

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
24 August 2006 (24.08.2006)

PCT

(10) International Publication Number
WO 2006/088664 A2

- (51) International Patent Classification: Not classified
- (21) International Application Number:
PCT/US2006/003843
- (22) International Filing Date: 1 February 2006 (01.02.2006)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
60/653,823 17 February 2005 (17.02.2005) US
- (63) Related by continuation (CON) or continuation-in-part (CIP) to earlier application:
US 60/653,823 (CIP)
Filed on 17 February 2005 (17.02.2005)
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- (81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, LY, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NG, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SM, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US (patent), UZ, VC, VN, YU, ZA, ZM, ZW.
- (84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IS, IT, LT, LU, LV, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).
- Published:**
— without international search report and to be republished upon receipt of that report
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.*

(54) Title: FLAVIVIRUS NS5A PROTEINS FOR THE TREATMENT OF HIV

(57) Abstract: GB virus C (GBV-C or hepatitis G virus) is a flavivirus that frequently leads to chronic viremia in humans. The invention provides compositions and methods involving a -GBV-C NS5A peptide or polypeptide for inhibiting and treating HIV infections.



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FLAVIVIRUS NS5A PROTEINS FOR THE TREATMENT OF HIV**BACKGROUND OF THE INVENTION**

5 The present application claims benefit of priority to U.S. Provisional Application Serial No. 60/653,823, filed February 17, 2005, the entire contents of which are hereby incorporated by reference.

The U.S. Government own rights in this invention pursuant to grant number AI58740 from NIH and merit grants awarded to Jack Stapleton and Jinhua Xiang from the Veterans Administration.

10

I. FIELD OF THE INVENTION

The present invention relates generally to the fields of molecular biology and virology. More particularly, it concerns methods and compositions to treat, inhibit or prevent HIV infection.

15 **II. DESCRIPTION OF RELATED ART****A. GB Virus Type C**

GB virus type C (GBV-C), also known as hepatitis G virus (HGV), is a virus whose genomic organization and nucleotide sequence places it in the *Flavivirus* family (Robertson *et al.*, 1998). It is the most closely related human virus to hepatitis 20 C virus (HCV) (Leary *et al.*, 1996; Linnen *et al.*, 1996; Simons *et al.*, 1995). It has been suggested that these viruses should be classified together with non-human GB-hepatitis agents as the hepacivirus genus. Although GBV-C was originally associated with post-transfusion hepatitis in humans (Linnen *et al.*, 1996), subsequent epidemiological studies indicated that it does not cause acute or chronic hepatitis 25 (Alter *et al.*, 1997a; Alter *et al.*, 1997b). In addition, experimental GBV-C infection of chimpanzees was not associated with acute hepatitis (Bukh *et al.*, 1998).

Persistent GBV-C viremia (as detected by RT-PCR) is common, with 0.9% to 3% of healthy U.S. blood donors and approximately 20%-30% of patients with HCV infection persistently infected with GBV-C (Dawson *et al.*, 1996; Feucht *et al.*, 1997; 30 Simons *et al.*, 1995a; Simons *et al.*, 1995b; Tacke *et al.*, 1997). Following infection,

about 80% of people clear their viremia, concomitantly developing antibody to the GBV-C E2 protein (Feucht *et al.*, 1997; Thomas *et al.*, 1998). Thus, it is estimated that approximately 20% of infected people remain viremic for long periods of time. GBV-C appears to be transmitted primarily by parenteral exposure (Simons *et al.*, 1995), although there are data suggesting that sexual and/or household transmission of GBV-C infection may occur (Akiyoshi *et al.*, 1999; de Martino *et al.*, 1998; Nerurkar *et al.*, 1998; Tanaka *et al.*, 1997; Wu *et al.*, 1997).

B. GBV-C and HIV

During progressive human immunodeficiency virus type 1 (HIV-1) infection, the virus-specific immune responses of an infected subject gradually deteriorate, leading to the development of acquired immunodeficiency syndrome (AIDS). Most infected patients do not exhibit overt clinical manifestations of the disease for six to ten years following initial infection, however, most individuals infected with HIV eventually die from conditions or infections; that the individual's immune system is no longer equipped to fight. While treatment for AIDS has been forthcoming, no effective cure has been reported. Thus, preventative and treatment options against HIV infection and the development of AIDS remain highly desirable.

GBV-C has been investigated in the context of HIV infection. The course of HIV-1 infection is extremely variable among infected individuals, although the reasons for this observation are not fully understood. Individuals whose HIV disease progresses slowly are often called long-term non-progressors (LTNPs). The prevalence of LTNPs varies from 1% to 25% of infected people, depending upon the definition used (reviewed in Easterbrook, 1999). There are no specific clinical criteria for LTNP. However, non-progression generally implies the absence of HIV-related clinical disease 10 or more years following infection and an absolute CD4 count of ≥ 500 cells/mm³ (Easterbrook, 1999). Evaluation of LTNP's has identified HIV isolates with deletions in key replicative genes (Deacon *et al.*, 1995) and host genetic factors, including specific HLA haplotypes (reviewed in reference Rowland-Jones, 1999). In some individuals, polymorphisms that result in absent or reduced expression of HIV co-receptors have been identified (Huang *et al.*, 1996). However, these findings are uncommon and thought to account for no more than one-third of LTNP's (Rowland-Jones, 1999).

Persistent GBV-C infection is common in humans, with infection rates of approximately 0.9% to 3% in healthy blood donors, 20-30% in HCV-positive people (Dawson *et al.*, 1996), and 35%-40% in HIV-positive individuals. GBV-C infection can persist for decades in the absence of any clinical morbidity or mortality. Among
5 immune-competent individuals, it is estimated that 60% to 75% of GBV-C-infected people clear the infection, concomitantly developing antibodies to the envelope glycoprotein E2 (Thomas *et al.*, 1998). It is also known that GBV-C can be propagated in cultures of peripheral blood mononuclear cells (PBMC's) (Fogeda *et al.*, 1999).

10 In 1998, Toyoda *et al.* found that hemophiliacs co-infected with HIV and GBV-C had a lower plasma HIV RNA concentration and a lower incidence of AIDS diagnoses compared to those infected with HIV alone (Toyoda *et al.*, 1998), although the differences were not statistically significant. In contrast, Sabin and colleagues found an increased rate of AIDS and death in hemophiliacs "exposed" to GBV-C
15 (Sabin *et al.*, 1998) compared to non-exposed individuals. This study included HIV-positive subjects who were either GBV-C viremic as determined by detection of GBV-C RNA in plasma, or HIV-infected people who were not viremic but were anti-GBV-C E2 antibody-positive. Although the mortality rate was higher among the GBV-C "exposed" individuals, the results were not statistically significant. Looking
20 at HIV-infected persons, Lefrère and colleagues reported a significant delay in the rate of CD4+ T cell decline, development of AIDS, and death in 23 HIV-positive individuals with GBV-C viremia compared to 72 HIV-infected people without GBV-C viremia (Lefrère *et al.*, 1999). In that study, HIV-infected individuals who were also GBV-C-positive were compared to HIV-infected individuals who were GBV-C-
25 negative. When these subjects were matched by age, sex, baseline HIV RNA load, and baseline CD4 T cell count, HIV disease progression appeared to be worse in GBV-C-negative subjects.

The interrelationship between HIV and GBV-C continues to be explored, with possible therapeutic aspects of GBV-C infection being examined.

SUMMARY OF THE INVENTION

A pharmaceutical composition comprising an isolated flavivirus NS5A peptide or polypeptide, or multiple flavivirus NS5A peptides from the same or different NS5A polypeptide. The NS5A polypeptide may be a full length NS5A polypeptide, a fusion polypeptide. The fusion may comprise a targeting signal, such as a nuclear targeting signal or a cell surface receptor (*e.g.*, a CD4 receptor). The NS5A peptide or polypeptide may be formulated in a lipid vehicle, such a liposome. The NS5A peptide or polypeptide may be formulated with an amphipathic peptide, an insect peptide, or pyrrolic acid. The flavivirus may be selected from the group consisting of DEN4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, GBV-C, HGV, HCV2a, HCV3a, HCV2b, HCV1a and HCV1b. The flavivirus NS5A peptide or polypeptide may residues 152-237 of GBV-C NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins, or may comprise domain II of HCV NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins. The NS5A peptide or polypeptide may be from a IFN-sensitive flavivirus, or a IFN-resistant flavivirus. The IFN-sensitive flavivirus or IFN-resistant flavivirus may be a GBV-C virus.

In yet another embodiment, there is provided a method for preventing or treating HIV infection comprising administering to a subject a composition comprising a flavivirus NS5A peptide or polypeptide. The flavivirus NS5A peptide or polypeptide comprises multiple flavivirus NS5A peptides, from the same or different NS5A polypeptides. The NS5A polypeptide may be a full length NS5A polypeptide, and may be a fusion polypeptide, for example, comprising a targeting signal, such as a nuclear targeting signal or a targeting signal that targets a cell surface receptor (*e.g.*, the CD4 receptor). The NS5A peptide or polypeptide may be formulated in a lipid vehicle, such a liposome. The NS5A peptide or polypeptide may be formulated with an amphipathic peptide, an insect peptide, or pyrrolic acid. The flavivirus may be selected from the group consisting of DEN4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, GBV-C, HGV, HCV2a, HCV3a, HCV2b, HCV1a and HCV1b. The flavivirus NS5A peptide or polypeptide may residues 152-237 of GBV-C NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins, or may comprise domain II of HCV NS5A, or the corresponding sequences thereto from

other flavivirus NS5A proteins. The NS5A peptide or polypeptide may be from a IFN-sensitive flavivirus, or a IFN-resistant flavivirus. The IFN-sensitive flavivirus or IFN-resistant flavivirus may be a GBV-C virus. The method may further comprise administration of at least a second anti-HIV therapy, before or after said flavivirus
5 NS5A peptide or polypeptide. The second anti-HIV therapy may be HAART therapy, or AZT therapy. The method may comprise multiple administrations of the composition.

In yet another embodiment, there is provided a method of reducing HIV replication in an HIV-infected cell comprising contacting said cell with a composition
10 comprising a flavivirus NS5A peptide or polypeptide. The cell may be a T lymphocyte. In still yet another embodiment, there is provided a method of inhibiting HIV infection of a cell comprising contacting said cell with a composition comprising a flavivirus NS5A peptide or polypeptide. The cell may be a T lymphocyte.

In still a further embodiment, there is provided a method for preventing or
15 treating HIV infection comprising administering to a subject a composition comprising an expression construct encoding a flavivirus NS5A peptide or polypeptide. The expression construct may be a viral expression construct, such as an adenovirus, a retrovirus, a lentivirus, an adeno-associated virus, a polyoma virus, a herpesvirus, or a pox virus. The expression construct may be a non-viral expression
20 construct, for example, dispersed in a lipid vehicle. The expression construct may encode an NS5A polypeptide or an NS5A peptide, a a full length NS5A polypeptide, a fusion polypeptide, for example, comprising a nuclear targeting signal or a signal that targets a cell surface receptor, e.g., the CD4 receptor. The flavivirus NS5A peptide or polypeptide may comprise residues 152-237 of GBV-C NS5A, or the
25 corresponding sequences thereto from other flavivirus NS5A proteins, or domain II of HCV NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins. The flavivirus may be selected from the group consisting of DEN4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, GBV-C, HGV, HCV2a, HCV3a, HCV2b, HCV1a and HCV1b.

30 The use of the word "a" or "an" when used in conjunction with the term "comprising" in the claims and/or the specification may mean "one," but it is also consistent with the meaning of "one or more," "at least one," and "one or more than

one.” Furthermore, where multiple steps of a method of process are cited, it is understood that the steps are not required to be performed in the particular order recited unless one of skill in the art is not be able to practice the method in a different order.

5 Other objects, features, and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become
10 apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

The following drawings form part of the present specification and are included to further demonstrate certain aspects of the present invention. The invention may be
15 better understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein.

FIG. 1 - NS5A expression in Jurkat cells. Jurkat cells containing GBV-C NS5A (A11a clone) expressed two immunoreactive proteins when incubated without doxycycline (Doxy). Rabbit anti-NS5A antisera was raised against NS5A expressed
20 in E.Coli. This antigen is shown as the positive control on lower panel (+), whereas the positive control [C(+)] top panel represents NS5A expressed in CHO cells; JICR In Press). Growth of cells in low levels of doxycycline for 48 hrs reduced NS5A protein levels to non-detectable levels. Doxy concentration in mg/ml.

FIGS. 2A-C - HIV replication in Jurkat cells with and without NS5A
25 **expression.** The amount of HIV p24 antigen released into culture supernatants from two cloned Jurkat cell lines expressing GBV-C NS5A (A11a and A11b) or vector control cell line (VC) 2 to 6 days following infection using an X-4 virus isolate (MOI of 0.1 (FIG. 2A) or 0.5 (FIG. 2B)). Cells were grown without doxycycline, and data represent the mean of 3 replicate samples. NS5A expression is shown (immunoblot of
30 cell lysates in triplicate) of A11a, A11b. VC = vector control cell lysate. Note that A11a has greater levels of NS5A expressed and HIV replication in these cells is lower

than in A11b (on day 6). Compared to vector control cells, HIV inhibition was significantly lower on day 6 ($p=0.02$).

FIG. 3A - Experimental Design. Jurkat cells with either vector control (VC) or NS5A were grown without doxycycline. Cells were divided, and half were maintained without doxycycline (NS5A was expressed) or with various doses of doxycycline (NS5A expression was differentially suppressed). Cells (VC and NS5A containing) were infected with X4 HIV-1 (clinical isolate) 48 hrs after dividing cells, and cell culture supernatants monitored 2, 4, and 6 days later for HIV p24 antigen.

FIG. 3B - Dose-dependent inhibition of HIV by NS5A. Experimental design described above. NS5A expression was suppressed by cell growth in various concentrations of doxycycline (0.01, 0.1 and 1 mg/ml as indicated) for 2 days prior to infection with HIV-1. HIV replication was monitored by measuring p24 antigen in culture supernatants. The amount of NS5A expression was directly correlated with inhibition of HIV expression in Jurkat-NS5A cells (left panel), and doxycycline had no effect in Jurkat-vector control cells (VC; right panel).

FIG. 4A - Experimental Design. Jurkat cells with NS5A were passaged with doxycycline to suppress NS5A expression. Cells were divided, and half were maintained without doxycycline (NS5A was expressed) or with 1 mg/ml doxycycline (NS5A was not expressed). NS5A containing cells were infected with X4 HIV-1 (clinical isolate) 96 hrs after cells were divided, and cell culture supernatants monitored 2, 4, and 6 days later for HIV p24 antigen.

FIG. 4B - GBV-C NS5A inhibits HIV replication. Data represent HIV replication (p24 antigen in supernatants). HIV replication was not different on days 1 and 2; however, by day 3, Jurkat-NS5A cells had 45% reduction in HIV p24 Ag 3 days post-infection (T-test, $P=0.03$), and by 56% - 60% on days 4, 5, and 6 ($p<0.001$ for each day). Infections were performed in triplicate, NS5A and vector control error bars shown. Results were similar for Jurkat cells without any plasmid DNA as well.

FIG. 5 - Release of SDF-1 into culture supernatants by Jurkat cells with or without GBV-C NS5A. SDF-1 in supernatants was measured by ELISA in triplicate at time points shown. There were no differences between Jurkat and vector

control Jurkat cells; however, SDF-1 increased in NS5A expressing cells on days 5 and 6 (T test, $P < 0.001$ for both days).

FIG. 6 - Jurkat cells expressing GBV-C NS5A have lower surface density of CXCR4 compared to Jurkat cells with a vector control. Jurkat cells that express NS5A or the vector control were grown without doxycycline, and analyzed by flow cytometry for surface expression of CXCR4 (shown above). Reproducibly, cells with NS5A demonstrated a 49.6% reduction in CXCR4 mean fluorescence ($p = 0.003$). No difference in CCR5 or CD4 was observed (data not shown). Preliminary microarray data demonstrated decreased levels of CXCR4 25 mRNA in NS5A expressing cells.

FIG. 7 - Release of caspase 3/7 into culture supernants following incubation with anti-Fas antibody (CH11). Jurkat cells with NS5A were relatively resistant to induction of apoptosis by CH11 when compared to Jurkat cells with the vector control. Similarly, spontaneous apoptosis was greater in Jurkat NS5A cells compared to vector control. Preliminary microarray data demonstrated NS5A-related increases in mRNA levels for TGFB1-induced anti-apoptotic factor 1 and chemokine ligand 25 mRNA. Both of these genes enhance resistance to apoptosis, and CC ligand 25 does so in CD4+ T cells.

FIG. 8 - Dose-Dependent HIV Inhibition by the NS5A Protein. Left panel – NS5A-expressing Jurkat cells; right panel – vector control Jurkat cells. HIV replication is measured by p24 antigen levels on days 2-6 (D2-D6). Amounts of doxycycline, which inversely relate to NS5A production, are shown in $\mu\text{g/ml}$.

FIG. 9 - Both IFN-R and IFN-S GBV-C NS5A Inhibit HIV Replication. HIV replication is measured by p24 antigen levels on days 2-6 (D2-D6). VC = vector control; GFP – vector expression GFP only; IFN-S = interferon sensitive NS5A; IFN-R = interferon resistant NS5A.

FIG. 10 - Anti-SDF-1 Blocks NSF5A-Mediated Inhibition of HIV Replication. Results reported as percent increase in HIV replication when cultures are incubated with anti-SDF-1 neutralizing antibody as compared to isotype control antibody. IFN-R = interferon resistant NS5A; IFN-R-GFP = interferon resistant NS5A linked to GFP; IFN-S-GFP = interferon sensitive NS5A linked to GFP.

FIGS. 11A-C – NS5A Inhibits Multiple Distinct HIV Strains. HIV replication is measured by p24 antigen levels on days 2-6 (D2-D6). VC-GFP = vector control expression GFP; IFN-R = interferon resistant NS5A; IFN-R-GFP = interferon resistant NS5A linked to GFP; IFN-S-GFP = interferon sensitive NS5A linked to GFP.

FIG. 12 – Fragments of NS5A That Have Been Stably Expressed in Jurkat Cells. Shaded bars indicate fusions or fragments.

FIG. 13 – Fragments of NS5A That Inhibit HIV Replication. Shaded bars inhibit HIV; open bars do not inhibit HIV.

FIG. 14 – Doxycycline-Repressed Expression of NS5A. Lanes are labeled with amounts of doxycycline ($\mu\text{g/ml}$); arrows indicate NS5A or mutants (μNS5A).

FIG. 15 – Percent Inhibition of HIV Replication by NS5A Fragments. Various fragments identified by residues of NS5A remaining, and their ability to inhibit HIV replication (by p24 antigen levels) on day 6 after infection.

FIG. 16 – Fragments of NS5A That Inhibit HIV Replication. Shaded bars inhibit HIV; open bars do not inhibit HIV; lined bars not yet tested.

FIG. 17 – R 152-237 NS5A inhibits HIV when expressed in Jurkat cells. By deduction, it appears that 152-182 is the minimal sequence required for HIV inhibition. Note that the latter sequence is conserved (IFN-R and IFN-S) GBV-C NS5A, and that there are multiple potential phosphorylation sites.

DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

Anti-retroviral medications suppress viral replication in HIV disease, yet they have failed to eradicate the virus from the body due to the multi-faceted nature of HIV infection, as well as the complexities of the immune system. Methods are being developed that both prevent infection and boost the immune system to keep it functioning at a level where it can assist in fighting HIV infection.

The present inventors have previously reported on methods and compositions for therapeutic and/or prophylactic treatment of HIV infection, including GBV-C envelope proteins, in particular GBV-C envelope protein E2 (E2). More specifically, the inventors have shown that HIV-infected subjects that are co-infected with GB virus C (GBV-C) typically have reduced mortality and slower progression to AIDS as compared to HIV-infected subjects without GBV-C co-infection (PCT/US2004/017706). Infection of peripheral blood mononuclear cells (PBMCs) with GBV-C and HIV results in inhibition of HIV-1 replication. GBV-C infection typically inhibits HIV by inducing β -chemokines and reducing expression of the HIV co-receptor CCR5, explaining part of the beneficial clinical findings of GBV-C on HIV disease progression. The inventors also described a therapeutic use for antibodies and/or binding agents that bind GBV-C proteins (*e.g.*, envelope proteins), in particular, the E2 protein, and similar antigens used for producing these antibodies or binding agents (PCT/US03/33925).

The inventors now demonstrate a unique role for the NS5A protein of GBV-C, as well as NS5A's from other flaviviruses, in the inhibition of HIV replication. Various aspects of the invention are described below.

III. FLAVIVIRUSES

A. Family

With a total of 69 pathogens in its ranks, *Flaviviridae* contains a myriad of viruses that cause disease in humans. Foremost among these is Yellow Fever Virus, the type virus of the *Flaviviridae*, from which the family begets its name (*flavus* in Latin means "yellow"). Flaviviruses have been subdivided by the ICTV into three genera: *Flavivirus*, *Pestivirus* and *Hepacivirus*.

The *Flavivirus* genus contains several dangerous viruses including yellow fever virus, dengue fever virus, and Japanese encephalitis (JE) virus. The *Pestivirus* genus is home to the three serotypes of bovine viral diarrhea, but no known human pathogens. The genus *Hepacivirus* consists of hepatitis C virus and its relatives.

5 *Flavivirus* genomes consist of a monopartite (*i.e.*, one piece of) linear, single-stranded, positive sense RNA. Because the RNA is positive sense, the nucleic acid itself is capable of instigating an infection in the appropriate host cells. The total genome can range from 10 to 11 kilobase pairs. The genome 3' terminus is not polyadenylated. The 5' end has a methylated nucleotide cap (allows for translation) or
10 a genome-linked protein (VPg). *Pestivirus* genomes are reported to be 12.5 kb in length. Like the *Flavivirus* genus, no poly-A tail exists on the 3' end of the RNA, however, *Pestivirus* genus members lack a 5' cap. In both genera, structural genes are found towards the 5' end of the RNA. Both the *Pestivirus* and *Hepacivirus* genera contain internal ribosomal entry sites (IRES) that provide a site of translation
15 initiation for host ribosomes. This is in contrast to the *Flavivirus* genus that uses the technique of ribosomal scanning to commence protein synthesis.

Under the EM, virions appear roughly as spheres (some experts say they're "pleomorphic"), 40-65 nm in diameter. What can be seen under the microscope is the virus's lipid envelope, which it obtains from host cells during egress (leaving the cell).
20 Underneath the envelope can be found an icosahedral capsid coat approximately 25-30 nm in diameter.

All members of the *Flavivirus* genus are transmitted by arthropods (*i.e.*, mosquitoes and ticks) while Hepatitis C is spread parenterally (*i.e.*, through contaminated bodily fluids). A key feature for viral transmission in Flaviviruses is
25 that they are capable of reproducing in their vector. Without the ability to replicate in the vector, they would not remain viable to be passed from one host to the next.

B. GBV-C

Like other members of the *Flaviviridae*, GBV-C is a positive-strand RNA virus that encodes a single long open reading frame (Leary *et al.*, 1996). GBV-C does
30 not cause acute or chronic hepatitis, yet it is the family member most closely related to HCV, the cause of hepatitis C. Sequences of GBV-C have been previously

reported, for example in U.S. Patent 5,874,563, which is specifically incorporated by reference. In particular, an infectious GBV-C clone has been described in the PCT application WO 01/77157, which is incorporated herein by reference.

The GBV-C polyprotein is predicted to be cleaved into two envelope proteins
5 (E1 and E2, referred to collectively as GBV-C envelope protein), an RNA helicase, a trypsin-like serine protease, and an RNA-dependent RNA polymerase. A major difference between GBV-C and HCV is in the amino terminus of the polyprotein. In many isolates, this region is truncated, and no core (or nucleocapsid) protein is present (Simons *et al.*, 1995; Xiang *et al.*, 1999). *In vitro* translation experiments
10 suggest that the AUG immediately upstream of the putative E1 protein is preferentially used to initiate translation, although there may be as many as four AUG's in frame with the polyprotein upstream of this AUG (Simons *et al.*, 1996).

The site of GBV-C replication has not been clearly identified, but it appears that replication in the hepatocyte, if it occurs, is not the primary source of virus in
15 infected individuals (Laskus *et al.*, 1998; Pessoa *et al.*, 1998; Seipp *et al.*, 1999). Recently, there were reports that human peripheral blood mononuclear cells (PBMC's) and interferon-resistant Daudi cells are permissive for GBV-C replication (Fogeda *et al.*, 1999; Shimizu, 1999). In addition, transient replication of GBV-C was described in MT-2 cells (a human T-cell line), and PH5CH (a human hepatocyte line
20 immortalized with simian virus 40 large T antigen) (Seipp *et al.*, 1999).

C. Other Flavivirus NS5A's

Other Flaviviruses contain NS5A's that can be used in accordance with the present invention. These viruses include DEN1-4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, HGV, HCV2a, HCV3a, HCV2b, HCV1a, HCV1c and
25 HCV1b.

IV. GBV-C POLYPEPTIDES

In certain aspects, the invention is directed to the the NS5A polypeptide of GBV-C virus, or a peptide or polypeptide derived there from. SEQ ID NO:2 shows
30 the NS5A translated product of SEQ ID NO:1 (cDNA). It is contemplated that the compositions and methods disclosed herein may be utilized to express all or part of

SEQ ID NO:2 and derivatives thereof. In certain embodiments, compositions of the invention may include the nucleic acids encoding the peptides as set forth in SEQ ID NO:1 or 3. Determination of which protein or DNA molecules inhibit HIV may be achieved using functional assays measuring HIV replication and infectivity, which are familiar to those of skill in the art. The structure of the various polypeptides or peptides can be modeled or resolved by computer modeling, NMR, or x-ray crystallography. Such structures may be used to engineer derivatives of the various NS5A protein.

Exemplary accession nos. (incorporated by reference) for other NS5A's are as follows:

<u>Virus</u>	<u>Accession No.</u>	<u>Virus</u>	<u>Accession No.</u>
West Nile	DQ318019	Dengue 1-4	AY66269
Yellow fever	NC002031	HCV 1a	AF011753
	AY603338	1b	AF333324
BVDV	AF502399	1c	D14853
Dengue 1-4	M878512	2a	D00944
	M14931	2b	D10988
	M20558	3a	AF046866

A. Variants of GBV-C NS5A Polypeptides

Embodiments of the invention include various GBV-C NS5A polypeptides, peptides, and derivatives thereof. Amino acid sequence variants of a polypeptide can be substitutional, insertional or deletion variants. Deletion variants lack one or more residues of the native protein that are not essential for function or immunogenic activity. Insertional mutants typically involve the addition of material at a non-terminal point in the polypeptide. This may include the insertion of an immunoreactive epitope or simply a single residue. Terminal additions, called fusion proteins, are discussed below.

Substitutional variants typically contain the exchange of one amino acid for another at one or more sites within the protein, and may be designed to modulate one

or more properties of the polypeptide, such as stability against proteolytic cleavage, without the loss of other functions or properties. Substitutions of this kind preferably are conservative, that is, one amino acid is replaced with one of similar shape and charge. Conservative substitutions are well known in the art and include, for example, the changes of: alanine to serine; arginine to lysine; asparagine to glutamine or histidine; aspartate to glutamate; cysteine to serine; glutamine to asparagine; glutamate to aspartate; glycine to proline; histidine to asparagine or glutamine; isoleucine to leucine or valine; leucine to valine or isoleucine; lysine to arginine; methionine to leucine or isoleucine; phenylalanine to tyrosine, leucine or methionine; serine to threonine; threonine to serine; tryptophan to tyrosine; tyrosine to tryptophan or phenylalanine; and valine to isoleucine or leucine.

The term "biologically functional equivalent" is well understood in the art and is further defined in detail herein. Accordingly, sequences that have between about 70% and about 80%; or more preferably, between about 81% and about 90%; or even more preferably, between about 91% and about 99%; of amino acids that are identical or functionally equivalent to the amino acids of GBV-C NS5A polypeptides, for example SEQ ID NO:2, provided the biological activity of the protein or peptide is maintained.

The term "functionally equivalent codon" is used herein to refer to codons that encode the same amino acid, such as the six codons for arginine or serine, and also refers to codons that encode biologically equivalent amino acids (see Table 1, below).

Certain embodiments of the invention include various peptides or polypeptides of the NS5A protein. For example, all or part of a GBV-C NS5A protein as set forth in SEQ ID NO:2 may be used in various embodiments of the invention. In certain embodiments, a fragment of the NS5A protein may comprise, but is not limited to about 5, about 6, about 7, about 8, about 9, about 10, about 11, about 12, about 13, about 14, about 15, about 16, about 17, about 18, about 19, about 20, about 21, about 22, about 23, about 24, about 25, about 26, about 27, about 28, about 29, about 30, about 31, about 32, about 33, about 34, about 35, about 36, about 37, about 38, about 39, about 40, about 41, about 42, about 43, about 44, about 45, about 46, about 47, about 48, about 49, about 50, about 51, about 52, about 53, about 54, about 55, about 56, about 57, about 58, about 59, about 60, about 61, about 62, about 63, about 64,

about 65, about 66, about 67, about 68, about 69, about 70, about 71, about 72, about 73, about 74, about 75, about 76, about 77, about 78, about 79, about 80, about 81, about 82, about 83, about 84, about 85, about 86, about 87, about 88, about 89, about 90, about 91, about 92, about 93, about 94, about 95, about 96, about 97, about 98, about 99, about 100, about 110, about 120, about 130, about 140, about 150, about 160, about 170, about 180, about 190, about 200, about 210, about 220, about 230, about 240, about 250, about 275, about 300, about 325, about 350, about 375, about 400, about 415, and any range derivable therein.

It also will be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids or 5' or 3' sequences, and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological activity (*e.g.*, immunogenicity) where protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region.

The following is a discussion based upon changing of the amino acids of an NS5A polypeptide or peptide to create an equivalent, or even an improved, second-generation molecule. For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid substitutions can be made in a protein sequence, and in its underlying DNA or RNA coding sequence, and nevertheless produce a protein with like properties. It is thus contemplated by the inventors that various changes may be made in the DNA or RNA sequences of genes or coding regions without appreciable loss of their biological utility or activity, as discussed herein. Table 1 shows the codons that encode particular amino acids.

TABLE 1 - CODON TABLE

Amino Acids			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like.

It also is understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U.S. Patent 4,554,101, incorporated herein by reference, states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

It is understood that an amino acid substituted for another having a similar hydrophilicity value still produces a biologically equivalent and immunologically equivalent protein.

In certain embodiments, an NS5A polypeptide may be a fusion protein. Fusion proteins may alter the characteristics of a given polypeptide, such as antigenicity or purification characteristics. A fusion protein is a specialized type of insertional variant. This molecule generally has all or a substantial portion of the native molecule, linked at the N- or C-terminus, to all or a portion of a second polypeptide. For example, fusions typically employ leader sequences from other species to permit the recombinant expression of a protein in a heterologous host. Another useful fusion includes the addition of an immunologically active domain, such as an antibody epitope, to facilitate purification of the fusion protein. Inclusion of a cleavage site at or near the fusion junction will facilitate removal of the extraneous polypeptide after purification. Other useful fusions include linking of functional domains, such as active sites from enzymes such as a hydrolase, glycosylation domains, cellular targeting signals, or transmembrane regions.

B. *In vitro* Production of NS5A Polypeptides or Peptides

Various types of expression vectors are known in the art that can be used for the production of protein products. Following transfection with a expression vector, a cell in culture, *e.g.*, a primary mammalian cell, a recombinant product may be prepared in various ways. A host cell strain may be chosen that modulates the expression of the inserted sequences, or that modifies and processes the gene product in the manner desired. Such modifications (*e.g.*, glycosylation) and processing (*e.g.*, cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cell lines or host systems can be chosen to insure the correct modification and processing of the foreign protein expressed. In order for the cells to be kept viable while *in vitro* and in contact with the expression construct, it is necessary to ensure that the cells maintain contact with the correct ratio of oxygen and carbon dioxide and nutrients but are protected from microbial contamination. Cell culture techniques are well documented (for exemplary methods see Freshney, 1992).

Animal cells can be propagated *in vitro* in two modes: as non-anchorage-dependent cells growing in suspension throughout the bulk of the culture or as

anchorage-dependent cells requiring attachment to a solid substrate for their propagation (*i.e.*, a monolayer type of cell growth).

Non-anchorage dependent or suspension cultures from continuous established cell lines are the most widely used means of large-scale production of cells and cell products. However, suspension cultured cells have limitations, such as tumorigenic potential and lower protein production than adherent cells.

In further aspects of the invention, other protein production methods known in the art may be used, including but not limited to prokaryotic, yeast, and other eukaryotic hosts such as insect cells and the like.

Because of their relatively small size, the peptides of the invention can also be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, (1984); Tam *et al.*, (1983); Merrifield, (1986); and Barany and Merrifield (1979), each incorporated herein by reference. Short peptide sequences, or libraries of overlapping peptides, usually from about 6 up to about 35 to 50 amino acids, which correspond to the selected regions described herein, can be readily synthesized and then screened in screening assays designed to identify reactive peptides.

C. Protein Purification

It may be desirable to purify NS5A polypeptides and peptides, or variants and derivatives thereof. Protein purification techniques are well known to those of skill in the art. These techniques involve, at one level, the crude fractionation of the cellular milieu to polypeptide and non-polypeptide fractions. Having separated the polypeptide from other proteins, the polypeptide of interest may be further purified using chromatographic and electrophoretic techniques to achieve partial or complete purification (or purification to homogeneity). Analytical methods particularly suited to the preparation of a pure peptide are ion-exchange chromatography, hydrophobic interaction chromatography, exclusion chromatography; polyacrylamide gel electrophoresis; isoelectric focusing. A particularly efficient method of purifying peptides is fast protein liquid chromatography (FPLC).

Certain aspects of the present invention concern the purification, and in particular embodiments, the substantial purification, of an encoded protein or peptide.

The term "purified protein or peptide" as used herein, is intended to refer to a composition, isolatable from other components, wherein the protein or peptide is purified to any degree relative to its naturally obtainable state. A purified protein or peptide therefore also refers to a protein or peptide, free from the environment in which it may naturally occur.

Generally, "purified" will refer to a protein or peptide composition that has been subjected to fractionation to remove various other components, and which composition substantially retains its expressed biological activity. Where the term "substantially purified" is used, this designation will refer to a composition in which the protein or peptide forms the major component of the composition, such as constituting about 50%, about 60%, about 70%, about 80%, about 90%, about 95% or more of the proteins in the composition.

Various methods for quantifying the degree of purification of the protein or peptide will be known to those of skill in the art in light of the present disclosure. These include, for example, determining the specific activity of an active fraction, or assessing the amount of polypeptides within a fraction by SDS/PAGE analysis. A preferred method for assessing the purity of a fraction is to calculate the specific activity of the fraction, to compare it to the specific activity of the initial extract, and to thus calculate the degree of purity, herein assessed by a "-fold purification number." The actual units used to represent the amount of activity will, of course, be dependent upon the particular assay technique chosen to follow the purification and whether or not the expressed protein or peptide exhibits a detectable activity.

There is no general requirement that the protein or peptide always be provided in their most purified state. Indeed, it is contemplated that less substantially purified products will have utility in certain embodiments. Partial purification may be accomplished by using fewer purification steps in combination, or by utilizing different forms of the same general purification scheme.

V. FLAVIVIRUS POLYNUCLEOTIDES

Certain embodiments of the invention include Flavivirus NS5A polynucleotides or nucleic acid molecules and fragments thereof. The

polynucleotides of the invention may be isolated and purified from Flavivirus or cells infected or transfected with Flavivirus polynucleotides. The term isolated indicating they are free or substantially free from total viral or cellular genomic RNA or DNA, and proteins. It is contemplated that an isolated and purified Flavivirus nucleic acid molecule may take the form of RNA or DNA. A Flavivirus nucleic acid molecule refers to an RNA or DNA molecule that is capable of yielding all or part of a Flavivirus NS5A from a transfected cell.

When the present application refers to the function or activity of an infectious Flavivirus that is encoded by a Flavivirus polynucleotide, it is meant that the polynucleotide encodes a molecule that has the ability to propagate an infectious Flavivirus virus particle from a cell. It is contemplated that a Flavivirus polynucleotide may refer to a Flavivirus RNA transcript that is able to propagate an infectious Flavivirus virus particle after introduction to a cell or to a Flavivirus expression construct, clone, or vector composed of double-stranded DNA or DNA/RNA hybrid that is similarly capable, or a doublestranded DNA that is similarly capable following *in vitro* transcription.

The term "cDNA" is intended to refer to DNA prepared using RNA as a template. The advantage of using a cDNA, as opposed to genomic RNA or an RNA transcript is stability and the ability to manipulate the sequence using recombinant DNA technology (*See* Maniatis, 1990; Ausubel, 1996). There may be times when the full or partial genomic sequence is preferred.

It also is contemplated that a given Flavivirus may be represented by natural variants or strains that have slightly different nucleic acid sequences but, nonetheless, encode the same viral polypeptides (see Table 1 above). Consequently, the present invention also encompasses derivatives of Flavivirus with minimal amino acid changes in its viral proteins, but that possesses the same activities.

The term "gene" is used for simplicity to refer to the nucleic acid giving rise to a functional protein, polypeptide, or peptide-encoding unit. As will be understood by those in the art, this functional term includes genomic sequences, cDNA sequences, and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, domains, peptides, fusion proteins, and mutants. The nucleic

acid molecule encoding Flavivirus may contain a contiguous nucleic acid sequence encoding one or more Flavivirus genes and regulatory regions and be of the following lengths: about 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 441, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710, 720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, 1000, 1010, 1020, 1030, 1040, 1050, 1060, 1070, 1080, 1090, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2100, 2200, 2300, 2400, 2500, 2600, 2700, 2800, 2900, 3000, 3100, 3200, 3300, 3400, 3500, 3600, 3700, 3800, 3900, 4000, 4100, 4200, 4300, 4400, 4500, 4600, 4700, 4800, 4900, 5000, 5100, 5200, 5300, 5400, 5500, 5600, 5700, 5800, 5900, 6000, 6100, 6200, 6300, 6400, 6500, 6600, 6700, 6800, 6900, 7000, 7100, 7200, 7300, 7400, 7500, 7600, 7700, 7800, 7900, 8000, 8100, 8200, 8300, 8400, 8500, 8600, 8700, 8800, 8900, 9000, 9100, 9200, 9300, 9400, 10,000 or more nucleotides, nucleosides, or base pairs. Such sequences may be identical or complementary to all or part of SEQ ID NO:1.

In particular embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating DNA sequences that encode Flavivirus NS5A polypeptides or peptides. Such vectors used in the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA or RNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol.

In a non-limiting example, one or more nucleic acid constructs may be prepared that include a contiguous stretch of nucleotides identical to or complementary to a Flavivirus genome. A nucleic acid construct may be about 50, 60, 70, 80, 90, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1,000, 2,000, 3,000, 4,000, 5,000, 6,000, 7,000, 8,000, 9,000, and 9,400, nucleotides in length, as well as

constructs of greater size, up to and including chromosomal sizes (including all intermediate lengths and intermediate ranges), given the advent of nucleic acids constructs such as a yeast artificial chromosome are known to those of ordinary skill in the art. It will be readily understood that “intermediate lengths” and “intermediate ranges,” as used herein, means any length or range including or between the quoted values (*i.e.*, all integers including and between such values). Non-limiting examples of intermediate lengths include about 11, about 12, about 13, about 16, about 17, about 18, about 19, *etc.*; about 21, about 22, about 23, *etc.*; about 31, about 32, *etc.*; about 51, about 52, about 53, *etc.*; about 101, about 102, about 103, *etc.*; about 151, about 152, about 153, *etc.*

The nucleic acid segments used in the present invention encompass biologically functional and/or immunogenically equivalent Flavivirus NS5A proteins and peptides. Such sequences may arise as a consequence of codon redundancy and functional equivalency that are known to occur naturally within nucleic acid sequences and the proteins thus encoded. Alternatively, functionally and immunologically equivalent proteins or peptides may be created via the application of recombinant DNA technology, in which changes in the protein structure may be engineered, based on considerations of the properties of the amino acids being exchanged. Changes designed by human may be introduced through the application of site-directed mutagenesis techniques, *e.g.*, to introduce improvements to the antigenicity of the protein.

A. Vectors Encoding Flavivirus

The present invention encompasses the use of vectors to encode for all or part of one or more Flavivirus NS5A polypeptides, including an infectious Flavivirus. The term “vector” is used to refer to a carrier nucleic acid molecule into which a nucleic acid sequence can be inserted for introduction into a cell where it can be replicated. A nucleic acid sequence can be “exogenous,” which means that it is foreign to the cell into which the vector is being introduced or that the sequence is homologous to a sequence in the cell but in a position within the host cell nucleic acid in which the sequence is ordinarily not found. Vectors include plasmids, cosmids, viruses (bacteriophage, animal viruses, and plant viruses), and artificial chromosomes (*e.g.*, YACs). In particular embodiments, gene therapy or immunization vectors are

contemplated. One of skill in the art would be well equipped to construct a vector through standard recombinant techniques, which are described in Maniatis *et al.*, 1990 and Ausubel *et al.*, 1996, both incorporated herein by reference.

The term “expression vector” or “expression construct” refers to a vector
5 containing a nucleic acid sequence coding for at least part of a gene product capable of being transcribed. In some cases, RNA molecules are then translated into a protein, polypeptide, or peptide. In other cases, these sequences are not translated, for example, in the production of antisense molecules or ribozymes. Expression vectors can contain a variety of “control sequences,” which refer to nucleic acid sequences
10 necessary for the transcription and possibly translation of an operably linked coding sequence in a particular host organism. In addition to control sequences that govern transcription and translation, vectors and expression vectors may contain nucleic acid sequences that serve other functions as well and are described *infra*. It is contemplated that an infectious Flavivirus particle of the present invention may arise
15 from a vector containing Flavivirus sequence or RNA encoding Flavivirus sequence into a cell. Either of these, or any other nucleic acid molecules of the present invention may be constructed with any of the following nucleic acid control sequences. Thus, the full-length RNA transcript may contain the benefit of recombinant DNA technology such that it contains exogenous control sequences or
20 genes.

1. Promoters and Enhancers

A “promoter” is a control sequence that is a region of a nucleic acid sequence at which initiation and rate of transcription are controlled. It may contain genetic elements at which regulatory proteins and molecules may bind such as RNA
25 polymerase and other transcription factors. The phrases “operatively positioned,” “operatively linked,” “under control,” and “under transcriptional control” means that a promoter is in a correct functional location and/or orientation in relation to a nucleic acid sequence to control transcriptional initiation and/or expression of that sequence. A promoter may or may not be used in conjunction with an “enhancer,” which refers
30 to a cis-acting regulatory sequence involved in the transcriptional activation of a nucleic acid sequence.

A promoter may be one naturally associated with a gene or sequence, as may be obtained by isolating the 5' non-coding sequences located upstream of the coding segment and/or exon. Such a promoter can be referred to as "endogenous." Similarly, an enhancer may be one naturally associated with a nucleic acid sequence, located either downstream or upstream of that sequence. Alternatively, certain advantages will be gained by positioning the coding nucleic acid segment under the control of a recombinant or heterologous promoter, which refers to a promoter that is not normally associated with a nucleic acid sequence in its natural environment. A recombinant or heterologous enhancer refers also to an enhancer not normally associated with a nucleic acid sequence in its natural environment. Such promoters or enhancers may include promoters or enhancers of other genes, and promoters or enhancers isolated from any other prokaryotic, viral, or eukaryotic cell, and promoters or enhancers not "naturally occurring," *i.e.*, containing different elements of different transcriptional regulatory regions, and/or mutations that alter expression. In addition to producing nucleic acid sequences of promoters and enhancers synthetically, sequences may be produced using recombinant cloning and/or nucleic acid amplification technology, including PCRTM, in connection with the compositions disclosed herein (see U.S. Patent 4,683,202 and U.S. Patent 5,928,906, each incorporated herein by reference). Furthermore, it is contemplated the control sequences that direct transcription and/or expression of sequences within non-nuclear organelles such as mitochondria, chloroplasts, and the like, can be employed as well.

Naturally, it will be important to employ a promoter and/or enhancer that effectively directs the expression of the nucleic acid segment in the cell type, organelle, and organism chosen for expression. Those of skill in the art of molecular biology generally know the use of promoters, enhancers, and cell type combinations for protein expression, for example, see Sambrook *et al.* (2001), incorporated herein by reference. The promoters employed may be constitutive, tissue-specific, inducible, and/or useful under the appropriate conditions to direct high level expression of the introduced DNA segment, such as is advantageous in the large-scale production of recombinant proteins and/or peptides. The promoter may be heterologous or exogenous, *i.e.*, from a different source the Flavivirus sequence. In some examples, a prokaryotic promoter is employed for use with *in vitro* transcription of a desired

sequence. Prokaryotic promoters for use with many commercially available systems include T7, T3, and Sp6.

Table 2 lists several elements/promoters that may be employed, in the context of the present invention, to regulate the expression of a gene. This list is not intended
5 to be exhaustive of all the possible elements involved in the promotion of expression but, merely, to be exemplary thereof. Table 3 provides examples of inducible elements, which are regions of a nucleic acid sequence that can be activated in response to a specific stimulus.

TABLE 2	
Promoter and/or Enhancer	
Promoter/Enhancer	References
Immunoglobulin Heavy Chain	Banerji <i>et al.</i> , 1983; Gilles <i>et al.</i> , 1983; Grosschedl <i>et al.</i> , 1985; Atchinson <i>et al.</i> , 1986, 1987; Imler <i>et al.</i> , 1987; Weinberger <i>et al.</i> , 1984; Kiledjian <i>et al.</i> , 1988; Porton <i>et al.</i> ; 1990
Immunoglobulin Light Chain	Queen <i>et al.</i> , 1983; Picard <i>et al.</i> , 1984
T-Cell Receptor	Luria <i>et al.</i> , 1987; Winoto <i>et al.</i> , 1989; Redondo <i>et al.</i> ; 1990
HLA DQ a and/or DQ β	Sullivan <i>et al.</i> , 1987
β -Interferon	Goodbourn <i>et al.</i> , 1986; Fujita <i>et al.</i> , 1987; Goodbourn <i>et al.</i> , 1988
Interleukin-2	Greene <i>et al.</i> , 1989
Interleukin-2 Receptor	Greene <i>et al.</i> , 1989; Lin <i>et al.</i> , 1990
MHC Class II 5	Koch <i>et al.</i> , 1989
MHC Class II HLA-DRA	Sherman <i>et al.</i> , 1989
β -Actin	Kawamoto <i>et al.</i> , 1988; Ng <i>et al.</i> ; 1989
Muscle Creatine Kinase (MCK)	Jaynes <i>et al.</i> , 1988; Horlick <i>et al.</i> , 1989; Johnson <i>et al.</i> , 1989
Prealbumin (Transthyretin)	Costa <i>et al.</i> , 1988
Elastase I	Omitz <i>et al.</i> , 1987
Metallothionein (MTII)	Karin <i>et al.</i> , 1987; Culotta <i>et al.</i> , 1989
Collagenase	Pinkert <i>et al.</i> , 1987; Angel <i>et al.</i> , 1987
Albumin	Pinkert <i>et al.</i> , 1987; Tronche <i>et al.</i> , 1989, 1990
α -Fetoprotein	Godbout <i>et al.</i> , 1988; Campere <i>et al.</i> , 1989
γ -Globin	Bodine <i>et al.</i> , 1987; Perez-Stable <i>et al.</i> , 1990
β -Globin	Trudel <i>et al.</i> , 1987
c-fos	Cohen <i>et al.</i> , 1987
c-HA-ras	Triesman, 1986; Deschamps <i>et al.</i> , 1985
Insulin	Edlund <i>et al.</i> , 1985
Neural Cell Adhesion Molecule (NCAM)	Hirsh <i>et al.</i> , 1990
α_1 -Antitrypsin	Latimer <i>et al.</i> , 1990
H2B (TH2B) Histone	Hwang <i>et al.</i> , 1990

TABLE 2	
Promoter and/or Enhancer	
Promoter/Enhancer	References
Mouse and/or Type I Collagen	Ripe <i>et al.</i> , 1989
Glucose-Regulated Proteins (GRP94 and GRP78)	Chang <i>et al.</i> , 1989
Rat Growth Hormone	Larsen <i>et al.</i> , 1986
Human Serum Amyloid A (SAA)	Edbrooke <i>et al.</i> , 1989
Troponin I (TN I)	Yutzey <i>et al.</i> , 1989
Platelet-Derived Growth Factor (PDGF)	Pech <i>et al.</i> , 1989
Duchenne Muscular Dystrophy	Klamut <i>et al.</i> , 1990
SV40	Banerji <i>et al.</i> , 1981; Moreau <i>et al.</i> , 1981; Sleight <i>et al.</i> , 1985; Firak <i>et al.</i> , 1986; Herr <i>et al.</i> , 1986; Imbra <i>et al.</i> , 1986; Kadesch <i>et al.</i> , 1986; Wang <i>et al.</i> , 1986; Ondek <i>et al.</i> , 1987; Kuhl <i>et al.</i> , 1987; Schaffner <i>et al.</i> , 1988
Polyoma	Swartzendruber <i>et al.</i> , 1975; Vasseur <i>et al.</i> , 1980; Katinka <i>et al.</i> , 1980, 1981; Tyndell <i>et al.</i> , 1981; Dandolo <i>et al.</i> , 1983; de Villiers <i>et al.</i> , 1984; Hen <i>et al.</i> , 1986; Satake <i>et al.</i> , 1988; Campbell and/or Villarreal, 1988
Retroviruses	Kriegler <i>et al.</i> , 1982, 1983; Levinson <i>et al.</i> , 1982; Kriegler <i>et al.</i> , 1983, 1984a, b, 1988; Bosze <i>et al.</i> , 1986; Miksicek <i>et al.</i> , 1986; Celander <i>et al.</i> , 1987; Thiesen <i>et al.</i> , 1988; Celander <i>et al.</i> , 1988; Chol <i>et al.</i> , 1988; Reisman <i>et al.</i> , 1989
Papilloma Virus	Campo <i>et al.</i> , 1983; Lusky <i>et al.</i> , 1983; Spandidos and/or Wilkie, 1983; Spalholz <i>et al.</i> , 1985; Lusky <i>et al.</i> , 1986; Cripe <i>et al.</i> , 1987; Gloss <i>et al.</i> , 1987; Hirochika <i>et al.</i> , 1987; Stephens <i>et al.</i> , 1987; Glue <i>et al.</i> , 1988
Hepatitis B Virus	Bulla <i>et al.</i> , 1986; Jameel <i>et al.</i> , 1986; Shaul <i>et al.</i> , 1987; Spandau <i>et al.</i> , 1988; Vannice <i>et al.</i> , 1988
Human Immunodeficiency Virus	Muesing <i>et al.</i> , 1987; Hauber <i>et al.</i> , 1988; Jakobovits <i>et al.</i> , 1988; Feng <i>et al.</i> , 1988; Takebe <i>et al.</i> , 1988; Rosen <i>et al.</i> , 1988; Berkhout <i>et al.</i> , 1989; Laspia <i>et al.</i> , 1989; Sharp <i>et al.</i> , 1989; Braddock <i>et al.</i> , 1989
Cytomegalovirus (CMV)	Weber <i>et al.</i> , 1984; Boshart <i>et al.</i> , 1985; Foecking <i>et al.</i> , 1986

TABLE 2	
Promoter and/or Enhancer	
Promoter/Enhancer	References
Gibbon Ape Leukemia Virus	Holbrook <i>et al.</i> , 1987; Quinn <i>et al.</i> , 1989

TABLE 3		
Inducible Elements		
Element	Inducer	References
MT II	Phorbol Ester (TFA) Heavy metals	Palmiter <i>et al.</i> , 1982; Haslinger <i>et al.</i> , 1985; Searle <i>et al.</i> , 1985; Stuart <i>et al.</i> , 1985; Imagawa <i>et al.</i> , 1987, Karin <i>et al.</i> , 1987; Angel <i>et al.</i> , 1987b; McNeall <i>et al.</i> , 1989
MMTV (mouse mammary tumor virus)	Glucocorticoids	Huang <i>et al.</i> , 1981; Lee <i>et al.</i> , 1981; Majors <i>et al.</i> , 1983; Chandler <i>et al.</i> , 1983; Lee <i>et al.</i> , 1984; Ponta <i>et al.</i> , 1985; Sakai <i>et al.</i> , 1988
β -Interferon	poly(rI)x poly(rc)	Tavernier <i>et al.</i> , 1983
Adenovirus 5 E2	E1A	Imperiale <i>et al.</i> , 1984
Collagenase	Phorbol Ester (TPA)	Angel <i>et al.</i> , 1987a
Stromelysin	Phorbol Ester (TPA)	Angel <i>et al.</i> , 1987b
SV40	Phorbol Ester (TPA)	Angel <i>et al.</i> , 1987b
Murine MX Gene	Interferon, Newcastle Disease Virus	Hug <i>et al.</i> , 1988
GRP78 Gene	A23187	Resendez <i>et al.</i> , 1988
α -2-Macroglobulin	IL-6	Kunz <i>et al.</i> , 1989
Vimentin	Serum	Rittling <i>et al.</i> , 1989
MHC Class I Gene H-2kb	Interferon	Blonar <i>et al.</i> , 1989
HSP70	E1A, SV40 Large T Antigen	Taylor <i>et al.</i> , 1989, 1990a, 1990b
Proliferin	Phorbol Ester-TPA	Mordacq <i>et al.</i> , 1989
Tumor Necrosis Factor	PMA	Hensel <i>et al.</i> , 1989

TABLE 3		
Inducible Elements		
Element	Inducer	References
Thyroid Stimulating Hormone α Gene	Thyroid Hormone	Chatterjee <i>et al.</i> , 1989

The identity of tissue-specific promoters or elements, as well as assays to characterize their activity, is well known to those of skill in the art. Examples of such regions include the human LIMK2 gene (Nomoto *et al.* 1999), the somatostatin receptor 2 gene (Kraus *et al.*, 1998), murine epididymal retinoic acid-binding gene (Lareyre *et al.*, 1999), human CD4 (Zhao-Emonet *et al.*, 1998), mouse alpha2 (XI) collagen (Tsumaki, *et al.*, 1998), D1A dopamine receptor gene (Lee, *et al.*, 1997), insulin-like growth factor II (Wu *et al.*, 1997), human platelet endothelial cell adhesion molecule-1 (Almendo *et al.*, 1996).

2. Initiation Signals and Internal Ribosome Binding Sites

A specific initiation signal also may be required for efficient translation of coding sequences. These signals include the ATG initiation codon or adjacent sequences. Exogenous translational control signals, including the ATG initiation codon, may need to be provided. One of ordinary skill in the art would readily be capable of determining this and providing the necessary signals. It is well known that the initiation codon must be "in-frame" with the reading frame of the desired coding sequence to ensure translation of the entire insert. The exogenous translational control signals and initiation codons can be either natural or synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements.

In certain embodiments of the invention, the use of internal ribosome entry sites (IRES) elements are used to create multigene, or polycistronic, messages. IRES elements are able to bypass the ribosome-scanning model of 5' methylated Cap dependent translation and begin translation at internal sites (Pelletier and Sonenberg, 1988). IRES elements from two members of the picornavirus family (polio and encephalomyocarditis) have been described (Pelletier and Sonenberg, 1988), as well an IRES from a mammalian message (Macejak and Sarnow, 1991). IRES elements can be linked to heterologous open reading frames. Multiple open reading frames can

be transcribed together, each separated by an IRES, creating polycistronic messages. By virtue of the IRES element, each open reading frame is accessible to ribosomes for efficient translation. Multiple genes can be efficiently expressed using a single promoter/enhancer to transcribe a single message (see U.S. Patent 5,925,565 and
5 5,935,819, herein incorporated by reference).

3. Multiple Cloning Sites

Vectors can include a multiple cloning site (MCS), which is a nucleic acid region that contains multiple restriction enzyme sites, any of which can be used in conjunction with standard recombinant technology to digest the vector. (See
10 Carbonelli *et al.*, 1999, Levenson *et al.*, 1998, and Cocea, 1997, incorporated herein by reference.) "Restriction enzyme digestion" refers to catalytic cleavage of a nucleic acid molecule with an enzyme that functions only at specific locations in a nucleic acid molecule. Many of these restriction enzymes are commercially available. Use of such enzymes is widely understood by those of skill in the art. Frequently, a vector is
15 linearized or fragmented using a restriction enzyme that cuts within the MCS to enable exogenous sequences to be ligated to the vector. "Ligation" refers to the process of forming phosphodiester bonds between two nucleic acid fragments, which may or may not be contiguous with each other. Techniques involving restriction enzymes and ligation reactions are well known to those of skill in the art of
20 recombinant technology.

4. Termination Signals

The vectors or constructs of the present invention will generally comprise at least one termination signal. A "termination signal" or "terminator" is comprised of the DNA sequences involved in specific termination of an RNA transcript by an RNA
25 polymerase. Thus, in certain embodiments a termination signal that ends the production of an RNA transcript is contemplated. A terminator may be necessary *in vivo* to achieve desirable message levels.

In eukaryotic systems, the terminator region may also comprise specific DNA sequences that permit site-specific cleavage of the new transcript to expose a
30 polyadenylation site. This signals a specialized endogenous polymerase to add a stretch of about 200 A residues (polyA) to the 3' end of the transcript. RNA

molecules modified with this polyA tail appear to more stable and are translated more efficiently. Thus, in other embodiments involving eukaryotes, it is preferred that that terminator comprises a signal for the cleavage of the RNA, and it is more preferred that the terminator signal promotes polyadenylation of the message. The terminator
5 and/or polyadenylation site elements can serve to enhance message levels and/or to minimize read through from the cassette into other sequences.

Terminators contemplated for use in the invention include any known terminator of transcription described herein or known to one of ordinary skill in the art, including but not limited to, for example, the termination sequences of genes,
10 such as for example the bovine growth hormone terminator or viral termination sequences, such as for example the SV40 terminator. In certain embodiments, the termination signal may be a lack of transcribable or translatable sequence, such as due to a sequence truncation.

5. Polyadenylation Signals

15 For expression, particularly eukaryotic expression, one will typically include a polyadenylation signal to effect proper polyadenylation of the transcript. The nature of the polyadenylation signal is not believed to be crucial to the successful practice of the invention, and/or any such sequence may be employed. Preferred embodiments include the SV40 polyadenylation signal and/or the bovine growth hormone
20 polyadenylation signal, convenient and/or known to function well in various target cells. Polyadenylation may increase the stability of the transcript or may facilitate cytoplasmic transport.

6. Origins of Replication

In order to propagate a vector in a host cell, it may contain one or more origins
25 of replication sites (often termed "ori"), which is a specific nucleic acid sequence at which replication is initiated. Alternatively, an autonomously replicating sequence (ARS) can be employed if the host cell is yeast.

7. Selectable and Screenable Markers

In certain embodiments of the invention, the cells containing a nucleic acid
30 construct of the present invention may be identified *in vitro* or *in vivo* by including a

marker in the expression vector. Such markers would confer an identifiable change to the cell permitting easy identification of cells containing the expression vector. Generally, a selectable marker is one that confers a property that allows for selection. A positive selectable marker is one in which the presence of the marker allows for its selection, while a negative selectable marker is one in which its presence prevents its selection. An example of a positive selectable marker is a drug resistance marker.

Usually the inclusion of a drug selection marker aids in the cloning and identification of transformants, for example, genes that confer resistance to neomycin, puromycin, hygromycin, DHFR, GPT, zeocin and histidinol are useful selectable markers. In addition to markers conferring a phenotype that allows for the discrimination of transformants based on the implementation of conditions, other types of markers including screenable markers such as GFP, whose basis is colorimetric analysis, are also contemplated. Alternatively, screenable enzymes such as herpes simplex virus thymidine kinase (*tk*) or chloramphenicol acetyltransferase (CAT) may be utilized. One of skill in the art would also know how to employ immunologic markers, possibly in conjunction with FACS analysis. The marker used is not believed to be important, so long as it is capable of being expressed simultaneously with the nucleic acid encoding a gene product. Further examples of selectable and screenable markers are well known to one of skill in the art.

B. Host Cells

As used herein, the terms "cell," "cell line," and "cell culture" may be used interchangeably. All of these terms also include their progeny, which refers to any and all subsequent generations. It is understood that all progeny may not be identical due to deliberate or inadvertent mutations. In the context of expressing a heterologous nucleic acid sequence, "host cell" refers to a prokaryotic or eukaryotic cell, and it includes any transformable organisms that is capable of replicating a vector and/or expressing a heterologous gene encoded by a vector. A host cell can, and has been, used as a recipient for vectors. A host cell may be "transfected" or "transformed," which refers to a process by which exogenous nucleic acid is transferred or introduced into the host cell. A transformed cell includes the primary subject cell and its progeny.

Host cells may be derived from prokaryotes or eukaryotes, depending upon whether the desired result is replication of the vector, expression of part or all of the vector-encoded nucleic acid sequences, or production of infectious viral particles. Numerous cell lines and cultures are available for use as a host cell, and they can be
5 obtained through the American Type Culture Collection (ATCC), which is an organization that serves as an archive for living cultures and genetic materials. An appropriate host can be determined by one of skill in the art based on the vector backbone and the desired result. A plasmid or cosmid, for example, can be introduced into a prokaryote host cell for replication of many vectors. Bacterial cells
10 used as host cells for vector replication and/or expression include DH5 α , JM109, and KC8, as well as a number of commercially available bacterial hosts such as SURE[®] Competent Cells and SOLOPACK[™] Gold Cells (STRATAGENE[®], La Jolla). Alternatively, bacterial cells such as *E. coli* LE392 could be used as host cells for phage viruses.

15 Examples of eukaryotic host cells for replication and/or expression of a vector include HeLa, NIH3T3, Jurkat, 293, Cos, CHO, Saos, and PC12. Many host cells from various cell types and organisms are available and would be known to one of skill in the art. Similarly, a viral vector may be used in conjunction with either an eukaryotic or prokaryotic host cell, particularly one that is permissive for replication
20 or expression of the vector.

C. Expression Systems

Numerous expression systems exist that comprise at least all or part of the compositions discussed above. Prokaryote- and/or eukaryote-based systems can be employed for use with the present invention to produce nucleic acid sequences, or
25 their cognate polypeptides, proteins and peptides. Many such systems are commercially and widely available.

The insect cell/baculovirus system can produce a high level of protein expression of a heterologous nucleic acid segment, such as described in U.S. Patents 5,871,986 and 4,879,236, both herein incorporated by reference, and which can be
30 bought, for example, under the name MAXBAC[®] 2.0 from INVITROGEN[®] and BACPACK[™] BACULOVIRUS EXPRESSION SYSTEM from CLONTECH[®].

Other examples of expression systems include STRATAGENE[®]'s COMPLETE CONTROL[™] Inducible Mammalian Expression System, which involves a synthetic ecdysone-inducible receptor, or its pET Expression System, an *E. coli* expression system. Another example of an inducible expression system is available from
5 INVITROGEN[®], which carries the T-REX[™] (tetracycline-regulated expression) System, an inducible mammalian expression system that uses the full-length CMV promoter. The Tet-On[™] and Tet-Off[™] systems from CLONTECH[®] can be used to regulate expression in a mammalian host using tetracycline or its derivatives. The implementation of these systems is described in Gossen *et al.*, 1992 and Gossen *et al.*,
10 1995, and U.S. Patent 5,650,298, all of which are incorporated by reference.

INVITROGEN[®] also provides a yeast expression system called the *Pichia methanolica* Expression System, which is designed for high-level production of recombinant proteins in the methylotrophic yeast *Pichia methanolica*. One of skill in the art would know how to express a vector, such as an expression construct, to
15 produce a nucleic acid sequence or its cognate polypeptide, protein, or peptide.

D. Introduction of Nucleic Acids into Cells

In certain embodiments, a nucleic acid may be introduced into a cell *in vitro* for production of polypeptides or *in vivo* for immunization purposes. There are a number of ways in which nucleic acid molecules such as expression vectors may be
20 introduced into cells. In certain embodiments of the invention, the expression vector comprises a Flavivirus infectious particle or engineered vector derived from a Flavivirus genome. In other embodiments, an expression vector known to one of skill in the art may be used to express a segment of a Flavivirus nucleic, which may be translated into a Flavivirus polypeptide or peptide. The ability of certain viruses to
25 enter cells via receptor-mediated endocytosis, to integrate into host cell genome and express viral genes stably and efficiently have made them attractive candidates for the transfer of foreign genes into mammalian cells (Ridgeway, 1988; Nicolas and Rubenstein, 1988; Baichwal and Sugden, 1986; Temin, 1986).

"Viral expression vector" is meant to include those vectors containing
30 sequences of that virus sufficient to (a) support packaging of the vector and (b) to express a polynucleotide that has been cloned therein. In this context, expression may

require that the gene product be synthesized. A number of such viral vectors have already been thoroughly researched, including adenovirus, adeno-associated viruses, retroviruses, herpesviruses, and vaccinia viruses.

Delivery may be accomplished *in vitro*, as in laboratory procedures for transforming cells lines, or *in vivo* or *ex vivo*, as in the treatment of certain disease states. One mechanism for delivery is via viral infection where the expression vector is encapsidated in an infectious viral particle. Several non-viral methods for the transfer of expression vectors into cultured mammalian cells also are contemplated by the present invention. These include calcium phosphate precipitation (Graham and Van Der Eb, 1973; Chen and Okayama, 1987; Rippe *et al.*, 1990) DEAE-dextran (Gopal, 1985), electroporation (Tur-Kaspa *et al.*, 1986; Potter *et al.*, 1984), direct microinjection (Harland and Weintraub, 1985), DNA-loaded liposomes (Nicolau and Sene, 1982; Fraley *et al.*, 1979) and lipofectamine-DNA complexes, cell sonication (Fechheimer *et al.*, 1987), gene bombardment using high velocity microprojectiles (Yang *et al.*, 1990), liposome (Ghosh and Bachhawat, 1991; Kaneda *et al.*, 1989) and receptor-mediated transfection (Wu and Wu, 1987; Wu and Wu, 1988). Some of these techniques may be successfully adapted for *in vivo* or *ex vivo* use.

In certain embodiments, the nucleic acid encoding a gene or genes may be stably integrated into the genome of the cell. This integration may be in the cognate location and orientation via homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the nucleic acid may be stably maintained in the cell as a separate, episomal segment of DNA. Such nucleic acid segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or in synchronization with the host cell cycle. How the expression vector is delivered to a cell and where in the cell the nucleic acid remains is dependent on the type of expression vector employed.

Transfer of a nucleic acid molecule may be performed by any of the methods mentioned above which physically or chemically permeabilize the cell membrane. This is particularly applicable for transfer *in vitro*, but it may be applied to *in vivo* use as well.

VI. ANTI-HIV THERAPIES

In certain embodiments, therapeutic methods will include administering to a patient or subject a composition comprising an antigen or an antibody derived from a Flavivirus NS5A peptide or polypeptide, such as human or humanized animal derived antibodies. In various embodiments, the treatment methods of the invention may be used in combination with other anti-HIV treatments, such as Flavivirus infection as a therapeutic or preventative treatment for AIDS. For exemplary compositions and methods see PCT application WO 01/77157, which is incorporated herein by reference.

As a therapeutic measure, a Flavivirus NS5A agent can be used to reduce the severity or progression of AIDS, including the prevention of AIDS in HIV-infected individuals. A reduction in severity or progression of AIDS includes, but is not limited to, prevention of or a reduction in the severity, duration, or discomfort associated with the following conditions: prolonged and unexplained fatigue; swollen glands; prolonged fever; chills; excessive sweating; swollen gums and mouth lesions; sore throat; cough; shortness of breath; constipation; diarrhea; symptoms of well-known opportunistic infections; Kaposi sarcomas; skin rashes or lesions; loss of appetite or weight loss; malaise; headaches; speech impairment; muscle atrophy; memory loss; reduced cognitive functioning; swelling of the joints; joint stiffness or pain; cold intolerance; pain or tenderness in bones; energy level; anxiety, stress, and tension; groin lump; pruritus; genital sores; blurred or decreased vision; diplopia; light sensitivity; pain in chest, sides, back, muscle or stomach; and seizures.

As a preventative measure, a patient may be administered a pharmaceutically acceptable composition comprising a Flavivirus NS5A peptide or polypeptide. This agent may be used in conjunction with infection of CD4+ T cells with Flavivirus or a recombinant version of Flavivirus to inhibit infection of these cells by HIV. Alternatively, treatment with the Flavivirus NS5A compositions of the present invention may effect a combination of preventative and therapeutic treatments insofar as infection of other cells in an HIV-infected subject's body is prevented or attenuated.

Inhibition of AIDS progression may be demonstrated by reduction of detectable HIV in the HIV-infected subject; maintaining a CD4 count above 200 for a longer than average period of time; maintaining a normal T cell count; or maintaining normal p24 antigen. The term "therapeutic benefit" or "therapeutic effect" used
5 throughout this application refers to anything that promotes or enhances the well-being of the subject with respect to the medical treatment of his/her condition, which includes treatment of HIV-infection (before the onset of AIDS), AIDS, as well as treatment of Hepatitis C. A list of nonexhaustive examples of this includes extension of the subject's life by any period of time; decrease or delay in the progression of
10 AIDS (HIV, as described above) or Hepatitis C; decrease in viral load of HIV or HCV; decrease in HIV replication; clearance of HIV or HCV viremia reduced transmission of HCV or HIV; decrease in liver damage or complications; and a decrease in pain to the subject that can be attributed to the subject's condition.

VII. COMBINATION THERAPIES

Of course it is understood that the method of the present invention, particularly administration of NS5A agents as treatment for an HIV-infected subject, may also be used in combination with the administration of traditional therapies. Alternatively, the compositions of the present invention may be given in combination with treatment or prevention of hepatitis C, such as α -interferon. Some such therapies
20 are described below.

In many clinical situations, it is advisable to use a combination of distinct therapies. Thus, it is envisioned that, in addition to the therapies described herein, one would also wish to provide to the patient more "standard" pharmaceutical anti-retroviral therapies. Examples of standard therapies are provided below.

Combinations may be achieved by administering to a patient a single composition or pharmacological formulation that includes both agents, or by administering to a patient two distinct compositions or formulations, at the same time, wherein one composition may include a Flavivirus NS5A, or expression construct encoding such, and the other includes the standard anti-retroviral therapy.
25 Alternatively, a Flavivirus-based therapeutic may precede or follow the other treatment by intervals ranging from minutes to weeks. In embodiments where the
30

other agent and NS5A are administered separately to the patient, one would generally ensure that a significant period of time did not expire between the time of each delivery, such that the agent and NS5A would still be able to exert an advantageously combined effect on the patient. In such instances, it is contemplated that one would
 5 administer to the patient both modalities within about 12-24 hours of each other and, more preferably, within about 6-12 hours of each other, with a delay time of only about 12 hours being most preferred. In some situations, it may be desirable to extend the time period for treatment significantly, however, where several days (2, 3, 4, 5, 6, or 7) to several weeks (1, 2, 3, 4, 5, 6, 7, or 8) lapse between the respective
 10 administrations.

It also is conceivable that more than one administration of a NS5A-based therapeutic agent will be desired. Various combinations may be employed, where a NS5A is "A" and the other agent is "B," as exemplified below:

A/B/A B/A/B B/B/A A/A/B B/A/A A/B/B B/B/B/A B/B/A/B
 15 A/A/B/B A/B/A/B A/B/B/A B/B/A/A B/A/B/A B/A/A/B B/B/B/A
 A/A/A/B B/A/A/A A/B/A/A A/A/B/A A/B/B/B B/A/B/B B/B/A/B

Other combinations are contemplated as well.

A. AZT

A well known, traditional therapy for the treatment of AIDS involves
 20 zovidovudine (AZT™ available from Burroughs Wellcome). This is one of a class of nucleoside analogues known as dideoxynucleosides which block HIV replication by inhibiting HIV reverse transcriptase. The anti-AIDS drug zidovudine (also known as AZT) may also be used in limited circumstances, mostly in combination with rifampin, as described by Burger *et al.* (1993).

25 The compositions and methods disclosed herein will be particularly effective in conjunction with other forms of therapy, such as AZT and/or protease inhibitors that are designed to inhibit viral replication, by maintaining desirable levels of white blood cells. This, in effect, buys the patient the time necessary for the anti-viral therapies to work.

B. HAART

New combination drug therapy has shown promising results in the treatment of HIV-infected patients. Treatment with potent anti-HIV drug combinations is referred to as "highly active anti-retroviral therapy" (HAART), and it has provided clinical improvement, longer survival, and improved quality of life for people infected with HIV during all four stages of HIV disease. Examples of HAART include a protease inhibitor (indinavir, nelfinavir, ritonavir, ritonavir/saquinavir, or saquinavir) combined with two nucleoside analogs (AZT/ddI, d4T/ddI, AZT/ddC, AZT/3TC, or d4T/3TC).

In many instances, it will be desirable to have multiple administrations of the inventive compositions and/or a vaccines, usually not exceeding six administrations or vaccinations, more usually not exceeding four vaccinations. In certain embodiments, one or more, usually at least about three administrations or vaccinations may be provided. The administrations or vaccinations will normally be at from two to twelve week intervals, more usually from three to five week intervals. Periodic boosters at intervals of 1-5 years, usually three years, will be desirable to maintain protective levels of the antibodies. The course of the immunization or treatment may be followed by standard antibody assays. The assays may be performed by labeling with conventional labels, such as radionuclides, enzymes, fluorescents, and the like. These techniques are well known and may be found in a wide variety of patents, such as U.S. Patents 3,791,932; 4,174,384 and 3,949,064, as illustrative of these types of assays.

The manner of application may be varied widely. Any of the conventional methods for administration of an antibody or vaccine are applicable. These are believed to include oral application on a solid physiologically acceptable base or in a physiologically acceptable dispersion, parenterally, by injection or the like. The dosage of the NS5A agent will depend on the route of administration and will vary according to the size of the host.

The NS5A agents and flavivirus nucleic acids of the invention may be formulated into a pharmaceutically acceptable composition, see below, or vaccine as neutral or salt forms. Pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the peptide) and those that are formed with

inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases
5 as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

The preparation of flavivirus NS5A agents as active ingredients is generally well understood in the art by analogy, as exemplified by U.S. Patents 6,479,243, 6,399,763, 5,714,153, 5,582,981, and 4,833,077, all incorporated herein by reference.
10 The preparation of vaccines that contain flavivirus sequences as active ingredients is generally well understood in the art by analogy, as exemplified by U.S. Patents 5,958,895, 6,004,799, and 5,620,896, all incorporated herein by reference.

VIII. PHARMACEUTICAL COMPOSITIONS AND ROUTES OF ADMINISTRATION

15 Pharmaceutical compositions including NS5A peptides and polypeptides will be formulated along the line of typical pharmaceutical drug and biological preparations. A discussion of formulations may be found in Remington's Pharmaceutical Sciences (1990). The percentage of active compound in any pharmaceutical preparation is dependent upon both the activity of the compound, in
20 this case ability of NS5A agents to inhibit HIV replication. Typically, such compositions should contain at least 0.1% active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 2 to about 60% of the weight of the unit. The amount of active compounds in such therapeutically useful compositions is such that a suitable dosage
25 will be obtained.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases, the form must be sterile and must be fluid to the extent that easy injection is possible. It must be stable under the
30 conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can

be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, phenylmercuric nitrate, m-cresol, and the like. In many cases, it will be preferable to use isotonic solutions, for example, sugars or sodium chloride.

10 Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate, and gelatin.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by sterile filtration. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying techniques that yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

The phrases "pharmaceutically acceptable" or "pharmacologically acceptable" refer to molecular entities and compositions that do not produce an adverse, allergic, or other untoward reaction when administered to an animal, or human, as appropriate. As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredients, its use in the therapeutic compositions is contemplated. Supplementary active ingredients, such as other anti-cancer agents, can also be incorporated into the compositions.

The active compounds of the present invention can be formulated for parenteral administration, *e.g.*, formulated for injection *via* the intravenous, intramuscular, intrathoracic, sub-cutaneous, or even intraperitoneal routes. Administration by i.v. or i.m. are specifically contemplated. Pharmaceutically acceptable salts, include the acid salts and those which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

In certain embodiments, it may be desirable to provide a continuous supply of therapeutic compositions to the patient. For intravenous or intraarterial routes, this is accomplished by drip system. For various approaches, delayed release formulations could be used that provided limited but constant amounts of the therapeutic agent over and extended period of time. For internal application, continuous perfusion may be preferred. This could be accomplished by catheterization followed by continuous administration of the therapeutic agent. The time period for perfusion would be selected by the clinician for the particular patient and situation, but times could range from about 1-2 hours, to 2-6 hours, to about 6-10 hours, to about 10-24 hours, to about 1-2 days, to about 1-2 weeks or longer. Generally, the dose of the therapeutic composition via continuous perfusion will be equivalent to that given by single or multiple injections, adjusted for the period of time over which the injections are administered. It is believed that higher doses may be achieved via perfusion, however.

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, sterile aqueous media that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage could be dissolved in 1 mL of isotonic NaCl solution and either added to

1000 mL of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, Remington's Pharmaceutical Sciences, 1990). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. The person responsible for administration will, in any event, determine the appropriate
5 dose for the individual subject.

An effective amount of the therapeutic composition is determined based on the intended goal. The term "unit dose" or "dosage" refers to physically discrete units suitable for use in a subject, each unit containing a predetermined-quantity of the therapeutic composition calculated to produce the desired responses, discussed above,
10 in association with its administration, *i.e.*, the appropriate route and treatment regimen. The quantity to be administered, both according to number of treatments and unit dose, depends on the protection desired.

Peptides or polypeptides may be administered in a dose that can vary from 0.01, 0.05, 0.1, 0.5, 1, 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100 mg/kg of weight to 50,
15 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200 mg/kg of weight in one or more daily, weekly, monthly, or yearly administrations during one or various days, weeks, months, or years. The antibodies can be administered by parenteral injection (intravenous, intraperitoneal, intramuscular, subcutaneous, intracavity or transdermic). For viral vectors, one generally will prepare a viral vector
20 stock. Depending on the kind of virus and the titer attainable, one will deliver 1 to 100, 10 to 50, 100-1000, or up to 1×10^4 , 1×10^5 , 1×10^6 , 1×10^7 , 1×10^8 , 1×10^9 , 1×10^{10} , 1×10^{11} , or 1×10^{12} infectious particles to the patient. Similar figures may be extrapolated for liposomal or other non-viral formulations by comparing relative uptake efficiencies. Formulation as a pharmaceutically acceptable composition is
25 discussed below.

In many instances, it will be desirable to have multiple administrations of the NS5A agent. The compositions of the invention may be administered 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more times. The administrations will normally be at from one to twelve week intervals, more usually from one to four week intervals. Periodic re-
30 administration will be desirable with recurrent exposure to the pathogen (*e.g.*, HIV). For example, an HIV positive mother would be re-inoculated prior to parturition from a second pregnancy.

Precise amounts and delivery regimen for the therapeutic composition also depend on the judgment of the practitioner and are peculiar to each individual. Factors affecting dose include physical and clinical state of the patient, the route of administration, the intended goal of treatment (alleviation of symptoms *versus* cure) and the potency, stability, and toxicity of the particular therapeutic substance.

In a particular embodiment of the invention, the NS5A agent may be entrapped in a liposome. Liposomes are vesicular structures characterized by a phospholipid bilayer membrane and an inner aqueous medium. Multilamellar liposomes have multiple lipid layers separated by aqueous medium. They form spontaneously when phospholipids are suspended in an excess of aqueous solution. The lipid components undergo self-rearrangement before the formation of closed structures and entrap water and dissolved solutes between the lipid bilayers (Ghosh and Bachhawat, 1991).

Current *in vivo* lipid delivery methods use subcutaneous, intradermal, intratumoral, or intracranial injection to avoid the toxicity and stability problems associated with cationic lipids in the circulation. The DOTAP:cholesterol lipid formulation is said to form a unique structure termed a "sandwich liposome." This formulation is reported to "sandwich" DNA between an invaginated bi-layer or 'vase' structure. Beneficial characteristics of these lipid structures include a positive ρ , colloidal stabilization by cholesterol, two dimensional DNA packing and increased serum stability.

The production of lipid formulations often is accomplished by sonication or serial extrusion of liposomal mixtures after (I) reverse phase evaporation (II) dehydration-rehydration (III) detergent dialysis and (IV) thin film hydration. Once manufactured, lipid structures can be used to encapsulate compounds that are toxic (chemotherapeutics) or labile (nucleic acids) when in circulation. Lipid encapsulation has resulted in a lower toxicity and a longer serum half-life for such compounds (Gabizon *et al.*, 1990).

VIII. EXAMPLES

The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventors to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

EXAMPLE 1

Cells: Tet-Off Jurkat cells (Clontech) were transfected with pTRE 2 Hyg plasmids containing full-length GBV-C NS5A (from the full-length GBV-C infectious clone AF121950) or with the vector only (control) by electroporation. Transfectants were selected using hygromycin, and cell lines cloned twice. Cells were grown in RPMI 1640 with or without doxycycline as recommended.

NS5A expression: Cells were lysed in RIPA buffer containing protease and phosphatase inhibitors and examined by immunoblot using rabbit anti-NS5A antisera.

HIV infections: Clinical and laboratory CXCR4 tropic strains of HIV-1 were used to infect Jurkat cell lines expressing NS5A or the vector control. Cells were maintained in doxycycline at various concentrations to inhibit NS5A expression. HIV replication was measured using culture supernatant p24 antigen ELISA.

Results: Jurkat cell lines stably expressing GBV-C NS5A protein were established, and expression of NS5A was regulated by doxycycline. HIV infection of cells expressing NS5A was decreased, as measured by p24 antigen compared to control cells or NS5A-containing cells grown in the presence of doxycycline. NS5A-expressing cells also had lower surface density of CXCR4 and released increased levels of the CXCR4 ligand SDF-1 into culture supernatants, which may account for observations. In addition, NS5A led to increased levels of two genes associated with resistance to apoptosis, and rendered cells resistant to Fas-mediated apoptosis. This

latter observation may account for slower decline in CD4 cell counts observed in HIV-infected people.

Summary: Jurkat cell lines stably expressing GBV-C NS5A protein (regulated by doxycycline) were established (FIG. 1). HIV infection of Jurkat cells that expressed GBV-C NS5A resulted in decreased levels of HIV replication (measured by p24 antigen) compared to vector control cells or cells in which NS5A expression was reduced by growth in doxycycline (FIGS. 2A-C). NS5A expressing cells had lower levels of CXCR4 on the surface than vector control cells, and also had reduced release of SDF-1 into culture supernatants, suggesting that NS5A expression may interact with chemokine receptor pathways (FIGS. 3A-B). Although part of the HIV inhibitory effect of GBV-C is mediated by E2 protein interactions with cellular receptor(s), the NS5A protein appears to also contribute to an anti-HIV effect of GBV-C (FIGS. 4A-B). GBV-C NS5A, like HCV, appears to have two phosphorylation states. HCV NS5A hyperphosphorylation has been reported by some to require NS4 expression. These data show that NS4 is not required for hyperphosphorylation of GBV-C NS5A (FIG. 5). NS5A led to relative resistance to Fas-mediated apoptosis, which could slow CD4 cell decline. HCV NS5A has previously been shown to be anti-apoptotic (FIG. 6). GBV-C infected PBMCs did not show decreased CXCR4 expression, although CCR5 expression was reduced. Because active GBV-C replication is required to produce NS5A, and active replication occurs in a small percentage of PBMCs, downregulation of CXCR4 would be missed. In contrast, E2 (structural protein) is released into supernatant and will have more widespread effects leading to decreased CCR5 (FIG. 7).

**EXAMPLE 2 – NS5A Inhibition of HIV Replication is Dose Dependent,
Independent of PKR and May Involve SDF-1**

Using Jurkat cells that are infected with HIV, doxycycline-repressable expression constructs encoding GBV-C NS5A were used to inhibit HIV replication. As shown in FIG. 8, the use of 0, 0.01, 0.1 and 1 µg of doxycycline produced decreasing amounts of HIV inhibition as measured by p24 antigen levels. No such effect was seen in control Jurkat cells.

The inventors previously showed that NS5A's from interferon resistant (IFN-R) GBV-C inhibited PKR-mediated phosphorylation of eIF2 α , which is part of the inteferon cascade, whereas interferon sensitive (IFN-S) did not. The NS5A's from IFN-R and IFN-S strains were compared for their respective abilities to inhibit HIV
5 replication. As shown in FIG. 9, both strains were equally effective at inhibiting HIV replication as measured by p24 antigen concentration. Thus, the effect is apparently not related to PKR-function.

Following treatment with of HIV-infected Jurkat cells with NS5A, SDF-1 is released into culture supernatants (data not shown). In an effort to determine whether
10 there is any link between SDF-1 and HIV inhibition, anti-SDF-1 antibodies were added to the Jurkat cell cultures. As shown in FIG. 10, there was an increase in HIV replication ranging from 50% to almost 60% as compared to untreated controls, thereby suggesting that SDF-1 might play a role.

EXAMPLE 3 – NS5A Inhibits Distinct Strains of HIV

15 Three different strains of HIV – ELI (X4) Clade D, JF (X4) Clade B and MN (X4) Clade B – were tested for replication in the presence of NS5A. As shown in FIGS. 11A-C, all were sensitive to NS5A-mediated inhibition.

EXAMPLE 4 – GBV-C NS5A Fragments Inhibit HIV Replication

A variety of NS5A fragments were tested for their ability to inhibit HIV
20 replication.

Methods. Stable Jurkat cell lines expressing PKR-inhibiting and non-inhibiting NS5A proteins were generated, as were a series of C-terminal deletion mutants (FIG. 12). These cell lines expressed NS5A proteins of 123, 161, 181, 250, 314, and 363 amino acids (aa). All constructs had stop codons after NS5A, followed
25 by the EMC IRES and GFP. A control cell line expressing only GFP served as a negative control, and NS5A and GFP expression were regulated by tetracycline. HIV replication was measured by p24 antigen release into culture supernatant or by measuring infectivity.

Results. Jurkat cell lines demonstrated regulated expression of NS5A and deletion mutants as shown by western blot and GFP expression. The NS5A and GFP genes were shown to remain linked by RT-PCR of cellular DNA from recombinant cell lines. Expression of either PKR-inhibiting or non-inhibiting NS5A proteins resulted in HIV inhibition (> 95% reduction in p24 antigen), thus the HIV- and PKR-inhibiting functions are independent. All deletion mutants of 250 aa or larger retained HIV-inhibiting effects, whereas those with 181 aa or smaller did not (FIG. 13).

EXAMPLE 5 – HCV and GBV-B Virus NS5A Proteins Inhibit HIV Replication in Jurkat Cells

Hepatitis C virus (HCV), like GBV-C, commonly infect humans. GBV-B is a primate virus that is closely related to both HCV and GBV-C. HCV and GBV-C NS5A proteins both inhibit PKR function. To further characterize the inventors work showing inhibition of X4-tropic HIV replication, they expressed HCV and GBV-B NS5A's and a series of GBV-C NS5A deletions in Jurkat cells, and measured the effect of these proteins on HIV replication.

Methods. Jurkat cell lines stably expressing HCV, GBV-B, and GBV-C NS5A proteins were generated, as were a series of GBV-C NS5A deletion mutants. All constructs had stop codons after NS5A, followed by the EMC IRES and GFP. A control cell line expressing only GFP served as a negative control, and NS5A and GFP expression were regulated by tetracycline. Plasmids were transfected using the Amaxa nucleofection method, cells were selected for growth in hygromycin and for GFP expression. HIV replication was measured by p24 antigen release into culture supernatant or by measuring infectivity.

Results. Jurkat cell lines demonstrated regulated expression of HCV, GBV-B GBV-C NS5A proteins and GBV-C deletion mutants as determined by GFP expression and when antibodies were available, by immunoblot (FIG. 14). For constructs in which antibodies were not available, the NS5A and GFP coding sequences were linked (detected by PCR of cellular DNA). Expression of HCV, GBV-B and GBV-C NS5A proteins resulted in HIV inhibition (> 95% reduction in p24 antigen), thus the inhibitory effect appears conserved between these three flaviviruses (FIG. 15). All deletion mutants containing GBV-C NS5A amino acids

(aa) between number 152 and 237 retained HIV-inhibiting effects, whereas those with C-terminal deletions containing ≤ 152 NS5A aa's did not inhibit HIV replication (FIG. 16). The effect was reduced in NS5A containing cells grown in doxycycline (NS5A expression turned off), but not in control cells grown in doxycycline. Thus, expression of GBV-C, GBV-B, and HCV NS5A proteins resulted in inhibition of CXCR4-tropic HIV replication in Jurkat cells, and the inhibitory effect requires GBV-C aa's 152-237, coinciding with domain II of the HCV NS5A protein. The region may possibly narrowed to 152-182 (FIG. 17).

* * * * *

All of the compositions and methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents that are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.

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The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated herein by reference.

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U.S. Patent 4,554,101

U.S. Patent 4,683,202

U.S. Patent 4,833,077

U.S. Patent 4,879,236

U.S. Patent 5,582,981

U.S. Patent 5,620,896

U.S. Patent 5,650,298

U.S. Patent 5,714,153

U.S. Patent 5,871,986

U.S. Patent 5,874,563

U.S. Patent 5,925,565

U.S. Patent 5,928,906

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CLAIMS

1. A pharmaceutical composition comprising an isolated flavivirus NS5A peptide or polypeptide.
2. The composition of claim 1, comprising a flavivirus NS5A peptide.
3. The composition of claim 2, comprising multiple flavivirus NS5A peptides.
4. The composition of claim 1, comprising a flavivirus NS5A polypeptide.
5. The composition of claim 4, wherein the flavivirus NS5A polypeptide is a full length NS5A polypeptide.
6. The composition of claim 4, wherein the flavivirus NS5A polypeptide is a fusion polypeptide.
7. The composition of claim 1, wherein the flavivirus NS5A peptide or polypeptide further comprises a targeting signal.
8. The composition of claim 7, wherein the targeting signal is a nuclear targeting signal.
9. The composition of claim 7, wherein the targeting signal targets a cell surface receptor.
10. The composition of claim 9, wherein the cell surface receptor is the CD4 receptor.
11. The composition of claim 1, wherein the flavivirus NS5A peptide or polypeptide is formulated in a lipid vehicle.
12. The composition of claim 11, wherein the lipid vehicle is a liposome.
13. The composition of claim 1, wherein said flavivirus NS5A peptide or polypeptide is formulated with an amphipathic peptide, an insect peptide, or pyrrhocoricin.

14. The composition of claim 1, wherein the flavivirus is selected from the group consisting of DEN4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, GBV-C, HGV, HCV2a, HCV3a, HCV2b, HCV1a and HCV1b.
15. The composition of claim 14, wherein the flavivirus is a GBV-C virus.
16. The composition of claim 3, wherein the multiple peptides are from the same NS5A polypeptide.
17. The composition of claim 3, wherein the multiple peptides are from different NS5A polypeptides.
18. The composition of claim 1, wherein the flavivirus NS5A peptide or polypeptide comprises residues 152-237 of GBV-C NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins.
19. The composition of claim 1, wherein the flavivirus NS5A peptide or polypeptide comprises domain II of HCV NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins.
20. The composition of claim 1, wherein the flavivirus NS5A peptide or polypeptide is from a IFN-sensitive flavivirus.
21. The composition of claim 1, wherein the flavivirus NS5A peptide or polypeptide is from a IFN-resistant flavivirus.
22. The composition of claim 20, wherein the IFN-sensitive flavivirus is a GBV-C virus.
23. The composition of claim 21, wherein the IFN-resistant flavivirus is a GBV-C virus.
24. A method for preventing or treating HIV infection comprising administering to a subject a composition comprising a flavivirus NS5A peptide or polypeptide.
25. The method of claim 24, wherein flavivirus NS5A peptide or polypeptide comprises a flavivirus NS5A peptide.

26. The method of claim 25, wherein the flavivirus NS5A peptide or polypeptide comprises multiple flavivirus NS5A peptides.
27. The method of claim 24, wherein the flavivirus NS5A peptide or polypeptide comprises a flavivirus NS5A polypeptide.
28. The method of claim 27, wherein the flavivirus NS5A polypeptide is a full length NS5A polypeptide.
29. The method of claim 24, wherein the flavivirus NS5A polypeptide is a fusion polypeptide.
30. The composition of claim 24, wherein the flavivirus NS5A peptide or polypeptide further comprises a targeting signal.
31. The method of claim 30, wherein the targeting signal is a nuclear targeting signal.
32. The method of claim 30, wherein the targeting signal targets a cell surface receptor.
33. The method of claim 32 wherein the cell surface receptor is the CD4 receptor.
34. The method of claim 24, wherein the flavivirus NS5A peptide or polypeptide is formulated in a lipid vehicle.
35. The method of claim 34, wherein the lipid vehicle is a liposome.
36. The method of claim 24, wherein the flavivirus NS5A peptide or polypeptide is formulated with an amphipathic peptide, an insect peptide, or pyrrolic acid.
37. The method of claim 24, wherein the flavivirus is selected from the group consisting of DEN4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, GBV-C, HGV, HCV2a, HCV3a, HCV2b, HCV1a and HCV1b.
38. The method of claim 37, wherein the flavivirus is GBV-C.
39. The method of claim 26, wherein the multiple peptides are from the same NS5A polypeptide.

40. The method of claim 26, wherein the multiple peptides are from different NS5A polypeptides.
41. The method of claim 27, wherein the flavivirus NS5A peptide or polypeptide comprises residues 152-237 of GBV-C NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins.
42. The method of claim 24, wherein the flavivirus NS5A peptide or polypeptide comprises domain II of HCV NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins.
43. The method of claim 24, wherein the flavivirus NS5A peptide or polypeptide is from a IFN-sensitive flavivirus.
44. The method of claim 24, wherein the flavivirus NS5A peptide or polypeptide is from a IFN-resistant flavivirus.
45. The method of claim 43, wherein the IFN-sensitive flavivirus is a GBV-C virus.
46. The method of claim 44, wherein the IFN-resistant flavivirus is a GBV-C virus.
47. The method of claim 24, further comprising administration of at least a second anti-HIV therapy.
48. The method of claim 47, wherein the second anti-HIV therapy is administered before said flavivirus NS5A peptide or polypeptide.
49. The method of claim 48, wherein the second anti-HIV therapy is administered after said flavivirus NS5A peptide or polypeptide.
50. The method of claim 47, wherein the second anti-HIV therapy is HAART therapy.
51. The method of claim 47, wherein the second anti-HIV therapy is AZT therapy.
52. The method of claim 24, wherein the composition is administered at least twice.

53. A method of reducing HIV replication in an HIV-infected cell comprising contacting said cell with a composition comprising a flavivirus NS5A peptide or polypeptide.
54. The method of claim 53, wherein the cell is a T lymphocyte.
55. A method of inhibiting HIV infection of a cell comprising contacting said cell with a composition comprising a flavivirus NS5A peptide or polypeptide.
56. The method of claim 55, wherein the cell is a T lymphocyte.
57. A method for preventing or treating HIV infection comprising administering to a subject a composition comprising an expression construct encoding a flavivirus NS5A peptide or polypeptide.
58. The method of claim 57, wherein the expression construct is a viral expression construct.
59. The method of claim 58, wherein the expression construct is an adenovirus, a retrovirus, a lentivirus, an adeno-associated virus, a polyoma virus, a herpesvirus, or a pox virus.
60. The method of claim 58, wherein the expression construct is a non-viral expression construct.
61. The method of claim 60, wherein the expression construct is dispersed in a lipid vehicle.
62. The method of claim 57, wherein the expression construct encodes an NS5A polypeptide.
63. The method of claim 57, wherein the expression construct encodes an NS5A peptide.
64. The method of claim 62, wherein the flavivirus NS5A polypeptide is a full length NS5A polypeptide.
65. The method of claim 62, wherein the flavivirus NS5A polypeptide is a fusion polypeptide.

66. The composition of claim 65, wherein the flavivirus NS5A peptide or polypeptide further comprises a targeting signal.
67. The method of claim 66, wherein the targeting signal is a nuclear targeting signal.
68. The method of claim 66, wherein the targeting signal targets a cell surface receptor.
69. The method of claim 68, wherein the cell surface receptor is the CD4 receptor.
70. The method of claim 57, wherein the flavivirus NS5A peptide or polypeptide comprises residues 152-237 of GBV-C NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins.
71. The method of claim 57, wherein the flavivirus NS5A peptide or polypeptide comprises domain II of HCV NS5A, or the corresponding sequences thereto from other flavivirus NS5A proteins.
72. The method of claim 57, wherein the flavivirus is selected from the group consisting of DEN4, YFV, TBEV, WNV, CSFV, BVDV, GBV-A, GBV-B, GBV-C, HGV, HCV2a, HCV3a, HCV2b, HCV1a and HCV1b.

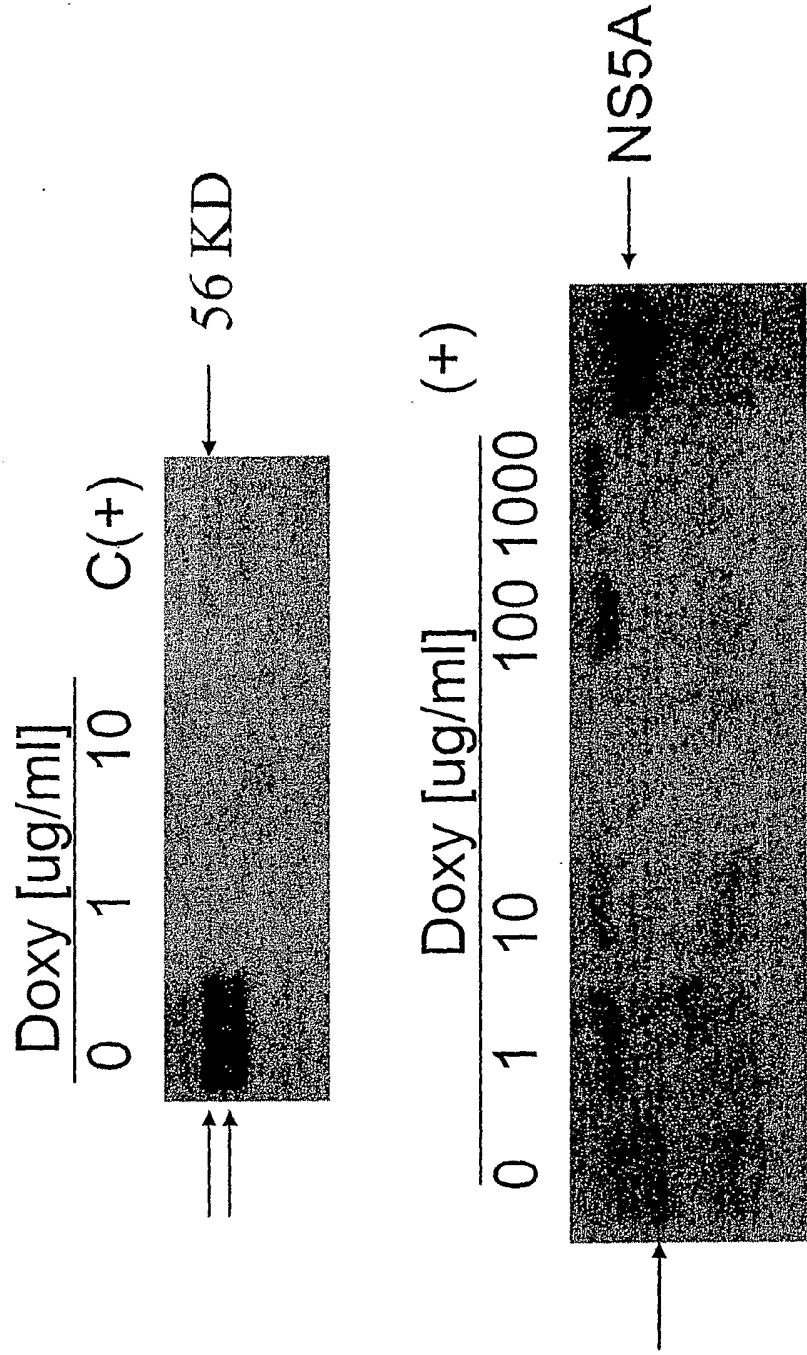


FIG. 1

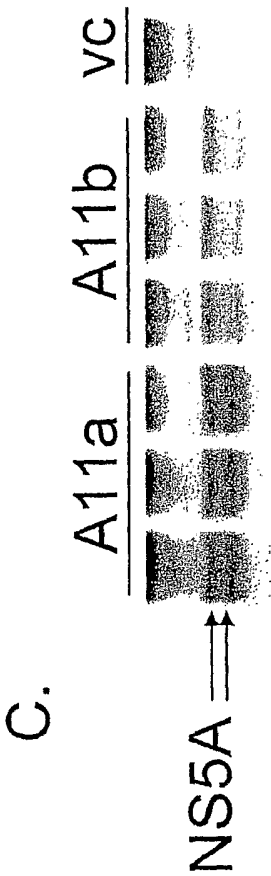
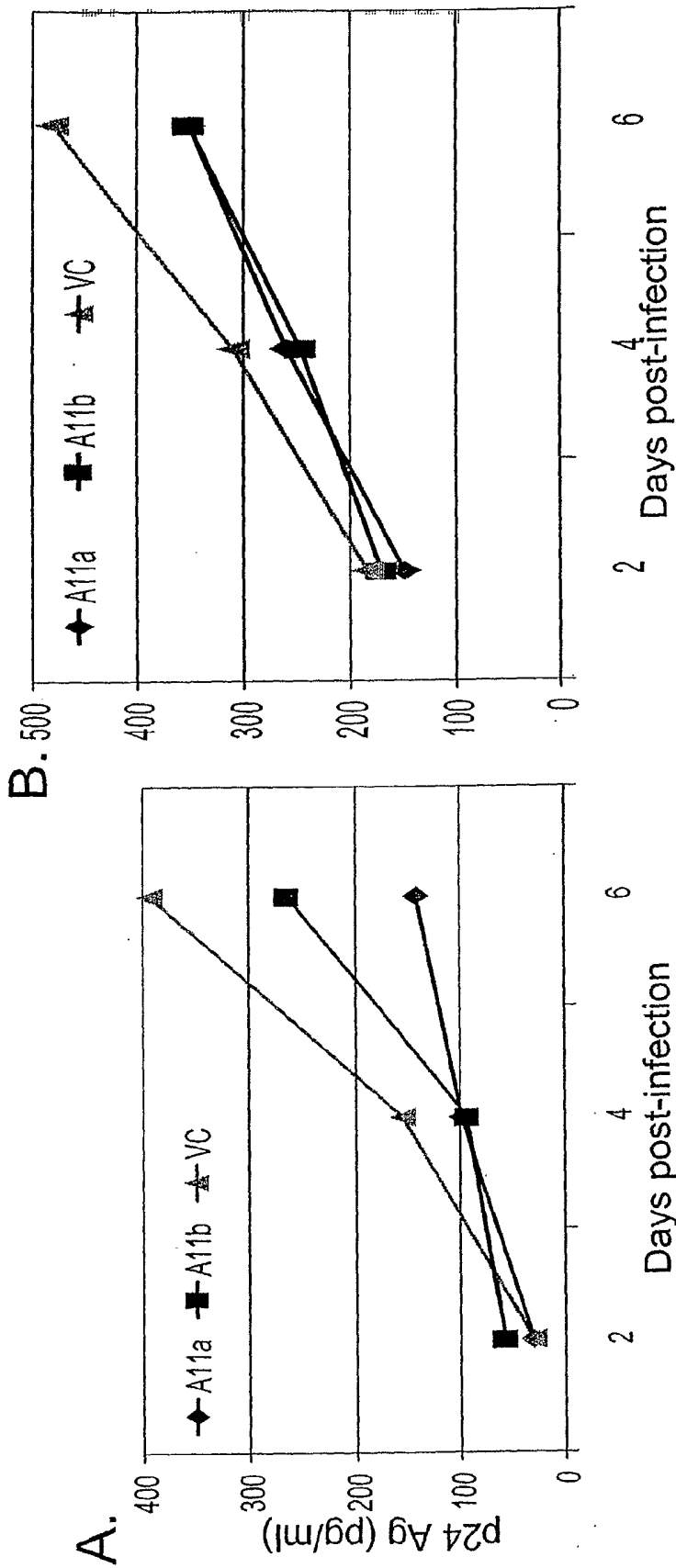


FIG. 2A-C

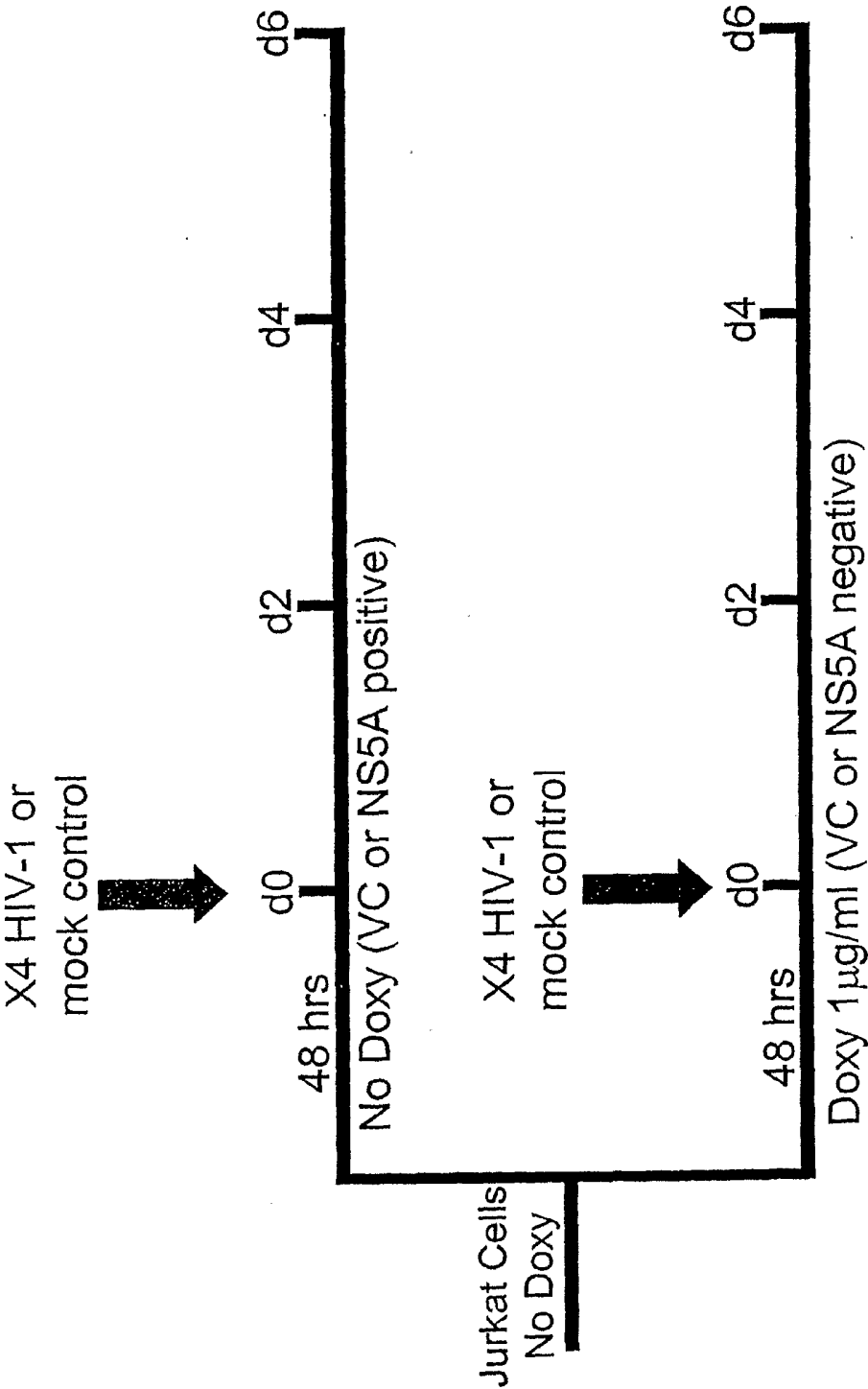


FIG. 3A

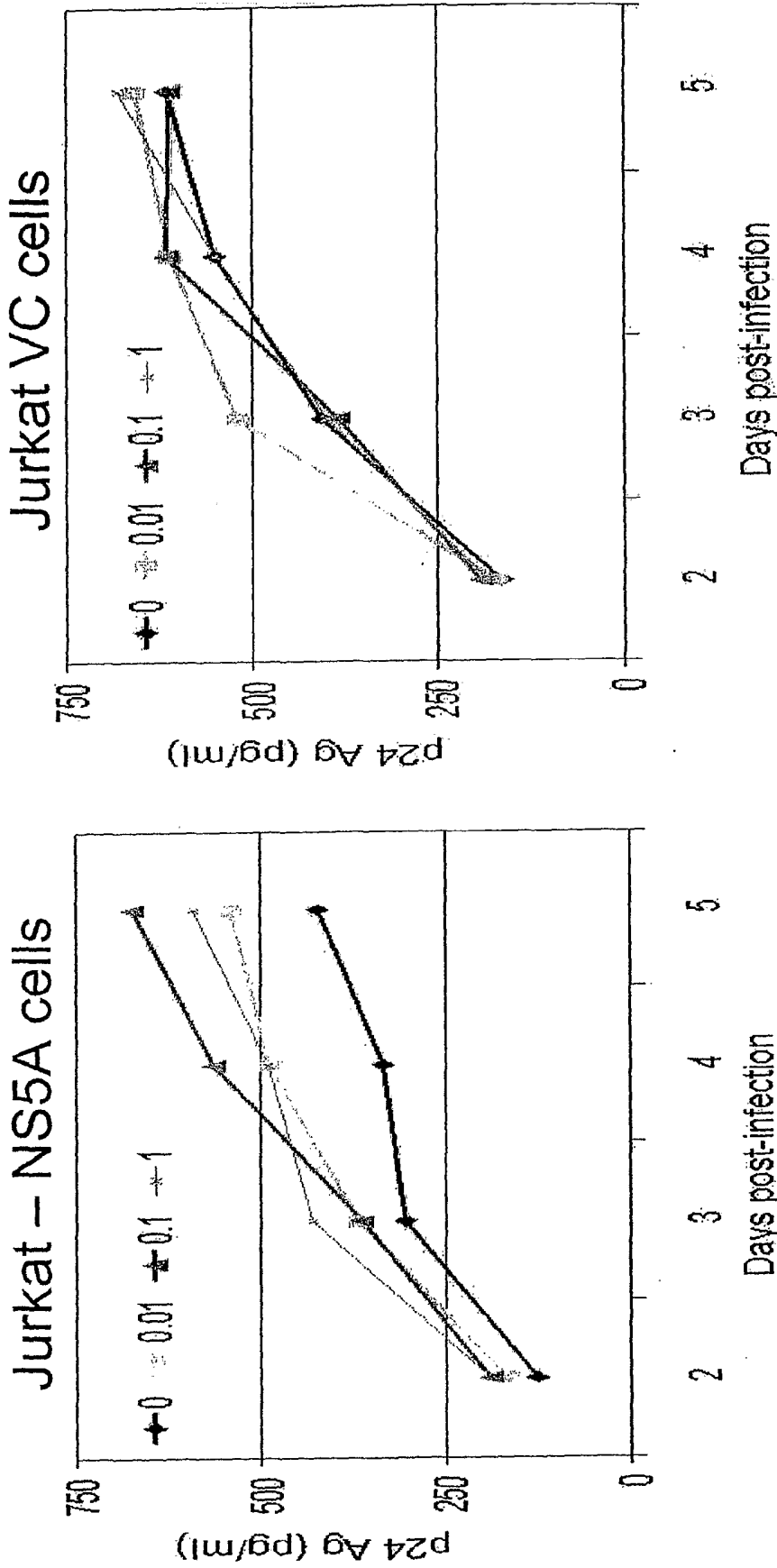


FIG. 3B

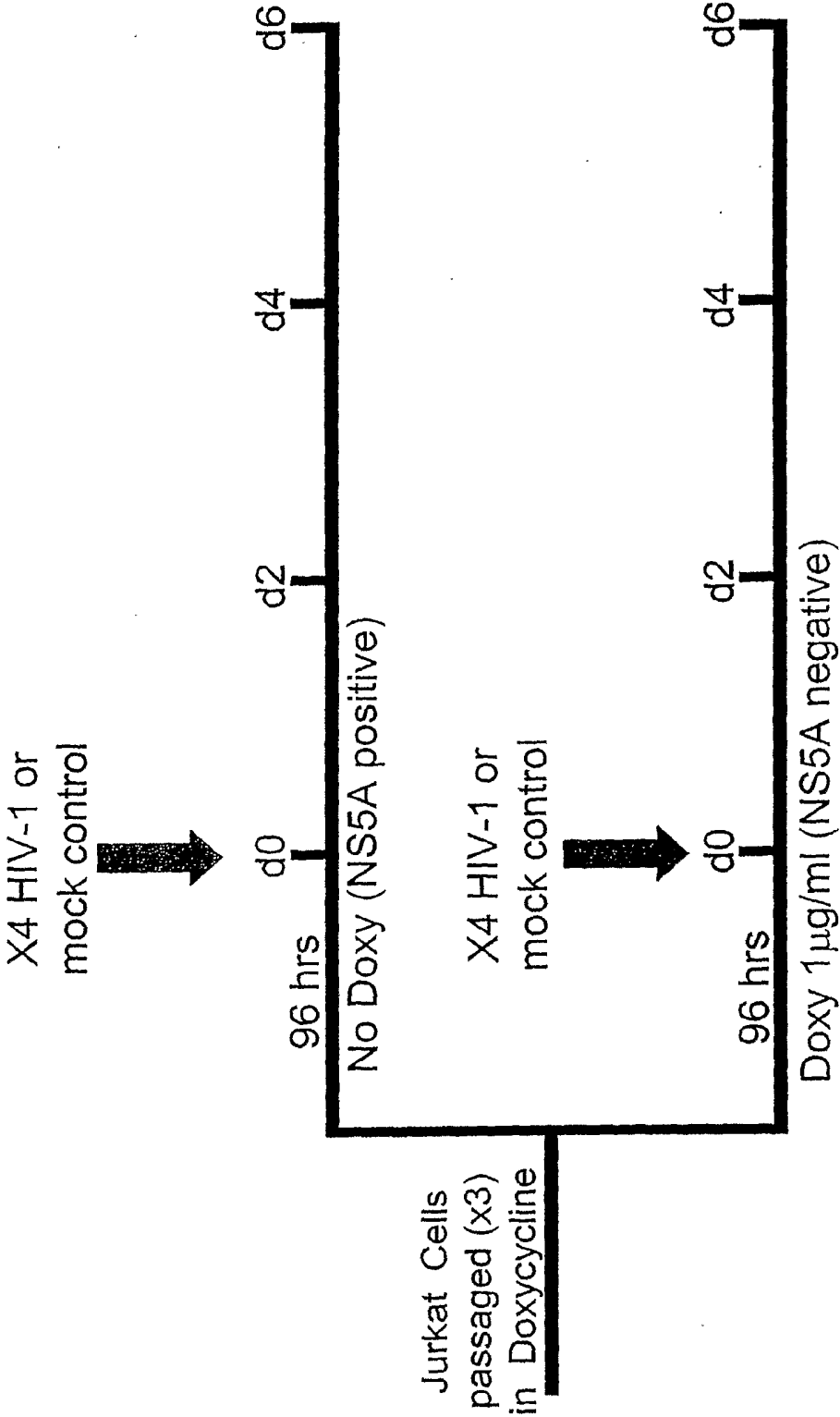


FIG. 4A

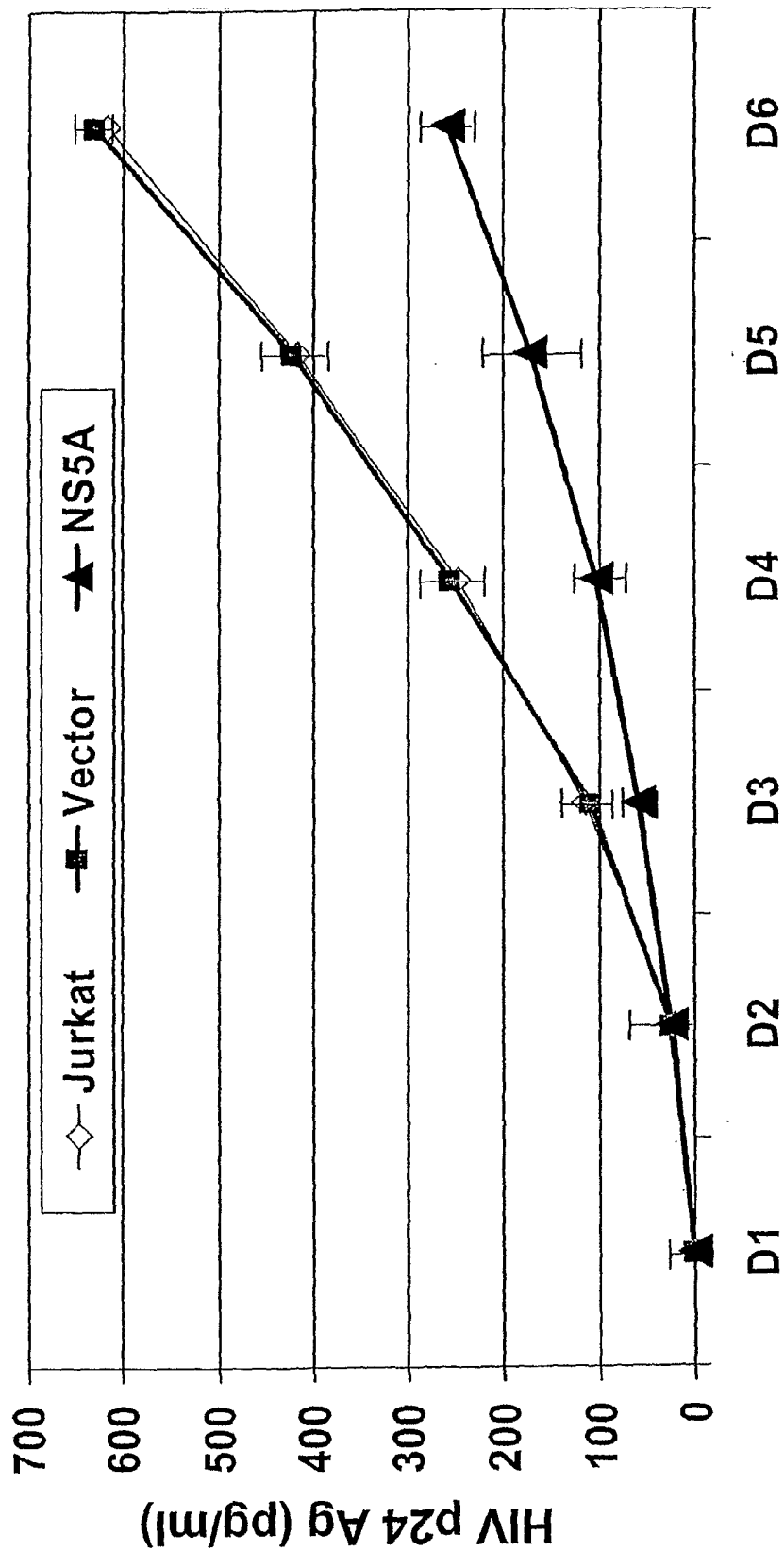


FIG. 4B

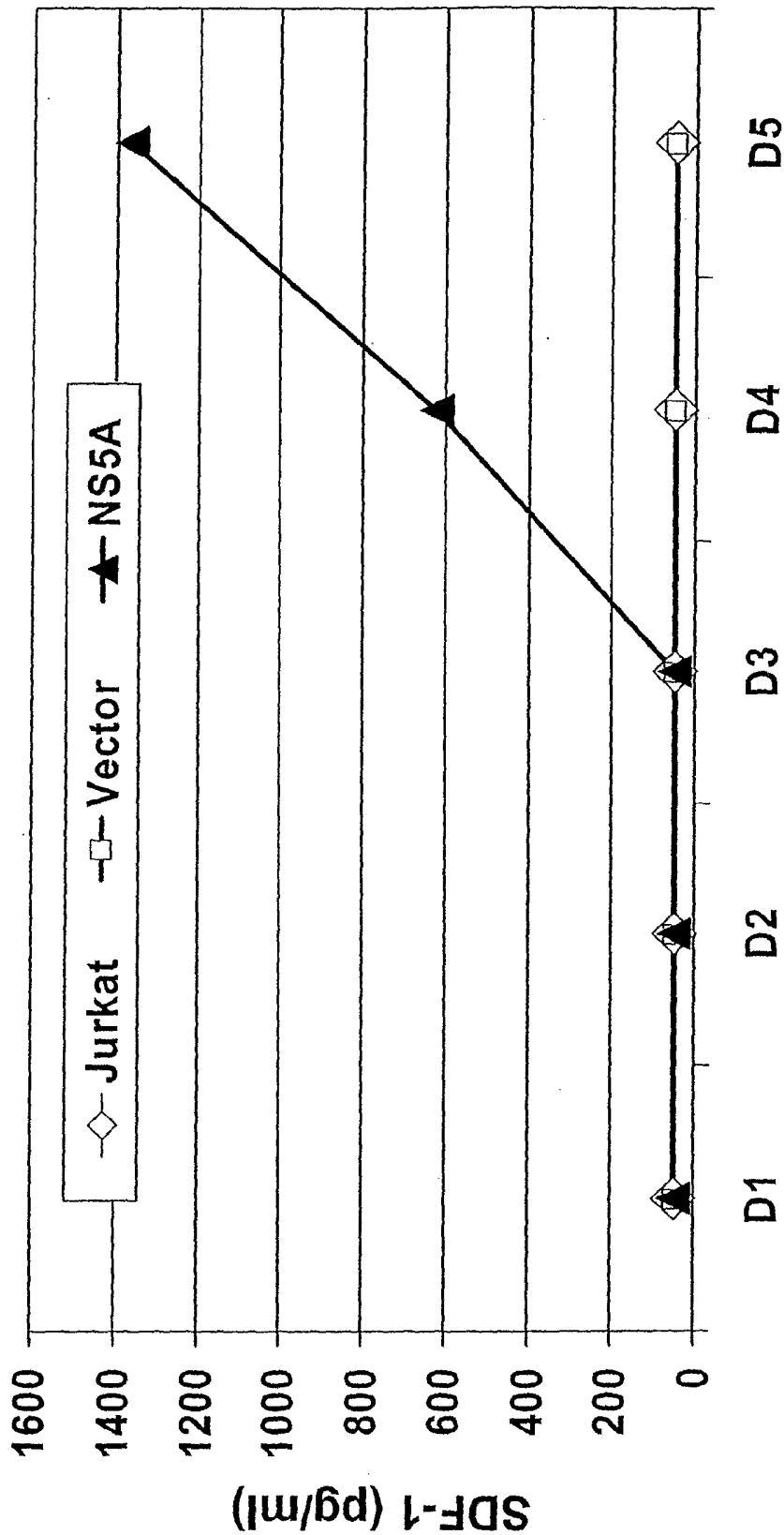


FIG. 5

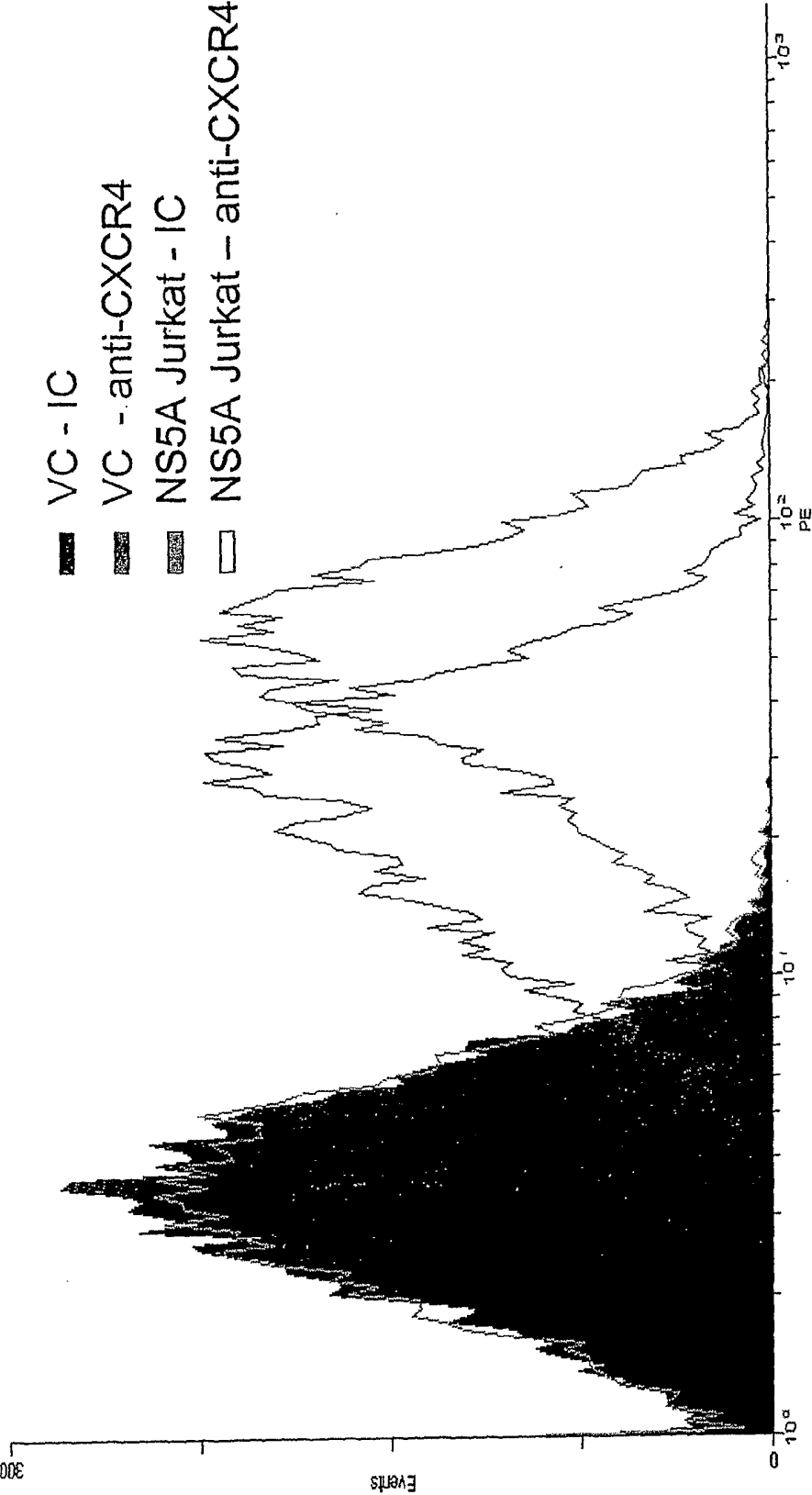


FIG. 6

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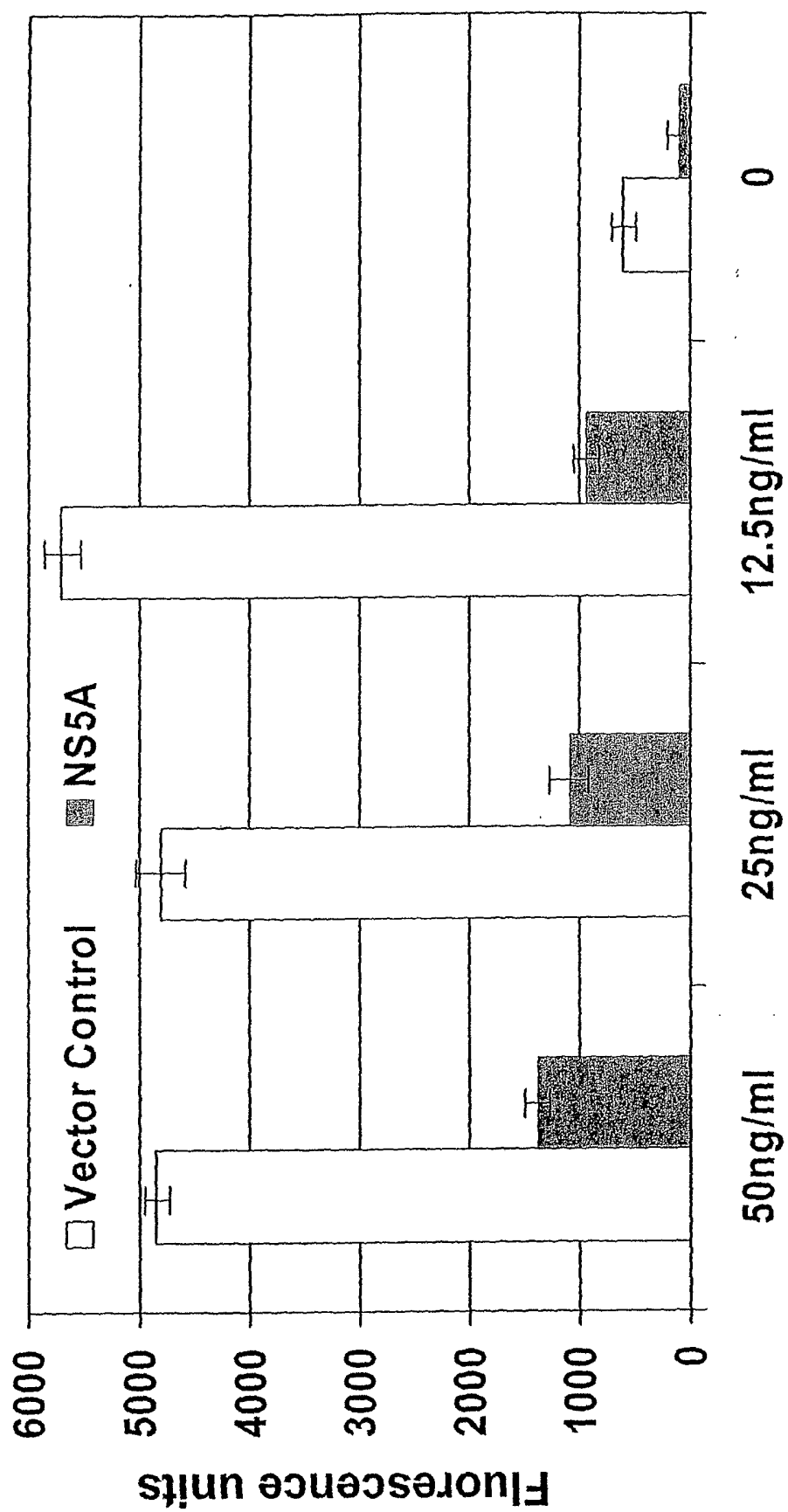


FIG. 7

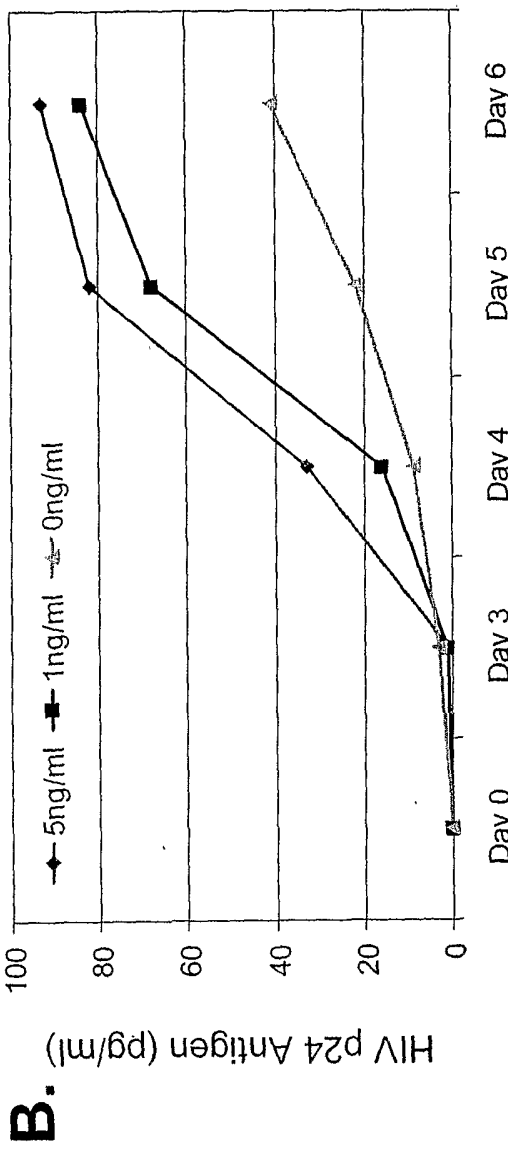
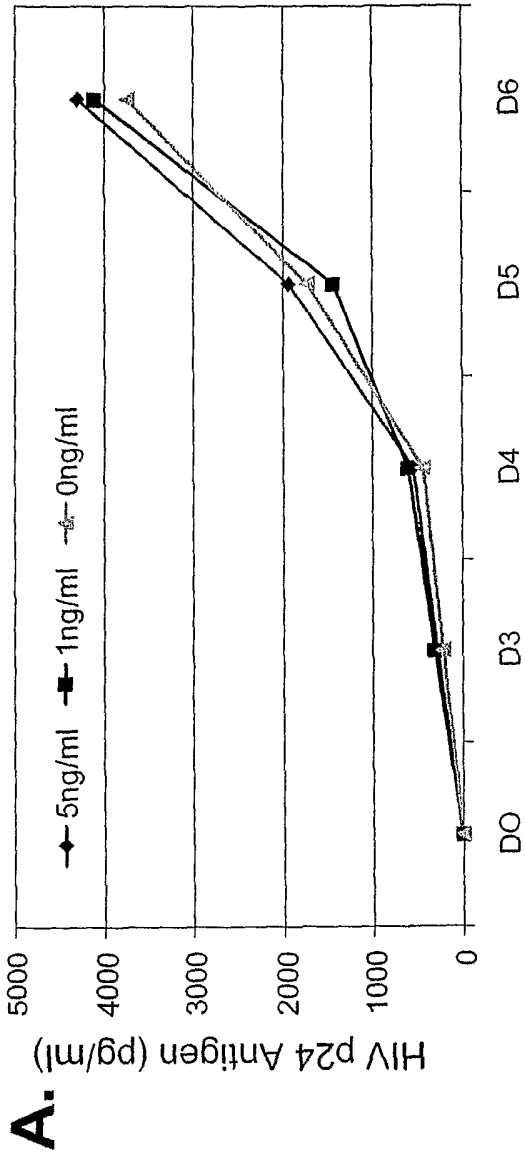


FIG. 8A-B

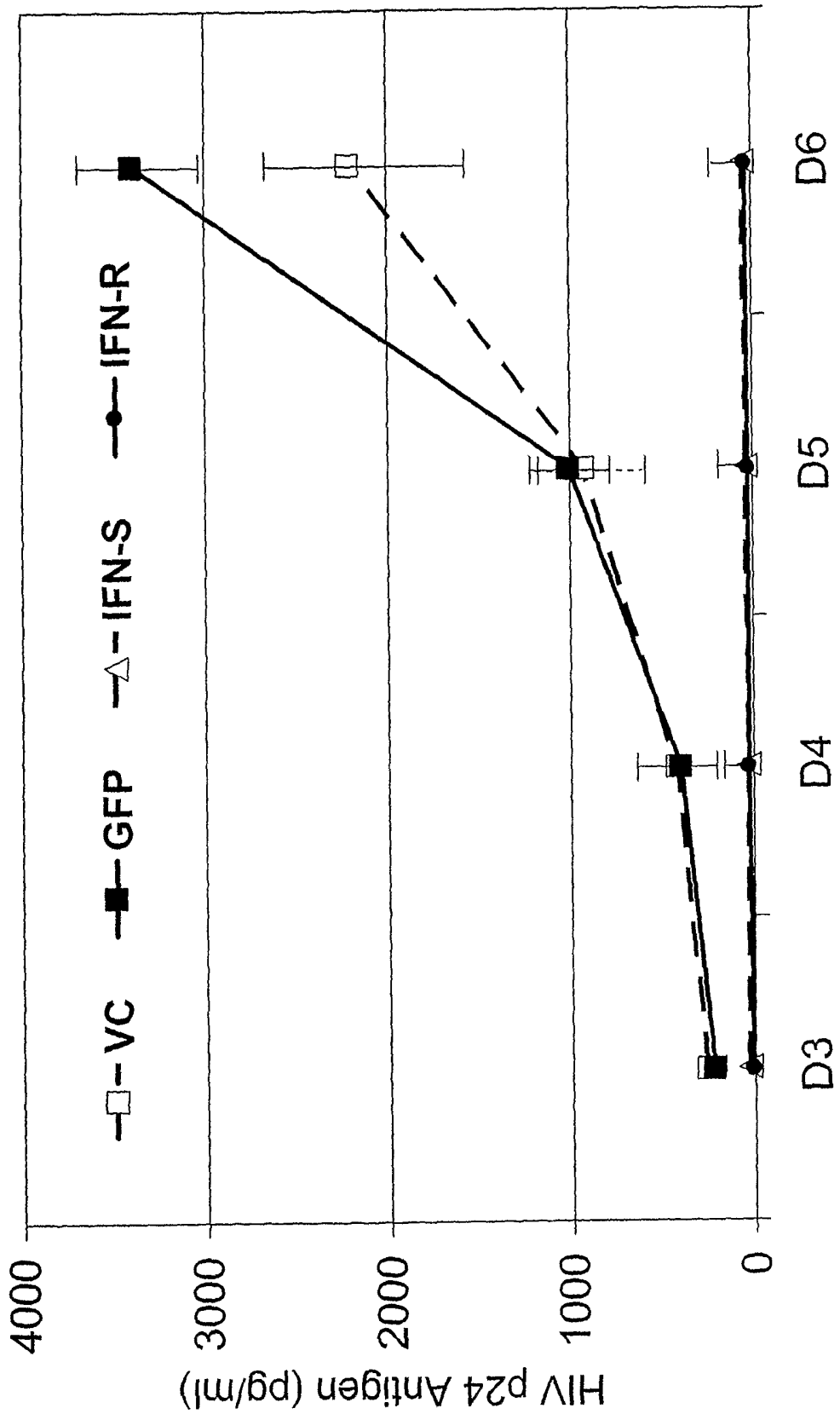


FIG. 9

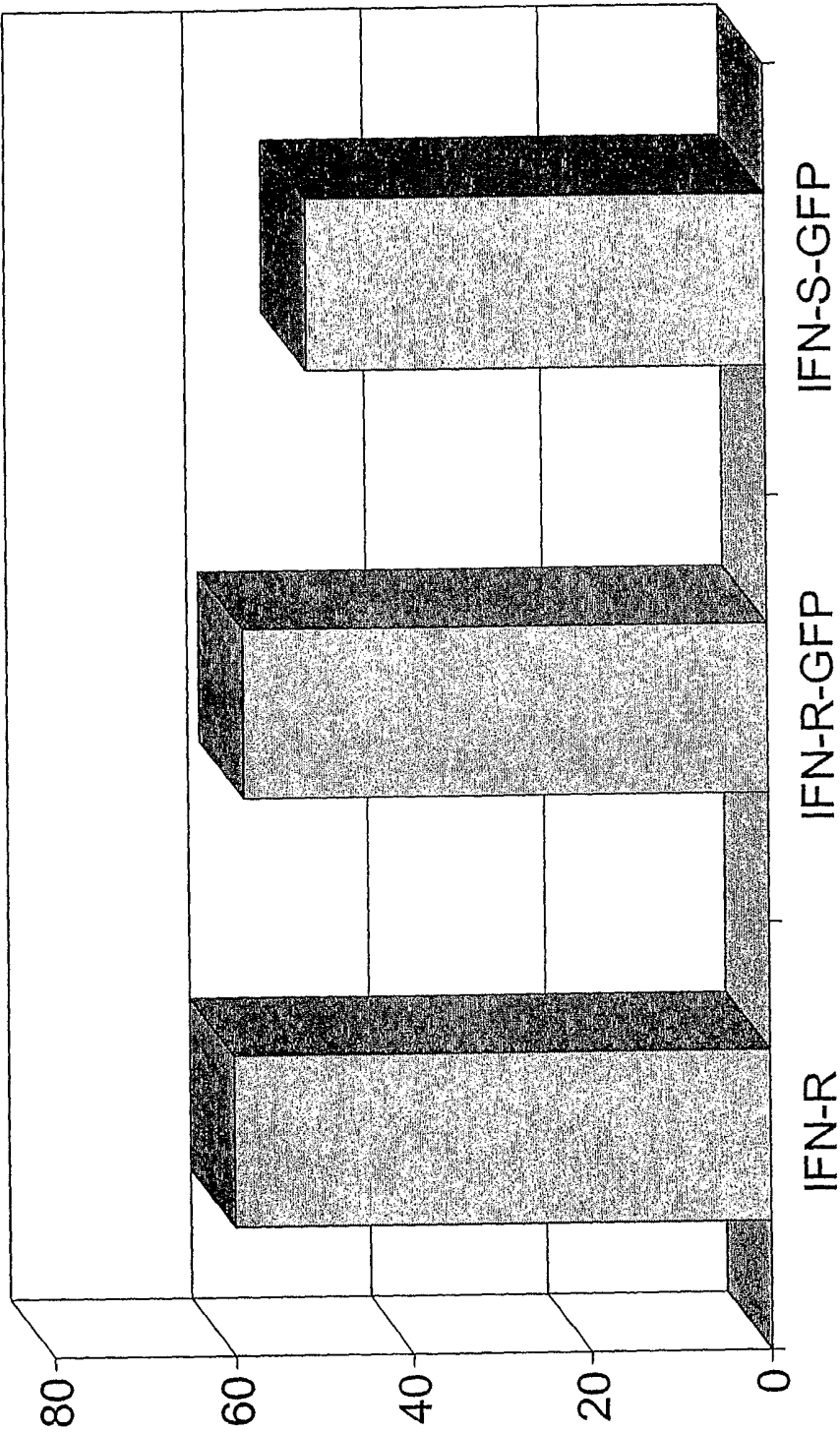


FIG. 10

NS5A Inhibits HIV MN (X4) Clade B

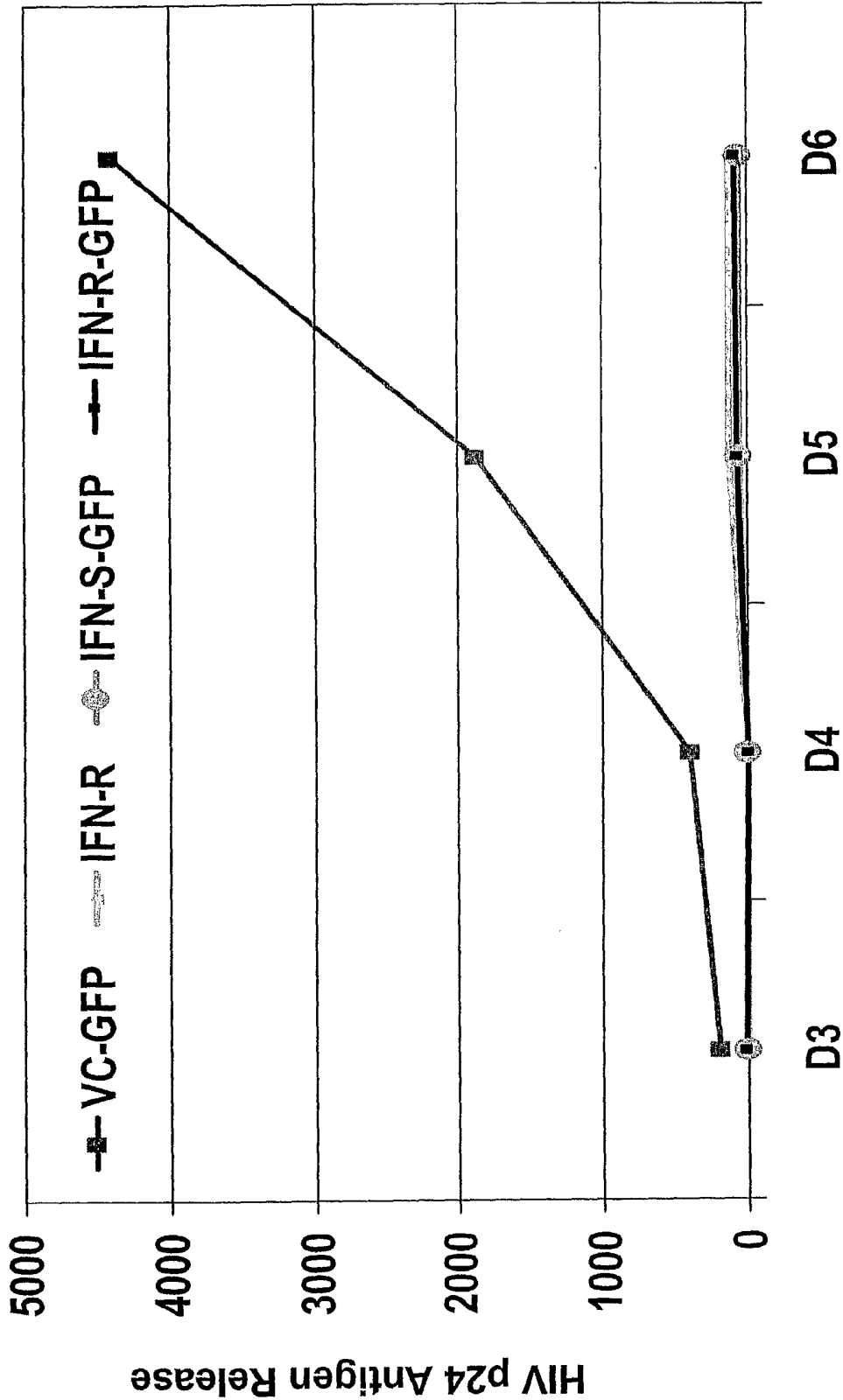


FIG. 11A

NS5A Inhibits HIV JF(X4) Clade B

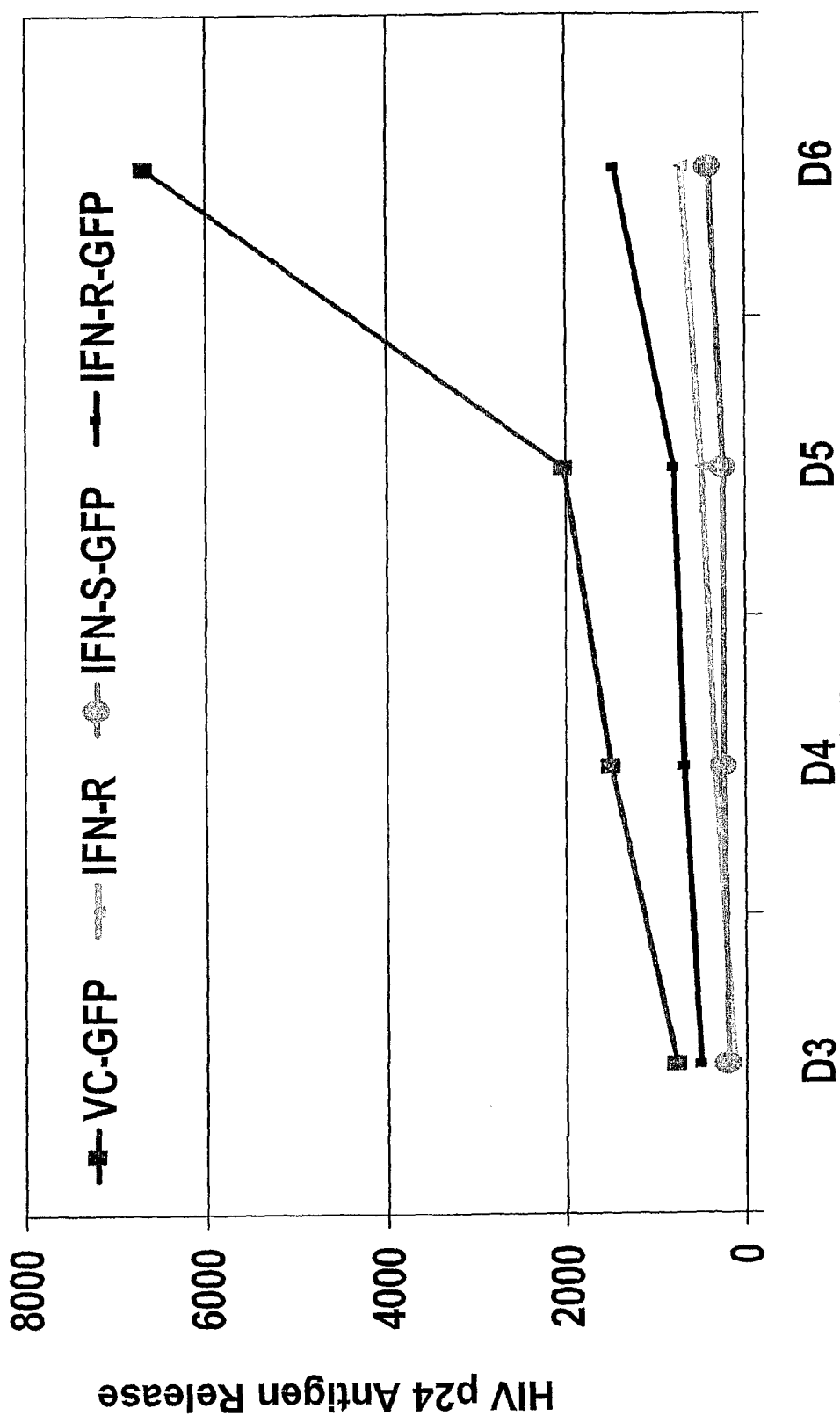


FIG. 11B

NS5A Inhibits HIV ELI(X4) Clade D

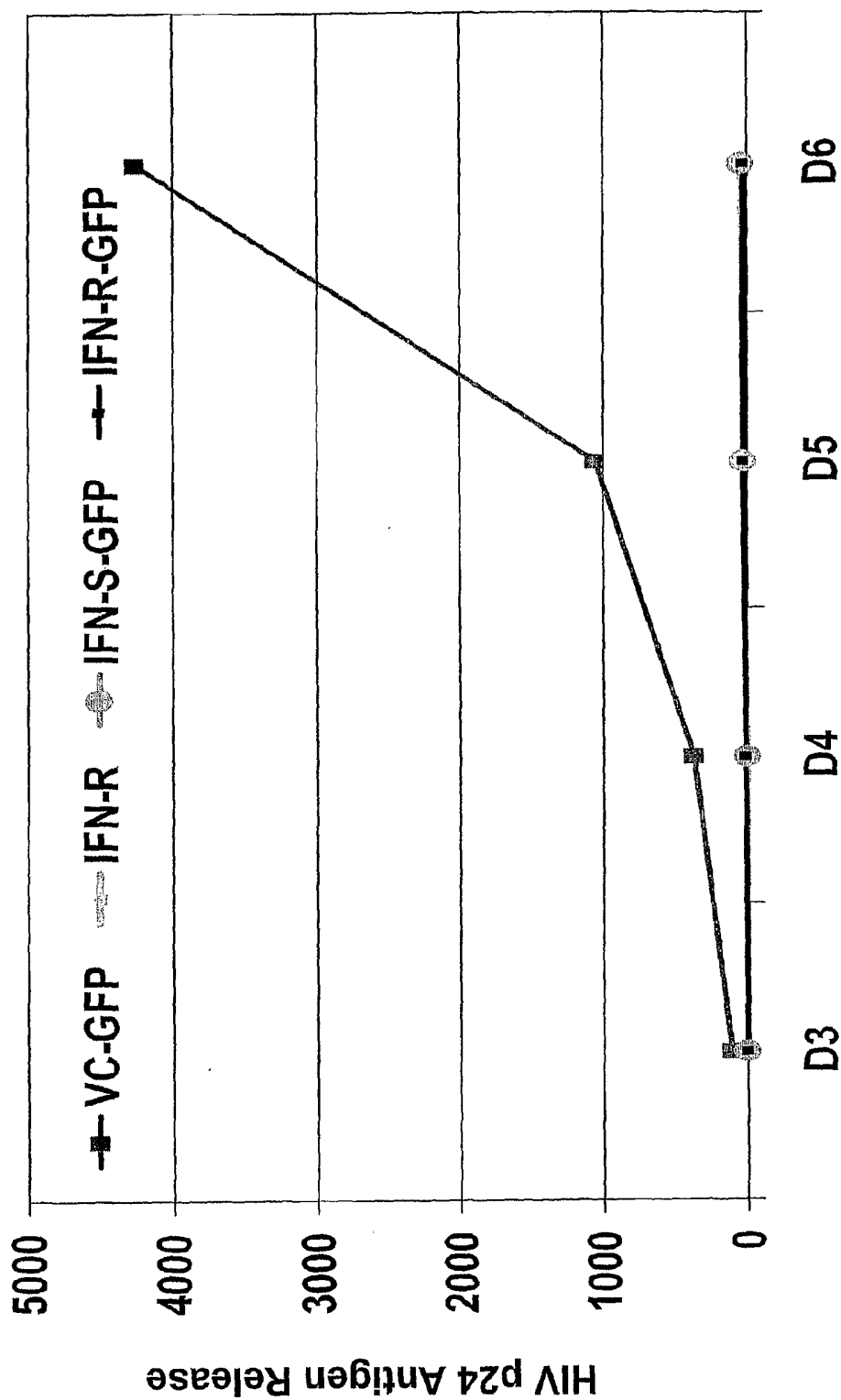


FIG. 11C

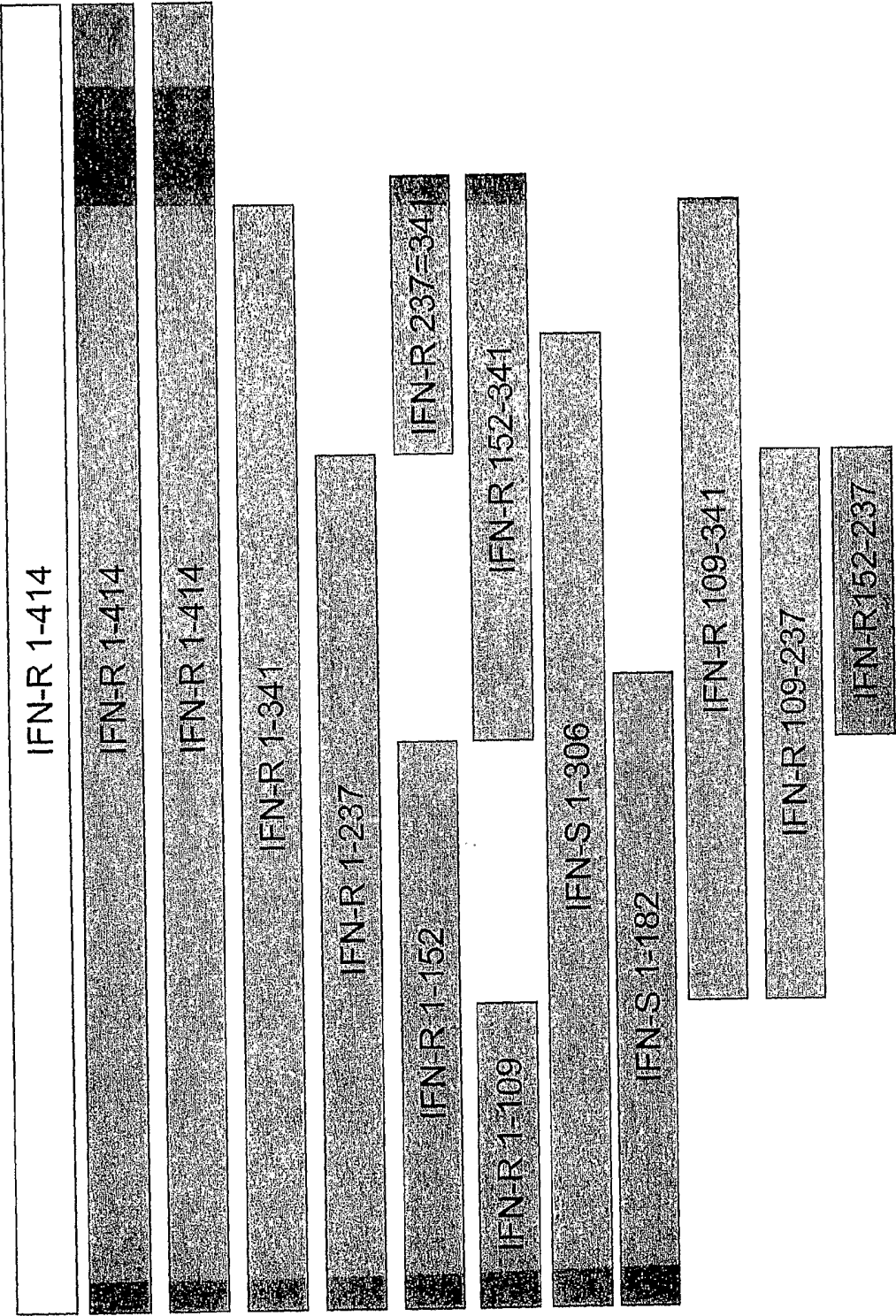


FIG. 12

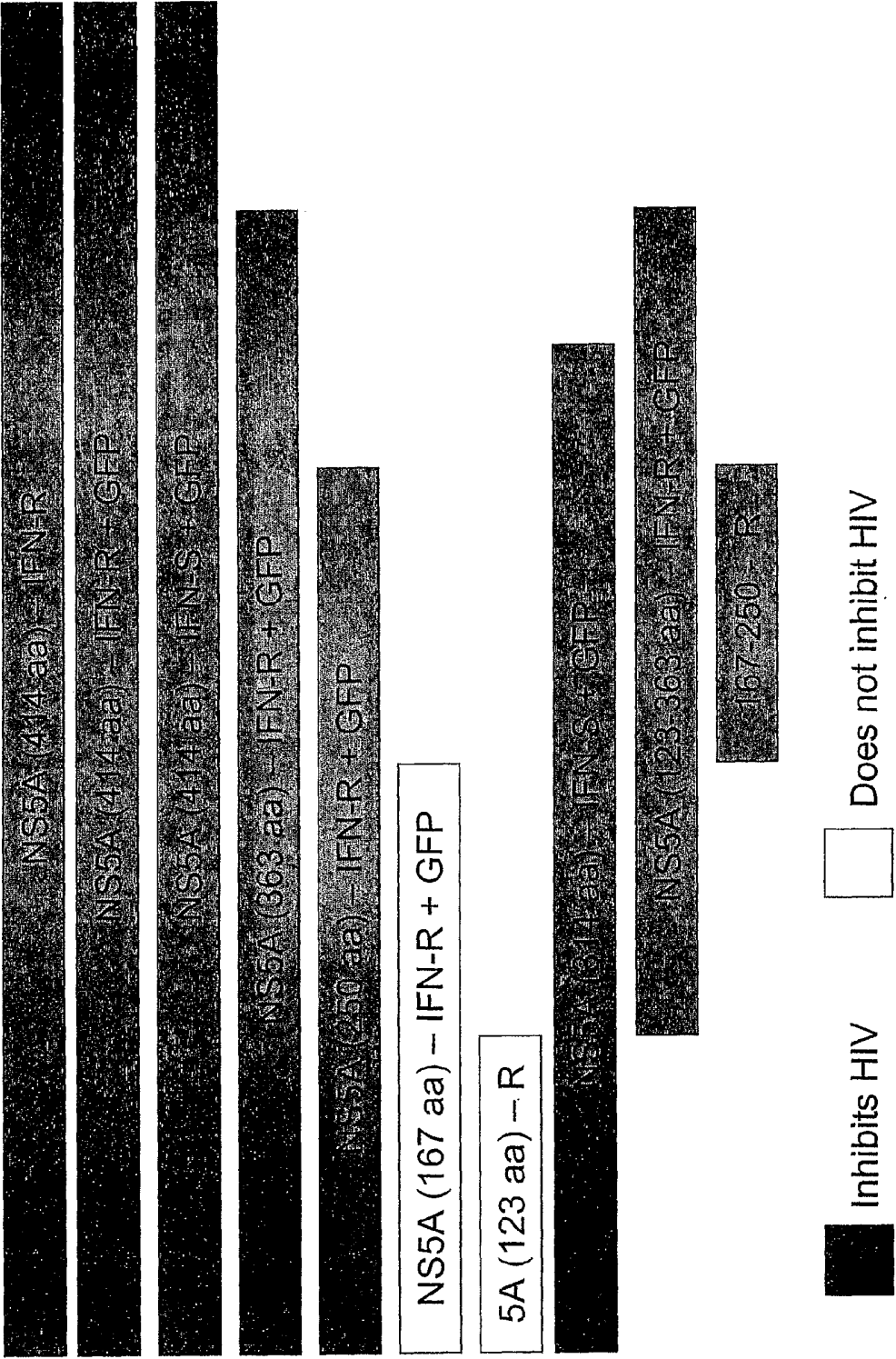


FIG. 13

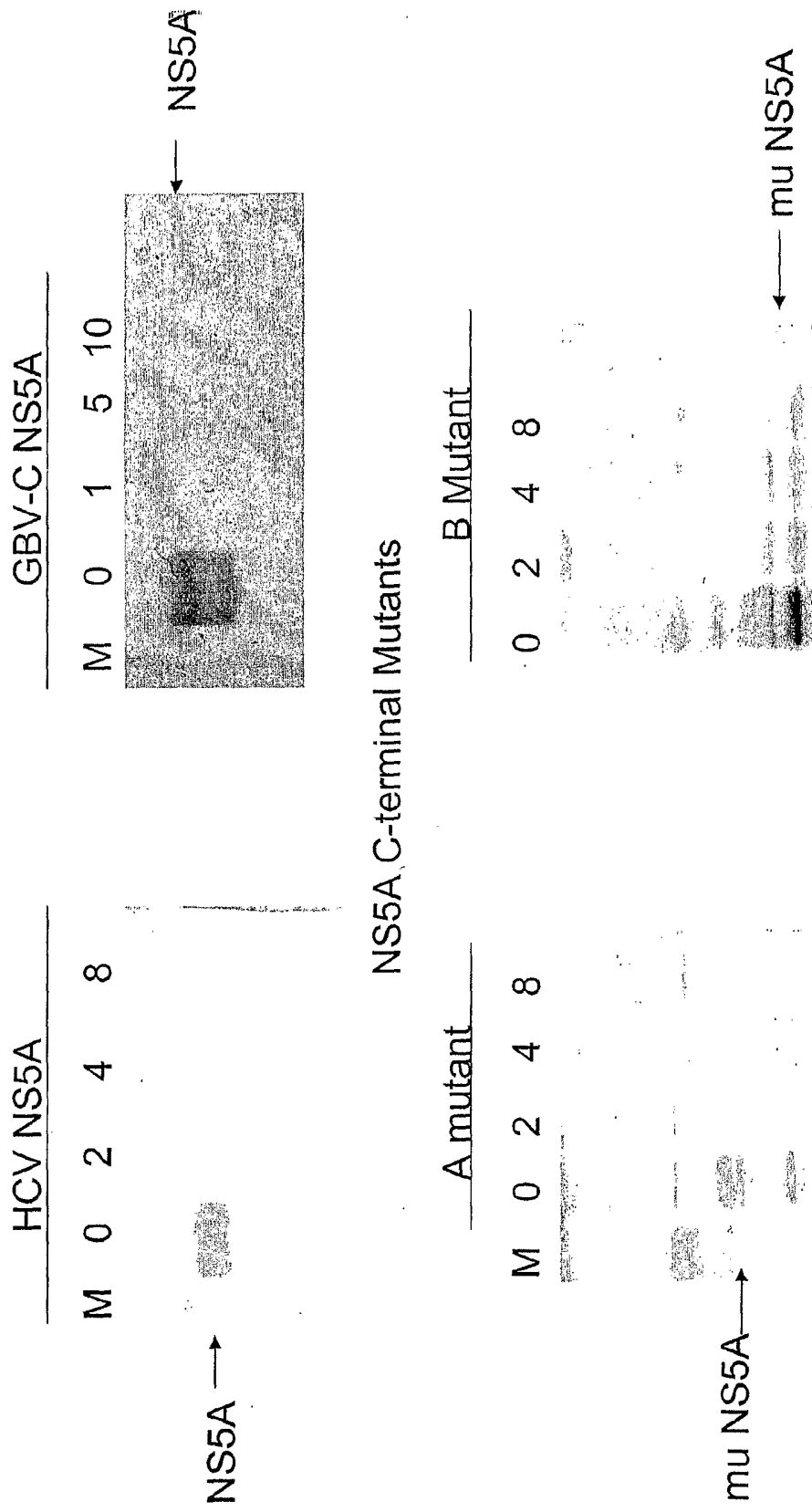


FIG. 14

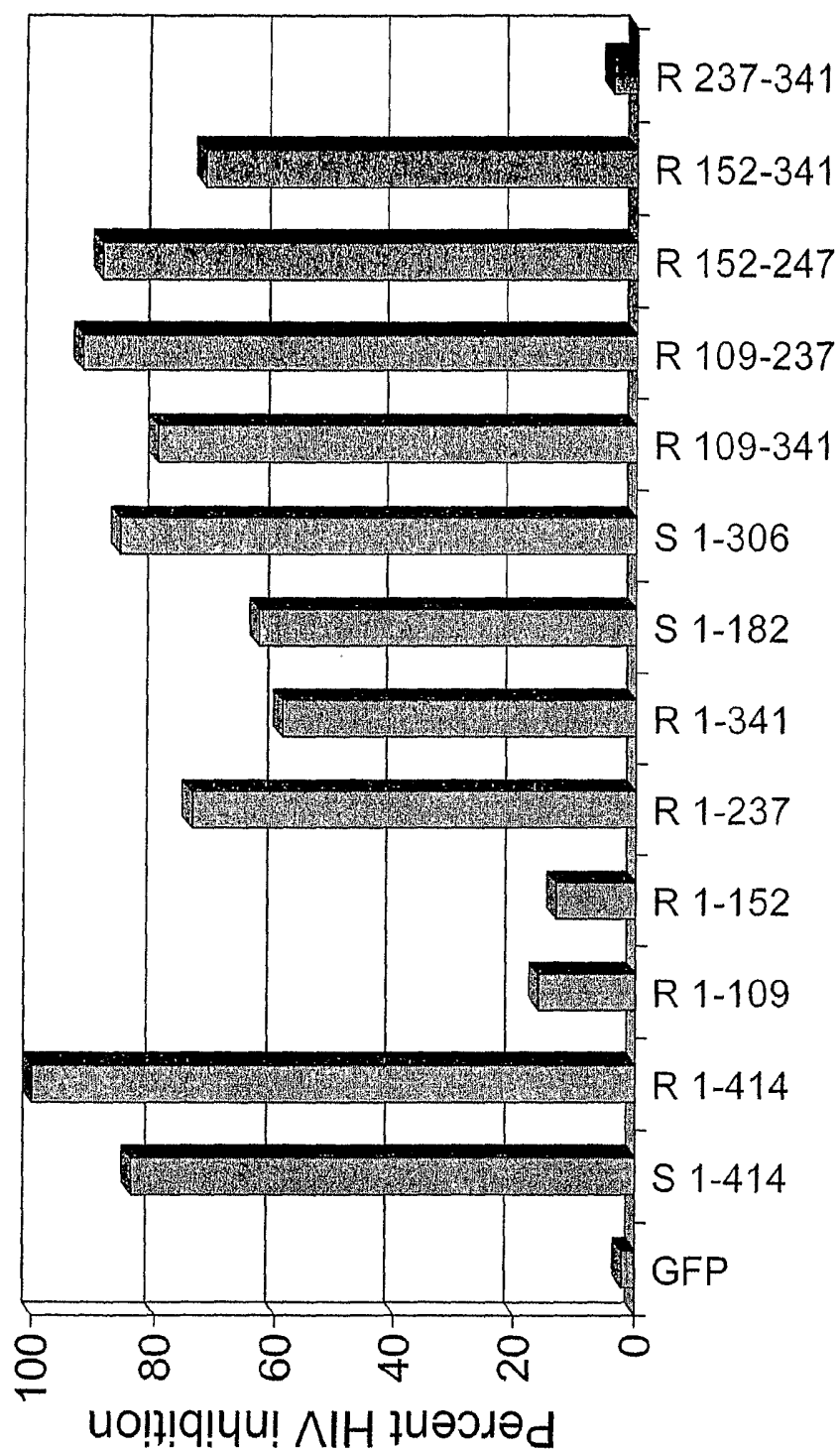


FIG. 15



FIG. 16

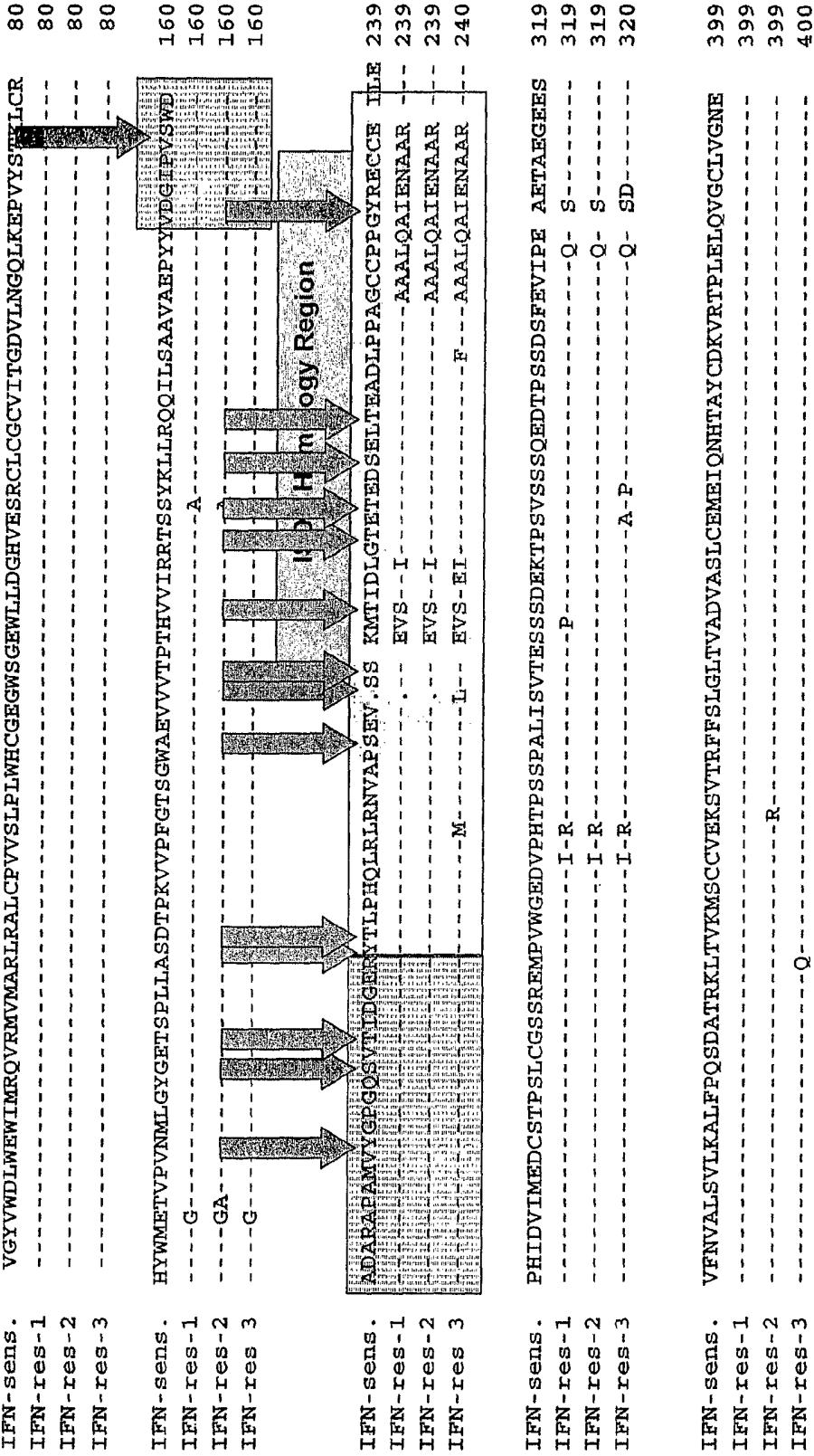


FIG. 17

SEQUENCE LISTING

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 XIANG, JINHUA
 CHANG, QING
 MCLINDEN, JAMES

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 Ala Arg Val Val Glu Cys Cys Val Met Ala Gly Glu Lys Ala Thr Thr
 790 795 800

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 His Met Gly Ser Phe Ser Arg Ala Val Lys Glu Arg Leu Leu Glu Trp
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 Ile Arg Asp Ala Ala Arg Thr Leu Ser Cys Gly Gln Cys Val Met Gly
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 950 955 960

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 6296
 Ser Gly Glu Trp Leu Leu Asp Gly His Val Glu Ser Arg Cys Leu Cys
 1910 1915 1920

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12

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    65          70          75          80
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    85          90          95
Val Ala Gly Ile Leu Gly Leu Gly Glu Val Tyr Ser Gly Val Leu Thr
    100         105         110
Val Gly Val Ala Leu Thr Arg Arg Val Tyr Pro Val Pro Asn Leu Thr
    115         120         125
Cys Ala Val Glu Cys Glu Leu Lys Trp Glu Ser Glu Phe Trp Arg Trp
    130         135         140
Thr Glu Gln Leu Ala Ser Asn Tyr Trp Ile Leu Glu Tyr Leu Trp Lys
    145         150         155         160
Val Pro Phe Asp Phe Trp Arg Gly Val Met Ser Leu Thr Pro Leu Leu
    165         170         175
Val Cys Val Ala Ala Leu Leu Leu Leu Glu Gln Arg Ile Val Met Val
    180         185         190
Phe Leu Leu Val Thr Met Ala Gly Met Ser Gln Gly Ala Pro Ala Ser
    195         200         205
Val Leu Gly Ser Arg Pro Phe Glu Ala Gly Leu Thr Trp Gln Ser Cys
    210         215         220
Ser Cys Arg Ser Asn Gly Ser Arg Val Pro Thr Gly Glu Arg Val Trp
    225         230         235         240
Glu Arg Gly Asn Val Thr Leu Leu Cys Asp Cys Pro Asn Gly Pro Trp
    245         250         255
Val Trp Val Pro Ala Leu Cys Gln Ala Ile Gly Trp Gly Asp Pro Ile
    260         265         270
Thr His Trp Ser His Gly Gln Asn Gln Trp Pro Leu Ser Cys Pro Gln
    275         280         285
Phe Val Tyr Gly Ala Val Ser Val Thr Cys Val Trp Gly Ser Val Ser
    290         295         300
Trp Phe Ala Ser Thr Gly Gly Arg Asp Ser Lys Val Asp Val Trp Ser
    305         310         315         320
Leu Val Pro Val Gly Ser Ala Ser Cys Thr Ile Ala Ala Leu Gly Ser
    325         330         335
Ser Asp Arg Asp Thr Val Val Glu Leu Ser Glu Trp Gly Ile Pro Cys
    340         345         350
Ala Thr Cys Ile Leu Asp Arg Arg Pro Ala Ser Cys Gly Thr Cys Val
    355         360         365
Arg Asp Cys Trp Pro Glu Thr Gly Ser Val Arg Phe Pro Phe His Arg
    370         375         380
Cys Gly Ala Gly Pro Arg Leu Thr Arg Asp Leu Glu Ala Val Pro Phe
    385         390         395         400
Val Asn Arg Thr Thr Pro Phe Thr Ile Arg Gly Pro Leu Gly Asn Gln
    405         410         415
Gly Arg Gly Asn Pro Val Arg Ser Pro Leu Gly Phe Gly Ser Tyr Thr
    420         425         430
Met Thr Lys Ile Arg Asp Ser Leu His Leu Val Lys Cys Pro Thr Pro
    435         440         445
Ala Ile Glu Pro Pro Thr Gly Thr Phe Gly Phe Phe Pro Gly Val Pro
    450         455         460
Pro Leu Asn Asn Cys Met Leu Leu Gly Thr Glu Val Ser Glu Val Leu
    465         470         475         480
Gly Gly Ala Gly Leu Thr Gly Gly Phe Tyr Glu Pro Leu Val Arg Arg
    485         490         495
Cys Ser Glu Leu Met Gly Arg Arg Asn Pro Val Cys Pro Gly Phe Ala
    500         505         510
Trp Leu Ser Ser Gly Arg Pro Asp Gly Phe Ile His Val Gln Gly His

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		515					520					525				
Leu	Gln	Glu	Val	Asp	Ala	Gly	Asn	Phe	Ile	Pro	Pro	Pro	Arg	Trp	Leu	
	530					535					540					
Leu	Leu	Asp	Phe	Val	Phe	Val	Leu	Leu	Tyr	Leu	Met	Lys	Leu	Ala	Glu	
545					550					555					560	
Ala	Arg	Leu	Val	Pro	Leu	Ile	Leu	Leu	Leu	Leu	Trp	Trp	Trp	Val	Asn	
				565					570					575		
Gln	Leu	Ala	Val	Leu	Xaa	Val	Xaa	Ala	Xaa	Xaa	Ala	Ala	Val	Ala	Gly	
			580				585						590			
Glu	Val	Phe	Ala	Gly	Pro	Ala	Leu	Ser	Trp	Cys	Leu	Gly	Leu	Pro	Phe	
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Val	Ser	Met	Ile	Leu	Gly	Leu	Ala	Asn	Leu	Val	Leu	Tyr	Phe	Arg	Trp	
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Met	Gly	Pro	Gln	Arg	Leu	Met	Phe	Leu	Val	Leu	Trp	Lys	Leu	Ala	Arg	
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Gly	Ala	Phe	Pro	Leu	Ala	Leu	Leu	Met	Gly	Ile	Ser	Ala	Thr	Arg	Gly	
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Arg	Thr	Ser	Val	Leu	Gly	Ala	Glu	Phe	Cys	Phe	Asp	Val	Thr	Phe	Glu	
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Val	Asp	Thr	Ser	Val	Leu	Gly	Trp	Val	Val	Ala	Ser	Val	Val	Ala	Trp	
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Ala	Ile	Ala	Leu	Leu	Ser	Ser	Met	Ser	Ala	Gly	Gly	Trp	Lys	His	Lys	
	690					695					700					
Ala	Ile	Ile	Tyr	Arg	Thr	Trp	Cys	Lys	Gly	Tyr	Gln	Xaa	Leu	Arg	Gln	
705					710					715					720	
Arg	Val	Val	Arg	Ser	Pro	Leu	Gly	Glu	Gly	Arg	Pro	Thr	Lys	Pro	Leu	
			725						730					735		
Thr	Ile	Ala	Trp	Cys	Leu	Ala	Ser	Tyr	Ile	Trp	Pro	Asp	Ala	Val	Met	
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Leu	Val	Val	Val	Ala	Met	Val	Leu	Leu	Phe	Gly	Leu	Phe	Asp	Ala	Leu	
		755					760					765				
Asp	Trp	Ala	Leu	Glu	Glu	Leu	Leu	Val	Ser	Arg	Pro	Ser	Leu	Arg	Arg	
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Leu	Ala	Arg	Val	Val	Glu	Cys	Cys	Val	Met	Ala	Gly	Glu	Lys	Ala	Thr	
785					790					795					800	
Thr	Val	Arg	Leu	Val	Ser	Lys	Met	Cys	Ala	Arg	Gly	Ala	Tyr	Leu	Phe	
				805					810					815		
Asp	His	Met	Gly	Ser	Phe	Ser	Arg	Ala	Val	Lys	Glu	Arg	Leu	Leu	Glu	
			820					825					830			
Trp	Asp	Ala	Ala	Leu	Glu	Xaa	Leu	Ser	Phe	Thr	Arg	Thr	Asp	Cys	Arg	
		835					840						845			
Ile	Ile	Arg	Asp	Ala	Ala	Arg	Thr	Leu	Ser	Cys	Gly	Gln	Cys	Val	Met	
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Gly	Leu	Pro	Val	Val	Ala	Arg	Arg	Gly	Asp	Glu	Val	Leu	Ile	Gly	Val	
865																

Gly Leu Ser Lys Gly Asp Lys Val Glu Leu Asp Val Ala Met Glu Val
 1010 1015 1020
 Ala Asp Phe Arg Gly Ser Ser Gly Ser Pro Val Leu Cys Asp Glu Gly
 1025 1030 1035 1040
 His Ala Val Gly Met Leu Val Ser Val Leu His Ser Gly Gly Arg Val
 1045 1050 1055
 Thr Ala Ala Arg Phe Thr Arg Pro Trp Thr Gln Val Pro Thr Asp Ala
 1060 1065 1070
 Lys Thr Thr Thr Glu Pro Pro Pro Val Pro Ala Lys Gly Val Phe Lys
 1075 1080 1085
 Glu Ala Pro Leu Phe Met Pro Thr Gly Ala Gly Lys Ser Thr Arg Val
 1090 1095 1100
 Pro Leu Glu Tyr Gly Asn Met Gly His Lys Val Leu Ile Leu Asn Pro
 1105 1110 1115 1120
 Ser Val Ala Thr Val Arg Ala Met Gly Pro Tyr Met Glu Arg Leu Ala
 1125 1130 1135
 Gly Lys His Pro Ser Ile Phe Cys Gly His Asp Thr Thr Ala Phe Thr
 1140 1145 1150
 Arg Ile Thr Asp Ser Pro Leu Thr Tyr Ser Thr Tyr Gly Arg Phe Leu
 1155 1160 1165
 Ala Asn Pro Arg Gln Met Leu Arg Gly Val Ser Val Val Ile Cys Asp
 1170 1175 1180
 Glu Cys His Ser His Asp Ser Thr Val Leu Leu Gly Ile Gly Arg Val
 1185 1190 1195 1200
 Arg Asp Val Ala Arg Gly Cys Gly Val Gln Leu Val Leu Tyr Ala Thr
 1205 1210 1215
 Ala Thr Pro Pro Gly Ser Pro Met Thr Gln His Pro Ser Ile Ile Glu
 1220 1225 1230
 Thr Lys Leu Asp Val Gly Glu Ile Pro Phe Tyr Gly His Gly Ile Pro
 1235 1240 1245
 Leu Glu Arg Met Arg Thr Gly Arg His Leu Val Phe Cys His Ser Lys
 1250 1255 1260
 Ala Glu Cys Glu Arg Leu Ala Gly Gln Phe Ser Ala Arg Gly Val Asn
 1265 1270 1275 1280
 Ala Ile Ala Tyr Tyr Arg Gly Lys Asp Ser Ser Ile Ile Lys Asp Gly
 1285 1290 1295
 Asp Leu Val Val Cys Ala Thr Asp Ala Leu Ser Thr Gly Tyr Thr Gly
 1300 1305 1310
 Asn Phe Asp Ser Val Thr Asp Cys Gly Leu Val Val Glu Glu Val Val
 1315 1320 1325
 Glu Val Thr Leu Asp Pro Thr Ile Thr Ile Ser Leu Arg Thr Val Pro
 1330 1335 1340
 Ala Ser Ala Glu Leu Ser Met Gln Arg Arg Gly Arg Thr Gly Arg Gly
 1345 1350 1355 1360
 Arg Ser Gly Arg Tyr Tyr Tyr Ala Gly Val Gly Lys Ala Pro Ala Gly
 1365 1370 1375
 Val Val Arg Ser Gly Pro Val Trp Ser Ala Val Glu Ala Gly Val Thr
 1380 1385 1390
 Trp Tyr Gly Met Glu Pro Asp Leu Thr Ala Asn Leu Leu Arg Leu Tyr
 1395 1400 1405
 Asp Asp Cys Pro Tyr Thr Ala Val Ala Ala Asp Ile Gly Glu Ala
 1410 1415 1420
 Ala Val Phe Phe Ala Gly Leu Ala Pro Leu Arg Met His Pro Asp Val
 1425 1430 1435 1440
 Ser Trp Ala Lys Val Arg Gly Val Asn Trp Pro Leu Leu Val Gly Val
 1445 1450 1455
 Gln Arg Thr Met Cys Arg Glu Thr Leu Ser Pro Gly Pro Ser Asp Asp
 1460 1465 1470
 Pro Gln Trp Ala Gly Leu Lys Gly Pro Asn Pro Val Pro Leu Leu Leu
 1475 1480 1485
 Arg Trp Gly Asn Asp Leu Pro Ser Lys Val Ala Gly His His Ile Val

1490	1495	1500
Asp Asp Leu Val Arg Arg Leu Gly Val Ala Glu Gly Tyr Val Arg Cys		
1505	1510	1515
Asp Ala Xaa Pro Ile Leu Met Val Gly Leu Ala Ile Ala Gly Gly Met		1520
	1525	1530
Ile Tyr Ala Ser Tyr Thr Gly Ser Leu Val Val Val Thr Asp Trp Asp		1535
	1540	1545
Val Lys Gly Gly Gly Asn Pro Leu Tyr Arg Ser Gly Asp Gln Ala Thr		1550
	1555	1560
Pro Gln Pro Val Val Gln Val Pro Pro Val Asp His Arg Pro Gly Gly		1565
	1570	1575
Glu Ser Ala Pro Arg Asp Ala Lys Thr Val Thr Asp Ala Val Ala Ala		1580
1585	1590	1595
Ile Gln Val Asn Cys Asp Trp Ser Val Met Thr Leu Ser Ile Gly Glu		1600
	1605	1610
Val Leu Thr Leu Ala Gln Ala Lys Thr Ala Glu Ala Tyr Ala Ala Thr		1615
	1620	1625
Ser Arg Trp Leu Ala Gly Cys Tyr Thr Gly Thr Arg Ala Val Pro Thr		1630
	1635	1640
Val Ser Ile Val Asp Lys Leu Phe Ala Gly Gly Trp Ala Ala Val Val		1645
	1650	1655
Gly His Cys His Ser Val Ile Ala Ala Ala Val Ala Ala Tyr Gly Ala		1660
1665	1670	1675
Ser Arg Ser Pro Pro Leu Ala Ala Ala Ala Ser Tyr Leu Met Gly Leu		1680
	1685	1690
Gly Val Gly Gly Asn Ala Gln Ala Arg Leu Ala Ser Ala Leu Leu Leu		1695
	1700	1705
Gly Ala Ala Gly Thr Ala Leu Gly Thr Pro Val Val Gly Leu Thr Met		1710
	1715	1720
Ala Gly Ala Phe Met Gly Gly Ala Ser Val Ser Pro Ser Leu Val Thr		1725
	1730	1735
Val Leu Leu Gly Ala Val Gly Gly Trp Glu Gly Val Val Asn Ala Ala		1740
1745	1750	1755
Ser Leu Val Phe Asp Phe Met Ala Gly Lys Leu Ser Thr Glu Asp Leu		1760
	1765	1770
Trp Tyr Ala Ile Pro Val Leu Thr Ser Pro Xaa Ala Gly Leu Ala Gly		1775
	1780	1785
Ile Ala Leu Gly Leu Val Leu Tyr Ser Ala Asn Asn Ser Gly Thr Thr		1790
	1795	1800
Thr Trp Leu Asn Arg Leu Leu Thr Thr Leu Pro Arg Ser Ser Cys Ile		1805
	1810	1815
Pro Asp Ser Tyr Phe Gln Gln Ala Asp Tyr Cys Asp Lys Val Ser Ala		1820
1825	1830	1835
Ile Val Arg Arg Leu Ser Leu Thr Arg Thr Val Val Ala Leu Val Asn		1840
	1845	1850
Arg Glu Pro Lys Val Asp Glu Val Gln Val Gly Tyr Val Trp Asp Leu		1855
	1860	1865
Trp Glu Trp Val Met Arg Gln Val Arg Met Val Met Ser Arg Leu Arg		1870
	1875	1880
Ala Leu Cys Pro Val Val Ser Leu Pro Leu Trp His Cys Gly Glu Gly		1885
	1890	1895
Trp Ser Gly Glu Trp Leu Leu Asp Gly His Val Glu Ser Arg Cys Leu		1900
1905	1910	1915
Cys Gly Cys Val Ile Thr Gly Asp Val Leu Asn Gly Gln Leu Lys Asp		1920
	1925	1930
Pro Val Tyr Ser Thr Lys Leu Cys Arg His Tyr Trp Met Gly Thr Val		1935
	1940	1945
Pro Val Asn Met Leu Gly Tyr Gly Glu Thr Ser Pro Leu Leu Ala Ser		1950
	1955	1960
Asp Thr Pro Lys Val Val Pro Phe Gly Thr Ser Gly Trp Ala Glu Val		1965
	1970	1975
		1980

Val Val Thr Pro Thr His Val Val Ile Arg Arg Thr Ser Cys Tyr Lys
 1985 1990 1995 2000
 Leu Leu Arg Gln Gln Ile Leu Ser Ala Ala Val Ala Glu Pro Tyr Tyr
 2005 2010 2015
 Val Asp Gly Ile Pro Val Ser Trp Glu Ala Asp Ala Arg Ala Pro Ala
 2020 2025 2030
 Met Val Tyr Gly Pro Gly Gln Ser Val Thr Ile Asp Gly Glu Arg Tyr
 2035 2040 2045
 Thr Leu Pro His Gln Leu Arg Met Arg Asn Val Ala Pro Ser Glu Val
 2050 2055 2060
 Ser Ser Glu Val Ser Ile Glu Ile Gly Thr Glu Thr Glu Asp Ser Glu
 2065 2070 2075 2080
 Leu Thr Glu Ala Asp Leu Pro Pro Ala Ala Ala Leu Gln Ala Ile
 2085 2090 2095
 Glu Asn Ala Ala Arg Ile Leu Glu Pro His Ile Asp Val Xaa Met Glu
 2100 2105 2110
 Asp Cys Ser Thr Pro Ser Leu Cys Gly Ser Ser Arg Glu Met Pro Val
 2115 2120 2125
 Trp Gly Glu Asp Ile Pro Arg Thr Pro Ser Pro Ala Leu Ile Ser Val
 2130 2135 2140
 Thr Glu Ser Ser Ser Asp Glu Lys Thr Leu Ser Val Thr Ser Ser Gln
 2145 2150 2155 2160
 Glu Asp Thr Pro Ser Ser Asp Ser Phe Glu Val Ile Gln Glu Ser Asp
 2165 2170 2175
 Thr Ala Glu Ser Glu Glu Ser Val Phe Asn Val Ala Leu Ser Val Leu
 2180 2185 2190
 Lys Ala Leu Phe Pro Gln Ser Asp Ala Thr Arg Lys Leu Thr Val Lys
 2195 2200 2205
 Met Ser Cys Cys Val Glu Lys Ser Val Thr Arg Phe Phe Ser Leu Gly
 2210 2215 2220
 Leu Thr Val Ala Asp Val Ala Ser Leu Cys Glu Met Glu Ile Gln Asn
 2225 2230 2235 2240
 His Thr Ala Tyr Cys Asp Lys Val Arg Thr Pro Leu Glu Leu Gln Val
 2245 2250 2255
 Gly Cys Leu Val Gly Asn Glu Leu Thr Phe Glu Cys Asp Lys Cys Glu
 2260 2265 2270
 Ala Arg Gln Glu Thr Leu Ala Ser Phe Ser Tyr Ile Trp Ser Gly Val
 2275 2280 2285
 Pro Leu Thr Arg Ala Thr Pro Ala Lys Pro Pro Val Val Arg Pro Val
 2290 2295 2300
 Gly Ser Leu Leu Val Ala Asp Thr Thr Lys Val Tyr Val Thr Asn Pro
 2305 2310 2315 2320
 Asp Asn Val Gly Arg Arg Val Asp Lys Val Thr Phe Trp Arg Ala Pro
 2325 2330 2335
 Arg Val His Asp Lys Phe Leu Val Asp Ser Ile Glu Arg Ala Arg Arg
 2340 2345 2350
 Ala Ala Gln Gly Cys Leu Ser Met Gly Tyr Thr Tyr Glu Glu Ala Ile
 2355 2360 2365
 Arg Thr Val Arg Pro His Ala Ala Met Gly Trp Gly Ser Lys Val Ser
 2370 2375 2380
 Val Lys Asp Leu Ala Thr Pro Ala Gly Lys Met Ala Val His Asp Arg
 2385 2390 2395 2400
 Leu Gln Glu Ile Leu Glu Gly Thr Pro Val Pro Phe Thr Leu Thr Val
 2405 2410 2415
 Lys Lys Glu Val Phe Phe Lys Asp Arg Lys Glu Glu Lys Ala Pro Arg
 2420 2425 2430
 Leu Ile Val Phe Pro Pro Leu Asp Phe Arg Ile Ala Glu Lys Leu Ile
 2435 2440 2445
 Leu Gly Asp Pro Gly Arg Val Ala Lys Ala Gly Val Gly Gly Ala Tyr
 2450 2455 2460
 Ala Phe Gln Tyr Thr Pro Asn Gln Arg Val Lys Glu Met Leu Lys Leu

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2465          2470          2475          2480
Trp Glu Ser Lys Lys Thr Pro Cys Ala Ile Cys Val Asp Ala Thr Cys
          2485          2490          2495
Phe Asp Ser Ser Ile Thr Glu Glu Asp Val Ala Leu Glu Thr Glu Leu
          2500          2505          2510
Tyr Ala Leu Ala Ser Asp His Pro Glu Trp Val Arg Ala Leu Gly Lys
          2515          2520          2525
Tyr Xaa Ala Ser Gly Thr Met Val Thr Pro Glu Gly Val Pro Val Gly
          2530          2535          2540
Glu Arg Tyr Cys Arg Ser Ser Gly Val Leu Thr Thr Ser Ala Ser Asn
2545          2550          2555          2560
Cys Leu Thr Cys Tyr Ile Lys Val Arg Ala Ala Cys Glu Arg Ile Gly
          2565          2570          2575
Leu Lys Asn Val Ser Leu Leu Ile Ala Gly Asp Asp Cys Leu Ile Val
          2580          2585          2590
Cys Glu Arg Pro Val Cys Asp Pro Cys Glu Ala Leu Gly Arg Thr Leu
          2595          2600          2605
Ala Ser Tyr Gly Tyr Ala Cys Glu Pro Ser Tyr His Ala Ser Leu Asp
          2610          2615          2620
Thr Ala Pro Phe Cys Ser Thr Trp Leu Ala Glu Cys Asn Ala Asp Gly
2625          2630          2635          2640
Xaa Arg His Phe Phe Leu Thr Thr Asp Phe Arg Arg Pro Leu Ala Arg
          2645          2650          2655
Met Ser Ser Glu Tyr Ser Asp Pro Met Ala Ser Ala Ile Gly Tyr Ile
          2660          2665          2670
Leu Leu Tyr Pro Trp Xaa Pro Ile Thr Arg Trp Val Ile Pro His
          2675          2680          2685
Val Leu Thr Cys Ala Ser Ser Arg Gly Gly Gly Thr Xaa Ser Asp Pro
          2690          2695          2700
Val Trp Cys Gln Val His Gly Asn Tyr Tyr Lys Phe Pro Leu Asp Lys
2705          2710          2715          2720
Leu Pro Asn Ile Ile Val Ala Leu His Gly Pro Ala Ala Leu Arg Val
          2725          2730          2735
Thr Ala Asp Thr Thr Lys Thr Lys Met Glu Ala Gly Lys Val Leu Ser
          2740          2745          2750
Asp Leu Lys Leu Pro Gly Leu Ala Val His Arg Lys Lys Ala Gly Ala
          2755          2760          2765
Leu Arg Thr Arg Met Leu Arg Ser Arg Gly Trp Ala Glu Leu Ala Arg
          2770          2775          2780
Gly Leu Leu Trp His Pro Gly Leu Arg Leu Pro Pro Glu Ile Ala
2785          2790          2795          2800
Gly Ile Pro Gly Gly Phe Pro Leu Ser Pro Pro Tyr Met Gly Val Val
          2805          2810          2815
His Gln Leu Asp Phe Thr Xaa Gln Arg Ser Arg Trp Arg Trp Leu Gly
          2820          2825          2830
Phe Leu Ala Leu Leu Ile Val Ala Leu Phe Gly
          2835          2840

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<210> 3

<211> 936

<212> DNA

<213> Hepatitis GB virus C

<220>

<221> CDS

<222> (1)..(936)

<400> 3

acc ata gcc gca ctg gga tct tcg gat cgc gac aca gtg gtt gag ctc 48

Thr 1	Ile	Ala	Ala	Leu 5	Gly	Ser	Ser	Asp	Arg 10	Asp	Thr	Val	Val	Glu 15	Leu	
tcc	gag	tgg	gga	att	ccc	tgc	gcc	act	tgt	atc	ctg	gac	agg	cgg	cct	96
Ser	Glu	Trp	Gly 20	Ile	Pro	Cys	Ala	Thr 25	Cys	Ile	Leu	Asp	Arg 30	Arg	Pro	
gcc	tcg	tgt	ggc	acc	tgt	gtg	agg	gac	tgc	tgg	ccc	gag	acc	ggg	tcg	144
Ala	Ser	Cys	Gly 35	Thr	Cys	Val	Arg	Asp 40	Cys	Trp	Pro	Glu	Thr 45	Gly	Ser	
gta	cgt	ttc	cca	ttc	cac	agg	tgt	ggc	gcg	gga	ccg	agg	ctg	acc	aga	192
Val	Arg 50	Phe	Pro	Phe	His 55	Arg	Cys	Gly	Ala	Gly 60	Pro	Arg	Leu	Thr	Arg	
gac	ctt	gag	gct	gtg	ccc	ttc	gtc	aat	agg	aca	act	ccc	ttc	acc	ata	240
Asp	Leu	Glu	Ala	Val	Pro	Phe	Val	Asn 70	Arg	Thr 75	Thr	Pro	Phe	Thr	Ile 80	
agg	ggg	ccc	ctg	ggc	aac	cag	ggg	cga	ggc	aac	ccg	gtg	cgg	tcg	ccc	288
Arg	Gly	Pro	Leu	Gly 85	Asn	Gln	Gly	Arg	Gly 90	Asn	Pro	Val	Arg	Ser	Pro 95	
ttg	ggt	ttt	ggg	tcc	tac	acc	atg	acc	aag	atc	cga	gac	tcc	tta	cac	336
Leu	Gly	Phe	Gly 100	Ser	Tyr	Thr	Met	Thr 105	Lys	Ile	Arg	Asp	Ser 110	Leu	His	
ttg	gtg	aaa	tgt	ccc	acc	cca	gcc	att	gag	cct	ccc	acc	gga	acg	ttt	384
Leu	Val	Lys 115	Cys	Pro	Thr	Pro	Ala	Ile 120	Glu	Pro	Pro	Thr 125	Gly	Thr	Phe	
ggg	ttc	ttc	cca	gga	gtc	ccc	ccc	ctt	aac	aac	tgc	atg	ctt	ctc	ggc	432
Gly	Phe	Phe	Pro	Gly	Val	Pro	Pro	Leu 135	Asn	Asn	Cys	Met 140	Leu	Leu	Gly	
act	gag	gtg	tca	gag	gta	ttg	ggt	ggg	gcg	ggc	ctc	act	ggg	ggg	ttt	480
Thr	Glu	Val	Ser	Glu	Val	Leu	Gly	Gly 150	Ala	Gly 155	Leu	Thr	Gly	Gly	Phe 160	
tac	gaa	cct	ctg	gtg	cgg	cgg	tgt	tca	gag	ctg	atg	ggt	cgg	cgg	aat	528
Tyr	Glu	Pro	Leu	Val 165	Arg	Arg	Cys	Ser	Glu 170	Leu	Met	Gly	Arg	Arg	Asn 175	
ccg	gtc	tgc	ccg	ggg	ttt	gca	tgg	ctc	tct	tcg	gga	cgg	cct	gat	ggg	576
Pro	Val	Cys	Pro	Gly 180	Phe	Ala	Trp	Leu 185	Ser	Ser	Gly	Arg	Pro 190	Asp	Gly	
ttc	ata	cat	gtt	cag	ggc	cac	ttg	cag	gag	gtg	gat	gcg	ggc	aac	ttc	624
Phe	Ile	His	Val	Gln	Gly	His	Leu 200	Gln	Glu	Val	Asp	Ala 205	Gly	Asn	Phe	
att	ccg	ccc	cca	cgc	tgg	ttg	ctc	ttg	gac	ttt	gta	ttt	gtc	ctg	tta	672
Ile	Pro	Pro	Pro	Arg	Trp	Leu	Leu 215	Leu	Asp	Phe	Val	Phe 220	Val	Leu	Leu	
tac	ctg	atg	aag	ctg	gca	gag	gca	cgg	ttg	gtc	ccg	ctg	atc	ctc	ctc	720
Tyr	Leu	Met	Lys	Leu	Ala	Glu	Ala	Arg	Leu	Val 235	Pro	Leu	Ile	Leu	Leu 240	
ctg	cta	tgg	tgg	tgg	gtg	aac	cag	ttg	gcg	gtc	ctt	gkt	gtg	scg	gct	768
Leu	Leu	Trp	Trp	Trp	Val	Asn	Gln	Leu	Ala	Val	Leu	Xaa	Val	Xaa	Ala	

	245	250	255	
gck crc gcc gcc gtg gct gga gag gtg ttt gcg gcc cct gcc ttg tcc				816
Xaa Xaa Ala Ala Val Ala Gly Glu Val Phe Ala Gly Pro Ala Leu Ser				
	260	265	270	
tgg tgt ctg gcc cta ccc ttc gtg agt atg atc ctg ggg cta gca aac				864
Trp Cys Leu Gly Leu Pro Phe Val Ser Met Ile Leu Gly Leu Ala Asn				
	275	280	285	
ctg gtg ttg tac ttc cgc tgg atg ggt cct caa cgc ctg atg ttc ctc				912
Leu Val Leu Tyr Phe Arg Trp Met Gly Pro Gln Arg Leu Met Phe Leu				
	290	295	300	
gtg ttg tgg aag ctc gct cgg ggg				936
Val Leu Trp Lys Leu Ala Arg Gly				
	305	310		

<210> 4

<211> 312

<212> PRT

<213> Hepatitis GB virus C

<220>

<221> MOD_RES

<222> (253)..(258)

<223> X = anything

<400> 4

Thr	Ile	Ala	Ala	Leu	Gly	Ser	Ser	Asp	Arg	Asp	Thr	Val	Val	Glu	Leu
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Ser	Glu	Trp	Gly	Ile	Pro	Cys	Ala	Thr	Cys	Ile	Leu	Asp	Arg	Arg	Pro
			20					25					30		

Ala	Ser	Cys	Gly	Thr	Cys	Val	Arg	Asp	Cys	Trp	Pro	Glu	Thr	Gly	Ser
		35					40					45			

Val	Arg	Phe	Pro	Phe	His	Arg	Cys	Gly	Ala	Gly	Pro	Arg	Leu	Thr	Arg
	50					55					60				

Asp	Leu	Glu	Ala	Val	Pro	Phe	Val	Asn	Arg	Thr	Thr	Pro	Phe	Thr	Ile
65					70					75					80

Arg	Gly	Pro	Leu	Gly	Asn	Gln	Gly	Arg	Gly	Asn	Pro	Val	Arg	Ser	Pro
				85					90					95	

Leu	Gly	Phe	Gly	Ser	Tyr	Thr	Met	Thr	Lys	Ile	Arg	Asp	Ser	Leu	His
			100					105					110		

Leu	Val	Lys	Cys	Pro	Thr	Pro	Ala	Ile	Glu	Pro	Pro	Thr	Gly	Thr	Phe
		115					120					125			

Gly	Phe	Phe	Pro	Gly	Val	Pro	Pro	Leu	Asn	Asn	Cys	Met	Leu	Leu	Gly
130						135					140				

Thr	Glu	Val	Ser	Glu	Val	Leu	Gly	Gly	Ala	Gly	Leu	Thr	Gly	Gly	Phe
145					150					155					160

Tyr	Glu	Pro	Leu	Val	Arg	Arg	Cys	Ser	Glu	Leu	Met	Gly	Arg	Arg	Asn
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

Cys

<210> 7
 <211> 13
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Synthetic Peptide

<400> 7
 Leu Thr Gly Gly Phe Tyr Glu Pro Leu Val Arg Arg Cys
 1 5 10

<210> 8
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Synthetic Peptide

<400> 8
 Phe Tyr Glu Pro Leu Val Arg Arg Cys
 1 5

<210> 9
 <211> 415
 <212> PRT
 <213> Hepatitis GB virus C

<400> 9
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 Leu Trp His Cys Gly Glu Gly Trp Ser Gly Glu Trp Leu Leu Asp Gly
 35 40 45
 His Val Glu Ser Arg Cys Leu Cys Gly Cys Val Ile Thr Gly Asp Val
 50 55 60
 Leu Asn Gly Gln Leu Lys Glu Pro Val Tyr Ser Thr Lys Leu Cys Arg
 65 70 75 80
 His Tyr Trp Met Gly Thr Val Pro Val Asn Met Leu Gly Tyr Gly Glu
 85 90 95
 Thr Ser Pro Leu Leu Ala Ser Asp Thr Pro Lys Val Val Pro Phe Gly
 100 105 110
 Thr Ser Gly Trp Ala Glu Val Val Val Thr Pro Thr His Val Val Ile
 115 120 125
 Arg Arg Thr Ser Ala Tyr Lys Leu Leu Arg Gln Gln Ile Leu Ser Ala
 130 135 140

Ala	Val	Ala	Glu	Pro	Tyr	Tyr	Val	Asp	Gly	Ile	Pro	Val	Ser	Trp	Asp	
145					150					155					160	
Ala	Asp	Ala	Arg	Ala	Pro	Ala	Met	Val	Tyr	Gly	Pro	Gly	Gln	Ser	Val	
				165					170					175		
Thr	Ile	Asp	Gly	Glu	Arg	Tyr	Thr	Leu	Pro	His	Gln	Leu	Arg	Leu	Arg	
			180					185					190			
Asn	Val	Ala	Pro	Ser	Glu	Val	Ser	Ser	Glu	Val	Ser	Ile	Asp	Ile	Gly	
		195					200					205				
Thr	Glu	Thr	Glu	Asp	Ser	Glu	Leu	Thr	Glu	Ala	Asp	Leu	Pro	Pro	Ala	
	210					215					220					
Ala	Ala	Ala	Leu	Gln	Ala	Ile	Glu	Asn	Ala	Ala	Arg	Ile	Leu	Glu	Pro	
225					230					235					240	
His	Ile	Asp	Val	Ile	Met	Glu	Asp	Cys	Ser	Thr	Pro	Ser	Leu	Cys	Gly	
			245					250						255		
Ser	Ser	Arg	Glu	Met	Pro	Val	Trp	Gly	Glu	Asp	Ile	Pro	Arg	Thr	Pro	
			260					265					270			
Ser	Ser	Pro	Ala	Leu	Ile	Ser	Val	Thr	Glu	Ser	Pro	Ser	Asp	Glu	Lys	
		275					280					285				
Thr	Pro	Ser	Val	Ser	Ser	Ser	Gln	Glu	Asp	Thr	Pro	Ser	Ser	Asp	Ser	
	290					295					300					
Phe	Glu	Val	Ile	Gln	Glu	Ser	Glu	Thr	Ala	Glu	Gly	Glu	Glu	Ser	Val	
305					310					315					320	
Phe	Asn	Val	Ala	Leu	Ser	Val	Leu	Lys	Ala	Leu	Phe	Pro	Gln	Ser	Asp	
				325					330					335		
Ala	Thr	Arg	Lys	Leu	Thr	Val	Lys	Met	Ser	Cys	Cys	Val	Glu	Lys	Ser	
			340					345					350			
Val	Thr	Arg	Phe	Phe	Ser	Leu	Gly	Leu	Thr	Val	Ala	Asp	Val	Ala	Ser	
		355					360					365				
Leu	Cys	Glu	Met	Glu	Ile	Gln	Asn	His	Thr	Ala	Tyr	Cys	Asp	Lys	Val	
	370					375					380					
Arg	Thr	Pro	Leu	Glu	Leu	Gln	Val	Gly	Cys	Leu	Val	Gly	Asn	Glu	Leu	
385					390					395					400	
Thr	Phe	Glu	Cys	His	Asn	Cys	Glu	Ala	Arg	Gln	Glu	Thr	Leu	Ala		
				405					410					415		