



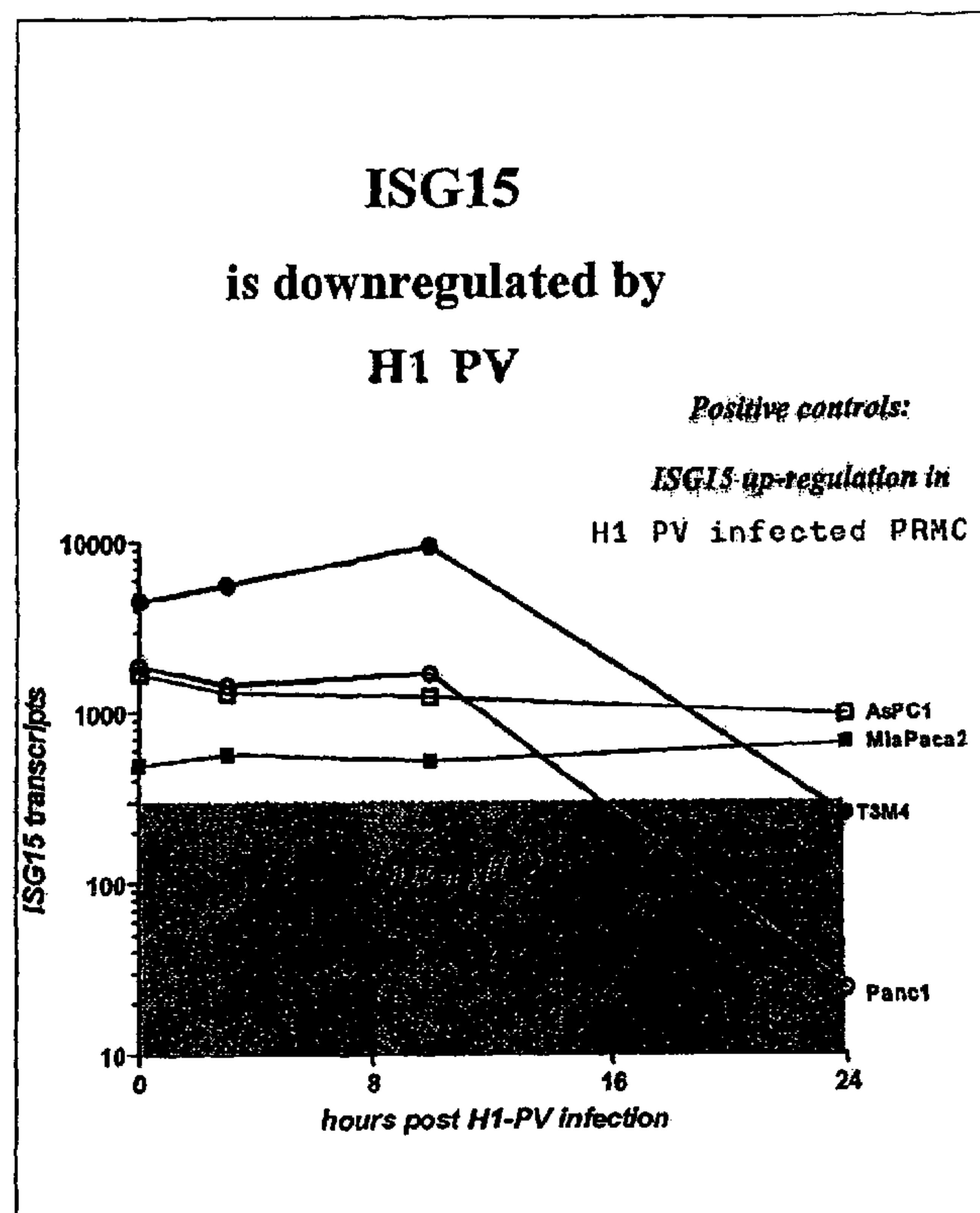
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(54) **Titre : PROCEDE DE DIAGNOSTIC POUR LA PREDICTION DE LA REPONSE D'UN PATIENT A UNE CHIMIO-VIROTHERAPIE
OU A UNE RADIO-VIROTHERAPIE**
(54) **Title: DIAGNOSTIC METHOD FOR PREDICTING THE RESPONSE OF A PATIENT TO CHEMOVIROTHErapy OR
RADIOVIROTHErapy**



(57) **Abrégé/Abstract:**

Described is a diagnostic method for predicting the response of a patient to chemovirotherapy or radiovirotherapy, comprising exposing primary tumor cells from a patient, e.g., tumor cells obtained from a brain tumor or pancreatic cancer, to (i) a parvovirus and/or (ii) a chemotherapeutic agent or radiotherapy, and determining the reduction of the expression or concentration of ISG15.

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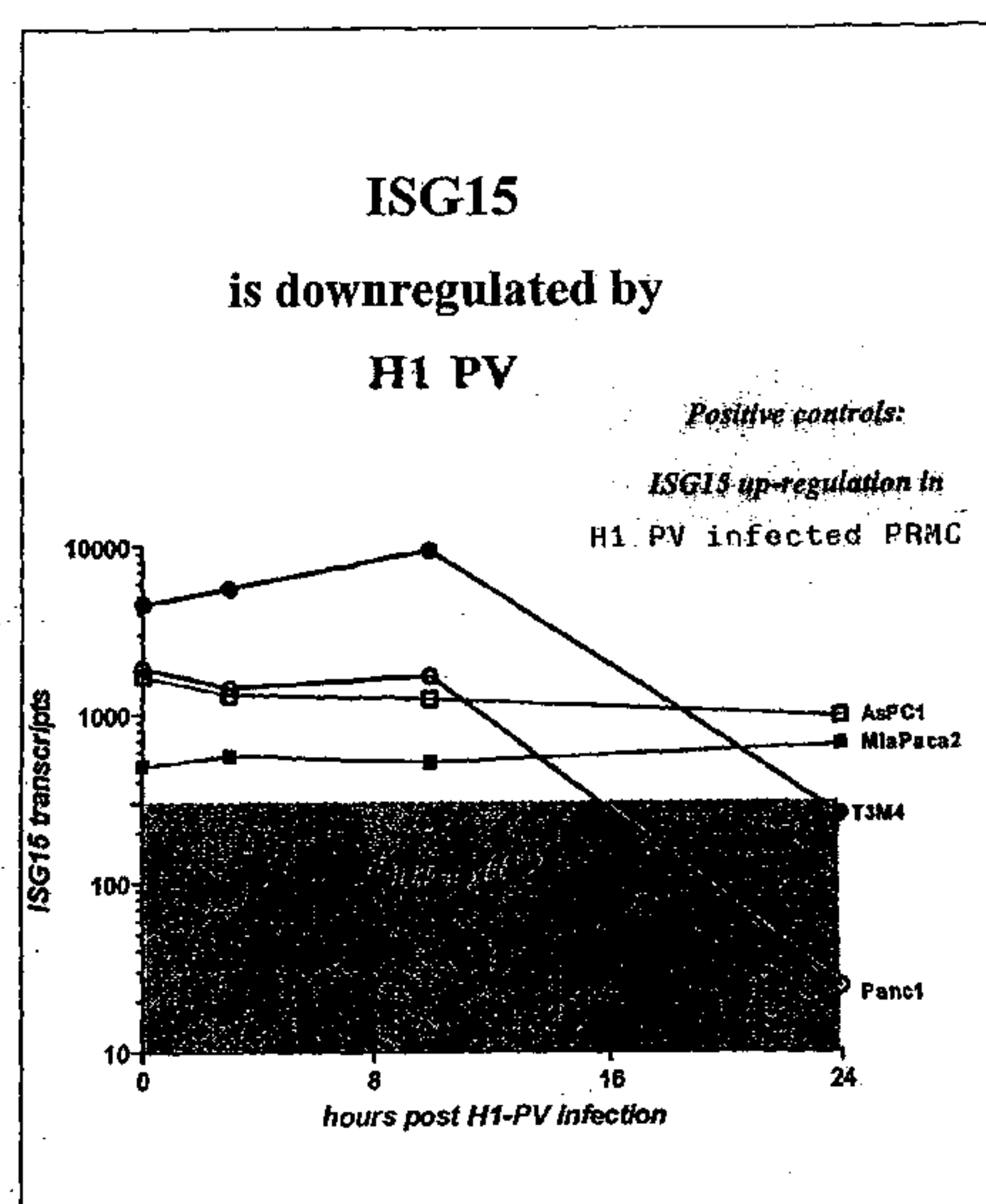
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(54) Title: DIAGNOSTIC METHOD FOR PREDICTING THE RESPONSE OF A PATIENT TO CHEMOVIROTHERAPY OR
RADIOVIROTHERAPY

Figure 3

(57) Abstract: Described is a diagnostic method for predicting the re-
sponse of a patient to chemovirotherapy or radiovirotherapy, comprising
exposing primary tumor cells from a patient, e.g., tumor cells obtained
from a brain tumor or pancreatic cancer, to (i) a parvovirus and/or (ii) a
chemotherapeutic agent or radiotherapy, and determining the reduction of
the expression or concentration of ISG15.

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**Diagnostic method for predicting the response of a patient to
chemovirotherapy or radiovirotherapy**

The present invention relates to a diagnostic method for
5 predicting the response of a patient to chemovirotherapy or
radiotherapy, comprising exposing primary tumor cells from a
patient to a (a) parvovirus and/or (b) a chemotherapeutic
agent or radiotherapy, and determining the reduction of the
expression or concentration of ISG15 (interferon stimulated
10 gene).

Cancer is the second leading cause of death in the United
States after cardiovascular disease. One in three Americans
will develop cancer in his or her lifetime, and one of every
15 four Americans will die of cancer. Malignant human gliomas
account for the largest number of human malignant brain
tumors. So far, the treatment of gliomas includes
neurosurgical techniques (resection or stereotactic
procedures), radiation therapy and chemotherapy. However,
20 despite these therapies gliomas are considered as incurable
as they fail to respond to ionizing radiation, chemotherapy
and surgical resection. In other words, with these therapies
only a very limited prolongation of lifespan of patients can
be achieved, i.e. despite these therapies, the average life
25 span after diagnosis is merely 12 to 16 months.

Pancreatic cancer is a malignant neoplasm of the pancreas.
Each year in the United States, about 43,000 individuals are
diagnosed with this condition and about 35,000 die from the
30 disease. The prognosis is relatively poor but has improved;
the three-year survival rate is now about thirty percent, but
less than 5 percent of those diagnosed are still alive five

years after diagnosis. Complete remission is still rather rare.

Biotherapeutics and especially oncolytic viruses have already
5 been applied in combination with chemotherapy for the
treatment of different types of cancer including PDAC (cancer
of pancreatic duct) (Kasuya et al., Cancer Gene Ther. 2005;
12: 725-36). However, the use of viruses as sensitizing
agents and the problem of predicting patient responsiveness
10 to chemovirotherapy were not successfully addressed until
now. In fact, previous *in vitro* and *in vivo* studies (Angelova
et al., Clin Cancer Res. 2009; 15: 511-9) showed that, e.g.,
pancreatic tumor cells differ in their responsiveness to
treatment with parvovirus (H1-PV) and also to diverse
15 combinations of H1-PV with the standard chemotherapeutic
gemcitabine. Unfortunately, molecular markers predictive for
response are missing. The potential efficacy is extrapolated
from the 'trial-and-error' experiments in animal models.
Since it is impossible to stratify patients into groups
20 according to the potential responsiveness, possibly effective
protocols fail if applied to the wrong target group - which
may profit from different application regimen.

In summary, inability to reliably predict the success of a
25 certain combinatorial treatment, e.g. to select potentially
responding patients, is a major obstacle for clinical
application of chemovirotherapy or radiovirotherapy.

Thus, the technical problem underlying the present invention
30 is to provide a diagnostic method for predicting the response
of a tumor patient to chemovirotherapy or radiotherapy.

The solution to said technical problem is achieved by providing the embodiments characterized herein. A novel chemovirotherapeutic protocol and a test allowing individual adjustment of doses and timing prior to
5 initiation of the treatment thus boosting the success rates of applied therapy was developed. It was found that H1-PV infection reduces ISG15 levels and thereby sensitises cancer cells for chemotherapy (e.g., with gemcitabine). Thus, measurement of ISG15 expression in
10 primary tumor cells exposed to different doses of H1-PV and gemcitabine over varying time periods allows to predict the response to potential therapy and to adjust individual doses of parvovirus (e.g., H1-PV) necessary for effective viro-sensitization (that is resensitising
15 cells towards chemotherapy following H1-PV infection). The experiments resulting in the present invention revealed that infection with H1-PV reduces the expression levels of ISG15 in a subset of human pancreatic cells. Thus the ability of H1-PV to down-regulate ISG15 can be
20 used (i) to design an individual treatment protocol first using H1-PV for sensitization of patients to consequently applied gemcitabine (or other chemotherapeutics) and (ii) to generate a screening tool which can predict potential response of patient derived primary tumor cells to this
25 particular approach prior to the initiation of the treatment. This approach can be expanded to radiovirotherapy since the IFN-related DNA damage resistance has been reported to relate to both chemo- and radioresistance (1-3). Benefits to patients and therapy-
30 financing agencies are apparent, i.e., the approach of the present invention significantly improves the objective responses, allows individualized therapy, and is cost saving.

Brief description of the drawings

Figure 1: Synergy of the combined treatment with H1-PV and gemzar

Figure 2: Downregulation of ISG15 by H-1PV (QRT-PCR)

5 See Example 2 for details

Figure 3: Downregulation of ISG15 by H-1PV

See Example 2 for details.

Figure 4: Comparison of the protocols of applying either Gemzar or H-1PV as first line treatment

10 See Example 2 for details.

Figure 5: Samples obtained during routine PDAC surgeries are(i) used to establish short- and long-term primary cultures, and simultaneously (ii) xenotransplanted into SCID mice and then expanded

15 See Example 3 for details.

Thus, the present invention relates to a diagnostic method for predicting the response of a patient to chemovirotherapy or radiovirotherapy, comprising

(a) exposing primary tumor cells of a tumor sample
20 obtained from a patient to different doses of (i) a parvovirus and/or

(ii) a chemotherapeutic agent or radiotherapy; and

(b) determining the reduction of the expression or concentration of ISG15 over varying time periods.

25 The present invention also relates to a diagnostic method for predicting the response of a patient to chemovirotherapy or radiovirotherapy, comprising

4a

(a) exposing primary tumor cells of a tumor sample obtained from a patient to different doses of (i) a parvovirus and (ii) a chemotherapeutic agent or radiotherapy;

5 (b) determining the reduction of the expression or concentration of ISG15 over varying time periods; and

(c) predicting the response of the patient to chemovirotherapy or radiovirotherapy based on the
10 result of step (b).

The present invention also relates to a diagnostic method for predicting the response of a patient to chemovirotherapy, comprising

15 (a) exposing primary tumor cells of a pancreatic tumor sample obtained from a patient to different doses of (i) parvovirus H1 and (ii) a chemotherapeutic agent being gemcitabine;

(b) determining the reduction of the expression or
20 concentration of ISG15 over varying time periods, wherein ISG15 reduction predicts increased sensitivity of the tumor to the chemovirotherapy; and

(c) predicting the response of the patient to
25 chemovirotherapy based on the result of step (b), wherein the reduction of the expression or concentration of ISG15 in the primary cells of (a) is predictive of the chemovirotherapy efficacy of inhibiting the growth of the patient's tumor cells.

30 Thus, the ISG15 expression in primary tumor cells exposed to different doses of, e.g., H1-PV and gemcitabine over varying

time periods allows to predict the response to potential therapy and to adjust individual doses of parvovirus

In an alternative embodiment, the present invention relates to a method of selecting a therapy modality for a patient afflicted with a tumor, comprising

- (a) exposing primary tumor cells of a tumor sample obtained from a patient to different doses of (i) a parvovirus and/or
- (ii) a chemotherapeutic agent or radiotherapy; and
- (b) determining the reduction of the expression or concentration of ISG15 over varying time periods.

The present invention also relates to a method of selecting a therapy modality for a patient afflicted with a tumor, comprising

- (a) exposing primary tumor cells of a tumor sample obtained from a patient to different doses of (i) a parvovirus and (ii) a chemotherapeutic agent or radiotherapy;
- (b) determining the reduction of the expression or concentration of ISG15 over varying time periods; and
- (c) selecting the therapy modality for the patient based on the result of step (b).

Based on the results of step (b) the response of a patient to chemovirotherapy/radiotherapy can be predicted and the most suitable therapy modality selected, i.e., both a cut-off value can be established and a test selects suitable patients and adjust an appropriate dose for these patients.

5a

The present invention also relates to a method of selecting a therapy modality for a patient afflicted with a pancreas tumor, comprising

- 5 (a) exposing primary tumor cells of a pancreatic tumor sample obtained from a patient to different doses of (i) parvovirus H1 and (ii) a chemotherapeutic agent being gemcitabine;
- 10 (b) determining the reduction of the expression or concentration of ISG15 over varying time periods, wherein ISG15 reduction predicts increased sensitivity of the tumor to the parvovirus H1 and gemcitabine; and
- 15 (c) selecting the therapy modality for the patient based on the result of step (b) wherein the therapy modality comprises a treatment with parvovirus H1 and/or gemcitabine.

The terms "chemovirotherapy" and "radiovirotherapys" as used herein, refer to a combination of (parvo)virotherapy with chemotherapy and radiotherapy, respectively. The
20 parvovirus can be administered prior to, simultaneously with or after administration of the chemotherapeutic agent or radiotherapy. Preferably, the parvovirus is administered prior to the chemotherapeutic agent or radiotherapy.

25 The term "tumor sample" as used herein, refers to a sample obtained from a patient. The tumor sample can be obtained from the patient by routine measures known to the person skilled in the art, i.e., biopsy (taken by aspiration or punctuation, excision or by any other
30 surgical method leading to biopsy or resected cellular material. For those areas not easily reached via an open biopsy, a surgeon can, through a

small hole made in the skull, use stereotaxic instrumentation to obtain a "closed" biopsy. Stereotaxic instrumentation allows the surgeon to precisely position a biopsy probe in three-dimensional space to allow access almost anywhere in the brain. Therefore, it is possible to obtain tissue for the diagnostic method of the present invention.

The term "tumor" is not limited to any stage, grade, histomorphological feature, invasiveness, aggressivity or malignancy of an affected tissue or cell aggregation. In particular stage 0 cancer, stage I cancer, stage II cancer, stage III cancer, stage IV cancer, grade I cancer, grade II cancer, grade III cancer, malignant cancer, primary carcinomas, and all other types of cancers, malignancies etc. are included.

The term "parvovirus" as used herein comprises wild-type or modified replication-competent derivatives thereof, as well as related viruses or vectors based on such viruses or derivatives. Suitable parvoviruses, derivatives, etc. as well as cells which can be used for actively producing said parvoviruses and which are useful for therapy, are readily determinable within the skill of the art based on the disclosure herein, without undue empirical effort. Examples of parvoviruses useful in the present invention include parvovirus H1 (H1-PV) or a related parvovirus such as LuIII, Mouse minute virus (MMV), Mouse parvovirus (MPV), Rat minute virus (RMV), Rat parvovirus (RPV) or Rat virus (RV).

ISG15 is an IFN-alpha/beta-induced ubiquitin-like protein that is conjugated to a wide array of cellular proteins through the sequential action of three conjugation enzymes that are also induced by IFN-alpha/beta. The amino acid

sequence of the protein as well as the nucleotide sequence of the gene encoding ISG15 are described in (4) and (5). Thus, the person skilled in the art can generate probes suitable for determining the expression and/or concentration of ISG15 according to standard methods. The person skilled in the art also knows routine methods for cultivating or maintaining primary tumor cells and incubating these cells with the parvovirus and/or chemotherapeutic agent.

10 The methods of the invention can be applied to any tumor. However, preferred tumors are brain tumor and pancreatic cancer.

Patients treatable by the combination of agents according to the invention include humans as well as non-human animals. Examples of the latter include, without limitation, animals such as cows, sheep, pigs, horses, dogs, and cats.

Chemotherapeutic agents useful for the purposes of the present invention include all chemical compounds that are effective in inhibiting tumor growth. The administration of chemotherapeutic agents can be accomplished in a variety of ways including systemically by the parenteral and enteral routes. Preferably, the parvovirus and the chemotherapeutic agent are administered as separate compounds. Examples of suitable chemotherapeutic agents include alkylating agents, for example, nitrogen mustards, ethyleneimine compounds and alkyl sulphonates; antimetabolites, for example, folic acid, purine or pyrimidine antagonists, mitotic inhibitors, for example, vinca alkaloids and derivatives of podophyllotoxin; cytotoxic antibiotics; compounds that damage or interfere with DNA expression; and growth factor receptor antagonists.

Particular examples of chemotherapeutic agents suitable for the combined therapy include cisplatin, dacarbazine (DTIC), dactinomycin, mechlorethamine (nitrogen mustard), streptozocin, cyclophosphamide, carmustine (BCNU), lomustine (CCNU), doxorubicin (adriamycin), daunorubicin, procarbazine, mitomycin, cytarabine, etoposide, methotrexate, 5-fluorouracil, vinblastine, vincristine, bleomycin, paclitaxel (taxol), docetaxel (taxotere), aldesleukin, asparaginase, busulfan, carboplatin, cladribine, dacarbazine, floxuridine, fludarabine, hydroxyurea, ifosfamide, leuprolide, megestrol, melphalan, mercaptopurine, plicamycin, mitotane, pegaspargase, pentostatin, pipobroman, plicamycin, streptozocin, tamoxifen, teniposide, testolactone, thioguanine, thiotepa, uracil mustard, vinorelbine, chlorambucil and combinations thereof. Particularly preferred chemotherapeutic agents are gemcitabine and temozolodine.

The expression of the gene encoding ISG15 and the concentration of the ISG15 protein can be assayed by standard methods known to the person skilled in the art. The nucleic acid sequence and derived amino acid sequence of ISG15 have been published (4). Preferred assays are based on hybridization or by PCR using appropriate probes/primer pairs, such as Northern blot analysis, reverse transcription polymerase chain reaction (RT-PCR), in situ hybridization, etc..

"Primer pairs" and "probes", within the meaning of the present invention, shall have the ordinary meaning of this term which is well known to the person skilled in the art of molecular biology. In a preferred embodiment of the invention "primer pairs" and "probes" shall be understood as being

polynucleotide molecules having a sequence identical, complementary, homologous, or homologous to the complement of regions of a target ISG15 protein which is to be detected. The primers/probes may be detectably labeled helpful in the
5 detection of the protein. Preferred labels are fluorescent labels, luminescent labels, radioactive labels and dyes.

Preferably, the concentration of the ISG15 protein is determined by using an antibody that specifically binds to
10 the ISG15 protein. Such an antibody can be generated using an ISG15 derived peptide or the entire protein as an immunogen.

The term "antibody" as used herein relates to any type of antibody known in the art. An antibody as used herein
15 includes intact immunoglobulin molecules, as well as fragments thereof, such as Fab, F(ab)₂, and Fv, which are capable of binding an epitope of IDH1. Typically, at least 6, 8, 10, or 12 contiguous amino acids are required to form an epitope. However, epitopes which involve non-contiguous amino
20 acids may require more, e.g., at least 15, 25, or 50 amino acids.

An antibody which specifically binds to ISG15 can be used in immunochemical assays, such as Western blots, ELISAs,
25 radioimmunoassays, immunohistochemical assays, immunoprecipitations, or other immunochemical assays known in the art. Various immunoassays can be used to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays are well
30 known in the art. Such immunoassays typically involve the measurement of complex formation between an immunogen and an antibody which specifically binds to the immunogen.

An antibody useful in the diagnostic method of the present invention can be raised according to well established methods, i.e., an ISG15 polypeptide or fragment thereof can be used to immunize a mammal, such as a mouse, rat, rabbit, 5 guinea pig, monkey, or human, to produce polyclonal antibodies. If desired, the (poly)peptide used as an immunogen can be conjugated to a carrier protein, such as bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin. Depending on the host species, various adjuvants 10 can be used to increase the immunological response. Such adjuvants include, but are not limited to, Freund's adjuvant, mineral gels (e.g., aluminum hydroxide), and surface active substances (e.g. lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, and 15 dinitrophenol). Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially useful.

Monoclonal antibodies which specifically bind to ISG15 can be 20 prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These techniques include, but are not limited to, the hybridoma technique, the human B cell hybridoma technique, and the EBV hybridoma technique [Kohler et al., 25 Nature 256 (1985), 495-7).

Antibodies useful in a method of the invention can be purified by methods well known in the art. For example, antibodies can be affinity purified by passage over a column 30 to which an ISG15 polypeptide is bound. The bound antibodies can then be eluted from the column using a buffer with a high salt concentration.

The invention is not limited to a particular immunoassay procedure, and therefore is intended to include both homogeneous and heterogeneous procedures. Exemplary immunoassays which can be conducted according to the invention include fluorescence polarization immunoassay (FPIA), fluorescence immunoassay (FIA), enzyme immunoassay (EIA), nephelometric inhibition immunoassay (NIA), enzyme linked immunosorbent assay (ELISA), and radioimmunoassay (RIA). An indicator moiety, or label group, can be attached to the subject antibodies and is selected so as to meet the needs of various uses of the method which are often dictated by the availability of assay equipment and compatible immunoassay procedures. General techniques to be used in performing the various immunoassays noted above are known to those of ordinary skill in the art.

In the method of the present invention which relates to the selection of a therapy modality for a patient afflicted with a tumor the term "therapy modality", inter alia, refers to a timely sequential or simultaneous administration of a parvovirus and a chemotherapeutic agent for cancer therapy. The administration of these can be performed in an adjuvant and/or neoadjuvant mode. The variation of the dose of the single agent, timeframe of application and frequency of administration within a defined therapy window depends on the reduction of the expression/level/activity of ISG15 after primary tumor cells of tumor samples obtained from a patient have been exposed to different doses of a parvovirus and/or a chemotherapeutic agent. However, the assay results obtained might even indicate that the patient may profit more from a different treatment regimen. Thus, the term "therapy modality" is not restricted to administration of a parvovirus and a chemotherapeutic agent.

The invention also provides a kit useful for carrying out a method of the invention, comprising an antibody that specifically binds to an ISG15 protein, or a probe or primer pair as described above, i.e., specifically hybridizing to
5 the ISG15 mRNA.

The present invention also relates to a kit comprising an antibody that specifically binds to ISG15 or a probe or primer specifically hybridizing to ISG15 mRNA; and an instruction protocol to carry out the method defined above.

10 The present invention also relates to a kit for predicting the response of a patient to chemovirotherapy comprising an antibody that specifically binds to ISG15 or a probe or primer specifically hybridizing to ISG15 mRNA; and an instruction protocol to carry out the method of any one of
15 claims 1 and 3-5.

The present invention also relates to a kit for selecting a therapy modality for a patient afflicted with a tumor, comprising an antibody that specifically binds to ISG15 or a probe or primer specifically hybridizing to ISG15 mRNA; and an instruction to
20 carry out the method of any one of claims 2-5.

The following examples illustrate the invention.

Example 1

Infection with H1-PV reduces the expression levels of ISG 15

Recent experiments revealed that infection with H1-PV reduces
25 the expression levels of the ISG 15 in a subset of human pancreatic cells. AsPC1, MiaPaCa2, Panc1 and T3M4 cell lines were seeded in 24 well plates and infected with H-1PV at MOIs of 1 and 10. At 3, 10 and 24 hrs post infection (hpi) cells were harvested in 300 µl of MagNA Pure LC mRNA lysis buffer

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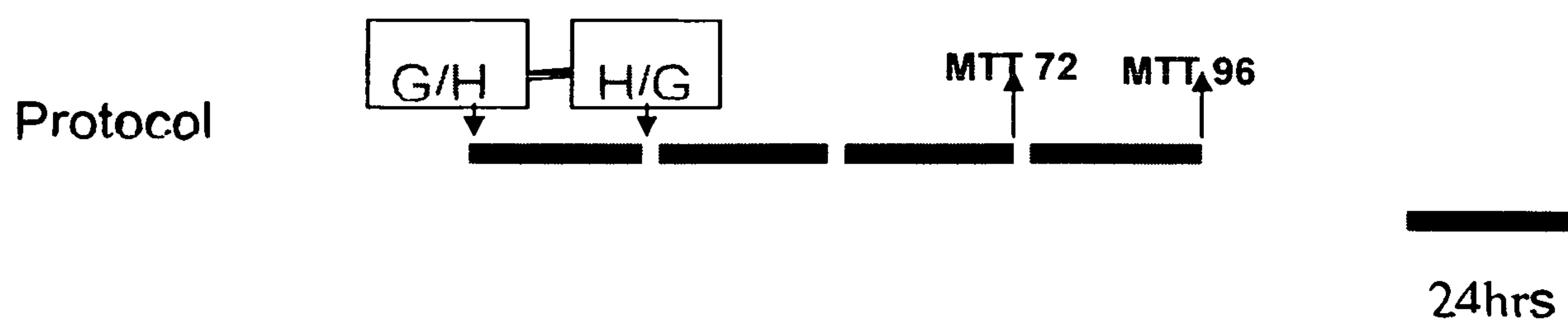
Roche) and after purification of mRNA subjected to QRT-PCR using primers for human ISG15, β -actin and H-1PV. The ISG15 copy numbers were drastically reduced in infected T3M4 and to a lesser extent in Panc1 cells (Fig. 2 and 3).

5

Example 2

Comparison of the protocols of applying either Gemzar or H-1PV as first line treatment

An initial experiment was performed to compare the protocols of applying either Gemzar or H-1PV as first line treatment with a 24 hrs difference. The above-mentioned pancreatic cell lines were plated in a 96 well plate and treated with MOIs 1 or 10 and an EC50 dose of Gemzar using the following scheme. An MTT cytotoxicity assay was performed at 72 and 96 hrs to assess the levels of cell growth inhibition.



10

The results of this initial experiment are shown on Fig. 4. The assumption concerning the improved effectiveness of the H/G (H-1PV-24h-Gemzar; H-G MOI 1 or H-G MOI 10) compared to the G/H (Gemzar-24h-H-1PV; G-H MOI 1 or G-H MOI 10) protocol could be confirmed in the case of T3M4 cells where the virus induces ISG15 reduction (see Fig 2 and 3).

20

Example 3

Selection of patients potentially responsive to the protocol based on use of H-1PV as an ISG15-dependent gemcitabine-sensitizer

25 Samples obtained during routine PDAC surgeries (n=6) will be
i) used to establish short- and long-term primary cultures,

and will be simultaneously ii) xenotransplanted into SCID mice (F0) and future expanded (F1/F2) (see Fig 5). The inhibitory effect of H-1PV on ISG15 expression will be tittered both *in vitro* using both QRT-PCRs and Western blots.

- 5 Rabbit Polyclonal Antibody to human ISG15 (Axxora: Boston) will be used as the primary antibody at 1:500 dilution and *in vivo*. A minimal effective dose of the virus will be established. The degree of tumor cell death will serve as functional read-out.

10

Surgically obtained PDAC material will be used to a establish QRT- or IHC-based screening tool enabling timely selection of patients potentially responsive to the protocol based on use of H-1PV as an ISG15-dependent gemcitabine-sensitizer.

List of references

1. An interferon-related gene signature for DNA damage resistance is a predictive marker for chemotherapy and radiation for breast cancer. Weichselbaum RR, Ishwaran H, Yoon T, Nuyten DS, Baker SW, Khodarev N, Su AW, Shaikh AY, Roach P, Kreike B, Roizman B, Bergh J, Pawitan Y, van de Vijver MJ, Minn AJ. Proc Natl Acad Sci U S A. 2008;105(47):18490-5.
5
2. Signal transducer and activator of transcription 1 regulates both cytotoxic and prosurvival functions in tumor cells. Khodarev NN, Minn AJ, Efimova EV, Darga TE, Labay E, Beckett M, Mauceri HJ, Roizman B, Weichselbaum RR. Cancer Res. 2007;67(19):9214-20.
10
15
3. STAT1 is overexpressed in tumors selected for radioresistance and confers protection from radiation in transduced sensitive cells. Khodarev NN, Beckett M, Labay E, Darga T, Roizman B, Weichselbaum RR. Proc Natl Acad Sci U S A. 2004;101(6):1714-9. Epub 2004 Jan 30.
20
4. Molecular characterization of the interferon-induced 15-kDa protein. Molecular cloning and nucleotide and amino acid sequence. Blomstrom DC, Fahey D, Kutny R, Korant BD, Knight E Jr. J Biol Chem. 1986;261(19):8811-6.
25
5. Production of ISG-15, an interferon-inducible protein, in human corneal cells. Taylor JL, D'Cunha J, Tom P, O'Brien WJ, Borden EC. J Interferon Cytokine Res. 1996;16(11):937-40.
30

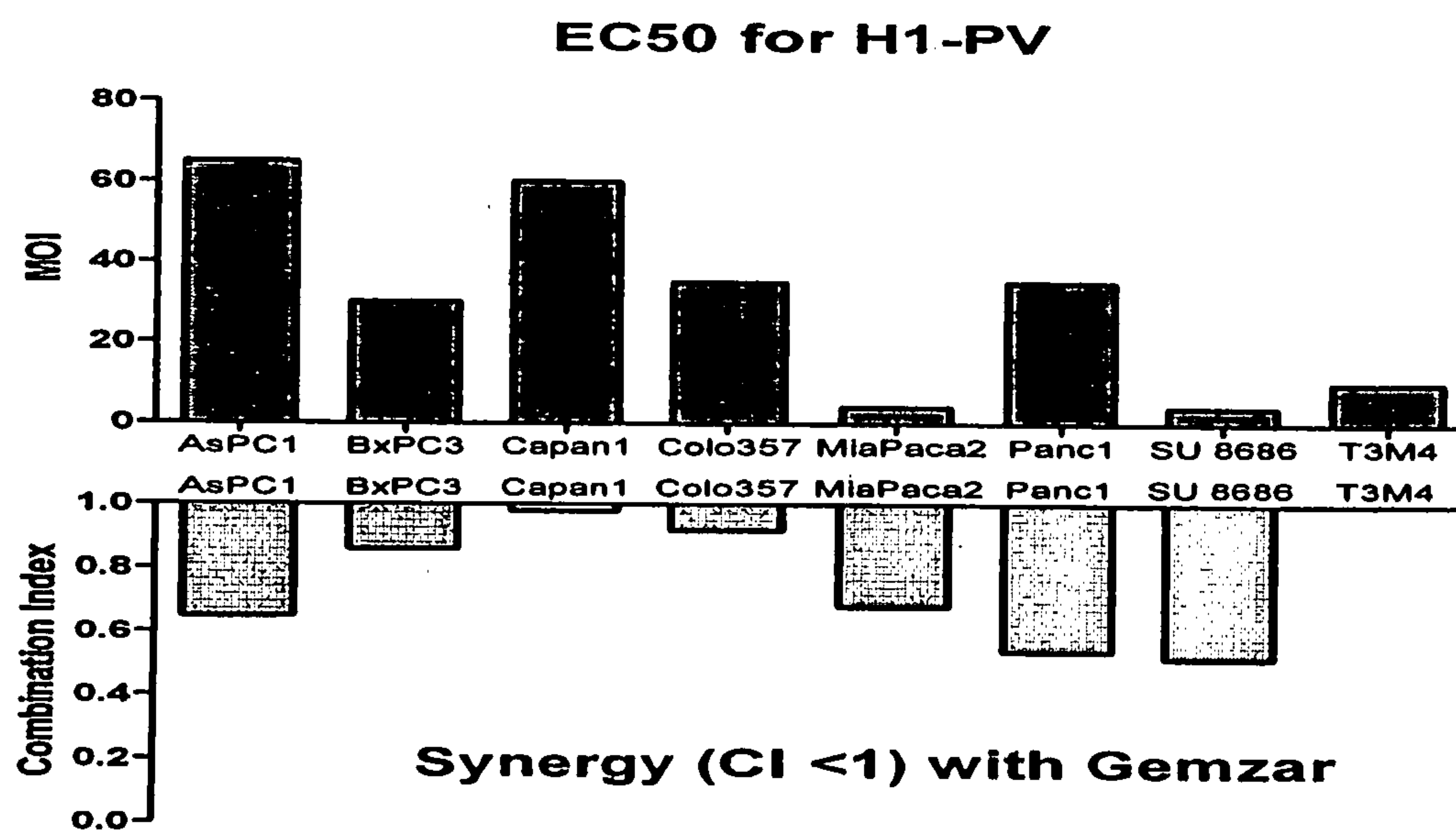
CLAIMS

1. A diagnostic method for predicting the response of a patient to chemovirotherapy, comprising
 - (a) exposing primary tumor cells of a pancreatic tumor sample obtained from a patient to different doses of (i) parvovirus H1 and (ii) a chemotherapeutic agent being gemcitabine;
 - (b) determining the reduction of the expression or concentration of ISG15 over varying time periods, wherein ISG15 reduction predicts increased sensitivity of the tumor to the chemovirotherapy; and
 - (c) predicting the response of the patient to chemovirotherapy based on the result of step (b), wherein the reduction of the expression or concentration of ISG15 in the primary cells of (a) is predictive of the chemovirotherapy efficacy of inhibiting the growth of the patient's tumor cells.
2. A method of selecting a therapy modality for a patient afflicted with a pancreas tumor, comprising
 - (a) exposing primary tumor cells of a pancreatic tumor sample obtained from a patient to different doses of (i) parvovirus H1 and (ii) a chemotherapeutic agent being gemcitabine; and
 - (b) determining the reduction of the expression or concentration of ISG15 over varying time periods, wherein ISG15 reduction predicts increased sensitivity of the tumor to the parvovirus H1 and gemcitabine; and
 - (c) selecting the therapy modality for the patient based on the result of step (b), wherein the therapy

modality comprises a treatment with parvovirus H1 and/or gemcitabine.

3. The method of claim 1 or 2, wherein the expression of ISG15 is determined on the mRNA level.
4. The method of claim 3, wherein the mRNA level is determined by a hybridization based method or by PCR.
5. The method of any one of claims 1 to 4, wherein the concentration of ISG15 is determined using an antibody that specifically binds to ISG15.

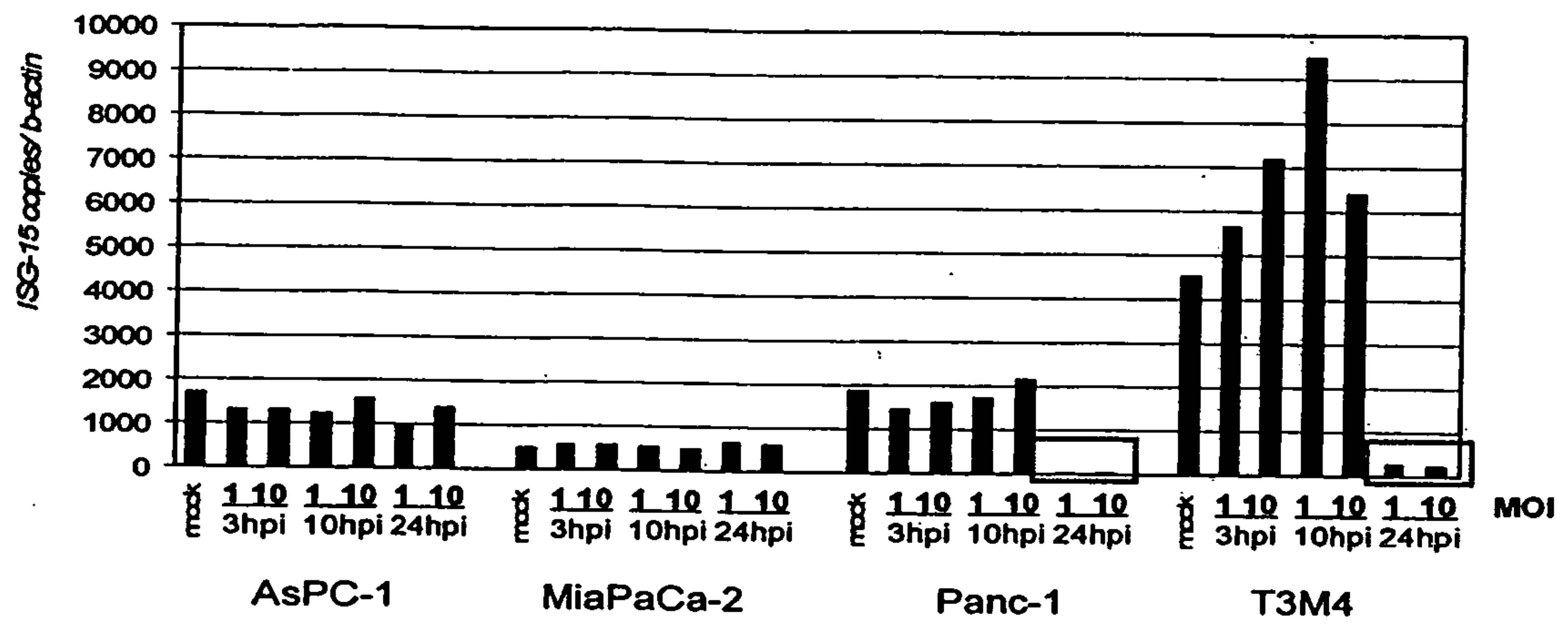
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Figure 1

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Fig. 2

Downregulation of ISG-15 by H-1PV; QRT-PCR



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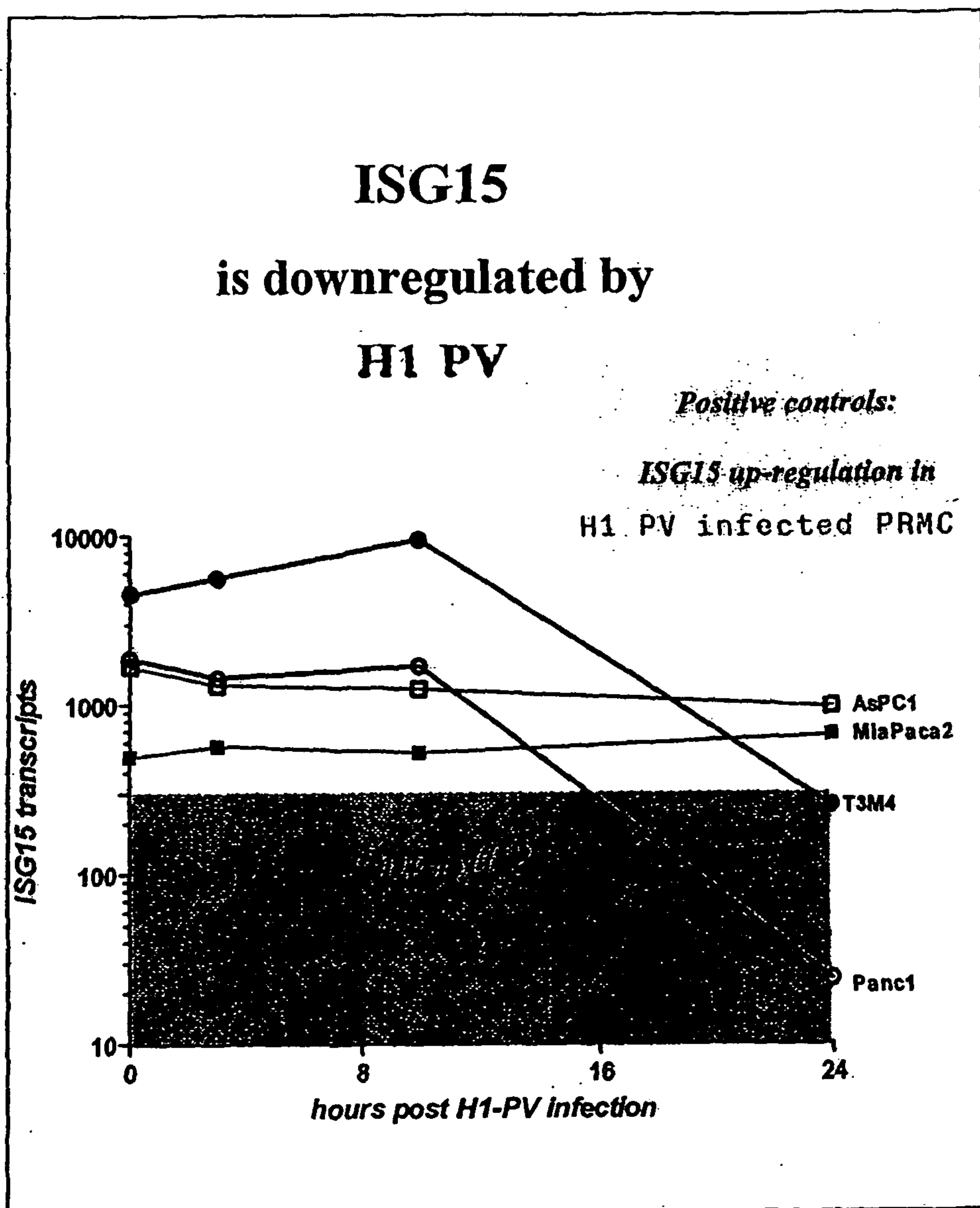
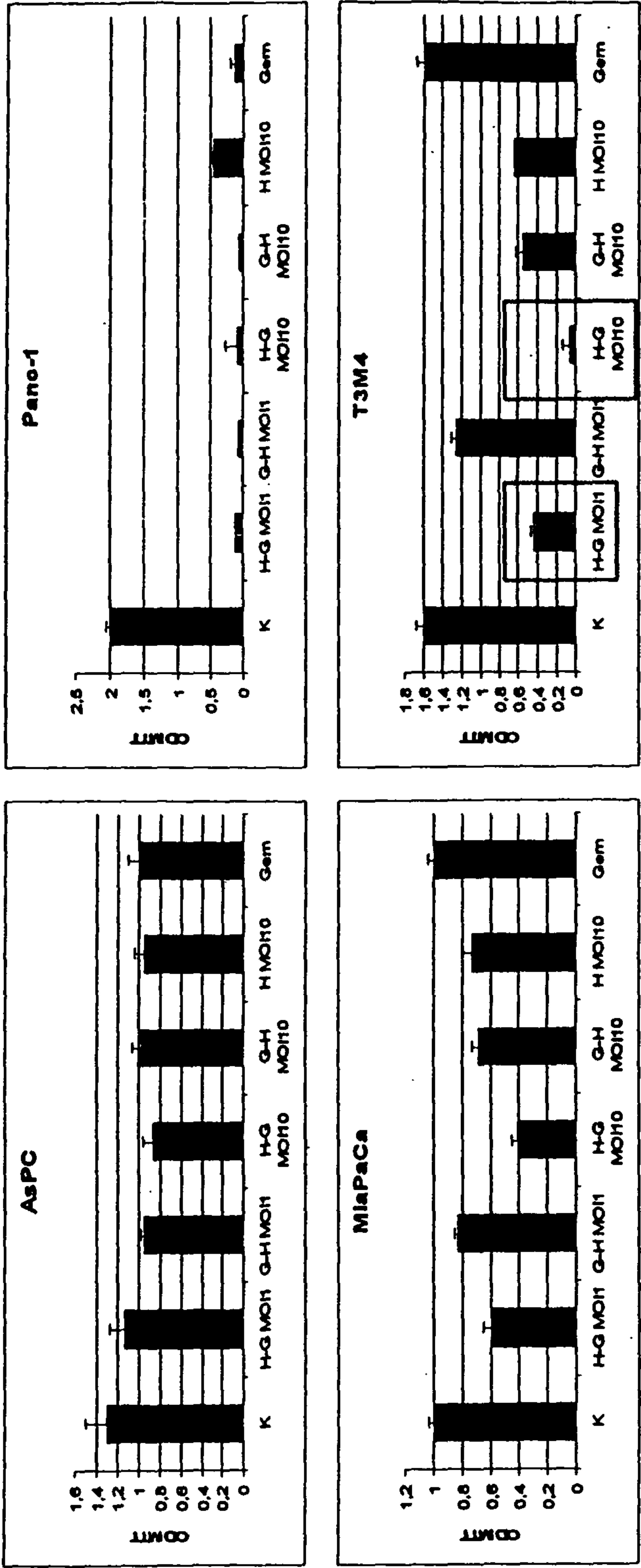
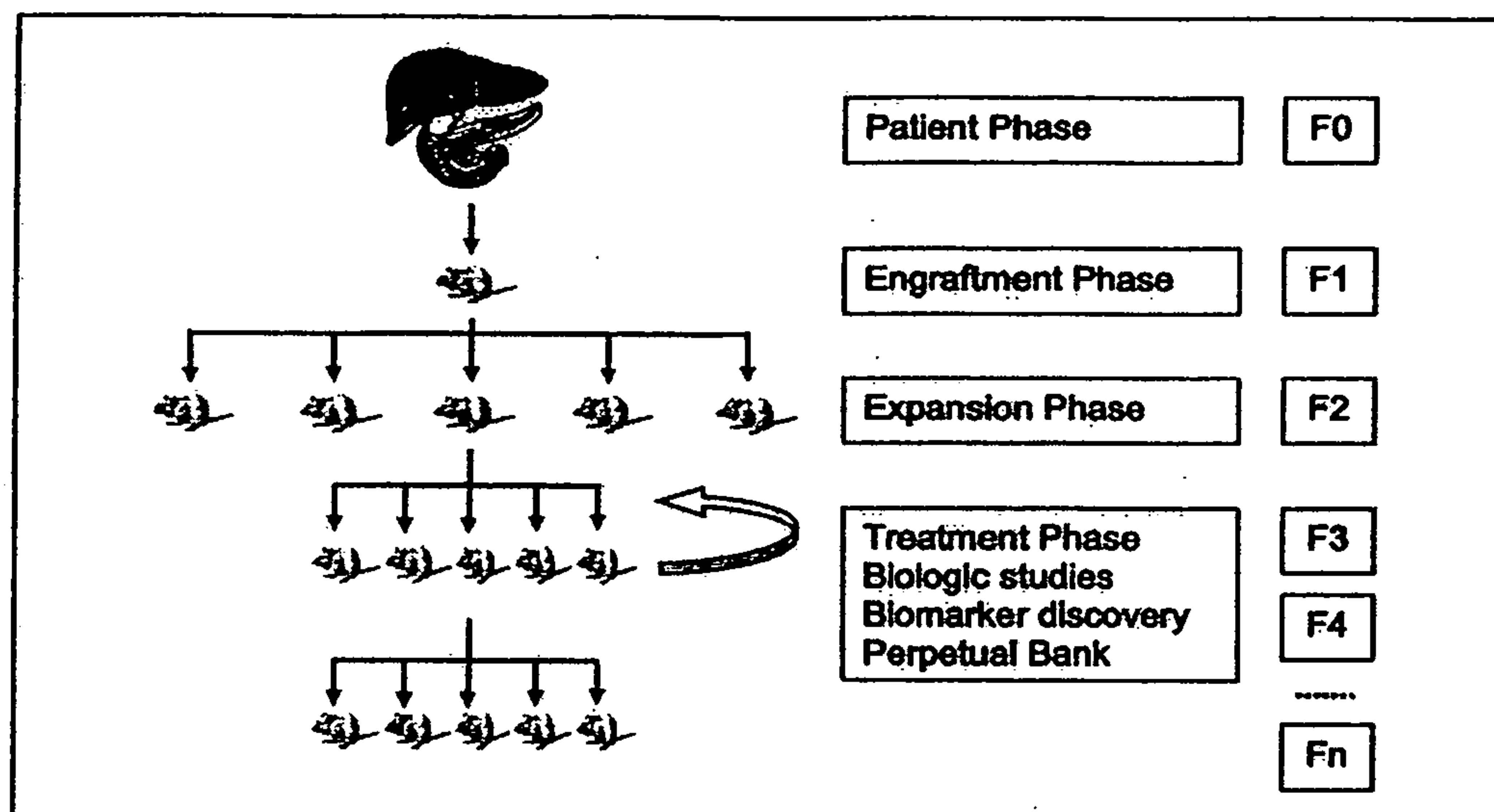
Figure 3

Fig. 4
MTT 96



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Figure 5

ISG15

is downregulated by

H1 PV

Positive controls:

ISG15 up-regulation in
H1 PV infected PRMC

