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(54) Title: COMPOSITIONS CONTAINING PLATELET CONTENTS

(57) Abstract: This document provides methods and materials relating to platelet lysates. For example, methods and materials for using platelet lysate compositions to grow adult stem cells, to differentiate adult stem cells, to grow primary cell cultures, to grow tumor stem cells, and to identify effective growth factors are provided.



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COMPOSITIONS CONTAINING PLATELET CONTENTS

BACKGROUND

1. Technical Field

5 This document relates to methods and materials involved in making and using growth factors, chemokines, and molecules responsible for growing, differentiating, or maintaining undifferentiated cells from normal human platelets (e.g., platelet lysates). For example, this document relates to methods and materials for manufacturing from platelets or platelet preparations (e.g., platelet apheresis preparations) those factors used to grow stem cells (e.g.,
10 adult stem cells) rapidly, to maintain them in an undifferentiated form, as an additive to media to differentiate stem cells (e.g., adult stem cells) in combination with other factors, to grow primary cell cultures (e.g., tumor cells and tumor cell lines), and to grow tumor cells with stem cell properties. This document also relates to methods and materials that can be used to identify, isolate, enrich, or optimize combinations of effective growth factors using
15 platelets as a source material. In addition, the document relates to platelet plasma culture supplements and compositions containing platelet contents.

2. Background Information

 Cell culture is involved in much of modern biology, drug discovery, and therapy. For
20 example, primary tumor cells can be cultured to obtain an antigen source that can be used in anti-tumor vaccines. Two general methods are used to culture primary brain tumor cells. Traditionally, cells are grown in minimal media supplemented with fetal bovine serum. Cells grown in this way can exhibit limited applicability as an effective antigen source because they often exhibit characteristics of differentiated glial cell subtypes with a reduced ability to
25 recapitulate the original tumor *in vivo*. Alternatively, cells can be grown in an enriched media supplemented with growth factors, EGF, and FGF. Cells grown in this environment can form neurospheres and can contain populations of tumor stem cells, which more closely recapitulate the phenotypic characteristics of the primary tumor. Although these cells can have reduced proliferative capacities, they are typically less differentiated, more tumorigenic,
30 and more antigenic than cells grown in fetal bovine serum (Lee *et al.*, *Cancer Cell.*, 9:391-403 (2006)). These approaches do not appear to generate primary tumor cultures with high efficiencies and do not appear to allow for the growth of cultures fast enough for many applications. While these methods can be used to generate cell cultures from malignant gliomas, these protocols typically include materials and methods not suitable for clinical use.

SUMMARY

This document provides methods and materials relating to the collection and use of molecules and growth factors found in platelets (e.g., platelet lysates or “PL”) or media containing platelets (e.g., a supernatant or filtrate from a platelet preparation such as a platelet apheresis preparation). For example, this document relates to methods and materials for making and using compositions containing platelet contents (e.g., platelet lysates and/or supernatants or filtrates from platelet-containing media). The compositions containing platelet contents provided herein can be used to promote enhanced growth of cells (e.g., normal adult progenitor cells, stem cells, and precursor cells). Examples of such cells include, without limitation, mesenchymal stromal cells, endothelial precursor cells, fibroblasts, and epithelial cells. The compositions containing platelet contents provided herein also can be used to enhance differentiation of progenitor cells, stem cells, and precursor cells into functional subtypes efficiently. In addition, the compositions containing platelet contents provided herein can be used to promote enhanced growth of undifferentiated stem cells such as tumor stem-like cells and adult mesenchymal stromal cells or to maintain cells as a mixture of stem cells and differentiated cells such as that found in primary tumor cultures and tumor cell lines. In some cases, the compositions containing platelet contents provided herein can be used to generate primary cell cultures (e.g., tumor cell lines) efficiently. As described herein, the compositions containing platelet contents provided herein (e.g., platelet lysates, supernatants from platelet-containing media, and filtrates from platelet-containing media) can be used to obtain superior growth kinetics when culturing cells (e.g., progenitor cells and tumor stem-like cells) as compared to cultures using supplementation from human serum, fetal bovine serum, or serum-free media with recombinant growth factor supplementation. The compositions containing platelet contents provided herein also can be manufactured in a manner that is acceptable for clinical use (e.g., meets current good manufacturing practice regulations).

This document also provides platelet plasma culture supplements and cell culture methods and materials for using platelet plasma culture supplements. The term “platelet plasma culture supplement” as used herein refers to a composition that contains (a) plasma (e.g., human plasma) and (b) platelet content that is in the form of a platelet lysate that contained greater than 3×10^8 platelets per mL (e.g., greater than 4×10^8 platelets per mL, greater than 5×10^8 platelets per mL, greater than 6×10^8 platelets per mL, greater than 7×10^8 platelets per mL, greater than 8×10^8 platelets per mL, greater than 9×10^8 platelets per

mL, or greater than 1×10^9 platelets per mL) before lysis or a medium that contained greater than 3×10^8 platelets per mL (e.g., greater than 4×10^8 platelets per mL, greater than 5×10^8 platelets per mL, greater than 6×10^8 platelets per mL, greater than 7×10^8 platelets per mL, greater than 8×10^8 platelets per mL, greater than 9×10^8 platelets per mL, or greater than 1×10^9 platelets per mL) before the platelets were removed. Such a medium can be a supernatant or filtrate obtained from a platelet preparation such as a platelet apheresis preparation or a large scale platelet isolation preparation. In some case, such supernatants or filtrates can be obtained from outdated platelet preparations (e.g., outdated platelet apheresis preparations). Outdated platelet preparations can be platelet preparations (e.g., platelet apheresis preparations) that have been obtained from a live human and stored between 20°C and 24°C for more than four days (e.g., more than five, more than six, seven, eight, nine, ten, 11, 12, 15, or more days). Typically, it takes 1-2 liters of blood having a platelet count of $150,000/\text{mm}^3$ to produce one unit of platelets having about 3×10^{11} platelets. Any volume of a preparation containing platelets at the above concentrations can be used. For example, 30 mL (e.g., two 15-mL tubes) 50 mL, 100 mL, 500 mL, or an entire unit of platelets can be used to obtain a platelet lysate or a medium (e.g., plasma) that contained greater than 3×10^8 platelets per mL.

A platelet plasma culture supplement provided herein can have the ability to establish glioma cultures. For example, minimal essential medium having 5 percent of a platelet plasma culture supplement provided herein can be capable of establishing, from newly diagnosed human glioblastoma multiforme biopsy tissue, human glioma cultures having greater than 3×10^7 cells within 60 days at 37°C (in a humidified incubator with 5% CO_2) with a success rate that is greater than 30 percent (e.g., greater than 35, 40, 45, 50, 55, 60, 65, or more percent). This success rate is based on the number of different newly diagnosed human glioblastoma multiforme biopsy tissue samples (i.e., samples for different humans) that can be used to establish a glioma culture, as opposed to the number of attempts using a single human's glioblastoma multiforme biopsy tissue sample multiple times.

The methods and materials provided herein can be used to grow adult stem cells rapidly, to differentiate stem cells (e.g., adult stem cells), to grow primary cell cultures (e.g., tumor cell lines), to grow tumor stem-like cells, and to identify effective growth factors. For example, the methods and materials provided herein can allow clinicians or medical personnel to develop patient-specific autologous cancer vaccines, while following FDA guidelines. Tumor cells such as malignant glioma cells grown using the methods and materials provided herein can maintain many aspects of neural tumor stem cell phenotypes

and can be enriched in tumor-specific antigens desired for recognition of host immune responses. The growth kinetics of cells grown using the methods and materials provided herein can allow clinicians to manufacture sufficient cellular material for multiple vaccinations in a short time frame dictated by current standard therapeutic regimens. Thus, the methods and materials provided herein can provide additional options for the expansion and use of patient specific tumor material for cell therapy.

Another importance of cell culture is the growing use of cells as drugs. In particular, stem cells (both adult and fetal) can be used in regenerative medicine. In such cases, speed of cell growth, fidelity of culture genetics, and clinical applicability can be involved in the success of the cell based therapies.

In general, one aspect of this documents features a platelet lysate composition comprising, or consisting essentially of, a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter. The lysed platelet preparation can be a lysed apheresis platelet preparation. The filter can be a 0.45 μm filter. The filter can be a 0.2 μm filter. The filtrate can be from the lysed platelet preparation being passed through a 0.45 μm filter and a 0.2 μm filter. The lysed platelet preparation can comprise supernatant from centrifugation of lysed platelets. The lysed platelets can be platelets lysed via a freeze/thaw cycle. The lysed platelets can be platelets lysed via at least two freeze/thaw cycles. The centrifugation can comprise a force between 2000 x g and 4000 x g for between 15 and 45 minutes. The centrifugation can comprise a force of about 3000 x g for about 30 minutes. The platelet lysate composition can comprise greater than 200 pg of VEGF polypeptide per mL. Culturing 1.4×10^6 mesenchymal stem cells with media containing about five percent of the platelet lysate composition can result in greater than 1.4×10^7 cells after three days. The mesenchymal stem cells can be adipose derived cells.

In another aspect, this document features a platelet lysate composition produced by filtering a lysed platelet preparation through a 0.45 μm or smaller filter. The lysed platelet preparation can be a lysed apheresis platelet preparation. The filter can be a 0.45 μm filter. The filter can be a 0.2 μm filter. The platelet lysate composition can be produced by filtering the lysed platelet preparation through a 0.45 μm filter and a 0.2 μm filter. The lysed platelet preparation can comprise supernatant from centrifugation of lysed platelets. The lysed platelets can be platelets lysed via a freeze/thaw cycle. The lysed platelets can be platelets lysed via at least two freeze/thaw cycles. The centrifugation can comprise a force between 2000 x g and 4000 x g for between 15 and 45 minutes. The centrifugation can comprise a force of about 3000 x g for about 30 minutes.

In another aspect, this document features a method for making a platelet lysate composition. The method comprises, or consists essentially of, filtering a lysed platelet preparation through a 0.45 μm or smaller filter. The lysed platelet preparation can be a lysed apheresis platelet preparation. The filter can be a 0.45 μm filter. The filter can be a 0.2 μm filter. The method can comprise filtering the lysed platelet preparation through a 0.45 μm filter and a 0.2 μm filter. The lysed platelet preparation can comprise supernatant from centrifugation of lysed platelets. The lysed platelets can be platelets lysed via a freeze/thaw cycle. The lysed platelets can be platelets lysed via at least two freeze/thaw cycles. The centrifugation can comprise a force between 2000 x g and 4000 x g for between 15 and 45 minutes. The centrifugation can comprise a force of about 3000 x g for about 30 minutes.

In another aspect, this document features a method for making a platelet lysate composition. The method comprises, or consists essentially of: (a) lysing platelets via one or more freeze/thaw cycles to obtain lysed platelets, (b) centrifuging the lysed platelets to obtain a supernatant, and (c) filtering the supernatant through a 0.45 μm or smaller filter to obtain a filtrate, wherein the filtrate is the platelet lysate composition. The lysing step can comprise at least two freeze/thaw cycles. The centrifuging step can comprise using a force between 2000 x g and 4000 x g for between 15 and 45 minutes. The centrifuging step can comprise a force of about 3000 x g for about 30 minutes. The filter can be a 0.45 μm filter. The filter can be a 0.2 μm filter. The filtering step can comprise filtering the supernatant through a 0.45 μm filter and a 0.2 μm filter. The platelets can be apheresis platelets.

In another aspect, this document features a method for expanding a cell population comprising, or consisting essentially of, culturing a first population of cells in the presence of medium comprising a platelet lysate composition under conditions wherein the first population of cells is expanded to a second population of cells having more cells than the first population, wherein the platelet lysate composition comprises a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter. The cells can be adult stem cells, primary tumor cells, or tumor stem cells.

In another aspect, this document features a method for differentiating stem cells. The method comprises, or consists essentially of, culturing the stem cells in the presence of differentiating medium comprising a platelet lysate composition under conditions wherein the stem cells differentiate, wherein the platelet lysate composition comprises a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter.

In another aspect, this document features a method for healing a wound. The method comprises, or consists essentially of, contacting the wound with a platelet lysate composition,

wherein the platelet lysate composition comprises a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter. The wound can be a cut, fistula, or diabetic ulcer. The contacting step can comprise using a stitch or glue comprising the platelet lysate composition to contact the wound. The contacting step can comprise spraying the platelet lysate composition onto the wound.

In another aspect, this document features a composition containing plasma and platelet contents from a platelet preparation passed through a 0.45 μm or smaller filter. The plasma can be human plasma. The platelet contents can be human platelet contents.

In another aspect, this document features a composition comprising medium (e.g., a minimal essential medium) and a platelet plasma culture supplement. The platelet plasma culture supplement can be a platelet lysate comprising plasma. The platelet plasma culture supplement can be a filtrate obtained from filtering a platelet apheresis preparation comprising plasma. The platelet apheresis preparation can be a platelet apheresis preparation collected from a human over three days earlier. The platelet apheresis preparation can be a platelet apheresis preparation collected from a human over five days earlier. The platelet apheresis preparation can be a platelet apheresis preparation collected from a human over six days earlier. The platelet plasma culture supplement can be a supernatant obtained from spinning a platelet apheresis preparation comprising plasma. The platelet apheresis preparation can be a platelet apheresis preparation collected from a human over ten days earlier.

In another aspect, this document features a method for obtaining a platelet plasma culture supplement comprising removing platelets or platelet debris from a platelet apheresis preparation such that at least some of the plasma within the platelet apheresis preparation is retained, thereby obtaining the platelet plasma culture supplement, wherein the platelet plasma culture supplement comprises the ability to induce differentiation of 1×10^6 adult stem cells into at least 1×10^3 progenitors of cardiomyocytes, adipocytes, osteoblasts, chondrocytes, myocytes, or nerve cells within 60 days when added to medium at a concentration of 5 percent as a base media with supplementation directed at the desired cell type.

In another aspect, this document features an isolated cell culture comprising MCGBM103 cells cultured in medium comprising a platelet plasma culture supplement.

In another aspect, this document features an isolated cell culture comprising MCGBM106 cells cultured in medium comprising a platelet plasma culture supplement.

In another aspect, this document features an isolated cell culture comprising cells cultured in medium comprising a platelet plasma culture supplement, wherein the cells grow

in the medium at a rate that is faster than the rate of control cells cultured in control medium comprising 10 percent fetal bovine serum. The cells and the control cells can be primary tumor cells. The cells and the control cells can be human primary glioma cells. The cells and the control cells can be stem cells. The cells and the control cells can be fibroblast cells. The medium and the control medium can be neurobasal medium or DMEM/F12 medium. 1×10^5 of the cells can grow in the medium to more than 1×10^6 of the cells faster than 1×10^5 of the control cells cultured in the control medium grow to 1×10^6 of the control cells. 1×10^5 of the cells can grow in the medium to more than 3×10^7 of the cells faster than 1×10^5 of the control cells cultured in the control medium grow to 3×10^7 of the control cells. The cells can be primary glioma cells, and 5×10^5 of the cells can grow in the medium to more than 1×10^7 of the cells within 30 days. The cells can be primary glioma cells, and 5×10^5 of the cells can grow in the medium to more than 1×10^7 of the cells within 60 days. The cells can be stem cells, and 1×10^5 of the cells can grow in the medium to more than 1×10^7 of the cells within 14 days. The cells can be stem cells, and 1×10^5 of the cells can grow in the medium to more than 1×10^7 of the cells within 20 days. The stem cells can be mesenchymal stem cells. The stem cells can be adipose-derived, mesenchymal stem cells. The cells can grow in the medium at a rate that is 1.5 times faster than the rate of the control cells cultured in the control medium.

In another aspect, this document features an isolated cell culture comprising tumor cells cultured in medium comprising a platelet plasma culture supplement, wherein the cells grow in the medium in a manner that is more genetically stable than when control cells are cultured in control medium comprising 10 percent fetal bovine serum. The tumor cells and the control cells can be primary tumor cells. The tumor cells and the control cells can be human primary glioma cells. The medium and the control medium can be neurobasal medium or DMEM/F12 medium. The tumor cells grown in the medium for four doublings can exhibit less than a five percent change in karyotype as compared to the tumor cells grown in the medium for one doubling. The tumor cells grown in the medium for four doublings can exhibit less than a three percent change in karyotype as compared to the tumor cells grown in the medium for one doubling. The tumor cells grown in the medium for ten doublings can exhibit less than a five percent change in karyotype as compared to the tumor cells grown in the medium for one doubling.

In another aspect, this document features an isolated cell culture comprising stem cells cultured in medium comprising a platelet plasma culture supplement, wherein the stem cells grow in the medium in a manner that is more genetically stable than when control stem

cells are cultured in control medium comprising 10 percent fetal bovine serum. The stem cells and the control stem cells can be mesenchymal stem cells. The stem cells and the control stem cells can be adipose-derived, mesenchymal stem cells. The medium and the control medium can be neurobasal medium or DMEM/F12 medium. The stem cells grown in the medium for four doublings can exhibit less than a five percent change in karyotype as compared to the stem cells grown in the medium for one doubling. The stem cells grown in the medium for four doublings can exhibit less than a three percent change in karyotype as compared to the stem cells grown in the medium for one doubling. The stem cells grown in the medium for ten doublings can exhibit less than a five percent change in karyotype as compared to the stem cells grown in the medium for one doubling.

In another aspect, this document features a method for obtaining a platelet plasma culture supplement. The method comprises, or consists essentially of, removing platelets or platelet debris from a platelet apheresis preparation such that at least some of the plasma within the platelet apheresis preparation is retained, thereby obtaining the platelet plasma culture supplement, wherein the platelet plasma culture supplement comprises the ability to induce proliferation of less than 1×10^6 adipose-derived, mesenchymal stem cells at passage five into at least 1×10^8 cells within 20 days when added to minimal essential medium at a concentration of 5 percent. The platelet plasma culture supplement can comprise the ability to induce proliferation of less than 1×10^6 adipose-derived, mesenchymal stem cells at passage five into at least 1×10^8 cells within 15 days when added to minimal essential medium at a concentration of 5 percent.

In another aspect, this document features a method for obtaining a platelet plasma culture supplement. The method comprises, or consists essentially of, removing platelets or platelet debris from a platelet apheresis preparation such that at least some of the plasma within the platelet apheresis preparation is retained, thereby obtaining the platelet plasma culture supplement, wherein minimal essential medium having 5 percent of the platelet plasma culture supplement is capable of establishing, from newly diagnosed human glioblastoma multiforme biopsy tissue, human glioma cultures having greater than 3×10^7 cells within 60 days at 37°C with a success rate that is greater than 30 percent. The success rate can be greater than 40 percent.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Although methods and materials similar or equivalent to those described herein can be used to practice the invention, suitable methods and materials are described below. All

publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

5 The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF THE DRAWINGS

10 Figure 1. Autologous primary neurospheres exhibited enhanced growth kinetics and characteristics of tumor stem cells. A. Morphology of a GBM tumor split and grown independently in Neural Stem Cell (NSC) media (neurospheres), HCTL#3 (mostly adherent cells in this particular cell culture; grown using 5% platelet lysate), and DMEM + 10% FBS (monolayer forming adherent cells). Light phase photomicrographs, 10X. B. Growth kinetics
15 of GBM tumor grown using the three different methods. C. A table listing staining results for neural stem cell and other cell surface markers. Cells were grown on coverslips, and nestin and SOX2 expression was analyzed by indirect immunofluorescence. All other markers were measured by staining cells for each and analyzing them using flow cytometry. +++ indicates >90% positive cells, ++ >50%, + >10%, and - indicates <1% positive cells. D. Cytogenetic
20 analysis of GBM083007 grown in the three different cultures. 20 metaphase cells were counted in each analysis.

Figure 2. Identification and monitoring of tumor associated antigens in autologous primary brain tumor cultures. Gene chip analysis was used to determine relative antigen expression in primary tumors (black bars), tumor stem cell cultures obtained using the Fine
25 protocol (Lee *et al.*, *Cancer Cell.*, 9:391-403 (2006)) (open bars; also known as NSC medium, Neural basal medium, neural stem cell medium), and brain tumor cultures using HCTL#1 (filled bars; grown using platelet lysate, and base media was DMEM/F12).

Figure 3 is a graph plotting the total number of viable GBM cells after one, two, three, four, or five days of being cultured with the media containing the indicated components.

30 Figure 4 is a graph plotting the total number of viable GBM cells after five days of being cultured with the indicated media.

Figure 5 is a graph plotting the total number of MSCs after three, four, or five days of being cultured with the media containing the indicated components.

Figure 6 is a graph plotting the number of GBM cells after three, six, and nine days of

being cultured with the Neurobasal Media (Invitrogen, Grand Island, NY) containing the indicated components.

Figure 7 is a graph plotting the percent of CD133, CD105, and CD90 positive GBM cells after being cultured with the Neural Basal Media containing the indicated components.

5 Figure 8 is a graph plotting the amount of VEGF (pg/mL) present in the indicated preparations. MM = minimally manipulated apheresis platelets from expired donations that were not frozen (3 bags were combined). PPP = Platelet poor plasma, which is a fraction of minimally manipulated apheresis platelets that were spun down at 3000g for five minutes and then spun at 10,000g for one minute. FT = freeze/thaw fraction, which is minimally
10 manipulated apheresis platelets that were subjected to two freeze/thaw cycles. PL = platelet lysate prepared as described in Example 2. MM, PPP, FT, and PL were obtained from the same starting material and were stored for over two months at 4°C.

Figure 9 demonstrates that the levels of growth factors (VEGF and PDGF) are consistent across different lots of manufactured platelet lysates. The levels of these growth
15 factors are significantly higher in the final product preparations versus other growth factor supplements like fetal bovine serum, fetal calf serum, human AB serum, and another commercially available platelet lysate preparation.

Figure 10 is a graph plotting the typical amounts of the indicated growth factors found in manufactured platelet lysates. Eleven manufactured platelet lysate products (Production
20 PL) were measured for the indicated growth factors by ELISA. The growth factor and the axis it uses are indicated below the columns. The amount of the indicated growth factors present within fetal bovine serum (FBS) was also measured.

Figure 11 is a graph plotting the amount of FGF (pg/mL) measured in platelet lysates produced as described herein (n=11) and in commercially available platelet lysates
25 (Cryocheck; n=4).

Figure 12 is a graph plotting the amount of PDGF-BB (ng/mL) measured in platelet lysates produced as described herein (n=11), human AB serum (HABS; n=4), fetal bovine serum (FBS; n=4), commercially available platelet lysates (Cryocheck; n=4), and fresh plasma without platelets (n=4).

30 Figure 13 is a graph plotting the amount of IGF-1 (ng/mL) measured in platelet lysates produced as described herein (n=11), human AB serum (HABS; n=4), fetal bovine serum (FBS; n=4), commercially available platelet lysates (Cryocheck; n=3), and fresh plasma without platelets (n=4).

Figure 14 is a graph plotting the amount of TGF- β (ng/mL) measured in platelet

lysates produced as described herein (n=11), human AB serum (HABS; n=4), fetal bovine serum (FBS; n=4), commercially available platelet lysates (Cryocheck; n=4), and fresh plasma without platelets (n=4).

Figure 15 is a graph plotting the amount of total protein (mg/mL) measured in platelet
5 lysates produced as described herein (n=11), human AB serum (HABS; n=4), fetal bovine serum (FBS; n=4), commercially available platelet lysates (Cryocheck PL; n=3), and fresh plasma without platelets (n=3).

Figure 16 contains a graph plotting the cell proliferation potential and presence of PDGF in PL fractionated using molecular weight cut-off filters. Cell proliferation calculated
10 the number of cells after 800 cells/well were plated and grown for four days containing the indicated component and a graph plotting the amount of PDGF-BB (ng/mL) measured by ELISA in the indicated component. HCTL PL = 5% PL using advanced MEM as the base media.

Figure 17 contains a graph (top) plotting the A_{280} for the indicated fractions of PL
15 isolated by size fractionation and a photograph (bottom) of a silver stained reducing gel containing the indicated fractions.

Figure 18 is a scanning electron photograph of complexes present with a platelet lysate produced as described in Example 2.

Figure 19 is a graph plotting the amount of the indicated cytokine or growth factor
20 present within platelet lysates.

Figure 20 contains four graphs comparing the levels of PDGF-BB, FGF, TGF- β 1, and IGF-1 present with platelet lysates produced as described in Example 2 (HCTL production) to the levels of PDGF-BB, FGF, TGF- β 1, and IGF-1 reported in the indicated references or bovine serum. Lange *et al.* is the Lange *et al.* reference (*Cellular Therapy and*
25 *Transplantation*, 1(2) December (2008)); Weibrich *et al.* is the Weibrich *et al.* reference (*Craniomaxillofac. Surg.*, 30:97-102 (2002)); and Doucet *et al.* is the Doucet *et al.* reference (*J. Cell Physiology*, 205: 228-236 (2005)).

Figure 21 is a graph plotting relative proliferation (absorbance) for MSCs cultured as indicated.

Figure 22 is a graph plotting the cell count for adult stem cells cultured for the
30 indicated number of days in medium (alpha-minimal essential medium (α -MEM)) supplemented with 5 percent of a platelet lysate produced as described in Example 2 or medium supplemented with 10 percent fetal calf serum.

Figure 23 is a graph plotting the population doublings for MSCs cultured in the indicated media.

Figures 24A is a table listing the results from culturing primary tumor cells with the indicated media, and Figures 24B, 24C, 24D, and 24E contain data regarding GBM103,
5 GBM106, GBM110, and GBM111 cell cultures, respectively.

Figure 25 contains four graphs plotting the number of cells at the indicated day for primary tumor cells grown in PL (AIM-V media supplemented with 5% platelet lysate as per example 2), NSC media (Neurobasal-A media with N2 and B27 supplements and EGF and FGF), or DMEM 10% FBS.

10 Figure 26 is a graph plotting the percent positive dendritic cells for the indicated marker of CD14+ monocytes cultured ex-vivo with 1% HABS in X-Vivo 15 media (XV-HABS), 1% platelet lysate-containing X-Vivo 15 media (XV-PL), or CellGro serum free media (no supplementation). All indications contained 2800 IU GM-CSF and 1000 IU IL-4 for the entire course of culture (six days). All conditions had TNF- α and PGE2 added on day
15 three, and the cells were phenotyped on day six. For XV-PL (3-6) the culture was incubated in HABS for the first three days and PL for the last two.

Figure 27 is a graph plotting the total MSCs after five days of culture with the indicated compositions.

Figure 28. Effect of culture condition on cell adherence. Tumors (in rows) were
20 physically and enzymatically digested and incubated in three culture conditions (NSC, neural stem cell; PL, platelet lysate; or FBS, fetal bovine serum). Cells were photographed and descriptions of the morphology listed.

Figure 29. PL promotes rapid growth of primary GBM cultures. Example growth kinetics of a primary tumor grown in each media supplement (left panel), and total cells
25 obtained after 20-30 days of culture for multiple tumor isolates (right panel). PL was superior in the number of cells grown independent of the individual tumor ($p=0.05$; $n=8$).

Figure 30 is a graph plotting cell growth observed with the indicated amount of platelet lysate (PL).

Figure 31. Presence of PDGF and FGF in PL after 48 days of incubation at different
30 temperatures. PL samples were incubated at the temperatures identified for four different lots of PL. Growth factors were measured by ELISA. Each dot represents the mean of three samples from that lot incubated at that temperature. Data plotted is the means from each of these lots. Bars are the mean of four lots, with changes represented by asterisk ($p<0.05$ for paired samples).

Figure 32 is a graph plotting the amount of PDFG and FGF in PL samples incubated at the indicated temperatures.

Figure 33 is a graph plotting the amount of PDFG in PL samples incubated at the indicated temperatures.

5 Figure 34 is a graph plotting the PDFG decay rate.

Figure 35 is contains the karyotype data for two mesenchymal stromal cell isolates cultured in either 10% FBS or 5% PL.

DETAILED DESCRIPTION

10 This document provides methods and materials related to compositions containing platelet contents (e.g., platelet lysates and/or supernatants or filtrates from platelet-containing media). For example, this document relates to methods and materials for making and using compositions containing platelet contents. Any appropriate source of platelets can be used to make a composition containing platelet contents. For example, apheresis platelets and
15 platelets derived from normal blood donation can be used as a source of platelets for making a platelet lysate composition, a supernatant or filtrate from a platelet-containing medium, or a platelet plasma culture supplement.

In one embodiment, a composition containing platelet contents (e.g., a platelet lysate) can be obtained as follows. Once obtained, platelets contents can be released using any
20 appropriate method including, without limitation, a single freeze/thaw cycle, repeated (e.g., 2, 3, 4, 5, or more) freeze/thaw cycles, detergent lysis, activation with thrombin, collagen, thromboxane A₂, ADP or other factors, and manipulation of ionic strength. In some cases, two freeze/thaw cycles can be used to obtain lysed platelets. Once lysed, the lysed platelet preparation can be centrifuged to obtain a supernatant. In general, the force of centrifugation
25 can be between 2000 x g and 5000 x g, and the duration can be between 10 minutes and 60 minutes. For example, a lysed platelet preparation can be centrifuged at about 3,000 x g for about 30 minutes. Once the supernatant is collected, it can be filtered. For example, the supernatant can be filtered through a 0.45 μm filter, a 0.2 μm filter, or a 0.45 μm filter followed by a 0.2 μm filter. The resulting filtrate can be used as a platelet lysate composition
30 without further processing or can be combined with heparin to form a platelet lysate composition.

A platelet lysate provided herein can be prepared without washing the platelets prior to lysing them. In such cases, the platelet lysate can include plasma and plasma components. For example, a platelet lysate provided herein can include albumin and/or thrombin at about

physiologic concentrations. In some cases, a platelet lysate provided herein can include platelet contents prepared from platelets lysed in the presence of plasma or a plasma-like composition.

In another embodiment, a composition containing platelet contents (e.g., a supernatant or filtrate from platelet-containing media) can be obtained as follows. Platelets can be maintained between 2°C and 42°C (e.g., between 2°C and 40°C, between 2°C and 38°C, between 2°C and 36°C, between 2°C and 30°C, between 5°C and 36°C, between 10°C and 36°C, between 15°C and 36°C, between 20°C and 30°C) for a period of time (e.g., two, three, four, five, or more days) in the presence of plasma without performing an active step designed to lyse the platelets. For example, a platelet preparation (e.g., outdated platelet preparation) obtained from an apheresis technique can be used without removing the plasma. Once obtained, the platelet preparation can be treated to remove platelets, platelet debris, or platelet ghosts, while obtaining the resulting medium that includes platelet contents and plasma components. For example, this resulting medium can be obtained by centrifugation and/or filtration. Once obtained, the resulting medium can be stored or used as a composition containing platelet contents or a platelet plasma culture supplement as described herein.

The compositions containing platelet contents (e.g., platelet lysate compositions) provided herein can be formulated with any appropriate medium to produce a culture medium having enhanced properties. Examples of media that can be supplemented with a composition provided herein include, without limitation, DMEM, RPMI, AIMV, X-VIVO15, and other defined serum free or serum requiring media. Any appropriate amount of a composition containing platelet contents provided herein can be added to a medium. For example, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more percent (e.g., vol/vol) of a medium can be a platelet lysate composition or a platelet plasma culture supplement provided herein. Such supplemented media can be used to promote enhanced growth of cells (e.g., normal adult progenitor cells, stem cells, precursor cells, fibroblasts cells, and mesenchymal stromal cells), to differentiate progenitor cells, stem cells, and precursor cells into functional subtypes efficiently, to promote enhanced growth of tumor stem cells in primary tumor cultures, to establish tumor cell lines, or to generate primary cell cultures (e.g., tumor cell lines) efficiently. As described herein, the compositions containing platelet contents (e.g., platelet lysate compositions) provided herein can be used to obtain superior growth kinetics when culturing cells (e.g., progenitor cells and tumor stem cells) as compared to cultures using supplementation from human serum, fetal bovine serum, or serum-free media with recombinant growth factor supplementation.

The compositions containing platelet contents (e.g., platelet lysate compositions) provided herein can be used to supplement media used to culture cells from any species including, without limitation, humans, monkeys, horses, dogs, cats, rats, or mice.

Any appropriate method can be used to derive differentiated cells from stem cells (e.g., mesenchymal stem cells). For example, differentiated cells can be derived from mesenchymal stem cells by incubating the mesenchymal stem cells with a composition (e.g., culture media) containing one or more factors together with a composition containing platelet contents (e.g., a platelet lysate composition) provided herein. The factors can be any type of factors such as polypeptides, steroids, hormones, and small molecules. Examples of such factors may include, without limitation, dexamethasone, EGF, FGF and BMP4. Stem cells can be incubated with a such compositions for 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, or 50 days. In some cases, a composition provided herein and used to promote cell growth or differentiation can be replaced every day or every 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, 50, or more days.

Once stem cells (e.g., mesenchymal stem cells) have been incubated with a composition provided herein or otherwise treated with differentiation factors, the state of differentiation can be monitored to determine whether or not the stem cells differentiated into differentiated cells having a desired phenotype. For example, a sample of cells can be collected and assessed using techniques such as Western blotting, fluorescence-activated cell sorting (FACS), immunostaining, laser confocal microscopy, and reverse transcription polymerase chain reaction (RT-PCR) techniques (e.g., quantitative RT-PCR).

This document also provides isolated cell cultures. For example, this document provides isolated primary tumor cell cultures such as a GBM106 cell culture or any other tumor cell culture identified in Figure 24A. It is noted that the isolated primary tumor cell cultures identified in Figure 24A can be designated with an "MC" prefix when established using a composition containing platelet contents provided herein. For example, the GBM106 cell culture obtained using a platelet lysate provided herein can be designated as MCGBM106. Likewise, the GBM110 cell culture obtained using a platelet lysate provided herein can be designated as MCGBM110, and so on. An isolated primary tumor cell culture provided herein can be obtained from a human patient and cultured in the presence of medium supplemented with a composition containing platelet contents. For example, a medium containing between 1 percent and 50 percent (e.g., between 1 percent and 45 percent, between 1 percent and 35 percent, between 1 percent and 25 percent, between 1 percent and

15 percent, between 1 percent and 10 percent, between 1 percent and 5 percent, between 5 percent and 50 percent, between 10 percent and 50 percent, between 20 percent and 50 percent, between 3 percent and 10 percent, or between 3 percent and 7 percent) of a platelet plasma culture supplement provided herein can be used to culture human primary glioma cells.

Cells (e.g., primary tumor cells) cultured as described herein can exhibit rapid growth kinetics (Figures 1 and 29), higher frequency of cell line establishment (Figures 24A-E), and stable genetics (Figure 1). With reference to Figure 1, it is noted that the karyotype of cells cultured with the HCTL#3 and the NSC media can be very similar, while the karyotype of the cells cultured with FBS can be different. This was done starting with a single tumor sample. Thus, the FBS cultured cells evolved away from the other two cell lines. The other two cell lines are very similar suggesting that these two reflect the progenitor tumor.

In some cases, primary tumor cells cultured as described herein can exhibit a constant doubling time (Figure 29). A constant doubling time can indicate that there is no significant lag time in establishing the culture. Thus, there are no other needed factors that require genetic rearrangements/or additives to have the cultures grow.

Primary tumor cells cultured as described herein also can exhibit the presence of cells with progenitor phenotypes. An example of SOX2 and CD133 expression in a primary tumor culture is provided in Figure 1. SOX2 and CD133 are markers of progenitor cells associated with tumor progenitor cells (tumor stem cells). Primary tumor cells cultured as described herein can be GMP compatible since the supplement (e.g., a composition containing platelet contents such as a platelet plasma culture supplement) can be GMP compatible.

In some cases, primary tumor cells cultured as described herein can be biologically relevant in that many tumors are associated with blood clots. While not being limited to any particular mode of action, a blood clot may act like an *in vivo* matrix for the collection of platelet supernatants. Thus, a composition containing platelet contents provided herein can mimic the normal biology of the tumor.

The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

EXAMPLES

Example 1 – Source of Platelets

All donors donating apheresis platelets fulfilled eligibility criteria as defined by AABB Standards for Blood Banks and Transfusion Service and the Food and Drug

Administration. Donors were screened using the Uniform Donor History Questionnaire (UDQ) and accompanying educational materials. This questionnaire is a screening document created by a coalition of regulatory, accrediting, and blood collecting institutions consisting of the Food and Drug Administration, Centers for Disease Control and Prevention, Armed Services Blood Program, National Heart Lung and Blood Institute, American Blood Resources Association, AABB, American Red Cross, and America's Blood Centers. Information concerning the UDQ can be found on the World Wide Web at "fda.gov/cber/dhq/dhq.htm."

All apheresis platelet donations were tested with the following infectious disease tests: (1) Serologic test for syphilis; (2) HCV EIA-hepatitis C virus antibody test, (3) HCV NAT-hepatitis C virus nucleic acid test, (4) HbsAg-hepatitis B surface antigen test, (6) Anti-HBc-hepatitis B Core antibody, (7) HIV-1/2 EIA-Human Immunodeficiency Virus 1/2 antibody test with ability to detect HIV 1 subgroup O; (8) HIV NAT-Human Immunodeficiency Virus nucleic acid test, (9) HTLV I/II EIA-Human T-Lymphotropic Virus Types I/II, (10) WNV NAT-West Nile Virus nucleic acid test, and (8) Anti-*T. cruzi*, (serologic test for Chagas disease) using FDA licensed procedures.

In addition to the above tests, all apheresis platelet products were also tested for bacterial contamination. Twenty-four hours after collection, the product was resuspended, and an 8 mL sample was collected. Four mL of this sample was inoculated into an anaerobic culture bottle, and 4 mL was inoculated into an aerobic bottle. These bottles were then placed into a BacT/ALERT[®] system (bioMérieux, Durham, NC, USA) within three hours of inoculation and monitored for CO₂ generation for 24 hours. If after 24 hours CO₂ production was not detected, the platelet products were released and made available for transfusion. The culture bottles continued to be monitored for the remaining shelf-life of the platelet product (total of five days; three additional days after release).

The platelet products that were released for manufacture of the growth media were collected from donors who fulfill donation criteria, were negative tests for the infectious diseases listed above, and exhibited no evidence of bacterial growth by their expiration date. Products from donors who failed to meet donor criteria, that exhibited positive infectious disease testing, or that produced cultures positive for bacteria were considered biohazardous waste. These products were quarantined and destroyed. They were not released for manufacture of a platelet lysate growth media. FDA tests and guidelines for release can change. However, platelets used for these purposes met FDA tests and guidelines current at the time of production.

Example 2 – Preparing Platelet Lysate from Apheresis Platelets

Apheresis platelets were obtained as described in Example 1. The apheresis platelets used were no more than four days past expiration. A single lot of platelet lysate consisted of ten individual apheresis platelet units, and one lot was used at a time to create a platelet lysate product. The processing for clinical grade reagents can be performed in a clean room suite. Ten individual apheresis platelet units were frozen at -70°C or colder. After being frozen for at least 24 hours, the units were removed from the freezer and allowed to thaw. The units were thawed at room temperature or at refrigerated temperatures. After thawing was complete, each unit was mixed by massaging the bag. Each thawed platelet bag was placed flat (to minimize breakage of tubing) in a supercold freezer (-70°C or colder) for a second freeze. After the apheresis platelet units were frozen for at least 24 hours for a second freeze, they were removed from the freezer and allowed to thaw. After the second thaw, the platelet product was aseptically transferred to 250 mL conical centrifuge tubes. The tubes were centrifuged for 30 minutes at 3000 x g for 30 minutes at room temperature using a Benchtop Centrifuge Sorvall Legend T. The resulting supernatants were transferred to 0.45-micron filter units (Pall Stericup, Catalog Number SCHV U05 RE; East Hills, NY or Nalgene Filter System, Catalog Number 167-0045; Rochester, NY) that were pre-fitted with one or more pre-filters (Glass Microfibre filters GF/B or GF/D used interchangeably; Whatman®, Florham Park, NJ). The filter unit was connected to a vacuum source and allowed to filter the product. If the product did not filter completely, the unfiltered product was transferred to another filter unit with a pre-filter. The filtrates from all of the 0.45-micron filter units were pooled and filtered through a 0.2-micron filter unit (Pall Stericup, Catalog Number SCHV U05 RE; East Hills, NY or Nalgene Filter System, Catalog Number 567-0020; Rochester, NY) that was pre-fitted with one or more pre-filters (Glass Microfibre filters GF/B or GF/D used interchangeably; Whatman®, Florham Park, NJ). The filter unit was connected to a vacuum source and allowed to filter the product. If the product did not completely filter, the unfiltered product was transferred to a second filter unit, and the process was repeated as needed. The 0.2-micron filtrates were combined into receiver bottles.

A plasma transfer set was connected to a 60-mL syringe, and the transfer set was spiked into a 2L blood bag. Using the syringe as a funnel, the filtered lysates from the apheresis platelet units were combined into the 2L bag. The contents were mixed well. Heparin (1000 U/mL) was added to the filtered platelet lysates to obtain a final concentration of two U/mL.

The lysates were divided into aliquots. The lysates were stored frozen at $\leq 20^{\circ}\text{C}$ or colder.

One of the aliquots containing the platelet lysate was used to perform the following tests to determine whether or not to release the platelet lysate preparation for use:

5 *Aerobic Culture.* One mL of platelet lysate was transferred to a Peds Bactec blood culture bottle (Becton, Dickinson and Company; Sparks, MD) that was used to test sterility.

10 *Anaerobic Culture.* Eight mL of platelet lysate was transferred to a Bactec Lytic/10 Anaerobic/F bottle (Becton, Dickinson and Company; Sparks, MD). Briefly, both aerobic and anaerobic bottles are loaded in the BACTEC 9240 instrument (Becton, Dickinson and Company; Sparks, MD) and monitored every four hours for 14 days. After 14 days, negative cultures are reported out as “No growth at 14 days”, positive cultures are subcultured and isolates identified.

15 *Endotoxin Assay.* One mL of platelet lysate was transferred to a sterile endotoxin-free tube that was used to perform an endotoxin assay. Briefly, a 1:50 dilution of Platelet Lysate to Limulus Amebocyte Lysate (LAL) Reagent Water was run on the Endosafe Portable Test System (PTS; Charles River, Wilmington, MA). The Endosafe PTS utilizes LAL kinetic chromogenic methodology to measure color intensity directly related to the endotoxin concentration in a sample. Each disposable cartridge contains precise amounts of licensed LAL reagent, chromogenic substrate, and control standard endotoxin. The result obtained
20 from each batch of Platelet Lysate must be <0.500 Endotoxin Units (EU)/mL.

25 *Cell Kinetics.* A batch (≥ 150 mL) of Platelet Lysate-5% (PL5%) media containing Advanced-MEM (120mL), GlutaMAX(1.2mL), Heparin (~ 0.24 mL), and 5% platelet lysate (6.4mL) was prepared. A vial of Previously frozen Mesenchymal Stem cells (reference cells) were thawed in a 37°C waterbath. Once thawed, the cells were placed in a sterile 50 mL tube with about 5 mL of the PL5% media. The tube was spun at $240 \times g$ for 5 minutes. The supernatant was removed from the tube, and one mL of the PL5% media was added to the cell pellet. A cell count was performed. The thawed cells were placed in one to two 175 cm^2 flasks with 50 mL of PL5% so that each flask contained 1.75×10^5 - 4.38×10^5 of the thawed cells. The flasks were incubated at 37°C in a 5 percent CO_2 incubator. The cells were
30 passaged using TrypLE™ (Invitrogen Corporation, Carlsbad, CA) after the flasks were confluent. The cells were combined, and a cell count performed. A population doubling calculation was performed.

Flow Cytometry. The cells from the cell kinetic assay were assessed using the following flow cytometry panel:

Tube #	FITC	PE
1	IgG1	IGg1
2	IgG2	IgG2
3	CD90	CD73
4	CD105	HLA-DR
5	CD44	HLA-ABC
6	CD45	CD14

5 Platelet lysates that were sterile, endotoxin-free, grew MSC with the expression profile of CD105, CD90, CD73, HLA-ABC positive and negative for CD14, CD45, and HLA-DR were released for clinical use and assigned an expiration date of two years from production.

Example 3 – Culturing Cells With Platelet Lysates

10 The phenotypic characteristics of GBM cells cultured with media containing platelet lysates were compared to those of GBM cells cultured using NSC media (Neurobasal media (Invitrogen, Grand Island NY); recombinant EGF and FGF (R&D Systems, Minneapolis, MN), N2 and B27 supplements (Invitrogen, Grand Island NY), glutamine and penicillin/streptomycin) or DMEM containing 10% FBS. The media containing platelet
15 lysate (HCTL#3) consisted of Neurobasal media supplement with 5% platelet lysate, glutamine, and penicillin/streptomycin. Cells grown in NSC media formed classical neurospheres enriched in tumor stem cells (Figure 1A). Cells grown in DMEM 10% FBS became adherent and were usually differentiated (Figure 1A). Cells grown in HCTL#3 resulted in a mixed population of free-floating neurospheres and adherent neuropheres with
20 superior growth kinetics (Figure 1A and 1B). Cells cultured in HCTL#3 also exhibited many aspects of neural tumor stem cells and were positive for CD133, nestin, and SOX2 (Figure 1C). In addition, cells grown in HCTL#3 were cytogenetically abnormal as often seen in primary GBM tumors (Figure 1D; Kitange *et al.*, *Curr. Opin. Oncol.*, 15(3):197-203 (2003)).

25 Additional results culturing primary tumor cells are presented in Figures 24A-E and 25. These additional results demonstrate that the platelet lysates provided herein can perform

better than fetal calf serum and a defined supplemented media (NSC) for growing primary tumor cells (e.g., gliomas) (Figures 24A-E). These results also demonstrate that platelet lysate-containing media can grow tumors in less time (Figure 25).

5 Tumor cells grown in media supplemented with manufactured platelet lysate were enriched for primary GBM-associated tumor antigens that were similar, if not superior, to cells grown in NSC media (Figure 2). As such, the PL supplemented media allowed for the generation of primary cell cultures from GBM patients that exhibited superior growth kinetics, characteristic tumor genetic rearrangements, populations of neural tumor stem cells, and enrichment in certain tumor antigens. The use of lysates from autologous tumor cultures
10 as antigen for DC immunotherapy offers the advantage of targeting multiple tumor antigens reflective of the patient's antigen profile, eliminating contaminating cells, and providing sufficient antigen for multiple immunizations. Preliminary research in GBM patients demonstrated the consistent ability to generate primary tumor cultures from small portions of resected tumor.

15 Cells cultured from a primary brain tumor were thawed and plated (3.5×10^4 cells/well) in one of the following culture media: Neurobasal media (Invitrogen, Grand Island, NY) plus 5% fully manufactured platelet lysate (platelet lysate), Neurobasal media containing 5 percent platelet lysate from a single freeze/thaw cycle without further processing (Min. Manipulated), Neurobasal media containing 5 percent human AB serum (HABS,
20 Sigma, St. Louis, MO), Neurobasal media alone, Neurobasal media containing 5 percent platelet rich plasma (PRP), Neurobasal media containing 5 percent platelet rich plasma that was sonicated (PRP-Sonicated), and Neurobasal media containing 5 percent of a commercially available platelet lysate (Cat. No. PNP-10, Lot No. PL14, Precision Biologic, Dartmouth, Nova Scotia). Equal numbers of cells were plated in duplicate for each
25 condition. The cells were cultured for five days, and the total viable number of cells was counted each day (Figure 3). The results from day 5 are presented in Figure 4. These results demonstrate that in this experiment, tumor cells grew the best in fully manufactured platelet lysate.

30 MSCs were thawed and grown in Advanced MEM (Invitrogen, Grand Island, NY) plus 5% HCTL manufactured platelet lysate until confluent. The cells were then passaged and split into Advance MEM with the following supplements: 5 percent (vol/vol) of a platelet lysate prepared as described in Example 2, platelet poor plasma (plt. poor plasma), whole platelets (whole plts.), or platelet exposed to two freeze/thaw cycles without further processing (Freeze/thaw plts.). The 5 percent platelet lysate was used as baseline with the

others being adjusted to provide an equivalent protein concentration as the 5 percent platelet lysate. The cells were allowed to equilibrate in their specific media for a few days before beginning the experiment. Once prepared, the MSC were plated at 2.5×10^4 cells/plate into T25 flasks with their appropriate medium. The total number of cells in each of three flasks for each condition was counted on days three, four, and five (Figure 5). These results demonstrate that MSCs grew the best in HCTL fully manufactured lysate.

A cell line from a primary tumor was thawed and plated similarly to the MSC growth experiment. Conditioned cells were then plated at 8×10^4 cells/plate in Neurobasal Media (Grand Island, NY) supplemented with either 5 percent (vol/vol) of a platelet lysate (PL) prepared as described in Example 2, platelet poor plasma (plt poor plasma or PPP), whole platelets (whole plts or WP), or platelet exposed to two freeze/thaw cycles without further processing (Freeze/thaw plts or FT). The 5 percent platelet lysate was used as baseline with the others being adjusted to provide an equivalent protein concentration as the 5 percent platelet lysate. The cells were allowed to equilibrate in their specific media for a few days before beginning the experiment. The total number of cells in each of three flasks for each condition was counted on days three, six, and nine (Figure 6). These results demonstrate that cells grew better in fully manufactured platelet lysate over 6 days.

On day 5, the cells from each flask for each particular condition type were pooled and analyzed by FACS for CD133 (two tubes were assessed), CD105 (one tube), and CD90 (one tube). The percent of positive cells for each marker was determined (Figure 7). These results demonstrate that cells grown in fully manufactured platelet lysate were enriched for CD133+ cells. CD133 expression has been documented as a marker for glioma-initiating stem cells (Singh *et al.*, *Cancer Res.*, Sept 15;63(18):5821-8 (2003)).

In another experiment, the amount of VEGF (pg/mL) present in various platelet preparations was determined (Figure 8). The platelet preparations included (1) minimally manipulated (MM) apheresis platelets from expired donations that were not frozen (3 bags were combined), (2) platelet poor plasma (PPP), which was a fraction of minimally manipulated apheresis platelets that were spun down at 3000g for five minutes and then spun at 10,000g for one minute, (3) a freeze/thaw fraction (FT), which contained minimally manipulated apheresis platelets that were subjected to two freeze/thaw cycles, (4) platelet lysate (PL) prepared as described in Example 2. The MM, PPP, FT, and PL preparations were obtained from the same starting material and were stored for over two months at 4°C. The PL preparation exhibited the highest level of VEGF (Figure 8). These results

demonstrate that our manufacturing process increases the concentration of this growth factor and is highest at the final step of manufacturing.

Subsequent experiments demonstrate that the levels of VEGF and PDGF are consistent across several different lots of platelet lysate manufactured in the HCTL (14244, 14453, 14569, and 14593) (Figure 9). In addition, these growth factors are significantly higher on a per volume basis than that of other growth supplements including fetal bovine serum, fetal calf serum, human AB serum, and another commercially available platelet lysate preparation (Cat. No. PNP-10, Lot No. PL14, Precision Biologic, Dartmouth, Nova Scotia).

In another experiment, six mesenchymal stromal cell isolates were split and grown using media supplemented with either 10% FBS or 5% PL. After 12 passages, the cells were submitted for karyotype analysis. Two of the six cultures grown in FBS had abnormal karyotype, while none of those lines grown in PL were abnormal. The karyotypes from the two abnormal and their analogous lines grown in PL are shown in Figure 35.

In addition to the initiation of primary tumor cultures derived from glioblastoma patients using PL, PL were used to initiate primary tumor cultures derived from surgical specimens obtained from patients with renal cell carcinoma.

Example 4 – Analysis of Platelet Lysates

Eleven lots of platelet lysates manufactured as described in Example 2 (Production PL) were analyzed to determine the amount of EGF, FGF, IGF-1, PDGF-BB, TGF- β , and VEGF present as measured by ELISA. The measured amounts were compared to those measured in one or more of the following: fetal bovine serum (FBS), human AB serum (HABS), a commercially available platelet lysate (Cryocheck; catalog number PNP-10; Precision BioLogic, Inc., Nova Scotia, Canada) and fresh plasma without platelets. The typical amounts of EGF, FGF, PDGF-BB, TGF- β , and VEGF within the manufactured platelet lots as compared to FBS are presented in Figure 10. The average amount of FGF within the manufactured platelet lots was about 180 pg/mL, while the average amount present within a commercially available platelet lysate was 80 pg/mL (Figure 11). The average amount of PDGF-BB within the manufactured platelet lots was about 8.5 ng/mL, while the average amount present within a commercially available platelet lysate was 2.9 ng/mL (Figure 12). The average amount of IGF-1 within the manufactured platelet lots was about 132 ng/mL, while the average amount present within a commercially available platelet lysate was 7.7 ng/mL (Figure 13). The average amount of TGF- β within the manufactured platelet lots was about 54 ng/mL, while the average amount present within a commercially available

platelet lysate was 37 ng/mL (Figure 14). The levels of PDGF-BB, FGF, TGF- β , and IGF-1 present in platelet lysates produced as described in Example 2 were compared to the levels of PDGF-BB, FGF, TGF- β , and IGF-1 reported in the literature or bovine serum (Figure 20).

The total amount of protein within eleven lots of platelet lysates manufactured as described in Example 2 was measured using a modified Bradford assay. The average amount of total protein within the manufactured platelet lots was about 49 mg/mL, while the average amount present within the commercially available platelet lysates was 0.03 mg/mL (Figure 15). The total amount of protein within the manufactured platelet lots was similar to the amounts measured in the fresh plasma without platelets, the fetal bovine serum, and the human AB serum (Figure 15).

These results suggest that growth factor concentration alone is not responsible for the effective growth stimulating properties of the platelet lysates provided herein. The mean of the levels of growth factors present with the platelet lysates provided herein was about 2 to 3 times that of the commercially available platelet lysate. However, the commercially available platelet lysate was not different than fetal bovine serum. In addition, the platelet lysates provided herein had about 3000 times the amount of protein than that of the commercially available platelet lysates, yet protein alone cannot account for the activity as the platelet lysates provided herein since they exhibited similar levels of total protein as that found in human AB serum.

The following experiments were performed to assess the size of the material within the platelet lysate provided herein that contains PDGF-BB and the ability to stimulate cell proliferation. Briefly, platelet lysates were produced from an outdated platelet concentrate from a single donor. The number of platelets for each lysing condition was equal, and the volume used for lysing was equal for all modifications and equal to the volume of the platelet concentrate used for the unmodified condition. The platelet lysates were filtered on size selection filters with specific molecular weight cut-offs. The filtrate (less than the size indicated) and the retentate (greater than the size indicated) were analysed for the presence of PDGF-BB by ELISA and the ability to stimulate cell proliferation. To assess cell proliferation, 96-well tissue culture plates seeded with 800 adult adipose derived MSC cells were used with Advanced MEM (Gibco) media supplemented with 2 units/mL heparin and a preparation of platelet lysate (5%; v/v). PDGF-BB has a molecular weight of about 30 KDa (30,000 MW protein).

PDGF-BB was observed only in the retentates including the >100,000 preparations suggesting that all of the PDGF-BB is bound to complexes greater than this size. Also,

proliferation occurred in cultures containing these larger (> 100,000 Da) complexes. These results indicate that there are high molecular weight complexes containing growth factors that can be responsible for the activity of the platelet lysates provided herein.

5 Growth factors and other small proteins were found in large complexes in the platelet lysates provided herein. A platelet lysate (0.5 mL), which was thawed, centrifuged at 3,000 x g, and filtered through a 0.2 mm filter, was applied to a 1.5 cm x 27 cm column packed with Sephadex G-150-120 (Pharmacia Fine Chemical Company) and equilibrated with 50 mM sodium phosphate, 150 mM NaCl, pH 7.4 buffer. The chromatography was developed at a flow rate of 25 mL/hour, and the A₂₈₀ of each fraction was measured. Column fractions were
10 analyzed by SDS-PAGE using a 4-15% Criterion gel (Bio-Rad Laboratories) and silver staining. This was a size exclusion analysis that resulted in the largest complexes being in the earliest fractions and the smaller complexes/single molecules being in the later fractions.

A large fraction of the platelet lysate flowed from the column right away (Figure 17, top). To see the size of these complexes, these fractions were run on a reducing gel (Figure
15 17, bottom). Early fractions contained proteins with a molecular weight less than 25,000 Da, suggesting that small proteins are bound to these large complexes.

A sample from platelet lysate manufactured as described in Example 2 was examined using a scanning electron microscope. As shown in Figure 18, several large complexes are readily observable. Many particles were observed up to 0.2 μm in size.

20 In another experiment, 1-4 lots of platelet lysates manufactured as described in Example 2 were assessed to determine the amount of 21 cytokines and growth factors present within the platelet lysates using a BioPlex 26-plex assay (Biorad, Hercules, CA) per manufacturer's directions. The results are presented in Figure 19.

25 Example 5 – Culturing Cells Using Platelet Lysates

MSCs were thawed and grown in a-MEM with 5% PL until confluent. The cells were then passaged and split into a-MEM medium only or a-MEM medium with the following supplements: 5 percent (vol/vol) Cryocheck, 10 percent (vol/vol) FBS, 5 percent (vol/vol) HABS, or 5 percent (vol/vol) of a platelet lysate prepared as described in Example 2.

30 Relative proliferation was assessed using absorbance measurements. The greatest amount of proliferation was observed with MSCs cultured in the presence of the platelet lysate prepared as described in Example 2 (Figure 21).

In other experiments, adipose tissue MSCs were observed to grow faster in the presence of 5 percent of a platelet lysate produced as described in Example 2 than in 10 percent fetal calf serum (Figures 22 and 23).

To determine the immune modulating effect of PL, PL was used as a supplement instead of human AB serum in culture conducive to the differentiation of CD14+ monocytes into the immune stimulating cells dendritic cells (described in Dietz *et al.*, *Transfusion*, 46(12):2083-9 (2006)). PL were incubated for the entire culture condition or for the last stage (day 3-6) of dendritic cell culture characteristic of generating active mature dendritic. The changes associated in the culture were measured with particular emphasis on the expression of CD80 and CD83; two characteristic markers of dendritic cell culture. No effect of the PL as a supplement was observed in these conditions. Thus, PL appears to have a benign effect on the immune response.

In another experiment, MSCs were plated in media, and cell growth was determined by counting. The media supplements were as follows: (a) none (media only), (b) PL #1 (platelet lysate manufactured in the presence of plasma), (c) washed PL #1 + plasma (platelet lysate manufactured after removal of plasma followed by adding back plasma), (d) PL #1 washed (platelet lysate manufactured after removal of plasma), (e) PL2 + Benzamidine (platelet lysate manufactured in the presence of plasma plus the clotting inhibitor benzamidine), and (f) washed platelets (PC2) in buffer (platelets manufactured in the absence of plasma). Cell proliferation was observed in cells cultured with PL #1 and cells cultured with washed PL #1 + plasma (Figure 27). Little or no proliferation was observed with cells cultured under the other conditions (Figure 27). These results suggest that both plasma and clotting factors can be required to produce an effective composition containing platelet contents.

Example 6 - High efficiency establishment of primary glioblastoma cultures for drug discovery, drug optimization, and anti-tumor vaccines

Patient-derived primary glioblastoma multiforme (GBM) cell cultures applicable for clinical translation are produced. The culture medium described herein containing platelet contents is used for a highly efficient establishment of tumor cell cultures. The medium is likely more relevant to tumor growth than other methods and can be optimized for clinical-scale production compliant with Good Manufacturing Practice (GMP) requirements.

The effect of media supplementation with platelet contents on tumor culture was tested by splitting primary tumors and incubating them in one of three culture methods. The

first culture included 10% fetal calf serum (FCS or FBS) in culture media. The second culture contained culture media used to culture tumor stem cells (Neural stem cell media, NSC; Piccirillo and Vescovi, *Expert Opinion on Biological Therapy*, 7(8):1129 (2007); Fan *et al.*, *Seminars in Cancer Biology*, 17(3):214 (2007)). The third culture media contained media supplemented with 5% human platelet derived media supplement (platelet lysate, PL).

The effect of media supplementation was clear (Figure 28). In response to the culture conditions, cells self-adhered forming spheres, preferentially adhered to the plastic forming monolayers, or cultures with mixed adherence. FBS cultures were uniformly adherent monolayers, with NSC media mostly having spheres or non-adherent cells. PL cultures existed within the complete spectrum of cell morphology. Cells continued in culture and were counted at each passage. PL was superior to other methods in the ability to generate large numbers of cells (Figure 29). Importantly, cells grown in PL demonstrated a constant growth rate for more than 10 doublings in the absence of an initial lag phase, suggesting an immediate proliferation response upon plating (Figure 29, left panel). Therefore, PL induced immediate and profound proliferation in many primary tumor cultures. Of the sixteen tumors cultured thus far, a 56% success rate of establishing primary cell growth was obtained, and a 44% rate of generating greater than 30 million cells in 60 days was obtained. Not a single culture grown in FBS could meet the latter requirement. Thirteen percent of the tumors grown in NSC conditions met these criteria. Interestingly, there was one cell culture that grew equally well in PL and NSC, and only one cell culture that grew uniquely in NSC. Overall, PL were greater than 3 times better at establishing primary tumor cultures than the next best method.

A criticism of FBS established cell cultures is that they are considered to have lost the undifferentiated nature of the *in vivo* tumor. To determine if cells cultured in PL have characteristics of tumor stem cells, the expression of nestin, Sox2, and CD133 was measured in PL cultured cells. The majority of cells in all cultures were nestin and Sox2 positive. Four of the seven cultures tested for CD133 expression had levels above background with one culture having more than 30% of the cells positive for CD133. These results suggest that indeed the cultures contain cells that have characteristics of tumor stem cells.

To determine the genetic stability of cultures grown in PL, human adipose derived mesenchymal stem cells from non-malignant donors were incubated in PL supplemented media. No abnormal karyotypes were found in up to twelve passages of cells. Thus, PL maintained the genetic integrity of normal cells in culture. GBM cultures were karyotyped and found to maintain a stable karyotype despite massive cell doublings (see, e.g., GBM106;

Figure 24C). In some cases, cultures consisted of stable subclones. The tumor cultures exhibited characteristic deletions (such as pten) and expressed tumor antigens such as EGFr, survivin, and IL13-r. Taken together, these results suggest that PL is a highly effective growth supplement for the establishment and proliferation primary tumor cultures.

5 An additional benefit of PL as a growth supplement is its source. Out-dated human platelets, approved-for-human-use, were used. This material was approved for therapeutic use with all the commensurate testing completed.

Thus, the methods and materials provided herein can allow to the generation of autologous tumor cells for use in patient specific vaccines, or other immune modulatory
10 protocols where patient specific material may be required. The efficient *in vitro* generation of tens of millions of cells from >40% of all GBM tumors can allow for many new approaches to cancer treatment, including (a) the possibility to generate sufficient patient specific materials for repeated vaccinations for immune therapy protocols, (b) the establishment of a primary tumor cell library for drug screening on cell cultures having
15 substantially fewer passages and more relevant material, (c) the development of specific drug screening protocols for patient specific optimization of chemotherapy, and (d) the cataloguing of the molecular profile of these primary tumor cultures and the association of these profiles with chemo-sensitivity.

A clinical trial is performed to confirm the use of autologous primary GBM cell
20 cultures, which are produced as described herein, as an antigen source for vaccine studies. Primary cell cultures are established from material obtained at surgery. Patients undergo standard care.

The patient begins vaccinations using the primary cell cultures combined with immune stimulating adjuvants. Possible adjuvants for a tumor vaccine include dendritic cells, alum,
25 GM-CSF, LPS, KLH etc. as well as other promising adjuvants.

In some cases, the generated cGMP material is used directly as tumor antigen for allogeneic donors. Patients have their tumors molecularly matched to the corresponding library, a match is identified, and is subsequently used for treatment.

To identify additional cancer treatments, the cell cultures produced as described
30 herein are screened for drug-mediated suppression of cell growth using compounds from the NIH clinical collection. This collection represents drugs used in early phase clinical trials with described adequate safety profiles and commercial availability. Drugs identified as candidates for cell suppression are tested *in vivo* on the same tumors growing in immune deficient mouse models. Likewise, drugs identified as candidates are matched to the

transcription profile or if the patient cells are growing fast enough, drugs tested on the patient material and used on the patient allowing patient specific drug screening and use.

Example 7 - Stability of Platelet Lysates

5 *Platelet Lysate*

Platelet lysate was manufactured as described in Example 2. Immediately after manufacture, PL was aliquotted into multiple vials and transferred to -70°C, 5°C, 25°C, or 37°C for long-term storage. Sufficient aliquots were transferred such that samples were maintained at a single temperature until analysis. Thus, the data represent samples stored at a constant temperature for the time indicated. Three independent lots of PL were analyzed for the presence of growth factors and two for the ability to proliferate MSC.

Analysis Methods

MSC Proliferation: Adipose derived, adult mesenchymal stromal cells (MSC) were obtained from fat collected as waste from gastric by-pass surgery. Fat was processed, and MSC were expanded and frozen until use in these assays. For these assays, MSC were thawed and allowed a single passage prior to plating for analysis. Growth was determined using the BioVision Quick Cell Proliferation Assay Kit II (Mountain View, CA). This assay is based on the cleavage of tetrazolium salt to formazin by cellular mitochondrial dehydrogenase. The amount of dye cleaved is proportional to live cells. Briefly, MSC were plated at 1×10^4 cells per well in RPMI1640 containing the designated amount of PL from the indicated conditions. The cells were incubated for 48 hours in 37°C, 5% CO₂ humidified incubator. The detection reagent was added (10 mL of WST) to each well. The plate was incubated for one hour, and the O.D. measured per manufacturer's directions.

25 Growth Factors: An ELISA test on PL that was stored at the indicated temperatures was used to determine the concentration of the growth factors of interest. All ELISAs were from R&D Systems (Minneapolis, MN) and used according to manufacturer's instructions.

Results

30 Growth of MSC in PL followed a typical dose response curve. Seventy percent of the capacity of MSC growth was observed using 20% of the typical amount of PL as a supplement. Thus, 1% PL media supplementation resulted in approximately 70% of the growth capacity.

Long-term storage of PL at 37°C reduced, but did not eliminate, MSC growth (Figure 30). MSC were cultured with different concentrations of PL that had been maintained at -70°C or 37°C for 48 days. There was a typical dose response curve observed for both PL incubated at -70°C and 37°C. Interestingly, the PL incubated for this long at body temperature maintained its ability to stimulate 70% of the expected growth of PL at a similar concentration.

To identify possible changes in growth factor concentrations associated with the reduction in growth of MSC, the levels of FGF and PDGF were measured in four lots of PL held at -70°C, 5°C, 25°C, and 37°C (Figures 31 and 32). These concentrations were measured immediately after production, and on days 7, 14, 21, and 48.

When PL was stored at 37°C for 48 days, FGF levels fell to 36.5 ± 5.8 pg/mL compared to baseline levels of 115 ± 29.5 and to 111.3 ± 26.8 when PL was stored for 48 days at -70°C. This reduction was significant ($p < 0.01$ and a reduction of approximately 32%; data represents mean from N=4 PL lots where each lot was measured using three independently stored samples). No reduction in concentrations of FGF was observed when it was stored for this amount of time at 5°C or 25°C.

Similarly, when PL was stored at 37°C for 48 days, PDGF levels fell to 2.8 ± 0.4 ng/mL compared to baseline levels of 10.2 ± 1.8 or 10.1 ± 2.6 when PL was stored for the same amount of time at -70°C. This reduction was significant ($p < 0.01$ and a reduction of approximately 27%). Additionally, a temperature dependent reduction in the amount of PDGF measured when the PL was stored at 25°C (mean= 6.0 ± 0.9 and $p = 0.02$) was observed. Thus, the loss in the growth factors approximates the loss in the MSC growth stimulating capacity. To calculate the shelf life, the changes in growth factors over time and temperature were used.

Estimation of shelf life

PDGF was the most sensitive to the effects of accelerated aging. Therefore, the shelf life results were derived from that data (Figure 33). The slope intercept method (Arrhenius equation), Q rule, and conservative approaches were used to estimate the shelf life of platelet lysates (Figure 34 and Table 1).

Table 1. Estimated shelf life of platelet lysates.

	For storage at -70°C	For storage at -20°C
Shelf life	24.1 years	Approximately 1 year (290.5 days)

Taken together, these results indicate that the compositions provided herein containing platelet contents have an estimated shelf life of greater than five years at -70°C and an estimated shelf life of about one year at -20°C.

5

OTHER EMBODIMENTS

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other
10 aspects, advantages, and modifications are within the scope of the following claims.

WHAT IS CLAIMED IS:

1. A platelet lysate composition comprising a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter.
2. The platelet lysate composition of claim 1, wherein said lysed platelet preparation is a lysed apheresis platelet preparation.
3. The platelet lysate composition of claim 1, wherein said filter is a 0.45 μm filter.
4. The platelet lysate composition of claim 1, wherein said filter is a 0.2 μm filter.
5. The platelet lysate composition of claim 1, wherein said filtrate is from said lysed platelet preparation being passed through a 0.45 μm filter and a 0.2 μm filter.
6. The platelet lysate composition of claim 1, wherein said lysed platelet preparation comprises supernatant from centrifugation of lysed platelets.
7. The platelet lysate composition of claim 6, wherein said lysed platelets are platelets lysed via a freeze/thaw cycle.
8. The platelet lysate composition of claim 7, wherein said lysed platelets are platelets lysed via at least two freeze/thaw cycles.
9. The platelet lysate composition of claim 6, wherein said centrifugation comprises a force between 2000 x g and 4000 x g for between 15 and 45 minutes.
10. The platelet lysate composition of claim 6, wherein said centrifugation comprises a force of about 3000 x g for about 30 minutes.
11. The platelet lysate composition of claim 1, wherein said platelet lysate composition comprises greater than 200 pg of VEGF polypeptide per mL.

12. The platelet lysate composition of claim 1, wherein culturing 1.4×10^6 mesenchymal stem cells with media containing about five percent of said platelet lysate composition results in greater than 1.4×10^7 cells after three days.
13. The platelet lysate composition of claim 1, wherein said mesenchymal stem cells are adipose derived cells.
14. A platelet lysate composition produced by filtering a lysed platelet preparation through a $0.45 \mu\text{m}$ or smaller filter.
15. The platelet lysate composition of claim 14, wherein said lysed platelet preparation is a lysed apheresis platelet preparation.
16. The platelet lysate composition of claim 14, wherein said filter is a $0.45 \mu\text{m}$ filter.
17. The platelet lysate composition of claim 14, wherein said filter is a $0.2 \mu\text{m}$ filter.
18. The platelet lysate composition of claim 14, wherein said platelet lysate composition is produced by filtering said lysed platelet preparation through a $0.45 \mu\text{m}$ filter and a $0.2 \mu\text{m}$ filter.
19. The platelet lysate composition of claim 14, wherein said lysed platelet preparation comprises supernatant from centrifugation of lysed platelets.
20. The platelet lysate composition of claim 19, wherein said lysed platelets are platelets lysed via a freeze/thaw cycle.
21. The platelet lysate composition of claim 20, wherein said lysed platelets are platelets lysed via at least two freeze/thaw cycles.

22. The platelet lysate composition of claim 19, wherein said centrifugation comprises a force between 2000 x g and 4000 x g for between 15 and 45 minutes.
23. The platelet lysate composition of claim 19, wherein said centrifugation comprises a force of about 3000 x g for about 30 minutes.
24. A method for making a platelet lysate composition, wherein said method comprises filtering a lysed platelet preparation through a 0.45 μm or smaller filter.
25. The method of claim 24, wherein said lysed platelet preparation is a lysed apheresis platelet preparation.
26. The method of claim 24, wherein said filter is a 0.45 μm filter.
27. The method of claim 24, wherein said filter is a 0.2 μm filter.
28. The method of claim 24, wherein said method comprises filtering said lysed platelet preparation through a 0.45 μm filter and a 0.2 μm filter.
29. The method of claim 24, wherein said lysed platelet preparation comprises supernatant from centrifugation of lysed platelets.
30. The method of claim 29, wherein said lysed platelets are platelets lysed via a freeze/thaw cycle.
31. The method of claim 30, wherein said lysed platelets are platelets lysed via at least two freeze/thaw cycles.
32. The method of claim 29, wherein said centrifugation comprises a force between 2000 x g and 4000 x g for between 15 and 45 minutes.
33. The method of claim 29, wherein said centrifugation comprises a force of about 3000 x g for about 30 minutes.

34. A method for making a platelet lysate composition, wherein said method comprises:
- (a) lysing platelets via one or more freeze/thaw cycles to obtain lysed platelets,
 - (b) centrifuging said lysed platelets to obtain a supernatant, and
 - (c) filtering said supernatant through a 0.45 μm or smaller filter to obtain a filtrate, wherein said filtrate is said platelet lysate composition.
35. The method of claim 34, wherein said lysing step comprises at least two freeze/thaw cycles.
36. The method of claim 34, wherein said centrifuging step comprises using a force between 2000 x g and 4000 x g for between 15 and 45 minutes.
37. The method of claim 34, wherein said centrifuging step comprises a force of about 3000 x g for about 30 minutes.
38. The method of claim 34, wherein said filter is a 0.45 μm filter.
39. The method of claim 34, wherein said filter is a 0.2 μm filter.
40. The method of claim 34, wherein said filtering step comprising filtering said supernatant through a 0.45 μm filter and a 0.2 μm filter.
41. The method of claim 34, wherein said platelets are apheresis platelets.
42. A method for expanding a cell population comprising culturing a first population of cells in the presence of medium comprising a platelet lysate composition under conditions wherein said first population of cells is expanded to a second population of cells having more cells than said first population, wherein said platelet lysate composition comprises a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter.

43. The method of claim 42, wherein said cells are adult stem cells, primary tumor cells, or tumor stem cells.
44. A method for differentiating stem cells, wherein said method comprises culturing said stem cells in the presence of differentiating medium comprising a platelet lysate composition under conditions wherein said stem cells differentiate, wherein said platelet lysate composition comprises a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter.
45. A method for healing a wound, wherein said method comprising contacting said wound with a platelet lysate composition, wherein said platelet lysate composition comprises a filtrate from a lysed platelet preparation passed through a 0.45 μm or smaller filter.
46. The method of claim 45, wherein said wound is a cut, fistula, or diabetic ulcer.
47. The method of claim 45, wherein said contacting step comprises using a stitch or glue comprising said platelet lysate composition to contact said wound.
48. The method of claim 45, wherein said contacting step comprises spraying said platelet lysate composition onto said wound.
49. A composition containing plasma and platelet contents from a platelet preparation passed through a 0.45 μm or smaller filter.
50. The composition of claim 49, wherein said plasma is human plasma.
51. The composition of claim 49, wherein said platelet contents are human platelet contents.
52. A composition comprising medium and a platelet plasma culture supplement.
53. The composition of claim 52, wherein said platelet plasma culture supplement is a platelet lysate comprising plasma.

54. The composition of claim 52, wherein said platelet plasma culture supplement is a filtrate obtained from filtering a platelet apheresis preparation comprising plasma.

55. The composition of claim 54, wherein said platelet apheresis preparation is a platelet apheresis preparation collected from a human over three days earlier.

56. The composition of claim 54, wherein said platelet apheresis preparation is a platelet apheresis preparation collected from a human over five days earlier.

57. The composition of claim 54, wherein said platelet apheresis preparation is a platelet apheresis preparation collected from a human over six days earlier.

58. The composition of claim 52, wherein said platelet plasma culture supplement is a supernatant obtained from spinning a platelet apheresis preparation comprising plasma.

59. The composition of claim 54, wherein said platelet apheresis preparation is a platelet apheresis preparation collected from a human over ten days earlier.

60. A method for obtaining a platelet plasma culture supplement comprising removing platelets or platelet debris from a platelet apheresis preparation such that at least some of the plasma within said platelet apheresis preparation is retained, thereby obtaining said platelet plasma culture supplement, wherein said platelet plasma culture supplement comprises the ability to induce differentiation of 1×10^6 adult stem cells into at least 1×10^3 progenitors of cardiomyocytes, adipocytes, osteoblasts, chondrocytes, myocytes, or nerve cells within 60 days when added to medium at a concentration of 5 percent as a base media with supplementation directed at the desired cell type.

61. An isolated cell culture comprising MCGBM103 cells cultured in medium comprising a platelet plasma culture supplement.

62. An isolated cell culture comprising MCGBM106 cells cultured in medium comprising a platelet plasma culture supplement.
63. An isolated cell culture comprising cells cultured in medium comprising a platelet plasma culture supplement, wherein said cells grow in said medium at a rate that is faster than the rate of control cells cultured in control medium comprising 10 percent fetal bovine serum.
64. The isolated cell culture of claim 63, wherein said cells and said control cells are primary tumor cells.
65. The isolated cell culture of claim 64, wherein said cells and said control cells are human primary glioma cells.
66. The isolated cell culture of claim 63, wherein said cells and said control cells are stem cells.
67. The isolated cell culture of claim 63, wherein said cells and said control cells are fibroblast cells.
68. The isolated cell culture of claim 63, wherein said medium and said control medium are neurobasal medium or DMEM/F12 medium.
69. The isolated cell culture of claim 63, wherein 1×10^5 of said cells grow in said medium to more than 1×10^6 of said cells faster than 1×10^5 of said control cells cultured in said control medium grow to 1×10^6 of said control cells.
70. The isolated cell culture of claim 63, wherein 1×10^5 of said cells grow in said medium to more than 3×10^7 of said cells faster than 1×10^5 of said control cells cultured in said control medium grow to 3×10^7 of said control cells.
71. The isolated cell culture of claim 63, wherein said cells are primary glioma cells, and wherein 5×10^5 of said cells grow in said medium to more than 1×10^7 of said cells within 30 days.

72. The isolated cell culture of claim 63, wherein said cells are primary glioma cells, and wherein 5×10^5 of said cells grow in said medium to more than 1×10^7 of said cells within 60 days.
73. The isolated cell culture of claim 63, wherein said cells are stem cells, and wherein 1×10^5 of said cells grow in said medium to more than 1×10^7 of said cells within 14 days.
74. The isolated cell culture of claim 63, wherein said cells are stem cells, and wherein 1×10^5 of said cells grow in said medium to more than 1×10^7 of said cells within 20 days.
75. The isolated cell culture of claim 73 or 74, wherein said stem cells are mesenchymal stem cells.
76. The isolated cell culture of claim 73 or 74, wherein said stem cells are adipose-derived, mesenchymal stem cells.
77. The isolated cell culture of claim 63, wherein said cells grow in said medium at a rate that is 1.5 times faster than the rate of said control cells cultured in said control medium.
78. An isolated cell culture comprising tumor cells cultured in medium comprising a platelet plasma culture supplement, wherein said cells grow in said medium in a manner that is more genetically stable than when control cells are cultured in control medium comprising 10 percent fetal bovine serum.
79. The isolated cell culture of claim 78, wherein said tumor cells and said control cells are primary tumor cells.
80. The isolated cell culture of claim 78, wherein said tumor cells and said control cells are human primary glioma cells.

81. The isolated cell culture of claim 78, wherein said medium and said control medium are neurobasal medium or DMEM/F12 medium.
82. The isolated cell culture of claim 78, wherein said tumor cells grown in said medium for four doublings exhibit less than a five percent change in karyotype as compared to said tumor cells grown in said medium for one doubling.
83. The isolated cell culture of claim 78, wherein said tumor cells grown in said medium for four doublings exhibit less than a three percent change in karyotype as compared to said tumor cells grown in said medium for one doubling.
84. The isolated cell culture of claim 78, wherein said tumor cells grown in said medium for ten doublings exhibit less than a five percent change in karyotype as compared to said tumor cells grown in said medium for one doubling.
85. An isolated cell culture comprising stem cells cultured in medium comprising a platelet plasma culture supplement, wherein said stem cells grow in said medium in a manner that is more genetically stable than when control stem cells are cultured in control medium comprising 10 percent fetal bovine serum.
86. The isolated cell culture of claim 85, wherein said stem cells and said control stem cells are mesenchymal stem cells.
87. The isolated cell culture of claim 78, wherein said stem cells and said control stem cells are adipose-derived, mesenchymal stem cells.
88. The isolated cell culture of claim 78, wherein said medium and said control medium are neurobasal medium or DMEM/F12 medium.
89. The isolated cell culture of claim 78, wherein said stem cells grown in said medium for four doublings exhibit less than a five percent change in karyotype as compared to said stem cells grown in said medium for one doubling.

90. The isolated cell culture of claim 78, wherein said stem cells grown in said medium for four doublings exhibit less than a three percent change in karyotype as compared to said stem cells grown in said medium for one doubling.

91. The isolated cell culture of claim 78, wherein said stem cells grown in said medium for ten doublings exhibit less than a five percent change in karyotype as compared to said stem cells grown in said medium for one doubling.

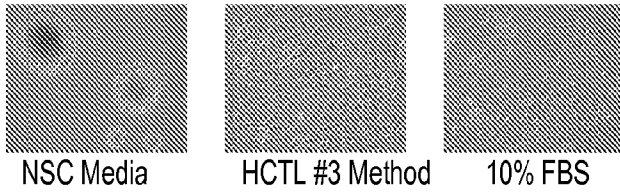
92. A method for obtaining a platelet plasma culture supplement comprising removing platelets or platelet debris from a platelet apheresis preparation such that at least some of the plasma within said platelet apheresis preparation is retained, thereby obtaining said platelet plasma culture supplement, wherein said platelet plasma culture supplement comprises the ability to induce proliferation of less than 1×10^6 adipose-derived, mesenchymal stem cells at passage five into at least 1×10^8 cells within 20 days when added to minimal essential medium at a concentration of 5 percent.

93. The method of claim 92, wherein said platelet plasma culture supplement comprises the ability to induce proliferation of less than 1×10^6 adipose-derived, mesenchymal stem cells at passage five into at least 1×10^8 cells within 15 days when added to minimal essential medium at a concentration of 5 percent.

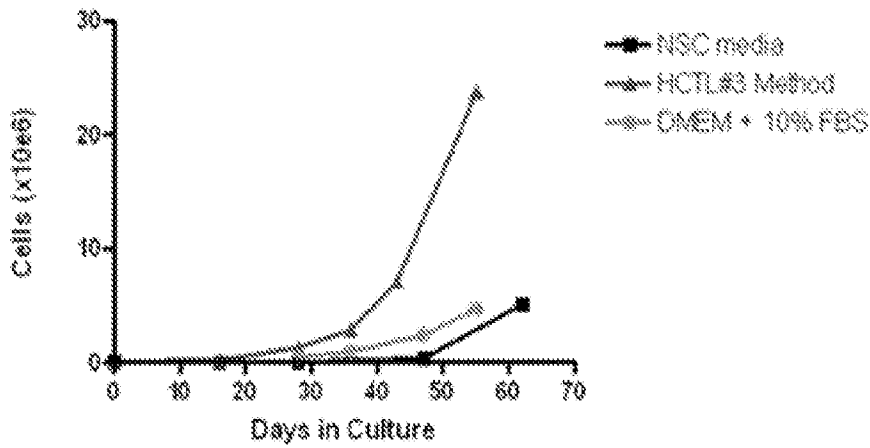
94. A method for obtaining a platelet plasma culture supplement comprising removing platelets or platelet debris from a platelet apheresis preparation such that at least some of the plasma within said platelet apheresis preparation is retained, thereby obtaining said platelet plasma culture supplement, wherein minimal essential medium having 5 percent of said platelet plasma culture supplement is capable of establishing, from newly diagnosed human glioblastoma multiforme biopsy tissue, human glioma cultures having greater than 3×10^7 cells within 60 days at 37°C with a success rate that is greater than 30 percent.

95. The method of claim 94, wherein said success rate is greater than 40 percent.

A



B



C

	CD133	Nestin	SOX2	CD45	CD34	CD90	CD105	HLA Class I	HLA Class II
NSC Media	0.51%	+++	+++	-	++	+++	-	+++	++
HCTL#3	15.58%	+++	++	-	++	+++	+	+++	+
10% FBS	0.45%	+++	++	-	+	+++	++	++	-

D

Neural stem cell media	73- 83,XXXX,add(1)(p22)x2,inv(3)(p23q26)x2,- 4,add(4)(p12), 5,+7,+7,add(9)(p13)x2,10,-10,-14,-14,-16,-16,-17,-17,-22,- 22[cp20]
HCTL #3 media	72- 82,XXXX,add(1)(p22)x2, 2,inv(3)(p23q32)x2,- 4,add(4)(p12), 5,+7,+7,add(9)(p13)x2,10,-10,-14,-14,-16,-16,-17,-17,-19,-22,- 22,+0- 2mar[cp20]
DMEM/10% FBS	69- 75,XXXX,add(1)(p22)x2, 2,inv(3)(p23q32)x2,-4,-5,- 6,+7,+7,add(9)(p13)x2,10,-10,-11,-12,-14,-14,- 15,-16,-16,-17,-17,-19,-22,- 22,+0- 1mar[cp20]

FIG. 1

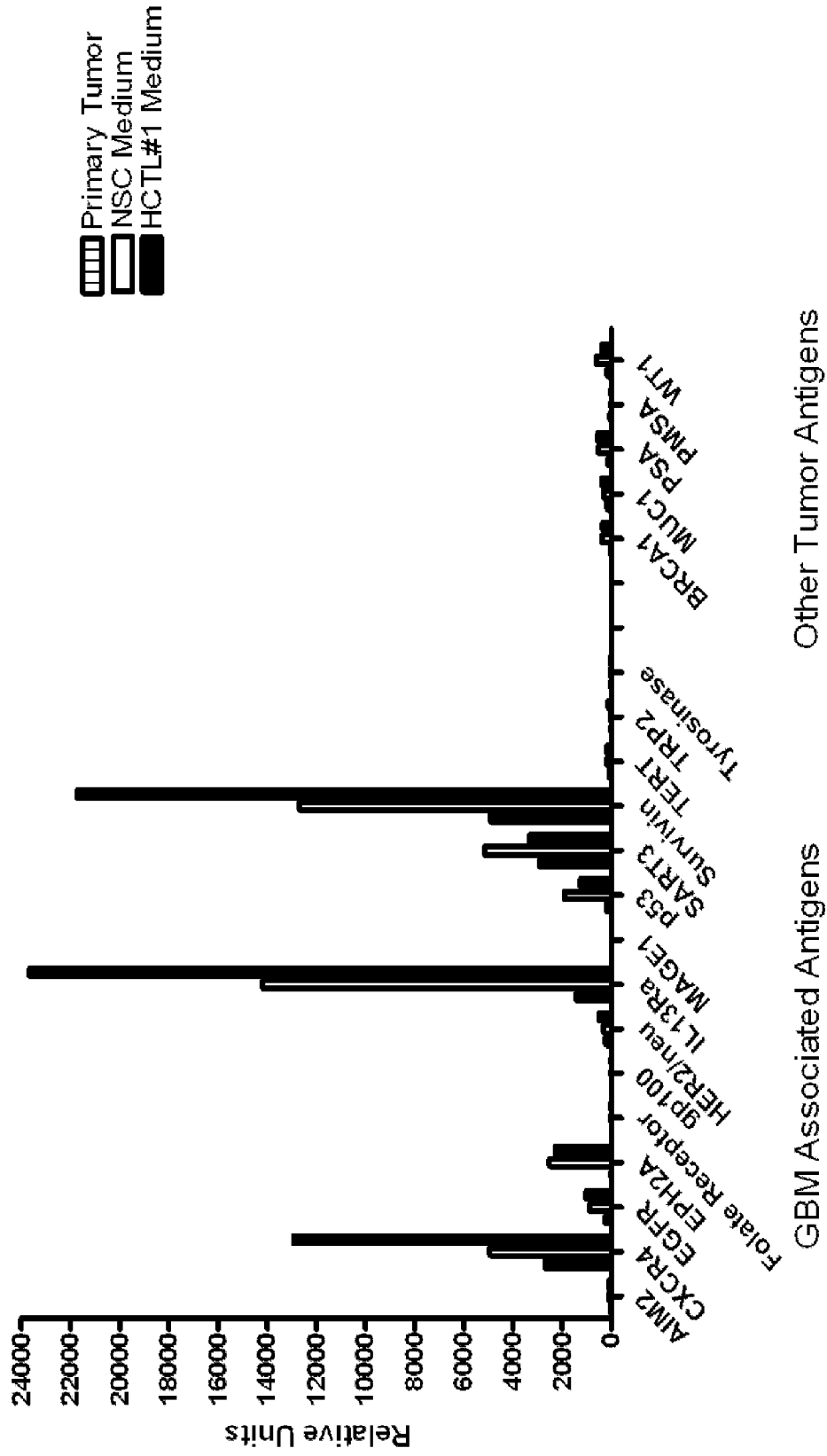


FIG. 2

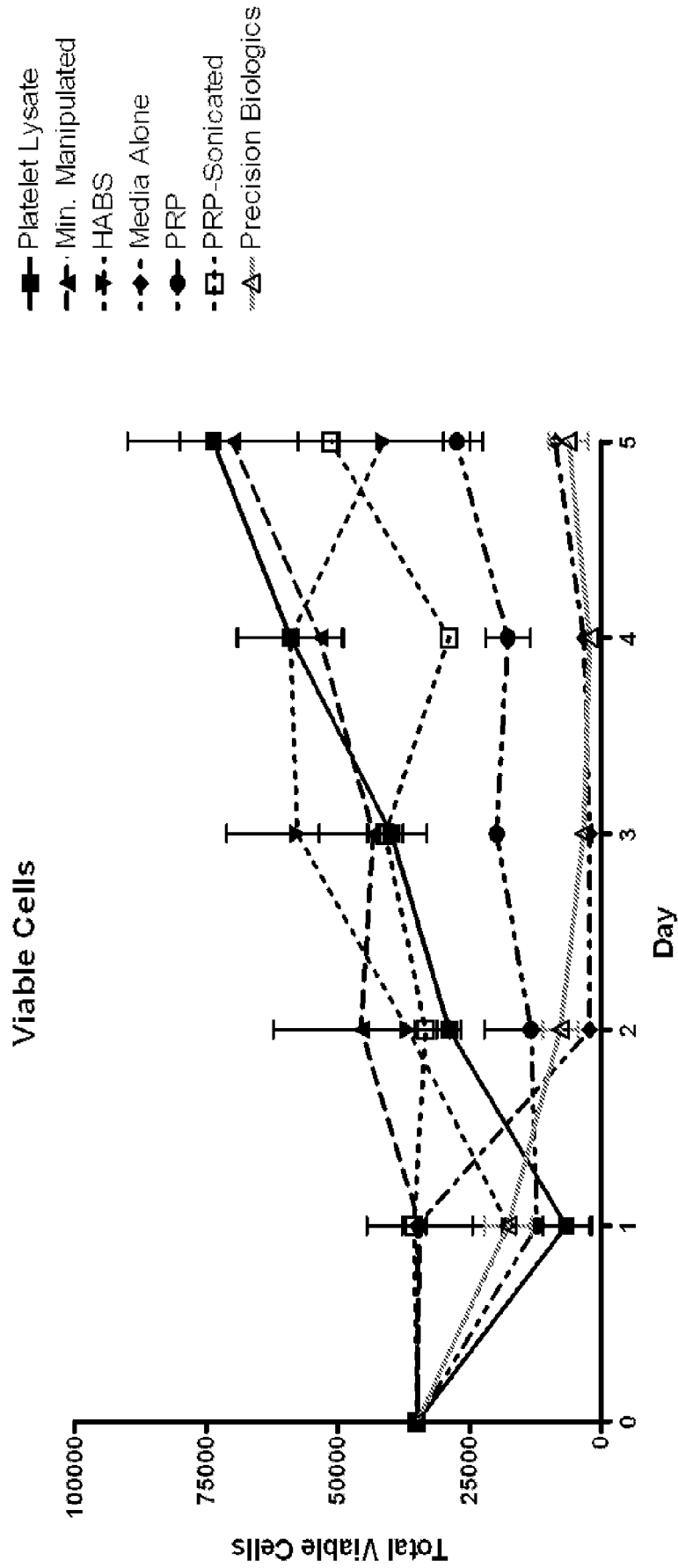


FIG. 3

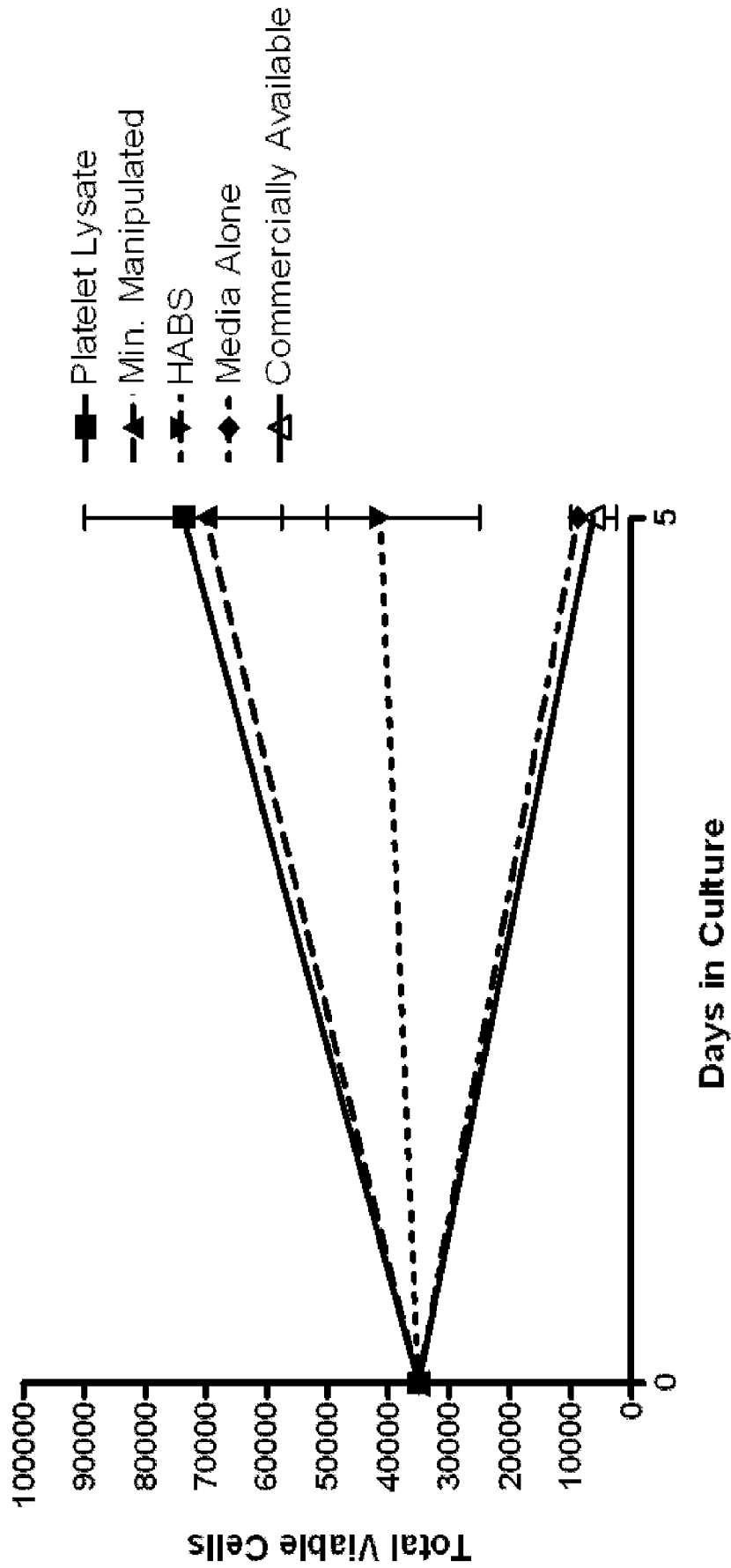


FIG. 4

MSC Growth Kinetics

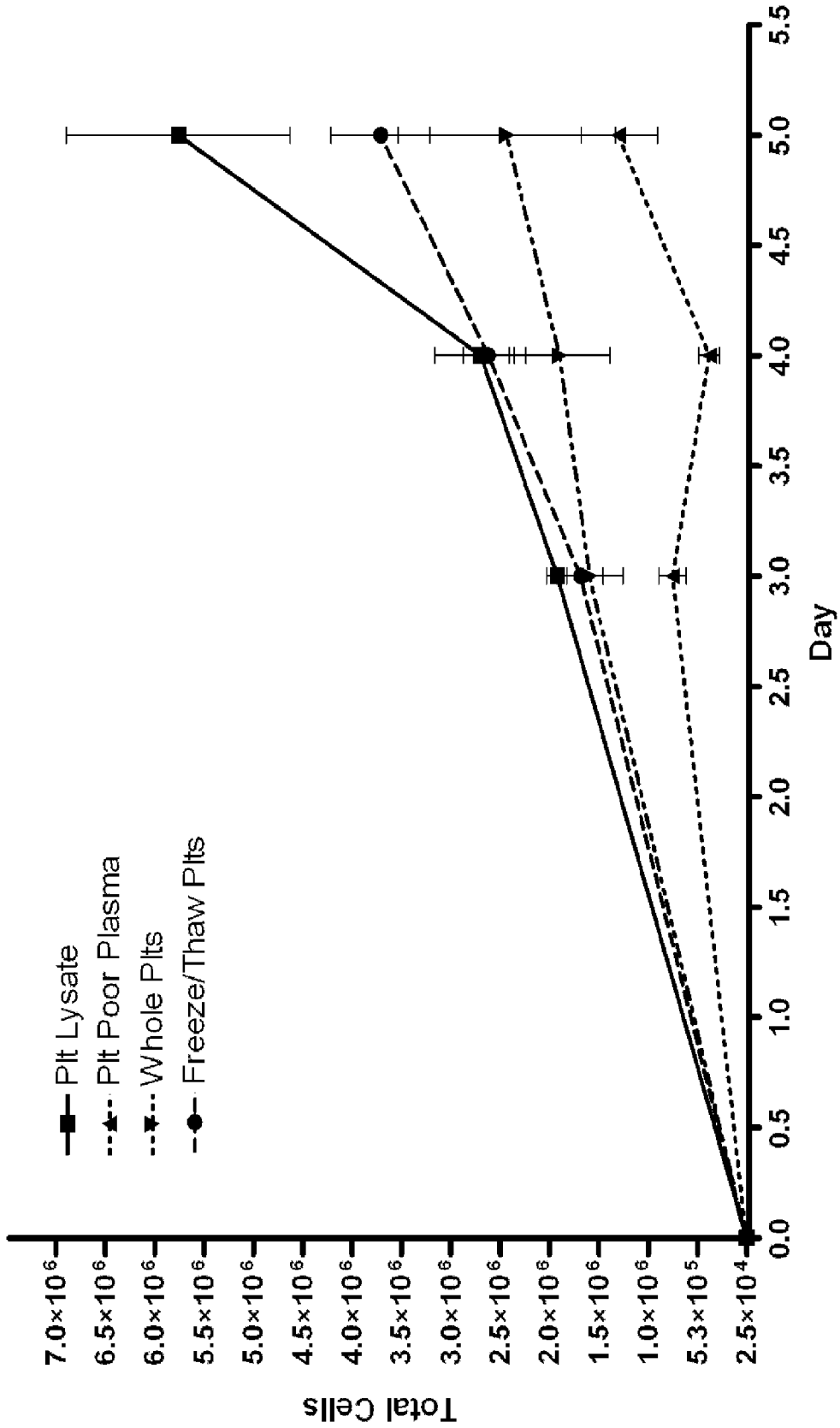


FIG. 5

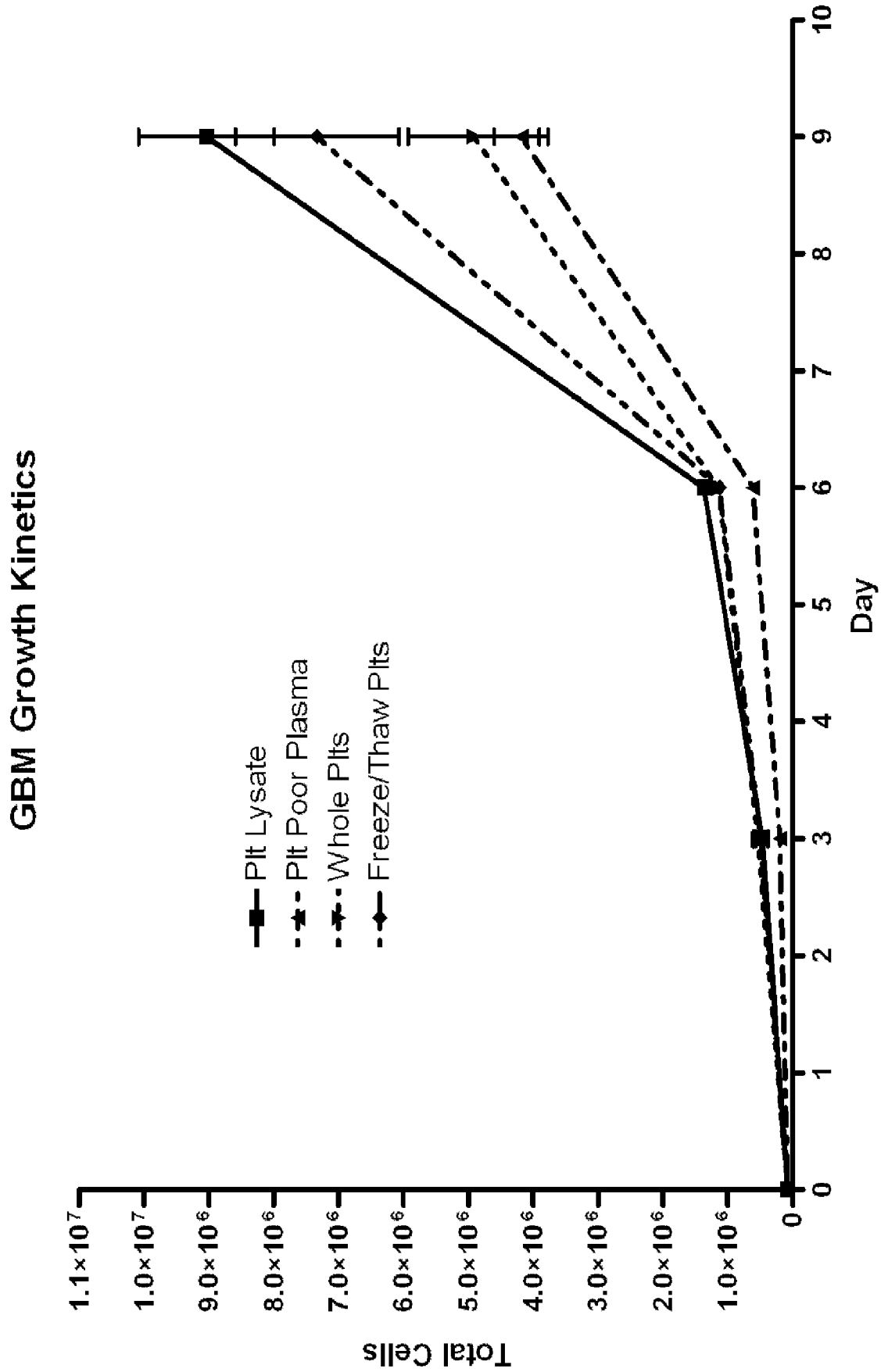


FIG. 6

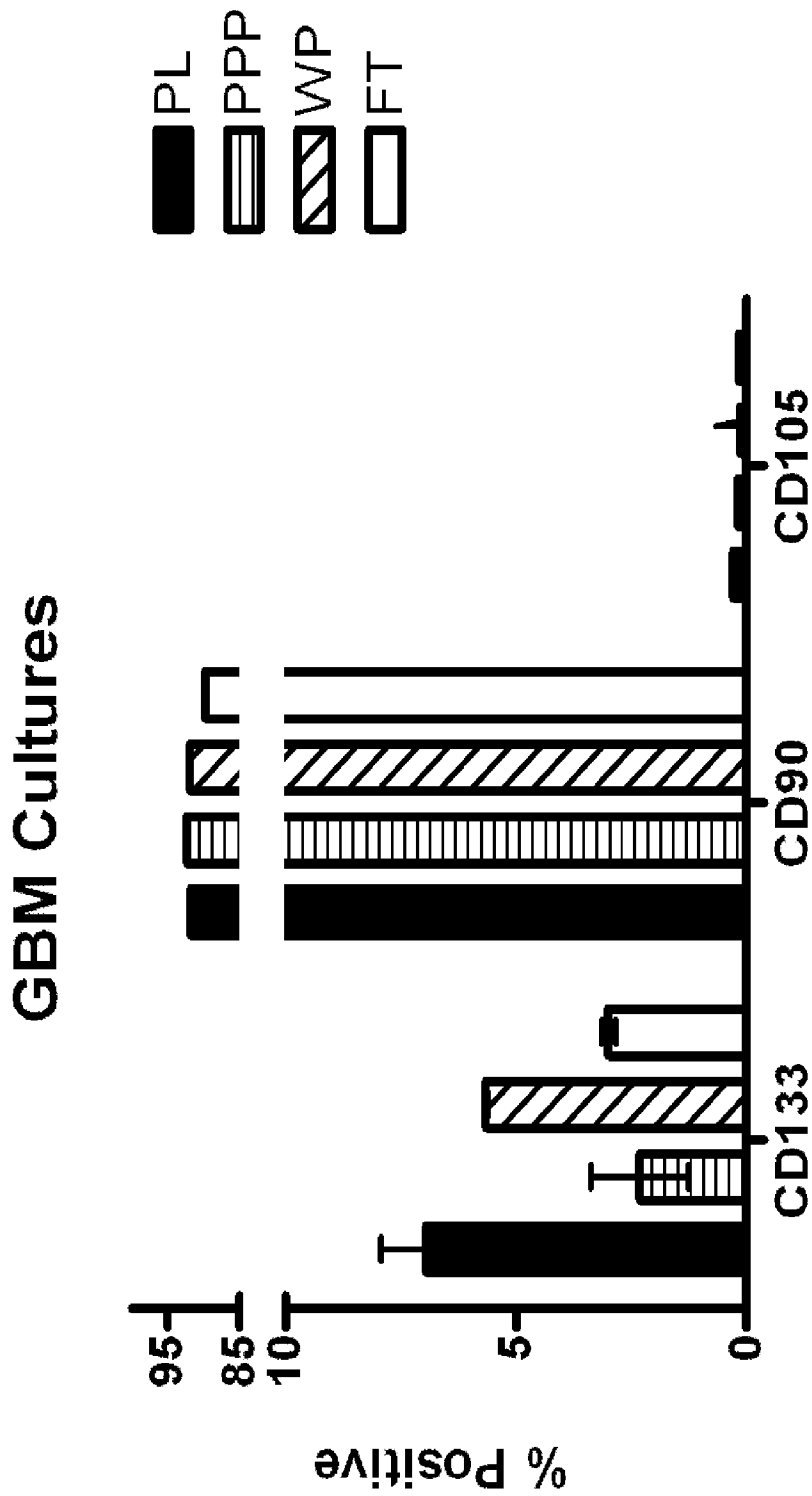


FIG. 7

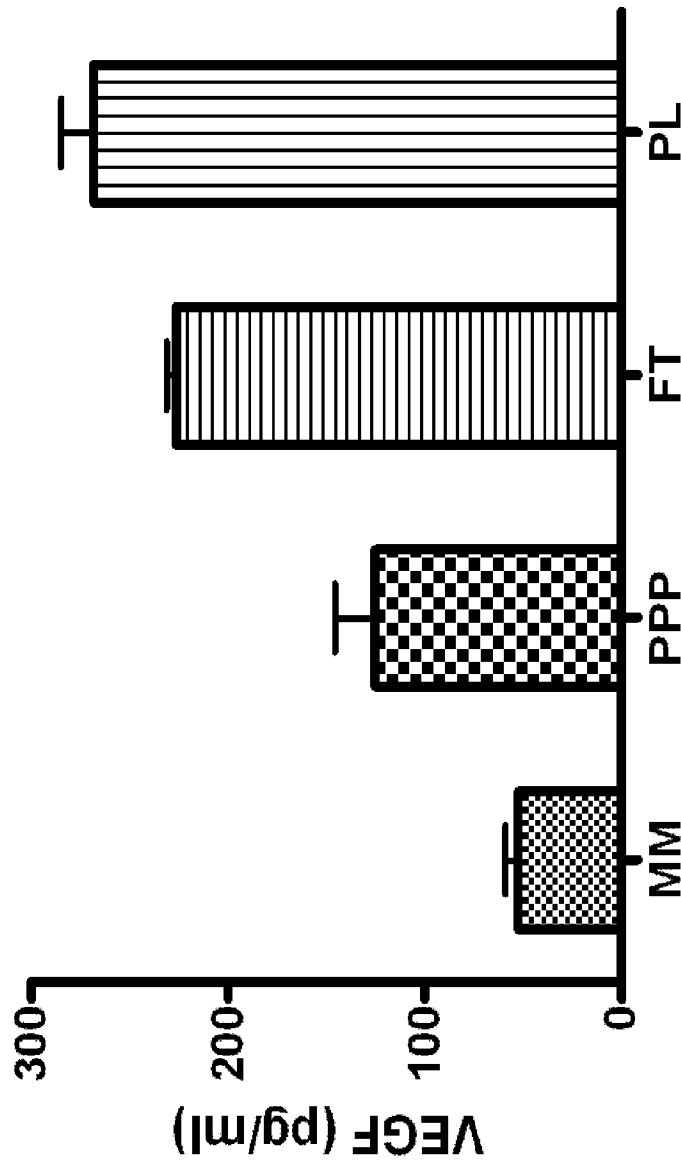


FIG. 8

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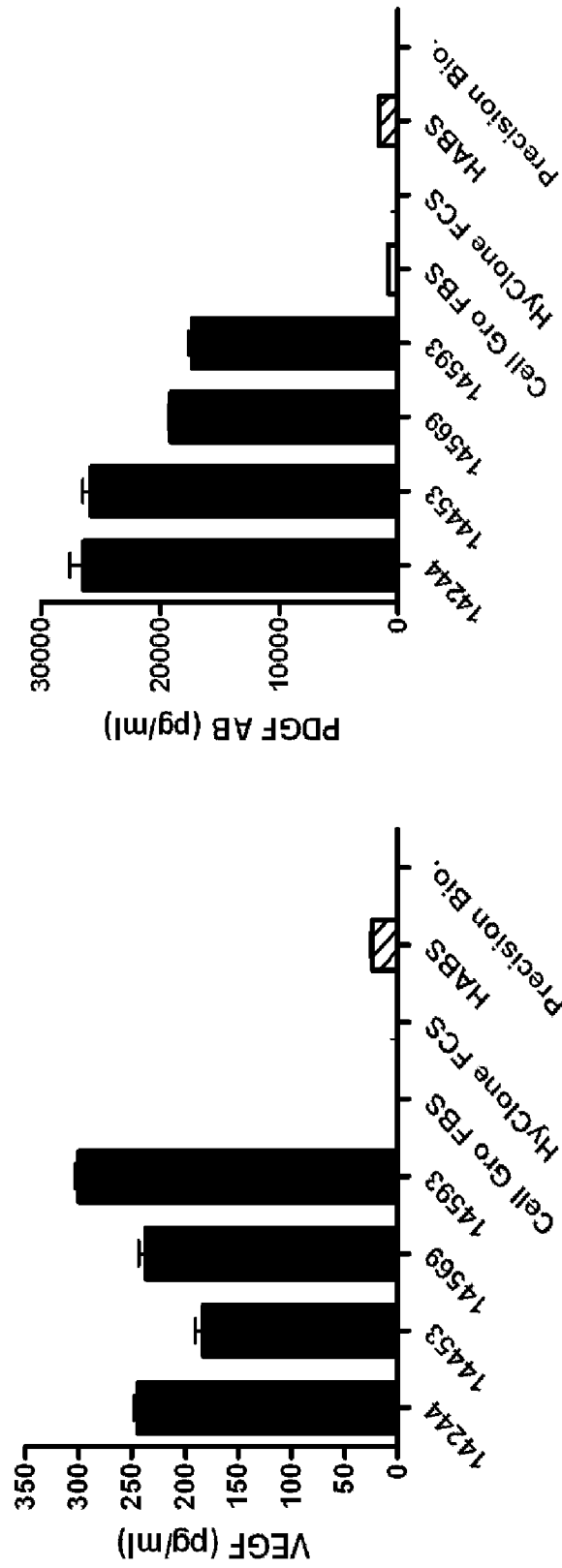


FIG. 9

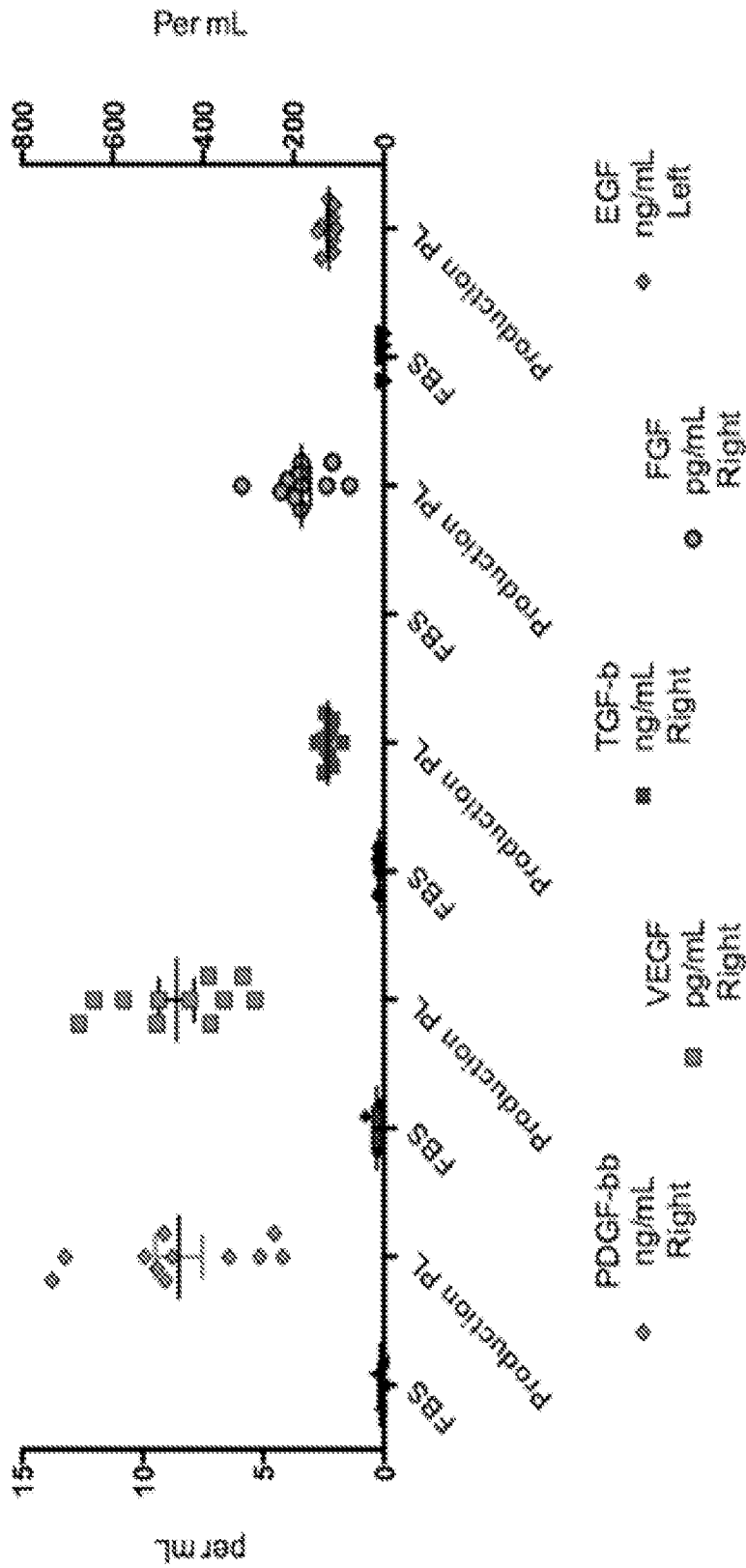


FIG. 10

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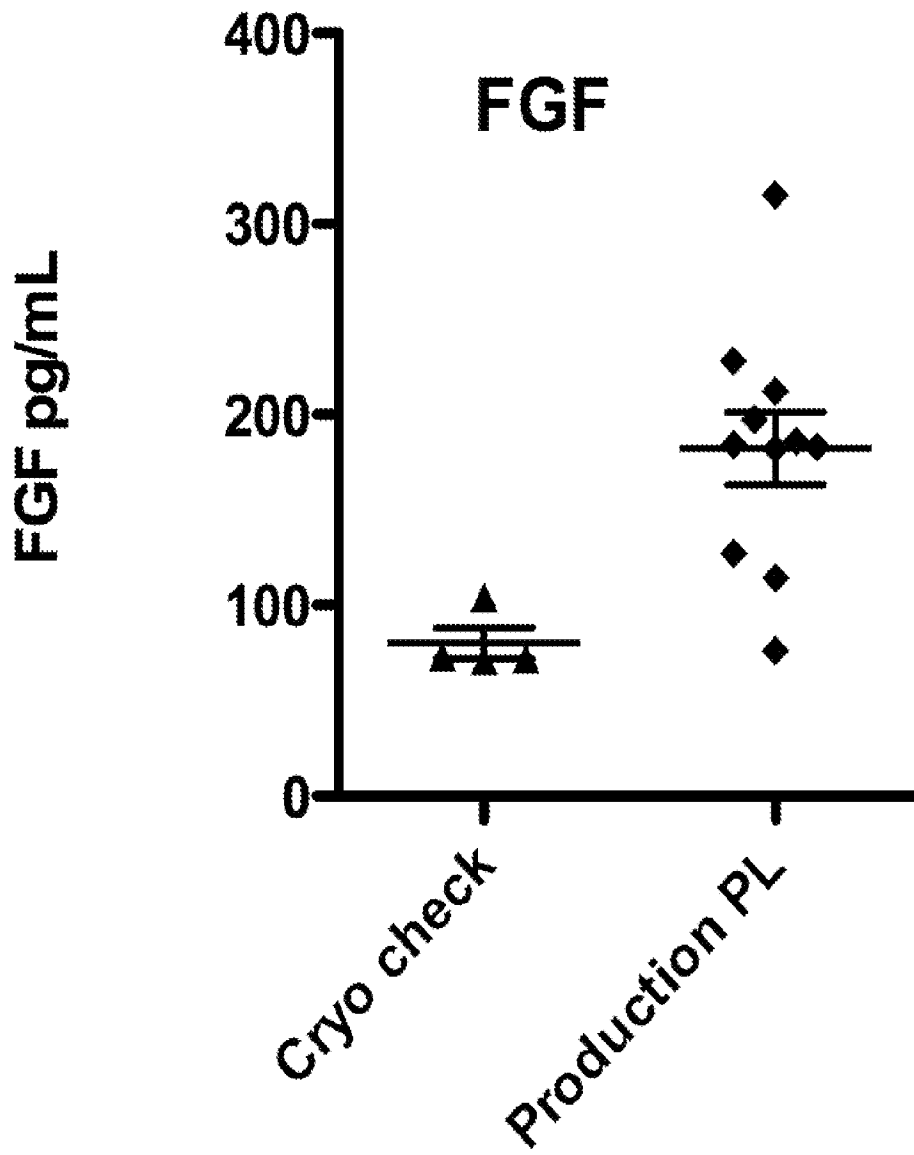


FIG. 11

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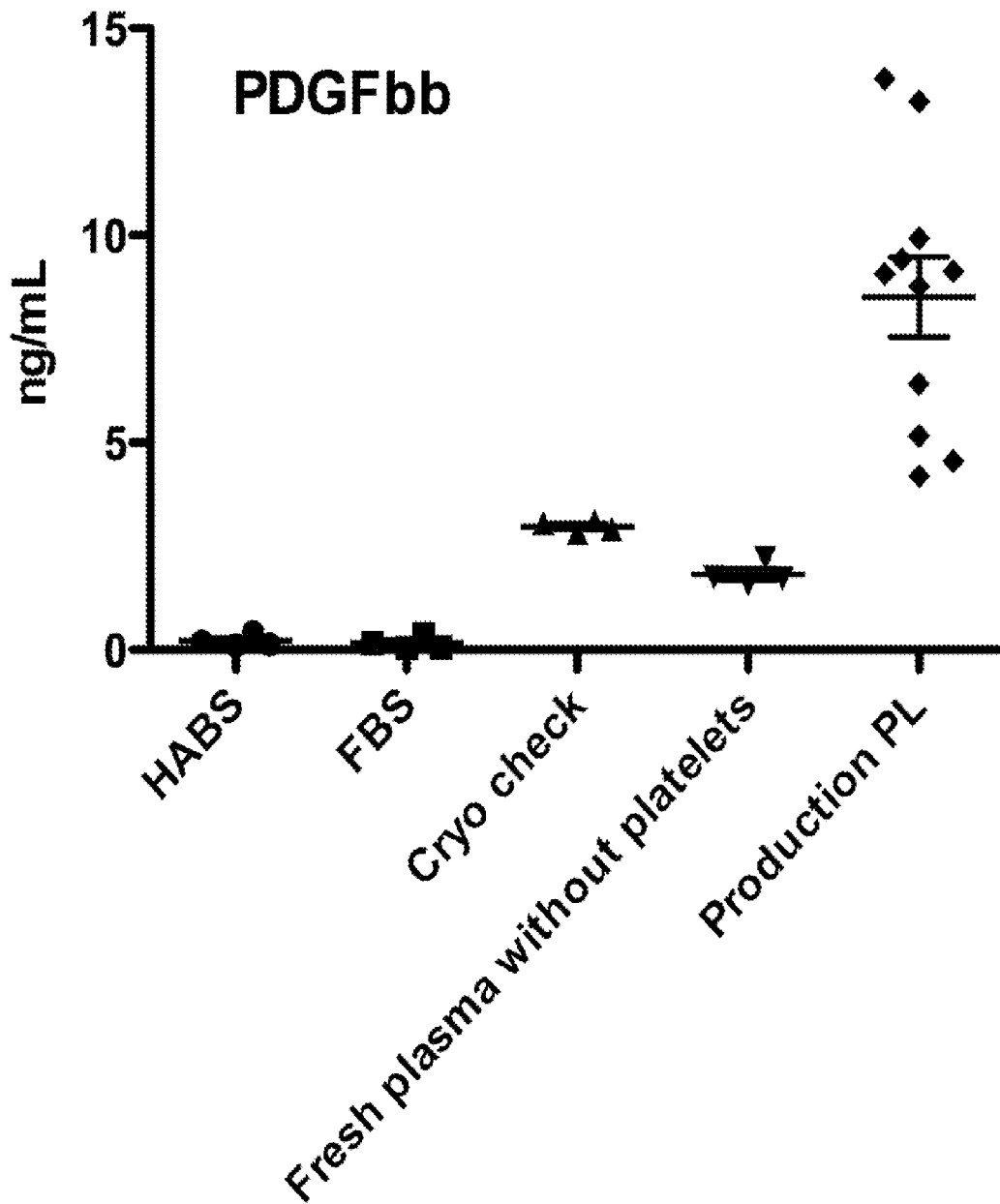


FIG. 12

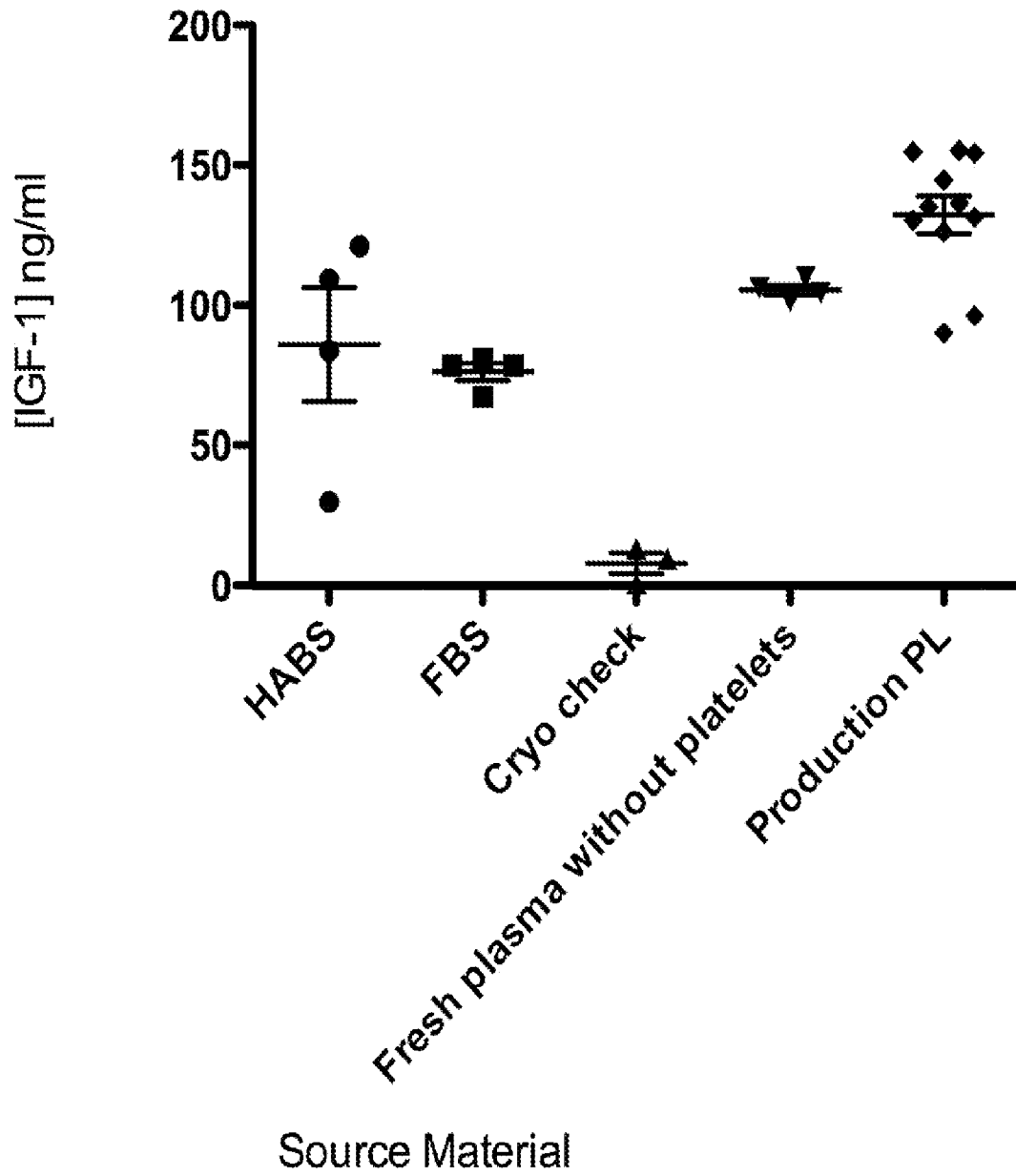


FIG. 13

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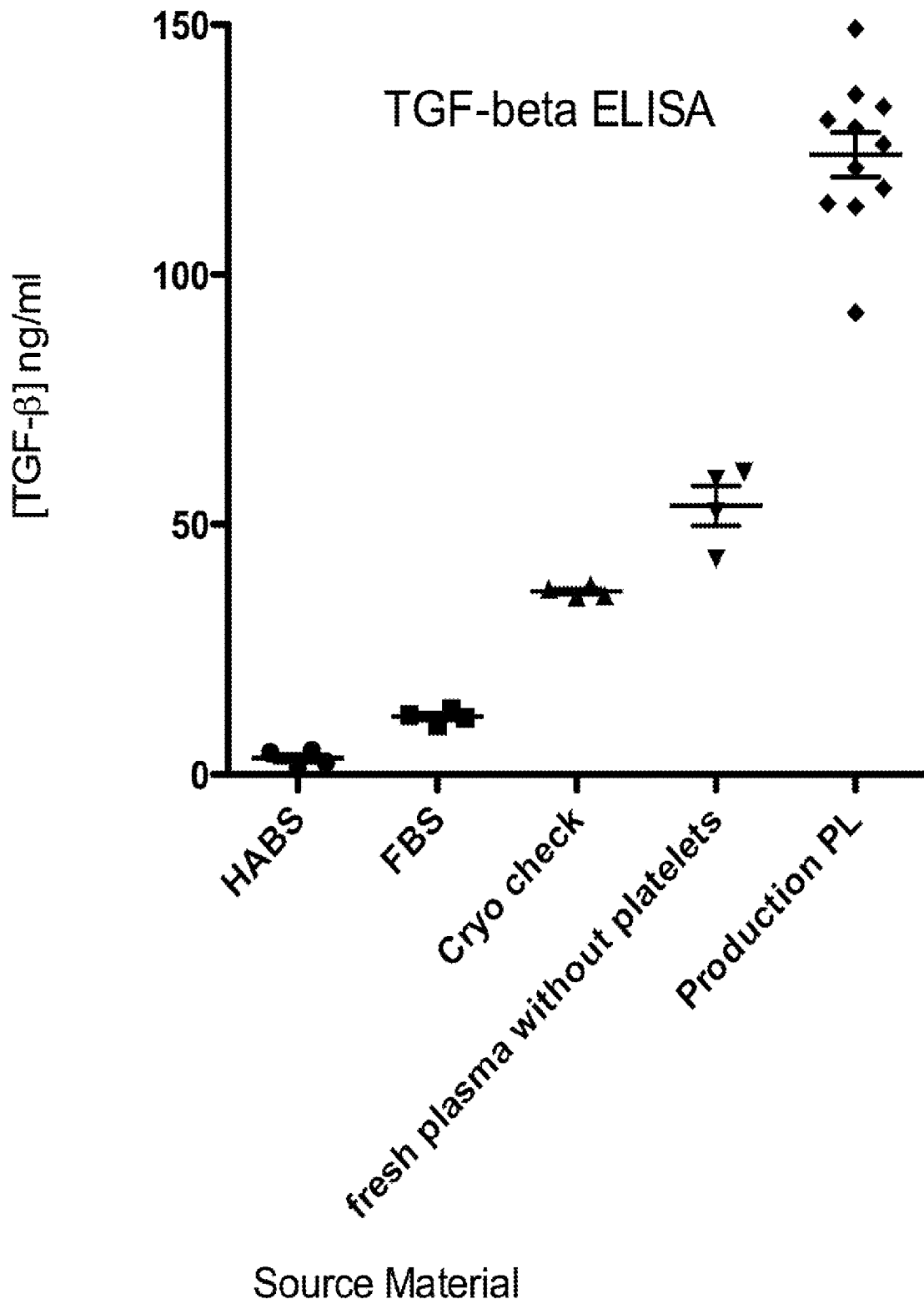


FIG. 14

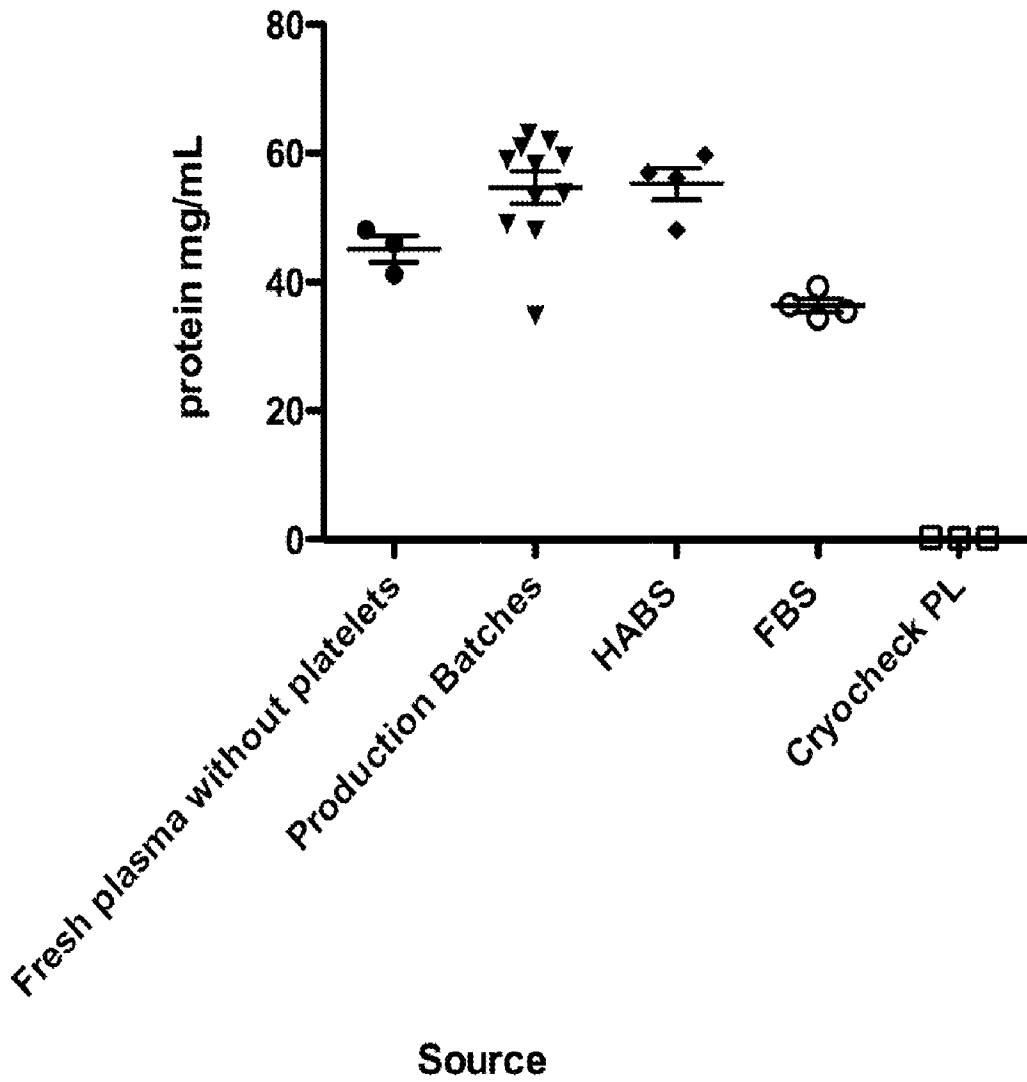


FIG. 15

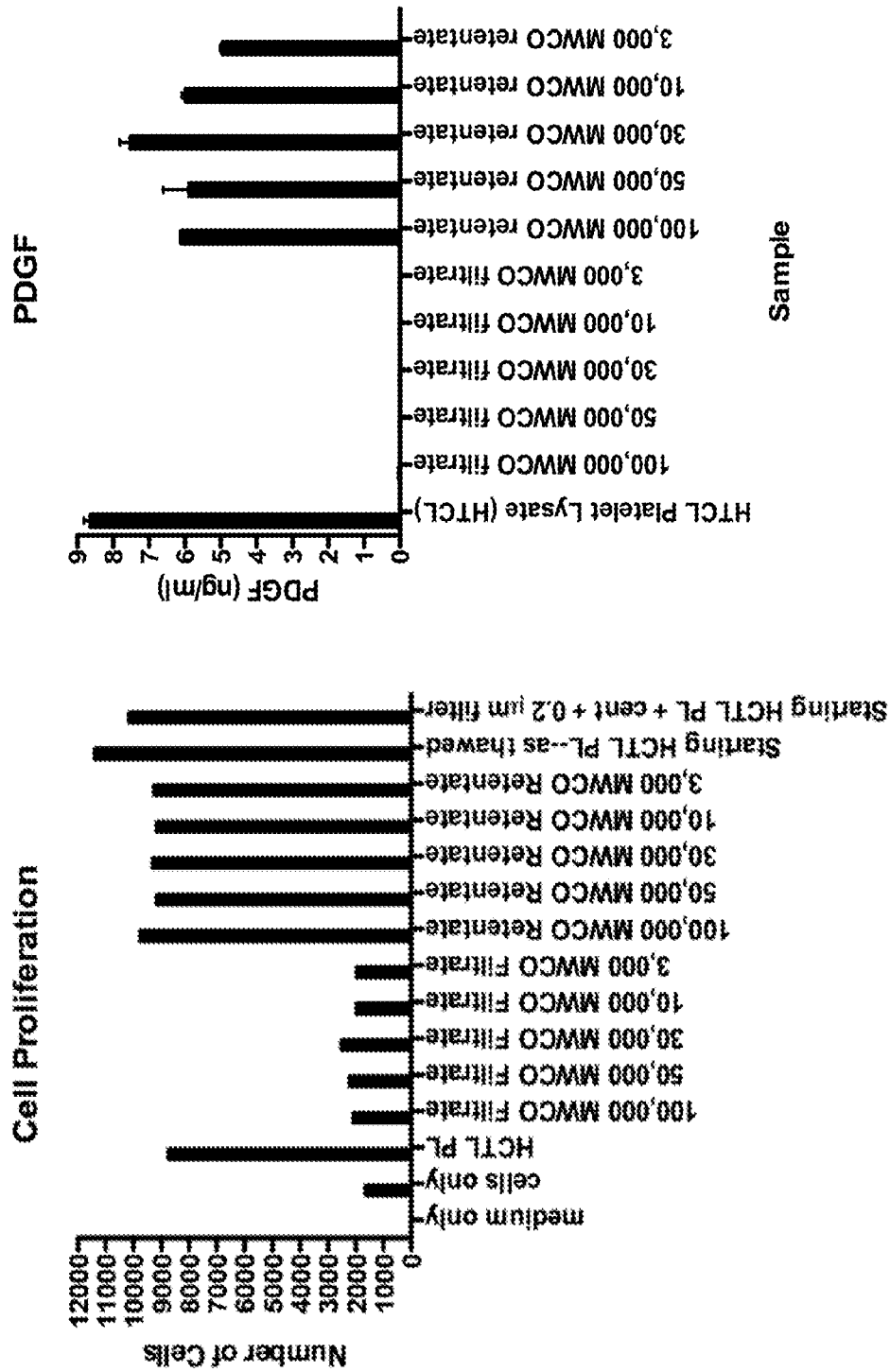


FIG. 16

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G-150: HCTL PL

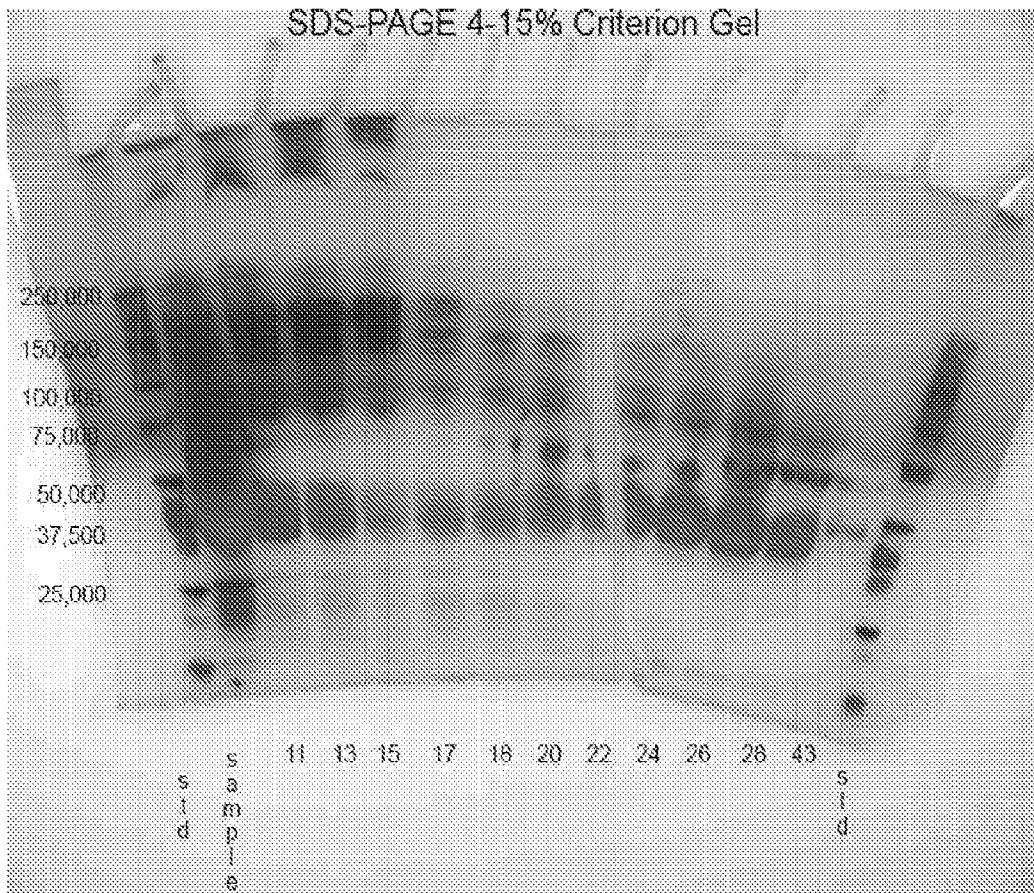
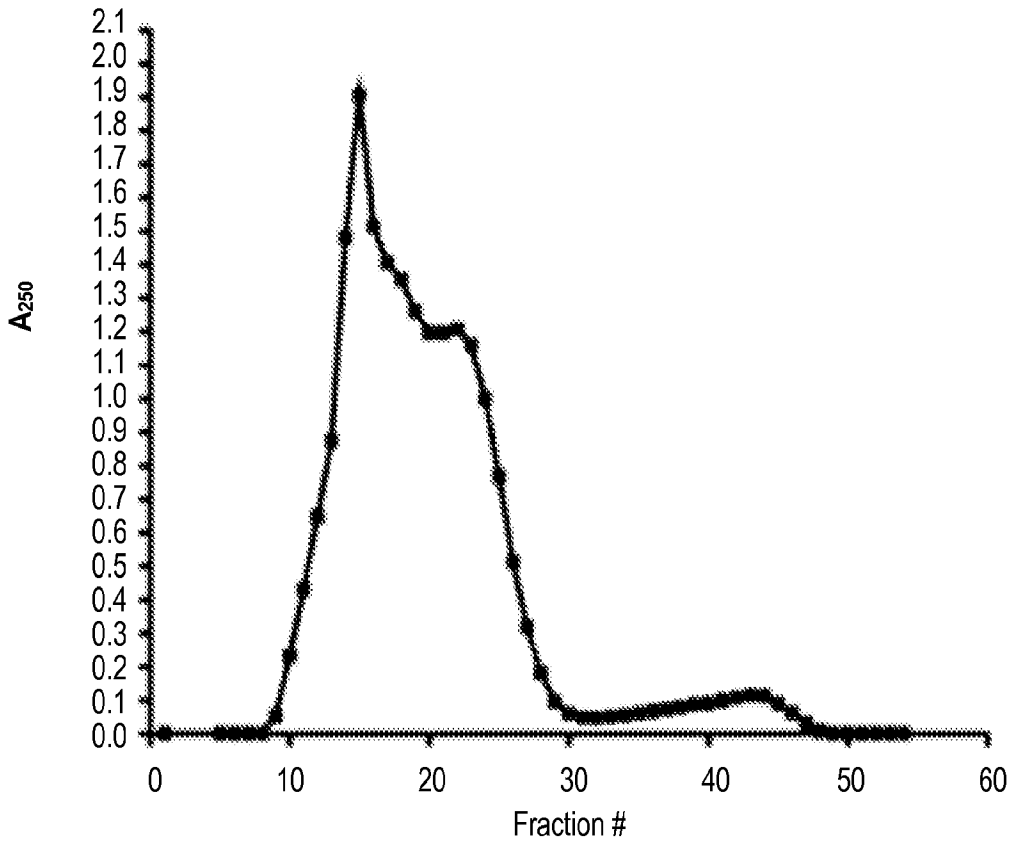


FIG. 17

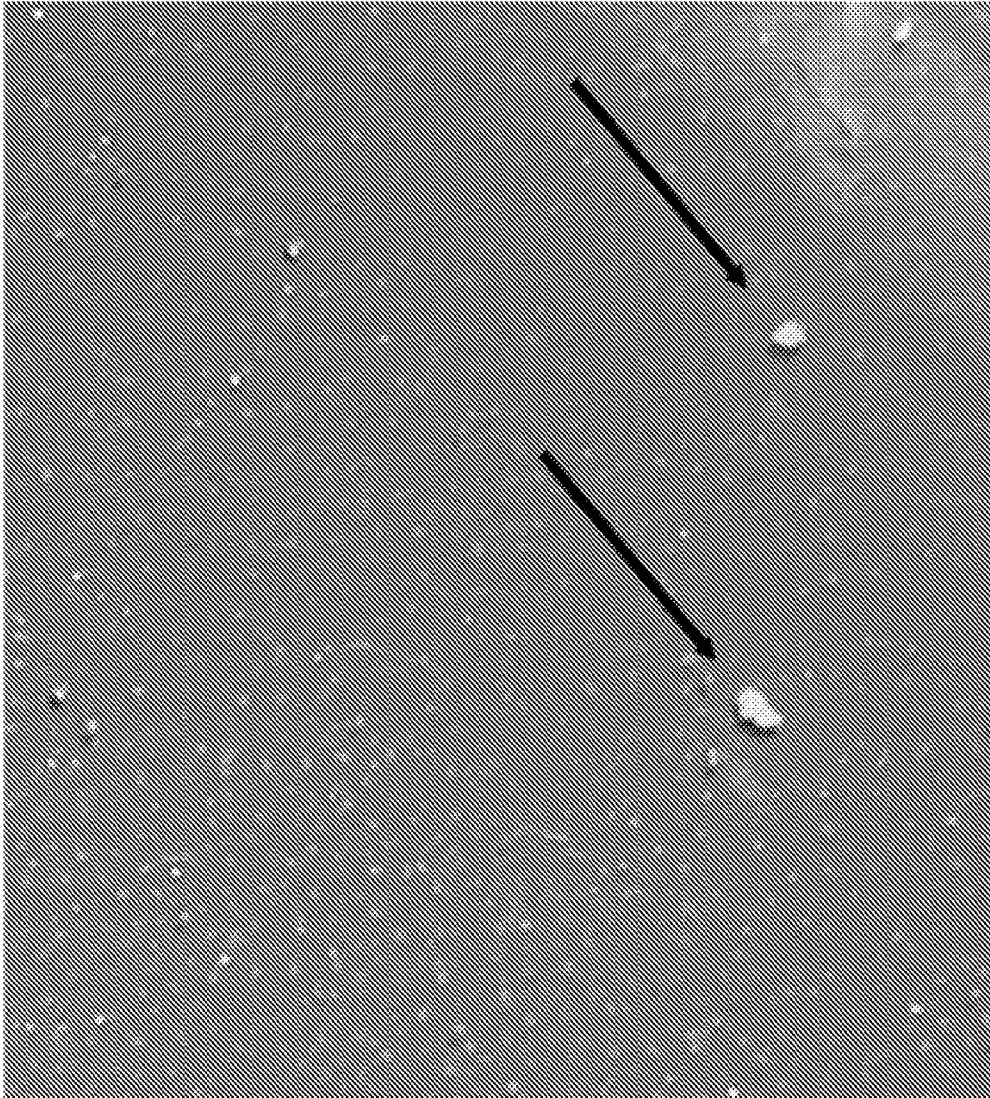


FIG. 18

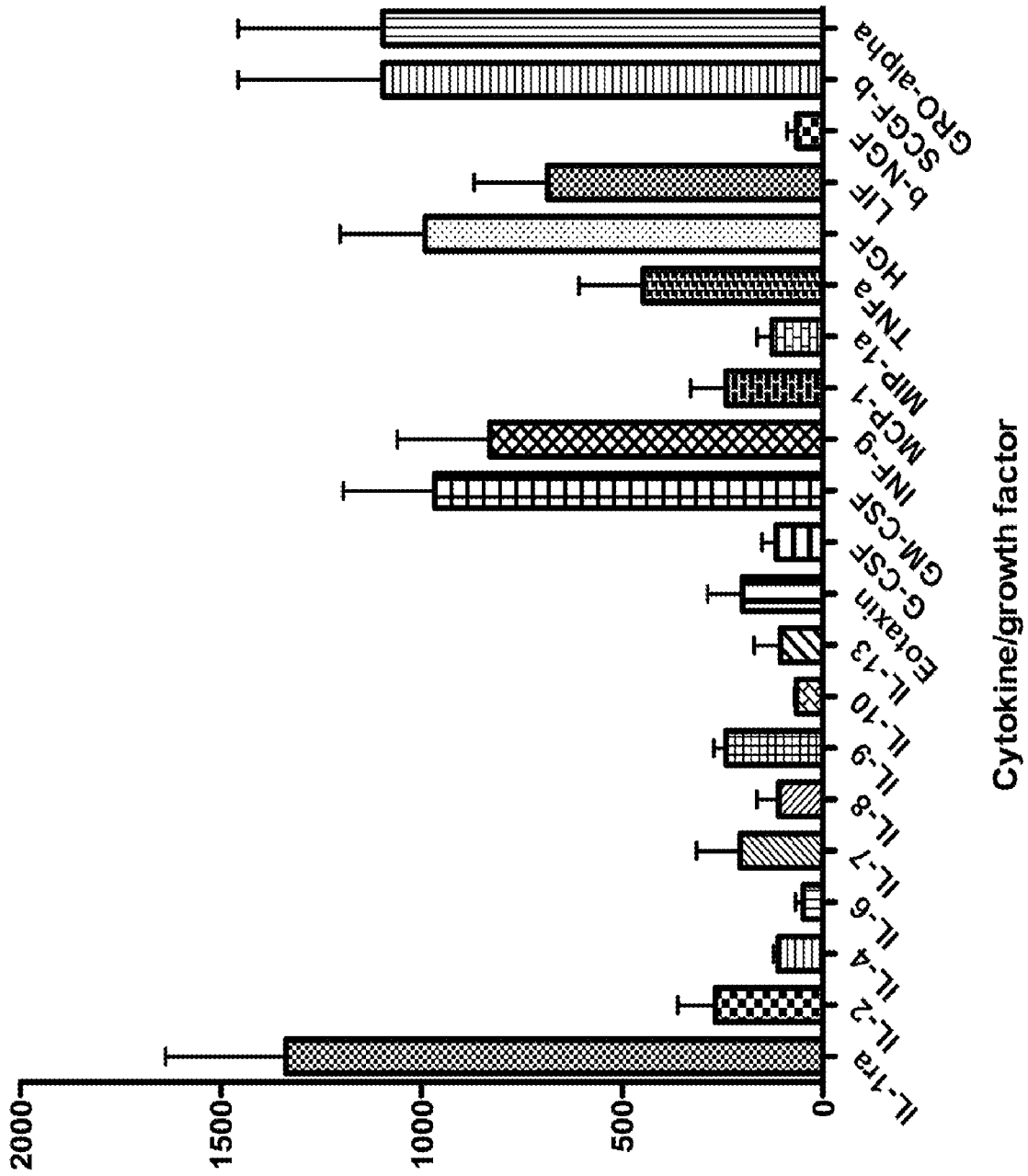
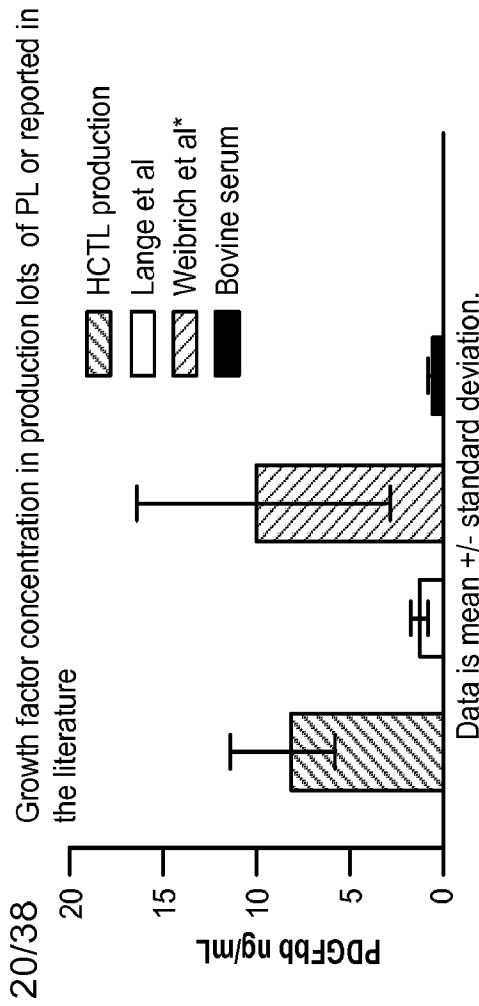


FIG. 19



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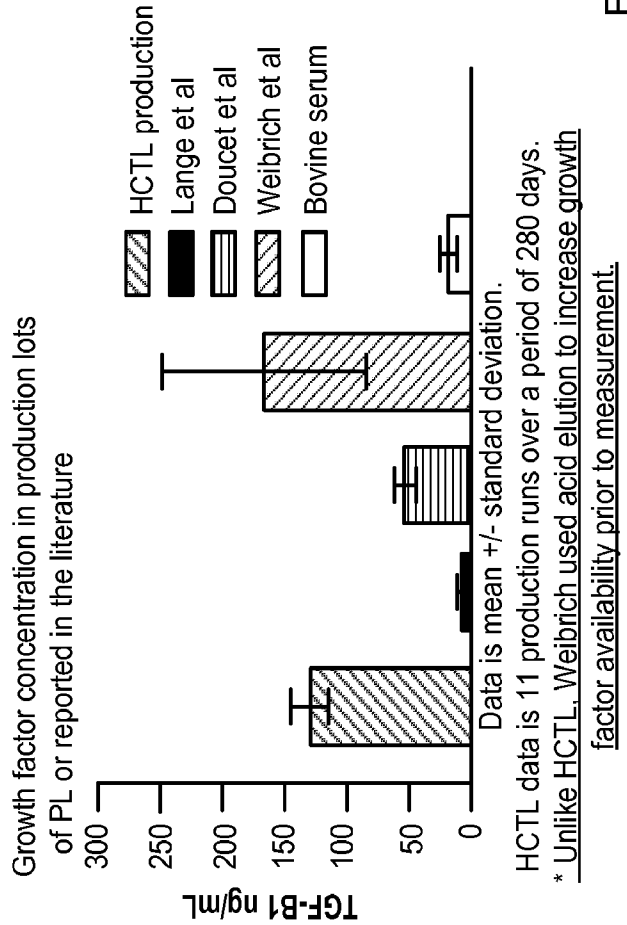
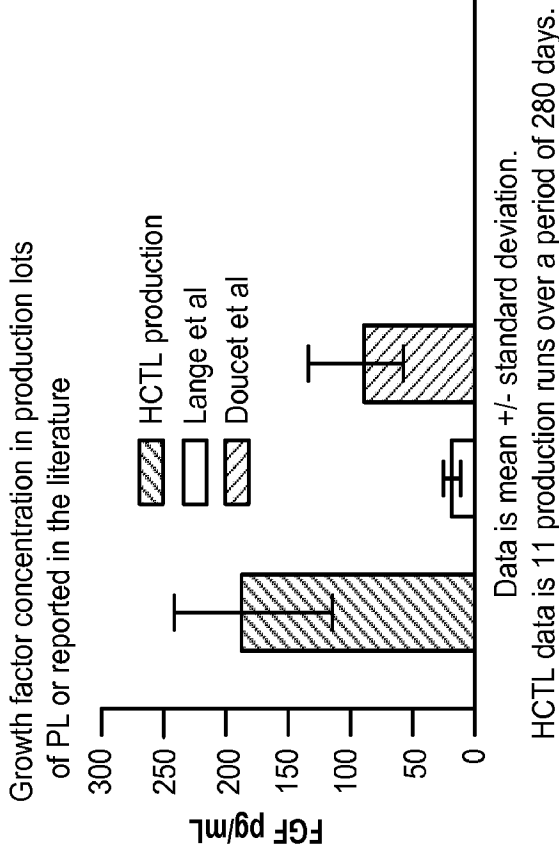
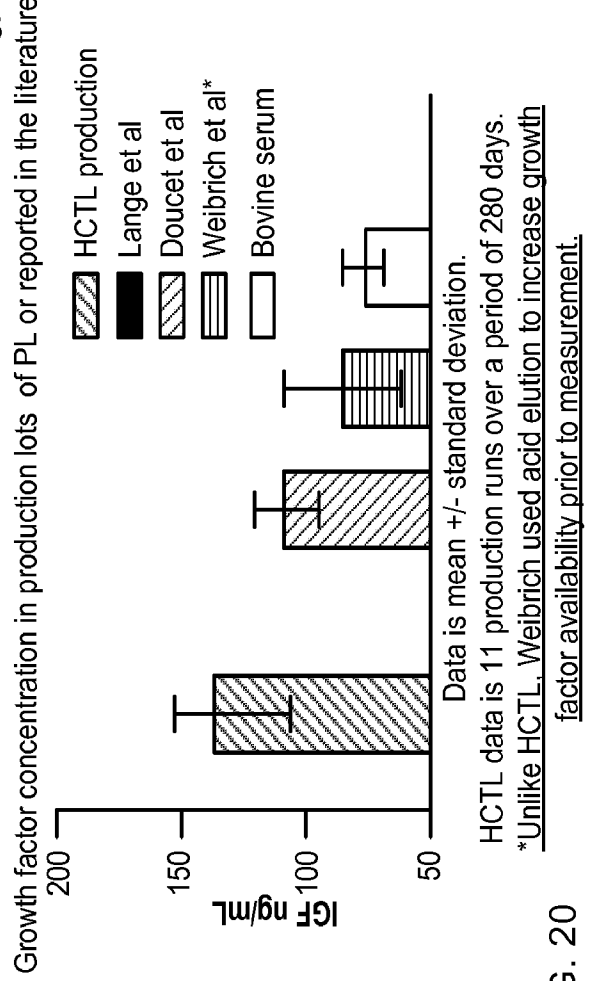


FIG. 20

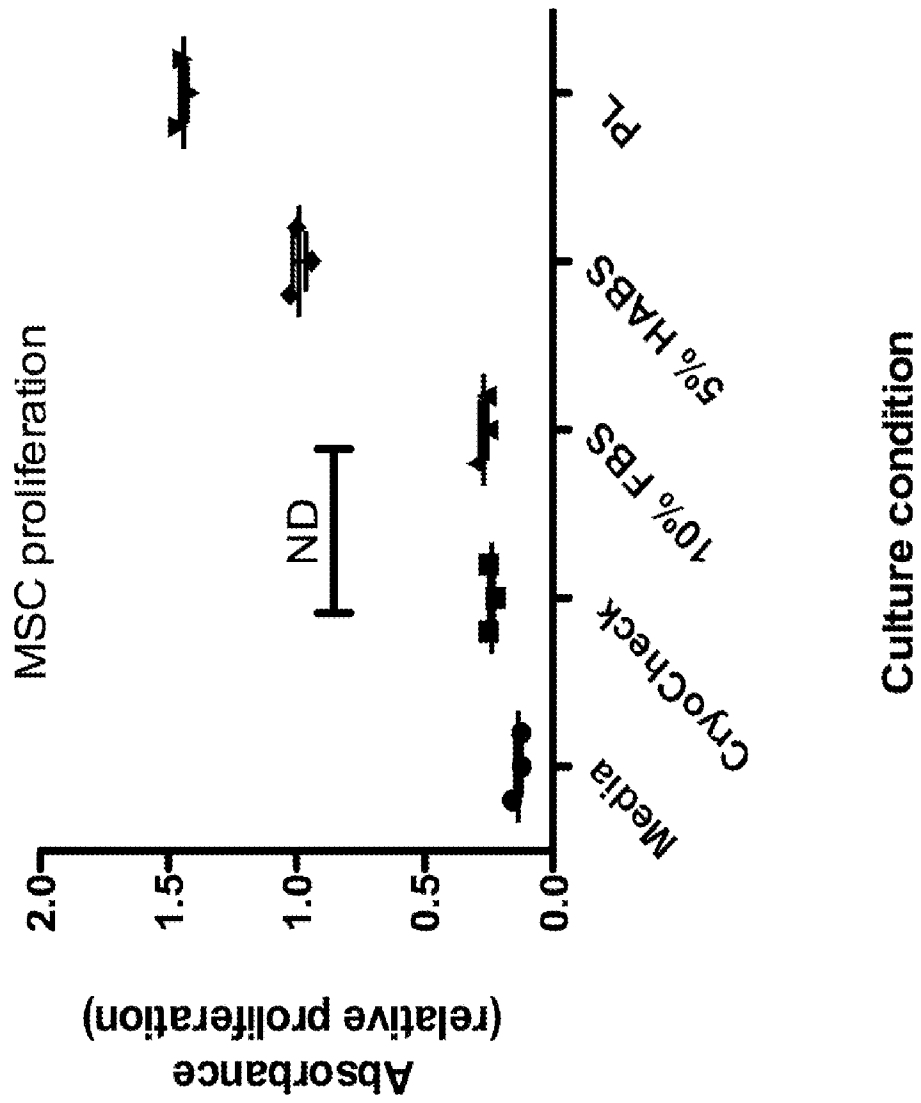
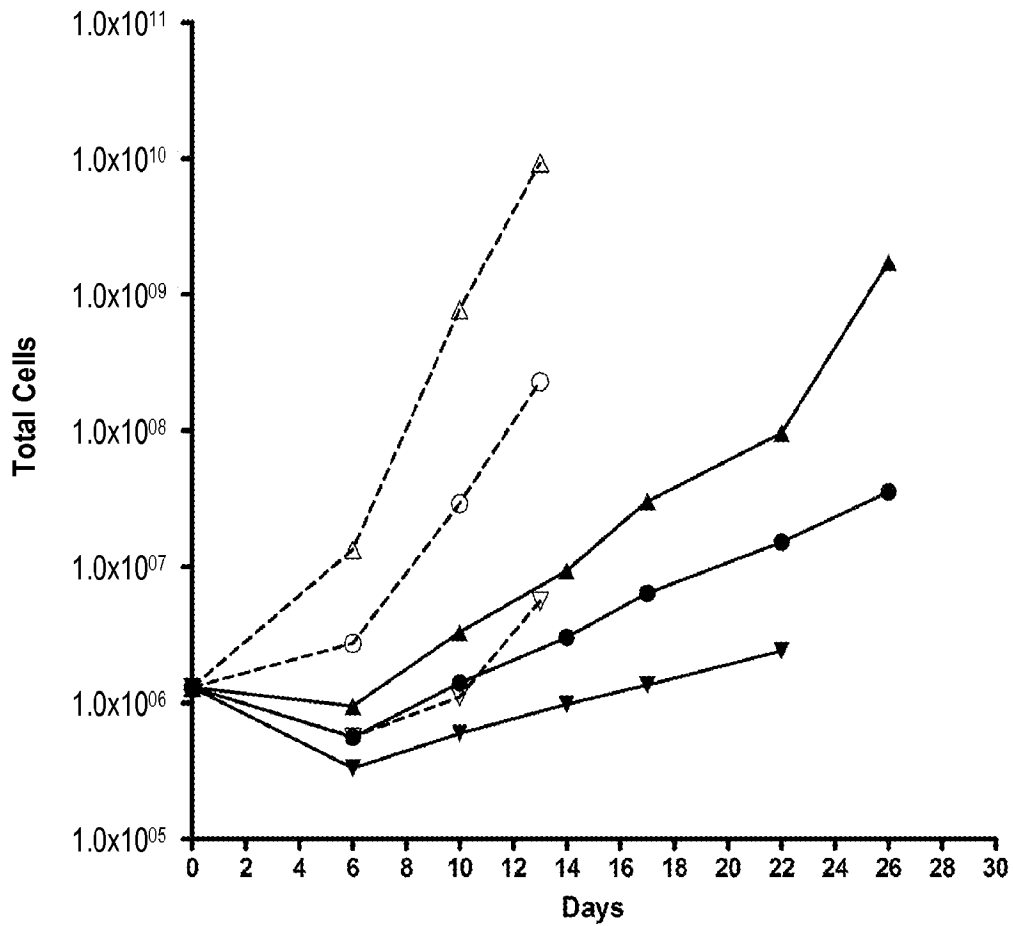


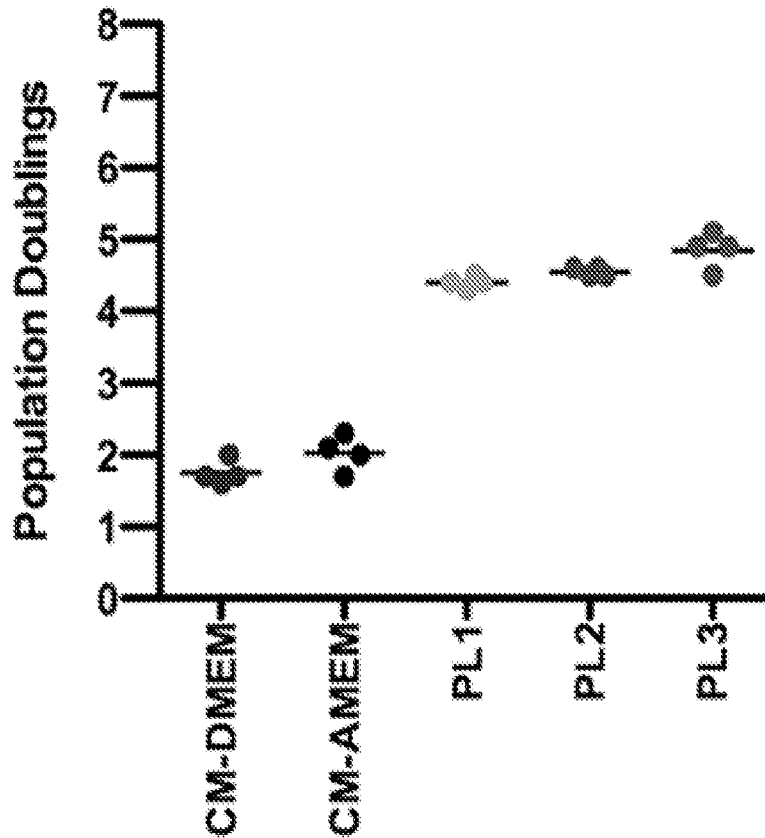
FIG. 21



Fat Derived MSCs grow faster in PL-5% Than CM
 Calculated growth rate of PL-5% (dotted line) vs CM (Solid Line). MSCs derived from normal adipose tissue and grown in PL-5% or CM. Triangles represent calculated 95% Confidence Interval.

FIG. 22

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MSC from adipose tissue were isolated, cultured in Complete Medium, CM (10% FCS +DMEM) for two days then frozen. After thaw the MSC were cultured in CM-DMEM, or 10% FCS+AMEM (CM-AMEM) or AMEM supplemented with one of three lots of platelet lysate. Data are replicates of four with the mean shown.

Data represent the number of population doublings during the second passage of a low density plating.

FIG. 23

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Tumor	Size	Conditions	30x106/60 days	CD133%	Cytogenetics
GBM2617-05	1.7	NSC/PL	N/N	21.2/0.12	
GBM2617-06	?2	NSC/PL	Y/Y	30/1	AB/AB
GBM083007	?1	NSC/PL/FBS	N/Y/N	0.5/15.6/0.5	AB/AB/AB
GBM091207	?1	NSC/PL/FBS	N/Y/N	3.3/38/2.3	AB/AB/AB
GBM2617-07	2.4	NSC/PL/FBS	N/N/N	ND	ND
GBM2617-11	1.5	NSC/PL/FBS	N/N/N	ND	AB
GBM2617-12	4.2	NSC/PL/FBS	N/N/N	ND	
GBM101	2.1	NSC/PL/FBS	N/N/N	ND	ND
GBM102	2.2	NSC/PL/FBS	Y/N/N	ND	ND
GBM103 CUSA	2.7	NSC/PL/FBS	N/Y/N	<1% each	AB/AB/AB
GBM106	2.2	NSC/PL/FBS	N/Y/N	14.1/.33/0	AB/AB/AB
GBM108	0.9	NSC/PL/FBS	N/N/N	ND/4.0/0	AB/AB/AB
GBM109 CUSA	3.2	NSC/PL/FBS	N/N/N	ND	ND
GBM110	0.4	NSC/PL/FBS	N/Y/N	14.8/0.2	(TBD)
GBM111 CUSA	3.0	NSC/PL/FBS	N/Y/N	5.3/4.7/0	AB/AB/AB
GBM112	1.2	NSC/PL/FBS	N/N/N	ND	ND

HCTL 7/16 meet vaccine criteria = 44%

NSC 2/16 meet vaccine criteria = 13%

FBS 0/15 meet vaccine criteria = 0%

An additional two cell lines grow in PL (2617-05 and GBM108) but did not meet criteria.

Tissue is essentially cut into fourths, 1 for RNA and 3 for culture conditions

General observations:

NSC: neurospheres, a couple of semiadherent

PL: monolayer and semiadherent, 1 neurosphere

FBS: monolayer

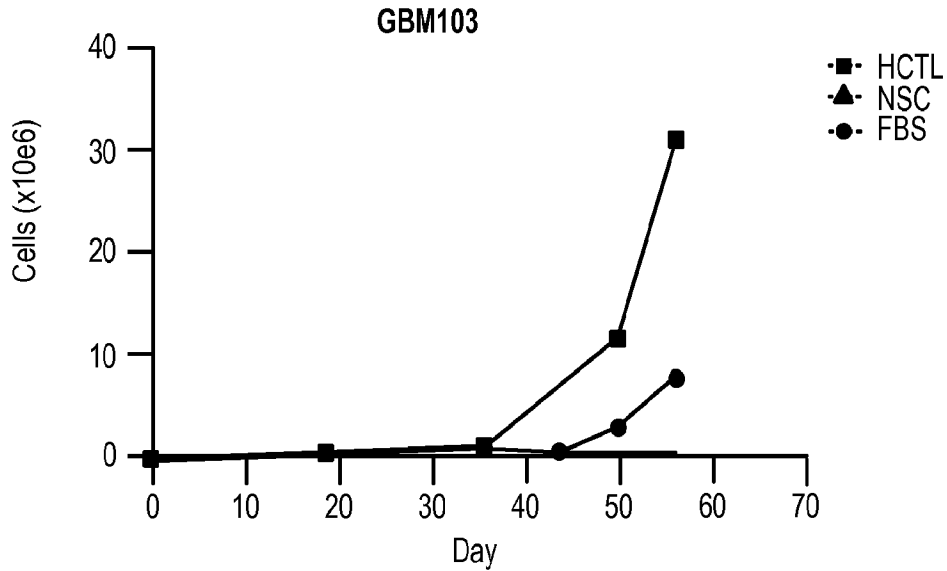
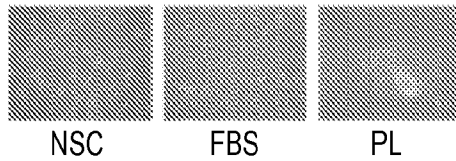
General phenotypic analysis

CD90+/CD45-/CD34lo/HLA ABC+/HLA DR-

FIG. 24A

GBM103

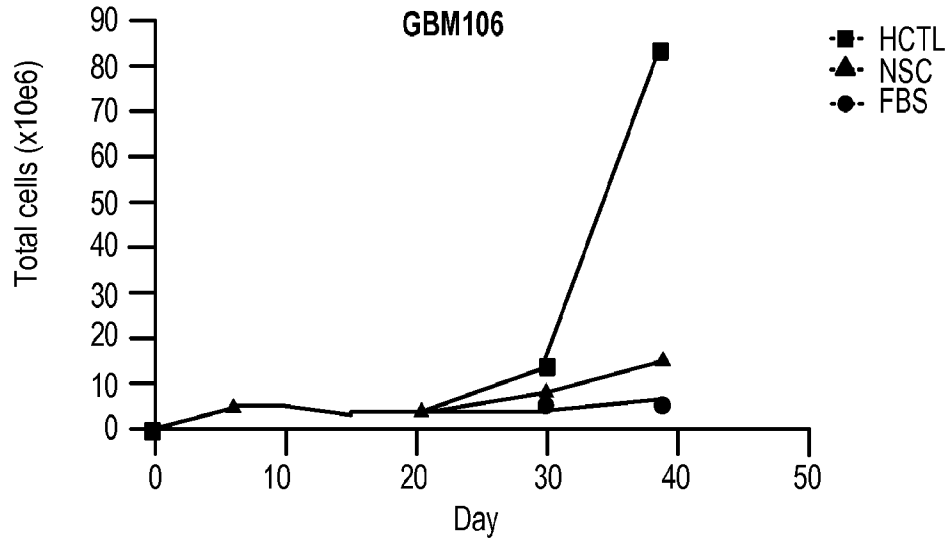
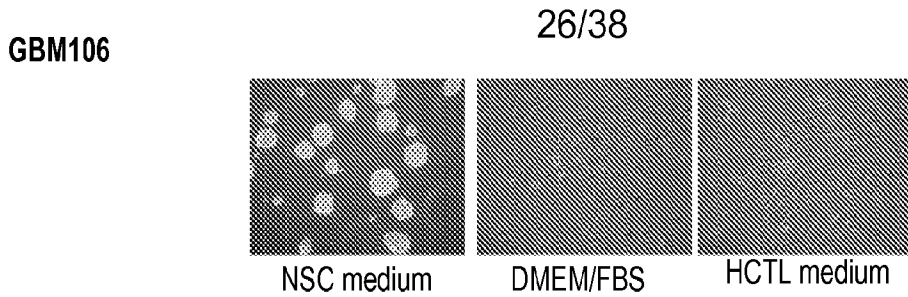
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GBM103	CD133	Nestin	SOX2	GFAP	β -III Tub	MBP	CD45	CD34	CD90	CD105	Class 2	Class 1
PL		++	+	++	++	-	-	-	+++	-	-	+++
NSC		+++	++	++	++	-	-	-	+++	-	-	+++
FBS		++	+	+	++	-	-	-	+++	-	-	+++

GBM103	
PL	74-83,XXYY,add(1)(q32),add(2)(q21)x2,-4-4,der(10)t(7;10)(p11.2;q22)x2,+11,-12,del(13)(q12q14)x2,-14,-14,-15,-16,-16,add(16)(p13.1)x2,der(17)add(17)p(11.2)del(17)(q11.2q21)x2,-19,-19,+0-2mar[cp20]
NSC	64-68,XXY,+Y,-4,-5,der(10)t(7;10)(p11.2;q22),inv(12)(p11.2q15)x2,del(13)(q12q14)x2,-14,der(14;14)(p13;p13),-15,-16,+17,der(17)add(17)p11.2,del(17)(q11.2q21)x2,-18,+19,-20,+21,+0-2mar[cp20]
FBS	61-73,XXY,+Y,+3,-4-6,+7,+8+10,der(10)t(7;10)p11.2;q22)x2,-11,inv(12)(p11.2q15)x2,+13,del(13)(q12q14)x2,-14,-16,der(17)add(17)(p11.2)del(17)(q11.2q21),-18,+20,i(21)(q10),-22,+0-2mar[cp20]

FIG. 24B



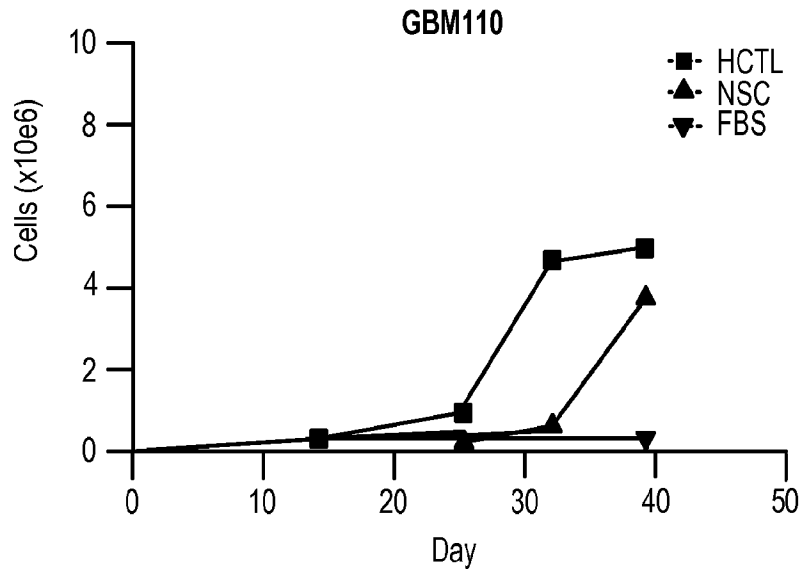
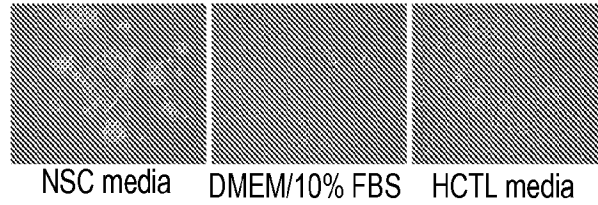
GBM106	CD133	Nestin	SOX2	GFAP	β -III Tub	MBP	CD45	CD34	CD90	CD105	Class 2	Class 1
PL		+++	+	+	+++	-	-	-	+++	-	-	+++
NSC		+++	++	+	+++	-	-	-	+++	-	-	+++
FBS		+++	-	+	+++	-	-	-	+++	-	-	+++

GBM106	
PL	50,XX,+4,+5,-6,+7,die(10;19)(q26;q13.4),+12,+15,der(20)t(7;20)(q11.2;q13.3),+22[5]/94,slx2[1]/50,XX,+4,+5,-6,+7,+7,add(10)q26,+12,die(12;19)(q24.3;q13.4),+15,psudie(18;18)(p11.2;p11.2),der(20)t(7;20),+22[3]/100,sl2x2[1]/50,XX,dis(3,19)(q29,q13.4),+5,-6,+7,+7,+12,+15,der(20)t(7;20),+22[1]/100,s13x2[1]/51,XX,+4,+5,-6,+7,+7,-10,+12,+15,add(19)(q13.3),der(20)t(7;20),+22[1]/100,sl4x2[5]/54,XX,+4,+5,+7,+7,+12,+12,+15,+15,psudie920;3)(q13.3;q21)+22[2]
NSC	48-63,XX,+4,+5 -6,+7,+7,-10,+12,+15, add(19)(q13.3.0,ser(20)t(7;20)(q11.2;q13.3),+22[cp14]
FBS	49-65,XX, add(2) (q33),+4,+5,-6,+7,+7,die(10;19)(q26;q13.4),+12,+12,+15,+15, add(19)(q13.3), add(20)(q13.1),der(20)t(7;20)(q11.2;q13.3),+22[cp9]

FIG. 24C

GBM110; SPORE # 3406

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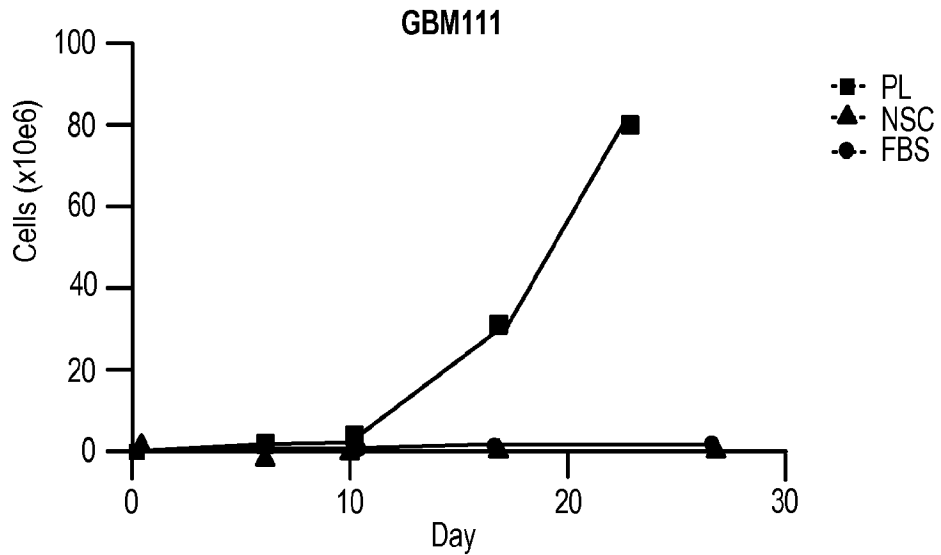
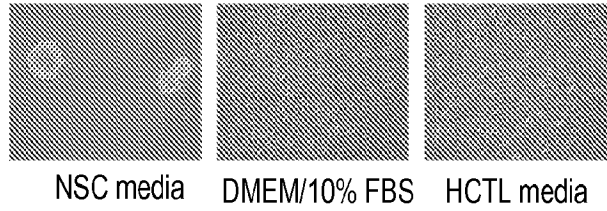
GBM110	CD133	Nestin	Sox2	CD45	CD34	CD90	CD105	Class 1
NSC	<1.0%	+	+	-	+	+++	-	+++
HCTL	<1.0%	+	-	-	+	+++	-	+++
FBS	<1.0%	+	-	-	+	+++	-	+++

GBM110	Karyotype
NSC media	75-84,XXXX,-X,-X,del(1)(q12),-5,-5,-6,+7,+7,+add(7),(q11.2)x2,t(7;18)(p13;p11.2)x2,-8,-10,-10,del(11)(p11.2p13)x2,-12,-13,-13,-14,-14,-15,-15,der(15;21)(q10;q10),i(15)(q10),i(16)(q10),-17,-17,-19,-19,-21,+22,+22,+der(22)t(1;22)(q21;q13),+0-2mar[cp18]/125-153,idemx2[cp2]
HCTL media	62-84,XXXX,-1,-2,-2,-3,-4,-5,-5,-6,+7,+7,+7,+add(7)(q11.2)x2,t(7;18)(p13;p11.2)x2,-8,-8,-10,-10,del(11),(p11.2p13)x2,-13,-13,-14,-14,-15,-15,-16,-17,-17,-18,-18,-19,+0-3mar[cp20]
10% FBS	73-83,XXXX,-X,-X,-2,-4,-4,inv(4),(q27q35);-5,del(5)(p11),der(5;15)(q10;q10),-6,+7,+7,+add(7)(q11.2)x2,t(7;18)(p13;p11.2)x2,-10,-10,del(11)(p11.2p13)x2,-13,-13,-14,-14,-15,-15,der(15;21)(q10;q10),i(15)(q10),i(16)(q10),-17,-17,-19,-19,+0-3mar[cp16]/141,idemx2,-i(15)[cp2]/46,XX[2]

FIG. 24D

GBM111; SPORE # 3407

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GBM111	CD133	Nestin	SOX2	GFAP	β -III Tub	MBP	CD45	CD34	CD90	CD105	Class 2	Class 1
PL	4.79	+	-	++	+	-	-	-	+++	-	-	+++
NSC	5.82	+	-	++	+	-	-	-	+++	-	-	+++
FBS	0.01	+	-	+	+	-	-	-	+++	-	-	+++

GBM111	Karyotype
NSC media	43-45,XY,t(1;19)(p34.3;q13.3),+7,-9,-10,inv(12)(p13q11),-13,-14,-17,+20,add(21)(p11.2),+1-3mar[cp4]/42-46,sl,t(3;6)(p21;p23)[cp4]/84-86,sdl1x2[cp3]/82-89,XXYY,t(1;19)x2,add(5)(p13),+7,+7,-9,-9,-10,inv(12)x2,-13,-13,-14,-14,+20,+20,add(21)(p11.2)x2,+1-2mar[cp9]
HCTL media	46,XY,t(1;19)(p34.3;q13.3),t(3;6)(p21;p23),+7,-10,inv(12)(p13q11),-13,-14,+20,add(21)(p11.2),+1-2mar[cp6]/84-92,idemx2[cp5]/82-88,XXYY,t(1;19)x2,-2,-4,-4,add(5)(p13),+7,+7,-9,-9,-10,inv(12)x2,-13,-13,-14,-14,+20,+20,-21,add(21)x2,+2-3mar[cp9]
10% FBS	54-64,XXY,add(X)(p22.1),+1,add(1)(p13),-3,-4,der(4)t(4;7)(q21;p13),+7,-8,i(8)(q10),-10,-11,psu die(11;15)(p15;p11.2),-12,-13,-14,-16,-17,-18,-18,+20,+21,add(21)(p11.2),-22,+1-4mar[cp20]

FIG. 24E

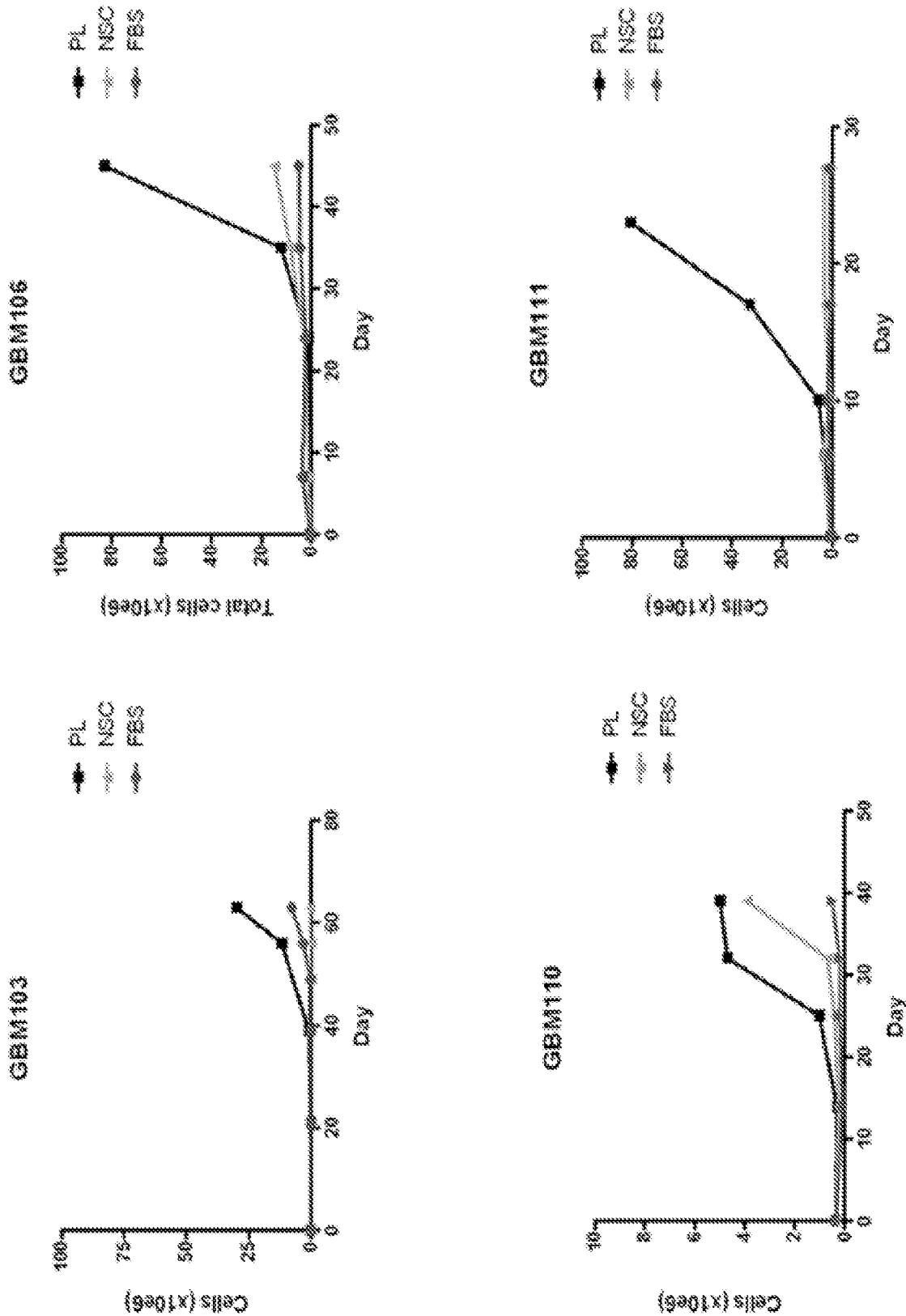


FIG. 25

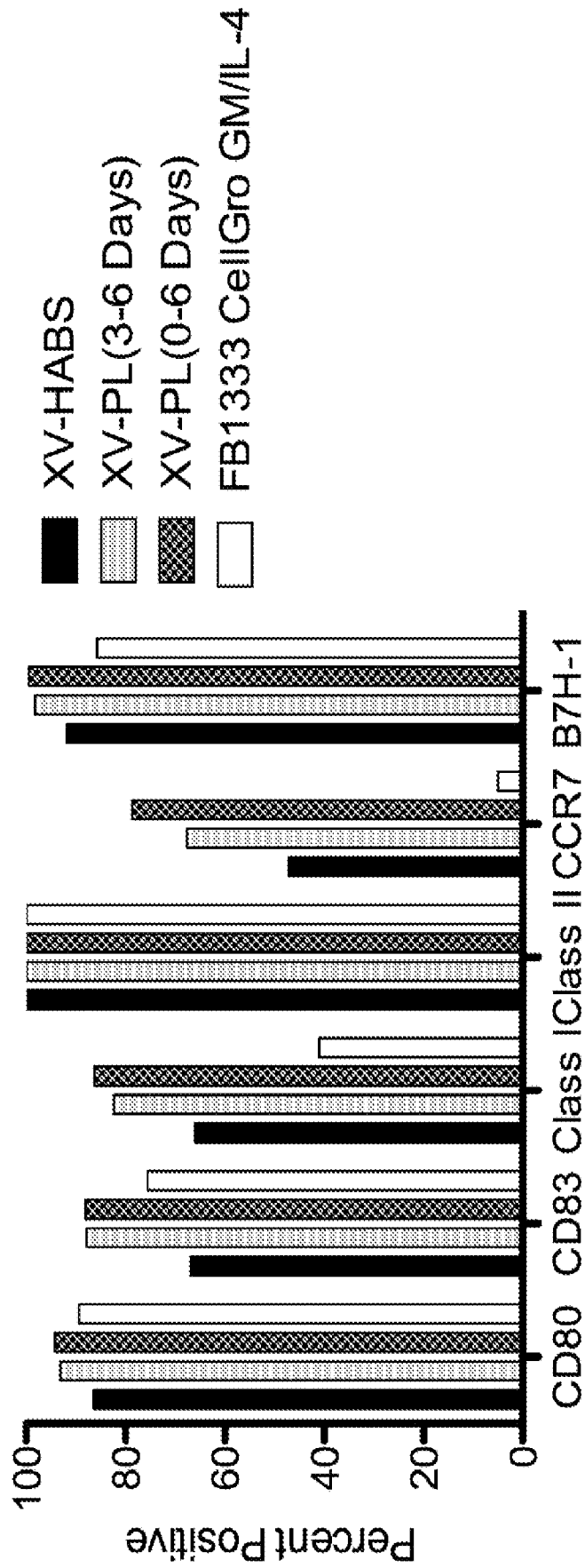


FIG. 26

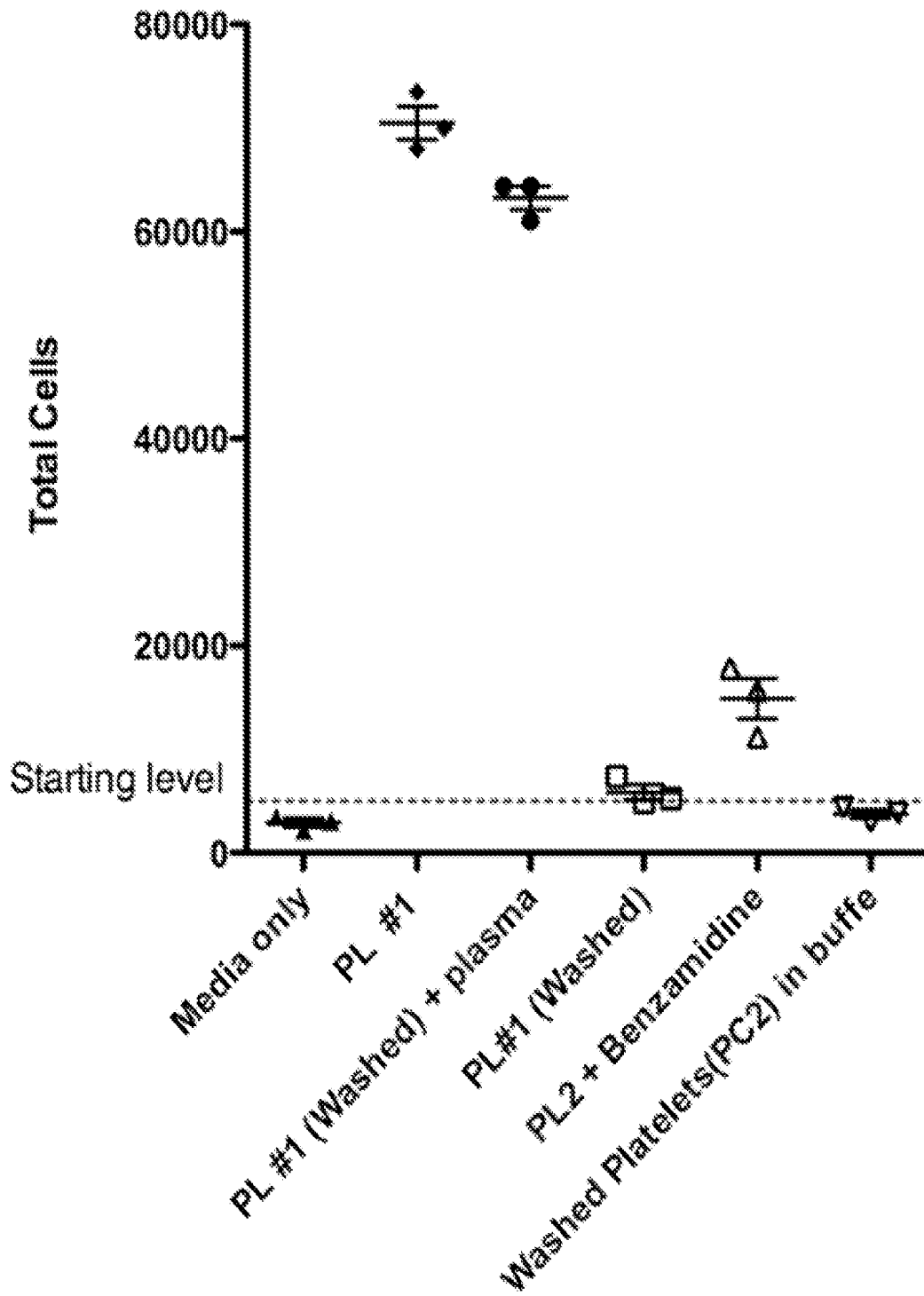


FIG. 27

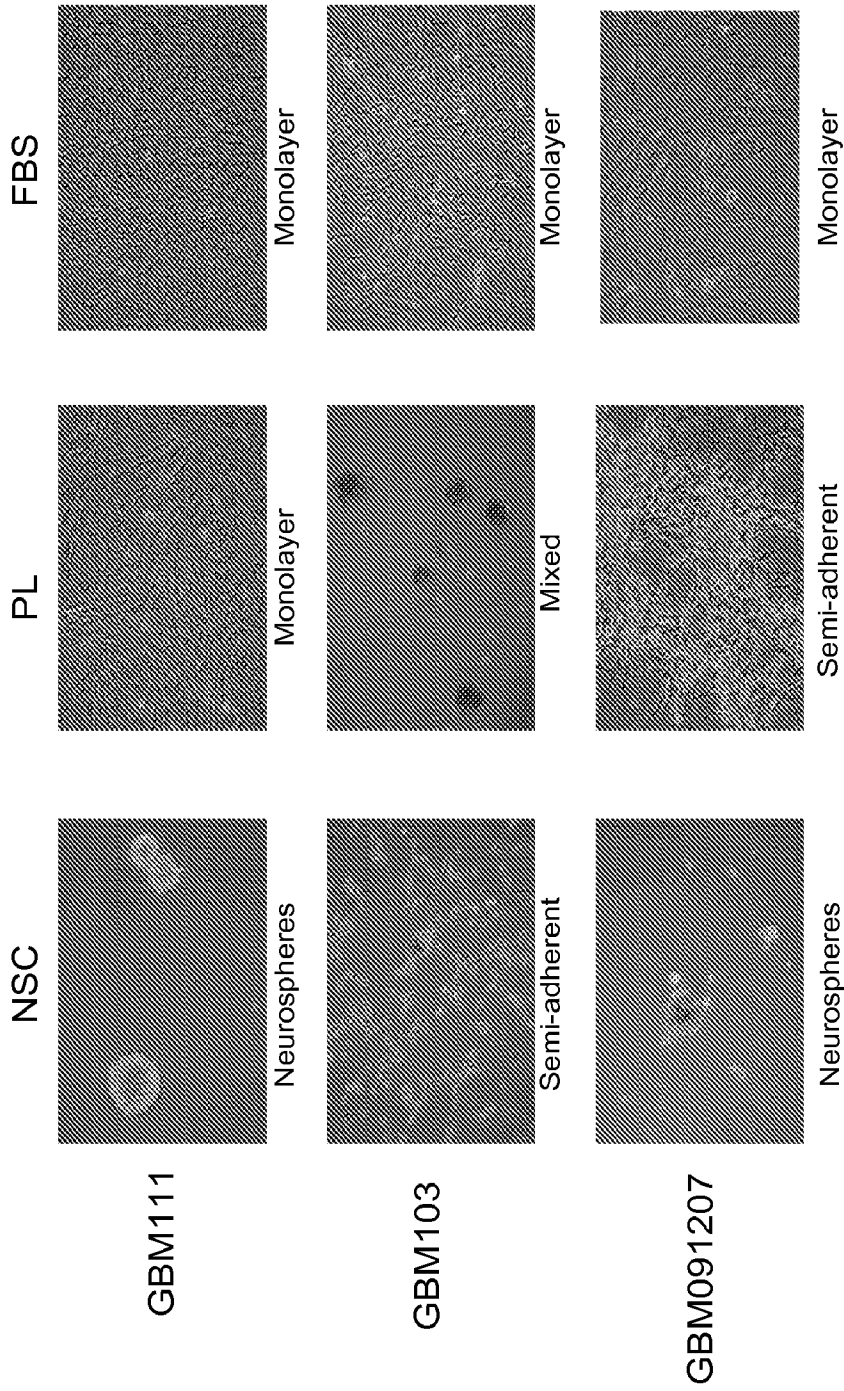


FIG. 28

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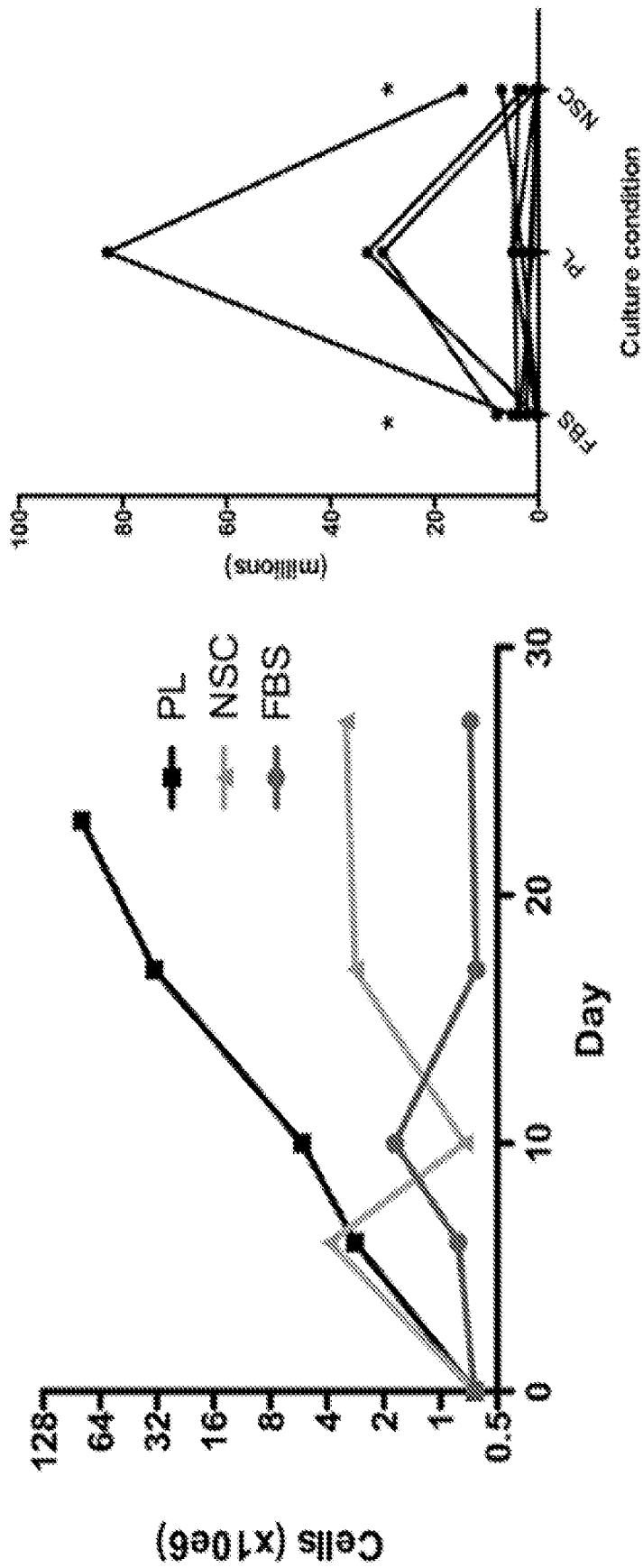


FIG. 29

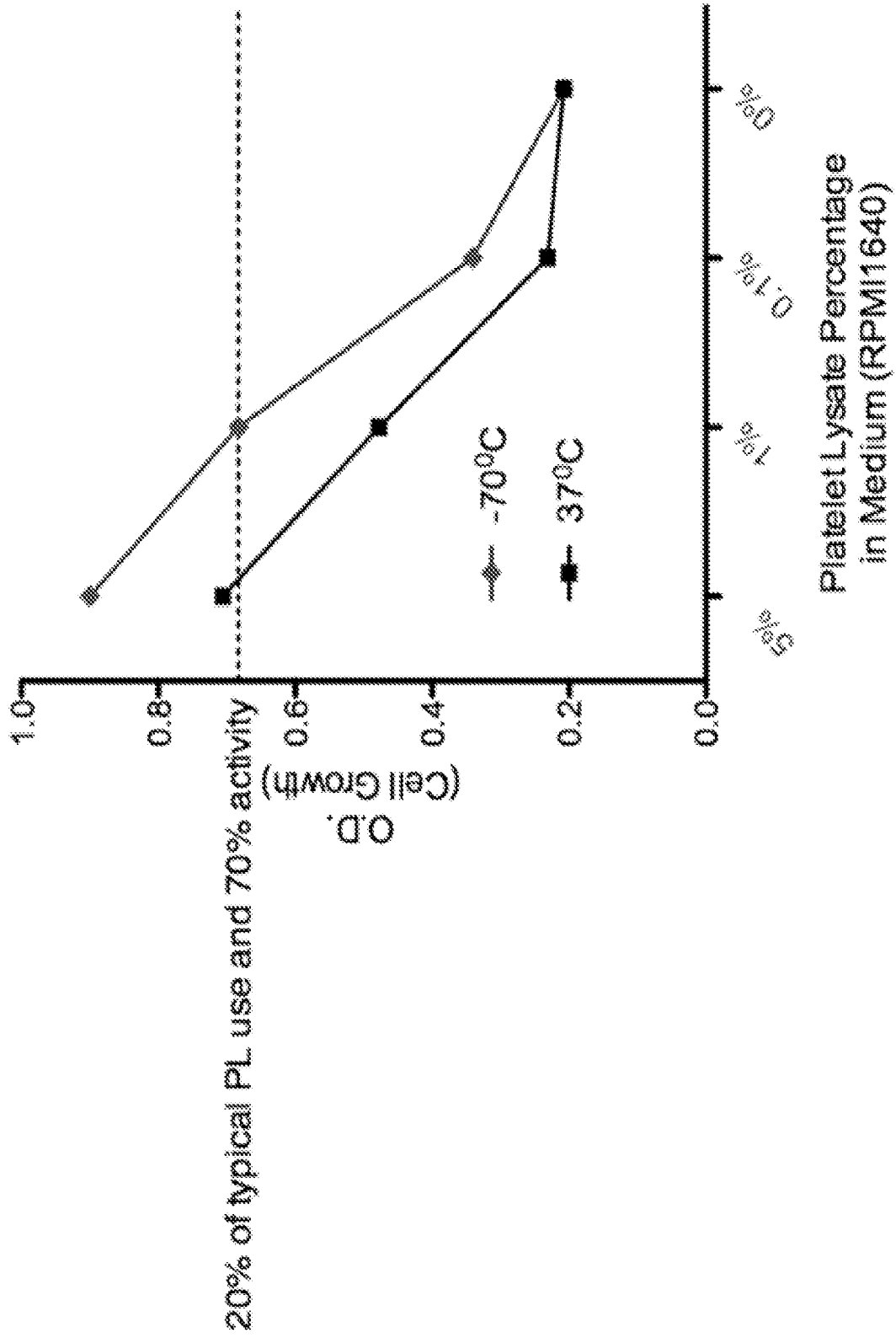


FIG. 30

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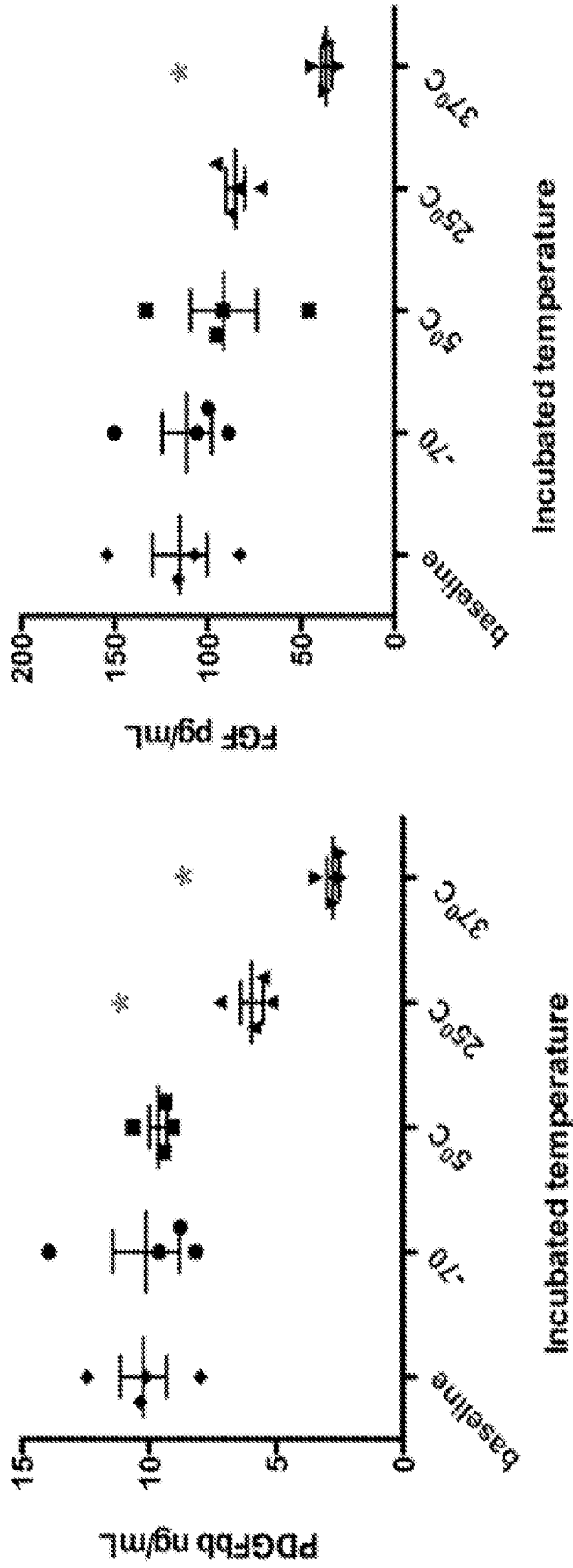


FIG. 31

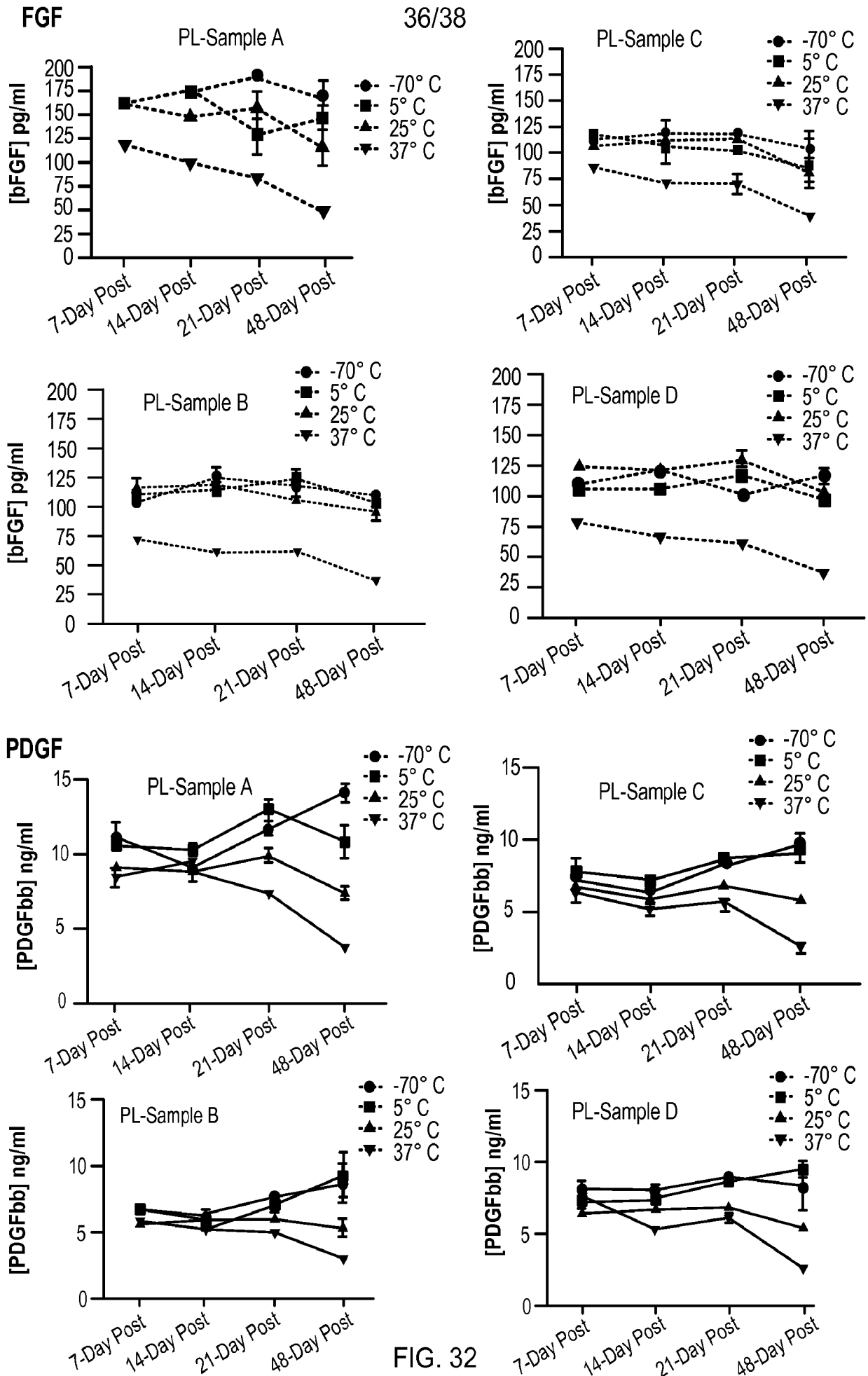


FIG. 32

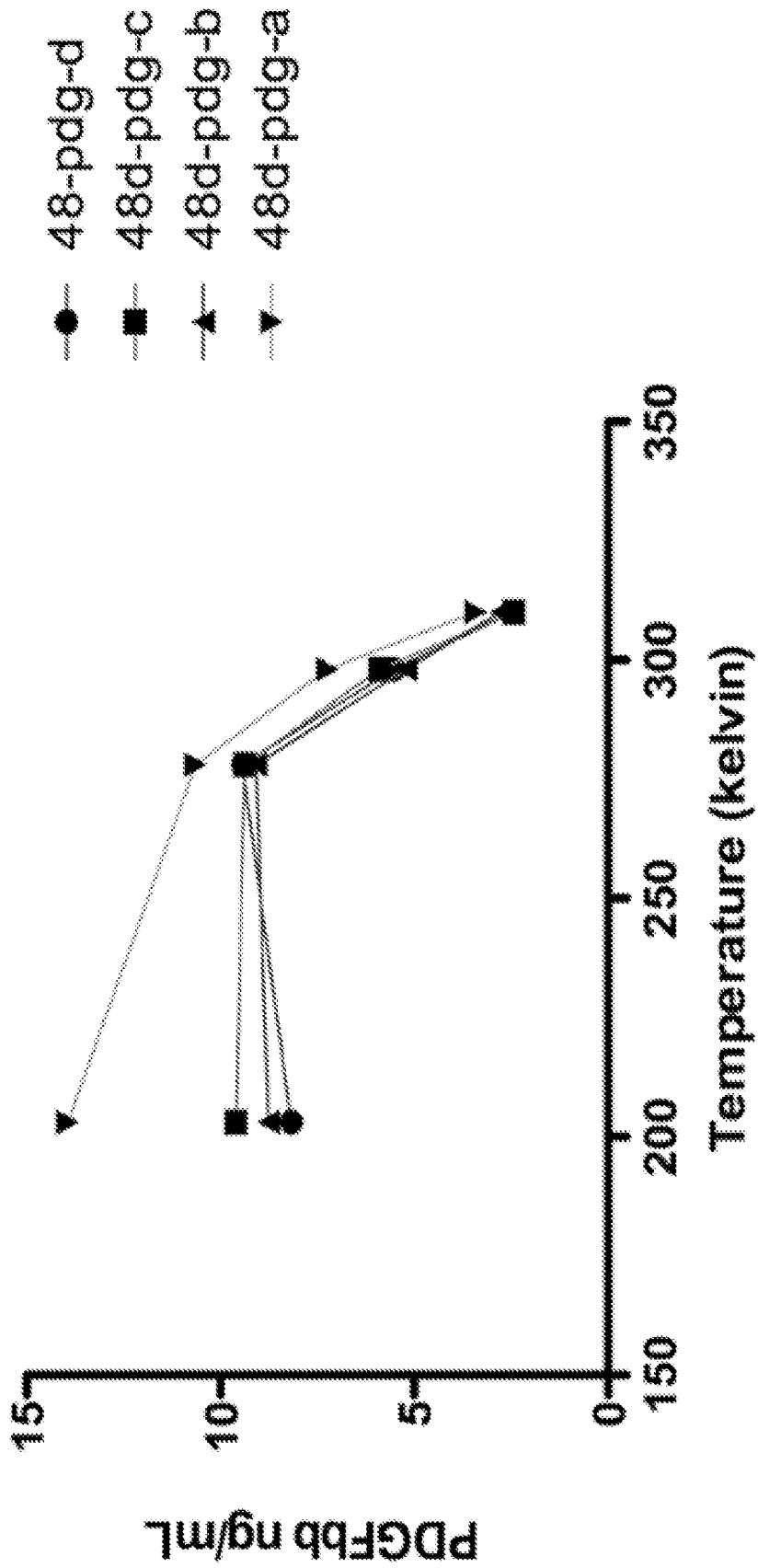


FIG. 33

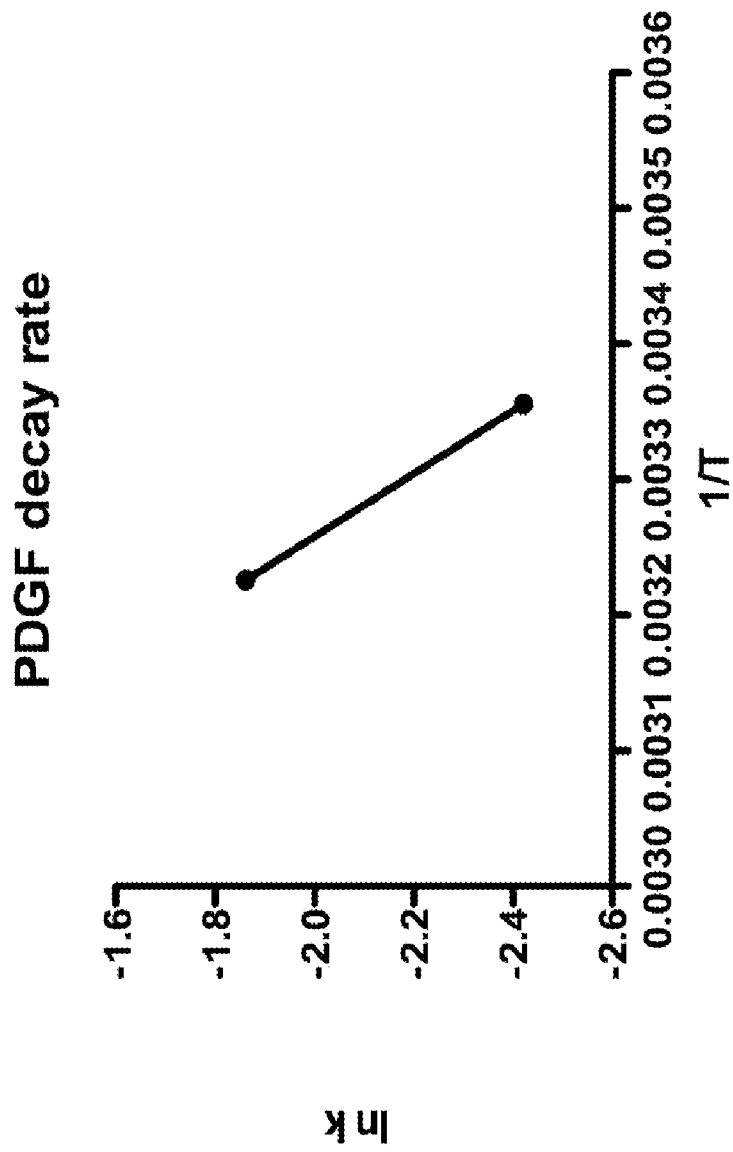


FIG. 34