



(86) Date de dépôt PCT/PCT Filing Date: 2005/12/19
(87) Date publication PCT/PCT Publication Date: 2006/06/22
(85) Entrée phase nationale/National Entry: 2007/06/04
(86) N° demande PCT/PCT Application No.: EP 2005/013954
(87) N° publication PCT/PCT Publication No.: 2006/063862
(30) Priorités/Priorities: 2004/12/17 (EP04029934.9);
2004/12/21 (US60/637,900)

(51) Cl.Int./Int.Cl. *C12N 15/82* (2006.01),
A01H 5/00 (2006.01)
(71) Demandeur/Applicant:
BAYER CROPS SCIENCE AG, DE
(72) Inventeurs/Inventors:
KOK-JACON, GERALDINE, BE;
VINCKEN, JEAN-PAUL, NL;
SUURS, LUC CJM, NL;
FROHBERG, CLAUS, DE;
VISSER, RICHARD GF, NL
(74) Agent: FETHERSTONHAUGH & CO.

(54) Titre : PLANTE TRANSFORMÉE EXPRIMANT UNE DEXTRANESUCRASE ET SYNTHÉTISANT UN AMIDON MODIFIÉ

(54) Title: TRANSFORMED PLANT EXPRESSING A DEXTRANSUCRASE AND SYNTHESIZING A MODIFIED STARCH

(57) **Abrégé/Abstract:**

The present invention relates to plant cells and plants, which are genetically modified, wherein the genetic modification leads to the expression in plastids of such plant cells and plants of an enzyme having the activity of a dextransucrase. Furthermore, the present invention relates to means and methods for the manufacture of such plant cells and plants. Plant cells and plants of this type synthesize a modified starch. The present invention therefore also relates to the starch synthesized by the plant cells and plants according to the invention as well as to methods for the manufacture of the starch and to the manufacture of starch derivatives of this modified starch.



(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
22 June 2006 (22.06.2006)

PCT

(10) International Publication Number
WO 2006/063862 A1

(51) International Patent Classification:
C12N 15/82 (2006.01) *A01H 5/00* (2006.01)

(74) Agent: KOSSMANN, Jochen; Bayer Bioscience GmbH,
Hermannswerder 20 a, 14473 Potsdam (DE).

(21) International Application Number:
PCT/EP2005/013954

(81) Designated States (*unless otherwise indicated, for every kind of national protection available*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, LY, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NG, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SM, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(22) International Filing Date:
19 December 2005 (19.12.2005)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
04029934.9 17 December 2004 (17.12.2004) EP
60/637,900 21 December 2004 (21.12.2004) US

(84) Designated States (*unless otherwise indicated, for every kind of regional protection available*): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IS, IT, LT, LU, LV, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

(71) Applicant (*for all designated States except US*): BAYER CROPSCIENCE GMBH [DE/DE]; Brüningstrasse 50, 65929 Frankfurt (DE).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): KOK-JACON, Geraldine [FR/BE]; 4 rue Renè Mènada, B-1320 Hamme-Mille (BE). VINCKEN, Jean-Paul [NL/NL]; Willebrordweg 23, NL-6871 ZS Renkum (NL). SUURS, Luc CJM [NL/NL]; Bernadottestraat 9, NL-6671 BM, Zetten (NL). FROHBERG, Claus [DE/DE]; Pilzwald 17, 14532 Kleinmachnow (DE). VISSER, Richard GF [NL/NL]; Emmalaan 17, NL-6721 ET Bennekom (NL).

Declaration under Rule 4.17:

— *of inventorship (Rule 4.17(iv))*

Published:

— *with international search report*

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: TRANSFORMED PLANT EXPRESSING A DEXTRANSUCRASE AND SYNTHESIZING A MODIFIED STARCH

(57) Abstract: The present invention relates to plant cells and plants, which are genetically modified, wherein the genetic modification leads to the expression in plastids of such plant cells and plants of an enzyme having the activity of a dextranucrase. Furthermore, the present invention relates to means and methods for the manufacture of such plant cells and plants. Plant cells and plants of this type synthesis a modified starch. The present invention therefore also relates to the starch synthesized by the plant cells and plants according to the invention as well as to methods for the manufacture of the starch and to the manufacture of starch derivatives of this modified starch.

WO 2006/063862 A1

DEMANDE OU BREVET VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET COMPREND PLUS D'UN TOME.

CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 50

NOTE : Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE THAN ONE VOLUME

THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 50

NOTE: For additional volumes, please contact the Canadian Patent Office

NOM DU FICHER / FILE NAME :

NOTE POUR LE TOME / VOLUME NOTE:

Transformed plant expressing a dextransucrase and synthesizing a modified starch

The present invention relates to plant cells and plants, which are
5 genetically modified, wherein the genetic modification leads to the
expression in plastids of such plant cells and plants of an enzyme having
the activity of a dextransucrase. Furthermore, the present invention relates
to means and methods for the manufacture of such plant cells and plants.
Plant cells and plants of this type synthesise a modified starch. The
10 present invention therefore also relates to the starch synthesised by the
plant cells and plants according to the invention as well as to methods for
the manufacture of the starch and to the manufacture of starch derivatives
of this modified starch.

15 With respect to the increasing significance which has recently been
ascribed to vegetal substances as regenerative sources of raw materials,
one of the objects of biotechnological research is to try to adapt vegetal
raw materials to the demands of the processing industry. In order to
enable the use of regenerative raw materials in as many areas as
20 possible, it is furthermore important to obtain a large variety of substances.
Apart from oils, fats and proteins, polysaccharides constitute the essential
regenerative raw materials derived from plants. Apart from cellulose,
starch maintains an important position among the polysaccharides, being
one of the most significant storage substances in higher plants.

25

Starch is deposited as granules in the chloroplasts of green leaves
(transitory starch) and in amyloplasts of tubers, roots and seeds (storage
starch) (Kossmann and Llyod 2000).

The polysaccharide starch is a polymer made up of chemically homogeneous basic components, namely the glucose molecules. However, it constitutes a highly complex mixture from various types of molecules which differ from each other in their degree of polymerization and in the degree of branching of the glucose chains. Therefore, starch is not a homogeneous raw material. One differentiates particularly between amylose-starch, a basically non-branched polymer made up of alpha-1,4-glycosidically branched glucose molecules, and amylopectin-starch which in turn is a complex mixture of various branched glucose chains. The branching results from additional alpha-1,6-glycosidic interlinkings.

In plant storage organs, starch biosynthesis takes place within the amyloplast and is the result of different reactions such as synthesis (polymerization of glucosyl residues), rearrangement and degradation, in which various starch synthases (E.C.2.4.1.21), transferases (branching (E.C.2.4.1.18) and disproportionating enzyme (E.C.2.4.1.25)), and hydrolytic enzymes (debranching enzyme (E.C.3.2.1.41)), respectively, play key roles.

In order to enable as wide a use of starch as possible, it seems to be desirable that plants be provided which are capable of synthesizing modified starch which is particularly suitable for various uses. One possibility to provide such plants--apart from breeding methods--is the specific genetic modification of the starch metabolism of starch-producing plants by means of recombinant DNA techniques.

Over the years, several studies have been done aimed at turning the amyloplast into a more versatile polysaccharide factory. For this purpose, several microbial enzymes have been equipped with a plastidial targeting transit, and their influence on starch structure and functionality has been

investigated.

Certain bacteria possess an array of enzymes, so-called glucansucrases, which can attach (contiguous) 1,6-linked or 1,3-linked glucosyl residues to maltodextrins. With few exceptions, glucansucrases are extracellular enzymes, which are produced by lactic acid bacteria such as *Leuconostoc mesenteroides* strains, oral Streptococci, and some species of *Lactobacillus* and *Lactococcus* (Robyt 1995; van Geel-Schutten et al. 1999). In addition, they are produced by other bacteria such as some of the *Neisseria* strains (Hehre et al. 1949). These strains are involved in different processes in nature. Some of the strains colonize the oral cavity of humans and animals and can induce the formation of dental caries. Other strains can invade the throat such as the commensal *Neisseria* species. Some *Lactobacillus* species increase the viscosity of fermented milk (de Vuyst and Degeest 1999).

The glucansucrases catalyse the polymerisation of glucose residues from sucrose, which leads to the production of a large variety of α -glucans with different sizes and structures, and composed of diverse linkage types.

Glucansucrases can be classified according to the structure of the glucan formed, and in particular the nature and frequency of the glucosidic linkages synthesized.

Dextran sucrose (DSR) (E.C.2.4.1.5) synthesizes a glucan, called dextran, mainly composed of α -(1 \rightarrow 6) glucosyl residues in the main linear chain and branched by variable proportions of α -(1 \rightarrow 2), α -(1 \rightarrow 3) or α -(1 \rightarrow 4) linkages (Jeanes et al., 1954; Sidebotham, 1975; Robyt, 1995).

Biosynthesis of dextrans is mediated by *Lactobacillus*, *Leuconostoc*, and *Streptococcus* bacteria in the presence of sucrose.

Nucleic acid sequences encoding dextransucrases are well known in the art and numerous different sequences have been identified for numerous different dextransucrases (GenBank Database).

Dextran produced by *Leuconostoc mesenteroides* NRRL B-512F is water-soluble, and consists of 95 % α -(1 \rightarrow 6) linkages in the main chain and 5 % α -(1 \rightarrow 3) side chains (van Cleve *et al.*, 1956). Its biosynthesis is mediated by a dextransucrase DSR-S (EC 2.4.1.5), which is a 1,527 amino-acid glucosyltransferase (Wilke-Douglas *et al.*, 1989; Monchois *et al.*, 1997). Its catalytic properties can be summarized as follows: after cleavage of sucrose, the glucose residue can be transferred to a growing dextran chain by the so-called two-site insertion mechanism, or to acceptor molecules (Robyt, 1995; Monchois *et al.*, 1999).

The sequence of the *Dsr-S* gene from *L. mesenteroides* NRRL B-512F has been reported in WO 89/12386 and by Quirasco *et al.* (1999); GenBank (Accession AJ271107).

Most glucansucrases share a common structure composed of four different regions: a signal peptide, a variable region, a catalytic domain, and a glucan-binding domain (GBD). (Monchois *et al.*, 1999, FEMS Microbiology Letters 177, 243-248; Monchois *et al.*, 1999, FEMS Microbiology Reviews 23, 131-151).

The signal peptide consists of 35-38 amino acids and is responsible for secretion of the sucrases, when expressed by their natural bacterial hosts. The signal peptide is followed by a variable region of 140-261 amino acids. Janecek *et al.* (2000) showed that conserved, long repeat elements (A-like repeats) are present in the downstream part of this region of dextransucrases of *Leuconostoc mesenteroides* NRRL B-512F (DSR-S), NRRL B-1299 (DSR-B). It has been hypothesized that these repeats can

play a role in glucan binding. Nevertheless, this region does not seem to be essential for enzyme activity because DSR-A, which is produced by *Leuconostoc mesenteroides* NRRL B-1299 and catalyses the formation of a polymer containing between 27 and 35% of α -(1 \rightarrow 2) branched linkages in addition to α -(1 \rightarrow 6) ones (Robyt et al, 1978), does not possess this region and is still active.

The catalytic domain is composed of about 900 amino acids and is highly conserved within the *Leuconostoc* and *Streptococcus* species (MacGregor et al. 1996). The catalytic domain is also called the sucrose-binding domain because it contains a catalytic triad of aspartic and glutamic acid residues that play an important role in binding and cleavage of sucrose molecules.

The glucan-binding domain is covering about 500 amino acids, and is composed of repeats named A,B,C, D that are defined by a consensus sequence (Monchois et al 1998, 1999). Nevertheless, the number and organization of these repeats is variable within glucansucrases, and it has been shown that the minimum number of these repeated units necessary to ensure glucan binding properties is different according to the enzymes, and more particularly is different for enzymes producing a soluble glucan than for those producing an insoluble one (Monchois et al., 1999)

The elongation of glucan chains by glucansucrases is quite different compared to that by starch synthases. First, the preferred substrate is sucrose instead of ADP-Glucose. Second, the glucose residues are added to the reducing end of a growing glucan chain by a so-called two-site insertion mechanism (Robyt 1995).

In addition, the branching of glucans does not take place by means of a branching enzyme as in starch biosynthesis, but by a so-called acceptor reaction catalyzed by the glucansucrases themselves (Robyt, 1995). The glucansucrase is thought to contain an acceptor-binding site that can bind
5 acceptor molecules such as the nascent glucan chains or maltodextrins (Su and Robyt, 1994).

The efficiency to catalyse acceptor reactions, particularly with starch polymers or maltodextrins is nevertheless unpredictable, as the structure-
10 function relationships underlying the acceptor reaction are not understood and is poorly documented. It seems nevertheless that the relative acceptor efficiency depends on the size of the acceptor molecules (Fu *et al.* 1990), and it is uncertain that amylopectine and amylose may be acceptor molecules for glucansucrases.

15 Starch polymer modification has been achieved by targeting the *Escherichia coli* glycogen synthase (GLGA) and the glycogen branching enzyme (GLGB) to the potato amyloplast (Shewmaker *et al.* 1994; Kortstee *et al.* 1996). In both cases, the natural balance of chain
20 elongation and branching was disturbed, resulting in starch granules with altered physical properties, and with more heavily branched polymers.

Attachment of novel glycosyl residues to starch polymers has also been an objective. For this purpose, a *Bacillus subtilis* levansucrase
25 (E.C.2.4.1.10) was introduced in potato tuber amyloplasts (Gerrits *et al.* 2001). Levansucrase can polymerize the fructose moiety of the donor substrate sucrose into a high molecular weight fructan. Nevertheless, the starch yield was severely compromised and the starch morphology was dramatically altered.

30

It has also been tried to convert starch in planta into high-value cyclic oligosaccharides, which can accommodate hydrophobic substances in their apolar cavity and can be used in various food and pharmaceutical applications. A cyclodextrin glycosyltransferase (CGTase; E.C.2.4.1.19) from *Klebsiella pneumoniae* was introduced into potato amyloplasts (Oakes *et al.* 1991) for cyclodextrin production. Only 0.01% of the endogenous starch was converted to the desired product, and this product was difficult to recover from the plant material.

These examples demonstrate that bacterial enzymes can be potentially powerful tools for starch modification, but that their performance in the plant are unpredictable beforehand (Kok-Jacob A. *et al.*, 2003).

The object of the present invention is therefore based on providing modified starch, new plant cells and/or plants, which synthesise such a modified starch, as well as methods for producing said plants.

DESCRIPTION OF FIGURES

Figure 1: map of plasmid pBinDsrS

Figure 2 : Modified starch granule morphology observed by scanning electron microscopy analysis performed on the starch of a selected transformant (dsrS30), compared to the starch of a wild-type Kardal plant (Kardal)

25

Therefore, the present invention relates to genetically modified plant cells or genetically modified plants characterized in that they show an enzymatic activity of a dextransucrase DSR-S protein in plastids and wherein said genetically modified plant cells or genetically modified plants synthesize a modified starch in comparison to starch synthesized by

30

corresponding non-genetically modified wild-type plant cells or wild type plants, respectively.

- 5 The term "genetically modified" or "transformed" refers to a plant cell or a plant having stably integrated in its genome at least one transgene. Preferentially, the transgene comprises a chimeric nucleic acid sequence comprising at least one element originating from an other organism than the transformed plant cell or transformed plant (heterologous transgene).
- 10 Particularly, the transgene is a recombinant transgene which comprises at least a promoter, a coding sequence and optionally a termination signal. More preferably the coding sequence of the recombinant transgene encodes a dextransucrase protein, most preferably a dextransucrase DSR-S protein.

15

- In conjunction with the present invention, the term "wild type plant cell" or "wild type plant" means that the plant cells or plants concerned were used as starting material for the manufacture of the plant cells according to the invention, i.e. their genetic information, apart from the introduced genetic
- 20 modification, corresponds to that of a plant cell according to the invention.

- In conjunction with the present invention, the term "corresponding" means that, in the comparison of several objects, the objects concerned that are compared with one another have been kept under the same conditions. In
- 25 conjunction with the present invention, the term "corresponding" in conjunction with "wild type plant cell" or "wild type plant" means that the plant cells or plants, which are compared with one another, have been raised under the same cultivation conditions and that they have the same cultivation age.

30

Here, within the framework of the present invention, the term "activity" means the expression of a transgene coding sequence and/or the presence of the protein encoded by a transgene coding sequence and/or the presence of the product produced by the the protein encoded by the transgene in the genetically modified plant cells or genetically modified plants, respectively.

The expression of a coding sequence of a transgene can, for example, be determined by measuring the quantity of transcripts of the transgene, e.g. using Northern blot analysis or RT-PCR.

The presence of a protein encoded by a transgene, which results in an activity of the respective protein in the genetically modified plant cells or genetically modified plants concerned, can, for example, be determined by immunological methods such as Western blot analysis, ELISA (Enzyme Linked Immuno Sorbent Assay) or RIA (Radio Immune Assay). In case the transgene encodes a dextranucrase DSR-S protein the presence of the DSR-S protein in genetically modified plant cells or genetically modified plants can be demonstrated, for example, with the help of native acrylamide gel electrophoresis. In doing so, plant cell or plant extracts containing proteins are first separated electrophoretically and, after incubation of the acrylamide gels in respective buffers containing sucrose, the acrylamide gels will be stained with the periodic acid-Shiff stain (Miller and Robyt, 1986, Analytical Biochemistry 156, 357-363; WO 00 47727).

The presence of the product dextran produced in plant cells according to the invention or plants according to the invention having been transformed with a nucleic acid sequence encoding a dextranucrase DSR-S protein can be demonstrated e.g. by immunological analysis according to the method described in example 3 of the present specification.

In conjunction with the present invention, the term "dextransucrase DSR-S protein" is to be understood as an enzyme catalysing the synthesis of dextran from sucrose. The dextran synthesized is composed of alpha-1,6-linked glucose units in the main chain and reveals alpha 1,3-linked side chains. Preferrably the dextransucrase protein of the invention synthesizes a dextran having around 5% alpha-1,3-linkages.

The term "dextransucrase DSR-S protein" is further defined as an enzyme having an identity of at least 70%, preferably at least 80%, more preferably at least 90%, and still more preferably at least 95% with the amino acid sequence identified under SEQ ID N0: 2 or SEQ ID N0: 4, and having the same catalytic properties than the enzyme encoded by the Dsr-S gene (accession AJ271107) from *L. mesenteroides*.

In conjunction with the present invention, the term "transgene" is understood to mean such a molecule that either does not occur naturally in the corresponding non-genetically modified wild type plant cells or non-genetically modified wildtype plants, or that does not occur naturally in the concrete spatial arrangement in non-genetically modified wild type plant cells or non-genetically modified wildtype plants, or that is localised at a place in the genome of the non-genetically modified wild type plant cell or non-genetically modified wildtype plant at which it does not occur naturally.

In conjunction with the invention the term "recombinant" means a nucleic acid molecule which consists of different elements, the combination or specific spatial arrangement of which does not occur naturally in plant cells or plants.

A large number of techniques are available for the introduction of DNA into a plant host cell. These techniques include the transformation of plant cells with T-DNA using *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes* as the transformation medium, the fusion of protoplasts,
5 injection, the electroporation of DNA, the introduction of DNA by means of the biolistic approach as well as other possibilities.

The use of agrobacteria-mediated transformation of plant cells has been intensively investigated and adequately described in EP 120516; Hoekema, IN: The Binary Plant Vector System Offsetdrukkerij Kanters
10 B.V., Alblasterdam (1985), Chapter V; Fraley et al., Crit. Rev. Plant Sci. 4, 1-46 and by An et al. EMBO J. 4, (1985), 277-287. For the transformation of potato, see Rocha-Sosa et al., EMBO J. 8, (1989), 29-33, for example.

The transformation of monocotyledonous plants by means of vectors
15 based on agrobacterium transformation has also been described (Chan et al., Plant Mol. Biol. 22, (1993), 491-506; Hiei et al., Plant J. 6, (1994) 271-282; Deng et al, Science in China 33, (1990), 28-34; Wilmink et al., Plant Cell Reports 11, (1992), 76-80; May et al., Bio/Technology 13, (1995), 486-492; Conner and Domisse, Int. J. Plant Sci. 153 (1992), 550-555;
20 Ritchie et al, Transgenic Res. 2, (1993), 252-265). An alternative system to the transformation of monocotyledonous plants is transformation by means of the biolistic approach (Wan and Lemaux, Plant Physiol. 104, (1994), 37-48; Vasil et al., Bio/Technology 11 (1993), 1553-1558; Ritala et al., Plant Mol. Biol. 24, (1994), 317-325; Spencer et al., Theor. Appl.
25 Genet. 79, (1990), 625-631), protoplast transformation, electroporation of partially permeabilised cells and the introduction of DNA by means of glass fibres. In particular, the transformation of maize has been described in the literature many times (cf. e.g. WO95/06128, EP0513849, EP0465875, EP0292435; Fromm et al., Biotechnology 8, (1990), 833-844;
30 Gordon-Kamm et al., Plant Cell 2, (1990), 603-618; Koziel et al.,

Biotechnology 11 (1993), 194-200; Moroc et al., Theor. Appl. Genet. 80, (1990), 721-726).

The successful transformation of other types of cereal has also already been described, for example for barley (Wan and Lemaux, see above; 5 Ritala et al., see above; Krens et al., Nature 296, (1982), 72-74) and for wheat (Nehra et al., Plant J. 5, (1994), 285-297). All the above methods are suitable within the framework of the present invention.

In conjunction with the present invention, the introduced nucleic acid may be integrated into the nuclear genome or into the plastidial genome of the 10 plant cell.

The classical way of transfecting plastids involves bombarding leaves with microprojectiles carrying DNA molecules (Svab et al., 1993). Today, stable plastid transfection is routinely performed in the tobacco 15 species *N. tabaccum* (Svab and Maliga, 1990; Svab et al., 1993). There has been recent progress in rice (Khan and Maliga, 1999), *Arabidopsis thaliana* (Sikdar et al., 1998), potato (Sidorov et al, 1999), colza (WO 00/39313), tomato (Ruf et al., 2001) and soybean (WO 04/053133). Examples of methods for obtaining transplastomic plants have been 20 described in Patent Application WO 04/055191.

Amongst other things, the plant cells according to the invention and the plants according to the invention can be differentiated from wild type plant cells and wild type plants respectively in that they contain at least one 25 copy of the foreign nucleic acid molecule stably integrated within their genome, wherein the foreign nucleic acid molecule encodes a dextranucrase DSR-S protein.

Furthermore, the plant cells according to the invention and the plants 30 according to the invention can preferably be differentiated from wild type

plant cells or wild type plants respectively by the following characteristic:
the plant cells according to the invention or plants according to the
invention have transcripts of the introduced nucleic acid molecules. These
can be verified, for example, by Northern blot analysis or by RT-PCR
5 (Reverse Transcription Polymerase Chain Reaction). Preferably, the plant
cells according to the invention and the plants according to the invention
contain a protein, which is coded by an introduced nucleic acid molecule.
This can be demonstrated by immunological methods, for example, in
particular by a Western Blot Analysis.

10

Furthermore the plant cells according to the invention and the plants
according to the invention can more preferably be differentiated from wild
type plant cells or wild type plants, respectively, by the characteristics that
they synthesize dextran. Preferrably the plant cells of the invention or the
15 plants of the invention produce dextran in their plastids.

The terms "starch which is modified in comparison to starch synthesized
by wild-type plant cells or "modified starch" mean a starch which, when
compared to starch synthesized in wild-type plants, differs for example in
20 its physico-chemical properties, the pastification behavior, the size and/or
the shape of the starch granule.

Compared with wild-type starch, such starch may be modified in particular
with respect to its viscosity and/or the gel formation properties of the glues
25 of this starch and/or its capability to be digested.

The modification in respect to the viscosity can be measured by several
means, and in particular by means of a Thermo Haake rheoscope
(Thermo Electron Cooperation) according to the manufacturer's
30 instructions or by means of a Rapid Visco Analyser (RVA), as for example

the Rapid Visco Analyser Super3 (Newport Scientific Pty Ltd, Investmet Support Group, Warriewod NSW 2102,Australia). The viscosity values are indicated in Centipoise (cP) in accordance with the manufacturer's operating manuals, which are incorporated into the description herewith by
5 reference.

A preferred way to determine the viscosity characteristics by means of a Rapid Visco Analyser (RVA) and the parameters which are used for the comparison of different samples are described in the general methods
10 (method 1) of the present invention.

An other preferred way to determine the viscometric profiles by means of a thermo Haake rheoscope is described in the general methods (method 2) of the present invention.

15 The determination of the gel formation properties of the glues of the starch (or gel strength) can be determined by means of a Texture Analyser, as for example the Texture Analyser TA-XT2 (Stable Micro Systems – Surrey, UK) in accordance with the manufacturer's operating manual, which is
20 incorporated into the description herewith by reference.

A preferred way to determine the gel formation properties of the glues of the starch by means of the Texture Analyser TA-XT2 is described in the general methods (method 3) of the present invention.

25 The capability to be digested can be determined by the determination of the percentage of digested starch, using the methodology of Englyst H.N. *et al.*, European Journal of Clinical Nutrition 4, Suppl.2, S33-S50, which is incorporated into the description herewith by reference, based on the
30 determination of resistant starches RS Type III, which is the indigestible

retrograded starch that is obtained, for example, by thermal and/or enzymatic treatment and then retrograded.

5 The method of Englyst can be modified in correspondence with the information on the determination of RS content in WO 00/02926, incorporated into the description herewith by reference. The resulting method is described in the general methods (method 4) of the present invention.

10 Further, the present invention relates to genetically modified plant cells or genetically modified plants of the invention characterized in that said plant cells or said plants, respectively, synthesize a modified starch which has a decreased end viscosity and/or an increased digestibility, in comparison to starch synthesized by wild-type plant cells.

15 In conjunction with the invention, the end viscosity can be measured by means of a rheoscope, particularly a Thermo Haake rheoscope or a Rapid Visco Analyser. Preferred methods for determination of the end viscosity are described in general methods (methods 1 and 2) of the present invention.

20

Preferably, the decrease of the end viscosity measured by means of a rheoscope, particularly a Thermo Haake rheoscope, is at least of 15 %, preferred at least of 20%, more preferred at least of 25%, most preferred at least of 30%.

25

In conjunction with the invention, the digestibility can be measured according to the method of Englyst, or by the method of Englyst modified according to WO 00/02926. A preferred method for the determination of the digestibility is described in general methods (method 4) of the present
30 invention.

Preferably, the increase in digestibility measured according to the method of Englyst or to the method of Englyst modified in correspondence with the information on the determination of RS content in WO 00/02926, is at least of 12%, preferred at least of 13%; more preferred at least of 14%, most preferred at least of 15%.

Furthermore, the invention relates to genetically modified plant cells according to the invention or genetically modified plants according to the invention, having integrated into its genome a transgene comprising linked to one another in a functional fashion in the direction of the transcription :

- a promoter sequence which initiates transcription in plant cells,
- a heterologous nucleic acid sequence encoding a dextranucrase DSR-S protein, and
- optionally a termination sequence which is active in plant cells.

15

In conjunction with the present invention, the term "genome" is to be understood to mean the totality of the genetic material present in a plant cell. It is known to the person skilled in the art that, as well as the cell nucleus, other compartments (e.g. plastids, mitochondrions) also contain genetic material.

20

In a preferred embodiment, the nucleic acid construct is integrated into the nuclear genome of the plant cell. Transport of the dextranucrase protein into a particular cellular compartment, such as plastid, may therefore be accomplished by the use of a transit peptide to target the cellular compartment of interest. The nucleic acid sequence encoding the transit peptide is inserted in front of the coding sequence. Sequences encoding a transit peptide may be derived from any nucleic acid sequence encoding a plant protein which is expressed in the cytoplasm and translocated to the cellular compartment of interest. The transit peptide can be identified by

30

comparing the messenger RNA encoding the particular polypeptide with the amino acid sequence of the mature protein. The amino acid sequences absent from the mature protein and coded for by the corresponding messenger RNA beginning at the initiation codon, usually a methionine, will normally be the transit peptide, or will normally contain the transit peptide. The skilled person will be able to determine sequences encoding transit peptides using a program for prediction of transit peptide, as for example Chloro 1.1 Server (Emanuelsson O. *et al*, 1999, Protein Science:8:978-984)

10

The transit peptide is the amino acid sequence capable of directing a protein joined to the transit peptide to a cellular compartment of interest and may be the whole naturally occurring (wild-type) transit peptide, a functional fragment thereof, a functional mutant thereof, or a chimeric transit peptide wherein at least two transit peptides are associated to each other or of parts of different transit peptides associated to each other in a functional manner. Such a chimeric transit peptide is reported as an optimised transit peptide in EP0508909 and EP0924299.

The nucleic acid encoding a transit peptide may be heterologous in respect to the nucleic acid sequence encoding the enzyme fused to it, meaning that the nucleic acid sequence encoding the transit peptide and the nucleic acid sequence encoding the enzyme to be directed to the plastids originate from different genes which again can originate from different species.

A transit peptide dedicated to target the enzyme translationally joined to it to a plastid, such as chloroplast or amyloplast, is called a plastidial transit peptide.

The present invention further relates to genetically modified plant cells of the invention or genetically modified plants of the invention having

integrated into its genome a nucleic acid construct comprising linked to one another in a functional fashion in the direction of the transcription :

- a promoter sequence which initiates transcription in plant cells,
- a heterologous nucleic acid sequence encoding a plastidial transit peptide translationally fused with
- a heterologous nucleic acid sequence encoding a dextranucrase DSR-S protein, and
- optionally a termination sequence which is active in plant cells.

The term "linked to one another in a functional fashion" means that the elements of the nucleic acid construct are linked to one another in such a way which permits the expression of the coding region.

In conjunction with the invention the term "translationally fused" shall mean a fusion of nucleic acid sequences in such a way that they represent a single open reading frame, which upon transcription leads to the production of a single messenger RNA encoding a single protein, when translated.

Plastidial transit peptides may be selected from the group comprising the transit peptide of a gene encoding a waxy protein (Klös gen et al, Mol Gen Genet. 217 (1989), 155-161), the ribulose biphosphate carboxylase small subunit (Wolter et al, Proc. Natl. Acad. Sci. USA 85 (1988), 846-850; Nawrath et al., Proc. Natl. Acad. Sci. 10 USA 91 (1994), 12760-12764), NADP-malate dehydrogenase (Gallardo et al., Planta 197 (1995), 324-332), Gluthation-reductase (Creissen et al., Plant J. 8 (1995), 167-175), EPSPS (US 5,188,642), and an optimised transit peptide described in EP0508909 and EP0924299. These examples are not limiting.

In a preferred embodiment, a nucleic acid encoding a plastidial transit peptide of the ferredoxin reductase gene is translationally fused with the nucleic acid sequence encoding a dextranucrase DSR-S protein (Smeekens et al, 1985; Pilon et al, 1995).

5

In a further preferred embodiment, the plastidial transit sequence of the ferredoxin reductase gene originates from spinach.

In an other preferred embodiment, a nucleic acid sequence encoding the optimised plastidial transit peptide described in EP0508909 and EP0924299 is translationally fused with the nucleic acid sequence encoding a dextranucrase DSR-S protein.

The technologies used for the construction of the nucleic acid construct of the invention are well known to the skilled person. As non-limiting examples, it is possible to mention the technologies described in Sambrook et al. (1989, *Molecular Cloning : A Laboratory Manual*, Nolan C. ed., New York: Cold Spring Harbor Laboratory Press).

Furthermore, plant and/or progeny thereof, which contain a plant cell according to the invention, are also the subject matter of the invention. Plants of this type can be produced from the plant cell according to the invention by regeneration, using methods known to the person skilled in the art, as for example methods described in "plant Cell Culture Protocols" 1999, edited by R.D. Hall, Humana Press, ISBN 0-89603-549-2.

In principle, the plants according to the invention can be plants of any plant species, i.e. both monocotyledonous and dicotyledonous plants. Preferably they are useful plants, i.e. plants, which are cultivated by

people for the purposes of food or for technical, in particular industrial purposes.

In a further preferred embodiment, the plant according to the invention is a starch-storing plant. The term "starch-storing plants" includes all plants
5 with starch-storing plant parts such as, for example, maize, rice, wheat, rye, oat, barley, cassava, potato, sago, mung bean, pea or sorghum. Preferred starch-storing plant parts are, for example, tubers, storage roots and grains containing an endosperm; tubers are particularly preferred;
10 tubers of potato plants are especially preferred.

In a further preferred embodiment, the present invention relates to a starch-storing plant according to the invention which is a potato plant.

15 In conjunction with the present invention, the term "potato plant" or "potato" means plant species of the genus *Solanum*, in particular tuber-producing species of the genus *Solanum* and especially *Solanum tuberosum*.

20 The present invention also relates to propagation material of plants according to the invention containing a plant cell according to the invention.

Here, the term "propagation material" includes those constituents of the
25 plant that are suitable for producing offspring by vegetative or sexual means. Cuttings, callus cultures, rhizomes or tubers, for example, are suitable for vegetative propagation. Other propagation material includes, for example, fruits, seeds, seedlings, protoplasts, cell cultures, etc. Preferably, the propagation material is seeds and particularly preferably
30 tubers.

In a further embodiment, the present invention relates to harvestable plant parts of plants according to the invention such as fruits, storage roots, blooms, buds, shoots or stems, preferably seeds or tubers, wherein these
5 harvestable parts contain plant cells according to the invention.

The present invention also relates to a method for the manufacture of genetically modified plants according to the invention wherein
a) a plant cell is transformed with a nucleic acid molecule comprising a
10 nucleic acid molecule encoding a dextransucrase DSR-S protein,
b) a plant is regenerated from a plant cell obtained in step a) and
c) if necessary, further plants are produced from the plants obtained in step b).

15 The plant cell obtained in step a) may be regenerated to whole plants according to methods known to the skilled person, as for example using the methods described in "plant Cell Culture Protocols" 1999, edited by R.D. Hall, Humana Press, ISBN 0-89603-549-2.

20 In a preferred method for the manufacture of genetically modified plant of the invention the nucleic acid molecule encoding the dextransucrase DSR-S protein in step a) is translationally fused with a nucleic acid molecule encoding a plastidic signal sequence.

25 The production of further plants according to Step (c) of the method according to the invention can be carried out, for example, by vegetative propagation (for example using cuttings, tubers or by means of callus culture and regeneration of whole plants) or by sexual propagation. Here, sexual propagation preferably takes place under controlled conditions, i.e.

selected plants with particular characteristics are crossed and propagated with one another.

The present invention also relates to a method for the manufacture of a
5 genetically modified plant according to the method disclosed above, wherein the nucleic acid molecule encoding a dextransucrase DSR-S protein is integrated into the plastidial genome of the plant.

The nucleic acid molecule encoding a dextransucrase DSR-S protein may
10 be from any desired origin, preferably the nucleic acid molecule encoding a dextransucrase originates from bacteria expressing such proteins.

More preferably, nucleic acid molecules used in the invention may encode a dextransucrase DSR-S protein from a bacteria selected from the group
15 consisting of *Leuconostoc*, *Lactobacillus* and *Streptococcus* bacteria.

Most preferably, nucleic acid molecules used in the invention may encode a dextransucrase DSR-S protein from *Leuconostoc mesenteroides*, especially preferably from *Leuconostoc mesenteroides* strain NRRL
20 B512F.

Nucleic acid molecules encoding a dextransucrase DSR-S protein used in the invention may be isolated e.g. from genomic DNA or DNA libraries produced from any origin, preferably from bacteria. Alternatively, they may
25 have been produced by means of recombinant DNA techniques (e.g. PCR) or by means of chemical synthesis. The identification and isolation of such nucleic acid molecules may take place by using the molecules according to the invention or parts of these molecules or, as the case may be, the reverse complement strands of these molecules, e.g. by
30 hybridization according to standard methods (see e.g. Sambrook et al.,

1989, Molecular Cloning, A Laboratory Manual, 2nd Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.).

As a probe for hybridization e.g. nucleic acid molecules may be used
5 which exactly or basically contain the nucleotide sequences indicated
under SEQ ID No. 1 or parts thereof. The fragments used as hybridization
probe may also be synthetic fragments which were produced by means of
the conventional synthesizing methods and the sequence of which is
basically identical with that of a nucleic acid molecule according to the
10 invention.

The molecules hybridizing to the nucleic acid molecules used in the
invention also comprise fragments, derivatives and allelic variants of the
above-described nucleic acid molecules which encode a dextranucrase
15 DSR-S protein. In this context, fragments are defined as parts of the
nucleic acid molecules, which are long enough in order to encode
proteins. In this context, the term derivatives means that the sequences of
these molecules differ from the sequences of the above-mentioned nucleic
acid molecules at one or more positions and that they exhibit a high
20 degree of homology to these sequences. Homology means a sequence
identity of at least 70% and still more preferably a sequence identity of
more than 90% and most preferably a sequence identity of more than
95%. The deviations occurring when comparing with the above-described
nucleic acid molecules might have been caused by deletion, substitution,
25 insertion or recombination.

Moreover, homology means that functional and/or structural equivalence
exists between the respective nucleic acid molecules or the proteins they
encode. The nucleic acid molecules, which are homologous to the above-

described molecules and represent derivatives of these molecules, are generally variations of these molecules, that constitute modifications which exert the same biological function: These variations may be naturally occurring variations, for example sequences derived from other bacteria, or mutations, whereby these mutations may have occurred naturally or they may have been introduced by means of a specific mutagenesis. Moreover the variations may be synthetically produced sequences. The allelic variants may be naturally occurring as well as synthetically produced variants or variants produced by recombinant DNA techniques.

10

In a preferred embodiment of the present invention the nucleic acid molecules encoding a dextransucrase DSR-S protein is chosen from the group consisting of:

- 15 a) Nucleic acid molecules, which encode a protein with the amino acid sequence given under Seq ID NO: 2 or SEQ ID N04;
- b) Nucleic acid molecules, which encode a protein, the amino acid sequence of which has an identity of at least 70% with the amino acid sequence given under SEQ ID NO: 2 or SEQ ID N04;
- 20 c) Nucleic acid molecules, comprising the nucleotide sequence shown under Seq ID NO:3 or a complementary sequence thereof;
- d) Nucleic acid molecules, the nucleic acid sequence of which has an identity of at least 70% with the nucleic acid sequences described under a) or c);
- 25 e) Nucleic acid molecules, the nucleotide sequence of which deviates from the sequence of the nucleic acid molecules identified under a), b), c) or d) due to the degeneration of the genetic code; and

f) Nucleic acid molecules, which represent fragments, allelic variants and/or derivatives of the nucleic acid molecules identified under a), b), c), d) or e).

5 In a further preferred embodiment of the invention, the nucleic acid molecules encoding a dextransucrase protein DSR-S encode a protein, the amino acid sequence of which has an identity of at least 70%, preferably at least 80%, more preferably at least 90%, and still more preferably at least 95% to the sequence Seq ID N0: 2 or SEQ IDN0:4.

10

In an other further preferred embodiment, the nucleic acid molecule encoding a dextransucrase protein DSR-S has a nucleic acid sequence with an identity of at least 70%, preferably at least 80%, more preferably at least 90%, and still more preferably at least 95% to the sequence Seq ID
15 N0:1 or SEQ ID N0: 3.

In conjunction with the present invention, the term "identity" is to be understood to mean the number of amino acids/nucleotides corresponding with the amino acids/nucleotides of other protein/nucleic acid, expressed
20 as a percentage. Identity is preferably determined by comparing the Seq. ID NO: 1, SEQ ID NO: 2, Seq. ID NO: 3 or SEQ ID NO: 4 with other protein/nucleic acid with the help of computer programs. If sequences that are compared with one another have different lengths, the identity is to be determined in such a way that the number of amino acids, which have the
25 shorter sequence in common with the longer sequence, determines the percentage quotient of the identity. Preferably, identity is determined by means of the computer program ClustalW, which is well known and available to the public (Thompson et al., Nucleic Acids Research 22 (1994), 4673-4680). ClustalW is made publicly available by Julie
30 Thompson (Thompson@EMBL-Heidelberg.DE) and Toby Gibson

(Gibson@EMBL-Heidelberg.DE), European Molecular Biology Laboratory, Meyerhofstrasse 1, D 69117 Heidelberg, Germany. ClustalW can also be downloaded from different Internet sites, including the IGBMC (Institut de Génétique et de Biologie Moléculaire et Cellulaire, B.P.163, 67404 Illkirch Cedex, France; ftp://ftp-igbmc.u-strasbg.fr/pub/) and the EBI (ftp://ftp.ebi.ac.uk/pub/software/) as well as from all mirrored Internet sites of the EBI (European Bioinformatics Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge CB10 1SD, UK).

Preferably, Version 1.8 of the ClustalW computer program is used to determine the identity between proteins according to the invention and other proteins. In doing so, the following parameters must be set: KTUPLE=1, TOPDIAG=5, WINDOW=5, PAIRGAP=3, GAOPEN=10, GAPEXTEND=0.05, GAPDIST=8, MAXDIV=40, MATRIX=GONNET, ENDGAPS(OFF), NOPGAP, NOHGAP.

Preferably, Version 1.8 of the ClustalW computer program is used to determine the identity between the nucleotide sequence of the nucleic acid molecules according to the invention, for example, and the nucleotide sequence of other nucleic acid molecules. In doing so, the following parameters must be set:

KTUPLE=2, TOPDIAGS=4, PAIRGAP=5, DNAMATRIX:IUB, GAOPEN=10, GAPEXT=5, MAXDIV=40, TRANSITIONS: unweighted.

Furthermore, identity means that functional and/or structural equivalence exists between the nucleic acid molecules concerned or the proteins coded by them. The nucleic acid molecules, which are homologous to the molecules described above and constitute derivatives of these molecules, are generally variations of these molecules, which constitute modifications, which execute the same biological function. At the same time, the variations can occur naturally, for example they can be sequences from other bacterial species, or they can be mutations, wherein these mutations

may have occurred in a natural manner or have been introduced by objective mutagenesis. The variations can also be synthetically manufactured sequences. The allelic variants can be both naturally occurring variants and also synthetically manufactured variants or variants
5 produced by recombinant DNA techniques. Nucleic acid molecules, which deviate from nucleic acid molecules according to the invention due to degeneration of the genetic code, constitute a special form of derivatives.

The use of nucleic acid molecules that encode a dextransucrase DSR-S
10 protein and the sequence of which differs from the nucleotide sequences of the above-mentioned molecules due to the degeneracy of the genetic code are also the subject-matter of the invention.

The invention also relates to the use of nucleic acid molecules showing a
15 sequence which is complementary to the whole or to a part of one of the above-mentioned sequences.

For expressing nucleic acid molecules described above, these are preferably linked with regulatory DNA sequences, which guarantee
20 initiation of transcription in plant cells. In particular, these include promoters. In general, any promoter that is active in plant cells is eligible for expression.

At the same time, the promoter can be chosen so that expression takes place constitutively or only in a certain tissue, at a certain stage of the
25 plant development or at a time determined by external influences. The promoter can be homologous or heterologous both with respect to the plant and with respect to the nucleic acid molecule.

Suitable promoters are, for example, the promoter of the 35S RNA of the cauliflower mosaic virus and the ubiquitin promoter from maize for
30 constitutive expression, the patatin promoter B33 (Rocha-Sosa et al.,

EMBO J. 8 (1989), 23-29) for tuber-specific expression in potatoes or a promoter, which only ensures expression in photosynthetically active tissues, e.g. the ST-LS1 promoter (Stockhaus et al., Proc. Natl. Acad. Sci. USA 84 (1987), 7943-7947; Stockhaus et al., EMBO J. 8 (1989), 2445-2451) or, for endosperm-specific expression of the HMG promoter from wheat, the USP promoter, the phaseolin promoter, promoters of zein genes from maize (Pedersen et al., Cell 29 (1982), 1015-1026; Quatroccio et al., Plant Mol. Biol. 15 (1990), 81-93), glutelin promoter (Leisy et al., Plant Mol. Biol. 14 (1990), 41-50; Zheng et al., Plant J. 4 (1993), 357-366; Yoshihara et al., FEBS Lett. 383 (1996), 213-218) or shrunken-1 promoter (Werr et al., EMBO J. 4 (1985), 1373-1380). However, promoters can also be used, which are only activated at a time determined by external influences (see for example WO 9307279). Promoters of heat-shock proteins, which allow simple induction, can be of particular interest here. Furthermore, seed-specific promoters can be used, such as the USP promoter from *Vicia faba*, which guarantees seed-specific expression in *Vicia faba* and other plants (Fiedler et al., Plant Mol. Biol. 22 (1993), 669-679; Bäumlein et al., Mol. Gen. Genet. 225 (1991), 459-467).

Promoters which are active in plastids of plant cells may be used if the nucleic acid construct of the invention is integrated in the plastidial genome of the plant cell. Among the promoters active in plastids of plant cells, by way of example, special mention can be made of the *psbA* gene which encodes the D1 polypeptide of PSII (Staub et al. 1993 EMBO Journal 12(2):601-606), and the constitutive *Prn* promoter which regulates the ribosomal RNA operon (Staub et al. 1992 Plant Cell 4:39-45).

Furthermore, a termination sequence (polyadenylation signal) can be present, which is used for adding a poly-A tail to the transcript. A function in the stabilisation of the transcripts is ascribed to the poly-A tail. Elements

of this type are described in the literature (cf. Gielen et al., EMBO J. 8 (1989), 23-29) and can be exchanged at will.

Plants obtainable by the method of the invention for the manufacture of a
5 plant according to the invention are a further embodiment of the invention.

Furthermore, the invention relates to vectors, especially plasmids, cosmids, viruses, bacteriophages and other vectors common in genetic engineering, which contain the above-mentioned nucleic acid molecules
10 encoding a dextranucrase DSR-S protein. Such vectors are preferably vectors which can be used for the transformation of plant cells. More preferably, they allow for the integration of the nucleic acid molecules of the invention into the nuclear or plastidial genome of the plant cell, if necessary in combination with flanking regulatory regions. Examples are
15 binary vectors which may be used in the Agrobacterium-mediated gene transfer, as for example pBIN20 binary vector (Hennegan and Danna, 1998). Examples of vectors which may be used for direct plastid transformation are given in WO 04/055191.

20 The plasmid comprising the heterologous nucleic acid to be introduced into the plant further will generally contain either a selectable marker or a reporter gene or both to facilitate identification and selection of transformed cells. Alternatively, the selectable marker may be carried on a separate vector and used in a co-transformation procedure. Both
25 selectable markers and reporter genes may be flanked with appropriate regulatory sequences to enable expression in plants. Useful selectable markers and reporter genes are well known in the art and include, for example, antibiotic and herbicide resistance genes, genes encoding beta-glucuronidase enzyme (Staub et al, 1993) or green fluorescent protein
30 (Sidorov et al, 1999) .

Specific examples of such genes are disclosed in Weising et al, 1988, Svab et al, 1993, White et al., Nucleic Acid Res. 18(4) :1062.

By using the nucleic acid molecule encoding a dextranucrase DSR-S
5 protein, it is now possible--by means of recombinant DNA techniques--to
interfere with the starch metabolism of plant cells or plants in a way so far
impossible. Thereby, the starch metabolism may be modified in such a
way that a modified starch is synthesized which e.g. is modified, compared
to the starch synthesized in corresponding non-genetically modified wild
10 ype plant cells or non-genetically modified wild type plants, respectively,
in its physico-chemical properties, the pastification behavior, the size
and/or the shape of the starch granule. Compared with wild-type starch,
such starch may be modified in particular with respect to its viscosity
and/or the gel formation properties of the glues of this starch and/or its
15 capability to be digested.

The present invention therefore also relates to modified starches
obtainable from plant cells according to the invention or plants according
to the invention, from propagation material according to the invention or
20 from harvestable plant parts according to the invention.

In a particularly preferred embodiment, the present invention relates to
modified potato starch.

The present invention further relates to a method for the manufacture of a
25 modified starch comprising the step of extracting the starch from a plant
cell according to the invention, from a plant according to the invention,
from harvestable parts of a plant according to the invention, or from a plant
obtainable by means of a method of the invention for the manufacture of a
plant according to the invention.

Preferably, such a method also comprises the step of harvesting the cultivated plants and/or starch-storing parts of such plants before extracting the starch. Most preferably, it further comprises the step of cultivating the plants of the invention before harvesting. Methods for the extraction of starch from plants or from starch-storing parts of plants are known to the skilled person. Methods for the extraction of starch from maize seeds have been described e.g. in Eckhoff et al. (Cereal Chem. 73 (1996) 54-57). The extraction of starch on an industrial level is usually achieved by the so-called wet-milling technique. Furthermore, methods for the extraction of starch from various other starch-storing plants have been described, e.g. in "Starch: Chemistry and Technology (Editor: Whistler, BeMiller and Paschall (1994), 2.sup.nd edition, Academic Press Inc. London Ltd; ISBN 0-12-746270-8; see e.g. chapter XII, page 412-468: maize and sorghum starches: production; by Watson; chapter XIII, page 469-479: tapioca, arrowroot and sago starches: production; by Corbishley and Miller; chapter XIV, page 479-490: potato starch: production and use; by Mitch; chapter XV, page 491 to 506: wheat starch: production, modification and use; by Knight and Oson; and chapter XVI, page 507 to 528: rice starch: production and use; by Rohmer and Klem). Appliances generally used for extracting starch from plant material are separators, decanters, hydrocyclones, spray dryers and cyclon driers.

Due to the expression of a nucleic acid molecule encoding a dextranucrase protein, the transgenic plant cells and plants described in the invention synthesize a starch which compared to starch synthesized in corresponding non-genetically modified wildtype plant cells or -genetically modified wildtype plants, respectively, is modified for example in its physico-chemical properties, the pastification behavior, the size and/or the shape of the starch granule. Compared with wildtype-starch, such starch may be modified in particular with respect to its viscosity and/or the gel formation properties of the glues of this starch and/or its capability to be

digested.

Thus, also the modified starch obtainable from the method according to the invention is the subject-matter of the present invention.

5

In a preferred embodiment of the invention the starch of the invention is a native starch.

In conjunction with the present invention, the term "native starch" means
10 that the starch is isolated from plants according to the invention, harvestable plant parts according to the invention or propagation material of plants according to the invention by methods known to the person skilled in the art.

15 The person skilled in the art knows that the characteristics of starch can be changed by thermal, chemical, enzymatic or mechanical derivation, for example. Derived starches are particularly suitable for different applications in the foodstuffs and/or non-foodstuffs sector. The starches according to the invention are better suited as a starting substance for the
20 manufacture of derived starches than conventional starches.

The present invention therefore also relates to a method for the production of a derived starch, wherein modified starch according to the invention or obtainable by means of a method according to the invention is derived
25 retrospectively.

In conjunction with the present invention, the term "derived starch" is to be understood to mean a modified starch according to the invention, the characteristics of which have been changed after isolation from vegetable
30 cells with the help of chemical, enzymatic, thermal or mechanical methods.

In a preferred embodiment of the present invention, the derived starch according to the invention is starch that has been heat-treated and/or acid-treated.

In a further preferred embodiment, the derived starches are starch ethers, in particular starch alkyl ethers, O-allyl ethers, hydroxylalkyl ethers, O-carboxymethyl ethers, nitrogen-containing starch ethers, phosphate-containing starch ethers or sulphur-containing starch ethers.

In a further preferred embodiment, the derived starches are cross-linked starches.

10 In a further preferred embodiment, the derived starches are starch graft polymers.

In a further preferred embodiment, the derived starches are oxidised starches.

In a further preferred embodiment, the derived starches are starch esters, 15 in particular starch esters, which have been introduced into the starch using organic acids. Particularly preferably these are phosphate, nitrate, sulphate, xanthate, acetate or citrate starches.

Methods for manufacturing derived starches according to the invention are 20 known to the person skilled in the art and are adequately described in the general literature. An overview on the manufacture of derived starches can be found, for example, in Orthoefer (in Corn, Chemistry and Technology, 1987, eds. Watson und Ramstad, Chapter 16, 479-499).

25 Derived starch obtainable by the method according to the invention for manufacturing a derived starch is also the subject matter of the present invention.

A further embodiment of the invention is the use of modified starch 30 according to the invention for the production of a derived starch.

The invention also relates to the use of a plant cell according to the invention, a plant according to the invention, harvestable parts of a plant according to the invention or a plant obtainable by means of a method of the invention, for the production of a modified starch.

The invention also relates to the use of a nucleic acid molecule encoding a dextranucrase DSR-S protein for the manufacture of a genetically modified plant cell according to the invention, a genetically modified plant according to the invention, propagation material according to the invention, or harvestable parts of plants according to the invention.

Furthermore the use of a nucleic acid sequence encoding a dextranucrase DSR-S protein for the production of a modified starch according to the invention is an embodiment of the invention.

General methods :

Method 1 : Determination of the viscosity characteristics by means of a Rapid Visco Analyser (RVA).

3 g of sample (for example flour) are taken up in 25 ml of H₂O (VE-type water, conductivity of at least 15 mega ohm) and used for the analysis in a Rapid Visco Analyser Super3 (Newport Scientific Pty Ltd., Investmet Support Group, Warriewood NSW 2102, Australia). The apparatus is operated following the manufacturer's instructions. The viscosity values are indicated in Centipoise (cP) in accordance with the manufacturer's operating manual, which is incorporated into the description herewith by reference. To determine the viscosity of the aqueous starch solution, the starch suspension is first stirred for 10 seconds at 960 rpm and subsequently heated at 50°C at a stirring speed of 160 rpm, initially for a minute (step 1). The temperature was then raised from 50°C to 95°C at a

heating rate of 12°C per minute (step 2). The temperature is held for 2.5 minutes at 95°C (step 3) and then cooled from 95°C to 50°C at 12°C per minute (step 4). In the last step (step 5), the temperature of 50°C is held for 2 minutes. The viscosity is determined during the entire duration.

- 5 After the program has ended, the stirrer is removed and the beaker covered. The gelatinized starch is now available for the texture analysis after 24 hours incubation at room temperature.

The profile of the RVA analysis contains parameters which are shown for
10 the comparison of different measurements and substances. In the context of the present invention, the following terms are to be understood as follows:

1. Maximum viscosity (RVA Max)

The maximum viscosity is understood as meaning the highest viscosity
15 value, measured in cP, obtained in step 2 or 3 of the temperature profile.

2. Minimum viscosity (RVA Min)

The minimum viscosity is understood as meaning the lowest viscosity value, measured in cP, observed in the temperature profile after the maximum viscosity. Normally, this takes place in step 3 of the temperature
20 profile.

3. Final viscosity (RVA Fin)

The final viscosity (or end viscosity) is understood as meaning the viscosity value, measured in cP, observed at the end of the measurement.

4. Setback (RVA Set)

25 What is known as the "setback" is calculated by subtracting the value of the final viscosity from that of the minimum occurring after the maximum viscosity in the curve.

5. Gelatinization temperature (RVA PT)

The gelatinization temperature is understood as meaning the point in time of the temperature profile where, for the first time, the viscosity increases drastically for a brief period.

5

Method 2 : Determination of the viscometric profiles by means of a Thermo Haake rheoscope.

Viscometric profiles from a 2 % starch suspension were determined by applying a small oscillating shear deformation at a frequency of 1 Hz, using a Thermo Haake rheoscope. The rheometer was equipped with parallel plate geometry (typ C70/1 Ti) and the gap size was 0.1 mm. The pasting profile of the 2 % starch-water (w/v) suspension was obtained by heating the suspension from 40 °C to 90 °C at a rate of 2 °C/min, where it was kept for 15 min followed by cooling to 20 °C at a rate of 2 °C/min and hold again for 15 min at 20 °C. The Tg (start gelatinization temperature), Tp (peak temperature) and viscosities were measured. Subsequently, from the retrogradated sample, an amplitude sweep was run at 10 Pa increasing to 1.000 Pa within 60 s to check that the measurements were made in the linear region, in which the amplitudes of stress and strain are proportional to each other.

Method 3 : Determination of the gel formation properties of the glues of the starch by means of a Texture Analyser TA-XT2.

The sample is gelatinized in the RVA apparatus in an aqueous suspension by means of a Rapid Visco Analyser (RVA) and subsequently stored for 24 hours at room temperature in a sealed container. The samples are fixed under the probe (round piston with planar surface) of a Texture Analyser TA-XT2 from Stable Micro Systems (Surrey, UK) and the gel strength was determined using the following parameters:

- Test speed 0.5 mm/s
- Depth of penetration 7 mm

- Contact surface 113 mm²
- Pressure 2 g.

Method 4 : Determination of digestibility of starch based on the determination of resistant starches RS Type III.

Resistant starches, RS, can be divided into the following types:

- RS type 1 Starch not accessible physically to digestion, for example partly milled plant cells (e.g. in muesli).
- RS type 2 Indigestible granular starch (starch grains), for example from raw potatoes, green bananas, etc.
- RS type 3 Indigestible retrograded starch that is obtained, for example, by thermal and/or enzymatic treatment and then retrograded.
- RS type 4 Indigestible, chemically modified starch that is formed, for example, by cross-bonding or esterification (acetylation, etc).

The determination of resistant starches RS Type III was obtained using the following steps:

a) Pancreatine/amyloglucosidase (AGS) treatment

Pancreatine/amyloglucosidase digestion buffer used:

0.1 M Na acetate pH 5.2

4 mM CaCl₂

Preparation of the enzyme solution:

12g pancreatine (Merck, Product no. 1.07130.1000) were stirred in 80 ml demineralised water (conductivity ca. 18 M ohm) for 10 min at 37 °C and then centrifuged for 10 min at 3000 rpm.

54 ml of the supernatant obtained after centrifugation were treated with 9.86 ml demineralised water and 0.14 ml amyloglucosidase (6000 u/ml, Sigma, Product no. A-3042).

Pancreatine/amyloglucosidase (AGS) digestion procedure

- 5 assays of the pancreatine/amyloglucosidae (AGS) digestion are prepared each time for each batch starch to be measured. No enzyme solution is later added to 2 of each of these 5 assays. The assays to which no enzyme solution is added are designated as reference and are used for determination of the recovery rate. The remaining 3 assays are designated as sample, later treated with enzyme solution and used for the determination of the RS content.
- 10 A number of reaction vessels which contain no starch were processed in parallel (blank samples). These blank samples which contain no starch are used for the determination amount of co-precipitated material (protein, salts).
- 15 The tare weight of 50 ml reaction vessels (Falcon tubes) was determined and then in each case ca. 200 mg of the starch are weighed in. 15 ml Na acetate buffer was added to each of the linear water-insoluble poly-alpha-1,4-D-glucan samples and the blanks samples, and 20 ml Na acetate buffer to each of the references (see above). These samples were pre-warmed to 37 °C.
- The reaction was initiated by the addition of 5 ml enzyme solution to each of the individual reaction vessels of the samples and the blank samples which were then shaken for 2 hours at 37 °C (200 rpm).
- The reaction was quenched by the addition of 5 ml glacial acetic acid (equilibrated to pH 3.0) and 80 ml technical ethanol to the samples, blank samples and the references.
- Precipitation of the starch from the reaction mixture was achieved by incubation of the quenched reaction assay at room temperature for 1 hour.

After sedimentation (centrifugation for 10 min at 2500 x g) the sediment of the individual assays obtained was washed twice with 80% ethanol to remove short-chain glucans and then freeze dried after cooling to -70°C . The samples were re-weighed and the weight differences used for the calculation of the "gravimetric" RS content.

b) Determination of the RS content

The following procedure was used for the determination of RS content of the individual batches of water-insoluble starch:

10

a) Determination of the water content of the individual sample batches of starch (wt.H₂O)

b) Determination of the tare weight of the individual reaction vessels for the respective samples (wt.RGP), references (wt.RGR) and the blank samples (wt.RGB).

15

c) Weighing ca. 200 mg of water-insoluble starch into the individual reaction vessels for samples (wt.P) and references (wt.R)

d) Calculation of the dry fraction of the weights for samples ($\text{wt.Ptr} = \text{wt.P} - \text{wt.H}_2\text{O}$) and references ($\text{wt.Rtr} = \text{wt.R} - \text{wt.H}_2\text{O}$)

20

e) Enzymatic digestion of the respective samples and blank samples. References are treated in the same way but without addition of the enzyme solution.

f) Precipitation, sedimentation, washing and freeze drying of the substances remaining in the reaction vessels of the samples, references and blank samples after the treatment described in e).

25

g) Weighing of the substances remaining in the reaction vessels of the samples (wt.PR_G), references (wt.RR_G) and blank samples (wt.BR_G), inclusive of reaction vessel after the treatment described in f).

- h) Calculation of the weight of the substances remaining in the reaction vessels of the samples ($\text{wt.Pnv} = \text{wt.PRg} - \text{wt.RGP}$), references ($\text{wt.Rnv} = \text{wt.RRG} - \text{wt.RGR}$) and the blank samples ($\text{wt.Bnv} = \text{wt.BRG} - \text{wt.RGB}$) after the treatment described under f).
- i) Determination of the water content of the substances remaining in the reaction vessels of samples (wt.H2OPnv), references (wt.H2ORnv) and the blank samples (wt.H2OBnv) after the treatment described under f).
- j) Calculation of the dry fraction of the substances remaining in the reaction vessels of the samples ($\text{wt.Pnvtr} = \text{wt.Pnv} - \text{wt.H2OPnv}$) references ($\text{wt.Rnvtr} = \text{wt.Rnv} - \text{wt.H2ORnv}$) and the blank samples ($\text{wt.Bnvtr} = \text{wt.Bnv} - \text{H2OBnv}$) after the treatment described under f).
- k) Determination of the corrected weights for the samples ($\text{wt.Pnvkor} = \text{wt.Pnvtr} - \text{wt.Bnvtr}$) and references ($\text{wt.Rnvkor} = \text{wt.Rnvtr} - \text{wt.Bnvtr}$)
- l) Calculation of the percentage fraction of the corrected weights of the water-insoluble starch remaining after enzymatic digestion relative to the dry weight of the starting amount of the samples ($\text{RSaP} = \text{wt.Pnvkor} / \text{wt.Ptr} \times 100$) and calculation of the percentage fraction of the corrected weights of the remaining water-insoluble starch of the references relative to the dry weight of the starting amounts of the references ($\text{RSaR} = \text{wt.Rnvkor} / \text{wt.Rtr} \times 100$).

- m) Determination of the mean value of the percentage fractions of the water-insoluble starch remaining after enzymatic digestion of the samples ($RSaPMW = n \times RSaP / n$) and determination of the mean values of the percentage fractions of the remaining water-insoluble starch of the references: (recovery rate; $RSaRMW = n \times RSaR / n$) where n is the number of sample and reference assays carried out for the respective batches of water-insoluble starch.
- n) Determination of the percentage RS content of the individual batches of water-insoluble starch by correction of the mean values of the percentage fractions of the water-insoluble starch remaining after enzymatic digestion with the recovery rate ($RS = RSaPMW / RSaRMW \times 100$).

The invention is specifically illustrated by the following examples which are not in any way limiting.

Example 1 : Cloning of a mature gene encoding a dextranucrase from *L. mesenteroides*.

An expression cassette containing the patatin promoter (Wenzler et al., 1989), the chloroplastic ferredoxin transit peptide (FD) from *Silene pratensis* (Smeekens et al., 1985; Pilon et al., 1995) fused to the NOS terminator was cloned into the pBluescript SK (pBS SK) plasmid, resulting in pPF.

In order to isolate nucleic acid molecules encoding a dextranucrase, a mature DsrS gene was amplified by PCR from the genomic DNA from *L. mesenteroides* NRRL B-512 F (WO 89/12386), with a forward primer containing a *Sma*I restriction site (5'-GCCTCATTTGCTCCCGGGACACCAAGT-3') and a reverse primer containing a *Nru*I restriction site (5'-TG GTGGTTCGCGAGTTATGCTGACACA-3') using the proofreading Pfu

turbo DNA polymerase (2.5 units/ μ l; Stratagene, UK) and cloned into the SmaI/EcoRV restriction sites of pPF between FD and the NOS terminator, resulting in pPFDsrS. pPFDsrS was digested with SacI and KpnI and ligated into a pBIN20 binary vector (Hennegan and Danna, 1998),
5 resulting in pBinDsrS. pBinDsrS is shown in Figure 1.

Example 2 : Transformation of potato plants

pBinDsrS was transformed into *Agrobacterium tumefaciens* strain LBA
10 4404 using electroporation (Takken et al., 2000). Internodal stem segments from two tetraploid potato cultivars (cv. Kardal (KD) and amylose-free (amf) mutant) were used for *Agrobacterium*-mediated transformation. Transformants were selected on plates with MS30 medium (Murashige and Skoog, 1962) containing kanamycin (100 mg/l). 30
15 transgenic, root forming, shoots were multiplied and were transferred to the greenhouse for tuber development. The mature tubers were harvested after 18 weeks.

Example 3 : Starch isolation and Immunological detection of 20 dextrans in tuber juices and gelatinized starches

Potato tubers were peeled and homogenized in a Sanamat Rotor (Spangenberg, The Netherlands). The resulting homogenate was allowed to settle overnight at 4 °C and the potato juice was decanted and stored at – 20 °C for characterization of soluble dextran polymers. The starch pellet
25 was washed three times with water and finally air-dried at room temperature for at least three days. The dried starch was powdered and stored at room temperature.

Presence of dextrans was investigated with enzyme-linked immunosorbent assay (ELISA) as described by Matsuda and Kabat (1989)

by using monoclonal α -(1,6) dextran antibodies (4.3F1 (groove-type) and 16.4.12E (cavity-type)) (Wang et al., 2002).

ELISA analyses show that dextran polymers were detected both in transgenic potato lines obtained from Kardal lines or amylose-free mutant,
5 then no dextran was detected in wildtype kardal lines or non-transgenic amylose-free mutant.

Example 4 : Impact of dextran expression on starch granule morphology, plant morphology, tuber number and yield.

10 Analysis of starch granule morphology was performed by light microscopy (LM) (Axiophot, Germany) equipped with a Sony colour video camera (CCD-Iris/RGB) and scanning electron microscopy (SEM, JEOL 6300F, Japan). For LM, the granules were stained with a 2 x diluted Lugol solution before visualisation.

15

Impact of dextran expression was scored by analyzing a population of 100 starch granules per selected transformants in triplicates compared to starch granules isolates from wildtype lines.

Both technology show a modified starch granule morphology for starch
20 isolated from transgenic lines expressing dextran. The granules of these transformants exhibited irregular surfaces, and round-protruded structures in comparison to starch granules isolated from wildtype lines.

Figure 2 shows the modified starch granule morphology observed by scanning electron microscopy analysis performed on the starch of a
25 selected transformant (dsrS30), compared to the starch of a wild-type Kardal plant (Kardal)

On an other hand, the morphology of plant expressing dextran showed no phenotypic alteration in comparison to KD wildtype plants. In addition, there was no correlation between the expression of dextran polymers and
30 alteration of tuber number and yield.

Example 5 : Impact of dextran expression on digestibility of starch.

The digestibility of starch has been determined using the method detailed in the general methods (method 4). The determination was based upon the method of Englyst (European Journal of Clinical Nutrition (1992) 46 (suppl.2), p.33-50) for the determination of resistant starches Type III, modified in correspondence with the information on the determination of RS content in WO 00 02926.

The following table show a significant increase in the percentage of the digested starch for the sample extracted from a selected transformant (DsrS30) compared to the starch of a wild-type Kardal plant (Kardal).

Results:

15	Percentage of digested starch (%) :									
		15	30	45	60	120	180	240	300	360 min
20	Kardal	3	4	5	7	13	20	27	33	38
	DsrS30	3	5	7	8	15	23	31	37	44
25	Kardal: average values from four independent measurements DsrS 30: average values from two independent measurements									
30										

Example 6 : Impact of dextran expression on viscosity of starch.

The viscometric profiles from a starch suspension obtained from a transformant (DsrS30) and from a wild-type Kardal plant (Kardal) have been selected by the mean of a Thermo Haake rheoscope, using the method described in the general methods (method 2).

The following table show a significant decrease in the final viscosity for the sample extracted from a selected transformant (DsrS30) compared to the starch of a wild-type Kardal plant (Kardal).

5 *Kardal and DSrS30: average values of two independent analyses, from a 2% starch solutions*

Starch Sample	T-Onset (°C)	T-Peak (°C)	Peak Viscosity (PaS)	Valley Viscosity (PaS)	End Viscosity (PaS)
Kardal	75,7	78,1	122	27	134
DsrS30	73,8	75,9	135,5	22,5	93

10

REFERENCES

- 15 * An et al. EMBO J. 4, (1985), 277-287
- * Bäumlein et al., Mol. Gen. Genet. 225 (1991), 459-467
- * Chan et al., Plant Mol. Biol. 22, (1993), 491-506;
- 20 * van Cleve, J.W., Schaefer, W.C. and Rist, C.E. 1956. J. Am. Chem. Soc. 78: 4435-4438.
- * Conner and Domisse, Int. J. Plant Sci. 153 (1992), 550-555
- 25 * Creissen et al., Plant J. 8 (1995), 167-175
- * De Vuyst and Degeest 1999 FEMS Microbiol Rev. 23 (2):153-77.
- 30 * Deng et al, Science in China 33, (1990), 28-34

- * Eckhoff et al. Cereal Chem. 73 (1996) 54-57
- * Emanuelsson O. *et al*, 1999, Protein Science:8:978-984
- 5 * Englyst H.N. et al., European Journal of Clinical Nutrition 4, Suppl.2, S33-S50
- * Fiedler et al., Plant Mol. Biol. 22 (1993), 669-679
- 10 * Fraley et al., Crit. Rev. Plant Sci. 4, 1-46
- * Fromm et al., Biotechnology 8, (1990), 833-844;
- * Fu D, Robyt JF (1990) Arch Biochem Biophys 283: 379-387
- 15 * Gallardo et al. (1995), Planta 197, 324-332
- * Gerrits, N., Turk, S.C.H.J., van Dun, K.P.M., Hulleman, S.H.D., Visser, R.G.F., Weisbeek, P.J. and Smeekens, S.C.M. 2001. Plant Physiol. 125: 926-934.
- 20 * Gielen et al. 1989, EMBO J. 8, 23-29
- * Gordon-Kamm et al. 1990, Plant Cell 2, 603-618;
- 25 * Hiei et al. (1994), Plant J. 6, 271-282
- * Hehre EJ, Hamilton DM, Carlson AS (1949). J Biol Chem 177: 267-279
- 30 * Hennegan, K.P. and Danna, K.J. (1998). Plant Mol. Biol. Rep. 16: 129-131.
- * Hoekema, IN: The Binary Plant Vector System Offsetdrukkerij Kanters B.V., Alblasterdam (1985), Chapter V
- 35 * Janecek S, Svensson B, Russell RRB (2000). FEMS Microbiol Lett 192: 53-57
- * Jeanes, A., Haynes, W.C., Wilham, C.A., Rankin, J.C., Melvin, E.H., Austin, M.J., Cluskey, J.E., Fisher, B.E., Tsuchiya, H.M. and Rist, C.E. (1954). J. Am. Chem. Soc. 76: 5041-5052.
- 40

- * Khan M.S. and Maliga P. (1999). Nat. Biotechnol. 17, 910-915
- * Klösgen et al (1989), Mol Gen Genet. 217, 155-161
- 5 * Kok-Jacob A., Ji Q., Vincken JP., and Visser RG. (2003) J. Plant Physiol;
160, 765-777
- * Kortstee AJ, Vermeesch AM, de Vries BJ, Jacobsen E, Visser RG.(
10 1996), Plant J. 10(1):83-90.
- * Kossmann and Llyod (2000), Crit. Rev. Bioch. Mol. Biol. 35 : 141-196
- 15 *Koziel et al., (1993) Biotechnology 11, 194-200;
- * Krens et al., Nature 296, (1982), 72-74
- * Leisy et al., Plant Mol. Biol. 14 (1990), 41-50
- 20 * MacGregor EA, Jespersen HM, Svensson B (1996). FEBS lett 378: 263-
266
- * Matsuda, T. and Kabat, E.A. 1989. J. Immunol. 142: 863-870.
- 25 * May et al., Bio/Technology 13, (1995), 486-492;
- * Monchois V, Remaud-Simeon M, Russell RRB, Monsan P and Willemot
RM. 1997. Appl. Microbiol. Biotechnol. 48 : 465-472
- 30 * Monchois V, Remaud-Simeon M, Monsan P and Willemot RM. 1998.
FEMS Microbiol. Lett. 159 : 307-315
- * Monchois et al., 1999, FEMS Microbiology Letters 177, 243-248;
- 35 * Monchois V, Willemot RM and Monsan P. 1999, FEMS Microbiology
Reviews 23, 131-151.
- * Moroc et al., Theor. Appl. Genet. 80, (1990), 721-726).
- 40 * Murashige, T. and Skoog, F. 1962. Physiol. Plant. 15: 473-497.
- * Nawrath et al., Proc. Natl. Acad. Sci. 10 USA 91 (1994), 12760-12764)

- * Nehra et al., Plant J. 5, (1994), 285-297
- * Oakes JV, Shewmaker CK, Stalker DM (1991). Biotechnol 9: 982-986
- 5 * Pedersen et al., Cell 29 (1982), 1015-1026;
- * Pilon, M., Wienk, H., Sips, W., de Swaaf, M., Talboom, I., van 't Hof, R.,
de Korte-Kool, G., Demel, R., Weisbeek, P. and de Kruijff, B. 1995. J. Biol.
10 Chem. 270: 3882-3893.
- * Quatroccio et al., Plant Mol. Biol. 15 (1990), 81-93
- * Quirasco *et al* (Appl. Environ. Microbiol., 65 (12), 5504-5509, 1999
- 15 * Ritala et al., Plant Mol. Biol. 24, (1994), 317-325;
- * Ritchie et al, Transgenic Res. 2, (1993), 252-265).
- 20 * Robyt, J.F. and Walseth, T.F. 1978. Carbohydr. Res. 61: 433-445.
- * Robyt, J.F. 1995. Adv. Carbohydr. Chem. Biochem. 51: 133-168.
- 25 * Rocha-Sosa et al., EMBO J. 8, (1989), 29-33
- * Ruf S., Hermann M., Berger I.J., Carrer H., and Bock R. (2001). Nat.
Biotechnol. 19 (9):870-875.
- 30 * Sambrook et al. (1989, Molecular Cloning : A Laboratory Manual, Nolan
C. ed., New York: Cold Spring Harbor Laboratory Press
- * Shewmaker CK, Boyer CD, Wiesenborn DP, Thompson DB, Boersig MR,
Oakes JV, Stalker DM. Plant Physiol. 1994 Apr;104(4):1159-66.
- 35 * Sidebotham, R.L. 1975. Dextran. Adv. Carbohydr. Chem. Biochem. 30:
371-444.
- * Sidorov V.A., Kasten D., Pang S.Z., Hajdukiewicz P.T., Staub J.M., and
40 Nehra N.S. (1999). Plant J. 19(2):209-216.
- * Sikdar S. R., et al. (1998). Plant Cell Reports 18:20-24.

- * Smeekens, S., van Binsbergen, J. and Weisbeek, P. 1985. Nucleic Acids Res. 13: 3179-3194
- 5 * Spencer et al., Theor. Appl. Genet. 79, (1990), 625-631
- * Staub et al. 1992 Plant Cell 4:39-45
- * Staub J.M., and Maliga P. (1993). EMBO J. 12 (2): 601-606.
- 10 * Stockhaus et al., Proc. Natl. Acad. Sci. USA 84 (1987), 7943-7947
- * Stockhaus et al., EMBO J. 8 (1989), 2445-2451
- 15 * Su D. and Robyt JF., Arch Biochem Biophys. 1994 Feb 1;308(2):471-6.
- * Svab Z., Hajdukiewicz P., and Maliga P. (1990). Proc. Natl. Acad. Sci. USA 87 (21):8526-8530.
- 20 * Svab Z., Maliga P. (1993) ; Proc. Natl. Acad. Sci. U S A Feb 1;90(3):913-7.
- * Takken, F.L.W., Luderer, R., Gabriëls, S.H.E.J., Westerink, N., Lu, R., de Wit, P.J.G.M. and Joosten, M.H.A.J. 2000. Plant J. 24: 275-283.
- 25 * Thompson et al., Nucleic Acids Research 22 (1994), 4673-4680
- * Van Geel-Schutten GH, Faber EJ, Smit E, Bonting K, Smith MR, Ten Brink B, Kamerling JP, Vliegenthart JF, Dijkhuizen L., Appl Environ Microbiol. 1999 Jul;65(7):3008-14.
- 30 * Vasil et al., Bio/Technology 11 (1993), 1553-1558;
- * Wan and Lemaux, Plant Physiol. 104, (1994), 37-48
- 35 * Wang, D., Liu, S., Trummer, B.J., Deng, C. and Wang, A. 2002. Nature Biotechnol. 20: 275-281.
- * Weising K, Schell J, Kahl G., Annu Rev Genet. 1988;22:421-77. Review
- 40 * Wenzler, H.C., Mignery, A., Fisher, L.M. and Park, W.D. 1989. Plant Mol. Biol. 12: 41-50.

- * Werr et al., EMBO J. 4 (1985), 1373-1380
- * Wilmink et al., Plant Cell Reports 11, (1992), 76-80;
- 5 * White et al., Nucleic Acid Res. 18(4) :1062.
- * Wolter et al, Proc. Natl. Acad. Sci. USA 85 (1988), 846-850
- 10 * Yoshihara et al., FEBS Lett. 383 (1996), 213-218
- * Zheng et al., Plant J. 4 (1993), 357-366

DEMANDE OU BREVET VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET COMPREND PLUS D’UN TOME.

CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 50

NOTE : Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE THAN ONE VOLUME

THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 50

NOTE: For additional volumes, please contact the Canadian Patent Office

NOM DU FICHER / FILE NAME :

NOTE POUR LE TOME / VOLUME NOTE:

Claims

1. A Genetically modified plant cell characterized in that it has an enzymatic activity of a dextranucrase DSR-S protein in their plastids and
5 wherein said genetically modified plant cell synthesizes a modified starch in comparison to starch synthesized by corresponding non-genetically modified wild-type plant cells.
2. A genetically modified plant cell according to claim 1 wherein the plant
10 cell synthesizes a modified starch which has a decreased end viscosity and/or an increased digestibility, in comparison to starch synthesized by corresponding non-genetically modified wild-type plant cells.
3. A plant and/or progeny thereof comprising a genetically modified plant
15 cell according to anyone of claims 1 or 2
4. A plant and/or progeny thereof according to claim 3, which is a starch-storing plant.
- 20 5. A plant and/or progeny thereof according to claim 4, which is a potato plant.
6. Propagation material of a plant according to anyone of claims 3 to 5, comprising plant cells according to anyone of claims 1 or 2
25
7. Harvestable parts of a plant according to anyone of claim 3 to 5, comprising plant cells according to anyone of claims 1 or 2
8. A method for the manufacture of a genetically modified plant according
30 to anyone of claims 3 to 5 wherein

- a) a plant cell is transformed with a nucleic acid molecule comprising a nucleic acid molecule encoding a dextranucrase DSR-S protein,
b) a plant is regenerated from a plant cell obtained in step a), and
c) if necessary, further plants are produced from the plants obtained in
5 step b).

9. A method according to claim 8, wherein the nucleic acid encoding the dextranucrase DSR-S protein in step a) is translationally fused with a nucleic acid molecule encoding a plastidial signal sequence.

10

10. A method according to claim 8, wherein the nucleic acid molecule encoding the dextranucrase DSR-S protein is integrated into the plastidial genome of the plant.

- 15 11. A method according to anyone of claims 8 to 10 or a plant cell according to any of claim 1 or 2, or a plant according to any of claim 3 to 5 wherein the nucleic acid molecule encoding a dextranucrase DSR-S protein is chosen from the group consisting of
a) Nucleic acid molecules, which encode a protein comprising the amino
20 acid sequence given under SEQ ID NO: 2 or SEQ ID N04;
b) Nucleic acid molecules, which encode a protein, the amino acid sequence of which has an identity of at least 70% with the amino acid sequence given under SEQ ID NO: 2 or SEQ ID N04;
c) Nucleic acid molecules, comprising the nucleotide sequence shown
25 under Seq ID NO:3 or a complementary sequence thereof;
d) Nucleic acid molecules, the nucleic acid sequence of which has an identity of at least 70% with the nucleic acid sequences described under a) or c);

e) Nucleic acid molecules, the nucleotide sequence of which deviates from the sequence of the nucleic acid molecules identified under a), b), c), or d) due to the degeneration of the genetic code; and

f) Nucleic acid molecules, which represent fragments, allelic variants
5 and/or derivatives of the nucleic acid molecules identified under a), b), c), d), or e).

12. A method for the manufacture of a modified starch comprising the step of extracting the starch from a plant cell according to anyone of claims 1 to
10 2, from a plant according to anyone of claims 3 to 5, from harvestable parts of a plant according to claim 7, or from a plant obtainable by means of a method of anyone of claims 8 to 11.

13. Modified starch obtainable by a method according to claim 12.
15

14. A method for the production of a derived starch, wherein modified starch according to claim 13 is derived.

15. Derived starch obtainable by a method according to claim 14.
20

16. Use of a plant cell according to anyone of claims 1 or 2, a plant according to anyone of the claims 3 to 5, harvestable parts of a plant according to claim 7 or a plant obtainable by means of a method of anyone of claims 8 to 11, for the production of a modified starch.
25

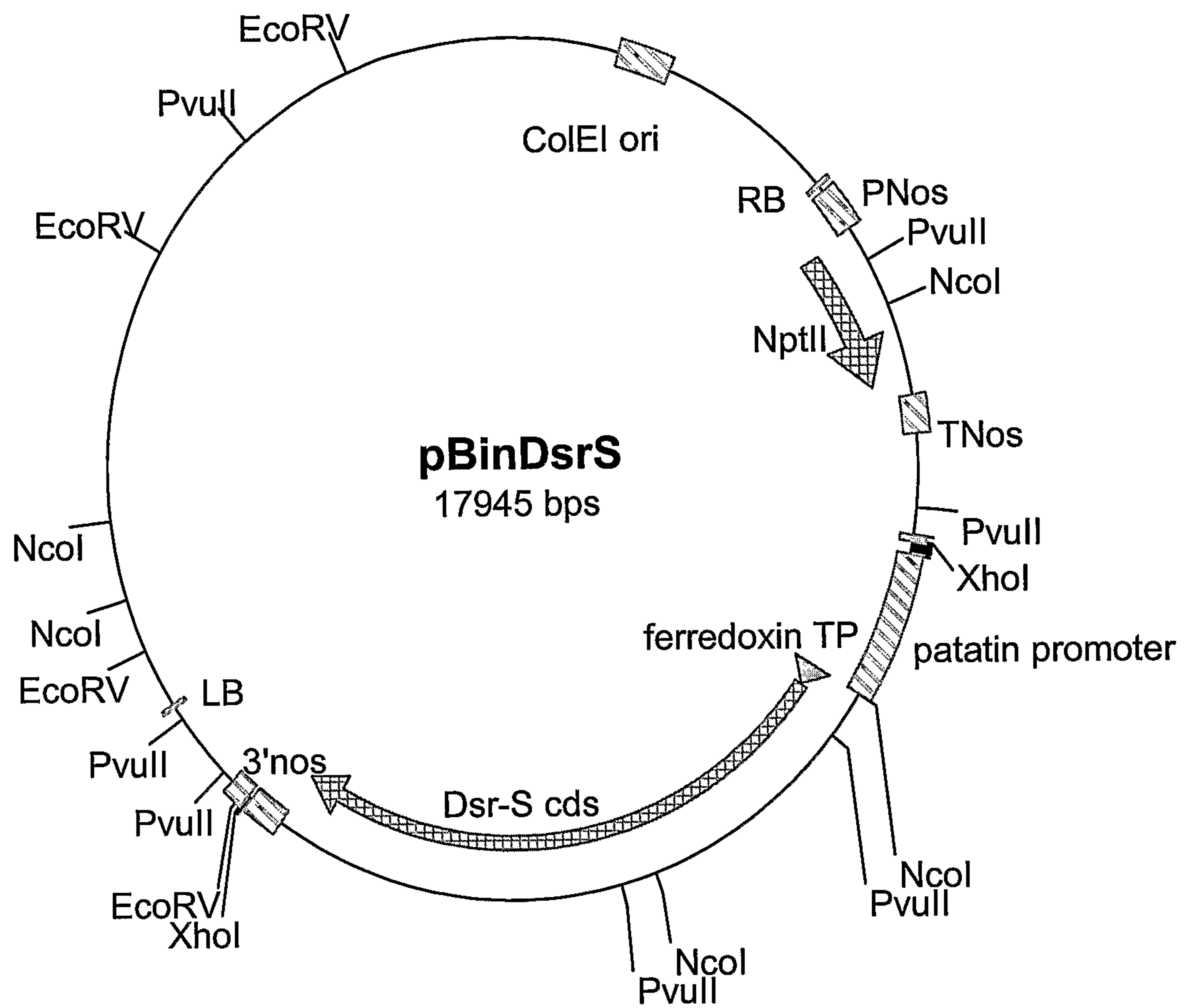
17. Use of a nucleic acid molecule encoding a dextransucrase DSR-S
protein for the manufacture of a genetically modified plant cell according to
anyone of claims 1 or 2, a genetically modified plant according to anyone
of claims 3 to 5, propagation material according to claim 6, or harvestable
5 parts of plants according to claim 7.

10

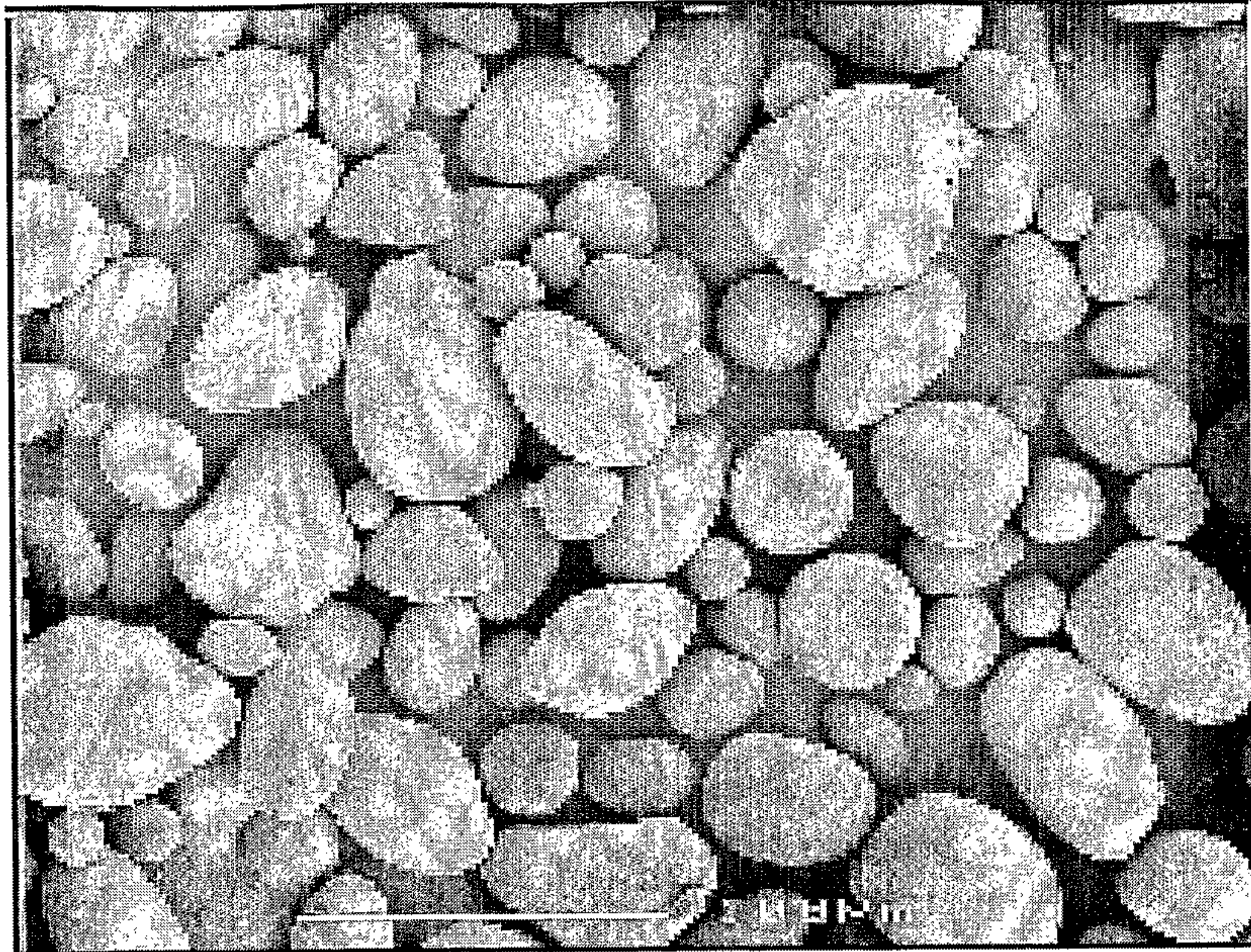
15

20

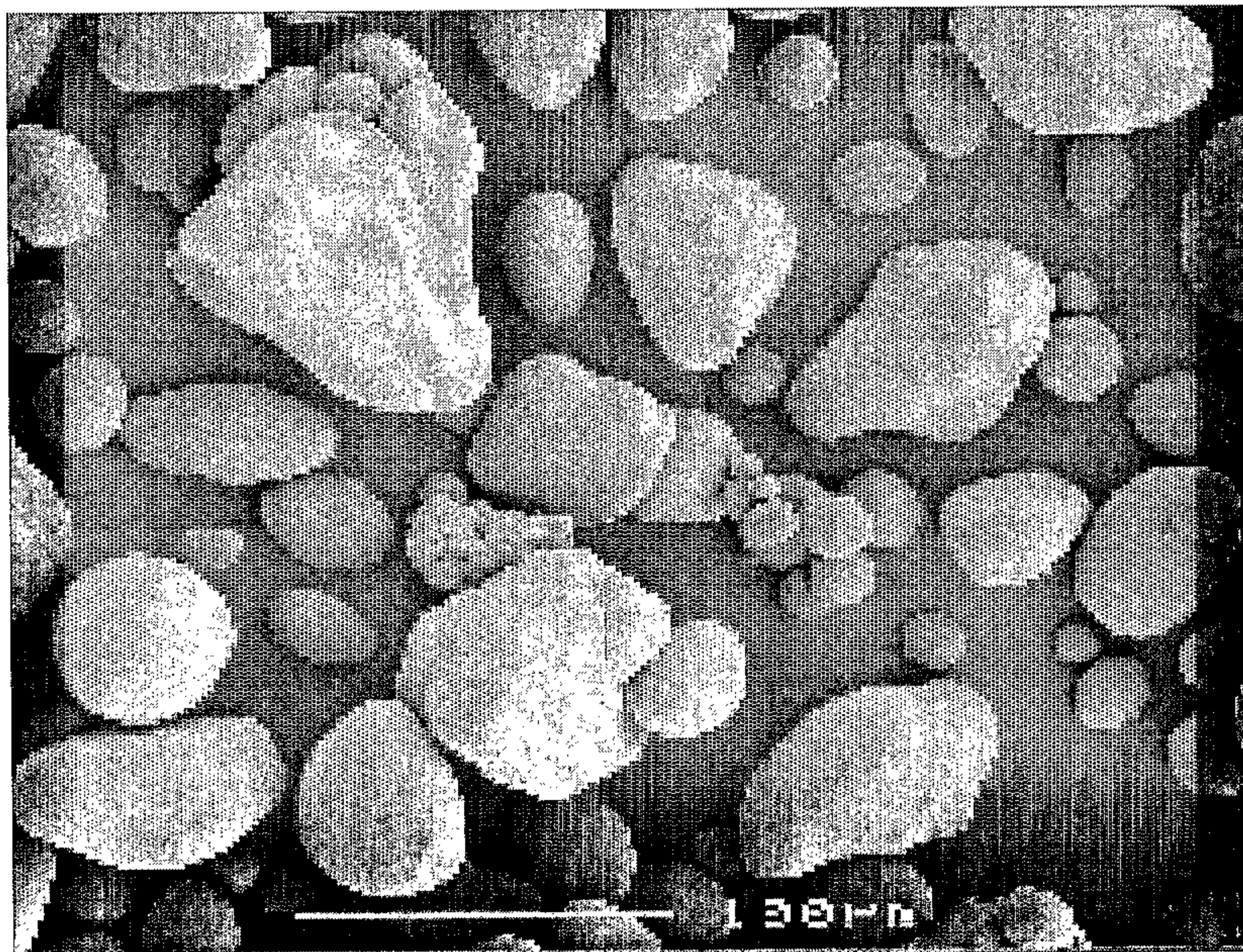
1/2

**FIG.1**

2/2



Kardal



Dsr5 30

FIG.2