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(54) Title: METHOD TO TREAT HEMOPHILIA BY HEPATIC GENE TRANSFER OF FACTOR VIII/IX WITH VESICLE VECTOR

(57) Abstract: Hemophilia is one of the most common genetic disorders. Standard therapies include transfusions with plasma products to provide clotting factors. The invention is a non-viral vesicle vector and method for the treatment of hemophilia. The vesicle vector contains the hepatitis B envelope protein to target the vesicle to the liver for delivery of an expression construct containing the coding sequence for factor VIII or IX driven by an appropriate promoter of factor VIII or IX protein.

**METHOD TO TREAT HEMOPHILIA BY HEPATIC GENE TRANSFER
OF FACTOR VIII/IX WITH VESICLE VECTOR**

CROSS-REFERENCES TO RELATED APPLICATIONS

5 This application claims the benefit of priority of United States provisional application Serial Number 60/286,314 filed April 25,2001 which is incorporated herein by reference in its entirety.

SEQUENCE LISTING

10 A sequence listing is submitted herewith under 35 C.F.R. §1.821 and is incorporated herein by reference.

BACKGROUND OF THE INVENTION

Hemophilia is one of the most common genetic disorders.

15 Hemophilia A caused by deficiency of Factor VIII occurs in about 1 in 5000 male births, while hemophilia B caused by a defect in Factor IX is around 1 in 30,000 male births. The prevalence is very general in all populations studied. Hemophilia has long been treated with clotting factor concentrates, but the aim of this therapy is to control bleeding and requires
20 lifelong repetitive intravenous infusions. Because of the increasing awareness of the risk of plasma derived products, the importance of the development of new and effective treatments is increased.

 Gene therapy approaches have been developed for the treatment of hemophilia. Hemophilia is a particularly attractive model for developing a
25 gene transfer approach for the treatment of disease. The proteins are well characterized, the genes are cloned and available, and there are large and small animal models of the disease. Moreover, there is no essential requirement for tissue specific delivery of the gene product and as protein function is regulated by activation of the protein; therefore, expression
30 levels of the protein need not be tightly regulated. Additionally, only a low level of protein expression is required for phenotypic correction of the disease. The major hurdle of treatment of hemophilia by gene therapy is

that the expression of the gene product must be sustained throughout the life of the individual; therefore, effective therapy would likely require re-administration of the gene therapy vector.

Clinical trials for the treatment of hemophilia using retroviral and adeno-associated viral (AAV) vectors are ongoing. Adenoviral and lentiviral vectors have been used experimentally. However, the problem with all of these viral vectors is that they have a limited capacity for nucleic acid and have been shown to elicit an immune response. The use of DNA or RNA with or without synthetic liposomes results in low efficiency gene transfer. Non-viral methods achieve only short term, non-targeted gene expression.

A novel, liver-specific vesicle vector expressing modified surface proteins of the hepatitis B virus was recently described by Yamada et al (2001a). The vesicles containing the hepatitis B membrane proteins are generated by the methods well known to those skilled in the art (Kuroda et al, 1992, and Yamada et al., 2001b, incorporated herein by reference). Briefly, a modified hepatitis B envelope (env) L protein, containing the pre-S1 + pre-S2 + S peptides, can be effectively generated in yeast by fusing the coding sequence for the chicken lysozyme signal sequence in frame to the beginning of the coding sequence for the modified env L protein (SEQ ID 1). The signal sequence directs the insertion of the proteins into the endoplasmic reticulum during translation. Protein rich vesicles bud from the endoplasmic reticulum and accumulate in the cytoplasm of the yeast cell. The vesicles are composed of lipid bilayers derived from the ER and the modified env L proteins as the major protein component. Particles formed by this method are very stable and can be easily purified through repetitive cesium chloride and sucrose gradients by methods well known to those skilled in the art.

Plasmid DNA can be incorporated into the env L containing particles by electroporation (Yamada et al. 2001a). Such DNA containing particles were demonstrated to facilitate entry of the DNA specifically into liver cells both in culture and upon systemic administration to nude mice in which human

hepatoma cells were transplanted. Yamada et al. (2001a) suggested that such a vesicle vector could be used for tissue specific delivery of nucleic acid and other compounds to the liver.

5

SUMMARY OF THE INVENTION

The invention is a non-viral vesicle vector for the treatment of hemophilia comprising a lipid bilayer containing a modified hepatitis B env L protein such that recognition of the S-peptide by the immune system is attenuated or abrogated, but the liver targeting signals are still exposed on the surface of the vesicle, and an expression construct for the expression of Factor VIII or IX for the treatment of hemophilia A or B, respectively. The expression construct may be single or double stranded DNA containing any of a number of promoters including, but not limited to general (e.g. cytomegalovirus, Rous sarcoma virus) and tissue specific (e.g. alpha fetoprotein, globulin, albumin, α 1-microglobulin) promoters. The construct may contain additional regulatory elements including, but not limited to enhancers, introns, poly A sequences, RNA targeting sequences. Sequences to promote replication of the plasmid including SV40 origin of replication can be included. Inverted terminal repeat (ITR) sequences from AAV can be included in the construct to promote expression cassette stability or to enhance integration into the host DNA with the AAV Rep protein. In lieu of ITR sequences, eukaryotic DNA transposon/transposases systems can be used to promote integration.

The invention is a method for the treatment of hemophilia by administration of the non-viral vesicle vector of the invention. The vesicle vector containing the nucleic acid construct with the appropriate coding sequence is administered intravenously or intraarterially. The individual is monitored for expression of the gene product of interest by detection of the protein or mRNA or by phenotypic recovery.

30

DETAILED DESCRIPTION AND PREFERRED EMBODIMENT

Hemophilia is one of the most common genetic disorders and is a result of a mutation or deletion in any of the clotting factors, most commonly Factor VIII or IX. Treatment requires the lifelong replacement of clotting factors which requires repetitive intravenous infusions and exposes patients to the dangers associated with plasma derived products.

Hemophilia is amenable to treatment with gene therapy for a number of reasons. First, the genes involved are cloned and available. Second, the proteins are well characterized and their activation is regulated by cleavage of the protein rather than at the transcriptional or translational level; therefore, the expression level does not need to be tightly regulated. Third, low levels of protein expression have been demonstrated to be sufficient for phenotypic recovery. Fourth, although the liver is the physiological site of production of most of the Factor VIII and IX, the site of production of the protein within the body is relatively unimportant. Fifth, a number of animal models are available for analysis of various therapies. However, to date no effective gene transfer vectors or methods for the treatment of hemophilia have been developed.

The invention is a vesicle vector for the treatment of hemophilia comprising a natural lipid vesicle preferably produced in yeast or insect cells, such as Sf9 cells, containing modified hepatitis B env L protein integrated into the membrane and an expression construct inside the vesicle for the expression of Factor VIII or IX. The vesicles are prepared by the vaccine production method of Kuroda (1992) further refined by Yamada (2001b). Briefly, the hepatitis B env L protein is composed of three regions: the 108- or 119-residue pre-S1 region involved in the direct interaction with hepatocytes, the 55-residue pre-S2 region associated with the polymerized albumin-mediated interaction and the major 226-residue S-protein region. Attempts to produce L protein in various eukaryotic cells had been unsuccessful, probably due to the presence of the N-terminus of

the pre-S1 peptide. The coding sequence of the N-terminus of the L protein was replaced by a chicken lysosome signal sequence to direct the translocation of the N-terminus through the endoplasmic reticulum (ER). The chimeric sequence was inserted into a yeast (*S. cerevisiae*) expression vector and inserted into yeast using a standard transformation protocol. 5 The chimeric L-protein was produced in abundance, up to 42% of the total yeast protein, and was properly inserted into the membrane. Vesicles budded off of the ER to form 23 nm spherical and filamentous particles containing the protein in the membrane of the vesicles. The yeast cells were disrupted with glass beads to release the vesicles. Vesicles were 10 purified by serial rounds of discontinuous cesium and sucrose gradients. Production and purification of vesicles from insect cells would be performed in a similar method. A crude membrane fraction could be prepared as with the yeast cells, by homogenization and differential 15 centrifugation. The fraction can be loaded onto cesium or sucrose gradients as with the yeast extract for purification of vesicles. The methods are amenable to inexpensive, large scale production of vesicles which is necessary for gene transfer. Vesicles are stable for long term storage at a low temperature but are unstable upon repeated freeze-thaw 20 cycles.

The vesicle vectors can be used for the delivery of any nucleic acid construct, single- or double-stranded DNA or RNA, or gene product to the liver. In a preferred embodiment of the invention, the nucleic acid is a double stranded DNA plasmid. The construct minimally contains the 25 coding sequence for human Factor VIII (SEQ ID 2) or IX (SEQ ID 3) for the treatment of hemophilia A or B respectively and a promoter to allow for transcription of the hemophilia gene. The construct may optionally contain additional regulatory and enhancer elements to modulate gene expression, intron and poly-A sequences to promote RNA processing and gene 30 expression, RNA targeting sequences, AAV-ITR or eukaryotic transposon

sequences to promote stabilization of expression cassettes and integration into the host genome and viral origin of replication sequences to promote amplification of the plasmid in host cells. Such sequences are well known to those skilled in the art. The number of elements that can be inserted
5 into the nucleic acid construct as the size is not limited by the requirements of a viral genome as is the case with many gene transfer protocols.

Any of a number of promoter sequences are known to be functional in liver cells. These include both non-tissue specific promoters such as CMV, RSV, ubiquitin, chicken β -actin and elongation factor (EF)-1 α ; and
10 tissue specific promoters such as alpha-fetoprotein, globulin, α 1-microglobulin and albumin.

AAV-ITR sequences may be incorporated into the construct flanking all of the coding and regulatory sequences, other than any origins of replication. The AAV-ITR sequences have been demonstrated to increase
15 the stability of transferred constructs in gene therapy protocols.

Alternatively, the AAV-ITR sequences may enhance integration into the human genome at a specific site with the cooperation of the AAV-Rep protein, which may be supplied by incorporation into the vesicles with the nucleic acid construct or by expression cassettes packaged into the same
20 vesicle.

Eukaryotic transposon sequences can be incorporated into the construct flanking all of the coding sequences and regulatory elements, similar to the AAV-ITR sequences. Transposase to promote integration may be expressed from the same expression cassette or from a separate
25 expression cassette packaged into the same vesicle.

Special considerations may be taken when expressing Factor VIII. Studies have demonstrated that human Factor VIII contains a sequence (nucleotides 1741 to 1771 in SEQ ID 2) that decreases heterologous expression of proteins (Fallaux et al., 1996). The sequence is AT-rich and
30 has been demonstrated to bind a nuclear factor and repress expression of

a reporter construct in cells. Deletion or random mutation of the sequence results in a non-functional Factor VIII. However, silent mutations that result in no change in the amino acid sequence of the gene product can be introduced into the coding sequence by methods well known to those skilled in the art to enhance expression of Factor VIII.

In a preferred embodiment, the nucleic acid construct of the invention is introduced into the vesicles by electroporation. The nucleic acid construct is mixed thoroughly with the vesicles, brought to a final volume in water and transferred to an electroporation cuvette. Voltage and resistance vary widely depending on the size (gap length) of the cuvette and the volume of material in the cuvette. Such parameters can be readily modified by methods well known to those skilled in the art to result in maximum transfer of nucleic acid into vesicles with minimum destruction of vesicles.

Alternatively the nucleic acid may be introduced into the vesicle by fusion with nucleic acid containing liposomes by methods well known to those skilled in the art (Dzau et al, 1996). The construct of the invention is encapsulated into liposomes prepared by vortexing. Liposomes may be composed of known phospholipids and membrane components (e.g. phosphatidyl-choline, cholesterol) or of commercially available proprietary mixtures of membrane components (e.g. Lipofectamine from Gibco-BRL). Nucleic acid encapsulated in liposomes will fuse with the yeast or insect cell derived vesicles upon incubation at 37°C for 10-30 minutes.

Alternatively, factor VIII or IX protein may be incorporated into the vesicle vector of the invention. Factor VIII (SEQ ID 4) and IX (SEQ ID 5) protein may be produced using any of a number of methods well known to those skilled in the art. A solution containing a high concentration of protein may be mixed with purified vesicles and subjected to osmotic shock or sonication to promote incorporation of the protein into the vesicles. Protein may also be incorporated into artificial membranes by

vortexing or sonication. The artificial membranes containing the protein can be fused with the hepatitis B vesicles.

The nucleic acid or protein containing non-viral vesicle vectors of the invention are administered to the individual intravenously or
5 intraarterially. To increase delivery, the vesicle vector can be administered directly into the hepatic or portal artery. The individual is monitored on regular intervals for the presence of factor VIII or IX or for phenotypic recovery. The amount of the non-viral vesicle to be administered would
10 depend on the strength of the promoter, integration sequences, number of plasmids per vesicle and a number of other considerations well known to those skilled in the art. As methods for monitoring the state of health of individuals are well known, an effective dose can be readily determined.

Although an exemplary embodiment of the invention has been
15 described above by way of example only, it will be understood by those skilled in the field that modifications may be made to the disclosed embodiment without departing from the scope of the invention, which is defined by the appended claims.

20 REFERENCES

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WE CLAIM:

10

CLAIMS

- 2 1. A non-viral vesicle vector comprising:
a vesicular membrane with hepatitis B envelope (env) protein
4 exposed on the vesicle surface and
a nucleic acid expression construct comprising a complete factor VIII
6 or factor IX coding sequence and a promoter sequence functional in liver
cells.
- 8 2. The vesicle vector of claim 1, wherein the env protein contains
10 mutations to reduce antigenicity.
- 12 3. The vesicle vector of claim 1, wherein the expression construct is
DNA.
- 14 4. The vesicle vector of claim 1, wherein the expression construct is
16 double stranded plasmid DNA.
- 18 5. The vesicle vector of claim 1, wherein the expression construct is
RNA.
- 20 6. The vesicle vector of claim 1, wherein the promoter is a non-
22 tissue specific promoter.
- 24 7. The vesicle vector of claim 6, wherein the non-tissue specific
promoter is selected from the group consisting of cytomegalovirus
26 promoter, Rous sarcoma virus promoter, ubiquitin promoter, chicken β -
actin promoter and elongation factor 1 α promoter.
- 28 8. The vesicle vector of claim 1, wherein the promoter is a liver
30 specific promoter.

- 32 9. The vesicle vector of claim 8, wherein the liver specific promoter
is selected from the group consisting of alpha-fetoprotein promoter,
34 globulin promoter, α 1-microglobulin and albumin.
- 36 10. The vesicle vector of claim 1, wherein the expression construct
comprises inverted terminal repeat sequences from adeno-associated virus
38 (AAV-ITR).
- 40 11. The vesicle vector of claim 1, wherein the expression construct
comprises eukaryotic transposon and transposase sequences.
42
12. The vesicle vector of claim 1, wherein the expression construct
44 comprises the coding sequence of factor VIII.
- 46 13. The vesicle vector of claim 12, wherein the factor VIII comprises
silent mutations to enhance expression.
48
14. The vesicle vector of claim 1, wherein the expression construct
50 comprises the coding sequence of factor IX.
- 52 15. A non-viral vesicle vector comprising:
a vesicular membrane with hepatitis B envelope (env) protein
54 exposed on the vesicle surface and
a protein comprising a complete factor VIII or factor IX.
56
16. The vesicle vector of claim 15, wherein the env protein contains
58 mutations to reduce antigenicity.
- 60 17. A method for treatment of hemophilia comprising:

administration into circulation of an individual with hemophilia a
62 non-viral vesicle vector comprising a vesicular membrane with hepatitis B
env protein exposed on the vesicle surface and
64 a nucleic acid expression construct comprising a complete factor VIII
or IX coding sequence and a promoter sequence functional in liver cells
66 and
monitoring the individual for amelioration of disease.

68

18. The method of claim 17, wherein administration into circulation
70 comprises intravenous administration.

72

19. The method of claim 17, wherein administration into circulation
comprises administration into a hepatic or portal artery

74

SEQUENCE LISTING

<110> Chien, Kenneth R
Hoshijima, Masahiko

<120> Method to treat hemophilia by hepatic gene transfer of Factor VIII/IX with vesicle vector

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<150> 60/286,314

<151> 2001-04-25

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<170> PatentIn version 3.1

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Trp Asp Tyr Met Gln Ser Asp Leu Gly Glu Leu Pro Val Asp Ala Arg
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Phe Pro Pro Arg Val Pro Lys Ser Phe Pro Phe Asn Thr Ser Val Val
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Tyr Lys Lys Thr Leu Phe Val Glu Phe Thr Asp His Leu Phe Asn Ile
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Ala Lys Pro Arg Pro Pro Trp Met Gly Leu Leu Gly Pro Thr Ile Gln
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Ala Glu Val Tyr Asp Thr Val Val Ile Thr Leu Lys Asn Met Ala Ser
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His Pro Val Ser Leu His Ala Val Gly Val Ser Tyr Trp Lys Ala Ser
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Tyr Leu Ser His Val Asp Leu Val Lys Asp Leu Asn Ser Gly Leu Ile
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Pro Glu Ile Glu Val Thr Trp Ala Lys Gln Gly Arg Thr Glu Arg
 1640 1645 1650

Leu Cys Ser Gln Asn Pro Pro Val Leu Lys Arg His Gln Arg Glu
 1655 1660 1665

Ile Thr Arg Thr Thr Leu Gln Ser Asp Gln Glu Glu Ile Asp Tyr
 1670 1675 1680

Asp Asp Thr Ile Ser Val Glu Met Lys Lys Glu Asp Phe Asp Ile
 1685 1690 1695

Tyr Asp Glu Asp Glu Asn Gln Ser Pro Arg Ser Phe Gln Lys Lys
 1700 1705 1710

Thr Arg His Tyr Phe Ile Ala Ala Val Glu Arg Leu Trp Asp Tyr
 1715 1720 1725

Gly Met Ser Ser Ser Pro His Val Leu Arg Asn Arg Ala Gln Ser

1730						1735								1740
Gly	Ser	Val	Pro	Gln	Phe	Lys	Lys	Val	Val	Phe	Gln	Glu	Phe	Thr
1745						1750					1755			
Asp	Gly	Ser	Phe	Thr	Gln	Pro	Leu	Tyr	Arg	Gly	Glu	Leu	Asn	Glu
1760						1765					1770			
His	Leu	Gly	Leu	Leu	Gly	Pro	Tyr	Ile	Arg	Ala	Glu	Val	Glu	Asp
1775						1780					1785			
Asn	Ile	Met	Val	Thr	Phe	Arg	Asn	Gln	Ala	Ser	Arg	Pro	Tyr	Ser
1790						1795					1800			
Phe	Tyr	Ser	Ser	Leu	Ile	Ser	Tyr	Glu	Glu	Asp	Gln	Arg	Gln	Gly
1805						1810					1815			
Ala	Glu	Pro	Arg	Lys	Asn	Phe	Val	Lys	Pro	Asn	Glu	Thr	Lys	Thr
1820						1825					1830			
Tyr	Phe	Trp	Lys	Val	Gln	His	His	Met	Ala	Pro	Thr	Lys	Asp	Glu
1835						1840					1845			
Phe	Asp	Cys	Lys	Ala	Trp	Ala	Tyr	Phe	Ser	Asp	Val	Asp	Leu	Glu
1850						1855					1860			
Lys	Asp	Val	His	Ser	Gly	Leu	Ile	Gly	Pro	Leu	Leu	Val	Cys	His
1865						1870					1875			
Thr	Asn	Thr	Leu	Asn	Pro	Ala	His	Gly	Arg	Gln	Val	Thr	Val	Gln
1880						1885					1890			
Glu	Phe	Ala	Leu	Phe	Phe	Thr	Ile	Phe	Asp	Glu	Thr	Lys	Ser	Trp
1895						1900					1905			
Tyr	Phe	Thr	Glu	Asn	Met	Glu	Arg	Asn	Cys	Arg	Ala	Pro	Cys	Asn
1910						1915					1920			
Ile	Gln	Met	Glu	Asp	Pro	Thr	Phe	Lys	Glu	Asn	Tyr	Arg	Phe	His
1925						1930					1935			
Ala	Ile	Asn	Gly	Tyr	Ile	Met	Asp	Thr	Leu	Pro	Gly	Leu	Val	Met
1940						1945					1950			

Ala Gln Asp Gln Arg Ile Arg Trp Tyr Leu Leu Ser Met Gly Ser
1955 1960 1965

Asn Glu Asn Ile His Ser Ile His Phe Ser Gly His Val Phe Thr
1970 1975 1980

Val Arg Lys Lys Glu Glu Tyr Lys Met Ala Leu Tyr Asn Leu Tyr
1985 1990 1995

Pro Gly Val Phe Glu Thr Val Glu Met Leu Pro Ser Lys Ala Gly
2000 2005 2010

Ile Trp Arg Val Glu Cys Leu Ile Gly Glu His Leu His Ala Gly
2015 2020 2025

Met Ser Thr Leu Phe Leu Val Tyr Ser Asn Lys Cys Gln Thr Pro
2030 2035 2040

Leu Gly Met Ala Ser Gly His Ile Arg Asp Phe Gln Ile Thr Ala
2045 2050 2055

Ser Gly Gln Tyr Gly Gln Trp Ala Pro Lys Leu Ala Arg Leu His
2060 2065 2070

Tyr Ser Gly Ser Ile Asn Ala Trp Ser Thr Lys Glu Pro Phe Ser
2075 2080 2085

Trp Ile Lys Val Asp Leu Leu Ala Pro Met Ile Ile His Gly Ile
2090 2095 2100

Lys Thr Gln Gly Ala Arg Gln Lys Phe Ser Ser Leu Tyr Ile Ser
2105 2110 2115

Gln Phe Ile Ile Met Tyr Ser Leu Asp Gly Lys Lys Trp Gln Thr
2120 2125 2130

Tyr Arg Gly Asn Ser Thr Gly Thr Leu Met Val Phe Phe Gly Asn
2135 2140 2145

Val Asp Ser Ser Gly Ile Lys His Asn Ile Phe Asn Pro Pro Ile
2150 2155 2160

Ile Ala Arg Tyr Ile Arg Leu His Pro Thr His Tyr Ser Ile Arg
2165 2170 2175

Ser Thr Leu Arg Met Glu Leu Met Gly Cys Asp Leu Asn Ser Cys
 2180 2185 2190

Ser Met Pro Leu Gly Met Glu Ser Lys Ala Ile Ser Asp Ala Gln
 2195 2200 2205

Ile Thr Ala Ser Ser Tyr Phe Thr Asn Met Phe Ala Thr Trp Ser
 2210 2215 2220

Pro Ser Lys Ala Arg Leu His Leu Gln Gly Arg Ser Asn Ala Trp
 2225 2230 2235

Arg Pro Gln Val Asn Asn Pro Lys Glu Trp Leu Gln Val Asp Phe
 2240 2245 2250

Gln Lys Thr Met Lys Val Thr Gly Val Thr Thr Gln Gly Val Lys
 2255 2260 2265

Ser Leu Leu Thr Ser Met Tyr Val Lys Glu Phe Leu Ile Ser Ser
 2270 2275 2280

Ser Gln Asp Gly His Gln Trp Thr Leu Phe Phe Gln Asn Gly Lys
 2285 2290 2295

Val Lys Val Phe Gln Gly Asn Gln Asp Ser Phe Thr Pro Val Val
 2300 2305 2310

Asn Ser Leu Asp Pro Pro Leu Leu Thr Arg Tyr Leu Arg Ile His
 2315 2320 2325

Pro Gln Ser Trp Val His Gln Ile Ala Leu Arg Met Glu Val Leu
 2330 2335 2340

Gly Cys Glu Ala Gln Asp Leu Tyr
 2345 2350

<210> 5
 <211> 461
 <212> PRT
 <213> Homo sapiens

<400> 5

Met Gln Arg Val Asn Met Ile Met Ala Glu Ser Pro Gly Leu Ile Thr
 1 5 10 15

Ile Cys Leu Leu Gly Tyr Leu Leu Ser Ala Glu Cys Thr Val Phe Leu
 20 25 30

Asp His Glu Asn Ala Asn Lys Ile Leu Asn Arg Pro Lys Arg Tyr Asn
 35 40 45

Ser Gly Lys Leu Glu Glu Phe Val Gln Gly Asn Leu Glu Arg Glu Cys
 50 55 60

Met Glu Glu Lys Cys Ser Phe Glu Glu Ala Arg Glu Val Phe Glu Asn
 65 70 75 80

Thr Glu Arg Thr Thr Glu Phe Trp Lys Gln Tyr Val Asp Gly Asp Gln
 85 90 95

Cys Glu Ser Asn Pro Cys Leu Asn Gly Gly Ser Cys Lys Asp Asp Ile
 100 105 110

Asn Ser Tyr Glu Cys Trp Cys Pro Phe Gly Phe Glu Gly Lys Asn Cys
 115 120 125

Glu Leu Asp Val Thr Cys Asn Ile Lys Asn Gly Arg Cys Glu Gln Phe
 130 135 140

Cys Lys Asn Ser Ala Asp Asn Lys Val Val Cys Ser Cys Thr Glu Gly
 145 150 155 160

Tyr Arg Leu Ala Glu Asn Gln Lys Ser Cys Glu Pro Ala Val Pro Phe
 165 170 175

Pro Cys Gly Arg Val Ser Val Ser Gln Thr Ser Lys Leu Thr Arg Ala
 180 185 190

Glu Thr Val Phe Pro Asp Val Asp Tyr Val Asn Ser Thr Glu Ala Glu
 195 200 205

Thr Ile Leu Asp Asn Ile Thr Gln Ser Thr Gln Ser Phe Asn Asp Phe
 210 215 220

Thr Arg Val Val Gly Gly Glu Asp Ala Lys Pro Gly Gln Phe Pro Trp
 225 230 235 240

Gln Val Val Leu Asn Gly Lys Val Asp Ala Phe Cys Gly Gly Ser Ile

245 250 255

Val Asn Glu Lys Trp Ile Val Thr Ala Ala His Cys Val Glu Thr Gly
260 265 270

Val Lys Ile Thr Val Val Ala Gly Glu His Asn Ile Glu Glu Thr Glu
275 280 285

His Thr Glu Gln Lys Arg Asn Val Ile Arg Ile Ile Pro His His Asn
290 295 300

Tyr Asn Ala Ala Ile Asn Lys Tyr Asn His Asp Ile Ala Leu Leu Glu
305 310 315 320

Leu Asp Glu Pro Leu Val Leu Asn Ser Tyr Val Thr Pro Ile Cys Ile
325 330 335

Ala Asp Lys Glu Tyr Thr Asn Ile Phe Leu Lys Phe Gly Ser Gly Tyr
340 345 350

Val Ser Gly Trp Gly Arg Val Phe His Lys Gly Arg Ser Ala Leu Val
355 360 365

Leu Gln Tyr Leu Arg Val Pro Leu Val Asp Arg Ala Thr Cys Leu Arg
370 375 380

Ser Thr Lys Phe Thr Ile Tyr Asn Asn Met Phe Cys Ala Gly Phe His
385 390 395 400

Glu Gly Gly Arg Asp Ser Cys Gln Gly Asp Ser Gly Gly Pro His Val
405 410 415

Thr Glu Val Glu Gly Thr Ser Phe Leu Thr Gly Ile Ile Ser Trp Gly
420 425 430

Glu Glu Cys Ala Met Lys Gly Lys Tyr Gly Ile Tyr Thr Lys Val Ser
435 440 445

Arg Tyr Val Asn Trp Ile Lys Glu Lys Thr Lys Leu Thr
450 455 460