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(54) Titre : DOMAINE EXTRACELLULAIRE DE HER2  
 (54) Title: HER2 EXTRACELLULAR DOMAIN

(57) **Abrégé/Abstract:**

An extracellular portion of the HER2 molecule, essentially free of transmembrane and cytoplasmic portions, which is antigenic in animals. Isolated DNA encoding the extracellular portion; an expression vector containing the isolated DNA; and a cell containing the expression vector. A process for producing the extracellular domain. A vaccine containing the extracellular domain.



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<p><b>(21) International Application Number:</b> PCT/US90/02697</p> <p><b>(22) International Filing Date:</b> 18 May 1990 (18.05.90)</p> <p><b>(30) Priority data:</b>  354,319                      19 May 1989 (19.05.89)                      US</p> <p><b>(71) Applicant:</b> GENENTECH, INC. [US/US]; 460 Point San Bruno Boulevard, South San Francisco, CA 94080 (US).</p> <p><b>(72) Inventors:</b> HUDZIAK, Robert, Michael ; 241 San Diego Avenue, Apartment #4, San Bruno, CA 94066 (US). SHEPARD, H., Michael ; 35 Delano Avenue, San Francisco, CA 94112 (US). ULLRICH, Axel ; D-8033 Martinsried (DE).</p>	<p><b>(74) Agent:</b> GREENBLUM, Neil, F.; Sandler, Greenblum &amp; Bernstein, 2920 South Glebe Road, Arlington, VA 22206 (US).</p> <p><b>(81) Designated States:</b> AT (European patent), BE (European patent), CA, CH (European patent), DE (European patent)*, DK (European patent), ES (European patent), FR (European patent), GB (European patent), IT (European patent), LU (European patent), NL (European patent), SE (European patent).</p> <p><b>Published</b>  <i>With international search report.</i>  <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>	
<p><b>(54) Title:</b> HER2 EXTRACELLULAR DOMAIN</p>		
<p><b>(57) Abstract</b></p> <p>An extracellular portion of the HER2 molecule, essentially free of transmembrane and cytoplasmic portions, which is antigenic in animals. Isolated DNA encoding the extracellular portion; an expression vector containing the isolated DNA; and a cell containing the expression vector. A process for producing the extracellular domain. A vaccine containing the extracellular domain.</p>		

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HER2 EXTRACELLULAR DOMAINBACKGROUND OF THE INVENTIONField of the Invention

The present invention is generally directed to the  
5 extracellular domain of p185<sup>HER2</sup>, a receptor-like protein  
which is encoded by the human homolog of the rat neu  
oncogene.

More specifically, the present invention is directed  
to a form of the extracellular domain which is essentially  
10 free of transmembrane and cytoplasmic domains, to the DNA  
encoding this form, and to a process for producing this  
form of the extracellular domain in a host cell.

Description of Background and Relevant Materials

Human epidermal growth factor receptor 2 (HER2, also  
15 known as NGL and human c-erbB-2, or ERBB2), is the human  
homolog of the rat proto-oncogene neu. HER2 encodes a  
1,255 amino acid tyrosine kinase receptor-like glycoprotein  
with homology to the human epidermal growth factor  
receptor. Although no ligand binding to this probable  
20 growth factor receptor has yet been isolated, the HER2 gene  
product, p185<sup>HER2</sup>, has the structural and functional  
properties of subclass I growth factor receptors (Yarden et  
al., Ann. Rev. Biochem., 57:443-478 (1988); Yarden et al.,  
Biochem., 27:3113-3119 (1988)).

25 The receptor tyrosine kinases all have the same  
general structural motif; an extracellular domain that  
binds ligand, and an intracellular tyrosine kinase domain  
that is necessary for signal transduction, or in aberrant



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cases, for transformation. These 2 domains are connected by a single stretch of approximately 20 mostly hydrophobic amino acids, called the transmembrane spanning sequence. This sequence is thought to play a role in transferring the signal generated by ligand binding from the outside of the cell to the inside. It has also been suggested to play a role in the proper positioning of the receptor in the plasma membrane.

Consistent with this general structure, the p185<sup>HER2</sup> glycoprotein, which is located on the cell surface, may be divided into three principle portions: an extracellular domain, or ECD (also known as XCD); a transmembrane spanning sequence; and a cytoplasmic, intracellular tyrosine kinase domain. While it is presumed that the extracellular domain is a ligand receptor, as stated above the corresponding ligand has not yet been identified.

The HER2 gene is of particular interest because its amplification has been correlated with certain types of cancer. Amplification of the HER2 gene has been found in human salivary gland and gastric tumor-derived cell lines, gastric and colon adenocarcinomas, and mammary gland adenocarcinomas. Semba et al., Proc. Natl. Acad. Sci. USA, 82:6497-6501 (1985); Yokota et al., Oncogene, 2:283-287 (1988); Zhou et al., Cancer Res., 47:6123-6125 (1987); King et al., Science, 229:974-976 (1985); Kraus et al., EMBO J., 6:605-610 (1987); van de Vijver et al., Mol. Cell. Biol., 7:2019-2023 (1987); Yamamoto et al., Nature, 319:230-234 (1986). Gene transfer experiments have shown that

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overexpression of HER2 will transform NIH 3T3 cells and also cause an increase in resistance to the toxic macrophage cytokine tumor necrosis factor. Hudziak et al., "Amplified Expression of the HER2/ERBB2 Oncogene Induces  
5 Resistance to Tumor Necrosis Factor Alpha in NIH 3T3 Cells", Proc. Natl. Acad. Sci. USA 85, 5102-5106 (1988).

Because amplification of the HER2 gene results in greatly increased numbers of the p185<sup>HER2</sup> protein residing on the surfaces of affected cells, there may be an  
10 interrelationship between increased amounts of p185<sup>HER2</sup> extracellular domain on the surfaces of affected cells and the resistance of these cells to treatment. It would therefore be highly desirable to be able to manipulate the p185<sup>HER2</sup> extracellular domain in order to investigate several  
15 possibilities for the treatment of conditions associated with amplification of the HER2 gene. These therapeutic modes relate not only to the extracellular domain, but also to the putative ligand, which it should be possible to isolate and characterize using the purified p185<sup>HER2</sup>  
20 extracellular domain.

#### SUMMARY OF THE INVENTION

The present invention is accordingly directed to an extracellular portion of the HER2 molecule containing at least 9 amino acids, and/or containing an immune epitope,  
25 which is essentially free of transmembrane and intracellular portions of the HER2 molecule. The extracellular portion may be substantially pure, or at least about 99% pure, and may extend to the entire



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extracellular portion of the HER2 molecule. Moreover, the extracellular portion may be antigenic in animals, and may be conjugated with a peptide having immunogenic properties; this peptide may contain an immune epitope.

5 In another embodiment, the present invention is directed to isolated DNA encoding the extracellular portion of the HER2 molecule. This isolated DNA terminates upstream of the DNA portion encoding the transmembrane domain of the HER2 molecule. The termination may occur at  
10 least 1 base pair upstream of the portion encoding the transmembrane domain of the HER2 molecule, and preferably occurs about 24 base pairs upstream of this portion.

The isolated DNA of the present invention encodes a sequence of at least 9 amino acids of the extracellular  
15 portion, and none of the transmembrane or intracellular portions of the HER2 molecule.

In a further embodiment, the present invention contemplates an expression vector, such as a plasmid or virus, containing the isolated DNA as described above, as  
20 well as a cell containing the expression vector. This cell may be eukaryotic or prokaryotic.

The present invention also extends to a process for producing an extracellular portion of the HER2 molecule, which includes the steps of ligating the isolated DNA as  
25 described above into an expression vector capable of expressing the isolated DNA in a suitable host; transforming the host with the expression vector;

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culturing the host under conditions suitable for expression of the isolated DNA and production of the extracellular portion; and isolating the extracellular portion from the host. The host cell may be a prokaryote, such as a bacterium, or a eukaryote.

In a yet further embodiment, the present invention extends to a vaccine comprising the extracellular portion of the HER2 molecule, which may be combined with suitable adjuvants.

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#### BRIEF DESCRIPTION OF FIGURES

Fig 1. HER2 expression vector and full-length and mutant HER2 proteins. The HER2 expression vector contained eukaryotic transcriptional units for the mouse dihydrofolate reductase (DHFR) cDNA and the bacterial neomycin phosphotransferase (neo) gene, both under SV40 early promoter control. Transcription of the full-length HER2 cDNA was also driven by the early SV40 promoter. The full-length HER2 protein contains an extracellular domain with two cysteine-rich clusters (hatched rectangle), separated by the transmembrane-spanning region (filled rectangle) from the intracellular tyrosine kinase domain (open rectangle). The mutant protein p185<sup>HER2 $\Delta$ TM</sup> has a deletion of 28 amino acids, including the transmembrane-spanning region. The truncated p185<sup>HER2XCD</sup> protein contains all N-terminal sequences up to 8 amino acids before the transmembrane-spanning region.

Fig. 2. Amplification of HER2 and HER2 $\Delta$ TM genes. Cell lines transfected with plasmids expressing wild type



or the  $\Delta$ TM mutant HER2 cDNAs were amplified to resistance to 400 nM methotrexate. Cultures were metabolically labeled with [ $^{35}$ S]-methionine and proteins immunoprecipitated with the G-H2CT17 antibody. Lane 1: CVN-transfected NIH 3T3 vector control line. Lanes 2 and 3: Parental and amplified HER2-3 line. Lanes 4, 5, and 6, 7: Parent and amplified lines derived from two independent clones, A1 and B2, of the  $\Delta$ TM mutant. The arrows indicate the positions expected for proteins of apparent molecular mass of 175 and 185 kDa.

Fig. 3. Autophosphorylation of p185<sup>HER2</sup> and p185<sup>HER $\Delta$ TM</sup> proteins. Triton X-100\* lysates of control, HER2-3<sub>400</sub>, and  $\Delta$ TM-expressing cell lines were prepared and immunoprecipitated with the G-H2CT17 antibody. The immune complexes were incubated in 50  $\mu$ l of HNTG, 5 mM MnCl<sub>2</sub> with 3  $\mu$ Ci [ $\gamma$ - $^{32}$ P] for 20 min, electrophoresed on a 7.5% polyacrylamide gel, and labeled bands visualized by autoradiography. Lane 1: CVN vector control. Lane 2: HER2-3<sub>400</sub> cells expressing full-length HER2. Lanes 3 and 4: Two independent lines expressing p185<sup>HER2 $\Delta$ TM</sup>. The arrows indicate the positions expected for proteins of apparent molecular mass of 66.2, 97, 175, and 185 kDa.

Fig 4. Secretion assay of  $\Delta$ TM mutants. Cell lines CVN, HER2-3<sub>400</sub>,  $\Delta$ TM-A1<sub>400</sub>, and  $\Delta$ TM-B2<sub>400</sub> were labeled with [ $^{35}$ S]-methionine overnight. Triton X-100 cell extracts were prepared and the labeling medium collected. Cells and cell-conditioned media were immunoprecipitated with G-H2CT17 antibody and analyzed on 7.5% SDS-PAGE gels.

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Lanes 1-4 are immunoprecipitations of cell extracts from the various lines, and lanes 5-8 are immunoprecipitations from the corresponding cell-conditioned media. Lanes 1 and 5: CVN vector control. Lanes 2 and 6: HER2-3<sub>400</sub> cell lines expressing full-length p185<sup>HER2</sup>. Lanes 3, 4 and 7, 8: ΔTM-A1<sub>400</sub> and ΔTM-B2<sub>400</sub> cell lines expressing mutant p185<sup>HER2ΔTM</sup>. The arrows indicate the positions expected for proteins of apparent molecular mass of 175 and 185 KDa.

Fig 5. Secretion of p185<sup>HER2XCD</sup> from 3T3 and CHO cells. NIH 3T3 and CHO cell lines expressing full-length and truncated p185<sup>HER2</sup> and vector controls were labeled with [<sup>35</sup>S]-methionine overnight. Cell extracts and cell-conditioned media were immunoprecipitated with anti-HER2 monoclonal antibody 3E8 and analyzed on 7.5% SDS-PAGE gels. Lanes 1 and 2: NIH 3T3 control cell line, extract and conditioned medium. Lanes 3 and 4: NIH 3T3 line A1 expressing p185<sup>HER2XCD</sup>, cells and medium. Lanes 5 and 6: NIH 3T3 line 3<sub>400</sub> expressing full-length p185<sup>HER2</sup>, cells and conditioned medium. Lanes 7 and 8: CHO control line, cell extract and conditioned medium. Lanes 9 and 10: CHO line 2, expressing p185<sup>HER2XCD</sup>, cells and conditioned medium. Lanes 11 and 12: CHO line HER2<sub>500</sub>, expressing full-length p185<sup>HER2</sup>, cells and conditioned medium. The arrows indicate the molecular mass of the indicated protein bands.

Fig 6. Increase in expression of p185<sup>HER2XCD</sup> with amplification. The CHO-derived cell line HER2XCD-2 was selected for growth in 500 nM and then 3000 nM methotrexate. The parent line, the two amplified

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derivatives, and control vector-transfected cells were labeled with [<sup>35</sup>S]-methionine. Cell extracts and cell-conditioned media were immunoprecipitated with the anti-HER2 monoclonal antibody 3E8 and analyzed on a 7.5% SDS-PAGE gel. Lanes 1 and 2: CVN cell extract and conditioned medium. Lanes 3 and 4: HER2XCD-2, unamplified cells and conditioned medium. Lanes 5 and 6: HER2XCD-2 amplified to resistance to 500 nM methotrexate, cells and conditioned medium. Lanes 7 and 8: HER2XCD-2 amplified to resistance to 3000 nM methotrexate, cells and conditioned medium. For comparative purposes, one-fifth as much sample of the 3000 nM line was loaded compared to the control, 0 nM, and 500 nM lines. The band intensities were quantitated with an LKB2202 laser densitometer. The arrows show the positions of proteins of apparent molecular mass of 88 and 103 KDa.

Fig 7. Biosynthesis of p185<sup>HER2XCD</sup>. The CHO line HER2XCD2<sub>3000</sub> was labeled with [<sup>35</sup>S]-methionine and cell extracts, and cell-conditioned media prepared. Lanes 1 and 2: Cell extract and cell-conditioned medium. Lanes 3 and 4: The same conditioned medium incubated or mock-incubated with endo H. Lanes 5 and 6: Cell extract and conditioned medium from cells treated with tunicamycin. The arrows show the positions expected for proteins of apparent molecular mass of 73, 88, and 103 KDa.

Fig 8. Morphology of NIH 3T3 cells transfected with HER2 and HER2 $\Delta$ TM expression constructs. A and D: Parental and amplified cells from NIH 3T3 cells transfected with



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vector alone. B and E: NIH 3T3 cells expressing p185<sup>HER2ΔTM</sup> (line A1), parent and amplified derivative selected for resistance to 400 nM methotrexate. C and F: NIH 3T3 cells expressing wild type p185<sup>HER2</sup> (line 3), parent and amplified derivative selected for resistance to 400 nM methotrexate.

Fig 9. Cell surface and cytoplasmic immunofluorescence staining of control, HER2, and HER2ΔTM lines. The top photos are intact cells labeled with anti-HER2 monoclonal antibody. The bottom photos are the same cell lines treated with 0.15% Triton X-100 detergent before addition of antibody. A and D: Control NIH 3T3 cells transfected with vector only. B and E: Cell line HER2ΔTM-A1<sub>400</sub>, expressing p185<sup>HER2ΔTM</sup>. C and F: Cell line HER2-3<sub>400</sub> expressing p185<sup>HER2</sup>.

Fig 10. Fluorescence-activated cell sorter histograms of control, HER2 and HER2ΔTM cells binding anti-p185<sup>HER2</sup> monoclonal antibody 4D5. Binding by the control antibody, 368, directed against human tissue plasminogen activator, light, broken line. Binding by the anti-HER2 antibody 4D5, dark unbroken line. Panel A: Control NIH 3T3 cells transfected with vector only. Panel B: Cell line HER2-3<sub>400</sub>, expressing p185<sup>HER2</sup>. Panel C: Cell line HER2ΔTM-A1<sub>400</sub> expressing p185<sup>ΔTM</sup>.

Fig 11. Biosynthesis of p185<sup>HER2</sup> and p185<sup>HER2ΔTM</sup> proteins. Cell lines HER2-3<sub>400</sub> and HER2ΔTM-A1<sub>400</sub> were labeled with [<sup>35</sup>S]-methionine and p185<sup>HER2</sup> and p185<sup>HER2ΔTM</sup> proteins collected by immunoprecipitation and analyzed on a 7.5% SDS-PAGE gel. Lane 1: Vector control. Lane 2: Untreated p185<sup>HER2ΔTM</sup>. Lanes

3 and 4: Aliquots of the same cell extract treated or mock-treated with endo H. Lane 5: Nonglycosylated p185<sup>HER2</sup> from cells treated with tunicamycin. Lane 6: Untreated p185<sup>HER2</sup>. Lanes 7 and 8: Aliquots of the same cell extract treated or mock-treated with endo H. Lane 9: Nonglycosylated p185<sup>HER2ΔTM</sup> from cells treated with tunicamycin. The arrows show the positions of proteins of apparent molecular weight of 175 and 185 kDa.

Fig. 12. Purification of the HER2 extracellular domain. Purified HER2 extracellular domain samples were analysed using PhastSystem\* SDS-Gel electrophoresis and silver stained protocols as recommended by Pharmacia. SDS polyacrylamide gel (10-15% gradient) electrophoretic analysis was performed according to Pharmacia protocol File No. 110. Silver staining was performed according to Pharmacia protocol File No. 210. Lane 1 contains molecular weight markers (BRL). Lane 2: Chinese Hamster Ovary Cell 15 X concentrate (1 microliter). Lanes 3 and 4: immunoaffinity purified HER2 extracellular domain (1.6 micrograms and 0.16 microgram, respectively). Lanes 5 and 6: immunoaffinity purified HER2 extracellular domain after DEAE chromatography (0.25 micrograms and 0.083 micrograms, respectively). Lanes 7 and 8: HER2 extracellular domain after formulation in PBS (0.32 micrograms and 0.082 micrograms, respectively).

Fig. 13. The predicted amino acid sequence of the HER2 extracellular domain, with the corresponding nucleic acid sequence. The boxed sequences show potential T-cell

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epitopes, using the algorithm developed by Margolit et al.,  
J. Immunol. 138:2213-2229(4) (1987).

#### DETAILED DESCRIPTION

It was initially hypothesized that removal of the  
5 transmembrane spanning sequence would yield a protein which  
would be secreted from the cell. As previously indicated,  
the transmembrane spanning sequence is principally composed  
of hydrophobic amino acids, which effectively anchor the  
protein in the cell membrane. Removal of this sequence  
10 would therefore be expected to permit passage of the  
protein through the membrane.

A first construct was accordingly prepared which  
deleted exactly in-frame the 22 amino acid transmembrane  
spanning sequence of HER2, and 3 amino acids on either side  
15 (Figure 1). The construct was prepared as follows:

The central EcoR1 fragment containing the trans-  
membrane spanning segment was cloned into the EcoR1 site of  
the bacteriophage vector M13 mp18 (Yanisch-Perron et al.,  
Gene, 33:103-119 (1985). The noncoding strand was used as  
20 template for oligonucleotide-directed mutagenesis. The  
construct deleted the transmembrane spanning sequence, and  
an additional 3 amino acids before and after.

Residues 651-678 were deleted by priming double  
stranded DNA synthesis with a 30 base pair oligonucleotide  
25 of sequence 5' CAG AGA GCC AGC CCT CAG CAG AAG ATC CGG 3'.  
The double stranded DNA was transformed into SR101 cells  
and mutants identified by hybridization to the same  
oligonucleotide 5' end labeled by polynucleotide kinase and

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[ $\gamma$ -<sup>32</sup>P] ATP (Amersham, 5000 Ci/mmol). An EcoR1 fragment containing the deletion was recombined into a plasmid expressing the HER2 cDNA, replacing the wild type sequence.

When expressed in NIH 3T3 cells, this mutant, designated HER2<sup>Δ</sup>, produced a polypeptide, designated p185<sup>HER2Δ</sup>, of apparent molecular weight 175 kD (Figure 2, lanes 5 and 7). Production took place at levels comparable to wild type p185<sup>HER2</sup> amplified to the same level of resistance to methotrexate (Figure 2, lane 3). The mutant proteins also retained an active tyrosine kinase activity.

In the presence of [ $\gamma$ -<sup>32</sup>P]-ATP, the mutant proteins (Figure 3, lanes 3 and 4) were autophosphorylated to the same extent as unaltered p185<sup>HER</sup> (Figure 3, lane 2). Figure 3 also shows autophosphorylated p185<sup>HER2Δ</sup>-related proteins of lower molecular weight than the complete protein. These smaller proteins may represent degradation products and, since they are not observed with p185<sup>HER2</sup>, could imply a difference in intracellular processing of the mutant form.

To determine whether the form lacking the trans-membrane sequence was secreted, cells were metabolically labeled with <sup>35</sup>S-methionine. The culture conditions used herein were as follows: cells were cultured in a 1:1 mixture of Dulbecco's modified Eagle's medium and Ham's nutrient mixture F-12 supplemented with glutamine (2 mM), penicillin (100 units/ml), streptomycin (100 ug/ml), and 10% serum. NIH 3T3-derived cell lines were cultured with calf serum (Hyclone). Chinese Hamster Ovary cells deficient in dihydrofolate reductase (CHO-DHFR) were



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cultured in fetal bovine serum (Gibco) supplemented with glycine (0.13 mM), hypoxanthine (0.11 mM), and thymidine (0.02 mM). (For selection of the transfected plasmid DHFR gene or to amplify introduced plasmids by methotrexate selection, the glycine, hypoxanthine, and thymidine were omitted and extensively dialyzed serum substituted for fetal bovine serum.)

Both cells and cell-conditioned medium were assayed for p185<sup>HER2</sup>. Figure 4 demonstrates that all p185<sup>HER2</sup> remained cell associated (lanes 2, 3, 4), and neither the wild type protein nor the mutant form was secreted (lanes 6, 7, 8).

Thus, contrary to expectations, deletion of the transmembrane spanning sequence was not sufficient to yield a secreted form of p185<sup>HER2</sup>.

The discovery that p185<sup>HER2ΔTM</sup> is not secreted suggested that perhaps there are sequences distal to the transmembrane spanning region that prevent passage of p185<sup>HER2</sup> through the plasma membrane. A second mutant was accordingly made that contained a UAA stop codon 8 amino acids before the beginning of the proposed transmembrane spanning sequence (Figure 1).

The second construct truncated p185<sup>HER2</sup> 8 amino acids before the start of the transmembrane spanning region at residue 645 by addition of a polypeptide chain-terminating TAA codon. The oligonucleotide 5' AAG GGC TGC CCC GCC GAG TAA TGA TCA CAG AGA GCC AGC CCT 3' was used to prime synthesis of double-stranded DNA from the same template used to construct the ΔTM mutant. Mutant plaques were

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identified by hybridization to the 5' end-labeled oligonucleotide, and confirmed by checking for the presence of a Bcl 1 site also introduced directly after the ochre codon. The chain-terminated mutant, designated HER2<sup>XCD</sup>, was then recombined into the HER2 cDNA expression plasmid. The structure of the plasmid and the 2 mutant HER2 derivatives is shown in Figure 1.

Secretion of the resulting form of p185<sup>HER2</sup>, designated p185<sup>HER2XCD</sup>, was assayed by first metabolically labeling the cells with <sup>35</sup>S-methionine, followed by immunoprecipitation of p185<sup>HER2</sup>-related proteins from both the cells and cell-conditioned media. In the immunoprecipitation procedure (Hudziak et al., Proc. Natl. Acad. Sci. USA, 84:7159-7163 (1987)), cells were harvested by trypsinization, counted electronically with a Coulter counter, and plated at least 6 hrs. before labeling. The plating medium was removed, cells washed with PBS, and the cells re-fed with methionine-free Dulbecco's modified minimal medium. [<sup>35</sup>S]-methionine (Amersham, 800 Ci/mmol, 29.6 TBq/mmol) was added at 100 uCi/6 cm plate in a volume of 3 ml. Cells were lysed at 4°C with 0.4 ml of HNEG lysis buffer per 6 cm plate. After 10 min, 0.8 ml of lysis dilution buffer (HNEG buffer with 1% bovine serum albumin, 0.1% Triton X-100 detergent) was added to each plate and the extracts were clarified by microcentrifugation for 5 min. Medium to be assayed for secretion of p185<sup>HER2</sup> related proteins was collected and clarified by microcentrifugation.



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Antibodies were added to cell extracts or conditioned medium and allowed to bind at 4°C for 2-4 h. The polyclonal antibody, G-H2CT17(0), recognizing the carboxy-terminal 17 amino acids of p185<sup>HER2</sup>, was used for characterization of cell lines expressing the transmembrane-deleted form of p185<sup>HER2</sup>. The monoclonal antibody 3E8, recognizing an epitope on the extracellular domain (Hudziak et al., Mol. Cell. Bio., 9:1165-1172 (1989)), was used at 8 ug/reaction to immunoprecipitate the truncated form. Seven ug of rabbit anti-mouse IgG was added to immunoprecipitations using this monoclonal to improve its binding to protein A-sepharose. Immune complexes were collected by absorption to protein A-sepharose beads and washed (Hudziak et al., Proc. Natl. Acad. Sci. USA, 85:5102-5106 (1988); Hudziak et al., Proc. Natl. Acad. Sci. USA, 84:7159-7163 (1987)). Proteins were separated on 7.5% sodium dodecyl sulphate-polyacrylamide gels (SDS-PAGE) and analyzed by autoradiography.

This revealed a form of p185<sup>HER2XCD</sup> of M<sub>r</sub> 88,000 kD that is associated with the cells (Figure 5, lanes 3 and 9); however, the cell-conditioned media from both the NIH 3T3 cells and Chinese hamster ovary-derived lines also contains larger amounts of a protein of M<sub>r</sub> 103,000, which is immunoprecipitated by anti-HER2 monoclonal antibody (Figure 5, lanes 4 and 10). Full length p185<sup>HER2</sup> was also expressed in both NIH 3T3 and CHO cells (Figure 5), lanes 5 and 11. There is no secretion of native p185<sup>HER2</sup> from either of these cell types (Figure 5, lanes 6 and 12).

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The larger size of the observed proteins in the cells and cell-conditioned medium (88,000 and 103,000, respectively) compared to the size predicted by the amino acid sequence (71,644) suggested that the truncated form was being glycosylated.

This was confirmed by treating the cells with the antibiotic tunicamycin, which prevents N-linked glycosylation. Treatment with tunicamycin resulted in the appearance of a cell-associated protein of  $M_r$  73,000, which is close to that predicted by the amino acid sequence (Figure 7, lane 5). It also almost completely inhibited secretion of p185<sup>HER2XCD</sup> into the medium (Figure 7, lane 6). Cell-conditioned medium from tunicamycin treated cells contains only small amounts of the mature 103,000 form, and none of the smaller forms (lane 6). This further suggests that secretion of p185<sup>HER2XCD</sup> is coupled to glycosylation.

The extent of glycosylation of the secreted form was investigated with the enzyme endoglycanase H (endo H, Boehringer Mannheim). This enzyme will hydrolyze asparagine-linked oligosaccharides of the high mannose type. High mannose oligosaccharides are biosynthetic intermediates in the glycosylation process. Final maturation of the carbohydrate side chains involves trimming off some mannose and addition of other sugars such as fucose. Such mature oligosaccharide side chains are resistant to endo H.

To determine if secreted p185<sup>HER2XCD</sup> is resistant to this enzyme, cell conditioned medium labeled with <sup>35</sup>S-methionine



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was immunoprecipitated. The immuno-precipitates were collected onto protein A sepharose beads and incubated with endo H. Neither mock incubated (lane 3) nor endo H-treated p185<sup>HER2XCD</sup> (lane 4) showed any decrease in mobility associated with hydrolysis of the glycosyl side chains, demonstrating that the glycosylation is complete.

Without being bound by any particular theory, these results taken together suggest that the cell-associated form of p185<sup>HER2XCD</sup> is an intermediate, and that fully mature glycosylated p185<sup>HER2</sup> extracellular domain is being synthesized and secreted. The lack of secretion of the p185<sup>HER2ΔTM</sup> protein could be hypothesized to result from the presence of processing information in the transmembrane spanning sequence which is necessary for Golgi transport and targeting of the plasma membrane; however, from these studies it appears instead that transport of tyrosine kinase receptor (or receptor-like) extracellular domain to the cell surface is coupled to proper glycosylation.

Therefore, insertion of the UAA stop codon 8 amino acids before the beginning of the proposed transmembrane spanning sequence yields a fully mature glycosylated p185<sup>HER2</sup> extracellular domain which is freely secreted by the cell.

Having succeeded in producing a secreted form of p185<sup>HER2</sup>, the next stage involved investigating whether the amount of secreted protein could be increased by gene amplification. Using the CHO-derived cell line, it was found that the amount of extracellular domain could be increased by methotrexate selection. The amount of

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secreted product increased 29-fold in cells selected for resistance to 500 nm methotrexate, and a further 4.4-fold by selection for resistance to 3000 nm methotrexate (Fig. 6).

5           Thus, a total increase of 128-fold in secreted p185<sup>HER2XCD</sup> was obtained when this cell line was amplified to resistance to 3000 nm methotrexate, making the production of relatively large quantities of p185<sup>HER2XCD</sup> possible.

10           To determine whether overexpression of p185<sup>HER2ATM</sup> results in cell transformation, DNA was introduced in mammalian cells by the CaHPO<sub>4</sub> coprecipitation method (Graham et al., Virology, 52:456-467 (1973)). Five ug of plasmid DNA was added to half-confluent plates of cells (6.0 cm) in 1 ml for 4-6 h. The DNA was removed and the cells shocked 15 with 20% (vol/vol) glycerol. After 2 days for phenotypic expression the selective agent geneticin was added at 400 ug/ml. Clones were picked using glass cloning cylinders with petroleum jelly for the bottom seal. The introduced plasmids were amplified by the methotrexate selection 20 procedure (Kaufman et al., J. Mol. Biol., 159:601-621 (1982)).

25           When the ATM mutant was expressed in NIH 3T3 cells, primary unamplified colonies after selection had the normal flat nontransformed phenotype (Figure 8, compare photo B with vector control alone, photo A). After the expression level was increased by methotrexate selection, the cells took on the refractile, spindle-shaped appearance of transformed cells and also grew piled up in irregular



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clumps (photo E). This observation is similar to our earlier findings with the unaltered HER2 cDNA (photos C and F, parent and amplified derivatives respectively), and suggests that high levels of expression of the mutant  $\Delta$ TM protein were also transforming.

The morphological changes seen at equivalent levels of amplification (400 nm methotrexate) are not as marked for the mutant, implying that the transforming potential of this form of p185<sup>HER2</sup> may be less. At higher levels of resistance to methotrexate, the  $\Delta$ TM cells become even more transformed in appearance.

The plasmid was also negative in a focus-forming assay whereas the wild type HER2 plasmid was positive, further indicating that the transforming potential of p185<sup>HER2 $\Delta$ TM</sup> protein is lower. Cells expressing high levels also displayed another property of the transformed phenotype, growth in soft agar. Colony formation in soft agar was determined by harvesting each line to be assayed with trypsin, counting the cells (Coulter counter), and plating 80,000 cells per 6-cm dish. The top layer consisted of 4 ml of 0.25% agar (Difco, "purified") over a bottom layer of 5 ml of 0.5% agar. Colonies were counted after 3-4 weeks. Cells from 2 independent clones plated in soft agar gave rise to soft agar colonies with an efficiency comparable to cells expressing the wild type HER2 gene:

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Table I  
Soft Agar Colony Formation

<u>Cell Line</u>	<u># of Soft Agar Colonies</u>
CVN	0
5 CVN <sub>400</sub>	0
HER2-3 <sub>0</sub>	5 +/- 1
HER2-3 <sub>400</sub>	208 +/- 27
ΔTM-A1 <sub>0</sub>	0
ΔTM-A1 <sub>400</sub>	205 +/- 62
10 ΔTM-B2 <sub>0</sub>	0
ΔTM-B2 <sub>400</sub>	205 +/- 13

Two control lines were used; NIH 3T3 cells transfected with a plasmid expressing only the neo and DHFR genes, and the same line amplified to resistance to 400 nM methotrexate. The number of soft agar colonies arising was determined for both parental and amplified lines of clones expressing either p185<sup>HER2</sup> or p185<sup>HER2ΔTM</sup> proteins. Each cell line was plated in triplicate and the value averaged.

Therefore, according to the present invention it has been determined that removal of only the transmembrane spanning sequence does not lead to secretion of p185<sup>HER2</sup>, unless the entire tyrosine kinase domain is also deleted. Removal of this domain results in proper glycosylation and secretion of the extracellular domain.

In order to obtain purified HER2 extracellular domain working material, Chinese Hamster Ovary Cell Harvest Fluid (CFF) containing recombinant HER2 ECD may be first concentrated by ultrafiltration, and then purified by immunoaffinity chromatography using a HER2 specific MAb



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coupled to CNBr activated Sepharose; other suitable immobilization supports may be used. Concentrated CCF is applied to the affinity column after filtration through a 0.2 micron Millipor\* filter. Purification cycles are performed as necessary until the desired amount of CCF is processed.

5

During each cycle of purification, the concentrated CCF is applied and the affinity column is washed to baseline with 0.5 M Tris buffer containing 0.15 M NaCl at a pH of approximately 7.5 (TB). HER2 extracellular domain is then eluted from the column with 0.1 M sodium citrate buffer containing 0.5 M NaCl at a pH of approximately 3.5. The affinity column eluant fractions containing HER2 ECD are pooled and neutralized. The immunoaffinity column is reequilibrated between each purification cycle with TB.

In a second step, the affinity column eluant is buffer exchanged into 25 ml of Tris buffer, at a pH of approximately 7.0 (TB2). The HER2 extracellular domain is then applied to a DEAE Sepharose Fast Flow\* column, and washed with TB2. The HER2 ECD is removed from the column by step or gradient salt elution in TB2 (containing up to 200 mM NaCl).

After DEAE chromatography, purified HER2 ECD fractions are pooled, exchanged into phosphate-buffered saline, and stored at 2-8° C. The resulting material is substantially pure, i.e., about 99% pure (see Fig. 12).

By means of the present invention it is accordingly possible to produce a secreted, glycosylated p185<sup>HER2</sup>

\*Trade-mark

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extracellular domain. This opens several possibilities for further research, as well as a broad range of potential therapeutic applications.

As previously stated, the HER2 gene is of particular interest because its amplification has been correlated with certain types of cancer. In a survey of 189 primary mammary gland adenocarcinomas, it was found that 30% contained amplifications of the HER2 gene. Slamon et al., "Human Breast Cancer: Correlation of Relapse and Survival with Amplification of the HER-2/neu Oncogene," Science 235, 177-182 (1987). Amplification was correlated with a negative prognosis and high probability of relapse.

This suggests that of the 120,000 women diagnosed with breast cancer each year, 36,000 will have HER2 amplification. Approximately half of these women, or about 15,000, may be expected to exhibit greater than 5-fold amplification, corresponding to nearly half of the 40,000 breast cancer-related deaths each year.

It has been demonstrated that a monoclonal antibody directed against the p185<sup>HER2</sup> extracellular domain specifically inhibits growth of breast tumor-derived cell lines overexpressing the HER2 gene product; prevents HER2-transformed NIH 3T3 cells from forming colonies in soft agar; and reduces the resistance to the cytotoxic effect of tumor necrosis factor alpha which accompanies HER2 overexpression. Hudziak et al., "p185<sup>HER2</sup> Monoclonal Antibody has Antiproliferative Effects In Vitro and Sensitizes Human Breast Tumor Cells to Tumor Necrosis



Factor", Mol. Cell. Biol. 9:1165-1172 (1989). See also,  
Drebin et al., "Inhibition of Tumor Growth by a Monoclonal  
Antibody Reactive with an Oncogene-Encoded Tumor Antigen",  
Proc. Natl. Acad. Sci. USA 83, 9129-9133 (1986) (in vivo  
5 treatment with anti-p185 monoclonal antibody asserted to  
inhibit tumorigenic growth of neu-transformed NIH 3T3 cells  
implanted in mice).

This effect presents the possibility that conditions  
characterized by amplification of the HER2 gene may be  
10 subject to treatment via Active Specific Immunotherapy.  
This therapeutic modality contemplates provoking an immune  
response in a patient by vaccination with an immunogenic  
form of the extracellular domain. The extracellular domain  
(or a derivative thereof, as discussed below) may be  
15 combined with a local adjuvant which is safe and effective  
in humans, such as alum, Bacillus calmette-Guerin (BCG),  
adjuvants derived from BCG cell walls, Detox\* (Ribi-  
immunochem), Syntex-1\*, or Corynebacterium partum.

Alternatively, systemic adjuvants, such as Interferon  
20 gamma, Interleukin 1, Interleukin 2, or Interleukin 6 may  
be suitable. An appropriate dose and schedule would be  
selected to maximize humoral and cell-mediated response.

It may also be possible to enhance an immune response  
by targeting the immunogen to the immune system, which  
25 could lead to more efficient capture of the antigen by  
antigen presenting cells, or by directing the immunogen so  
that it is presented by MHC Class 1 molecules, since these  
usually induce a T-cell response.

\*Trade-mark.

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In addition to Active Specific Immunotherapy, it should be possible to use the purified extracellular domain to isolate and characterize the putative ligand. The HER2 ligand may be used in turn to deliver toxin to tumor cells which are overexpressing HER2, such as by molecular fusion of the ligand with toxin, or by chemical cross-linking. Alternatively, patients overexpressing HER2 may be vaccinated with HER2 ligand conjugated to, or in combination with, a suitable adjuvant.

10 A patient overexpressing HER2 will also presumably be overexpressing the HER2 ligand. The ligand-HER2 binding interaction, which is likely to contribute to tumor growth, may be inhibited by blocking free ligand in the patient's serum. This blocking can be accomplished by treating the patient with the HER2 extracellular domain, which will proceed to bind free HER2 ligand, thereby preventing the ligand from binding to the HER2 receptor site.

15 Rather than using the HER2 extracellular domain per se, it may be more desirable to use a derivative which has an increased affinity for the ligand, and/or which has an increased half-life in vivo. Cross-linking on cells is known to improve binding affinity, suggesting that artificial cross-linking can be used to improve the binding ability of the HER2 extracellular domain. The half-life of the extracellular domain in serum can be improved by, for example, fusing the extracellular domain with other molecules present in the serum which are known to have a



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long half-life, such as the Fc-portion of an immunoglobulin molecule.

The present invention has of necessity been discussed herein by reference to certain specific methods and materials. It is to be understood that the discussion of these specific methods and materials in no way constitutes any limitation on the scope of the present invention, which extends to any and all alternative materials and methods suitable for accomplishing the ends of the present invention.

THE EMBODIMENTS OF THE INVENTION IN WHICH AN EXCLUSIVE PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. A composition comprising an extracellular portion of the HER2 molecule  
5 comprising at least 9 amino acids and/or an immune epitope, essentially free of transmembrane and intracellular portions of said HER2 molecule, and in substantially pure form, for use in Active Specific Immunotherapy.
2. A composition according to claim 1, wherein the extracellular portion of the HER2  
10 molecule has a purity of at least about 99%.
3. A composition according to claim 1 or claim 2, comprising the entire extracellular portion of the HER2 molecule.
- 15 4. A composition according to any one of claims 1 to 3, wherein the extracellular portion of the HER2 molecule is conjugated with a peptide having immunogenic properties.
5. A composition according to claim 4, wherein said peptide comprises an immune  
20 epitope.
6. A composition according to any one of claims 1 to 5 further comprising an adjuvant.
- 25 7. A composition according to claim 6 wherein the adjuvant comprises any of alum, Bacillus calmette-Guerin (BCG), a BCG cell wall derivative, Detox, *Corynebacterium parvum*, interferon gamma, interleukin 1, interleukin 2, Syntex-1 and interleukin 6.



8. Use of an extracellular portion of the HER2 molecule comprising at least 9 amino acids and/or an immune epitope, essentially free of transmembrane and intraocular portions of said HER2 molecule, in the manufacture of a composition for treatment of a patient by Active Specific Immunotherapy.
- 5
9. Use according to claim 8, wherein the composition comprises the entire extracellular portion of the HER2 molecule.
10. Use according to claim 8 or claim 9 wherein the extracellular portion of the HER2 molecule is conjugated with a peptide having immunogenic properties.
- 10
11. Use according to claim 10 wherein said peptide comprises an immune epitope.
12. Use according to any one of claims 8 to 11, wherein the composition comprises an adjuvant.
- 15
13. Use according to claim 12, wherein the adjuvant comprises any of alum, Bacillus calmette-Guerin (BCG), a BCG cell wall derivative, Detox, *Corynebacterium parvum*, interferon gamma, interleukin 1, interleukin 2, Syntex-1 and interleukin 6.
- 20
14. Use of an effective amount of an extracellular portion of the human HER2 receptor to treat a patient via Active Specific Immunotherapy to provoke a cell-mediated immune response to the HER2 receptor in the said patient.
- 25
15. Use according to claim 14 wherein the patient has, or is at risk of acquiring, cancer.
16. Use according to claim 15 wherein cells from a tissue of a patient have increased numbers of the HER2 receptor residing on the surfaces thereof, relative to normal cells of the same tissue type.
- 30

17. Use according to claim 16 wherein the tissue is a mammary gland tissue.
18. Use according to claim 17 wherein the patient is a woman.
- 5 19. Use according to claim 16 wherein the tissue is salivary gland tissue.
20. Use according to claim 16 wherein the tissue is gastric tissue.
21. Use according to claim 16 the tissue is colon tissue.
- 10 22. Use according to claim 14 which further comprises the use of an adjuvant.
23. Use according to claim 22 wherein the adjuvant is selected from the group consisting of alum, Bacillus calmette-Guerin (BCG), a BCG cell wall derivative, Detox, *Corynebacterium parvum*, interferon gamma, interleukin 1, interleukin 2 and interleukin 6.
- 15 24. Use according to claim 14 which elicits both a humoral and cell-mediated response in the patient.
- 20 25. Use according to claim 14 wherein the extracellular portion is fused to a heterologous molecule.
26. Use according to claim 25 wherein the heterologous molecule comprises the Fc-portion of an immunoglobulin molecule.
- 25 27. Use of an effective amount of an extracellular portion of the human HER2 receptor to provoke a humoral and cell-mediated immune response to the HER2 receptor in a patient.

30



28. Use according to claim 27 wherein the patient has, or is at risk of acquiring, cancer.
29. Use according to claim 27 wherein cells from a tissue of the patient have increased numbers of the HER2 receptor residing on the surfaces thereof, relative to normal cells of the same tissue.
- 5
30. Use according to claim 29 wherein the tissue is mammary gland tissue.
31. Use according to claim 29 wherein the tissue is salivary gland tissue.
- 10
32. Use according to claim 29 wherein the tissue is gastric tissue.
33. Use according to claim 29 wherein the tissue is colon tissue.
- 15
34. Use according to claim 27 which further comprises the use of an adjuvant.
35. Use according to claim 27 wherein the extracellular portion is fused to a heterologous molecule.
- 20
36. Use according to claim 27 which induces a T-cell response to the HER2 receptor.
37. Use according to claim 14 which induces a T-cell response to the HER2 receptor.
38. Use of an effective amount of an isolated extracellular portion of the human HER2 receptor to treat a human patient by Active Specific Immunotherapy to provoke a cell-mediated immune response to the HER2 receptor in the said patient.
- 25
39. Use of an effective amount of an isolated extracellular portion of the human HER2 receptor to provoke a humoral and cell-mediated immune response to the HER2 receptor in a human patient.
- 30

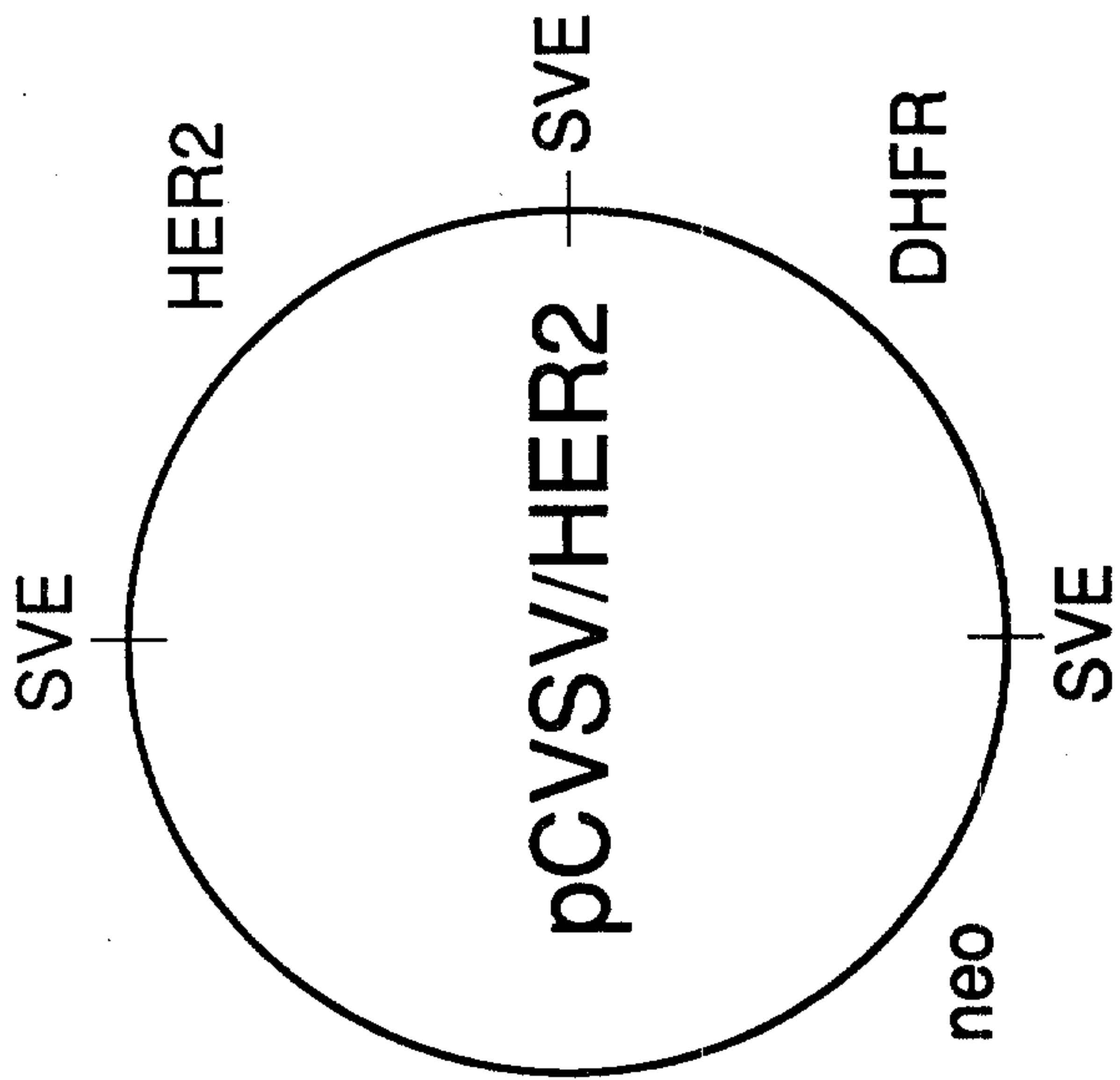
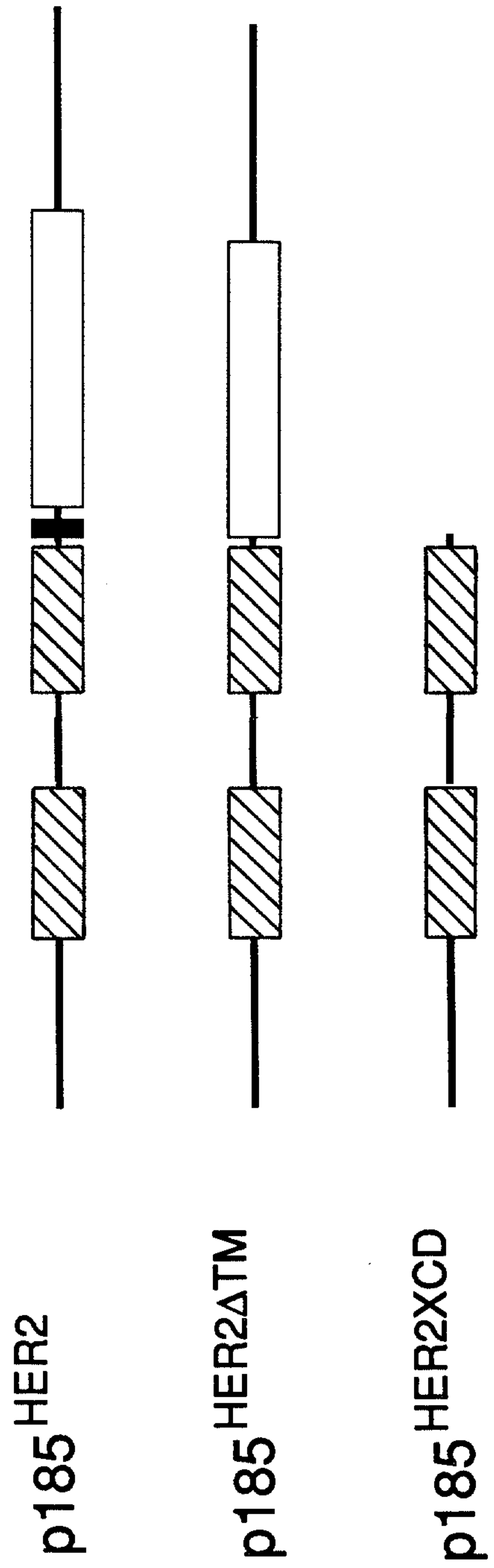
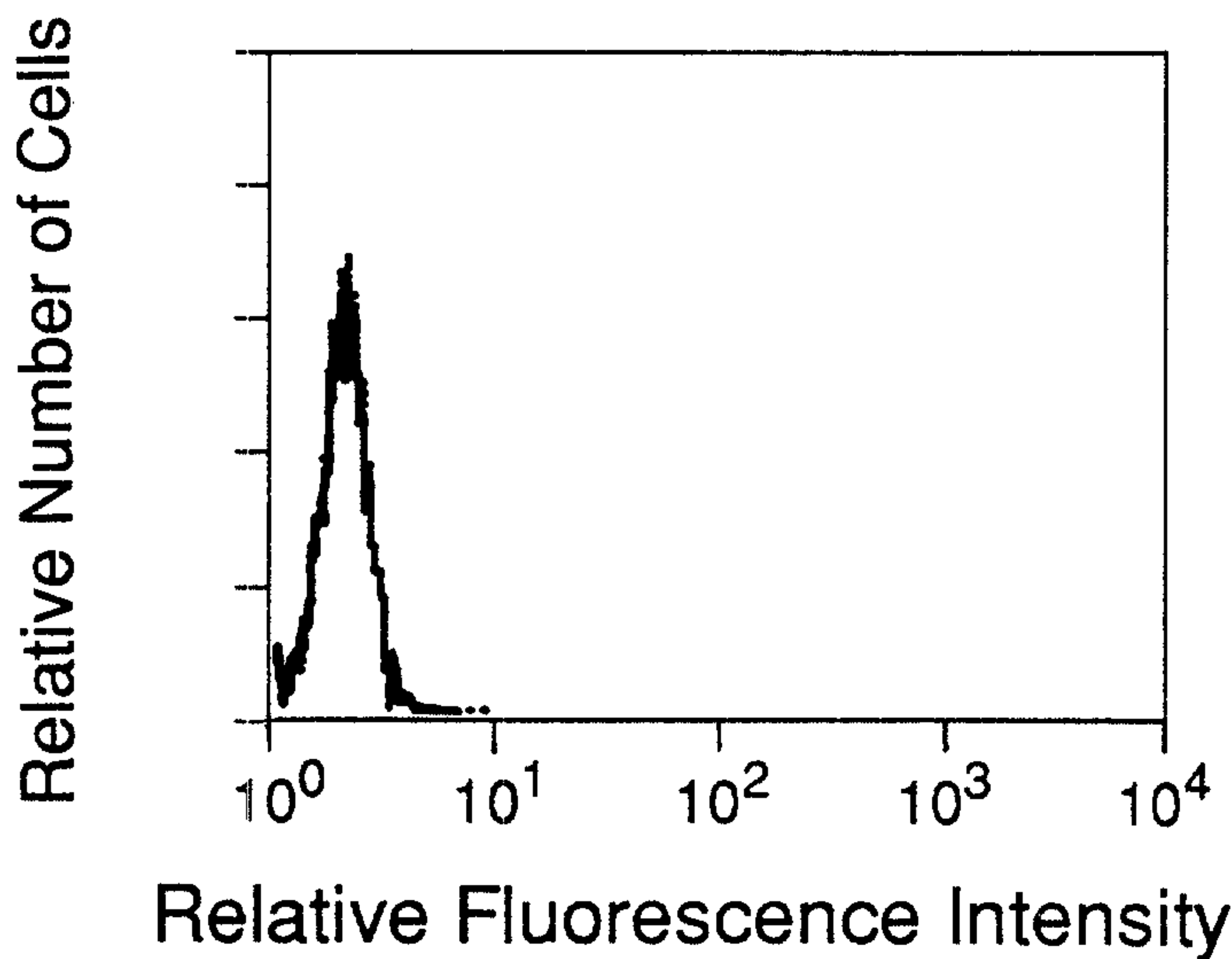


FIG. 1A

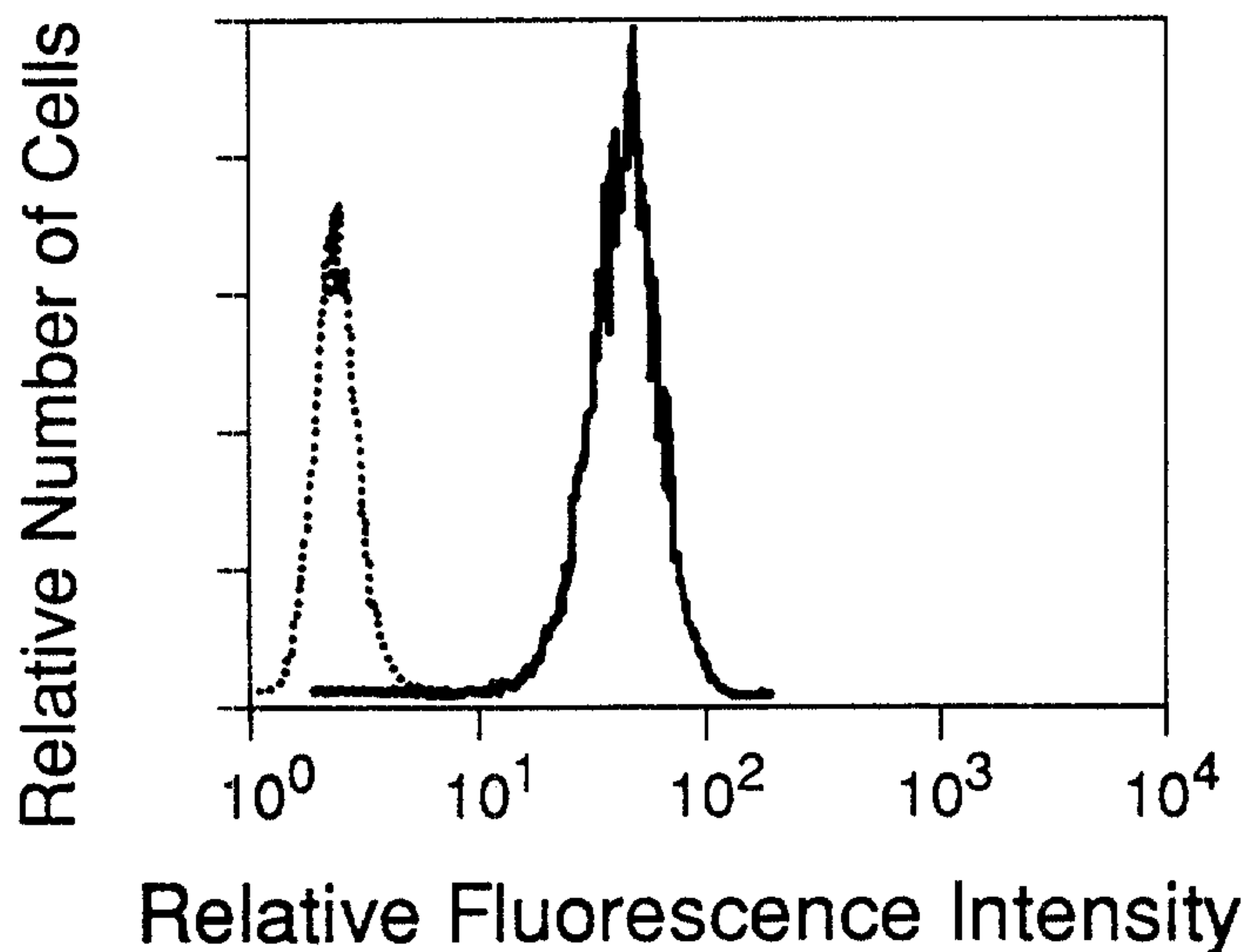
FIG. 1B



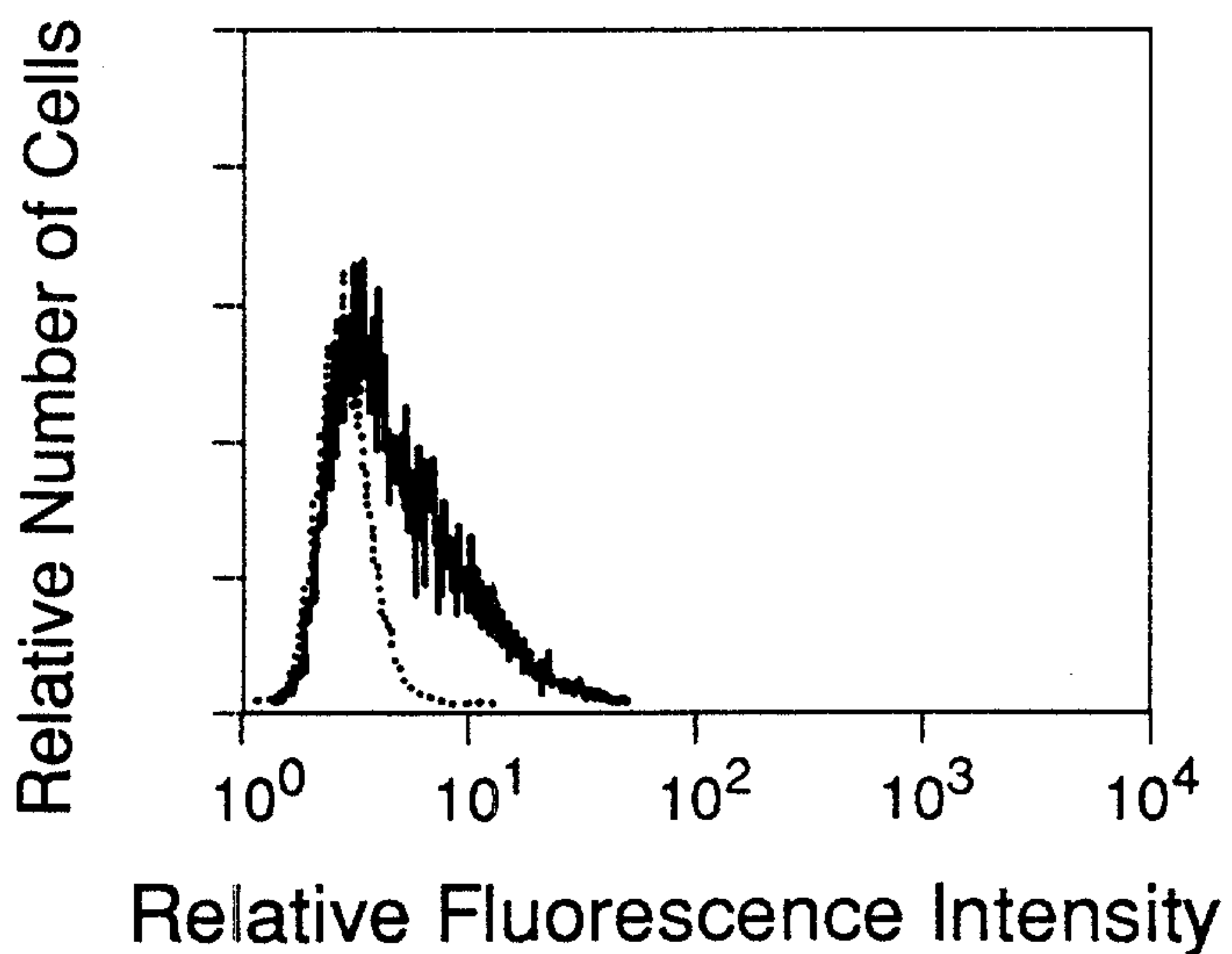




**FIG. 10A**



**FIG. 10B**



**FIG. 10C**

**A1**

POTENTIAL T-CELL EPITOPES IN THE HER2 EXTRACELLULAR DOMAIN

1 SER THR GLN VAL CYS THR GLY THR ASP MET LYS LEU ARG LEU PRO ALA SER PRO GLU THR 20  
 AGC ACC CAA GUG UGC ACC GGC ACA GAC AUG AAG CUG CGG CUC CCU GCC AGU CCC GAG ACC  
 30 LEU ASP MET LEU ARG HIS LEU TYR GLN GLY CYS 40  
 CAC CUG GAC AUG CUC CGC CAC CUC UAC CAG GGC UGC GLN VAL VAL GLN GLY ASN LEU GLU  
 LEU THR TYR LEU PRO THR ASN ALA SER LEU SER PHE LEU GLN ASP ILE GLN GLU VAL GLN 60  
 CUC ACC UAC CUG CCC ACC AAU GCC AGC CUG UCC UUC CUG CAG GAU AUC CAG GAG GUG CAG  
 70 GLY TYR VAL LEU ILE ALA HIS ASN GLN VAL ARG GLN VAL PRO LEU GLN ARG LEU ARG ILE 80  
 GGC UAC GUG CUC AUC GCU CAC AAC CAA GUG AGG CAG GUC CCA CUG CAG AGG CUG CGG AUU  
 90 VAL ARG GLY THR GLN LEU PHE PHE LEU PHE ALA LEU ALA VAL LEU ASP ASN GLY ASP 100  
 GUG CGA GGC ACC CAG CUC UUU GAG GAC AAC UAU GCC CUG CUG GUC CUA GAC AAU GGA GAC  
 110 PRO LEU ASN ASN THR THR PRO VAL THR GLY ALA SER PRO GLY GLY LEU ARG GLU LEU GLN 120  
 CCG CUG AAC AAU ACC ACC CCU GUC ACA GGG GCC UCC CCA GGA GGC CUG CGG GAG CUG CAG  
 130 LEU ARG SER LEU THR GLU ILE LEU LYS GLY GLY VAL LEU ILE GLN ARG ASN PRO GLN LEU 140  
 CUU CGA AGC CUC ACA GAG AUC UUG AAA GGA GGG GUC UUG AUC CAG CGG AAC CCC CAG CUC  
 150 CYS TYR GLN ASP THR ILE LEU TRP LYS ASP ILE PHE HIS LYS ASN ASN GLN LEU ALA LEU  
 UGC UAC CAG GAC ACG AUU UUG UGG AAG GAC AUC UUC CAC AAG AAC AAC CAG CUG GCU CUC 160

FIG. 13A

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170  
 THR LEU ILE ASP THR ASN ARG SER ARG SER ARG ALA CYS HIS PRO CYS SER PRO MET CYS LYS GLY  
 ACA CUG AUA GAC ACC AAC CGC UCU CGG GCC UGC CAC CCC UGU UCU CCG AUG UGU AAG GGC 180

190  
 SER ARG CYS TRP GLY GLU GLY SER SER GLU ASP CYS GLN SER LEU THR ARG THR VAL CYS ALA  
 UCC CGC UGC UGC GGA GAG GAG UCU GAG GAU UGU CAG AGC AGC CUG ACU GUC UGU GUC UGU GCC 200

210  
 GLY GLY CYS ALA ARG CYS LYS GLY PRO LEU PRO THR ASP CYS CYS HIS GLU GLN CYS ALA  
 GGU GGC UGU GCC CGC UGC UGC AAG GGG CCA CUG CCC ACU GAC UGC UGC CAU GAG CAG UGU GCU 220

230  
 ALA GLY CYS THR GLY PRO LYS HIS SER ASP CYS LEU ALA CYS LEU HIS PHE ASN HIS SER  
 GCC GGC UGC ACG GGC CAC CCC AAG CAC UCU GAC UGC CUG GCC UGC CUC CAC UUC AAC CAC AGU 240

250  
 GLY ILE CYS GLU LEU LEU LEU LEU VAL THR TYR ASN THR ASP THR PHE GLU SER  
 GGC AUC UGU GAG CUG CAC HIS CYS PRO ALA LEU LEU VAL THR ACC ACC UAC ACA GAC ACU GAG UCC 260

270  
 MET PRO ASN PRO GLU TYR THR PHE PHE GLY ALA SER CYS VAL THR ALA CYS PRO TYR  
 AUG CCC AAU CCC GAG GGC CGG UAU ACA UUC GGC GCC AGC UGU GUG ACU GCC UGU CCC UAC 280

290  
 ASN TYR LEU SER THR ASP VAL GLY SER CYS THR LEU VAL CYS PRO LEU HIS ASN GLN GLU  
 AAC UAC CUU UCU ACG GAC GUG GGA UCC UGC ACC CUC GUC UGC CCC CUG CAC AAC CAA GAG 300

310  
 VAL THR ALA GLU ASP GLY THR GLN ARG CYS GLU LYS CYS SER LYS PRO CYS ALA ARG VAL  
 GUG ACA GCA GAG GAU GGA ACA CAG CGG UGU GAG AAG UGC AGC AAG CCC UGU GCC CGA GUG 320

330  
 CYS TYR GLY LEU GLY MET GLU HIS LEU ARG GLU VAL ARG ALA VAL THR SER ALA ASN ILE  
 UGC UAU GGU CUG GGC AUG GAG CAC UUG CGA GAG GUG AGG GCA GUU ACC AGU GCC AAU AUC 340

FIG. 13B





540	GLU	GLU	CYS	ARG	VAL	LEU	GLN	GLY	LEU	PRO	ARG	GLU	TYR	VAL	ASN	ALA	ARG	HIS	CYS	LEU
	GAG	GAA	UGC	CGA	GUA	CUG	CAG	GGG	CUC	CCC	AGG	GAG	UAU	GUG	AAU	GCC	AGG	CAC	UGU	UUG
550	PRO	CYS	HIS	PRO	GLU	CYS	GLN	PRO	GLN	ASN	GLY	SER	VAL	THR	CYS	PHE	GLY	PRO	GLU	ALA
	CCG	UGC	CAC	CCU	GAG	UGU	CAG	CCC	CAG	AAU	GGC	UCA	GUG	ACC	UGU	UUU	GGA	CCG	GAG	GCU
570	ASP	GLN	CYS	VAL	ALA	CYS	ALA	HIS	TYR	LYS	ASP	PRO	PRO	PHE	CYS	VAL	ALA	ARG	CYS	PRO
	GAC	CAG	UGU	GUG	GCC	UGU	GCC	CAC	UAU	AAG	GAC	CCU	CCC	UUC	UGC	GUG	GCC	CGC	UGC	CCC
590	SER	GLY	VAL	LYS	PRO	ASP	LEU	SER	TYR	MET	PRO	ILE	TRP	LYS	PHE	PRO	ASP	GLU	GLU	GLY
	AGC	GGU	GUG	AAA	CCU	GAC	CUC	UCC	UAC	AUG	CCC	AUC	UGG	AAG	UUU	CCA	GAU	GAG	GAG	GGC
610	ALA	CYS	GLN	PRO	CYS	PRO	ILE	ASN	CYS	THR	HIS	SER	CYS	VAL	ASP	LEU	ASP	ASP	LYS	GLY
	GCA	UGC	CAG	CCU	UGC	CCC	AUC	AAC	UGC	ACC	CAC	UCC	UGU	GUG	GAC	CUG	GAU	GAC	AAG	GGC
624	CYS	PRO	ALA	GLU																
	UCC	CCC	GCC	GAG																

FIG. 13D

UNSCANNABLE ITEM

RECEIVED WITH THIS APPLICATION

(ITEM ON THE 10TH FLOOR ZONE 5 IN THE FILE PREPARATION SECTION)

# 2055441

DOCUMENT REÇU AVEC CETTE DEMANDE

NE POUVANT ÊTRE BALAYÉ

(DOCUMENT AU 10 IÈME ÉTAGE AIRE 5 DANS LA SECTION DE LA  
PRÉPARATION DES DOSSIERS)