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ABSTRACT

This invention provides methods, compositions and articles of manufacture for inhibiting the onset of and treating glomerular injury. The instant invention is based on the blockade of RAGE and/or RAGE G82S function.

RAGE-RELATED METHODS AND COMPOSITIONS FOR TREATING GLOMERULAR INJURY

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Throughout this application, various publications are referenced. Full citations for these publications may be found immediately preceding the claims. The disclosures of these publications are hereby incorporated by reference into this application in order to more fully describe the state of the art as of the date of the invention described and claimed herein.

Background of the Invention

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Primary or secondary focal segmental glomerulosclerosis (FSGS) encompasses a range of diseases characterized by glomerular and tubulointerstitial fibrosis that often progress, unhaltingly, to irreversible renal scarring and failure in human subjects (1). Secondary cases of 20 FSGS may emerge in the face of chronic disease (hemodynamic, immunologic or metabolic). However, in both cases of primary and secondary disease, despite study, is definitive years of there no many 25 understanding of the molecular mechanisms that underlie As such, insights into means to these disorders. prevent/treat these disorders have not been elucidated.

Key steps in identifying rational therapeutic targets for these diseases, however, may emerge from animal studies. Development of FSGS by agents that incite pathways linked to glomerular fibrosis and hyperpermeability are useful as a means to track the early, initiating events and the later, amplified consequences of proteinuria and renal scarring. In

this context, multiple studies have employed administration of agents such as puromycin or adriamycin (ADR) to rats, to induce processes analogous to human FSGS in the kidney (2-4). In addition, other studies in rats have included the induction of Passive Heymann Nephritis as a means to induce irreversible glomerular injury (5). Overall, these studies in rats have been frustrated by the inability to precisely link activation of specific cells to the pathogenesis and/or progression of GS upon disease induction.

A paucity of mouse models existed for the study of FSGS-like diseases until the first description of ADR-induced toxicity in mice (6-7). In 2000, Wang and colleagues reported on the impact of ADR up to 42 days (6 weeks) after administration of ADR (9). Male BALB/c mice, 20 to 25 gm, were injected with ADR, 10.5 mg/kg, by IV injection. These investigators carefully followed the time course of events in the ADR-treated mice and observed the following (9).

First, overt proteinuria developed in all mice by day 5. Proteinuria persisted throughout 6 weeks of study. Only 35.7% of mice developed hematuria but 53.6% developed leukocyturia.

Second, levels of serum albumin were consistently lower in ADR-treated mice vs controls beginning one week after ADR treatment.

Third, creatinine clearance declined with time and was significantly decreased compared to control mice 4 weeks post-ADR.

Fourth, by week 6, tubular atrophy and intratubular cast formation with tubulointerstitial expansion had occurred and was widely seen in the cortex. Extensive FGS and severe interstitial fibrosis and inflammation were present. Global sclerosis was observed in many glomeruli.

Fifth, by EM, effacement of foot processes of podocytes had occurred. At week 1, effacement was segmental, but global by week 6. Control mice failed to demonstrate any epithelial cell abnormalities at any point.

Importantly, in this study, cellular infiltration and inflammation were examined.

15 Sixth, at early and later times after ADR, CD4+ and CD8+ T cells, and macrophages were significantly increased in the kidneys of ADR-treated mice. These cell types were found both in the interstitium as well 20 as in the glomeruli after injury. Infiltration of inflammatory cells was noted quite early after ADR, within the first 24 hours, and persisted for up to These findings support the premise weeks after ADR. that inflammation, at least in part, contributes as an 25 early trigger, and/or late progression factor in the molecular pathways leading to sustained glomerular perturbation, fibrosis and albuminuria that converge to cause renal dysfunction.

These studies highlighted that even 6 weeks after ADR, progressive renal injury, proteinuria and decreased creatinine clearance were features of the disease. In addition, new insights into proinflammatory mechanisms into the disease process were uncovered by time course

examination of cellular infiltration after ADR. Other studies have, in fact, confirmed inflammatory cell infiltration into the ADR-treated kidney (9). Indeed, the observation that human FSGS is typified by differentiation of podocytes into MP-like cells, along with inflammatory cell infiltration from the periphery (MP and T lymphocytes) in the interstitium, periglomerular regions and glomeruli (1, 10-12) is compatible with the concept put forth in the ADR-induced murine model of FSGS, that is, it is plausible that inflammatory stimuli importantly contribute to the pathogenesis and/or progression of FSGS.

In parallel with progressive renal dysfunction and 15 scarring in primary or secondary FSGS syndromes in human subjects (and murine models), injury depletion of glomerular podocytes, eventuating podocyte "insufficiency" and capillary collapse, have been implicated as important steps in the development 20 of FSGS (13, 14). In most cases of nephrotic syndrome, podocyte foot process effacement is considered an early manifestation of injury, and is followed by a continuum characterized damage of progressive podocyte vacuolization, pseudocyst formation, detachment of 25 podocytes from the GBM; processes that lead to irreversible loss/apoptosis of podocytes (15).

Key evidence that podocytes are not mere bystanders, but rather active participants in molecular pathways of injury, was highlighted by recent studies in TGF-ß overexpressing transgenic mice. In those mice, marked upregulation of Smad 7 was observed in damaged podocytes. Both TGF-ß and Smad7 were associated with apoptosis in cultured podocytes. In the former case,

activation of p38 MAP kinase and caspase-3 were key intermediary steps in TGF-ß-induced apoptosis. In the latter case, suppressed nuclear translocation of the cell survival factor NF-kB led to Smad7-induced podocyte apoptosis (16). These studies highlight the concept that activation of cell signalling and modulation of gene expression in the podocyte may be early events in the development of FSGS, and thus, may contribute to the pathogenesis of this disease.

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It is important to note that the concept of key roles for podocytes in the pathogenesis/progression of glomerular dysfunction have parallels in diabetes. Diabetes is a highly complex environment in which such as 15 multiple contributing pathways, Advanced Glycation of accumulation/activation Endproducts, activation of PKC, especially the Bisoform, as well as hyperglycemia itself are implicated in the pathogenesis of this disorder (17-19). Evidence 20 is accumulating that podocytes are perturbed early in diabetes, and that their products, such as VEGF, may contribute to cellular dysfunction in this disorder (20-25). As in FSGS and FSGS-like disorders, the case for the podocyte as bystander vs contributory agent to 25 the pathogenesis and progression of glomerular injury remains to be rigorously tested.

Although inhibiting RAGE has been implicated in treating symptoms of diabetes (35), the literature does

not provide a basis for concluding that inhibiting the binding of RAGE to its ligands could play a role in treating or preventing glomerular injury.

Summary of the Invention

ligand thereof.

This invention provides a method for inhibiting the onset of a glomerular injury in a subject comprising administering to the subject a prophylactically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

This invention further provides a method for treating a glomerular injury in a subject comprising administering to the subject a therapeutically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

This invention further provides a method for inhibiting the onset of glomerulosclerosis, proteinuria or albunuria in a subject comprising administering to the subject a prophylactically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

This invention further provides a method for treating glomerulosclerosis, proteinuria or albunuria in a subject comprising administering to the subject a therapeutically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a

This invention further provides an article of 30 manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label

indicating a use for the agent for inhibiting the onset of glomerular injury in a subject.

This invention further provides an article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for inhibiting the onset of glomerulosclerosis, proteinuria or albuminuria in a subject.

This invention further provides an article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for treating a glomerular injury in a subject.

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article of Finally, invention provides an this manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the 25 packaging material has affixed thereto use for the agent for treating indicating a glomerulosclerosis, proteinuria or albuminuria in a subject.

Brief Description of the Figures

Figure 1. Administration of ADR to BALB/c mice: effects of sRAGE. BALB/c mice were treated with ADR or control (saline). ADR-treated mice received sRAGE or PBS. At 2 weeks after ADR, kidney wt/body wt ratio and mesangial area & mesangial/glomerular fraction determined. N=5 mice/group. Statistical considerations are indicated in the figures.

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Figure 2. Administration of ADR to BALB/c mice: effects of sRAGE. BALB/c mice were treated with ADR or control (saline). ADR-treated mice received sRAGE or PBS. At 6 weeks after ADR, kidney wt/body wt ratio and mesangial area & mesangial/glomerular fraction determined. N=5 mice/group. Statistical considerations are indicated in the figures.

Figure 3. Blockade of RAGE suppresses albuminuria after administration of ADR. At 2 and 6 weeks after ADR, urine albumin/creatinine ratio was determined. N=5 mice/condition. N=5 mice/condition. Statistical considerations are indicated in the figure.

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Detailed Description of the Invention

Definitions

5 "Agent" shall include, without limitation, an organic compound, a nucleic acid, a polypeptide, a lipid, and a carbohydrate. Agents include, for example, agents which are known with respect to structure and/or function, and those which are not known with respect to

10 structure or function.

"Antibody" shall include, by way of example, both naturally occurring and non-naturally occurring antibodies. Specifically, this term includes polyclonal and monoclonal antibodies, and antigen-binding fragments thereof. Furthermore, this term includes chimeric antibodies and wholly synthetic antibodies, and antigen-binding fragments thereof.

20 As used herein, "inhibit," when used in connection with the binding between RAGE and/or RAGE G82S with a ligand thereof, shall mean to reduce such binding. In one embodiment, "inhibit" shall mean to eliminate such binding.

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"Inhibiting" the onset of a disorder shall mean either lessening the likelihood of the disorder's onset, or preventing the onset of the disorder entirely. In the preferred embodiment, inhibiting the onset of a disorder means preventing its onset entirely.

"Subject" shall mean any animal, such as a human, nonhuman primate, mouse, rat, guinea pig or rabbit.

"Treating" a disorder shall mean slowing, stopping or reversing the disorder's progression. In the preferred embodiment, treating a disorder means reversing the disorder's progression, ideally to the point of eliminating the disorder itself. As used herein, ameliorating a disorder and treating a disorder are equivalent.

Embodiments of the Invention

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This invention provides methods for inhibiting the onset of and treating glomerular injury. This invention is based on the surprising discovery of a correlation between suppressing glomerular injury in a non-diabetic subject and blocking RAGE and/or RAGE G82S function.

Specifically, this invention provides a method for inhibiting the onset of a glomerular injury in a subject comprising administering to the subject a prophylactically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

In one embodiment of the instant method, the glomerular injury is associated with reduced removal of toxins.

In another embodiment, the glomerular injury is associated with glomerulosclerosis. In a further embodiment, the glomerular injury is associated with proteinuria. In yet a further embodiment, the glomerular injury is associated with albuminuria.

In the preferred embodiment of the instant method, the subject is human. In one embodiment the subject is

afflicted with diabetes. In another embodiment of the instant method, the subject has been afflicted with diabetes for less than 20 years. In a further afflicted embodiment, the subject is not 5 diabetes. In yet a further embodiment, the subject is receiving or is about to receive a chemotherapy drug. In yet a further embodiment, the chemotherapy drug is In yet a further embodiment, adriamycin. chemotherapy drug is selected from the following: 5interferon; Alpha Actinomycin D; 10 fluorouracil; Bleomycin; Cisplatin; Cyclophosphamide; Dexamethasone; Doxorubicin; Epoetin alfa; Etoposide; Interleukin-2; alfa; Herceptin; Interferon Interleukin-11; Methotrexate; Neupogen; Nitrogen Paclitaxel; Prednisolone; Prednisone; · 15 Mustard; Tamoxifen; Thalidomide; Rituximab; PROCRIT; Vinblastine; and Vincristine. Additional chemotherapy drugs are envisioned, and are listed in chemocare.com (http://www.chemocare.com/bio/default.sps).

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In one embodiment of the instant method, the agent is soluble RAGE. In another embodiment, the agent is soluble RAGE G82S. In a further embodiment, the agent is an antibody directed to RAGE. In yet a further embodiment, the agent is an antibody directed to RAGE G82S.

This invention further provides a method for treating a glomerular injury in a subject comprising administering to the subject a therapeutically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

In one embodiment of the instant method, the glomerular injury is associated with reduced removal of toxins. In another embodiment, the glomerular injury is associated with glomerulosclerosis. In a further embodiment, the glomerular injury is associated with proteinuria. In yet a further embodiment, the glomerular injury is associated with albuminuria.

In the preferred embodiment of the instant method, the 10 subject is human. In one embodiment, the subject is not afflicted with diabetes. In another embodiment, the subject is receiving or is about to receive a chemotherapy drug. In a further embodiment, the chemotherapy drug is adriamycin. In yet a further 15 embodiment, the chemotherapy drug is selected from the following: 5-fluorouracil; Actinomycin D; interferon; Bleomycin; Cisplatin; Cyclophosphamide; Dexamethasone; Doxorubicin; Epoetin alfa; Etoposide; Gleevec; Herceptin; Interferon alfa; Interleukin-2; 20 Interleukin-11; Methotrexate; Neupogen; Mustard; Paclitaxel; Prednisolone; Prednisone; PROCRIT; Rituximab; Tamoxifen; Thalidomide; Vinblastine; and Vincristine. Additional chemotherapy drugs are envisioned, and are listed in chemocare.com 25 (http://www.chemocare.com/bio/default.sps).

In one embodiment of the instant method, the agent is soluble RAGE. In another embodiment, the agent is soluble RAGE G82S. In a further embodiment, the agent 30 is an antibody directed to RAGE. In yet a further embodiment, the agent is an antibody directed to RAGE G82S.

This invention further provides a method for inhibiting the onset of glomerulosclerosis, proteinuria or albunuria in a subject comprising administering to the subject a prophylactically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

In the preferred embodiment of the instant method, the subject is human. In one embodiment the subject is 10 afflicted with diabetes. In another embodiment of the instant method, the subject has been afflicted with diabetes for less than 20 years. In a further embodiment, the subject is not afflicted with diabetes. In yet a further embodiment, the subject is receiving 15 or is about to receive a chemotherapy drug. In yet a embodiment, the chemotherapy drug further In yet a further embodiment, the adriamycin. chemotherapy drug is selected from the following: 5-Actinomycin D; Alpha fluorouracil; 20 Bleomycin; Cisplatin; Cyclophosphamide; Dexamethasone; Epoetin alfa; Etoposide; Doxorubicin; Herceptin; Interferon alfa; Interleukin-2; Interleukin-Nitrogen Mustard; Neupogen; 11; Methotrexate; Prednisolone; Prednisone; PROCRIT; Paclitaxel; 25 Rituximab; Tamoxifen; Thalidomide; Vinblastine; and Vincristine. Additional chemotherapy drugs in chemocare.com envisioned, and are listed (http://www.chemocare.com/bio/default.sps).

30 In one embodiment of the instant method, the agent is soluble RAGE. In another embodiment, the agent is soluble RAGE G82S. In a further embodiment, the agent is an antibody directed to RAGE. In yet a further

embodiment, the agent is an antibody directed to RAGE G82S. $\,$

This invention further provides a method for treating glomerulosclerosis, proteinuria or albunuria in a subject comprising administering to the subject a therapeutically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof.

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In the preferred embodiment of the instant method, the subject is human. In one embodiment, the subject is not afflicted with diabetes. In another embodiment, the subject is receiving or is about to receive a 15 chemotherapy drug. In a further embodiment, the chemotherapy drug is adriamycin. In yet a further embodiment, the chemotherapy drug is selected from the following: 5-fluorouracil; Actinomycin D; Alpha interferon; Bleomycin; Cisplatin; Cyclophosphamide; 20 Dexamethasone; Doxorubicin; Epoetin alfa; Etoposide; Gleevec; Herceptin; Interferon alfa; Interleukin-2; Interleukin-11; Methotrexate; Neupogen; Nitrogen Mustard; Paclitaxel; Prednisolone; Prednisone; PROCRIT; Rituximab; Tamoxifen; Thalidomide; Vinblastine; Additional chemotherapy drugs 25 Vincristine. are envisioned, and are listed in chemocare.com (http://www.chemocare.com/bio/default.sps).

In one embodiment of the instant method, the agent is soluble RAGE. In another embodiment, the agent is soluble RAGE G82S. In a further embodiment, the agent is an antibody directed to RAGE. In yet a further embodiment, the agent is an antibody directed to RAGE G82S.

Determining a therapeutically or prophylactically effective amount of agent can be done based on animal data using routine computational methods. In one 5 embodiment, the therapeutically or prophylactically effective amount contains between about lng and about lg of protein, as applicable. In another embodiment, the effective amount contains between about 10ng and about 100mg of protein, as applicable. In a further 10 embodiment, the effective amount contains between about 100ng and about 10mg of the protein, as applicable. a yet a further embodiment, the effective amount contains between about $1\mu g$ and about 1mg of the protein, as applicable. In a yet a further embodiment, 15 the effective amount contains between about $10\mu g$ and about $100\mu g$ of the protein, as applicable. In a yet a further embodiment, the effective amount contains between about $100\mu g$ and about 10mg of the protein, as applicable. In yet a further embodiment, the effective 20 amount of agent, wherein the agent is soluble RAGE, is administered to the subject at a rate from about 2µg/kg/hr to about 100µg/kg/hr (e.g. about 5, 10, 25, 50 or $75\mu g/kg/hr$).

25 In this invention, administering agents can be effected or performed using any of the various methods and delivery systems known to those skilled in the art. performed, for be administering can orally, via implant, transmucosally, intravenously, 30 transdermally, intramuscularly, and subcutaneously. following delivery systems, which employ a number of routinely used pharmaceutical carriers, representative of the many embodiments envisioned for administering the instant compositions.

Injectable drug delivery systems include solutions, suspensions, gels, microspheres and polymeric injectables, and can comprise excipients such as solubility-altering agents (e.g., ethanol, propylene glycol and sucrose) and polymers (e.g., polycaprylactones and PLGA's). Implantable systems include rods and discs, and can contain excipients such as PLGA and polycaprylactone.

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Oral delivery systems include tablets and capsules. These can contain excipients such as binders (e.g., hydroxypropylmethylcellulose, polyvinyl pyrilodone, other cellulosic materials and starch), diluents (e.g., lactose and other sugars, starch, dicalcium phosphate and cellulosic materials), disintegrating agents (e.g., starch polymers and cellulosic materials) and lubricating agents (e.g., stearates and talc).

Transmucosal delivery systems include patches, tablets, suppositories, pessaries, gels and creams, and can contain excipients such as solubilizers and enhancers (e.g., propylene glycol, bile salts and amino acids), and other vehicles (e.g., polyethylene glycol, fatty acid esters and derivatives, and hydrophilic polymers such as hydroxypropylmethylcellulose and hyaluronic acid).

Dermal delivery systems include, for example, aqueous and nonaqueous gels, creams, multiple emulsions, microemulsions, liposomes, ointments, aqueous and nonaqueous solutions, lotions, aerosols, hydrocarbon bases and powders, and can contain excipients such as solubilizers, permeation enhancers (e.g., fatty acids,

fatty acid esters, fatty alcohols and amino acids), and hydrophilic polymers (e.g., polycarbophil and polyvinylpyrolidone). In one embodiment, the pharmaceutically acceptable carrier is a liposome or a transdermal enhancer.

Solutions, suspensions and powders for reconstitutable delivery systems include vehicles such as suspending agents (e.g., gums, zanthans, cellulosics and sugars), humectants (e.g., sorbitol), solubilizers (e.g., ethanol, water, PEG and propylene glycol), surfactants (e.g., sodium lauryl sulfate, Spans, Tweens, and cetyl pyridine), preservatives and antioxidants (e.g., parabens, vitamins E and C, and ascorbic acid), anticaking agents, coating agents, and chelating agents (e.g., EDTA).

In one embodiment of this invention, the delivery system used comprises more than water alone, or more than buffer alone.

of invention further provides an article This manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE 25 and/or RAGE G82S and a ligand thereof, wherein the has affixed thereto a label packaging material indicating a use for the agent for inhibiting the onset of glomerular injury in a subject. This invention further provides an article of manufacture comprising a 30 packaging material having therein an agent inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent

for inhibiting the onset of glomerulosclerosis, proteinuria or albuminuria in a subject.

In the preferred embodiment of the instant articles of 5 manufacture, the subject is human. In one embodiment the subject is afflicted with diabetes. In another embodiment of the instant methods, the subject has been afflicted with diabetes for less than 20 years. .In a further embodiment, the subject is not afflicted with 10 diabetes. In yet a further embodiment, the subject is receiving or is about to receive a chemotherapy drug. In yet a further embodiment, the chemotherapy drug is adriamycin. In yet a further embodiment, the chemotherapy drug is selected from the following: 5-15 fluorouracil; Actinomycin D; Alpha interferon; Bleomycin; Cisplatin; Cyclophosphamide; Dexamethasone; Doxorubicin; Epoetin alfa; Etoposide; Gleevec; Herceptin; Interferon alfa; Interleukin-2; Interleukin-11; Methotrexate; Neupogen; Nitrogen Mustard; 20 Paclitaxel; Prednisolone; Prednisone; PROCRIT; Rituximab; Tamoxifen; Thalidomide; Vinblastine; and Vincristine. Additional chemotherapy drugs are envisioned, and are listed in chemocare.com (http://www.chemocare.com/bio/default.sps).

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In one embodiment of the instant articles of manufacture, the agent is soluble RAGE. In another embodiment, the agent is soluble RAGE G82S. In a further embodiment, the agent is an antibody directed to RAGE. In yet a further embodiment, the agent is an antibody directed to RAGE G82S.

This invention further provides an article of manufacture comprising a packaging material having

therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for treating a glomerular injury in a subject. Finally, this invention provides an article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for treating glomerulosclerosis, proteinuria or albuminuria in a subject.

In the preferred embodiment of the instant articles of 15 manufacture, the subject is human. In one embodiment, the subject is not afflicted with diabetes. In another embodiment, the subject is receiving or is about to receive a chemotherapy drug. In a further embodiment, the chemotherapy drug is adriamycin. In yet a further 20 embodiment, the chemotherapy drug is selected from the following: 5-fluorouracil; Actinomycin D; Alpha interferon; Bleomycin; Cisplatin; Cyclophosphamide; Dexamethasone; Doxorubicin; Epoetin alfa; Etoposide; Gleevec; Herceptin; Interferon alfa; Interleukin-2; 25 Interleukin-11; Methotrexate; Neupogen; Nitrogen Mustard; Paclitaxel; Prednisolone; Prednisone; PROCRIT; Rituximab; Tamoxifen; Thalidomide; Vinblastine; and Additional chemotherapy drugs Vincristine. are listed in chemocare.com envisioned, and 30 (http://www.chemocare.com/bio/default.sps).

In one embodiment of the instant articles of manufacture, the agent is soluble RAGE. In another embodiment, the agent is soluble RAGE G82S. In a

further embodiment, the agent is an antibody directed to RAGE. In yet a further embodiment, the agent is an antibody directed to RAGE G82S.

5 This invention is illustrated in the Experimental Details section which follows. This section is set forth to aid in an understanding of the invention but is not intended to, and should not be construed to, limit in any way the invention set forth in the claims which follow.

Experimental Details

Methods

5

Animal studies

Male BALB/c mice at the age of six weeks received one intravenous dose of adriamycin (ADR), 10.5 mg/kg.

10 Immediately after injection of ADR, mice received once daily administration of murine soluble RAGE, the extracellular ligand binding domain of RAGE, 100 µg per day, beginning immediately at the time of ADR treatment, and continued until the day of sacrifice.

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Morphologic studies

Dissected kidneys were fixed in buffered formalin (10%) overnight and then routinely processed for light 20 microscopy. Fixed paraffin-embedded tissues were cut (3 µm thick) and mounted on slides coated with 3aminopropyltriethoxy silane (Sigma) followed incubation at 37°C overnight. Light microscopic views after staining with periodic acid Schiff (PAS) were 25 scanned into a computer and the quantification of areas of mesangial matrix and glomerulus was performed using microscope and image analysis Zeiss (MediaCybernetics). To calculate mesangial area, only nuclei-free regions were included. Forty glomeruli from 30 each animal were selected at random on the stained sections (20 from the outer region and 20 from the inner region). Morphometry was performed investigators blinded to the experimental protocol.

Functional studies

Twenty-four hour urine collection was obtained from each animal using metabolic cages. Urine albumin and creatinine were determined using Albuwell M and creatinine assays from Exocell (Philadelphia, PA) according to the manufacturer's instructions.

10 Statistical analysis

The mean ± standard error (SE) of the mean is reported. Statistical significance (defined as p<0.05) was determined by ANOVA. Where indicated, post-hoc analysis was employed using Dunnett's t-test using StatView 4.0 (Abacus Concepts, Inc., Berkeley, CA).

Results

20 RAGE and cellular activation

It was in the context of roles for inflammatory cells and podocytes in the pathogenesis of FSGS that a role for Receptor for AGE (RAGE) was first speculated. RAGE is a multiligand member of the immunoglobulin superfamily of cell surface molecules (26-27) that engages distinct molecules; ligand-RAGE interaction activates cell signalling pathways (such as NF-kB; p44/p42, p38 and SAPK/JNK MAP kinases; cdc42/rac; and JAK/STAT, for example) (28-33) that are required for RAGE-mediated effects. Importantly, deletion of the cytosolic tail of RAGE imparts a dominant negative effect in cultured cells and in vivo.

RAGE is principally expressed in the podocyte in the glomerulus

The findings have demonstrated that the principal site of RAGE expression in the glomerulus is the podocyte, at low levels in homeostasis (34); podocyte RAGE expression is upregulated in human and murine diabetes (34).

10 To address the concept that RAGE may be involved in the pathogenesis of ADR-mediated FSGS, a single injection of ADR, 10.5 mg/kg, to male BALB/c mice at age 6 weeks was administered. ADR-treated mice received once daily murine soluble RAGE, the administration of 15 extracellular ligand binding domain of RAGE, 100 µg per day, beginning immediately at the time of treatment, and continued until the day of sacrifice. Other ADR-treated mice received vehicle, PBS. At 2 and 6 weeks after ADR, kidney weight/body weight ratios 20 were significantly decreased in sRAGE-treated vs. PBStreated mice. Examination of mesangial area at 2 and 6 weeks after ADR revealed that in a time-dependent manner, ADR administration was associated with increased mesangial area, and increased mesangial 25 matrix/glomerular area fraction by PAS staining (Fig. 1&2, respectively). At 2 and 6 weeks, administration of sRAGE resulted in significantly decreased mesangial area and mesangial/glomerular area compared with PBS treatment (Fig. 1&2, respectively).

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The key test of these concepts was the degree to which blockade of RAGE would suppress the development of albuminuria. Mice were placed in metabolic cages and 24 hr urine collected. Urine levels of albumin and

creatinine were determined; results are reported as µg albumin/µg creatinine. At 2 weeks after ADR, PBStreated mice displayed an ≈10-fold increase in urine albumin/creatinine compared to saline-treated mice not ADR (809.55±365.85 vs. 85.78±17.56 5 receiving albumin/creatinine; p<0.01) (Fig. 3). receiving ADR and sRAGE, levels of albumin/creatinine were markedly reduced (191.08±49.93; p<0.05 vs. PBStreated mice receiving ADR) (Fig. 3). At six weeks, 10 the results were similarly striking. PBS-treated mice receiving ADR displayed urine albumin/creatinine of 1,362.96±987.97 vs 84.47±49.93 in control mice not receiving ADR; p<0.01 (Fig. 3). In the presence of sRAGE, ADR-mediated albuminuria was significantly 15 reduced, to 249.76±283.19 μg albumin/creatinine; p<0.01 vs PBS/ADR (Fig. 3).

Taken together, these findings strongly support the hypothesis that RAGE activation importantly contributes to mechanisms linked to glomerular injury. Administration of soluble RAGE afforded significant protection against the morphologic and functional indices of glomerular injury upon administration of glomerulosclerosis-inducing agents. RAGE blockade is proposed as a new means to prevent glomerular injury in this class of diseases.

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What is claimed is:

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1. Use of a prophylactically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof for the manufacture of a medicament for inhibiting the onset of a glomerular injury in a subject.

- 2. Use of a therapeutically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof for the manufacture of a medicament for treating a glomerular injury in a subject.
- 3. Use of a prophylactically effective amount of an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof for the manufacture of a medicament for inhibiting the onset of glomerulosclerosis, proteinuria or albunuria in a subject.
- Use of a therapeutically effective amount of an agent that inhibits binding between RAGE and/or RAGE
 G82S and a ligand thereof for the manufacture of a medicament to treat glomerulosclerosis, proteinuria or albunuria in a subject.
- 5. An article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for inhibiting the onset of glomerular injury in a subject.

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6. An article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affized thereto a label indicating a use for the agent for inhibiting the onset of glomerulosclerosis, proteinuria or albuminuria in a subject.

- 7. An article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for treating a glomerular injury in a subject.
- 8. An article of manufacture comprising a packaging material having therein an agent that inhibits binding between RAGE and/or RAGE G82S and a ligand thereof, wherein the packaging material has affixed thereto a label indicating a use for the agent for treating glomerulosclerosis, proteinuria or albuminuria in a subject.
 - 9. The use of claim 1 or 2 wherein the glomerular injury is associated with reduced removal of toxins from the subject.
- 10. The use of claim 1 or 2 wherein the glomerular injury is associated with glomerulosclerosis.
 - 11. The use of claim 1 or 2 wherein the glomerular injury is associated with proteinuria.
 - 12. The use of claim 1 or 2 wherein the glomerular injury is associated with albuminuria.

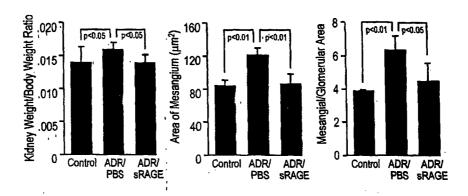
13. The use of claim 1, 2, 3, or 4 wherein the subject is human.

- 14. The article of claim 5, 6, 7, or 8 wherein the subject is human.
- 5 15 The use of claim 13 wherein the subject is afflicted with diabetes
 - 16. The article of claim 14 wherein the subject is afflicted with diabetes.
- 17. The use of claim 15 as it depends on claim 1 or 3 wherein the subject has been afflicted with diabetes for less than 20 years.
 - 18. The article of claim 16 as it depends on claim 5 or 6 wherein the subject has been afflicted with diabetes for less than 20 years.
- 15 19. The use of claim 13, wherein the subject is not afflicted with diabetes.
 - 20. The article of claim 14, wherein the subject is not afflicted with diabetes.
- 21. The use of claim 13, wherein the subject is receiving or is about to receive a chemotherapy drug.
 - 22. The article of claim 14, wherein the subject is receiving or is about to receive a chemotherapy drug.
- 25 23. The use of claim 21 wherein the chemotherapy drug is adriamycin.

24. The article of claim 22, wherein the chemotherapy drug is adriamycin.

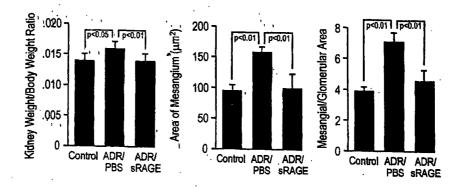
- 25. The use of claim 1, 2, 3, or 4 wherein the agent is soluble RAGE.
- 5 26. The article of claim 5, 6, 7, or 8 wherein the agent is soluble RAGE.
 - 27. The use of claim 1, 2, 3, or 4 wherein the agent is an antibody directed to RAGE.
- 28. The article of claim 5, 6, 7, or 8 wherein the agent is an antibody directed to RAGE.
 - 29. The use of claim 1, 2, 3, or 4 wherein the agent is an antibody directed to RAGE G82S.
 - 30. The article of claim 5, 6, 7, or 8 wherein the agent is an antibody directed to RAGE G82S.

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Figure 1



2/3

Figure 2



3/3

Figure 3.

