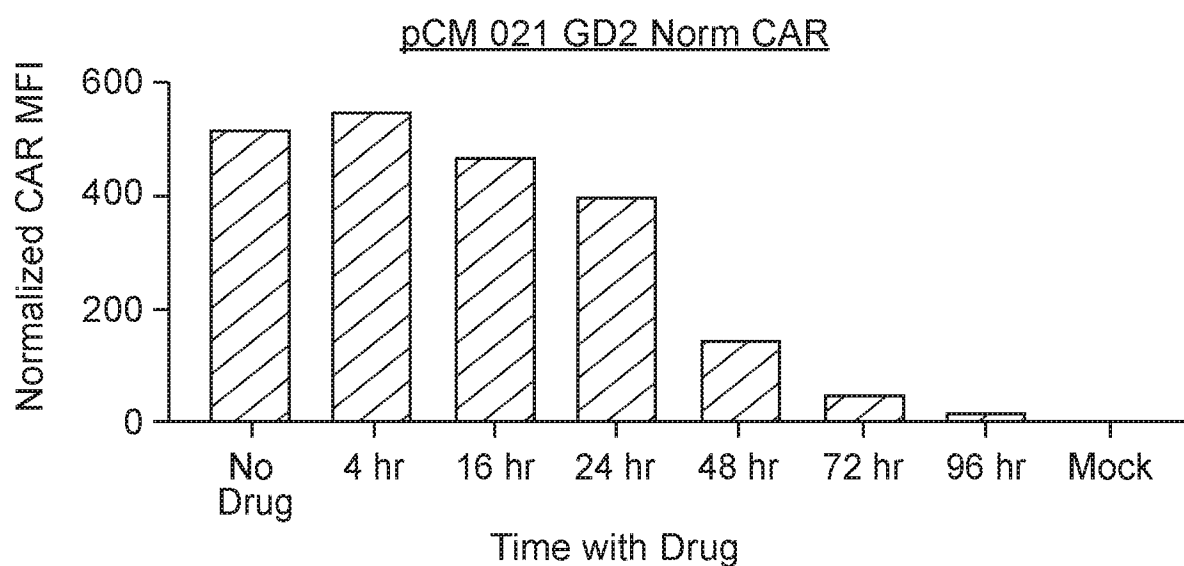
21/33

FIG. 17

A



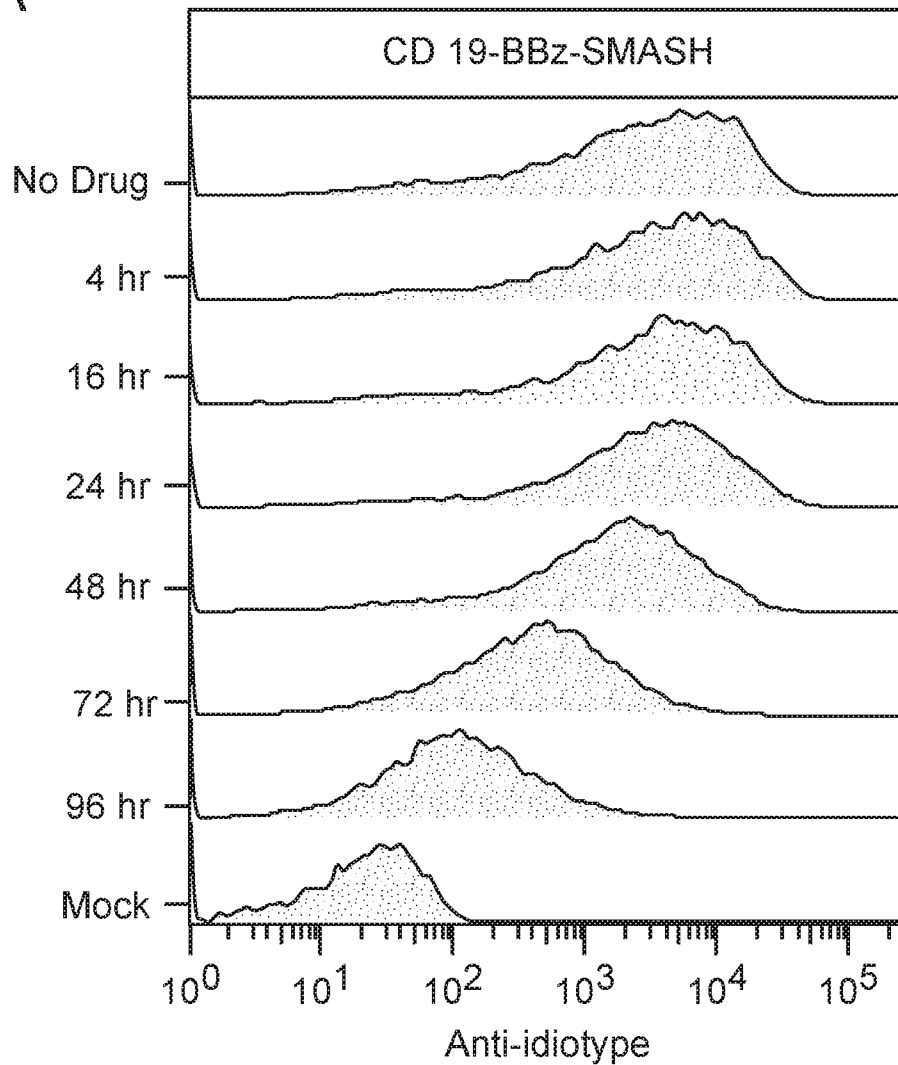
B



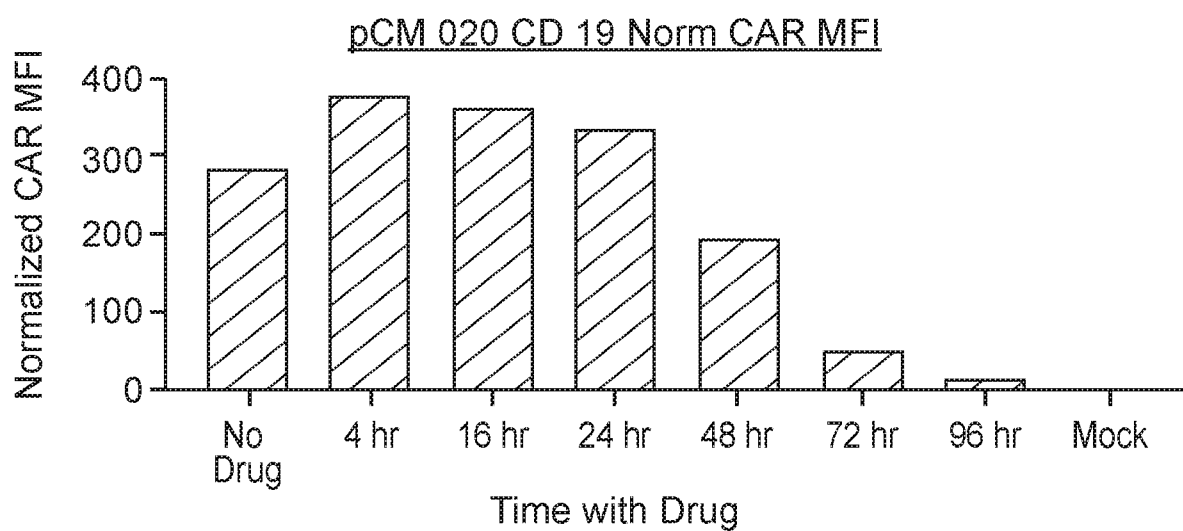
22/33

FIG. 18

A

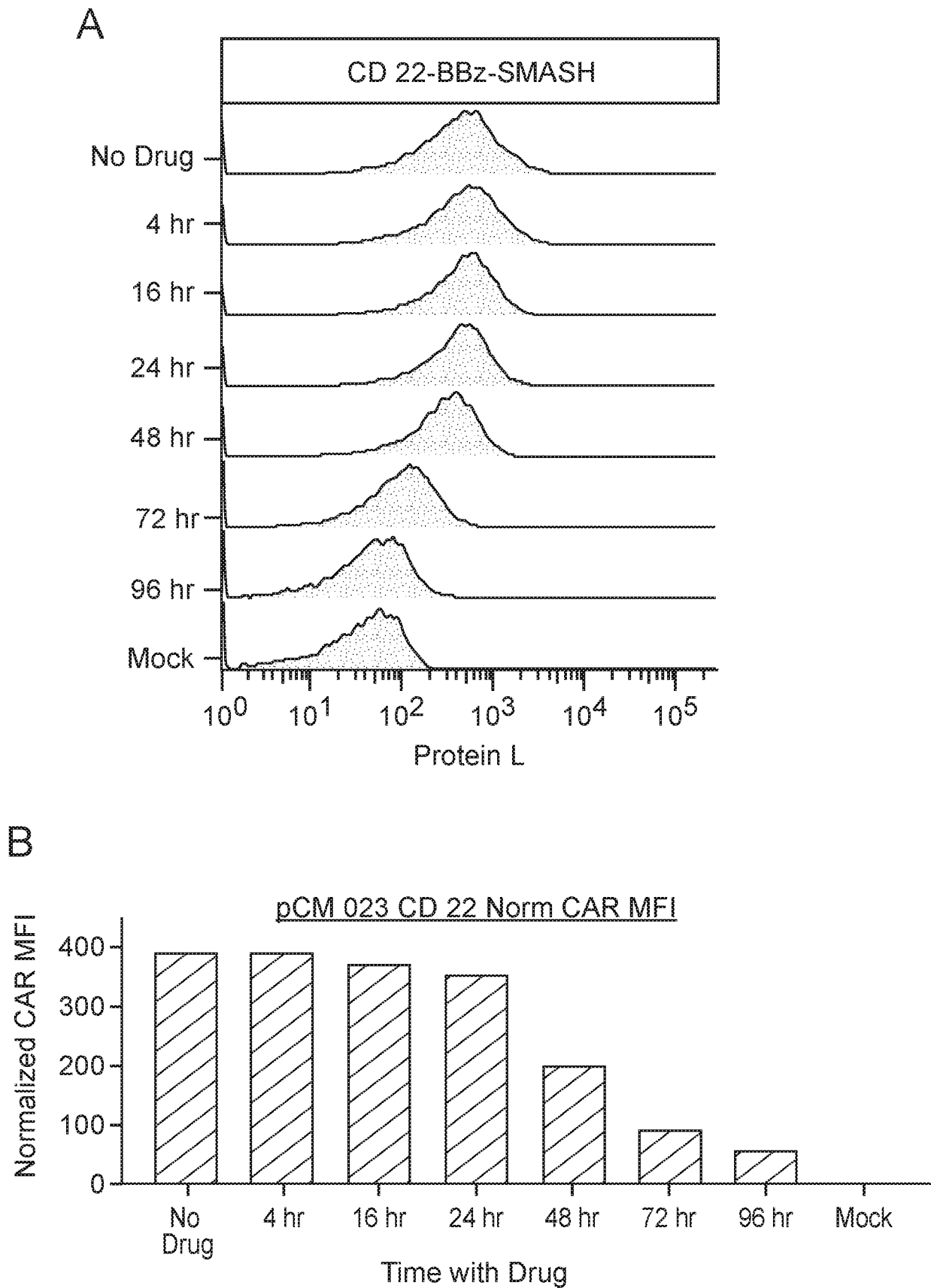


B



23/33

FIG. 19



24/33

FIG. 20

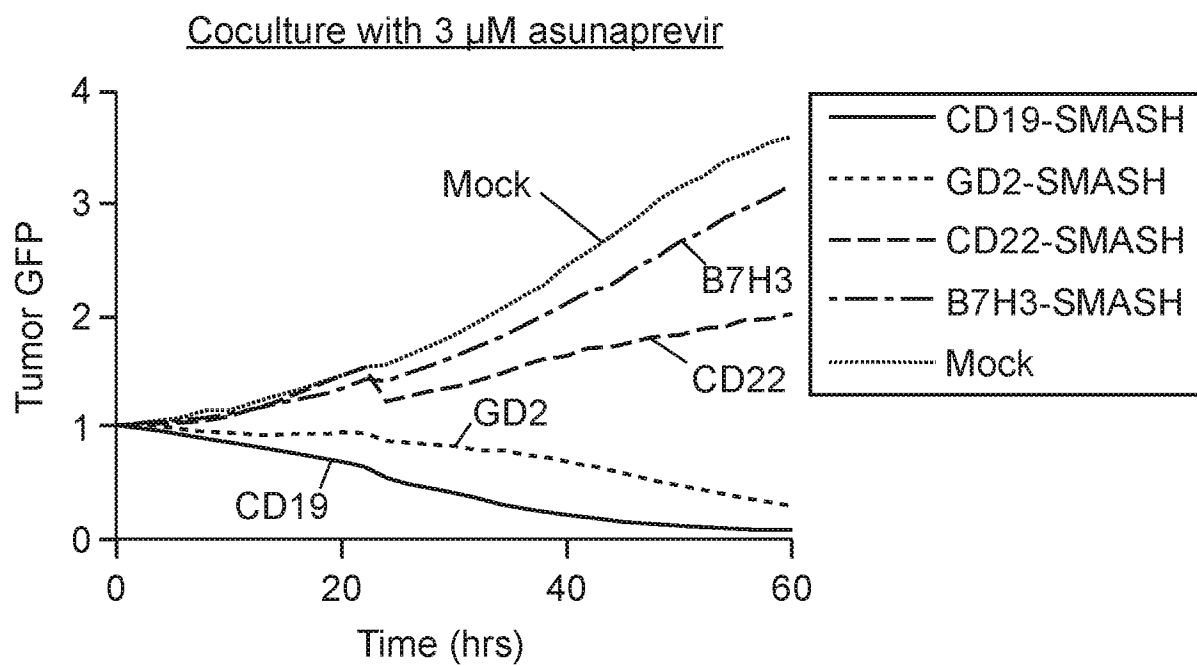


FIG. 21

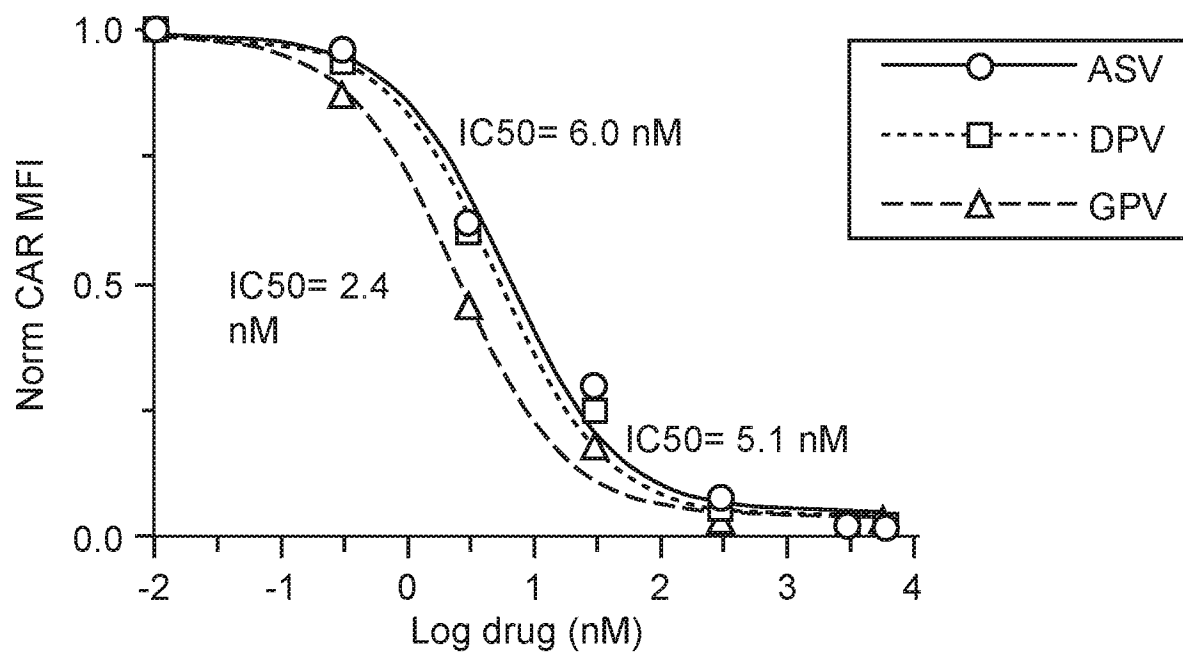
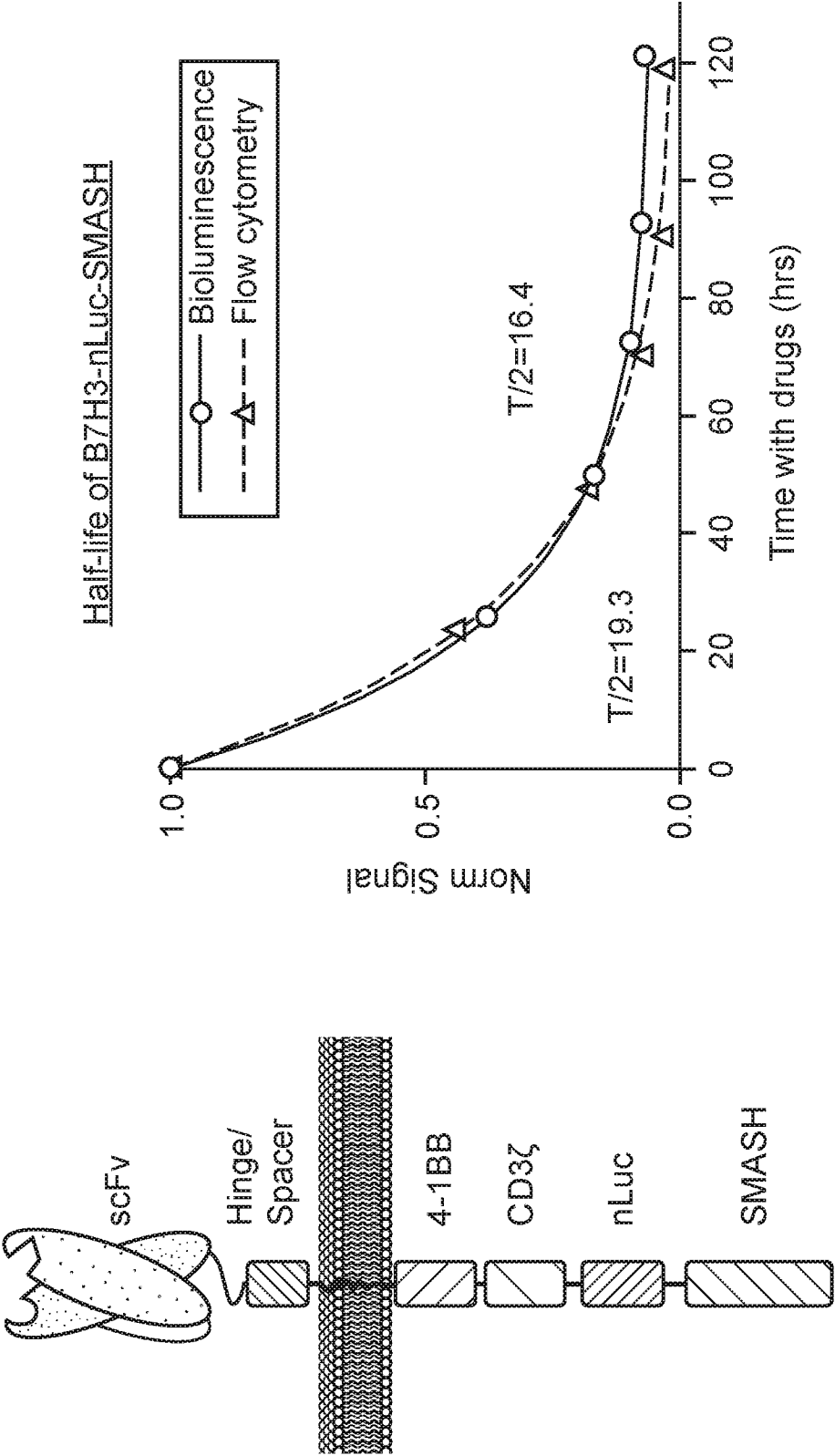
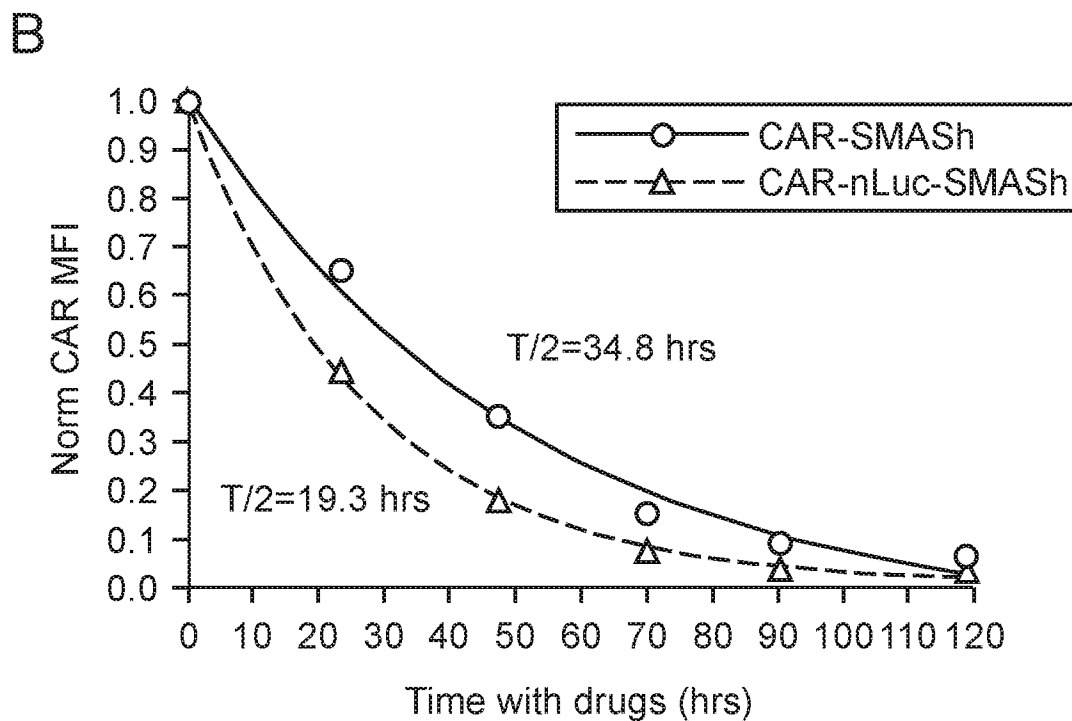
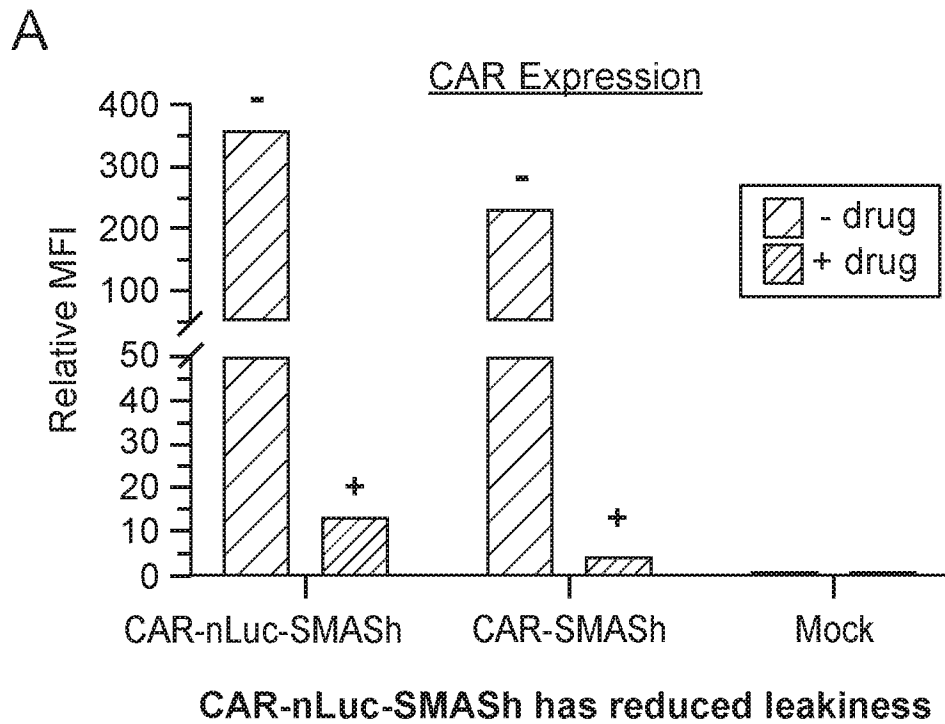


FIG. 22



26/33

FIG. 23



CAR-nLuc-SMASH has faster drug response time

FIG. 24

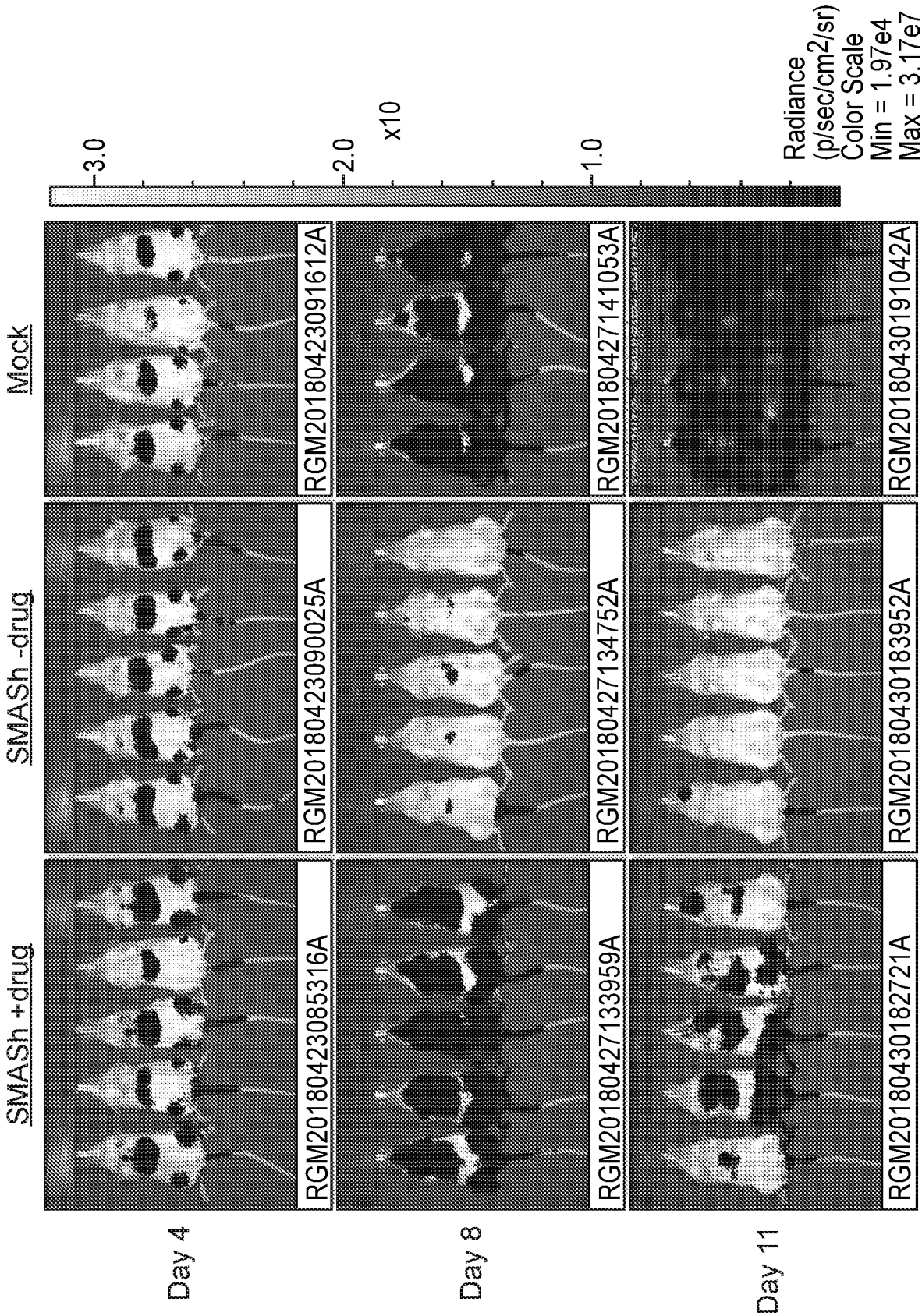


FIG. 24 (Cont.)

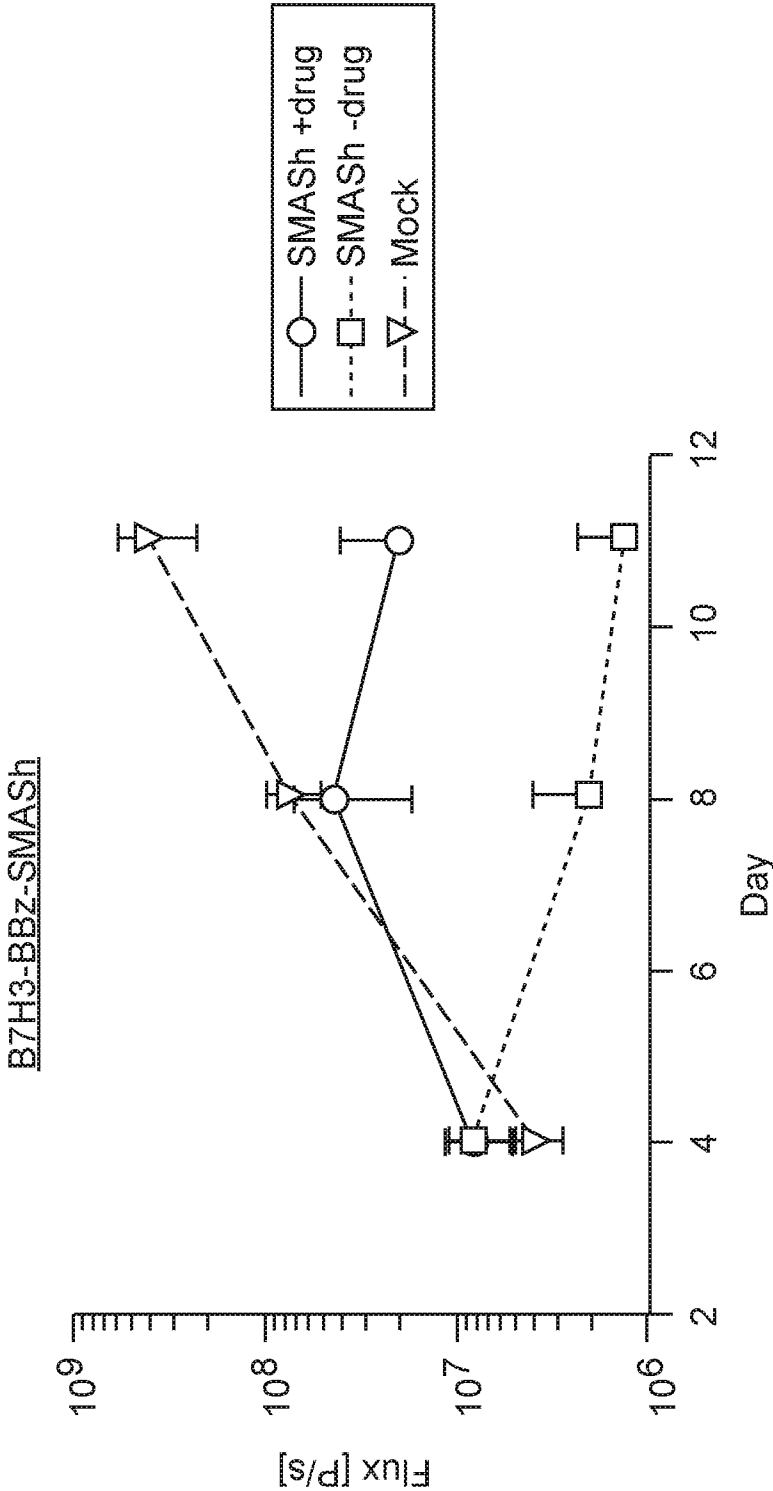
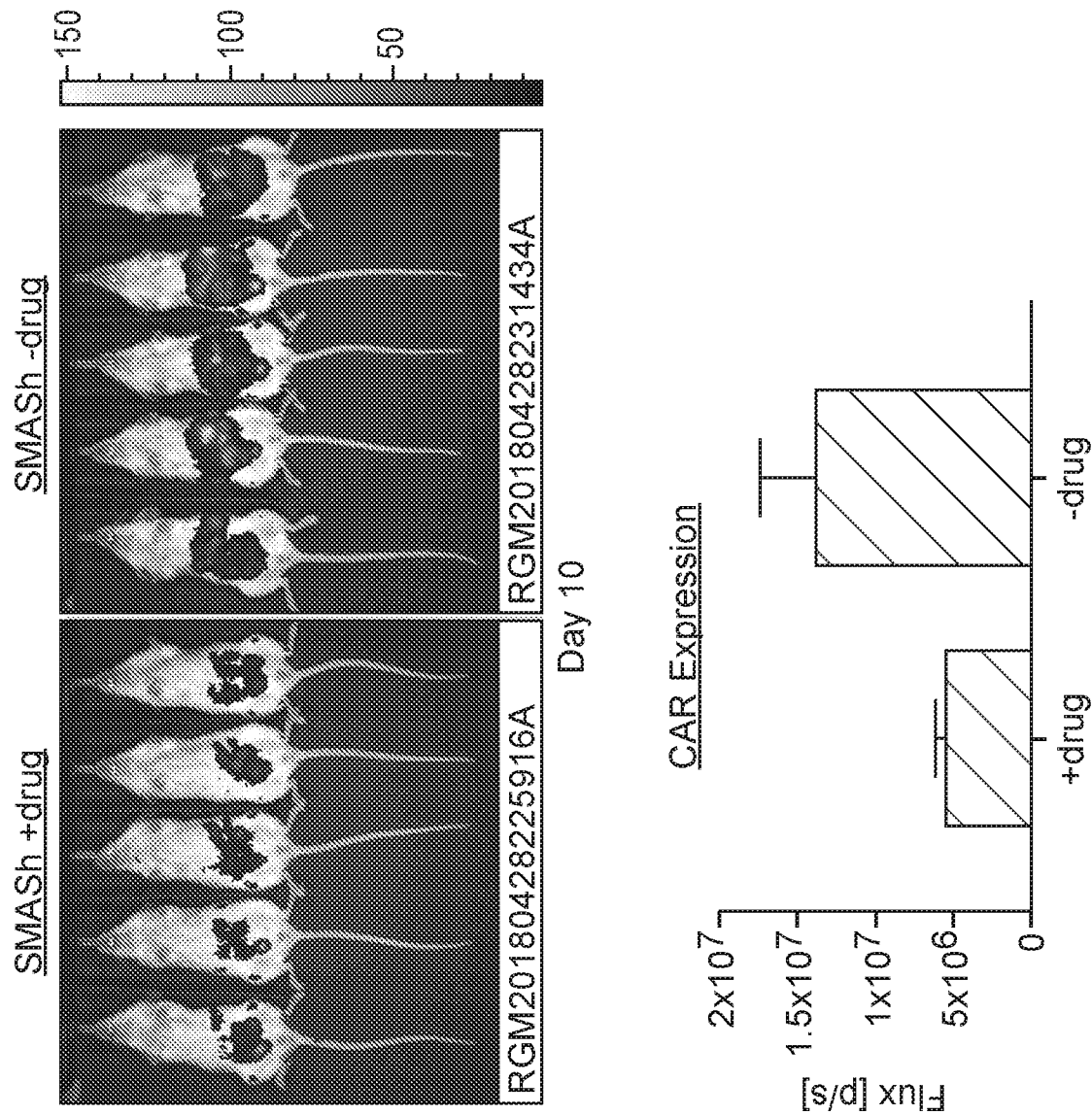
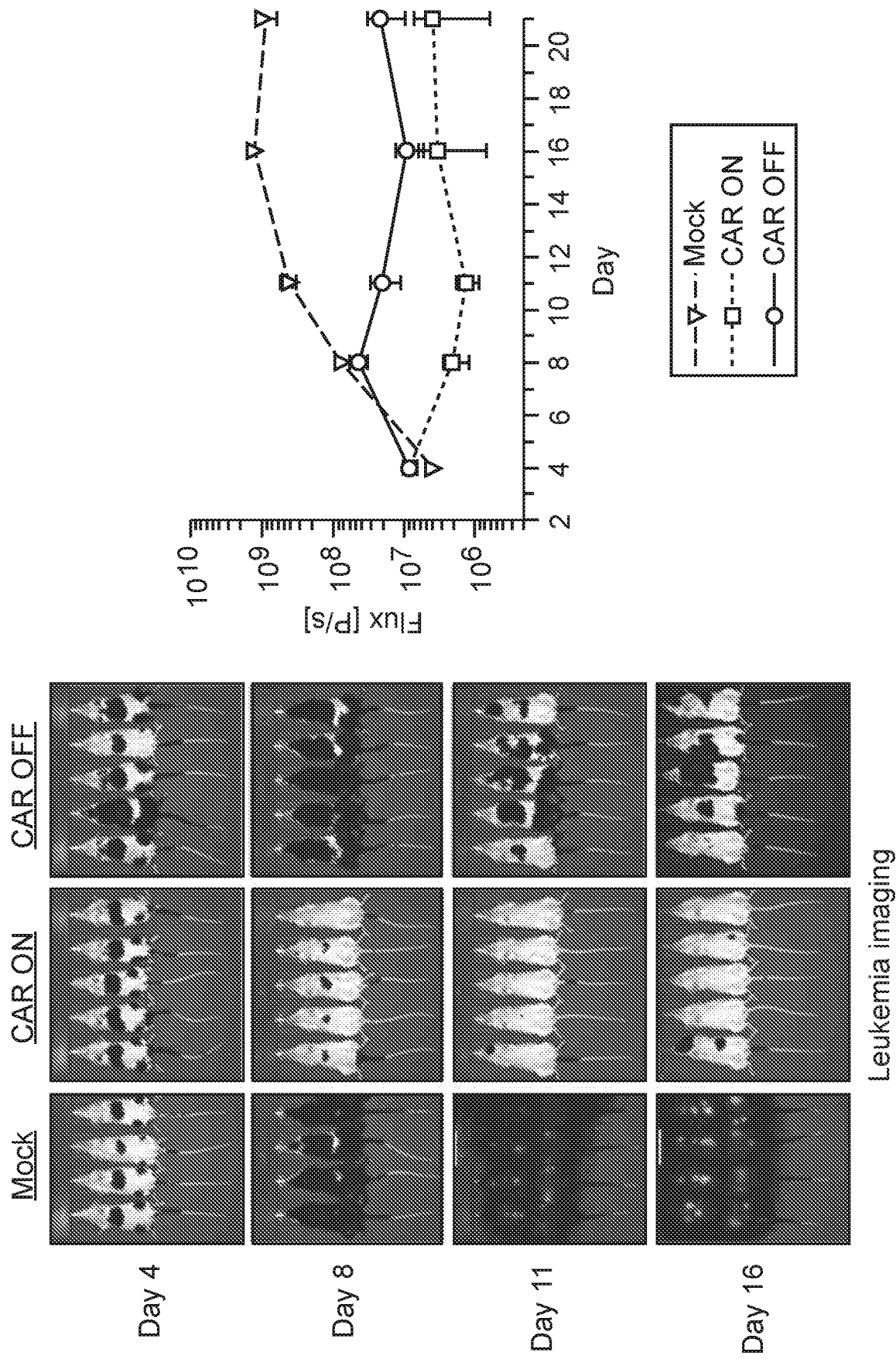


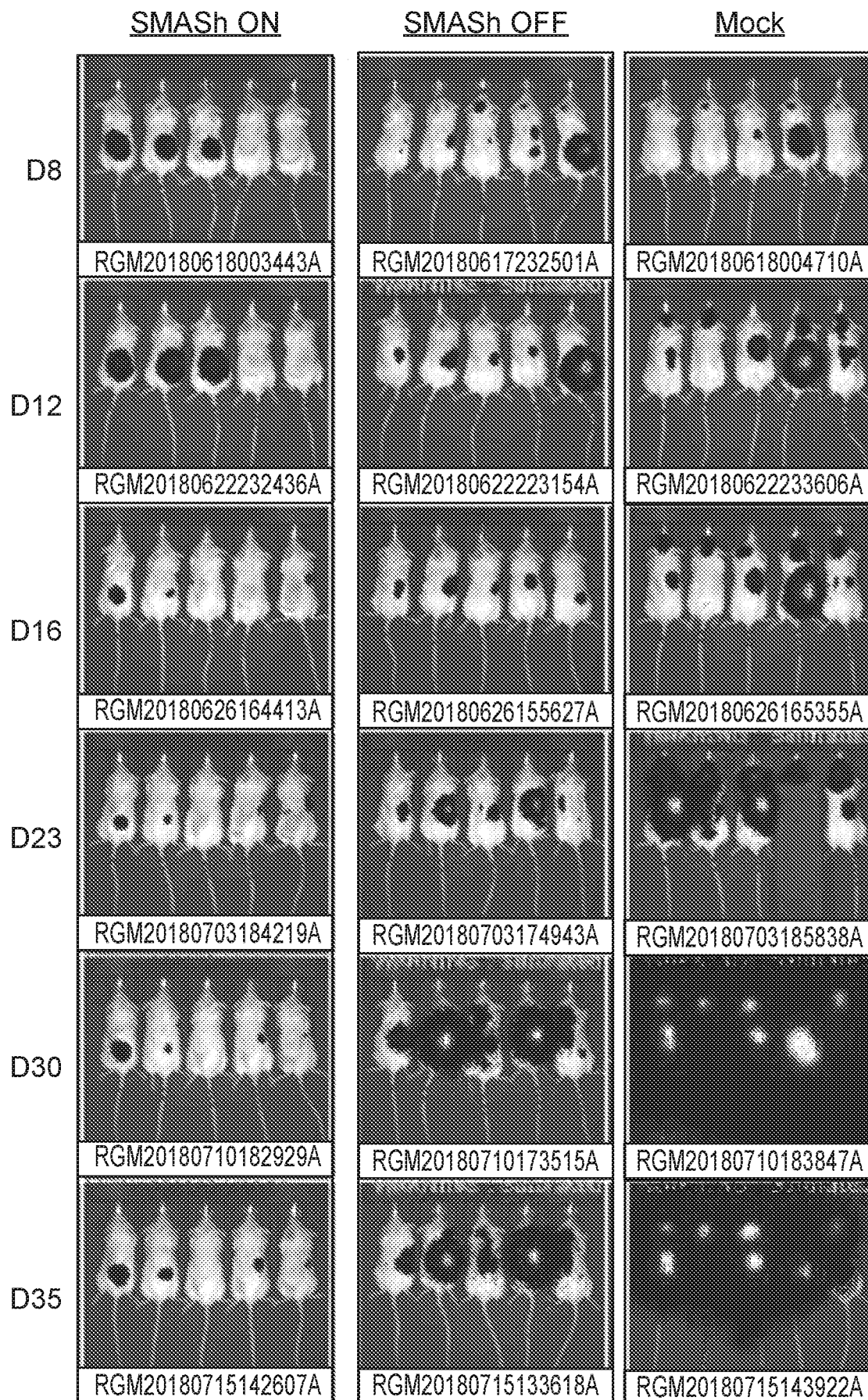
FIG. 25



30/33

FIG. 26



31/33
FIG. 27

32/33

FIG. 27 (Cont.)

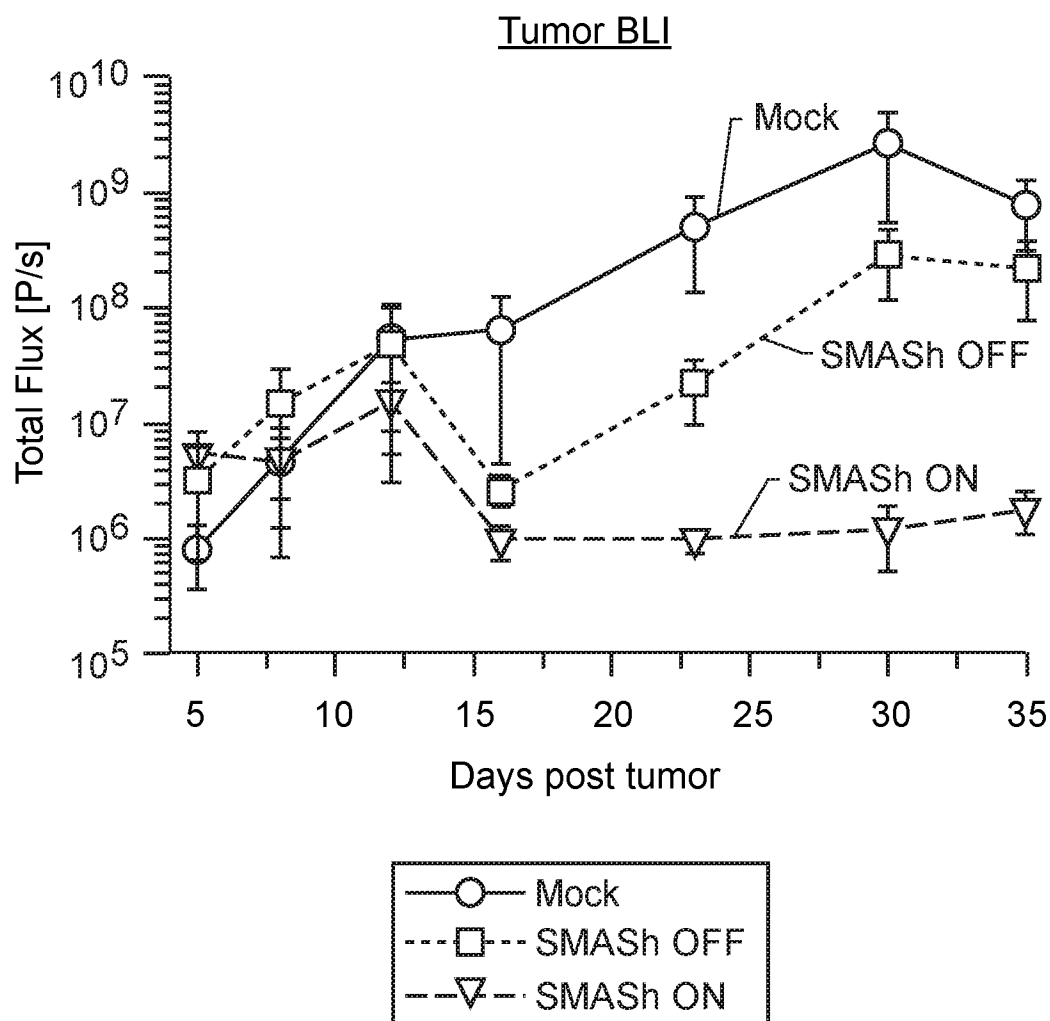
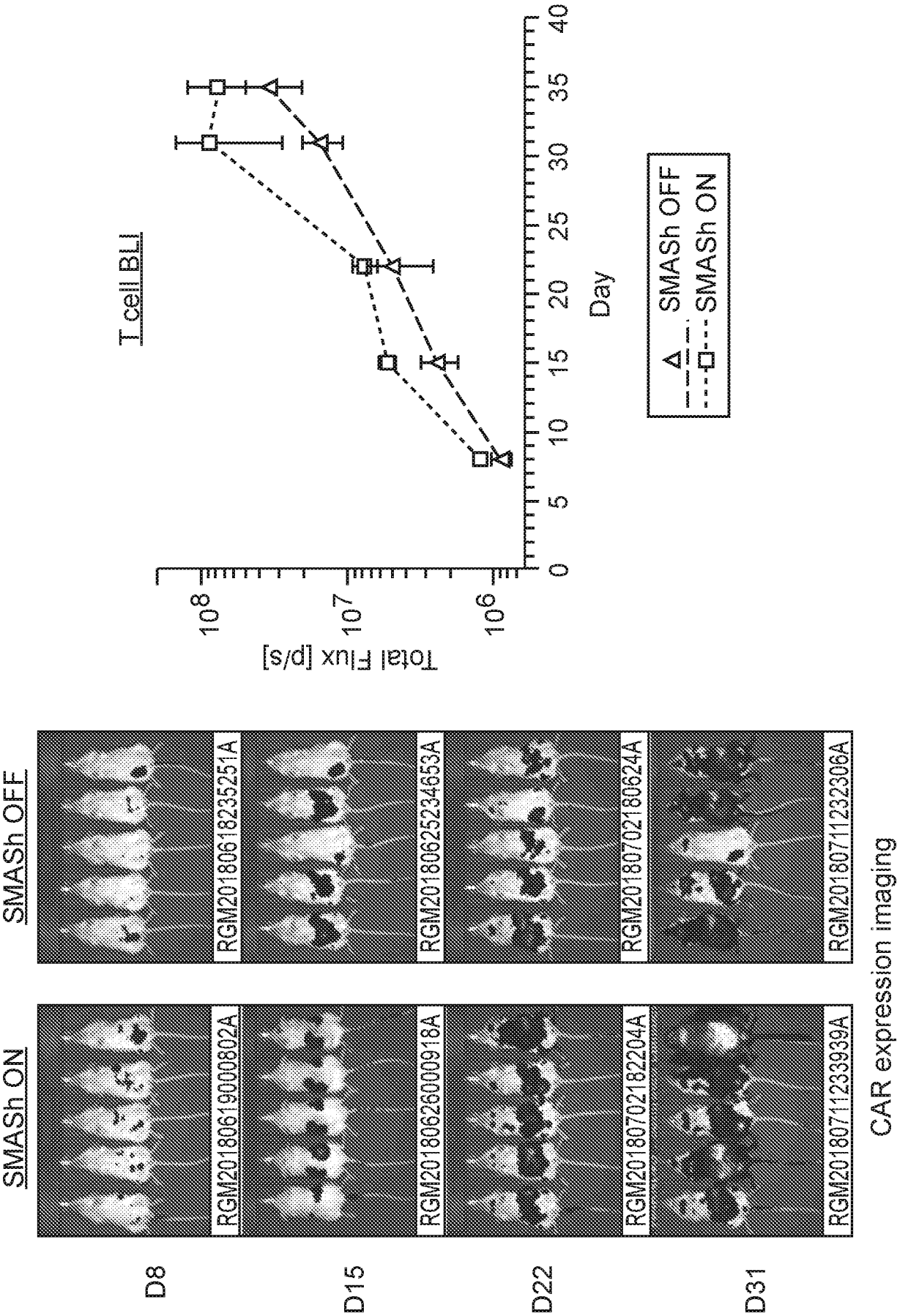


FIG. 28



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 19/40572

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - C07K 14/705; C07K 14/725; C07K 19/00; C12N 15/85 (2019.01)

CPC - C07K 2319/50; C07K 2319/95; C07K 14/7051; C07K 2317/622; C07K 2319/02; C07K 2319/03

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

See Search History Document

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

See Search History Document

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

See Search History Document

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X ---	WO 2018/121602A1 (TIANJIN TIANRUI BIOLOGICAL TECHNOLOGY CO., LTD.) 5 July 2018 (05.07.2018). [published in Chinese language with Google Chinese to English translation provided]. Especially PDF pg 6 para 6, PDF pg 7 para 2, PDF pg 13 para 4-5; sheet 5 fig 8.	1, 2, 4, 6/4 ----- 3, 5, 6/5
Y	WO 2017/004022 A2 (BOARD OF TRUSTEES OF THE LELAND STANFORD JUNIOR UNIVERSITY) 5 January 2017 (05.01.2017). Especially pg 2 ln 18-28, pg 57 ln 18-25, sheet 2 fig 1B.	3, 5, 6/5

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier application or patent but published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search 17 September 2019	Date of mailing of the international search report 16 OCT 2019
Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, Virginia 22313-1450 Facsimile No. 571-273-8300	Authorized officer: Lee W. Young PCT Helpdesk: 571-272-4300 PCT OSP: 571-272-7774

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 19/40572

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☒ Claims Nos.: 7-69
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.